

Karl-Johan Malmberg

## Effective immunotherapy against cancer

### A question of overcoming immune suppression and immune escape?

Received: 2 October 2003 / Accepted: 9 October 2003 / Published online: 28 July 2004  
© Springer-Verlag 2004

**Abstract** During the last decade, the breakthroughs in understanding of the molecular mechanisms responsible for immune activation and the advent of recombinant DNA technologies have changed the view on immunotherapy from “a dream scenario” to becoming a clinical reality. It is now clear that both cellular immunity comprising T and NK cells, as well as strategies based on antibodies, can provide strong antitumoral effects, and evidence is emerging that these strategies may also cure patients with previously incurable cancers. However, there are still a number of issues that remain unresolved. Progress in immunotherapy against cancer requires a combination of new, improved clinical protocols and strategies for overcoming mechanisms of immune escape and tumor-induced immune suppression. This review discusses some of the salient issues that still need to be resolved, focusing on the role of oxidative stress and the use of antioxidants to alleviate the immune hyporesponsiveness induced by reactive oxygen species (ROS).

**Keywords** Immune escape · Immune suppression · Immunotherapy · Oxidative stress

#### Abbreviations

HLA Human leukocyte antigen  
KIR Killer cell immunoglobulin-like receptor  
NKR Natural killer cell receptor

ROS Reactive oxygen species  
TAA Tumor-associated antigen

#### Introduction

##### Tumor immunoediting

In the early 20th century, the immunologist Paul Ehrlich [37] first conceived of the idea that tumors arise continuously in our bodies and that the immune system scans for and eradicates these tumors before they become clinically manifested. Experimental evidence that tumors could be repressed by the immune system came from the use of tumor transplantation models in the mid-20th century [51]. The results from these models strongly suggested the existence of tumor-associated antigens (TAAs) and formed the basis of the “immun-surveillance theory” postulated by Sir Macfarlane Burnet and Lewis Thomas [17, 18, 150]. According to this theory, spontaneously developing carcinomas are efficiently eliminated by the immune system recognizing specific determinants expressed by the tumor cells. Burnet claimed that the presence of such a tumor scanning system was an evolutionary necessity and as important for survival as the protection against infections. Soon, a critique against this concept was aroused. Experimental data challenged predictions of the immun-surveillance theory. There was no increased incidence of spontaneous or chemically induced tumors in athymic, nude mice, as compared to wild-type animals even though long observation periods of up to 420 days were employed [129, 145]. If tumors eventually appeared in immunocompromised mice these were often virally induced lymphomas [68]. All these lines of evidence argued against immunological surveillance. However, recent evidence from knockout (KO) mice lacking components of the IFN- $\gamma$  signaling pathways, perforin, or

This work is part of the Symposium in Writing “Tumor escape from the immune response,” published in vol 53.

K.-J. Malmberg  
Center for Infectious Medicine (CIM), F59,  
Department of Medicine, Karolinska Institutet,  
Karolinska University Hospital Huddinge,  
S-14186 Stockholm, Sweden  
E-mail: kalle.malmberg@medhs.ki.se  
Tel.: +46-8-58589635  
Fax: +46-8-7467637

recombination activating gene 1/2 (RAG-1/2), has revived the discussion of a potential immunosurveillance against tumors [33]. These mice display enhanced sensitivity to methylcholanthrene (MCA)-induced tumor formation when compared with wild-type mice. RAG-2-deficient mice also developed spontaneous epithelial tumors of predominantly intestinal origin. Further support of immunosurveillance against tumors in humans is provided by the positive correlation between the presence of lymphocytes in a tumor as detected by immune histochemistry, and increased survival of patients with colorectal cancer [106]. The experimental revival of the immunosurveillance concept has extended the phenomenon, and it is now referred to as “tumor immunoediting” [33]. Tumor immunoediting includes both immunosurveillance and immune escape. It refers to the protective capacity of the immune system against tumors (i.e., immunosurveillance) and to the consequences of the proposed immune selection pressure (i.e., immune escape). However, the process of tumor immunoediting is somewhat distinct from immune escape, as it includes the observation that tumors that grow in immunocompromised hosts become more immunogenic and are easily rejected when transplanted into immunocompetent mice. In the absence of immune selection, tumors may develop an immunogenic phenotype. Evidence for immune escape upon immune-mediated selection pressure is abundant both in mice and humans. Re-passage of transplantable tumors through immunocompetent hosts leads to the development of tumor variants with reduced immunogenicity [157, 158]. In humans, the immune selection pressure combined with the genetic instability of tumor cells frequently gives rise to deletions and/or altered expression of several molecules important for immune recognition, such as components of the antigen processing and presentation pathway [45, 139]. Several aspects of the original version of the immunosurveillance theory were probably incorrect, and most tumor immunologists would probably agree that it is not a primary property of the immune system to fight cancer. From an evolutionary perspective, there is no advantage to protecting a species from a disease occurring after fertile age [171]. Most tumors arise late in life when the evolutionary pressure cannot exert its influence. Nevertheless, as outlined above, most tumors are antigenic and can be eradicated by the immune system given the right circumstances [162]. Lack of immunogenicity is therefore a result of the tumor’s inability to induce an immune response. Understanding the particular conditions under which antitumor immune responses can be raised is the main challenge for cancer immunotherapy.

---

### **Immunotherapy against cancer**

Although there are anecdotes telling of immunization attempts against cancer performed during the

18th century, it is not until the last decade that the antigens and the T-cell receptors responsible for immune recognition of tumors have been identified. Still, the progress of vaccination against cancer is dependent on new insights in the fields of basic cellular and molecular immunology. Fundamental properties of the immune system such as formation of long-term memory, tolerance, immunodominance, and anergy remain to be fully understood. The use of mature dendritic cells (DCs) both to break tolerance when trying to induce tumor immunity [107] and to induce peripheral tolerance in the case of autoimmune disorders [156] clearly demonstrates that more knowledge is needed to elucidate the basic mechanisms of immune activation.

### **Targeting specific tumor antigens with cancer vaccines**

Based on the discovery and identification of tumor-associated antigens, it is now possible to target tumors specifically by antibodies or by cytotoxic T lymphocytes (CTLs) [123]. This may be achieved either by specific vaccination or adoptive transfer of *in vitro* generated antitumor CTLs. Several different strategies of vaccination have been tried, including delivery of tumor antigens in the form of peptides, proteins, naked DNA, or whole tumor cells [35, 124, 152]. Vaccines based on DCs pulsed or transduced with tumor antigens have also been tested with varying degrees of success [38]. Altogether these efforts clearly demonstrate that it is possible to induce tumor-specific T-cell responses and in a few patients even objective clinical responses. Herein, the immense literature on the development of cancer vaccines will not be reviewed. The discussion will be limited to some of the obstacles for effective immunization against cancer. Most defined tumor antigens are products of normal genes that are overexpressed or selectively expressed in tumor tissues [47]. Thus, one of the major hurdles for effective vaccination is to overcome the central and peripheral tolerance to these “self” antigens. The existing T-cell repertoire specific for self-antigens is limited to low quality “low avidity” T cells that may recognize a peptide-pulsed target but not endogenously presented antigens on tumor cells [34]. Nevertheless, it has been shown that immunization can be effective even though the response is limited to low avidity CTLs [104]. Attempts have also been made to create more immunogenic antigens by molecular manipulation. By inserting an amino acid change in the peptide epitope, it is possible to produce an antigen that binds more strongly to the relevant HLA molecule and therefore stands a higher chance of breaking tolerance against self-proteins [124]. Vaccination of melanoma patients with a modified peptide, gp100<sub>209–217</sub> (210 M), from the gp100 melanocyte differentiation antigen was able to induce an immune response that recognized the native peptide antigen and HLA-A2-matched tumors. In combination with IL-2, this peptide induced objective clinical responses in 13 out of 42 treated patients [124].

## Bone marrow transplantation and adoptive immunotherapy

Although vaccination against cancer is a promising approach, some of the most encouraging data regarding immunotherapy against cancer stems from studies employing adoptive transfer of tumor reactive T cells and perhaps also of NK cells. Bone marrow transplantation (BMT) has recently been used as immunotherapy of solid tumors with promising results [21]. The intersection of BMT and more specific immunotherapy based on the knowledge of defined antigens is particularly interesting, as illustrated by two recent studies. First, Dummer et al. [32] exploited the homeostatic proliferation phase of T cells occurring after sublethal irradiation (conditioning) to induce strong CD8 T-cell responses against tumor-associated self-antigens. This was based on the concept that homeostatic proliferation occurring in situations of lymphopenia is driven by recognition of self-MHC/peptide ligands similar to those that control positive selection of T cells in the thymus [93]. Adoptively transferred T cells, educated during the homeostatic reconstitution in lymphopenic and tumor-bearing animals, grew to become tumor specific. Similarly, Dudley and colleagues recently reported remarkable clinical responses after adoptive transfer of tumor-specific, tumor-infiltrating lymphocytes (TILs) into melanoma patients that were conditioned with low doses of chemotherapy [31]. Around 50% of the patients experienced objective clinical responses. Other studies provide further evidence that the posttransplant period is optimal for redirecting immune responses toward tumor antigens [6, 149]. The reasons for the beneficial effect of lymphopenia during immunotherapy remain unclear. It may involve the presence of homeostatic proliferation, but other factors such as depletion of immunoregulatory CD4<sup>+</sup>CD25<sup>+</sup> T cells may also contribute. In another study, T-cell clones specific for the Melan-A/MART-1 or the gp100 melanoma differentiation antigens were repeatedly infused into patients with metastatic melanoma along with low doses of IL-2 [168]. Objective tumor responses were seen along with preferential localization of the infused T cells to the tumor sites. However, adoptively transferred T cells are rapidly eliminated from the circulation and the effect of adoptive immunotherapy is therefore short lived. This may be due to homeostatic forces contracting the lymphocyte population to normal levels. Alternatively, *in vitro* expanded T cells may have a limited lifespan due to lymphocyte senescence [112]. In other models, EBV-specific T cells were shown to have the potential of long-term survival when adoptively transferred to a recipient [57]. This property was recently exploited by expressing a chimeric tumor-specific TCR in EBV-specific T cells [127]. These T cells survived long-term *in vivo* and provided a source of T cells capable of killing both EBV-infected cells and tumor cells expressing the antigen recognized by the chimeric receptor. It will be important to define the factors that influence the functional

