

NK cell-mediated targeting of human cancer and possibilities for new means of immunotherapy

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Abstract Insights into the molecular basis for natural killer (NK) cell recognition of human cancer have been obtained in recent years. Here, we review current knowledge on the molecular specificity and function of human NK cells. Evidence for NK cell targeting of human tumors is provided and new strategies for NK cell-based immunotherapy against human cancer are discussed. Based on current knowledge, we foresee a development where more cancers may be subject to treatment with drugs or other immunomodulatory agents affecting NK cells, either directly or indirectly. We also envisage a possibility that certain forms of cancers may be subject to treatment with adoptively transferred NK cells, either alone or in combination with other therapeutic interventions.

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Abbreviations

AICD	Activation induced cell death
ADCC	Antibody mediated cellular cytotoxicity
AML	Acute myeloid leukemia
ATG	Anti-thymocyte globulin
ATM	Ataxia telangiectasia, mutated
ATR	ATM- and Rad3-related
CR	Complete remission
DC	Dendritic cell
DLI	Donor lymphocyte infusion
EBV	Epstein-Barr virus
Fas L	Fas ligand
Flt-3 L	Flt-3 ligand
GMP	Good manufacturing practice
GVH	Graft-versus-host
GVT	Graft-versus-tumor
GVHD	Graft-versus-host-disease
HLA	Human leukocyte antigen
IFN	Interferon
KIR	Killer-Ig-like receptors
LAK	Lymphokine activated killer
MHC	Major histocompatibility complex
NCR	Natural cytotoxicity receptor
NK	Natural killer
SCF	Stem cell factor
SCT	Stem cell transplantation
TNF	Tumor necrosis factor
Treg	Regulatory T
TRAIL	TNF-related apoptosis-inducing ligand
UL	Unique long

Introduction

Natural killer (NK) cells represent a unique subset of lymphocytes, distinct from T and B cells. In an integrative manner with other immune cells, they contribute to host anti-microbial and anti-tumor immunity. NK cells were originally identified on a functional basis, because of their ability to lyse certain tumor cells *in vitro* without the requirement for prior immune sensitization of the host. This initial observation gave them their name [58]. Today, these cells are well characterized with respect to their origin, differentiation, receptor repertoire, and effector functions [53, 65, 77, 83]. Here, we discuss current insights into NK cell-recognition of human cancer, and discuss several possible future therapeutic modalities involving NK cells, either directly or indirectly.

Phenotype, numbers and tissue location of human NK cells

Natural killer cells account for 5–15% of human peripheral blood lymphocytes. In addition to peripheral blood, they are found in the bone marrow, spleen and lymph nodes as well as in specific organs such as the liver and lungs. They are activated by cytokines and/or by interactions with specific molecules expressed on target cells [13, 65, 83]. This leads to induction of effector functions, including cytokine production and cellular cytotoxicity [77]. Upon activation, NK cells produce cytokines and chemokines and can exert strong cytolytic effector functions [78, 94]. Human NK cells are broadly defined as CD3⁻CD56⁺ lymphocytes. They can be further sub-divided into two main functional subsets based on their surface expression of CD56. CD56^{bright} NK cells have potent immunoregulatory properties, and CD56^{dim} NK cells have potent cytotoxic functions [26, 37]. These functions are; however, not strictly confined to each respective subset; a certain degree of functional overlap exists. The CD56^{dim} NK cells express high levels of Fc γ RIIIA (CD16) allowing them to mediate ADCC. Although insights into NK cell development has been obtained, it is still not fully clear at which stage of differentiation CD56^{bright} and CD56^{dim} NK cells separate from each other [30, 36].

NK cells in host responses against tumors

Since their discovery, a large number of studies have demonstrated NK cell killing of many different types of murine and human tumor cell lines *in vitro* [117, 131]. Several experimental studies in mice have demonstrated a role for NK cells in the eradication of grafted murine tumor cell lines [2, 68]. Furthermore, NK cells have been shown to be involved in rejection responses against experimentally induced and spontaneously developing tumors in mice

[119]. Human NK cells adoptively transferred to mice also participate in the rejection of grafted human tumors [47]. Direct evidence for NK cell targeting of human cancer has come from studies of NK cell interactions with primary tumor cells tested for susceptibility to NK cells lysis *ex vivo*. Such approaches have recently been taken for, e.g., neuroblastomas, ovarian carcinomas, and multiple myelomas [18, 21, 33]. Evidence for NK cell targeting of human tumors has also come from clinical studies in settings of SCT and adoptive transfer of NK cells to cancer patients [70, 76, 99, 101]. It should be noted, however, that despite the large amount of studies that demonstrate the ability of NK cells to target tumor cells *in vitro* and *in vivo* in experimental models [117, 131], there is still only limited evidence for clinical efficacy of activated NK cells administered to patients with cancer.

All tumors are, however, not susceptible to NK cell-mediated lysis. A variety of reasons may underlie this resistance, including specific properties of some tumor cells themselves as well as effects imposed by certain tumors on NK cells. Data from both experimental models and from studies of human cancer have demonstrated tumor cell evasion from NK cells [50, 73, 74]. This includes the intriguing observations of, e.g., high MHC class I expression of some metastasizing human tumors and/or loss of ligands for NK cell activation receptors on other tumors [28, 55, 103]. This may argue for selection of tumor cell mutants or tumor cell modulation during an ongoing NK cell-mediated response. Down-regulation of NK cell-activating receptors can be mediated by, e.g., TGF- β , which selectively down-regulates the surface expression of some NK cell-activating receptors [20]. Likewise, NK cell interactions with tumor target cells may specifically down-regulate the expression of activating receptors, e.g., down-regulation of DNAM-1 upon recognition of PVR-expressing ovarian carcinomas ([33]; our own unpublished results). Finally, tumor cells may also restrain NK cell effector function by promoting the expansion of CD4⁺CD25⁺ Treg cells. Recent evidence points to a critical role for Treg cells in dampening NK cell immune responses by suppression of homeostatic proliferation, cytotoxicity, and IL-12-mediated IFN- γ production [43, 118].

Molecular interactions involved in NK cell targeting of tumors

The identification of NK cell germ-line-encoded activation and inhibitory receptors has in large uncovered the molecular mechanisms used by NK cells to recognize tumor cells (Table 1). This knowledge emerges from early observations that NK cell cytotoxicity is triggered by tumor cells lacking expression of certain (or all) self-MHC class I molecules [44, 52, 57, 68], a phenomenon

Table 1 Activation and inhibitory receptors expressed on freshly isolated human peripheral blood NK cells (Adopted from ref [13]. For some receptors, ligands are not known or not well characterized)

Receptor ^a	Type of receptor	Signaling motif	Ligand
CD16 (FcγRIIIA)	Activation	ITAM ^b	IgG
NKp30 (CD337)	Activation	ITAM	
NKp46 (CD335)	Activation	ITAM	Viral hemagglutinin
KIR2DS1-2	Activation	ITAM	HLA-C (low affinity)
KIR2DS3-6	Activation	ITAM	
KIR3DS1	Activation	ITAM	
NKG2C (CD94/159c)	Activation	ITAM	HLA-E
NKG2D (CD314)	Activation	Non-ITAM	ULBPs, MICA, MICB
2B4 (CD244)	Activation	Non-ITAM	CD48
CD2	Activation	Non-ITAM	LFA-3 (CD58)
CRACC (CD319)	Activation	Non-ITAM	CRACC (CD319)
NTB-A	Activation	Non-ITAM	NTB-A
DNAM-1 (CD226)	Activation	Non-ITAM	PVR (CD155), CD112
CD7	Activation	Non-ITAM	SECTM1, Galectin
CD59	Activation	Non-ITAM	C8, C9
BY55 (CD160)	Activation	Non-ITAM	HLA-C
KIR2DL4 (CD158d)	Activation	Non-ITAM	HLA-G (soluble)
CD44	Activation	Non-ITAM	Hyaluronan
LFA-1 (α L β 2, CD11 α /18)	Activation	Integrin	ICAM-1-5
MAC-1 (α M β 2, CD11 β /18)	Activation	Integrin	ICAM-1, iC3b, Fibrinogen
CD11c/18	Activation	Integrin	ICAM-1, iC3b
VLA-4 (α 4 β 1, CD49d/29)	Activation	Integrin	VCAM-1, Fibronectin
VLA-5 (α 5 β 1, CD49e/29)	Activation	Integrin	Fibronectin
KIR2DL1 (CD158a)	Inhibitory	ITIM ^c	HLA-C group 2
KIR2DL2/3	Inhibitory	ITIM	HLA-C group 1
KIR2DL5	Inhibitory	ITIM	
KIR3DL1	Inhibitory	ITIM	HLA-B alleles
KIR3DL2	Inhibitory	ITIM	HLA-A alleles
LIR-1/ILT2 (CD85j)	Inhibitory	ITIM	Multiple HLA class I alleles
NKG2A (CD94/CD159a)	Inhibitory	ITIM	HLA-E
KLRG1	Inhibitory	ITIM	E/N/P-cadherin
NKR-P1 (CD161)	Inhibitory	ITIM	LLT1
Siglec-7 (CD328)	Inhibitory	ITIM	sialic acid
Siglec-9 (CD329)	Inhibitory	ITIM	sialic acid
Irp60 (CD300a)	Inhibitory	ITIM	

