REVIEW

Raquel Tarazona · Javier G. Casado · Rocío Soto Olga DelaRosa · Esther Peralbo · Luis Rioja José Peña · Rafael Solana

Expression of NK-associated receptors on cytotoxic T cells from melanoma patients: a two-edged sword?

Received: 1 December 2003 / Accepted: 27 January 2004 / Published online: 4 May 2004 © Springer-Verlag 2004

Abstract The coexistence of tumor progression with a tumor-specific immune response constitutes a major paradox of tumor immunity. During the last decade, the presence of cytotoxic T lymphocytes (CTLs) recognising melanoma-associated antigens has been unequivocally demonstrated in numerous different in vivo and in vitro models. However, most often these melanoma-specific T lymphocytes do not control tumor growth. Several mechanisms that involve changes in melanoma phenotype and/or in T-cell differentiation and function could explain the inability of the immune response to control melanoma. In the last few years it has been demonstrated that cellular cytotoxicity is the result of a balance between activating signals triggered by the TCR and costimulatory molecules and inhibitory signals triggered by inhibitory receptors expressed by the CTL. Because the final outcome of the immune response against melanoma depends on the balance between activating and inhibitory signals, the expression de novo on melanoma cells of ligands for inhibitory NKRs and the down-regulation of costimulatory molecules may favor the escape of tumor cells from immunosurveillance. In this paper we review how altered expression of molecules required for T-cell costimulation could result in impaired lysis of melanoma. The modulation of antimelanoma T-cell responses by a group of receptors originally described on NK cells (NK-associated receptors) but which are now

This article forms part of the Symposium in Writing "Tumor escape from the immune response," published in Vol. 53

R. Tarazona (🖂)

Department of Physiology, Immunology Unit,

Veterinary Faculty, University of Extremadura, Cáceres, Spain E-mail: rtarazon@unex.es

R. Soto · L. Rioja

known also to be expressed on a subset of cytolytic effector cells is reviewed. We hypothesize that the expression of ligands for NKRs on melanoma cells may contribute to T-cell-mediated immune responses against melanoma either enhancing or inhibiting activation and differentiation to effector cells. Blocking inhibitory receptors or increasing activating receptors could result in new strategies to improve T-cell-mediated rejection of melanoma.

Keywords CD28 · CD94/NKG2 · Costimulation · KIR · NKR · Tumor escape · Tumor immunity

Abbreviations

CTL	cytotoxic T lymphocyte		
Ig	immunoglobulin		
ILT	immunoglobulin-like transcripts		
ITAM	immunoreceptor	tyrosine-based	activation
	motif		
ITIM	immunoreceptor	tyrosine-based	inhibition
	motif		
KIR	killer cell immunoglobulin-like receptors		
NK cell	natural killer cell		
NKR	natural killer cell-associated receptor		
TCR	T-cell receptor	-	

Introduction

Cytolytic lymphocytes play a pivotal role in protection from tumors. Several strategies have been developed to enhance tumor recognition, including induction of MHC molecules by cytokines. In the last decade, it has been reported that cytotoxic T-cell activation and effector function, upon recognition of specific peptides presented by MHC class I, may be regulated by the expression of natural killer (NK) cell–associated receptors (NKRs) specific for major histocompatibility complex (MHC) class I molecules [13]. NKRs were first identified on NK cells as a regulatory mechanism of

J. G. Casado · O. DelaRosa · E. Peralbo · J. Peña · R. Solana (⊠) Department of Cellular Biology, Physiology and Immunology, Faculty of Medicine, University of Córdoba, Córdoba, Spain E-mail: rsolana@uco.es

Department of Plastic Surgery, Reina Sofia University Hospital, Córdoba, Spain

cytotoxicity [47, 62, 63, 81, 88, 146]. Although most NKRs are not present on high percentages of T cells, a subpopulation of T cells that expresses those receptors and that corresponds to cells with a memory/effector phenotype has been identified [86]. The majority of T cells expressing NKRs are included within the CD8⁺CD28⁻ T-cell subset, characterized by higher cytotoxic capacity, diminished proliferative response, shortened telomeres, and low levels of telomerase activity [116, 125]. The functional significance of the expression of ligands for NKRs and costimulatory molecules on melanoma cells and their correlation with their susceptibility to lysis by T lymphocytes and NK cells has not yet been fully established. In addition, the study of the role of several cytokines as modulators of NKRs and costimulatory molecule expression on both melanoma and lymphocytes may open new avenues for successful immunotherapy against melanoma.

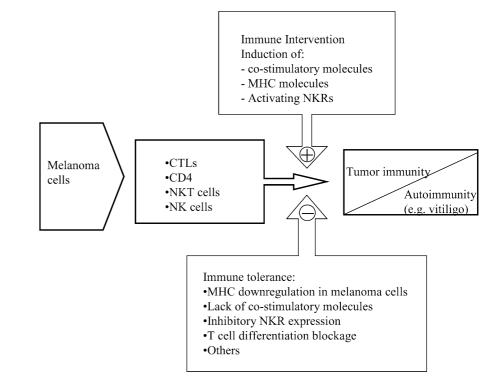
Immune response against melanoma

Different components of humoral and cellular immune responses are involved in the elimination of tumor cells, although apparently paradoxical, tumor immunity frequently coexists with tumor progression (Fig. 1). Even though cellular immune responses against tumors are generally mediated by T lymphocytes, the role of other components of the immune response such as NK cells, NKT cells, antibodies, or phagocytes in the elimination of melanoma cells has also been defined in different experimental models.

Thus, antibodies against melanoma have been identified in many patients but their role in the anti-

Fig. 1 Immune responses against melanoma. Different cell types participate in the rejection of tumor cells leading to tumor immunity. A potential consequence of tumor immunity is autoimmunity. Immune intervention can enhance tumor immunity and overcome the immune tolerance status mediated by several escape mechanisms tumoral immune response has not been fully clarified. It has been reported that the induction of antibodies against melanoma antigens after vaccination may contribute to the therapeutic efficacy by a mechanism primarily involving antibody destruction of tumor cells by antibody-dependent cellular cytotoxicity (ADCC) through the engagement of Fc receptors [76]. Furthermore, antibody-coated tumor cells could also be killed by a process involving opsonization and phagocytosis. However, the role of macrophages in the immune response against melanoma is mainly related to their role as antigen-presenting cells and to their capacity to secrete several effector molecules such as tumor necrosis factor α (TNF- α) and interleukin (IL)-12 that may participate in melanoma rejection [54].

Natural killer (NK) cells are cytolytic cells that also contribute to the immune responses against tumors as they kill some cancer cells without prior sensitization and without a requirement for MHC restriction [95, 145]. NK cells are CD3-negative lymphocytes and, within NK cells, two subsets have been defined according to the level of expression of CD56. The CD56^{dim} NK-cell subset is characterized by higher cytotoxic capacity than the CD56^{bright} subset, which represents a minority subset and is characterized by the production of greater amounts of cytokines [24]. NK-cell cytotoxicity has been found to be diminished in patients with metastatic melanoma and could reflect the immunosuppressed state associated with advanced tumors [60]. NK cytotoxicity is controlled by the balance of activating and inhibitory signals mediated by different receptors some of which bind to MHC class I molecules on target cells [13, 47, 124, 146]. As will be discussed in this review, these receptors can play a significant role in



tumor immunity because of their expression by a subset of cytotoxic T cells.

T-cell-mediated immune response against melanoma

Cellular immunity plays a pivotal role in the immune response against melanoma and an active T-cell-mediated immunity against melanoma-associated antigens has been demonstrated in different in vivo and in vitro models [102]. During the last decade, significant progress has been made in the identification and characterization of several MHC class I-restricted melanoma-associated antigens recognized by CTLs. These antigens belong to three main groups: cancer-testis-specific antigens (e.g., MAGE, BAGE, GAGE, PRAME, and NY-ESO-1), melanocyte differentiation antigens (e.g., tyrosinase, Melan-A/MART-1, gp100, TRP-1, and TRP-2), and mutated or aberrantly expressed antigens (e.g., MUM-1, CDK4, beta-catenin, gp100-in4, p15, and N-acetylglucosaminyl transferase V). $CD4^+$ T cells can also recognize several MHC class II-restricted epitopes from melanoma antigens [46, 72, 128, 129, 131]. The discovery of new antigens recognized by T cells has allowed a better understanding of the cellular and molecular bases of the immune response against melanoma. Because many of these melanoma-associated antigens are selfproteins, T-cell effector function is frequently found to be weak or absent as a consequence of immune tolerance, which allows melanoma cells to escape from immune responses [35].

The development of MHC class I–peptide tetrameric complexes to study antigen-specific T cells ex vivo by flow cytometry has significantly increased our understanding of the immune response against melanoma antigens and has allowed the quantification and phenotypic characterization of specific $CD8^+$ T cells [70, 94]. At the same time, the improvement of technology for the in vitro expansion of specific T cells allows the functional characterization of different T-cell subsets [17]. Such approaches have shown that different melanoma-associated antigens may stimulate a differential T-cell response with selective involvement of either CD4 or CD8 T cells [50].

Melanoma-specific T cells: differentiation from naïve to effector and memory cells

Differentiation of naïve $CD8^+$ T cells toward effector/ memory T cells is required for an effective immune response against melanoma. Frequently, CTLs from melanoma patients show a low cytotoxic capacity, and it has been postulated by several authors that this defect could be the consequence of an impaired maturation from memory/effector lymphocytes to terminally differentiated cytotoxic effector lymphocytes [2, 35, 70, 90].

