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# Strengthening the Evidence for a Causal Link between Type 2 Diabetes Mellitus and Pancreatic Cancer: Insights from Two-Sample and Multivariable Mendelian Randomization

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**Abstract:** This two-sample Mendelian randomization (MR) study was conducted to investigate the causal associations between type 2 diabetes mellitus (T2DM) and the risk of pancreatic cancer (PaCa), as this causal relationship remains inconclusive in existing MR studies. The selection of instrumental variables for T2DM was based on two genome-wide association study (GWAS) meta-analyses from European cohorts. Summary-level data for PaCa were extracted from the FinnGen and UK Biobank databases. Inverse variance weighted (IVW) and four other robust methods were employed in our MR analysis. Various sensitivity analyses and multivariable MR approaches were also performed to enhance the robustness of our findings. In the IVW and Mendelian Randomization Pleiotropy RESidual Sum and Outlier (MR-PRESSO) analyses, the odds ratios (ORs) for each 1-unit increase in genetically predicted log odds of T2DM were approximately 1.13 for PaCa. The sensitivity tests and multivariable MR supported the causal link between T2DM and PaCa without pleiotropic effects. Therefore, our analyses suggest a causal relationship between T2DM and PaCa, shedding light on the potential pathophysiological mechanisms of T2DM's impact on PaCa. This finding underscores the importance of T2DM prevention as a strategy to reduce the risk of PaCa.

**Keywords:** two-sample Mendelian randomization; type 2 diabetes mellitus; pancreatic cancer; FinnGen; UK Biobank



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## 1. Introduction

Pancreatic cancer (PaCa) ranks as the 12th most common cancer and is the 6th leading cause of cancer-related mortality worldwide [1,2]. Although there have been advancements in the treatment of PaCa recently, the 5-year survival rate remains low, at approximately 9% in 2018 [3]. The low survival rate of PaCa can largely be attributed to the majority of diagnoses occurring at locally advanced stages or once metastasis has happened. Diagnosing PaCa at an early stage is challenging, especially in patients who exhibit no specific symptoms. According to Global Cancer Observatory (GLOBOCAN) estimations [1,4], a trend towards an increase in PaCa incidence (+78.3% with 399,571 new cases) and mortality (+81.9% with 382,260 deaths) is projected from 2022 to 2045. Due to the high incidence rate and persistently low survival rate of PaCa, compounded by the absence of a screening program for PaCa, preventing PaCa by the noted risk factors becomes crucial.

Previous literature reviews [3,5–9] have summarized numerous potential risk factors for PaCa, including cigarette smoking, heavy alcohol consumption, obesity, chronic pancreatitis, type 2 diabetes mellitus (T2DM), hepatitis B, cholecystectomy, periodontal disease, aging, male gender, African American ethnicity, non-O blood type, family history, certain hereditary syndromes, and germline mutation. Among these risk factors, T2DM stands out as a growing global health crisis, with an increasingly higher prevalence and disease burden being reported [10–12]. In 2017, the global prevalence rate of T2DM was

6059 cases per 100,000 individuals [10]. In our previous UK Biobank cohort study [13,14], a history of diabetes mellitus (DM) was found to at least double the risk of PaCa compared to participants without a DM history [13,14]. Additionally, by calculating the population attributable fraction (PAF), it was revealed that 6% of PaCa cases could be eliminated by avoiding DM [13] in the UK Biobank cohort. Notably, T2DM represents around 90% to 95% of all identified DM cases [15]. Therefore, the ongoing rise in T2DM prevalence worldwide is expected to contribute to an increase in the incidence rate of PaCa.

Despite numerous observational studies [13,15–18] revealing an association between T2DM and an increased risk of PaCa, the causal relationship between T2DM and PaCa remains controversial in Mendelian randomization (MR) studies. MR studies are important for investigating causal relationships, and T2DM was suggested to be causally associated with PaCa in some two-sample Mendelian randomization (2SMR) studies [19,20]. However, other 2SMR studies [21,22] did not observe a causal relationship between T2DM and the risk of PaCa. The null results may be caused by the limited number of instrumental variables (IVs), which could attenuate the statistical power. On the other hand, in studies observing causal relationships, the potential for overestimation still needs to be considered, as some included SNPs may exert pleiotropic effects on BMI, and some results may be influenced by linkage disequilibrium (LD). Therefore, in this study, we attempt to use comprehensive IVs, avoid SNPs located near genes reported to be associated with BMI, and also check for LD in the IV selection process to enhance the robustness of the results.

MR is one of the epidemiological approaches that adopt genetic variants as IVs to explore causal relationships between exposures and outcomes [23–25]. The advantage of MR over traditional observational studies lies in the genetic variants randomly assorted at conception [23–25]; however, there are three critical assumptions [26]. These include IVs that are robustly associated with the exposure (relevance assumption [26]), are not linked with any confounders (independence assumption [26]), and influence the outcome solely through the exposure (exclusion restriction assumption [26]). Consequently, MR has the capability to minimize confounding and reduce bias from reverse causation, offering insights comparable to those obtained from randomized controlled trials [23–25]. 2SMR is a variation in the MR approach [27,28]. Unlike traditional MR studies, which often require individual-level data on both genetic variants related to the exposure and outcome traits from the same participant dataset, 2SMR can utilize summary-level data from separate genome-wide association studies (GWAS) for both the exposure and the outcome [27,28].

To further explore the underlying mechanisms of PaCa and to elevate awareness of prevention education in public health strategies, understanding the causal relationship between T2DM and PaCa is important. The consistency regarding whether the genetic liability to T2DM is causally related to PaCa remains absent in previous studies [19–22]. Consequently, we conducted a 2SMR study to comprehensively utilize the most up-to-date GWAS summary data and two large population-based datasets, including FinnGen and UK Biobank, to investigate the associations of genetic liability to T2DM with PaCa.

## 2. Results

### 2.1. Selection of Instrumental Variables

According to the IV selection criteria mentioned in Section 4.2, 414, 423, and 423 SNPs were chosen from the FinnGen, UKBB, and combined FinnGen and UKBB GWAS summary data in the comprehensive model analysis. The details of all selected IVs are provided in Supplementary Tables S1–S3. In this study, the proportion of variance explained (PVE) or  $R^2$  by IVs used in the FinnGen, UKBB, and combined FinnGen and UKBB studies were 39.16%, 40.18%, and 40.31%, respectively. The F-statistics for these IVs were 933.97, 958.92, and 961.29. Two SNPs near the *FTO* gene (rs78020297 and rs1421085) were removed in the restricted model analysis.

## 2.2. MR Analysis

In our 2SMR analysis, the inverse variance weighted (IVW) method was designated as the principal method due to its higher statistical efficacy. Additionally, four robust methods, including the Mendelian randomization-Egger (MR-Egger), the weighted median (WM), the weighted mode (WMO), and the Mendelian Randomization Pleiotropy RESidual Sum and Outlier (MR-PRESSO), were employed as complementary approaches to evaluate the genetic causal associations between T2DM and PaCa risk.

In the FinnGen dataset, a causal association between T2DM and PaCa risk was indicated by both the IVW ( $p = 0.033$ ) and MR-PRESSO ( $p = 0.029$ ) methods (Table 1). For a one-unit increase in the log-transformed odds of T2DM, the OR of PaCa risk was estimated at 1.102 (95% CI = 1.008–1.204) by the IVW method and 1.097 (95% CI = 1.010–1.191) by the MR-PRESSO method, respectively (Table 1). The MR-PRESSO global test identified two outlier SNPs; consequently, these outliers were corrected in our MR-PRESSO analysis. After eliminating two SNPs near the *FTO* gene in the restricted model, the causal link between T2DM and PaCa risk remained significant in both the IVW ( $p = 0.046$ ) and MR-PRESSO ( $p = 0.031$ ) methods (Table 1) in the FinnGen dataset. With each unit increment in the log-transformed odds of T2DM, the OR for PaCa risk was 1.095 (95% CI = 1.001–1.198) and 1.094 (95% CI = 1.008–1.187) using the IVW method and MR-PRESSO method, respectively. Five outlier SNPs were detected in the MR-PRESSO global test; therefore, these outliers were removed in our MR-PRESSO analysis.

Within the UKBB dataset, the WM ( $p = 0.022$ ), IVW ( $p = 0.001$ ), and MR-PRESSO ( $p = 0.005$ ) methods revealed a causal relationship between T2DM and PaCa risk (Table 1). These methods indicated a 23.7% increase (OR = 1.237, 95% CI = 1.031–1.482) for WM, an 18.5% increase in the odds of PaCa risk (OR = 1.185, 95% CI = 1.068–1.315) for IVW, and a 16.2% increase (OR = 1.162, 95% CI = 1.048–1.288) for MR-PRESSO, per one-unit increase in the log-transformed odds of T2DM (Table 1). After eliminating two SNPs near the *FTO* gene in the restricted model, the WM ( $p = 0.041$ ), IVW ( $p = 0.002$ ), and MR-PRESSO ( $p = 0.007$ ) methods persistently demonstrated a causal link between T2DM and PaCa risk, as shown in Table 1. A one-unit rise in the log-transformed odds of T2DM correlated with a 23% increase (OR = 1.23, 95% CI = 1.008–1.501) in the WM method, an 18% elevation in PaCa risk odds (OR = 1.179, 95% CI = 1.061–1.310) in the IVW method, and a 15.6% rise (OR = 1.156, 95% CI = 1.042–1.284) in the MR-PRESSO method (Table 1).

