

Commentary

# Considerations for exerkin research focusing on the response to exercise training

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García-Hermoso and colleagues<sup>1</sup> recently published a systematic literature review and meta-analysis on exercise training-induced changes in exerkin concentrations in type 2 diabetes mellitus patients, providing a contemporary view on how exerkins respond to exercise training. That review prompted us to highlight 2 additional considerations that should be taken into account when studying the response of exerkins to exercise training. Firstly, whether exerkins can exhibit discordant responses to acute exercise compared to exercise training, and secondly, the need to consider the residual effects of the most recent exercise bout.

The term “exerkin” has been operationally defined as “a signaling moiety released in response to acute exercise and/or chronic exercise, exerting its effects through endocrine, paracrine, and/or autocrine pathways”, and encompasses “a broad range of signaling moieties, including cytokines, nucleic acids (microRNA, mRNA, and mitochondrial DNA), lipids, and metabolites, which are frequently driven by cell-specific extracellular vesicle secretion”.<sup>2</sup> To date, most exerkin research has focused on molecules that have circulating concentrations that are increased in response to acute exercise.<sup>2,3</sup> Whether there are changes in resting concentrations of exerkins as a consequence of exercise training is an important question for the field. For example, health benefits conveyed by exerkins may be limited to the effects of acute exercise whereby exerkins function through stimulating transient changes in tissue metabolism via signal transduction and/or altered gene expression. Alternatively, exerkins could play a more comprehensive role in conveying the benefits of exercise whereby exercise training induces altered exerkin profiles at rest that lead to more persistent changes in tissue metabolism and/or signaling, thereby creating a more prolonged time course of benefit.<sup>3</sup>

## Discordance between the response to acute exercise and exercise training

Some exerkins are pleiotropic molecules in the sense that they can be released from different cell types under different conditions/stimuli and elicit varying physiological effects depending on the context of their release. For example, interleukin-6 (IL-6) is released from activated macrophages<sup>4</sup> and adipocytes within adipose tissue to elicit proinflammatory effects,<sup>5</sup> and consequent insulin resistance.<sup>4,5</sup> IL-6 is also released from contracting skeletal muscle during acute exercise, with resultant effects including potentiating insulin secretion and action, enhancing glucose uptake, and increasing fat oxidation.<sup>6</sup> In contrast to acute exercise where circulating IL-6 concentrations increase, García-Hermoso and colleagues’ analysis<sup>1</sup> found that circulating IL-6 concentrations decrease at rest in response to exercise training. In fact, the exercise training-induced decreases in resting IL-6 concentrations were positively correlated with a decrease in hemoglobin A1c ( $\beta = 0.44$ ,  $p = 0.012$ ). Therefore, despite the increase in circulating IL-6 stimulated by acute exercise contributing to whole body glucose control during and after acute exercise, a decrease in resting IL-6 concentration in response to exercise training is also associated with improvements in glucose control.<sup>6</sup> Release of IL-6 from skeletal muscle is negligible at rest.<sup>7</sup> Therefore the decreases in circulating IL-6 elicited by exercise training is likely caused by lower IL-6 release from non-muscle sources such as adipose tissue<sup>8</sup> which when present and indicative of chronic, subclinical inflammation can contribute to tissue-specific insulin resistance.<sup>9</sup> A similar example is fetuin-A, a liver- and adipose tissue-derived glycoprotein whose elevated concentration is associated with insulin resistance.<sup>10</sup> Circulating concentration of fetuin-A is transiently elevated by a single bout of exercise before returning to baseline after 24 h,<sup>11</sup> yet is decreased in response to exercise training.<sup>1</sup>

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These examples illustrate that an exerkine may exhibit discordant responses to acute exercise compared to exercise training, with the discordance being characterized by aspects such as the direction of change in concentration, the cell- or tissue-specific source of the exerkine, and the metabolic consequence of exerkine action. Another important implication is that characterisation of the acute exerkine response cannot be used to infer the response to exercise training, and especially if there are several cell/tissue sources of that exerkine, and several possible mechanisms of action. This point is salient given that several recent studies report a comprehensive characterisation of the exerkine response to acute exercise,<sup>12,13</sup> yet the implications for metabolic health and adaptations to exercise training for many candidate exerkines remain to be described.

### The residual influence of the most recent acute exercise bout

Acute exercise bouts produce short-term improvements in markers of cardiovascular and metabolic health, such as reduced blood pressure and circulating triglycerides, and increased insulin sensitivity.<sup>9</sup> These benefits can persist for periods of ~16–48 h,<sup>14–17</sup> but many benefits are not augmented or attenuated with a period of exercise training.<sup>9,18</sup> These observations demonstrate that acute exercise is capable of conveying discrete benefits that are independent from responses or adaptations to exercise training, an effect previously termed as “last bout effects”.<sup>18</sup> From a methodological perspective, when investigating the effects of exercise training specifically, the presence of protracted residual influences of a recent acute exercise bout may be a confounding factor in studies where the timing of the post-training blood sample has not accounted for this phenomenon. An example would be when post-training blood samples are taken <24 h after the last exercise bout of the training intervention. In a recent multi-omic investigation, most candidate exerkines identified to change in response to acute exercise trended back towards their baseline concentration ~1 h after exercise, yet some continue to increase at 1 h after exercise.<sup>12</sup> Few studies have characterized the exerkine response beyond 3–8 h after exercise, but illustrative examples include 2 microRNA (out of 7 total transcripts measured) were increased,<sup>19</sup> and 6 metabolites were altered (5 decreased and 1 increased),<sup>20</sup> in circulation at 24 h after exercise.<sup>19,20</sup> Therefore, we have previously suggested that it may be useful to obtain blood samples at least 24 h after exercise cessation when investigating the influence of exercise training on exerkines.<sup>3</sup> We carefully examined the studies cited in Supplementary Table 2 by García-Hermoso et al.<sup>1</sup> and found that only 15 of the 41 trials analyzed reported the proximity to acute exercise from which samples were obtained.