T-cell differentiation from naïve to effector cytolytic T lymphocytes is an area of intense study by many

investigators. Although no consensus on the transition from naïve to memory cells has yet been established, four different stages of maturation have been defined within CD8⁺ T cells according to their expression of CCR7 and CD45RA molecules [108]. Whereas CD8 naïve cells express both CCR7 and CD45RA, central memory cells are CCR7⁺CD45RA⁻, and effector memory cells CCR7⁻CD45RA⁻. A fourth stage of differentiation that constitutes terminally differentiated cytotoxic effector cells is defined by the lack of CCR7 expression, the reacquisition of CD45RA and large amounts of perforin [108]. The relationship between central memory and effector/memory T cells has been further analyzed in humans [7] and mice [130, 143]. In mice, a linear differentiation model has been proposed by Wherry et al. in which effector/memory cells are derived from effector cells, and central/memory cells are derived from effector/memory cells [143]. This model has not been confirmed in humans, and recent results by Baron et al. suggest that most effector memory and central memory cells are independently generated [7].

Different stages of T-cell maturation, from naïve $(CD45RA^+CCR7^+)$ to terminally differentiated (CD45RA⁺CCR7⁻) CD8⁺ T cells, have been observed in melanoma-specific T cells obtained from patients [3, 56, 134]. Dunbar et al. [33] found that Melan-A-specific $CD8^+$ T cells from melanoma patients may have a CCR7⁺CD45RA⁺ naïve phenotype associated with a defective immune response to Melan-A peptide ex vivo. In contrast, a group of melanoma patients with metastases were found to possess Melan-A-specific CD8⁺ T cells with a CCR7⁻CD45RO⁺CD45RA⁻ effector memory phenotype and were able to respond to Melan-A peptide. Tyrosinase-specific T cells with an effector/ memory phenotype have also been found in bone marrow from melanoma patients at similar or higher levels than in peripheral blood [71].

In melanoma patients, CD8⁺ T cells from tumor-free lymph nodes showed a naïve phenotype characterized mainly by the expression of CCR7 and CD45RA [3]. A similar phenotype was found in a large fraction of CD8⁺ T cells specific for melanocyte differentiation antigens or tumor-restricted antigens obtained from tumor-invaded lymph nodes. In contrast, preterminally differentiated CTLs (CCR7⁻CD45RA⁻CD27⁺CD28⁻ perforin⁺) were observed in tumor-invaded lymph nodes from a small fraction of melanoma patients, but no evidence for tumor regression was found in these patients. This may suggest a lack of terminally differentiated CD8⁺ T cells [3, 90], but interestingly, in vitro treatment with IL-2 or IL-15 promoted differentiation to melanoma-specific effector T cells [3].

Antitumor activity of NKT cells

Recently, natural killer T (NKT) cells have been implicated in defense against tumors [26, 65, 66, 113]. NKT cells represent a novel T-cell lineage characterized by the restricted expression of an invariant TCR α chain encoded by $V\alpha 24/J\alpha Q$ gene segments in humans and $V\alpha 14/J\alpha 281$ in mice. NKT cells constitute a small subpopulation of T cells (0.1-0.5% of peripheral blood lymphocytes) that have been frequently confused with NK cells because they share several phenotypic and functional characteristics [45]. Because NKT cells can express NKRs (e.g., CD161, CD56) it was also suggested that they could represent the subset of NKR⁺ cytotoxic T cells. However, detailed analysis of these cells has demonstrated that NKT and NKR⁺ cytotoxic T cells represent different T-cell subpopulations [29]. NKT cells react with CD1d and, after stimulation, produce high levels of IL-4 and interferon (INF)- γ [37]. The specific ligand of murine Va14⁺ and human Va24⁺ NKT cells is α -galactosylceramide (α GalCer) which is presented by the CD1d molecule [45]. Recently, a ganglioside named GD3 that is presented by CD1d molecules has been found to be expressed on several human tumors including melanoma. GD3 expression on normal tissues is low or absent. Furthermore, NKT cells from mice immunized with a human melanoma cell line expressing GD3 recognized human GD3 cross-presented by mouse APCs in a CD1d-restricted fashion, resulting in production of IL-4, IFN- γ , and IL-10 [144].

It has been observed that NKT cells exert protective immunity against melanoma in vivo and in vitro. In a murine model, dendritic cells pulsed with α GalCer have been shown to be effective against tumor development in vivo by means of activating Val4 NKT cells and increasing their cytolytic capacity [64, 132]. Studies in vitro using human $V\alpha 24^+$ NKT cells from melanoma patients have shown that aGalCer-stimulated NKT cells did not display cytolytic activity against melanoma cell lines (autologous or allogeneic) but were able to suppress proliferation of melanoma cell lines cocultured with activated V α 24⁺ NKT cells. This effect was mediated by the induction of cytokine release (IFN- γ and to a lesser extent IL-12) rather than by direct cytotoxicity [66]. Together, these results indicate that α GalCer can induce antitumor effects that can be mediated by increased cytolytic activity or by cytokine release depending on the experimental model used.

Balance between activating and inhibitory signals

As mentioned above, the final outcome of T-cell-mediated immune responses against melanoma depends on a balance between activating and inhibitory signals. Activation of T lymphocytes requires two signals, the first mediated by T-cell receptor (TCR) recognition of the specific peptide in the context of an appropriate MHC molecule and the second mediated by costimulatory receptors on T cells that interact with their specific ligands on antigen-presenting cells or target cells. Interaction of CD28 on T cells with members of the B7 family (e.g., CD80 and CD86) on target cells delivers the second signal for an efficient T-cell activation after TCR- mediated recognition of the specific ligand [78]. In addition to CD28-B7, other receptors have recently been defined that also enhance T-cell responses and may be important in tumor immunity. Manipulation of the immune system through induction of costimulatory molecules could lead to enhanced immune responses against tumors [136].

NKRs expressed on T cells can mediate negative as well as positive signals, resulting in melanoma growth promotion or suppression, respectively. The role in tumor rejection of signals mediated by activating NKRs has not been characterized. In contrast, interaction of inhibitory NKRs on the effector cells with their ligands on tumor cells leads to inhibition of TCR-mediated cytotoxicity against melanoma [6, 137]. Modulation of this balance between activating and inhibitory signals invites a new strategy for enhancing effective immunity against melanoma.

Expression of costimulatory molecules by T cells from melanoma patients

Delivery of a costimulatory signal through engagement of CD28 on T cells by members of the B7 family (e.g., CD80 [B7.1], CD86 [B7.2]) on antigen-presenting cells is essential for full T-cell activation, potentially leading to tumor elimination. The outcome of T-cell costimulation through B7 molecules involves a combination of activating and inhibitory signals from CD28 and CTLA-4 molecules [20, 36, 112]. Although the expression of B7 family molecules in nonlymphoid tissues is mostly not constitutive, it is sometimes inducible by cytokines such as IFN- γ , IFN- α , IFN- β , or TNF- α [112]. It should be also considered that recognition of melanoma antigens by tumor-infiltrating lymphocytes induces the release of several cytokines such as granulocyte-macrophage colony-stimulating factor (GM-CSF), TNF- α , and IFN- γ that may have clinical relevance [49, 109].

Frequently, T-cell responses in melanoma patients are defective [70], and tumor-associated antigens expressed by melanoma cells are poorly immunogenic due, in part, to the lack of expression of costimulatory molecules by human melanoma cells [10, 19, 43, 48] that render melanoma-specific CTLs anergic in vivo [70]. Thus, it has been reported that the expression of CD80 and CD86 molecules in spontaneously regressing primary melanomas improved T-cell-mediated antitumor immunity, whereas these molecules were absent in metastatic melanoma and consequently favored the escape of melanoma cells from immune surveillance [30]. On the other hand, others have found no significant correlation between CD80 and CD86 expression on melanoma metastases and regression of the tumor, time to progression or survival [11].

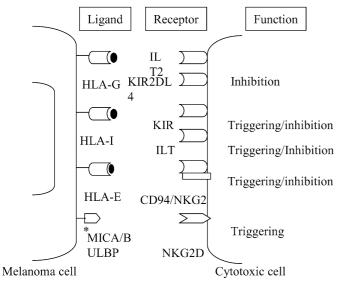
It has also been reported that transfection of melanoma cells with the costimulatory molecules CD80 or CD86 improves immune responses mediated by both $CD4^+$ and $CD8^+$ T cells [14, 34, 74, 80]. Transfection of

HLA-A*0201 melanoma cell lines with both the IL-2 and B7-1 (CD80) genes also resulted in an improved response by specific CTLs. Transfected melanoma cells displayed higher immunogenicity even with low antigen expression and were able to stimulate cytotoxicity and IFN- γ release by autologous and HLA-A*0201–compatible allogeneic T cells [79].

Although the CD28-CD80/CD86 and CTLA4-CD80/ CD86 pathways of costimulatory and inhibitory signals are the best characterized, other new members of the B7/CD28 superfamily have recently been identified. One involves the molecule B7 h (also known as ICOSL, LI-COS, GL50, B7RP-1, and B7H2) a member of the B7 family of costimulatory molecules, which interacts with inducible costimulatory molecule (ICOS). ICOS is expressed on activated T cells and it has been found that expression of B7 h on tumor cells enhanced CTL response in vivo [36, 75, 112, 139]. Recently, B7-H1 (PD-L1) has been identified as a new member of the B7 family of costimulatory molecules. B7-H1 molecules are involved in the negative regulation of the immune responses through engagement of the PD-1 (programmed death-1) receptor on activated T and B cells [18]. Interestingly, B7-H1 expression has been found in several tumors including melanomas. Furthermore, B7-H1 expression may be induced by IFN- γ treatment of tumor cell lines [32]. In vitro experiments showed that the interaction of B7-H1 with its ligand induces apoptosis of activated tumor-reactive T cells and may favor the growth of B7-1⁺ tumors in vivo [32]. The expression of B7-H1 on tumor cells can represent an escape mechanism from the T-cell immune response, and blockade of

Fig. 2A,B Inhibitory and activating NKRs. Expression and function of human NKRs on cytotoxic cells and their potential ligands on melanoma cells. A HLA class I–specific receptors. B Non-HLAspecific receptors

A) HLA class I-specific NKRs and their ligands



*MHC related molecules

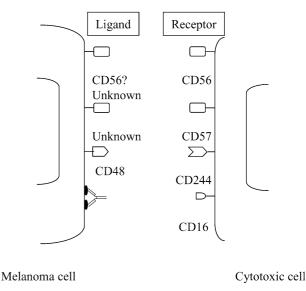
The human NK-like cell line YT-Indy constitutively expresses CD28 that triggers NK-mediated lysis of target cells expressing B7. By transfection of KIR2DL2 on YT-Indy cells we have analyzed the role of inhibitory NKRs on CD28-mediated lysis. Interaction of KIR2DL2 on YT-Indy cells with its ligand, HLA-Cw3, on target cells inhibited CD28-dependent cytotoxicity. This inhibition was blocked by the addition of anti-KIR2DL2 or anti-HLA class I mAb, whereas it was not affected by the addition of other control mAb [126]. CD28-mediated lysis of CD80-transfected melanoma cells by YT-Indy cells has also been observed and this effect was inhibited by interaction of the inhibitory receptor KIR2DL2 on YT-Indy with its ligand on melanoma cells (J.G. Casado et al., unpublished data).

