



The tRNA^{Val} half: A strong endogenous Toll-like receptor 7 ligand with a 5'-terminal universal sequence signature

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Toll-like receptors (TLRs) are crucial components of the innate immune system. Endosomal TLR7 recognizes single-stranded RNAs, yet its endogenous ssRNA ligands are not fully understood. We previously showed that extracellular (ex-) 5'-half molecules of tRNA^{HisGUG} (the 5'-tRNA^{HisGUG} half) in extracellular vesicles (EVs) of human macrophages activate TLR7 when delivered into endosomes of recipient macrophages. Here, we fully explored immunostimulatory ex-5'-tRNA half molecules and identified the 5'-tRNA^{ValCAC/AAC} half, the most abundant tRNA-derived RNA in macrophage EVs, as another 5'-tRNA half molecule with strong TLR7 activation capacity. Levels of the ex-5'-tRNA^{ValCAC/AAC} half were highly up-regulated in macrophage EVs upon exposure to lipopolysaccharide and in the plasma of patients infected with *Mycobacterium tuberculosis*. The 5'-tRNA^{ValCAC/AAC} half-mediated activation of TLR7 effectively eradicated bacteria infected in macrophages. Mutation analyses of the 5'-tRNA^{ValCAC/AAC} half identified the terminal GUUU sequence as a determinant for TLR7 activation. We confirmed that GUUU is the optimal ratio of guanosine and uridine for TLR7 activation; microRNAs or other RNAs with the terminal GUUU motif can indeed stimulate TLR7, establishing the motif as a universal signature for TLR7 activation. These results advance our understanding of endogenous ssRNA ligands of TLR7 and offer insights into diverse TLR7-involved pathologies and their therapeutic strategies.

tRNA half | TLR7 | short non-coding RNA | macrophage | Valine

One of the fundamental properties of the immune system is to initiate immune responses against invasive pathogens by recognizing conserved pathogen-associated molecular patterns (PAMPs) (1, 2). The immune system can also recognize intracellular molecules of host cells released by stressed or injured tissues, so-called damage-associated molecular patterns (DAMPs), such as histones, self-DNAs, or -RNAs (3–5). Toll-like receptors (TLRs) and other pathogen-recognition receptors in the innate immune system detect PAMPs and DAMPs, initiating protective responses (6, 7). Among the 10 TLRs characterized in humans, TLR1, -2, -4, -5, -6, and -10 localize to the cell surface (surface TLRs), while TLR3, -7, -8, and -9 localize to intracellular compartments such as endosomes (endosomal TLRs). While the surface TLRs mainly recognize bacterial and fungal cell wall components, viral envelope proteins, and protozoal components, endosomal TLRs are nucleic acid-sensing receptors; TLR7 and -8 are specialized in detecting single-stranded RNAs (ssRNAs), while TLR3 and -9 sense double-stranded RNAs (dsRNAs) and single-stranded DNAs, respectively (8). ssRNA recognition by TLR7 or -8 results in the recruitment of the adaptor protein MyD88, leading to IRF7- and NF- κ B-mediated transcription and downstream induction of interferon and cytokine production (6, 8).

While “foreign” ssRNAs from bacteria or viruses have been well studied as the ligands of TLR7 and -8 (9–14), “endogenous” ssRNA ligands from host cells have not been fully characterized yet. To be recognized by TLR7 and -8, ssRNAs should be present inside the endosomes where TLR7 and -8 reside, which can be achieved via extracellular vesicle (EV)-mediated delivery of ssRNAs to endosomes. Indeed, certain EV-contained extracellular (ex-) microRNAs (miRNAs) encapsulated within EVs are transported to the endosomes of recipient cells where they activate TLR7 and -8 (15–18). This miRNA-mediated TLR7/8 activation is relevant not only to immune response (19, 20) but also to neurodegeneration (15), cancers (21, 22), autoimmunity (23, 24), and various other diseases (25).

Although miRNAs have dominated current research on both ex-short noncoding RNAs (sncRNAs) and endogenous ssRNA ligands of TLRs, recent advances in our understanding of “previously hidden” sncRNAs have widened the pool of the candidate ssRNA molecules of TLR7/8 ligands. sncRNAs generally possess either a hydroxyl group (OH), a phosphate (P), or a 2',3'-cyclic phosphate (cP) at their termini (26). To date, most sncRNA sequencing

Significance

Toll-like receptor 7 (TLR7) is vital for innate immunity, recognizing single-stranded RNAs (ssRNAs) as ligands. While microRNAs have been the primary focus of endogenous TLR7 ligand research, recent advances in short noncoding RNA biology have expanded the range of candidate ssRNA molecules. This study comprehensively explored extracellular (ex-) transfer RNA (tRNA)-derived RNAs in extracellular vesicles (EVs) secreted from human macrophages and identified the 5'-tRNA^{ValCAC/AAC} half as a potent activator of TLR7. The terminal GUUU sequence of the 5'-tRNA^{ValCAC/AAC} half was identified as a universal signature for TLR7 activation. In patients infected with *Mycobacterium tuberculosis*, the levels of the extracellular 5'-tRNA^{ValCAC/AAC} half are drastically upregulated. These findings provide insights into a variety of TLR7-related pathologies and potential therapeutic strategies.

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studies have relied on a standard small RNA-seq method in which 5'- and 3'-adaptors (AD) can be ligated only to the 5'-P and 3'-OH ends of RNAs; thus, current sncRNA analyses significantly underrepresent non-miRNA-sncRNAs lacking the 5'-P or 3'-OH ends (26–28). This notion is especially important when it comes to the sequencing analyses of ex-sncRNAs because many ex-sncRNAs lack 5'-P or 3'-OH (29–31). Pretreatment of RNAs with T4 polynucleotide kinase (T4 PNK) to convert the RNA termini to 5'-P/3'-OH-ends (thus rendering them available for 5'-/3'-AD ligation) is required to capture whole ex-sncRNAs (29–31). In our sequencing study of sncRNAs in EVs secreted from human monocyte-derived macrophages (HMDMs), T4 PNK treatment was required to amplify the majority of the cDNAs (32). Treating with a mutant form of T4 PNK, which is deficient in 3'-dephosphorylation activity, resulted in a dramatic reduction of cDNA yield (32), indicating that the majority of sncRNAs within HMDM EVs contain 3'-P or cP, while miRNAs and other 3'-OH-containing RNAs constitute only a minor proportion. Among the EV-sncRNAs, 5'-tRNA halves were found to be one of the major species, comprising over 93% of tRNA-derived sncRNAs in HMDM-secreted EVs (32).

tRNA halves are the most abundant class of tRNA-derived sncRNAs. In mammalian cells, they are generated from angiogenin (ANG)-mediated anticodon cleavage of tRNAs, which can be induced by various biological factors/phenomena such as stress stimuli (33–35) and sex-hormone signaling pathways (36). tRNA halves are expressed as functional sncRNAs that promote stress granule formation (37), regulate translation (33, 38), and promote cell proliferation (36). Our previous study found that mycobacterial infection and accompanying surface TLR activation lead to upregulation of NF- κ B-mediated *ANG* gene transcription, inducing the accumulation of tRNA halves in HMDMs (32). Then, the 5'-tRNA halves are selectively and abundantly packaged into EVs of HMDMs. Among ex-5'-tRNA halves in HMDM EVs, the ex-5'-tRNA^{HisGUG} half is abundantly accumulated (>210 times more than miR-150, the most abundant ex-miRNA) and delivered to endosomes of recipient HMDMs, where it strongly activates TLR7 (32). The strength of its activity was equal to or even more than that of HIV-derived ssRNA40, a widely used positive control ssRNA known as a strong activator of endosomal TLRs (9), suggesting that the 5'-tRNA^{HisGUG} half has the capacity to produce an immune response (32). While these findings suggest that 5'-tRNA halves serve as immune activators (39), only the 5'-tRNA^{HisGUG} half had been demonstrated to activate endosomal TLR.

In this study, by widely exploring immunostimulatory ex-5'-tRNA half molecules, we identified the 5'-tRNA^{ValCAC/AAC} half as another strong TLR7 activator. We further investigated sequence determinants within these 5'-tRNA halves for TLR7 activation to shed light on why only the 5'-tRNA^{HisGUG} half and the 5'-tRNA^{ValCAC/AAC} half exhibit strong activity. Our findings provide a fuller understanding of the role of endogenously produced 5'-tRNA half molecules in activating the endosomal TLR pathway and suggest the wide existence of other endogenous ssRNA molecules that function as ligands for endosomal TLRs.

Results

The 5'-tRNA^{ValCAC/AAC} Half Activates TLR7. Our previous sequencing of T4 PNK-treated sncRNAs from HMDM-secreted EVs revealed 5'-tRNA half as the most major class of ex-tRNA-derived sncRNAs (32). To explore active 5'-tRNA halves in endosomal TLR stimulation, we focused on abundant 5'-tRNA half species whose read numbers in the sequencing data of HMDM

EV samples were >1% of total 5'-tRNA half reads (*SI Appendix, Table S1*). Given that the human genome encodes 55 cytoplasmic (cyto) tRNA isoacceptors, each with distinct anticodon sequences (40), the 17 species of 5'-tRNA halves that were extracted originate from a specific subset of cyto tRNA isoacceptors: cyto tRNA^{ValCAC/AAC}, tRNA^{HisGUG}, tRNA^{GlyGCC/CCC}, and tRNA^{GluCUC} (the 5'-tRNA half-derived sequences of some isodecoders of tRNA^{ValCAC} and tRNA^{ValAAC}, or tRNA^{GlyGCC} and tRNA^{GlyCCC}, are identical and thus indistinguishable). The four cyto tRNAs were in aggregate the sources of over 90% of the identified EV-5'-tRNA halves (*SI Appendix, Table S2*). Among 5'-tRNA halves from these tRNAs, the 5'-tRNA^{HisGUG} half has already been shown to activate TLR7, while the 5'-tRNA^{GluCUC} half was inactive (32). We further explored the activity of the rest of the major 5'-tRNA halves, 5'-tRNA^{ValCAC/AAC} half, and 5'-tRNA^{GlyGCC/CCC} half, whose most abundant sequences are derived from nucleotide position [np: as per the nucleotide numbering system specific to tRNAs (41)] 1 to 33 and 1 to 32, respectively (*SI Appendix, Tables S1 and S3*).

