

Preprints are preliminary reports that have not undergone peer review. They should not be considered conclusive, used to inform clinical practice, or referenced by the media as validated information.

Enhanced Multiscale Human Brain Imaging by Semi-supervised Digital Staining and Serial Sectioning Optical Coherence Tomography

Lei Tian leitian@bu.edu Boston University https://orcid.org/0000-0002-1316-4456 Shiyi Cheng **Boston University** Shuaibin Chang Boston University https://orcid.org/0000-0003-3416-7290 Yunzhe Li University of California, Berkeley Anna Novoseltseva Department of Biomedical Engineering, Boston University Sunni Lin **Boston University Yicun Wu Boston University** Jiahui Zhu **Boston University** Ann Mckee **Boston University Douglas Rosene Boston University** Hui Wang Department of Radiology, Massachusetts General Hospital, A.A. Martinos Center for Biomedical Imaging Irving Bigio **Boston University David Boas Boston University**

Article

Keywords:

Posted Date: March 21st, 2024

DOI: https://doi.org/10.21203/rs.3.rs-4014687/v1

License: © ④ This work is licensed under a Creative Commons Attribution 4.0 International License. Read Full License

Additional Declarations: (Not answered)

1		
23	Title	Enhanced Multiscale Human Brain Imaging by Semi-supervised Digital Staining and
4		Serial Sectioning Optical Coherence Tomography
5		
6 7	Autho	
8	Autilo	Shiyi Cheng ^{1†} , Shuaibin Chang ^{1†} , Yunzhe Li ^{2†} , Anna Novoseltseva ³ , Sunni Lin ^{1,3} , Yicun
9		Wu ⁴ , Jiahui Zhu ¹ , Ann C. McKee ^{5,6,7,8,9} , Douglas L. Rosene ¹⁰ , Hui Wang ¹¹ , Irving J.
10		Bigio ^{1,5,12} , David A. Boas ^{1,5,12} , Lei Tian ^{1,5,12} $^{\circ}$
12	Affilia	ations
13 14		¹ Department of Electrical and Computer Engineering, Boston University, 8 St Mary's St, Boston, MA, 02215, USA.
15 16		² Department of Electrical Engineering and Computer Sciences, University of California, Cory Hall, Berkeley, California, 94720, USA.
17 18		³ Department of Biomedical Engineering, Boston University, 44 Cummington Mall, Boston MA, 02215, USA.
19 20		⁴ Department of Computer Science, Boston University, 665 Commonwealth Ave, Boston, MA, 02215, USA.
21 22		⁵ Boston University Alzheimer's Disease Research Center and CTE Center, Boston University, Chobanian and Avedisian School of Medicine, Boston, MA, 02118, USA.
23 24		⁶ Department of Neurology, Boston University, Chobanian and Avedisian School of Medicine, Boston, MA, 02118, USA.
25 26		⁷ VA Boston Healthcare System, U.S. Department of Veteran Affairs, Jamaica Plain, MA, 02130, USA.
27 28		⁸ Department of Psychiatry and Ophthalmology, Boston University School of Medicine, Boston, MA, 02118, USA.
29 30		⁹ Department of Pathology and Laboratory Medicine, Boston University School of Medicine, Boston, MA, 02118, USA.
31 32		¹⁰ Department of Anatomy & Neurobiology, Boston University Chobanian & Avedisian School of Medicine, Boston, Massachusetts, USA.
33 34		¹¹ Athinoula A. Martinos Center for Biomedical Imaging, Department of Radiology, Massachusetts General Hospital/Harvard Medical School, Charlestown, MA, 02129, USA
35		¹² Neurophotonics Center, Boston University, Boston, MA, 02215, USA.
36		[†] These authors contributed equally to this work.
37 38 39		*Email: leitian@bu.edu *Telephone: (617)353-1334

40 Abstract

A major challenge in neuroscience is to visualize the structure of the human brain at 41 different scales. Traditional histology reveals micro- and meso-scale brain features, but 42 43 suffers from staining variability, tissue damage and distortion that impedes accurate 3D 44 reconstructions. Here, we present a new 3D imaging framework that combines serial 45 sectioning optical coherence tomography (S-OCT) with a deep-learning digital staining 46 (DS) model. We develop a novel semi-supervised learning technique to facilitate DS model training on weakly paired images. The DS model performs translation from S-OCT to 47 48 Gallyas silver staining. We demonstrate DS on various human cerebral cortex samples with 49 consistent staining quality. Additionally, we show that DS enhances contrast across cortical 50 layer boundaries. Furthermore, we showcase geometry-preserving 3D DS on cubic-51 centimeter tissue blocks and visualization of meso-scale vessel networks in the white matter. 52 We believe that our technique offers the potential for high-throughput, multiscale imaging 53 of brain tissues and may facilitate studies of brain structures.

56 Introduction

54 55

57 The human brain consists of an estimated 86 billion neurons (1), which form intricate 58 connections and networks that underlie the complex functions. To gain new insights into 59 the brain, major efforts have recently been made to develop multiscale imaging technologies for visualizing anatomical structures with microscopic resolution across cubic centimeters 60 of tissue. The most widely used techniques for visualizing anatomical and neuronal 61 62 structures are based on histological staining. Gallyas silver staining is used to characterize 63 myelin content and neuronal structures, as well as to identify pathological features of neurodegenerative diseases in human brain tissue (2, 3). To create a high-resolution 3D 64 65 model of the cytoarchitecture, the BigBrain project (4) reconstructed a whole human brain with more than 7000 histological sections, which involves slicing the tissue into 20-um 66 sections, staining with silver halide to reveal cellular and fiber structures, and registering 67 68 the slices in 3D. However, these histological staining processes are generally complex, labor-intensive, time-consuming, and prone to experimental error and staining variability. 69 Furthermore, the slicing, mounting, dehydration, and staining inevitably cause tissue 70 71 damage and slice-specific distortions, which can limit the accuracy of 3D alignment and 72 reconstruction of structures at the micron scale (5, 6). Therefore, there is a growing need for 73 developing 3D pathology imaging techniques, especially label-free techniques that can 74 provide high-resolution 3D visualizations of brain tissues with minimal tissue damage and 75 distortion, and that can reduce the need for physical staining (PS) (7-10).

Optical coherence tomography (OCT) is a label-free imaging technique that allows high-76 77 resolution 3D visualization and quantification of intrinsic optical properties of tissue, such 78 as the scattering coefficient and back-scattering coefficient (11, 12). Recently, OCT has 79 shown great promise in brain imaging applications, such as visualizing single neurons (13), fiber tracts (14), and the laminar structure of the cerebral cortex in the human brain (15, 16). 80 81 While traditionally limited by light penetration, serial sectioning OCT (S-OCT) integrates 82 OCT with a vibratome slicer to enable 3D imaging of cubic centimeters of tissue (17). S-83 OCT permits straightforward and accurate 3D high-resolution reconstruction of large-scale 84 brain anatomy, microstructures, and tractography (17-19) with minimal tissue distortion. This is achieved through the use of a serial imaging protocol (20), where OCT imaging of 85 86 the top $\sim 150 \,\mu m$ thick tissue is alternated with the slicing off of the superficial tissue, thus 87 reducing cutting-induced distortion after imaging. This enables accurate reconstruction of 88 the complex 3D structures of brain tissues without requiring sophisticated inter-slice

registration algorithms. Despite its ability to routinely generate large-scale volumetric brain
imaging data, S-OCT still requires considerable expertise to identify and annotate
anatomical and neuronal features for further analysis (11, 14, 17, 21). Our goal is to augment
S-OCT with a digital staining (DS) technique that enables straightforward 3D histology on
large-scale human brain tissues.

94 In the past few years, deep learning methods have revolutionized the field of DS, which 95 aims to transform label-free images into histological staining-like images using a 96 computational model (22). DS offers a fast and low-cost alternative to conventional PS 97 methods. Several DS models have been developed that transform different pairs of inputoutput imaging modalities. However, most existing DS methods rely on supervised learning 98 99 methods, which requires paired images of the tissue slice with and without staining for 100 model training. To ensure accurate DS results, cross-modal registration between the image 101 pairs with pixel-level accuracy is crucial (23–26). However, obtaining such image pairs is difficult and often involves sophisticated image registration procedures (22, 24). To 102 overcome this challenge, some recent studies have explored unsupervised image translation 103 104 models for DS, which only need unpaired collections of images from the two modalities for model training (8, 27-30). The most popular unsupervised method is CycleGAN (31), 105 which comprises two sets of generators and discriminators that enforce cycle consistency 106 107 and content preservation for the image translation task. A recent improvement over CycleGAN is Contrastive Unpaired Translation (CUT) (32), which uses contrastive learning 108 109 to achieve better structural and content preservation with only one set of generator and discriminator, and has demonstrated superior performance in DS tasks (28). However, these 110 unsupervised models still lag behind supervised models in terms of accuracy (22). 111

Here we present a new *semi-supervised* learning framework for DS using a *limited amount* 112 of weakly paired image data. As a proof-of-concept demonstration, we use our DS model 113 to translate S-OCT images to Gallyas silver staining. Our DS model consists of two novel 114 modules that address several challenges in our technique. Our main model is based on the 115 116 CUT framework to perform DS using unpaired training data. This module combines contrastive learning and adversarial learning to address the lack of paired imaging data since 117 the physically stained images were obtained from unordered adjacent brain tissue sections 118 to the OCT-imaged sections and were confounded by tissue damage and distortion during 119 the staining process. 120

To improve the accuracy of the unsupervised model, we augment it with semi-supervision 121 from two auxiliary tasks. Firstly, we devise a pseudo-supervised learning module by training 122 the DS network on a pseudo-paired training dataset that is generated using our previously 123 124 established biophysical model. Our previous work has revealed a linear correlation between 125 the OCT scattering coefficients (SC) and the optical density (OD) computed from the Gallyas silver stained image (21). Based on this similarity prior, this module learns to 126 127 translate the generated OD back to the Gallyas silver stain, acting as a proxy supervision for 128 learning the translation from OCT-SC to Gallyas silver stain. This naturally pixel-aligned 129 pseudo supervision augments the training data, enabling training the DS model effectively despite the limited data available to our task due to the scarcity of the human brain samples. 130 Additionally, when combined with the adversarial learning component in the CUT 131 132 backbone, the domain gap between the OCT-SC images and OD maps are effectively mitigated by the mechanism of domain-adversarial training (33). Secondly, we develop an 133 134 unsupervised cross-modality image registration module that aligns the adjacent Gallyas image with the OCT-SC image. This module enables the DS model to utilize the geometric 135

similarity information provided by the adjacent slices, thereby guiding the image translation
process. To train the registration network effectively, we introduce a novel two-stage,
multiscale training strategy. It allows the network to learn image registration at the "global"
whole slide image (WSI) scale, while simultaneously learning image translation at the
"local" image patch scale. Furthermore, this novel training strategy facilitates collaborative
training between the DS model and the registration model, leading to more effective
enforcement of high-quality DS results.