properties and the survival of *in vitro*-expanded T cells to be used for adoptive immunotherapy against cancer. Yet another breakthrough comes from the potential of NK cells in the setting of allogeneic bone marrow transplantation [128]. NK cells are negatively regulated by inhibitory HLA class I-binding receptors including the killer cell immunoglobulin-like receptors (KIRs) [75]. Briefly, it was demonstrated that the alloreactivity of NK cells, as governed by a donor-recipient KIR-HLA mismatch, correlated with increased survival, more successful grafting, and decreased risk of graft-versus-host disease (GVHD). The mechanism behind the latter observation was dissected in a mouse model revealing that the alloreactive NK cells eliminated GVHD by killing the host antigen-presenting cells, thus preventing these cells from presenting host antigens to donor T cells which initiates the GVH reaction [143]. The striking survival advantage for patients with AML who received KIR mismatched bone marrow transplants clearly demonstrates that the immune system is capable of eradicating tumor cells and suggests that NK cells provide an excellent cellular source for adoptive immunotherapy. Alternative strategies of adoptive immunogene therapy that are based on gene transfer into immune effector cells that are infused back to the patients are also very promising. For examples it was possible to eradicate systemic B-cell tumors using human T cells that were genetically targeted to the CD19 antigen [16]. Other approaches involve transfection of human PBMCs with selected T-cell receptors with specificity for tumor-associated antigens (TAAs) [22]. This strategy has advantages compared with conventional adoptive T-cell therapy. For instance, one can select pre-made constructs with specificities for the tumor of each patient and avoid the difficulties associated with T-cell cloning. Although results from different approaches of immunotherapy bring hope for a successful future for this field, it is clear that much improvement is needed before immunotherapy will become a powerful therapeutic tool in the clinic. Several issues that need to be considered will be discussed below.

### Timing the immunotherapeutic approach with stage of disease

Perhaps the most important explanation for why immunotherapy has not been more successful in the clinical setting relates to the timing of immunotherapeutic intervention with respect to the disease progression. To date, most vaccinations against cancer have been performed in the late stages of the disease when all other treatment modalities have failed. This is a major difference compared with vaccines against viral or bacterial antigens, as patients with advanced cancer are often immune suppressed and not as responsive to immunization (see further below and references [66, 113]). Furthermore, most vaccines against infectious diseases are prophylactic and allow the immune system

to be prepared already at the onset of infection. This is in sharp contrast to cancer vaccines performed in “late stage” disease when a large number of tumor cells are present. Although there are striking examples of regression of large tumors [125, 126], it is likely that vaccines against cancer will operate more efficiently when administered early during tumor progression or directly after conventional treatment modalities, such as surgery or chemotherapy. One example of this possibility comes from a vaccination study against lymphoma with an idio-type protein vaccine in combination with GM-CSF [10]. Eight out of 11 patients that were in their first clinical complete remission after standard chemotherapy but still had molecular signs of disease, as determined by the presence of cells with the 14;18 translocation, went into molecular remission upon vaccination. Similar mechanisms may account for the curing potential of the graft-versus-leukemia (GVL) effect seen in BMT against acute leukemias. The GVL effect of T and NK cells follows upon chemotherapy, and patients are often in complete remission before being subjected to the bone marrow transplantation. Thus, the immune system may efficiently eradicate minimal residual disease. In situations of larger tumor burden, adoptive transfer of high numbers of T or NK cells may be needed.

---

### Impediments to immunotherapy

As immunotherapy becomes more efficient, the immunological selection pressure against tumor cells will increase. In combination with the inherited genetic instability of tumor cells, this will lead to the development of immune escape variants [65]. Below, several ways by which tumors may escape the immune system will be discussed with references to the current thinking on how to counteract these mechanisms. This review will give a particular emphasis on tumor-induced immune suppression and the role of oxidative stress for decreased T-cell function, as this may constitute a substantial impediment for effective immunotherapy against cancer. It should be stressed that the development of tumor-induced immune suppression is perhaps more of a reflection of the large tumor burden in “late stage” disease, than a consequence of the rules of the microevolution and selection pressure.

#### Loss of antigen processing and presentation

The presentation of antigens in the context of HLA molecules is crucial both during T-cell priming and during the effector phase of an adaptive immune response. Alterations in antigen processing and presentation are commonly seen both during viral infection and in malignancies. Viruses have adapted to the immune-mediated selection pressure in numerous ingenious ways,

affecting almost every part of the biochemistry of the MHC presentation pathway [169]. For instance, Epstein-Barr viruses (EBVs) express a protein (EBNA-1) that carries a repeated sequence (Gly-Ala repeat) that makes it inaccessible for proteasomal degradation into peptides [81]. The herpes simplex virus (HSV)-derived protein ICP47 and the cytomegalovirus (CMV)-derived protein US6 interfere with TAP and prevent peptide translocation into the ER [42]. In tumors, the antigen presentation pathway is disrupted as a consequence of mutations and/or deletions of one or several genes encoding components of the antigen-processing machinery. Complete HLA loss is a common event in several murine and human tumors [44, 140]. Total loss of HLA class I is usually associated with alteration of  $\beta_2m$  expression as the MHC class I molecule is not properly assembled in the absence of this molecule. Mutations of  $\beta_2m$  along with loss of heterozygosity (LOH) of the second allele has been described and may be associated with progression of disease in the course of adoptive immunotherapy [120]. However, also the loss of proteasomal subunits such as LMP-2 and LMP-7, as well as of peptide transporters TAP1 and TAP2, leads to total loss of HLA class I and was reported in several different tumor types including cervical carcinoma, small cell lung carcinoma, non-small cell lung cancer, prostate carcinoma, and renal cell carcinoma [140]. Tumors often display a selective loss of HLA haplotype, locus, or allele. Haplotype losses may be the result of LOH on chromosome 6 [118], whereas several mechanisms may account for the locus down-regulation. Sequential loss of several alleles in successive metastasis from a single patient who underwent immunotherapy suggests that immune-mediated selection pressure plays a pivotal role in selecting the HLA loss variants [78]. A study of several melanoma cell lines demonstrated low or no expression of gene products of the HLA-C locus [92]. An interesting aspect of the loss of certain HLA alleles comes from the study of Ikeda et al. [60] where they demonstrate that tumor cells that had lost HLA-Cw7 upon progression could be sensitized to tumor antigen-specific CTLs. These CTLs were incapable of killing tumors derived from an earlier biopsy, before any loss of HLA had occurred, although they expressed the same tumor antigen. The reason turned out to be that the expression of HLA-Cw7 by early tumors enabled ligation of inhibitory KIR, expressed by the CTLs. Thus, loss of HLA resulted in increased sensitivity to CTLs. Similarly, tumor cells that lose HLA class I should become more sensitive to NK-cell-mediated lysis because they fulfill the criteria of “the missing self hypothesis” [86]. Indeed, this has been reported to be the case. Loss of  $\beta_2m$  leads to increased NK sensitivity and effective tumor elimination [62]. However, NK cells may express inhibitory NK receptors (iNKR) belonging to different families that only partially overlap, and the failure of one receptor-ligand system may be compensated for by another; i.e., lack of HLA-C/KIR signaling may be compensated for by the HLA-E : CD94/NKG2C

interaction [88]. Furthermore, NK cells are dependent also on activation signals, which is why lack of inhibition is not always enough. The balance of activating and inhibitory signals to NK and T cells will be discussed below.

#### Antigenic loss variants

An unfortunate consequence of efficient immunotherapy is that the immune system selects for tumor variants that are no longer recognized, i.e., that have lost the targeted antigen. Antigenic variation is the hallmark of many successful pathogens and may be the most difficult obstacle for the induction of long-lasting immunity against many infectious diseases including HIV [46]. This phenomenon was recently demonstrated in the study of Yee et al. [168], where the histology of recurrent metastasis following adoptive transfer of Mart-1-specific T cells was examined. In three out of five patients studied, the recurrent metastasis lacked expression of Mart-1 while expression of gp100 and tyrosinase remained intact. Similar observations have been made in the course of peptide and DC immunizations [151]. These results strongly suggest that one has to target multiple tumor antigens, or perhaps antigens that are crucial for the growth of the tumor cells. In the advent of antigen loss variants, the tumor cells would stop dividing and thereby lose their malignant potential. There are several reported successful attempts of immunotherapy using multiepitope vaccines. For instance it was reported that one could obtain immune responses to several epitopes by using genetic constructs coding for a series of epitopes ordered in a “string-of-beads” fashion [153]. However, the use of several epitopes together leads to yet unpredictable patterns of immunodominance. Immune responses to immunodominant epitopes within a multiepitope construct may undermine responses to subdominant epitopes. The factors governing immunodominance are largely unknown but will be important to elucidate, as they influence the efficacy of multiepitope vaccines as a solution for the escape of antigenic loss variants.

