

Review

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Eosinophil granulocytes in chronic inflammatory respiratory diseases and CRSwNP: Function, immunological basis, and clinical significance

Felix Klimek¹, Christoph Bergmann², Jan Hagemann³, Mandy Cuevas⁴, Sven Becker⁵, Oliver Pfaar⁶, Ingrid Casper¹, and Ludger Klimek¹

¹Center for Rhinology and Allergology, Wiesbaden, ²Practice for Ear, Nose and Throat disease, Clinic RKM 740, Düsseldorf, ³Department for Otolaryngology, University Medical Center Mainz, Mainz, ⁴Department for Otolaryngology, University Hospital Carl Gustav Carus, TU Dresden, Dresden, ⁵Department for Otolaryngology, University Hospital Tübingen, Tübingen, and ⁶Section of Rhinology and Allergy, Department of Otorhinolaryngology, Head and Neck Surgery, University Hospital Marburg, Philipps-Universität Marburg, Marburg, Germany

Key words

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Abstract. Introduction: Eosinophils play an important regulatory and immunomodulatory role in airway mucosa and have antiparasitic and antiviral properties as well as pro-inflammatory effects that may also cause persistence of inflammation with tissue remodeling. The number of eosinophils and the detection of specific mediators in biological samples from, e.g., blood, nasal secretions, and bronchial fluid can serve as biomarkers that reflect the underlying pathophysiology of certain diseases, predict treatment success, and detect therapy effects. Materials and methods: A literature search was conducted to determine the immunologic basis, mode of action, clinical significance, and available evidence for therapeutic approaches using eosinophil-targeted monoclonal antibodies by searching Medline, Pubmed, and the national and international trial database (ClinicalTrials.gov) and guideline registries as well as the Cochrane Library. Human studies published on the topic in the period up to and including 10/2023 were considered. Results: Based on the international literature and previous experience, the results are summarized, and

recommendations are given. Conclusion: The important role of eosinophils in immunological processes in the airway mucosa is comprehensively analyzed and can serve as a basis for current and future treatment approaches.

Introduction

Eosinophils develop both beneficial and harmful activities as part of immune responses, they are involved in the maintenance of health and homeostasis, but also in the development of diseases. Their role in the pathophysiology of various allergic and non-allergic diseases has been known for decades [1, 2]. This has led to the development of a wide range of therapies focusing on eosinophils, either acting non-specifically by inhibiting various upstream or downstream immune pathways or specifically by eosinophil-targeted biologics [3, 4].

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Correspondence to: Professor Dr. med. L. Klimek, Center for Rhinology and Allergology, An den Quellen 10, 65183 Wiesbaden, Germany, Ludger.Klimek@Allergiezentrum.org

Recently, several previously unknown physiological functions of eosinophils have been identified, consisting of important regulatory and immunomodulatory, anti-inflammatory, antiparasitic, and antiviral properties [5, 6]. On the other hand, the involvement of pro-inflammatory eosinophils in the initiation, progression, and persistence of inflammation with tissue remodeling is well known and has been documented for many decades – especially in the upper and lower respiratory tract. Therefore, the eosinophil count in biological samples from different body compartments such as blood, nasal secretions, or bronchial fluid can serve as a biomarker that reflects the underlying pathophysiology of certain diseases, can roughly predict treatment success, and detect therapy effects [3, 4, 7]. The precise definition of eosinophilia and the distinction between an actual pathological condition and hypereosinophilia as an epiphenomenon are crucial for the correct interpretation and use of eosinophils as biomarkers in clinical practice.

Origin and life cycle

Eosinophils are part of the innate immune system and belong to the white blood cell family [8]. These cells were first described by Paul Ehrlich in the 19th century [9], who gave his name to the German regulatory authority in vaccines, the Paul Ehrlich Institut in Langen. Eosinophils have a characteristic bilobed nucleus and large granules that can be intensely stained with the dye eosin, which gave the cells their name. The granules contain several enzymes and cationic proteins, including peroxidases, lysosomal enzymes, and the major basic protein (MBP). Eosinophils originate from the bone marrow, where they are formed from a myeloid precursor that they share with basophils [10]. At the myelocyte stage, the precursor cells stop dividing and enter a maturation phase of ~ 4 days, during which the cells mature into functional granulocytes [11].

This process is regulated by cytokine receptors (e.g., CD131-containing receptors: CD116/CD131, CD123/CD131, and CD125/CD131, which bind to GM-CSF, IL-3, and IL-5, respectively [12]), alarmin receptors (e.g.,

ST2, which binds to IL-33) [13], and specific transcription factors (e.g., GATA1/2 and C/EBPs) [14]. Subsequently, mature eosinophils are released from the bone marrow and can be detected in small numbers in the peripheral blood (~ 50 – 150 cells/mL blood; 1 – 3% of total leukocytes) in homeostasis [15]. The possibility of *in situ* eosinophilopoiesis has also been described [16]. In homeostasis, the half-life of eosinophils in peripheral blood is unknown but is estimated to be 11 – 63 hours [17, 18, 19].

In a healthy state, eosinophils can be detected in various tissues such as the intestine and fatty tissue, where they fulfill various homeostatic functions. An increased number of pre-activated eosinophils can be found in the peripheral blood and in inflamed target tissues in various diseases, especially but by no means only in allergic diseases [20]. In addition to the classic allergic respiratory diseases asthma and rhinitis, which are associated with eosinophil infiltration of the target organs, there is a broad spectrum of non-allergic diseases (e.g., non-allergic eosinophilic asthma and eosinophilic bronchitis, eosinophilic granulomatosis with polyangiitis, chronic rhinosinusitis with nasal polyps (CRSwNP), non-allergic eosinophilic rhinitis, eosinophilic esophagitis (EoE), anaphylaxis, etc.) [21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33]. They can be accompanied by high eosinophil counts, which can be detectable both in the blood and in the tissue.

IL-5 is a central cytokine for the life cycle of eosinophils as it is firstly a growth factor for eosinophil progenitor cells, secondly is involved in the mobilization of eosinophils from the bone marrow, and thirdly plays an important role in their activation and their colonization in target tissues [34]. However, the presence of IL-5 is not solely crucial for the development of eosinophils, as IL-5 knockout mice still exhibit activated eosinophils [35].

A study with a monoclonal antibody against IL-5 (mepolizumab) showed a significant decrease in eosinophils in the peripheral blood and inflamed tissue in patients with EoE but had no effect on the eosinophil count in the duodenum [36]. Similarly, treatment with mepolizumab significantly reduced the number of eosinophils in peripheral blood and sputum [37], but did not lead to a significant reduction in eosinophils in airway tissue and eosinophil activation

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markers such as MBP [38]. This may be due to the fact that a certain subset of airway resident eosinophils do not respond to IL-5 [39]. There is a growing consensus that IL-5 plays an important role in reactive eosinophilia, whereas it appears to be less important for homeostatic eosinophils in tissues. A detailed and comprehensive overview of all migration and activation factors of eosinophils as well as their mediators and receptors can be found in a recent review [40].