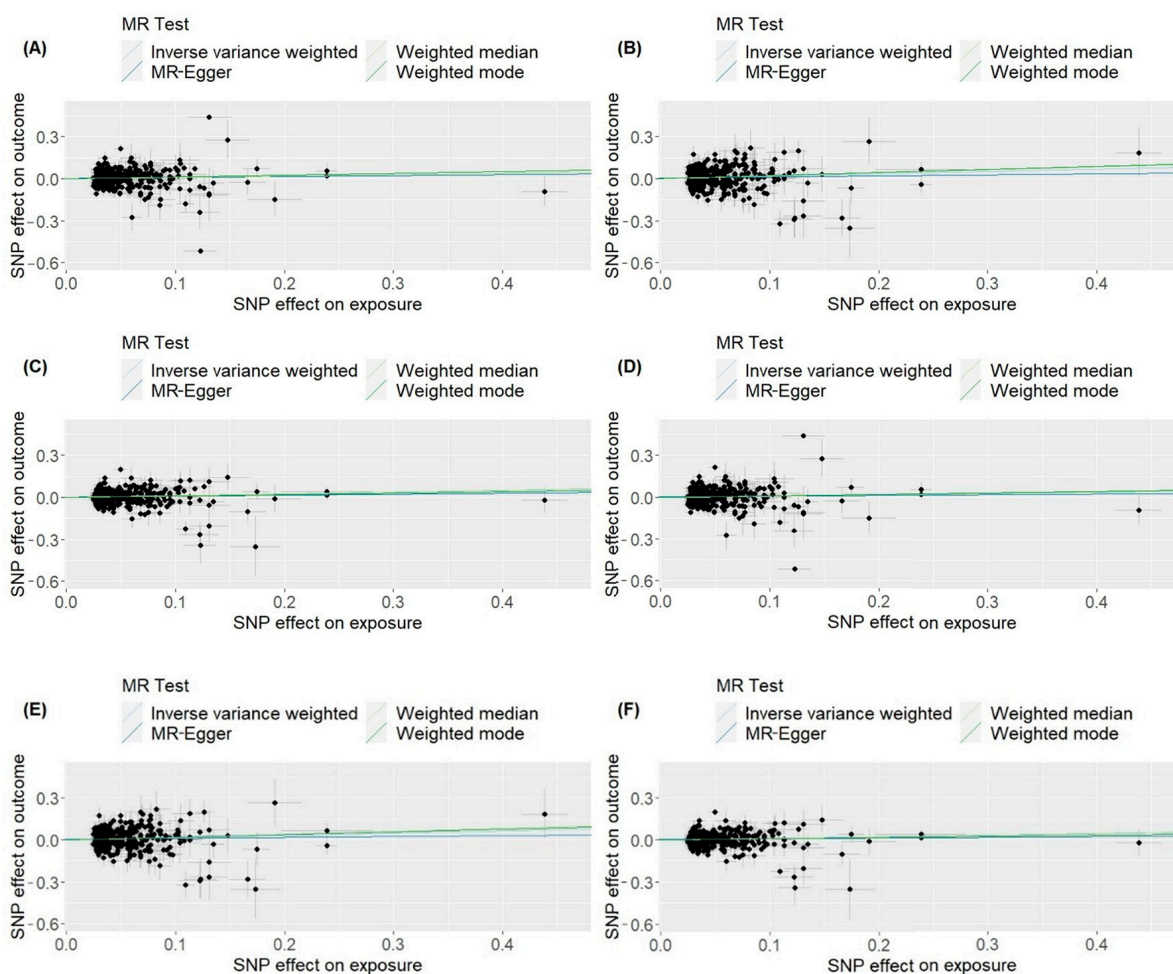
For the combined FinnGen and UKBB dataset, the causal effect of T2DM on PaCa risk was demonstrated significantly using the WM ( $p = 0.017$ ), IVW ( $p = 0.001$ ), and MR-PRESSO ( $p < 0.001$ ) approaches (Table 1). With a one-unit increase in the log-transformed odds of T2DM, the OR of PaCa risk was elevated by 15.1% (OR = 1.151, 95% CI = 1.025–1.293) in the WM approach, 13.1% (OR = 1.131, 95% CI = 1.052–1.216) in the IVW method, and 12.7% (OR = 1.127, 95% CI = 1.056–1.204) in the MR-PRESSO method (Table 1). Two outlier SNPs were detected in the MR-PRESSO global test; therefore, these outliers were removed in our MR-PRESSO analysis. After the two SNPs were removed in the restricted model, the IVW ( $p = 0.002$ ) and MR-PRESSO ( $p = 0.001$ ) methods still significantly highlighted the causal effect of T2DM on PaCa risk, as shown in Table 1. For each unit increase in the log-transformed odds of T2DM, there was a 12.5% increase in PaCa risk odds (OR = 1.125, 95% CI = 1.046–1.211) according to the IVW method, and a 12.4% rise (OR = 1.124, 95% CI = 1.053–1.201) using the MR-PRESSO method. The MR-PRESSO global test detected five outlier SNPs; hence, these outliers were eliminated in our MR-PRESSO analysis.

**Table 1.** The two-sample Mendelian randomization (MR) analysis results from the comprehensive and restricted models.

Two-Sample Mendelian Randomization Analysis in the Comprehensive Model						
PaCa Outcome Source	MR Method	nsnp	OR	p-Value	Forest Plot	95% CI
FinnGen	MR-Egger	414	1.072	0.461		0.891–1.291
	WM	414	1.100	0.230		0.942–1.285
	IVW	414	1.102	<b>0.033</b>		1.008–1.204
	WMO	414	1.135	0.274		0.905–1.425
	MR-PRESSO after outlier-corrected	412	1.097	<b>0.029</b>		1.010–1.191
UKBB	MR-Egger	423	1.079	0.498	0.866–1.345	
	WM	423	1.237	<b>0.022</b>	1.031–1.482	
	IVW	423	1.185	<b>0.001</b>	1.068–1.315	
	WMO	423	1.241	0.114	0.950–1.620	
	MR-PRESSO	423	1.162	<b>0.005</b>	1.048–1.288	
FinnGen+UKBB	MR-Egger	423	1.074	0.359	0.922–1.250	
	WM	423	1.151	<b>0.017</b>	1.025–1.293	
	IVW	423	1.131	<b>0.001</b>	1.052–1.216	
	WMO	423	1.108	0.202	0.947–1.298	
	MR-PRESSO after outlier-corrected	421	1.127	<b>&lt;0.001</b>	1.056–1.204	
Two-sample Mendelian randomization analysis in the restricted model						
PaCa outcome source	MR Method	nsnp	OR	p-val	Forest plot	95% CI
FinnGen	MR-Egger	412	1.053	0.593		0.872–1.270
	WM	412	1.100	0.242		0.938–1.290
	IVW	412	1.095	<b>0.046</b>		1.001–1.198
	WMO	412	1.112	0.317		0.903–1.369
	MR-PRESSO after outlier-corrected	407	1.094	<b>0.031</b>		1.008–1.187
UKBB	MR-Egger	421	1.064	0.589	0.850–1.330	
	WM	421	1.230	<b>0.041</b>	1.008–1.501	
	IVW	421	1.179	<b>0.002</b>	1.061–1.310	
	WMO	421	1.213	0.163	0.925–1.589	
	MR- PRESSO	421	1.156	<b>0.007</b>	1.042–1.284	
FinnGen+UKBB	MR-Egger	421	1.056	0.491	0.905–1.232	
	WM	421	1.122	0.063	0.994–1.266	
	IVW	421	1.125	<b>0.002</b>	1.046–1.211	
	WMO	421	1.108	0.183	0.953–1.287	
	MR-PRESSO after outlier-corrected	416	1.124	<b>0.001</b>	1.053–1.201	

PaCa: pancreatic cancer; MR: Mendelian randomization; nsnp: number of single nucleotide polymorphisms; OR: odds ratio; 95% CI: 95% confidence interval; UKBB: UK Biobank; MR-Egger: Mendelian randomization-Egger; WM: weighted median; IVW: inverse variance weighted; WMO: weighted mode; MR-PRESSO: Mendelian Randomization Pleiotropy RESidual Sum and Outlier. A bold font indicates statistical significance—a *p*-value < 0.05.

In the scatter plot (Figure 1), the direction of the causal effect of T2DM on PaCa risk was consistently depicted across all MR analysis approaches in the FinnGen, UKBB, and combined FinnGen and UKBB datasets, both in the comprehensive and restricted models.

