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REVIEW

The double life of a chemotherapy drug: Immunomodulatory functions of gemcitabine in cancer

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Alaina C. Larson^{1,2} | Kenadie R. Doty^{1,2} | Joyce C. Solheim^{1,2,3,4}

¹Eppley Institute for Research in Cancer & Allied Diseases, University of Nebraska Medical Center, Omaha, Nebraska, USA

²Fred & Pamela Buffett Cancer Center, University of Nebraska Medical Center, Omaha, Nebraska, USA

³Department of Biochemistry & Molecular Biology, University of Nebraska Medical Center, Omaha, Nebraska, USA

⁴Department of Pathology, Microbiology, & Immunology, University of Nebraska Medical Center, Omaha, Nebraska, USA

Correspondence

Joyce C. Solheim, Eppley Institute for Research in Cancer and Allied Diseases, University of Nebraska Medical Center, 986805 Nebraska Medical Center, Omaha, NE 68198, USA. Email: jsolheim@unmc.edu

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Abstract

Although the development of immunotherapies has been revolutionary in the treatment of several cancers, many cancer types remain unresponsive to immune-based treatment and are largely managed by chemotherapy drugs. However, chemotherapeutics are not infallible and are frequently rendered ineffective as resistance develops from prolonged exposure. Recent investigations have indicated that some chemotherapy drugs have additional functions beyond their normative cytotoxic capacity and are in fact immune-modifying agents. Of the pharmaceuticals with identified immune-editing properties, gemcitabine is well-studied and of interest to clinicians and scientists alike. Gemcitabine is a chemotherapy drug approved for the treatment of multiple cancers, including breast, lung, pancreatic, and ovarian. Because of its broad applications, relatively low toxicity profile, and history as a favorable combinatory partner, there is promise in the recharacterization of gemcitabine in the context of the immune system. Such efforts may allow the identification of suitable immunotherapeutic combinations, wherein gemcitabine can be used as a priming agent to improve immunotherapy efficacy in traditionally insensitive cancers. This review looks to highlight documented immunomodulatory abilities of one of the most wellknown chemotherapy agents, gemcitabine, relating to its influence on cells and proteins of the immune system.

KEYWORDS

cancer biology, chemotherapy, immunology, pancreatic cancer

1 **INTRODUCTION**

The immune system is a network comprised of cells, proteins, tissues, and organs which function to protect the host from foreign pathogens and tumor development. Although the immune system has intrinsic anticancer properties, it is not infallible and notably liable for failure in the case of those individuals who succumb to cancer.¹ The reality of this insufficiency prompted the development of therapies to reinvigorate immune reactivity, aptly called immunotherapies, and includes the likes of vaccines, cytokines, adoptive T-cell transfer, and monoclonal antibodies (e.g., immune checkpoint inhibitors).² The scope of these immunotherapies, much like the components of the immune

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system itself, is widespread and encompasses agents that both invoke and inhibit the immune response as modes of therapeutic management.^{3–6} Immunotherapies, particularly immune checkpoint inhibitors, have revolutionized treatment and patient outcomes for several cancer types, such as Hodgkin's lymphoma, melanoma, non-small cell lung cancer, and renal cancer.^{7–14} Despite these promising developments, multiple other cancers have not displayed sensitivity to tested immunotherapies in the current setting. This unresponsive phenotype is often derived from insufficient mutational burden, inhibitory immune checkpoint expression patterns, and defects in antigen presentation.^{15–18} Immunotherapy-ineligible patients instead rely on transiently effective, resistance-prone chemotherapy drugs as the backbone of disease management.^{19–21}

As the need for improved treatment options intensifies, attention has turned to "old-school" chemotherapies as an untapped source of potential. Although chemotherapeutics were initially approved for their cancer-killing properties, some are now known to be potent immunomodulators, capable of suppressing or invigorating the immune system.^{22–27} The repurposing of these old school drugs with their novel immune-editing abilities may allow for improved antitumor outcomes in the form of combination therapies. Specifically, chemotherapeutic immunomodulators can be paired with immunotherapies, wherein the chemotherapy drug can act as a priming agent and allow for enhanced immunotherapeutic efficacy in cancers that are normally unresponsive. One such drug that exhibits both chemotherapeutic and immunomodulatory abilities is gemcitabine.

Gemcitabine is a chemotherapy drug initially approved by the Food and Drug Administration in 1996 for the treatment of locally advanced and metastatic pancreatic cancer,^{28,29} and it remains active in clinical use today for the treatment of breast, non-small cell lung, and ovarian cancer, in addition to pancreatic cancer.³⁰ Gemcitabine's use as a monotherapy is infrequent, and it is most often prescribed in combination regimens with other chemotherapies, including platinum and taxol-based agents.^{31–34} Gemcitabine's reputation as a favorable combination partner is derived in part from its limited toxicity, with adverse effects like myelosuppression, hair loss, nausea, and vomiting reported as mild or rarely of clinical significance.³⁵

Gemcitabine belongs to the antimetabolite class of chemotherapeutics, joining pharmaceuticals like 5-fluorouracil and methotrexate.³⁶ Its cytotoxic function is exerted via masked chain termination in which gemcitabine, *masking* as a nucleoside (deoxycytidine), is mistakenly incorporated into the DNA strand. Following addition of a single nucleotide, DNA polymerase is released, the replication fork collapses, and the cell succumbs to apoptosis (Figure 1A).^{37,38} The fork collapse is accompanied by ataxia telangiectasia mutated and Rad3-related (ATR) pathway activation, and it has been demonstrated that inhibition of ATR interferes with PD-L1 upregulation that would otherwise occur via interferon regulatory factor 1 (IRF1) signaling.^{39,40} In a concurrent mechanism, gemcitabine covalently binds to the active site of ribonucleotide reductase (RNR), an enzyme responsible for conversion of ribonucleotides (NTPs) to deoxyribonucleotides (dNTPs). Impairment of RNR function disrupts dNTP levels, thereby improving gemcitabine's propensity for DNA incorporation in a process called self-potentiation (Figure 1B).³⁸

Multiple studies have indicated that gemcitabine holds not only cytotoxic capabilities but also boasts additional immune-modifying functions. In fact, gemcitabine treatment has demonstrated immuno-altering properties across several cancer types. For example, gemcitabine increases expression of immune system proteins like MHC class I chain-related protein A and B (MICA/B), major histocompatibility class I (MHC-I), programmed deathligand 1 and 2 (PD-L1, PD-L2), calreticulin (CRT), and others.^{24,41,42} The immunomodulatory abilities of gemcitabine extend beyond alterations of tumor cells themselves-this drug also influences both the behaviors and relative abundance of several immune cell populations (Figure 2).^{43–45} Because of its potential to improve patient sensitivity to immunotherapy-mediated intervention, gemcitabine's immune-editing capacity relative to immune cells and tumor cells will be highlighted within this review.

2 | IMMUNOMODULATORY EFFECTS OF GEMCITABINE ON IMMUNE CELLS

Tumor cells are not the sole perpetrators of malignant behaviors; rather, they require and extort neighboring immune cells to further their proliferative and migratory objectives. Thus, the presence and abundance of certain tumor-promoting immune cells within the microenvironment can have consequences for disease progression.⁴⁶ Finding mechanisms either to deplete such immunosuppressive cells or to increase the presence of antitumor immune cells is an ongoing area of investigation.⁴⁶ Pharmaceutical intervention may hold the answer to such an immunological dilemma and is described in the context of gemcitabine in the section below.

2.1 | Regulatory T cells

Regulatory T cells, or as they are commonly denoted, Tregs, play an important role in regulation of the





FIGURE 1 Gemcitabine's chemotherapeutic mechanisms of action. The figure depicts the primary modes by which gemcitabine induces cancer cell death (others exist which are not described). (A) Gemcitabine in its prodrug form enters the cell via a nucleoside transporter. In the cytoplasm, gemcitabine is converted to its nucleoside-mimicking active state. DNA polymerase incorporates gemcitabine into the DNA chain and an additional nucleotide is added, securing gemcitabine in the DNA strand (masked chain termination). The polymerase is unable to proceed which promotes fork collapse and cell death. (B) Gemcitabine occupies the active site of RNR. RNR inhibition disrupts dNTP pools, increasing the likelihood of gemcitabine's incorporation into DNA (self-potentiation), and initiating cell death by masked chain termination. dNTP, deoxyribonucleotide triphosphate; GEM, gemcitabine; NTP, ribonucleotide triphosphate; RNR, ribonucleotide reductase.

immune response via release of suppressive cytokines and inhibition of T cell proliferation.⁴⁷⁻⁴⁹ Normally, these immunosuppressive functions are critical for the prevention of autoimmunity, promotion of selftolerance, and maintenance of homeostasis.⁵⁰ However, while such immunoregulation is protective in a healthy host system, it can become pathological in the context of cancer. Consequently, Tregs' infiltration is often characterized as a blockage to effective tumor immunity and thus a poor prognostic factor in several solid tumor cancers.^{51–53} Identifying mechanisms to facilitate the depletion or inactivation of Tregs may allow for cancer management by restoring the antitumor immune response.