Fig. 1. Overview of the proposed OCT DS technique. (A) Data acquisition and DS model.
 S-OCT alternates between 3D imaging and tissue sectioning to acquire a stack of block-face
 OCT images, which are then processed to compute the scattering coefficient (OCT-SC) map
 stack. Sectioned sample slices are physically stained and imaged. The DS neural network is
 trained from a few weakly-aligned pairs of OCT-SC and Gallyas silver-stained images. (B)
 After the DS model is trained, it can perform inference on completely new slices of OCT SC images for volumetric DS.

We present our DS pipeline for data acquisition and deep learning model training in Fig. 151 1A. We use S-OCT to obtain label-free volumetric data of human brain samples. We then 152 process the OCT data to calculate the SC maps (11) (see details in Methods). Next, we 153 154 develop a deep learning DS model that transforms OCT-SC images into Gallyas silver stain images. We choose OCT-SC as the input for the DS model instead of the raw OCT 155 measurements because SC measures the intrinsic optical properties of the tissue and 156 eliminates the inhomogeneity in the raw OCT intensity by using a nonlinear model-fitting 157 process (11). Moreover, a biophysical model from our previous work showed that OCT-SC 158 mainly depends on the contribution of myelin content, which is captured by the OD of the 159 160 Gallyas silver staining (21). We hypothesize that the correlation between these two modalities can be leveraged to create a more accurate image-to-image mapping using a deep 161 learning model. During S-OCT, we also collect a few unordered tissue slices that are 162 163 physically stained for DS model training and evaluation. The deep learning model is trained on a few weakly-aligned pairs of OCT-SC and Gallyas silver stained WSIs. The inference 164 stage of the DS model is shown in Fig. 1B. After the model is trained, it can be applied on 165 166 any OCT-SC maps to enable 3D neurohistology on cubic centimeters of brain tissue and visualize mesoscopic brain structures. 167

First, we present the OCT DS results on single-section tissues from various cerebral cortex
samples and compare them with PS results from adjacent sections. We demonstrate that DS
exploits the quantitative nature of OCT-SC and thus can produce consistent staining quality

- 171 across different samples. Compared to PS, DS reveals comparable mesoscopic (~10 µm) structures in different tissue regions without introducing staining variability across samples 172 173 and experiments. In addition, we show that DS enhances contrast across cortical layer 174 boundaries and can consistently differentiate cortical layers IV, V and VI. Next, we show a 3D-rendered volumetric DS result on a cubic centimeter-scale tissue block that was not used 175 176 for training the DS model. The result shows geometry-preserving 3D staining on large-scale 177 brain tissue and visualization of vessel structure in the white matter region. Finally, we 178 showcase a pilot study on the generalization performance of our method - we apply the DS 179 model trained on cortex regions to samples from other anatomical regions acquired from 180 different OCT setups.
- In summary, we present a novel deep learning technique for DS of OCT images for large-181 scale human brain imaging. Our method allows direct visualization of important mesoscopic 182 183 3D brain features, including myeloarchitecture of the cerebral cortex and main 3D blood vessel network in the white matter, with contrast that closely resembles Gallyas-silver 184 staining. Our method has several advantages over traditional PS, such as reducing staining 185 186 variability, preserving complex brain 3D geometry and facilitating volume generation across cubic centimeters of tissue. Our method also improves the interpretability of the 187 188 label-free OCT modality for brain imaging. However, our method also faces some 189 limitations that originated from our current S-OCT system, such as artifacts from image 190 stitching (12, 14), uneven tissue sectioning, speckle noise, and limited lateral and axial 191 resolution due to the SC model fitting. Although our technique is sensitive to fiber structures 192 in the gray matter, the speckle noise and limited resolution resulted in discontinuities and 193 grainy artifacts in the DS results. We expect that these issues will likely be overcome by future generations of high-resolution S-OCT systems (34, 35) and improved processing 194 195 algorithms. Despite current limitations, we believe that our semi-supervised learning-based 196 DS framework is broadly useful to other bioimaging modalities and DS applications. 197 Furthermore, our work has significant implications for quantitative volumetric 198 neuropathology. The integration of DS techniques with S-OCT has great potential for high-199 throughput, multiscale human brain imaging. The data generated from this technique could 200 help better understand the meso- and micro-structure of brain tissues and their role in disease development, and ultimately enhance our knowledge of the brain's structure and function. 201

202

203 204

Results

205 206

Digital staining by semi-supervised learning using weakly-paired images

We formulate the DS task as a weakly-paired image translation problem because we do not have access to pixel-aligned image pairs of OCT-SC and PS images. To achieve better performance than fully unsupervised methods, we exploit the side information provided by the structural and content similarity between the adjacent sections in the imaging data, as well as a biophysical model for linking OCT-SC and the contrast in Gallyas silver stain in a semi-supervised deep learning framework.



213
214
215
216
217
218
219
220
221
222
223

224

Fig. 2. The training framework of our DS neural network model. (A) The backbone of the DS network *G* is built on the CUT framework, which combines contrastive learning and adversarial learning. The input is a 2D OCT-SC map *X* and the output is a digitally stained image G(X) that is compared with a PS image *Y* from an adjacent slice. (B) Auxiliary pseudo-supervised learning task. The biophysical module computes the optical density OD(Y) of the PS image *Y*, which is fed as an input to *G*. The digitally stained OD image G(OD(Y)) is compared with the original PS image *Y* during training. (C) Auxiliary unsupervised cross-modality image registration task. We alternate between optimizing *G* and a registration network *R* under different image scales. We fix *R* while updating *G*, which provides more informative supervision for *R* in the next iteration. We use patch-wise losses for training *G*, and whole slide image (WSI) losses for training *R*.

- 225 The training framework of our DS network consists of several novel learning components, as shown in Fig. 2. Based on the CUT framework as the backbone (32), the DS model uses 226 a mix of adversarial loss and contrastive loss in the unpaired image setting, as shown in Fig. 227 228 2A. The adversarial learning measures the perceptual similarity of the generated DS images and the PS images. It tries to reduce the gap between the high-dimensional distributions of 229 230 the DS and PS images such that the generated DS images are perceptually indistinguishable from the PS images. The contrastive loss uses self-supervised patch-wise learning to ensure 231 232 structural consistency between the OCT-SC and DS images. It maximizes mutual information and provides self-guidance for content preservation. The combination of 233 234 contrastive loss and adversarial loss enables high-quality DS images that preserve the 235 content and structures of the OCT-SC images.
- 236To improve upon the unsupervised CUT framework, we propose a semi-supervised learning237method. Our method leverages augmented pseudo pairs generated by a biophysical model238and registered cross-modality image pairs that are dynamically adjusted by a learnable239registration network. The intuition is that using additional auxiliary supervision enhances240the learnability, efficiency and accuracy of the model compared to unsupervised learning.241Crucially, our semi-supervised method does not require any exact paired PS and OCT-SC242images during training.
- In Fig. 2B, we introduce the pseudo-supervised learning auxiliary task to enhance the 243 unpaired image translation for DS of OCT-SC images. We first compute the OD maps from 244 the PS images and then utilize the OD - PS image pairs to train the DS model in a pseudo-245 supervised manner. This approach proves effective because the OD image exhibits similar 246 image contrast and feature distribution as the OCT-SC across various cortical regions. 247 Additionally, the OCT-SC demonstrates an approximate linear relationship with the OD of 248 the Gallyas silver stain (21). Furthermore, since the OD map is naturally pixel-aligned with 249 the PS image, it facilitates supervised learning and provides additional semi-supervision and 250 alignment constraints for the main DS model. However, the inherent disparities in image 251 252 features and intensity value distributions between the OD map and the OCT-SC image result in a domain gap, which limits the accuracy of the trained DS model when relying solely on 253 this auxiliary task. Our insight is that when this task is combined with the adversarial 254 255 learning component in the CUT backbone, it enables domain adaptation similar to the domain-adversarial training framework (33). The performance on the OCT-SC image is 256 ensured by penalizing the perceptual differences between the DS images generated from the 257 258 OCT-SC image and the OD map using the adversarial loss. By leveraging both the pseudosupervised learning and adversarial learning components, we effectively bridge the domain 259 gap and improve the accuracy of the DS model for OCT-SC image translation. 260
- In Fig. 2C, we illustrate the second auxiliary task for aligning the PS image, the OCT-SC 261 image, and the DS image using a registration network. This registration module undergoes 262 two training stages: pre-training and fine-tuning. During the pre-training stage, the 263 264 registration module operates on the WSI scale. It predicts a deformation field that indicates the pixel-wise displacement vectors required for non-rigid transformation. To facilitate 265 cross-modal self-supervised registration, we utilize the OD map as a surrogate for the OCT-266 SC image and learn a deformation field between the OD map and the input OCT-SC image. 267 268 This result is used as an initial estimate for the deformation between the PS image and the matching OCT-SC image. By leveraging our biophysical model, we bootstrap the 269 270 challenging self-supervised cross-modality image registration problem in this pre-training stage. The subsequent fine-tuning of the registration model aims to provide pixel-wise 271

