

Prediction of Carboxylesterase 1-mediated In Vivo Drug Interaction between Methylphenidate and Cannabinoids using Static and Physiologically Based Pharmacokinetic Models[§]

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ABSTRACT

The use of cannabis products has increased substantially. Cannabis products have been perceived and investigated as potential treatments for attention-deficit/hyperactivity disorder (ADHD). Accordingly, co-administration of cannabis products and methylphenidate (MPH), a first-line medication for ADHD, is possible. Oral MPH undergoes extensive presystemic metabolism by carboxylesterase 1 (CES1), a hepatic enzyme which can be inhibited by two prominent cannabinoids, Δ^9 -tetrahydrocannabinol (THC) and cannabidiol (CBD). This prompts further investigation into the likelihood of clinical interactions between MPH and these two cannabinoids through CES1 inhibition. In the present study, inhibition parameters were obtained from a human liver S9 system and then incorporated into static and physiologically-based pharmacokinetic (PBPK) models for prediction of potential clinical significance. The inhibition of MPH hydrolysis by THC and CBD was reversible, with estimated unbound inhibition constants ($K_{i,u}$) of 0.031 and 0.091 μM , respectively. The static model predicted a mild increase in MPH exposure by concurrent THC (34%) and CBD (94%) from smoking a cannabis cigarette and ingestion of prescriptive

CBD, respectively. PBPK models suggested no significant interactions between single doses of MPH and CBD (2.5 – 10 mg/kg) when administered simultaneously, while a mild interaction (area under drug concentration-time curve increased by up to 55% and maximum concentration by up to 45%) is likely if multiple doses of CBD (10 mg/kg twice daily) are administered. In conclusion, the pharmacokinetic disposition of MPH can be potentially influenced by THC and CBD under certain clinical scenarios. Whether the magnitude of predicted interactions translates into clinically relevant outcomes requires verification in an appropriately designed clinical study.

SIGNIFICANCE STATEMENT

This work demonstrated a potential mechanism of drug-drug interactions between methylphenidate (MPH) and two major cannabinoids (Δ^9 -tetrahydrocannabinol [THC] and cannabidiol [CBD]) not previously reported. We predicted a mild interaction between MPH and THC when the cannabinoid exposure occurred via cannabis smoking. Mild interactions between MPH and CBD were predicted with multiple oral administrations of CBD.

Introduction

Cannabis products are prepared from plants within the *Cannabis* genus which contain a number of unique compounds (cannabinoids) that can regulate various biologic processes. Most cannabis products contain the major psychoactive constituent Δ^9 -tetrahydrocannabinol (THC) and are frequently used recreationally. The use of cannabis has increased dramatically in the US both recreationally and medically, which appears to coincide with the growing number of states legalizing its use (Azofeifa et al., 2016). In addition to its recreational use, the use of

cannabis for treatment of certain diseases and palliative care has attracted attention in the medical community (Bridgeman and Abazia, 2017). Notably, cannabidiol (CBD), the major non-psychoactive cannabinoid, has been formulated as a highly purified oral solution (Epidiolex) and received approval by the US Food and Drug Administration (FDA) for treatment of two severe forms of epilepsy (Lennox-Gastaut syndrome and Dravet syndrome) in pediatric patients.

With an increase in interest and use of cannabis, the likelihood of concomitant use of cannabis/cannabinoids and conventional medications also increases and with it, a higher risk of drug-drug interactions (DDIs). Importantly, medical cannabis users are typically treating chronic medical conditions that are only partially responsive to conventional medications and may be more likely to use prescription medications with medical cannabis (Caputi and Humphreys, 2018). We recently identified THC, CBD, and cannabinol as potent in vitro inhibitors of carboxylesterase 1 (CES1), and THC and CBD were predicted to pose a risk of DDI when consumed concurrently with CES1 substrate medications (Qian et al., 2019a; Qian et al., 2019b).

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ABBREVIATIONS: ADHD, attention-deficit/hyperactivity disorder; AUC, area under drug concentration-time curve; CBD, cannabidiol; CES1, carboxylesterase 1; C_{max} , maximum concentration; DDI, drug-drug interaction; FDA, US Food and Drug Administration; f_m , fraction metabolized by individual enzymes; $f_{u,inc}$, unbound fraction in incubation mixture; HLS9, human liver S9; $IC_{50,u}$, unbound inhibitor concentration that achieves 50% inhibition; IR-MPH, immediate release methylphenidate; K_i , inhibition constant; $K_{i,u}$, unbound inhibition constant; K_m , the Michaelis-Menten constant; LC-MS/MS, liquid chromatography-tandem mass spectrometry; MPH, methylphenidate; P450, cytochrome P450; PBPK, physiologically based pharmacokinetics; PE, prediction error; PK, pharmacokinetics; RA, ritalinic acid; TDI, time-dependent inhibition; THC, Δ^9 -tetrahydrocannabinol V_{max} , the maximum metabolite formation rate.

CES1 is an esterase predominantly expressed in the liver and is responsible for the majority of hepatic hydrolytic activity (Imai et al., 2006; Crow et al., 2007; Taketani et al., 2007; Basit et al., 2020). When examined by the absolute abundance in the liver, the expression of CES1 exceeds that of any major phase I or phase II drug-metabolizing enzymes (i.e., cytochrome P450s (P450s) and UDP-glucuronosyltransferases) suggesting its essential role in the disposition of both endogenous compounds and xenobiotics (He et al., 2019). Indeed, therapeutic agents serving as CES1 substrates include one or more medications from almost all drug classes (Her and Zhu, 2020; Qian and Markowitz, 2020).

Methylphenidate (MPH) is almost exclusively eliminated by CES1 (Faraj et al., 1974; Redalieu et al., 1982; Sun et al., 2004; Zhu et al., 2008). A typical MPH formulation is a racemic mixture of *d*- and *l*-threo enantiomers (i.e., *d*-MPH and *l*-MPH), with *d*-MPH being the biologically active isomer (Patrick et al., 1987). MPH is highly soluble and has almost complete absorption after oral administration (Faraj et al., 1974). However, the absolute oral bioavailability of MPH is relatively low (~20% and 5% for *d*-MPH and *l*-MPH, respectively), suggesting extensive presystemic metabolism (Srinivas et al., 1993). After oral administration, the majority of MPH is recovered in the urine in the form of its inactive metabolite ritalinic acid (RA) (Faraj et al., 1974; Patrick et al., 1981; Redalieu et al., 1982), and this clearance pathway (with preference on *l*-MPH) was identified as metabolism by CES1 (Sun et al., 2004; Patrick et al., 2007; Zhu et al., 2008). Accordingly, impairment of CES1 activity by either genetic polymorphism or concurrent administration with medications serving as metabolic inhibitors of CES1 may result in an increase in the exposure to MPH. This hypothesis finds support in several clinical studies (Patrick et al., 2007; Patrick et al., 2013; Zhu et al., 2017). A schematic representing inhibition of CES1-mediated hydrolysis of MPH by THC and CBD was shown in Fig. 1.

MPH is prescribed as a first-line medication for patients with attention-deficit/hyperactivity disorder (ADHD) (Conners, 2002; Piper et al., 2018; Wolraich et al., 2019), a neuropsychiatric disorder that has an onset typically in childhood but can persist well into adulthood (Wilens and Spencer, 2010; Wolraich et al., 2019). Concomitant use of MPH with cannabis products is likely to occur under several circumstances. “Self-medication” with cannabis products for ADHD symptoms is reported to be relatively common (Mitchell et al., 2016), and the potential therapeutic effects of cannabis products in ADHD have been

investigated and reported (Strohbeck-Kuehner et al., 2008; Cooper et al., 2017; Hupli, 2018; Hergenrather et al., 2020). In addition, over-the-counter availability of CBD makes it very accessible to the general public as well as ADHD patients (Smith et al., 2021).

In the present study, we assessed the inhibition kinetics of MPH metabolism by the major cannabinoids THC and CBD using an established *in vitro* human liver S9 (HLS9) system and predicted the magnitude of clinical interactions by both static and physiologically based pharmacokinetic (PBPK) models.

Materials and Methods

Materials. Racemic MPH, phenacetin, and bovine serum albumin (BSA) were purchased from MilliporeSigma (St. Louis, MO). THC, CBD, and RA analytical standards were purchased from Cayman Chemical (Ann Arbor, MI). HLS9 pooled from 200 donors (XTreme 200) was purchased from Sekisui Xeno-Tech (Kansas City, KS). HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid) was purchased from Thermo Fisher Scientific, Inc. (Waltham, MA). All other used chemicals were of the highest analytical grade.