Gemcitabine exposure is known to influence Tregs across several models. A modest but significant increase in survival was illustrated in an orthotopic pancreatic cancer mouse model, which demonstrated reduced levels of Tregs following gemcitabine treatment.²³ The increase in survival appeared to be in part derived from the depletion of Tregs, as the administered gemcitabine dosage was suboptimal and did not affect the volume of the primary tumor.²³ In peripheral blood samples from mesothelioma patients, gemcitabine exposure was associated with decreased Treg proliferation.⁵⁴ In conjunction, a study conducted by Eriksson et al. observed a higher percentage of circulating Tregs in chemonaïve pancreatic cancer patients in comparison with healthy donors.⁴⁴ However, treatment with gemcitabine reduced the average percent of peripheral blood Tregs in these patients.⁴⁴ In addition, the mean effector: Treg ratio of this patient cohort was augmented during the gemcitabine cycle, suggesting that gemcitabine likely increased the favorable effector T cell population, lessened the number of circulating, suppressive Tregs, or expedited an advantageous combination of both.44



FIGURE 2 Pro-immune effects of gemcitabine in cancer. The figure highlights gemcitabine's positive immunomodulatory properties on immune cells and immune-related proteins reported in at least one type of cancer. Gemcitabine reduced levels of circulating (MDSCs, Tregs) and intratumoral (Tregs) immunosuppressive cell populations in various cancer models. In addition, gemcitabine intensified the antitumor abilities of NK cells, including their tumor infiltration and lytic activity. The proliferative potential of NK cells was also improved by gemcitabine in patients with malignant mesothelioma. In pancreatic cancer patients treated with gemcitabine, the number of DCs in circulation increased. Gemcitabine also improved the cross-priming of CD8⁺ T cells in tumor-bearing mice. The proliferative capacity of effector T cells was not diminished after gemcitabine treatment in pancreatic cancer patients, and tumor infiltration of CD8⁺ and CD4⁺ effector T cells was stimulated by gemcitabine in a mouse model of ovarian cancer. Gemcitabine augmented expression of immune-promoting proteins, including surface CRT on cancer cells. Plasma membrane-resident CRT can act as an "eat me" signal and induce engulfment by phagocytic cells. Increased expression of the MHC-I complex, a cytotoxic T cell-activating molecule, has also been documented in cancer models following gemcitabine exposure. [Green arrows indicate signal activation.] CRT, calreticulin; DC, dendritic cell; MDSC, myeloid-derived suppressor cell; MHC-I, major histocompatibility complex class I; NK, natural killer cell; Treg, regulatory T cell.

2.2 | Myeloid-derived suppressor cells, monocytes, and macrophages

Like Tregs, myeloid-derived suppressor cells (MDSCs) are well known for their immune-quelching activities.⁵⁵ Both immature cells of granulocytic and monocytic lineage are classified as MDSCs and exert their immuno-suppressive functions primarily through inhibition of T-cell activation, but also by mitigating macrophage-mediated cytokine secretion and NK cell cytotoxic-ity.^{56–58} The role of MDSCs in maintaining homeostatic conditions remains somewhat ambiguous, albeit these cells are known to promote favorable immunosup-pression in semi-allogeneic states of pregnancy and transplantation.^{59,60} Under cancerous conditions, the

immunosuppressive functions of MDSCs are exploited by tumors in attempts to avoid immune detection and maintain their proliferative agenda. Circulating MDSC levels are often enhanced in several cancer types, including melanoma, pancreatic cancer, and squamous cell carcinoma of the head and neck.^{61–63}

Interestingly, gemcitabine appears to reduce MDSC levels. In a study conducted by Le et al., mice were subcutaneously injected in the flank with murine mammary tumor cells.⁶⁴ Because MDSCs can accumulate in peripheral lymphoid organs, the population of MDSCs in the spleen was evaluated after 3 weeks.⁶⁴ Upon harvesting, MDSCs had amassed within the spleen and accounted for approximately 30% of splenic cells in the untreated group.⁶⁴ However, MDSCs in the spleen were depleted to 10% in mice that received weekly gemcitabine treatments.⁶⁴ The findings of this study were similar to those of another group which showed that gemcitabine decreased the MDSC population localized within the spleen in mice bearing large mesothelioma or lung tumors (flank, subcutaneously inoculated).⁶⁵ Despite the gemcitabine-induced MDSC loss, there was no corresponding decrease in effector CD4⁺ or CD8⁺ T cells within the spleens of these animals.⁶⁵ Gemcitabine treatment also affects MDSC populations within human patients. Eriksson et al. showed that gemcitabine administration reduced peripheral MDSCs in patients with pancreatic cancer to levels near that of the healthy donor cohort.⁴⁴ Similar results were published in a study of individuals diagnosed with recurrent breast cancer who were prescribed gemcitabine.⁶⁶ Both prior and post-chemotherapy administration, the percentage of circulating MDSCs was evaluated.⁶⁶ Levels of MDSCs in the peripheral blood significantly decreased following gemcitabine treatment.⁶⁶ Furthermore, the percentage of circulating MDSCs after the gemcitabine cycle was not statistically different from that of the healthy volunteer group, indicating gemcitabine favorably modulates MDSC levels in refractory breast cancer cases.⁶⁶

Monocytes, and upon their differentiation, macrophages, play a critical part in mediating inflammation, namely through phagocytosis, secretion of cytokines, and activation of the adaptative immune response.⁶⁷ Phenotypically distinct subtypes of monocytes and macrophages have unique roles in the suppression or promotion of tumor development.⁶⁸ In patients with advanced pancreatic cancer, Soeda et al. reported an increase in the absolute number and percentage of circulating CD14⁺ monocytes after gemcitabine treatment.⁶⁹ However, because additional phenotypic markers were not used to further stratify these monocytes (e.g., HLA-DR^{low/neg}),⁶⁸ it is difficult to infer the clinical significance. Gemcitabine's pro-tumor effect extends to macrophages, as this chemotherapy drug has been observed to facilitate intratumoral infiltration by anti-inflammatory M2 macrophages.⁷⁰ Mice with orthotopic xenograft pancreatic tumors were found to have more macrophages within the tumors if the mice had been treated with gemcitabine, and conditioned culture media from gemcitabine-exposed pancreatic cancer cells induced polarization of macrophages to the M2 phenotype.⁷⁰ In a mouse model of breast cancer, gemcitabine treatment stimulated monocyte development, and subsequently increased the presence of CCR2⁺ monocytes and macrophages in the lungs, thereby potentiating tumor metastasis.⁷¹

Neutrophils are often credited as the initial responders in sites of injury, inflammation, or infection.⁷² In cancer, the pro-tumor and antitumor effects of neutrophils remain inconclusive.⁷² Gemcitabine monotherapy, as well Cancer Medicine

as its combination with other chemotherapies, may induce levels of neutropenia (and thrombocytopenia) that can be of clinical concern.⁷³⁻⁷⁵ Surprisingly, incidence of chemotherapy-induced neutropenia (CIN) has been correlated with improved survival in cancer patients,⁷⁶ and for gemcitabine-treated pancreatic cancer patients, earlyonset CIN was identified as a predictor of more favorable prognosis.⁷⁷

2.3 | Natural killer cells

Natural killer cells, or NK cells as they are commonly called, are a population of lymphoid cells which were discovered in the 1960s.⁷⁸ As indicated by their name, this cell type is *naturally* cytotoxic, requiring no previous antigen exposure to induce lysis of a target cell.⁷⁸⁻⁸⁰ NK cell-mediated lysis and granule secretion are primarily regulated through the presence and/or absence of various receptors and molecules on the surface of target cells.⁸¹⁻⁸⁵ This cytolytic capacity is critical to the host's defense against cells that have been either virally infected or malignantly transformed.⁸⁶ The presence of NK cell populations, which have corresponding antitumor abilities, is considered to be a favorable phenotype in an array of solid tumors.⁸⁷

Notably, gemcitabine has demonstrated an ability to influence NK cell populations in cancer . Gürlevik et al. established an R0 resectable transgenic mouse model of pancreatic cancer in which mice developed a single, pancreas-specific tumor.45 Following R0 resection, gemcitabine was administered in a localized fashion to the pancreas remnants.⁴⁵ Mice that received adjuvant gemcitabine exhibited enhanced infiltration of NK cells at the resection margins in comparison to the untreated control group.⁴⁵ Furthermore, this gemcitabine-perpetuated increase in NK cells was therapeutically relevant as depletion of NK cells enhanced local disease recurrence in this cohort.⁴⁵ Gemcitabine also stimulated NK cell-mediated cytotoxicity in a study conducted by Zhang et al. Splenic NK cells were purified from subcutaneous tumor-burdened mice and cocultured in vitro with lung tumor cells.⁴³ The NK cells obtained from the gemcitabine-treated mice lysed a higher percentage of tumor cells than those collected from the untreated mice, suggesting that gemcitabine positively enhances NK cell activity.43 Gemcitabine also appears to modulate the proliferative capacity of circulating NK cells in individuals with malignant mesothelioma.54 Mesothelioma patients received first-line platinum-pemetrexed chemotherapy, and those without progressive disease were later given either gemcitabine as maintenance therapy or basic supportive care (BSC) treatment.⁵⁴ NK cell proliferation was significantly upregulated in the cohort which received

maintenance gemcitabine treatment in comparison to the BSC group.⁵⁴

2.4 | Cytotoxic and helper alpha beta T cells, gamma delta T cells, dendritic cells, and B cells

The function of the adaptive immune system is carried out by the activities of several cell populations, including T cells, dendritic cells (DCs), and B cells. Alpha beta T cells and gamma delta T cells are distinguished based on their expression of the alpha beta T cell receptor or gamma delta T cell receptor, respectively.⁸⁸ Within the alpha beta designation, CD8⁺ cytotoxic T cells initiate direct cell-mediated lysis upon recognition of MHC-I-bound antigens, while CD4⁺ helper T cells release pro-inflammatory cytokines after identification of MHC-II-displayed antigens.¹ DCs and B cells are both professional antigen-presenting cells and promote activation of T cells, though B cells are mainly distinguished for their secretion of antibodies.¹