To deliver the 5'-tRNA halves into the endosomes of HMDMs, the cells were transfected with the RNAs using the cationic liposome 1,2-dioleoyloxy-3-trimethylammonium-propane (DOTAP). DOTAP, serving as an EV mimic, actively facilitates the delivery of encapsulated RNAs into the endosomes of recipient cells and has been widely adopted for the endosomal delivery of RNAs in previous studies (32, 42–44). As a negative control, an inactive mutant of 20-nucleotide (nt) HIV-1-derived ssRNA termed ssRNA41 (9, 32) was also transfected. As shown in Fig. 1*A*, transfections of the 5'-tRNA^{ValCAC/AAC} half, as well as the 5'-tRNA^{HisGUG} half, enhanced the levels of TNF α and IL-1 β mRNAs, whereas transfections of the 5'-tRNA^{GlyGCC/CCC} half and ssRNA41 did not. We further observed the induction of the secretion of cytokines, such as TNF α , IL-1 β , IL-12p40, and IL-6, into the culture medium upon the transfection of the 5'-tRNA^{ValCAC/AAC} half and the 5'-tRNA^{HisGUG} half (Fig. 1*B*). The upregulation of cytokine expression upon DOTAP-mediated transfection of the two 5'-tRNA halves was not observed in both of the two clones of *TLR7* knockout (KO) HMDMs whose *TLR7* expression was completely depleted (32) (Fig. 1*C*). Collectively, these results identified the 5'-tRNA^{ValCAC/AAC} half, in addition to the 5'-tRNA^{HisGUG} half, as a TLR7-stimulating molecule, while the 5'-tRNA^{GlyGCC/CCC} half has been confirmed to be inactive in endosomal TLR stimulation.

Earlier studies have shown that posttranscriptional modifications in tRNAs or other RNAs can affect their activity in stimulating endosomal TLRs (45–47). Among the two active 5'-tRNA halves, our previous study has demonstrated that the 5'-tRNA^{HisGUG} half containing endogenous modifications [dihydrouridine (D) at np 16, 19, and 20, and pseudouridine (Ψ) at np 32] maintains a similar potency to its unmodified version in stimulating TLR7. Regarding the 5'-tRNA^{ValCAC/AAC} half, based on the information from mature tRNA^{ValCAC/AAC} (48), it is expected to contain the following three species of modifications: Ψ at np 13 and 27; D at np 16, 19, and 20; and 2-methylguanosine (m²G) at np 26. As shown in Fig. 1*D*, the 5'-tRNA^{ValCAC/AAC} half containing all these modifications exhibited approximately 30 to 40% reduced activity compared to its unmodified version. These results suggest that the modified endogenous 5'-tRNA^{ValCAC/AAC} half is still a potent TLR7 activator, although the modifications negatively influence the activity.

We further examined whether the 5'-tRNA half-mediated pathways contribute to bacterial elimination. After DOTAP-mediated transfection of the 5'-tRNA^{HisGUG} half or 5'-tRNA^{ValCAC/AAC} half, HMDMs were infected with *Escherichia coli*. Subsequent analysis

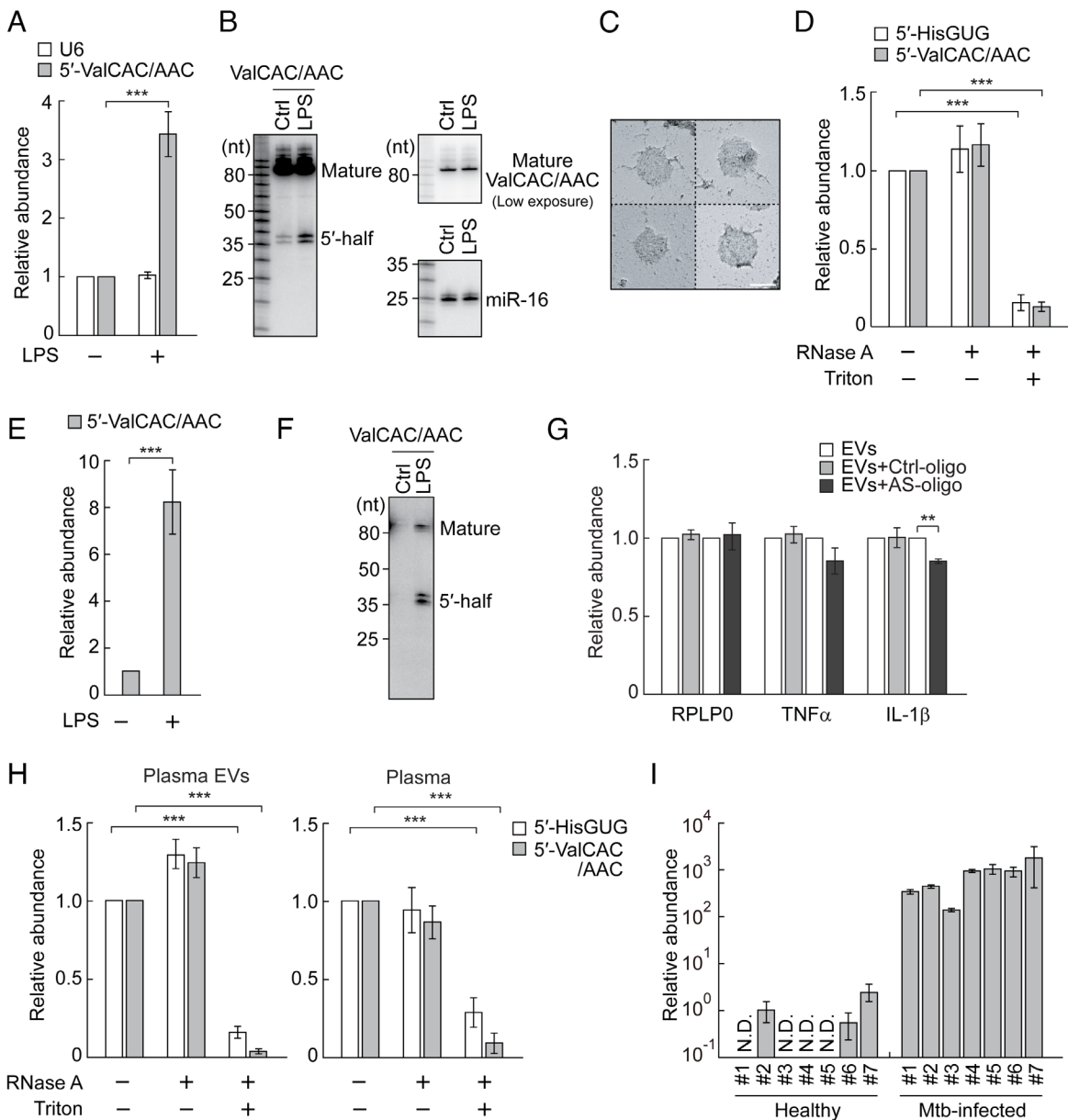


Fig. 2. Enhanced accumulation of the ex-5'-tRNA^{Val}CAC/AAC half in EVs from LPS-treated HMDMs and in plasma from Mtb-infected patients. (A) Total RNAs from HMDMs treated with LPS were subjected to TaqMan RT-qPCR for the 5'-tRNA^{Val}CAC/AAC half. U6 snRNA was quantified as a control. The quantified RNA levels were normalized to the levels of 5S rRNA. (B) Total RNAs from HMDMs treated with LPS were subjected to northern blot for the 5'-tRNA^{Val}CAC/AAC half and its corresponding mature tRNA. miR-16 was analyzed as a control. (C) Transmission electron microscopic evaluation of the isolated EVs. Four representative EV images are shown. (Scale bar, 100 nm.) (D) HMDM-secreted EVs were treated with RNase A and/or Triton X-100 and then subjected to TaqMan RT-qPCR for quantification of 5'-tRNA halves. (E and F) Total RNAs from HMDM-secreted EVs were subjected to TaqMan RT-qPCR (E) and northern blot (F) for the 5'-tRNA^{Val}CAC/AAC half. (G) EVs from LPS-treated HMDMs were mixed with DOTAP-fused AS-oligo of the 5'-tRNA^{Val}CAC/AAC half or Ctrl-oligo with scrambled sequences and applied to recipient HMDMs, followed by quantification of the indicated mRNAs. (H) Human plasma and its isolated EVs were treated with RNase A and/or Triton X-100 and then subjected to TaqMan RT-qPCR for quantification of 5'-tRNA halves. (I) RNAs isolated from plasma samples of healthy individuals or Mtb-infected patients were subjected to TaqMan RT-qPCR for the 5'-tRNA^{Val}CAC/AAC half. The quantified 5'-tRNA half levels were normalized to spike-in RNA levels. The relative abundances to the healthy sample #2 (set as 1) are shown.

and western blot, verified the successful isolation of HMDM EVs as in our previous study (32). EVs treated solely with RNase A and those left untreated showed comparable amplification signals in TaqMan RT-qPCR for the 5'-tRNA^{Val}CAC/AAC half. In contrast, EVs treated with both RNase A and a detergent exhibited significantly diminished signals (Fig. 2D), confirming that the 5'-tRNA^{Val}CAC/AAC half is located inside the EVs, rather than present as non-EV contaminants. LPS treatment up-regulated the levels of the 5'-tRNA^{Val}CAC/AAC half not only in HMDMs (Fig. 2A and B) but also in their EVs (Fig. 2E and F); the fold change of the upregulation was more evident in EVs than in HMDMs. In our northern blot analyses, the lower 33 nt band of

the 5'-tRNA^{Val}CAC/AAC half is more abundant than its longer 34 nt band in EVs (Fig. 2F), while the 34 nt band is more prominent than the 33 nt band in HMDMs (Fig. 2B). These results are consistent with our sequencing data (32), and suggest selective packaging of the 33 nt version of the 5'-tRNA^{Val}CAC/AAC half into EVs. Collectively, these results suggest that the 5'-tRNA^{Val}CAC/AAC half is an immune-responsive molecule whose accumulation and packaging to EVs are induced by surface TLR activation.