- 272 weak-supervision for the DS model. In this stage, we employ an alternate training approach that involves collaborative learning between the DS model and the registration model. When 273 the DS model is fixed, the registration model is trained at the WSI scale to address global 274 geometry correction. When the registration model is fixed, the DS model is trained at the 275 image patch scale to provide sufficient samples for local translation learning. This 276 277 unsupervised cross-modality image registration module enables the DS model to learn improved local color tone mapping from unaligned imaging modalities without the need for 278 279 explicit supervision.
- 280 Overall, our DS framework augments unpaired image translation with pseudo supervised learning and unsupervised cross-modality image registration. The total loss function used 281 for training is the weighted sum of the four objectives derived from the main image 282 283 translation task and two auxiliary tasks. Our method achieves superior performance over 284 other baseline methods, including CycleGAN, CUT and FastCUT in terms of DS quality and accuracy, as shown in Supplementary Materials (SM) Section 1, Section 2, Fig. S1 and 285 Fig. S3. Additional details about the network structure, training procedures and quantitative 286 evaluations are described in Methods and SM Section 3, 4, 9 and 10. 287

Digital staining enhances mesoscopic brain structures and provides high staining uniformity

- We present the ability of our DS technique to preserve the mesoscopic brain structures and achieve uniform staining of cerebral cortex sections from post-mortem human brains. We use two groups of PS imaging results as comparative references: one group consists of WSIs of well-stained sections, and the other group consists of WSIs of less-ideally-stained sections.
- In Fig. 3A, we present the OCT-SC, DS, and well-stained PS images of adjacent sections 295 from the human cerebral cortex, arranged from left to right. The DS images show that our 296 technique can accurately capture various brain structures that match the PS images, such as 297 298 cortical layers, myelin fibers, and vessel blobs. The DS and PS images share similar 299 contrast, with white matter (WM) regions appearing as dark brown or black and gray matter (GM) regions appearing as white, while the OCT-SC image has the opposite contrast. 300 Within the gray matter, the infra layers also appear to be darker than supra layers, consistent 301 with the PS images. These correspondence in mesoscale structures validate that our DS 302 model can reliably and accurately learn this general inverse mapping between OCT-SC and 303 304 PS images.
- In the zoom-in regions, we present the images on different types of cortex regions, including 305 gyral crest regions marked as 1 and 3 and sulcal fundus regions marked as 2 and 4, from the 306 three modalities: OCT-SC, DS and PS. In region 1, the structures of radial myelin fiber 307 308 bundles at scales of about 10-20 µm are shown as dark brown tubular features in both DS and PS images, especially in the GM region. By comparing OCT-SC and DS images, we 309 310 can see that the image content is consistent, which indicates that the ability of resolving fine features is primarily limited by the input OCT-SC data. Despite the limitations of resolution 311 and speckle noise in the OCT data, the orientation of fiber bundle traces and the intensity 312 313 distribution according to cortical layers can still be discerned in the DS results. Similar 314 patterns are also evident in zoom-in regions 3 and 4, where the local intensity variation is visible in the GM regions, although the fiber bundles are less distinct in OCT-SC and DS 315 316 images than in the PS images. In region 2, the supra cortical layers (I-III), infra layers (IV, V, VI) and WM are clearly distinguished by the white, light brown, and dark brown bands, 317



Fig. 3. DS results on OCT-SC of tissue slices and comparisons with PS images. Cases include (A) ideal staining samples; (B) non-uniform staining and understanding samples. ROI 1, 3, 5, 7 are gyral crest regions and 2, 4, 6, 8 are sulcal fundus regions.VS: "vessel space". Scale bars are 1 mm.

respectively. The black line structure near the top of the PS image indicates smaller vessels, which are also visible in the DS image at the same locations. The zoom-in regions 1, 2 and in PS show small white blob or tubeness features especially in the WM regions. In PS, these white blobs represent the empty space previously occupied by vessels which are lost

318 319

320 321

322

due to slicing and washing steps during staining. In contrast, the white blobs in DS images
primarily represent the space within vascular walls and perivascular space which appear
smaller since no slicing or physical staining is performed on OCT-SC images. Those
features are generally referred to as VS ("vessel space") in Fig. 3. These visualizations
demonstrate that our DS model can faithfully reveal ~20 µm scale brain structures.

A major advantage of DS over PS is stain uniformity. To demonstrate this, we present three 332 333 types of images in Fig. 3B from the less-ideal PS group that comprises most of our PS data. One inherent limitation of traditional histological staining is the variability across different 334 335 sample regions and experiments. Despite our careful sample preparation and staining procedures, the staining result is influenced by many confounding factors of the chemical 336 reaction and uniformity of the staining quality is challenging to ensure. In Fig. 3B, the 337 rightmost column of the first row shows a PS example with over- and non-uniform staining 338 339 (in particular along the vertical directions); the second row shows a PS example with understaining. 340

- We select two gyral crest regions (marked as 5 and 7) and two sulcal fundus regions (marked 341 as 6 and 8) to provide in-depth analysis. The PS images in regions 5 and 6 are over-stained, 342 while the PS images in regions 7 and 8 are under-stained. In region 5, the DS and OCT-SC 343 344 images show clear ridges corresponding to cortical layer V, but the PS image shows a dark brown shade due to over-staining. In region 6, which is a sulcal fundus region with less 345 visible cortical layers, the DS image shows a clear boundary between WM and GM regions, 346 but the PS image shows an ambiguous boundary. Small vessel blobs are also more visible 347 in the DS image than in the PS image. In region 7, which is a gyral crest region, the DS 348 image shows dark ridge features corresponding to cortical layer IV and V, but the PS image 349 does not show these features due to under-staining. Additional examples are shown in SM 350 351 Section 5 and Fig. S4.
- The superior stain uniformity demonstrated by our DS results across different sections is enabled by the OCT-SC that extracts a normalized quantity based on a physics model that reflects the intrinsic property of the brain tissue. This stain uniformity will be a great advantage during anatomical and pathological evaluations. A limitation of our current OCT-SC curve fitting model is that it reduces the spatial resolution (lateral: 6 µm raw OCT measurement, 12 µm fitted SC map; axial: 6 µm raw OCT measurement, 150 µm fitted SC map), which limits the ability to resolve fine fiber structures.

359 Digital staining enables reliable cortical layer differentiation and layer thickness 360 quantification

- We demonstrate the capability of DS-OCT to reliably distinguish cortical layers with comparable or even better sensitivity than PS, thanks to the uniform DS quality as discussed before. We identify cortical layers IV, V and VI by the displayed fiber density (*36, 37*), since these layers are more prominent than layers I, II and III in most of our samples. We provide additional examples of DS layer visualization and compare them with well-stained and less-ideal stained PS samples in SM Section 6 and Fig. S5. We also show how the layer thickness can be consistently quantified in our DS images.
- Figure 4A shows the WSIs of the DS result and the reference PS of an adjacent brain slice. The DS image clearly reveals the curved double-band structures above the WM region, which are stained in dark brown. These features indicate higher myelin fiber density that are characteristic in cortical layer IV and V (*37*). Consistent image contrast variations for

the laminar structures are observed in the DS result. In contrast, the double-band structures are less visible around some of the gyral regions and the contrast is less distinct in the PS image. Figure 4B shows zoom-ins from a gyral crest region and a sulcus region of the three modalities, corresponding to the regions marked by the green box and red box in Fig. 4A respectively. The OCT-SC and DS images have a strong correlation in their intensity variations. The DS image consistently shows the double-band features in the GM region, while the PS image often fails to reveal them due to over- or under-staining.



Fig. 4. Comparisons results of layer differentiation and thickness estimation in DS results. (A) The DS and PS WSIs from a cortex tissue section. (B) Zoom-in ROIs of inverted OCT-SC, DS and PS modalities marked in green and red boxes in (A) and normalized intensity profiles aggregates along white dotted lines. (C) Manually annotated layers IV/V/VI labeled in three colors and estimated local thickness. Statistics of thickness are visualized in box plot and grouped by gyral crest and sulcus regions. ROI is the zoom-in of the dotted blue box from (A).

Next, we demonstrate the improved contrast between cortical layers in DS by plotting the 387 388 average intensity (across the three color channels) along the white dotted lines in Fig. 4B. The right panel shows the normalized profiles over a 3.5-mm depth range, where blue, green 389 and red represent OCT-SC, DS and PS modalities, respectively. We manually marked the 390 391 boundaries of layer IV, V and VI with dotted vertical lines in four different colors. In both gyrus and sulcus regions, the DS profiles show the highest contrast (measured by the 392 difference between the maximum and minimum values) in layer IV and V among the three 393 394 modalities, which facilitates identifying the layer boundaries. When comparing OCT-SC

372

395and PS with DS, the DS enhances the intensity variations at the boundary between layer IV396and V. This reduces any confusion when distinguishing between these two layers.397Comparing the profiles between OCT-SC and DS in different layers suggests that our DS398model works beyond our approximate linear biophysical model (21) and increases the local399contrast by a nonlinear mapping function expressed by our neural network.