The importance of T cells to the therapeutic efficacy of gemcitabine was made evident by the discovery that this drug is ineffective against tumors in athymic nude mice.⁸⁹ Gemcitabine contributes to cross-priming of DCs (by inducing tumor cell apoptosis and other mechanisms), and thereby causes antigen-specific stimulation of CD8⁺ T cells.^{90,91} Furthermore, gemcitabine has been observed to increase the number of DCs in peripheral blood samples from pancreatic cancer patients.⁶⁹

Plate et al. evaluated the kinetics of gemcitabine's impact on immune cell populations and found that initial drops in T and B cell numbers were reversed as treatment of pancreatic cancer patients progressed.⁹² Anti-CD3 antibody stimulation of peripheral blood mononuclear cells obtained from gemcitabine-treated pancreatic cancer patients showed no diminution of T cell proliferation capability in comparison to controls.^{44,93} In a mouse model, gemcitabine caused some reduction of CD4⁺ and CD8⁺ T cell numbers, but a more than double impairment of B cell proliferation.⁹⁴ This gemcitabine-mitigated B cell activity was accompanied by a substantial reduction of the antibody response to a model tumor antigen that was expressed by mesothelioma tumor cells.⁹⁴

Ovarian tumor-bearing mice that received gemcitabine had more CD4⁺ and CD8⁺ T cells infiltrating the tumors.⁹⁵ Gemcitabine has been noted to have positive effects on CD8⁺ cytotoxic T cell antitumor activity, possibly as a consequence of diminishing the viability of MDSCs in the tumor microenvironment,⁶⁵ as well as by its abovementioned ability to enhance T cell priming. In the context of mesothelioma, gemcitabine treatment of patients led to increased expression of co-stimulatory molecules by CD4⁺ helper T cells and CD8⁺ cytotoxic T cells, indicating phenotypic changes in these cell populations.⁵⁴

In addition to the alpha beta T cell receptor-expressing CD4⁺ and CD8⁺ T cells, the gamma-delta T cell receptorexpressing subset can also have antitumor activities. However, gamma delta T cell-mediated cytotoxicity is independent of antigen presentation by MHC molecules.⁹⁶ Shimizu et al. showed that low-dose gemcitabine pretreatment augmented the cytolytic activity of zoledronic acidstimulated gamma delta T cells against urinary bladder cancer cells, and this combination also reduced tumor burden in vivo.⁹⁷ In a study of patients treated with gemcitabine alone or with both gemcitabine and gamma delta T cell therapy, no statistically significant survival differences were found.⁹⁸ However, there was a correlation between the persistence of gamma delta T cells in the blood and the absence of disease recurrence, suggesting that a clinical trial with a larger enrollment of patients would possibly yield more evidence of efficacy for this combination therapy.⁹⁸

3 | IMMUNOMODULATORY EFFECTS OF GEMCITABINE ON CANCER CELLS

Although off-target effects certainly exist, the intended quarries of chemotherapeutics (gemcitabine included) are the cancer cells themselves.^{99,100} As such, it is not surprising that a number of the immunomodulatory effects of gemcitabine manifest within the tumor. Tumor cells employ multiple tactics to evade immune surveillance, for example, the downregulation and upregulation of proteins that promote and curtail the immune response, respectively.^{101–103} Thus, pharmaceutical-induced expression of proteins that can restore the immune response is an active area of interest. Gemcitabine's impact on several of these immune-related proteins is described below.

3.1 | Major histocompatibility complex class I

Major histocompatibility complex class I (MHC-I) is a molecule that is expressed at the surface of all nucleated cells and has a critical function in the presentation of abnormal peptides to CD8⁺ cytotoxic T cells.¹⁰⁴ Recognition of these peptides as atypical (e.g., virus- or tumor-derived) induces T cell-mediated lysis against the infected or malignant cell.¹⁰⁵ Therefore, MHC-I is a vital component in instigating an immune response under pathological conditions. As such, many tumors downregulate expression of MHC-I as a means of immune escape.^{106–110} Thus, finding ways to fully restore surface expression of this molecule could play a major role in reinvigorating the T cell-mediated antitumor immune response. Gemcitabine has demonstrated a propensity to increase expression of MHC-I in several human cancer cell lines, including pancreatic, colon, breast, lung, and cholangiocarcinoma.^{24,111-114} Our own work in human pancreatic cancer cell lines demonstrated that gemcitabine increased MHC-I mRNA and protein levels, as well as cell surface expression and stability.¹¹⁵ Of note, we also observed that gemcitabine modified MHC-I-displayed peptides on a pancreatic cancer cell line and improved these peptides' predicted affinity and immunogenicity.¹¹⁵ Liu et al. and Principe et al. showed that gemcitabine's stimulation of MHC-I protein expression was recapitulated in murine models of lung cancer and pancreatic cancer, respectively.^{24,114} In vivo analysis revealed that while gemcitabine alone could enhance MHC-I expression in murine pancreatic tumors, it was not sufficient to promote effector T-cell infiltration. However, with the triple combination of gemcitabine, an immune checkpoint inhibitor, and a TGF-β-signaling inhibitor, T cell penetration into the tumors was not only restored, but the mice also exhibited increased overall survival.²⁴ These experiments strengthen the potential clinical relevance of gemcitabine's immunomodulatory capacity and indicate its ability to act successfully in combination therapies.

3.2 | Calreticulin

Calreticulin (CRT) is a calcium-binding, sarcoplasmic reticulum (SR) and endoplasmic reticulum (ER)-resident chaperone protein.^{116,117} This protein harnesses many functions and is involved in regulation of calcium homeostasis, intracellular signaling, gene expression, and assistance in protein folding.¹¹⁸⁻¹²⁰ Under physically or chemically induced cellular stress, CRT can be translocated from the ER to the cell surface.^{121,122} Following its incorporation into the plasma membrane, CRT acts as a damage-associated molecular response (DAMP) or "eatme" signal at the surface of the cell, initiating phagocyticmediated engulfment.¹²³ Elevated expression of surface CRT is correlated with improved clinical outcomes and considered a favorable prognostic factor in acute myeloid leukemia, ovarian cancer, and non-small cell lung cancer.^{124–126} Thus, the translocation of CRT is a desired contributor to the antitumor immune response, and of interest in this review, potentially achievable through gemcitabine exposure. In vitro analysis of murine bladder and pancreatic cancer cell lines revealed gemcitabine's capacity to increase surface expression of CRT.⁴¹ Such results were _Cancer Medicine

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mirrored in human cholangiocarcinoma and lung cancer cells, as well as a murine model of lung cancer.^{43,127,128} Interestingly, Smith et al. observed that human pancreatic cancer cells cultured in medium with gemcitabine not only had increased surface expression of CRT but also a higher frequency of engulfment via monocyte-derived dendritic cells, though it cannot be stated that this was specifically due to CRT upregulation.¹¹³ Nonetheless, because membrane-bound CRT is a potent DAMP and inducer of immune cell-mediated death, combining gemcitabine and other immune-enhancing therapies could lead to strategies to increase cancer-directed phagocytosis.

3.3 | PD-L1

Programmed death-ligand 1 (PD-L1) was the first discovered ligand of the immune cell-expressed programmed cell death protein 1 (PD-1).¹²⁹ The interaction of PD-1 and PD-L1 promotes the inactivation of immune cells, thereby decreasing their proliferation and capacity for cytokine production.¹³⁰ PD-L1 is categorized as an *immune checkpoint*, and it is critical in the regulation of self-tolerance and homeostasis. However, this checkpoint has a nefarious connotation in the setting of cancer, perpetuating T cell impairment and subduing immune-mediated responses against malignant cells.^{131,132} Expression of PD-L1 is significantly increased in many solid tumors, including nasopharyngeal carcinoma as well as bladder, breast, and gastric cancers.¹³³⁻¹³⁵ PD-L1 upregulation is characterized as a poor prognostic factor and affiliated with lower survival rates in the aforementioned cancers.¹³³⁻¹³⁵ However, PD-L1 overexpression on tumor cells can also be advantageous in terms of enhancing the efficacy of monoclonal antibody therapies targeting this protein. Immune checkpoint inhibitors, such as anti-PD-L1 therapeutics, have been clinically successful and paramount in the restoration of antitumor T cell responses.^{136,137} Thus, paradoxical as it may seem, the inadvertent upregulation of PD-L1 by pharmaceuticals could be beneficial in conversion of a traditionally insensitive cancer to one that is immunotherapeutically targetable. Gemcitabine has been observed to increase PD-L1 across several models of pancreatic cancer, including established cell lines, primary cell line-derived xenografts, murine cell lines, and a mouse model of pancreatic cancer.^{24,138,139} For ovarian cancer cell lines, gemcitabine stimulated mRNA levels and surface expression of PD-L1 by threefold and sixfold, respectively.⁹⁵ Furthermore, a study by Jung et al. observed that gemcitabine enhanced expression of PD-L1 by a human colorectal cancer cell line and significantly improved tumor infiltration and binding of an anti-PD-L1 antibody in vivo.¹⁴⁰ Therefore, gemcitabine-stimulated

PD-L1 expression may promote advantageous responses in the clinic via immune checkpoint combinations.

4 | POTENTIAL MECHANISMS

The propensity for gemcitabine (and other chemotherapeutics with similar activities) to engage in both cytotoxic and immunomodulatory behavior remains an active area of exploration. With such an array of immunomodifications induced by gemcitabine, it is unlikely that a singular channel is responsible. Instead, this section will highlight several potential mechanisms by which gemcitabine may influence the immunophenotype of cells.

