

Biomarkers in inflammatory bowel disease: a practical guide

Jennie Clough, Michael Colwill, Andrew Poullis, Richard Pollok, Kamal Patel and Sailish Honap 

Ther Adv Gastroenterol

2024, Vol. 17: 1–19

DOI: 10.1177/
17562848241251600

© The Author(s), 2024.
Article reuse guidelines:
[sagepub.com/journals-](https://sagepub.com/journals-permissions)
permissions

Abstract: Inflammatory bowel disease (IBD), comprising ulcerative colitis (UC) and Crohn's disease (CD), is a costly condition in terms of morbidity and healthcare utilization, with an increasing prevalence now approaching 1% in the Western world. Endoscopic assessment of IBD remains the gold standard for diagnosis, evaluation of treatment response and determination of post-operative recurrence, but is expensive and invasive. Biomarkers can facilitate non-invasive disease assessment, with C-reactive protein and faecal calprotectin as the most widely available biomarkers in current clinical practice. This narrative review summarizes the evidence for their use in both UC and CD and offers practical guidance for healthcare providers taking into account the limitations of biomarker interpretation. We present evidence for the future use of novel biomarkers in IBD and discuss how biomarker discovery could deliver the goal of precision medicine in IBD.

Plain language summary

Biomarkers in inflammatory bowel disease: a practical guide

Inflammatory bowel disease (IBD) is a term used to describe two conditions, ulcerative colitis (UC) and Crohn's disease (CD). These two diseases cause inflammation of the bowel, which can lead to diarrhoea, abdominal pain and bleeding from the back passage. The best way of assessing how active a patient's IBD is, is by performing a camera test called a colonoscopy. However, having a colonoscopy is inconvenient, comes with some risks to the patient, and uses a lot of healthcare resources. 'Biomarkers' are proteins detectable in body fluids (such as blood, poo and urine) which can give information to medical staff about how active a patient's disease is, without the need for colonoscopy. In this article, we give guidance about how best to use these tests, and when they might not be so useful. We also discuss new biomarkers and ways in which they could be used in the future to predict which treatments patients might respond to best.

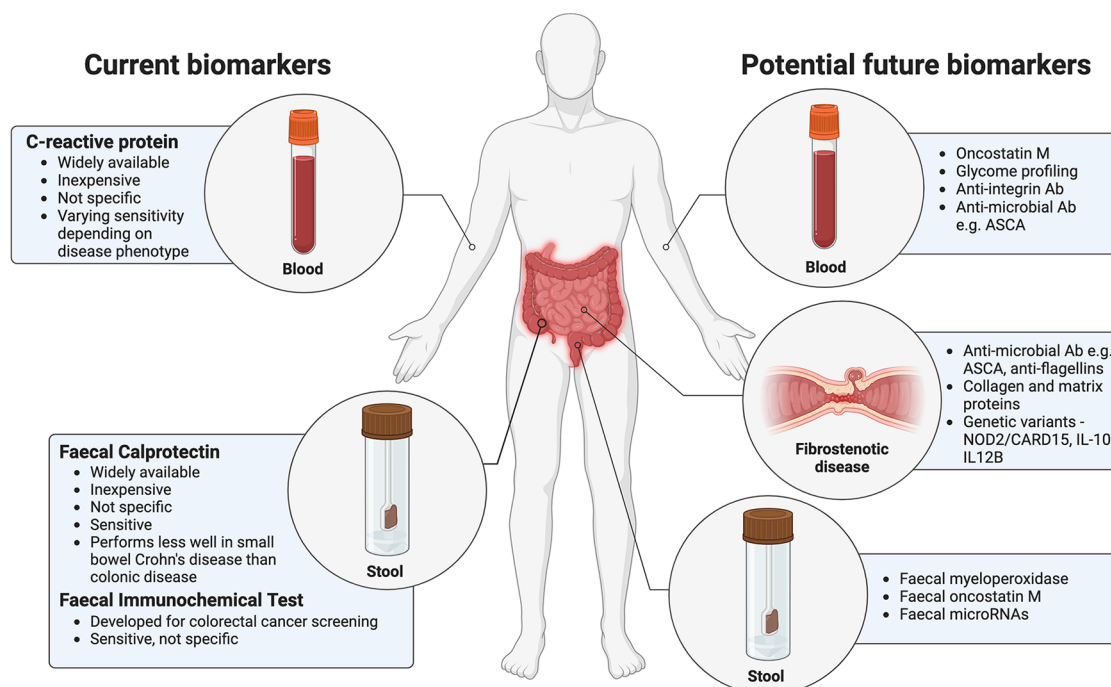
Keywords: biomarker, C-reactive protein, Crohn's disease, disease monitoring, faecal calprotectin, precision medicine, ulcerative colitis

Received: 13 February 2024; revised manuscript accepted: 12 April 2024.

Correspondence to:
Sailish Honap
St George's University
Hospitals NHS Foundation
Trust, London, UK
School of Immunology
and Microbial Sciences,
King's College London,
London, UK
INFINY Institute, Nancy
University Hospital,
Vandœuvre-lès-Nancy,
France
shonap@nhs.net
Jennie Clough
St George's University
Hospitals NHS Foundation
Trust, London, UK
School of Immunology
and Microbial Sciences,
King's College London,
London, UK
Michael Colwill
Andrew Poullis
Kamal Patel
St George's University
Hospitals NHS Foundation
Trust, London, UK
Richard Pollok
St George's University
Hospital NHS Foundation
Trust
Institute of Infection and
Immunity, St George's
University, London, UK

Biomarkers in inflammatory bowel disease: a practical guide

A narrative review of current and potential future biomarkers for diagnosis, prognostication and disease monitoring



Visual Abstract

Introduction

Inflammatory bowel disease (IBD) is a chronic immune-mediated disease affecting the gastrointestinal tract and comprises two main subtypes: ulcerative colitis (UC) and Crohn's disease (CD).^{1,2} The exact cause of IBD remains unknown, but it is believed that in genetically predisposed individuals, an environmental trigger initiates an inappropriate intestinal immune response.³ The overall global prevalence is expected to rise to approximately 1% in the coming decades.⁴

Currently available therapeutic agents attenuate an array of pro-inflammatory cytokines and prevent leucocyte trafficking to the site of inflammation by inhibiting sphingosine-1-phosphate receptors and integrins.^{5,6} The goal of these therapies is to induce clinical and endoscopic remission, reduce the risks of complications and the need for surgical intervention, and improve the quality of life for patients. The gold standard in the assessment of IBD activity is endoscopy, usually through colonoscopy.⁷ However, this procedure is costly and invasive with associated risks to the patient. Therefore,

the use of biomarkers to non-invasively assess disease activity, response to therapy and recurrence of disease has become commonplace. Biomarkers are defined by the National Institute of Health as 'a characteristic that is objectively measured and evaluated as an indication of normal biologic processes, pathogenic processes or pharmacologic responses to a therapeutic intervention'.⁸

Biomarkers can be collected from sources including serum, urine, stool or tissue. Whilst the number of biomarkers available to clinicians has increased in recent years, particularly driven by the growth of metabolomics, genomics and proteomics, not all biomarkers are useful or available to the practicing clinician in everyday practice.⁹⁻¹² An ideal biomarker is sensitive and specific to the observed outcome, available without the need for invasive collection, relevant to underlying pathophysiology, responsive to treatment, useful in prognostication, cost-effective and acceptable to the patient.¹³ For clinicians, there are also further considerations such as availability, turn-around time for testing and robustness of the analytic method used.

While the search for the optimal biomarker in IBD continues, the most widely available biomarkers in current clinical practice include serum and stool testing with C-reactive protein (CRP) and faecal calprotectin (FCP). The use of other faecal markers, for example faecal lactoferrin, is less widespread. CRP and FCP are frequently used by primary care clinicians to differentiate between IBD and irritable bowel syndrome (IBS)¹⁴ and by IBD clinicians to evaluate symptoms and monitor response to therapy. There are also newer genetic biomarkers that may have a future role to play in IBD, such as the NUDT15 codon that predicts adverse effects from thiopurines.¹⁵ Despite significant collaborative research efforts towards biomarker identification, the recent PRedicting Outcomes For Crohn's dIsease using a moLecular biomarkEr (PROFILE) trial of a blood-based biomarker did not show clinical utility in identifying patients with CD at risk of a more severe disease course at diagnosis, for example.^{16,17}

All biomarkers have individual strengths and limitations, and effective clinical use requires nuance and careful interpretation. We present a narrative review of the current literature to provide a practical guide to assist clinicians in day-to-day practice and explore the future of biomarker use in IBD.

Methods

To identify relevant articles for this narrative review, a MEDLINE literature search was conducted through the PubMed platform for articles published in the English language from inception until March 2024. The following search terms were used 'inflammatory bowel disease', 'Crohn's disease', 'ulcerative colitis', 'biomarker(s)', 'C-reactive protein', and 'calprotectin'. Secondary references of the retrieved articles were reviewed to identify publications not captured by the electronic search.

C-reactive protein

CRP and the inflammatory response

First discovered in the 1930s, CRP is a pentameric acute-phase protein that is primarily synthesized in the liver but also by smooth muscle cells, lymphocytes, adipocytes, macrophages and

endothelial cells.¹⁸ In response to infectious stimuli or tissue damage, cytokines including interleukin (IL)-6 and IL-1 β are produced leading to the secretion, primarily by hepatocytes, of CRP into the plasma. CRP binds to C1q molecules to activate the complement pathway, as well as binding *via* Fc receptors to IgG resulting in the release of further pro-inflammatory cytokines.¹⁹ It also plays a role in innate immunity by binding to phosphocholine expressed on the surface of bacterial cells, activating complement-induced phagocytosis and apoptosis. Studies have also shown that circulating CRP breaks down into monomeric subunits which can exert pro-inflammatory effects through activation of monocytes, endothelial cells, platelets and neutrophils.²⁰ CRP serum concentrations can increase by up to 1000-fold within 24–72 h of some bacterial infections, but once the stimulus ends levels rapidly decrease over 18–20 h.¹⁸ As well as infection and tissue damage, CRP is found to be elevated in a multitude of inflammatory conditions such as rheumatoid arthritis, some cardiovascular disease and IBD.

CRP testing is widely available in primary and secondary care, with results available within minutes to hours, acceptable to patients and cost-effective for assessing inflammation. However, CRP is limited as a biomarker in IBD by its lack of specificity, with its expression upregulated in numerous infective and inflammatory pathologies, thus limiting its usefulness in distinguishing between IBD and other differential diagnoses. Its utility in IBD is largely as an adjunct to clinical and endoscopic findings.