^a KIR, NKG2C, CD2, NKG2A, LIR-1, NKR-G1, NKR-P1, Siglec-7, and Siglec-9 are only expressed by subsets of NK cells

^b Immunoreceptor tyrosine-based activation motif

^c Immunoreceptor tyrosine-based inhibition motif

referred to as “missing-self” recognition [69]. This observation led to the identification of NK cell inhibitory receptors that recognize MHC class I molecules [56, 79]. However, sensing the absence of self-MHC class I molecules is not sufficient to cause target cell killing by NK cells. NK cells also need stimulation by target cell-ligands to trigger activation via specific receptors. The identification of the latter remained elusive until rather recently [65, 83]. Thus, we now know that NK cell recognition of tumors is tightly regulated by processes involving the integration of signals delivered from multiple activating and inhibitory receptors [13, 65, 81, 83, 132].

NK cell activation receptors in tumor recognition

One important group of human NK cell activation receptors is the so-called NCRs: NKp46, NKp30, and NKp44 [90, 130]. Two of these, NKp46 and NKp30, are constitutively expressed on all peripheral blood NK cells, whereas NKp44 is induced and expressed by IL-2-activated NK cells and some T cells. Despite considerable effort to characterize their ligands, their constituents on tumor cells remain poorly defined [8, 92]. However, the nuclear factor HLA-B-associated transcript 3 (BAT3) was shown to be released from tumor cells and to bind NKp30. BAT-3 trig-

gered NKp30-mediated NK cell cytotoxicity and thus represents the first identified cellular ligand for any of the natural cytotoxicity receptors [8, 92]. The NCRs have a major role in NK cell-mediated lysis of various human tumor cell lines including melanomas, carcinomas, neuroblastomas, and myeloid or lymphoblastic leukemias, as well as EBV-transformed B cells [8]. Other well-characterized activation receptors are NKG2D and DNAM-1 [4, 9]. NKG2D recognizes the stress inducible molecules MICA and MICB as well as the UL16-binding proteins. NKG2D ligands are expressed on a number of human epithelial tumor and leukemic cell lines and have a significant role in rendering these cells susceptible to NK cell-mediated lysis. DNAM-1 recognizes PVR (CD155) and Nectin-2 (CD122). These ligands are highly expressed in human carcinomas, melanomas, and neuroblastomas [24, 27, 80]. CD16 on NK cells binds the Fc-portion of IgG on opsonized cells, thus mediating ADCC. In addition, several other receptors, including 2B4 (CD244), NTBA, NKp80, CD2, CD11a/CD18, and CD59, have important co-activating or co-stimulatory functions in NK cell activation and tumor cell recognition [13].

NK cell inhibitory receptors in tumor recognition

Activation of NK cells is under control by inhibitory receptors [65, 81, 83]. Inhibitory receptors bind classical and/or non-classical MHC class I molecules. These molecules are normally expressed on most healthy cells in the body, but are often lost upon transformation or during tumor evolution [75]. In humans, KIR and CD94-NKG2A play major roles as HLA-class I-specific inhibitory NK cell receptors. KIR recognizes groups of HLA-A, -B, and -C alleles [82, 86, 124], whereas CD94-NKG2A/B receptors recognize HLA-E molecules [12]. Individuals differ in the number and type of inherited KIR genes, and specific KIR gene products are expressed on distinct subsets of NK cells. Thus, many NK cells express only a few of many possible types of receptors. Most NK cells, however, express at least one inhibitory receptor that is specific for a self-MHC class I ligand. The pattern of KIRs expressed creates a system allowing NK cells to detect cells lacking expression of single MHC class I alleles [86]. As discussed below, this diversity is potentially beneficial in settings of SCT and adoptive NK cell-based immunotherapy against cancer.

Relative importance of different NK cell activation receptors in NK cell tumor targeting

The relative importance of different NK cell activation receptors and their ligands in recognizing primary human tumors is only partially known. Interestingly, efficient natural cytotoxicity by resting (e.g., not preactivated by cyto-

kines) NK cells usually requires co-activation by several types of receptors [13, 14]. In contrast, engagement of LFA-1 is sufficient to induce cytotoxicity by IL-2 activated NK cells [14]. Translated to the context of killing tumor cells, these findings indicate that a tumor cell might elicit activation of an unstimulated NK cell by expressing array of ligands for several activating receptors, any one of which alone would be incapable of triggering a response. For an NK cell activated by, e.g., IL-2, fewer qualitatively distinct receptor–ligand interactions might suffice for tumor cell recognition and killing provided MHC class I molecules do not confer inhibition. In some situations, the activation signals may override the inhibitory signals mediated by MHC class I molecules, as has been demonstrated, e.g., for NKG2D-mediated triggering of some MHC class I expressing tumor cell lines [23, 31]. The final fate of the NK cells is also critically dependent—on not only—the type of ligands but also their relative density for the integration of signals from activating and inhibitory signals.

NK cell-effector functions involved in anti-tumor responses

Like cytotoxic T cells, NK cells possess different effector mechanisms by which they mount anti-tumor responses [127]. Many of them have been explored extensively in experimental models. NK cells use two major mechanisms to induce target cell apoptosis, the granule exocytosis and the death receptor pathway [114]. Granulae exocytosis involves the release of perforin and granzymes [121]. The death receptor pathway is largely mediated by members of the TNF superfamily, in which Fas L, TNF- α , and TRAIL are key apoptosis-inducing members [105, 114]. NK cells can also produce many different types of cytokines (e.g., IFN- γ , TNF- α , and GM-CSF) as well as chemokines, at least some of which have a direct effect on tumors. The best-studied cytokine in this respect is IFN- γ . IFN- γ can decrease tumor proliferation and metabolic activity and inhibit angiogenesis [116]. IFN- γ produced by NK cells might also play a role in the regulation of killing by death receptors, either by down-regulating anti-apoptotic proteins, or by up-regulating caspases that are essential for death receptor-mediated apoptosis. IFN- γ induces type-1 immunity and may by this means counteract tumor escape mechanisms actively promoted by cancer cells and Treg cells through the secretion of type-2 cytokines [118].

Activation of NK cells to enhance anti-tumor responses

Given the fact that NK cells in some situations can target human cancers, it may be of interest to activate endogenous NK cells to induce more potent anti-tumor responses. It may also advantageous to activate NK cells *ex vivo* prior

to adoptive immunotherapy. Several cytokines affect NK cell differentiation and activation such as IL-2, IL-12, IL-15, IL-18, IL-21, and type-1 IFNs [5, 25]. Upon cytokine stimulation, NK cells proliferate, produce cytokines, and upregulate effector molecules such as adhesion molecules, NKp44, perforin, granzymes, FasL, and TRAIL. IL-2 stimulates NK cell progenitors and mature NK cells, and induces the production of NK cell effector molecules, enhancing NK cytolytic activity. In addition, IL-2 augments NK cell degranulation via a syntaxin 11-independent pathway [15], and may reduce the dependency of co-activation for some receptors implicated in tumor cell lysis [14]. Recent evidence suggests a unique role for IL-15 in the differentiation, proliferation, survival, and activation of NK cells [84, 126]. IL-15 may also protect lymphocytes from IL-2 induced AICD [96, 126]. This cytokine synergizes with Flt3-L and SCF in inducing human CD56^{bright} NK cells [30, 36]. IL-12 and IL-18 act late during the NK cell differentiation, and synergistically enhance cytotoxicity against tumor targets and IFN- γ production by NK cells [66]. IL-21 is of particular interest in its ability to stimulate cytotoxic CD56^{dim} NK cells and to enhance NK cell cytotoxicity [10, 87].