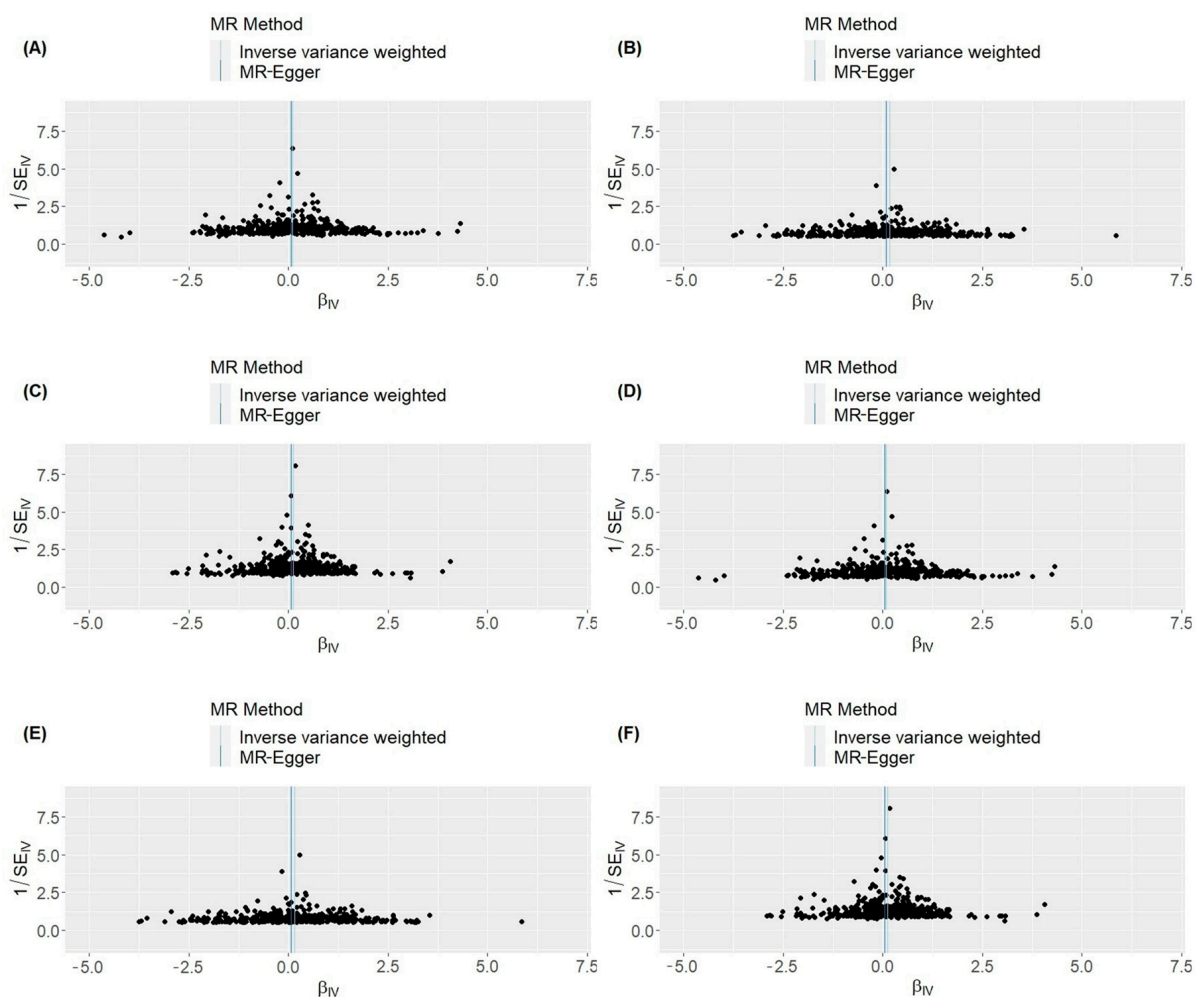


**Figure 1.** Scatter plots for two-sample Mendelian randomization analyses of the causal effect of type 2 diabetes mellitus on pancreatic cancer. (A) Comprehensive model of the FinnGen dataset. (B) Comprehensive model of the UK Biobank dataset. (C) Comprehensive model of the FinnGen and UK Biobank combined dataset. (D) Restricted model of the FinnGen dataset. (E) Restricted model of the UK Biobank dataset. (F) Restricted model of the FinnGen and UK Biobank combined dataset. MR: Mendelian randomization; SNP: single nucleotide polymorphism; IVW: inverse variance weighted; MR-Egger: Mendelian randomization-Egger; WM: weighted median; WMO: weighted mode.

### 2.3. Sensitivity Analysis

The MR-Egger regression intercept analysis revealed no horizontal pleiotropy in the FinnGen, UKBB, and combined FinnGen and UKBB datasets, both in the comprehensive and restricted models (Supplementary Table S4). Furthermore, in our funnel plot visualization (Figure 2), general symmetry suggests the absence of horizontal pleiotropy. In Cochran's Q test for IVW, heterogeneity ( $p < 0.05$ ) was observed in the analyses of the FinnGen and the combined FinnGen and UKBB datasets (Supplementary Table S4). According to the leave-one-out analysis, the sequential removal of each IV did not impact the causal relationship between T2DM and PaCa, nor did it affect the OR. Furthermore, the leave-one-out test did not reveal any potential outliers or evidence of horizontal pleiotropy. The details of the leave-one-out test are listed in Supplementary Tables S5–S10. In the MR

Steiger directionality test, the variance explained in the outcome is less than that in the exposure, confirming the correct causal direction to be true (Supplementary Table S11).



**Figure 2.** Funnel plot for two-sample Mendelian randomization analyses of the causal effect of type 2 diabetes mellitus on pancreatic cancer. (A) Comprehensive model of the FinnGen dataset. (B) Comprehensive model of the UK Biobank dataset. (C) Comprehensive model of the FinnGen and UK Biobank combined dataset. (D) Restricted model of the FinnGen dataset. (E) Restricted model of the UK Biobank dataset. (F) Restricted model of the FinnGen and UK Biobank combined dataset. MR: Mendelian randomization;  $\beta_{IV}$ : beta coefficient of each instrumental variable, which indicates the estimated effect of each SNP on the exposure variable;  $1/SE_{IV}$ : the inverse of the standard error of the  $\beta_{IV}$  estimation, which indicates the precision or uncertainty of these estimates.

#### 2.4. Multivariable Mendelian Randomization (MR)

In the multivariable MR analysis, the observed association between T2DM and PaCa remained significant after adjusting for body mass index (BMI) and waist circumference (OR = 1.485,  $p < 0.001$ , 95%CI = 1.228–1.796) (Table 2).

**Table 2.** The multivariable Mendelian randomization analysis for type 2 diabetes mellitus adjusted by body mass index and waist circumference.

Exposure	nsnp	Multivariable MR			
		OR	<i>p</i> -Value	95% Lower CI	95% Upper CI
BMI	43	2.753	0.395	0.267	28.365
T2DM	43	1.485	<b>&lt;0.001</b>	1.228	1.796
Waist circumference	43	0.190	0.266	0.010	3.545

MR: Mendelian randomization; nsnp: number of single nucleotide polymorphisms; OR: odds ratio; CI: confidence interval; BMI: body mass index; T2DM: type 2 diabetes mellitus. A bold font indicates statistical significance—a *p*-value < 0.05.

### 3. Discussion

To explore the causal relationship between T2DM and PaCa, we performed a 2SMR analysis using two large T2DM genome-wide association meta-analyses [29,30] and PaCa cases from the FinnGen and UK Biobank datasets. The IVW method, along with four other robust methods, were utilized in the MR analysis. Sensitivity analyses, the MR Steiger directionality test, and multivariable MR were conducted to strengthen our results. Our findings revealed that genetic liability to T2DM was associated with a higher PaCa risk. Our 2SMR results provided evidence of causal associations between T2DM and PaCa.

The previous two MR studies [21,22] did not observe a causal relationship between genetic liability to T2DM and the risk of PaCa. In the study by Carreras-Torres et al. [21], 44 IVs for T2DM were identified from the previous genetic fine-mapping studies [31,32]. PaCa cases were obtained from the Pancreatic Cancer Cohort Consortium (PanScan) and the Pancreatic Cancer Case-Control Consortium (PanC4). The limited number of IVs, possibly due to the reliance on non-current T2DM GWAS meta-analyses, may have led to reduced statistical power, making it difficult to detect a genuine causal effect. In another study by Chen et al. [22], 231 SNPs associated with T2DM were selected from a genome-wide association meta-analysis [30], with PaCa cases identified from FinnGen, UK Biobank, and PanScan. Despite observing no significant association between genetic liability to T2DM and PaCa in their primary IVW approach, a multivariable MR analysis adjusted for BMI showed that T2DM was associated with a higher OR of 1.19 (95% CI = 1.01–1.40) in the UK Biobank dataset. Given these inconsistent results, further examination is required to investigate whether genetic liability to T2DM is linked to PaCa.

Our results align with two previous 2SMR studies [19,20], indicating a causal association between T2DM and PaCa. In the 2SMR study [19], 83 IVs were selected from the 2018 GWAS meta-analysis [33]. The PaCa cases were obtained from the PanScan and the PanC4. The IVW approach revealed a borderline association between T2DM and PaCa with an OR of 1.09 (95% CI = 1.00–1.19, *p*-value = 0.05) in their restricted MR model. The other 2SMR study [20] used 295 IVs from the DIAGRAM [29] consortium and excluded SNPs in or near the *FTO* gene region. PaCa cases were identified within the UK Biobank. Their results suggested a causal association between genetic liability to T2DM and PaCa, with an OR of 1.12 (95% CI = 1.03–1.21) using IVW methods. An umbrella review [34] that included only three studies [19–21] also indicated potential causal associations between genetically predicted T2DM and PaCa. However, the potential for overstated strength and pleiotropic bias in the studies [19,20] still needs to be addressed. For instance, the selected IVs in the prior study [19] included SNPs related to the *FTO* gene, and linkage disequilibrium was not accounted for during the IV selection process in the latter study [20]. In our study, our results are likely to be more robust and well-substantiated by circumventing potential overestimation and pleiotropic effects during the IV selection process.

Our findings from the 2SMR study strengthen the evidence for a causal association between T2DM and PaCa and further support previous observational studies [13,14,17,18,35]. In our two previous UK Biobank cohort studies [13,14], the OR of PaCa was 2.08 and 2.57 in participants with a history of DM compared to controls without a history of DM.

Additionally, both new-onset and long-term DM have been reported to approximately double the risk of PaCa [17,18,35]. An umbrella review [34] also revealed a pooled OR of roughly 2 for PaCa risk among patients with T2DM compared to controls.