CRP in the diagnosis of IBD

CRP is often used in primary care to screen for underlying inflammatory pathology, and with regard to gastrointestinal symptoms, it is effective at distinguishing between inflammatory and functional disease.²¹ However, a review identified that up to 25% of patients with active CD did not mount a CRP response and early work from St Mark's Hospital (UK) found CRP to be elevated in only 50% of patients with UC.²² Genetic polymorphism has also been described as a source of inter-patient variability in CRP.²³ Exclusion of IBD, therefore, should not be made based solely on a normal CRP but in combination with clinical assessment and other markers with better sensitivity.

CRP in disease monitoring

In patients with a known diagnosis of IBD, CRP is commonly used in clinical practice to provide a non-invasive marker of disease activity. However, its accuracy varies based on many clinical factors including whether the patient suffers from UC or CD, and the extent of their disease.^{24,25} The correlation between disease activity in CD and CRP is stronger in CD than in UC²⁶; however, this is dependent upon the disease severity and location. A Korean study of 435 patients found that an elevated CRP was more likely to be seen in ileocolonic or colonic CD compared to patients with isolated ileal disease.²⁷

Even in the setting of a normal CRP, many patients with CD have active disease. One study identified that 92.9% of patients with an elevated Crohn's Disease Activity Index (CDAI) and a normal CRP had active mucosal disease at endoscopy, although these lesions were deemed to correspond only to mildly active disease [Crohn's Disease Endoscopic Index of Severity (CDEIS) ≤ 6]. It has been demonstrated that a normal CRP is negatively associated with hospitalization and the need for surgery, indicating that a normal CRP is suggestive of the absence of severely active CD.²⁸ Conversely, asymptomatic patients with an elevated CRP (so-called 'silent IBD') are at a seven-fold higher risk of having worse disease trajectories²⁹ and a two-fold higher risk of hospitalization.²⁸ Whilst escalating CD therapy based upon a single biomarker is not an advisable strategy, it should prompt physicians to consider other modalities of disease assessment.

CRP also has utility in acute presentations of CD, when a significantly elevated CRP can be indicative of a complication such as perforation, abscess formation or peri-anal collection and may guide the need for subsequent radiological or endoscopic investigation.

The performance of CRP in assessing disease activity in UC is inferior to FCP when correlating with endoscopic appearances.³⁰ A 2019 study found that CRP did not correlate well with low-grade mucosal disease activity, defined as Mayo endoscopic sub-score (MES) of 0 or 1.³¹ An elevated CRP was associated with MES 2–3 disease but only in left-sided or pan-colonic disease. The investigators also examined other serum biomarkers – albumin, erythrocyte sedimentation rate, white blood cell count and platelet count

– but none were found to have any statistically significant correlation with MES. Therefore, whilst CRP may be useful in identifying those with moderate to severe and extensive disease, it does not have a role in proctitis or mild disease, and assessment for these patients should be based upon clinical assessment and other biomarkers such as FCP.

CRP does have a crucial role, however, in the assessment of acute severe UC (ASUC), together with clinical, radiographic and endoscopic evaluation. European Crohn's and Colitis Organisation (ECCO) guidelines state that patients with a significantly elevated CRP >30 mg/L in association with bloody diarrhoea with a stool frequency of >6 /day have developed ASUC and require admission for intensive treatment with either intravenous steroids, infliximab or ciclosporin.³² The ECCO guidelines, based upon work by Truelove and Witts and the Oxford criteria, state that non-responders or patients with a worsening clinical picture at day 3, including a static or worsening CRP, have an 85% chance of requiring colectomy during admission.³³ Whilst more recent data have suggested the rate of colectomy may not be this high,³⁴ a recent study found a clear correlation between CRP and deep ulceration seen at endoscopy³⁵ which itself represents a higher colectomy risk. The CRP:albumin ratio (CAR) has also been identified as a useful biomarker in ASUC, with a CAR >0.85 at day 3 of admission predictive of steroid-refractory ASUC and the need for rescue therapy.³⁶ Furthermore, in patients who had responded to medical management, a CAR of >0.37 at discharge was predictive of the need for colectomy within 12 months.³⁷ CRP therefore remains a key part of overall clinical assessment and decision-making regarding treatment escalation and the need for surgery in ASUC.

CRP in assessing treatment response

As above, a significant proportion of patients with CD will not mount an elevated CRP despite endoscopically and clinically active disease, and CRP is therefore not a useful monitoring tool in these patients. Overall, a persistently elevated CRP is associated with therapy failure,^{38,39} whereas a fall in CRP is correlated with clinical response.^{40,41} For patients in remission, a prospective 2010 study found that CRP can predict relapse but it was less sensitive and specific compared to FCP in this role.⁴² It has also

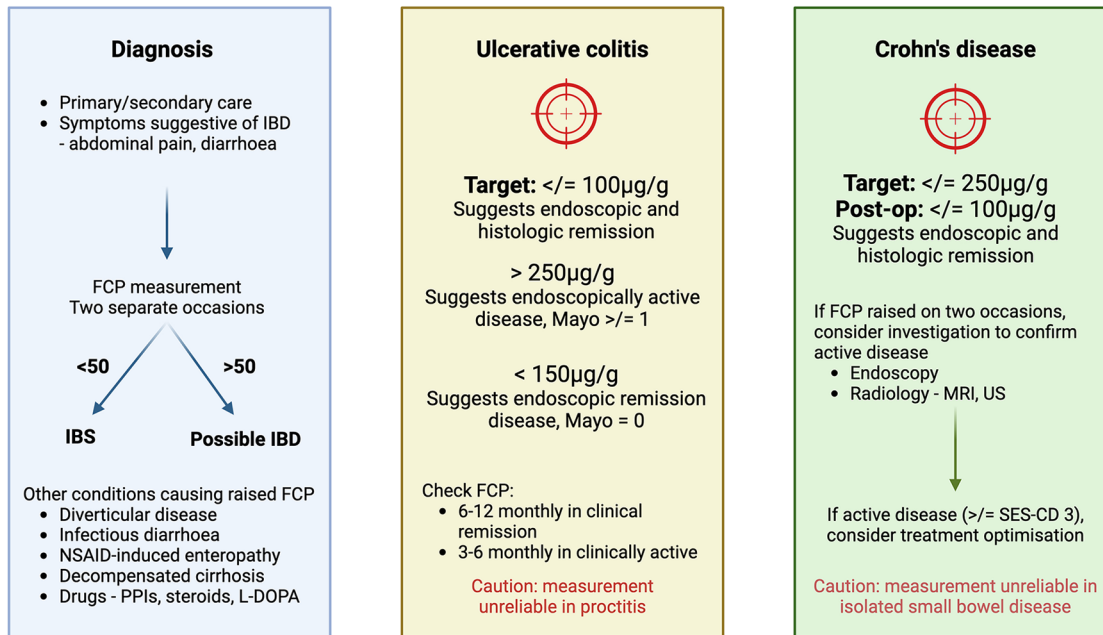


Figure 1. A guide to use and interpretation of FCP testing in clinical practice.

Source: Image created in BioRender.

IBD, inflammatory bowel disease; IBS, irritable bowel syndrome; FCP, faecal calprotectin; MRI, magnetic resonance imaging; NSAID, non-steroidal anti-inflammatory; PPI, proton pump inhibitor; SES-CD, simple endoscopic score in Crohn's disease; US, ultrasound.

been demonstrated that even in asymptomatic individuals, persistently elevated CRP is associated with a higher rate of hospitalization.²⁸ A persisting CRP can therefore be an indicator that further assessment and consideration of treatment modification may be required in CD.

In patients with moderate to severe UC, data suggest that CRP correlates with response to therapy. A 3-year follow-up study of 72 patients with a partial Mayo score (PMS) of 4–9 who were initiated on infliximab found that responders, defined as a PMS ≤ 2 at week 14, had a significantly lower CRP at week 2 compared to partial and non-responders.⁴³ CRP appears to be less effective in predicting relapse, with a study of 74 patients with clinically and endoscopically inactive UC demonstrating that remission CRP was not predictive of those who would flare.⁴⁴

CRP in the prediction of post-operative recurrence

There is conflicting data with regard to the ability of CRP to predict post-operative recurrence in CD. A 2010 study found that persistently elevated CRP was associated with post-operative

recurrence in a small cohort of 12 patients.⁴⁵ However, as discussed earlier, this persistent CRP may simply reflect a more aggressive and severe disease which is known to be a risk factor for post-operative recurrence. Conversely, a 2011 study of 24 patients with CD who were randomized to receive infliximab or placebo post-operatively found that whilst there was a general trend between elevated CRP and relapse, there was no statistical correlation between CRP and endoscopically disease recurrence as assessed by the Rutgeerts score.⁴⁶ Given this conflicting data and small study cohorts, clinical, endoscopic and radiological investigation assessment remains essential in this patient group.

Faecal calprotectin

Faecal calprotectin structure and function

First identified in the 1980s, FCP is a non-invasive biomarker measured in stool, permitting the detection of gut inflammation.^{47,48} Calprotectin (CP) is part of a highly conserved family of calcium-binding S100 leucocyte proteins, composed of two monomers, S100A8 and S100A9.⁴⁹ The monomers can form heterodimers and tetramers

in a calcium-dependent manner, with each heterodimer possessing transition metal binding sites.⁵⁰ CP is found in abundance in the cytosol of neutrophils and is constitutively expressed by monocytes, dendritic cells, activated macrophages and squamous mucosal epithelium.⁴⁷ Importantly, expression of CP is induced during inflammation, with bacterial lipopolysaccharide, tumour necrosis factor- α (TNF α) and IL-1 β (IL-1 β) able to drive CP expression.^{51,52}

The S100A8/S100A9 CP complex controls several functions involved in the control of intracellular pathways of innate immune cells, including modulation of cytoskeletal rearrangement to permit leucocyte recruitment and facilitation of arachidonic acid transport to sites of inflammation.⁵³ Arachidonic acid is a potent inflammatory mediator that has been associated with inflammation and tissue damage in active IBD.⁵⁴ The S100A8/S100A9 complex is readily secreted, triggering neutrophil chemotaxis and endothelial adhesion.⁴⁷ Free CP promotes the expression of both pro-inflammatory and anti-inflammatory mediators, including IL-1 β , IL-6, IL-10 and TNF α ,^{55,56} and regulates cell proliferation, differentiation and apoptosis.⁵⁷ Once tissue damage has been initiated at the mucosal surface, CP release is perpetuated by transcriptional induction of the S100A8 and S100A9 subunits in epithelial cells.⁴⁷ During unresolved inflammation, CP itself can contribute to ongoing mucosal injury in the gut.⁵⁸

FCP in the diagnosis of IBD

Studies in healthy individuals have identified an FCP range between 10 and 50 $\mu\text{g/g}$, although this varies slightly depending on the study population and the assay used.^{59,60} With the development of FCP detection capabilities in 1992, FCP became the first stool biomarker able to discriminate between inflammatory and non-inflammatory gastrointestinal diseases.^{21,61} FCP testing is inexpensive, widely available in primary and secondary care settings, and CP remains stable at room temperature in stool for at least 3 days, reducing the complexities of sample handling and transport.⁶² A summary guide on the use and interpretation of FCP testing in IBD is presented in Figure 1.