General In Vitro Assay Conditions. In the *in vitro* assay, racemic MPH was incubated with HLS9 which efficiently converted MPH into its metabolite RA. The formation of RA was monitored as a surrogate of MPH metabolism. The incubation mixture contained varying concentrations of MPH, 250 µg/ml of HLS9, 0.2% BSA, and 20 mM of HEPES. Consistent with previously reported studies (Sun et al., 2004; Aresti-Sanz et al., 2020), significant spontaneous hydrolysis of MPH was observed in various buffered aqueous solutions tested in our study. The least amount of RA was spontaneously formed in HEPES, and, accordingly, it was chosen as the buffer system for the studies. Similarly, BSA was used, which has the effect of limiting the relative fraction of MPH hydrolyzed spontaneously compared with MPH hydrolyzed by enzymes. The reaction was initiated by addition of HLS9 to the incubation mixture at 37°C for a period of 20 minutes. Our preliminary results indicated that the reaction was linear with respect to the selected enzyme concentration and incubation time. After 20 minutes, the reactions were terminated by adding acetonitrile (4-fold volume of the incubation mixture) containing 2.5 µM phenacetin as the internal standard and 1% formic acid. Formic acid was added to increase the stability of MPH during sample analysis (Aresti-Sanz et al., 2020). The resulting mixture was vortexed and centrifuged at 16,100 × g for 10 minutes to precipitate the protein. The supernatant was transferred out and diluted with water/acetonitrile containing 1% formic acid before liquid chromatography-tandem mass spectrometry (LC-MS/MS) analysis.

Evaluation of Inhibition Mechanism. The mechanism of inhibitory effects exerted by THC and CBD on MPH hydrolysis was determined in a time-dependent inhibition (TDI) assay. In this assay, samples were divided into either the preincubation group or control group, and there was an extra preincubation period prior to the typical incubation period described in the “General In Vitro Assay Conditions” section. In the preincubation period, HLS9 was incubated either with (preincubation group) or without (control group) inhibitor (i.e., THC and CBD, final concentration 0–31.8 µM) for 30 minutes. Once the preincubation period ended, MPH was added to both groups to initiate the reactions. Inhibitor was compensated to the control group accordingly to match the inhibitor concentrations between preincubation and control groups. Both groups contained the same mixture components in the incubation period: 75 µM substrate (MPH), 250 µg/ml HLS9, 0.2% BSA, 20 mM HEPES, and 1% (v/v) DMSO. THC and CBD have limited aqueous solubility and were thereby predissolved in DMSO. The DMSO concentrations (v/v) in all samples were adjusted to 0.5% during the preincubation period and 1% during the incubation period to control potential bias introduced by this organic solvent. The incubation lasted for 20 minutes, after which the reactions were terminated by addition of acetonitrile (4-fold incubation volume) containing 2.5 µM phenacetin and 1% formic acid. The resulting mixture was then prepared for LC-MS/MS analysis as described above in the “General In Vitro Assay Conditions” section. This study was conducted in a single experiment with triplicate samples.

Since some spontaneous hydrolysis of MPH during incubation was observed, the measured RA formation was corrected by control samples with identical experimental setup and MPH concentrations but without HLS9. The resulting rate of RA formation was expressed as a percentage ratio (R_c) of the one in sample

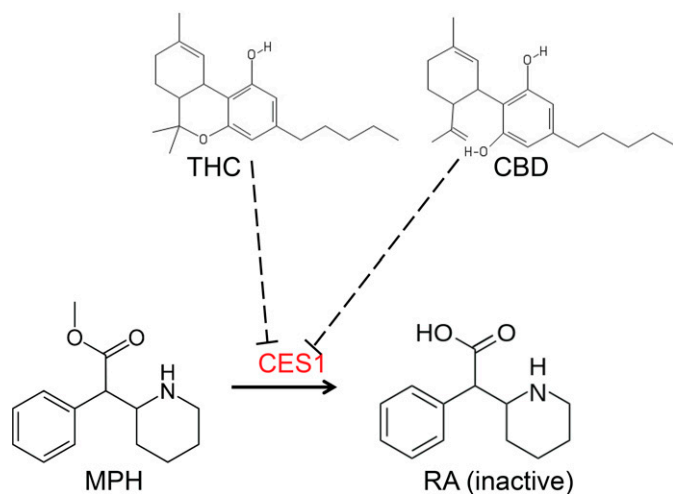


Fig. 1. Chemical structures of MPH, RA, THC, and CBD and hypothesis of CES1-mediated interactions between MPH and THC/CBD. Dashed lines with flat arrows represent inhibition.

without inhibitor. A nonlinear regression analysis (eq. 1) was performed using R_v as the dependent variable and unbound concentrations ($[I_u]$) of either THC or CBD as the independent variable:

$$R_v = 100 \times \left(1 - \frac{I_{max} \cdot [I_u]^b}{[I_u]^b + IC_{50,u}^b} \right) \quad (1)$$

where the iterated parameters were: $IC_{50,u}$, the unbound half maximal inhibition concentration; I_{max} , the maximal degree of inhibition; and b , a shape exponent. The estimated $IC_{50,u}$ was further converted to $IC_{50,u}$ (unbound inhibitor concentration that achieves 50% R_v) using eq. 2:

$$IC_{50,u} = IC_u / (2I_{max} - 1)^{1/b} \quad (2)$$

A TDI (i.e., irreversible inhibition of mechanism) is concluded when a decrease of ≥ 1.5 -fold $IC_{50,u}$ is observed with a 30-minute preincubation (Grimm et al., 2009).

Inhibition Kinetics of MPH Metabolism by THC and CBD. The inhibition of MPH hydrolysis by THC and CBD was further investigated in in vitro assays in which the kinetic parameters of the inhibition were estimated for determination of inhibition types and potency. The assay conditions and procedures have been described in the "General In Vitro Assay Conditions" section with the exception of inclusion of inhibitors (i.e., THC and CBD). The incubation mixture comprised combinations of varying MPH concentrations (0, 25, 50, 100, 200, and 400 μ M) and cannabinoid concentrations (0–19.1 μ M for THC and 0–25.4 μ M for CBD). The final DMSO concentration (v/v) was 1%. Incubation mixtures containing 0–400 μ M MPH and 1% DMSO without the presence of either cannabinoid were used as control samples. The reactions were initiated by addition of HLS9 into the incubation mixture at 37°C and lasted for 20 minutes. Afterward, acetonitrile (4-fold incubation volume) containing 2.5 μ M phenacetin and 1% formic acid was added to terminate the reactions. The resulting mixture was then prepared for LC-MS/MS analysis as described above in the "General In Vitro Assay Conditions" section. This study was conducted in three independent experiments with duplicate samples in each.

Similar to the TDI study, the RA formed in the control samples without HLS9 was subtracted from the one measured in the samples with the same substrate concentrations to account for spontaneous hydrolysis of MPH. The kinetic parameters were estimated without weighting using a mixed competitive-non-competitive model described by eq. 3 (Segel, 1975; Greenblatt et al., 2011):

$$V = \frac{V_{max} \cdot [S]}{K_m \left(1 + \frac{[I_u]}{K_{i,u}} \right) + [S] \left(1 + \frac{[I_u]}{\alpha \cdot K_{i,u}} \right)} \quad (3)$$

The dependent variable (V) is the formation rate of RA and the independent variables included MPH ($[S]$) and unbound cannabinoid ($[I_u]$) concentrations. The iterated parameters were: V_{max} , the maximum metabolite formation rate; K_m , the Michaelis-Menten constant; $K_{i,u}$, the unbound inhibition constant for inhibitor which is an indicator of inhibition potency; and α , an indicator of the inhibition type. An estimated α value approaching positive infinity

indicates competitive type of inhibition, while a value equal to 1 suggests a noncompetitive type of inhibition. The inhibition type was further verified by Lineweaver-Burk plots.

Binding of THC and CBD in In Vitro Assay. Excessive nonspecific binding of cannabinoids to both protein and test tube wall has been previously documented (Garrett and Hunt, 1974; Patilea-Vrana and Unadkat, 2019; Qian et al., 2019b). Given that 0.2% BSA was included in the in vitro incubation, ignoring this factor will result in an underestimated inhibition potency of the tested cannabinoids. Accordingly, the unbound concentrations of THC and CBD in the in vitro assay were calculated using the tube adsorption method (Patilea-Vrana and Unadkat, 2019). In the binding assay, two groups (protein group and buffer group) of samples were prepared. In the protein group, the composition of the incubation mixture (i.e., 250 μ g/ml of HLS9, cannabinoids, 0.2% BSA, and 20 mM of HEPES) was identical to that used in the kinetic study described above with the exception of MPH which was not added. In the buffer group, the incubation mixture contained all the components in the protein group except for HLS9 and BSA. The mixture was placed in a water bath maintained at 37°C and incubated for 20 minutes. Afterward, in each group, samples were further split into two treatments. In the first treatment (Treatment A), the incubation mixture was transferred to a twofold volume of acetonitrile (containing phenacetin as the internal standard) in a new tube. In the second treatment (Treatment B), a same twofold volume of acetonitrile solution was directly added to the incubation mixture. Addition of acetonitrile was presumed to recover essentially all of the cannabinoid adsorbed to the tube wall. The resulting mixture was vortexed and centrifuged at 16,100 \times g for 10 minutes. The supernatant was further diluted with acetonitrile/water and subject to LC-MS/MS analysis. This study was conducted in three independent experiments with duplicate samples each.

