FOCUSSED RESEARCH REVIEW

# Novel insights into the molecular mechanisms of HLA class I abnormalities

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Abstract Alterations in the MHC class I surface antigens represent one mechanism of tumor cells to escape from natural or immunotherapy-induced antitumor immune responses. In order to restore MHC class I expression, knowledge about the underlying molecular mechanisms of MHC class I defects in different tumor types is required. In most cases, abnormalities of MHC class I expression are reversible by cytokines suggesting a deregulation rather than structural abnormalities of members of the antigenprocessing and presentation machinery (APM). The impaired expression of APM components could be controlled at the epigenetic, transcriptional and/or posttranscriptional level. Furthermore, a direct link between altered transcription factor binding, interferon signal transduction and MHC class I APM component expression has been shown, which might be further associated with cell cycle progression. This information will not only give novel insights into the (patho) physiology of the antigen-processing and presenting pathway, but will help in the future to design effective T cell-based immunotherapies.

Keywords MHC class I antigens · Antigen processing · Tumors - Gene regulation - Signal transduction

### Abbreviations



Tumors represent heterogeneous diseases with a variable clinical cause, but a high frequency of recurrences. Although a number of ongoing clinical trials provide promise for the implementation of immunotherapies for the treatment of cancer patients and a number of different immunotherapeutic strategies alone or in combination with irradiation or chemotherapy have been explored, the general efficacies of these therapies are not as potent as expected. This might be due to changes in the tumor host interaction, which is mediated by professional antigen-presenting cells (APC), such as B cells and dendritic cells (DC), effector cells like T,

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NK and NKT cells, tumor stroma and endothelium as well as immune suppressive cells including regulatory T cells (Treg) and myeloid-derived suppressor cells (MDSC). The tumor microenvironment may provide protection of tumors against potent T cell responses by increasing for example the frequency of Treg and MDSC [[1,](#page-4-0) [2\]](#page-4-0).

In general, there exists interplay between immune and tumor cells, which profoundly influences each other. An association between host immune responses and prognosis has been described in a variety of tumor types, which is also influenced by the tumor microenvironment, consisting of a heterogeneous mix of cellular and non-cellular components. The immune cells are able to recognize tumor cells, thereby leading to their apoptosis and tumor rejection. This activity is mainly mediated by T cells, NK cells as well as macrophages. During disease progression, host immune responses have been biased against antitumor immune responses. Tumors could not be recognized by immune cells resulting in tumor cell proliferation and in the induction of an immune suppressive microenvironment, which negatively interferes with the systemic and local adaptive immune responses. Lastly, this could lead to the induction of immune escape variants.

#### Different immune escape strategies of tumors

There exist different strategies how tumors evade the immune system. These include defects in the elucidation or maintenance of an effective antitumor responses such as insufficient antigen processing by DC, poor recruitment of or impaired activation of effector cells, the secretion of immune suppressive factors, like cytokines, prostaglandine and growth factors, the expression of molecules of negative regulatory pathways, such as B7-H1, B7-H3 and B7-H4 and the non-classical HLA-G antigen, increased frequency of Treg and MDSC, altered metabolite expression, modulation of the pH as well as downregulation or lack of the expression of classical HLA class I molecules and signal transduction molecules (Fig. 1). Although there exists evidence for improved responses at earlier disease stages, not only activated effector  $CD8<sup>+</sup>$  T cells, but also tumor antigen (TA)specific T cells could be induced. For the antitumoral T cell responses, the interaction of the T cell receptors with their specific HLA class I complex is required. Therefore, the HLA class I antigen-processing machinery (APM) plays a crucial role in mediating immune responses by the generation and expression of the trimeric HLA class I,  $\beta_2$ -microglobuline ( $\beta_2$ -m) and peptide complex. The APM pathway consists of four major steps. The first step involves the peptide generation and peptide trimming, which is mainly mediated by the proteasome and cytosolic peptidases. Endogeneously synthesized proteins are ubiquitinated and