FCP correlates with the number of neutrophils present in the intestinal lumen and, whilst

sensitive for the detection of gut inflammation, is not able to discriminate between different inflammatory aetiologies. FCP is also elevated, for example, in infective gastroenteritis, with levels correlating with disease severity in *Salmonella*, *Campylobacter* and *Clostridia* infections.^{63–65} Elevated FCP levels can also be seen in the setting of colonic malignancy,⁶⁶ diverticular diseases,⁶⁷ necrotizing enterocolitis,⁶⁸ graft-versus-host disease⁶⁹ and non-steroidal anti-inflammatory (NSAID) enteropathy.⁷⁰ High FCP values can also be seen in non-intestinal pathology, including decompensated liver cirrhosis⁷¹ and pneumonia,⁷² most likely as a consequence of altered intestinal microbiota and bacterial translocation.

Despite its lack of specificity, FCP has utility in excluding a wide range of inflammatory gut disorders. This makes it especially useful in the setting of primary care, where FCP testing is recommended in national guidelines to differentiate between IBS and IBD – conditions with significant symptom overlap.⁷³ A level greater than 50 $\mu\text{g/g}$ on two occasions is deemed to warrant further invasive testing with colonoscopy and/or bowel imaging, with a recent meta-analysis indicating a pooled sensitivity of 85.8% and specificity of 91.7% for the diagnosis of IBD at this threshold.⁷⁴ Although higher levels (>250 $\mu\text{g/g}$) may be more suggestive of active intestinal inflammation,⁷⁵ a study examining the 12-month outcome of indeterminate FCP levels (50–249 $\mu\text{g/g}$) noted an 8% chance of developing IBD compared with 1% in those <50 $\mu\text{g/g}$.⁷⁶ However, interpretation of slightly elevated FCP concentrations should be made with care, as common drugs including proton pump inhibitors,⁷⁷ NSAIDs,⁷⁰ glucocorticoids⁷⁸ and levodopa⁷⁹ may also induce CP expression.

FCP in disease monitoring

FCP correlates well with endoscopic IBD activity, particularly in the setting of colonic inflammation, with low levels seen in patients with endoscopic and histological remission.^{80,81} It has shown superiority over CRP in predicting endoscopic disease activity,⁸² and is increasingly used for patients in clinical remission to predict disease relapse, and to monitor response to therapy in active disease. The Effect of Tight Control Management on Crohn's disease study

demonstrated that a treat-to-target approach based on FCP results was superior to treatment escalation based on symptoms alone,⁸³ and the International Organisation for the Study of Inflammatory Bowel Disease has published recommendations as part of the Selecting Therapeutic Targets in IBD consensus advising a target FCP of $<150\mu\text{g/g}$ as a goal of treatment.⁷

A challenge in devising treat-to-target strategies in IBD is the lack of evidence exploring the effect of targeting various FCP thresholds on long-term clinical outcomes, with different expert groups proposing different thresholds. Although the use of different thresholds for UC and CD cohorts offers a more nuanced approach, studies suggest that a target of $<250\mu\text{g/g}$ for both groups is a reasonable long-term strategy,^{80,84} which may be more achievable for clinicians managing patients with IBD outside of specialist centres.

Evidence suggests that a reduction in FCP, as well as a target below a certain threshold, has prognostic significance, with FCP able to predict long-term clinical outcomes when measured 12 weeks after initiation of biologic treatment.⁸⁵ In a study of response to anti-TNF α , FCP $<300\mu\text{g/g}$ or a 50% decrease in FCP at weeks 12–14 was predictive of clinical and endoscopic remission.⁸⁶ Whilst individual FCP trajectories are highly heterogeneous over the longitudinal course of the disease, distinct patterns can be seen in FCP trends in patients with CD.⁸⁷ Elevated FCP trajectories were associated with a longer time from diagnosis to initiation of biologic therapy and smoking at diagnosis.

An FCP $>150\mu\text{g/g}$ can predict post-operative recurrence in CD with a sensitivity of approximately 70%,⁸⁸ although current guidelines recommend colonoscopy at 6 months post-operatively for visualization of the anastomosis and calculation of prognostic scoring (Rutgeerts score) to guide further treatment. FCP has also been shown to be a valid method of assessing disease activity in pregnant patients with IBD.⁸⁹ Many clinicians will use both CRP and FCP for the assessment of IBD activity in routine clinical practice. Research supports the combination of available biomarkers as a valid disease monitoring strategy, with a raised CRP and FCP better able to predict outcomes in infliximab-treated patients than using either marker alone.^{84,90}

Limitations of FCP testing

Despite its widespread adoption in the diagnosis and monitoring of IBD, FCP has limitations. Studies have demonstrated marked intra-individual variation in FCP measurement over a few days,⁹¹ which could hinder decision-making strategies based on an isolated sample. However, the variability appears to be greatest in subjects with high levels of CP, which may reduce the clinical relevance of such variation.⁶² Factors such as diet and exercise have been shown to affect day-to-day and within-day FCP variation,⁹² although FCP appears to be homogeneously distributed within a stool sample and a single stool ‘punch’ is, therefore, an adequate sampling strategy.⁹¹ Local policy is to request that patients submit a sample taken from the first bowel motion of the day, to minimize this variability and to ensure the concentration of FCP could be expected to be highest given its accumulation overnight.⁶² Whilst most patients find FCP testing acceptable, sample return rates are highly variable.^{93–95} Most studies of FCP testing have been performed in secondary care settings, limiting their applicability to primary care.

FCP is more sensitive in assessing disease activity in UC than CD⁹⁶ and is limited in its ability to accurately detect disease activity in patients with isolated ileal CD.⁹⁷ Within UC, disease extent affects FCP interpretation, with patients with proctitis exhibiting a poor correlation between FCP concentration and endoscopic activity.⁹⁸ Furthermore, a minority of patients do not appear to mount a detectable FCP increase even in the presence of endoscopically active disease,⁹⁹ and disease assessment and monitoring therefore needs to be personalized for any given patient.

Although FCP is stable within stool for up to 7 days in the presence of calcium, co-existing mucus or blood in patients with active IBD can influence FCP levels.⁹² FCP may undergo oxidative cross-linking *in vivo*, increasing its susceptibility to proteolytic degradation and leading to underestimation of its true levels in commercial assays.¹⁰⁰ Variability can also be seen based on which assay is used for FCP measurement, with the enzyme-linked immunosorbent assay (ELISA) technique deemed to produce the most robust results.¹⁰¹ However, ELISA testing is time-consuming to perform, meaning tests are often run in batches, which may delay the availability of results.¹⁰¹ Automated ELISA tests are now

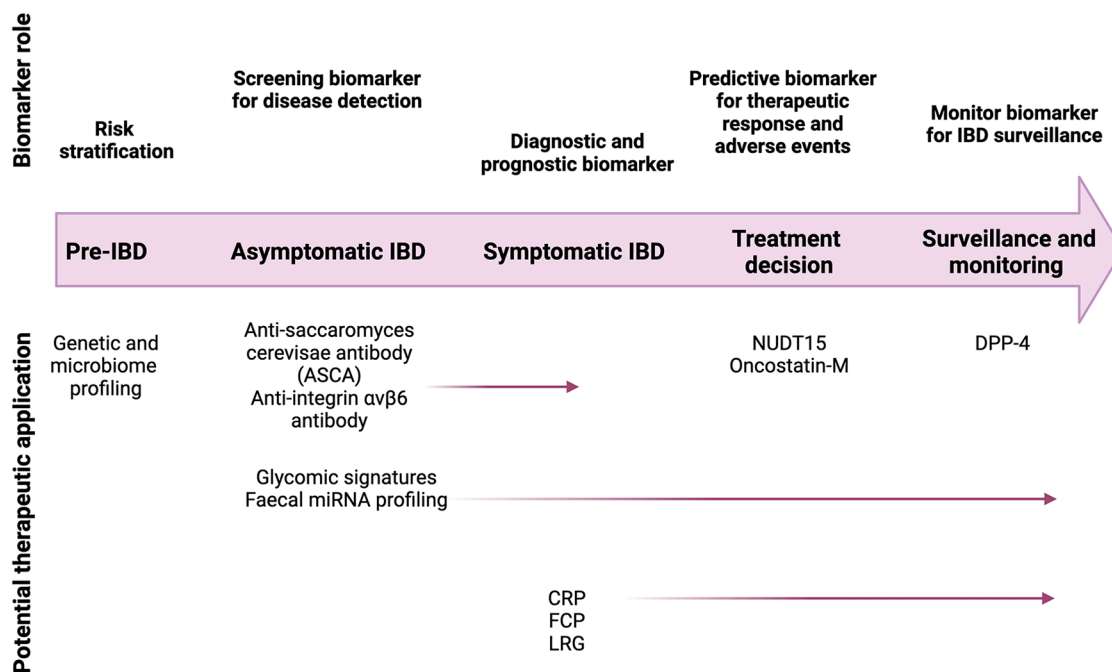


Figure 2. The potential role of biomarkers in the course of IBD, and examples of existing or novel biomarkers which could perform these roles.

Source: Figure created in BioRender.

CRP, C-reactive protein; DPP-4, dipeptidyl peptidase-4; FCP, faecal calprotectin; LRG, leucine-rich alpha-2 glycoprotein.

available permitting individual sample analysis, with data suggesting comparable accuracy to traditional ELISA testing.^{102,103} Newer point-of-care (POC) and home FCP tests have been developed using lateral flow immune assays, with a sample reader able to provide either quantitative or semi-quantitative results.^{30,104} POC tests have been shown to deliver rapid FCP results which correlate well with endoscopic disease activity,³⁰ but they are costly and their use may rely on the use of smartphone applications, limiting their accessibility to certain patient populations. Importantly, longitudinal samples from the same patient should only be compared where they have been analysed using the same FCP assay.