Cytokines and other immunomodulatory drugs in cancer treatment

Cytokines have successfully been applied in the treatment of several human cancers and, in some instances, the mechanisms of action are through direct or indirect activation of NK cells [35, 115]. Several clinical trials have assessed the effects of IL-2 administration on activation of NK cells in patients with cancer [97, 115]. Unfortunately, high doses of IL-2 are associated with significant toxicity [38]. Irrespectively, IL-2 is frequently used in lower doses to promote NK cell activity in vivo. This cytokine has more recently also been used together with monoclonal antibodies that mediate ADCC to enhance NK cell activity [95]. In many respects, IL-15 may be a better cytokine. IL-15 is more efficient than IL-2 in expanding the NK cell compartment [46, 85, 125]. It promotes survival of NK cells and protects NK cells from AICD. However, high doses of IL-15 may be needed in vivo to obtain effective anti-tumor effects [60, 125]. New insight into the role of early hematopoietic growth factors, such as c-kit ligand and flt-3 ligand, and their synergy with IL-15 in the development of human NK cells in the bone marrow, will likely permit studies of additional cytokine combinations for expansion of NK cells for clinical use. Nevertheless, better knowledge about how best to use cytokines for optimal activation of NK cells, either alone or in combination with other immune interventions [115], is clearly needed. It still remains to be studied whether the impressive results

obtained in mice, using individual and combinations of cytokines to activate NK cells to kill tumor cells, can be translated to humans. Apart from specific cytokines and/or growth factors, broad activators of immune function implicated in anti-tumor immunity, may also stimulate NK cells. For instance, in myeloma, NK cell activity has been shown to increase in response to thalidomide and its analogue lenalidomide, explaining, in part, the mechanism of action of this drug [51]. Immunostimulatory DNA complexes have also been shown to enhance in vivo anti-tumor activity, mediated, at least in part, through the activation of NK cells [39]. Imatinib mesylate (Gleevec), a specific inhibitor of tyrosine kinase receptors may also lead to host-anti tumor effects mediated by the innate immune system and a new type of immune cell, referred to as IFN-producing killer dendritic cells (IKDC) that resemble natural killer cells [7, 113, 123]. Finally, NK cells may also contribute to the clinical efficacy of bacillus Calmette-Guerin (BCG) treatment of bladder cancer [11].

NK cell-mediated anti-tumor responses in settings of allogeneic SCT

Allogeneic hematopoietic SCT is an established treatment strategy for several hematological malignancies. An intentional mismatch between donor KIRs and recipient HLA ligands in hematopoietic SCT is predicted to allow for a graft anti-tumor effect by NK cells that develop in the recipient. In pioneering studies by Velardi and collaborators, adult patients with AML undergoing haploidentical SCT showed greatly improved disease-free survival time and low relapse rates when a KIR-ligand mismatch prevailed [100, 101]. This effect occurred in the absence of donor T cells, which had been removed prior to the transplant. Notably, hematopoietic grafts with NK cell alloreactivity in the GvH direction also had increased rates of bone marrow engraftment and reduced rates of clinically significant GvHD. The latter effect may be caused by donor NK cell lysis of host antigen-presenting cells, impairing alloreactive donor T cell priming [101, 109]. Since the initial report by Velardi and collaborators [101], numerous studies have addressed the role of KIR-ligand mis-match in different settings of transplantation [34, 102]. Some, but not all, studies have demonstrated a beneficial role of NK cell alloreactivity. Discrepancies in outcome among the studies may depend on the transplantation protocol including differences such as the type of preconditioning, dose of stem cells, degree of T cell depletion, and post-transplantation immunosuppression. Further studies are warranted to better understand the conditions that steer NK cell maturation and receptor-acquisition following SCT, particularly relative to alloreactivity and GvT effects.

NK cell-mediated anti-tumor responses in settings of DLI following SCT

Unfortunately, many patients relapse after hematopoietic SCT. DLIs can induce a direct and potent GvT effect in some of these patients [63, 64, 112]. The major risk of DLI is GvHD, which may be a severe, even lethal complication. To minimize the risk of GvHD, modified strategies have been developed such as partially T cell-depleted DLI. In haploidentical SCT, studies have been initiated where purified donor NK cells have been used as DLIs with the aim to consolidate engraftment and induce GvL effects [88, 89]. In a related study, patients with solid tumors undergoing allogeneic SCT were infused with donor-derived *ex vivo* expanded NK cells (Dilber and collaborators, submitted). Although, no firm conclusions can be made regarding the clinical efficacy of NK cell-based DLI at this stage, available data indicate that NK cell infusions are safe and may generate anti-tumor responses. Since normal tissues do not generally express ligands for activating NK cell receptors, alloreactive NK cells should not normally cause GvHD [8, 45]. The development of NK cell-based DLI represents new possibilities for treating relapses in patients undergoing haploidentical or cord blood SCT.

The use of NK cells in pre-SCT conditioning

Experimental data have demonstrated that NK cells can be administered directly after conditioning but before SCT. The potential benefits are threefold. First, NK cell-mediated GvL effects could enhance anti-tumor activity and reduce risk for relapse. Second, NK cell-mediated depletion of recipient DC could prevent the development of acute GvHD and perhaps allow higher numbers of alloreactive T cells in the graft, thus avoiding death from infections in the early post-transplant period [72, 99, 101]. Finally, NK cells may facilitate engraftment and promote a fast immune reconstitution, thereby reducing the need for myeloablative regimens and shortening the neutropenic period.

Adoptive immunotherapy using short term *ex vivo* activated NK cells

Because of the lack of significant clinical effects with past protocols based on autologous “LAK” cells or NK cells [16, 67, 98], and because of the promising effects observed in haploidentical T cell-depleted SCT, focus shifted toward the potential of using allogeneic (potentially alloreactive) NK cells in adoptive cell therapy. In recent studies, Miller and collaborators infused haploidentical NK cells together with IL-2 to 43 non-transplanted patients with advanced cancer. Low and high intensity preparative regimens were tested. The high-dose lymphodepleting regimen resulted in

long-term survival and expansion of donor-derived NK cells *in vivo*. The *in vivo* expansion was associated with increased levels of endogenous IL-15, possibly driving the proliferation of donor NK cells. Notably, one of these patients manifested a preferential expansion of the alloreactive NK cell subset. In general, these donor NK cell infusions were feasible and tolerated without unexpected toxicity. Moreover, 5/19 patients with AML achieved CR with this protocol. Only four of the 19 patients had donors with a predictable alloreactive NK cell repertoire. Interestingly, of these four patients, three achieved CR [76].

Adoptive immunotherapy using NK cell lines and *ex vivo* expanded NK cells

In parallel with the foregoing studies, adoptive transfers are also being done with the NK-92 cell line [120]. This cell line can be grown continuously under GMP-conditions, express many NK cell activation receptors and low levels of KIRs, and displays significant cytotoxicity toward many tumor targets. Following irradiation, more than 20 patients with advanced renal cell carcinoma and malignant melanomas have received NK-92 cells [59]. In general, these infusions are well tolerated and have yielded anti-tumor effects in some cases. Furthermore, this cell line is easily modified genetically which opens up interesting possibilities for future therapeutic trials. Whether adoptive immunotherapy with this cell line will produce more substantial clinical responses remains to be seen.