The pathophysiological mechanisms linking T2DM to PaCa are complex and multifaceted, encompassing insulin resistance and hyperinsulinemia [16,36–38], persistent hyperglycemia [39–42], chronic inflammation [16,43,44], alterations in gut microbiota [45–49], and dysregulated adipokine secretion [16,50–54]. Insulin resistance in T2DM leads to hyperinsulinemia, potentially promoting tumor growth directly by acting on insulin receptors or indirectly by increasing levels of insulin-like growth factor-1 (IGF-1), both of which can stimulate cell proliferation and inhibit apoptosis to enhance cell proliferation pathways [36–38]. Additionally, hyperglycemia provides an energy-rich environment for cancer cells, inducing oxidative stress and leading to DNA damage [39–42]. Concurrently, chronic inflammation associated with T2DM creates a pro-inflammatory environment conducive to pancreatic carcinogenesis [43,44]. Some inflammatory mediators or cytokines, such as interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF- $\alpha$ ), can promote tumor growth and metastasis [43,44]. Changes in the gut microbiota related to T2DM may influence systemic inflammation and metabolic profiles, which may affect cancer development through alterations in bile acid metabolism and the release of metabolites that may have carcinogenic properties [45–49]. Moreover, altered adipokine secretion, characterized by increased leptin and decreased adiponectin levels due to adiposity in T2DM, may facilitate cancer progression through pro-inflammatory and anti-apoptotic effects [50–54]. These interconnected pathways underscore the complex relationship between T2DM and PaCa.

On the other hand, DM may also be a consequence of PaCa [16,55]. For instance, type 3c diabetes (T3cDM), defined as diabetes secondary to pancreatic exocrine disease [16,55], includes cases resulting from PaCa. Therefore, examining the potential for reverse causation is also crucial. In this study, the results of the MR Steiger directionality test verified the correct direction of causality between T2DM and PaCa, thereby enhancing confidence in our causal inference result. Obesity has been widely recognized as a significant risk factor for T2DM [56,57]. To mitigate the confounding bias, SNPs in the *FTO* gene, which are strongly associated with obesity [58], were excluded from the restricted model. Additionally, multivariable MR analyses were conducted with adjustments for BMI and waist circumference. The results from the restricted model 2SMR and multivariable MR robustly support our causal inference, demonstrating that it is not influenced by the confounding factor of obesity.

Recognizing the causal link between T2DM and PaCa can raise awareness about targeting T2DM for pancreatic cancer prevention. Understanding these pathophysiological mechanisms could potentially pave the way for developing targeted prevention and treatment strategies.

This study possesses several strengths. Our 2SMR study design effectively mitigates biases due to confounding, pleiotropy, and reverse causality. Firstly, the IVs for T2DM were obtained from two large European GWAS meta-analyses, enabling us to identify the latest and most comprehensive SNPs related to T2DM and ensuring sufficient statistical power. Additionally, focusing on the European population helped avoid population structure bias in our results. Secondly, the total and average instrument strengths in this study were assessed using the PVE and the F-statistic, which diminished the bias from weak instruments. Furthermore, power calculations for our 2SMR study indicated a robust power estimation. Thirdly, our IVs selection process was high quality and stringent, ensuring all chosen SNPs met the genome-wide significance threshold, were independent of LD, had harmonized strand orientation, and did not include any ambiguous palindromic SNPs. Fourthly, in addition to the main IVW approach, we employed WM, WMO, MR-Egger, and MR-PRESSO as robust methods to minimize potential biases, including pleiotropy, weak instrument bias, and other sources of analytical distortion, thereby enhancing the reliability and validity of the causal inferences drawn from our study. Fifthly, our sensitivity analyses, including the MR-Egger regression intercept test, funnel plots, and leave-one-out



test, all confirmed the absence of significant horizontal pleiotropy. Sixthly, the MR Steiger directionality test was performed to ensure the causality direction from T2DM to PaCa was correctly identified. Last but not least, to mitigate the potential pleiotropy associated with obesity, SNPs near the *FTO* gene were excluded from our restricted model. Moreover, multivariable MR analyses were conducted, confirming the causal association between T2DM and PaCa by adjusting for BMI and waist circumference.

Nonetheless, some limitations still need to be considered in this study. Firstly, both the IVs and the outcome data were obtained from European population-based datasets. Hence, this limits our findings from being generalized to other populations. Secondly, heterogeneity among IVs was detected in some FinnGen analyses using Cochran's Q test for the IVW method. However, the MR-Egger test and other sensitivity analyses did not indicate pleiotropy. In the MR-PRESSO test, causal inference remained significant even after correcting for outliers. Additionally, in the comprehensive model, the WM approach also revealed a significant effect in the combined FinnGen and UK Biobank analysis. Therefore, the likelihood of bias in the results due to pleiotropic effects or invalid IVs is reduced. Lastly, although disease diagnoses are defined by ICD codes and electronic healthcare records, there still exists the possibility of detection bias among T2DM and PaCa cases. Nevertheless, our results are derived from two large GWAS meta-analyses and two extensive population-based datasets, which may overcome individual errors or biases in disease identification.

## 4. Materials and Methods

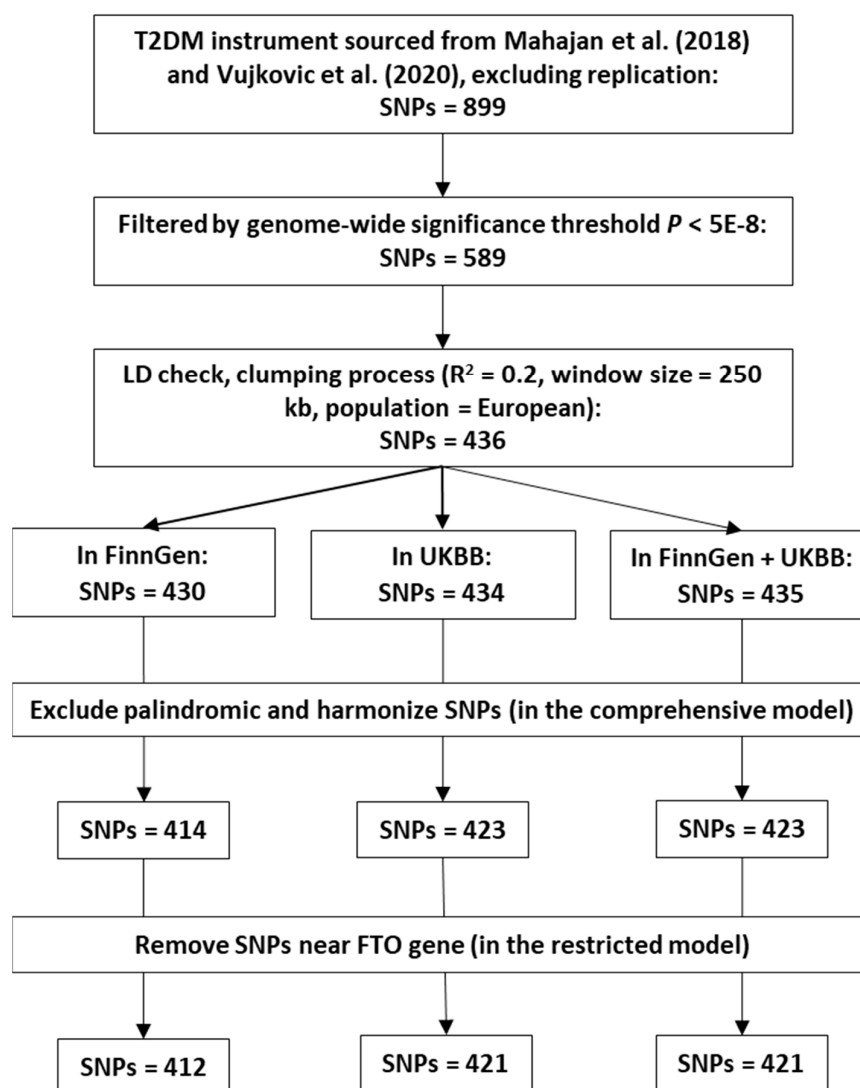
### 4.1. Study Design

The 2SMR study was used to investigate the causal relationship between T2DM and PaCa. We utilized publicly available summary data from genome-wide association studies (GWAS), including the FinnGen, the UKBB, and two genome-wide association meta-analyses [29,30]. Our research adhered to the three critical assumptions of MR [59]: (1) a strong association between the genetic instruments and T2DM; (2) no association of these instruments with confounding variables; and (3) the exclusive influence of these instruments on PaCa through T2DM.