Gemcitabine's seemingly selective reduction of immunosuppressive cell populations may be in part due to the proliferative nature of these cells. Both MDSCs and Tregs have the potential for rapid expansion,^{141–143} and thus may be more prone to gemcitabine incorporation than other immune populations. Suzuki et al., showed that gemcitabine increased apoptosis of MDSCs (Gr-1⁺/CD11b⁺), but did not affect non-suppressive lymphocyte populations in vitro.⁶⁵ In a similar vein, intratumoral Treg populations underwent higher rates of cellular division compared to conventional T cell populations in a pancreatic cancer model, and thus their reduction by gemcitabine was more likely due to preferential proliferative-targeting by this drug.²³

Transcriptional regulation is often shared between the immune system's "on switch" (e.g., MHC-I) and the corresponding "off switch" (e.g., PD-L1) to prevent extended and unintentional reactivity beyond the initial immune response. For example, interferon-sensitive response elements (ISREs) located in the promoters of the PD-L1 gene and MHC-I-associated genes confer their sensitivity to interferon-mediated signaling.^{144,145} Likewise, nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB)binding sites make these genes' susceptible to NF- κ B regulation (e.g., via TNFα signaling).^{144,146} In a murine model of pancreatic cancer, gemcitabine upregulated secretion of several cytokines, including interferon gamma (IFN γ) and tumor necrosis factor alpha (TNF α),²⁴ both of which are known transcriptional inducers of MHC-I and PD-L1.^{144,146} Thus, it is plausible that in vivo gemcitabine instigates cytokine-mediated signaling to modify expression of immune-related proteins.

This ability for gemcitabine to stimulate secretion of inflammatory cytokines may be a byproduct of its inherent chemotherapeutic mechanisms (i.e., DNA damage and nucleotide pool disruption). In nasopharyngeal carcinoma, MHC-I induction by gemcitabine and a gemcitabine/ cisplatin combination was found to be dependent on the STING type I interferon-dependent pathway,¹⁴⁷ suggesting that chemotherapy-induced DNA fragmentation triggers inflammatory cytokine production. Nucleotide depletion by gemcitabine has also been shown to stimulate expression of several interferon-stimulated genes (ISGs).¹⁴⁸ Activity of these ISGs was inhibited by reintroduction of certain nucleotides with gemcitabine co-treatment, indicating that it is the nucleotide inhibition by gemcitabine which invokes expression of these ISGs.¹⁴⁸ Stimulation of immune-associated genes via pharmaceutical-induced disruption of nucleotide pools has been previously observed and could be a concomitant effect of the infection mimicry state induced by these drugs.^{149–152}

The signaling events which relay nucleotide loss to these genes requires additional investigation. Of note, a recent report by Mullens et al., revealed that stimulation of MHC-I associated genes by a nucleotide-disrupting drug was abrogated through inhibition of positive transcription elongation factor b (P-TEFb), a protein necessitated for efficient transcription.¹⁵³ Thus, it appears that drug-induced nucleotide depletion is dependent on the activity of P-TEFb, but discerning whether this is a conserved mechanism among agents with similar modes of action (e.g., gemcitabine) will require further exploration.

5 | CONCLUSIONS AND PERSPECTIVES

Chemotherapy has proven to be a front-line method of disease management for most cancer types. However, many forms of cancer suffer from high refractory rates or chemoresistance, and there is a continued need to identify courses of action to improve prognosis beyond the canonical neoadjuvant and adjuvant therapies.¹⁵⁴ Such a reality has prompted a reinvestigation into the mechanisms of standard chemotherapy drugs and brought forth additional immune-editing abilities for several of these anticancer drugs.¹⁵⁵ It is believed that the immunostimulatory properties of chemotherapy drugs, such as gemcitabine, can be used to enhance the efficacy of immunotherapies against cancers that are traditionally insensitive. Gemcitabine exerts favorable impacts on both immune cell populations and tumor cells, including depletion of Tregs and MDSCs, infiltration of NK cells and effector T cells, as well as invigoration of immune-stimulating surface CRT and MHC-I expression (Figure 2). Within the last 3 years, more than 15 clinical trials have evaluated immunotherapy combination strategies which include gemcitabine (Table 1).^{156–171} Although the results varied between trial settings, cancer types, and patient cohorts, positive outcomes were documented within the published trials. For example, the combination of gemcitabine/nab-paclitaxel and an immune checkpoint inhibitor (anti-PD-1) improved 1-year survival

Phase and no. of patients enrolled	Cancer	Chemotherapy/SOC	Immunotherapy/target	Primary outcome	Identifier no.	Reference
Phase II—128 patients	Biliary tract cancer (advanced)	Gemcitabine/cisplatin	 Durvalumab (anti-PDL1) Tremelimumab (anti-CTLA-4) Durvalumab/tremelimumab 	The ORR from the chemotherapy +durvalumab was 72%, from chemotherapy+durvalumab+trem- limumab it was 70%, and it was 50% for those who received chemotherapy then chemotherapy +durvalumab+tremlimumab.	NCT03875235	156
Phase II—38 patients	Biliary tract cancer (advanced)	Gemcitabine/oxaliplatin	1. Camrelizumab (anti-PD-1)	Combination improved PFS by 2.6 months when compared to historical averages. OS was also increased by the combination therapy (extended by 3.3 months). Combination therapy was a tolerable regimen.	NCT03486678	157
Phase II—105 patients	Pancreatic (metastatic)	Gemcitabine/ nab-paclitaxel	 Nivolumab (anti-PD-1) Sotigalimab (anti-CD40, agonist) Nivolumab and sotigalimab 	Combination of nivolumab + chemotherapy extended 1-year OS (57.7%) from the historical average (35%), $n = 34$). Other combinations did not improve 1-year OS compared to historical numbers.	NCT03214250	158
Phase II—180 patients	Pancreatic (metastatic)	Gemcitabine/ nab-paclitaxel	 Durvalumab/ tremelimumab (anti-PD-L1/ anti-CTLA-4) 	Combination therapy did not improve survival.	NCT02879318	159
Phase II—68 patients	Biliary tract cancer (advanced)	Gemcitabine/cisplatin	1. Nivolumab (anti-PD-1)	Combination had similar PFS (6 months, 59.4%) as standard chemotherapy treatment. However, OS was improved by combination therapy (2 years, 35.4%) when compared to historical averages (15%–22%), indicating there is a subset of patients who may benefit.	NCT03101566	160
Phase III—184 patients	Large B cell lymphoma (refractory or early relapsed)	R-DHAP/R-ICE/R-GDP	 Lisocabtagene maraleucel (autologous, CD19-directed CAR T cell therapy) 	Inclusion of CAR T cell therapy increased median event-free survival (10.1 months) in comparison to the standard of care group (2.3 months).	NCT03575351	161
Phase Ib—21 patients	Pancreatic (advanced)	Gemcitabine	1. Ipilimumab (anti-CTLA-4)	Combination therapy was a tolerable regimen.	NCT01473940	162
Phase I—42 patients	Pancreatic (advanced, treatment refractory)	Gemcitabine	 Defactinib/pembrolizumab (FAK inhibitor/anti-PD-1) 	Combination therapy was a tolerable regimen.	NCT02546531	163,164
Phase Ib—23 patients	Head and neck squamous cell carcinoma (advanced)	Gemcitabine/cisplatin	1. Toripalimab (anti-PD-1)	Combination therapy was a tolerable regimen.	NCT04947241	165 (Continues)

TABLE 1 Completed clinical trials testing gemcitabine and immunotherapy combinations.^a

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Phase and no. of patients enrolled	Cancer	Chemotherapy/SOC	Immunotherapy/target	Primary outcome	Identifier no.	Reference
Phase II—32 patients	Biliary tract cancer (metastatic or unresectable)	Gemcitabine/cisplatin	1. Nivolumab (anti-PD-1)	Of the patients who were evaluable, the ORR was 55.6%, with 18.6% obtaining a CR.	NCT03311789	166
Phase III—303 patients	Pancreatic (advanced)	Gemcitabine/ nab-paclitaxel	 Algenpantucel-L (vaccine; allogeneic pancreatic cancer cells engineered to express murine a(1,3)GT) 	Median OS (14.9 months) was not improved from that of the standard of care group (14.3 months).	NCT01836432	167
Phase II—27 patients	Primary mediastinal B cell lymphoma (relapsed/refractory)	Gemcitabine/vinorelbine/ PLD	1. Camrelizumab (anti-PD-1)	Combination therapy was tolerable. The ORR of the combination therapy was 74%, with 56% achieving CR. Historical numbers not provided; disease is rare.	NCT03346642	168
Phase I/II—32 patients	Pancreatic (resected)	Gemcitabine (adjuvant)	 TG01GM-CSF (vaccine; 7 RAS peptides representing the 7 most common KRAS codon 12 and 13 oncogenic mutations/granulocyte macrophage colony-stimulating factor) 	Combination therapy increased median OS (33.3months) compared to historical data (17.1–26.5 months) for adjuvant gemcitabine, which was the SOC at the time. TG01/ GM-CSF is safe to use in conjunction with chemotherapy. A positive immune response (defined as a DTH response and/or a positive T cell proliferation assay) was observed for >90% of patients.	NCT02261714	69 I
Phase Ib—20 patients	Squamous non-small cell lung cancer (advanced)	Gemcitabine/cisplatin	1. Sintilimab (anti-PD-1)	Combination therapy was a tolerable regimen.	NCT02937116	170
Phase II—71 patients	Ovarian cancer (relapsed, but platinum and gemcitabine-sensitive)	Gemcitabine/carboplatin	 DCVAC/OvCa (vaccine; autologous DCs pulsed with human OV-90 and SK-OV-3 ovarian cancer cells pretreated with high hydrostatic pressure for the induction of immunogenic cell death) 	Combination therapy was a tolerable regimen but did not extend median PFS compared to chemotherapy alone arm (11.3 months vs. 9.5 months, respectively). However, it did improve median OS by 13.4 months).	NCT02107950	171
Abbreviations: α[1,3]GT, al associated antigen 4; DCs, <i>c</i> adhesion kinase; PD-1, pro <u></u> Rat sarcoma; R-DHAP, ritu:	pha-1,3-galactosyltransfera: lendritic cells; DTH, delaye: grammed cell death protein ximab/dexamethasone/cyti	se; CAR, chimeric antigen recep d type hypersensitivity response 1; PD-L1, programmed death-li arabine/cisplatin; R-GDP, rituxi	tor; CD19, cluster of differentiation 19; CD4(;; GM-CSF, granulocyte macrophage colony- gand 1; PFS, progression-free survival; PLD, mab/gemcitabine/dexamethasone/cisplatin;	 cluster of differentiation 40; CR, complete respons stimulating factor; KRAS, Kirsten rat sarcoma viral pegylated liposomal doxorubicin; ORR, overall resp R-ICE, rituximab/itosfamide/carboplatin/etoposide 	ie; CTLA-4, cytotox oncogene homolog onse rate; OS, over ; SOC, standard of	ic T-lymphocyte- FAK, focal all survival; RAS, care.