Fig. 1 Different immune escape mechanisms of tumors. Tumors exert different strategies to escape immune cell surveillance. In addition, the frequency and function of immune cells are altered by the tumor microenvironment, which also results in an impaired anti tumor immune response

then degraded by the multi-catalytic proteasome complex consisting of the constitutive and interferon (IFN)- $\gamma$ -inducible subunits, the low molecular weight proteins (LMP)2, -7 and -10. The proteasome yields peptides with a correct C-terminus and an extended N-terminus, which is further trimmed by cytosolic peptidases. Then, the peptides are transported in an ATP- and sequence-dependent manner from the cytosol into the endoplasmic reticulum (ER). In the ER the MHC class I molecules are assembled, which is assisted by various chaperones, such as calnexin, calreticulin and in particular tapasin, which facilitates peptide loading onto MHC class I molecules [[3\]](#page-4-0). This trimeric complex is then transported via the trans-Golgi to the cell surface and there exposed to  $CD8<sup>+</sup>$  cytotoxic T lymphocytes (CTL).

# Alterations in MHC class I antigens and APM components in tumors

It has been shown by many different groups including ours that MHC class I abnormalities could occur at a relatively high frequency in many solid and hematopoietic tumors [\[4–6](#page-4-0)]. A total MHC class I antigen loss, MHC class I downregulation, a selective loss or downregulation of MHC class I allospecificities have been described, which are further associated with disease progression and reduced patients' survival (Table [1\)](#page-2-0) [[7–9\]](#page-4-0). This is often accompanied by an impaired expression of a single or various APM component(s)  $[10-12]$ . For example, an altered expression of LMP2, LMP7, TAP1 and  $\beta_2$ -m was found in renal cell carcinoma (RCC), prostate cancers and melanoma when compared to adjacent normal tissues [\[13–15](#page-4-0)]. The loss of these APM components was associated with a disease progression and/or an early disease recurrence of patients

<span id="page-2-0"></span>



[\[16](#page-4-0)]. In addition, MHC class I APM component expression appears to be tissue- and tumor subtype-specific regulated and differentially expressed during differentiation process and cell cycle progression.

#### Molecular mechanisms of APM downregulation

In order to understand the molecular mechanisms of MHC class I loss or downregulation, structural alterations and control mechanisms involved in the deregulation of diverse APM components were determined (Fig. 2) [[7–9\]](#page-4-0). Structural abnormalities of APM components represent a relatively rare event and appear to be tumor specific, for example mutations, deletion and/or loss of heterozygosity of  $\beta_2$ -m and the HLA class I heavy chains (HC) have been mainly described in colorectal carcinoma and melanoma, but not in RCC [[17–24\]](#page-4-0). Mutations in TAP have only been found in two melanoma cell lines, in non-small lung carcinoma and leiomyosarcomas, whereas mutations in tapasin and LMPs were detected in neuroblastoma



Fig. 2 Molecular mechanisms of MHC class I APM downregulation. The loss or downregulated expression of APM components could occur at the distinct levels of the MHC class I APM pathway and involves transcriptional, posttranscriptional and epigenetic processes resulting in a deregulated expression, whereas structural alterations like mutations, deletions and LOH are rare events

(B. S. unpublished data) [\[13,](#page-4-0) [25](#page-4-0), [26](#page-5-0)]. The defective APM component expression could be restored by gene transfer resulting in increased MHC class I surface antigen levels, enhanced antitumoral responses and survival [[27,](#page-5-0) [28](#page-5-0)]. Thus, the major molecular mechanisms of impaired MHC class I APM component expression appear to be caused by their de-regulation as already demonstrated for virustransformed cells [[29–33\]](#page-5-0). In addition, cytokine-mediated upregulation of MHC class I APM components further strengthened their deregulation [[13,](#page-4-0) [34](#page-5-0), [35\]](#page-5-0). This could occur either at the epigenetic expression, transcriptional and/or posttranscriptional level. Methylation and histone modifications of APM components appear to be a rare event with a frequency comparable to that of mutations and have been only described for TAP1,  $\beta_2$ -m and HLA class I HC [\[36–39](#page-5-0)]. In contrast, transcriptional and/or posttranscriptional control of all major APM components was detected in tumors of distinct histology. Although in most cases APM-specific mRNA and protein were coordinately downregulated, some tumor cells expressed high levels of TAP, LMP, tapasin and ERAP mRNA, but lacked respective protein expression [[13,](#page-4-0) [14](#page-4-0)]. Interestingly, a posttranscriptional downregulation of TAP2 in cells with TAP1 loss due to a mutation in TAP1 leading to an earlier stop codon was reported [\[13](#page-4-0)]. It was speculated that the mutation-mediated lack of TAP1 expression directly destabilises TAP2 protein. In order to prove this hypothesis, cells were left untreated or treated with different proteasome inhibitors like epoxomycin and MG-132. An increased ubiquitination and degradation of TAP2 in the absence of the proteasome inhibitors were found. These data were further confirmed using shRNA plasmids for TAP1 demonstrating a concordant TAP2 downregulation by TAP1 silencing resulting in reduced TAP function and downregulated MHC class I surface expression.