Manipulation of costimulation through B7 molecule interactions with their different receptors on effector cells is therefore expected to provide a useful model not only to further our understanding of immune responses against melanoma but also for the development of new immunotherapy strategies.

Inhibitory and activating MHC class I–specific NK receptors on T cells in melanoma patients

NKRs were firstly described in NK cells and in a subset of CTLs and play an important role in regulating cytotoxicity (for review, see [13]). In humans, three families of genes encode these HLA class I–specific receptors (Fig. 2A). The first family identified is called the killer cell immunoglobulin-like receptor (KIR) which consists of type I transmembrane molecules belonging to the immunoglobulin (Ig) superfamily, and interacts with HLA class I molecules to regulate NK- and T-cell

B) Non-HLA-specific receptors and their ligands



function. KIRs with long cytoplasmic tails contain immunoreceptor tyrosine-based inhibitory motifs (ITIM) (e.g., KIR2DL and KIR3DL) and inhibit lysis of target cells expressing their MHC class I molecule ligands [87], and also inhibit cytokine production by the effector cells [27]. In contrast, KIR with short cytoplasmic tails (e.g., KIR2DS) can activate NK-cell cytotoxicity toward target cells expressing an appropriate MHC class I ligand [89] through interactions with the adaptor molecule DAP12, a molecule expressed as a homodimer that contains an immunoreceptor tyrosine-based activation motif (ITAM) [69]. A second group of receptors also belonging to the Ig superfamily are named immunoglobulin-like transcripts (ILTs) and are expressed predominantly in monocytes, NK cells, and also on a subpopulation T cells. KIR and ILT ligands include several classical HLA class I molecules and the nonclassical HLA-G molecule. C-type lectin family members constitute the third family of HLA-specific NK receptors and are composed of heterodimers of CD94 covalently associated with a member of the NKG2 family of molecules. The ligand for CD94/NKG2A (inhibitory) and CD94/NKG2C (activating) receptors is the nonclassical class I molecule HLA-E. The inhibitory members of this family contain ITIM in their cytoplasmic tails and the activating receptors associate with DAP12 [13]. It has been repeatedly demonstrated that KIR, ILT, and C-type lectin receptors (CD94:NKG2) can modulate T-cell cytotoxicity [86].

Predominance of stimulatory signals mediated by activating NKRs has been observed in autoimmune diseases. Thus, CD4⁺ T cells from rheumatoid arthritis patients frequently lack the costimulatory molecule CD28 but have an increased expression of activating forms of KIRs (CD158j). These receptors were functional and enhanced IFN- γ production after T-cell receptor stimulation. These results suggest that activating KIRs in T cells function as costimulatory molecules and may contribute to the autoreactivity observed in rheumatoid arthritis [92, 114].

NKG2D is another C-type lectin-activating receptor that is expressed by both NK cells and CTLs in humans and by all NK cells and activated T cells in mice [103]. NKG2D recognizes MICA and MICB that are stressinduced cell-surface MHC class I-related molecules. Interestingly, MICA and MICB expression is frequently up-regulated on tumor cell lines and primary tumors of epithelial origin [110, 138], and shedding of MIC has been observed in epithelial tumors [39]. Expression of MIC has been recently reported in primary and metastatic melanomas, and NKG2D expression was also found in lymphocytes infiltrating the tumor [138]. NKG2D/MIC interaction induces endocytosis and degradation of NKG2D. Furthermore, defective expression of NKG2D has been linked to the presence of tumor-associated MIC that impairs NK- and T-cellmediated immune responses and constitutes a new immune evasion mechanism [39]. NKG2D also recognizes a new family of MHC class I-related molecules named

UL16-binding proteins (ULBPs). Four different ULBP proteins (ULBP1–4) have been identified so far and are characterized by their ability to bind to the cytomegalovirus (CMV) glycoprotein UL16 [25, 57, 119]. These results suggest that NKG2D/MIC interactions may be involved in NK-cell-mediated as well as T-cell-mediated cytotoxicity to melanoma cells [82, 99]. Furthermore, NKG2D-mediated NK-cell activation may be inhibited by the engagement of inhibitory NKRs [99].

The expression of classical MHC class I molecules on melanoma cells is required for TCR-mediated recognition by specific T cells and the down-regulation of HLA class I molecules in melanoma cells is an important tumor escape mechanism (for review, see Garrido et al. in this Symposium in Writing). Furthermore, although its in vivo relevance has not been defined, the engagement of MHC class I-specific activating receptors might enhance T-cell-mediated immune responses against melanoma cells.

In contrast, as discussed above, MHC class I molecules on melanoma cells can also be recognized by inhibitory receptors that would block T-cell activation and then favor tumor escape. The analysis on tumor cells of HLA-A, HLA-B, HLA-C, and nonclassical MHC molecules such as HLA-E and HLA-G that are recognized by KIR, CD94/NKG2, and ILT receptors, respectively, has been limited by the absence of monoclonal antibodies. Even though MHC class I down-regulation in tumor cells is a well-known escape mechanism to avoid T-cell recognition, frequently tumor cells are also resistant to NK-mediated lysis and the expression of nonclassical HLA molecules recognized by inhibitory receptors has been postulated to be involved in this process.

In a recent study, the expression of HLA-E molecules was analyzed in a panel of tumor cells. The expression of HLA-E on the cell surface of tumor cell lines correlated inversely with the expression of other HLA class I molecules and was related to the availability of β_2 -microglobulin. Thus, HLA-E expression could be induced in an HLA class I-negative melanoma cell line by transfection of the β_2 -microglobulin gene [77]. Therefore, in contrast with total loss of HLA class I molecules, partial loss of HLA-A, HLA-B, or HLA-C class I molecules (allelic or haplotype HLA loss) may at the same time inhibit CTL responses by deleting the restriction element and also inhibit NK-cell-mediated cytotoxicity by expressing HLA-E molecules. Furthermore, HLA-E expression on tumor cells could inhibit specific CTL-mediated cytotoxicity by interaction with inhibitory NKRs (CD94/NKG2A) [77].

An increased expression of NKRs, including HLAspecific receptors, on CD8⁺ T lymphocytes has been observed in several clinical conditions that involve chronic activation of the immune system (e.g., HIV infection, as well as tumor patients) [86, 116, 125]. Furthermore, the expression of NKRs in T cells has been associated with cells that have undergone a process of replicative senescence after multiple rounds of cell division probably as a consequence of chronic stimulation [15, 124].

Recently, the expression of several NKRs on T cells has been analyzed in melanoma patients. However, the proportion of NKR⁺ T cells showed great variation between individuals, and it was therefore difficult to establish significant differences compared with healthy donors [115, 116]. The functional capabilities of NKRs expressed on T cells from melanoma patients have been assessed against melanoma cells. Thus, HLA-specific inhibitory NKRs have been shown to inhibit lysis of melanoma cells mediated by specific CTLs transfected with the inhibitory receptor KIR3DL1 [6]. It has also been reported that melanoma-specific CTLs can express inhibitory NKRs in vivo that impair their cytolytic activity [51, 53, 98, 115].

Ikeda et al. found the expression of an inhibitory NKR (KIR2DL2) on melanoma-specific CTLs that was able to inhibit cytolysis upon interaction with its ligand (HLA-Cw7) on melanoma cells [53]. The expression of C-type lectin-like NKRs has been also analyzed in melanoma patients. Triggering of CD94/NKG2A inhibitory receptor by its ligand on melanoma cells induced inhibition of CTL-mediated lysis of melanoma cells [115]. Both inhibitory and activating forms of CD94/NKG2 receptors have been found in MART-1–specific T cells in different areas of a primary melanoma. Inhibitory CD94/NKG2-A receptors were found exclusively in the vitiligo-like areas, whereas both the inhibitory receptors and the activating CD94/NKG2-C isoforms were present within the tumor [98].

In vitro cultures of $CD8^+$ T cells primed with melanocyte differentiation antigens, showed that a fraction of $CD8^+$ T cells from healthy donors expressed both activating and inhibitory forms of NKRs, but only inhibitory receptors display functional activity inhibiting effector functions of $CD8^+$ T cells [51]. Furthermore, phenotypic characterization of melanoma-specific $CD8^+$ T lymphocytes expanded in vitro has allowed the identification of changes in several NKRs after stimulation [51, 117] (Casado et al. unpublished results).

We have previously discussed NKR expression as effector molecules that may trigger or inhibit CD8⁺ Tcell function [86, 124]. However, recent data also support the notion that NKR expression may correlate with the differentiation stage of T cells [147]. Although NKR expression is preferentially observed in the CD8⁺CD28⁻ T-cell subset, we have found that other subsets of CD28⁺ T lymphocytes can also express NKRs [19], and we suggest that these subsets correspond to intermediate maturation stages from "naïve" to cytolytic effector cells. Thus, a differential expression of KIR and ILT2 receptors in CTLs that correlates with transition from effector to memory lymphocytes has been recently demonstrated [147]. KIR expression is associated with acquisition of a memory phenotype and resistance to apoptosis induced by antigenic activation. In contrast, ILT2 expression correlates with effector cytotoxic cells. Recently, KIR [133] and CD94/NKG2A [42] expression Expression of non-MHC-specific NK receptors on T cells in melanoma patients

differentiation as targets for possible immunotherapies.