Faecal immunochemical testing

The faecal immunochemical test (FIT) measures faecal haemoglobin concentrations using a specific antibody and is widely used in primary care settings to predict colorectal cancer risk and the need for referral for endoscopic examination.^{105,106} However, FIT is also able to sensitively detect mucosal inflammation and occult luminal blood loss, suggesting a role as a potential biomarker in IBD.^{107–109} A prospective study demonstrated

that a combination of FIT < 100 ng/mL and FCP < 250 µg/g was strongly predictive of mucosal healing, though demonstrated better performance in UC than CD.¹¹⁰

Potential advantages of FIT over FCP testing include lower cost, higher throughput and the benefit of automation over ELISA-based assays.¹¹¹ However, as with FCP testing, a limitation of FIT testing is its lack of specificity, with elevations also seen in systemic inflammatory disease and other inflammatory gastrointestinal pathologies, as well as colorectal cancer.¹¹² Similarly, FIT testing does not appear to perform as well in the detection of small bowel CD compared to colonic IBD.¹¹⁰ This may relate to the optimization of FIT for the detection of colorectal malignancies, and it is notable that FIT is less sensitive for the detection of right-sided colonic lesions.¹¹³

The future of biomarkers in IBD

Of the biomarkers initially identified through research, few make it through clinical testing and validation to become available for use in clinical practice. For those that do, the timeline is long

and arduous, with an estimate of over 10 years from discovery to clinical use. Proposed biomarkers frequently do not perform as anticipated during clinical validation studies, with the recently published PROFILE study demonstrating the challenges in biomarker development and implementation.¹¹⁴ Collaborative research efforts identified a 17-gene blood-based biomarker based on CD8⁺ T-cell transcriptional signatures which were able to categorize patients into two groups associated with higher ('IBDhi') or lower ('IBDlo') risk of treatment escalation.¹¹⁵ Initial validation cohorts confirmed the ability of the prognostic biomarker to identify patients at risk of a more aggressive disease course,¹¹⁶ but in a larger randomized-controlled prospective CD cohort the biomarker was not able to predict patients most likely to benefit from early advanced therapy.¹¹⁴

Whilst currently available biomarkers are primarily used for differentiation of IBD from functional pathology and disease monitoring, future biomarkers could have a role in risk stratification of subjects without disease, as well as screening of asymptomatic individuals for IBD (Figure 2). Artificial intelligence (AI)-based methods are likely to expand the horizon for biomarker discovery, enabling the integration of multimodal data from existing datasets to discover new biomarkers.¹¹⁷ We describe a series of novel biomarkers at various stages of the discovery pipeline, as well as newer techniques for evaluating existing biomarkers.

Novel biomarkers

Oncostatin M. Oncostatin M (OSM) is part of the IL-6 cytokine family and signals through a receptor complex to induce JAK-STAT or P13K-Akt pathway signalling, depending on the cell type and environmental conditions.¹¹⁸ A role for the OSM signalling axis in the pathogenesis of IBD was first suggested with the discovery of a disease-susceptibility polymorphism within the OSM receptor locus,¹¹⁹ and cytokine expression panels have identified OSM as the most highly and consistently expressed cytokine in the inflamed mucosa of patients with IBD.¹²⁰ OSM is proposed to act as an inflammatory amplifier and a driver of disease chronicity.

Newly diagnosed patients with both UC and CD demonstrate increased mucosal expression of

OSM compared to control subjects, and elevated serum OSM was able to predict post-operative CD recurrence 6 months after surgery with greater accuracy than FCP.¹²¹ Elevated colonic OSM and OSM receptor expression were associated with a worse disease prognosis in terms of the requirement to escalate biologic therapy, and high pre-treatment mucosal OSM expression was strongly associated with primary non-response to anti-TNF.¹²⁰ Interestingly, serum OSM levels were also elevated in first-degree relatives of IBD patients,¹²¹ although further work is required to define whether this is predictive of the development of future IBD in these subjects. OSM is also detectable in faeces and has been shown to predict endoscopic disease activity both on its own and in combination with FCP.¹²²

Glycome profiling. Glycans are sequences of carbohydrates conjugated to proteins and lipids. Most secreted proteins are glycosylated through post-translational modification, and glycans play an essential role in the regulation of biological processes including protein folding, immune cell migration and adhesion and pathogen recognition.¹²³ Studies have demonstrated that aberrant glycosylation is associated with numerous inflammatory diseases, including IBD, with the serum *N*-glycome a possible source for biomarker discovery.¹²⁴ Compared to healthy cohorts, IBD patients exhibit a significant decrease in levels of galactosylation and sialylation, as well as altered glycan complexity.¹²⁵ Glycomic signatures generated through ultra-high performance liquid chromatography from the serum of IBD patients obtained at the time of diagnosis were able to predict the need for IBD treatment escalation, with the potential for utility in guiding treatment decisions.¹²⁶ Changes to the glycosylation profile of faecal mucins are also evident in patients with CD compared to healthy controls, suggesting a role in non-invasive monitoring.¹²⁷

Leucine-rich alpha 2-glycoprotein. Leucine-rich alpha-2 glycoprotein (LRG) is predominantly derived from neutrophils, macrophages, gut epithelial cells and hepatocytes in response to elevated TNF α , IL-1 β , IL-6 and IL-22.¹²⁸ Elevated LRG levels have been reported in IBD patients with clinically and endoscopically active disease, and other inflammatory disorders including rheumatoid arthritis, systemic lupus erythematosus and primary biliary cholangitis.^{129–131}

In studies in patients with UC, serum LRG levels were correlated with endoscopic disease activity but were not able to outperform standard FCP testing.¹³¹ In patients with CD, the performance of serum LRG testing appears to be equivalent to that of CRP and FCP in identifying those with endoscopically active disease. Importantly, serum LRG could predict mucosal healing in both patients with UC and CD with normal CRP levels, suggesting a valuable role as a serum biomarker with superior performance over CRP.¹³¹ In addition, levels of serum correlate well with active small bowel CD¹³²; a situation in which FCP alone performs less well.

Faecal myeloperoxidase. Whilst FCP has been extensively studied as a biomarker, additional faecal neutrophil markers may also play a role in IBD monitoring. Myeloperoxidase (MPO) is an abundant neutrophil enzyme that plays a vital role in killing bacteria through the production of hypochlorous acid, but this can also promote inflammatory tissue damage.¹³³ The proposed benefit of testing faecal MPO (fMPO) is that it is not susceptible to oxidative proteolysis, which can reduce the measure of FCP in stool samples thus underestimating the inflammatory burden.¹⁰⁰ Elevated fMPO has been described in small historic studies, particularly in the setting of UC, but these studies demonstrated considerable variability in fMPO measurement.^{134–136} A larger study ($n=172$) including patients with both UC and CD demonstrated similar performance characteristics to FCP in the ability of fMPO to predict moderate-to-severely active IBD, with elevated fMPO levels predictive of a more severe disease course and need for treatment escalation within 12 months of follow-up.¹³⁷ Longer-term follow-up indicated that a raised fMPO is associated with long-term IBD outcomes over a 24-month period, with a combination of baseline CRP, FCP, fMPO and clinical symptom score giving the most accurate prediction of a complicated disease course.⁸⁴

Faecal microRNAs. Micro (mi)-RNAs are small non-coding RNAs detectable in extracellular fluids. Evidence suggests that miRNA dysregulation in IBD could contribute to intestinal inflammation through increased fibrosis, activation of Nuclear factor kappa B (NF- κ B) signalling and altered autophagy.¹³⁸ miRNAs are resistant to degradation and easy to detect through existing laboratory techniques, making them attractive as potential biomarkers.¹³⁹ Faecal miRNA profiling has

demonstrated distinctly different composition in subjects with CD compared to healthy control subjects, with miR-223 and miR-1246 present at high levels in the stool of subjects with active IBD.¹⁴⁰ miR-223 can be detected in both serum and faeces and correlates well with clinical disease activity scores in CD.¹⁴¹ Future work may enable the identification of specific miRNA signatures associated with specific IBD phenotypes or treatment-refractory states to guide treatment decisions.

Antibodies for the detection of IBD. Integrins are key proteins involved in cell adhesion and are comprised of an α and a β chain, with 24 combinations of chains identified.¹⁴² The $\alpha\beta6$ protein is expressed on epithelial cells, with a key role in maintaining epithelial barrier integrity.¹⁴³ Anti-integrin $\alpha\beta6$ antibodies may have a role in the pathophysiology of UC, with 92% of patients with UC testing positive compared to 5.2% healthy control subjects. The presence of the $\alpha\beta6$ antibody had a high specificity and sensitivity for the diagnosis of UC, with a positive correlation between antibody titre and Mayo score, suggesting it may be useful as a biomarker for both diagnosis and disease monitoring.¹⁴³ Antibodies to oligomannosidic epitopes of the yeast *Saccharomyces cerevisiae* (ASCA) have been associated with the development of CD, and when used in combination with a negative perinuclear antineutrophil cytoplasmic autoantibody test demonstrated a high sensitivity and specificity for the diagnosis of CD.¹⁴⁴

Biomarkers for the prediction of fibrostenosing CD. Biomarkers which could be used singly or in combination to predict those at risk of fibrostenotic complications of CD could have clinical utility in stratifying patients most likely to benefit from aggressive treatment escalation. A systematic review identified 35 distinct markers of intestinal fibrosis, which were subsequently categorized into serum ($n=20$), genetic ($n=9$) and histopathology markers ($n=8$).¹⁴⁵ Serum markers included anti-microbial antibodies (including ASCA) and anti-flagellins,^{146,147} collagen and matrix proteins,^{148,149} and miRNAs.¹⁵⁰ The *NOD2/CARD* mutations have been established as genetic variants associated with stricturing disease,¹⁵¹ with additional genetic associations including IL-12B polymorphisms and IL-10 variants.¹⁵² Whilst numerous potential markers of fibrotic CD have been identified, there is significant heterogeneity in their performance and none have yet undergone clinical validation.