In Fig. 4C, we further demonstrate straightforward segmentation and thickness 400 401 quantification of cortical layers IV, V and VI using our DS result (see details in Methods), which can provide valuable information for many neuropathological studies (17, 38, 39). 402 403 The top panel shows the zoom-in region of the dotted blue box in Fig. 4A, where we manually labeled the boundaries of the three cortical layers. We estimated the layer 404 405 thicknesses from the binary mask obtained from cortical layer segmentation using an 406 algorithm from our previous work (17). We chose two gyral crest regions and a sulcus region indicated by the white boxes in the binary mask image. The bottom panel displays 407 the box plot of the local layer thickness statistics in gyrus and sulcus regions. We observed 408 a similar pattern of variation in layer thickness for layer IV, V and VI in the sulcus, gyrus 409 410 and the entire cortical regions. The median local thickness of layer IV, V and VI were 300 μm, 540 μm and 480 μm respectively. We also observed a significant reduction in layer 411 thickness in all three layers in the sulcus regions compared to the gyrus regions, in 412 413 agreement with the literature (40, 41). The median thickness of layer IV, V and VI were 410 μm, 630 μm and 580 μm respectively in the gyrus regions, and were 250 μm, 370 μm and 414 310 µm respectively in the sulcus regions. 415

416 Volumetric digital staining on cubic centimeter-scale brain tissue

Next, we showcase volumetric staining on cubic centimeter-scale brain tissue enabled by
our technique that combines S-OCT and DS. Our technique significantly reduces tissue
distortion and misalignment during the 3D reconstruction process suffered by the traditional
3D pathology technique. We demonstrate 3D DS on a 4 cm × 5 cm × 1.2 cm brain tissue
block that was not used for training our DS model. We show that our method can preserve
the intricate 3D brain structures in both GM and WM regions. Moreover, we visualize the
3D vessel network in the WM.

424 In Fig. 5A, we present a 3D visualization of the DS output on the whole tissue block in the top panel. The DS model takes as input a z-stack of around a hundred slices of OCT-SC 425 images. Each OCT-SC slice, which has a size of 4 cm \times 5 cm, is processed separately and 426 427 fed to the DS model. The DS output images are then directly stacked along the z-axis to 428 create the digitally stained volume. Consistent with the 2D results, the 3D DS volume generates white and dark-brown colors that correspond to GM and WM regions 429 430 respectively. We can also observe a smooth transition of these GM and WM boundaries 431 along the z direction, which reflects the preservation of 3D geometries of the brain structures. In Fig. 5B, we display several orthogonal cross-sectional views of the DS 432 volume. The overall color tone and contrast variations match with the 2D results in Fig. 3. 433 Small white blobs and tubes within the WM region indicate the vessel space. These results 434 435 are consistent with 2D DS results that have been verified with PS references, and partly confirm the generalization ability of our DS model on unseen large-scale brain samples. 436 437 Moreover, the X-Z cross section also shows several continuous features along the depth, 438 such as intricate brain folding structures, double-band cortical layers, and small tubular vessels. This again illustrates the 3D geometry preservation feature of our DS technique. 439



Fig. 5. 3D visualization and cross-sections views of the DS results on a large unseen tissue block. (A) The DS output images are stacked along the z-axis to render the whole digitally stained volume as well as segmented WM regions. (B) Orthogonal cross-sectional views of the DS volume. (C) Two zoom-in regions of vessel structures in yellow and green boxes from (A) are shown on the left. Three orthogonal maximum intensity projections (MIP) of the DS volume are shown on the right. All scale bars are 5 mm.

447 To further illustrate the ability of our DS technique to preserve the 3D geometry of mesoscale brain structures, we present a 3D visualization of a centimeter-scale network of 448 449 vessel space which is not visible in 2D PS images. Besides the GM and WM contrast, our DS volume also shows several continuous white tubular structures corresponding to blood 450 vessels in the top panel of Fig. 5A. In the bottom panel of Fig. 5A, we show the segmented 451 452 DS volume displaying only the WM region, where the white tubular structures are more 453 prominent and not masked by the GM. In Fig. 5C, we highlight two regions in yellow and green boxes. The vessel spaces in those regions are rendered with more transparency and 454 reveal the branching and connectivity of the vessel network. On the right panel of Fig. 5C, 455 three orthogonal maximum intensity projections (MIP) of the DS volume further 456 demonstrate the preservation of the 3D vessel structures. We note that the axial continuity 457 of our DS volume is currently limited by the axial resolution (150 µm) imposed by our SC 458 fitting model, which we aim to improve in the future. Being able to image brain samples as 459 large as $4 \times 4 \times 1$ cm³ (34), we can easily extend the aforementioned analysis to large brain 460 areas with uniform and enhanced contrasts, which could greatly improve the throughput of 461 brain anatomy study. 462

463 Generalization to unseen anatomical regions

440

464 To further demonstrate the generalization capability of our trained DS model, we conducted 465 a pilot study on different anatomical regions that were imaged by a different S-OCT setup 466 not seen during training. We used the same fitting model to generate the OCT-SC image in 467 Fig. 6, which shows a sample from the hippocampus region acquired by a different S-OCT 468 setup. Since our SC fitting model extracts an intrinsic tissue property and is relatively 469 insensitive to variations in hardware platforms and sample conditions, it ensures the 470 robustness of our DS method. The DS image is inferred by directly inputting the OCT-SC 471 to the previously trained model. Figure 6 shows the OCT-SC and DS images, and the reference PS image of an adjacent section from left to right. We roughly aligned the field of 472 views of the DS and PS images using a rigid transformation. On a large scale, the DS process 473 474 successfully transforms the image contrast to match the anatomical structures found in the PS image. By comparing with the anatomy of hippocampus (42), we can identify the alveus 475 476 (AL) and/or fimbria fomix (FF) layer at the top, the stratum pyramidale (SP) layer beneath them, and the stratum radiatum (SR), stratum moleculare (SM) and the dentate gyrus (DG) 477 478 layers that encase the Cornu Ammonis areas (CA1-CA4) of dense neurons. Importantly, in 479 CA1-CA4 areas, we found bright spots in OCT-SC images, which are transformed to brown 480 spots in the DS images. These structures correlate strongly with the brown spots seen in the PS image and are likely individual neuron somas. More examples of generalization results 481 can be found in SM Section 7 and Fig. S6. 482



Fig. 6. DS-OCT generalization performance on a hippocampus tissue slice. Examples
of OCT-SC, DS and PS images (of adjacent sections) on one sample from the Hippocampus
region are shown. SP: Stratum Pyramidale; AL: Alveus; FF: Fimbria Fomix; SR: Stratum
Radiatum; SM: Stratum Moleculare; DG: Dentate Gyrus; CA: *Cornu Ammonis*.

488 Such generalization agrees with our previous work that discovered a universal correlation 489 between optical scattering and myelin density across the human brain (21). This suggests 490 that a DS-OCT model, even if trained on limited regions of the human brain, may be 491 effectively employed in other unseen regions. This significantly decreases the training effort 492 compared to those that rely on transfer learning.

493

494 **Discussion**

495 In summary, we developed a novel semi-supervised learning technique for DS of OCT 496 images for large-scale volumetric visualization and analysis on human brain tissue samples. 497 Our technique works by integrating label-free S-OCT imaging and an advanced deep

497 Our technique works by integrating label-free S-OCT imaging and an advanced deep

498 learning DS model. The S-OCT enables imaging of cubic centimeter-scale brain tissues and 499 preserves complex 3D tissue geometry across sections. Our semi-supervised learning 500 method bypasses the need for paired unstained and stained images and can achieve high-501 quality DS using a limited amount of weakly paired image data for model training. Our deep learning model is built on an unsupervised CUT model backbone, which is augmented with 502 503 two auxiliary tasks. The pseudo-supervised module reduces the data requirement for model training by exploiting the correlation between the OCT-SC and the OD of Gallyas silver 504 505 stain. The unsupervised cross-modality image registration module exploits the structural 506 information between the adjacent tissue sections. By working with a fitted tissue property, 507 namely the SC, from the raw OCT measurement as the input to the deep learning model, it greatly enhances the uniformity and generalizability of the DS results. This is highlighted 508 509 by our volumetric DS result on cubic centimeter-scale brain tissue block and on unseen anatomical regions from different OCT systems. We believe our OCT DS technique is a 510 511 promising solution for large-scale human brain imaging for comprehension characterization 512 of brain structure across scales.

513 We envision that our deep learning framework holds great potential for a wide range of applications in the field of DS. There is a growing demand for exploring semi-supervised 514 learning approaches to effectively harness the wealth of information contained in unpaired 515 516 or weakly paired biomedical images. Obtaining pairs of images with labels and without labels can be a challenge in many biomedical contexts. However, it is often easier to obtain 517 images of samples with slight distortions or adjacent sections. To leverage these types of 518 519 datasets, our method leveraged a novel inverse mapping technique, going from stained images to label-free modalities, and generated pairs of images that were pixel-aligned to 520 serve as augmented supervision. Furthermore, we introduced a novel cross-modality 521 522 registration algorithm to correct for sample distortions and account for the complex geometries of the samples. As a result, our enhanced semi-supervised learning framework 523 facilitates more straightforward training on datasets that may be naturally acquired from 524 525 routine staining experiments, even when those datasets are only weakly paired. In essence, incorporating semi-supervised methods can significantly enhance the efficiency of the "data 526 collection-training-validation" cycle in the development of digital staining models. 527

528 We discuss some of the main limitations that affect the quality of S-OCT images and the DS method based on them. The first limitation is the data processing pipeline of OCT 529 imaging. Coherent scattering results in speckle noise, which manifests as randomly 530 531 distributed fine-grained dark or white spots in OCT and the fitted SC images. These speckle artifacts do not necessarily correspond to the actual cortical structures in PS images, as 532 533 shown in Fig. 3 and SM Fig. S4. Consequently, visualizing and digitally staining small 534 vessels, capillaries, and fine axonal fiber structures become challenging. Moreover, the current resolution of our OCT-SC data is insufficient to resolve delicate structures like 535 single neurons. To address this limitation, a possible future direction is to optimize the 536 537 processing pipeline of OCT-SC with deep learning techniques to achieve higher imaging quality. For example, self-supervised learning algorithms for speckle suppression can be 538 developed by utilizing a customized blind-spot denoising network and a speckle statistics 539 model (43). Enhancing the resolution of SC can be explored by employing a deep learning 540 model similar to (44) to learn a more accurate and robust fitting model without the need for 541 542 local-averaging. These improvements can increase the robustness and resolution of our 543 method, enabling us to capture finer neuronal structures. The second limitation pertains to 544 stitching artifacts that cannot be fully corrected in our current DS model, thereby affecting the quality of WSI image, as observed in Fig. 3, 4 and SM Fig. S6. To address this issue, it 545