To estimate unbound fractions of THC and CBD in the incubation mixture, a binding model (Supplemental Fig. 1) was proposed as a simplification of the binding process, where the cannabinoid was assumed to be in unbound form ($[S]$), bound to the tube wall (SW_b in the buffer group and SW_p in the protein group), or bound to the protein (SP) and rapid equilibrium among these three forms was assumed. The unbound fractions of THC and CBD over the range of concentrations studied in the in vitro assays were not considered solubility limited.

The binding between cannabinoid and tube wall was studied in the buffer group in the absence of protein. The binding data were described by the following equation using nonlinear regression analysis:

$$SW_b = \frac{W_T \cdot [S]}{\frac{1}{K_w} + [S]} \quad (4)$$

where SW_b and $[S]$ represented the cannabinoid bound to the tube wall and unbound cannabinoid concentration, respectively. The estimated parameters were: K_w , the binding constant between cannabinoid and tube wall; and W_T , the total tube wall ($W + SW_b$) available for cannabinoid binding.

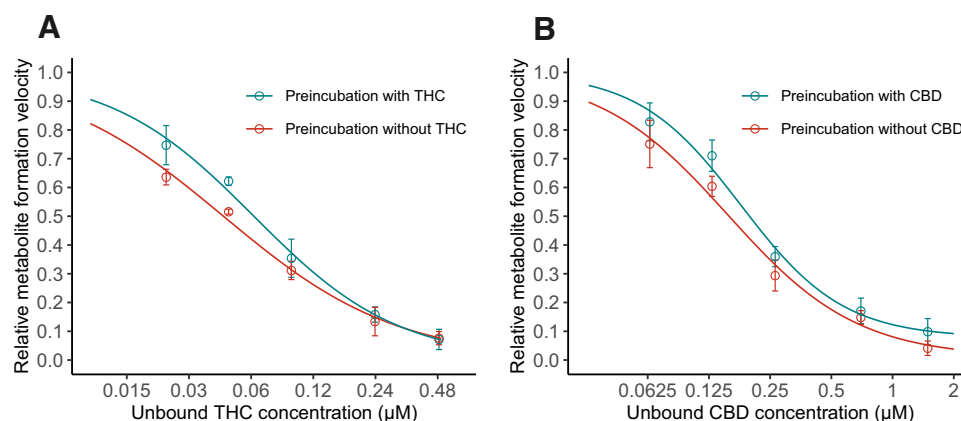


Fig. 2. Inhibition curves for THC (A) and CBD (B) in time-dependent inhibition study. HLS9 (250 μ g/ml) was preincubated with or without cannabinoids for 30 minutes before addition of MPH. The formation rate of metabolite (ritalinic acid) from MPH was expressed as a relative value over the control sample without addition of cannabinoids. Data points represent the mean (\pm S.D.) of triplicate samples from a single experiment. Lines represent model prediction.

TABLE 1

Inhibition of MPH hydrolysis by THC and CBD in time-dependent studies.

Cannabinoids	IC _{50,u} (μM)		A/B
	Preincubation: 0 min (A)	Preincubation: 30 min (B)	
THC	0.0441 ± 0.0074	0.0618 ± 0.0097	0.71
CBD	0.157 ± 0.027	0.198 ± 0.029	0.79

IC_{50,u}, unbound inhibitor concentration that achieves 50% activity. IC_{50,u} values were presented as estimate ± SE (uncertainty) from nonlinear regression analysis using data from a single experiment with three replicate samples.

When the protein was added (the protein group), the free cannabinoid cannot be directly measured. Instead, the measured concentration in solution included both free cannabinoid (S) and the fraction bound to the protein (SP). By contrast, the fraction of cannabinoid bound to tube wall (SW_p) could still be measured (difference in cannabinoid concentration between Treatments A and B). The binding process could be described by a new equation integrating the protein component [derivation of equation provided in (Supplemental Appendix A)]:

$$SW_p = \frac{\left(S_T + W_T + \frac{K_p \cdot [P]}{K_w} + \frac{1}{K_w}\right) - \sqrt{\left(S_T + W_T + \frac{K_p \cdot [P]}{K_w} + \frac{1}{K_w}\right)^2 - 4W_T \cdot S_T}}{2} \quad (5)$$

The dependent variable was the measured cannabinoid bound to the tube wall (SW_p), and the independent variable was the total cannabinoid concentration (S_T = S + SW_p + SP) in the incubation mixture, which was measured as the cannabinoid concentration in the sample from the protein group with Treatment B. W_T and K_w had been estimated in eq. 4 and were used as constants here. K_p represented the binding constant between cannabinoid and protein. [P] was another variable representing the free concentration of protein in the incubation mixture. Since [P] could not be measured, the term K_p × [P] was estimated as a single parameter with the assumption that [P] was constant regardless of cannabinoid concentrations. This assumption appeared to be valid based on the large amount of BSA (0.2%) added in the incubation mixture.

It was determined that neither THC nor CBD could be completely recovered with Treatment B. Adsorption onto the inner layer of tube wall might be a potential cause. Consequently, in our analysis, empirical equations were employed to account for the observed loss of cannabinoids.

Once the parameters were estimated, the unbound concentrations of cannabinoid ([S]) in the incubation mixture of inhibition studies could be calculated utilizing an equation derived from eq. 4 by assuming same interaction between cannabinoid and tube wall regardless of the presence of protein:

$$[S] = \frac{SW_p}{K_w \cdot (W_T - SW_p)} \quad (6)$$

where K_w and W_T are estimated from eq. 4, and SW_p can be calculated from eq. 5 based on the total cannabinoid added to the incubation (i.e., S_T).

LC-MS/MS Analysis. The concentrations of RA formed during in vitro incubations and cannabinoids from the in vitro binding assay were determined by LC-MS/MS. The system consisted of a Shimadzu high-performance liquid chromatography system (Shimadzu, Kyoto, Japan) coupled to an AB Sciex API 3000 triple-quadrupole mass spectrometer (Applied Biosystems, Foster City, CA). Chromatographic separation was achieved on an analytical column (Aqua, 50 × 2.0 mm, 5 μm, Phenomenex, Inc., Torrance, CA). A gradient mobile phase was employed, comprised of water with 0.1% formic acid as the aqueous phase and methanol as the organic phase. The total flow rate of the mobile phase was 0.25 ml/min. Mass spectrometry was performed in positive mode with

electrospray as the ionization method. The monitored mass transitions for RA, THC, CBD, and the internal standard phenacetin were m/z 220 → 84, 315 → 193,

315 → 193, and 180 → 110, respectively. The lowest standards used in our study were 5 nM, 79 nM, and 79 nM for RA, THC, and CBD, respectively. The range and linearity of calibration standards and the accuracy and precision of low, median, and high standards for each analyte are provided in (Supplemental Table 1). Representative chromatograms of RA, THC, and CBD are provided in (Supplemental Fig. 2).

Prediction of Clinical Interactions by Static Models. To evaluate the possibility of interactions between MPH and tested cannabinoids rising to clinical significance, a ratio of intrinsic clearance (R₁) was calculated by eq. 7 (FDA Center for Drug Evaluation and Research, 2020):

$$R_1 = 1 + \frac{I_{max,u}}{K_{i,u}} \quad (7)$$

where I_{max,u} is the maximal unbound physiological concentrations of THC and CBD at steady-state, and K_{i,u} is the unbound inhibition constant obtained from in vitro study.

R₁ was then compared with a cut-off value of 1.02 as recommended in the FDA guidance referenced above.