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TABLE 1 (Continued)

^aOnly published results with documented National Clinical Trial (NCT) identifier numbers from January 2020 to May 2023 are included. Combination studies conducted prior to 2020 or that are currently ongoing were excluded from the table. Active clinical trials can be found at Clinical Trialsgov.

from historical averages in patients with metastatic pancreatic cancer.¹⁵⁸ Thus, there is continued interest in investigating multi-therapy approaches and identifying the cancer types and patient populations most responsive to these combinations.

Although this review has focused mostly on the positive immune-promoting effects of gemcitabine, it is important to note that some of its modulatory properties could be classified as suppressive and detrimental. The neutropenia and thrombocytopenia induced in cancer patients by gemcitabine can potentially lead to mortality, yet they are also statistically associated with improved survival.^{172,173} Despite gemcitabine's propensity to reduce MDSCs in vivo,⁶⁵ it may also activate the inflammasome pathway in these cells and accelerate tumor expansion through release of inflammatory cytokines.¹⁷⁴ As noted above, gemcitabine has also been implicated in increasing tumor infiltration by M2polarized macrophages, a phenotype that is typically regarded as tumor-promoting.⁷⁰ In addition, as mentioned earlier in this review, gemcitabine treatment is correlated with increased production of monocytes, which by homing to sites such as the lungs and differentiating to immunosuppressive macrophages, can assist metastasis.⁷¹

Subsequently, further exploration must be conducted to accurately estimate gemcitabine's beneficial modulation of immune parameters from potential disadvantageous effects. Various investigations have sought biomarker correlates of gemcitabine treatment and patient outcomes,^{175,176} and some studies have included hematological and immunological factors.^{173,177} For example, Blomstrand et al. profiled blood proteins and cells in pancreatic cancer patients and evaluated whether any were prognostic for prolonged survival.¹⁷³ The occurrence of thrombocytopenia was a positive prognostic marker for progressionfree survival, but myelosuppression had neither positive nor negative linkage to survival in this setting.¹⁷³ Another example of a potential immune-related biomarker is endoplasmic reticulum aminopeptidase 2 (ERAP2), which correlates with worse prognosis in pancreatic cancer patients and its expression in pancreatic cancer cells is increased by gemcitabine treatment.¹⁷⁷ ERAP2 is a protein involved in the processing of peptide ligands for binding to MHC-I,¹⁷⁸ but it also has a role in the PI3K/AKT/mTOR pathway, and whether its immunological function and/or its cell signaling function is the rationale for its correlation with poor prognosis is not well understood. Bauer et al. showed that gemcitabine therapy decreased DC vaccine impact on B cell and CD8⁺ T cell responses, but the combination of the DC vaccine and gemcitabine still was clinically beneficial,¹⁷⁹ showing the difficulty of establishing specific immunological biomarkers that correlate with overall clinical prowess. In the future, more in-depth knowledge of the immunological events occurring after gemcitabine treatment, and

their association with outcomes, may not only influence selection of immunotherapy partners, but also aid in the establishment of dosages and administration scheduling to achieve optimal responses and mitigate potential suppressive effects in the clinical setting.

AUTHOR CONTRIBUTIONS

Alaina C. Larson: Conceptualization (lead); formal analysis (lead); funding acquisition (equal); project administration (equal); writing – original draft (lead); writing – review and editing (equal). Joyce C. Solheim: Funding acquisition (equal); project administration (equal); supervision (lead); writing – review and editing (equal). Kenadie R. Doty: Writing – review and editing (equal).

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CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were generated for this review.

ORCID

Joyce C. Solheim D https://orcid.org/0000-0002-6313-8213

REFERENCES

- Marshall JS, Warrington R, Watson W, Kim HL. An introduction to immunology and immunopathology. *Allergy Asthma Clin Immunol.* 2018;14(Suppl 2):49.
- 2. Marshall HT, Djamgoz MBA. Immuno-oncology: emerging targets and combination therapies. *Front Oncol.* 2018;8:315.
- Showalter A, Limaye A, Oyer JL, et al. Cytokines in immunogenic cell death: applications for cancer immunotherapy. *Cytokine*. 2017;97(123):123-132.
- 4. Yang JC, Rosenberg SA. Adoptive T-cell therapy for cancer. *Adv Immunol.* 2016;130:279-294.
- Gatti-Mays ME, Redman JM, Collins JM, Bilusic M. Cancer vaccines: enhanced immunogenic modulation through therapeutic combinations. *Hum Vaccin Immunother*. 2017;13(11): 2561-2574.

- Rogers LM, Veeramani S, Weiner GJ. Complement in monoclonal antibody therapy of cancer. *Immunol Res.* 2014;59(1-3):203-210.
- Ansell SM, Lesokhin AM, Borrello I, et al. PD-1 blockade with Nivolumab in relapsed or refractory Hodgkin's lymphoma. N Engl J Med. 2015;372(4):311-319.
- Ramchandren R, Domingo-Domènech E, Rueda A, et al. Nivolumab for newly diagnosed advanced-stage classic Hodgkin lymphoma: safety and efficacy in the phase II CheckMate 205 study. JCO. 2019;37(23):1997-2007.
- 9. Porter DL, Levine BL, Kalos M, Bagg A, June CH. Chimeric antigen receptor-modified T cells in chronic lymphoid leukemia. *New Engl J Med.* 2011;365(8):725-733.
- Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med.* 2010;363(8):711-723.
- Hodi FS, O'Day SJ, McDermott DF, et al. Erratum-improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med.* 2010;363(8):711-723.
- Wolchok JD, Kluger H, Callahan MK, et al. Nivolumab plus ipilimumab in advanced melanoma. *New Engl J Med.* 2013;369(2):122-133.
- Wolchok JD, Kluger H, Callahan MK, et al. Erratum-Nivolumab plus ipilimumab in advanced melanoma. N Engl J Med. 2013;369(2):122-133.
- 14. Motzer RJ, Rini BI, McDermott DF, et al. Nivolumab for metastatic renal cell carcinoma: results of a randomized phase II trial. *J Clin Oncol.* 2015;33(13):1430-1437.
- 15. Xu Y, Wan B, Chen X, et al. The association of PD-L1 expression with the efficacy of anti-PD-1/PD-L1 immunotherapy and survival of non-small cell lung cancer patients: a metaanalysis of randomized controlled trials. *Transl Lung Cancer Res.* 2019;8:413-428.
- Samstein RM, Lee CH, Shoushtari AN, et al. Morris LGT. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. *Nature*. 2019;51(202):202-206.
- Shin DS, Zaretsky JM, Escuin-Ordinas H, et al. Primary resistance to PD-1 blockade mediated by JAK1/2 mutations. *Cancer Discov.* 2017;7(2):188-201.
- Patel SJ, Sanjana NE, Kishton RJ, et al. Identification of essential genes for cancer immunotherapy. *Nature*. 22017;548(7669): 537-542.
- 19. Santos ES. Treatment options after first-line immunotherapy in metastatic NSCLC. *Expert Rev Anticancer Ther.* 2020;20(3): 221-228.
- Chamoto K, Hatae R, Honjo T. Current issues and perspectives in PD-1 blockade cancer immunotherapy. *Int J Clin Oncol.* 2020;25(5):790-800.
- Sohal DPS, Kennedy EB, Cinar P, et al. Metastatic pancreatic cancer: ASCO guideline update. J Clin Oncol. 2020;38(27): 3217-3230.
- Wada S, Yoshimura K, Hipkiss EL, et al. Cyclophosphamide augments antitumor immunity: studies in an autochthonous prostate cancer model. *Cancer Res.* 2009;69(10):4309-4318.
- Shevchenko I, Karakhanova S, Soltek S, et al. Low-dose gemcitabine depletes regulatory T cells and improves survival in the orthotopic Panc02 model of pancreatic cancer. *Int J Cancer*. 2013;133(1):98-107.
- 24. Principe DR, Narbutis M, Kumar S, et al. Long-term gemcitabine treatment reshapes the pancreatic tumor microenvironment

and sensitizes murine carcinoma to combination immunotherapy. *Cancer Res.* 2020;80(15):3101-3115.