### 4.2. Genetic Instrument Selection

The selection of IVs for T2DM in this study was based on two genome-wide association meta-analyses [29,30]. The first, known as the DIAGRAM consortium by Mahajan et al. [29], encompassed 74,124 T2DM cases and 824,006 controls of European descent. Second, a genome-wide association meta-analysis by Vujkovic et al. [30] involved 228,499 T2DM cases and 1,178,783 controls of multi-ancestry. The following criteria were applied for the selection of IVs: (1) Initially, we identified single nucleotide polymorphisms (SNPs) from the genome-wide association meta-analyses conducted by Mahajan et al. [29] and Vujkovic et al. [30]. SNPs identified as replication variants were selected for possessing a smaller  $p$ -value ( $n = 899$ ). (2) SNPs that met the genome-wide statistical significance threshold ( $p < 5 \times 10^{-8}$ ) were selected as instrumental variables for T2DM ( $n = 589$ ). (3) The clumping threshold for linkage disequilibrium (LD) was set at  $r^2 = 0.2$  within a 250 kb window [60], using the 1000 Genomes European Panel as the reference. Based on this threshold, 436 SNPs were confirmed as independent. (4) A total of 430 SNPs, 434 SNPs, and 435 SNPs were available in the FinnGen, UKBB, and a combination of FinnGen and UKBB datasets, respectively. (5) To mitigate issues related to the orientation of strands, we flipped the reverse strand to the forward strand, harmonized the effect of SNPs on exposure and outcome, and dealt with the palindromic SNPs. Ambiguous palindromic SNPs that exhibited a minor allele frequency (MAF) greater than 0.42 were discarded [61]. Finally, 414 SNPs in the FinnGen dataset, 423 SNPs in the UKBB dataset, and 423 SNPs in a combined FinnGen and UKBB dataset were selected for comprehensive model analysis. (6) Genetic variants near the *FTO* gene were reported to be associated with BMI [58]. Therefore, to mitigate potential pleiotropic effects, SNPs in the vicinity of the *FTO* gene

were excluded from our restricted model analysis ( $n = 412$  (FinnGen),  $n = 421$  (UKBB), and  $n = 421$  (FinnGen + UKBB)). The flowchart of the IV selection is shown in Figure 3.



**Figure 3.** The flowchart for type 2 diabetes mellitus (T2DM) instrument selection. T2DM instrument sourced from Mahajan et al. (2018) [29] and Vujkovic et al. (2020) [30].

#### 4.3. Outcome Data Source

The GWAS summary data for PaCa used in our study were sourced from the R10 release of the FinnGen Consortium [62,63], including both the FinnGen and UK Biobank GWAS summary data. For detailed information on the web browser, please refer to: <https://public-metareresults-fg-ukbb.finnngen.fi/> (accessed on 5 January 2024). In the search browser [63], the phenotype “Malignant neoplasm of pancreas (excluding all other cancers in controls)” was used, including 1626 cases and 314,193 controls in FinnGen and 936 cases and 400,294 controls in the UK Biobank. In the FinnGen dataset, PaCa cases were identified using codes from ICD-8, ICD-9, and ICD-10, as well as surgery codes and medication purchase codes. In the UK Biobank, PaCa cases were diagnosed using codes from ICD-9 and ICD-10, surgery records, and self-reported information.

#### 4.4. The Strength of the Selection of Instrumental Variables and Power Calculations

To avoid bias from weak instruments in this study, we adopted the proportion of variance explained (PVE) known as  $R^2$  for assessing total strength [64] and the F-statistic

for measuring average instrument strength [64]. The PVE in the exposure is explained by the selected genetic variants. Generally, a higher PVE is preferable, as it significantly enhances the effectiveness of an MR analysis [64]. The F-statistic was proposed to assess IV strength [65,66]. A commonly used cutoff value is 10 [65,66]; an F-statistic less than 10 indicates weak instruments. The formulas we adopted for the PVE [67] and the F-statistic [67,68] are as follows:

$$PVE = 2 \times EAF \times (1 - EAF) \times \beta^2 \quad (1)$$

$$F\text{-statistic} = \frac{PVE \times (N - K - 1)}{(1 - PVE) \times (K)} \quad (2)$$

where  $\beta$  denotes the beta coefficient for the exposure of the SNP according to the GWAS summary data,  $EAF$  represents the effect allele frequency of the SNP,  $N$  refers to the total number of samples, and  $K$  is the number of IVs.

In this study, the PVE of 436 SNPs after LD clumping was 41.4%, and the F-statistic was 986.35. Both the total and average instrument strengths are considered good.

The power calculation for two-sample MR analysis was conducted using an online tool (<https://shiny.cnsgenomics.com/mRnd/> (accessed on 5 January 2024)) [69,70], and the outcomes are presented in Supplementary Table S12. The variance explained by the genetic instruments associated with T2DM adopted in FinnGen, UKBB, and combined FinnGen and UKBB studies was 39.16%, 40.18%, and 40.31%, respectively.

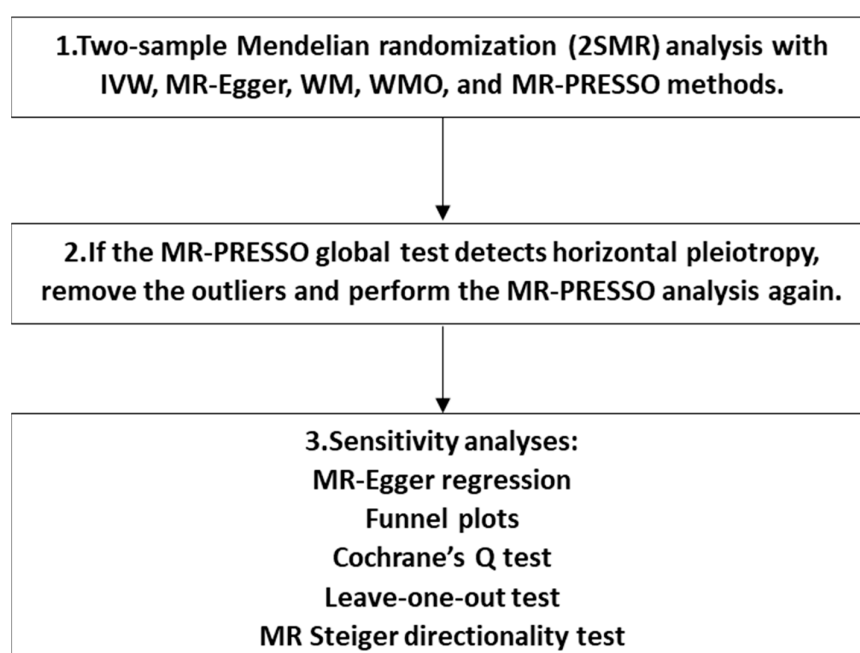
#### 4.5. Two-Sample Mendelian Randomization (2SMR) Analysis

Numerous MR methods were applied in our 2SMR analysis, including IVW [71] and four other robust methods: the MR-Egger method [72], the WM [73] method, the WMO method [74], and the MR-PRESSO method [75]. The most widely used method is the IVW method, which aggregates Wald estimates of each SNP to derive a comprehensive overall effect estimate [71]. This approach hinges on a crucial prerequisite: the IVs employed in the analysis must be valid and satisfy the three assumptions of MR [71]. Due to its efficiency with valid IVs, the IVW method is often recommended as the primary method for analysis [61]. Nonetheless, to detect potential pleiotropic effects, other robust MR methods should also be performed [61].

The MR-Egger method calculates the causal effect by using the average pleiotropic effect as the intercept [61]. However, its accuracy relies on the indirect effects being unrelated to the exposure, a concept referred to as the Instrument Strength Independent of Direct Effect (InSIDE) [72]. In the WM method, it is assumed that a majority of the variants used are valid instruments (known as the majority valid assumption) [73]. The effect estimate of each genetic variant on the outcome is weighted according to its association with the exposure. The median is then determined. The advantage of the WM method lies in its ability to handle invalid IVs and outliers [61]. Regarding the WMO method, it operates under the assumption that more variants estimate the true causal effect than any other quantity (known as the plurality valid assumption) [61]. Similar to the WM method, weights are assigned to each variant. However, the WMO method utilizes the mode rather than the median. The benefit of the WMO method is its ability to withstand invalid IVs and outliers [74]. MR-PRESSO builds upon the IVW method and includes a global test, an outlier test, and a distortion test [75]. The MR-PRESSO global test is capable of identifying horizontal pleiotropy [61]. If horizontal pleiotropy is detected, it is corrected through outlier removal. Additionally, the distortion test is used to assess whether there are notable disparities in effects before and after the outlier correction process [75].

Hence, in our study, the IVW method was designated as the primary approach, while the other four robust methods were employed as complementary methods. We first performed MR analysis with all the above-selected IVs. If the MR-PRESSO global test identified horizontal pleiotropy, the outliers would be eliminated, and the MR-PRESSO analysis would be repeated. In addition, sensitivity analyses were performed. The MR-

Egger regression intercept analysis was conducted to examine horizontal pleiotropy, with a  $p$ -value  $< 0.05$  considered as evidence of horizontal pleiotropy. Funnel plots were also created for pleiotropy direction detection, where an asymmetrical or skewed pattern may indicate horizontal pleiotropy is present [72]. Furthermore, heterogeneity was assessed through Cochran's Q test, where a  $p$ -value  $< 0.05$  would be considered an indication of heterogeneity. Moreover, a leave-one-out test was executed, systematically removing each SNP to mitigate the potential heterogeneity and consolidate the stability of the estimated causal effect in our study. Ultimately, the MR Steiger directionality test [76] was employed to ascertain the direction of causality by assessing whether the variance explained in the outcome is less than that in the exposure. Our 2SMR analysis was in accordance with the recommendations provided in the STROBE-MR statement [77]. Detailed information is listed in Supplementary Table S13. The flowchart of the 2SMR analysis process is shown in Figure 4.



**Figure 4.** The flowchart of the two-sample Mendelian randomization (2SMR) analysis.

#### 4.6. Multivariable Mendelian Randomization (MR)

To mitigate the effects of pleiotropy and reduce bias due to confounding from obesity, we also conducted multivariable Mendelian randomization (MR). Multivariable MR [78] can assess the influence of multiple exposures on the same outcome. Thus, we utilized multivariable MR to explore the causal relationship between T2DM and PaCa by adjusting for BMI and waist circumference. Initially, the 436 T2DM IVs through LD clumping were searched on the PhenoScanner [79,80] website. Of these, 45 SNPs were found to have overlapping traits with “Body Mass Index” and “Waist Circumference”. After harmonizing the SNPs, 43 SNPs were ultimately selected as IVs for our multivariable MR analysis.