Functions

Traditionally, eosinophils have been described as important cells of the innate immune defense against multicellular parasites, especially helminths. This is largely based on observations of eosinophilia in the context of parasitic diseases and the killing of parasites by eosinophils and their toxic granules *in vitro* [41].

However, profound functional mechanisms in which eosinophils are involved are only partially understood yet and may also differ from species to species, as eosinophils only make a variable contribution to parasite killing in mouse models, for example [42]. One of the most important tasks of eosinophils is to maintain tissue homeostasis in various organs such as the lungs, nose and paranasal sinuses, gastrointestinal tract, thymus, and adipose tissue. They support the normal function of the immune system (immune tolerance), promote fertility and prevent obesity and bronchial hyperreactivity [43, 44, 45]. This is mainly due to the fact that these eosinophils are found in numerous healthy human tissues (see overview) [6] and in mouse models of metabolism [46], endometriosis [47], and other tissue functions [48]. The mechanisms underlying these functions are discussed in more detail below.

Mechanisms of eosinophil activation and function

Eosinophils express a variety of receptors on their surface, which have been described in detail in a recent review [8]. Several receptors are important for therapeutic

targeting of specific eosinophil-related diseases. Eosinophils express three receptors with a common β -chain (GM-CSF, IL-3 and IL-5 receptor), all of which are involved in the control of the eosinophil life cycle, including survival. In addition, the eosinophil-specific Siglec-8 is also involved in their survival. Several eosinophil receptors are involved in adhesion to the endothelium (for example, L-selectin, Mac-1/CD11b/CD18, VLA-4/CD49d) and chemotaxis (for example, CCR3/CD193, C5aR/CD88, platelet activation receptor) [49]. In addition, several surface receptors are associated with eosinophil activation (for example, Fc γ RII/CD32A, Fc α R/CD89, CR3 - CD11b, glucan receptors) [40, 50, 51, 52, 53].

Eosinophils are highly cytotoxic due to their intracellular mediators, which can become a potential threat to the host tissue if they are not adequately controlled. Pre-activation or priming is a crucial mechanism in this process. Under homeostatic conditions, eosinophils tend to be refractory cells that can only respond to high concentrations of activators, such as opsonized particles. However, when eosinophils are exposed to specific cytokines or other immune mediators *in vitro*, even at low concentrations, they can rapidly (within 1 – 15 minutes, depending on the priming agent) transition to a pre-activated phenotype that makes them highly susceptible to these targets [20, 54]. Priming can also occur in patients with eosinophil-mediated diseases [55].

Eosinophils have a whole arsenal of cytotoxic effector mechanisms that are mainly exerted extracellularly, i.e., within the synapse between the cell and its major (i.e., parasitic) targets. These functions include the abundant production of reactive oxygen species by a membrane-bound NADPH oxidase (NOX-2) [56], the degranulation of highly cytotoxic granular proteins (for example, MBP, eosinophil peroxidase, eosinophil cationic protein (ECP), or eosinophil-derived neurotoxin (EDN)) [57] or peptides (e.g., polycationic peptides) into the synapse and the killing of extracellular targets by the formation of eosinophilic extracellular traps (EET) [58]. In addition, eosinophils produce a variety of cytokines (e.g. IL-4, IL-5, and GM-CSF) and bioactive lipid mediators (e.g., leukotriene C4 and platelet activating factor), which are released upon activation [59, 60, 61].

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Eosinophils are relatively inert or refractory in the bloodstream and in tissues. When activated by receptors, they release their cytotoxic components and cause tissue damage. Eosinophils harbor a unique secretory organelle, the so-called crystalloid granule, which contains MBP in high concentrations and leads to the formation of a crystalline core. The content of the crystalloid granules can only be released by eosinophils through degranulation. There are several types of degranulation in eosinophils, most of which fall into the category of classical exocytosis with SNARE-mediated membrane fusion (including compound exocytosis and piecemeal degranulation, the latter occurring mainly in allergic inflammation) [60, 62, 63, 64, 65, 66]. In addition, free eosinophil granules can be released as intact, membrane-bound organelles by a form of necrotic release also known as cytolysis [67]. Eosinophils also release DNA into the extracellular space during EET formation. The molecular mechanism of this process is not yet fully understood [58, 68]. EET formation occurs independently of degranulation, although granule proteins have been detected on DNA strands [69]. The association of granule products with DNA has been suggested to occur both before [58, 70] and after their release [71]. EETs have been shown to increase the viscosity of nasal secretions in patients with chronic rhinosinusitis [72]. In addition, EETs have also been associated with Charcot-Leyden crystals, which in turn have been associated with eosinophilia in the past [73]. Furthermore, eosinophil-derived Charcot-Leyden crystals in the mucus of asthma patients play a role in allergic inflammation, goblet cell metaplasia, IgE synthesis, and bronchial hyperreactivity [74].

In addition to their involvement in the defense against parasites, eosinophils are also involved in other aspects of immunity as outlined below.

Antiviral functions

Eosinophils can inactivate viruses. Granular proteins of eosinophils such as ECP and the neurotoxin EDN have RNase activity that can inhibit viral replication in situ. This antiviral effect may be lost in patients with allergic asthma [64] and this may explain

why viral infections often precede exacerbations of allergic asthma. Eosinophils possess several pathogen-related receptors that can recognize viral antigens (for example, toll-like receptors TLR-3, -7, -9 and RIG-I receptor), produce several cytokines with antiviral effects (for example, IL-2, IL-12, and IFN- γ), express costimulatory molecules (for example, CD80, CD86, CD28, CD40) and are actively involved in the presentation of viral antigens to CD8 T lymphocytes [75]. Possible defense mechanisms of eosinophils against SARS-CoV-2 in COVID-19 have also been described [76]. While eosinopenia has been identified as a prospective screening, diagnostic, and prognostic tool, the actual role of eosinophils in lung pathology in this infection is unclear [77]. Eosinopenia has been shown to be an early, appropriate diagnostic and screening tool for COVID-19 infection [78, 79] and a prognostic marker for disease severity and unfavorable outcome in patients with COVID-19 pneumonia [80, 81, 82, 83]. Interestingly, eosinophilia (especially in asthma patients treated with inhaled corticosteroids) has been associated with a better outcome of COVID-19 disease [84]. On the other hand, studies analyzing the outcome of COVID-19 in patients with severe asthma under biologic therapy showed inconsistent results – also regarding SARS-CoV-2 vaccinations [85, 86, 87, 88, 89, 107].

