

## Protective Activity of Thymosin Against Opportunistic Infections in Animal Models

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**Summary.** *Animal models for opportunistic infections were developed by using mice immunosuppressed by 5-FU. These mice were susceptible to various microorganisms, while normal mice had greater tolerance to such microbial infections. In these models, thymosin  $\alpha_1$  was found to protect mice against lethal infections with *Candida albicans*, *Listeria monocytogenes*, *Pseudomonas aeruginosa*, and *Serratia marcescens* when it was administered during 5-FU treatment prior to the infections. Thymosin  $\alpha_1$  was effective in some infections at 0.4–400  $\mu\text{g}/\text{kg}/\text{day}$  IP, about  $1/100$  of the dose required for thymosin fraction 5. Activity was also demonstrated against *L. monocytogenes* and *Ps. aeruginosa* by counting the viable bacteria in the liver after infection. The protective activity against *Candida*, elimination of which macrophages were essential, was abrogated by anti-thymocyte serum and/or carrageenan, indicating that thymosin  $\alpha_1$  serves to maintain the functions of macrophages by reducing the damage to T cells by 5-FU. On the other hand, the activity against *Pseudomonas* infection was not affected by anti-thymocyte serum or carrageenan. It is probable that thymosin  $\alpha_1$  also exerts its effect on neutrophils without participation of T cells and macrophages.*

### Introduction

The thymus gland has been shown to play an important role in the development, growth, and function of lymphoid systems through a hormonal mechanisms. One of the thymic hormones, thymosin fraction 5 [10], stimulates T cell development and corrects some immunodeficiencies resulting from the lack of thymus functions [14]. Considering the importance of T cells in immunoregulatory systems, thymosin may be expected to be useful as a pharmaceutical agent for immunodeficiency diseases that are caused by or accompany the aberration of these systems. In fact, thymosin fraction 5, a partially purified material with some 40 peptides, has been shown to be effective in some primary immunodeficiency diseases [19].

It is well known that patients in the secondary immunodeficient state, either due to disease itself or as the consequence of therapies they have received, are highly susceptible to microbial infections, with serious consequences. In cancer patients, chemotherapy and radiotherapy generally decrease immunocompetence. Such defects make the patients readily susceptible to infection with common pathogens (opportunistic

infection) and probably also contribute to the growth of metastatic tumors. Specifically, infections are the major cause of mortality and morbidity in cancer patients [2, 11, 16]. Therefore, it is of interest to study whether fractions 5 and its component peptide  $\alpha_1$  [6] are useful to remedy such defects.

For this reason, we have explored and established animal models for opportunistic infections to assess the activity of fraction 5 and thymosin  $\alpha_1$  given with immunosuppressive cytostatics prior to challenge with opportunistic pathogens. Thymosin fraction 5 and thymosin  $\alpha_1$  were found to protect mice against lethal infections with four different types of microorganisms in such models when the same cytostatic, 5-fluorouracil, was used. Furthermore, we have provided some insight into the mechanisms of action of thymosin  $\alpha_1$  against infection with two different types of pathogens in immunosuppressed mice using anti-thymocyte serum [12] and carrageenan [3], which depress T lymphocytes and macrophages, respectively. Two of the pathogens used were intracellular parasites, *Candida albicans* and *Listeria monocytogenes*, which are eliminated from the infected host mainly by macrophages and T cells [4]. The other is an extracellular parasite, *Pseudomonas aeruginosa*, which is eliminated mainly by neutrophils [17]. These studies suggest that thymosin  $\alpha_1$  might affect not only T cells but also other cell populations, such as neutrophils, whose functions are damaged by 5-FU.

In this report we describe the details of thymosin  $\alpha_1$  activity against opportunistic infections and its probable mode of action. A part of the study has been presented orally elsewhere [18].

### Materials and Methods

**Animals.** Female ddY, ICR, and C57BL/6 mice (6 weeks old) used in these studies were obtained from Shizuoka Agricultural Cooperative Association for Laboratory Animals, Hamamatsu, Japan.

**Microorganisms.** *Candida albicans* ATCC 10231 was purchased from ATCC and *Listeria monocytogenes* EGD was kindly provided by Prof. K. Nomoto, Kyushu University. *Pseudomonas aeruginosa* 5E81-1 (gentamicin-resistant) and *Serratia marcescens* 5A412-1 were clinical isolates. Culture media were nutrient broth (Difco) supplemented with 0.5% glucose for *Candida*, trypticase soy broth (Difco) for *Listeria*, and heart infusion broth (Difco) for *Pseudomonas* and *Serratia*. The inocula were prepared from overnight cultures

and dilutes in saline. Mice were infected with 0.2 ml of the microorganism in each case, by the IV route and at the doses indicated in Tables 1–8 and Fig. 1.