Since both R₁ values for THC and CBD exceeded the cut-off value, the magnitude of DDI was further evaluated using a static mechanistic model. In the model, a ratio of area under the plasma concentration-time curve (AUCR) of MPH in the presence to absence of cannabinoids was predicted using the equation provided in the FDA guidance document with minor modification (FDA Center for Drug Evaluation and Research, 2020):

$$AUCR = \frac{1}{1 + \frac{[I]_h}{K_{i,u}} \cdot f_m + (1 - f_m)} \quad (8)$$

where [I]_h is the maximum unbound concentration of cannabinoid in the liver. f_m is the fraction of MPH that is metabolized by CES1. An f_m value of 80% was used based on a PK study in which the radio-labeled metabolites of MPH excreted in urine were measured (Faraj et al., 1974). K_{i,u} is the unbound inhibition constant estimated from the in vitro inhibition kinetic study. The intestinal component of a typical static model was not considered due to the unknown fraction of MPH available after intestinal metabolism. Per FDA guidance, this fraction is assumed to be 1 when the data are not available, which consequently leads to inattention to DDI in intestine (FDA Center for Drug Evaluation and Research, 2020). Notably, minimal influence from inhibition of intestinal CES1 is expected due to the relatively low abundance of CES1 as compared with that in the liver (Taketani et al., 2007; Basit et al., 2020).

For both THC and CBD, only drug concentrations achieved systemically (i.e., plasma/serum/blood concentrations) were used as the surrogates for [I]_h in eq. 8. Although the reported maximum concentration of CBD was achieved by the oral route (Taylor et al., 2018), a maximum unbound concentration of CBD at portal vein was not estimated in the static model due to a paucity of information around the fraction of absorption, intestinal bioavailability, and absorption rate of CBD with this formulation. Instead, PBPK models were constructed for a more mechanistic prediction of clinical interactions between MPH and CBD.

Prediction of Clinical Interactions by PBPK Models. To obtain a more mechanistic prediction of DDIs between MPH and CBD under various clinical scenarios, PBPK models were built for both immediate-release (IR) MPH (Ritalin) and an oral solution formulation of CBD (Epidiolex). Pharmacokinetic (PK) studies of MPH and CBD using human subjects were searched in the public domain and summarized. Only PK data with Ritalin and Epidiolex were collected with the exception of data with intravenous administration route. A total of 17

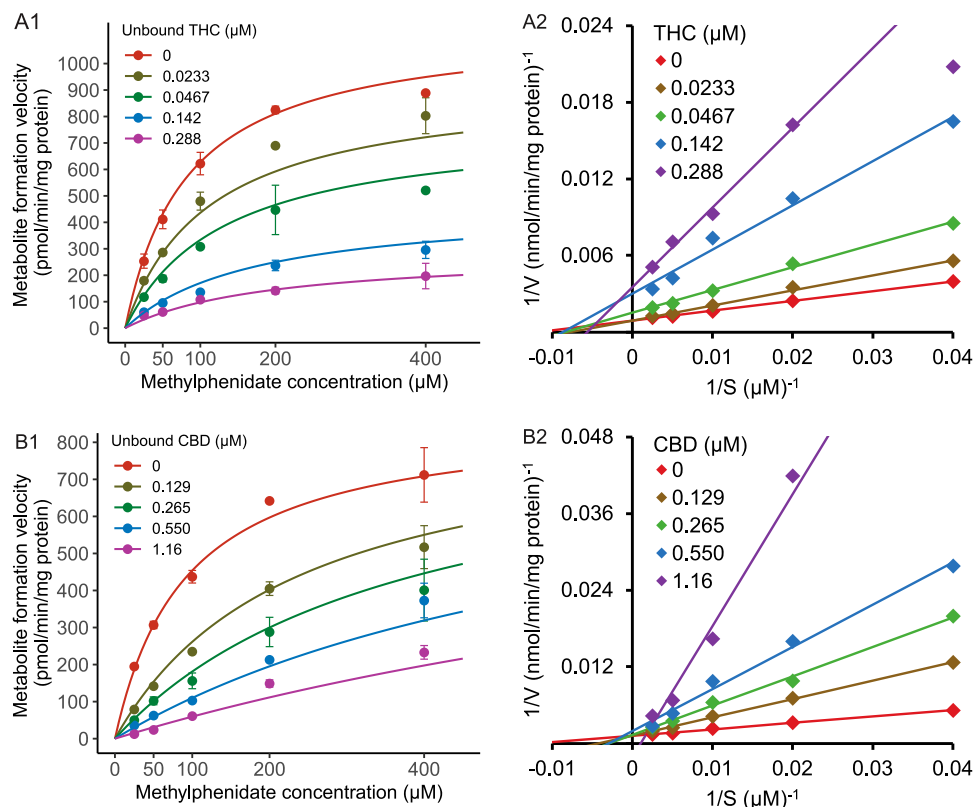
TABLE 2

Determination of kinetic parameters for inhibition of MPH hydrolysis by THC and CBD.

Cannabinoids	K _m (μM)	K _{i,u} (μM)	α	V _{max} (pmol/min/mg protein)
THC	85.7 ± 12.9	0.031 ± 0.003	16.4 ± 14.7	1056 ± 77
CBD	90.6 ± 10.1	0.091 ± 0.004	>9999	940 ± 150

All values were presented as mean ± S.D. of three independent experiments with duplicate samples.

Fig. 3. Representative analyses of inhibition kinetics and Lineweaver-Burk plots for THC (A1, A2) and CBD (B1, B2). Incubation was conducted using 250 $\mu\text{g/ml}$ HLS9 with combinations of varying concentrations of MPH and cannabinoids. Data were from one of three independent experiments with duplicate samples. Points represent mean values (\pm S.D.) of duplicate samples.



(21 datasets) and 5 (8 datasets) studies were selected for MPH and CBD, respectively. The demographics of subjects in each study was summarized in (Supplemental Tables 2 and 3).

A “middle-out” approach was implemented during model development for both MPH and CBD. The system-specific parameters were mainly derived from the default database of the PBPK software, and drug-specific parameters were obtained from various sources as well as software prediction (Supplemental Tables 4 and 5). Generally, data from intravenous administrations were used first to inform the in vivo clearance and volume of distribution. Components such as dissolution, intestinal permeability, and presystemic metabolism were then added and refined using the datasets with oral formulation. A general workflow of the model development and verification is summarized in (Supplemental Fig. 3).

For MPH, enzyme kinetic data from a reported in vitro study were incorporated (Sun et al., 2004). f_m of MPH by CES1 was fixed at 80% based on the fraction of MPH excreted as RA in urine after both intravenous and oral dose of radio-labeled MPH (Faraj et al., 1974). The remaining MPH was presumed to be cleared by P450 enzymes with unidentified individual isoforms. In the PBPK model, a dummy non-CES1 enzyme was created to account for the remaining 20% clearance of MPH. The intrinsic clearance of MPH by CES1 and the non-CES1 enzyme were first calculated based on the in vivo plasma clearance of MPH observed from a study with intravenous dosing route (Srinivas et al., 1993). However, PK studies with oral doses consistently showed shorter terminal half-lives than the only available intravenous data (Supplemental Fig. 4). Therefore, the data with intravenous administration were not used in the model development. Instead, an in silico lipophilicity value was used without optimization to inform drug distribution and the V_{max} values were optimized based on the datasets with oral formulation. Features of competitive inhibition of CES1 by *d*- and *l*-MPH was also incorporated into the model, and the K_m values obtained in vitro (Sun et al., 2004) were used as K_i . The abundance of CES1 in the liver and intestine was obtained from a proteomics study (Basit et al., 2020).

For CBD, CYP3A4, CYP2C19, and CYP2C9 are the major enzymes responsible for its oxidative metabolism (Jiang et al., 2011; Beers et al., 2021). The f_m of CBD by the respective isoforms have been reported based on an in vitro CBD depletion assay (Beers et al., 2021). These f_m values are 0.54, 0.31, and 0.15 for CYP3A4, CYP2C19, and CYP2C9, respectively. A similar f_m value (0.46) for

CYP3A4 can be estimated (Supplemental Appendix B) based on the relative CBD exposure (area under drug concentration-time curve, AUC) after an oral administration of 1:1 THC:CBD either with or without a strong CYP3A4 inhibitor (ketoconazole) (Stott et al., 2013). In this study, the f_m values reported by Beers et al. were used. Then, further based on the in vivo plasma clearance of CBD (74.4 L/h) after an intravenous dose (Ohlsson et al., 1986), the intrinsic clearance of CBD by CYP3A4, CYP2C19, and CYP2C9 were calculated as 4.62, 15.1, and 1.44 $\mu\text{l}/\text{min}/\text{pmol}$ protein, respectively. The details of the back-calculation method are provided in (Supplemental Appendix B). Meanwhile, mechanism-based inhibition of CYP3A4 and CYP2C19 and reversible inhibition of CYP2C9 by CBD have been reported (Bansal et al., 2020), which was also incorporated into our PBPK model. The hepatic abundance of CYP3A4, CYP2C19, and CYP2C9 was set using system-default, while their abundance in intestine was scaled based on a proteomics study (Drozdzik et al., 2018). There is a paucity of information regarding the dissolution and absorption of CBD, which appears to be solubility limited. Accordingly, two Weibull functions were empirically implemented to describe this process.