#### The Fas “counterattack”

The Fas “counterattack” refers to the possibility that tumor cells attack and kill T cells that home in to the tumor. This concept was based on the observation that FasL-expressing sertoli cells were not rejected when transplanted into allogeneic mice [163]. It was concluded that FasL expression was a key feature of immunological privilege supported by the finding of FasL expression in the eye [49]. This held great promise for future clinical applications as FasL-expressing transplants would be tolerated by the recipient. The first example of FasL expression on tumors was in colon cancer, rapidly followed by studies of melanoma [52]. FasL expression had

functional consequences, as it induced apoptosis of Fas-expressing Jurkat T cells and tumor-infiltrating lymphocytes. FasL expression was also reported on ovarian carcinoma leading to signaling defects and apoptosis of tumor-associated lymphocytes [117]. More recently, the concept of a tumor-mediated counterattack has been revisited [119]. When trying to transplant FasL-expressing islet  $\beta$ -cells, one observed granulocyte infiltration and a rapid rejection of the transplant [2]; hence the opposite to tolerance. Similar observations have been made in tumor models in which FasL-expressing tumors were rapidly rejected [4]. Furthermore, several researchers failed to see expression of FasL on tumors previously reported to express this molecule [39]. The Fas-mediated apoptosis of infiltrating lymphocytes seemed rather to be the consequence of T-cell activation and activation-induced cell death [170]. The Fas death receptor signaling pathway may instead be a very important component of host defense against tumors [24]. Not surprisingly, escape variants that are resistant to death receptor signaling have evolved (discussed below).

#### Escaping death receptor signaling

Two death receptor signaling systems that contribute to the immunosurveillance of tumors are FasL and TNF-related apoptosis-inducing ligand (TRAIL) [24, 147]. Death receptors have cytoplasmic sequences—death domains—which transmit the apoptotic signal upon ligation. This triggers a cascade of caspases involving caspase 8 (also known as FLICE), caspase 3, and caspase 6 and 7. Tumor cells may become resistant to death receptor signaling by expressing an inhibitor of caspase 8—cellular FLICE inhibitory protein (FLIP) [29]. Overexpression of FLIP was shown to mediate tumor cell escape in vivo [96]. Tumors also display loss of Fas expression which renders them resistant to apoptosis [74]. Alterations may appear further downstream in the death receptor signaling pathway, including functional impairment of FADD and caspase 10 by inactivation mutations [142]. Similarly, tumors escape death receptor signaling by losing the expression of TRAIL receptors or downstream signaling components [56]. Yet another intriguing way for tumors to escape recognition by CTLs and NK cells is by overexpressing PI-9 (in mice known as SPI-6) [95]. PI-9 is a serine protease inhibitor that inactivates granzyme B which is one of the important components of granule-mediated CTL killing [154]. One suggested physiological role of SPI-6 is to protect activated DCs from being killed by the induced CTLs during a Th1 response [97].

#### Engaging inhibition-blocking activation

The balance between activation and inhibition of T and NK cells is dependent on the quality of a number of

receptor–ligand interactions. Evidence suggests that tumors may evade T- and NK-cell recognition by shifting the balance toward inhibition. Ectopic expression of the nonclassical HLA class I molecule, HLA-G, has been described in several different tumors including melanoma, renal cell carcinoma, breast carcinoma, cutaneous lymphoma, and ovarian carcinoma [59, 77, 91, 111]. HLA-G expression protects tumor cells from lysis by T and NK cells via ligation of inhibitory receptors LIR-1 and/or KIR2DL4. The HLA-G leader sequence–derived peptide (Gsp) binds another nonclassical HLA class I molecule, HLA-E, and thereby promotes a strong interaction with CD94/NKG2 receptors expressed on NK cells and a subset of T cells [91, 160]. It has been demonstrated that the strength of the inhibitory signal depends on properties of the leader sequence peptide that is bound to the HLA-E molecule [160]. While some peptides, including the HLA-G leader sequence–derived peptide, mediate strong interaction with the CD94/NKG2 heterodimer and thereby promote significant signaling, other peptides seem to silence this pathway [100]. HLA-G also shuts down other effector functions such as the production of IFN- $\gamma$  and Fc $\gamma$  receptor–mediated activation of LIR-1–expressing macrophages [23]. Soluble HLA-G1 was furthermore demonstrated to induce apoptosis of CD8<sup>+</sup> T cells via a Fas-FasL dependent mechanism [41], perhaps constituting a refined version of the counterattack concept. The secreted form of HLA-G1 induced the expression of functional FasL in activated T cells, and this was shown to be the result of the interaction between the conserved  $\alpha 3$  domain of the HLA-G1 molecule and the CD8 molecule. Similarly, soluble HLA class I molecules, including the nonclassical HLA-E and HLA-F molecules, as well as alleles of HLA-C, were shown to induce apoptosis of NK cells upon ligation of activating receptors [144]. Again, this was dependent on the up-regulation of FasL in stimulated NK cells resulting in suicide along with killing of neighboring Fas-expressing NK cells (Fratri-cide). An alternative way for tumors to avoid being killed by NK and T cells is by shedding decoy ligands for activating receptors. Tumors were found to secrete soluble MICA, which is the ligand for the activatory receptor NKG2D [50]. It was demonstrated that soluble MICA is frequently found in sera of cancer patients at levels sufficient to cause down-regulation of NKG2D on tumor-infiltrating and circulating  $\alpha\beta$  and  $\gamma\delta$  T cells and NK cells. As NKG2D is involved in direct recognition of MICA-expressing tumors, as well as in costimulating T-cell responses, this had functional consequences, down-modulating the reactivity and cytokine production capacity of antitumor CTLs. The expression of ligands that have the capacity to turn off the function of immune effector cells could potentially abolish an otherwise effective T- or NK-cell–mediated tumor recognition. Such inhibitory pathways can be targeted. For instance, it has been shown that mouse NK cells that were preincubated with mAbs against inhibitory receptors had a more potent antitumor effect when

adoptively transferred into tumor-bearing mice [69]. Alternatively, one may select or genetically engineer NK- and T-cell clones to express less of inhibitory receptors and perhaps more of activating receptors, prior to adoptive transfer. Another interesting possibility would be to target the nonclassical HLA class I molecules that are ectopically expressed by tumors. This could be done either by humanized specific antibodies or by generating HLA-G or HLA-E allo-specific T cells. With an increasing knowledge of the receptor–ligand interactions that govern T- and NK-cell activation, it may be possible to augment antitumor responses and dampen undesirable reactivity of the immune system.

#### Suppression of antitumor responses by regulatory T cells

Recently, a subset of T cells, coexpressing CD4 and CD25 (IL-2R  $\alpha$  chain), with unique immune regulatory properties was described [130]. These regulatory T cells (T<sub>reg</sub>) seem to play a pivotal role in immune homeostasis, protecting the host from several T-cell–mediated autoimmune disorders including type 1 diabetes, hypothyroidism due to thyroiditis, and pernicious anemia due to gastritis [122]. CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells dramatically suppress the function and proliferation of CD4<sup>+</sup>CD25<sup>-</sup> and CD8<sup>+</sup> T cells [115]. Much of the immune regulatory function of these cells has been attributed to their capacity to secrete immune-suppressive cytokines such as TGF- $\beta$  and IL-10 which may severely inhibit CTL responses as will be further described below [122]. Although beneficial in protecting the host from autoimmune disorder, regulatory T cells may also dampen antitumor responses. When T<sub>reg</sub> were depleted in mice, transplantable tumors were efficiently rejected by the host immune system [141]. T<sub>reg</sub> has been shown to infiltrate several types of human tumors, including non-small cell lung cancer, and pancreas, breast, and ovarian carcinomas [85, 165]. Regulatory T cells are also found at a higher frequency in the peripheral blood of cancer patients and may induce peripheral ignorance of tumor cells, facilitating metastatic spread of the disease [85]. However, in a recent report by Edinger et al. [36], a novel application of regulatory T cells was demonstrated. Cotransplantation of T<sub>reg</sub> with allogeneic BM could inhibit graft-versus-host reaction of transplanted T cells while preserving their graft-versus-tumor potential, thus curing animals with leukemia and lymphoma. T<sub>reg</sub> inhibited the massive proliferation of alloreactive T cells associated with the onset of GVHD without affecting their function.

#### Tumor-induced immune suppression

Mice with experimental tumors and patients with cancer show evidence of decreased immunological potency [66]. This is manifested by decreased proliferation, decreased Th1 cytokine production, and a poor cytolytic capacity

of TILs and PBLs. On a molecular level, this is associated with decreased levels of signaling molecules such as CD3 $\zeta$ , p56<sup>lck</sup>, p59<sup>lyn</sup>, ZAP70, JAK3, and STAT5, and the blocking of nuclear factor- $\kappa$ B [71, 72, 101, 114, 159]. The underlying mechanisms for immune dysfunction in cancer patients are probably multifactorial. A large number of factors, including old age and cachexia, contribute to the poorly functioning immune system in cancer patients [112]. However, the role of the tumor itself was nicely demonstrated in mice with fibrosarcoma [132]. Upon inoculation with tumors, these mice became immune suppressed, as manifested by impaired antitumor reactivity of T cells associated with decreased expression of signaling molecules. This phenotype was completely reversed following surgical removal of the tumor, unmasking a population of primed T cells that were capable of rejecting a subsequent tumor challenge.

### Mechanism of tumor-induced immune suppression

The unresponsiveness of the immune system in patients with cancer has been attributed to the secretion of immune-suppressive cytokines such as IL-10 and TGF- $\beta$ . Indeed, and as mentioned above, these are the cytokines that may be responsible for the suppressive effect of regulatory T cells. IL-10 is frequently detected in high amounts in cancer patients [116]. This cytokine may skew immune responses away from a protective and antitumor Th1 response [64]. IL-10 also inhibits differentiation, maturation and functional status of DCs [25], thus interfering with the induction of antitumor responses. Furthermore, IL-10 may suppress T-cell-mediated immunity by down-regulating the function of TAP molecules and the expression of MHC class I molecules on target cells [131]. The immune-suppressive properties of IL-10 have been adopted by the Epstein-Barr virus which has a gene product (BCRF1) with striking homology to the mammalian IL-10 sequence [103]. TGF- $\beta$  is another cytokine that inhibits activation, proliferation, and activity of lymphocytes [40]. This cytokine is often found at high levels in malignancies and is associated with poor prognosis and lack of response to immunotherapy [48].