In order to determine the transcriptional activity of MHC class I APM components, the various APM promoters were cloned into a luciferase (luc) expression vector and the APM promoter activity was determined upon transient transfection of the APM-luc constructs into different human tumor cells. In parallel, the mRNA and protein expression of the distinct components was analyzed. Using this approach, a transcriptional as well as posttranscriptional regulation in particular of the LMP subunits, TAP1/2 and tapasin, was observed in melanoma cells. So far, it is not solved, which transcription factors negatively or positively interfere with the APM component expression in tumors. Detailed analysis of the transcription factor binding sites (TFBS) using APM promoter constructs in which the TFBS were altered by site directed mutagenesis will give information about the factors involved in the tumor-mediated suppression of APM components.

## Novel insights into MHC class I APM component regulation

By performing cDNA arrays of  $TAP1<sup>+</sup>$  transfectants and TAP1<sup>-</sup> counterparts, an altered expression pattern of genes involved in the IFN signal transduction pathway as well as in the cellular metabolism was detected (B. S. unpublished data). Therefore, a link between the IFN signaling and/or metabolism with proper APM component expression was postulated. So far, there exists no information whether the altered energy metabolism affects APM components, which might be an interesting topic to investigate. In order to test the association of IFN signaling with APM components, human melanoma cell lines were screened for the loss of the IFN- $\gamma$  inducibility of MHC surface antigens [\[40](#page-5-0)]. Indeed, some melanoma cells lack IFN- $\gamma$  inducibility of MHC molecules. To characterize the mechanisms of IFN resistance, the expression and function of diverse IFN- $\gamma$  signal transduction pathway components was analyzed. Upon IFN- $\gamma$ -binding to its specific receptors, a number of proteins were induced including the janus kinase (JAK)1, JAK2 and the signal transducer and activation (STAT)1. A loss of IFN- $\gamma$ , but not of IFN- $\alpha$  and tumor necrosis factor (TNF)-a inducibility of MHC class I antigens, was found in melanoma cells, suggesting that the lack of IFN- $\gamma$  inducibility of MHC class I molecules was not due to defects in APM component expression. The IFN- $\gamma$  resistance in these melanoma cells was caused by a deletion of the JAK2 gene on chromosome 9, whereas upstream and downstream genes were still present and expressed. The loss of JAK2 not only resulted in the lack of JAK2 mRNA and protein expression, but was also directly associated with a downregulation of constitutive APM component expression as determined for TAP1, TAP2, tapasin as well as  $\beta_2$ -m [\[35](#page-5-0)]. JAK2 overexpression into the JAK2-deficient melanoma cells upregulated MHC class I surface expression as well as basal APM component expression and also restored their IFN- $\gamma$  inducibility. Vice versa treatment of cells with JAK2 inhibitors as well as with JAK2-specific shRNA resulted in a downregulation or loss of HLA class I antigens as well as APM component expression.

# In vitro models of oncogenic transformation as suitable tools for analyzing the mechanisms of deficient APM component expression

In order to characterize the molecular mechanisms of transcriptional regulation of APM components and MHC class I abnormalities, oncogene-transformed cells served as models. Downregulation of MHC class I surface antigens was found upon mos-, myc- and ras- as well as HER-2/neumediated transformation of murine and/or human cells,