Several techniques have been developed for *ex vivo* expansion of NK cells [59]. A few of these protocols allow the expansion of NK cell-enriched cellular populations under GMP-conditions [17]. Using such protocols, it has been demonstrated that NK cells can be expanded *ex vivo*, also from tumor-bearing patients [3, 48]. The latter opens up for expansion of autologous NK cells for adoptive immunotherapy, as strategy that may be developed further despite the earlier disappointments with autologous LAK cell therapy in the 1980s to patients with advanced solid tumors [67, 98]. Expansion protocols provide greater numbers of NK cells to be used for adoptive therapy that might be desired in some situations. For such expansions to be effective, it is important that the expansion of NK cells *ex vivo* is not associated with phenotypic changes, lineage deviation, and/or selective expansion of specific subsets, such that their anti-tumor function will be affected. Another aspect to consider, apart from consequences of activation and proliferation, is that *in vitro* manipulation does not alter the NK cells' ability to mediate cell–cell interactions, trafficking, and homing to desired location. With respect to autologous NK cells, one may predict that they may be more effective in situations where tumors express low levels of MHC class I molecules.

Future possibilities and strategies for adoptive NK cell therapy

We have recently described critical questions that must be considered for the development of successful NK cell-based adoptive immunotherapy [70]. Below, we briefly discuss some issues with respect to the possible advantages, but also difficulties, of using allogeneic NK cells in future settings of adoptive NK cell-based immunotherapy.

Since autologous NK cells are inhibited by self-MHC class I molecules, allogeneic NK cells may, in certain situations, represent a better cellular population for adoptive immunotherapy *in vivo*. The latter choice applies particularly to situations in which tumor targets express normal levels of MHC class I molecules in combination with low or only moderate expression of ligands for activating receptors. NK cell alloreactivity depends on “missing” KIR-ligands (MHC class I) in the recipient. However, although NK cell alloreactivity is predicted by genetic algorithms based on KIR- and HLA-genotyping, the numbers of alloreactive NK cells in a given donor vary significantly, from below 1 to 60 % of the NK cells (our unpublished observations). Predicting the effectiveness of therapy may thus be based on assessment of the NK cell repertoire and selection of a donor with the largest alloreactive NK cell subset. This is feasible in KIR haplotype-A donors who lack activating KIR; in such donors, genetic analysis can be combined with phenotypic analysis using multi-parameter flow cytometry.

A prerequisite for survival of the infused cells is that they are not rejected by the recipient’s immune system. If donor-derived NK cells are infused at the time of transplantation they may engraft along with the stem cells because of the pre-transplant conditioning. However, rejection of allogeneic NK cells represents a major challenge for specific NK cell therapy in the absence of myeloablative conditioning. It is likely that some type of conditioning will be required for effective transfer of allogeneic NK cells. Apart from preventing rejection, such regimens may also eradicate regulatory T cells that could otherwise interfere with the proliferation and function of the donor derived NK cells [42]. Moreover, there is reduced competition for growth factors during the homeostatic proliferation that follows lymphodepletion and the surge of cytokines, including IL-15, may promote proliferation, *in vivo* survival, and expansion of the infused NK cells. Indeed, in the studies by Miller and collaborators [76], NK cell expansion was dependent on an intense preparative regimen (high-dose cyclophosphamide/fludarabine). The latter regimen is similar to that used recently by Rosenberg and colleagues to induce homeostatic proliferation of adoptively transferred T cells [32]. As understanding of the conditions required for engraftment of NK cells improves, dosing of the prepar-

ative regimen will be more precise and the risks associated with high-dose myeloablative treatments will decrease.

Strategies to activate and/or modulate NK cells *in vivo* in patients with cancer

An exciting possibility that has now reached the stage of clinical trials in humans is to block inhibitory KIRs with monoclonal antibodies, thereby augmenting tumor cell recognition by NK cells [108]. Such reagents could ideally be used, e.g., in the treatment of hematopoietic cancers that are not amenable to SCT. Preclinical evidence in mouse models has demonstrated that this strategy may enhance antitumor activity in autologous [61] and allogeneic settings [62]. Along the same lines, the modulation of activating ligands on tumor cells may also improve the efficacy of NK cell recognition. NK cells may also be genetically engineered prior to adoptive transfer to the patients. One interesting possibility is to stably over-express chimeric receptors recognizing ligands expressed by tumors combined with signaling components that trigger NK cell function [106]. For example, NK cells engineered to express chimeric anti-CD19-CD3 signaling receptors became highly cytotoxic against autologous leukemia cells [54]. Genetically modified NK-92 cells expressing a chimeric antigen receptor specific for the tumor-associated ErbB2 (HER2/neu) antigen specifically lysed ErbB2-expressing tumor cells that were completely resistant to cytolysis by parental NK-92 cells [122].

Which human cancers may be targeted by NK cells?

With respect to tumor cell types, it is already evident from studies performed *in vitro* and even in some clinical studies that certain tumor types may be better suited than others for NK cell based immunotherapy. The presence on human tumors of ligands for activating receptors provides an important prerequisite for NK cell activation, and thus for the potential of achieving good clinical results [80]. An illustration of this is the inefficient NK cell killing of lymphoid compared to myeloid leukemias that may be caused, at least in part, by the absence of LFA-1 ligand expression [100]. Likewise, low expression of MHC class I molecules, particularly in situations where KIR-ligand mismatching (“missing-self”-reactivity) does not prevail, is also important. Most immunotherapy trials have been performed in patients with significant tumor burdens, where conventional therapies were ineffective. The best clinical setting for most cellular therapies including NK cell-based immunotherapy is probably when the tumor burden is small, i.e., in minimal residual disease [111]. NK cell therapy against large solid tumors represents special problems including not only the size of the tumor *per se* but also the presumed necessity of

NK cells to infiltrate the tumor. Despite the knowledge gained so far about the mechanisms that control trafficking of NK cells, we still know too little about the requirements for NK cell homing to and infiltration of tumors. It is known, however, that chemokines are required to attract NK cells to tumor sites. NK cells express a wide array of chemokine receptors on their cell surface. Different NK cell subsets can be identified based upon chemokine receptor expression and the pattern of expression is likely highly dependent upon the activation status of the NK cells [94].

Combination therapies involving NK cells may develop into promising treatment options for some human cancers

In all likelihood, combination therapies with NK cells representing one important mediator will become ever more important in the future. Ligands for the activating receptor NKG2D are up-regulated by genotoxic stress and stalled DNA replication, through activation of major DNA damage checkpoint pathways initiated by ATM or ATR protein kinases [40, 41]. Thus, the response to DNA damage alerts the immune system to the presence of potentially dangerous cells. Since several of the currently used chemotherapeutic drugs as well as ionizing irradiation act via the DNA damage response pathway, a mild preconditioning using these drugs and/or local ionizing irradiation might sensitize tumor cells to immune recognition, leading to synergistic anti-tumor effects. Similarly, new generation cancer drugs such as the proteasome inhibitors and the histone deacetylase inhibitors can upregulate the death receptor DR5, sensitizing tumor cells to TRAIL-mediated killing by NK cells [71, 104, 128]. Histone deacetylase inhibitors induce MICA/B expression [110]. Imatinib mesylate, previously discussed as a potential stimulator of innate immunity to tumors, was also shown to influence the expression and shedding of the activating NK cell ligand MICA on Bcr/Abl positive targets [22]. Thus, although NKG2D expression on NK cells is restored upon Imatinib treatment, this may lead to decreased tumor targeting because of reduced MICA expression [6, 22].

As has been discussed, NK cells are major effectors in mediating ADCC. Rituximab (Mabthera), a chimeric mouse/human antibody that recognize the CD20 antigen expressed on mature B lymphocytes [93] is currently given alone or combined with chemotherapy to patients with non-Hodgkin's lymphomas. One mechanism of this antibody's action is the induction of ADCC mediated by NK cells [29, 129]. Trials combining Rituximab with IL-2 to activate and expand the pool of NK cells available for ADCC are under way [91]. Several other antibodies are being evaluated in clinical practices and for many of them such as, e.g., Herceptin, at least part of their effector mechanism seem to be mediated by NK cells via ADCC [1, 19]. These and other

findings suggest the possibility of using antibodies in conjunction with adoptive NK cell immunotherapy or NK cell stimulation-based protocols. A related therapeutic approach is the use of bispecific antibodies to promote NK cell targeting of tumors. Experimental and clinical data suggest that bispecific antibodies can be beneficial in tumor treatment. One approach is the use of antibodies specific for CD16 on NK cells and CD19 on B cell lymphomas or HER2/neu on breast cancers to target tumors expressing these, respective antigens [107]. Interestingly, clinical responses have been observed in patients with Hodgkin's lymphoma treated with bispecific antibodies against CD16 and CD30 [49].