### Suppressor macrophages and free radicals

In the mid-1970s, macrophages from tumor-bearing hosts were found to inhibit proliferative responses of splenocytes in a non-antigen-dependent manner [67]. Splenocytes from tumor-bearing individuals had proliferative responses that were 90% below normal. However, if these "suppressor macrophages" were removed, the proliferation was completely restored. Adoptive transfer of macrophages from tumor-bearing mice facilitated tumor growth in the recipient [167]. Several human tumor types, including carcinomas of the breast, bladder, and cervix, show an association between the

levels of infiltration of tumor-associated macrophages and poor prognosis [13]. Such protumor effects by macrophages may depend on secretion of proangiogenic cytokines and enzymes [76], secretion of immune suppressive cytokines [87], and release of free radicals including NO [1] and hydrogen peroxide [138]. Free radicals may act on immune cells and render them hyporesponsive or even apoptotic [66]. It is important to stress that the action of H<sub>2</sub>O<sub>2</sub> may be beneficial to the host as a component of innate immunity against bacteria and other pathogens [7]. There is also abundant evidence for a potent proapoptotic and antitumor role for free radicals [102]. Furthermore, radiation therapy is dependent on the generation of free radicals and their capacity to induce DNA damage and apoptosis of tumor cells [11]. Thus, the role of tumor-associated macrophages in tumor progression is controversial.

### Consequences for immune cell function and signaling

Early studies suggested that macrophages inhibited T- and NK-cell function by releasing hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) [138]. The addition of catalase, an enzyme that specifically catalyzes the reduction of H<sub>2</sub>O<sub>2</sub>, rescued NK cells from the inhibitory effects of activated macrophages [138]. This was further supported by other investigators showing that histamine, an inhibitor of H<sub>2</sub>O<sub>2</sub> production, could rescue NK cells from the detrimental effects of activated macrophages [54]. Later, it was shown that several other features of tumor-induced immune suppression, including loss of Th1 cytokine production, and decreased expression of signaling molecules including the CD3 $\zeta$  chain, were induced by activated or tumor-infiltrating macrophages [3, 73]. These effects could be mimicked by exogenous H<sub>2</sub>O<sub>2</sub> and were restored in the presence of catalase, an enzyme that catalyzes the reduction of H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O + O<sub>2</sub>. Interestingly, tumor cells have also been found to produce large amounts of hydrogen peroxide [146], which could potentially induce T-cell dysfunction of tumor-infiltrating T cells. The hyporesponsiveness induced by oxidative stress has been investigated in several models. Hydrogen peroxide, added exogenously to freshly isolated and noncultured PBLs, selectively targeted Th1 cytokine production in CD45RO<sup>+</sup> memory/activated T cells [89]. The loss of Th1 cytokine production in CD45RO<sup>+</sup> T cells was associated with a block of NF- $\kappa$ B activation. This suggested that H<sub>2</sub>O<sub>2</sub> inhibited proximal signaling events in CD45RO<sup>+</sup> T cells, leading to diminished activation of NF- $\kappa$ B and impaired production of cytokines regulated by this transcription factor [8]. Although not shown, the reason for such selective targeting of CD45RO<sup>+</sup> cells may be related to the role of the CD45 molecule in T-cell receptor signaling. CD45 is a protein tyrosine phosphatase (PTP) that preferentially removes phosphates bound to regulatory tyrosine residues (C-terminal tyrosine residues) on src family kinases. This is essential for T-cell activation and

sustained TCR signaling as it allows lck and fyn to be phosphorylated at their active sites [155]. CD45 exists as several isoforms due to alternative splicing of three consecutive exons (4, 5, and 6, designated A, B, and C) in the extra cellular domain [155]. Naïve T cells predominantly express the larger CD45RA isoforms and, over the course of 3 to 5 days after activation, they switch to express the smallest CD45RO isoform [14]. Compatible with the rapid onset of a memory response, CD45RO<sup>+</sup> CD4 and CD8 T cells are highly sensitive to antigen and secrete a broad range of cytokines upon stimulation [108]. Interestingly, this responsiveness to antigen correlates with the colocalization of the CD45RO isoform with CD4/CD8 and the TCR at the cell surface [28, 30, 79, 80]. The association of the CD45RO isoform with other signaling molecules may be responsible for the more efficient response to antigen by memory T cells. However, it may also explain the enhanced sensitivity of this subset to H<sub>2</sub>O<sub>2</sub>. It is well known that H<sub>2</sub>O<sub>2</sub> functions as a PTP inhibitor [19]. It can therefore be speculated that H<sub>2</sub>O<sub>2</sub> interferes with proper TCR signaling, by blocking the PTP activity of CD45RO. Consequently NF- $\kappa$ B activation and cytokine production is blocked. Such inhibition of CD45 PTP activity would have less functional consequence for CD45RA<sup>+</sup> cells, as the CD45RA molecule is less active in promoting TCR signaling due to its distant localization from the T-cell receptor [79]. Another recently described phenomenon that may help in understanding the selective targeting of memory/activated T cells by oxidative stress is the enhanced efficiency by which the CD45RO molecule forms inactive homodimers [166]. This is particularly interesting as H<sub>2</sub>O<sub>2</sub>-mediated inhibition of PTP activity is known to involve formation and stabilization of homodimers [15]. It should be stressed though that the H<sub>2</sub>O<sub>2</sub>-mediated inhibition of other PTPs in T cells may instead directly promote kinase activity and induce signaling [61]. Indeed, there is a renewed interest in the role of H<sub>2</sub>O<sub>2</sub> as a second messenger [121]. For instance, H<sub>2</sub>O<sub>2</sub> may mimic the effects of an antigen, leading to stimulation of resting B cells. As BCR engagement leads to H<sub>2</sub>O<sub>2</sub> production stimulating neighboring BCR in an antigen-independent, paracrine fashion, it was suggested that this results in a rapid amplification of the BCR signal [121]. Furthermore, anti-CD3 stimulation of mature T cells leads to production of ROS including hydrogen peroxide and superoxide anion [27]. It has been proposed that the oxidative burst by macrophages serves the sole purpose of stimulating T and B cells [121]. Along these lines, several papers have reported that hydrogen peroxide activates NF- $\kappa$ B in lymphocytes and induces production of IL-2 [134, 136]. However, other studies have shown that IL-2 production upon T-cell activation is independent of ROS production [27]. Recently a sharp analysis of the molecular mechanisms underlying ROS-induced signaling defects in hyporesponsive cells was reported [20]. T cells that were cocultured with activated neutrophils exhibited decreased expression of CD3 $\zeta$ , p56<sup>lck</sup>,

and LAT. Another signaling molecule, PI3 K, was spared, indicating a differential effect of oxidative stress on different components of the T-cell signaling machinery. Two sequential effects of oxidative stress were described: protein modification and protein degradation. While p56<sup>lck</sup> and LAT were sensitive to protein degradation, CD3 $\zeta$  was relatively resistant to degradation. In contrast, loss of the CD3 $\zeta$  molecule was shown to be dependent on protein modification. This was demonstrated using two antibodies with distinct binding motifs on the CD3 $\zeta$  molecule. When using an antibody recognizing the cytoplasmic domain of the molecule, CD3 $\zeta$  expression was decreased in a dose-dependent manner by coculture with ROS-producing granulocytes. However, the recognition of CD3 $\zeta$  by an antibody directed toward the membrane proximal domain was unaffected. Moreover, protein modification seemed to be the rate-limiting step for induction of hyporesponsiveness as function could not be restored by blocking degradation.

The complexity of redox regulation of immune responses and the dual properties of hydrogen peroxide as a second messenger and a negative regulator of immune reactivity are interesting. Perhaps several important properties of hydrogen peroxide can be attributed to its capacity to shut down protein tyrosine phosphatases, leading to activation or inhibition of signaling cascades depending on which tyrosine residues are available for the targeted PTP.

#### Induction of systemic immune suppression?

Reviewed above is some of the evidence suggesting that oxidative stress may play a role in tumor-induced immune suppression. The chain of evidence includes the role of hydrogen peroxide producing tumor-infiltrating macrophages/granulocytes, and one may argue that such effects could only operate locally in the tumor environment. As several reports also describe signs of immune suppression in peripheral blood of patients with advanced cancer, the mechanisms behind systemic immune suppression have been sought. In a recent study the existence of activated and H<sub>2</sub>O<sub>2</sub>-secreting granulocytes in PBLs correlated with functional and molecular defects in peripheral T cells of patients with advanced cancer [135]. The release of free radicals by granulocytes may be the principal mechanism for generating a state of persistent oxidative stress in cancer patients. For instance, it has been shown that colorectal cancer patients have lower serum concentrations of free radical scavengers such as vitamin E and C along with increased levels of 8-oxoGua, which serves as a DNA marker for oxidative stress [43]. Excluding the unlikely explanation that universally, patients and normal individuals had different eating habits, it may be concluded that the decreased levels of scavengers resulted from enhanced consumption by the state of oxidative stress present in the patients. One may not cure cancer by altering the



redox status in patients with malignancies. However, it may be very important to relieve patients from oxidative stress in order to make them responsive to immunotherapy with the ultimate goal of curing their disease.