which is accompanied by an impaired APM component expression [\[41–45](#page-5-0)]. However, MHC class I surface expression could be restored by IFN- $\gamma$  treatment in these cases. The mechanisms of this oncogene-mediated downregulation could be due to a dysregulation mediated by specific factors and/or repressors modulating the APM component expression. HER-2/neu belongs to the epidermal growth factor (EGF) receptor family (HER-1, -2, -3, -4), has receptor tyrosine kinase activity and represents the preferred dimer partner of the different HERs. So far, no soluble ligands for HER-2/neu have been identified. HER-2/neu is physiologically expressed in epithelial cells, whereas gene amplification and/or overexpression of HER-2/neu occurs in solid tumors, such as breast, lung and colon carcinoma and is often associated with disease progression, poor patients' outcome and impaired cell recognition [\[46](#page-5-0)]. The HER-2/neu-mediated transformation of murine fibroblasts not only caused an enhanced proliferation, development of foci, anchorage-independent cell growth and tumor formation, but also suppressed MHC class I surface antigens, which is mediated by a transcriptional downregulation of various APM components [\[43](#page-5-0)]. Furthermore, an inverse correlation of HER-2/neu and MHC class I APM component expression was found upon analysis of HER-2/ neu<sup>+</sup> versus HER-2/neu<sup>-</sup> mammary carcinoma lesions. These data were confirmed by shRNA-mediated downregulation of HER-2/neu in human tumor cells, resulting in an increased APM component and MHC class I surface expression [[47](#page-5-0)].

So far, transcriptional regulators of the APM have been not well characterized. The role of PML controlling APM expression is controversially discussed [\[48](#page-5-0)], although a concordant regulation of PML and HLA class I antigens was described in prostate cancer [\[49](#page-5-0)]. Analysis of the activity of wild-type and mutant APM promoters in HER-2/  $neu$ <sup>-</sup> and HER-2/neu<sup>+</sup> cells revealed a downregulation of the activity of the various APM promoters in the presence of HER-2/neu. Regarding the tapasin promoter activity, a mutation of the p300 and E2F1 restored the HER-2/neumediated suppression of tapasin promoter activity to levels excluding that of  $HER-2/neu^-$  cells. Thus, both transcription factors appear to be important for the HER-2/neumediated tapasin downregulation [[50\]](#page-5-0). These data were further confirmed by a siRNA approach inhibiting E2F1, which was accompanied by an upregulation of tapasin expression and consequently also MHC class I surface antigen expression [\[50](#page-5-0)]. E2F1 is a member of the large E2F family of highly redundant transcription factors and is often overexpressed in human tumors of distinct histologies. This is further associated with an aberrant cell cycle progression since E2F1 targets genes involved in cell cycle regulation. Our recent data postulated a link between cell cycle control and MHC class I surface expression. However, in order to

<span id="page-4-0"></span>support and confirm that an altered cell cycle progression might play an important role in tumor-associated downregulation of MHC class I APM component expression, combined analyses of cell cycle phases with MHC class I APM molecules should be performed.

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Conflict of interest The authors declare that they have no conflict of interest.