Other anti-infective effects of eosinophils

The prominent role of eosinophils in parasitic infections is well known. These effects include antigen presentation and modulation of the T-cell reaction. They modulate the production of IgE and the production of mucus by goblet cells. In addition, their granular proteins are directly involved in the killing and neutralization of parasites [42, 90]. On the other hand, eosinophils can also have a deleterious effect in certain parasitic infections, which can contribute to tissue damage [91]. Eosinophils also play a role in the complex defense against selected bacteria. Although their phagocytic activity and bacterial killing is lower than that of neutrophils, they contribute to the elimination of selected bacteria, while their granu-

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lar proteins and enzymes help to neutralize bacterial proteins [92, 93]. The formation of extracellular traps by eosinophils (stimulated by various mediators, for example, thymic stromal lymphopoietin) is an important phenomenon in bacterial killing [94]. Finally, eosinophils also exert antifungal activities. They use their CD11b surface receptor to recognize β -glucan – a major component of the cell wall of fungi [53]. Proteases released by fungi activate protease-activated receptors in eosinophils, resulting in the release of various cytokines. In addition, eosinophils can probably inactivate fungal spores [95].

Modulation of inflammation and fibrosis

It is often not known that eosinophils also have regulatory or even anti-inflammatory properties. In mast cell-induced inflammation, for example, they can modulate the harmful effects of mast cell activation by oxidative deamination of histamine and enzymatic inactivation of other mast cell inflammatory mediators [96]. Eosinophils can also suppress T cells (“regulatory eosinophils”) [97]. In fibrogenesis, eosinophils play a pathophysiological role by releasing TGF- β and thus stimulate collagen production by parenchymal cells [98].

Hypereosinophilic syndrome (according to Schwaab et al. [99])

Blood eosinophils are measured by taking a differential blood count. For better comparability, it should be carried out under standardized conditions and without recent physical stress. A distinction is made between relative and absolute hypereosinophilia: the limit values of 0.5 G/L or 6% in the differential blood count from whole blood apply here. Blood hypereosinophilia can also be graded from mild to moderate to severe and in terms of its duration – transient, episodic, persistent. In order to make a diagnosis of hypereosinophilia in the blood, two abnormal samples must be taken more than 2 weeks apart.

Hypereosinophilia can also be detected in organs other than the blood. Tissue hypereosinophilia is diagnosed by clinical-pathological assessment of tissue samples if eosinophils or their markers are found extensively in the tissue (bone marrow: > 20% of nucleated cells). Tissue damage is not necessarily present in this case; however, if this is the case, a diagnosis of (tissue-specific) hypereosinophilic syndrome (Schwaab et al. [99]) should be made, depending on whether or not blood hypereosinophilia is also present at the same time.

Eosinophilia is associated with a variety of diseases that have different underlying causes and can affect different organs [99]. The diagnostic approach to hypereosinophilia is facilitated by the established subdivision into primary and secondary (= reactive) hypereosinophilia [100], which have been further refined according to updated classifications [101, 102]. Recently, new, refined diagnostic criteria and a classification of eosinophilic diseases have been proposed [103]:

- Familial (hereditary) hypereosinophilia – is often diagnosed in childhood and is sometimes associated with immunodeficiencies;
- Hypereosinophilia of unknown significance – without familial clustering, underlying pathology, associated molecular (genetic) abnormalities or organ damage caused by hypereosinophilia;
- Secondary (reactive) hypereosinophilia – non-clonal eosinophilia due to various immunoregulatory mechanisms;
- Primary (clonal, neoplastic) hypereosinophilia – caused by malignant degenerated eosinophils.

Conclusion

In addition to the well-characterized pro-inflammatory and disease-promoting effects of eosinophils in the context of chronic inflammatory diseases – including in the upper and lower respiratory tract – these cells also have homeostatic, anti-inflammatory, and anti-infectious activities.

These have often not been adequately considered to date, but these properties must be taken into account when selecting

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therapies that affect eosinophils in different ways. Therapeutic effects can range from a reduction in the number of cells to the complete depletion of eosinophils.

It is important to note that there has been considerable progress in therapies targeting eosinophils and that these therapies have become much more targeted and precise due to the availability of eosinophil (IL-5)-targeting monoclonal antibodies (mAbs, biologics) [104, 105, 106].

All biologics approved to date have shown a positive treatment effect and improved disease burden in patients with eosinophil-related diseases. However, studies on predictors of a good clinical response to these biologics are still insufficient and, particularly in patients with severe eosinophilic disease, there appears to be involvement and simultaneous activation of multiple signaling pathways. Phenotyping patients based on eosinophil granulocytes in the blood may therefore not be accurate enough for targeted endotyping. In CRSwNP and also in asthma, for example, the use of blood eosinophils as the only biomarker often proved to be insufficient for selecting the right drug for the right patient or for efficient monitoring of therapeutic response.

Therefore, it is important to understand the underlying immunology of CRSwNP and possibly establish immunoendotyping of patients to select the most appropriate treatment.