Several receptors, usually expressed on NK cells, recognizing ligands others than MHC class I molecules, are also expressed by a subpopulation of T cells and, to some extent, may contribute to cytotoxicity or other effector functions (Fig. 2B). One of these receptors is CD56, an isoform of the neuronal cell-adhesion molecule (NCAM), that is involved in homotypic and heterotypic binding of cytotoxic cells to target cells, although its precise function, either on NK cells or on T cells, remains elusive [68]. CD56 expression on NK cells defines two different subsets according to the cell-surface density of this marker, CD56^{dim} and CD56^{bright} NK cells, with differential phenotype and function [24, 96, 127]. In contrast to NK cells, CD56 expression on T cells is not constitutive. It has been observed that $CD56^+$ T cells are almost absent at birth and in adults are found as oligoclonal expansions [101]. Cord blood CD3⁺ cells can acquire CD56 expression after culture with IL-15 and the expanded CD3⁺CD56⁺ T cells have a $CD8^+CD25^+$ IFN- γ^+ phenotype, with 40% being $\gamma\delta$ T cells [23]. Furthermore, the percentage of $CD8^+CD56^+$ T cells increased with age (DelaRosa, unpublished results) but is diminished in HIV infection [125]. CD8⁺CD56⁺ T cells generally contain high amounts of intracellular perforin and granzyme B, and CD56 expression on $CD8^+$ T cells correlates with cytolytic activity [101]. The finding that CD8⁺CD28⁻DR⁻ T cells have low telomerase activity and frequently express CD56 [118] further supports the idea that $CD56^+$ T cells have an effector/memory phenotype. CD56 expression has been demonstrated in several tumors of the nervous system, melanomas, and other epithelial cancers suggesting that CD56 molecules may be involved in tumor biology [52, 105]. CD56-mediated homotypic adhesion between CD56⁺ tumor cell lines and NK cells has been shown [93, 122].

CD57 is an oligosaccharide found on many cell-surface glycoproteins that has been implicated in cell-to-cell and cell-to-extracellular matrix adhesion. CD57 is preferentially expressed in NK cells and in a subset of T cells. The expansion of the CD8⁺CD28⁻CD57⁺ T-cell subset is age-associated probably due to repeated antigenic stimulation [83]. Several studies report the association between viral infections (e.g., human cytomegalovirus) and increased numbers of CD8⁺CD57⁺ T cells [107, 142]. Furthermore, CD57 expression on virusspecific $CD8^+$ T cells is related to a state of replicative senescence with low proliferative capacity and short telomeres [15]. CD57⁺ T cells are also frequently found in tumor patients, generally as a monoclonal or oligoclonal expansions, have a low rate of turnover and low expression of the apoptotic marker CD95 [120], and exert higher cytotoxic activity ex vivo than their CD57⁻ counterparts in an anti-CD3-redirected assay [135]. It has also been found that most NKR⁺ T cells are included within the CD28⁻CD57⁺ subset [84]. This CD57⁺ T-cell subset comprises the majority of the CD56⁺ T cells previously described, which represent differentiated effector T cells [101]. Glycoproteins containing carbohydrate epitopes recognized by antibodies against CD57 can also be found in several tumors including melanoma. Moreover, CD57 expression on melanoma cells has been associated with metastatic potential and seems to have a function in intercellular adhesion [58, 123].

CD244 (2B4 or C1.7) is a cell-surface glycoprotein related to CD2 that belongs to the Ig-like superfamily and is involved in T- and NK-cell activation [21, 59]. CD244 is expressed on the surface of all human NK cells, a subset of CD8⁺ T cells, monocytes, and basophils. In NK cells, CD244 ligation can induce redirected lysis [21, 67]. It has been reported that CD244 expression on CD8⁺ T cells overlaps with activation markers such as granzyme B and perforin [117]. CD244 expression can be induced in vitro on CD8⁺ T cells by anti-CD3 cross-linking, and its expression constitutes a marker for activated/memory CD8⁺ T cells [100]. Furthermore, recent studies show that CD244 activation causes an increased expression of matrix metalloproteinase-2 and may induce degradation of extracellular matrices promoting lymphocyte invasion of tumors [21]. CD48, the ligand for CD244, is a ubiquitously expressed molecule in humans [16, 91]. Until now, expression of CD48 has not been reported in melanoma cells; however, it has recently been shown that CD244-CD48 interactions can occur directly between T cells and induce proliferative responses of neighboring T cells [61]. Furthermore, transfection of CD48 into poorly immunogenic melanoma cells enhances the immune response [73]. In melanoma patients, CD244 was found to be upregulated in Melan-A-specific T cells, although its role in promoting T-cell effector function has not been established [117]. In addition, inhibitory receptors as KIR2DL1 or CD94/NKG2 can block CD244 activation [141].

CD16 is a low-affinity Fc receptor for IgG (Fc γ RIII) that mediates ADCC by binding to the Fc portion of antibodies. CD16 is mainly expressed in a subset of NK cells (CD56^{dim}) and in a small subpopulation of CD3⁺ lymphocytes. Ligation of CD16 stimulates not only cytotoxicity but also cytokine production [1] and proliferation [140]. In animal models, CD16 has been

involved in both passive and active immunization against melanoma [22].

CD161 (NKRP1) belongs to the C-type lectin family and is expressed by NK cells and a subpopulation of T cells. While the expression of the majority of NKRs on T cells is essentially restricted to CD8⁺ T cells, CD161 is expressed on both CD4 and CD8 T cells. Moreover, expression of CD161 is a characteristic of NKT cells [121, 124]. CD161 cross-linking triggers cytotoxicity. Although its ligand remains to be identified, it has been postulated that this receptor recognizes carbohydrate structures expressed on target cells [4].

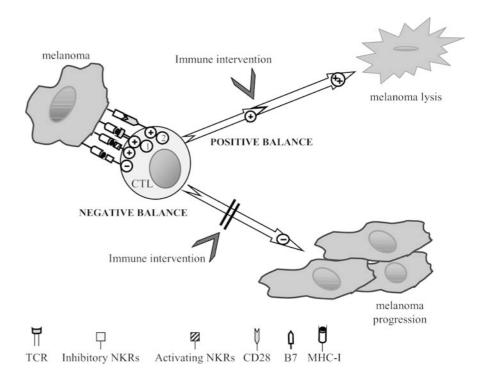
Natural cytotoxicity receptors (NCRs) expressed in NK cells have recently been identified. These receptors belong to the Ig-superfamily but their ligands are not HLA class I molecules. NKp46, NKp44, and NKp30 were the first members identified and are characterized by their capacity to trigger natural cytotoxicity. NCRs associate with signal-transducing polypeptides containing ITAM. The expression of NCR seems to be restricted to NK cells and, so far, none of these receptors has been identified in T cells.

Several groups, including our own, have found that the expression of CD56, CD57, CD244, or CD16 in T cells is associated with an activated/effector phenotype [28, 38, 101, 116, 117]. NKR expression is mainly restricted to the CD8⁺CD28⁻CD56⁺ T-cell subset that represents the mature effector cytolytic subset responsible for the immune response against viral infection and tumors [116, 125].

Different studies show that several cytokines may modulate the expression of HLA-specific NKRs on T lymphocytes. Thus, TGF- β [12, 41], IL-10 [106], and IL-15 [85] have been implicated in the induction of CD94/ NKG2A receptors on T cells. Thus, IL-15 produced by melanoma cells induces the expression of inhibitory NKRs on specific T cells and may constitute a new escape mechanism [8]. Recently, the induction of CD94/ NKG2A receptors has also been observed after IL-12 treatment. Because IL-12 is frequently used for the in vitro induction of antigen-specific T cells, the expression of the inhibitory receptor CD94/NKG2A may have functional relevance [31]. In contrast, expression of the triggering receptor CD161 is also up-regulated by IL-12 [5].

Conclusions

A major paradox of tumor immunity is that tumor progression frequently, if not always, coexists with tumor-specific immune responses (Fig. 1). Numerous studies have been performed to investigate possible tumor escape mechanisms from immunosurveillance. Thus, MHC class I down-regulation by tumor cells represents a well-known major evasion mechanism. Expression of HLA class I has been found to be altered in human melanoma, including total absence of HLA class I, loss of expression of particular allotypes, or loss Fig. 3 Inhibitory/activating balance. Different receptors participate in the immune response against melanoma mediated by CTLs. Positive signals mediated by TCR and costimulatory molecules may be enhanced by the expression of activating NKRs on CTLs and their ligands on melanoma. By contrast, inhibitory signals triggered by inhibitory NKRs after interaction with their ligands will allow melanoma escape from immunosurveillance



of heterozygosity (LOH) leading to an HLA haplotype loss [97, 104].

Total loss of HLA class I molecules would interfere with CTL recognition but would render melanoma cells susceptible to NK-cell-mediated cytotoxicity. A recent report correlates inversely the expression of HLA-E on the cell surface of tumor cell lines with the expression of other HLA class I molecules [77]. Thus, partial loss of HLA class I molecules (allelic or haplotype HLA loss) may inhibit CTL responses if the restriction element is lost and may favor the expression of HLA-E molecules that can be recognized by inhibitory NKRs (CD94/ NKG2A) on T cells or NK cells.

The expression of surface molecules on melanoma cells, such as adhesion molecules (e.g., intercellular adhesion molecule 1 [ICAM-1]) has been associated with aggressive tumors and defective immune responses. ICAM-1 shedding by melanoma cells can inhibit conjugate formation between T-cell clones and the autologous melanoma cells and abrogated the MHC-restricted killing of the melanoma by T-cell clones. In addition, melanoma cells induced anergy of CD4⁺ T cells due to lack of a costimulatory signal mediated by B7/CD28 interactions [9].