Considering the abundant accumulation of the 5'-tRNA^{Val}CAC/AAC half in HMDM EVs, we further analyzed the activity of the EV-contained, endogenous 5'-tRNA^{Val}CAC/AAC half in stimulating TLR7 by utilizing antisense oligonucleotide (AS-oligo) of the

5'-tRNA^{ValCAC/AAC} half and control oligonucleotide (Ctrl-oligo) with scrambled sequences. Application of the isolated HMDM EVs to recipient HMDMs up-regulated the levels of TNF α and IL-1 β mRNA as described in our previous study (32). The AS-oligo reduced the EV-induced upregulation of cytokine mRNAs, while Ctrl-oligo did not show such activity (Fig. 2G). The reduction by the AS-oligo was not as pronounced as that observed for the 5'-tRNA^{HisGUG} half in our previous study (32). This difference might be due to the inherently lower TLR7 stimulation activity of the endogenous 5'-tRNA^{ValCAC/AAC} half compared to the 5'-tRNA^{HisGUG} half. While posttranscriptional modifications do not adversely affect the TLR7 stimulation activity of the 5'-tRNA^{HisGUG} half (32), they reduce the activity of the 5'-tRNA^{ValCAC/AAC} half (Fig. 1D). Despite this, our findings suggest that the endogenous 5'-tRNA^{ValCAC/AAC} half, incorporated in EVs, still possesses significant potential to induce cytokine production by stimulating TLR7.

We further examined the accumulation of the 5'-tRNA^{ValCAC/AAC} half in human plasma samples. Our upgraded version of TaqMan RT-qPCR for limited amounts of RNA materials (49) successfully detected the 5'-tRNA^{ValCAC/AAC} half in the EVs isolated from human plasma (Fig. 2H). As in the case of HMDM EVs, drastic reduction of amplification signals upon treatments with both RNase A and detergent, but not with RNase A treatment alone, confirmed the presence of the 5'-tRNA^{ValCAC/AAC} half inside the plasma EVs. As the quantification of the 5'-tRNA half using plasma (not the isolated plasma EVs) exhibited similar amplification patterns (Fig. 2H), the detected 5'-tRNA^{ValCAC/AAC} half molecules were anticipated to predominantly reside within plasma EVs. Subsequently, we quantified the abundance of the 5'-tRNA^{ValCAC/AAC} half in the plasma samples from healthy individuals or patients infected with Mtb. Considering the potential effects of sex hormones (36) and aging (50) on tRNA half expression, we restricted our analysis to males aged 30 to 35 y, aligning with the parameters of our prior research (32, 49). During the RNA extraction process, we included a synthetic control RNA as a spike-in to serve as a reference for normalization. As shown in Fig. 2I, the levels of the plasma 5'-tRNA^{ValCAC/AAC} half were drastically enhanced by approximately 1,000-fold in Mtb-infected patients compared to healthy individuals. These findings demonstrate that the upregulation and secretion of the 5'-tRNA^{ValCAC/AAC} half during the immune response occur not only in cell culture conditions but also in the plasma of patients infected with microbes, indicating its relevance in actual pathological contexts.

The Presence of Successive Uridine Sequences in Single-Stranded Regions of 5'-tRNA Halves Is Not Sufficient for TLR7 Activation. Considering that the 5'-tRNA^{HisGUG} half and the 5'-tRNA^{ValCAC/AAC} half are active but the 5'-tRNA^{GluCUC} half and the 5'-tRNA^{GlyGCC/CCC} half are inactive in TLR7 stimulation, we attempted to characterize sequence determinants required for TLR7 stimulation. TLR7 possesses two ligand-binding sites for guanosine and ssRNA, respectively (10). Successive uridine sequences (such as UU and UUU) in ligand ssRNAs have been shown to be important for TLR7-binding (51), and indeed, both of the two active 5'-tRNA halves contain successive Us in single-stranded regions in their secondary structures; the 5'-tRNA^{HisGUG} half contains one UU in a loop and one UU at the 3'-end, while the 5'-tRNA^{ValCAC/AAC} half contains two UUs in loops and one UUU at the 5'-end (*SI Appendix, Fig. S2 A and B*). To explore other active 5'-tRNA halves identified in HMDM-secreted EVs, we screened the most abundant sequences of the ex-5'-tRNA halves for the presence of successive Us in single-stranded regions (*SI Appendix, Table S4*), leading us to select four 5'-tRNA halves

(5'-tRNA^{AsnGUU} half, 5'-tRNA^{GluUUC} half, 5'-tRNA^{PheGAA} half, and 5'-tRNA^{ProAGG/UGG/CGG} half) with more than two UUs in single-stranded regions (*SI Appendix, Fig. S2C*) for examination of their activity in endosomal TLR stimulation. As shown in Fig. 3A and B, none of the four 5'-tRNA halves showed the strong activity of cytokine induction upon their DOTAP-mediated transfection, while the two active 5'-tRNA halves served as positive controls, suggesting that the presence of successive Us in the single-stranded region is not sufficient for TLR7 activation.

The Presence of Terminal GUUU Sequences in 5'-tRNA Halves Is Necessary for TLR7 Activation. To further explore sequence determinants of active 5'-tRNA halves, we made various mutants of the 5'-tRNA^{HisGUG} half and the 5'-tRNA^{ValCAC/AAC} half whose successive Us were replaced with As (Fig. 4A and *SI Appendix, Fig. S2 A and B*). To keep the secondary structures of the mutants identical to those of corresponding wild-type molecules, Us were replaced with G or C when replacement with A affected the secondary structure. The mutant 5'-tRNA^{HisGUG} half lacking both of the two UUs (H1) did not activate TLR7 (Fig. 4B), indicating the necessity of successive Us. The mutant 5'-tRNA^{HisGUG} half lacking either one of the two UUs (H2 and H3) showed significantly reduced activity (Fig. 4B), suggesting that two UUs are required to strongly activate TLR7. As in the case of the 5'-tRNA^{HisGUG} half, the mutant 5'-tRNA^{ValCAC/AAC} half lacking three successive Us (V1) did not show TLR7 stimulation activity (Fig. 4C). However, the mutant retaining UUU at the 5'-end but lacking two UUs in loop regions (V2) activated TLR7 as strongly as the wild-type molecule, while the mutants containing only one UU in loop regions (V3 and V4) did not strongly activate TLR7 (Fig. 4C). The replacement of the 5'-terminal G (next to the UUU sequence) with C (V5) abolished the activity (Fig. 4C), suggesting that 5'-terminal GUUU sequences are necessary for TLR7 stimulation activity. Replacement of the UUU with UU (V6) also abolished the activity (Fig. 4C), indicating that three Us are required and two Us are not enough to stimulate TLR7. Collectively, these results suggest that the 5'-terminal GUUU sequences in the 5'-tRNA^{ValCAC/AAC} half are important determinants for TLR7 stimulation activity.

To test whether 3'-terminal UUUU sequences also act as the determinant, we added U to 3'-terminal UU sequences in the mutant H2 to make a 5'-tRNA^{HisGUG} half containing 3'-terminal UUUG sequences but lacking any other successive U sequences (mutant H4, Fig. 4A and *SI Appendix, Fig. S2A*). As shown in Fig. 4B, the changes partially restored the TLR7 stimulation activity. This suggests that not only the 5'-terminal GUUU but also the 3'-terminal UUUG could be important determinants for TLR7 activation, although changing 3'-terminal UUG sequences of the wild-type 5'-tRNA^{HisGUG} half to UUUG (mutant H5, *SI Appendix, Fig. S2A*) resulted in TLR7 stimulation activity that was comparable to, but not greater than, that of the wild-type molecule (*SI Appendix, Fig. S3*).

The Presence of Terminal GUUU Sequences Could Be Sufficient for Endosomal TLR Stimulation. Although it has been reported that mature, full-length tRNAs accumulate in EVs (52), it is noteworthy that the full-length tRNA^{ValCAC/AAC} cannot activate TLR7 (Fig. 5A–C). Unlike the 5'-tRNA^{ValCAC/AAC} whose 5'-terminal GUUU is located in the single-stranded region, the 5'-terminal GUUU sequences in the full-length tRNA form base-pairs in the acceptor stem, suggesting that the terminal GUUU sequences need to exist as ssRNAs to act as TLR7 ligands. Given the terminal GUUU sequences are important for TLR7