Concluding remarks

We foresee a development where certain forms of cancer may become targets for therapeutic interventions involving NK cells. In addition to adoptive-based NK cell immunotherapy, we have highlighted different ways to enhance NK cell activity and the possibility of exploring combinatorial treatment strategies where NK cells mediate direct or indirect anti-tumor effects in future cancer treatments. In the latter respect, induction of NK cell activation ligands on tumor targets is one interesting aspect. Clearly, many questions and obstacles need to be met, and practical issues often play a central role in all strategies of human cell therapy. The effort required to translate advances at the bench to the clinic shall not be underestimated. Standardization of protocols will be needed to allow for future comparisons between different trials. Nonetheless, we believe that NK cell may likely find a role in future therapies against some human cancers, either alone or in combination with other therapies.

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References

1. Adams GP, Weiner LM (2005) Monoclonal antibody therapy of cancer. *Nat Biotechnol* 23:1147–1157
2. Algarra I, Ohlen C, Perez M, Ljunggren HG, Klein G, Garrido F, Karre K (1989) NK sensitivity and lung clearance of MHC-class-I-deficient cells within a heterogeneous fibrosarcoma. *Int J Cancer* 44:675–680
3. Alici E, Sutlu T, Bjorkstrand B, Gilljam M, Stellan B, Nahi H, Quezada HC, Gahrton G, Ljunggren HG, Dilber MS (2008) Autologous anti-tumor activity by NK cells expanded from myeloma patients using GMP-compliant components. *Blood* (in press)

4. Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, Spies T (1999) Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. *Science* 285:727–729
5. Becknell B, Caligiuri MA (2005) Interleukin-2, interleukin-15, and their roles in human natural killer cells. *Adv Immunol* 86:209–239
6. Boissel N, Rea D, Tieng V, Dulphy N, Brun M, Cayuela JM, Rousselot P, Tamouza R, Le Bouteiller P, Mahon FX, Steinle A, Charron D et al (2006) BCR/ABL oncogene directly controls MHC class I chain-related molecule A expression in chronic myelogenous leukemia. *J Immunol* 176:5108–5116
7. Borg C, Terme M, Taieb J, Menard C, Flament C, Robert C, Maruyama K, Wakasugi H, Angevin E, Thielemans K, Le Cesne A, Chung-Scott V et al (2004) Novel mode of action of c-kit tyrosine kinase inhibitors leading to NK cell-dependent antitumor effects. *J Clin Invest* 114:379–388
8. Bottino C, Castriconi R, Moretta L, Moretta A (2005) Cellular ligands of activating NK receptors. *Trends Immunol* 26:221–226
9. Bottino C, Castriconi R, Pende D, Rivera P, Nanni M, Carnemolla B, Cantoni C, Grassi J, Marcenaro S, Reymond N, Vitale M, Moretta L et al (2003) Identification of PV9 (CD155) and Nectin-2 (CD112) as cell surface ligands for the human DNAM-1 (CD226) activating molecule. *J Exp Med* 198:557–567
10. Brady J, Hayakawa Y, Smyth MJ, Nutt SL (2004) IL-21 induces the functional maturation of murine NK cells. *J Immunol* 172:2048–2058
11. Brandau S, Riemensberger J, Jacobsen M, Kemp D, Zhao W, Zhao X, Jocham D, Ratliff TL, Bohle A (2001) NK cells are essential for effective BCG immunotherapy. *Int J Cancer* 92:697–702
12. Braud VM, Allan DS, O’Callaghan CA, Soderstrom K, D’Andrea A, Ogg GS, Lazetic S, Young NT, Bell JI, Phillips JH, Lanier LL, McMichael AJ (1998) HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. *Nature* 391:795–799
13. Bryceson YT, March ME, Ljunggren HG, Long EO (2006) Activation, coactivation, and costimulation of resting human natural killer cells. *Immunol Rev* 214:73–91
14. Bryceson YT, March ME, Ljunggren HG, Long EO (2006) Synergy among receptors on resting NK cells for the activation of natural cytotoxicity and cytokine secretion. *Blood* 107:159–166
15. Bryceson YT, Rudd E, Zheng C, Edner J, Ma D, Wood SM, Bechensteen AG, Boelens JJ, Celkan T, Farah RA, Hultenby K, Winiarski J et al (2007) Defective cytotoxic lymphocyte degranulation in syntaxin-11 deficient familial hemophagocytic lymphohistiocytosis 4 (FHL4) patients. *Blood* 110:1906–1915
16. Burns LJ, Weisdorf DJ, DeFor TE, Vesole DH, Repka TL, Blazar BR, Burger SR, Panoskaltis-Mortari A, Keever-Taylor CA, Zhang MJ, Miller JS (2003) IL-2-based immunotherapy after autologous transplantation for lymphoma and breast cancer induces immune activation and cytokine release: a phase I/II trial. *Bone Marrow Transplant* 32:177–186
17. Carlens S, Gilljam M, Chambers BJ, Aschan J, Guven H, Ljunggren HG, Christensson B, Dilber MS (2001) A new method for in vitro expansion of cytotoxic human CD3-CD56+ natural killer cells. *Hum Immunol* 62:1092–1098
18. Carlsten M, Bjorkstrom NK, Norell H, Bryceson Y, van Hall T, Baumann BC, Hanson M, Schedvins K, Kiessling R, Ljunggren HG, Malmberg KJ (2007) DNAX accessory molecule-1 mediated recognition of freshly isolated ovarian carcinoma by resting natural killer cells. *Cancer Res* 67:1317–1325
19. Carter PJ (2006) Potent antibody therapeutics by design. *Nat Rev Immunol* 6:343–357
20. Castriconi R, Cantoni C, Della Chiesa M, Vitale M, Marcenaro E, Conte R, Biassoni R, Bottino C, Moretta L, Moretta A (2003) Transforming growth factor beta 1 inhibits expression of NKp30 and NKG2D receptors: consequences for the NK-mediated killing of dendritic cells. *Proc Natl Acad Sci USA* 100:4120–4125
21. Castriconi R, Dondero A, Corrias MV, Lanino E, Pende D, Moretta L, Bottino C, Moretta A (2004) Natural killer cell-mediated killing of freshly isolated neuroblastoma cells: critical role of DNAX accessory molecule-1-poliovirus receptor interaction. *Cancer Res* 64:9180–9184
22. Cebo C, Da Rocha S, Wittnebel S, Turhan AG, Abdelali J, Cailat-Zucman S, Bourhis JH, Chouaib S, Caignard A (2006) The decreased susceptibility of Bcr/Abl targets to NK cell-mediated lysis in response to imatinib mesylate involves modulation of NKG2D ligands, GM1 expression, and synapse formation. *J Immunol* 176:864–872
23. Cerwenka A, Baron JL, Lanier LL (2001) Ectopic expression of retinoic acid early inducible-1 gene (RAE-1) permits natural killer cell-mediated rejection of a MHC class I-bearing tumor in vivo. *Proc Natl Acad Sci USA* 98:11521–11526
24. Chang CC, Ferrone S (2006) NK cell activating ligands on human malignant cells: molecular and functional defects and potential clinical relevance. *Semin Cancer Biol* 16:383–392
25. Colucci F, Caligiuri MA, Di Santo JP (2003) What does it take to make a natural killer? *Nat Rev Immunol* 3:413–425
26. Cooper MA, Fehniger TA, Caligiuri MA (2001) The biology of human natural killer-cell subsets. *Trends Immunol* 22:633–640
27. Costello RT, Fauriat C, Sivori S, Marcenaro E, Olive D (2004) NK cells: innate immunity against hematological malignancies? *Trends Immunol* 25:328–333
28. Costello RT, Sivori S, Marcenaro E, Lafage-Pochitaloff M, Mozziconacci MJ, Reviron D, Gastaut JA, Pende D, Olive D, Moretta A (2002) Defective expression and function of natural killer cell-triggering receptors in patients with acute myeloid leukemia. *Blood* 99:3661–3667
29. Dall’Ozzo S, Tartas S, Paintaud G, Cartron G, Colombat P, Bardos P, Watier H, Thibault G (2004) Rituximab-dependent cytotoxicity by natural killer cells: influence of FCGR3A polymorphism on the concentration–effect relationship. *Cancer Res* 64:4664–4669
30. Di Santo JP (2006) Natural killer cell developmental pathways: a question of balance. *Annu Rev Immunol* 24:257–286
31. Diefenbach A, Jensen ER, Jamieson AM, Raulet DH (2001) Rae1 and H60 ligands of the NKG2D receptor stimulate tumour immunity. *Nature* 413:165–171
32. Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, Topalian SL, Sherry R, Restifo NP, Hubicki AM, Robinson MR, Raffeld M et al (2002) Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Science* 298:850–854
33. El-Sherbiny YM, Meade JL, Holmes TD, McGonagle D, Mackie SL, Morgan AW, Cook G, Feyler S, Richards SJ, Davies FE, Morgan GJ, Cook GP (2007) The requirement for DNAM-1, NKG2D, and Nkp46 in the natural killer cell-mediated killing of myeloma cells. *Cancer Res* 67:8444–8449
34. Farag SS, Bacigalupo A, Eapen M, Hurley C, Dupont B, Caligiuri MA, Boudreau C, Nelson G, Oudshoorn M, van Rood J, Velardi A, Maiers M et al (2006) The effect of KIR ligand incompatibility on the outcome of unrelated donor transplantation: a report from the center for international blood and marrow transplant research, the European blood and marrow transplant registry, and the Dutch registry. *Biol Blood Marrow Transplant* 12:876–884
35. Farag SS, Caligiuri MA (2004) Cytokine modulation of the innate immune system in the treatment of leukemia and lymphoma. *Adv Pharmacol* 51:295–318
36. Farag SS, Caligiuri MA (2006) Human natural killer cell development and biology. *Blood Rev* 20:123–137