Many of the antigens presented by melanoma cells are unaltered self-proteins and may induce autotolerance. Several strategies have been developed to overcome poor immunogenicity and tumor evasion of CTL-mediated destruction. It has been observed that induction of HLA class I expression on melanoma cells can restore specific CTL responses [111]. In contrast, increased HLA I expression in melanoma cells could induce the activation of HLA-specific inhibitory NKRs leading to inhibition of lysis both in NK cells and T cells. On the other hand, activating HLA-specific receptors could be expressed both in NK cells and T cells. The final balance between activating and inhibitory signals will generate an effective immune response or tumor progression.

Loss of costimulatory molecules [19, 43, 44] or defective function of TCR signaling molecules has been reported in melanoma patients [40, 148]. The induction of several costimulatory molecules has been also attempted in order to increase tumor immunity against melanoma cells. Transfection of melanoma cells with CD80/CD86 or CD48 molecules increases tumor recognition by CTLs. It is interesting to consider that NKRs can also inhibit CD28/CD80 and CD244/CD48 interactions [126, 141].

Studies on new antigenic determinants capable of generating adequate tumor immunity constitute a priority area in melanoma research. In this context, the expression on melanoma cells of ligands for inhibitory NKRs expressed on effector T cells may represent an escape mechanism used by these tumor cells. In contrast, the expression of ligands for activating or costimulatory receptors may contribute to the efficient recognition and killing of melanoma cells (Fig. 3). Blocking inhibitory receptors a new strategy with therapeutic potential.

Acknowledgements This work was supported by grants QLRT-2001-00668 (Outcome and Impact of Specific Treatment in European Research on Melanoma, OISTER) and QLK6-CT2002-02283 (T cells in Ageing, T-CIA) from the 5th Framework Program of the European Union, FIS01/0478, FIS03/1383 (to R.S.); FIS00/0853 (to R.T.) from the Ministry of Health; SAF2003-05184 (to R.T.) from the Ministry of Science and Technology; and 03/2 (to R.T.) from the "Consejería de Sanidad y Consumo" Junta de Extremadura (Spain).

References

- Anegon I, Cuturi MC, Trinchieri G, Perussia B (1988) Interaction of Fc receptor (CD16) ligands induces transcription of interleukin 2 receptor (CD25) and lymphokine genes and expression of their products in human natural killer cells. J Exp Med 167:452
- 2. Anichini A, Molla A, Mortarini R, Tragni G, Bersani I, Di Nicola M, Gianni AM, Pilotti S, Dunbar R, Cerundolo V, Parmiani G (1999) An expanded peripheral T cell population to a cytotoxic T lymphocyte (CTL)-defined, melanocyte-specific antigen in metastatic melanoma patients impacts on generation of peptide-specific CTLs but does not overcome tumor escape from immune surveillance in metastatic lesions. J Exp Med 190:651
- Anichini A, Scarito A, Molla A, Parmiani G, Mortarini R (2003) Differentiation of CD8+ T cells from tumor-invaded and tumor-free lymph nodes of melanoma patients: role of common gamma-chain cytokines. J Immunol 171:2134
- 4. Arase N, Arase H, Park SY, Ohno H, Ra C, Saito T (1997) Association with FcRgamma is essential for activation signal through NKR-P1 (CD161) in natural killer (NK) cells and NK1.1+ T cells. J Exp Med 186:1957
- Azzoni L, Zatsepina O, Abebe B, Bennett IM, Kanakaraj P, Perussia B (1998) Differential transcriptional regulation of CD161 and a novel gene, 197/15a, by IL-2, IL-15, and IL-12 in NK and T cells. J Immunol 161:3493
- Bakker AB, Phillips JH, Figdor CG, Lanier LL (1998) Killer cell inhibitory receptors for MHC class I molecules regulate lysis of melanoma cells mediated by NK cells, gamma delta T cells, and antigen-specific CTL. J Immunol 160:5239
- Baron V, Bouneaud C, Cumano A, Lim A, Arstila TP, Kourilsky P, Ferradini L, Pannetier C (2003) The repertoires of circulating human CD8(+) central and effector memory T cell subsets are largely distinct. Immunity 18:193
- Barzegar C, Meazza R, Pereno R, Pottin-Clemenceau C, Scudeletti M, Brouty-Boye D, Doucet C, Taoufik Y, Ritz J, Musselli C, Mishal Z, Jasmin C, Indiveri F, Ferrini S, Azzarone B (1998) IL-15 is produced by a subset of human melanomas, and is involved in the regulation of markers of melanoma progression through juxtacrine loops. Oncogene 16:2503
- Becker JC, Brocker EB (1995) Lymphocyte-melanoma interaction: role of surface molecules. Recent Results Cancer Res 139:205
- Becker JC, Brabletz T, Czerny C, Termeer C, Brocker EB (1993) Tumor escape mechanisms from immunosurveillance: induction of unresponsiveness in a specific MHC-restricted CD4+ human T cell clone by the autologous MHC class II+ melanoma. Int Immunol 5:1501
- Bernsen MR, Hakansson L, Gustafsson B, Krysander L, Rettrup B, Ruiter D, Hakansson A (2003) On the biological relevance of MHC class II and B7 expression by tumour cells in melanoma metastases. Br J Cancer 88:424
- Bertone S, Schiavetti F, Bellomo R, Vitale C, Ponte M, Moretta L, Mingari MC (1999) Transforming growth factorbeta-induced expression of CD94/NKG2A inhibitory receptors in human T lymphocytes. Eur J Immunol 29:23
- Borrego F, Kabat J, Kim DK, Lieto L, Maasho K, Pena J, Solana R, Coligan JE (2002) Structure and function of major histocompatibility complex (MHC) class I specific receptors expressed on human natural killer (NK) cells. Mol Immunol 38:637
- Brady MS, Lee F, Eckels DD, Ree SY, Latouche JB, Lee JS (2000) Restoration of alloreactivity of melanoma by transduction with B7.1. J Immunother 23:353
- 15. Brenchley JM, Karandikar NJ, Betts MR, Ambrozak DR, Hill BJ, Crotty LE, Casazza JP, Kuruppu J, Migueles SA, Connors M, Roederer M, Douek DC, Koup RA (2003) Expression of CD57 defines replicative senescence and antigen-induced apoptotic death of CD8+ T cells. Blood 101:2711

- Brown MH, Boles K, van der Merwe PA, Kumar V, Mathew PA, Barclay AN (1998) 2B4, the natural killer and T cell immunoglobulin superfamily surface protein, is a ligand for CD48. J Exp Med 188:2083
- Bullock TN, Mullins DW, Colella TA, Engelhard VH (2001) Manipulation of avidity to improve effectiveness of adoptively transferred CD8(+) T cells for melanoma immunotherapy in human MHC class I-transgenic mice. J Immunol 167:5824
- Carreno BM, Collins M (2002) The B7 family of ligands and its receptors: new pathways for costimulation and inhibition of immune responses. Annu Rev Immunol 20:29
- Casado JG, DelaRosa O, Peralbo E, Tarazona R, Solana R (2003) CD28 downregulation and expression of NK associated receptors on T cells in aging and situations of chronic activation of the immune system. In: Pawelec G (ed) Basic biology and clinical impact of immunosenescence. Elsevier, Amsterdam, p 123
- Chambers CA, Kuhns MS, Egen JG, Allison JP (2001) CTLA-4-mediated inhibition in regulation of T cell responses: mechanisms and manipulation in tumor immunotherapy. Annu Rev Immunol 19:565
- Chuang SS, Kim MH, Johnson LA, Albertsson P, Kitson RP, Nannmark U, Goldfarb RH, Mathew PA (2000) 2B4 stimulation of YT cells induces natural killer cell cytolytic function and invasiveness. Immunology 100:378
- 22. Clynes R, Takechi Y, Moroi Y, Houghton A, Ravetch JV (1998) Fc receptors are required in passive and active immunity to melanoma. Proc Natl Acad Sci U S A 95:652
- Cookson S, Reen D (2003) IL-15 drives neonatal T cells to acquire CD56 and become activated effector cells. Blood 102:2195
- 24. Cooper MA, Fehniger TA, Caligiuri MA (2001) The biology of human natural killer-cell subsets. Trends Immunol 22:633
- 25. Cosman D, Mullberg J, Sutherland CL, Chin W, Armitage R, Fanslow W, Kubin M, Chalupny NJ (2001) ULBPs, novel MHC class I-related molecules, bind to CMV glycoprotein UL16 and stimulate NK cytotoxicity through the NKG2D receptor. Immunity 14:123
- 26. Cui J, Shin T, Kawano T, Sato H, Kondo E, Toura I, Kaneko Y, Koseki H, Kanno M, Taniguchi M (1997) Requirement for Valpha14 NKT cells in IL-12-mediated rejection of tumors. Science 278:1623
- D'Andrea A, Chang C, Phillips JH, Lanier LL (1996) Regulation of T cell lymphokine production by killer cell inhibitory receptor recognition of self HLA class I alleles. J Exp Med 184:789
- De Maria A, Moretta L (2000) HLA-class I-specific inhibitory receptors in HIV-1 infection. Hum Immunol 61:74
- DelaRosa O, Tarazona R, Casado JG, Alonso C, Ostos B, Pena J, Solana R (2002) Valpha24+ NKT cells are decreased in elderly humans. Exp Gerontol 37:213
- 30. Denfeld RW, Dietrich A, Wuttig C, Tanczos E, Weiss JM, Vanscheidt W, Schopf E, Simon JC (1995) In situ expression of B7 and CD28 receptor families in human malignant melanoma: relevance for T-cell-mediated anti-tumor immunity. Int J Cancer 62:259
- Derre L, Corvaisier M, Pandolfino MC, Diez E, Jotereau F, Gervois N (2002) Expression of CD94/NKG2-A on human T lymphocytes is induced by IL-12: implications for adoptive immunotherapy. J Immunol 168:4864
- 32. Dong H, Strome SE, Salomao DR, Tamura H, Hirano F, Flies DB, Roche PC, Lu J, Zhu G, Tamada K, Lennon VA, Celis E, Chen L (2002) Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. Nat Med 8:793
- 33. Dunbar PR, Smith CL, Chao D, Salio M, Shepherd D, Mirza F, Lipp M, Lanzavecchia A, Sallusto F, Evans A, Russell-Jones R, Harris AL, Cerundolo V (2000) A shift in the phenotype of melan-A-specific CTL identifies melanoma patients with an active tumor-specific immune response. J Immunol 165:6644