#### Alleviating immune suppression with antioxidants

Epidemiological studies on the effect of dietary factors in prevention of cancer have largely centered on fat and vitamin intake. Evidence indicates that an increased intake of fat and red meat is associated with a higher risk of colorectal, breast, and prostate cancer [12]. In contrast, high consumption of fruits and vegetables is associated with a reduced risk of several cancers including, lung, oral, pancreas, larynx, esophagus, bladder, and stomach. A molecular explanation for this epidemiological observation is lacking, although it is likely that multiple components of the diet are responsible. A role for vitamin E in decreasing the incidence of prostate cancer was suggested in a large primary prevention study that is being followed up in the recently launched Study of Selenium and Vitamin E Cancer Prevention Trial (SELECT) [26, 53]. Vitamin E is a naturally occurring lipid-soluble antioxidant that acts as a scavenger and terminates the cascade of free radical formation induced by toxic and carcinogenic compounds [83]. It is found at high concentrations in membranes of T cells and is essential for normal immune function. Deficiency of vitamin E is associated with increased sensitivity to infection and incidence of tumors [9]. Supplementation of high doses of dietary vitamin E improves the compromised immune function in elderly and in patients with HIV infection [82, 99]. Similar results have been obtained in animal models of aging mice and rats, with evidence of enhanced proliferative capacity and IL-2 production after intake of vitamin E-enriched diets [148]. We recently investigated the possibility of enhancing the immune function in patients with advanced (Dukes' C and D) colorectal cancer by reconstituting their antioxidant defenses [90]. This was tested in a pilot clinical trial in which patients were given high doses of dietary vitamin E along with vitamin C and selenium at the recommended daily intake (RDA) doses for a period of 2 weeks. Vitamin C and selenium were included in the treatment regimen to recycle vitamin E to its reduced state and thus allow optimal function of vitamin E [9]. Colorectal cancer patients provided a suitable study group as several features of immune suppression were previously reported in these patients. For instance, they exhibit a decrease in signaling transduction molecules of peripheral T cells that is associated with severity of disease [94, 105]; there is evidence for reduced CD4 counts leading to altered CD4/CD8 ratios [5]; and Th1 cytokine production is significantly impaired [55]. Interestingly, surgical removal of the tumor burden leads to a normalization of the cytokine production capacity indicating that the immune suppression is specifically induced by the tumor

[55]. In 10 out of 12 treated patients, we observed an increase in the number of T cells capable of producing IL-2 in response to PMA/Ionomycin [90]. The enhanced IL-2 production after vitamin E treatment may be the central mechanism behind the immune-stimulatory effects of vitamin E. IL-2 can reverse T-cell anergy and has a potent capacity to induce T-cell proliferation via both auto and paracrine loops acting at the IL-2 receptor expressed by activated T cells [137]. Decreased CD4/CD8 ratios have been associated with immunological dysfunction in patients with AIDS [109] and different types of cancer, including multiple myeloma [70], colorectal carcinoma [5], and Hodgkin's disease [63]. Increased apoptosis of CD4<sup>+</sup> T cells may also contribute to the transient immunodeficiency that follows allogeneic BMT [84]. The altered CD4/CD8 ratios in multiple myeloma and colorectal carcinoma correlated with the stage of disease, being more reduced in advanced stages [5, 70]. Others have demonstrated that TILs display a decreased CD4/CD8 ratio while the balance of lymphocyte subsets remained unaltered in peripheral blood [133]. Evidence from human and mouse studies, suggests that vitamin E increases CD4 counts [109]. In our study, vitamin E significantly increased the overall CD4/CD8 ratios in colorectal cancer patients. This may be of particular importance for the individuals with CD4 counts and CD4/CD8 ratios much below normal. Indeed, a stratified analysis of the five patients with CD4/CD8 ratios below the reference interval for healthy individuals revealed a significant increase after vitamin E treatment. CD4<sup>+</sup> T helper cells are essential for induction of CD8<sup>+</sup> cytolytic T-lymphocyte immunity due to their ability to drive the maturation of DCs [98]. There is experimental evidence that peptide-based vaccination may become more efficient if tumor antigen-specific CD4<sup>+</sup> T-cell-mediated responses are concurrently induced by including HLA class II tumor peptides in the vaccine [110]. The search for such HLA class II-restricted epitopes within previously known and new tumor-associated antigens has been intensified during the last couple of years [164]. Furthermore, CD4 T cells can have a more direct role in eliminating tumors during the effector phase [58]. For any of these mechanisms to be efficient, one is dependent on the presence of functional CD4<sup>+</sup> T cells. Antioxidants such as vitamin E may set the stage for efficient immunotherapy against cancer by inducing Th1 production and restoring CD4 counts. Future strategies may also involve genetic transfer of antioxidant genes such as catalase into adoptively transferred tumor-specific T cells to allow optimal function during the effector phase at the tumor site.

---

#### Conclusions

Since the description of the first tumor antigens by Pierre Van der Bruggen and Thierry Boon in 1991 [161], we have been facing a new era of tumor immunotherapy.

Along with a tremendous increase in the understanding of basic principles of immunity and novel technologies, we now have the tools to ask fundamental questions of immune-mediated tumor recognition in clinical trials. A better understanding of immune escape mechanisms and the mechanism underlying tumor-induced immune suppression will help in designing novel and more efficient protocols for immunotherapy. However, as for all currently available treatments for cancer, one will have to define the responsive patient groups (types of cancer), the optimal stage of disease, and the ideal combination with other treatment modalities. It is likely that different strategies of immunotherapy will be beneficial under different circumstances. The promising results from several clinical trials of adoptive immunotherapy including transfer of tumor-specific CTLs and the GVL effect of allogeneic BMT [21, 31, 128] indicate that treatment of advanced stages of cancer with large tumor burden may require adoptive transfer of high numbers of tumor-reactive T and NK cells.

**Acknowledgements** This work was supported by grants from the Swedish Foundation for Strategic Research, the Swedish Society for Medical Research and the Swedish Cancer Society.

## References

- Albina JE, Abate JA, Henry WL Jr (1991) Nitric oxide production is required for murine resident peritoneal macrophages to suppress mitogen-stimulated T cell proliferation: role of IFN-gamma in the induction of the nitric oxide-synthesizing pathway. *J Immunol* 147:144
- Allison J, Georgiou HM, Strasser A, Vaux DL (1997) Transgenic expression of CD95 ligand on islet beta cells induces a granulocytic infiltration but does not confer immune privilege upon islet allografts. *Proc Natl Acad Sci U S A* 94:3943
- Aoe T, Okamoto Y, Saito T (1995) Activated macrophages induce structural abnormalities of the T cell receptor-CD3 complex. *J Exp Med* 181:1881
- Arai H, Gordon D, Nabel EG, Nabel GJ (1997) Gene transfer of Fas ligand induces tumor regression in vivo. *Proc Natl Acad Sci U S A* 94:13862
- Arista MC, Callopoli A, De Franceschi L et al (1994) Flow cytometric study of lymphocyte subsets in patients at different stages of colorectal carcinoma. *Dis Colon Rectum* 37:S30
- Asavaroengchai W, Kotera Y, Mule JJ (2002) Tumor lysate-pulsed dendritic cells can elicit an effective antitumor immune response during early lymphoid recovery. *Proc Natl Acad Sci U S A* 99:931
- Babior BM, Kipnes RS, Curnutte JT (1973) Biological defense mechanisms: the production by leukocytes of superoxide, a potential bactericidal agent. *J Clin Invest* 52:741
- Baeuerle PA, Henkel T (1994) Function and activation of NF-kappa B in the immune system. *Annu Rev Immunol* 12:141
- Beharka A, Redican S, Leka L, Meydani SN (1997) Vitamin E status and immune function. *Methods Enzymol* 282:247
- Bendandi M, Gocke CD, Kobrin CB et al (1999) Complete molecular remissions induced by patient-specific vaccination plus granulocyte-monocyte colony-stimulating factor against lymphoma. *Nat Med* 5:1171
- Biaglow JE, Mitchell JB, Held K (1992) The importance of peroxide and superoxide in the X-ray response. *Int J Radiat Oncol Biol Phys* 22:665
- Biesalski HK (2002) Meat and cancer: meat as a component of a healthy diet. *Eur J Clin Nutr* 56[Suppl 1]:S2
- Bingle L, Brown NJ, Lewis CE (2002) The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. *J Pathol* 196:254
- Birkeland ML, Johnson P, Trowbridge IS, Pure E (1989) Changes in CD45 isoform expression accompany antigen-induced murine T-cell activation. *Proc Natl Acad Sci USA* 86:6734
- Blanchetot C, Tertoolen LG, den Hertog J (2002) Regulation of receptor protein-tyrosine phosphatase alpha by oxidative stress. *Embo J* 21:493
- Brentjens RJ, Latouche JB, Santos E et al (2003) Eradication of systemic B-cell tumors by genetically targeted human T lymphocytes co-stimulated by CD80 and interleukin-15. *Nat Med* 9:279
- Burnet FM (1957) Cancer—a biological approach. *BMJ* i:841
- Burnet FM (1970) The concept of immunological surveillance. *Prog Exp Tumor Res* 13:1
- Caselli A, Marzocchini R, Camici G et al (1998) The inactivation mechanism of low molecular weight phosphotyrosine-protein phosphatase by H<sub>2</sub>O<sub>2</sub>. *J Biol Chem* 273:32554
- Cemerski S, van Meerwijk JP, Romagnoli P (2003) Oxidative-stress-induced T lymphocyte hyporesponsiveness is caused by structural modification rather than proteasomal degradation of crucial TCR signaling molecules. *Eur J Immunol* 33:2178
- Childs R, Chernoff A, Contentin N et al (2000) Regression of metastatic renal-cell carcinoma after nonmyeloablative allogeneic peripheral-blood stem-cell transplantation. *N Engl J Med* 343:750
- Clay TM, Custer MC, Sachs J, Hwu P, Rosenberg SA, Nishimura MI (1999) Efficient transfer of a tumor antigen-reactive TCR to human peripheral blood lymphocytes confers anti-tumor reactivity. *J Immunol* 163:507
- Colonna M, Samaridis J, Cella M et al (1998) Human myelomonocytic cells express an inhibitory receptor for classical and nonclassical MHC class I molecules. *J Immunol* 160:3096
- Davidson WF, Giese T, Fredrickson TN (1998) Spontaneous development of plasmacytoid tumors in mice with defective Fas-Fas ligand interactions. *J Exp Med* 187:1825
- De Smedt T, Van Mechelen M, De Becker G, Urbain J, Leo O, Moser M (1997) Effect of interleukin-10 on dendritic cell maturation and function. *Eur J Immunol* 27:1229
- DeFrancesco L (2001) Prostate cancer prevention trial launched. *Nat Med* 7:1076
- Devadas S, Zaritskaya L, Rhee SG, Oberley L, Williams MS (2002) Discrete generation of superoxide and hydrogen peroxide by T cell receptor stimulation: selective regulation of mitogen-activated protein kinase activation and fas ligand expression. *J Exp Med* 195:59
- Dianzani U, Luqman M, Rojo J et al (1990) Molecular associations on the T cell surface correlate with immunological memory. *Eur J Immunol* 20:2249
- Djerbi M, Screpanti V, Catrina AI, Bogen B, Biberfeld P, Grandien A (1999) The inhibitor of death receptor signaling, FLICE-inhibitory protein defines a new class of tumor progression factors. *J Exp Med* 190:1025
- Dornan S, Sebestyen Z, Gamble J et al (2002) Differential association of CD45 isoforms with CD4 and CD8 regulates the actions of specific pools of p56lck tyrosine kinase in T cell antigen receptor signal transduction. *J Biol Chem* 277:1912
- Dudley ME, Wunderlich JR, Robbins PF et al (2002) Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Science* 298:850
- Dummer W, Niethammer AG, Baccala R et al (2002) T cell homeostatic proliferation elicits effective antitumor autoimmunity. *J Clin Invest* 110:185
- Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD (2002) Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol* 3:991