37. Farag SS, Fehniger TA, Ruggeri L, Velardi A, Caligiuri MA (2002) Natural killer cell receptors: new biology and insights into the graft-versus-leukemia effect. *Blood* 100:1935–1947
38. Fehniger TA, Cooper MA, Caligiuri MA (2002) Interleukin-2 and interleukin-15: immunotherapy for cancer. *Cytokine Growth Factor Rev* 13:169–183
39. Fujii H, Trudeau JD, Teachey DT, Fish JD, Grupp SA, Schultz KR, Reid GS (2007) In vivo control of acute lymphoblastic leukemia by immunostimulatory CpG oligonucleotides. *Blood* 109:2008–2013
40. Gasser S, Orsulic S, Brown EJ, Raulet DH (2005) The DNA damage pathway regulates innate immune system ligands of the NKG2D receptor. *Nature* 436:1186–1190
41. Gasser S, Raulet D (2006) The DNA damage response, immunity and cancer. *Semin Cancer Biol* 16:344–347
42. Gattinoni L, Powell DJ Jr, Rosenberg SA, Restifo NP (2006) Adoptive immunotherapy for cancer: building on success. *Nat Rev Immunol* 6:383–393
43. Ghiringhelli F, Menard C, Terme M, Flament C, Taieb J, Chaput N, Puig PE, Novault S, Escudier B, Vivier E, Lécésne A, Robert C et al (2005) CD4+CD25+ regulatory T cells inhibit natural killer cell functions in a transforming growth factor-beta-dependent manner. *J Exp Med* 202:1075–1085
44. Glas R, Sturmhofel K, Hammerling GJ, Karre K, Ljunggren HG (1992) Restoration of a tumorigenic phenotype by beta 2-microglobulin transfection to EL-4 mutant cells. *J Exp Med* 175:843–846
45. Gonzalez S, Groh V, Spies T (2006) Immunobiology of human NKG2D and its ligands. *Curr Top Microbiol Immunol* 298:121–138
46. Gosselin J, Tomolu A, Gallo RC, Flamand L (1999) Interleukin-15 as an activator of natural killer cell-mediated antiviral response. *Blood* 94:4210–4219
47. Guimaraes F, Guven H, Donati D, Christensson B, Ljunggren HG, Bejarano MT, Dilber MS (2006) Evaluation of ex vivo expanded human NK cells on antileukemia activity in SCID-beige mice. *Leukemia* 20:833–839
48. Guven H, Gilljam M, Chambers BJ, Ljunggren HG, Christensson B, Kimby E, Dilber MS (2003) Expansion of natural killer (NK) and natural killer-like T (NKT)-cell populations derived from patients with B-chronic lymphocytic leukemia (B-CLL): a potential source for cellular immunotherapy. *Leukemia* 17:1973–1980
49. Hartmann F, Renner C, Jung W, da Costa L, Tembrink S, Held G, Sek A, Konig J, Bauer S, Kloft M, Pfreundschuh M (2001) Anti-CD16/CD30 bispecific antibody treatment for Hodgkin's disease: role of infusion schedule and costimulation with cytokines. *Clin Cancer Res* 7:1873–1881
50. Hayakawa Y, Smyth MJ (2006) Innate immune recognition and suppression of tumors. *Adv Cancer Res* 95:293–322
51. Hayashi T, Hideshima T, Akiyama M, Podar K, Yasui H, Raje N, Kumar S, Chauhan D, Treon SP, Richardson P, Anderson KC (2005) Molecular mechanisms whereby immunomodulatory drugs activate natural killer cells: clinical application. *Br J Haematol* 128:192–203
52. Hoglund P, Ljunggren HG, Ohlen C, Ahrlund-Richter L, Scangos G, Bieberich C, Jay G, Klein G, Karre K (1988) Natural resistance against lymphoma grafts conveyed by H-2Dd transgene to C57BL mice. *J Exp Med* 168:1469–1474
53. Huntington ND, Voshchenrich CA, Di Santo JP (2007) Developmental pathways that generate natural-killer-cell diversity in mice and humans. *Nat Rev Immunol* 7:703–714
54. Imai C, Iwamoto S, Campana D (2005) Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. *Blood* 106:376–383
55. Jager MJ, Hurks HM, Levitskaya J, Kiessling R (2002) HLA expression in uveal melanoma: there is no rule without some exception. *Hum Immunol* 63:444–451
56. Karlhofer FM, Ribaud RK, Yokoyama WM (1992) MHC class I alloantigen specificity of Ly-49+ IL-2-activated natural killer cells. *Nature* 358:66–70
57. Karre K, Ljunggren HG, Piontek G, Kiessling R (1986) Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy. *Nature* 319:675–678
58. Kiessling R, Klein E, Wigzell H (1975) Natural killer cells in the mouse. I. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Specificity and distribution according to genotype. *Eur J Immunol* 5:112–117
59. Klingemann HG (2005) Natural killer cell-based immunotherapeutic strategies. *Cytotherapy* 7:16–22
60. Kobayashi H, Dubois S, Sato N, Sabzevari H, Sakai Y, Waldmann TA, Tagaya Y (2005) Role of trans-cellular IL-15 presentation in the activation of NK cell-mediated killing, which leads to enhanced tumor immunosurveillance. *Blood* 105:721–727
61. Koh CY, Blazar BR, George T, Welniak LA, Capitini CM, Raziuddin A, Murphy WJ, Bennett M (2001) Augmentation of anti-tumor effects by NK cell inhibitory receptor blockade in vitro and in vivo. *Blood* 97:3132–3137
62. Koh CY, Ortaldo JR, Blazar BR, Bennett M, Murphy WJ (2003) NK-cell purging of leukemia: superior antitumor effects of NK cells H2 allogeneic to the tumor and augmentation with inhibitory receptor blockade. *Blood* 102:4067–4075
63. Kolb HJ, Mittermuller J, Clemm C, Holler E, Ledderose G, Brehm G, Heim M, Wilmanns W (1990) Donor leukocyte transfusions for treatment of recurrent chronic myelogenous leukemia in marrow transplant patients. *Blood* 76:2462–2465
64. Kolb HJ, Simoes B, Schmid C (2004) Cellular immunotherapy after allogeneic stem cell transplantation in hematologic malignancies. *Curr Opin Oncol* 16:167–173
65. Lanier LL (2005) NK cell recognition. *Annu Rev Immunol* 23:225–274
66. Lauwerys BR, Garot N, Renauld JC, Houssiau FA (2000) Cytokine production and killer activity of NK/T-NK cells derived with IL-2, IL-15, or the combination of IL-12 and IL-18. *J Immunol* 165:1847–1853
67. Law TM, Motzer RJ, Mazumdar M, Sell KW, Walther PJ, O'Connell M, Khan A, Vlavis V, Vogelzang NJ, Bajorin DF (1995) Phase III randomized trial of interleukin-2 with or without lymphokine-activated killer cells in the treatment of patients with advanced renal cell carcinoma. *Cancer* 76:824–832
68. Ljunggren HG, Karre K (1985) Host resistance directed selectively against H-2-deficient lymphoma variants. Analysis of the mechanism. *J Exp Med* 162:1745–1759
69. Ljunggren HG, Karre K (1990) In search of the 'missing self': MHC molecules and NK cell recognition. *Immunol Today* 11:237–244
70. Ljunggren HG, Malmberg KJ (2007) Prospects for the use of NK cells in immunotherapy of human cancer. *Nat Rev Immunol* 7:329–339
71. Lundqvist A, Abrams SI, Schrupp DS, Alvarez G, Suffredini D, Berg M, Childs R (2006) Bortezomib and decapeptide sensitize tumors to tumor necrosis factor-related apoptosis-inducing ligand: a novel method to potentiate natural killer cell tumor cytotoxicity. *Cancer Res* 66:7317–7325
72. Lundqvist A, McCoy JP, Samsel L, Childs R (2007) Reduction of GVHD and enhanced anti-tumor effects after adoptive infusion of alloreactive Ly49-mismatched NK-cells from MHC-matched donors. *Blood* 109:3603–3606
73. Malmberg KJ (2004) Effective immunotherapy against cancer: a question of overcoming immune suppression and immune escape? *Cancer Immunol Immunother* 53:879–892