Dipeptidyl peptidase 4. Dipeptidyl peptidase (DPP)-4 is nearly ubiquitously expressed and serves an essential role in many metabolic functions, including the regulation of glucose metabolism and the activation of cytokines, chemokines and neuropeptides involved in inflammation.¹⁵³ DPP-4 inhibitors, principally prescribed for glycaemic control, have been shown to suppress inflammation and alleviate oxidative stress.¹⁵⁴ However, the role of DPP-4 in the pathogenesis of IBD is unclear. Whilst some studies demonstrate a reduced risk of IBD in patients receiving DPP-4 inhibitors,¹⁵⁵ others report an increased IBD risk in association with these agents.¹⁵⁶ In IBD patients, serum DPP-4 appears to be inversely correlated with clinically active disease, although initial studies lacked robust endoscopic assessment.¹⁵⁷

New techniques for measuring existing biomarkers

In addition to the advent of novel biomarkers to define disease activity and drug response, new patient-centred ways of measuring existing biomarkers are emerging. A wearable sensor device with the ability to measure CRP and IL-1 β secreted into eccrine sweat has been developed, allowing real-time monitoring of these inflammatory biomarkers and early flare detection.¹⁵⁸ FCP measurement using the standard ELISA technique is frequently time-consuming, with a typical turnaround time of several days. Rapid POC tests use lateral flow chromatography to provide semi-quantitative results within 30 min and show reasonable agreement with ELISA results.¹⁵⁹ Newer methods in which a smartphone application scans a faecal sample to calculate FCP concentration have been proposed and validated, allowing patients to obtain an immediate FCP result in their own homes.¹⁶⁰ Rather than providing a stool sample, which patients can find unpleasant and inconvenient, FCP can also be measured in colonic mucous swabbed from the anus after defecation.¹⁶¹ Furthermore, the use of urinary biomarkers in IBD is being explored, which may be more acceptable to patients than stool sample provision.¹⁶²

Conclusion

Despite the promise of advances in biomarker discovery, most clinicians currently find themselves limited to the use of CRP and FCP in routine clinical practice. It is important, therefore, to understand the strengths and limitations of these

commonly used biomarkers, and the guidance offered in this article is designed to support practicing gastroenterologists. Where doubt exists about variance in performance of biomarkers, or a lack of corroboration between clinical symptoms and biomarker results, endoscopy remains the gold standard tool for assessment of endoscopic healing and is still frequently used to guide treatment decisions. The growing availability of intestinal ultrasound as a POC tool for assessing disease activity is also expected to enhance decision-making in IBD care.^{163,164}

A number of groups are currently underserved by existing biomarkers, including patients with isolated ileal CD, and patients with proctitis, in whom neither CRP nor FCP perform well in disease assessment and monitoring. Future research should focus on evaluation biomarkers that could accurately predict treatment response and outcomes in these populations, especially in the case of small bowel CD where disease is not easily endoscopically accessible. In addition, there is no current access outside of the research setting for biomarkers that can quantify the risk of developing IBD in a susceptible individual, nor screen for IBD detection in asymptomatic individuals.

Precision medicine, which seeks to target therapies by evaluating genetic factors and biomarkers to identify the most active inflammatory pathways in a given patient, is a major goal of future IBD care delivery.⁹ Despite international research efforts, we still have some way to go before we are truly able to deliver personalized medicine in IBD – no currently available biomarker has been robustly validated in predicting response to individual advanced therapies. This represents a significant challenge in an era of ever-expanding IBD therapy, where there is little evidence to guide clinician and patient decision-making in treatment selection and sequencing. With the ever-increasing complexity of available data, AI is likely to play a valuable role in integrating genetic, transcriptomic, proteomic and metabolomic outputs to help achieve this goal,^{10,12,117} driven by advances in the affordability and availability of technology.

Declarations

Ethics approval and consent to participate
Not applicable.

Consent for publication

Not applicable.

Author contributions

Jennie Clough: Project administration; Writing – original draft; Writing – review & editing.

Michael Colwill: Writing – original draft; Writing – review & editing.

Andrew Poullis: Supervision; Writing – review & editing.

Richard Pollok: Supervision; Writing – review & editing.

Kamal Patel: Supervision; Writing – review & editing.

Sailish Honap: Conceptualization; Supervision; Writing – review & editing.

Acknowledgements

None.

Funding

The authors received no financial support for the research, authorship and/or publication of this article.

Competing interests

JC has received a travel grant from Galapagos and has contributed to an advisory board for AbbVie. MC has received a travel grant from Celltrion. AP has no competing interests. RP has contributed to an advisory board for Galapagos. KVP reports payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events from AbbVie, DrFalk, Janssen, PredictImmune and Takeda; support for attending meetings or travel from AbbVie, Ferring, Janssen and Tillotts; and participation on a data safety monitoring board or advisory board for AbbVie, Galapagos and Janssen. SH has received speaker, consultant, advisory board member fees and/or has received travel grants from Pfizer, Janssen, AbbVie, Takeda, Ferring, Galapagos, Lilly and Pharmacosmos.

Availability of data and materials

Not applicable.

ORCID iD

Sailish Honap  <https://orcid.org/0000-0001-6657-2763>

References

1. Le Berre C, Honap S and Peyrin-Biroulet L. Ulcerative colitis. *Lancet* 2023; 402: 571–584.
2. Torres J, Mehandru S, Colombel JF, *et al.* Crohn's disease. *Lancet* 2017; 389: 1741–1755.
3. De Souza HSP and Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol* 2016; 13: 13–27.
4. Kaplan GG and Windsor JW. The four epidemiological stages in the global evolution of inflammatory bowel disease. *Nat Rev Gastroenterol Hepatol* 2021; 18: 56–66.
5. Liu J, Di B and Xu L. Recent advances in the treatment of IBD: targets, mechanisms and related therapies. *Cytokine Growth Factor Rev* 2023; 71–72: 1–12.
6. Cai Z, Wang S and Li J. Treatment of inflammatory bowel disease: a comprehensive review. *Front Med* 2021; 8: 1–24.
7. Turner D, Ricciuto A, Lewis A, *et al.* STRIDE-II: an update on the Selecting Therapeutic Targets in Inflammatory Bowel Disease (STRIDE) Initiative of the International Organization for the Study of IBD (IOIBD): determining therapeutic goals for treat-to-target strategies in IBD. *Gastroenterology* 2021; 160: 1570–1583.
8. Aronson JK and Ferner RE. Biomarkers – a general review. *Curr Protoc Pharmacol* 2017; 76: 9.23.1–9.23.17.
9. Lamb CA, Saifuddin A, Powell N, *et al.* The future of precision medicine to predict outcomes and control tissue remodeling in inflammatory bowel disease. *Gastroenterology* 2022; 162: 1525–1542.
10. Mao R and Chen M. Precision medicine in IBD: genes, drugs, bugs and omics. *Nat Rev Gastroenterol Hepatol* 2022; 19: 81–82.
11. Pinu FR, Goldansaz SA and Jaine J. *Translational metabolomics: current challenges and future opportunities.* *Metabolites* 2019; 9: 108.
12. Atreya R and Neurath MF. Biomarkers for personalizing IBD therapy: the quest continues. *Clin Gastroenterol Hepatol* 2024: 1–12. Epub ahead of print. DOI: 10.1016/j.cgh.2024.01.026
13. Sakurai T and Saruta M. Positioning and usefulness of biomarkers in inflammatory bowel disease. *Digestion* 2023; 104: 30–41.
14. Waugh N, Cummins E, Royle P, *et al.* Faecal calprotectin testing for differentiating amongst

- inflammatory and non-inflammatory bowel diseases: systematic review and economic evaluation. *Health Technol Assess (Rockv)* 2013; 17: xv–xix.
15. Walker GJ, Harrison JW, Heap GA, *et al.* Association of genetic variants in NUDT15 with thiopurine-induced myelosuppression in patients with inflammatory bowel disease. *JAMA* 2019; 321: 753–761.
 16. Parkes M, Noor NM, Dowling F, *et al.* PRedicting Outcomes for Crohn's dIsease using a moLecular biomarkEr (PROFILE): protocol for a multicentre, randomised, biomarker-stratified trial. *BMJ Open* 2018; 8: 1–7.
 17. Parigi TL, D'Amico F, Abreu MT, *et al.* Difficult-to-treat inflammatory bowel disease: results from an international consensus meeting. *Lancet Gastroenterol Hepatol* 2023; 8: 853–859.
 18. Sproston NR and Ashworth JJ. Role of C-reactive protein at sites of inflammation and infection. *Front Immunol* 2018; 9: 1–11.
 19. Pradhan AD, Manson JE, Rifai N, *et al.* C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA* 2001; 286: 327–334.
 20. Ullah N and Wu Y. Regulation of conformational changes in C-reactive protein alters its bioactivity. *Cell Biochem Biophys* 2022; 80: 595–608.
 21. Menees SB, Powell C, Kurlander J, *et al.* A meta-analysis of the utility of C-reactive protein, erythrocyte sedimentation rate, fecal calprotectin, and fecal lactoferrin to exclude inflammatory bowel disease in adults with IBS. *Am J Gastroenterol* 2015; 110: 444–454.
 22. Shine B, Berghouse L, Jones JEL, *et al.* C-reactive protein as an aid in the differentiation of functional and inflammatory bowel disorders. *Clin Chim Acta* 1985; 148: 105–109.
 23. Jones J, Loftus EV, Panaccione R, *et al.* Relationships between disease activity and serum and fecal biomarkers in patients with Crohn's disease. *Clin Gastroenterol Hepatol* 2008; 6: 1218–1224.
 24. Saverymuttu SH, Hodgson HJF, Chadwick VS, *et al.* Differing acute phase responses in Crohn's disease and ulcerative colitis. *Gut* 1986; 27: 809–813.
 25. Gross V, Andus T, Caesar I, *et al.* Evidence for continuous stimulation of interleukin-6 production in Crohn's disease. *Gastroenterology* 1992; 102: 514–519.
 26. Fagan EA, Dyck RF, Maton PN, *et al.* Serum levels of C-reactive protein in Crohn's disease and ulcerative colitis. *Eur J Clin Invest* 1982; 12: 351–359.
 27. Yang DH, Yang SK, Park SH, *et al.* Usefulness of C-reactive protein as a disease activity marker in Crohn's disease according to the location of disease. *Gut Liver* 2015; 9: 80–86.
 28. Click B, Vargas EJ, Anderson AM, *et al.* Silent Crohn's disease: asymptomatic patients with elevated C-reactive protein are at risk for subsequent hospitalization. *Inflamm Bowel Dis* 2015; 21: 2254–2261.
 29. Bhattacharya A, Rao BB, Koutroubakis IE, *et al.* Silent Crohn's disease predicts increased bowel damage during multiyear follow-up: the consequences of under-reporting active inflammation. *Inflamm Bowel Dis* 2016; 22: 2665–2671.
 30. Lobatón T, Rodríguez-Moranta F, Lopez A, *et al.* A new rapid quantitative test for fecal calprotectin predicts endoscopic activity in ulcerative colitis. *Inflamm Bowel Dis* 2013; 19(4): 1034–1042.
 31. Sonoyama H, Kawashima K and Ishihara S. Capabilities of fecal calprotectin and biomarkers as surrogate endoscopic markers. *J Clin Biochem Nutr* 2019; 64: 265–270.
 32. Harbord M, Eliakim R, Bettenworth D, *et al.* Third European evidence-based consensus on diagnosis and management of ulcerative colitis. Part 2: current management. *J Crohn's Colitis* 2017; 11: 769–784.
 33. Truelove SC and Witts LJ. Cortisone in ulcerative colitis. *Br Med J* 1955; 2: 1386.
 34. Moore AC and Bressler B. Acute severe ulcerative colitis: the Oxford criteria no longer predict in-hospital colectomy rates. *Dig Dis Sci* 2020; 65: 576–580.
 35. Rivière P, Le Chevillier A, Rullier A, *et al.* Deep ulcers are associated with increased C-reactive protein in active ulcerative colitis. *Dig Liver Dis* 2023; 55: 1194–1200.
 36. Gibson DJ, Hartery K, Doherty J, *et al.* CRP/albumin ratio: an early predictor of steroid responsiveness in acute severe ulcerative colitis. *J Clin Gastroenterol* 2018; 52: e48–e52.
 37. Choy MC, Seah D, Gorelik A, *et al.* Predicting response after infliximab salvage in acute severe ulcerative colitis. *J Gastroenterol Hepatol* 2018; 33: 1347–1352.