#### References

- 1. Nagaraj S, Gabrilovich DI (2010) Myeloid-derived suppressor cells in human cancer. Cancer J 16(4):348–353. doi[:10.1097/](http://dx.doi.org/10.1097/PPO.0b013e3181eb3358) [PPO.0b013e3181eb3358](http://dx.doi.org/10.1097/PPO.0b013e3181eb3358)
- 2. Lu T, Ramakrishnan R, Altiok S, Youn JI, Cheng P, Celis E, Pisarev V, Sherman S, Sporn MB, Gabrilovich D (2011) Tumorinfiltrating myeloid cells induce tumor cell resistance to cytotoxic T cells in mice. J Clin Invest. doi[:10.1172/JCI45862](http://dx.doi.org/10.1172/JCI45862)
- 3. Scholz C, Tampe R (2009) The peptide-loading complex—antigen translocation and MHC class I loading. Biol Chem 390(8): 783–794. doi[:10.1515/BC.2009.069](http://dx.doi.org/10.1515/BC.2009.069)
- 4. Ferrone S, Marincola FM (1995) Loss of HLA class I antigens by melanoma cells: molecular mechanisms, functional significance and clinical relevance. Immunol Today 16(10):487–494
- 5. Campoli M, Ferrone S (2008) HLA antigen changes in malignant cells: epigenetic mechanisms and biologic significance. Oncogene 27(45):5869–5885. doi:[10.1038/onc.2008.273](http://dx.doi.org/10.1038/onc.2008.273)
- 6. Aptsiauri N, Carretero R, Garcia-Lora A, Real LM, Cabrera T, Garrido F (2008) Regressing and progressing metastatic lesions: resistance to immunotherapy is predetermined by irreversible HLA class I antigen alterations. Cancer Immunol Immunother 57(11):1727–1733. doi:[10.1007/s00262-008-0532-3](http://dx.doi.org/10.1007/s00262-008-0532-3)
- 7. Seliger B (2008) Molecular mechanisms of MHC class I abnormalities and APM components in human tumors. Cancer Immunol Immunother 57(11):1719–1726
- 8. Garrido F, Algarra I, Garcia-Lora AM (2010) The escape of cancer from T lymphocytes: immunoselection of MHC class I loss variants harboring structural-irreversible ''hard'' lesions. Cancer Immunol Immunother 59(10):1601–1606. doi[:10.1007/](http://dx.doi.org/10.1007/s00262-010-0893-2) [s00262-010-0893-2](http://dx.doi.org/10.1007/s00262-010-0893-2)
- 9. Garrido C, Algarra I, Maleno I, Stefanski J, Collado A, Garrido F, Garcia-Lora AM (2010) Alterations of HLA class I expression in human melanoma xenografts in immunodeficient mice occur frequently and are associated with higher tumorigenicity. Cancer Immunol Immunother 59(1):13–26. doi[:10.1007/s00262-009-](http://dx.doi.org/10.1007/s00262-009-0716-5) [0716-5](http://dx.doi.org/10.1007/s00262-009-0716-5)
- 10. Seliger B, Schreiber K, Delp K, Meissner M, Hammers S, Reichert T, Pawlischko K, Tampe R, Huber C (2001) Downregulation of the constitutive tapasin expression in human tumor cells of distinct origin and its transcriptional upregulation by cytokines. Tissue Antigens 57(1):39–45
- 11. Restifo NP, Esquivel F, Kawakami Y, Yewdell JW, Mule JJ, Rosenberg SA, Bennink JR (1993) Identification of human cancers deficient in antigen processing. J Exp Med 177(2):265–272
- 12. Cabrera CM, Lopez-Nevot MA, Jimenez P, Garrido F (2005) Involvement of the chaperone tapasin in HLA-B44 allelic losses in colorectal tumors. Int J Cancer 113(4):611–618. doi: [10.1002/ijc.20526](http://dx.doi.org/10.1002/ijc.20526)
- 13. Seliger B, Ritz U, Abele R, Bock M, Tampe R, Sutter G, Drexler I, Huber C, Ferrone S (2001) Immune escape of melanoma: first evidence of structural alterations in two distinct components of the MHC class I antigen processing pathway. Cancer Res 61(24):8647–8650
- 14. Kamphausen E, Kellert C, Abbas T, Akkad N, Tenzer S, Pawelec G, Schild H, van Endert P, Seliger B (2010) Distinct molecular mechanisms leading to deficient expression of ER-resident aminopeptidases in melanoma. Cancer Immunol Immunother 59(8):1273–1284. doi:[10.1007/s00262-010-0856-7](http://dx.doi.org/10.1007/s00262-010-0856-7)