34. Dutoit V, Taub RN, Papadopoulos KP et al (2002) Multi-epitope CD8(+) T cell response to a NY-ESO-1 peptide vaccine results in imprecise tumor targeting. *J Clin Invest* 110:1813
35. Eaton JD, Perry MJ, Nicholson S et al (2002) Allogeneic whole-cell vaccine: a phase I/II study in men with hormone-refractory prostate cancer. *BJU Int* 89:19
36. Edinger M, Hoffmann P, Ermann J et al (2003) CD4(+)CD25(+) regulatory T cells preserve graft-versus-tumor activity while inhibiting graft-versus-host disease after bone marrow transplantation. *Nat Med* 9:1144
37. Ehrlich P (1909) Ueber den jetzigen stand der karzinomforschung. *Ned Tijdschr Geneesk* 5
38. Engleman EG (2003) Dendritic cell-based cancer immunotherapy. *Semin Oncol* 30:23
39. Favre-Felix N, Fromentin A, Hammann A, Solary E, Martin F, Bonnotte B (2000) Cutting edge: the tumor counterattack hypothesis revisited: colon cancer cells do not induce T cell apoptosis via the Fas (CD95, APO-1) pathway. *J Immunol* 164:5023
40. Fontana A, Frei K, Bodmer S et al (1989) Transforming growth factor-beta inhibits the generation of cytotoxic T cells in virus-infected mice. *J Immunol* 143:3230
41. Fournel S, Aguerre-Girr M, Huc X et al (2000) Cutting edge: soluble HLA-G1 triggers CD95/CD95 ligand-mediated apoptosis in activated CD8+ cells by interacting with CD8. *J Immunol* 164:6100
42. Fruh K, Ahn K, Djaballah H et al (1995) A viral inhibitor of peptide transporters for antigen presentation. *Nature* 375:415
43. Gackowski D, Banaszkiwicz Z, Rozalski R, Jawien A, Oliniski R (2002) Persistent oxidative stress in colorectal carcinoma patients. *Int J Cancer* 101:395
44. Garrido F, Festenstein H, Schirmacher V (1976) Further evidence for depression of H-2 and Ia-like specificities of foreign haplotypes in mouse tumour cell lines. *Nature* 261:705
45. Garrido F, Ruiz-Cabello F, Cabrera T et al (1997) Implications for immunosurveillance of altered HLA class I phenotypes in human tumours. *Immunol Today* 18:89
46. Gaschen B, Taylor J, Yusim K et al (2002) Diversity considerations in HIV-1 vaccine selection. *Science* 296:2354
47. Gilboa E (1999) The makings of a tumor rejection antigen. *Immunity* 11:263
48. Gorsch SM, Memoli VA, Stukel TA, Gold LI, Arrick BA (1992) Immunohistochemical staining for transforming growth factor beta 1 associates with disease progression in human breast cancer. *Cancer Res* 52:6949
49. Griffith TS, Brunner T, Fletcher SM, Green DR, Ferguson TA (1995) Fas ligand-induced apoptosis as a mechanism of immune privilege. *Science* 270:1189
50. Groh V, Wu J, Yee C, Spies T (2002) Tumour-derived soluble MIC ligands impair expression of NKG2D and T-cell activation. *Nature* 419:734
51. Gross L (1943) Intradermal immunisation of C3H mice against a sarcoma that originated in an animal of the same line. *Cancer Res* 3:326
52. Hahne M, Rimoldi D, Schroter M et al (1996) Melanoma cell expression of Fas(Apo-1/CD95) ligand: implications for tumor immune escape. *Science* 274:1363
53. Heinonen OP, Albanes D, Virtamo J et al (1998) Prostate cancer and supplementation with alpha-tocopherol and beta-carotene: incidence and mortality in a controlled trial. *J Natl Cancer Inst* 90:440
54. Hellstrand K, Hermodsson S (1986) Histamine H2-receptor-mediated regulation of human natural killer cell activity. *J Immunol* 137:656
55. Heriot AG, Marriott JB, Cookson S, Kumar D, Dalglish AG (2000) Reduction in cytokine production in colorectal cancer patients: association with stage and reversal by resection. *Br J Cancer* 82:1009
56. Hersey P, Zhang XD (2001) How melanoma cells evade trail-induced apoptosis. *Nat Rev Cancer* 1:142
57. Heslop HE, Ng CY, Li C et al (1996) Long-term restoration of immunity against Epstein-Barr virus infection by adoptive transfer of gene-modified virus-specific T lymphocytes. *Nat Med* 2:551
58. Hung K, Hayashi R, Lafond-Walker A, Lowenstein C, Pardoll D, Levitsky H (1998) The central role of CD4(+) T cells in the antitumor immune response. *J Exp Med* 188:2357
59. Ibrahim EC, Guerra N, Lacombe MJ et al (2001) Tumor-specific up-regulation of the nonclassical class I HLA-G antigen expression in renal carcinoma. *Cancer Res* 61:6838
60. Ikeda H, Lethe B, Lehmann F et al (1997) Characterization of an antigen that is recognized on a melanoma showing partial HLA loss by CTL expressing an NK inhibitory receptor. *Immunity* 6:199
61. Jin YJ, Friedman J, Burakoff SJ (1998) Regulation of tyrosine phosphorylation in isolated T cell membrane by inhibition of protein tyrosine phosphatases. *J Immunol* 161:1743
62. Kambayashi T, Michaelsson J, Fahlen L et al (2001) Purified MHC class I molecules inhibit activated NK cells in a cell-free system in vitro. *Eur J Immunol* 31:869
63. Kandil A, Bazarbashi S, Mourad WA (2001) The correlation of Epstein-Barr virus expression and lymphocyte subsets with the clinical presentation of nodular sclerosing Hodgkin disease. *Cancer* 91:1957
64. Kemp RA, Ronchese F (2001) Tumor-specific Tc1, but not Tc2, cells deliver protective antitumor immunity. *J Immunol* 167:6497
65. Khong HT, Restifo NP (2002) Natural selection of tumor variants in the generation of "tumor escape" phenotypes. *Nat Immunol* 3:999
66. Kiessling R, Wasserman K, Horiguchi S et al (1999) Tumor-induced immune dysfunction. *Cancer Immunol Immunother* 48:353
67. Kirchner H, Chused TM, Herberman RB, Holden HT, Lavrin DH (1974) Evidence of suppressor cell activity in spleens of mice bearing primary tumors induced by Moloney sarcoma virus. *J Exp Med* 139:1473
68. Klein G (1973) Immunological surveillance against neoplasia. *Harvey Lect* 69:71-102
69. Koh CY, Blazar BR, George T et al (2001) Augmentation of antitumor effects by NK cell inhibitory receptor blockade in vitro and in vivo. *Blood* 97:3132
70. Koike M, Sekigawa I, Okada M et al (2002) Relationship between CD4(+)/CD8(+) T cell ratio and T cell activation in multiple myeloma: reference to IL-16. *Leuk Res* 26:705
71. Kolenko V, Wang Q, Riedy MC et al (1997) Tumor-induced suppression of T lymphocyte proliferation coincides with inhibition of Jak3 expression and IL-2 receptor signaling: role of soluble products from human renal cell carcinomas. *J Immunol* 159:3057
72. Kono K, Rensing ME, Brandt RM et al (1996) Decreased expression of signal-transducing zeta chain in peripheral T cells and natural killer cells in patients with cervical cancer. *Clin Cancer Res* 2:1825
73. Kono K, Salazar-Onfray F, Petersson M et al (1996) Hydrogen peroxide secreted by tumor-derived macrophages down-modulates signal-transducing zeta molecules and inhibits tumor-specific T cell- and natural killer cell-mediated cytotoxicity. *Eur J Immunol* 26:1308
74. Landowski TH, Qu N, Buyuksal I, Painter JS, Dalton WS (1997) Mutations in the Fas antigen in patients with multiple myeloma. *Blood* 90:4266
75. Lanier LL (1998) NK cell receptors. *Annu Rev Immunol* 16:359
76. Leek RD, Harris AL, Lewis CE (1994) Cytokine networks in solid human tumors: regulation of angiogenesis. *J Leukoc Biol* 56:423
77. Lefebvre S, Antoine M, Uzan S et al (2002) Specific activation of the non-classical class I histocompatibility HLA-G antigen and expression of the ILT2 inhibitory receptor in human breast cancer. *J Pathol* 196:266