- 25. Wan S, Pestka S, Jubin RG, Lyu YL, Tsai YC, Liu LF. Chemotherapeutics and radiation stimulate MHC class I expression through elevated interferon-beta signaling in breast cancer cells. *PLoS One*. 2012;7(3):e32542.
- Ohtsukasa S, Okabe S, Yamashita H, Iwai T, Sugihara K. Increased expression of CEA and MHC class I in colorectal cancer cell lines exposed to chemotherapy drugs. *J Cancer Res Clin Oncol.* 2003;129(12):719-726.
- 27. Grabosch S, Bulatovic M, Zeng F, et al. Cisplatin-induced immune modulation in ovarian cancer mouse models with distinct inflammation profiles. *Oncogene*. 2019;38(13):2380-2393.
- Food, Administration D. DRUGS@FDA: FDA-approved drugsreview of 020509. Approval of Date(s) and History, Letters, Labels. Reviews for NDA 1996;020509. https://www.accessdata. fda.gov/scripts/cder/daf/index.cfm?event=overview.process& ApplNo=020509
- 29. King RS. Gemcitabine. New first-line therapy for pancreatic cancer. *Cancer Pract*. 1996;4(6):353-354.
- Toschi L, Finocchiaro G, Bartolini S, Gioia V, Cappuzzo F. Role of gemcitabine in cancer therapy. *Future Oncol.* 2005;1(1):7-17.
- Renschler MF. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *New Engl J Med.* 2013;369(18):1691-1703.
- 32. Pfisterer J, Plante M, Vergote I, et al. Gemcitabine plus carboplatin compared with carboplatin in patients with platinumsensitive recurrent ovarian cancer: an intergroup trial of the AGO-OVAR, the NCIC CTG, and the EORTC GCG. J Clin Oncol. 2006;24(29):4699-4707.
- 33. Sederholm C, Hillerdal G, Lamberg K, et al. Phase III trial of gemcitabine plus carboplatin versus single-agent gemcitabine in the treatment of locally advanced or metastatic non-smallcell lung cancer: the Swedish lung cancer study group. *J Clin Oncol.* 2005;23(33):8380-8388.
- 34. von der Maase H, Hansen SW, Roberts JT, et al. Gemcitabine and cisplatin versus methotrexate, vinblastine, doxorubicin, and cisplatin in advanced or metastatic bladder cancer: results of a large, randomized, multinational, multicenter, phase III study. *J Clin Oncol.* 2000;18(17):3068-3077.
- Aapro MS, Martin C, Hatty S. Gemcitabine-a safety review. Anticancer Drugs. 1998;9(3):191-201.
- Kaye SB. New antimetabolites in cancer chemotherapy and their clinical impact. *Br J Cancer*. 1998;78(Suppl 3):1-7.
- Gandhi V, Legha J, Chen F, Hertel LW, Plunkett W. Excision of 2',2'-difluorodeoxycytidine (gemcitabine) monophosphate residues from DNA. *Cancer Res.* 1996;56(19):4453-4459.
- de Sousa CL, Monteiro G. Gemcitabine: metabolism and molecular mechanisms of action, sensitivity and chemoresistance in pancreatic cancer. *Eur J Pharmacol.* 2014;741:8-16.
- 39. Sato H, Niimi A, Yasuhara T, et al. DNA double-strand break repair pathway regulates PD-L1 expression in cancer cells. *Nat Commun.* 2017;8:1751.
- Mouw KW, Konstantinopoulos PA. From checkpoint to checkpoint: DNA damage ATR/Chk1 checkpoint signalling elicits PD-L1 immune checkpoint activation. *Brit. J Cancer*. 2018;118:933-935.
- 41. Hayashi K, Nikolos F, Lee YC, et al. Tipping the immunostimulatory and inhibitory DAMP balance to harness immunogenic cell death. *Nat Commun*. 2020;11(1):6299.

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- 42. Morisaki T, Onishi H, Koya N, et al. Combinatorial cytotoxicity of gemcitabine and cytokine-activated killer cells in hepatocellular carcinoma via the NKG2D-MICA/B system. *Anticancer Res.* 2011;31(7):2505-2510.
- 43. Zhang X, Wang D, Li Z, et al. Low-dose gemcitabine treatment enhances immunogenicity and natural killer cell-driven tumor immunity in lung cancer. *Front Immunol.* 2020;11:331.
- Eriksson E, Wenthe J, Irenaeus S, Loskog A, Ullenhag G. Gemcitabine reduces MDSCs, tregs and TGFβ-1 while restoring the teff/treg ratio in patients with pancreatic cancer. *J Transl Med.* 2016;14:282.
- 45. Gürlevik E, Fleischmann-Mundt B, Brooks J, et al. Administration of Gemcitabine after Pancreatic Tumor Resection in mice induces an antitumor immune response mediated by natural killer cells. *Gastroenterology*. 2016;151(2):338-350.e7.
- Zamarron BF, Chen W. Dual roles of immune cells and their factors in cancer development and progression. *Int J Biol Sci.* 2011;7(5):651-658.
- Nakamura K, Kitani A, Strober W. Cell contact-dependent immunosuppression by CD4(+)CD25(+) regulatory T cells is mediated by cell surface-bound transforming growth factor beta. J Exp Med. 2001;194(5):629-644.
- 48. Rubtsov YP, Rasmussen JP, Chi EY, et al. Regulatory T cellderived interleukin-10 limits inflammation at environmental interfaces. *Immunity*. 2008;28(4):546-558.
- Thornton AM, Shevach EM. CD4+CD25+ immunoregulatory T cells suppress polyclonal T cell activation in vitro by inhibiting interleukin 2 production. *J Exp Med.* 1998;188(2):287-296.
- Smigiel KS, Srivastava S, Stolley JM, Campbell DJ. Regulatory T-cell homeostasis: steady-state maintenance and modulation during inflammation. *Immunol Rev.* 2014;259(1):40-59.
- Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors in cancer therapy: a focus on T-regulatory cells. *Immunol Cell Biol.* 2018;96(1):21-33.
- Bates GJ, Fox SB, Han C, et al. Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. *J Clin Oncol.* 2006;24(34):5373-5380.
- Sasada T, Kimura M, Yoshida Y, Kanai M, Takabayashi A. CD4+CD25+ regulatory T cells in patients with gastrointestinal malignancies: possible involvement of regulatory T cells in disease progression. *Cancer*. 2003;98(5):1089-1099.
- 54. Dammeijer F, De Gooijer CJ, van Gulijk M, et al. Immune monitoring in mesothelioma patients identifies novel immunemodulatory functions of gemcitabine associating with clinical response. *EBioMedicine*. 2021;64:103160.
- Gabrilovich DI, Bronte V, Chen SH, et al. The terminology issue for myeloid-derived suppressor cells. *Cancer Res.* 2007;67(1):425-426.
- Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Cross-talk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. *J Immunol.* 2007;179(2):977-983.
- Melani C, Chiodoni C, Forni G, Colombo MP. Myeloid cell expansion elicited by the progression of spontaneous mammary carcinomas in c-erbB-2 transgenic BALB/c mice suppresses immune reactivity. *Blood.* 2003;102(6):2138-2145.
- Li H, Han Y, Guo Q, Zhang M, Cao X. Cancer-expanded myeloidderived suppressor cells induce anergy of NK cells through membrane-bound TGF-beta 1. *J Immunol.* 2009;182(1):240-249.

- 59. Köstlin N, Kugel H, Spring B, et al. Granulocytic myeloid derived suppressor cells expand in human pregnancy and modulate T-cell responses. *Eur J Immunol.* 2014;44(9):2582-2591.
- Luan Y, Mosheir E, Menon MC, et al. Monocytic myeloidderived suppressor cells accumulate in renal transplant patients and mediate CD4(+) Foxp3(+) Treg expansion. Am J Transplant. 2013;13(12):3123-3131.
- Poschke I, Mougiakakos D, Hansson J, Masucci GV, Kiessling R. Immature immunosuppressive CD14+HLA-DR-/low cells in melanoma patients are Stat3hi and overexpress CD80, CD83, and DC-sign. *Cancer Res.* 2010;70(11):4335-4345.
- Khaled YS, Ammori BJ, Elkord E. Increased levels of granulocytic myeloid-derived suppressor cells in peripheral blood and tumour tissue of pancreatic cancer patients. *J Immunol Res.* 2014;2014(879897):1-9.
- 63. Chen WC, Lai CH, Chuang HC, Lin PY, Chen MF. Inflammationinduced myeloid-derived suppressor cells associated with squamous cell carcinoma of the head and neck. *Head Neck*. 2017;39(2):347-355.
- 64. Le HK, Graham L, Cha E, Morales JK, Manjili MH, Bear HD. Gemcitabine directly inhibits myeloid derived suppressor cells in BALB/c mice bearing 4T1 mammary carcinoma and augments expansion of T cells from tumor-bearing mice. *Int Immunopharmacol.* 2009;9(7):900-909.
- Suzuki E, Kapoor V, Jassar AS, Kaiser LR, Albelda SM. Gemcitabine selectively eliminates splenic gr-1+/CD11b+ myeloid suppressor cells in tumor-bearing animals and enhances antitumor immune activity. *Clin Cancer Res.* 2005;11(18):6713-6721.
- 66. Gonda K, Shibata M, Ohtake T, et al. Myeloid-derived suppressor cells are increased and correlated with type 2 immune responses, malnutrition, inflammation, and poor prognosis in patients with breast cancer. *Oncol Lett.* 2017;14(2):1766-1774.
- 67. Parihar A, Eubank TD, Doseff AI. Monocytes and macrophages regulate immunity through dynamic networks of survival and cell death. *J Innate Immun.* 2010;2(3):204-215.
- 68. Chen X, Li Y, Xia H, Chen YH. Monocytes in tumorigenesis and tumor immunotherapy. *Cells*. 2023;12(13):1673.
- Soeda A, Morita-Hoshi Y, Makiyama H, et al. Regular dose of gemcitabine induces an increase in CD14+ monocytes and CD11c+ dendritic cells in patients with advanced pancreatic cancer. *Jpn J Clin Oncol.* 2009;37(12):797-806.
- 70. Deshmukh SK, Tyagi N, Khan MA, et al. Gemcitabine treatment promotes immunosuppressive microenvironment in pancreatic tumors by supporting the infiltration, growth, and polarization of macrophages. *Sci Rep.* 2018;8(1):12000.
- 71. Wu C, Zhong Q, Shrestha R, et al. Reactive myelopoiesis and FX-expressing macrophages triggered by chemotherapy promote cancer lung metastasis. *JCI Insight*. 2023;8(9):e167499.
- 72. Wu L, Saxena S, Singh RK. Neutrophils in the tumor microenvironment. *Adv Exp Med Biol*. 2020;1224:1-20.
- Blomstrand H, Scheibling U, Bratthäll C, Green H, Elander NO. Real world evidence on gemcitabine and nab-paclitaxel combination chemotherapy in advanced pancreatic cancer. *BMC Cancer*. 2019;19(1):40.
- 74. Ito G, Kawakami K, Aoyama T, et al. Risk factors for severe neutropenia in pancreatic cancer patients treated with gemcitabine/nab-paclitaxel combination therapy. *PLoS One.* 2021;16(7):e0254726.
- 75. Shaw JL, Nielson CM, Park JK, Marongiu A, Soff GA. The incidence of thrombocytopenia in adult patients receiving