#### 4.7. Statistical Analysis

A significance level below 0.05 was considered statistically significant, with statistical significance determined by 95% confidence intervals (95% CI) not including one. In this study, the statistical analyses were conducted using R software [81] (version 4.3.2, R Development Core Team, Vienna, Austria). The TwoSampleMR R package [27] and MR-PRESSO R package [82] were employed for all 2SMR analyses, utilizing functions such as `harmonise_data`, `mr`, `mr_presso`, `mr_heterogeneity`, `mr_pleiotropy_test`, `mr_singlesnp`,

mr\_leaveoneout, mr\_scatter\_plot, mr\_forest\_plot, mr\_funnel\_plot, mv\_multiple, and directionality\_test [83].

#### 4.8. Ethics

All data analyzed in this study were obtained from publicly available GWAS summary datasets. The original GWAS had received approval from the relevant ethics committee. This study did not collect any new data; hence, further ethical approval was not necessary.

### 5. Conclusions

Our findings indicate that a causal relationship between T2DM and PaCa has been established in the 2SMR and multivariable MR studies. This discovery should therefore be used to enhance awareness and the implementation of early prevention and detection strategies for PaCa. These strategies include managing diabetes as a preventive measure against PaCa and emphasizing the importance of controlling blood sugar levels and other metabolic risk factors. Additionally, increasing public awareness of the causal link between T2DM and PaCa could underscore the significance of lifestyle interventions, such as diet habits, physical activity, and weight management, not only for diabetes management but also for reducing the risk of PaCa. Furthermore, understanding the causal pathways could lead to discovering biomarkers and developing pharmacological strategies to improve treatment outcomes and enable more personalized treatment plans. Therefore, further studies are necessary to elucidate the precise pathophysiological mechanisms involved. Ultimately, all these efforts aim to reduce the incidence and mortality associated with PaCa, highlighting the role of T2DM in the development of PaCa and the potential approaches to preventing this disease.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms25094615/s1>. Reference [84] is cited in the Supplementary Materials.

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**Informed Consent Statement:** Not applicable. No human data were collected in this study.

**Data Availability Statement:** The datasets analyzed during the current study are available in the FinnGen repository (<https://r10.finnngen.fi/> (accessed on 5 January 2024)). The summary statistics can be accessed by applying at: <https://elomake.helsinki.fi/lomakkeet/124935/lomake.html> (accessed on 5 January 2024).

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## References

1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA A Cancer J. Clin.* **2021**, *71*, 209–249. [CrossRef] [PubMed]
2. Ferlay, J.E.M.; Lam, F.; Laversanne, M.; Colombet, M.; Mery, L.; Piñeros, M.; Znaor, A.; Soerjomataram, I.; Bray, F. *Global Cancer Observatory: Cancer Today*; International Agency for Research on Cancer: Lyon, France, 2020. Available online: <https://gco.iarc.who.int/today> (accessed on 6 February 2024).
3. Rawla, P.; Sunkara, T.; Gaduputi, V. Epidemiology of Pancreatic Cancer: Global Trends, Etiology and Risk Factors. *World J. Oncol.* **2019**, *10*, 10–27. [CrossRef] [PubMed]
4. Ferlay, J.E.M.; Lam, F.; Colombet, M.; Mery, L.; Pineros, M.; Znaor, A.; Soerjomataram, I. *Global Cancer Observatory: Cancer Tomorrow*; International Agency for Research on Cancer: Lyon, France, 2020. Available online: [http://gco.iarc.fr/tomorrow/graphic-isotype?type=1&population=900&mode=population&sex=0&age\\_group=value&apc\\_male=0&apc\\_female=0](http://gco.iarc.fr/tomorrow/graphic-isotype?type=1&population=900&mode=population&sex=0&age_group=value&apc_male=0&apc_female=0) (accessed on 7 February 2024).
5. Klein, A.P. Pancreatic cancer epidemiology: Understanding the role of lifestyle and inherited risk factors. *Nat. Rev. Gastroenterol. Hepatol.* **2021**, *18*, 493–502. [CrossRef]
6. Hu, J.X.; Zhao, C.F.; Chen, W.B.; Liu, Q.C.; Li, Q.W.; Lin, Y.Y.; Gao, F. Pancreatic cancer: A review of epidemiology, trend, and risk factors. *World J. Gastroenterol.* **2021**, *27*, 4298–4321. [CrossRef] [PubMed]
7. Cai, J.; Chen, H.; Lu, M.; Zhang, Y.; Lu, B.; You, L.; Zhang, T.; Dai, M.; Zhao, Y. Advances in the epidemiology of pancreatic cancer: Trends, risk factors, screening, and prognosis. *Cancer Lett.* **2021**, *520*, 1–11. [CrossRef] [PubMed]
8. Zhao, Z.; Liu, W. Pancreatic Cancer: A Review of Risk Factors, Diagnosis, and Treatment. *Technol. Cancer Res. Treat.* **2020**, *19*, 1533033820962117. [CrossRef] [PubMed]
9. McGuigan, A.; Kelly, P.; Turkington, R.C.; Jones, C.; Coleman, H.G.; McCain, R.S. Pancreatic cancer: A review of clinical diagnosis, epidemiology, treatment and outcomes. *World J. Gastroenterol.* **2018**, *24*, 4846–4861. [CrossRef]
10. Khan, M.A.B.; Hashim, M.J.; King, J.K.; Govender, R.D.; Mustafa, H.; Al Kaabi, J. Epidemiology of Type 2 Diabetes—Global Burden of Disease and Forecasted Trends. *J. Epidemiol. Glob. Health* **2020**, *10*, 107–111. [CrossRef] [PubMed]
11. Singer, M.E.; Dorrance, K.A.; Oxenreiter, M.M.; Yan, K.R.; Close, K.L. The type 2 diabetes ‘modern preventable pandemic’ and replicable lessons from the COVID-19 crisis. *Prev. Med. Rep.* **2022**, *25*, 101636. [CrossRef]
12. Burden of diabetes and hyperglycaemia in adults in the Americas, 1990–2019: A systematic analysis for the Global Burden of Disease Study 2019. *Lancet Diabetes Endocrinol.* **2022**, *10*, 655–667. [CrossRef]
13. Ke, T.M.; Lophatananon, A.; Muir, K.R. Risk Factors Associated with Pancreatic Cancer in the UK Biobank Cohort. *Cancers* **2022**, *14*, 4991. [CrossRef] [PubMed]
14. Ke, T.M.; Lophatananon, A.; Muir, K.R. An Integrative Pancreatic Cancer Risk Prediction Model in the UK Biobank. *Biomedicines* **2023**, *11*, 3206. [CrossRef] [PubMed]
15. CDC: Diabetes Fast Facts. Available online: <https://www.cdc.gov/diabetes/basics/quick-facts.html#:~:text=Diabetes%20Fast%20Facts,-Diabetes%20Fast%20Facts&text=Type%20%20diabetes%20accounts%20for,for%20approximately%205-10%25> (accessed on 14 February 2024).
16. Andersen, D.K.; Korc, M.; Petersen, G.M.; Eibl, G.; Li, D.; Rickels, M.R.; Chari, S.T.; Abbruzzese, J.L. Diabetes, Pancreatogenic Diabetes, and Pancreatic Cancer. *Diabetes* **2017**, *66*, 1103–1110. [CrossRef]
17. Elena, J.W.; Stepilowski, E.; Yu, K.; Hartge, P.; Tobias, G.S.; Brotzman, M.J.; Chanock, S.J.; Stolzenberg-Solomon, R.Z.; Arslan, A.A.; Bueno-De-Mesquita, H.B.; et al. Diabetes and risk of pancreatic cancer: A pooled analysis from the pancreatic cancer cohort consortium. *Cancer Causes Control* **2013**, *24*, 13–25. [CrossRef] [PubMed]
18. Song, S.; Wang, B.; Zhang, X.; Hao, L.; Hu, X.; Li, Z.; Sun, S. Long-Term Diabetes Mellitus Is Associated with an Increased Risk of Pancreatic Cancer: A Meta-Analysis. *PLoS ONE* **2015**, *10*, e0134321. [CrossRef]
19. Lu, Y.; Gentiluomo, M.; Lorenzo-Bermejo, J.; Morelli, L.; Obazee, O.; Campa, D.; Canzian, F. Mendelian randomisation study of the effects of known and putative risk factors on pancreatic cancer. *J. Med. Genet.* **2020**, *57*, 820–828. [CrossRef] [PubMed]
20. Yuan, S.; Kar, S.; Carter, P.; Vithayathil, M.; Mason, A.M.; Burgess, S.; Larsson, S.C. Is Type 2 Diabetes Causally Associated With Cancer Risk? Evidence From a Two-Sample Mendelian Randomization Study. *Diabetes* **2020**, *69*, 1588–1596. [CrossRef] [PubMed]
21. Carreras-Torres, R.; Johansson, M.; Gaborieau, V.; Haycock, P.C.; Wade, K.H.; Relton, C.L.; Martin, R.M.; Davey Smith, G.; Brennan, P. The Role of Obesity, Type 2 Diabetes, and Metabolic Factors in Pancreatic Cancer: A Mendelian Randomization Study. *J. Natl. Cancer Inst.* **2017**, *109*, dxj012. [CrossRef] [PubMed]
22. Chen, J.; Yuan, S.; Fu, T.; Ruan, X.; Qiao, J.; Wang, X.; Li, X.; Gill, D.; Burgess, S.; Giovannucci, E.L.; et al. Gastrointestinal Consequences of Type 2 Diabetes Mellitus and Impaired Glycemic Homeostasis: A Mendelian Randomization Study. *Diabetes Care* **2023**, *46*, 828–835. [CrossRef]
23. Lee, K.; Lim, C.Y. Mendelian Randomization Analysis in Observational Epidemiology. *J. Lipid Atheroscler.* **2019**, *8*, 67–77. [CrossRef]
24. Sanderson, E.; Glymour, M.M.; Holmes, M.V.; Kang, H.; Morrison, J.; Munafò, M.R.; Palmer, T.; Schooling, C.M.; Wallace, C.; Zhao, Q.; et al. Mendelian randomization. *Nat. Rev. Methods Primers* **2022**, *2*, 6. [CrossRef] [PubMed]
25. Burgess, S.; Foley, C.N.; Allara, E.; Staley, J.R.; Howson, J.M.M. A robust and efficient method for Mendelian randomization with hundreds of genetic variants. *Nat. Commun.* **2020**, *11*, 376. [CrossRef] [PubMed]