- 546 may be possible to incorporate a structural prior constraint into our DS training framework, which will potentially yield better correction of these artifacts. The last limitation involves 547 the imperfect registration component in our DS model. The fitting depth range we utilized 548 549 for SC (150 µm) is larger than the physical sectioning thickness of PS images (50 µm). Furthermore, during staining experiments, sample destruction may occur, introducing 550 551 imaging content mismatch. However, our registration learning only corrects for global-scale 552 geometric distortion between adjacent sections and does not account for potential content 553 mismatch between weakly-paired images. Consequently, the registration process fails to 554 generate pixel-aligned image data, as seen in SM Fig. S2. To tackle this issue, further 555 improvements to the deep learning framework may consider methods to address content 556 mismatch.
- 557 It is worth noting that our training and testing images comprise a mix of normal control and 558 neurodegenerative human brain samples, which hinders the model's ability to learn the distinctions between normal and diseased brain images. To expand our work towards 559 distinguishing between normal and diseased cases, one needs to acquire images from a 560 561 larger set of brain samples for both conditions. Additionally, we plan to incorporate multimodality input, such as polarization information, into our DS model to increase the imaging 562 sensitivity to birefringence structures, including myelin fibers (17, 19). Another promising 563 564 modality we aim to integrate with the S-OCT is two photon microscopy, which allows imaging of small vessels and myelin fibers based on autofluorescence contrast with reduced 565 noise and improved resolution (34). 566

567 Materials and Methods

568 569 Serial-sectioning OCT (S-OCT)

The S-OCT microscope was described previously (34). We used a swept light source 570 (AxsunTech) with 100 kHz swept rate, a central wavelength of 1310 nm, and a spectral full 571 width half maximum (FWHM) of 110nm, yielding an axial resolution of 5.6 \Box m in brain 572 tissue (n=1.4). We used a free-space interferometer and quarter wave plate (OWP) to 573 illuminate the sample with circularly polarized light, and used two balanced detectors for 574 measuring orthogonally polarized reflection light. A variable retarder (VR) placed in the 575 sample arm was used to compensate for the system birefringence and to recover precise 576 measurement of sample birefringence. To sample the spectrum in even-k space, we input 577 578 the k-clock of the light source into a high-speed digitizer (ATS9350, AlazarTech), afterwards real-time FFT was carried out using a Graphic Processing Unit (RTX4000, 579 580 NVIDIA), and the spatial-domain data was trimmed to only save the first 1 mm depth. The post-objective power was measured to be 3.7 mW, achieving a 95dB SNR in both 581 582 polarization channels. We used 1×1 mm² FOV with 3 µm lateral step size and 30% overlap between tiles. The sample was mounted on XYZ motorized stages which translated the 583 sample to image the whole surface as well as between the vibratome and objective. After 584 block-face imaging, a custom vibratome cut off the 50 µm slices with 0.3mm/s cutting speed 585 and 3000 rotations per minute (RPM) blade vibrating frequency. 586

587 **Optical properties estimation with S-OCT**

588 Tissue optical properties were extracted by following a previously established procedure to 589 analyze the reflectance intensities of OCT using a nonlinear fitting method (11, 12). To 590 summarize, spatial parametrization is first applied to the confocal parameter across a 3D 591 OCT image to constrain and reduce the degrees of freedom in the nonlinear coefficient 592 fitting problem, resulting in improved confidence in the estimated optical properties of the 593 sample. Afterwards, a nonlinear least-squares solver is used to estimate the back-scattering 594 and scattering coefficients from the nonlinear reflectance-vs-depth over about 150 μ m 595 depth. All curve fitting was performed in MATLAB. After extracting the optical properties 596 for each image tile, the tiles were stitched using the coordinates generated during the 597 volumetric reconstruction with ImageJ software (45).

598 Sample preparation

599 For the training phase, we used a set of 15 samples obtained from the Boston University 600 Alzheimer's Disease Research Center brain bank. These samples consisted of five cases 601 with stage VI Alzheimer's disease (AD), five cases with stage III and IV Chronic Traumatic 602 Encephalopathy (CTE), and five age-matched normal control cases. To ensure 603 representation across the thickness of the tissue, we selected one slice per millimeter for this 604 study.

605For the pilot generalization study, we used OCT data from five samples obtained from two606human brains. These samples were collected at the Massachusetts General Hospital Autopsy607Suite and encompassed various brain regions, including the cerebellum, hippocampus,608somatosensory cortex, superior frontal cortex, and middle temporal area 21. The subjects609from whom these samples were obtained had no history of neurological deficits and had a610mean age of 53.5 ± 12.0 years, with one male and one female.

All samples were fixed by immersion in 10% formalin for at least two months. The post-611 mortem interval did not exceed 24 hours. Prior to imaging, the samples were washed in 1X 612 phosphate buffered saline for a month to remove residual fixation agents and then embedded 613 in 4.5% agarose for tissue support (46). During embedding, the brain blocks were warmed 614 to 65 °C to allow sufficient penetration of agarose into the deep sulcus. During imaging, the 615 616 brain tissue blocks were mounted in a water bath filled with Deionized (DI) water. The DI water was changed every day to remove the debris from cutting that could degrade the OCT 617 image quality. Following data collection, the tissue slices were stored in 1X PBS with an 618 619 antibacterial agent (sodium azide) at a temperature of 4 °C. To maintain the sequence of the slices, each slice was stored in an individual glass vial. 620

621 Gallyas silver staining and imaging

A total of 35 brain slices were obtained from 15 samples for our study. To ensure anatomical diversity, at least two slices were taken per sample, with each slice being separated in depth by 1 mm. These slices had a thickness of 50 μ m and were mounted onto gelatin-coated slides. Gallyas staining protocol, as described by Pistorio (2), was then employed to process the samples. Modifications were made to the impregnation and bleaching time to accommodate the increased thickness of the samples.

Following the staining process, the samples were captured in brightfield mode using a $10 \times$ objective (NA=0.4) and an RGB camera. We utilized the VS-120 slide scanner designed for $75 \times 25 \text{ mm}^2$ slides for this purpose. The exposure time was set at 1.73 ms, and the pixel size was 0.7 µm with a $1 \times 1 \text{ mm}^2$ FOV. For wider samples, imaging was conducted using the BZ-X microscope under similar settings. The resulting images can be opened in Olympus Olyvia software and exported as TIFF images for further processing.

634 Image processing

635Our image dataset consists of two types of images: PS images from the slide scanner and636OCT-SC images computed from S-OCT. We first inspected all the PS images visually and

637 excluded the ones that had low staining quality or artifacts in the training dataset. We 638 selected 9 out of 35 PS WSIs for training our DS model. The PS WSIs had different sizes 639 depending on the tissue sample, but they were around the median scale of $36 \text{ mm} \times 48 \text{ mm}$ with the pixel size of 1.9 µm. To generate the weakly-paired training dataset, we manually 640 paired the PS images with the OCT-SC images that had the most similar appearance. Since 641 the sectioning thickness (50 µm) of PS samples did not match the fitting thickness used for 642 643 OCT-SC images (150 μ m) and the depth information of PS samples was not recorded, we 644 can only pair the PS with the closest adjacent OCT-SC image sample by qualitatively assessing the similarity of tissue features. We then downsampled the PS images using 645 bicubic interpolation by a scale factor of 6.32 to match the 12 µm pixel size in OCT-SC 646 images. We also cropped or padded the PS images to have the same image size as the 647 corresponding OCT-SC images, which was around 3000×4000 pixels for each sample. We 648 performed this procedure on all PS images when we compared them with the OCT-SC 649 650 images side-by-side in our results.

The PS images undergo several preprocessing steps to minimize the effects of sample and 651 652 staining variations before they are used for training. The preprocessing steps include background removal, intensity normalization and color transfer. The background removal 653 654 eliminates the unwanted image artifacts in PS image and is done by interactive image segmenter in MATLAB. The intensity normalization adjusts the PS images to balance the 655 656 varying illumination levels across different imaging experiments. The brightest pixel (Ir, Ig, Ib) is used to estimate the illuminant color and the image is scaled by the constant 657 $(1/I_r, 1/I_g, 1/I_b)$ for each color channel, followed by a range normalization to map the 658 overall image value range to [0, 1]. The color transfer uses Reinhard method (47) to 659 standardize the staining color variations among experiment, sample and imaging conditions 660 given a target PS image with a relatively ideal staining as reference. 661

662 The OCT-SC images obtained from the fitting algorithm show some artifacts mainly in the background areas and near the sharp boundaries of the vessel regions, because the algorithm 663 664 assumes a constant SC value for the 150 μ m imaging thickness (11). To reduce the background noise and correct the over-smoothed values near the vessel edges, the OCT-SC 665 images are further processed by several steps. First, the background is removed by applying 666 667 a histogram-based thresholding method using the triangle algorithm (48), followed by a sequence of smoothing morphological operations such as erosion, small object removal and 668 dilation. Next, the pixels with zero values in the masked image are identified as defective 669 and are replaced by the local median. Then, the edges of the vessel regions are detected 670 using a difference-of-Gaussian (DoG) filter and thresholding. Finally, the outlier regions 671 with small values compared to the local maximum are segmented and combined with the 672 673 edge mask. The combined mask is smoothed by similar morphological filters, and the values in the mask are replaced by the local maximum. The preprocessing pipeline is implemented 674 in Python using the basic filters and morphological operators from scikit-image package 675 (48). 676

To generate the training image dataset, we used PyTorch to create a parallel processing module that can split the WSIs of different image sizes into smaller patches during training on the fly. This allows us to dynamically update the intermediate image tensors that can be input to different parts of deep learning models to train at different image scales. The WSIs dataset with different sizes can then be directly handled by a custom data loader for standard-size tensor operation. We first pad the WSI to the size of multiple integers of patch size, and then use the tensor unfolding method in PyTorch to cut the image tensor using a sliding window into smaller tensors stacked in the batch dimension. The inverse stitching
operation is done similarly using the tensor folding methods.