chemotherapy for solid tumors or hematologic malignancies. *Eur J Haematol.* 2021;106(5):662-672.

- Shitara K, Matsuo K, Oze I, et al. Meta-analysis of neutropenia or leukopenia as a prognostic factor in patients with malignant disease undergoing chemotherapy. *Cancer Chemother Pharmacol.* 2011;68(2):301-307.
- Chen Y, Shi Y, Yan H, Wang YR, Dai GH. Timing of chemotherapy-induced neutropenia: the prognostic factor in advanced pancreatic cancer patients treated with gemcitabine/gemcitabine-based chemotherapy. *Oncotarget*. 2017;8(39):66593-66600.
- Rosenau W, Moon HD. Lysis of homologous cells by sensitized lymphocytes in tissue culture. J Natl Cancer Inst. 1961;27:471-483.
- Herberman RB, Nunn ME, Holden HT, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic and allogeneic tumors. II. Characterization of effector cells. *Int J Cancer*. 1975;16(2):230-239.
- Kiessling R, Klein E, Pross H, Wigzell H. Natural killer cells in the mouse. II. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Characteristics of the killer cell. *Eur J Immunol.* 1975;5(2):117-121.
- Velotti F, Palmieri G, D'Ambrosio D, Piccoli M, Frati L, Santoni A. Differential expression of granzyme a and granzyme B proteases and their secretion by fresh rat natural killer cells (NK) and lymphokine-activated killer cells with NK phenotype (LAK-NK). *Eur J Immunol.* 1992;22(4):1049-1053.
- Jamieson AM, Diefenbach A, McMahon CW, Xiong N, Carlyle R. The role of the NKG2D immunoreceptor in immune cell activation and natural killing. *Immunity*. 2002;17(1):19-29.
- Kärre K, Ljunggren HG, Piontek G, Kiessling R. Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy. *Nature*. 1986;319(6055):675-678.
- Vitale M, Bottino C, Sivori S, et al. NKp44, a novel triggering surface molecule specifically expressed by activated natural killer cells, is involved in non-major histocompatibility complexrestricted tumor cell lysis. *J Exp Med.* 1998;187(12):2065-2072.
- Huard B, Karlsson L, Triebel F. KIR down-regulation on NK cells is associated with down-regulation of activating receptors and NK cell inactivation. *Eur J Immunol.* 2001;31(6):1728-1735.
- Trinchieri G. Biology of natural killer cells. Adv Immunol. 1989;47:187-376.
- Nersesian S, Schwartz SL, Grantham MSR, et al. NK cell infiltration is associated with improved overall survival in solid cancers: a systematic review and meta-analysis. *Transl Oncol.* 2021;14(1):100930.
- Morath A, Schamel WW. αβ and γδ T cell receptors: similar but different. *J Leukoc Biol.* 2020;107(6):1045-1055.
- 89. Piadel K, Dalgleish AG, Smith PL. Gemcitabine in the era of cancer immunotherapy. *J Clin Haematol*. 2020;1:107-120.
- Nowak AK, Lake RA, Marzo AL, et al. Induction of tumor cell apoptosis in vivo increases tumor antigen cross-presentation, cross-priming rather than cross-tolerizing host tumor-specific CD8 T cells. *J Immunol*. 2003;170(10):4905-4913.
- McDonnell AM, Joost Lesterhuis W, Khong A, et al. Restoration of defective cross-presentation in tumors by gemcitabine. *Oncoimmunology*. 2015;4(5):e1005501.
- 92. Plate JM, Plate AE, Shott S, Bograd S, Harris JE. Effect of gemcitabine on immune cells in subjects with adenocarcinoma of the pancreas. *Cancer Immunol Immunother*. 2005;54(9):915-925.

- 93. Homma Y, Taniguchi K, Nakazawa M, et al. Changes in the immune cell population and cell proliferation in peripheral blood after gemcitabine-based chemotherapy for pancreatic cancer. *Clin Transl Oncol.* 2014;16(3):330-335.
- Nowak AK, Robinson BWS, Lake RA. Gemcitabine exerts a selective effect on the humoral immune response: implications for combination chemo-immunotherapy. *Cancer Res.* 2002;62(8):2353-2358.
- 95. Peng J, Hamanishi J, Matsumura N, et al. Chemotherapy induces programmed cell death-ligand 1 overexpression via the nuclear factor-κB to foster an immunosuppressive tumor microenvironment in ovarian cancer. *Cancer Res.* 2015;75(23):5034-5045.
- Park JH, Lee HK. Function of γδ T cells in tumor immunology and their application to cancer therapy. *Exp Mol Med*. 2021;53(3):318-327.
- 97. Shimizu T, Tomogane M, Miyashita M, Ukimura O, Ashihara E. Low dose gemcitabine increases the cytotoxicity of human Vγ9Vδ2 T cells in bladder cancer cells in vitro and in an orthotopic xenograft model. *Oncoimmunology*. 2018;7(5):e1424671.
- Aoki T, Matsushita H, Hoshikawa M, Hasegawa K, Kokudo N, Kakimi K. Adjuvant combination therapy with gemcitabine and autologous gamma-delta T-cell transfer in patients with curatively resected pancreatic cancer. *Cytotherapy*. 2017;19(4):473-485.
- Mini E, Novili S, Caciagli B, Landini I, Mazzei T. Cellular pharmacology of gemcitabine. Ann Oncol. 2006;17(Suppl 5):7-12.
- 100. Bagnyukova TV, Serebriiskii IG, Zhou Y, Hopper-Borge EA, Golemis EA, Bagnyukova I. Chemotherapy and signaling: how can targeted therapies supercharge cytotoxic agents? *Cancer Biol Ther.* 2010;10(9):839-853.
- 101. Natali PG, Nicotra MR, Bigotti A, et al. Selective changes in expression of HLA class I polymorphic determinants in human solid tumors. *Proc Natl Acad Sci USA*. 1989;86(17):6719-6723.
- 102. Rivoltini L, Barracchini KC, Viggiano V, et al. Quantitative correlation between HLA class I allele expression and recognition of melanoma cells by antigen-specific cytotoxic T lymphocytes. *Cancer Res.* 1995;55(14):3149-3157.
- 103. Jacobs JF, Idema AJ, Bol KF, et al. Regulatory T cells and the PD-L1/PD-1 pathway mediate immune suppression in malignant human brain tumors. *Neuro Oncol.* 2009;11(4):394-402.
- 104. Natarajan K, Li H, Mariuzza RA, Margulies DH. MHC class I molecules, structure and function. *Rev Imunogenet*. 1999;1(1):32-46.
- Blum JS, Wearsch PA, Cresswell P. Pathways of antigen processing. *Annu Rev Immunol*. 2013;31:443-473.
- 106. Maleno I, López-Nevot MA, Cabrera T, Salinero J, Garrido F. Multiple mechanisms generate HLA class I altered phenotypes in laryngeal carcinomas: high frequency of HLA haplotype loss associated with loss of heterozygosity in chromosome region 6p21. *Cancer Immunol Immunother*. 2002;51(7):389-396.
- 107. Maleno I, Cabrera CM, Cabrera T, et al. Distribution of HLA class I altered phenotypes in colorectal carcinomas: high frequency of HLA haplotype loss associated with loss of heterozygosity in chromosome region 6p21. *Immunogenetics*. 2004;56(4):244-253.
- 108. Feenstra M, Veltkamp M, Kuik J, Slootweg P, Tweel J, et al. HLA class I expression and chromosomal deletions at 6p and 15q in head and neck squamous cell carcinomas. *Tissue Antigens*. 1999;54(3):235-245.
- 109. Garrido MA, Rodriguez T, Zinchenko S, et al. HLA class I alterations in breast carcinoma are associated with a high

frequency of the loss of heterozygosity at chromosomes 6 and 15. *Immunogenetics*. 2018;70(10):647-659.