- 15. Atkins D, Breuckmann A, Schmahl GE, Binner P, Ferrone S, Krummenauer F, Storkel S, Seliger B (2004) MHC class I antigen processing pathway defects, ras mutations and disease stage in colorectal carcinoma. Int J Cancer 109(2):265–273. doi[:10.1002/](http://dx.doi.org/10.1002/ijc.11681) [ijc.11681](http://dx.doi.org/10.1002/ijc.11681)
- 16. Meissner M, Reichert TE, Kunkel M, Gooding W, Whiteside TL, Ferrone S, Seliger B (2005) Defects in the human leukocyte antigen class I antigen processing machinery in head and neck squamous cell carcinoma: association with clinical outcome. Clin Cancer Res 11(7):2552–2560. doi[:10.1158/1078-0432.CCR-04-](http://dx.doi.org/10.1158/1078-0432.CCR-04-2146) [2146](http://dx.doi.org/10.1158/1078-0432.CCR-04-2146)
- 17. Yang T, McNally BA, Ferrone S, Liu Y, Zheng P (2003) A single-nucleotide deletion leads to rapid degradation of TAP-1 mRNA in a melanoma cell line. J Biol Chem 278(17):15291– 15296. doi[:10.1074/jbc.M300954200](http://dx.doi.org/10.1074/jbc.M300954200)
- 18. Yang T, Lapinski PE, Zhao H, Zhou Q, Zhang H, Raghavan M, Liu Y, Zheng P (2005) A rare transporter associated with antigen processing polymorphism overpresented in HLAlow colon cancer reveals the functional significance of the signature domain in antigen processing. Clin Cancer Res 11(10):3614–3623. doi: [10.1158/1078-0432.CCR-04-1804](http://dx.doi.org/10.1158/1078-0432.CCR-04-1804)
- 19. Vermeulen CF, Jordanova ES, ter Haar NT, Kolkman-Uljee SM, de Miranda NF, Ferrone S, Peters AA, Fleuren GJ (2007) Expression and genetic analysis of transporter associated with antigen processing in cervical carcinoma. Gynecol Oncol 105(3):593–599. doi:[10.1016/j.ygyno.2007.02.016](http://dx.doi.org/10.1016/j.ygyno.2007.02.016)
- 20. Maleno I, Lopez-Nevot MA, Cabrera T, Salinero J, Garrido F (2002) Multiple mechanisms generate HLA class I altered phenotypes in laryngeal carcinomas: high frequency of HLA haplotype loss associated with loss of heterozygosity in chromosome region 6p21. Cancer Immunol Immunother 51(7): 389–396. doi:[10.1007/s00262-002-0296-0](http://dx.doi.org/10.1007/s00262-002-0296-0)
- 21. Hayashi T, Kobayashi Y, Kohsaka S, Sano K (2006) The mutation in the ATP-binding region of JAK1, identified in human uterine leiomyosarcomas, results in defective interferongamma inducibility of TAP1 and LMP2. Oncogene 25(29):4016– 4026
- 22. Chen HL, Gabrilovich D, Tampe R, Girgis KR, Nadaf S, Carbone DP (1996) A functionally defective allele of TAP1 results in loss of MHC class I antigen presentation in a human lung cancer. Nat Genet 13(2):210–213. doi[:10.1038/ng0696-210](http://dx.doi.org/10.1038/ng0696-210)
- 23. Chang CC, Ogino T, Mullins DW, Oliver JL, Yamshchikov GV, Bandoh N, Slingluff CL Jr, Ferrone S (2006) Defective human leukocyte antigen class I-associated antigen presentation caused by a novel beta2-microglobulin loss-of-function in melanoma cells. J Biol Chem 281(27):18763–18773. doi:[10.1074/jbc.M511](http://dx.doi.org/10.1074/jbc.M511525200) [525200](http://dx.doi.org/10.1074/jbc.M511525200)
- 24. Cabrera CM, Jimenez P, Cabrera T, Esparza C, Ruiz-Cabello F, Garrido F (2003) Total loss of MHC class I in colorectal tumors can be explained by two molecular pathways: beta2-microglobulin inactivation in MSI-positive tumors and LMP7/TAP2 downregulation in MSI-negative tumors. Tissue Antigens 61(3): 211–219
- 25. Seliger B, Bock M, Ritz U, Huber C (2002) High frequency of a non-functional TAP1/LMP2 promoter polymorphism in human tumors. Int J Oncol 20(2):349–353
- <span id="page-5-0"></span>26. Hodson I, Bock M, Ritz U, Brenner W, Huber C, Seliger B (2003) Analysis of the structural integrity of the TAP2 gene in renal cell carcinoma. Int J Oncol 23(4):991–999