In both models, the lipophilicity of the compounds was input to enhance the prediction of a number of parameters, such as intestinal and tissue permeability. For calculation of drug distribution, the method described by Rodgers and Rowland (Rodgers and Rowland, 2006; Rodgers and Rowland, 2007) was selected for MPH, and the method described by Schmitt (Schmitt, 2008) was selected for CBD. As aforementioned, the lipophilicity value of MPH was used without optimization, while the lipophilicity value of CBD was optimized due to a substantial underprediction of drug exposure when compared with observed data with intravenous administration (Ohlsson et al., 1986). Selection of lipophilicity for optimization was guided by physiologic plausibility, sensitivity analysis (Supplemental Fig. 5), and the ability of the new estimate to describe observed data (Supplemental Fig. 6). The optimized lipophilicity value (3.15) is different from the reported in silico LogP of CBD (approximately 5.9) (Nelson et al., 2020). However, it has been noted that methods for drug distribution may not be very reliable for highly lipophilic compounds (Rodgers and Rowland, 2007). It has been documented that predicted distribution may be vulnerable to experimental error, uncertainty from in silico predictions, and overprediction of tissue distribution for lipophilic compounds

TABLE 3
Model evaluation for MPH.

Dose	N	AUC (ng·h/ml)			C _{max} (ng/ml)			
		Mean observed	Mean predicted	PE (%)	Mean observed	Mean predicted	PE (%)	
<i>d/dl</i> -MPH								
Calibration Datasets								
(Midha et al., 2001)	40 mg Ritalin	24	59.1	90.8	53.7	11.7	16.9	45.1
(Midha et al., 2001)	40 mg Ritalin (fed)	24	68.1	103	51.2	14.3	15.6	9.1
(Patrick et al., 2007)	0.3 mg/kg Ritalin	20	82.9	54.5	-34.3	15.3	8.5	-44.4
(Spencer et al., 2006)	40 mg Ritalin	12	81.9 ^b	97.1	18.5	14.1	15.7	11.3
(Teo et al., 2004)	20 mg Focalin	15	121	96.1	-20.5	23.7	18.1	-23.7
(Teo et al., 2004)	20 mg Focalin (fed)	15	132	99.6	-24.5	22.1	15.5	-30.0
(Patrick et al., 2013)	0.3 mg/kg Ritalin	24	52.1	53.4	2.5	10.1	8.2	-18.8
(Patrick et al., 2013)	0.15 mg/kg Focalin	24	53.7	53.4	-0.6	10.7	8.2	-23.4
(Parasrampur et al., 2007)	50 mg Ritalin	49	106	125	17.9	21.1	24.0	13.7
(Parasrampur et al., 2007)	90 mg Ritalin	49	195	226	15.9	38.8	43.2	11.3
Verification Datasets								
(Hysek et al., 2014)	60 mg Ritalin	15	175 ^a	172	-1.9	30.4	32.4	6.6
(Abbas et al., 2016)	40 mg ^c Ritalin	29	132	202	52.6	15.6	22.9	46.8
(Adjei et al., 2014)	25 mg ^d Ritalin	24	282	188	-33.3	29.1	18.0	-38.1
(DeVane et al., 2000)	10 mg Ritalin	6	23.7 ^a	19.7	-16.9	4	4.5	12.5
(Koehm et al., 2010)	20 mg Ritalin	9	35	43	22.9	7	7.1	1.4
(Meyer et al., 2000)	20 mg Ritalin	20	35.8	48.9	36.5	7.6	8.8	15.9
(Patrick et al., 1989)	10 mg ^e Ritalin	18	42.0	45.3	7.8	6.4	4.8	-25.3
(Spencer et al., 2012)	40 mg ^f Ritalin	26	179 ^a	185	3.3	25.4	22.6	-10.8
(Stage et al., 2017)	10 mg Ritalin	16	21.4 ^g	23.2 ^g	18.7	5 ^g	3.9 ^g	-19.2
(Wong et al., 1998)	40 mg Ritalin	21	93.6	90.1	-3.7	17.8	16.4	-7.9
Weighted PE (%)								
11.7								
<i>l</i> -MPH								
(Wong et al., 1998)	40 mg Ritalin	21	1.51	1.33	-11.9	0.821	0.54	-34.2
(Patrick et al., 2013)	0.3 mg/kg Ritalin	24	0.77	0.72	-6.5	0.18	0.22	22.2
(Patrick et al., 2007)	0.3 mg/kg Ritalin	20	0.82 ^{a,b}	0.80	-2.4	0.22 ^b	0.25	13.6
Weighted PE (%)								
-7.0								

PK parameters shown as arithmetic mean values unless otherwise noted.

^a AUC_{0-last}

^b Obtained from a non-compartmental analysis of digitized population-level data.

^c Two doses administered 6 hours apart.

^d Three doses administered 4 hours apart.

^e Two doses administered 5 hours apart.

^f Two doses administered 4 hours apart.

^g Values shown as median.

(Pearce et al., 2017). Therefore, we consider the optimization of lipophilicity for CBD justifiable.

Overall, the PBPK models were developed based on training datasets (10 for MPH and 3 for CBD). The developed PBPK models were further verified on other datasets (10 for MPH and 4 for CBD) that were not used in model development. For model evaluation, simulations were performed with virtual populations (100 subjects each) and study designs matching the reported clinical studies. PK parameters from simulation were compared with the ones observed. Prediction error (PE) was calculated as below for model evaluation (Khalil and Laer, 2014):

$$PE = \frac{Y_{predicted} - Y_{observed}}{Y_{observed}} \times 100\% \quad (9)$$

where $Y_{predicted}$ and $Y_{observed}$ represent the PK parameters summarized from simulation and observed from reported studies, respectively. PE is further weighted by number of study subjects, and a weighted PE within $\pm 25\%$ is considered acceptable.

Once the models for MPH and CBD were verified, both models were integrated into a joint model for prediction of DDI under various clinical scenarios in which immediate release (IR)-MPH (Ritalin) and the oral solution of CBD (Epidiolex) were co-administered. Inhibition of CES1 by CBD was described by the $K_{i,u}$ value obtained from this study. The non-CES1 enzymes are assumed not subject to inhibition by CBD. A virtual population comprising 100 subjects (50% female) with an age range of 18–55 were created and used in all simulations. The AUC and maximum concentration (C_{max}) of the biologically active enantiomer *d*-MPH were selected as the endpoints and compared with the one in which racemic MPH was given alone. The significance of interactions was assessed based on the changes in *d*-MPH AUC and C_{max} as perpetrated by co-administered CBD.

Software and Statistical Methods. The built-in “nl” function (NL2SOL algorithm) in R 3.6.0 (R Foundation for Statistical Computing, Vienna, Austria) was employed in the nonlinear regression analysis of the data from nonspecific binding, TDI, and inhibition kinetic studies. The model performance was evaluated by diagnostic residual plots, parameter uncertainty and plausibility, and visual check. Noncompartmental analysis was performed utilizing the “PKNCA” package in R 3.6.0. AUC was calculated using a linear trapezoidal method. The “ggplot2” package in R 3.6.0 was used for generating plots. Lineweaver-Burk plots were made using Microsoft Excel (Redmond, WA). The PBPK models were developed and verified using PK-Sim and Mobi version 9.1 (Open Systems Pharmacology, <http://www.open-systems-pharmacology.org>) software. A subset of physicochemical parameters of MPH and CBD (Supplemental Tables 4 and 5) were predicted by using either PK-Sim or ADMET Predictor v9.5.0. (Simulation Plus). Parameter optimization was performed utilizing the “Parameter Identification” module in PK-Sim and the Monte-Carlo algorithm was used in estimation.