**Anti-Thymocyte Serum (ATS).** Anti-thymocyte serum was prepared by the method described by Levey and Medawar [12]. Three rabbits each received 1 ml IV of a suspension of thymocytes ( $10^9$  cells) from C57BL/6 mice, and booster injections of the same doses of thymocytes on two occasions at intervals of 2 weeks. Serum was obtained from the rabbits at 1 week after the last treatment. The sera (50 ml) were treated at 56° C for 30 min, and absorbed at 4° C for 60 min with liver sediments prepared by homogenization of 50 g liver from C57BL/6 mice and then washing 10 times with phosphate-buffered saline. The neutralization titer of ATS thus obtained was determined to be 1:320 ~ 1:640 by cytotoxicity against thymocytes from C57BL/6 mice with guinea pig complement. Inhibition of the mitogenic response of spleen cells by injection of the ATS was determined by measuring incorporation of  $^3\text{H}$ -thymidine [7].

**Treatment with 5-Fluorouracil and Thymosin.** Strain ddY mice were pretreated daily for 8 or 10 days with 0.2 ml each of thymosin  $\alpha_1$  (0.4 ~ 400  $\mu\text{g}/\text{kg}/\text{day}$ ) or thymosin fraction 5 (4 mg/kg/day) and 5-FU (25 mg/kg/day) in saline by the IP route. At 24 h after the last treatment the mice were infected with microorganisms. Thymosin fraction 5 and  $\alpha_1$  peptide were dissolved in saline, divided into one-dose portions, and kept at -20° C. They were used immediately after thawing.

**In vivo Growth.** Each group of three ddY mice which had been treated daily for 7 days with saline, 5-FU (25 mg/kg/day), or thymosin  $\alpha_1$  (40  $\mu\text{g}/\text{kg}/\text{day}$ ) by the IP route was infected by *L. monocytogenes* or *Ps. aeruginosa* at 24 h after the last treatment. The mice were then sacrificed by excessive bleeding after mild anesthesia. The liver was aseptically removed, rinsed in saline, and homogenized. The homogenate, diluted with saline, was spread on heart infusion agar plates. After 24 h incubation, the number of viable bacteria was determined. The data are presented as the mean of the value obtained in three mice.

**Compounds.** Thymosin fraction 5, prepared from calf thymus, and chemically synthesized  $\alpha_1$  peptide were kindly supplied by Drs A. Ramel and J. Meienhofer, Hoffmann-La Roche Inc., Nutley, NJ, USA, respectively.

**Statistical Analysis.** Analyses of mortality differences were compared by the Mann-Whitney *U*-test (Rank sum test). Differences were considered to be significant when probability (*P*) values < 0.05 were obtained.

## Results

### Suppression by Treatment with 5-FU of Host Defense System Against Microbial Infection

Some microbes (bacteria, fungi, and viruses) that are not pathogenic in healthy individuals with normal immunocompetence often cause infections in immunosuppressed individuals, with serious consequences (opportunistic infections). Similar phenomena were demonstrated in mice immunosuppressed by a cytostatic, 5-FU, prior to infection.

When mice were treated daily 8–10 times with 5-FU (25 mg/kg/day) by the IP route, various immune parameters

**Table 1.** LD<sub>50</sub> of various microorganisms in normal and 5-FU-treated mice

| Microorganism                        | LD <sub>50</sub> (No. of microorganisms) <sup>a</sup> |                   |
|--------------------------------------|---|-------------------|
|                                      | Normal mice   | 5-FU-treated mice |
| <i>Listeria monocytogenes</i> EGD    | $2 \times 10^4$                                       | $5 \times 10^2$   |
| <i>Candida albicans</i> ATCC 10231   | $2 \times 10^6$                                       | $1 \times 10^5$   |
| <i>Pseudomonas aeruginosa</i> 5E81-1 | $2 \times 10^7$                                       | $1 \times 10^4$   |
| <i>Serratia marcescens</i> 5A412-1   | $8.6 \times 10^7$                                     | $4 \times 10^4$   |

<sup>a</sup> ddY mice pretreated daily for 8 days with saline or 5-FU (25 mg/kg/day, IP) were infected with various doses of microorganisms by the IV route. LD<sub>50</sub> was determined by the method of Reed and Muench [15]

**Table 2.** Effect of pretreatment with thymosin  $\alpha_1$  on lethal infection with *L. monocytogenes* or *C. albicans* in 5-FU-treated mice