38. Consigny Y, Modigliani R, Colombel JF, *et al.* A simple biological score for predicting low risk of short-term relapse in Crohn's disease. *Inflamm Bowel Dis* 2006; 12: 551–557.
39. Bitton A, Dobkin PL, Edwardes MD, *et al.* Predicting relapse in Crohn's disease: a biopsychosocial model. *Gut* 2008; 57: 1386–1392.
40. Rutgeerts P. Subanalysis from a phase 3 study on the evaluation of natalizumab in active Crohn's disease therapy-1 (ENACT-1). *Gut* 2003; 52: 239.
41. Boirivant M, Leoni M, Tariciotti D, *et al.* The clinical significance of serum C reactive protein levels in Crohn's disease. Results of a prospective longitudinal study. *J Clin Gastroenterol* 1988; 10: 401–405.
42. Kallel L, Ayadi I, Matri S, *et al.* Fecal calprotectin is a predictive marker of relapse in Crohn's disease involving the colon: a prospective study. *Eur J Gastroenterol Hepatol* 2010; 22: 340–345.
43. Iwasa R, Yamada A, Sono K, *et al.* C-reactive protein level at 2 weeks following initiation of infliximab induction therapy predicts outcomes in patients with ulcerative colitis: a 3 year follow-up study. *BMC Gastroenterol* 2015; 15: 1–7.
44. Bitton A, Peppercorn MA, Antonioli DA, *et al.* Clinical, biological, and histologic parameters as predictors of relapse in ulcerative colitis. *Gastroenterology* 2001; 120: 13–20.
45. Sorrentino D, Paviotti A, Terrosu G, *et al.* Low-dose maintenance therapy with infliximab prevents postsurgical recurrence of Crohn's disease. *Clin Gastroenterol Hepatol* 2010; 8: 591–599.e1.
46. Regueiro M, Kip KE, Schraut W, *et al.* Crohn's disease activity index does not correlate with endoscopic recurrence one year after ileocolonic resection. *Inflamm Bowel Dis* 2011; 17: 118–126.
47. Jukic A, Bakiri L, Wagner EF, *et al.* Calprotectin: from biomarker to biological function. *Gut* 2021; 70: 1978–1988.
48. Andersson KB, Sletten K, Berntzen HB, *et al.* The leucocyte L1 protein: identity with the cystic fibrosis antigen and the calcium-binding MRP-8 and MRP-14 macrophage components. *Scand J Immunol* 1988; 28: 241–245.
49. Zimmer DB, Eubanks JO, Ramakrishnan D, *et al.* Evolution of the S100 family of calcium sensor proteins. *Cell Calcium* 2013; 53: 170–179.
50. Zyguel EM and Nolan EM. Transition metal sequestration by the host-defense protein calprotectin. *Annu Rev Biochem* 2018; 87: 621–643.
51. Hu BS, Harrison C, Xu K, *et al.* Induction of the chemotactic S100 protein, CP-10, in monocyte/macrophages by lipopolysaccharide. *Blood* 1996; 87: 3919–3929.
52. Xu K and Geczy CL. IFN- γ and TNF regulate macrophage expression of the chemotactic S100 protein S100A8. *J Immunol* 2000; 164: 4916–4923.
53. Vogl T, Ludwig S, Goebeler M, *et al.* MRP8 and MRP14 control microtubule reorganization during transendothelial migration of phagocytes. *Blood* 2004; 104: 4260–4268.
54. Nielsen OH, Ahnfelt-Rønne I and Elmgreen J. Abnormal metabolism of arachidonic acid in chronic inflammatory bowel disease: enhanced release of leucotriene B4 from activated neutrophils. *Gut* 1987; 28: 181–185.
55. Simard JC, Cesaro A, Chapeton-Montes J, *et al.* S100A8 and S100A9 induce cytokine expression and regulate the NLRP3 inflammasome via ROS-dependent activation of NF- κ B(1.). *PLoS One* 2013; 8: e72138.
56. Cesaro A, Anceriz N, Plante A, *et al.* An inflammation loop orchestrated by S100A9 and calprotectin is critical for development of arthritis. *PLoS One* 2012; 7: e45478.
57. Yang J, Anholts J, Kolbe U, *et al.* Calcium-binding proteins S100A8 and S100A9: investigation of their immune regulatory effect in myeloid cells. *Int J Mol Sci* 2018; 19: 1833.
58. Zhang X, Wei L, Wang J, *et al.* Suppression colitis and colitis-associated colon cancer by anti-S100a9 antibody in mice. *Front Immunol* 2017; 8: 1774.
59. Jha AK, Chaudhary M, Dayal VM, *et al.* Optimal cut-off value of fecal calprotectin for the evaluation of ulcerative colitis: an unsolved issue? *JGH Open* 2018; 2: 207–213.
60. Srinivas M, Eyre R, Ellis R, *et al.* PTU-243 faecal calprotectin (FC) assays: comparison of four assays with clinical correlation. *Gut* 2012; 61(Suppl. 2): A284.3–A285.
61. Røseth AG, Fagerhol MK, Aadland E, *et al.* Assessment of the neutrophil dominating protein calprotectin in feces: a methodologic study. *Scand J Gastroenterol* 1992; 27: 793–798.

62. Lasson A, Stotzer PO, öhmanb L, *et al.* The intra-individual variability of faecal calprotectin: a prospective study in patients with active ulcerative colitis. *J Crohn's Colitis* 2015; 9: 26–32.
63. Chen CC, Huang JL, Chang CJ, *et al.* Fecal calprotectin as a correlative marker in clinical severity of infectious diarrhea and usefulness in evaluating bacterial or viral pathogens in children. *J Pediatr Gastroenterol Nutr* 2012; 55: 541–547.
64. Nielsen HL, Engberg J, Ejlersen T, *et al.* Evaluation of fecal calprotectin in *Campylobacter concisus* and *Campylobacter jejuni/coli* gastroenteritis. *Scand J Gastroenterol* 2013; 48: 633–635.
65. Drózdź M, Biesiada G, Pituch H, *et al.* The level of fecal calprotectin significantly correlates with *Clostridium difficile* infection severity. *Folia Med Cracov* 2019; 59: 53–65.
66. Lehmann FS, Trapani F, Fueglistaler I, *et al.* Clinical and histopathological correlations of fecal calprotectin release in colorectal carcinoma. *World J Gastroenterol* 2014; 20: 4994–4999.
67. Tursi A, Brandimarte G, Elisei W, *et al.* Faecal calprotectin in colonic diverticular disease: a case-control study. *Int J Colorectal Dis* 2009; 24: 49–55.
68. Pergialiotis V, Konstantopoulos P, Karampetsou N, *et al.* Calprotectin levels in necrotizing enterocolitis: a systematic review of the literature. *Inflamm Res* 2016; 65: 847–852.
69. Malik MN, Rafae A, Durer C, *et al.* Fecal calprotectin as a diagnostic and prognostic biomarker for gastrointestinal graft versus host disease: a systematic review of literature. *Cureus* 2019; 11: 2–7.
70. Tibble JA, Sigthorsson G, Foster R, *et al.* High prevalence of NSAID enteropathy as shown by a simple faecal test. *Gut* 1999; 45: 362–326.
71. Gundling F, Schmidtler F, Hapfelmeier A, *et al.* Fecal calprotectin is a useful screening parameter for hepatic encephalopathy and spontaneous bacterial peritonitis in cirrhosis. *Liver Int* 2011; 31: 1406–1415.
72. Gong SS, Fan YH, Han QQ, *et al.* Nested case-control study on risk factors for opportunistic infections in patients with inflammatory bowel disease. *World J Gastroenterol* 2019; 25: 2240–2250.
73. Arasaradnam RP, Brown S, Forbes A, *et al.* Guidelines for the investigation of chronic diarrhoea in adults: British Society of Gastroenterology, 3rd edition. *Gut* 2018; 67: 1380–1399.
74. Dajti E, Frazzoni L, Iascione V, *et al.* Systematic review with meta-analysis: diagnostic performance of faecal calprotectin in distinguishing inflammatory bowel disease from irritable bowel syndrome in adults. *Aliment Pharmacol Ther* 2023; 58: 1120–1131.
75. Dhaliwal A, Zeino Z, Tomkins C, *et al.* Utility of faecal calprotectin in inflammatory bowel disease (IBD): what cut-offs should we apply? *Frontline Gastroenterol* 2015; 6: 14–19.
76. McFarlane M, Chambers S, Malik A, *et al.* Clinical outcomes at 12 months and risk of inflammatory bowel disease in patients with an intermediate raised fecal calprotectin: a 'real-world' view. *BMJ Open* 2016; 6: 1–6.
77. Lundgren D, Eklöf V, Palmqvist R, *et al.* Proton pump inhibitor use is associated with elevated faecal calprotectin levels. A cross-sectional study on subjects referred for colonoscopy. *Scand J Gastroenterol* 2019; 54: 152–157.
78. Hsu K, Passey RJ, Endoh Y, *et al.* Regulation of S100A8 by glucocorticoids. *J Immunol* 2005; 174: 2318–2326.
79. Weis S, Schwartz A, Unger MM, *et al.* Effect of Parkinson's disease and related medications on the composition of the fecal bacterial microbiota. *npj Park Dis* 2019; 5: 1–9.
80. Lin JF, Chen JM, Zuo JH, *et al.* Meta-analysis: fecal calprotectin for assessment of inflammatory bowel disease activity. *Inflamm Bowel Dis* 2014; 20: 1407–1415.
81. Zittan E, Kelly OB, Kirsch R, *et al.* Low fecal calprotectin correlates with histological remission and mucosal healing in ulcerative colitis and colonic Crohn's disease. *Inflamm Bowel Dis* 2016; 22: 623–630.
82. Mosli MH, Zou G, Garg SK, *et al.* C-reactive protein, fecal calprotectin, and stool lactoferrin for detection of endoscopic activity in symptomatic inflammatory bowel disease patients: a systematic review and meta-analysis. *Am J Gastroenterol* 2015; 110: 802–819.
83. Colombel J-F, Panaccione R, Bossuyt P, *et al.* Effect of tight control management on Crohn's disease (CALM): a multicentre, randomised, controlled phase 3 trial. *Lancet* 2018; 390: 2779–2789.
84. Swaminathan A, Borichevsky GM, Frampton CM, *et al.* Comparison of fecal calprotectin and myeloperoxidase in predicting outcomes in