74. Malmberg KJ, Ljunggren HG (2006) Escape from immune- and nonimmune-mediated tumor surveillance. *Semin Cancer Biol* 16:16–31
75. Mendez R, Ruiz-Cabello F, Rodriguez T, Del Campo A, Paschen A, Schadendorf D, Garrido F (2007) Identification of different tumor escape mechanisms in several metastases from a melanoma patient undergoing immunotherapy. *Cancer Immunol Immunother* 56:88–94
76. Miller JS, Soignier Y, Panoskatsis-Mortari A, McNearney SA, Yun GH, Fautsch SK, McKenna D, Le C, Defor TE, Burns LJ, Orchard PJ, Blazar BR et al (2005) Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer. *Blood* 105:3051–3057
77. Moretta A, Bottino C, Mingari MC, Biassoni R, Moretta L (2002) What is a natural killer cell? *Nat Immunol* 3:6–8
78. Moretta A, Marcenaro E, Sivori S, Della Chiesa M, Vitale M, Moretta L (2005) Early liaisons between cells of the innate immune system in inflamed peripheral tissues. *Trends Immunol* 26:668–675
79. Moretta A, Vitale M, Bottino C, Orengo AM, Morelli L, Augugliaro R, Barbaresi M, Ciccone E, Moretta L (1993) P58 molecules as putative receptors for major histocompatibility complex (MHC) class I molecules in human natural killer (NK) cells. Anti-p58 antibodies reconstitute lysis of MHC class I-protected cells in NK clones displaying different specificities. *J Exp Med* 178:597–604
80. Moretta L, Bottino C, Pende D, Castriconi R, Mingari MC, Moretta A (2006) Surface NK receptors and their ligands on tumor cells. *Semin Immunol* 18:151–158
81. Moretta L, Bottino C, Pende D, Vitale M, Mingari MC, Moretta A (2004) Different checkpoints in human NK-cell activation. *Trends Immunol* 25:670–676
82. Moretta L, Moretta A (2004) Killer immunoglobulin-like receptors. *Curr Opin Immunol* 16:626–633
83. Moretta L, Moretta A (2004) Unravelling natural killer cell function: triggering and inhibitory human NK receptors. *Embo J* 23:255–259
84. Mrozek E, Anderson P, Caligiuri MA (1996) Role of interleukin-15 in the development of human CD56+ natural killer cells from CD34+ hematopoietic progenitor cells. *Blood* 87:2632–2640
85. Ozdemir O, Ravindranath Y, Savasan S (2005) Mechanisms of superior anti-tumor cytotoxic response of interleukin 15-induced lymphokine-activated killer cells. *J Immunother* (1997) 28:44–52
86. Parham P (2005) MHC class I molecules and KIRs in human history, health and survival. *Nat Rev Immunol* 5:201–214
87. Parrish-Novak J, Dillon SR, Nelson A, Hammond A, Sprecher C, Gross JA, Johnston J, Madden K, Xu W, West J, Schrader S, Burkhead S et al (2000) Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. *Nature* 408:57–63
88. Passweg JR, Stern M, Koehl U, Uharek L, Tichelli A (2005) Use of natural killer cells in hematopoietic stem cell transplantation. *Bone Marrow Transplant* 35:637–643
89. Passweg JR, Tichelli A, Meyer-Monard S, Heim D, Stern M, Kuhne T, Favre G, Gratwohl A (2004) Purified donor NK-lymphocyte infusion to consolidate engraftment after haploidentical stem cell transplantation. *Leukemia* 18:1835–1838
90. Pessino A, Sivori S, Bottino C, Malaspina A, Morelli L, Moretta L, Biassoni R, Moretta A (1998) Molecular cloning of NKp46: a novel member of the immunoglobulin superfamily involved in triggering of natural cytotoxicity. *J Exp Med* 188:953–960
91. Pitini V, Arrigo C, Naro C, Altavilla G (2007) Interleukin-2 and lymphokine-activated killer cell therapy in patients with relapsed B-cell lymphoma treated with rituximab. *Clin Cancer Res* 13:5497
92. Pogge von Strandmann E, Simhadri VR, von Tresckow B, Sasse S, Reiners KS, Hansen HP, Rothe A, Boll B, Simhadri VL, Borchmann P, McKinnon PJ, Hallek M et al (2007) Human leukocyte antigen-B-associated transcript 3 is released from tumor cells and engages the NKp30 receptor on natural killer cells. *Immunity* 27:965–974
93. Reff ME, Carner K, Chambers KS, Chinn PC, Leonard JE, Raab B, Newman RA, Hanna N, Anderson DR (1994) Depletion of B cells in vivo by a chimeric mouse human monoclonal antibody to CD20. *Blood* 83:435–445
94. Robertson MJ (2002) Role of chemokines in the biology of natural killer cells. *J Leukoc Biol* 71:173–183
95. Roda JM, Joshi T, Butchar JP, McAlees JW, Lehman A, Tridandapani S, Carson WE III (2007) The activation of natural killer cell effector functions by cetuximab-coated, epidermal growth factor receptor positive tumor cells is enhanced by cytokines. *Clin Cancer Res* 13:6419–6428
96. Rodella L, Zamai L, Rezzani R, Artico M, Peri G, Falconi M, Facchini A, Pelusi G, Vitale M (2001) Interleukin 2 and interleukin 15 differentially predispose natural killer cells to apoptosis mediated by endothelial and tumour cells. *Br J Haematol* 115:442–450
97. Rosenberg SA (2000) Interleukin-2 and the development of immunotherapy for the treatment of patients with cancer. *Cancer J Sci Am* 6(Suppl 1):S2–S7
98. Rosenberg SA, Lotze MT, Muul LM, Leitman S, Chang AE, Ettinghausen SE, Matory YL, Skibber JM, Shiloni E, Vetto JT et al (1985) Observations on the systemic administration of autologous lymphokine-activated killer cells and recombinant interleukin-2 to patients with metastatic cancer. *N Engl J Med* 313:1485–1492
99. Ruggeri L, Aversa F, Martelli MF, Velardi A (2006) Allogeneic hematopoietic transplantation and natural killer cell recognition of missing self. *Immunol Rev* 214:202–218
100. Ruggeri L, Capanni M, Casucci M, Volpi I, Tosti A, Perruccio K, Urbani E, Negrin RS, Martelli MF, Velardi A (1999) Role of natural killer cell alloreactivity in HLA-mismatched hematopoietic stem cell transplantation. *Blood* 94:333–339
101. Ruggeri L, Capanni M, Urbani E, Perruccio K, Shlomchik WD, Tosti A, Posati S, Rogaia D, Frassoni F, Aversa F, Martelli MF, Velardi A (2002) Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. *Science* 295:2097–2100
102. Ruggeri L, Mancusi A, Burchielli E, Aversa F, Martelli MF, Velardi A (2007) Natural killer cell alloreactivity in allogeneic hematopoietic transplantation. *Curr Opin Oncol* 19:142–147
103. Salih HR, Rammensee HG, Steinle A (2002) Cutting edge: down-regulation of MICA on human tumors by proteolytic shedding. *J Immunol* 169:4098–4102
104. Sayers TJ, Brooks AD, Koh CY, Ma W, Seki N, Raziuddin A, Blazar BR, Zhang X, Elliott PJ, Murphy WJ (2003) The proteasome inhibitor PS-341 sensitizes neoplastic cells to TRAIL-mediated apoptosis by reducing levels of c-FLIP. *Blood* 102:303–310
105. Screpanti V, Wallin RP, Grandien A, Ljunggren HG (2005) Impact of FASL-induced apoptosis in the elimination of tumor cells by NK cells. *Mol Immunol* 42:495–499
106. Sentman CL, Barber MA, Barber A, Zhang T (2006) NK cell receptors as tools in cancer immunotherapy. *Adv Cancer Res* 95:249–292
107. Shahied LS, Tang Y, Alpaugh RK, Somer R, Greenspon D, Weiner LM (2004) Bispecific minibodies targeting HER2/neu and CD16 exhibit improved tumor lysis when placed in a divalent tumor antigen binding format. *J Biol Chem* 279:53907–53914
108. Sheridan C (2006) First-in-class cancer therapeutic to stimulate natural killer cells. *Nat Biotechnol* 24:597