| Pretreatment <sup>a</sup> | Infection <sup>b</sup> at day 0       | Survival/tested at day |       |       | <i>P</i> value <sup>c</sup> ( <i>U</i> -test) |
|---------------------------|---------------------------------------|------------------------|-------|-------|---|
|                           |                                       | 4                      | 8     | 15    |   |
| Thymosin (IP)             | 5-FU (IP)                             |                        |       |       |   |
| <b>Expt 1.</b>            |                                       |                        |       |       |   |
| <i>L. monocytogenes</i>   |                                       |                        |       |       |   |
| Control (saline)          | +                                     | -                      | 10/10 | 10/10 | 10/10   |
|                           | -                                     | +                      | 10/10 | 10/10 | 9/10  |
|                           | +                                     | +                      | 9/10  | 0/10  | 0/10  |
| Thymosin $\alpha_1$       | 4 $\mu\text{g}/\text{kg}/\text{day}$  | +                      | 10/10 | 7/10  | 6/10  |
|                           | 40 $\mu\text{g}/\text{kg}/\text{day}$ | +                      | 10/10 | 6/10  | 4/10  |
| <b>Expt 2.</b>            |                                       |                        |       |       |   |
| <i>C. albicans</i>        |                                       |                        |       |       |   |
| Control (saline)          | +                                     | -                      | 7/7   | 7/7   | 7/7   |
|                           | -                                     | +                      | 7/7   | 7/7   | 7/7   |
|                           | +                                     | +                      | 2/7   | 2/7   | 1/7   |
| Thymosin Fr. 5            | 4 mg/kg/day                           | +                      | 6/7   | 5/7   | 4/7   |
| Thymosin $\alpha_1$       | 4 $\mu\text{g}/\text{kg}/\text{day}$  | +                      | 5/7   | 5/7   | 3/7   |
|                           | 40 $\mu\text{g}/\text{kg}/\text{day}$ | +                      | 7/7   | 6/7   | 5/7   |

<sup>a</sup> Pretreated daily for 8 days (expt 1) or 10 days (expt 2)

<sup>b</sup> Infected with *L. monocytogenes* ( $1 \times 10^3$ , IV, expt 1) or *C. albicans* ( $2 \times 10^5$ , IV, expt 2) at day 0

<sup>c</sup> Compared with the control mice pretreated with 5-FU and then infected

were decreased. In such mice, the infections with microorganisms used were lethal at doses at least 20 times lower than those in normal mice (Table 1). Decrease in the host defense systems was particularly obvious against *Pseudomonas* and *Serratia* infections. The daily dose of 5-FU used in the experiments, 25 mg/kg IP, was found to be approximately in the optimal range for the treatment of animal tumors in our system, as reported by other investigators [9].

#### Protective Activity of Thymosin Against Infection with Intracellular Parasites

Since thymosin  $\alpha_1$  is known to affect the population of T cells, the effect of this agent was first examined against *L. monocytogenes* and *C. albicans*, for which T cells are known to participate in the elimination from infected hosts.

As shown in Table 2 (expt 1), mice immunosuppressed by 5-FU were susceptible to *Listeria*, while normal mice had quite high tolerance. On the other hand, when thymosin  $\alpha_1$  was administered with 5-FU prior to the infection, the mice largely survived. The activity of thymosin  $\alpha_1$  was further demonstrated by measuring the number of the viable bacteria in the liver of mice pretreated with 5-FU after infection at a relatively high dose of bacteria (Table 3). The number of bacteria was higher 5 h after infection in mice pretreated with 5-FU than that in normal mice. On the other hand, pretreatment with thymosin  $\alpha_1$  (40  $\mu\text{g}/\text{kg}/\text{day}$ , IP) together with 5-FU reduced the number to a lower level than that in normal mice. However, when the clearance rate of bacteria from blood was measured during the first 30 min after infection there was no significant difference between these groups (data not shown). The treatment

**Table 3.** Effect of thymosin  $\alpha_1$  on the growth of *L. monocytogenes* in 5-FU treated mice

| Treatment <sup>a</sup>                | No. of bacteria in liver <sup>b</sup> |
|---------------------------------------|---------------------------------------|
| Saline (control)                      | $1.7 \pm 0.1 \times 10^6$             |
| 5-FU                                  | $> 5 \times 10^7$                     |
| 5-FU + thymosin $\alpha_1$            |                                       |
| 4 $\mu\text{g}/\text{kg}/\text{day}$  | $4.8 \pm 0.7 \times 10^5$             |
| 40 $\mu\text{g}/\text{kg}/\text{day}$ | $4.7 \pm 0.4 \times 10^5$             |

<sup>a</sup> Daily treatment (days -7 ~ -1) prior to infection with *L. monocytogenes* ( $5 \times 10^6$ , IV)

<sup>b</sup> The number of the viable bacteria in liver at 5 h after the infection

regimen for thymosin  $\alpha_1$  was investigated as shown in Table 4. Long-term treatment with thymosin  $\alpha_1$  seemed to be required for protective activity against *Listeria*, although a few treatments at high doses before the infection also showed a significant activity.

Thymosin was next examined for its activity against *Candida* infection in the immunosuppressed mice. As shown in Table 2 (expt. 2), thymosin fraction 5 and thymosin  $\alpha_1$  protected mice from the lethal infection to a similar extent at doses of 4 mg/kg/day and 40  $\mu\text{g}/\text{kg}/\text{day}$ , respectively. Although not studied in detail, thymosin  $\alpha_1$  appeared to be about 100 times more active than thymosin fraction 5 on a per weight basis. The activity of thymosin  $\alpha_1$  was also observed in a different strain (ICR) of mice at similar dose ranges (data not shown).