For creating a 3D visualization of the DS images that show the volumetric digital staining 686 results, we change the white-color background of the DS images to black, so that only the 687 688 sample region is visible. This is done by converting the DS color images to grayscale and applying a triangle method threshold to select the foreground pixels. Then, a morphology 689 690 smoothing operation is performed to remove any noise or artifacts. To extract the WM masks from the DS gravscale images for highlighting the WM regions in the sample, we 691 692 use a histogram thresholding method based on the minimum method (48) and apply another 693 morphology smoothing operation. The pixels that are not part of the WM masks are set to zero, and the resulting images are stacked in a volume for 3D visualization. The 3D viewer 694 695 in ImageJ (45) is used to display the volume. More details on the image processing 696 procedures are provided in SM Section 8 and Fig. S7.

697 Semi-supervised deep learning framework

- The proposed framework combines generative adversarial learning, contrastive learning,
 pseudo-supervised learning based on self-generated image pairs based on a biophysical
 model, and unsupervised cross-modality image registration.
- We denote the OCT-SC images as X and the PS images as Y. The main framework consists 701 of a DS network G and a registration network R. The DS network G transforms grayscale 702 OCT-SC images X into color images that resemble the color and contrast of PS images Y. 703 The registration network R takes pairs of unaligned images X and Y as input and outputs a 704 705 deformation field $\phi = R(X, Y)$ that can be applied to resample and register Y to X. We use an auxiliary discriminator network D to enforce structural similarity between the output DS 706 707 and reference PS images by adversarial learning. We also apply contrastive learning to ensure structural consistency between the input OCT-SC and output DS images using a fully 708 709 connected network f.
- Our framework operates on two different image scales: WSI scale (denoted by upper case 710 letters) and image patch scale (denoted by lower case letters). R is trained on WSIs, which 711 have a size of about 3000×4000 pixels. G, D, f are trained on image patches, which have a 712 size of 512×512 pixels. We design an efficient image processing module to either split (X, 713 714 Y) into patches (x, y) or stitch patches back to WSIs, as detailed in the Image Processing section. The CUT framework (32) is used to jointly train the networks G, D, and f during 715 the training phase. Additionally, G undergoes a pseudo-supervised training scheme and an 716 alternating training process with R, which are explained below. 717
- The objective of the adversarial learning module is to enhance the perceptual similarity 718 719 between the DS output G(x, y) and the target modality PS images y. This is achieved by 720 using an auxiliary discriminator D. The role of D is to learn to differentiate between the 721 desired modality y and the generated images G(x). During the training of D, the PS images y are assigned the label 1, indicating that they are "true" images. On the other hand, the 722 generated images G(x) are assigned the label 0, indicating that they are "false" images. The 723 least-squares generative adversarial network (GAN) loss $L_{GAN}(D)$ is employed to measure 724 725 the extent to which D's outputs align with the binary labels assigned to both y and G(x). 726 This loss function is minimized when D becomes proficient at distinguishing between y and G(x). Conversely, when training G, the $L_{GAN}(G)$ loss is utilized to promote the fidelity of 727 the generated images G(x). Minimizing this loss prompts G to effectively deceive the 728

729discriminator D. The training process alternates between two steps: first, G is fixed while D730is updated using the $L_{GAN}(D)$ loss, and then D is fixed while G is updated using the731 $L_{GAN}(G)$ loss:

$$L_{\text{GAN}}(D) = E_{y}[(D(y) - 1)^{2}] + E_{x}[D^{2}(G(x))]$$
(1)

(2)

733
$$L_{\text{GAN}}(G) = E_x \left[\left(D(G(x)) - 1 \right)^2 \right]$$

732

734 The contrastive learning module ensures that the image structures and content present in x is preserved when it is translated to G(x). We implement G with a ResNet model and treats 735 736 the first half of the ResNet layers as the encoder and the remaining layers as the decoder. The encoder Genc transforms images from both domains into a common latent space, and 737 the decoder G_{dec} generates translated images from latent vectors. To formulate the multi-738 739 layer patch-wise contrastive loss, we adopt the approach in (32) to sample the encoded 740 feature maps from both x and G(x). Each layer and spatial location in the feature map stack 741 corresponds to a patch of the input image that covers the corresponding receptive field. We 742 extract multiple layers of the encoded feature maps, randomly sample the spatial locations and apply a fully connected network f to obtain a stack of latent features $\hat{z}_{s,l} = f(G_{enc}^{s,l}(x))$, 743 where s is the spatial index within [1, S] and l is the selected layer within [1, L]. We do the 744 same processing on image G(x): $\hat{z}_{s,l} = f(G_{enc}^{s,l}(G(x)))$ Then we compute a PatchNCE loss 745 746 using a cross-entropy contrastive loss:

747
$$L_{\text{PatchNCE}}(G, f, x) = E_x \sum_{l=1}^{L} \sum_{s=1}^{S} \log\left(\frac{\exp(z_{s,l} \cdot \hat{z}_{s,l})}{\sum_{t=1}^{S} \exp(z_{s,l} \cdot \hat{z}_{t,l})}\right)$$
(3)

748 This loss function encourages the latent representations of image patches from x and G(x) that belong to the same spatial location to have similar content to be close in the feature 749 space, while pushing away the representations of patches that are uncorrelated or have 750 751 different content. By this internal negative sampling scheme in the feature space, the model learns to contrast positive and negative pairs of patches based on their content similarity, 752 which maximizes the mutual information between the input image x and the output image 753 G(x). This provides a self-supervised signal for preserving the content of the image during 754 755 the transformation.

The training procedure for pseudo-supervised learning is formulated as a pixel-wise loss function that minimizes the discrepancy between the digital stained OD images G(OD(Y))and the physical Gallyas-silver stain (PS) images Y. This loss function aims to guide G to learn a mapping that translates images from the OD modality to the PS modality. By doing so, it provides a "proxy supervision" for learning the mapping from OCT-SC modality to the PS modality. To facilitate this training, we first compute the OD of image Y by

762
$$OD(Y) = -\frac{1}{3} \sum_{c=R,G,B} \log_{10} Y_c$$
(4)

763Subsequently, we extract patches OD(y) and y from the processed WSIs and employ an764auxiliary pseudo-supervised loss, defined as:

765
$$L_{\text{Pseudo}}(G) = E_y \|G(\text{OD}(y) - y)\|_1$$
 (5)

766 However, there exists a mismatch in the intensity values between X and OD(Y). This domain gap between the input modalities hinders the model's direct generalization on X if it is solely 767 trained on pairs of OD(Y) and Y. To address this issue, we first apply histogram equalization 768 to the WSIs of OD(Y) and X before feeding them into G. This normalization step aims to 769 align the distribution of intensity range. However, we found that this transformation alone 770 is insufficient in mitigating the domain gap. As a result, this learning module is further 771 combined with the adversarial learning module in the CUT backbone to mitigate the domain 772 773 gap between OCT-SC and OD.

774 The cross-modality image registration module is trained in two stages. In the first stage, we 775 pre-train the registration network R on WSIs of X, Y and OD(Y). The registration network R takes weakly-paired X and Y as input and predicts a deformation field $\phi = R(X, Y)$ that 776 indicates the pixel-wise displacement vectors needed to perform the non-rigid 777 transformation. To formulate a cross-modal self-supervised registration loss L_{reg}^{l} , we use 778 OD(Y) as a surrogate of Y and exploit its correlation with the input OCT-SC image X. By 779 minimizing the difference between the registered OD(Y) and X, we indirectly learn the 780 781 deformation between Y and X. This training is enabled by a differentiable resampling layer that performs image warping denoted by •. We also add a total variation (TV) regularization 782 783 term to encourage the smoothness of the learned deformation field. The registration loss during this pre-training stage is computed at the WSI scale as follows: 784

$$L_{\text{reg}}^{I}(R) = E_{X,Y} \| X - \phi \circ OD(Y) \|_{1} + \| \phi \|_{TV}$$
(6)

786 where $\|\phi\|_{TV}$ is the total variation norm calculated as:

785

799

787
$$\|\phi\|_{TV} = \sum_{i,j} \sqrt{|\phi_{i+1} - \phi_{i,j}|^2 + |\phi_{i,j+1} - \phi_{i,j}|^2}$$
(7)

In the second fine-tuning stage, we train R and G in an alternating and collaborative manner. The purpose of fine-tuning R is to provide pixel-wise weak-supervision between the registered Y and the DS image G(x), which in turn helps to fine-tune the DS network G. Using the coarsely trained G, we can produce G(x) that has the same image modality as the PS image Y and use a pixel-wise loss function to perform training. We implement the following scheme for alternating training. When we fix G, we train R by comparing the registered PS image Y and the DS image G(X) at the WSI scale using the loss function

795
$$L_{\text{reg}}^{II}(R) = E_{X,Y} \|G(X) - \phi \circ Y\|_1 + \|\phi\|_{TV}$$
(8)

796 When we fix *R*, we crop the intermediate registered WSI $\phi \circ Y$ into patches $\phi_y \circ y$ and train 797 *G* at the patch scale by comparing the registered PS image patch and the DS image patch 798 *G*(*x*) using the loss function

$$L_{\text{reg}}^{II}(G) = E_{x,y} \left\| G(x) - \phi_y \circ y \right\|_1$$
(9)

800Additional details about the deep learning framework and individual model architectures801are provided in SM Section 9, 10 and Fig. S8, S9 and S10.