109. Shlomchik WD, Couzens MS, Tang CB, McNiff J, Robert ME, Liu J, Shlomchik MJ, Emerson SG (1999) Prevention of graft versus host disease by inactivation of host antigen-presenting cells. *Science* 285:412–415
110. Skov S, Pedersen MT, Andresen L, Straten PT, Woetmann A, Odum N (2005) Cancer cells become susceptible to natural killer cell killing after exposure to histone deacetylase inhibitors due to glycogen synthase kinase-3-dependent expression of MHC class I-related chain A and B. *Cancer Res* 65:11136–11145
111. Slavin S (2005) Allogeneic cell-mediated immunotherapy at the stage of minimal residual disease following high-dose chemotherapy supported by autologous stem cell transplantation. *Acta Haematol* 114:214–220
112. Slavin S, Naparstek E, Nagler A, Ackerstein A, Samuel S, Kapelushnik J, Brautbar C, Or R (1996) Allogeneic cell therapy with donor peripheral blood cells and recombinant human interleukin-2 to treat leukemia relapse after allogeneic bone marrow transplantation. *Blood* 87:2195–2204
113. Smyth MJ (2006) Imatinib mesylate—uncovering a fast track to adaptive immunity. *N Engl J Med* 354:2282–2284
114. Smyth MJ, Cretney E, Kelly JM, Westwood JA, Street SE, Yagita H, Takeda K, van Dommelen SL, Degli-Esposti MA, Hayakawa Y (2005) Activation of NK cell cytotoxicity. *Mol Immunol* 42:501–510
115. Smyth MJ, Cretney E, Kershaw MH, Hayakawa Y (2004) Cytokines in cancer immunity and immunotherapy. *Immunol Rev* 202:275–293
116. Smyth MJ, Crowe NY, Pellicci DG, Kyparissoudis K, Kelly JM, Takeda K, Yagita H, Godfrey DI (2002) Sequential production of interferon-gamma by NK1.1(+) T cells and natural killer cells is essential for the antimetastatic effect of alpha-galactosylceramide. *Blood* 99:1259–1266
117. Smyth MJ, Hayakawa Y, Takeda K, Yagita H (2002) New aspects of natural-killer-cell surveillance and therapy of cancer. *Nat Rev Cancer* 2:850–861
118. Smyth MJ, Teng MW, Swann J, Kyparissoudis K, Godfrey DI, Hayakawa Y (2006) CD4+CD25+ T regulatory cells suppress NK cell-mediated immunotherapy of cancer. *J Immunol* 176:1582–1587
119. Street SE, Hayakawa Y, Zhan Y, Lew AM, MacGregor D, Jamieson AM, Diefenbach A, Yagita H, Godfrey DI, Smyth MJ (2004) Innate immune surveillance of spontaneous B cell lymphomas by natural killer cells and gammadelta T cells. *J Exp Med* 199:879–884
120. Tam YK, Martinson JA, Doligosa K, Klingemann HG (2003) Ex vivo expansion of the highly cytotoxic human natural killer-92 cell-line under current good manufacturing practice conditions for clinical adoptive cellular immunotherapy. *Cytotherapy* 5:259–272
121. Trapani JA, Smyth MJ (2002) Functional significance of the perforin/granzyme cell death pathway. *Nat Rev Immunol* 2:735–747
122. Uherek C, Tonn T, Uherek B, Becker S, Schnierle B, Klingemann HG, Wels W (2002) Retargeting of natural killer-cell cytolytic activity to ErbB2-expressing cancer cells results in efficient and selective tumor cell destruction. *Blood* 100:1265–1273
123. Ullrich E, Bonmort M, Mignot G, Chaput N, Taieb J, Menard C, Viaud S, Tursz T, Kroemer G, Zitvogel L (2007) Therapy-induced tumor immunosurveillance involves IFN-producing killer dendritic cells. *Cancer Res* 67:851–853
124. Wagtmann N, Biassoni R, Cantoni C, Verdiani S, Malnati MS, Vitale M, Bottino C, Moretta L, Moretta A, Long EO (1995) Molecular clones of the p58 NK cell receptor reveal immunoglobulin-related molecules with diversity in both the extra- and intracellular domains. *Immunity* 2:439–449
125. Waldmann TA (2006) The biology of interleukin-2 and interleukin-15: implications for cancer therapy and vaccine design. *Nat Rev Immunol* 6:595–601
126. Waldmann TA, Dubois S, Tagaya Y (2001) Contrasting roles of IL-2 and IL-15 in the life and death of lymphocytes: implications for immunotherapy. *Immunity* 14:105–110
127. Wallace ME, Smyth MJ (2005) The role of natural killer cells in tumor control—effectors and regulators of adaptive immunity. *Springer Semin Immunopathol* 27:49–64
128. VanOosten RL, Moore JM, Karacay B, Griffith TS (2005) Histone deacetylase inhibitors modulate renal cell carcinoma sensitivity to TRAIL/Apo-2L-induced apoptosis by enhancing TRAIL-R2 expression. *Cancer Biol Ther* 4:1104–1112
129. Weng WK, Levy R (2003) Two immunoglobulin G fragment C receptor polymorphisms independently predict response to rituximab in patients with follicular lymphoma. *J Clin Oncol* 21:3940–3947
130. Vitale M, Bottino C, Sivori S, Sanseverino L, Castriconi R, Marcenaro E, Augugliaro R, Moretta L, Moretta A (1998) NKp44, a novel triggering surface molecule specifically expressed by activated natural killer cells, is involved in non-major histocompatibility complex-restricted tumor cell lysis. *J Exp Med* 187:2065–2072
131. Wu J, Lanier LL (2003) Natural killer cells and cancer. *Adv Cancer Res* 90:127–156
132. Zamai L, Ponti C, Mirandola P, Gobbi G, Papa S, Galeotti L, Cocco L, Vitale M (2007) NK cells and cancer. *J Immunol* 178:4011–4016