#### Protective Mechanisms against *Candida albicans*

Since macrophages and T cells are known to be essential for elimination of *C. albicans* [4], treatment with thymosin  $\alpha_1$  appears to reduce the damage caused to the populations and/or functions of such cells by 5-FU. In an attempt to examine the above possibility, the effect of thymosin  $\alpha_1$  was studied in C57BL/6 mice treated with ATS. As shown in Table 5 (expt. 1), treatment with ATS during the thymosin treatment abolished the protective activity against *Candida* infection. When normal mice (C57BL/6) were infected once IV with 0.2 ml anti-thymocyte serum, the mitogenic response measured by <sup>3</sup>H-thymidine incorporation of spleen cells to Con A (3  $\mu\text{g}/\text{ml}$ ) was reduced to 4.5%, while that to LPS (60  $\mu\text{g}/\text{ml}$ ) was reduced to 54.4% compared with the response of spleen cells from control mice. These data suggest that T cells participate in the protective activity of thymosin  $\alpha_1$ .

Treatment with carrageenan 1 day before the infection increased susceptibility in all groups, including normal mice, and consequently the protective activity of thymosin  $\alpha_1$  was abrogated (Table 5, expt. 2). The presence of functional macrophages is quite important in preventing the *Candida* infection. On the other hand, when carrageenan was given prior to the treatment with thymosin  $\alpha_1$  and/or 5-FU, the protective activity of thymosin  $\alpha_1$  was retained (Table 5, expt. 3). Thymosin  $\alpha_1$  may serve to maintain the function of macrophages, which is damaged by carrageenan and 5-FU, probably through reducing the damage to the T cells by 5-FU.

**Table 4.** Effect of treatment schedule for thymosin  $\alpha_1$  on lethal infection by *Listeria* in mice treated with 5-FU<sup>a</sup>

| Treatment with thymosin $\alpha_1$ ( $\mu\text{g}/\text{kg}/\text{day}$ ) (schedule) | Treatment with 5-FU (day -8 ~ -1) | Survival/tested at day 15 | P value (U-test) |
|--|-----------------------------------|---------------------------|------------------|
| Saline   | -                                 | 10/10                     |                  |
| Saline   | +                                 | 2/10                      |                  |
| 4 $\times$ 1 (day -1)  | +                                 | 2/10                      |                  |
| 40 $\times$ 1  | +                                 | 3/10                      |                  |
| 400 $\times$ 1   | +                                 | 6/10                      | < 0.02           |
| 4 $\times$ 3 (day -3 ~ -1)   | +                                 | 4/10                      |                  |
| 40 $\times$ 3  | +                                 | 6/10                      | < 0.02           |
| 400 $\times$ 3   | +                                 | 7/10                      | < 0.02           |
| 4 $\times$ 5 (day -5 ~ -1)   | +                                 | 6/10                      | < 0.02           |
| 40 $\times$ 5  | +                                 | 9/10                      | < 0.005          |
| 40 $\times$ 8 (day -8 ~ -1)  | +                                 | 9/10                      | < 0.005          |

<sup>a</sup> Mice treated with 5-FU and thymosin  $\alpha_1$  at day indicated were infected with *L. monocytogenes* ( $1 \times 10^3$ , IV) at day 0

**Table 5.** Effect of anti-thymocyte serum or carrageenan on the protective activity of thymosin  $\alpha_1$  against *C. albicans* in 5-FU-treated mice