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References

- [1] Blanchard C, Rothenberg ME. Biology of the eosinophil. *Adv Immunol.* 2009; *101*: 81-121. [CrossRef PubMed](#)
- [2] Chusid MJ. Eosinophils: Friends or Foes? *J Allergy Clin Immunol Pract.* 2018; *6*: 1439-1444. [CrossRef PubMed](#)
- [3] Hillas G, Fouka E, Papaioannou AI. Antibodies targeting the interleukin-5 signaling pathway used as add-on therapy for patients with severe eosinophilic asthma: a review of the mechanism of action, efficacy, and safety of the subcutaneously administered agents, mepolizumab and benralizumab. *Expert Rev Respir Med.* 2020; *14*: 353-365. [CrossRef PubMed](#)
- [4] Simon D, Simon H-U. Therapeutic strategies for eosinophilic dermatoses. *Curr Opin Pharmacol.* 2019; *46*: 29-33. [CrossRef PubMed](#)
- [5] Abdala-Valencia H, Coden ME, Chiarella SE, Jacobsen EA, Bochner BS, Lee JJ, Berdnikovs S. Shaping eosinophil identity in the tissue contexts of development, homeostasis, and disease. *J Leukoc Biol.* 2018; *104*: 95-108. [CrossRef PubMed](#)
- [6] Marichal T, Mesnil C, Bureau F. Homeostatic Eosinophils: Characteristics and Functions. *Front Med (Lausanne).* 2017; *4*: 101. [CrossRef PubMed](#)
- [7] Diamant Z, Vijverberg S, Alving K, Bakirtas A, Bjermer L, Custovic A, Dahlen SE, Gaga M, Gerth van Wijk R, Giacco SD, Hamelmann E, Heaney LG, Heffler E, Kalayci Ö, Kostikas K, Lutter R, Olin AC, Sergejeva S, Simpson A, Sterk PJ, et al. Toward clinically applicable biomarkers for asthma: An EAACI position paper. *Allergy.* 2019; *74*: 1835-1851. [CrossRef PubMed](#)
- [8] Klion AD, Ackerman SJ, Bochner BS. Contributions of Eosinophils to Human Health and Disease. *Annu Rev Pathol.* 2020; *15*: 179-209. [CrossRef PubMed](#)
- [9] Ehrlich P. Beiträge zur Kenntnis der granulierten Bindegewebszellen und der eosinophilen Leukocythen in Leinenband der Zeit. *Arch. Anat. Phys. Berlin*; 1879. p. 166-169.
- [10] Gauvreau GM, Denburg JA. Human mast cell and basophil/eosinophil progenitors. *Methods Mol Biol.* 2015; *1220*: 59-68. [CrossRef PubMed](#)
- [11] Hassani M, Tak T, van Aalst C, van Nederveen S, Tesselaar K, Vrizekoop N, Koenderman L. Differential effects of short- and long-term treatment with mepolizumab on eosinophil kinetics in blood and sputum in eosinophilic asthma. *iScience.* 2021; *24*: 102913. [CrossRef PubMed](#)
- [12] Geijsen N, Koenderman L, Coffey PJ. Specificity in cytokine signal transduction: lessons learned from the IL-3/IL-5/GM-CSF receptor family. *Cytokine Growth Factor Rev.* 2001; *12*: 19-25. [CrossRef PubMed](#)
- [13] Johnston LK, Hsu CL, Krier-Burris RA, Chhiba KD, Chien KB, McKenzie A, Berdnikovs S, Bryce PJ. IL-33 Precedes IL-5 in Regulating Eosinophil Commitment and Is Required for Eosinophil Homeostasis. *J Immunol.* 2016; *197*: 3445-3453. [CrossRef PubMed](#)
- [14] Mack EA, Pear WS. Transcription factor and cytokine regulation of eosinophil lineage commitment. *Curr Opin Hematol.* 2020; *27*: 27-33. [CrossRef PubMed](#)
- [15] Hartl S, Breyer MK, Burghuber OC, Ofenheimer A, Schrott A, Urban MH, Agusti A, Studnicka M, Wouters EFM, Breyer-Kohansal R. Blood eosinophil count in the general population: typical values and potential confounders. *Eur Respir J.* 2020; *55*: 1901874. [CrossRef PubMed](#)
- [16] Dorman SC, Efthimiadis A, Babirad I, Watson RM, Denburg JA, Hargreave FE, O'Byrne PM, Sehmi R. Sputum CD34+IL-5Ralpha+ cells increase after allergen: evidence for in situ eosinophilopoiesis. *Am J Respir Crit Care Med.* 2004; *169*: 573-577. [CrossRef PubMed](#)
- [17] Steinbach KH, Schick P, Trepel F, Raffler H, Döhrmann J, Heilgeist G, Heltzel W, Li K, Past W, van der Woerd-de Lange JA, Theml H, Fliedner TM, Begemann H. Estimation of kinetic parameters of neutrophilic, eosinophilic, and basophilic granulocytes in human blood. *Blut.* 1979; *39*: 27-38. [CrossRef PubMed](#)
- [18] Walle AJ, Parwaresch MR. Estimation of effective eosinopoiesis and bone marrow eosinophil reserve capacity in normal man. *Cell Tissue Kinet.* 1979; *12*: 249-255. [CrossRef PubMed](#)
- [19] Willebrand R, Voehringer D. Regulation of eosinophil development and survival. *Curr Opin Hematol.* 2017; *24*: 9-15. [CrossRef PubMed](#)
- [20] Koenderman L. Priming: a critical step in the control of eosinophil activation. In: Lee JJ, Rosenberg HF (eds). *Eosinophils in Health and Disease*. Amsterdam; Elsevier Acad. Press: 2013. p. 170-179.
- [21] Bölke G, Church MK, Bergmann K-C. Comparison of extended intervals and dose reduction of omalizumab for asthma control. *Allergo J Int.* 2019; *28*: 1-4. [CrossRef](#)
- [22] Ciprandi G, Schiavetti I, Ricciardolo FLM. Patients with asthma consulting an allergist differ from those consulting a pulmonologist. *Allergo J Int.* 2023; *32*: 154-155. [CrossRef](#)
- [23] Greve J, Kinaciyan T, Maurer M, Dillenburger B, Recke A, Schöffl C. Expert consensus on prophylactic treatment of hereditary angioedema. *Allergo J Int.* 2022; *31*: 233-242. [CrossRef](#)
- [24] Klimek L, Casper I, Bergmann K-C, Biedermann T, Bousquet J, Hellings P, Jung K, Merk H, Olze H, Mösges R, Schlenker W, Gröger M, Ring J, Chaker A, Pfaar O, Wehrmann W, Zuberbier T, Becker S. Die Therapie der allergischen Rhinitis in der Routineversorgung: evidenzbasierte Nutzenbewertung der kombinierten Anwendung mehrerer