- inflammatory bowel disease. *Inflamm Bowel Dis* 2024; 1–9.
85. Haisma SM, Verkade HJ, Scheenstra R, *et al.* Time-to-reach target calprotectin level in newly diagnosed patients with inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2019; 69: 466–473.
 86. Sollelis E, Quinard RM, Bouguen G, *et al.* Combined evaluation of biomarkers as predictor of maintained remission in Crohn's disease. *World J Gastroenterol* 2019; 25: 2354–2364.
 87. Constantine-Cooke N, Monterrubio-Gómez K, Plevris N, *et al.* Longitudinal fecal calprotectin profiles characterize disease course heterogeneity in Crohn's disease. *Clin Gastroenterol Hepatol* 2023; 21: 2918–2927.e6.
 88. Tham YS, Yung DE, Fay S, *et al.* Fecal calprotectin for detection of postoperative endoscopic recurrence in Crohn's disease: systematic review and meta-analysis. *Therap Adv Gastroenterol* 2018; 11: 1–12.
 89. Kammerlander H, Nielsen J, Kjeldsen J, *et al.* Fecal calprotectin during pregnancy in women with moderate-severe inflammatory bowel disease. *Inflamm Bowel Dis* 2018; 24: 839–848.
 90. Magro F, Estevinho MM, Catalano G, *et al.* How many biomarker measurements are needed to predict prognosis in Crohn's disease patients under infliximab? – a prospective study. *United Eur Gastroenterol J* 2023; 11: 531–541.
 91. Cremer A, Ku J, Amininejad L, *et al.* Variability of faecal calprotectin in inflammatory bowel disease patients: an observational case-control study. *J Crohn's Colitis* 2019; 13: 1372–1379.
 92. Poullis A, Foster R, Shetty A, *et al.* Bowel inflammation as measured by fecal calprotectin: a link between lifestyle factors and colorectal cancer risk. *Cancer Epidemiol Biomarkers Prev* 2004; 13: 279–284.
 93. Lecky DM, Hawking MKD and McNulty CAM. Patients' perspectives on providing a stool sample to their GP: a qualitative study. *Br J Gen Pract* 2014; 64: e684–e693.
 94. Schultze A, Akmatov MK, Andrzejak M, *et al.* Comparison of stool collection on site versus at home in a population-based study Feasibility and participants' preference in Pretest 2 of the German National Cohort. *Bundesgesundheitsblatt Gesundheitsforschung Gesundheitsschutz* 2014; 57: 1264–1269.
 95. Buisson A, Gonzalez F, Poullienot F, *et al.* Comparative acceptability and perceived clinical utility of monitoring tools: a nationwide survey of patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2017; 23: 1425–1433.
 96. Costa F, Mumolo MG, Ceccarelli L, *et al.* Calprotectin is a stronger predictive marker of relapse in ulcerative colitis than in Crohn's disease. *Gut* 2005; 54: 364–368.
 97. Sipponen T, Savilahti E, Kolho KL, *et al.* Crohn's disease activity assessed by fecal calprotectin and lactoferrin: correlation with Crohn's disease activity index and endoscopic findings. *Inflamm Bowel Dis* 2008; 14: 40–46.
 98. Sakuraba A, Nemoto N, Hibi N, *et al.* Extent of disease affects the usefulness of fecal biomarkers in ulcerative colitis. *BMC Gastroenterol* 2021; 21: 1–8.
 99. Bjarnason I. The use of fecal calprotectin in inflammatory bowel disease. *Gastroenterol Hepatol* 2017; 13: 53–56.
 100. Hoskin TS, Crowther JM, Cheung J, *et al.* Oxidative cross-linking of calprotectin occurs in vivo, altering its structure and susceptibility to proteolysis. *Redox Biol* 2019; 24: 101202.
 101. D'Amico F, Nancey S, Danese S, *et al.* A practical guide for faecal calprotectin measurement: myths and realities. *J Crohn's Colitis* 2021; 15: 152–161.
 102. Kraemer A, Bulgakova T, Schukina O, *et al.* Automated fecal biomarker profiling – a convenient procedure to support diagnosis for patients with inflammatory bowel diseases. *Clin Lab* 2020; 66: 1249–1259.
 103. De Sloovere MMW, De Smet D, Baert FJ, *et al.* Analytical and diagnostic performance of two automated fecal calprotectin immunoassays for detection of inflammatory bowel disease. *Clin Chem Lab Med* 2017; 55: 1435–1446.
 104. Schulz C, Wex T, Schütte K, *et al.* Validation of two calprotectin rapid tests in daily routine. *Clin Lab* 2016; 62: 1249–1254.
 105. Morikawa T, Kato J, Yamaji Y, *et al.* A comparison of the immunochemical fecal occult blood test and total colonoscopy in the asymptomatic population. *Gastroenterology* 2005; 129: 422–428.
 106. Park DI, Ryu S, Kim YH, *et al.* Comparison of guaiac-based and quantitative immunochemical fecal occult blood testing in a population at average risk undergoing colorectal cancer screening. *Am J Gastroenterol* 2010; 105: 2017–2025.
 107. Mooiweer E, Fidder HH, Siersema PD, *et al.* Fecal hemoglobin and calprotectin

- are equally effective in identifying patients with inflammatory bowel disease with active endoscopic inflammation. *Inflamm Bowel Dis* 2014; 20: 307–314.
108. Inokuchi T, Kato J, Hiraoka S, *et al.* Fecal immunochemical test versus fecal calprotectin for prediction of mucosal healing in Crohn's disease. *Inflamm Bowel Dis* 2016; 22: 1078–1085.
 109. Nakarai A, Kato J, Hiraoka S, *et al.* Evaluation of mucosal healing of ulcerative colitis by a quantitative fecal immunochemical test. *Am J Gastroenterol* 2013; 108: 83–89.
 110. Ma C, Lumb R, Walker EV, *et al.* Noninvasive fecal immunochemical testing and fecal calprotectin predict mucosal healing in inflammatory bowel disease: a prospective cohort study. *Inflamm Bowel Dis* 2017; 23: 1643–1649.
 111. Kato J, Hiraoka S, Nakarai A, *et al.* Fecal immunochemical test as a biomarker for inflammatory bowel diseases: can it rival fecal calprotectin? *Intest Res* 2016; 14: 5–14.
 112. Noh CK, Lee E, Park B, *et al.* A positive faecal immunochemical test result and its association with the incidence of rheumatoid arthritis, systemic lupus erythematosus, and psoriatic arthritis: an analysis of one-million national colorectal cancer screening programme results. *BMC Med* 2022; 20: 1–14.
 113. Haug U, Knudsen AB, Brenner H, *et al.* Is fecal occult blood testing more sensitive for left- versus right-sided colorectal neoplasia? A systematic literature review. *Expert Rev Mol Diagn* 2011; 11: 605–616.
 114. Noor NM, Lee JC, Bond S, *et al.* A biomarker-stratified comparison of top-down versus accelerated step-up treatment strategies for patients with newly diagnosed Crohn's disease (PROFILE): a multicentre, open-label randomised controlled trial. *Lancet Gastroenterol Hepatol* 2024; 1253: 1–13.
 115. Lee JC, Lyons PA, McKinney EF, *et al.* Gene expression profiling of CD8 + T cells predicts prognosis in patients with Crohn disease and ulcerative colitis. *J Clin Invest* 2011; 121: 4170–4179.
 116. Biasci D, Lee JC, Noor NM, *et al.* A blood-based prognostic biomarker in IBD. *Gut* 2019; 68: 1386–1395.
 117. Prelaj A, Miskovic V, Zanitti M, *et al.* Artificial intelligence for predictive biomarker discovery in immuno-oncology: a systematic review. *Ann Oncol* 2023; 35: 29–65.
 118. Hermanns HM. Oncostatin M and interleukin-31: cytokines, receptors, signal transduction and physiology. *Cytokine Growth Factor Rev* 2015; 26: 545–558.
 119. Sabino J, Verstockt B, Vermeire S, *et al.* New biologics and small molecules in inflammatory bowel disease: an update. *Therap Adv Gastroenterol* 2019; 12: 1–14.
 120. West NR, Hegazy AN, Owens BMJ, *et al.* Oncostatin M drives intestinal inflammation and predicts response to tumor necrosis factor-neutralizing therapy in patients with inflammatory bowel disease. *Nat Med* 2017; 23: 579–589.
 121. Verstockt S, Verstockt B, MacHiels K, *et al.* Oncostatin M is a biomarker of diagnosis, worse disease prognosis, and therapeutic nonresponse in inflammatory bowel disease. *Inflamm Bowel Dis* 2021; 27: 1564–1575.
 122. Cao Y, Dai Y, Zhang L, *et al.* Combined use of fecal biomarkers in inflammatory bowel diseases: oncostatin m and calprotectin. *J Inflamm Res* 2021; 14: 6409–6419.
 123. Hart GW and Copeland RJ. Glycomics hits the big time. *Cell* 2010; 143: 672–676.
 124. Theodoratou E, Campbell H, Ventham NT, *et al.* The role of glycosylation in IBD. *Nat Rev Gastroenterol Hepatol.* 2014; 11: 588–600.
 125. Clerc F, Novokmet M, Dotz V, *et al.* Plasma N-glycan signatures are associated with features of inflammatory bowel diseases. *Gastroenterology* 2018; 155: 829–843.
 126. Shubhakar A, Jansen BC, Adams AT, *et al.* Serum N-glycomic biomarkers predict treatment escalation in inflammatory bowel disease. *J Crohn's Colitis* 2023; 17: 919–932.
 127. Masselot CR, Cordier C, Marsac B, *et al.* Fecal mucin O-glycans as novel biomarkers in inflammatory bowel diseases. *Inflamm Bowel Dis* 2023; 29: E12.
 128. Naka T and Fujimoto M. LRG is a novel inflammatory marker clinically useful for the evaluation of disease activity in rheumatoid arthritis and inflammatory bowel disease. *Immunol Med* 2018; 41: 62–67.
 129. Serada S, Fujimoto M, Terabe F, *et al.* Serum leucine-rich alpha-2 glycoprotein is a disease activity biomarker in ulcerative colitis. *Inflamm Bowel Dis* 2012; 18: 2169–2179.
 130. Shinzaki S, Matsuoka K, Iijima H, *et al.* Leucine-rich Alpha-2 glycoprotein is a serum biomarker of mucosal healing in ulcerative colitis. *J Crohn's Colitis* 2017; 11: 84–91.