- 34. Fenton RG, Turcovski-Corrales SM, Taub DD (1998) Induction of melanoma antigen-specific cytotoxic T lymphocytes in vitro by stimulation with B7-expressing human melanoma cell lines. J Immunother 21:95
- Ferrone S, Finerty JF, Jaffee EM, Nabel GJ (2000) How much longer will tumour cells fool the immune system? Immunol Today 21:70
- Frauwirth KA, Thompson CB (2002) Activation and inhibition of lymphocytes by costimulation. J Clin Invest 109:295
- Godfrey DI, Hammond KJ, Poulton LD, Smyth MJ, Baxter AG (2000) NKT cells: facts, functions and fallacies. Immunol Today 21:573
- Gritzapis AD, Dimitroulopoulos D, Paraskevas E, Baxevanis CN, Papamichail M (2002) Large-scale expansion of CD3(+)CD56(+) lymphocytes capable of lysing autologous tumor cells with cytokine-rich supernatants. Cancer Immunol Immunother 51:440
- Groh V, Wu J, Yee C, Spies T (2002) Tumour-derived soluble MIC ligands impair expression of NKG2D and T-cell activation. Nature 419:734
- 40. Guarini A, Riera L, Cignetti A, Montacchini L, Massaia M, Foa R (1997) Transfer of the interleukin-2 gene into human cancer cells induces specific antitumor recognition and restores the expression of CD3/T-cell receptor associated signal transduction molecules. Blood 89:212
- Guerra N, Benlhassan K, Carayol G, Guillard M, Pardoux C, Chouaib S, Caignard A (1999) Effect of tumor growth factorbeta on NK receptor expression by allostimulated CD8+ T lymphocytes. Eur Cytokine Netw 10:357
- Gunturi A, Berg RE, Forman J (2003) Preferential survival of CD8 T and NK cells expressing high levels of CD94. J Immunol 170:1737
- 43. Hakansson A, Gustafsson B, Krysander L, Hjelmqvist B, Rettrup B, Hakansson L (1999) On down-regulation of the immune response to metastatic malignant melanoma. Cancer Immunol Immunother 48:253
- 44. Hakansson A, Hakansson L, Gustafsson B, Krysander L, Rettrup B, Ruiter D, Bernsen MR (2002) Biochemotherapy of metastatic malignant melanoma: on down-regulation of CD28. Cancer Immunol Immunother 51:499
- 45. Hammond KJ, Pelikan SB, Crowe NY, Randle-Barrett E, Nakayama T, Taniguchi M, Smyth MJ, van Driel IR, Scollay R, Baxter AG, Godfrey DI (1999) NKT cells are phenotypically and functionally diverse. Eur J Immunol 29:3768
- 46. Heinzel S, Rea D, Offringa R, Pawelec G (2001) The self peptide annexin II (208–223) presented by dendritic cells sensitizes autologous CD4+ T lymphocytes to recognize melanoma cells. Cancer Immunol Immunother 49:671
- Held W, Coudert JD, Zimmer J (2003) The NK cell receptor repertoire: formation, adaptation and exploitation. Curr Opin Immunol 15:233
- Hersey P, Si Z, Smith MJ, Thomas WD (1994) Expression of the co-stimulatory molecule B7 on melanoma cells. Int J Cancer 58:527
- 49. Hom SS, Schwartzentruber DJ, Rosenberg SA, Topalian SL (1993) Specific release of cytokines by lymphocytes infiltrating human melanomas in response to shared melanoma antigens. J Immunother 13:18
- Houghton AN, Gold JS, Blachere NE (2001) Immunity against cancer: lessons learned from melanoma. Curr Opin Immunol 13:134
- 51. Huard B, Karlsson L (2000) A subpopulation of CD8 + T cells specific for melanocyte differentiation antigens expresses killer inhibitory receptors (KIR) in healthy donors: evidence for a role of KIR in the control of peripheral tolerance. Eur J Immunol 30:1665
- 52. Huttenbach Y, Prieto VG, Reed JA (2002) Desmoplastic and spindle cell melanomas express protein markers of the neural crest but not of later committed stages of Schwann cell differentiation. J Cutan Pathol 29:562
- 53. Ikeda H, Lethe B, Lehmann F, van Baren N, Baurain JF, De Smet C, Chambost H, Vitale M, Moretta A, Boon T,

Coulie PG (1997) Characterization of an antigen that is recognized on a melanoma showing partial HLA loss by CTL expressing an NK inhibitory receptor. Immunity 6:199

- 54. Imaizumi K, Kawabe T, Ichiyama S, Kikutani H, Yagita H, Shimokata K, Hasegawa Y (1999) Enhancement of tumoricidal activity of alveolar macrophages via CD40-CD40 ligand interaction. Am J Physiol 277:L49-L57
- 55. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N (2002) Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. Proc Natl Acad Sci U S A 99:12293
- 56. Jager E, Hohn H, Necker A, Forster R, Karbach J, Freitag K, Neukirch C, Castelli C, Salter RD, Knuth A, Maeurer MJ (2002) Peptide-specific CD8+ T-cell evolution in vivo: response to peptide vaccination with Melan-A/MART-1. Int J Cancer 98:376
- 57. Jan CN, Sutherland CL, Lawrence WA, Rein-Weston A, Cosman D (2003) ULBP4 is a novel ligand for human NKG2D. Biochem Biophys Res Commun 305:129
- 58. Jaques AJ, Opdenakker G, Rademacher TW, Dwek RA, Zamze SE (1996) The glycosylation of Bowes melanoma tissue plasminogen activator: lectin mapping, reaction with anti-L2/ HNK-1 antibodies and the presence of sulphated/glucuronic acid containing glycans. Biochem J 316 (Pt 2):427
- Johnson LA, Goldfarb RH, Mathew PA (2000) Regulation of IFN-gamma production following 2B4 activation in human NK cells. In Vivo 14:625
- 60. Jovic V, Konjevic G, Radulovic S, Jelic S, Spuzic I (2001) Impaired perforin-dependent NK cell cytotoxicity and proliferative activity of peripheral blood T cells is associated with metastatic melanoma. Tumori 87:324
- Kambayashi T, Assarsson E, Chambers BJ, Ljunggren HG (2001) Cutting edge: Regulation of CD8(+) T cell proliferation by 2B4/CD48 interactions. J Immunol 167:6706
- 62. Karlhofer FM, Ribaudo RK, Yokoyama WM (1992) MHC class I alloantigen specificity of Ly-49 + IL-2-activated natural killer cells. Nature 358:66
- Karre K, Ljunggren HG, Piontek G, Kiessling R (1986) Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy. Nature 319:675
- 64. Kawano T, Cui J, Koezuka Y, Toura I, Kaneko Y, Sato H, Kondo E, Harada M, Koseki H, Nakayama T, Tanaka Y, Taniguchi M (1998) Natural killer-like nonspecific tumor cell lysis mediated by specific ligand-activated Valpha14 NKT cells. Proc Natl Acad Sci U S A 95:5690
- 65. Kawano T, Nakayama T, Kamada N, Kaneko Y, Harada M, Ogura N, Akutsu Y, Motohashi S, Iizasa T, Endo H, Fujisawa T, Shinkai H, Taniguchi M (1999) Antitumor cytotoxicity mediated by ligand-activated human V alpha24 NKT cells. Cancer Res 59:5102
- 66. Kikuchi A, Nieda M, Schmidt C, Koezuka Y, Ishihara S, Ishikawa Y, Tadokoro K, Durrant S, Boyd A, Juji T, Nicol A (2001) In vitro anti-tumour activity of alpha-galactosylceramide-stimulated human invariant Valpha24+NKT cells against melanoma. Br J Cancer 85:741
- 67. Kubin MZ, Parshley DL, Din W, Waugh JY, Davis-Smith T, Smith CA, Macduff BM, Armitage RJ, Chin W, Cassiano L, Borges L, Petersen M, Trinchieri G, Goodwin RG (1999) Molecular cloning and biological characterization of NK cell activation-inducing ligand, a counterstructure for CD48. Eur J Immunol 29:3466
- Lanier LL, Chang C, Azuma M, Ruitenberg JJ, Hemperly JJ, Phillips JH (1991) Molecular and functional analysis of human natural killer cell-associated neural cell adhesion molecule (N-CAM/CD56). J Immunol 146:4421
- Lanier LL, Corliss BC, Wu J, Leong C, Phillips JH (1998) Immunoreceptor DAP12 bearing a tyrosine-based activation motif is involved in activating NK cells. Nature 391:703
- Lee PP, Yee C, Savage PA, Fong L, Brockstedt D, Weber JS, Johnson D, Swetter S, Thompson J, Greenberg PD, Roederer M, Davis MM (1999) Characterization of circulating T cells