| Pretreatment <sup>a</sup> |      |                  |  | Infection <sup>d</sup><br>with <i>Candida</i> | Survival/tested at day |     |     | P value ( <i>U</i> -test) <sup>e</sup> |
|---------------------------|------|------------------|--|---|------------------------|-----|-----|--|
|                           |      |                  |  |   | 4                      | 8   | 15  |  |
| <i>Expt 1</i>             |      |                  |  |   |                        |     |     |  |
| Saline                    |      |                  |  | +   | 7/7                    | 7/7 | 7/7 |  |
| Saline                    | 5-FU |                  |  | -   | 7/7                    | 7/7 | 7/7 |  |
| Saline                    | 5-FU |                  |  | +   | 2/7                    | 1/7 | 1/7 |  |
| Saline                    | 5-FU | ATS <sup>b</sup> |  | +   | 4/7                    | 2/7 | 1/7 |  |
| Thymosin $\alpha_1$       |      |                  |  | +   | 6/7                    | 6/7 | 6/7 | < 0.001                                |
| Thymosin $\alpha_1$       | 5-FU | ATS              |  | +   | 4/7                    | 3/7 | 3/7 | NS                                     |
| <i>Expt 2</i>             |      |                  |  |   |                        |     |     |  |
| Saline                    |      |                  |  | +   | 8/8                    | 8/8 | 8/8 |  |
| Saline                    | 5-FU |                  |  | -   | 8/8                    | 8/8 | 8/8 |  |
| Saline                    | 5-FU |                  |  | +   | 6/7                    | 3/7 | 2/7 |  |
| Saline                    |      | Ca <sup>c</sup>  |  | +   | 3/8                    | 1/8 | 1/8 |  |
| Saline                    | 5-FU | Ca               |  | +   | 3/7                    | 1/7 | 1/7 |  |
| Thymosin $\alpha_1$       | 5-FU |                  |  | +   | 8/8                    | 8/8 | 8/8 | < 0.006                                |
| Thymosin $\alpha_1$       | 5-FU | Ca               |  | +   | 3/8                    | 3/8 | 3/8 | NS                                     |
| <i>Expt 3</i>             |      |                  |  |   |                        |     |     |  |
| Saline                    |      |                  |  | +   | 6/6                    | 6/6 | 6/6 |  |
| Saline                    | 5-FU |                  |  | -   | 7/7                    | 7/7 | 7/7 |  |
| Saline                    | 5-FU |                  |  | +   | 6/7                    | 4/7 | 2/7 |  |
| Saline                    |      | Ca <sup>c</sup>  |  | +   | 7/7                    | 5/7 | 3/7 |  |
| Saline                    | 5-FU | Ca               |  | +   | 4/6                    | 2/6 | 1/6 |  |
| Thymosin $\alpha_1$       | 5-FU |                  |  | +   | 6/6                    | 6/6 | 6/6 | < 0.006                                |
| Thymosin $\alpha_1$       | 5-FU | Ca               |  | +   | 6/6                    | 4/6 | 4/6 | < 0.005                                |

<sup>a</sup> C57BL/6 mice (expt 1) or ddY mice (expts 2 and 3) were pretreated with 5-FU (25 mg/kg) or thymosin  $\alpha_1$  (40  $\mu$ g/kg) daily for 10 days from days -10 to -1

<sup>b</sup> ATS (0.2 ml, IV) was injected at days -11, -8, -5, -3, and -1

<sup>c</sup> Carrageenan (Ca, 200 mg/kg, IP) was injected at day -1 (expt 2) or -11 (expt 3)

<sup>d</sup> Infected with *C. albicans* (expt 1,  $2.5 \times 10^5$ ; expts 2 and 3,  $4 \times 10^5$ ) by the IP route at day 0

<sup>e</sup> Compared with the control mice pretreated with 5-FU, or 5-FU plus Ca or ATS, and then infected  
NS, not significant

**Table 6.** Effect of pretreatment with thymosin  $\alpha_1$  on lethal infection with *Ps. aeruginosa* or *S. marcescens* in 5-FU treated mice

| Pretreated (day -10 ~ -1) <sup>a</sup> |                    |      |  | Infection<br>at day 0 <sup>b</sup> | Survival/tested at day |       |       | P value ( <i>U</i> -test) <sup>c</sup> |
|--|--------------------|------|--|------------------------------------|------------------------|-------|-------|--|
|  |                    |      |  |                                    | 4                      | 8     | 15    |  |
| Thymosin $\alpha_1$                    |                    | 5-FU |  |                                    |                        |       |       |  |
| <i>Expt 1.</i>                         |                    |      |  |                                    |                        |       |       |  |
| <i>Ps. aeruginosa</i>                  |                    |      |  |                                    |                        |       |       |  |
| Control (saline)                       |                    |      |  | -                                  | 10/10                  | 10/10 | 10/10 |  |
|  |                    |      |  | +                                  | 10/10                  | 10/10 | 10/10 |  |
|  |                    |      |  | +                                  | 0/10                   | 0/10  | 0/10  |  |
| Thymosin $\alpha_1$                    | 4 $\mu$ g/kg/day   |      |  | +                                  | 4/10                   | 4/10  | 4/10  | < 0.0001                               |
|  | 40 $\mu$ g/kg/day  |      |  | +                                  | 8/10                   | 8/10  | 8/10  | < 0.0001                               |
| <i>Expt 2.</i>                         |                    |      |  |                                    |                        |       |       |  |
| <i>S. marcescens</i>                   |                    |      |  |                                    |                        |       |       |  |
| Control (saline)                       |                    |      |  | -                                  | 10/10                  | 10/10 | 10/10 |  |
|  |                    |      |  | +                                  | 10/10                  | 10/10 | 10/10 |  |
|  |                    |      |  | +                                  | 1/10                   | 1/10  | 1/10  |  |
| Thymosin $\alpha_1$                    | 0.4 $\mu$ g/kg/day |      |  | +                                  | 6/10                   | 5/10  | 4/10  | < 0.085                                |
|  | 4 $\mu$ g/kg/day   |      |  | +                                  | 6/10                   | 6/10  | 6/10  | < 0.016                                |
|  | 40 $\mu$ g/kg/day  |      |  | +                                  | 7/10                   | 7/10  | 7/10  | < 0.013                                |