- Wirkstoffe. *Allergologie*. 2020; 43: 476-488. [CrossRef](#)
- [25] Klimek L, Förster-Ruhrmann U, Beule AG, Chaker AM, Hagemann J, Klimek F, Casper I, Huppertz T, Hoffmann TK, Dazert S, Deitmer T, Olze H, Strieth S, Wrede H, Schlenter W, Welkoborsky H-J, Woltenberg B, Bergmann C, Cuevas M, Beutner C, et al. Indicating biologics for chronic rhinosinusitis with nasal polyps (CRSwNP). *Allergo J Int*. 2022; 31: 149-160. [CrossRef](#)
- [26] Koennecke M, Klimek L, Mullol J, Gevaert P, Woltenberg B. Subtyping of polyposis nasi: phenotypes, endotypes and comorbidities. *Allergo J Int*. 2018; 27: 56-65. [CrossRef PubMed](#)
- [27] Luperto P, Masieri S, Cavaliere C, Compalati E, Ciprandi G, Frati F. Nasal cytology identifies allergic rhinitis phenotypes for managing allergen immunotherapy in clinical practice. *Allergo J Int*. 2022; 31: 51-55. [CrossRef](#)
- [28] Ong KY. What's new in the Global Initiative for Asthma 2018 report and beyond. *Allergo J Int*. 2019; 28: 63-72. [CrossRef](#)
- [29] Pitsios C, Pantavou K, Terreehorst I, Cianferoni A, Nowak-Wegzyn A, Vidal C, Vassilopoulou E, Papachristodoulou M, Tsigkrelis GP, Bonovas S, Nikolopoulos GK. Use of allergy tests to identify dietary and environmental triggers of eosinophilic esophagitis: protocol for a systematic review. *Allergo J Int*. 2020; 29: 280-283. [CrossRef](#)
- [30] Ring J, Beyer K, Biedermann T, Bircher A, Fischer M, Fuchs T, Heller A, Hoffmann F, Hutegger I, Jakob T, Klimek L, Kopp MV, Kugler C, Lange L, Pfaar O, Rietschel E, Rueff F, Schnadt S, Seifert R, Stöcker B, et al. Guideline (S2k) on acute therapy and management of anaphylaxis: 2021 update: S2k-Guideline of the German Society for Allergology and Clinical Immunology (DGAKI), the Medical Association of German Allergologists (AeDA), the Society of Pediatric Allergology and Environmental Medicine (GPA), the German Academy of Allergology and Environmental Medicine (DAAU), the German Professional Association of Pediatricians (BVKJ), the Society for Neonatology and Pediatric Intensive Care (GNPI), the German Society of Dermatology (DDG), the Austrian Society for Allergology and Immunology (ÖGAI), the Swiss Society for Allergy and Immunology (SGAI), the German Society of Anaesthesiology and Intensive Care Medicine (DGAI), the German Society of Pharmacology (DGP), the German Respiratory Society (DGP), the patient organization German Allergy and Asthma Association (DAAB), the German Working Group of Anaphylaxis Training and Education (AGATE). *Allergo J Int*. 2021; 30: 1-25. [CrossRef PubMed](#)
- [31] Rothe T. A century of "intrinsic asthma". *Allergo J Int*. 2018; 27: 215-219. [CrossRef](#)
- [32] Sennekamp J, Lehmann E, Joest M. Improved IgG antibody diagnostics of hypersensitivity pneumonitis and pulmonary mycoses by means of newly evaluated serum antibody ranges and frequencies using IgG ImmunoCAP™. *Allergo J Int*. 2022; 31: 172-182. [CrossRef](#)
- [33] Simon D, Straumann A, Schoepfer AM, Simon HU. Current concepts in eosinophilic esophagitis. *Allergo J Int*. 2017; 26: 258-266. [CrossRef PubMed](#)
- [34] Takatsu K, Tominaga A. Interleukin 5 and its receptor. *Prog Growth Factor Res*. 1991; 3: 87-102. [CrossRef PubMed](#)
- [35] Nishinakamura R, Miyajima A, Mee PJ, Tybulewicz VL, Murray R. Hematopoiesis in mice lacking the entire granulocyte-macrophage colony-stimulating factor/interleukin-3/interleukin-5 functions. *Blood*. 1996; 88: 2458-2464. [CrossRef PubMed](#)
- [36] Conus S, Straumann A, Bettler E, Simon HU. Mepolizumab does not alter levels of eosinophils, T cells, and mast cells in the duodenal mucosa in eosinophilic esophagitis. *J Allergy Clin Immunol*. 2010; 126: 175-177. [CrossRef PubMed](#)
- [37] Leckie MJ, ten Brinke A, Khan J, Diamant Z, O'Connor BJ, Walls CM, Mathur AK, Cowley HC, Chung KF, Djukanovic R, Hansel TT, Holgate ST, Sterk PJ, Barnes PJ. Effects of an interleukin-5 blocking monoclonal antibody on eosinophils, airway hyper-responsiveness, and the late asthmatic response. *Lancet*. 2000; 356: 2144-2148. [CrossRef PubMed](#)
- [38] Flood-Page PT, Menzies-Gow AN, Kay AB, Robinson DS. Eosinophil's role remains uncertain as anti-interleukin-5 only partially depletes numbers in asthmatic airway. *Am J Respir Crit Care Med*. 2003; 167: 199-204. [CrossRef PubMed](#)
- [39] Mesnil C, Raulier S, Paulissen G, Xiao X, Birrell MA, Pirotin D, Janss T, Starkl P, Ramery E, Henket M, Schleich FN, Radermecker M, Thielemans K, Gillet L, Thiry M, Belvisi MG, Louis R, Desmet C, Marichal T, Bureau F. Lung-resident eosinophils represent a distinct regulatory eosinophil subset. *J Clin Invest*. 2016; 126: 3279-3295. [CrossRef PubMed](#)
- [40] Gigon L, Fettelet T, Yousefi S, Simon D, Simon HU. Eosinophils from A to Z. *Allergy*. 2023; 78: 1810-1846. [CrossRef PubMed](#)
- [41] O'Connell EM, Nutman TB. Eosinophilia in Infectious Diseases. *Immunol Allergy Clin North Am*. 2015; 35: 493-522. [CrossRef PubMed](#)
- [42] Klion AD, Nutman TB. The role of eosinophils in host defense against helminth parasites. *J Allergy Clin Immunol*. 2004; 113: 30-37. [CrossRef PubMed](#)
- [43] Bochner BS. The eosinophil: For better or worse, in sickness and in health. *Ann Allergy Asthma Immunol*. 2018; 121: 150-155. [CrossRef PubMed](#)
- [44] Krishack PA, Louviere TJ, Decker TS, Kuzel TG, Greenberg JA, Camacho DF, Hrusch CL, Sperling AI, Verhoef PA. Protection against *Staphylococcus aureus* bacteremia-induced mortality depends on ILC2s and eosinophils. *JCI Insight*. 2019; 4: e124168. [CrossRef PubMed](#)
- [45] Shah K, Ignacio A, McCoy KD, Harris NL. The emerging roles of eosinophils in mucosal homeostasis. *Mucosal Immunol*. 2020; 13: 574-583. [CrossRef PubMed](#)
- [46] Rao RR, Long JZ, White JP, Svensson KJ, Lou J, Lokurkar I, Jedrychowski MP, Ruas JL, Wrann CD, Lo JC, Camera DM, Lachey J, Gygi S, Seehra J, Hawley JA, Spiegelman BM. Meteorin-like is a hormone that regulates immune-adipose interactions to increase beige fat thermogenesis. *Cell*. 2014; 157: 1279-1291. [CrossRef PubMed](#)
- [47] da Silva FR, Soares Thimoteo D, Ferraz Carbonel A, Fuchs LFP, da Silva Sasso GR, Dos Santos Simões R, Invitti A, Ramos Vieira R, de Souza Ferreira LP, da Silva RA, Lima PDA, Soares Júnior JM, de Jesus Simões M, Bertoncini CRA. Histomorphometric analysis of the endometrium in an ectopic