26. Richmond, R.C.; Davey Smith, G. Mendelian Randomization: Concepts and Scope. *Cold Spring Harb. Perspect. Med.* **2022**, *12*, a040501. [[CrossRef](#)] [[PubMed](#)]
27. Hemani, G.; Zheng, J.; Elsworth, B.; Wade, K.H.; Haberland, V.; Baird, D.; Laurin, C.; Burgess, S.; Bowden, J.; Langdon, R.; et al. The MR-Base platform supports systematic causal inference across the human phenome. *Elife* **2018**, *7*, e34408. [[CrossRef](#)] [[PubMed](#)]
28. Hartwig, F.P.; Davies, N.M.; Hemani, G.; Davey Smith, G. Two-sample Mendelian randomization: Avoiding the downsides of a powerful, widely applicable but potentially fallible technique. *Int. J. Epidemiol.* **2016**, *45*, 1717–1726. [[CrossRef](#)] [[PubMed](#)]
29. Mahajan, A.; Taliun, D.; Thurner, M.; Robertson, N.R.; Torres, J.M.; Rayner, N.W.; Payne, A.J.; Steinthorsdottir, V.; Scott, R.A.; Grarup, N.; et al. Fine-mapping type 2 diabetes loci to single-variant resolution using high-density imputation and islet-specific epigenome maps. *Nat. Genet.* **2018**, *50*, 1505–1513. [[CrossRef](#)] [[PubMed](#)]
30. Vujkovic, M.; Keaton, J.M.; Lynch, J.A.; Miller, D.R.; Zhou, J.; Tcheandjieu, C.; Huffman, J.E.; Assimes, T.L.; Lorenz, K.; Zhu, X.; et al. Discovery of 318 new risk loci for type 2 diabetes and related vascular outcomes among 1.4 million participants in a multi-ancestry meta-analysis. *Nat. Genet.* **2020**, *52*, 680–691. [[CrossRef](#)]
31. Morris, A.P.; Voight, B.F.; Teslovich, T.M.; Ferreira, T.; Segrè, A.V.; Steinthorsdottir, V.; Strawbridge, R.J.; Khan, H.; Grallert, H.; Mahajan, A.; et al. Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat. Genet.* **2012**, *44*, 981–990. [[CrossRef](#)] [[PubMed](#)]
32. Gaulton, K.J.; Ferreira, T.; Lee, Y.; Raimondo, A.; Mägi, R.; Reschen, M.E.; Mahajan, A.; Locke, A.; Rayner, N.W.; Robertson, N.; et al. Genetic fine mapping and genomic annotation defines causal mechanisms at type 2 diabetes susceptibility loci. *Nat. Genet.* **2015**, *47*, 1415–1425. [[CrossRef](#)]
33. Xue, A.; Wu, Y.; Zhu, Z.; Zhang, F.; Kemper, K.E.; Zheng, Z.; Yengo, L.; Lloyd-Jones, L.R.; Sidorenko, J.; Wu, Y.; et al. Genome-wide association analyses identify 143 risk variants and putative regulatory mechanisms for type 2 diabetes. *Nat. Commun.* **2018**, *9*, 2941. [[CrossRef](#)]
34. Pearson-Stuttard, J.; Papadimitriou, N.; Markozannes, G.; Cividini, S.; Kakourou, A.; Gill, D.; Rizos, E.C.; Monori, G.; Ward, H.A.; Kyrgiou, M.; et al. Type 2 Diabetes and Cancer: An Umbrella Review of Observational and Mendelian Randomization Studies. *Cancer Epidemiol. Biomark. Prev.* **2021**, *30*, 1218–1228. [[CrossRef](#)] [[PubMed](#)]
35. Gupta, S.; Vittinghoff, E.; Bertenthal, D.; Corley, D.; Shen, H.; Walter, L.C.; McQuaid, K. New-onset diabetes and pancreatic cancer. *Clin. Gastroenterol. Hepatol.* **2006**, *4*, 1366–1372, quiz 1301. [[CrossRef](#)] [[PubMed](#)]
36. Toledo, F.G.S.; Chari, S.; Yadav, D. Understanding the Contribution of Insulin Resistance to the Risk of Pancreatic Cancer. *Am. J. Gastroenterol.* **2021**, *116*, 669–670. [[CrossRef](#)] [[PubMed](#)]
37. Wang, F.; Herrington, M.; Larsson, J.; Permert, J. The relationship between diabetes and pancreatic cancer. *Mol. Cancer* **2003**, *2*, 4. [[CrossRef](#)] [[PubMed](#)]
38. Deng, J.; Guo, Y.; Du, J.; Gu, J.; Kong, L.; Tao, B.; Li, J.; Fu, D. The Intricate Crosstalk Between Insulin and Pancreatic Ductal Adenocarcinoma: A Review From Clinical to Molecular. *Front. Cell Dev. Biol.* **2022**, *10*, 844028. [[CrossRef](#)] [[PubMed](#)]
39. Duan, W.; Shen, X.; Lei, J.; Xu, Q.; Yu, Y.; Li, R.; Wu, E.; Ma, Q. Hyperglycemia, a neglected factor during cancer progression. *Biomed. Res. Int.* **2014**, *2014*, 461917. [[CrossRef](#)] [[PubMed](#)]
40. Giri, B.; Dey, S.; Das, T.; Sarkar, M.; Banerjee, J.; Dash, S.K. Chronic hyperglycemia mediated physiological alteration and metabolic distortion leads to organ dysfunction, infection, cancer progression and other pathophysiological consequences: An update on glucose toxicity. *Biomed. Pharmacother.* **2018**, *107*, 306–328. [[CrossRef](#)] [[PubMed](#)]
41. Ramteke, P.; Deb, A.; Shepal, V.; Bhat, M.K. Hyperglycemia Associated Metabolic and Molecular Alterations in Cancer Risk, Progression, Treatment, and Mortality. *Cancers* **2019**, *11*, 1402. [[CrossRef](#)]
42. Sato, K.; Hikita, H.; Myojin, Y.; Fukumoto, K.; Murai, K.; Sakane, S.; Tamura, T.; Yamai, T.; Nozaki, Y.; Yoshioka, T.; et al. Hyperglycemia enhances pancreatic cancer progression accompanied by elevations in phosphorylated STAT3 and MYC levels. *PLoS ONE* **2020**, *15*, e0235573. [[CrossRef](#)]
43. Shadhu, K.; Xi, C. Inflammation and pancreatic cancer: An updated review. *Saudi J. Gastroenterol.* **2019**, *25*, 3–13. [[CrossRef](#)]
44. Padoan, A.; Plebani, M.; Basso, D. Inflammation and Pancreatic Cancer: Focus on Metabolism, Cytokines, and Immunity. *Int. J. Mol. Sci.* **2019**, *20*, 676. [[CrossRef](#)]
45. Sammallahti, H.; Kokkola, A.; Rezasoltani, S.; Ghanbari, R.; Asadzadeh Aghdaei, H.; Knuutila, S.; Puolakkainen, P.; Sarhadi, V.K. Microbiota Alterations and Their Association with Oncogenomic Changes in Pancreatic Cancer Patients. *Int. J. Mol. Sci.* **2021**, *22*, 12978. [[CrossRef](#)] [[PubMed](#)]
46. Li, Q.; Jin, M.; Liu, Y.; Jin, L. Gut Microbiota: Its Potential Roles in Pancreatic Cancer. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 572492. [[CrossRef](#)] [[PubMed](#)]
47. Chai, Y.; Huang, Z.; Shen, X.; Lin, T.; Zhang, Y.; Feng, X.; Mao, Q.; Liang, Y. Microbiota Regulates Pancreatic Cancer Carcinogenesis through Altered Immune Response. *Microorganisms* **2023**, *11*, 1240. [[CrossRef](#)]
48. Wei, M.Y.; Shi, S.; Liang, C.; Meng, Q.C.; Hua, J.; Zhang, Y.Y.; Liu, J.; Zhang, B.; Xu, J.; Yu, X.J. The microbiota and microbiome in pancreatic cancer: More influential than expected. *Mol. Cancer* **2019**, *18*, 97. [[CrossRef](#)]
49. Yang, Q.; Zhang, J.; Zhu, Y. Potential Roles of the Gut Microbiota in Pancreatic Carcinogenesis and Therapeutics. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 872019. [[CrossRef](#)] [[PubMed](#)]
50. Parida, S.; Siddharth, S.; Sharma, D. Adiponectin, Obesity, and Cancer: Clash of the Bigwigs in Health and Disease. *Int. J. Mol. Sci.* **2019**, *20*, 2519. [[CrossRef](#)]