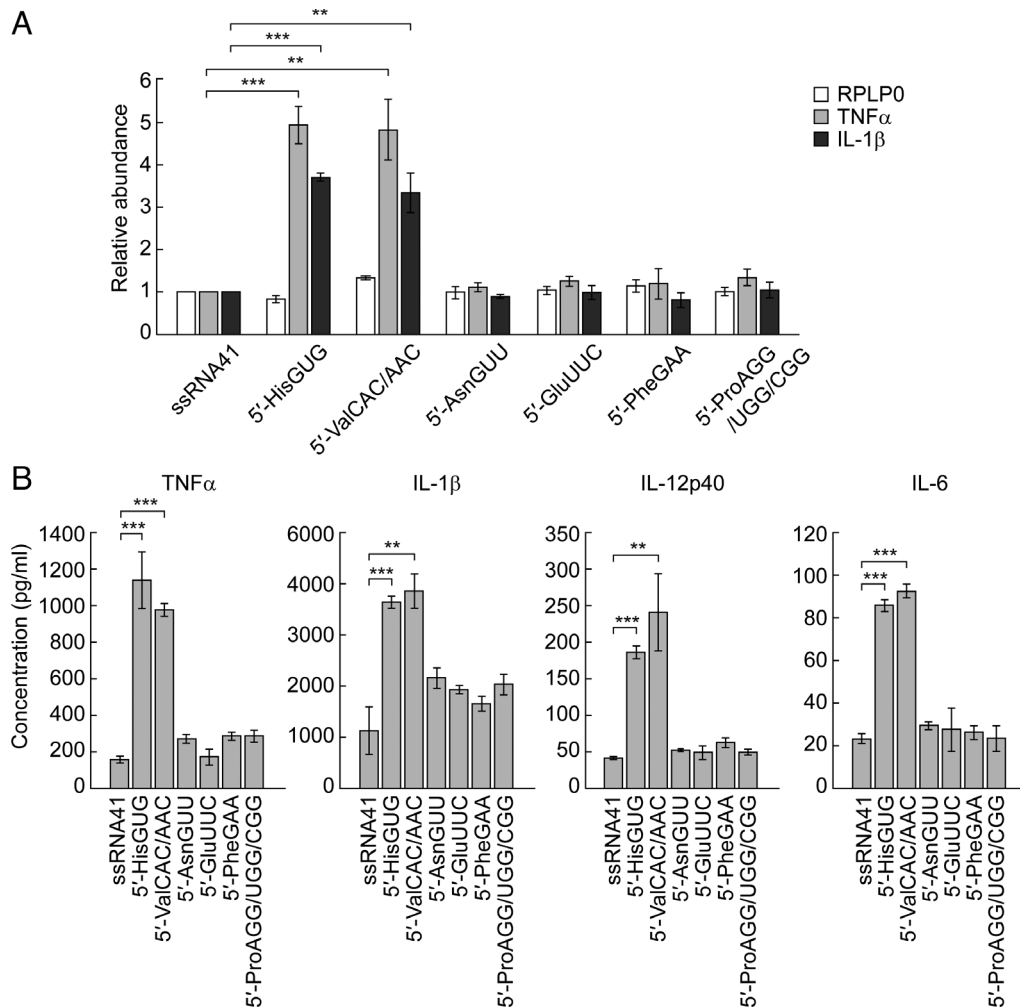


Fig. 3. The 5'-tRNA^{HisGUG} half and the 5'-tRNA^{ValCAC/AAC} half are the only 5'-tRNA halves that strongly activate endosomal TLR. (A) The indicated 5'-tRNA halves and ssRNA41 (negative control) were subjected to DOTAP-mediated transfection into HMDMs. Total RNAs from the cells were subjected to RT-qPCR for the indicated mRNAs. (B) After the RNA transfection, the culture medium was subjected to measurement of the concentration of the indicated cytokines.

activation, we further explored endogenous RNA molecules containing terminal GUUU sequences; our screening of human miRNA sequences extracted three miRNAs (miR-122-5p, miR-548ah-5p, and miR-552-5p) as those that contain 5'-terminal GUUU or 3'-terminal UUUG sequences. As shown in Fig. 5 A–C, DOTAP-mediated transfection of all three miRNAs induced cytokine secretions, suggesting that all of the examined miRNAs are active in endosomal TLR stimulation. These results suggested the universality of the single-stranded terminal GUUU sequences to act as a TLR7 stimulator.

GUUU Is the Best Ratio of Guanosine and Uridine for Endosomal TLR Activation. The ligands for the first and second binding pockets of TLR7 are G nucleotide and ssRNAs with successive Us, respectively (10, 51). To determine the optimal ratio of G and U nucleotides, we investigated the activity of 33-nt ssRNAs with repeat sequences of GU, GUU (GU₂), GUUU (GU₃), GUUUU (GU₄), GUUUUU (GU₅), or poly U with terminal G residues (GU₃1), as well as poly U without any G residues (U₃₃; Fig. 6A). The GUUU repeats exhibited the highest activity in endosomal TLR stimulation following DOTAP transfection (Fig. 6B). Replacement of some GUUU sequences in the GUUU repeats to GU repeats (GU₃-A-C; Fig. 6A) led to the reduction of the activity (Fig. 6C), confirming that the GUUU repeats are the sequences with the optimal G/U sequence signature for endosomal

TLR activation. The activity of poly U (U₃₃) was significantly weaker than that of the GUUU repeats (Fig. 6B), suggesting that the presence of G nucleotides in ssRNA, which would become the ligand of the first binding pocket of TLR7, is important. To explore the role of the length of GUUU repeats in TLR7 activation, we analyzed the activity of ssRNAs with varying numbers of GUUU repeats: GUUU (a single repeat), Oligo-GU₃-3 (3 repeats), Oligo-GU₃-5 (5 repeats), Oligo-GU₃-7 (7 repeats), and Oligo-GU₃ (8 repeats). As shown in Fig. 6D, an increase in GUUU repeat numbers corresponded to enhanced TLR7 stimulation activity. To determine whether a greater quantity of shorter RNAs could offset their lower activity, we further performed the experiments with the adjusted amounts to match the molar quantity of GUUU sequences in Oligo-GU₃. Specifically, we used 8, 2.7, 1.6, and 1.1 times the molar amounts of GUUU, Oligo-GU₃-3, Oligo-GU₃-5, and Oligo-GU₃-7, respectively, compared to Oligo-GU₃. Despite the equivalent amounts of GUUU molecules, the longer RNAs consistently demonstrated higher activity (Fig. 6D), underscoring the significant impact of RNA length on TLR7 stimulation activity. Interestingly, the activity of the 5'-tRNA^{ValCAC/AAC} half was found to be stronger than that of the GUUU repeats with the same length (Oligo-GU₃, Fig. 6E), likely due to unidentified molecular factors. The activity of the 5'-tRNA^{ValCAC/AAC} half in our experiments at a specific time point was further shown to be stronger than that of R848, an agonist of TLR7 and -8 which

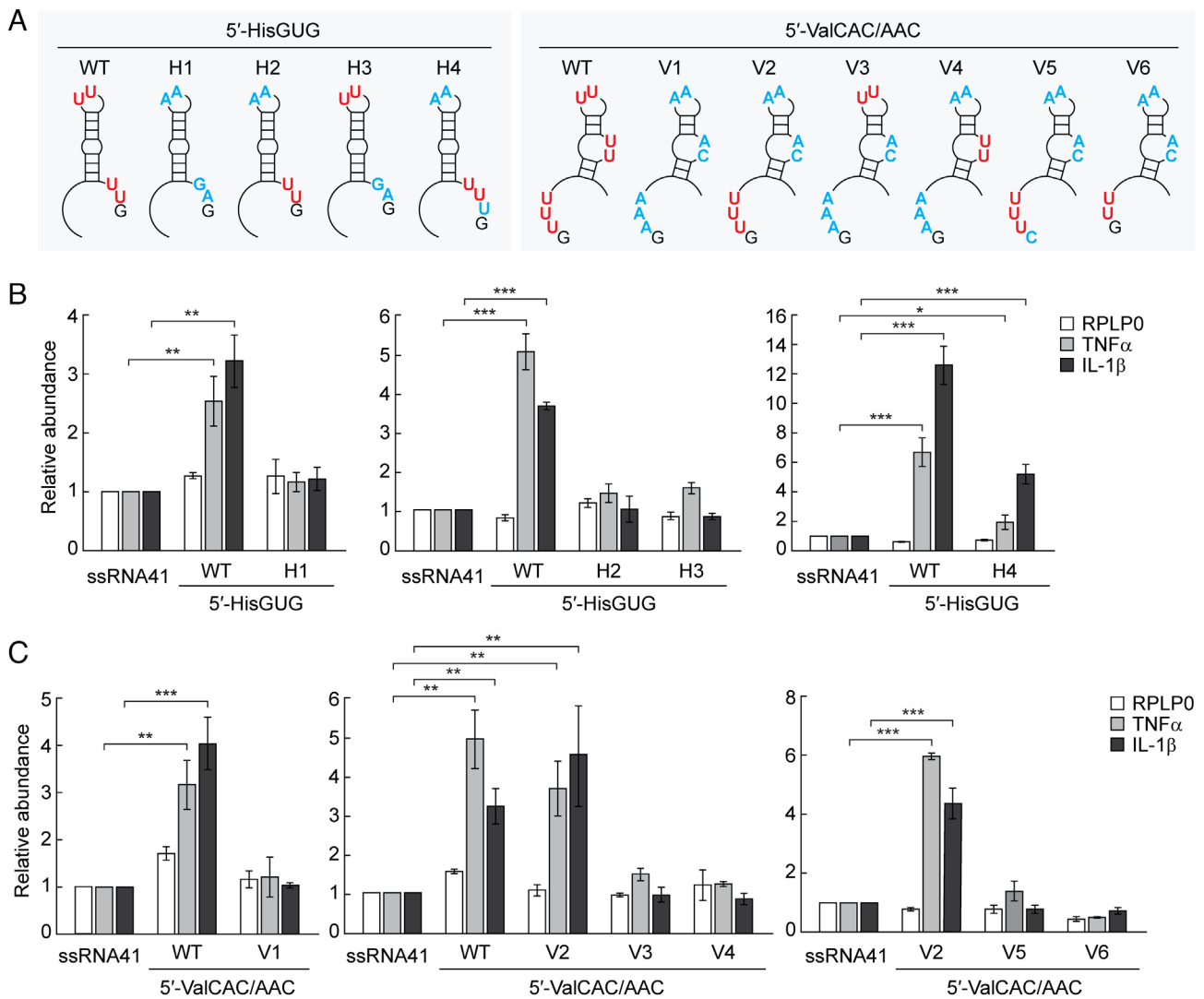


Fig. 4. Determining sequence elements of 5'-tRNA halves required for TLR7 activation. (A) Positions of mutations in respective mutants in the 5'-tRNA^{HisGUG} half and the 5'-tRNA^{ValCAC/AAC} half (the mutated nucleotides are shown in blue). (B and C) The mutants of 5'-tRNA^{HisGUG} half (B) or 5'-tRNA^{ValCAC/AAC} half (C) were subjected to DOTAP-mediated transfection into HMDMs. Total RNAs from the cells were subjected to RT-qPCR for the indicated mRNAs.

have been utilized for therapies of cancer and infectious diseases (53–56) (Fig. 6E).