<sup>a</sup> Pretreated daily for 10 days by the IP route

<sup>b</sup> Infected with *Ps. aeruginosa* ( $5 \times 10^4$ , IV) or *S. marcescens* ( $5 \times 10^5$ , IV) at day 0

<sup>c</sup> Compared with the control mice pretreated with 5-FU, and then infected

**Table 7.** Effect of ATS or carrageenan on the protective activity of thymosin  $\alpha_1$  against *Ps. aeruginosa*

| Pretreatment <sup>a</sup> |      |     |   | Infection <sup>b</sup><br>with<br><i>Pseudomonas</i> | Survival/tested at day |     |          | P value (U-test) <sup>c</sup> |
|---------------------------|------|-----|---|--|------------------------|-----|----------|-------------------------------|
|                           |      |     |   |  | 4                      | 8   | 15       |                               |
| <i>Expt 1</i>             |      |     |   |  |                        |     |          |                               |
| Saline                    | —    |     | + | 8/8  | 8/8                    | 8/8 |          |                               |
| Saline                    | 5-FU |     | — | 8/8  | 8/8                    | 8/8 |          |                               |
| Saline                    | 5-FU |     | + | 1/8  | 1/8                    | 1/8 |          |                               |
| Saline                    | 5-FU | ATS | + | 3/8  | 1/8                    | 1/8 |          |                               |
| Thymosin $\alpha_1$       | 5-FU |     | + | 7/8  | 7/8                    | 7/8 | < 0.025  |                               |
| Thymosin $\alpha_1$       | 5-FU | ATS | + | 8/8  | 8/8                    | 8/8 | < 0.0001 |                               |
| <i>Expt 2</i>             |      |     |   |  |                        |     |          |                               |
| Saline                    | —    |     | + | 7/7  | 7/7                    | 7/7 |          |                               |
| Saline                    | 5-FU |     | — | 7/7  | 7/7                    | 7/7 |          |                               |
| Saline                    |      | Ca  | + | 6/7  | 5/7                    | 5/7 |          |                               |
| Saline                    | 5-FU |     | + | 2/7  | 0/7                    | 0/7 |          |                               |
| Saline                    | 5-FU | Ca  | + | 0/8  | 0/8                    | 0/8 |          |                               |
| Thymosin $\alpha_1$       | 5-FU |     | + | 5/7  | 5/7                    | 5/7 | < 0.001  |                               |
| Thymosin $\alpha_1$       | 5-FU | Ca  | + | 5/7  | 5/7                    | 5/7 | < 0.0001 |                               |

<sup>a</sup> C57BL/6 mice (expt 1) or ddY mice (expt 2) were pretreated with 5-FU or thymosin  $\alpha_1$  (40  $\mu\text{g}/\text{kg}/\text{day}$ ) daily for 10 days from days  $-10$  to  $-1$ . ATS (0.2 ml, IV) was injected at days  $-11$ ,  $-8$ ,  $-5$ ,  $-3$ , and  $-1$ . Carrageenan (Ca, 200 mg/kg, IP) was injected at day  $-1$

<sup>b</sup> Infected with *Ps. aeruginosa* ( $5 \times 10^4$  for expt 1, or  $2 \times 10^4$  for expt 2) by the IV route at day 0

<sup>c</sup> Compared with control mice pretreated with 5-FU, or 5-FU plus ATS or Ca; then infected

### Protective Activity Against Infections with Extracellular Parasites

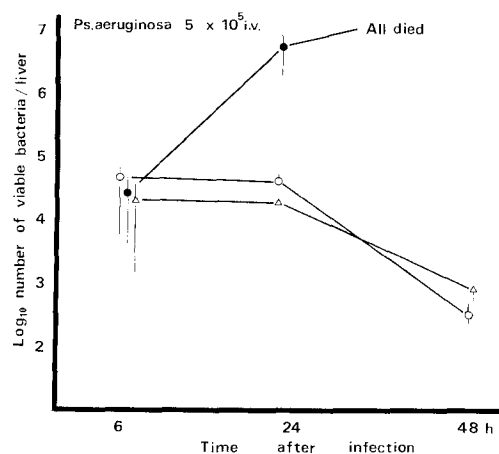
In an attempt to evaluate whether the activity of thymosin  $\alpha_1$  could be useful in other types of opportunistic infections, thymosin  $\alpha_1$  was examined for its activity against infections with extracellular parasites such as *Ps. aeruginosa* and *S. marcescens*. For elimination of these extracellular parasites from infected hosts, neutrophils are known to be essential [17]. As shown in Table 6, thymosin  $\alpha_1$  protected mice from lethal infection with these bacteria, as observed against *Listeria* and *Candida* infections. Furthermore, the activity of thymosin  $\alpha_1$  against *Pseudomonas* was also demonstrated when the number of viable bacteria in liver was compared (Fig. 1). In mice pretreated with 5-FU the viable cells increased, while in mice pretreated with thymosin  $\alpha_1$  and 5-FU it decreased, as observed in normal mice.