- 27. Qin Z, Schwartzkopff J, Pradera F, Kammertoens T, Seliger B, Pircher H, Blankenstein T (2003) A critical requirement of interferon gamma-mediated angiostasis for tumor rejection by CD8? T cells. Cancer Res 63(14):4095–4100
- 28. Kallfelz M, Jung D, Hilmes C, Knuth A, Jaeger E, Huber C, Seliger B (1999) Induction of immunogenicity of a human renalcell carcinoma cell line by TAP1-gene transfer. Int J Cancer 81(1):125-133. doi[:10.1002/\(SICI\)1097-0215\(19990331\)81:1](http://dx.doi.org/10.1002/(SICI)1097-0215(19990331)81:1%3c125::AID-IJC21%3e3.0.CO;2-2)<125:: AID-IJC21>[3.0.CO;2-2](http://dx.doi.org/10.1002/(SICI)1097-0215(19990331)81:1%3c125::AID-IJC21%3e3.0.CO;2-2)
- 29. Verweij MC, Koppers-Lalic D, Loch S, Klauschies F, de la Salle H, Quinten E, Lehner PJ, Mulder A, Knittler MR, Tampe R, Koch J, Ressing ME, Wiertz EJ (2008) The varicellovirus UL49.5 protein blocks the transporter associated with antigen processing (TAP) by inhibiting essential conformational transitions in the  $6 + 6$  transmembrane TAP core complex. J Immunol 181(7): 4894–4907
- 30. Oosten LE, Koppers-Lalic D, Blokland E, Mulder A, Ressing ME, Mutis T, van Halteren AG, Wiertz EJ, Goulmy E (2007) TAP-inhibiting proteins US6, ICP47 and UL49.5 differentially affect minor and major histocompatibility antigen-specific recognition by cytotoxic T lymphocytes. Int Immunol 19(9):1115– 1122. doi:[10.1093/intimm/dxm082](http://dx.doi.org/10.1093/intimm/dxm082)
- 31. Koppers-Lalic D, Verweij MC, Lipinska AD, Wang Y, Quinten E, Reits EA, Koch J, Loch S, Marcondes Rezende M, Daus F, Bienkowska-Szewczyk K, Osterrieder N, Mettenleiter TC, Heemskerk MH, Tampe R, Neefjes JJ, Chowdhury SI, Ressing ME, Rijsewijk FA, Wiertz EJ (2008) Varicellovirus UL 49.5 proteins differentially affect the function of the transporter associated with antigen processing, TAP. PLoS Pathog 4(5):e1000080
- 32. Griffin BD, Verweij MC, Wiertz EJ (2010) Herpesviruses and immunity: the art of evasion. Vet Microbiol 143(1):89–100. doi: [10.1016/j.vetmic.2010.02.017](http://dx.doi.org/10.1016/j.vetmic.2010.02.017)
- 33. Croft NP, Shannon-Lowe C, Bell AI, Horst D, Kremmer E, Ressing ME, Wiertz EJ, Middeldorp JM, Rowe M, Rickinson AB, Hislop AD (2009) Stage-specific inhibition of MHC class I presentation by the Epstein-Barr virus BNLF2a protein during virus lytic cycle. PLoS Pathog 5(6):e1000490. doi[:10.1371/](http://dx.doi.org/10.1371/journal.ppat.1000490) [journal.ppat.1000490](http://dx.doi.org/10.1371/journal.ppat.1000490)
- 34. Rodriguez T, Mendez R, Del Campo A, Jimenez P, Aptsiauri N, Garrido F, Ruiz-Cabello F (2007) Distinct mechanisms of loss of IFN-gamma mediated HLA class I inducibility in two melanoma cell lines. BMC Cancer 7:34. doi:[10.1186/1471-2407-7-34](http://dx.doi.org/10.1186/1471-2407-7-34)
- 35. Respa A, Bukur J, Ferrone S, Pawelec G, Zhao Y, Wang E, Marincola FM, Seliger B (2011) Association of IFN-gamma signal transduction defects with impaired HLA class I antigen processing in melanoma cell lines. Clin Cancer Res 17(9):2668– 2678
- 36. Setiadi AF, Omilusik K, David MD, Seipp RP, Hartikainen J, Gopaul R, Choi KB, Jefferies WA (2008) Epigenetic enhancement of antigen processing and presentation promotes immune recognition of tumors. Cancer Res 68(23):9601–9607
- 37. Setiadi AF, David MD, Seipp RP, Hartikainen JA, Gopaul R, Jefferies WA (2007) Epigenetic control of the immune escape mechanisms in malignant carcinomas. Mol Cell Biol 27(22): 7886–7894
- 38. Sers C, Kuner R, Falk CS, Lund P, Sueltmann H, Braun M, Buness A, Ruschhaupt M, Conrad J, Mang-Fatehi S, Stelniec I,