Discussion

Although miRNAs have been the best-explored endogenous ssRNA ligands for endosomal TLRs thus far (16–18), 3'-OH-containing RNAs, such as miRNAs, are the minor species of ex-sncRNAs (29, 32, 57). On the other hand, it is increasingly acknowledged that tRNA-derived sncRNAs are a plentiful type of ex-RNAs (39, 58), and cP-containing 5'-tRNA half molecules are the most abundant class of ex-tRNA-derived sncRNAs (32). Although recent studies have provided evidence that certain tRNA halves exist in the form of nicked mature tRNAs (59, 60), the scarcity of 3'-tRNA halves in HMDM EVs (32) and in the plasma of Mtb-infected patients (57) suggests that 5'- and 3'-tRNA halves can be regulated through distinct and independent pathways. This aligns well with multiple studies that have reported a much higher accumulation of 5'-tRNA halves compared to 3'-tRNA halves in various cells and tissues (61–65). The much greater abundance of ex-5'-tRNA halves compared to that of miRNAs (32) suggests ex-5'-tRNA halves form a more significant, biologically relevant class of endogenous ligands of immune receptors. Upon conducting a thorough investigation

of ex-5'-tRNA halves in EVs secreted by HMDMs, we have identified the 5'-tRNA^{ValCAC/AAC} half as the most abundant and as an immunostimulatory molecule activating TLR7. This finding represents a second example of an immune-active 5'-tRNA half, with the 5'-tRNA^{HisGUG} half being the first identified (32). We propose that, like cytokines, these 5'-tRNA halves may function as “immune activators” by stimulating TLR7 upon delivery to endosomes in neighboring immune cells and other types of cells via EV-mediated cell–cell communication.

Surface TLR activation induces the accumulation of the 5'-tRNA^{ValCAC/AAC} half in HMDMs and their secreted EVs. The expressional induction and secretion were not limited to cell culture settings but further occurred in plasma samples of Mtb-infected patients which showed dramatic enrichment of the 5'-tRNA^{ValCAC/AAC} half by an approximately 1,000-fold. The enhanced levels of the 5'-tRNA^{ValCAC/AAC} half, as well as the 5'-tRNA^{HisGUG} half (49), and their activity in TLR7 stimulation could be involved in the immune response against Mtb infection. In fact, the genetic polymorphisms in the *TLR7* gene, which reduce its activity, increase susceptibility to Mtb infection (66), and upregulation or stimulation of TLR7 in macrophages suppress Mtb growth (67, 68). In addition to Mtb infection, a dramatic increase in the levels of serum ex-5'-tRNA halves has also been observed in patients infected with hepatitis B

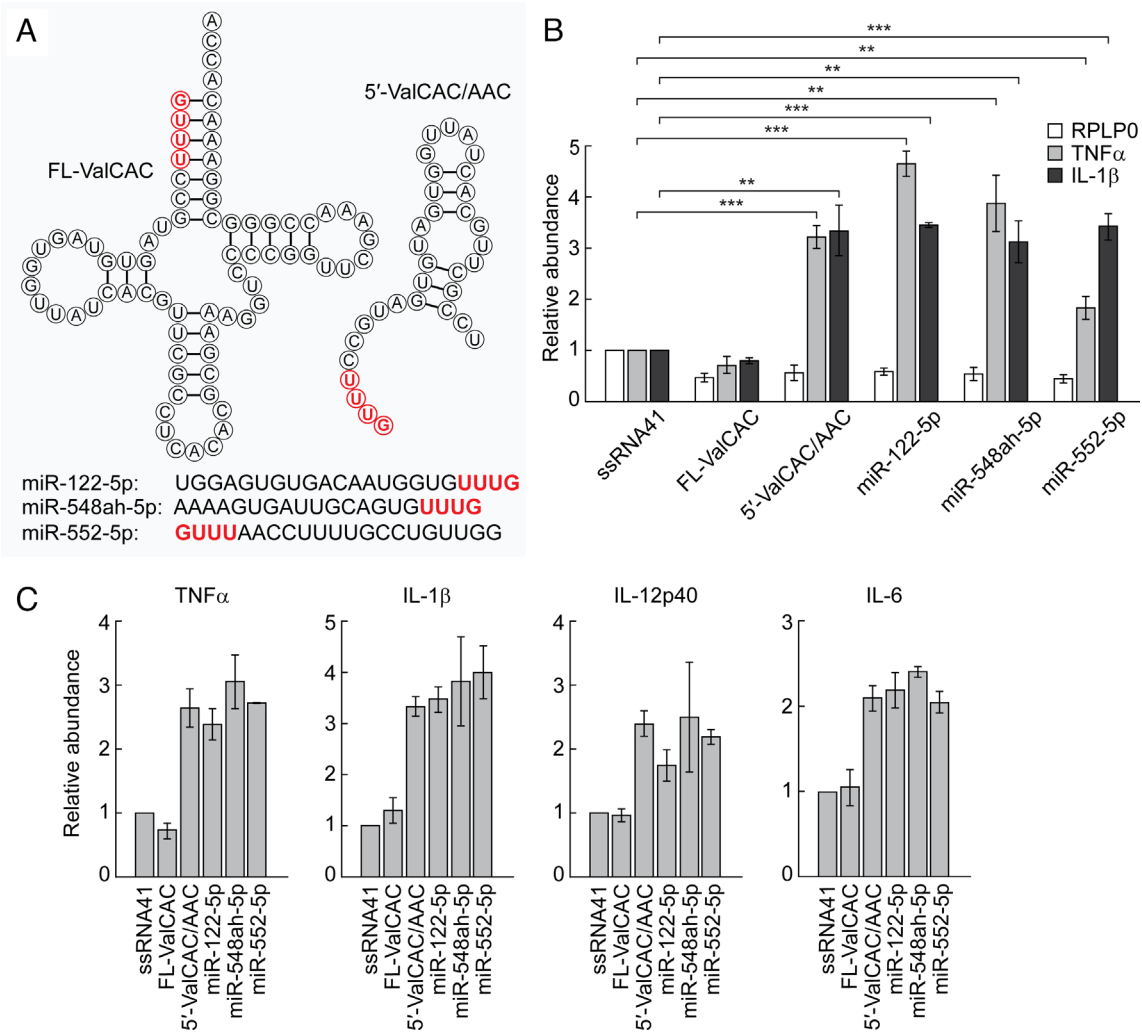


Fig. 5. Requirements of terminal GUUU sequences for TLR7 activation. (A) Sequences of the indicated snRNAs. (B and C) The indicated RNAs were subjected to DOTAP-mediated transfection into HMDMs. Total RNAs from the cells were subjected to RT-qPCR for the indicated mRNAs (B) or culture medium was subjected to measurement of concentration of the indicated cytokines (C).

virus infection (69), and the infections with respiratory syncytial virus (RSV) and *Rickettsia* have been reported to induce the expression of 5'-tRNA halves (70–72). Therefore, the induction of 5'-tRNA halves and their secretion as ex-5'-tRNA halves could potentially be a universal phenomenon across various infectious diseases. Further investigations may establish 5'-tRNA half molecules as potential circulating biomarkers for the noninvasive evaluation of infection severity and immune response status.

Full-length tRNA^{ValCAC/AAC} was incapable of stimulating TLR7. Shortening it by anticodon cleavage into less-rigid 5'-half molecules to expose the 5'-terminal GUUU sequence as the single-stranded region is crucial to producing immunostimulatory snRNAs. We identified ANG as the enzyme which produces immune-responsive tRNA halves in HMDMs (32). An increase in ANG levels has been observed upon RSV and *Rickettsia* infections (72, 73). Further research is warranted on immune-responsive ribonucleases, which cleave tRNAs and other RNAs to produce ex-sncRNAs. ANG-mediated cleavage leaves a cP in 3'-cleavage products (74), and thus 5'-tRNA halves contain a cP (36). Other unidentified immune-responsive ribonucleases, if any, may also produce cP-containing RNAs, because cP-containing RNAs (and possibly 3'-P-containing RNAs) are the majority of ex-sncRNAs (29, 32). The formation of cP by

such enzymes could contribute to TLR7 activation, because cP-containing guanosine (G-cP) exhibits significantly higher affinity compared to guanosine without cP, and thus G-cP is proposed as an endogenous ligand for the first binding site of TLR7 (51). The cP-formation is also implicated in the activation of TLR8 (75). Given that this study employed synthetic RNAs lacking cP in the analysis of TLR7 stimulation activity, further research is necessary to elucidate the potential impact of cP in tRNA halves on the activation of TLR7.

Posttranscriptional modifications of RNA molecules, such as 2'-O-methylation, m⁶A, m⁵C, m⁵U, s²U, and Ψ , have been recognized for their inhibitory effects on the activation of TLR7 and -8 (45, 46, 76–78). The 5'-tRNA^{ValCAC/AAC} half containing modifications showed a reduced TLR7 stimulation activity compared to its unmodified version. Although this experiment was performed under the premise that tRNA half molecules possess modifications identical to those of their corresponding mature tRNAs, and that their modification status remains consistent both within cells and in EVs, further research is required to confirm these assumptions. Despite this, the modified version still retained some level of TLR7 stimulation activity, and it is worth noting that the 5'-tRNA^{ValCAC/AAC} half is the most abundantly accumulated tRNA-derived snRNAs in HMDM EVs. Further, our experiments

TLR7 is involved in various biological processes and diseases (87–92). For example, *TLR7* mutations are associated with the symptom severity of COVID-19 (93–95). TLR7 has also been implicated in the progression of Parkinson's disease (96), Alzheimer's disease (15, 97, 98), and autoimmune diseases such as systemic lupus erythematosus (99) and atherosclerosis (100, 101). Although the growing evidence of TLR7 involvement in the pathogenesis of noninfectious diseases highlights the need for a better understanding of endogenous ssRNA ligands of immune receptors, their exploration and characterization are still at an initial stage, and we are just beginning to appreciate the previously hidden classes of sncRNAs not detected by standard RNA-seq methods. We expect that the 5'-tRNA^{His}GUG half and the 5'-tRNA^{Val}CAC/AAC half are just the early examples of immune receptor-stimulating endogenous ssRNAs, and other ex-sncRNAs would be found to be similarly immunostimulatory through the TLR7 or -8 axis. Beyond tRNA-derived sncRNAs, our first genome-wide identification of cP-containing sncRNAs revealed abundant expression of rRNA- and mRNA-derived sncRNAs in various tissues (50), and, indeed, rRNA- and mRNA-derived sncRNAs have been shown to be abundantly secreted as ex-RNAs (30, 57, 102, 103). Updated sncRNA sequencing methods to address the issue of reverse transcription impairment by posttranscriptional modifications, such as PANDORA-seq (104) and TGIRT-seq (105), present viable options for future exploration of immunostimulatory ex-sncRNAs, especially those focusing on heavily modified sncRNAs such as tRNA- and rRNA-derived sncRNAs. The exploration should review studies looking into the characteristics of ssRNA ligands of TLR7 and -8, including terminal GUUU sequence signatures for TLR7 activation that this study revealed. Further research into endogenous ssRNA ligands of TLR7 and -8 would help our fuller understanding of the immune system in the body's response to diverse pathologies.