### Protective Mechanism Against *Ps. aeruginosa*

The mode of action of thymosin  $\alpha_1$  against *Ps. aeruginosa* infection in the immunosuppressed mice was also investigated in a similar manner to that used in the case of *Candida* infection. While the treatment with ATS or carrageenan abrogated the thymosin activity and increased susceptibility to *Candida* infection, such treatments did not affect the thymosin activity against lethal infection with *Ps. aeruginosa* in the immunosuppressed mice (Table 7). Treatment with thymosin  $\alpha_1$  may exert a direct effect on neutrophils or their precursors, and maintain or recover their immunological function.

### Effect of Thymosin $\alpha_1$ on Infection in Normal Mice

Treatment with thymosin  $\alpha_1$  prevented the bacterial growth to some extent, as seen when the growth of *Listeria* in liver was measured after the infection at a relatively low dose of the bacteria (Table 8). Both control and treated groups survived with such dosages. However, for lethal infection of normal



**Fig. 1.** Effect of thymosin  $\alpha_1$  on the growth of *Ps. aeruginosa* in 5-FU treated mice. ddY Mice were treated daily for 7 days with saline, 5-FU, or 5-FU and thymosin  $\alpha_1$  (40  $\mu\text{g}/\text{kg}$ ) prior to infection with *Ps. aeruginosa* ( $5 \times 10^5$ , IV) (○) saline; (●) saline, 5-FU; (△) thymosin  $\alpha_1$ , 5-FU

**Table 8.** Effect of thymosin  $\alpha_1$  on growth of *L. monocytogenes* in normal mice

| Treatment <sup>a</sup>                                   | No. of bacteria/liver             |                   |                   |
|--|-----------------------------------|-------------------|-------------------|
|  | Days after infection <sup>b</sup> |                   |                   |
|  | 1                                 | 2                 | 3                 |
| Control (saline)   | $2.2 \times 10^6$                 | $1.3 \times 10^6$ | $7.7 \times 10^4$ |
| Thymosin $\alpha_1$ 4 $\mu\text{g}/\text{kg}/\text{day}$ | $1.1 \times 10^5$                 | $1.0 \times 10^6$ | $1.9 \times 10^5$ |
| 40 $\mu\text{g}/\text{kg}/\text{day}$                    | $7.3 \times 10^3$                 | $4.7 \times 10^4$ | $2.5 \times 10^3$ |

<sup>a</sup> Daily treatment with saline or thymosin  $\alpha_1$  from days  $-7$  to  $-1$

<sup>b</sup> Infected with *L. monocytogenes* ( $2 \times 10^3$ , IV) at day 0

mice with much higher doses of *Candida* and *Listeria*, thymosin  $\alpha_1$  showed essentially no activity (data not shown).

## Discussion

The present study deals with the effect of thymosin  $\alpha_1$  against opportunistic infections in mice made immunodeficient by administration of a cytostatic, 5-FU. In these models, co-administration of thymosin  $\alpha_1$  with 5-FU was found to protect mice from lethal infection with all opportunistic pathogens tested so far, i.e., *C. albicans*, *L. monocytogenes*, *Ps. aeruginosa*, and *S. marcescens*. The activity of thymosin  $\alpha_1$  was reproducible and the minimum dose showing the activity was quite low (about 4–40  $\mu\text{g}/\text{kg}/\text{day}$ ), which was about  $1/100$  of the dose required for fraction 5. Both this dose range and the difference between thymosin  $\alpha_1$  and fraction 5 were similar to those observed by Ohta (unpublished work), who assessed the thymosin activity by measuring the restoration of DTH response in mice immunosuppressed by 5-FU. Some activity was also observed in normal mice, but the activity was much clearer in immunosuppressed mice.

Since thymosin  $\alpha_1$  has been reported to affect mainly T cells, it is reasonable that thymosin  $\alpha_1$  could protect mice from the lethal infection with *Candida* and *Listeria*, since T cells participate either directly or indirectly in their elimination from infected hosts [4]. The participation of T cells in the thymosin activity against *Candida* was proved by the fact that the activity was abrogated by treatment with ATS. In addition, treatment with carrageenan increases the susceptibility of normal mice to *Candida*, and consequently abrogated the thymosin activity, indicating that macrophages are crucially important for elimination of the fungi. On the other hand, when carrageenan was given prior to the treatment with thymosin  $\alpha_1$  the activity remained. Thymosin  $\alpha_1$  may serve to maintain the function of macrophages, which is damaged by carrageenan and/or 5-FU, through an effect on T cells. Since it is known that macrophages are activated by sensitized T cells or their products, lymphokines, and that thymosin  $\alpha_1$  affects the differentiation of T cells [1, 5] and enhances the production of lymphokines such as MIF [6] and interferon [8], this is likely.