802 Image analysis

The layer differentiation analysis is primarily performed using the open-source ImageJ 803 software package. The line profiles are computed by selecting the rectangular region in the 804 805 center region of interest (ROI) and aggregating the intensity value along the horizontal direction. Those profiles are then normalized to [0, 1] by their individual value range for 806 807 visual comparisons. The cortical layer boundaries are manually annotated by identifying the 808 local maxima and edges according to (36, 37). The layer segmentation on the larger ROI is performed by manual annotation on layer IV, V and VI. We used the built-in local thickness 809 810 estimation function to generate the local thickness map and calculated the box plot for the 811 thickness distribution using Matlab. Two Gyral crest ROIs and one Sulcus ROI are manually selected. Additional details about the analysis methods for the myelin fibers and vessel 812 quantification are provided in SM Section 11 and Fig. S11. 813

814 **Code availability**

818 819

820

821 822

834

815 We have open sourced our codebase with training/testing script and pre-trained model 816 weights on our GitHub repository: <u>https://github.com/bu-cisl/DS-OCT</u>, which will be 817 made public upon publication.

Data availability

All data are available in the main text or the supplementary materials.

Acknowledgments: The authors acknowledge funding support from:

- 823 National Institutes of Health grant R01 EY032163 (SC, LT)
- 824 National Institutes of Health grant U54 NS115266 (AM)
- 825 National Institutes of Health grant P30 AG072978 (AM)
- 826 National Institutes of Health grant U19 AG068753 (AM)
- 827 National Institutes of Health grant R01 NS125307 (DR)
- 828 National Institutes of Health grant R01 AG075727 (DR, IB)
- 829 National Institutes of Health grant RF1 AG062831 (DR)
- 830 Boston University Kilachand Fund Award (IB, DR, DB, LT)
- 831 National Institutes of Health grant R00 EB023993 (HW)
- 832 National Institutes of Health grant R01 NS128843 (HW)
- 833 National Institutes of Health grant U01 MH117023 (SC, AN, DB)

835 Author contributions: L.T., D.A.B and I.J.B. conceived the original idea of this work. H.W. provided initial OCT data and processing algorithms for prototyping. D.L.R. provided 836 experimental supervision and guidance, chemical material, and laboratory support for 837 Gallvas silver stain. A.C.M. provided valuable brain samples and initial Gallvas silver 838 839 stained image data for prototyping. Shiyi C. constructed the algorithmic pipeline for image processing, neural network design, training and testing, image analysis and results 840 evaluation, and performed Gallyas silver staining (GSS) experiment with A.N.. Shuaibin C. 841 constructed the S-OCT hardware setup, performed brain sample sectioning and imaging 842 experiment, constructed scattering coefficient fitting and data preprocessing algorithms. 843 844 Y.L. provided innovative ideas on modifications and refinement on the computational framework and performed extensive image data analysis. A.N. performed the GSS 845 experiment with Shivi C. and completed bright field imaging and image acquisition. S.L. 846 constructed and optimized CUT and FastCUT neural networks for baseline model 847 848 comparison. Y.W. and J.Z. constructed and optimized CycleGAN network for baseline model comparison. All authors contributed to the writing of the paper and insightful 849 850 discussions on the result analysis.

851 852 853		Competing interests: The authors declare that they have no competing interests.			
854 855	Refe	References			
856 857	1.	S. Herculano-Houzel, The remarkable, yet not extraordinary, human brain as a scaled-up primate brain and its associated cost. <i>Proc. Natl. Acad. Sci.</i> 109 , 10661–10668 (2012).			
858 859	2.	A. L. Pistorio, S. H. Hendry, X. Wang, A modified technique for high-resolution staining of myelin. <i>J. Neurosci. Methods</i> 153 , 135–146 (2006).			
860 861	3.	N. Kuninaka, M. Kawaguchi, M. Ogawa, A. Sato, K. Arima, S. Murayama, Y. Saito, Simplification of the modified Gallyas method. <i>Neuropathology</i> 35 , 10–15 (2015).			
862 863 864 865	4.	K. Amunts, C. Lepage, L. Borgeat, H. Mohlberg, T. Dickscheid, MÉ. Rousseau, S. Bludau, PL. Bazin, L. B. Lewis, AM. Oros-Peusquens, N. J. Shah, T. Lippert, K. Zilles, A. C. Evans, BigBrain: An Ultrahigh-Resolution 3D Human Brain Model. <i>Science</i> 340 , 1472–1475 (2013).			
866 867 868 869	5.	P. A. Yushkevich, B. B. Avants, L. Ng, M. Hawrylycz, P. D. Burstein, H. Zhang, J. C. Gee, "3D Mouse Brain Reconstruction from Histology Using a Coarse-to-Fine Approach" in <i>Biomedical Image Registration</i> , J. P. W. Pluim, B. Likar, F. A. Gerritsen, Eds. (Springer, Berlin, Heidelberg, 2006) <i>Lecture Notes in Computer Science</i> , pp. 230–237.			
870 871	6.	S. Osechinskiy, F. Kruggel, Slice-to-Volume Nonrigid Registration of Histological Sections to MR Images of the Human Brain. <i>Anat. Res. Int.</i> 2011 , 287860 (2011).			
872 873 874	7.	E. Min, S. Ban, J. Lee, A. Vavilin, S. Baek, S. Jung, Y. Ahn, K. Park, S. Shin, S. Han, H. Cho, W. Lee-Kwon, J. Kim, C. J. Lee, W. Jung, Serial optical coherence microscopy for label-free volumetric histopathology. <i>Sci. Rep.</i> 10 , 6711 (2020).			
875 876 877	8.	R. Cao, S. D. Nelson, S. Davis, Y. Liang, Y. Luo, Y. Zhang, B. Crawford, L. V. Wang, Label-free intraoperative histology of bone tissue via deep-learning-assisted ultraviolet photoacoustic microscopy. <i>Nat. Biomed. Eng.</i> 7 , 124–134 (2023).			
878 879 880	9.	Y. Zhang, L. Kang, W. Yu, V. T. C. Tsang, T. T. W. Wong, Three-dimensional label-free histological imaging of whole organs by microtomy-assisted autofluorescence tomography. <i>iScience</i> 25 , 103721 (2022).			
881 882 883	10.	Y. Sun, S. You, X. Du, A. Spaulding, Z. G. Liu, E. J. Chaney, D. R. S. Jr, M. Marjanovic, H. Tu, S. A. Boppart, Real-time three-dimensional histology-like imaging by label-free nonlinear optical microscopy. <i>Quant. Imaging Med. Surg.</i> 10 , 2177190–2172190 (2020).			
884 885 886	11.	H. Wang, C. Magnain, S. Sakadžić, B. Fischl, D. A. Boas, Characterizing the optical properties of human brain tissue with high numerical aperture optical coherence tomography. <i>Biomed. Opt. Express</i> 8 , 5617–5636 (2017).			
887 888 889 890 891	12.	J. Yang, I. A. Chen, S. Chang, J. Tang, B. Lee, K. Kılıç, S. Sunil, H. Wang, D. Varadarajan, C. Magnain, SC. Chen, I. Costantini, F. Pavone, B. Fischl, D. A. Boas, Improving the characterization of ex vivo human brain optical properties using high numerical aperture optical coherence tomography by spatially constraining the confocal parameters. <i>Neurophotonics</i> 7 , 045005 (2020).			

893 894		S. Rockland, Colocalization of neurons in optical coherence microscopy and Nissl-stained histology in Brodmann's area 32 and area 21. <i>Brain Struct. Funct.</i> 224 , 351–362 (2019).
895 896 897 898 898	14.	J. Yang, S. Chang, I. A. Chen, S. Kura, G. A. Rosen, N. A. Saltiel, B. R. Huber, D. Varadarajan, Y. Balbastre, C. Magnain, SC. Chen, B. Fischl, A. C. McKee, D. A. Boas, H. Wang, Volumetric Characterization of Microvasculature in Ex Vivo Human Brain Samples By Serial Sectioning Optical Coherence Tomography. <i>IEEE Trans. Biomed. Eng.</i> 69 , 3645–3656 (2022).
900 901 902	15.	C. Magnain, J. C. Augustinack, M. Reuter, C. Wachinger, M. P. Frosch, T. Ragan, T. Akkin, V. J. Wedeen, D. A. Boas, B. Fischl, Blockface histology with optical coherence tomography: a comparison with Nissl staining. <i>NeuroImage</i> 84 , 524–533 (2014).
903 904 905	16.	C. Magnain, J. C. Augustinack, E. Konukoglu, M. P. Frosch, S. Sakadžić, A. Varjabedian, N. Garcia, V. J. Wedeen, D. A. Boas, B. Fischl, Optical coherence tomography visualizes neurons in human entorhinal cortex. <i>Neurophotonics</i> 2 , 015004 (2015).
906 907 908 909	17.	H. Wang, C. Magnain, R. Wang, J. Dubb, A. Varjabedian, L. S. Tirrell, A. Stevens, J. C. Augustinack, E. Konukoglu, I. Aganj, M. P. Frosch, J. D. Schmahmann, B. Fischl, D. A. Boas, as-PSOCT: Volumetric microscopic imaging of human brain architecture and connectivity. <i>NeuroImage</i> 165 , 56–68 (2018).
910 911	18.	C. J. Liu, K. E. Williams, H. T. Orr, T. Akkin, Visualizing and mapping the cerebellum with serial optical coherence scanner. <i>Neurophotonics</i> 4 , 011006 (2016).
912 913	19.	H. Wang, J. Zhu, T. Akkin, Serial optical coherence scanner for large-scale brain imaging at microscopic resolution. <i>NeuroImage</i> 84 , 1007–1017 (2014).
914 915	20.	A. Odgaard, K. Andersen, F. Melsen, H. J. G. Gundersen, A direct method for fast three- dimensional serial reconstruction. <i>J. Microsc.</i> 159 , 335–342 (1990).
916 917 918	21.	S. Chang, D. Varadarajan, J. Yang, I. A. Chen, S. Kura, C. Magnain, J. C. Augustinack, B. Fischl, D. N. Greve, D. A. Boas, H. Wang, Scalable mapping of myelin and neuron density in the human brain with micrometer resolution. <i>Sci. Rep.</i> 12 , 363 (2022).
919 920	22.	B. Bai, X. Yang, Y. Li, Y. Zhang, N. Pillar, A. Ozcan, Deep learning-enabled virtual histological staining of biological samples. <i>Light Sci. Appl.</i> 12 , 57 (2023).
921 922 923	23.	Y. Rivenson, T. Liu, Z. Wei, Y. Zhang, K. de Haan, A. Ozcan, PhaseStain: the digital staining of label-free quantitative phase microscopy images using deep learning. <i>Light Sci. Appl.</i> 8 , 23 (2019).
924 925 926 927	24.	Y. Rivenson, H. Wang, Z. Wei, K. de Haan, Y. Zhang, Y. Wu, H. Günaydın, J. E. Zuckerman, T. Chong, A. E. Sisk, L. M. Westbrook, W. D. Wallace, A. Ozcan, Virtual histological staining of unlabelled tissue-autofluorescence images via deep learning. <i>Nat. Biomed. Eng.</i> 3 , 466–477 (2019).
928 929 930 931	25.	K. de Haan, Y. Zhang, J. E. Zuckerman, T. Liu, A. E. Sisk, M. F. P. Diaz, KY. Jen, A. Nobori, S. Liou, S. Zhang, R. Riahi, Y. Rivenson, W. D. Wallace, A. Ozcan, Deep learning-based transformation of H&E stained tissues into special stains. <i>Nat. Commun.</i> 12 , 4884 (2021).