78. Lehmann F, Marchand M, Hainaut P et al (1995) Differences in the antigens recognized by cytolytic T cells on two successive metastases of a melanoma patient are consistent with immune selection. *Eur J Immunol* 25:340
79. Leitenberg D, Boutin Y, Lu DD, Bottomly K (1999) Biochemical association of CD45 with the T cell receptor complex: regulation by CD45 isoform and during T cell activation. *Immunity* 10:701
80. Leitenberg D, Novak TJ, Farber D, Smith BR, Bottomly K (1996) The extracellular domain of CD45 controls association with the CD4-T cell receptor complex and the response to antigen-specific stimulation. *J Exp Med* 183:249
81. Levitskaya J, Coram M, Levitsky V et al (1995) Inhibition of antigen processing by the internal repeat region of the Epstein-Barr virus nuclear antigen-1. *Nature* 375:685
82. Liang B, Chung S, Araghiniknam M, Lane LC, Watson RR (1996) Vitamins and immunomodulation in AIDS. *Nutrition* 12:1
83. Liebler DC (1993) The role of metabolism in the antioxidant function of vitamin E. *Crit Rev Toxicol* 23:147
84. Lin MT, Tseng LH, Frangoul H et al (2000) Increased apoptosis of peripheral blood T cells following allogeneic hematopoietic cell transplantation. *Blood* 95:3832
85. Liyanage UK, Moore TT, Joo HG et al (2002) Prevalence of regulatory T cells is increased in peripheral blood and tumor microenvironment of patients with pancreas or breast adenocarcinoma. *J Immunol* 169:2756
86. Ljunggren HG, Karre K (1990) In search of the 'missing self': MHC molecules and NK cell recognition [see comments]. *Immunol Today* 11:237
87. Loercher AE, Nash MA, Kavanagh JJ, Platsoucas CD, Freedman RS (1999) Identification of an IL-10-producing HLA-DR-negative monocyte subset in the malignant ascites of patients with ovarian carcinoma that inhibits cytokine protein expression and proliferation of autologous T cells. *J Immunol* 163:6251
88. Long EO, Rajagopalan S (2002) Stress signals activate natural killer cells. *J Exp Med* 196:1399
89. Malmberg KJ, Arulampalam V, Ichihara F et al (2001) Inhibition of activated/memory (CD45RO(+)) T cells by oxidative stress associated with block of NF-kappaB activation. *J Immunol* 167:2595
90. Malmberg KJ, Lenkei R, Petersson M et al (2002) A short-term dietary supplementation of high doses of vitamin E increases T helper 1 cytokine production in patients with advanced colorectal cancer. *Clin Cancer Res* 8:1772
91. Malmberg KJ, Levitsky V, Norell H et al (2002) IFN-gamma protects short-term ovarian carcinoma cell lines from CTL lysis via a CD94/NKG2A-dependent mechanism. *J Clin Invest* 110:1515
92. Marincola FM, Shamamian P, Simonis TB et al (1994) Locus-specific analysis of human leukocyte antigen class I expression in melanoma cell lines. *J Immunother Emph Tumor Immunol* 16:13
93. Marrack P, Bender J, Hildeman D et al (2000) Homeostasis of alpha beta TCR + T cells. *Nat Immunol* 1:107
94. Matsuda M, Petersson M, Lenkei R et al (1995) Alterations in the signal-transducing molecules of T cells and NK cells in colorectal tumor-infiltrating, gut mucosal and peripheral lymphocytes: correlation with the stage of the disease. *Int J Cancer* 61:765
95. Medema JP, de Jong J, Peltenburg LT et al (2001) Blockade of the granzyme B/perforin pathway through overexpression of the serine protease inhibitor PI-9/SPI-6 constitutes a mechanism for immune escape by tumors. *Proc Natl Acad Sci U S A* 98:11515
96. Medema JP, de Jong J, van Hall T, Melief CJ, Offringa R (1999) Immune escape of tumors in vivo by expression of cellular FLICE-inhibitory protein. *J Exp Med* 190:1033
97. Medema JP, Schuurhuis DH, Rea D et al (2001) Expression of the serpin serine protease inhibitor 6 protects dendritic cells from cytotoxic T lymphocyte-induced apoptosis: differential modulation by T helper type 1 and type 2 cells. *J Exp Med* 194:657
98. Melief CJ, Van Der Burg SH, Toes RE, Ossendorp F, Offringa R (2002) Effective therapeutic anticancer vaccines based on precision guiding of cytolytic T lymphocytes. *Immunol Rev* 188:177
99. Meydani SN, Meydani M, Blumberg JB et al (1997) Vitamin E supplementation and in vivo immune response in healthy elderly subjects: a randomized controlled trial. *JAMA* 277:1380
100. Michaelsson J, Teixeira De Matos C, Achour A, Lanier LL, Karre K, Soderstrom K (2002) A signal peptide derived from hsp60 binds HLA-E and interferes with CD94/NKG2A recognition. *J Exp Med* 196:1403
101. Mizoguchi H, O'Shea JJ, Longo DL, Loeffler CM, McVicar DW, Ochoa AC (1992) Alterations in signal transduction molecules in T lymphocytes from tumor-bearing mice. *Science* 258:1795
102. Mizutani H, Tada-Oikawa S, Hiraku Y, Oikawa S, Kojima M, Kawanishi S (2002) Mechanism of apoptosis induced by a new topoisomerase inhibitor through the generation of hydrogen peroxide. *J Biol Chem* 277:30684
103. Moore KW, Vieira P, Fiorentino DF, Trounstein ML, Khan TA, Mosmann TR (1990) Homology of cytokine synthesis inhibitory factor (IL-10) to the Epstein-Barr virus gene BCRF1. *Science* 248:1230
104. Morgan DJ, Kreuwel HT, Fleck S, Levitsky HI, Pardoll DM, Sherman LA (1998) Activation of low avidity CTL specific for a self epitope results in tumor rejection but not autoimmunity. *J Immunol* 160:643
105. Mulder WM, Bloemena E, Stukart MJ, Kummer JA, Wagstaff J, Scheper RJ (1997) T cell receptor-zeta and granzyme B expression in mononuclear cell infiltrates in normal colon mucosa and colon carcinoma. *Gut* 40:113
106. Naito Y, Saito K, Shiiba K et al (1998) CD8+ T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. *Cancer Res* 58:3491
107. Nestle FO, Alijagic S, Gilliet M et al (1998) Vaccination of melanoma patients with peptide- or tumor lysate-pulsed dendritic cells. *Nat Med* 4:328
108. Novak TJ, Farber D, Leitenberg D, Hong SC, Johnson P, Bottomly K (1994) Isoforms of the transmembrane tyrosine phosphatase CD45 differentially affect T cell recognition. *Immunity* 1:109
109. Odeleye OE, Watson RR (1991) The potential role of vitamin E in the treatment of immunologic abnormalities during acquired immune deficiency syndrome. *Prog Food Nutr Sci* 15:1
110. Ossendorp F, Mengede E, Camps M, Filius R, Melief CJ (1998) Specific T helper cell requirement for optimal induction of cytotoxic T lymphocytes against major histocompatibility complex class II negative tumors. *J Exp Med* 187:693
111. Paul P, Cabestre FA, Le Gal FA et al (1999) Heterogeneity of HLA-G gene transcription and protein expression in malignant melanoma biopsies. *Cancer Res* 59:1954
112. Pawelec G, Barnett Y, Forsey R et al (2002) T cells and aging, January 2002 update. *Front Biosci* 7:d1056
113. Pawelec G, Heinzel S, Kiessling R, Muller L, Ouyang Q, Zeuthen J (2000) Escape mechanisms in tumor immunity: a year 2000 update. *Crit Rev Oncog* 11:97
114. Pericle F, Kirken RA, Bronte V, Sconocchia G, DaSilva L, Segal DM (1997) Immunocompromised tumor-bearing mice show a selective loss of STAT5a/b expression in T and B lymphocytes. *J Immunol* 159:2580
115. Piccirillo CA, Shevach EM (2001) Cutting edge: control of CD8+ T cell activation by CD4+ CD25+ immunoregulatory cells. *J Immunol* 167:1137