Results

Nonspecific Binding of THC and CBD in the In Vitro Incubation Mixture. In pure buffer, THC and CBD present in the solution at the end of incubation was substantially lower than the amount added (Supplemental Fig. 7). Their concentrations increased in accordance with the increasing amount added, suggesting solubility was not the main cause of the observed loss of cannabinoids in solution. Based on the nonlinear feature of the increase, a saturable process was indicated, which matched our hypothesis of nonspecific binding to tube wall. In buffer containing proteins (BSA and HLS9), the majority of the added cannabinoids were recovered in the solution (Supplemental Fig. 7),

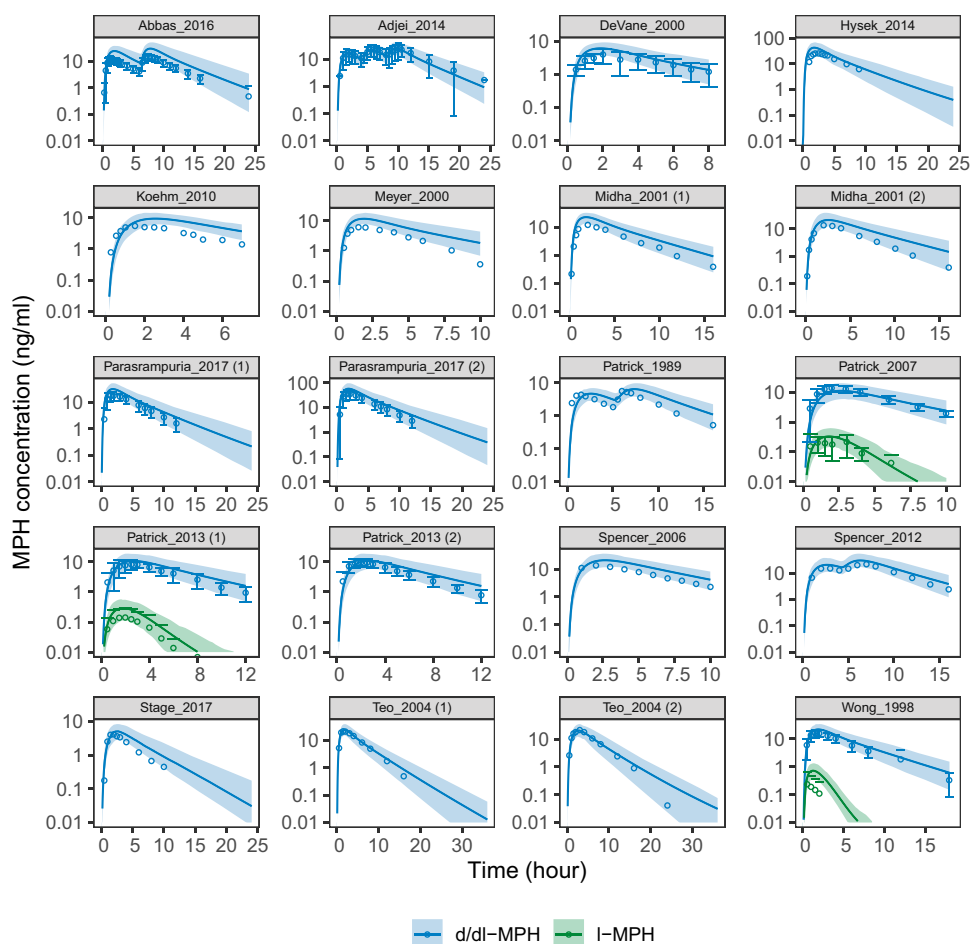


Fig. 4. PBPK model simulated and observed concentration-time profiles of MPH after oral administration. The simulated PK profiles were compared with the observed data reported in literature for assessment of model performance. Demographics of the simulated virtual individuals ($n = 100$) were matched to ones reported in the respective study. Solid lines and shades represent mean and 90% (5th–95th) prediction interval of simulated profiles, respectively. Points and error bars represent mean and S.D. of the observed data, respectively. Median values (both simulated and observed) were summarized for the Stage_2017 study.

indicating extensive protein binding. The data were well described by the proposed binding model (eq. 4 and 5, [Supplemental Fig. 1]). The unbound fractions of THC and CBD tested in the incubation mixture ($f_{u,inc}$) were then calculated (Supplemental Table 6). For THC, the added total concentrations (1.59–31.8 μM) in the incubation mixture converted to unbound concentrations in the range of 0.0233–0.487 μM ($f_{u,inc}$ approximated 1.5%). For CBD, the added total concentrations (1.59–31.8 μM) in the mixture converted to unbound concentrations in the range of 0.0640–1.49 μM ($f_{u,inc}$ approximated 4.4%).

THC and CBD Reversibly Inhibited MPH Hydrolysis. The hydrolysis of MPH mediated by HLS9 was inhibited by THC and

CBD both in a concentration-dependent manner (Fig. 2). The calculated $\text{IC}_{50,u}$ values for THC and CBD at the presence and absence of tested cannabinoids during a 30-minute preincubation period are summarized in Table 1. Based on the observation that the inhibition of MPH hydrolysis was not potentiated (i.e., lower $\text{IC}_{50,u}$) by preincubation with either THC or CBD, reversible mechanism of inhibition (i.e., not time-dependent) was concluded for both cannabinoids.

In Vitro Kinetic Study on Inhibition of MPH Hydrolysis by THC and CBD. The kinetic data were well described by the mixed competitive-noncompetitive model. The estimated parameters are

TABLE 4
Model evaluation for CBD.

Dose (Epidiolex)	N	AUC (ng-h/ml)			C_{max} (ng/ml)			
		Mean observed	Mean predicted	PE (%)	Mean observed	Mean predicted	PE (%)	
Calibration Datasets								
(Taylor et al., 2018)	1500 mg	6	1470 ^{a,b}	1486 ^a	1.1	260 ^b	214	-17.7
(Taylor et al., 2018)	6000 mg	6	3490 ^{a,b}	3076 ^a	-11.9	610 ^b	453	-25.7
(Tayo et al., 2020)	200 mg	8	435 ^{a,b}	648 ^a	49.0	118 ^b	87.0	-26.3
Verification Datasets								
(Taylor et al., 2019)	200 mg	8	522 ^{a,b}	548 ^a	5.0	96.4 ^b	86.2	-10.6
(Crockett et al., 2020)	750 mg	29	1490 ^b	1526	2.4	154 ^b	152	-1.3
(Taylor et al., 2018)	3000 mg	6	2540 ^{a,b}	2608 ^a	2.7	418 ^b	386	-7.7
(Taylor et al., 2018)	4500 mg	6	3100 ^{a,b}	2968 ^a	-4.3	677 ^b	441	-34.9
Weighted PE (%)					6.2			-12.3

PK parameters are shown as geometric mean values.

^a AUC_{0-last}

^b Obtained from a non-compartmental analysis of digitized population-level data due to the discrepancies observed between the reported PK parameters and concentration-time plots.

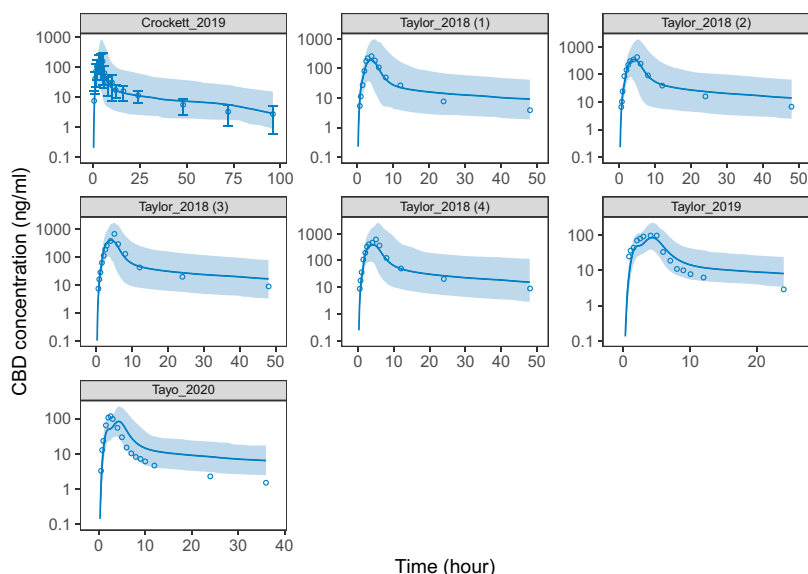


Fig. 5. PBPK model simulated and observed concentration-time profiles of CBD after oral administration. The simulated PK profiles were compared with the observed data reported in literature for assessment of model performance. Demographics of the simulated virtual individuals ($n = 100$) were matched to ones reported in the respective study. Solid lines and shades represent geometric mean and 90% (5th–95th) prediction interval of simulated profiles, respectively. Points and error bars represent geometric mean and S.D. of the observed data, respectively.

listed in Table 2 and representative kinetic profiles are shown in Fig. 3. The $K_{i,u}$ values for THC and CBD were 0.031 and 0.091 μM , respectively. The estimated α values and the Lineweaver-Burk plots (Fig. 3) suggested mixed but closer to noncompetitive inhibition type for THC and competitive inhibition type for CBD.

Prediction of Clinical Interactions between MPH and Cannabinoids by Static Models. Literature reporting the human PK of THC and CBD with various cannabis products and dosing routes were reviewed to extract the maximum attainable systemic exposure to these two cannabinoids. For THC, the highest exposure was reported in a study by Huestis and colleagues, where a mean THC plasma C_{max} of 0.515 μM was achieved in healthy subjects ($n = 6$) after smoking a cannabis cigarette containing 33.8 mg THC (Huestis et al., 1992).