Klimek F, Bergmann C, Hagemann J, Cuevas M, Becker S, Pfaar O, Casper I, Klimek L.
Eosinophil granulocytes in chronic inflammatory respiratory diseases and CRSwNP: Function, immunological basis, and clinical significance. *Allergol Select*. 2024; 8: 40-50.
DOI 10.5414/ALX02469E

- model of endometriosis in mice. *Gynecol Endocrinol.* 2022; 38: 874-878. [CrossRef PubMed](#)
- [48] Lee JJ, Jacobsen EA, McGarry MP, Schleimer RP, Lee NA. Eosinophils in health and disease: the LIAR hypothesis. *Clin Exp Allergy.* 2010; 40: 563-575. [CrossRef PubMed](#)
- [49] Bochner BS, Schleimer RP. The role of adhesion molecules in human eosinophil and basophil recruitment. *J Allergy Clin Immunol.* 1994; 94: 427-438, quiz 439. [CrossRef PubMed](#)
- [50] Knol EF, Tackey F, Tedder TF, Klunk DA, Bickel CA, Sterbinsky SA, Bochner BS. Comparison of human eosinophil and neutrophil adhesion to endothelial cells under nonstatic conditions. Role of L-selectin. *J Immunol.* 1994; 153: 2161-2167. [Cross-Ref PubMed](#)
- [51] Koenderman L, Hermans SW, Capel PJ, van de Winkel JG. Granulocyte-macrophage colony-stimulating factor induces sequential activation and deactivation of binding via a low-affinity IgG Fc receptor, hFc gamma RII, on human eosinophils. *Blood.* 1993; 81: 2413-2419. [CrossRef PubMed](#)
- [52] Monteiro RC, Hostoffer RW, Cooper MD, Bonner JR, Gartland GL, Kubagawa H. Definition of immunoglobulin A receptors on eosinophils and their enhanced expression in allergic individuals. *J Clin Invest.* 1993; 92: 1681-1685. [CrossRef PubMed](#)
- [53] Yoon J, Ponikau JU, Lawrence CB, Kita H. Innate antifungal immunity of human eosinophils mediated by a beta 2 integrin, CD11b. *J Immunol.* 2008; 181: 2907-2915. [CrossRef PubMed](#)
- [54] Koenderman L, van der Bruggen T, Schweizer RC, Warringa RA, Coffey P, Caldenhoven E, Lammers JW, Raaijmakers JA. Eosinophil priming by cytokines: from cellular signal to in vivo modulation. *Eur Respir J Suppl.* 1996; 22: 119s-125s. [PubMed](#)
- [55] Bracke M, van de Graaf E, Lammers JW, Coffey PJ, Koenderman L. In vivo priming of FcalphaR functioning on eosinophils of allergic asthmatics. *J Leukoc Biol.* 2000; 68: 655-661. [CrossRef PubMed](#)
- [56] Lacy P, Abdel-Latif D, Steward M, Musat-Marcu S, Man SF, Moqbel R. Divergence of mechanisms regulating respiratory burst in blood and sputum eosinophils and neutrophils from atopic subjects. *J Immunol.* 2003; 170: 2670-2679. [CrossRef PubMed](#)
- [57] Hagan SP, Rosenberg HF, Moqbel R, Phipps S, Foster PS, Lacy P, Kay AB, Rothenberg ME. Eosinophils: biological properties and role in health and disease. *Clin Exp Allergy.* 2008; 38: 709-750. [CrossRef PubMed](#)
- [58] Mukherjee M, Lacy P, Ueki S. Eosinophil Extracellular Traps and Inflammatory Pathologies-Untangling the Web! *Front Immunol.* 2018; 9: 2763. [CrossRef PubMed](#)
- [59] Bandeira-Melo C, Weller PF. Eosinophils and cysteinyl leukotrienes. *Prostaglandins Leukot Essent Fatty Acids.* 2003; 69: 135-143. [CrossRef PubMed](#)
- [60] Davoine F, Lacy P. Eosinophil cytokines, chemokines, and growth factors: emerging roles in immunity. *Front Immunol.* 2014; 5: 570. [CrossRef PubMed](#)
- [61] Tool AT, Koenderman L, Kok PT, Blom M, Roos D, Verhoeven AJ. Release of platelet-activating factor is important for the respiratory burst induced in human eosinophils by opsonized particles. *Blood.* 1992; 79: 2729-2732. [CrossRef PubMed](#)
- [62] Lacy P. The role of Rho GTPases and SNAREs in mediator release from granulocytes. *Pharmacol Ther.* 2005; 107: 358-376. [CrossRef PubMed](#)
- [63] Lacy P, Logan MR, Bablitz B, Moqbel R. Fusion protein vesicle-associated membrane protein 2 is implicated in IFN-gamma-induced piecemeal degranulation in human eosinophils from atopic individuals. *J Allergy Clin Immunol.* 2001; 107: 671-678. [CrossRef PubMed](#)
- [64] Logan MR, Lacy P, Bablitz B, Moqbel R. Expression of eosinophil target SNAREs as potential cognate receptors for vesicle-associated membrane protein-2 in exocytosis. *J Allergy Clin Immunol.* 2002; 109: 299-306. [CrossRef PubMed](#)
- [65] Logan MR, Lacy P, Odemuyiwa SO, Steward M, Davoine F, Kita H, Moqbel R. A critical role for vesicle-associated membrane protein-7 in exocytosis from human eosinophils and neutrophils. *Allergy.* 2006; 61: 777-784. [CrossRef PubMed](#)
- [66] Willetts L, Felix LC, Jacobsen EA, Puttagunta L, Condjella RM, Zellner KR, Ochkur SJ, Kim JD, Luo H, Lee NA, Lee JJ, Moqbel R, Lacy P. Vesicle-associated membrane protein 7-mediated eosinophil degranulation promotes allergic airway inflammation in mice. *Commun Biol.* 2018; 1: 83. [Cross-Ref PubMed](#)
- [67] Radonjic-Hoesli S, Wang X, de Graauw E, Stoeckle C, Styp-Rekowska B, Hlushchuk R, Simon D, Spaeth PJ, Yousefi S, Simon HU. Adhesion-induced eosinophil cytolysis requires the receptor-interacting protein kinase 3 (RIPK3)-mixed lineage kinase-like (MLKL) signaling pathway, which is counterregulated by autophagy. *J Allergy Clin Immunol.* 2017; 140: 1632-1642. [CrossRef PubMed](#)
- [68] Simon H-U, Yousefi S, Germic N, Arnold IC, Haczk A, Karaulov AV, Simon D, Rosenberg HF. The Cellular Functions of Eosinophils: Collegium Internationale Allergologica (CIA) Update 2020. *Int Arch Allergy Immunol.* 2020; 181: 11-23. [CrossRef PubMed](#)
- [69] Yousefi S, Gold JA, Andina N, Lee JJ, Kelly AM, Kozlowski E, Schmid I, Straumann A, Reichenbach J, Gleich GJ, Simon HU. Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. *Nat Med.* 2008; 14: 949-953. [CrossRef PubMed](#)
- [70] Nogawa H, Suzuki H, Kawabata Y, Ota T, Yuki Y, Katagiri Y, Hino T, Yanagawa N, Ueki S. An unusual case of eosinophilic lung disease with multiple cyst formation. *Respir Med Case Rep.* 2020; 31: 101300. [CrossRef PubMed](#)
- [71] Fettlelet T, Gigon L, Karaulov A, Yousefi S, Simon HU. The Enigma of Eosinophil Degranulation. *Int J Mol Sci.* 2021; 22: 7091. [CrossRef PubMed](#)
- [72] Ueki S, Konno Y, Takeda M, Moritoki Y, Hirokawa M, Matsuwaki Y, Honda K, Ohta N, Yamamoto S, Takagi Y, Wada A, Weller PF. Eosinophil extracellular trap cell death-derived DNA traps: their presence in secretions and functional attributes. *J Allergy Clin Immunol.* 2016; 137: 258-267. [CrossRef PubMed](#)
- [73] Ueki S, Miyabe Y, Yamamoto Y, Fukuchi M, Hirokawa M, Spencer LA, Weller PF. Charcot-Leyden Crystals in Eosinophilic Inflammation: Active Cytolysis Leads to Crystal Formation. *Curr Allergy Asthma Rep.* 2019; 19: 35. [CrossRef PubMed](#)
- [74] Persson EK, Verstraete K, Heyndrickx I, Gevaert E, Aegerter H, Percier JM, Deswarte K, Verschueren KHG, Dansercoer A, Gras D, Chanez P, Bachert C,