Krapfenbauer U, Poustka A, Schafer R (2009) Down-regulation of HLA Class I and NKG2D ligands through a concerted action of MAPK and DNA methyltransferases in colorectal cancer cells. Int J Cancer 125(7):1626–1639. doi[:10.1002/ijc.24557](http://dx.doi.org/10.1002/ijc.24557)

- 39. Khan AN, Gregorie CJ, Tomasi TB (2008) Histone deacetylase inhibitors induce TAP, LMP, Tapasin genes and MHC class I antigen presentation by melanoma cells. Cancer Immunol Immunother 57(5):647–654. doi:[10.1007/s00262-007-0402-4](http://dx.doi.org/10.1007/s00262-007-0402-4)
- 40. Epperson DE, Arnold D, Spies T, Cresswell P, Pober JS, Johnson DR (1992) Cytokines increase transporter in antigen processing-1 expression more rapidly than HLA class I expression in endothelial cells. J Immunol 149(10):3297–3301
- 41. Vertegaal AC, Kuiperij HB, Houweling A, Verlaan M, van der Eb AJ, Zantema A (2003) Differential expression of tapasin and immunoproteasome subunits in adenovirus type 5- versus type 12-transformed cells. J Biol Chem 278(1):139–146. doi[:10.1074/](http://dx.doi.org/10.1074/jbc.M206267200) [jbc.M206267200](http://dx.doi.org/10.1074/jbc.M206267200)
- 42. Seliger B, Harders C, Lohmann S, Momburg F, Urlinger S, Tampe R, Huber C (1998) Down-regulation of the MHC class I antigen-processing machinery after oncogenic transformation of murine fibroblasts. Eur J Immunol 28(1):122–133. doi[:10.1002/](http://dx.doi.org/10.1002/(SICI)1521-4141(199801)28:01<122::AID-IMMU122>3.0.CO;2-F) (SICI)1521-4141(199801)28:01<122::AID-IMMU122&#62; [3.0.CO;2-F](http://dx.doi.org/10.1002/(SICI)1521-4141(199801)28:01<122::AID-IMMU122>3.0.CO;2-F)
- 43. Herrmann F, Lehr HA, Drexler I, Sutter G, Hengstler J, Wollscheid U, Seliger B (2004) HER-2/neu-mediated regulation of components of the MHC class I antigen-processing pathway. Cancer Res 64(1):215–220
- 44. Griffioen M, Peltenburg LT, van Oorschot DA, Schrier PI (1995) C-myc represses transiently transfected HLA class I promoter sequences not locus-specifically. Immunobiology 193(2–4):238– 247
- 45. Ashrafi GH, Tsirimonaki E, Marchetti B, O'Brien PM, Sibbet GJ, Andrew L, Campo MS (2002) Down-regulation of MHC class I by bovine papillomavirus E5 oncoproteins. Oncogene 21(2): 248–259. doi:[10.1038/sj.onc.1205008](http://dx.doi.org/10.1038/sj.onc.1205008)
- 46. Mimura K, Ando T, Poschke I, Mougiakakos D, Johansson CC, Ichikawa J, Okita R, Nishimura MI, Handke D, Krug N, Choudhury A, Seliger B, Kiessling R (2011) T cell recognition of HLA-A2 restricted tumor antigens is impaired by the oncogene HER2. Int J Cancer 128(2):390–401
- 47. Choudhury A, Charo J, Parapuram SK, Hunt RC, Hunt DM, Seliger B, Kiessling R (2004) Small interfering RNA (siRNA) inhibits the expression of the Her2/neu gene, upregulates HLA class I and induces apoptosis of Her2/neu positive tumor cell lines. Int J Cancer 108(1):71–77. doi[:10.1002/ijc.11497](http://dx.doi.org/10.1002/ijc.11497)
- 48. Zheng P, Guo Y, Niu Q, Levy DE, Dyck JA, Lu S, Sheiman LA, Liu Y (1998) Proto-oncogene PML controls genes devoted to MHC class I antigen presentation. Nature 396(6709):373–376. doi:[10.1038/24628](http://dx.doi.org/10.1038/24628)
- 49. Zhang H, Melamed J, Wei P, Cox K, Frankel W, Bahnson RR, Robinson N, Pyka R, Liu Y, Zheng P (2003) Concordant downregulation of proto-oncogene PML and major histocompatibility antigen HLA class I expression in high-grade prostate cancer. Cancer Immun 3:2
- 50. Bukur J, Herrmann F, Handke D, Recktenwald C, Seliger B (2010) Identification of E2F1 as an important transcription factor for the regulation of tapasin expression. J Biol Chem 285(40): 30419–30426