51. Wang, Q.; Wang, H.; Ding, Y.; Wan, M.; Xu, M. The Role of Adipokines in Pancreatic Cancer. *Front. Oncol.* **2022**, *12*, 926230. [[CrossRef](#)] [[PubMed](#)]
52. Brocco, D.; Florio, R.; De Lellis, L.; Veschi, S.; Grassadonia, A.; Tinari, N.; Cama, A. The Role of Dysfunctional Adipose Tissue in Pancreatic Cancer: A Molecular Perspective. *Cancers* **2020**, *12*, 1849. [[CrossRef](#)]
53. Chang, H.-H.; Eibl, G. Obesity-Induced Adipose Tissue Inflammation as a Strong Promotional Factor for Pancreatic Ductal Adenocarcinoma. *Cells* **2019**, *8*, 673. [[CrossRef](#)]
54. Kim, J.; Oh, C.M.; Kim, H. The Interplay of Adipokines and Pancreatic Beta Cells in Metabolic Regulation and Diabetes. *Biomedicines* **2023**, *11*, 2589. [[CrossRef](#)]
55. Hart, P.A.; Bellin, M.D.; Andersen, D.K.; Bradley, D.; Cruz-Monserrate, Z.; Forsmark, C.E.; Goodarzi, M.O.; Habtezion, A.; Korc, M.; Kudva, Y.C.; et al. Type 3c (pancreatogenic) diabetes mellitus secondary to chronic pancreatitis and pancreatic cancer. *Lancet Gastroenterol. Hepatol.* **2016**, *1*, 226–237. [[CrossRef](#)]
56. Ruze, R.; Liu, T.; Zou, X.; Song, J.; Chen, Y.; Xu, R.; Yin, X.; Xu, Q. Obesity and type 2 diabetes mellitus: Connections in epidemiology, pathogenesis, and treatments. *Front. Endocrinol.* **2023**, *14*, 1161521. [[CrossRef](#)]
57. Maggio, C.A.; Pi-Sunyer, F.X. Obesity and type 2 diabetes. *Endocrinol. Metab. Clin. North. Am.* **2003**, *32*, 805–822. [[CrossRef](#)] [[PubMed](#)]
58. Frayling, T.M.; Timpson, N.J.; Weedon, M.N.; Zeggini, E.; Freathy, R.M.; Lindgren, C.M.; Perry, J.R.; Elliott, K.S.; Lango, H.; Rayner, N.W.; et al. A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. *Science* **2007**, *316*, 889–894. [[CrossRef](#)] [[PubMed](#)]
59. Lawlor, D.A.; Harbord, R.M.; Sterne, J.A.; Timpson, N.; Davey Smith, G. Mendelian randomization: Using genes as instruments for making causal inferences in epidemiology. *Stat. Med.* **2008**, *27*, 1133–1163. [[CrossRef](#)] [[PubMed](#)]
60. Garfield, V.; Farmaki, A.E.; Fatemifar, G.; Eastwood, S.V.; Mathur, R.; Rentsch, C.T.; Denaxas, S.; Bhaskaran, K.; Smeeth, L.; Chaturvedi, N. Relationship Between Glycemia and Cognitive Function, Structural Brain Outcomes, and Dementia: A Mendelian Randomization Study in the UK Biobank. *Diabetes* **2021**, *70*, 2313–2321. [[CrossRef](#)]
61. Burgess, S.; Davey Smith, G.; Davies, N.M.; Dudbridge, F.; Gill, D.; Glymour, M.M.; Hartwig, F.P.; Kutalik, Z.; Holmes, M.V.; Minelli, C.; et al. Guidelines for performing Mendelian randomization investigations: Update for summer 2023. *Wellcome Open Res.* **2019**, *4*, 186. [[CrossRef](#)]
62. Kurki, M.I.; Karjalainen, J.; Palta, P.; Sipilä, T.P.; Kristiansson, K.; Donner, K.M.; Reeve, M.P.; Laivuori, H.; Aavikko, M.; Kaunisto, M.A.; et al. FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature* **2023**, *613*, 508–518. [[CrossRef](#)] [[PubMed](#)]
63. FinnGen R10 + UKBB. Available online: <https://public-metareresults-fg-ukbb.finnngen.fi/about> (accessed on 29 January 2024).
64. Garfield, V.; Salzman, A.; Burgess, S.; Chaturvedi, N. A Guide for Selection of Genetic Instruments in Mendelian Randomization Studies of Type 2 Diabetes and HbA1c: Toward an Integrated Approach. *Diabetes* **2023**, *72*, 175–183. [[CrossRef](#)] [[PubMed](#)]
65. Andrews, I.; Stock, J.H.; Sun, L. Weak Instruments in Instrumental Variables Regression: Theory and Practice. *Annu. Rev. Econ.* **2019**, *11*, 727–753. [[CrossRef](#)]
66. Staiger, D.; Stock, J.H. Instrumental Variables Regression with Weak Instruments. *Econometrica* **1997**, *65*, 557–586. [[CrossRef](#)]
67. You, R.; Chen, L.; Xu, L.; Zhang, D.; Li, H.; Shi, X.; Zheng, Y.; Chen, L. High Level of Uromodulin Increases the Risk of Hypertension: A Mendelian Randomization Study. *Front. Cardiovasc. Med.* **2021**, *8*, 736001. [[CrossRef](#)]
68. Burgess, S.; Thompson, S.G. Avoiding bias from weak instruments in Mendelian randomization studies. *Int. J. Epidemiol.* **2011**, *40*, 755–764. [[CrossRef](#)]
69. Brion, M.J.; Shakhbuzov, K.; Visscher, P.M. Calculating statistical power in Mendelian randomization studies. *Int. J. Epidemiol.* **2013**, *42*, 1497–1501. [[CrossRef](#)]
70. mRnd: Power Calculations for Mendelian Randomization. Available online: <https://shiny.cnsgenomics.com/mRnd/> (accessed on 30 January 2024).
71. Burgess, S.; Butterworth, A.; Thompson, S.G. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet. Epidemiol.* **2013**, *37*, 658–665. [[CrossRef](#)]
72. Bowden, J.; Davey Smith, G.; Burgess, S. Mendelian randomization with invalid instruments: Effect estimation and bias detection through Egger regression. *Int. J. Epidemiol.* **2015**, *44*, 512–525. [[CrossRef](#)]
73. Bowden, J.; Davey Smith, G.; Haycock, P.C.; Burgess, S. Consistent Estimation in Mendelian Randomization with Some Invalid Instruments Using a Weighted Median Estimator. *Genet. Epidemiol.* **2016**, *40*, 304–314. [[CrossRef](#)]
74. Hartwig, F.P.; Davey Smith, G.; Bowden, J. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. *Int. J. Epidemiol.* **2017**, *46*, 1985–1998. [[CrossRef](#)]
75. Verbanck, M.; Chen, C.Y.; Neale, B.; Do, R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat. Genet.* **2018**, *50*, 693–698. [[CrossRef](#)]
76. Hemani, G.; Tilling, K.; Davey Smith, G. Orienting the causal relationship between imprecisely measured traits using GWAS summary data. *PLoS Genet.* **2017**, *13*, e1007081. [[CrossRef](#)]
77. Skrivankova, V.W.; Richmond, R.C.; Woolf, B.A.R.; Davies, N.M.; Swanson, S.A.; VanderWeele, T.J.; Timpson, N.J.; Higgins, J.P.T.; Dimou, N.; Langenberg, C.; et al. Strengthening the reporting of observational studies in epidemiology using mendelian randomisation (STROBE-MR): Explanation and elaboration. *BMJ* **2021**, *375*, n2233. [[CrossRef](#)]



78. Sanderson, E. Multivariable Mendelian Randomization and Mediation. *Cold Spring Harb. Perspect. Med.* **2021**, *11*, a038984. [[CrossRef](#)]
79. Kamat, M.A.; Blackshaw, J.A.; Young, R.; Surendran, P.; Burgess, S.; Danesh, J.; Butterworth, A.S.; Staley, J.R. PhenoScanner V2: An expanded tool for searching human genotype-phenotype associations. *Bioinformatics* **2019**, *35*, 4851–4853. [[CrossRef](#)]
80. PhenoScanner V2. Available online: <http://www.phenoscanter.medschl.cam.ac.uk/> (accessed on 5 February 2024).
81. R Software. Available online: <https://www.r-project.org/> (accessed on 30 January 2024).
82. MR-PRESSO R Package. Available online: <https://github.com/rondolab/MR-PRESSO> (accessed on 30 January 2024).
83. TwoSampleMR: Mendelian Randomization with GWAS Summary Data. Available online: <https://mrcieu.github.io/TwoSampleMR/index.html> (accessed on 30 January 2024).
84. Skrivankova, V.W.; Richmond, R.C.; Woolf, B.A.R.; Yarmolinsky, J.; Davies, N.M.; Swanson, S.A.; VanderWeele, T.J.; Higgins, J.P.T.; Timpson, N.J.; Dimou, N.; et al. Strengthening the Reporting of Observational Studies in Epidemiology using Mendelian Randomization (STROBE-MR) Statement. *JAMA* **2021**, *326*, 1614–1621. [[CrossRef](#)]

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