- Gonçalves A, Van Gorp H, De Haard H, Blanchetot C, Saunders M, Hammad H, Savvides SN, Lambrecht BN. Protein crystallization promotes type 2 immunity and is reversible by antibody treatment. *Science*. 2019; 364: eaaw4295. [CrossRef PubMed](#)
- [75] Rodrigo-Muñoz JM, Gil-Martínez M, Sastre B, Del Pozo V. Emerging Evidence for Pleiotropism of Eosinophils. *Int J Mol Sci*. 2021; 22: 7075 [CrossRef PubMed](#)
- [76] Rodrigo-Muñoz JM, Sastre B, Cañas JA, Gil-Martínez M, Redondo N, Del Pozo V. Eosinophil Response Against Classical and Emerging Respiratory Viruses: COVID-19. *J Investig Allergol Clin Immunol*. 2021; 31: 94-107. [CrossRef PubMed](#)
- [77] Jesenak M, Banovcin P, Diamant Z. COVID-19, chronic inflammatory respiratory diseases and eosinophils-Observations from reported clinical case series. *Allergy*. 2020; 75: 1819-1822. [Cross-Ref PubMed](#)
- [78] Myari A, Papapetrou E, Tsaousi C. Diagnostic value of white blood cell parameters for COVID-19: Is there a role for HFLC and IG? *Int J Lab Hematol*. 2022; 44: 104-111. [CrossRef PubMed](#)
- [79] Soni M. Evaluation of eosinopenia as a diagnostic and prognostic indicator in COVID-19 infection. *Int J Lab Hematol*. 2021; 43 (Suppl 1): 137-141. [CrossRef PubMed](#)
- [80] Cazzaniga M, Fumagalli LAM, D'angelo L, Cerino M, Bonfanti G, Fumagalli RM, Schiavo G, Lorini C, Lainu E, Terragni S, Chiarelli M, Scarazzati C, Bonato C, Zago M. Eosinopenia is a reliable marker of severe disease and unfavourable outcome in patients with COVID-19 pneumonia. *Int J Clin Pract*. 2021; 75: e14047. [CrossRef PubMed](#)
- [81] Eijmael M, Janssens N, le Cessie S, van Dooren Y, Koster T, Karim F. Coronavirus disease 2019 and peripheral blood eosinophil counts: a retrospective study. *Infection*. 2021; 49: 1325-1329. [Cross-Ref PubMed](#)
- [82] Klimek L, Novak N, Hamelmann E, Werfel T, Wagenmann M, Taube C, Bauer A, Merk H, Rabe U, Jung K, Schlenker W, Ring J, Chaker A, Wehrmann W, Becker S, Mülleneisen N, Nemat K, Czech W, Wrede H, Brehler R, et al. Severe allergic reactions after COVID-19 vaccination with the Pfizer/BioNTech vaccine in Great Britain and USA: Position statement of the German Allergy Societies: Medical Association of German Allergologists (AeDA), German Society for Allergology and Clinical Immunology (DGAKI) and Society for Pediatric Allergology and Environmental Medicine (GPA). *Allergo J Int*. 2021; 30: 51-55. [CrossRef PubMed](#)
- [83] Ring J, Beyer K, Biedermann T, Bircher A, Fischer M, Fuchs T, Heller A, Hoffmann F, Huttegger I, Jakob T, Klimek L, Kopp MV, Kugler C, Lange L, Pfaar O, Rietschel E, Rueff F, Schnadt S, Seifert R, Stöcker B, et al. Messages for patients and relatives from the 2021 update of the guideline on acute therapy and management of anaphylaxis. *Allergo J Int*. 2021; 30: 243-248. [CrossRef PubMed](#)
- [84] Zein JG, Strauss R, Attaway AH, Hu B, Milinovich A, Jawhari N, Chamat SS, Ortega VE. Eosinophilia Is Associated with Improved COVID-19 Outcomes in Inhaled Corticosteroid-Treated Patients. *J Allergy Clin Immunol Pract*. 2022; 10: 742-750.e14. [CrossRef PubMed](#)
- [85] Eger K, Hashimoto S, Braunstahl GJ, Brinke AT, Patberg KW, Beukert A, Smeenk F, van der Sar-van der Brugge S, Weersink EJM, Bel EH. Poor outcome of SARS-CoV-2 infection in patients with severe asthma on biologic therapy. *Respir Med*. 2021; 177: 106287. [CrossRef PubMed](#)
- [86] Francis CHR, Hearn AP, Ratnakumar S, Taylor A, Duckitt J, Ahmed U, Dhariwal J, Nanzer AM, Jackson DJ. COVID-19 in the absence of eosinophils: The outcome of confirmed SARS-CoV-2 infection whilst on treatment with benralizumab. *Allergy*. 2022; 77: 2558-2560. [CrossRef PubMed](#)
- [87] Klimek L, Bergmann KC, Brehler R, Pfützner W, Zuberbier T, Hartmann K, Jakob T, Novak N, Ring J, Merk H, Hamelmann E, Ankermann T, Schmidt S, Untersmayr E, Hötzenecker W, Jensen-Jarolim E, Brockow K, Mahler V, Worm M. Practical handling of allergic reactions to COVID-19 vaccines: A position paper from German and Austrian Allergy Societies AeDA, DGAKI, GPA and ÖGAI. *Allergo J Int*. 2021; 30: 79-95. [CrossRef PubMed](#)
- [88] Klimek L, Hagemann J, Döge J, Freudelsperger L, Cuevas M, Klimek F, Hummel T. Olfactory and gustatory disorders in COVID-19. *Allergo J Int*. 2022; 31: 243-250. [CrossRef PubMed](#)
- [89] Rost J, Langhein S, Bartel D, Bonertz A, Mahler V. Good manufacturing practice- and good distribution practice-compliant cold storage and refrigerated transport of allergen products: what is important? *Allergo J Int*. 2022; 31: 36-42. [CrossRef](#)
- [90] Huang L, Appleton JA. Eosinophils in Helminth Infection: Defenders and Dupes. *Trends Parasitol*. 2016; 32: 798-807. [CrossRef PubMed](#)
- [91] Cadman ET, Thyse KA, Bearder S, Cheung AY, Johnston AC, Lee JJ, Lawrence RA. Eosinophils are important for protection, immunoregulation and pathology during infection with nematode microfilariae. *PLoS Pathog*. 2014; 10: e1003988. [Cross-Ref PubMed](#)
- [92] Linch SN, Gold JA. The role of eosinophils in non-parasitic infections. *Endocr Metab Immune Disord Drug Targets*. 2011; 11: 165-172. [CrossRef PubMed](#)
- [93] Ondari E, Calvino-Sanles E, First NJ, Gestal MC. Eosinophils and Bacteria, the Beginning of a Story. *Int J Mol Sci*. 2021; 22: 8004. [CrossRef PubMed](#)
- [94] Morshed M, Yousefi S, Stöckle C, Simon HU, Simon D. Thymic stromal lymphopoietin stimulates the formation of eosinophil extracellular traps. *Allergy*. 2012; 67: 1127-1137. [CrossRef PubMed](#)
- [95] Figueiredo RT, Neves JS. Eosinophils in fungal diseases: An overview. *J Leukoc Biol*. 2018; 104: 49-60. [CrossRef PubMed](#)
- [96] Austen KF. Homeostasis of effector systems which can also be recruited for immunologic reactions. *J Immunol*. 1978; 121: 793-805. [CrossRef PubMed](#)
- [97] Lingblom C, Andersson J, Andersson K, Wennerås C. Regulatory Eosinophils Suppress T Cells Partly through Galectin-10. *J Immunol*. 2017; 198: 4672-4681. [CrossRef PubMed](#)
- [98] Takemura N, Kurashima Y, Mori Y, Okada K, Ogino T, Osawa H, Matsuno H, Aayam L, Kaneto S, Park EJ, Sato S, Matsunaga K, Tamura Y, Ouchi Y, Kumagai Y, Kobayashi D, Suzuki Y, Yoshioka Y, Nishimura J, Mori M, et al. Eosinophil depletion suppresses radiation-induced small intestinal fibrosis. *Sci Transl Med*. 2018; 10: eaan0333. [CrossRef PubMed](#)
- [99] Schwaab J, Lübke J, Reiter A, Metzgeroth G. Idiopathic hypereosinophilic syndrome – diagnosis