- 71. Letsch A, Keilholz U, Assfalg G, Mailander V, Thiel E, Scheibenbogen C (2003) Bone marrow contains melanomareactive CD8 + effector T cells and, compared with peripheral blood, enriched numbers of melanoma-reactive CD8 + memory T cells. Cancer Res 63:5582
- 72. Li K, Adibzadeh M, Halder T, Kalbacher H, Heinzel S, Muller C, Zeuthen J, Pawelec G (1998) Tumour-specific MHC-class-II-restricted responses after in vitro sensitization to synthetic peptides corresponding to gp100 and Annexin II eluted from melanoma cells. Cancer Immunol Immunother 47:32
- 73. Li Y, Hellstrom KE, Newby SA, Chen L (1996) Costimulation by CD48 and B7-1 induces immunity against poorly immunogenic tumors. J Exp Med 183:639
- 74. Li Y, McGowan P, Hellstrom I, Hellstrom KE, Chen L (1994) Costimulation of tumor-reactive CD4+ and CD8+ T lymphocytes by B7, a natural ligand for CD28, can be used to treat established mouse melanoma. J Immunol 153:421
- 75. Liu X, Bai XF, Wen J, Gao JX, Liu J, Lu P, Wang Y, Zheng P, Liu Y (2001) B7H costimulates clonal expansion of, and cognate destruction of tumor cells by, CD8(+) T lymphocytes in vivo. J Exp Med 194:1339
- 76. Maio M, Fonsatti E, Lamaj E, Altomonte M, Cattarossi I, Santantonio C, Melani C, Belli F, Arienti F, Colombo MP, Parmiani G (2002) Vaccination of stage IV patients with allogeneic IL-4- or IL-2-gene-transduced melanoma cells generates functional antibodies against vaccinating and autologous melanoma cells. Cancer Immunol Immunother 51:9
- 77. Marin R, Ruiz-Cabello F, Pedrinaci S, Mendez R, Jimenez P, Geraghty DE, Garrido F (2003) Analysis of HLA-E expression in human tumors. Immunogenetics 54:767
- Marincola FM, Wang E, Herlyn M, Seliger B, Ferrone S (2003) Tumors as elusive targets of T-cell-based active immunotherapy. Trends Immunol 24:335
- 79. Mazzocchi A, Melani C, Rivoltini L, Castelli C, Del Vecchio M, Lombardo C, Colombo MP, Parmiani G (2001) Simultaneous transduction of B7-1 and IL-2 genes into human melanoma cells to be used as vaccine: enhancement of stimulatory activity for autologous and allogeneic lymphocytes. Cancer Immunol Immunother 50:199
- 80. McCarthy DO, Glowacki N, Schell K, Emler CA, Albertini MR (2000) Antigenicity of human melanoma cells transfected to express the B7-1 co-stimulatory molecule (CD80) varies with the level of B7-1 expression. Cancer Immunol Immunother 49:85
- McQueen KLParham P (2002) Variable receptors controlling activation and inhibition of NK cells. Curr Opin Immunol 14:615
- Menier C, Riteau B, Carosella ED, Rouas-Freiss N (2002) MICA triggering signal for NK cell tumor lysis is counteracted by HLA-G1-mediated inhibitory signal. Int J Cancer 100:63
- Merino J, Martinez-Gonzalez MA, Rubio M, Inoges S, Sanchez-Ibarrola A, Subira ML (1998) Progressive decrease of CD8high+ CD28+ CD57- cells with ageing. Clin Exp Immunol 112:48
- 84. Mingari MC, Schiavetti F, Ponte M, Vitale C, Maggi E, Romagnani S, Demarest J, Pantaleo G, Fauci AS, Moretta L (1996) Human CD8 + T lymphocyte subsets that express HLA class I-specific inhibitory receptors represent oligoclonally or monoclonally expanded cell populations. Proc Natl Acad Sci U S A 93:12433
- 85. Mingari MC, Ponte M, Bertone S, Schiavetti F, Vitale C, Bellomo R, Moretta A, Moretta L (1998) HLA class I-specific inhibitory receptors in human T lymphocytes: interleukin 15induced expression of CD94/NKG2A in superantigen- or alloantigen-activated CD8+ T cells. Proc Natl Acad Sci U S A 95:1172
- Mingari MC, Ponte M, Vitale C, Bellomo R, Moretta L (2000) Expression of HLA class I-specific inhibitory receptors

in human cytolytic T lymphocytes: a regulated mechanism that controls T-cell activation and function. Hum Immunol 61:44

- 87. Moretta A, Moretta L (1997) HLA class I specific inhibitory receptors. Curr Opin Immunol 9:694
- 88. Moretta A, Vitale M, Bottino C, Orengo AM, Morelli L, Augugliaro R, Barbaresi M, Ciccone E, Moretta L (1993) P58 molecules as putative receptors for major histocompatibility complex (MHC) class I molecules in human natural killer (NK) cells. Anti-p58 antibodies reconstitute lysis of MHC class I-protected cells in NK clones displaying different specificities. J Exp Med 178:597
- Moretta A, Sivori S, Vitale M, Pende D, Morelli L, Augugliaro R, Bottino C, Moretta L (1995) Existence of both inhibitory (p58) and activatory (p50) receptors for HLA-C molecules in human natural killer cells. J Exp Med 182:875
- 90. Mortarini R, Piris A, Maurichi A, Molla A, Bersani I, Bono A, Bartoli C, Santinami M, Lombardo C, Ravagnani F, Cascinelli N, Parmiani G, Anichini A (2003) Lack of terminally differentiated tumor-specific CD8+ T cells at tumor site in spite of antitumor immunity to self-antigens in human metastatic melanoma. Cancer Res 63:2535
- Nakajima H, Cella M, Langen H, Friedlein A, Colonna M (1999) Activating interactions in human NK cell recognition: the role of 2B4-CD48. Eur J Immunol 29:1676
- 92. Namekawa T, Snyder MR, Yen JH, Goehring BE, Leibson PJ, Weyand CM, Goronzy JJ (2000) Killer cell activating receptors function as costimulatory molecules on CD4+CD28null T cells clonally expanded in rheumatoid arthritis. J Immunol 165:1138
- Nitta T, Yagita H, Sato K, Okumura K (1989) Involvement of CD56 (NKH-1/Leu-19 antigen) as an adhesion molecule in natural killer-target cell interaction. J Exp Med 170:1757
- Palmowski M, Salio M, Dunbar RP, Cerundolo V (2002) The use of HLA class I tetramers to design a vaccination strategy for melanoma patients. Immunol Rev 188:155
- Park SH, Kyin T, Bendelac A, Carnaud C (2003) The contribution of NKT cells, NK cells, and other gamma-chaindependent non-T non-B cells to IL-12-mediated rejection of tumors. J Immunol 170:1197
- 96. Parrish-Novak J, Dillon SR, Nelson A, Hammond A, Sprecher C, Gross JA, Johnston J, Madden K, Xu W, West J, Schrader S, Burkhead S, Heipel M, Brandt C, Kuijper JL, Kramer J, Conklin D, Presnell SR, Berry J, Shiota F, Bort S, Hambly K, Mudri S, Clegg C, Moore M, Grant FJ, Lofton-Day C, Gilbert T, Rayond F, Ching A, Yao L, Smith D, Webster P, Whitmore T, Maurer M, Kaushansky K, Holly RD, Foster D (2000) Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. Nature 408:57
- 97. Paschen A, Mendez RM, Jimenez P, Sucker A, Ruiz-Cabello F, Song M, Garrido F, Schadendorf D (2003) Complete loss of HLA class I antigen expression on melanoma cells: a result of successive mutational events. Int J Cancer 103:759
- 98. Pedersen LO, Vetter CS, Mingari MC, Andersen MH, thor SP, Brocker EB, Becker JC (2002) Differential expression of inhibitory or activating CD94/NKG2 subtypes on MART-1reactive T cells in vitiligo versus melanoma: a case report. J Invest Dermatol 118:595
- 99. Pende D, Cantoni C, Rivera P, Vitale M, Castriconi R, Marcenaro S, Nanni M, Biassoni R, Bottino C, Moretta A, Moretta L (2001) Role of NKG2D in tumor cell lysis mediated by human NK cells: cooperation with natural cytotoxicity receptors and capability of recognizing tumors of nonepithelial origin. Eur J Immunol 31:1076
- 100. Peritt D, Sesok-Pizzini DA, Schretzenmair R, Macgregor RR, Valiante NM, Tu X, Trinchieri G, Kamoun M (1999) C1.7 antigen expression on CD8 + T cells is activation dependent: increased proportion of C1.7+CD8+ T cells in HIV-1-infected patients with progressing disease. J Immunol 162:7563
- Pittet MJ, Speiser DE, Valmori D, Cerottini JC, Romero P (2000) Cutting edge: cytolytic effector function in human cir-

culating CD8+ T cells closely correlates with CD56 surface expression. J Immunol 164:1148