Materials and Methods

In Vitro RNA Synthesis. The synthetic RNAs used in this study are shown in [SI Appendix, Table S3](#) [the information on sncRNA "license plates" (106) and names via tDRnamer (107) are included]. These RNAs were synthesized by an in vitro transcription reaction as described previously (32). dsDNA templates were synthesized using PrimeSTAR GXL DNA Polymerase (Takara Bio) and the primers shown in [SI Appendix, Table S5](#). The templates were then subjected to an in vitro transcription reaction with T7 RNA polymerase (New England Biolabs) at 37 °C for 4 h. For the RNAs whose 5'-terminal nucleotide is not G, the in vitro synthesized RNAs contained the ribozyme sequence to generate a mature 5'-end as described previously (108). After incubation for in vitro RNA synthesis, the reaction mixture was further incubated for three cycles at 90 °C for 2.5 min and 37 °C for 15 min, allowing the ribozyme reaction. The synthesized RNAs were then gel-purified using denaturing PAGE with single-nucleotide resolution. For full-length tRNA^{Val}CAC, we performed annealing by incubating it in the annealing buffer, consisting of 50 mM Tris-HCl (pH 8.0) and 100 mM MgCl₂ at 70 °C for 3 min, followed by incubation at 37 °C for 20 min. The 5'-tRNA^{Val}CAC/AAC half containing posttranscriptional modifications (the 5'-ValCAC/AAC-Mod in [SI Appendix, Table S3](#)) was synthesized by Integrated DNA Technologies.

Cell Culture, TLR4 Activation, RNA and R848 Delivery, and Measurement of Cytokine Concentration. THP-1 human acute monocytic leukemia cells (American Type Culture Collection) were cultured in RPMI 1640 medium (Corning) with 10% FBS and differentiated into HMDMs using phorbol 12-myristate 13-acetate (PMA; Sigma-Aldrich), as described previously (32, 109). For activation of TLR4, HMDMs were cultured with a medium containing 100 ng/mL of LPS from *E. coli* O111:B4 (Sigma-Aldrich) for 12 h as described previously (32). *TLR7* KO THP-1 cell lines, whose TLR7 expression is completely depleted, were previously generated using a CRISPR/Cas9 approach (32). Before transfection, the cells were primed with 100 units/mL of interferon γ (Thermo Fisher Scientific) for 18 to 24 h

(42). To deliver RNAs to endosomes, we used the cationic liposome DOTAP (Sigma-Aldrich) as previously described (32, 110, 111). In brief, 230 pmol of synthetic RNAs or Resiquimod (R848, InvivoGen) were mixed with 60 μ L of HBS buffer and 15 μ L of DOTAP reagent and incubated for 15 min. The RNA-DOTAP solution was then added to 1 mL RPMI 1640 medium with 2% FBS, followed by incubation of the cells for 16 h. Cytokine concentrations of the cultured medium of HMDMs were measured by Multiplexing LASER Bead Technology (Eve Technologies) or ELISA kit (R&D Systems), as previously described (32).

EVs and Their Treatments. EVs were isolated from the culture medium of LPS-treated HMDMs according to an ultracentrifugation-based method. As described previously (32), the abundant presence of EVs in the EV solution was confirmed by NTA and transmission electron microscopy (JEM-2100, JEOL) at Materials Characterization Core at Drexel University. To confirm the presence of EV-RNAs, the isolated EVs were incubated with PureLink RNase A (200 ng/ μ L, Thermo Fisher Scientific) with or without 0.1% Triton X-100 at 37 °C for 30 min.

EV-Mediated RNA Delivery to Endosomes. The experiments using EVs and antisense oligonucleotides were performed as described previously (32). AS-oligo or Ctrl-oligo with scrambled sequences ([SI Appendix, Table S3](#)) was first infused with DOTAP as described above. EVs isolated from LPS-treated HMDMs were mixed with the DOTAP-oligonucleotide solution and then were applied to recipient HMDMs, followed by incubation for 16 h. To eliminate possible effects of potential endotoxin (LPS) contamination, EVs isolated from LPS-treated HMDMs were incubated with 10 mg/mL polymyxin B (PMB) (Sigma-Aldrich) at 4 °C for 1 h prior to adding to HMDMs.

Quantification of RNAs by TaqMan RT-qPCR and Standard RT-qPCR. Total RNA was isolated from the cells and EVs using TRIzol (Bioline). TaqMan RT-qPCR for specific quantification of 5'-tRNA halves was performed as described previously (32, 35, 36, 50, 112). Briefly, to remove cP from 5'-tRNA halves, total RNA was treated with T4 polynucleotide kinase (T4 PNK; New England Biolabs), followed by ligation to a 3'-RNA AD by T4 RNA ligase (T4 Rnl; Thermo Fisher Scientific). Ligated RNA was then subjected to TaqMan RT-qPCR using the One Step PrimeScript RT-PCR Kit (Takara Bio), 200 nM of a TaqMan probe targeting the boundary of the target RNA and the 3'-AD, and forward and reverse primers. The TaqMan probe and primer sequences are shown in [SI Appendix, Table S6](#). Although both 33-nt and 34-nt versions of the tRNA^{Val}CAC/AAC half are present in HMDMs and their EVs, the TaqMan probe designed for this study specifically targets the 33-nt version. Due to limited amounts of starting RNA materials, for quantification of plasma 5'-tRNA halves, we utilized an upgraded, more efficient version of TaqMan RT-qPCR in which T4 PNK treatment and 3'-AD ligation are simultaneously performed in a one-step reaction (49). For quantification of mRNAs by standard RT-qPCR, total RNA was treated with DNase I (Promega) and subjected to reverse transcription using RevertAid Reverse Transcriptase (Thermo Fisher Scientific) and a reverse primer. The synthesized cDNAs were then subjected to PCR using 2 \times qPCR Master Mix (Bioland Scientific) and forward and reverse primers. Sequences of the primers used are shown in [SI Appendix, Table S7](#).

Northern Blot. Northern blot was performed as previously described (36) with the following antisense probes: 5'-tRNA^{Val}CAC/AAC half, 5'-GCGAACGTGATAACCTAC-3'; and miR-16, 5'-GCCAATATTACGTGCTGCTA-3'.

Bacterial Infection and Invasion Assay. After 16 h of DOTAP transfection of RNAs, HMDMs (1 \times 10⁶ cells) were plated on 6-well plates and were incubated with *E. coli* [100 multiplicities of infection (MOI)] in RPMI 1640 (no antibiotics) for 60 min at 37 °C. HMDMs were then washed with PBS three times and incubated with RPMI 1640 containing high concentration (3 \times) of penicillin-streptomycin (Thermo Fisher Scientific) at 37 °C for 60 min. Subsequently, the medium was replaced with RPMI 1640 containing normal concentration (1 \times) of penicillin-streptomycin, followed by further incubation at 37 °C for 24 h. HMDMs were then washed and lysed with 0.5% Triton X-100. Intracellular bacteria were enumerated by plating on LB agar plates.

Ethical Approval, Human Plasma Samples, and RNA Isolation. The Office of Human Research of Thomas Jefferson University approved our use of patient samples without private information in accordance with all federal, institutional,

and ethical guidelines. We obtained the plasma samples from a biological specimen company, BioIVT, without receiving patients' information. The plasma samples were derived from healthy or Mtb-infected males aged 30 to 35 y. RNA isolation from the plasma samples was performed as described previously (32, 49). First, 500 μ L plasma was centrifuged at 16,060 g for 5 min, and then 400 μ L supernatant was mixed with 1 fmol of the spike-in control RNA (*SI Appendix, Table S3*) and subjected to RNA extraction using TRIzol LS Reagent (Thermo Fisher Scientific). The extracted RNAs were further subjected to purification using the miRNeasy Mini Kit (Qiagen).

Data, Materials, and Software Availability. There are no data underlying this work.