- and treatment. *Allergo J Int.* 2022; 31: 251-256. [CrossRef](#)
- [100] Simon D, Simon H-U. Eosinophilic disorders. *J Allergy Clin Immunol.* 2007; 119: 1291-1300, quiz 1301-1302. [CrossRef PubMed](#)
- [101] Gotlib J. World Health Organization-defined eosinophilic disorders: 2017 update on diagnosis, risk stratification, and management. *Am J Hematol.* 2017; 92: 1243-1259. [CrossRef PubMed](#)
- [102] Valent P, Gleich GJ, Reiter A, Roufosse F, Weller PF, Hellmann A, Metzgeroth G, Leiferman KM, Arock M, Sotlar K, Butterfield JH, Cerny-Reiterer S, Mayerhofer M, Vandenberghe P, Haferlach T, Bochner BS, Gotlib J, Horny HP, Simon HU, Klion AD. Pathogenesis and classification of eosinophil disorders: a review of recent developments in the field. *Expert Rev Hematol.* 2012; 5: 157-176. [CrossRef PubMed](#)
- [103] Valent P, Klion AD, Roufosse F, Simon D, Metzgeroth G, Leiferman KM, Schwaab J, Butterfield JH, Sperr WR, Sotlar K, Vandenberghe P, Hoermann G, Haferlach T, Moriggl R, George TI, Akin C, Bochner BS, Gotlib J, Reiter A, Horny HP, et al. Proposed refined diagnostic criteria and classification of eosinophil disorders and related syndromes. *Allergy.* 2023; 78: 47-59. [CrossRef PubMed](#)
- [104] Gschwend A, Helbling A, Feldmeyer L, Mani-Weber U, Meincke C, Heidemeyer K, Bossart S, Jörg L. Treatment with IL5-/IL-5 receptor antagonists in drug reaction with eosinophilia and systemic symptoms (DRESS). *Allergo J Int.* 2022; 32: 1-8. [CrossRef PubMed](#)
- [105] Kendziora B, Frey J, Reinholz M, Ruëff F, Oppel E, Zuberbier T, Hartmann D, Schlager JG, French LE. Efficacy and safety of medications for antihistamine-refractory chronic spontaneous urticaria: a systematic review and network meta-analysis. *Allergo J Int.* 2023; 32: 83-92. [CrossRef](#)
- [106] Miranda J, Plácido JL, Amaral L. A case of protracted eosinopenia after a single subcutaneous dose of benralizumab. *Allergo J Int.* 2022; 32: 1-2. [CrossRef PubMed](#)
- [107] Pfaar O, Klimek L, Hamelmann E, Kleine-Tebbe J, Taube C, Wagenmann M, Werfel T, Brehler R, Novak N, Mülleneisen N, Becker S, Worm M. COVID-19 vaccination of patients with allergies and type-2 inflammation with concurrent antibody therapy (biologics) - A Position Paper of the German Society of Allergology and Clinical Immunology (DGAKI) and the German Society for Applied Allergology (AeDA). *Allergol Select.* 2021; 5: 140-147. [CrossRef PubMed](#)