For CBD, the highest exposure (plasma $C_{\text{max}} = 2.33 \mu\text{M}$) was observed after healthy volunteers ($n = 9$) took the second oral dose of 750 mg twice daily Epidiolex (Taylor et al., 2018). The unbound plasma C_{max} ($I_{\text{max,u}}$) values were further calculated by multiplying the C_{max} by the unbound fraction of THC (0.028) (Garrett and Hunt, 1974) and CBD (0.04) in plasma (FDA Center for Drug Evaluation and Research, 2018b). Based on the $I_{\text{max,u}}$ and $K_{i,u}$ values, The R_1 values were calculated as 1.46 and 2.54 for THC and CBD, respectively, suggesting possible clinical DDI and necessity of further investigations. The likelihood of clinical DDIs was then evaluated by a static mechanistic model, which predicted AUCR values of 1.34 and 1.94 for THC and CBD, respectively.

Development of PBPK models for MPH and CBD. The MPH model was developed and verified using datasets shown in Table 3. The weighted PE for both *d*-MPH (11.7% for AUC and -0.8% for C_{max}) and *l*-MPH (-7.0% for AUC and 1.3% for C_{max}) were within the prespecified acceptance range. Plots of model-predicted versus observed concentration-time profiles for all MPH datasets are provided in Fig. 4. The CBD model was developed and verified using datasets shown in Table 4. The weighted PE were 6.2% for AUC and -12.3% for C_{max} , which also met prespecified acceptance criteria. Plots of model-predicted versus observed concentration-time profiles for all CBD datasets are provided in Fig. 5.

Prediction of Clinical Interactions between MPH and CBD by PBPK Models. To evaluate the likelihood of a clinical interaction between MPH and CBD, virtual scenarios in which single doses of IR-MPH (10 mg) and prescriptive oral solution of CBD (0 – 10 mg/kg) were administered simultaneously were simulated. The AUC and C_{max} of MPH caused by highest concomitant CBD dose (10 mg/kg) increased by 22% and 10%, respectively (Fig. 6).

CBD dose (10 mg/kg) was then simulated with a time interval either before or after the MPH dose to explore effects of dosing time. A maximum magnitude of interactions was predicted when the CBD dose was given between 1 and 4 hours prior to the MPH dose (Fig. 6). Under this scenario, the changes in MPH AUC and C_{max} could be above 31% and 24%, respectively.

Finally, multiple doses of *dl*-MPH (10 mg twice daily with a 4-hour interval) and CBD (0, 5 or 10 mg/kg every 12 hours) were simulated for 7 days to assess the outcomes from long-term co-administration. An increasing trend of interaction was predicted over the simulated duration (Fig. 7). On Day 7, as compared with treatment with placebo (i.e., 0 mg/kg CBD), concomitant use of 5 mg/kg CBD increased MPH AUC₂₄ (AUC over 24-hour interval) and C_{max} by 38% and 31%, respectively. Similarly, the higher simulated CBD dose (10 mg/kg) exerted a mild influence, with a predicted increase in MPH AUC₂₄ and C_{max} by 55% and 45%, respectively.

Discussion

In the present study, potent reversible inhibition of MPH hydrolysis by THC and CBD with $K_{i,u}$ values in the nanomolar range were identified in vitro. Since MPH is almost exclusively metabolized by CES1, the observed inhibitory effects are presumed to be mediated by inhibition on CES1. This observation is consistent with our prior work utilizing recombinant CES1 (Qian et al., 2019b). The essential role CES1 plays in the PK of MPH is most obviously demonstrated by an approximately 150% increase in MPH exposure observed in subjects carrying loss-of-function *CES1* variants as compared with their non-carrier peers (Patrick et al., 2007; Zhu et al., 2008; Stage et al., 2017). In addition, a recently published study revealed an inverse correlation between plasma CES1 (an hypothesized indicator of hepatic CES1 activity) and MPH exposure (Shi et al., 2021). Although essentially no direct clinical assessments have been conducted to assess the influence of CES1 inhibitors on the PK of co-administered CES1 substrate drugs, reduced CES1 activity associated with loss-of-function *CES1* variants has been documented to significantly increase drug exposures in patients treated with a number of CES1 substrate drugs, including MPH (Zhu et al., 2008; Stage et al., 2017), clopidogrel (Lewis et al., 2013; Tarkiainen et al., 2015), and oseltamivir (Tarkiainen et al., 2012). Similarly, impairment of CES1 activity by concomitant drugs is expected to increase exposure to MPH and potentially lead to a higher likelihood

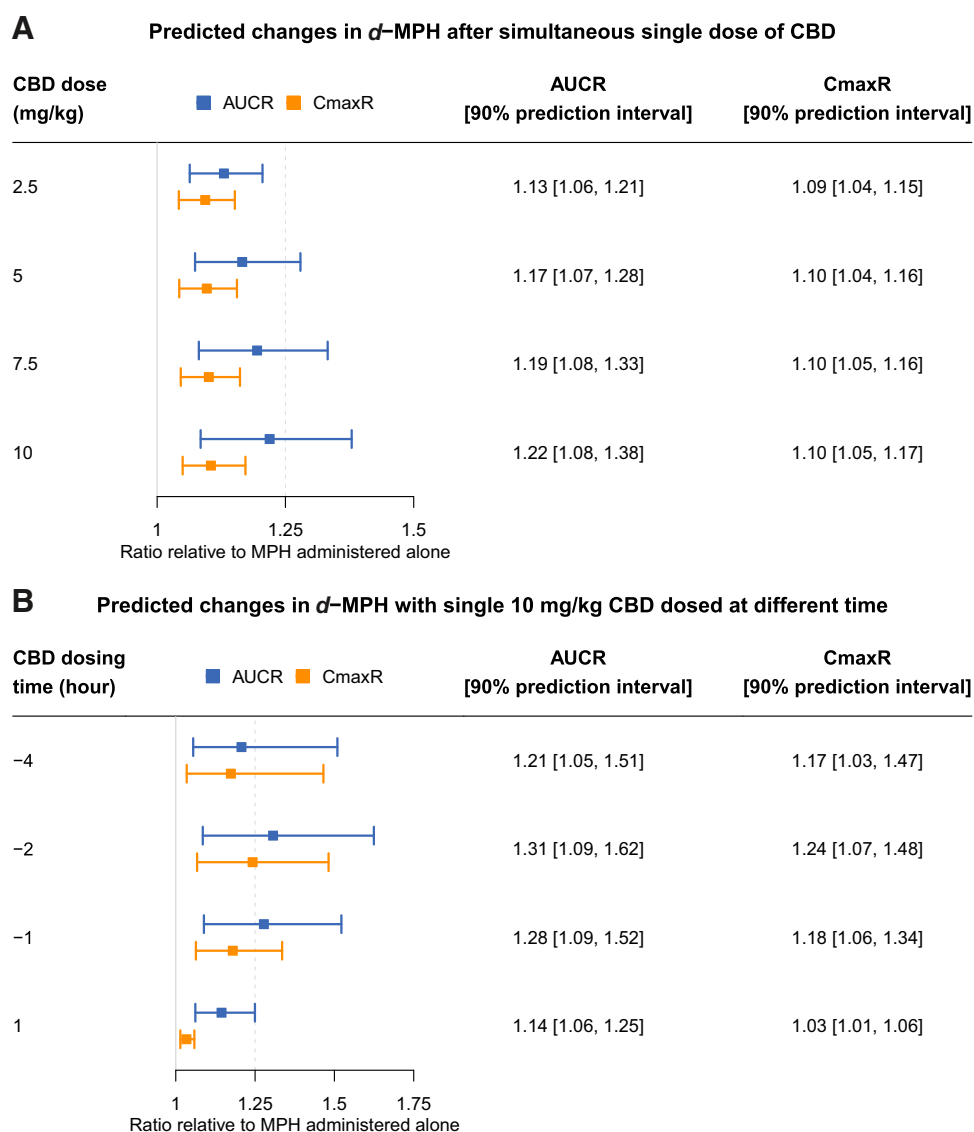


Fig. 6. Simulated outcomes of administration of MPH either simultaneously (A) or with a time difference (B) to a single dose of CBD. The IR formulation of MPH (Ritalin) and the prescriptive oral solution formulation of CBD (Epidiolex) were used in simulation. Under the scenarios of simultaneous administration (A), different strengths of CBD dose (2.5–10 mg/kg) were simulated. Under the scenarios of MPH and CBD administered at different time (B), the dose of CBD was fixed at 10 mg/kg. The resulting influence of CBD on the exposure to *d*-MPH were summarized as ratios of AUC and C_{max} as compared with MPH administered alone. The ratios were reported in the forms of geometric means and 90% prediction interval of the simulated 100 virtual subjects.

and severity of adverse effects (such as insomnia and blood pressure and heart rate increase) that are typically associated with higher doses of MPH (Stein et al., 2003; Nissen, 2006; Food and Drug Administration, 2021).