This finding highlights a flaw in methodological and/or reporting approach that is quite pervasive in experimental studies of the response of exerkines to exercise training. Not reporting the proximity of the most recent exercise bout for blood samples taken after an exercise training intervention impacts the confidence with which results can be specifically attributed to the effect of that exercise training.

### Concluding remarks

This recent systematic review by García-Hermoso et al.<sup>1</sup> provides an important contribution to an important aspect of the ever-expanding field of exerkine research. The findings reported provide thought-provoking insights into how exerkines are presently understood to respond to exercise training, and we intend our comments to provide useful considerations for planning and interpretation of future experiments in this field.

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### Authors' contributions

IAJD contributed to the conceptualization, formal analysis, and writing of the original draft; BE contributed to the conceptualization, writing (review and editing), and supervision, as well as funding acquisition. Both authors have read and approved the final version of the manuscript, and agree with the order of presentation of the authors.

### Competing interests

Both authors declare that they have no competing interests.

### References

- García-Hermoso A, Ramírez-Vélez R, Díez J, González A, Izquierdo M. Exercise training-induced changes in exerkine concentrations may be relevant to the metabolic control of type 2 diabetes mellitus patients: A systematic review and meta-analysis of randomized controlled trials. *J Sport Health Sci* 2023;**12**:147–57.
- Chow LS, Gerszten RE, Taylor JM, et al. Exerkines in health, resilience and disease. *Nat Rev Endocrinol* 2022;**18**:273–89.
- Darragh IAJ, O'Driscoll L, Egan B. Exercise training and circulating small extracellular vesicles: Appraisal of methodological approaches and current knowledge. *Front Physiol* 2021;**12**:738333. doi:10.3389/fphys.2021.738333.
- Harford KA, Reynolds CM, McGillicuddy FC, Roche HM. Fats, inflammation and insulin resistance: Insights to the role of macrophage and T-cell accumulation in adipose tissue. *Proc Nutr Soc* 2011;**70**:408–17.
- Han MS, White A, Perry RJ, et al. Regulation of adipose tissue inflammation by interleukin 6. *Proc Natl Acad Sci U S A* 2020;**117**:2751–60.
- Pedersen BK, Febbraio MA. Interleukin-6 does have a beneficial role in insulin sensitivity and glucose homeostasis. *J Appl Physiol* (1985) 2007;**102**:814–6.
- Steensberg A, van Hall G, Osada T, Sacchetti M, Saltin B, Klarlund Pedersen B. Production of interleukin-6 in contracting human skeletal muscles can account for the exercise-induced increase in plasma interleukin-6. *J Physiol* 2000;**529**:237–42.
- Kawanishi N, Yano H, Yokogawa Y, Suzuki K. Exercise training inhibits inflammation in adipose tissue via both suppression of macrophage infiltration and acceleration of phenotypic switching from M1 to M2 macrophages in high-fat-diet-induced obese mice. *Exerc Immunol Rev* 2010;**16**:105–18.
- Booth FW, Roberts CK, Laye MJ. Lack of exercise is a major cause of chronic diseases. *Compr Physiol* 2012;**2**:1143–211.

10. Stefan N, Hennige AM, Staiger H, et al.  $\alpha$ 2-Heremans-Schmid glycoprotein/fetuin-A is associated with insulin resistance and fat accumulation in the liver in humans. *Diabetes Care* 2006;**29**:853–7.
11. Ren G, Bowers RL, Kim T, et al. Serum fetuin-A and Ser312 phosphorylated fetuin-A responses and markers of insulin sensitivity after a single bout of moderate intensity exercise. *Physiol Rep* 2021;**9**:e14773. doi:10.14814/phy2.14773.
12. Contrepolis K, Wu S, Moneghetti KJ, et al. Molecular choreography of acute exercise. *Cell* 2020;**181**:1112–30.
13. Wei W, Riley NM, Lyu X, et al. Organism-wide, cell-type-specific secretome mapping of exercise training in mice. *Cell Metab* 2023;**35**:1261–79.
14. Gill JM, Mees GP, Frayn KN, Hardman AE. Moderate exercise, postprandial lipaemia and triacylglycerol clearance. *Eur J Clin Invest* 2001;**31**:201–7.
15. Malkova D, Evans RD, Frayn KN, Humphreys SM, Jones PR, Hardman AE. Prior exercise and postprandial substrate extraction across the human leg. *Am J Physiol Endocrinol Metab* 2000;**279**:E1020–8.
16. Mikines KJ, Sonne B, Farrell PA, Tronier B, Galbo H. Effect of physical exercise on sensitivity and responsiveness to insulin in humans. *Am J Physiol* 1988;**254**:E248–59.
17. Perseghin G, Price TB, Petersen KF, et al. Increased glucose transport-phosphorylation and muscle glycogen synthesis after exercise training in insulin-resistant subjects. *N Engl J Med* 1996;**335**:1357–62.
18. Haskell WL, JB. Wolffe Memorial Lecture. Health consequences of physical activity: Understanding and challenges regarding dose–response. *Med Sci Sports Exerc* 1994;**26**:649–60.
19. Mooren FC, Viereck J, Krüger K, Thum T. Circulating microRNAs as potential biomarkers of aerobic exercise capacity. *Am J Physiol Heart Circ Physiol* 2014;**306**:H557–63.
20. Cendali F, D’Alessandro A, Nemkov T. Dried blood spot characterization of sex-based metabolic responses to acute running exercise. *Anal Sci Adv* 2023;**4**:37–48.