- 102. Ramirez-Montagut T, Turk MJ, Wolchok JD, Guevara-Patino JA, Houghton AN (2003) Immunity to melanoma: unraveling the relation of tumor immunity and autoimmunity. Oncogene 22:3180
- 103. Raulet DH (2003) Roles of the NKG2D immunoreceptor and its ligands. Nat Rev Immunol 3:781
- 104. Real LM, Jimenez P, Kirkin A, Serrano A, Garcia A, Canton J, Zeuthen J, Garrido F, Ruiz-Cabello F (2001) Multiple mechanisms of immune evasion can coexist in melanoma tumor cell lines derived from the same patient. Cancer Immunol Immunother 49:621
- 105. Reed JA, Finnerty B, Albino AP (1999) Divergent cellular differentiation pathways during the invasive stage of cutaneous malignant melanoma progression. Am J Pathol 155:549
- 106. Romero P, Ortega C, Palma A, Molina IJ, Pena J, Santamaria M (2001) Expression of CD94 and NKG2 molecules on human CD4(+) T cells in response to CD3-mediated stimulation. J Leukoc Biol 70:219
- 107. Rowbottom AW, Garland RJ, Lepper MW, Kaneria SS, Goulden NJ, Oakhill A, Steward CG (2000) Functional analysis of the CD8+CD57+ cell population in normal healthy individuals and matched unrelated T-cell-depleted bone marrow transplant recipients. Br J Haematol 110:315
- 108. Sallusto F, Lenig D, Forster R, Lipp M, Lanzavecchia A (1999) Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. Nature 401:708
- 109. Schwartzentruber DJ, Topalian SL, Mancini M, Rosenberg SA (1991) Specific release of granulocyte-macrophage colonystimulating factor, tumor necrosis factor-alpha, and IFNgamma by human tumor-infiltrating lymphocytes after autologous tumor stimulation. J Immunol 146:3674
- 110. Seliger B, Abken H, Ferrone S (2003) HLA-G and MIC expression in tumors and their role in anti-tumor immunity. Trends Immunol 24:82
- 111. Serrano A, Tanzarella S, Lionello I, Mendez R, Traversari C, Ruiz-Cabello F, Garrido F (2001) Rexpression of HLA class I antigens and restoration of antigen-specific CTL response in melanoma cells following 5-aza-2'-deoxycytidine treatment. Int J Cancer 94:243
- 112. Sharpe AHFreeman GJ (2002) The B7-CD28 superfamily. Nature Rev Immunol 2:116
- 113. Smyth MJ, Thia KY, Street SE, Cretney E, Trapani JA, Taniguchi M, Kawano T, Pelikan SB, Crowe NY, Godfrey DI (2000) Differential tumor surveillance by natural killer (NK) and NKT cells. J Exp Med 191:661
- 114. Snyder MR, Lucas M, Vivier E, Weyand CM, Goronzy JJ (2003) Selective activation of the c-Jun NH2-terminal protein kinase signaling pathway by stimulatory KIR in the absence of KARAP/DAP12 in CD4+ T cells. J Exp Med 197:437
- 115. Speiser DE, Pittet MJ, Valmori D, Dunbar R, Rimoldi D, Lienard D, MacDonald HR, Cerottini JC, Cerundolo V, Romero P (1999) In vivo expression of natural killer cell inhibitory receptors by human melanoma-specific cytolytic T lymphocytes. J Exp Med 190:775
- 116. Speiser DE, Valmori D, Rimoldi D, Pittet MJ, Lienard D, Cerundolo V, MacDonald HR, Cerottini JC, Romero P (1999) CD28-negative cytolytic effector T cells frequently express NK receptors and are present at variable proportions in circulating lymphocytes from healthy donors and melanoma patients. Eur J Immunol 29:1990
- 117. Špeiser DE, Colonna M, Ayyoub M, Cella M, Pittet MJ, Batard P, Valmori D, Guillaume P, Lienard D, Cerottini JC, Romero P (2001) The activatory receptor 2B4 is expressed in vivo by human CD8+ effector alpha beta T cells. J Immunol 167:6165
- 118. Speiser DE, Migliaccio M, Pittet MJ, Valmori D, Lienard D, Lejeune F, Reichenbach P, Guillaume P, Luscher I, Cerottini JC, Romero P (2001) Human CD8(+) T cells expressing HLA-DR and CD28 show telomerase activity and are distinct from cytolytic effector T cells. Eur J Immunol 31:459

- 119. Sutherland CL, Chalupny NJ, Schooley K, VandenBos T, Kubin M, Cosman D (2002) UL16-binding proteins, novel MHC class I-related proteins, bind to NKG2D and activate multiple signaling pathways in primary NK cells. J Immunol 168:671
- 120. Sze DM, Giesajtis G, Brown RD, Raitakari M, Gibson J, Ho J, Baxter AG, Fazekas dSG, Basten A, Joshua DE (2001) Clonal cytotoxic T cells are expanded in myeloma and reside in the CD8(+)CD57(+)CD28(-) compartment. Blood 98:2817
- 121. Takahashi T, Nieda M, Koezuka Y, Nicol A, Porcelli SA, Ishikawa Y, Tadokoro K, Hirai H, Juji T (2000) Analysis of human V alpha 24+ CD4+ NKT cells activated by alphaglycosylceramide-pulsed monocyte-derived dendritic cells. J Immunol 164:4458
- 122. Takasaki S, Hayashida K, Morita C, Ishibashi H, Niho Y (2000) CD56 directly interacts in the process of NCAM-positive target cell-killing by NK cells. Cell Biol Int 24:101
- 123. Tang NE, Luyten GP, Mooy CM, Naus NC, de Jong PT, Luider TM (1996) HNK-1 antigens on uveal and cutaneous melanoma cell lines. Melanoma Res 6:411
- 124. Tarazona R, DelaRosa O, Alonso C, Ostos B, Espejo J, Pena J, Solana R (2000) Increased expression of NK cell markers on T lymphocytes in aging and chronic activation of the immune system reflects the accumulation of effector/senescent T cells. Mech Ageing Dev 121:77
- 125. Tarazona R, DelaRosa O, Casado JG, Torre-Cisneros J, Villanueva JL, Galiani MD, Pena J, Solana R (2002) NK-associated receptors on CD8 T cells from treatment-naive HIV-infected individuals: defective expression of CD56. AIDS 16:197
- 126. Tarazona R, Borrego F, Galiani MD, Aguado E, Pena J, Coligan JE, Solana R (2002) Inhibition of CD28-mediated natural cytotoxicity by KIR2DL2 does not require p56(lck) in the NK cell line YT-Indy. Mol Immunol 38:495
- 127. Tarazona R, Casado JG, DelaRosa O, Torre-Cisneros J, Villanueva JL, Sanchez B, Galiani MD, Gonzalez R, Solana R, Pena J (2002) Selective depletion of CD56(dim) NK cell subsets and maintenance of CD56(bright) NK cells in treatment-naïve HIV-1-seropositive individuals. J Clin Immunol 22:176
- 128. Topalian SL, Rivoltini L, Mancini M, Markus NR, Robbins PF, Kawakami Y, Rosenberg SA (1994) Human CD4+ T cells specifically recognize a shared melanoma-associated antigen encoded by the tyrosinase gene. Proc Natl Acad Sci U S A 91:9461
- 129. Topalian SL, Gonzales MI, Parkhurst M, Li YF, Southwood S, Sette A, Rosenberg SA, Robbins PF (1996) Melanomaspecific CD4+ T cells recognize nonmutated HLA-DR-restricted tyrosinase epitopes. J Exp Med 183:1965
- Tough DF (2003) Deciphering the relationship between central and effector memory CD8+ T cells. Trends Immunol 24:404
- 131. Touloukian CE, Leitner WW, Topalian SL, Li YF, Robbins PF, Rosenberg SA, Restifo NP (2000) Identification of a MHC class II-restricted human gp100 epitope using DR4-IE transgenic mice. J Immunol 164:3535
- 132. Toura I, Kawano T, Akutsu Y, Nakayama T, Ochiai T, Taniguchi M (1999) Cutting edge: inhibition of experimental tumor metastasis by dendritic cells pulsed with alpha-galactosylceramide. J Immunol 163:2387
- 133. Ugolini S, Arpin C, Anfossi N, Walzer T, Cambiaggi A, Forster R, Lipp M, Toes RE, Melief CJ, Marvel J, Vivier E (2001) Involvement of inhibitory NKRs in the survival of a subset of memory-phenotype CD8 + T cells. Nat Immunol 2:430
- 134. Valmori D, Scheibenbogen C, Dutoit V, Nagorsen D, Asemissen AM, Rubio-Godoy V, Rimoldi D, Guillaume P, Romero P, Schadendorf D, Lipp M, Dietrich PY, Thiel E, Cerottini JC, Lienard D, Keilholz U (2002) Circulating tumor-reactive CD8(+) T cells in melanoma patients contain a CD45RA(+)CCR7(-) effector subset exerting ex vivo tumor-specific cytolytic activity. Cancer Res 62:1743

- 135. Van den Hove LE, Van Gool SW, Vandenberghe P, Boogaerts MA, Ceuppens JL (1998) CD57+/CD28- T cells in untreated hemato-oncological patients are expanded and display a Th1-type cytokine secretion profile, ex vivo cytolytic activity and enhanced tendency to apoptosis. Leukemia 12:1573
- 136. Vesosky B, Hurwitz AA (2003) Modulation of costimulation to enhance tumor immunity. Cancer Immunol Immunother 52:663
- 137. Vetter CS, Straten PT, Terheyden P, Zeuthen J, Brocker EB, Becker JC (2000) Expression of CD94/NKG2 subtypes on tumor-infiltrating lymphocytes in primary and metastatic melanoma. J Invest Dermatol 114:941
- 138. Vetter CS, Groh V, thor SP, Spies T, Brocker EB, Becker JC (2002) Expression of stress-induced MHC class I related chain molecules on human melanoma. J Invest Dermatol 118:600
- 139. Wallin JJ, Liang L, Bakardjiev A, Sha WC (2001) Enhancement of CD8 + T cell responses by ICOS/B7 h costimulation. J Immunol 167:132
- 140. Warren HS, Kinnear BF (1999) Quantitative analysis of the effect of CD16 ligation on human NK cell proliferation. J Immunol 162:735
- 141. Watzl C, Stebbins CC, Long EO (2000) NK cell inhibitory receptors prevent tyrosine phosphorylation of the activation receptor 2B4 (CD244). J Immunol 165:3545

- 142. Weekes MP, Wills MR, Mynard K, Hicks R, Sissons JG, Carmichael AJ (1999) Large clonal expansions of human virus-specific memory cytotoxic T lymphocytes within the CD57+ CD28- CD8+ T-cell population. Immunology 98:443
- 143. Wherry EJ, Teichgraber V, Becker TC, Masopust D, Kaech SM, Antia R, von Andrian UH, Ahmed R (2003) Lineage relationship and protective immunity of memory CD8 T cell subsets. Nat Immunol 4:225
- 144. Wu DY, Segal NH, Sidobre S, Kronenberg M, Chapman PB (2003) Cross-presentation of disialoganglioside GD3 to natural killer T cells. J Exp Med 198(1):173–181
- 145. Wu J, Lanier LL (2003) Natural killer cells and cancer. Adv Cancer Res 90:127
- 146. Yokoyama WM, Plougastel BF (2003) Immune functions encoded by the natural killer gene complex. Nat Rev Immunol 3:304
- 147. Young NT, Uhrberg M, Phillips JH, Lanier LL, Parham P (2001) Differential expression of leukocyte receptor complexencoded Ig-like receptors correlates with the transition from effector to memory CTL. J Immunol 166:3933
- 148. Zea AH, Curti BD, Longo DL, Alvord WG, Strobl SL, Mizoguchi H, Creekmore SP, O'Shea JJ, Powers GC, Urba WJ (1995) Alterations in T cell receptor and signal transduction molecules in melanoma patients. Clin Cancer Res 1:1327