116. Pisa P, Halapi E, Pisa EK et al (1992) Selective expression of interleukin 10, interferon gamma, and granulocyte-macrophage colony-stimulating factor in ovarian cancer biopsies. *Proc Natl Acad Sci U S A* 89:7708
117. Rabinowich H, Reichert TE, Kashii Y, Gastman BR, Bell MC, Whiteside TL (1998) Lymphocyte apoptosis induced by Fas ligand- expressing ovarian carcinoma cells. Implications for altered expression of T cell receptor in tumor-associated lymphocytes. *J Clin Invest* 101:2579
118. Ramal LM, Maleno I, Cabrera T et al (2000) Molecular strategies to define HLA haplotype loss in microdissected tumor cells. *Hum Immunol* 61:1001
119. Restifo NP (2000) Not so Fas: re-evaluating the mechanisms of immune privilege and tumor escape. *Nat Med* 6:493
120. Restifo NP, Marincola FM, Kawakami Y, Taubenberger J, Yannelli JR, Rosenberg SA (1996) Loss of functional beta 2-microglobulin in metastatic melanomas from five patients receiving immunotherapy. *J Natl Cancer Inst* 88:100
121. Reth M (2002) Hydrogen peroxide as second messenger in lymphocyte activation. *Nat Immunol* 3:1129
122. Roncarolo MG, Bacchetta R, Bordignon C, Narula S, Levings MK (2001) Type 1 T regulatory cells. *Immunol Rev* 182:68
123. Rosenberg SA (1999) A new era for cancer immunotherapy based on the genes that encode cancer antigens. *Immunity* 10:281
124. Rosenberg SA, Yang JC, Schwartzentruber DJ et al (1998) Immunologic and therapeutic evaluation of a synthetic peptide vaccine for the treatment of patients with metastatic melanoma. *Nat Med* 4:321
125. Rosenberg SA, Yang JC, Topalian SL et al (1994) Treatment of 283 consecutive patients with metastatic melanoma or renal cell cancer using high-dose bolus interleukin 2. *JAMA* 271:907
126. Rosenberg SA, Yang JC, White DE, Steinberg SM (1998) Durability of complete responses in patients with metastatic cancer treated with high-dose interleukin-2: identification of the antigens mediating response. *Ann Surg* 228:307
127. Rossig C, Bollard CM, Nuchtern JG, Rooney CM, Brenner MK (2002) Epstein-Barr virus-specific human T lymphocytes expressing antitumor chimeric T-cell receptors: potential for improved immunotherapy. *Blood* 99:2009
128. Ruggeri L, Capanni M, Urbani E et al (2002) Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. *Science* 295:2097
129. Rygaard J, Povlsen CO (1974) The mouse mutant nude does not develop spontaneous tumours: an argument against immunological surveillance. *Acta Pathol Microbiol Scand [B] Microbiol Immunol* 82:99
130. Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M (1995) Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol* 155:1151
131. Salazar-Onfray F, Charo J, Petersson M et al (1997) Down-regulation of the expression and function of the transporter associated with antigen processing in murine tumor cell lines expressing IL-10. *J Immunol* 159:3195
132. Salvadori S, Martinelli G, Zier K (2000) Resection of solid tumors reverses T cell defects and restores protective immunity. *J Immunol* 164:2214
133. Santin AD, Hermonat PL, Ravaggi A et al (2001) Phenotypic and functional analysis of tumor-infiltrating lymphocytes compared with tumor-associated lymphocytes from ascitic fluid and peripheral blood lymphocytes in patients with advanced ovarian cancer. *Gynecol Obstet Invest* 51:254
134. Schmidt KN, Amstad P, Cerutti P, Baeuerle PA (1996) Identification of hydrogen peroxide as the relevant messenger in the activation pathway of transcription factor NF-kappa B. *Adv Exp Med Biol* 387:63
135. Schmielau J, Finn OJ (2001) Activated granulocytes and granulocyte-derived hydrogen peroxide are the underlying mechanism of suppression of t-cell function in advanced cancer patients. *Cancer Res* 61:4756
136. Schreck R, Rieber P, Baeuerle PA (1991) Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1. *Embo J* 10:2247
137. Schwartz RH (2003) T cell anergy. *Annu Rev Immunol* 21:305-334
138. Seaman WE, Gindhart TD, Blackman MA, Dalal B, Talal N, Werb Z (1981) Natural killing of tumor cells by human peripheral blood cells: suppression of killing in vitro by tumor-promoting phorbol diesters. *J Clin Invest* 67:1324
139. Seliger B, Maeurer MJ, Ferrone S (1997) TAP off—tumors on. *Immunol Today* 18:292
140. Seliger B, Maeurer MJ, Ferrone S (2000) Antigen-processing machinery breakdown and tumor growth. *Immunol Today* 21:455
141. Shimizu J, Yamazaki S, Sakaguchi S (1999) Induction of tumor immunity by removing CD25+CD4+ T cells: a common basis between tumor immunity and autoimmunity. *J Immunol* 163:5211
142. Shin MS, Park WS, Kim SY et al (1999) Alterations of Fas (Apo-1/CD95) gene in cutaneous malignant melanoma. *Am J Pathol* 154:1785
143. Shlomchik WD, Couzens MS, Tang CB et al (1999) Prevention of graft versus host disease by inactivation of host antigen-presenting cells. *Science* 285:412
144. Spaggiari GM, Contini P, Dondero A et al (2002) Soluble HLA class I induces NK cell apoptosis upon the engagement of killer-activating HLA class I receptors through FasL-Fas interaction. *Blood* 100:4098
145. Stutman O (1974) Tumor development after 3-methylcholanthrene in immunologically deficient athymic-nude mice. *Science* 183:534
146. Szatrowski TP, Nathan CF (1991) Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res* 51:794
147. Takeda K, Hayakawa Y, Smyth MJ et al (2001) Involvement of tumor necrosis factor-related apoptosis-inducing ligand in surveillance of tumor metastasis by liver natural killer cells. *Nat Med* 7:94
148. Tanaka J, Fujiwara H, Torisu M (1979) Vitamin E and immune response, I: enhancement of helper T cell activity by dietary supplementation of vitamin E in mice. *Immunology* 38:727
149. Teshima T, Mach N, Hill GR et al (2001) Tumor cell vaccine elicits potent antitumor immunity after allogeneic T-cell-depleted bone marrow transplantation. *Cancer Res* 61:162
150. Thomas L (1959) Cellular and humoral aspects of the hypersensitive states. Hoeber-Harper, New York
151. Thurner B, Haendle I, Roder C et al (1999) Vaccination with mage-3A1 peptide-pulsed mature, monocyte-derived dendritic cells expands specific cytotoxic T cells and induces regression of some metastases in advanced stage IV melanoma. *J Exp Med* 190:1669
152. Timmerman JM, Singh G, Hermanson G et al (2002) Immunogenicity of a plasmid DNA vaccine encoding chimeric idiotypic in patients with B-cell lymphoma. *Cancer Res* 62:5845
153. Toes RE, Hoeben RC, van der Voort EI et al (1997) Protective anti-tumor immunity induced by vaccination with recombinant adenoviruses encoding multiple tumor-associated cytotoxic T lymphocyte epitopes in a string-of-beads fashion. *Proc Natl Acad Sci U S A* 94:14660
154. Trapani JA, Sutton VR, Smyth MJ (1999) CTL granules: evolution of vesicles essential for combating virus infections. *Immunol Today* 20:351

155. Trowbridge IS, Thomas ML (1994) CD45: an emerging role as a protein tyrosine phosphatase required for lymphocyte activation and development. *Annu Rev Immunol* 12:85
156. Turley SJ (2002) Dendritic cells: inciting and inhibiting autoimmunity. *Curr Opin Immunol* 14:765
157. Urban JL, Holland JM, Kripke ML, Schreiber H (1982) Immunoselection of tumor cell variants by mice suppressed with ultraviolet radiation. *J Exp Med* 156:1025
158. Uyttenhove C, Van Snick J, Boon T (1980) Immunogenic variants obtained by mutagenesis of mouse mastocytoma P815, I: rejection by syngeneic mice. *J Exp Med* 152:1175
159. Uzzo RG, Clark PE, Rayman P et al (1999) Alterations in NFkappaB activation in T lymphocytes of patients with renal cell carcinoma. *J Natl Cancer Inst* 91:718
160. Vales-Gomez M, Reyburn HT, Erskine RA, Lopez-Botet M, Strominger JL (1999) Kinetics and peptide dependency of the binding of the inhibitory NK receptor CD94/NKG2-A and the activating receptor CD94/NKG2-C to HLA-E. *Embo J* 18:4250
161. Van der Bruggen P, Traversari C, Chomez P et al (1991) A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. *Science* 254:1643
162. Van Pel A, Boon T (1982) Protection against a nonimmunogenic mouse leukemia by an immunogenic variant obtained by mutagenesis. *Proc Natl Acad Sci U S A* 79:4718
163. Vaux DL (1995) Immunology: ways around rejection. *Nature* 377:576
164. Wang RF (2002) Enhancing antitumor immune responses: intracellular peptide delivery and identification of MHC class II-restricted tumor antigens. *Immunol Rev* 188:65
165. Woo EY, Chu CS, Goletz TJ et al (2001) Regulatory CD4(+)CD25(+) T cells in tumors from patients with early-stage non-small cell lung cancer and late-stage ovarian cancer. *Cancer Res* 61:4766
166. Xu Z, Weiss A (2002) Negative regulation of CD45 by differential homodimerization of the alternatively spliced isoforms. *Nat Immunol* 3:764
167. Yamagishi H, Pellis NR, Macek C, Kahan BD (1980) Changes in spleen morphology and lymphoid cell activity during tumor progression. *Eur J Cancer* 16:1417
168. Yee C, Thompson JA, Byrd D et al (2002) Adoptive T cell therapy using antigen-specific CD8+ T cell clones for the treatment of patients with metastatic melanoma: in vivo persistence, migration, and antitumor effect of transferred T cells. *Proc Natl Acad Sci U S A* 99:16168
169. Yewdell JW, Hill AB (2002) Viral interference with antigen presentation. *Nat Immunol* 3:1019
170. Zaks TZ, Chappell DB, Rosenberg SA, Restifo NP (1999) Fas-mediated suicide of tumor-reactive T cells following activation by specific tumor: selective rescue by caspase inhibition. *J Immunol* 162:3273
171. Zinkernagel RM (2001) Immunity against solid tumors? *Int J Cancer* 93:1