For THC, smoking a cannabis cigarette (Huestis et al., 1992) was predicted to achieve sufficient systemic concentrations to alter the disposition of MPH. An increase in MPH exposure (AUC) by 34% was predicted using the static model, suggesting a mild magnitude of DDI. For CBD, the highest drug exposure was conferred by use of Epidiolex due to its relatively large oral CBD doses, and an increase of 94% in MPH exposure was predicted by the static model. PBPK models were then developed to provide a more mechanistic insight into the interactions between MPH and CBD. As suggested by the PBPK model, a single dose of oral solution CBD, even at the maximum dose in its label, is not likely to cause any significant impact on concurrent IR-MPH. However, a mild DDI might be expected under the scenario where the CBD dose was given 1–4 hours prior to the MPH dose. One potential explanation for this phenomenon is the

relatively slow dissolution/absorption process of CBD with the prescriptive oral solution formulation. Notably, CBD has a relatively long half-life (Ohlsson et al., 1986; Crockett et al., 2020) and accordingly, drug accumulation is expected after multiple doses. In addition, mechanism-based inhibition of CYP3A4 and CYP2C19 by CBD has been reported (Bansal et al., 2020), which poses a potential of autoinhibition since CBD is also metabolized by these two enzymes (Jiang et al., 2013; Beers et al., 2021). Therefore, a higher CBD exposure may be expected at steady state, which prompted us to assess the scenarios under multiple CBD doses. Consequently, a higher but still mild inhibition of MPH clearance caused by a 7-day treatment of twice daily 10 mg/kg CBD was predicted.

The scenarios of co-administration of MPH and medical/recreational cannabis products are likely. A positive association between childhood ADHD and cannabis use disorder was found (Lee et al., 2011). As more states in the US are expected to legalize recreational use of cannabis, the chance of concomitant consumption of MPH and THC is growing. In addition, utilization of cannabis products for treatment of ADHD symptoms

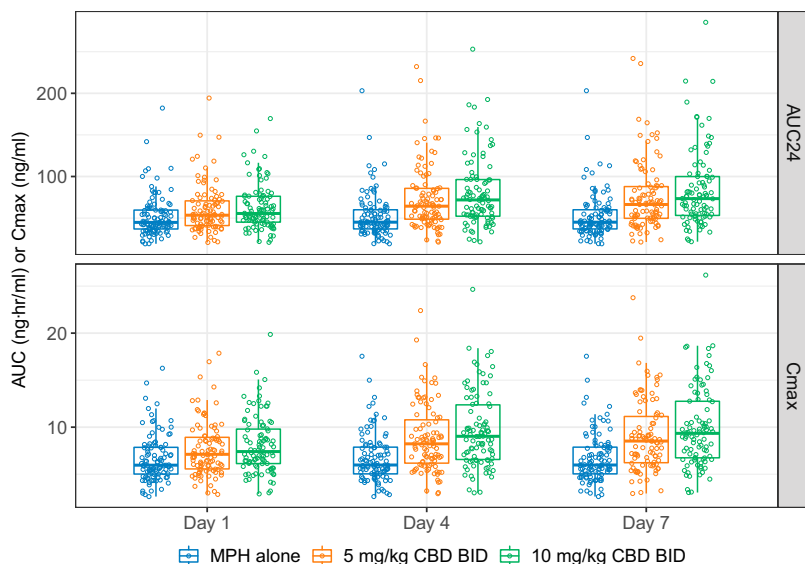


Fig. 7. Simulated changes in MPH exposure when co-administered with multiple doses of CBD for 7 days. The IR formulation of MPH (Ritalin) and the prescriptive oral solution formulation of CBD (Epidiolex) were used in simulation. In the simulation, MPH was administered twice daily with a time interval of 4 hours between the first and second doses, while CBD (0, 5, and 10 mg/kg) was given every 12 hours. The AUC_{24} and C_{max} of *d*-MPH were summarized on Day 1, 4, and 7. Open circles represent individual parameter values of the simulated 100 virtual subjects. Horizontal bars in the boxplots represent the 5th, 50th, and 95th percentiles of the simulated population.

has garnered attention from both the lay public and the research community (Mitchell et al., 2016; Cooper et al., 2017; Mansell et al., 2021). It should be noted that the potency of cannabis has risen dramatically since those earlier studies from which we estimated the physiologically achievable concentrations (i.e., C_{max}) of THC (Chandra et al., 2019). Accordingly, the exposure to THC on an individual via cannabis abuse is likely higher presently than what was reported in earlier studies.

Although CBD content is relatively low in conventional preparations of cannabis (Mehmedic et al., 2010), relatively pure formulations can be accessed by either prescribed Epidiolex or by various over-the-counter CBD products derived from industrial hemp. The over-the-counter CBD products are particularly concerning since their use is not regulated by healthcare professionals and marked differences have been observed between the actual and labeled CBD content of these products (Gurley et al., 2020). These factors have added an additional layer of uncertainty surrounding the risk of DDI between MPH and CBD beyond the clinical scenarios with IR-MPH and Epidiolex evaluated in this work. Moreover, CBD exposure is significantly increased when consumed after a meal (Taylor et al., 2018), which makes consideration of food effects necessary in designing future clinical DDI studies.

Our study has several limitations. First, the PBPK models were developed based on population-level data from the published literature. Consequently, certain features of individual data were overlooked. In particular, covariates that can help explain the observed PK variability were not examined. In the evaluation of PBPK models, we have observed individual PE in the range of around $\pm 50\%$ for several datasets and were not able to further investigate the cause of this inter-study variability due to lack of individual-level data and subject demographics. Second, this work only assessed DDI potentials associated with the IR formulation of MPH. A variety of extended-release and modified-release formulations are now more widely prescribed (Cortese et al., 2017; Steingard et al., 2019). As illustrated in Fig. 6, the relative timing of MPH and CBD dosing may influence the magnitude of their interactions. Therefore, the findings in the current work may not be fully extrapolated to scenarios in which one of the modified-release formulations of MPH is used. Third, the potential for additional contribution to DDI by THC or the metabolites of CBD were not evaluated in this study. We previously identified significant inhibition of CES1 by THC but minimal inhibition by two of its major metabolites (Qian et al., 2019b). Moreover, in the CBD PBPK model, the remaining 20% clearance of MPH was considered exempted from inhibition by CBD due to uncertainty

surrounding the individual enzymes involved. These assumptions potentially lead to an underprediction of DDI magnitude.

In this study, the estimated $K_{i,u}$ values of THC and CBD reflect the unbound concentrations after correction for nonspecific binding using the tube adsorption method (Patilea-Vrana and Unadkat, 2019). Employing a different system (recombinant CES1 as enzyme source and oseltamivir as the substrate), we previously reported 11- to 17-fold higher K_i values for THC (0.541 μM) and CBD (0.974 μM) (Qian et al., 2019b). This difference may be partially explained by the non-specific binding, which was not considered in the previous work. The $f_{u,inc}$ (approximately 1.5% for THC and 4.4% for CBD) we calculated from binding assay were in a similar range to the reported plasma protein binding (Garrett and Hunt, 1974; FDA Center for Drug Evaluation and Research, 2018a; Bansal et al., 2022). Furthermore, other methods (e.g., ultracentrifugation) have been employed in similar studies, and differences do exist among the $f_{u,inc}$ values calculated by different methods (Basit et al., 2020; Bansal et al., 2022). Notably, whether consideration of nonspecific binding can translate into a more accurate prediction of DDI needs further confirmation through clinical study.

In conclusion, we determined the reversible inhibition mechanism of THC and CBD on MPH metabolism and estimated the inhibition potency as measured by $K_{i,u}$ values. The static model suggests mild increase in MPH exposure caused by exposure to either THC from smoking cannabis or by orally administered CBD. PBPK models were developed and used for simulation of various clinical scenarios. The simulation suggests that concomitant administration of MPH and a single oral dose of prescriptive CBD is generally free of DDI concerns even at the highest strength (10 mg/kg) of CBD. However, a mild increase in MPH exposure (AUC by up to 55% and C_{max} by up to 45%) caused by inhibition of CES1 by CBD is possible under scenarios where CBD is administered long term and particularly a few hours prior to the MPH dose. A clinical study is warranted to confirm and evaluate the clinical relevance of the findings in this work.

Authorship Contributions

Participated in research design: Qian, Markowitz.

Conducted experiments: Qian.

Performed data analysis: Qian, Markowitz.

Wrote or contributed to the writing of the manuscript: Qian, Markowitz.

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