1. R. Medzhitov, Origin and physiological roles of inflammation. *Nature* **454**, 428–435 (2008).
2. O. Takeuchi, S. Akira, Pattern recognition receptors and inflammation. *Cell* **140**, 805–820 (2010).
3. H. Kono, K. L. Rock, How dying cells alert the immune system to danger. *Nat. Rev. Immunol.* **8**, 279–289 (2008).
4. T. Gong, L. Liu, W. Jiang, R. Zhou, DAMP-sensing receptors in sterile inflammation and inflammatory diseases. *Nat. Rev. Immunol.* **20**, 95–112 (2020).
5. B. C. Remick, M. M. Gaidt, R. E. Vance, Effector-triggered immunity. *Annu. Rev. Immunol.* **41**, 453–481 (2023).
6. T. Kawasaki, T. Kawai, Toll-like receptor signaling pathways. *Front. Immunol.* **5**, 461 (2014).
7. J. Zindl, P. Kubes, DAMPs, PAMPs, and LAMPs in immunity and sterile inflammation. *Annu. Rev. Pathol.* **15**, 493–518 (2020).
8. X. Tan, L. Sun, J. Chen, Z. J. Chen, Detection of microbial infections through innate immune sensing of nucleic acids. *Annu. Rev. Microbiol.* **72**, 447–478 (2018).
9. F. Heil *et al.*, Species-specific recognition of single-stranded RNA via Toll-like receptor 7 and 8. *Science* **303**, 1526–1529 (2004).
10. Z. Zhang *et al.*, Structural analysis reveals that Toll-like receptor 7 is a dual receptor for guanosine and single-stranded RNA. *Immunity* **45**, 737–748 (2016).
11. J. M. Lund *et al.*, Recognition of single-stranded RNA viruses by Toll-like receptor 7. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 5598–5603 (2004).
12. J. Melchjorsen *et al.*, Activation of innate defense against a paramyxovirus is mediated by RIG-I and TLR7 and TLR8 in a cell-type-specific manner. *J. Virol.* **79**, 12944–12951 (2005).
13. K. Triantafyllou *et al.*, Human cardiac inflammatory responses triggered by Coxsackie B viruses are mainly Toll-like receptor (TLR) 8-dependent. *Cell Microbiol.* **7**, 1117–1126 (2005).
14. M. P. Gantier *et al.*, Genetic modulation of TLR8 response following bacterial phagocytosis. *Hum. Mutat.* **31**, 1069–1079 (2010).
15. S. M. Lehmann *et al.*, An unconventional role for miRNA: Let-7 activates Toll-like receptor 7 and causes neurodegeneration. *Nat. Neurosci.* **15**, 827–835 (2012).
16. M. Fabbri, A. Paoe, F. Calore, R. Galli, C. M. Croce, A new role for microRNAs, as ligands of Toll-like receptors. *RNA Biol.* **10**, 169–174 (2013).
17. X. Chen, H. Liang, J. Zhang, K. Zen, C. Y. Zhang, microRNAs are ligands of Toll-like receptors. *RNA* **19**, 737–739 (2013).
18. R. Bayraktar, M. T. S. Bertilaccio, G. A. Calin, The interaction between two worlds: MicroRNAs and Toll-like receptors. *Front. Immunol.* **10**, 1053 (2019).
19. Y. Feng *et al.*, Extracellular microRNAs induce potent innate immune responses via TLR7/MyD88-dependent mechanisms. *J. Immunol.* **199**, 2106–2117 (2017).
20. L. Zou *et al.*, Brain innate immune response via miRNA-TLR7 sensing in polymicrobial sepsis. *Brain Behav. Immun.* **100**, 10–24 (2022).
21. M. Fabbri, TLRs as miRNA receptors. *Cancer Res.* **72**, 6333–6337 (2012).
22. L. Casadei *et al.*, Exosome-derived miR-25-3p and miR-92a-3p stimulate liposarcoma progression. *Cancer Res.* **77**, 3846–3856 (2017).
23. W. C. Wu, S. J. Song, Y. Zhang, X. Li, Role of extracellular vesicles in autoimmune pathogenesis. *Front. Immunol.* **11**, 579043 (2020).
24. V. Salvi *et al.*, Exosome-delivered microRNAs promote IFN- α secretion by human plasmacytoid DCs via TLR7. *JCI Insight* **3**, e98204 (2018).
25. M. Groot, H. Lee, Sorting mechanisms for microRNAs into extracellular vesicles and their associated diseases. *Cells* **9**, 1044 (2020).
26. M. Shigematsu, T. Kawamura, Y. Kirino, Generation of 2',3'-cyclic phosphate-containing RNAs as a hidden layer of the transcriptome. *Front. Genet.* **9**, 562 (2018).
27. M. Shigematsu, Y. Kirino, Making invisible RNA visible: Discriminative sequencing methods for RNA molecules with specific terminal formations. *Biomolecules* **12**, 611 (2022).
28. J. Gumas, Y. Kirino, Challenges inherent to the sequencing and quantification of short non-coding RNAs and how to address them. *Clin. Transl. Discovery* **2**, e93 (2022).
29. M. D. Giraldez *et al.*, Phospho-RNA-seq: A modified small RNA-seq method that reveals circulating mRNA and lncRNA fragments as potential biomarkers in human plasma. *EMBO J.* **38**, e101695 (2019).
30. K. M. Akat *et al.*, Detection of circulating extracellular mRNAs by modified small-RNA-sequencing analysis. *JCI Insight* **5**, e127317 (2019).
31. Y. Qin *et al.*, High-throughput sequencing of human plasma RNA by using thermostable group II intron reverse transcriptases. *RNA* **22**, 111–128 (2016).
32. K. Pawar, M. Shigematsu, S. Sharbati, Y. Kirino, Infection-induced 5'-half molecules of tRNA^{His}GUG activate Toll-like receptor 7. *PLoS Biol.* **18**, e3000982 (2020).
33. S. Yamasaki, P. Ivanov, G. F. Hu, P. Anderson, Angiogenin cleaves tRNA and promotes stress-induced translational repression. *J. Cell Biol.* **185**, 35–42 (2009).
34. H. Fu *et al.*, Stress induces tRNA cleavage by angiogenin in mammalian cells. *FEBS Lett.* **583**, 437–442 (2009).
35. M. Shigematsu, Y. Kirino, Oxidative stress enhances the expression of 2',3'-cyclic phosphate-containing RNAs. *RNA Biol.* **17**, 1060–1069 (2020).
36. S. Honda *et al.*, Sex hormone-dependent tRNA halves enhance cell proliferation in breast and prostate cancers. *Proc. Natl. Acad. Sci. U.S.A.* **112**, E3816–E3825 (2015).
37. M. M. Emara *et al.*, Angiogenin-induced tRNA-derived stress-induced RNAs promote stress-induced stress granule assembly. *J. Biol. Chem.* **285**, 10959–10968 (2010).
38. P. Ivanov, M. M. Emara, J. Villen, S. P. Gygi, P. Anderson, Angiogenin-induced tRNA fragments inhibit translation initiation. *Mol. Cell* **43**, 613–623 (2011).
39. J. Gumas, Y. Kirino, Extracellular tRNA-derived RNAs as emerging activators of endosomal Toll-like receptors: A narrative review. *ExRNA* **5**, 0002 (2023).
40. P. P. Chan, T. M. Lowe, tRNAdb: A database of transfer RNA genes detected in genomic sequence. *Nucleic Acids Res.* **37**, D93–D97 (2009).
41. M. Sprinzl, C. Horn, M. Brown, A. Ioudovitch, S. Steinberg, Compilation of tRNA sequences and sequences of tRNA genes. *Nucleic Acids Res.* **26**, 148–153 (1998).
42. M. P. Gantier *et al.*, TLR7 is involved in sequence-specific sensing of single-stranded RNAs in human macrophages. *J. Immunol.* **180**, 2117–2124 (2008).
43. M. Sioud, Single-stranded small interfering RNA are more immunostimulatory than their double-stranded counterparts: A central role for 2'-hydroxyl uridines in immune responses. *Eur. J. Immunol.* **36**, 1222–1230 (2006).
44. A. Forsbach *et al.*, Identification of RNA sequence motifs stimulating sequence-specific TLR8-dependent immune responses. *J. Immunol.* **180**, 3729–3738 (2008).
45. K. Kariko, M. Buckstein, H. Ni, D. Weissman, Suppression of RNA recognition by Toll-like receptors: The impact of nucleoside modification and the evolutionary origin of RNA. *Immunity* **23**, 165–175 (2005).
46. S. Jockel *et al.*, The 2'-O-methylation status of a single guanosine controls transfer RNA-mediated Toll-like receptor 7 activation or inhibition. *J. Exp. Med.* **209**, 235–241 (2012).
47. S. Gehrig *et al.*, Identification of modifications in microbial, native tRNA that suppress immunostimulatory activity. *J. Exp. Med.* **209**, 225–233 (2012).
48. E. Y. Chen, B. A. Roe, Sequence studies on human placenta tRNAVal: Comparison with the mouse myeloma tRNAVal. *Biochem. Biophys. Res. Commun.* **78**, 631–640 (1977).
49. T. Kawamura, M. Shigematsu, Y. Kirino, In vitro production and multiplex quantification of 2',3'-cyclic phosphate-containing 5'-tRNA half molecules. *Methods* **203**, 335–341 (2022).
50. M. Shigematsu, K. Morichika, T. Kawamura, S. Honda, Y. Kirino, Genome-wide identification of short 2',3'-cyclic phosphate-containing RNAs and their regulation in aging. *PLoS Genet.* **15**, e1008469 (2019).
51. Z. Zhang *et al.*, Structural analyses of Toll-like receptor 7 reveal detailed RNA sequence specificity and recognition mechanism of agonistic ligands. *Cell Rep.* **25**, 3371–3381.e5 (2018).
52. M. J. Shurtleff *et al.*, Broad role for YBX1 in defining the small noncoding RNA composition of exosomes. *Proc. Natl. Acad. Sci. U.S.A.* **114**, E8987–E8995 (2017).
53. S. Y. Kim *et al.*, Lyophilizable and multifaceted Toll-like receptor 7/8 agonist-loaded nanoemulsion for the reprogramming of tumor microenvironments and enhanced cancer immunotherapy. *ACS Nano* **13**, 12671–12686 (2019).
54. D. J. Dowling *et al.*, Development of a TLR7/8 agonist adjuvant formulation to overcome early life hyporesponsiveness to DTap vaccination. *Sci. Rep.* **12**, 16860 (2022).
55. J. Lee *et al.*, Activation of anti-hepatitis C virus responses via Toll-like receptor 7. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 1828–1833 (2006).
56. H. Nian *et al.*, R-848 triggers the expression of TLR7/8 and suppresses HIV replication in monocytes. *BMC Infect. Dis.* **12**, 5 (2012).
57. J. Gumas, T. Kawamura, M. Shigematsu, Y. Kirino, Immunostimulatory short non-coding RNAs in the circulation of patients with tuberculosis infection. *Mol. Ther. Nucleic Acids* **35**, 1–15 (2024).
58. J. P. Tosar, A. Cayota, Extracellular tRNAs and tRNA-derived fragments. *RNA Biol.* **17**, 1149–1167 (2020).
59. B. Costa *et al.*, Nicked tRNAs are stable reservoirs of tRNA halves in cells and biofluids. *Proc. Natl. Acad. Sci. U.S.A.* **120**, e2216330120 (2023).
60. X. Chen, S. L. Wolin, Transfer RNA halves are found as nicked tRNAs in cells: Evidence that nicked tRNAs regulate expression of an RNA repair operon. *RNA* **29**, 620–629 (2023).
61. J. M. Dhabhi *et al.*, 5' tRNA halves are present as abundant complexes in serum, concentrated in blood cells, and modulated by aging and calorie restriction. *BMC Genomics* **14**, 298 (2013).
62. Z. Su, C. Kuscu, A. Malik, E. Shibata, A. Dutta, Angiogenin generates specific stress-induced tRNA halves and is not involved in tRF-3-mediated gene silencing. *J. Biol. Chem.* **294**, 16930–16941 (2019).
63. X. Mo *et al.*, Lactate induces production of the tRNA(His) half to promote B-lymphoblastic cell proliferation. *Mol. Ther.* **28**, 2442–2457 (2020).
64. S. Krishna *et al.*, Dynamic expression of tRNA-derived small RNAs define cellular states. *EMBO Rep.* **20**, e47789 (2019).
65. L. Chen *et al.*, 5' Half of specific tRNAs feeds back to promote corresponding tRNA gene transcription in vertebrate embryos. *Sci. Adv.* **7**, eabh0494 (2021).
66. Y. F. Lai *et al.*, Functional polymorphisms of the TLR7 and TLR8 genes contribute to Mycobacterium tuberculosis infection. *Tuberculosis (Edinb)* **98**, 125–131 (2016).
67. M. Bao, Z. Yi, Y. Fu, Activation of TLR7 inhibition of Mycobacterium tuberculosis survival by autophagy in RAW 264.7 macrophages. *J. Cell Biochem.* **118**, 4222–4229 (2017).
68. H. J. Lee *et al.*, TLR7 stimulation with imiquimod induces selective autophagy and controls Mycobacterium tuberculosis growth in mouse macrophages. *Front. Microbiol.* **11**, 1684 (2020).