13. C. Magnain, J. C. Augustinack, L. Tirrell, M. Fogarty, M. P. Frosch, D. Boas, B. Fischl, K.

892

932 933 934 935	26.	Y. Winetraub, E. Yuan, I. Terem, C. Yu, W. Chan, H. Do, S. Shevidi, M. Mao, J. Yu, M. Hong, E. Blankenberg, K. E. Rieger, S. Chu, S. Aasi, K. Y. Sarin, A. de la Zerda, OCT2Hist: Non-Invasive Virtual Biopsy Using Optical Coherence Tomography. medRxiv [Preprint] (2021). https://doi.org/10.1101/2021.03.31.21254733.
936 937 938	27.	X. Li, G. Zhang, H. Qiao, F. Bao, Y. Deng, J. Wu, Y. He, J. Yun, X. Lin, H. Xie, H. Wang, Q. Dai, Unsupervised content-preserving transformation for optical microscopy. <i>Light Sci. Appl.</i> 10 , 44 (2021).
939 940 941 942 943	28.	K. B. Ozyoruk, S. Can, B. Darbaz, K. Başak, D. Demir, G. I. Gokceler, G. Serin, U. P. Hacisalihoglu, E. Kurtuluş, M. Y. Lu, T. Y. Chen, D. F. K. Williamson, F. Yılmaz, F. Mahmood, M. Turan, A deep-learning model for transforming the style of tissue images from cryosectioned to formalin-fixed and paraffin-embedded. <i>Nat. Biomed. Eng.</i> 6 , 1407–1419 (2022).
944 945 946 947	29.	M. Combalia, J. Pérez-Anker, A. García-Herrera, L. Alos, V. Vilaplana, F. Marqués, S. Puig, J. Malvehy, "Digitally Stained Confocal Microscopy through Deep Learning" in <i>Proceedings of The 2nd International Conference on Medical Imaging with Deep Learning</i> (PMLR, 2019; https://proceedings.mlr.press/v102/combalia19a.html), pp. 121–129.
948 949 950	30.	L. Kang, X. Li, Y. Zhang, T. T. W. Wong, Deep learning enables ultraviolet photoacoustic microscopy based histological imaging with near real-time virtual staining. <i>Photoacoustics</i> 25 , 100308 (2022).
951 952 953 954 955	31.	JY. Zhu, T. Park, P. Isola, A. A. Efros, "Unpaired Image-To-Image Translation Using Cycle-Consistent Adversarial Networks" in <i>Proceedings of the IEEE International</i> <i>Conference on Computer Vision</i> (2017; https://openaccess.thecvf.com/content_iccv_2017/html/Zhu_Unpaired_Image-To- Image_Translation_ICCV_2017_paper.html), pp. 2223–2232.
956 957 958 959	32.	T. Park, A. A. Efros, R. Zhang, JY. Zhu, "Contrastive Learning for Unpaired Image-to- Image Translation" in <i>Computer Vision – ECCV 2020</i> , A. Vedaldi, H. Bischof, T. Brox, J M. Frahm, Eds. (Springer International Publishing, Cham, 2020) <i>Lecture Notes in Computer</i> <i>Science</i> , pp. 319–345.
960 961 962 963 964	33.	Y. Ganin, E. Ustinova, H. Ajakan, P. Germain, H. Larochelle, F. Laviolette, M. Marchand, V. Lempitsky, "Domain-Adversarial Training of Neural Networks" in <i>Domain Adaptation in Computer Vision Applications</i> , G. Csurka, Ed. (Springer International Publishing, Cham, 2017; http://link.springer.com/10.1007/978-3-319-58347-1_10) <i>Advances in Computer Vision and Pattern Recognition</i> , pp. 189–209.
965 966 967 968 969	34.	S. Chang, J. Yang, A. Novoseltseva, X. Fu, C. Li, SC. Chen, J. C. Augustinack, C. Magnain, B. Fischl, A. C. Mckee, D. A. Boas, I. A. Chen, H. Wang, Multi-Scale Label-free Human Brain Imaging with Integrated Serial Sectioning Polarization Sensitive Optical Coherence Tomography and Two-Photon Microscopy. bioRxiv [Preprint] (2023). https://doi.org/10.1101/2023.05.22.541785.
970 971	35.	D. Varadarajan, C. Magnain, M. Fogarty, D. A. Boas, B. Fischl, H. Wang, A novel algorithm for multiplicative speckle noise reduction in ex vivo human brain OCT images.

 9/1
 algorithm for multiplicative speck

 972
 NeuroImage 257, 119304 (2022).

973 974	36.	N. Palomero-Gallagher, K. Zilles, Cortical layers: Cyto-, myelo-, receptor- and synaptic architecture in human cortical areas. <i>NeuroImage</i> 197 , 716–741 (2019).
975 976	37.	R. Turner, Myelin and Modeling: Bootstrapping Cortical Microcircuits. <i>Front. Neural Circuits</i> 13 , 34 (2019).
977 978	38.	D. J. Lin, K. L. Hermann, J. D. Schmahmann, The Diagnosis and Natural History of Multiple System Atrophy, Cerebellar type. <i>Cerebellum Lond. Engl.</i> 15 , 663–679 (2016).
979 980	39.	G. M. Halliday, Re-evaluating the glio-centric view of multiple system atrophy by highlighting the neuronal involvement. <i>Brain</i> 138 , 2116–2119 (2015).
981 982	40.	J. DeFelipe, L. Alonso-Nanclares, J. I. Arellano, Microstructure of the neocortex: comparative aspects. <i>J. Neurocytol.</i> 31 , 299–316 (2002).
983 984 985 986 987	41.	K. Wagstyl, S. Larocque, G. Cucurull, C. Lepage, J. P. Cohen, S. Bludau, N. Palomero-Gallagher, L. B. Lewis, T. Funck, H. Spitzer, T. Dickscheid, P. C. Fletcher, A. Romero, K. Zilles, K. Amunts, Y. Bengio, A. C. Evans, BigBrain 3D atlas of cortical layers: Cortical and laminar thickness gradients diverge in sensory and motor cortices. <i>PLOS Biol.</i> 18 , e3000678 (2020).
988 989 990	42.	C. CC. Pang, C. Kiecker, J. T. O'Brien, W. Noble, R. CC. Chang, Ammon's Horn 2 (CA2) of the Hippocampus: A Long-Known Region with a New Potential Role in Neurodegeneration. <i>The Neuroscientist</i> 25 , 167–180 (2019).
991 992 993	43.	A. B. Molini, D. Valsesia, G. Fracastoro, E. Magli, Speckle2Void: Deep Self-Supervised SAR Despeckling With Blind-Spot Convolutional Neural Networks. <i>IEEE Trans. Geosci. Remote Sens.</i> 60 , 1–17 (2022).
994 995	44.	R. Liu, S. Cheng, L. Tian, J. Yi, Deep spectral learning for label-free optical imaging oximetry with uncertainty quantification. <i>Light Sci. Appl.</i> 8 , 102 (2019).
996 997	45.	C. A. Schneider, W. S. Rasband, K. W. Eliceiri, NIH Image to ImageJ: 25 years of image analysis. <i>Nat. Methods</i> 9 , 671–675 (2012).
998 999	46.	J. Fan, T. M. Dawson, V. L. Dawson, Cell Death Mechanisms of Neurodegeneration. <i>Adv. Neurobiol.</i> 15 , 403–425 (2017).
1000 1001	47.	E. Reinhard, M. Adhikhmin, B. Gooch, P. Shirley, Color transfer between images. <i>IEEE Comput. Graph. Appl.</i> 21 , 34–41 (2001).
1002 1003	48.	S. van der Walt, J. L. Schönberger, J. Nunez-Iglesias, F. Boulogne, J. D. Warner, N. Yager, E. Gouillart, T. Yu, scikit-image: image processing in Python. <i>PeerJ</i> 2 , e453 (2014).
1004		

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

• SupplementaryMaterials.pdf