131. Yasutomi E, Inokuchi T, Hiraoka S, *et al.* Leucine-rich alpha-2 glycoprotein as a marker of mucosal healing in inflammatory bowel disease. *Sci Rep* 2021; 11: 1–11.
132. Kawamoto A, Takenaka K, Hibiya S, *et al.* Serum Leucine-rich α 2 glycoprotein: a novel biomarker for small bowel mucosal activity in Crohn's disease. *Clin Gastroenterol Hepatol* 2022; 20: e1196–e1200.
133. Winterbourn CC, Kettle AJ and Hampton MB. Reactive oxygen species and neutrophil function. *Annu Rev Biochem* 2016; 85: 765–792.
134. Masoodi I, Dutta U, Vaiphei K, *et al.* Evaluation of fecal myeloperoxidase as a biomarker of disease activity and severity in ulcerative colitis. *Dig Dis Sci* 2012; 57: 1336–1340.
135. Saiki T. Myeloperoxidase concentrations in the stool as a new parameter of inflammatory bowel disease. *Kurume Med J* 1998; 45: 69–73.
136. Peterson CGB, Lampinen M, Hansson T, *et al.* Evaluation of biomarkers for ulcerative colitis comparing two sampling methods: fecal markers reflect colorectal inflammation both macroscopically and on a cellular level. *Scand J Clin Lab Invest* 2016; 76: 393–401.
137. Swaminathan A, Borichevsky GM, Edwards TS, *et al.* Faecal myeloperoxidase as a biomarker of endoscopic activity in inflammatory bowel disease. *J Crohn's Colitis* 2022; 16: 1862–1873.
138. Cao B, Zhou X, Ma J, *et al.* Role of MiRNAs in inflammatory bowel disease. *Dig Dis Sci* 2017; 62: 1426–1438.
139. Mitchell PS, Parkin RK, Kroh EM, *et al.* Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008; 105: 10513–10518.
140. Verdier J, Breunig IR, Ohse MC, *et al.* Faecal micro-RNAs in inflammatory bowel diseases. *J Crohn's Colitis*. 2020; 14: 110–117.
141. Zhang J, Guo Z, Wang Z, *et al.* Fecal miR-223 is a noninvasive biomarker for estimating Crohn's disease activity. *Immun Inflamm Dis* 2023; 11: 1–8.
142. Abram CL and Lowell CA. The ins and outs of leukocyte integrin signaling. *Annu Rev Immunol* 2009; 27: 339–362.
143. Kuwada T, Shiokawa M, Kodama Y, *et al.* Identification of an anti-integrin α v β 6 autoantibody in patients with ulcerative colitis. *Gastroenterology* 2021; 160: 2383–2394.e21.
144. Vermeire S, Peeters M, Vlietinck R, *et al.* Anti-Saccharomyces cerevisiae antibodies (ASCA), phenotypes of IBD, and intestinal permeability: a study in IBD families. *Inflamm Bowel Dis* 2001; 7: 8–15.
145. Steiner CA, Berinstein JA, Louissaint J, *et al.* Biomarkers for the prediction and diagnosis of fibrostenosing Crohn's disease: a systematic review. *Clin Gastroenterol Hepatol* 2022; 20: 817–846.
146. Schoepfer AM, Schaffer T, Mueller S, *et al.* Phenotypic associations of Crohn's disease with antibodies to flagellins A4-Fla2 and Fla-X, ASCA, p-ANCA, PAB, and NOD2 mutations in a Swiss Cohort. *Inflamm Bowel Dis* 2009; 15: 1358–1367.
147. Papadakis KA, Yang H, Ippoliti A, *et al.* Anti-flagellin (CBir1) phenotypic and genetic Crohn's disease associations. *Inflamm Bowel Dis* 2007; 13: 524–530.
148. van Haaften WT, Mortensen JH, Karsdal MA, *et al.* Misbalance in type III collagen formation/degradation as a novel serological biomarker for penetrating (Montreal B3) Crohn's disease. *Aliment Pharmacol Ther* 2017; 46: 26–39.
149. Bourgonje AR, Alexdottir MS, Otten AT, *et al.* Serological biomarkers of type I, III and IV collagen turnover are associated with the presence and future progression of stricturing and penetrating Crohn's disease. *Aliment Pharmacol Ther* 2022; 56: 675–693.
150. Lewis A, Mehta S, Hanna LN, *et al.* Low serum levels of microRNA-19 are associated with a stricturing Crohn's disease phenotype. *Inflamm Bowel Dis* 2015; 21: 1926–1934.
151. Heresbach D, Gicquel-Douabin V, Birebent B, *et al.* NOD2/CARD15 gene polymorphisms in Crohn's disease: a genotype-phenotype analysis. *Eur J Gastroenterol Hepatol* 2004; 16: 55–62.
152. Henckaerts L, Van Steen K, Verstreken I, *et al.* Genetic risk profiling and prediction of disease course in Crohn's disease patients. *Clin Gastroenterol Hepatol* 2009; 7: 972–980.e2.
153. Perry C, Kapur N and Barrett TA. DPP-4 as a novel biomarker for inflammatory bowel disease: Is it ready for clinical use? *Inflamm Bowel Dis* 2020; 26: 1720–1721.
154. Zhang J, Chen Q, Zhong J, *et al.* DPP-4 inhibitors as potential candidates for antihypertensive therapy: improving vascular inflammation and assisting the action of traditional antihypertensive drugs. *Front Immunol* 2019; 10: 1–12.

155. Kim SC, Schneeweiss S, Glynn RJ, *et al.* Dipeptidyl peptidase-4 inhibitors in type 2 diabetes may reduce the risk of autoimmune diseases: a population-based cohort study. *Ann Rheum Dis* 2015; 74: 1968–1975.
156. Abrahami D, Douros A, Yin H, *et al.* Dipeptidyl peptidase-4 inhibitors and incidence of inflammatory bowel disease among patients with type 2 diabetes: population based cohort study. *BMJ* 2018; 360: 1–8.
157. Pinto-Lopes P, Afonso J, Pinto-Lopes R, *et al.* Serum dipeptidyl peptidase 4: a predictor of disease activity and prognosis in inflammatory bowel disease. *Inflamm Bowel Dis* 2020; 26: 1707–1719.
158. Jagannath B, Lin KC, Pali M, *et al.* A sweat-based wearable enabling technology for realtime monitoring of il-1 β and crp as potential markers for inflammatory bowel disease. *Inflamm Bowel Dis* 2020; 26: 1533–1542.
159. Kok L, Elias SG, Witteman BJM, *et al.* Diagnostic accuracy of point-of-care fecal calprotectin and immunochemical occult blood tests for diagnosis of organic bowel disease in primary care: the cost-effectiveness of a decision rule for abdominal complaints in primary care (CEDAR) study. *Clin Chem* 2012; 58: 989–998.
160. Vinding KK, Elsberg H, Thorkilgaard T, *et al.* Fecal calprotectin measured by patients at home using smartphones – a new clinical tool in monitoring patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2016; 22: 336–344.
161. Nooredinvand HA and Poullis A. Emerging role of colorectal mucus in gastroenterology diagnostics. *World J Gastroenterol* 2022; 28: 1220–1225.
162. Baldan-Martin M, Chaparro M and Gisbert JP. Systematic review: urine biomarker discovery for inflammatory bowel disease diagnosis. *Int J Mol Sci* 2023; 24: 10159.
163. Novak KL, Nylund K, Maaser C, *et al.* Expert Consensus on optimal acquisition and development of the International Bowel Ultrasound Segmental Activity Score [IBUS-SAS]: a reliability and inter-rater variability study on intestinal ultrasonography in Crohn's disease. *J Crohn's Colitis* 2021; 15: 609–616.
164. Shivaji UN, Segal JP, Plumb AA, *et al.* Intestinal ultrasonography: a useful skill for efficient, non-invasive monitoring of patients with IBD using a clinic-based point-of-care approach. *Frontline Gastroenterol* 2022; 13: 447–451.

Appendix

Abbreviations

AI	artificial intelligence
ASUC	acute severe ulcerative colitis
CAR	C-reactive protein-to-albumin ratio
CD	Crohn's disease
CDAI	Crohn's disease activity index
CDEIS	Crohn's disease endoscopic index of severity
CRP	C-reactive protein
ECCO	European Crohn's and Colitis Organisation
FIT	faecal immunochemical testing
fMPO	faecal myeloperoxidase
HBI	Harvey-Bradshaw index
IBD	inflammatory bowel disease
IBS	irritable bowel syndrome
IL	interleukin
JAK	janus kinase
LRG	leucine-rich alpha-2 glycoprotein
MES	Mayo endoscopic sub-score
PMS	partial Mayo score
POC	point of care
UC	ulcerative colitis
FCP	faecal calprotectin
TNF α	tumour necrosis factor alpha

Visit Sage journals online
journals.sagepub.com/
home/tag

 Sage journals