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69. Y. Zhang *et al.*, Identification and characterization of an ancient class of small RNAs enriched in serum associating with active infection. *J. Mol. Cell Biol.* **6**, 172–174 (2014).
70. J. Zhou *et al.*, Identification of two novel functional tRNA-derived fragments induced in response to respiratory syncytial virus infection. *J. Gen. Virol.* **98**, 1600–1610 (2017).
71. J. Deng *et al.*, Respiratory syncytial virus utilizes a tRNA fragment to suppress antiviral responses through a novel targeting mechanism. *Mol. Ther.* **23**, 1622–1629 (2015).
72. B. Gong *et al.*, Compartmentalized, functional role of angiogenin during spotted fever group rickettsia-induced endothelial barrier dysfunction: Evidence of possible mediation by host tRNA-derived small noncoding RNAs. *BMC Infect. Dis.* **13**, 285 (2013).
73. Q. Wang *et al.*, Identification and functional characterization of tRNA-derived RNA fragments (tRFs) in respiratory syncytial virus infection. *Mol. Ther.* **21**, 368–379 (2013).
74. R. Shapiro, J. F. Riordan, B. L. Vallee, Characteristic ribonucleolytic activity of human angiogenin. *Biochemistry* **25**, 3527–3532 (1986).
75. W. Greulich *et al.*, TLR8 is a sensor of RNase T2 degradation products. *Cell* **179**, 1264–1275.e13 (2019).
76. K. Rimbach, S. Kaiser, M. Helm, A. H. Dalpke, T. Eigenbrod, 2'-O-methylation within bacterial RNA acts as suppressor of TLR7/TLR8 activation in human innate immune cells. *J. Innate. Immun.* **7**, 482–493 (2015).
77. P. Keller *et al.*, Double methylation of tRNA-U54 to 2'-O-methylthymidine (Tm) synergistically decreases immune response by Toll-like receptor 7. *Nucleic Acids Res.* **46**, 9764–9775 (2018).
78. I. Freund, T. Eigenbrod, M. Helm, A. H. Dalpke, RNA modifications modulate activation of innate Toll-like receptors. *Genes (Basel)* **10**, 92 (2019).
79. B. Kuhle, Q. Chen, P. Schimmel, tRNA renovatio: Rebirth through fragmentation. *Mol. Cell* **83**, 3953–3971 (2023).
80. Q. Chen, T. Zhou, Emerging functional principles of tRNA-derived small RNAs and other regulatory small RNAs. *J. Biol. Chem.* **299**, 105225 (2023).
81. B. Vanwalscappel, T. Tada, N. R. Landau, Toll-like receptor agonist R848 blocks Zika virus replication by inducing the antiviral protein viperin. *Virology* **522**, 199–208 (2018).
82. B. Bahmani *et al.*, Intratumoral immunotherapy using platelet-cloaked nanoparticles enhances antitumor immunity in solid tumors. *Nat. Commun.* **12**, 1999 (2021).
83. K. A. Michaelis *et al.*, The TLR7/8 agonist R848 remodels tumor and host responses to promote survival in pancreatic cancer. *Nat. Commun.* **10**, 4682 (2019).
84. R. Lu *et al.*, Formulation and preclinical evaluation of a Toll-like receptor 7/8 agonist as an anti-tumoral immunomodulator. *J. Control Release* **306**, 165–176 (2019).
85. R. Seki, K. Nishizawa, Use of TLR9 and TLR7/8 agonists in combination with D-galactosamine in exploring models for distinct severities of systemic inflammation relative to liver injury. *Physiol. Res.* **69**, 1125–1129 (2020).
86. N. M. Zahr, Q. Zhao, R. Goodcase, A. Pfefferbaum, Systemic administration of the TLR7/8 agonist resiquimod (R848) to mice is associated with transient, in vivo-detectable brain swelling. *Biology (Basel)* **11**, 274 (2022).
87. N. A. Lind, V. E. Rael, K. Pestal, B. Liu, G. M. Barton, Regulation of the nucleic acid-sensing Toll-like receptors. *Nat. Rev. Immunol.* **22**, 224–235 (2022), 10.1038/s41577-021-00577-0.
88. K. Miyake *et al.*, Nucleic acid sensing by Toll-like receptors in the endosomal compartment. *Front. Immunol.* **13**, 941931 (2022).
89. S. Trivedi, E. L. Greidinger, Endosomal Toll-like receptors in autoimmunity: Mechanisms for clinical diversity. *Therapy* **6**, 433–442 (2009).
90. N. Kano, G. H. Ong, D. Ori, T. Kawai, Pathophysiological role of nucleic acid-sensing pattern recognition receptors in inflammatory diseases. *Front. Cell Infect. Microbiol.* **12**, 910654 (2022).
91. K. E. Crump, S. E. Sahingur, Microbial nucleic acid sensing in oral and systemic diseases. *J. Dent. Res.* **95**, 17–25 (2016).
92. L. He *et al.*, Nucleic acid sensing pattern recognition receptors in the development of colorectal cancer and colitis. *Cell Mol. Life Sci.* **74**, 2395–2411 (2017).
93. T. Asano *et al.*, X-linked recessive TLR7 deficiency in ~1% of men under 60 years old with life-threatening COVID-19. *Sci. Immunol.* **6**, eabl4348 (2021).
94. X. Solanich *et al.*, Genetic screening for TLR7 variants in young and previously healthy men with severe COVID-19. *Front. Immunol.* **12**, 719115 (2021).
95. C. Fallarini *et al.*, Association of Toll-like receptor 7 variants with life-threatening COVID-19 disease in males: Findings from a nested case-control study. *Elife* **10**, e67569 (2021).
96. M. Campolo *et al.*, TLR7/8 in the pathogenesis of Parkinson's disease. *Int. J. Mol. Sci.* **21**, 9384 (2020).
97. S. Frank, E. Copanaki, G. J. Burbach, U. C. Muller, T. Deller, Differential regulation of toll-like receptor mRNAs in amyloid plaque-associated brain tissue of aged APP23 transgenic mice. *Neurosci. Lett.* **453**, 41–44 (2009).
98. P. Demby *et al.*, Human endogenous retrovirus HERV-K(HML-2) RNA causes neurodegeneration through Toll-like receptors. *JCI Insight* **5**, e131093 (2020).
99. G. J. Brown *et al.*, TLR7 gain-of-function genetic variation causes human lupus. *Nature* **605**, 349–356 (2022).
100. G. Karadimou *et al.*, Low TLR7 gene expression in atherosclerotic plaques is associated with major adverse cardio- and cerebrovascular events. *Cardiovasc. Res.* **113**, 30–39 (2017).
101. M. Salagianni *et al.*, Toll-like receptor 7 protects from atherosclerosis by constraining "inflammatory" macrophage activation. *Circulation* **126**, 952–962 (2012).
102. M. Li *et al.*, Analysis of the RNA content of the exosomes derived from blood serum and urine and its potential as biomarkers. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **369**, 20130502 (2014).
103. J. Jia *et al.*, Distinct extracellular RNA profiles in different plasma components. *Front. Genet.* **12**, 564780 (2021).
104. J. Shi *et al.*, PANDORA-seq expands the repertoire of regulatory small RNAs by overcoming RNA modifications. *Nat. Cell Biol.* **23**, 424–436 (2021).
105. H. Xu, J. Yao, D. C. Wu, A. M. Lambowitz, Improved TGIRT-seq methods for comprehensive transcriptome profiling with decreased adapter dimer formation and bias correction. *Sci. Rep.* **9**, 7953 (2019).
106. V. Pliatsika, P. Loher, A. G. Telonis, I. Rigoutsos, MINTbase: A framework for the interactive exploration of mitochondrial and nuclear tRNA fragments. *Bioinformatics* **32**, 2481–2489 (2016).
107. A. D. Holmes *et al.*, A standardized ontology for naming tRNA-derived RNAs based on molecular origin. *Nat. Methods* **20**, 627–628 (2023).
108. P. Fechter, J. Rudinger, R. Giege, A. Theobald-Dietrich, Ribozyme processed tRNA transcripts with unfriendly internal promoter for T7 RNA polymerase: Production and activity. *FEBS Lett.* **436**, 99–103 (1998).
109. K. Pawar, C. Hanisch, S. E. Palma Vera, R. Einspanier, S. Sharbati, Down regulated lncRNA MEG3 eliminates mycobacteria in macrophages via autophagy. *Sci. Rep.* **6**, 19416 (2016).
110. M. Fabbri *et al.*, MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc. Natl. Acad. Sci. U.S.A.* **109**, E2110–E2116 (2012).
111. K. Honda *et al.*, Spatiotemporal regulation of MyD88-IRF-7 signalling for robust type-I interferon induction. *Nature* **434**, 1035–1040 (2005).
112. S. Honda *et al.*, The biogenesis pathway of tRNA-derived piRNAs in Bombyx germ cells. *Nucleic Acids Res.* **45**, 9108–9120 (2017).