Thymosin  $\alpha_1$  also showed activity against different types of bacteria, *Pseudomonas* and *Serratia*. For elimination of such bacteria, neutrophils are known to be essential [17]. In contrast to infection with *Candida*, the treatment with ATS or carrageenan could not abrogate the activity of thymosin  $\alpha_1$  against *Pseudomonas* infection. These results indicate that thymosin  $\alpha_1$  might exert some effect on neutrophils or their progenitor cells, either directly or indirectly. This possibility was supported by Ohta and Yagi, who showed that treatment with thymosin  $\alpha_1$  restored the granulocyte-macrophage colony-forming capacity of bone marrow cells in 5-FU-treated mice [13]. Further studies are required to establish this interesting possibility.

In another series of experiments, thymosin  $\alpha_1$  was shown also to be effective in preventing the rapid death otherwise caused by the inoculation of leukemic cells to immunosuppressed mice (Y. Umeda et al. 1982, unpublished work). Both

these results indicate the usefulness of this agent in cancer as an adjuvant to conventional therapies.

## References

1. Ahmed A, Smith AH, Wong DM, Thurman GB, Goldstein AL (1978) In vitro induction of Lyt surface markers on precursor cells incubated with thymosin polypeptides. *Cancer Treat Reports* 62: 1739
2. Bodey GP (1975) Infections in cancer patients. *Cancer Treat Rev* 2: 89
3. Catanzaro PJ, Schwarts HJ, Graham RC (1971) Spectrum and possible mechanism of carrageenan cytotoxicity. *Am J Pathol* 64: 387
4. Dobias B (1964) specific and nonspecific immunity in *Candida* infections. *Acta Med Scand* 176: 79
5. Goldschneider I, Ahmed A, Bollum FJ, Goldstein AL (1981) Induction of terminal deoxynucleotidyl transferase and Lyt antigens with thymosin: Identification of multiple subsets of prothymocytes in mouse bone marrow and spleen. *Proc Natl Acad Sci USA* 78: 2469
6. Goldstein AL, Low TLK, McAdoo M, McClure J, Thurman G, Rossio J, Lai C, Chang D, Wang S, Marvey C, Ramel AH, Meienhofer J (1977) Thymosin  $\alpha_1$ : Isolation and sequence analysis of an immunologically active thymic polypeptide. *Proc Natl Acad Sci USA* 74: 725
7. Hartzman RJ, Bach ML, Bach FH, Thurman GB, Sell KW (1972) Precipitation of radioactively labeled sample: A semiautomatic multiple processor. *Cell Immunol* 4: 182
8. Haung K-Y, Kind PD, Jagoda EM, Goldstein AL (1981) Thymosin treatment modulates production of interferon. *J Int Res* 1: 411
9. Heidelberger C, Griesbach L, Cruz O, Schnitzer RJ, Grunberg E (1958) Fluorinated pyrimidines, VI. Effects of 5-fluorouridine and 5-fluoro-2'-deoxyuridine on transplanted tumors. *Proc Soc Exp Biol Med* 97: 470
10. Hooper JA, MaDaniel MC, Thurman GB, Cohen GH, Schulof RS, Goldstein AL (1975) The purification and properties of bovine thymosin. *Ann NY Acad Sci* 249: 125
11. Ketchel SJ, Rodriguez V (1978) Acute infections in cancer patients. *Semin Oncol* 5: 167
12. Levey RH, Medawar RB (1966) Nature and mode of action of antilymphocyte antiserum. *Proc Natl Acad Sci USA* 56: 1130
13. Ohta Y, Yagi Y (1982) Effect of thymosin  $\alpha_1$  on growth and differentiation of hematopoietic stem cells. (Abstract) *Proc Jap Can Assoc. The 41th Annu Meet* 392
14. Pazmino NH, Ihle JN, Goldstein AL (1978) Induction in vivo and in vitro of terminal deoxynucleotidyl transferase by thymosin in bone marrow cells from athymic mice. *J Exp Med* 147: 708
15. Reed LJ, Muench H (1938) A simple method of estimating fifty per cent endpoint. *Am J Hyg* 27: 493
16. Singer C, Kaplan MH, Armstrong D (1977) Bacteremia and fungemia complicating neoplastic disease. *Am J Med* 62: 731
17. Tatsukawa K, Mitsuyama M, Takeya K, Nomoto K (1979) Differing contribution of polymorphonuclear cells and macrophages to protection of mice against *Listeria monocytogenes* and *Pseudomonas aeruginosa*. *J Gen Microbiol* 115: 161
18. Umeda Y, Nakamura J, Ishitsuka H, Yagi Y (1981) Protective mechanisms of thymosin  $\alpha_1$  against opportunistic infections. (Abstract) *Proc Jap Soc Immunol* 11: 222
19. Wara DW, Goldstein AL, Doyle W, Ammann AJ (1975) Thymosin activity in patients with cellular immunodeficiency. *N Engl J Med* 292: 70

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