- 110. Maleno I, Romero JM, Cabrera T, et al. LOH at 6p21.3 region and HLA class I altered phenotypes in bladder carcinomas. *Immunogenetics*. 2006;58(7):503-510.
- 111. Sawasdee N, Thepmalee C, Sujjitjoon J, et al. Gemcitabine enhances cytotoxic activity of effector T-lymphocytes against chemo-resistant cholangiocarcinoma cells. *Int Immunopharmacol.* 2020;78:106006.
- 112. Gravett AM, Trautwein N, Stevanović S, Dalgleish AG, Copier J. Gemcitabine alters the proteasome composition and immunopeptidome of tumour cells. *Oncoimmunology*. 2018;7(6):e1438107.
- 113. Smith PL, Yogaratnam Y, Samad M, Kasow S, Dalgleish AG. Effect of gemcitabine based chemotherapy on the immunogenicity of pancreatic tumour cells and T-cells. *Clin Transl Oncol.* 2021;23(1):110-121.
- 114. Liu WM, Fowler DW, Smith P, Dalgleish AG. Pre-treatment with chemotherapy can enhance the antigenicity and immunogenicity of tumours by promoting adaptive immune responses. *Br J Cancer.* 2010;102(1):115-123.
- 115. Larson AC, Knoche SM, Brumfield GL, et al. Gemcitabine modulates HLA-I regulation to improve tumor antigen presentation by pancreatic cancer cells. *Int J Mol Sci.* 2024; 25(6):3211.
- 116. Ostwald TJ, MacLennan DH. Isolation of a high affinity calcium-binding protein from sarcoplasmic reticulum. *J Biol Chem.* 1974;249(3):974-979.
- 117. Milner RE, Baksh S, Shemanko C, et al. Calreticulin, and not calsequestrin, is the major calcium binding protein of smooth muscle sarcoplasmic reticulum and liver endoplasmic reticulum. *J Biol Chem.* 1991;266(11):7155-7165.
- 118. Camacho P, Lechleiter JD. Calreticulin inhibits repetitive intracellular Ca2+ waves. *Cell*. 1995;82(5):765-771.
- 119. Michalak M, Burns K, Andrin C, et al. Endoplasmic reticulum form of calreticulin modulates glucocorticoid-sensitive gene expression. *J Biol Chem.* 1996;271(46):29436-29445.
- 120. Spiro RG, Zhu Q, Bhoyroo V, Söling HD. Definition of the lectin-like properties of the molecular chaperone, calreticulin, and demonstration of its copurification with endomannosidase from rat liver Golgi. *J Biol Chem.* 1996;271(19):11588-11594.
- Obeid M, Panaretakis T, Joza N, et al. Calreticulin exposure is required for the immunogenicity of gamma-irradiation and UVC light-induced apoptosis. *Cell Death Differ*. 2007;14(10):1848-1850.
- 122. Panaretakis T, Joza N, Modjtahedi N, et al. The co-translocation of ERp57 and calreticulin determines the immunogenicity of cell death. *Cell Death Differ*. 2008;15(9):1499-1509.
- 123. Gardai SJ, McPhillips KA, Frasch SC, et al. Cell-surface calreticulin initiates clearance of viable or apoptotic cells through transactivation of LRP on the phagocyte. *Cell*. 2005;123(2):321-334.
- 124. Fucikova J, Truxova I, Hensler M, et al. Calreticulin exposure by malignant blasts correlates with robust anticancer immunity and improved clinical outcome in AML patients. *Blood*. 2016;128(26):3113-3124.
- 125. Kasikova L, Hensler M, Truxova I, et al. Calreticulin exposure correlates with robust adaptive antitumor immunity and favorable prognosis in ovarian carcinoma patients. *J Immunother Cancer*. 2019;7(1):312.
- 126. Fucikova J, Becht E, Iribarren K, et al. Calreticulin expression in human non-small cell lung cancers correlates with increased

accumulation of antitumor immune cells and favorable prognosis. *Cancer Res.* 2016;76(7):1746-1756.

- 127. Koido S, Kan S, Yoshida K, et al. Immunogenic modulation of cholangiocarcinoma cells by chemoimmunotherapy. *Anticancer Res.* 2014;34(11):6353-6361.
- 128. Solari J, Filippi-Chiela E, Pilar ES, et al. Damage-associated molecular patterns (DAMPs) related to immunogenic cell death are differentially triggered by clinically relevant chemotherapeutics in lung adenocarcinoma cells. *BMC Cancer*. 2020;20(1):474.
- 129. Dong H, Zhu G, Tamada K, Chen L. B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat Med.* 1999;5(12):1365-1369.
- Saunders PA, Hendrycks VR, Lidinsky WA, Woods ML. PD-L2:PD-1 involvement in T cell proliferation, cytokine production, and integrin-mediated adhesion. *Eur J Immunol*. 2005;35(12):3561-3569.
- 131. Qin G, Wang X, Ye S, et al. NPM1 upregulates the transcription of PD-L1 and suppresses T cell activity in triple-negative breast cancer. *Nat Commun*. 2020;11(1):1669.
- 132. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci USA*. 2002;99(19):12293-12297.
- 133. Nakanishi J, Wada Y, Matsumoto K, Azuma M, Kikuchi K, Ueda S. Overexpression of B7-H1 (PD-L1) significantly associates with tumor grade and postoperative prognosis in human urothelial cancers. *Cancer Immunol Immunother*. 2007;56(8):1173-1182.
- 134. Wang Q, Liu F, Liu L. Prognostic significance of PD-L1 in solid tumor: an updated meta-analysis. *Medicine*. 2017;96(18):e6369.
- 135. Zhang J, Fang W, Qin T, et al. Co-expression of PD-1 and PD-L1 predicts poor outcome in nasopharyngeal carcinoma. *Med Oncol.*. 2015;32(3):86.
- 136. Mognol GP, Spreafico R, Wong V, et al. Exhaustion-associated regulatory regions in CD8+ tumor-infiltrating T cells. *Proc Natl Acad Sci USA*. 2017;114(13):E2776-E2785.
- 137. Sun L, Zhang L, Yu J, et al. Clinical efficacy and safety of anti-PD-1/PD-L1 inhibitors for the treatment of advanced or metastatic cancer: a systematic review and meta-analysis. *Sci Rep.* 2020;10(1):2083.
- 138. Doi T, Ishikawa T, Okayama T, et al. The JAK/STAT pathway is involved in the upregulation of PD-L1 expression in pancreatic cancer cell lines. *Oncol Rep.* 2017;37(3):1545-1554.
- Azad A, Yin Lim S, D'Costa Z, et al. PD-L1 blockade enhances response of pancreatic ductal adenocarcinoma to radiotherapy. *EMBO Mol Med.* 2017;9(2):167-180.
- 140. Jung KH, Park JW, Lee JH, Moon SH, Cho YS, Lee KH. 89Zrlabeled anti-PD-L1 antibody PET monitors gemcitabine therapy-induced modulation of tumor PD-L1 expression. J Nucl Med. 2021;62(5):656-664.
- 141. Smigiel KS, Richards E, Srivastava S, et al. CCR7 provides localized access to IL-2 and defines homeostatically distinct regulatory T cell subsets. *J Exp Med*. 2013;211(1):121-136.
- 142. Guan Q, Blankstein AR, Anjos K, et al. Functional myeloidderived suppressor cell subsets recover rapidly after allogeneic hematopoietic stem/progenitor cell transplantation. *Biol Blood Marrow Transplant*. 2015;21(7):1205-1214.
- 143. Cassetta L, Bruderek K, Skrzeczynska-Moncznik J, et al. Differential expansion of circulating human MDSC subsets in

patients with cancer, infection and inflammation. *JImmunother Cancer*. 2020;8(2):e001223.

- 144. Taylor BC, Balko JM. Mechanisms of MHC-I downregulation and role in immunotherapy response. *Front Immunol*. 2022;13:844866.
- 145. Kong SK, Kim BS, Lim H, Kim HJ, Kim YS. Dissection of PD-L1 promoter reveals differential transcriptional regulation of PD-L1 in VHL mutant clear cell renal cell carcinoma. *Lab Invest.* 2022;102(4):352-362.
- 146. Antonangeli F, Natalini A, Garassino MC, Sica A, Santoni A, Rosa F. Regulation of PD-L1 expression by NF-κB in cancer. *Front Immunol.* 2020;11:584626.
- 147. Lv J, Wei Y, Yin JH, et al. The tumor immune microenvironment of nasopharyngeal carcinoma after gemcitabine plus cisplatin treatment. *Nat Med.* 2023;29(6):1424-1436.
- 148. Lee K, Kim DE, Jang KS, Kim SJ, Cho S, Kim C. Gemcitabine, a broad-spectrum antiviral drug, suppresses enterovirus infections through innate immunity induced by the inhibition of pyrimidine biosynthesis and nucleotide depletion. *Oncotarget*. 2017;8(70):115315-115325.
- 149. Lucas-Hourani M, Dauzonne D, Jorda P, et al. Inhibition of pyrimidine biosynthesis pathway suppresses viral growth through innate immunity. *PLoS Pathog.* 2013;9(10):e1003678.
- 150. Sprenger HG, MacVicar T, Bahat A, et al. Cellular pyrimidine imbalance triggers mitochondrial DNA-dependent innate immunity. *Nat Metab.* 2021;3(5):636-650.
- 151. Pan Q, de Ruiter PE, Metselaar HJ, et al. Mycophenolic acid augments interferon-stimulated gene expression and inhibits hepatitis C virus infection in vitro and in vivo. *Hepatology*. 2012;55(6):1673-1683.
- 152. Chung DH, Golden JE, Adcock RS, et al. Discovery of a broad-spectrum antiviral compound that inhibits pyrimidine biosynthesis and establishes a type 1 interferonindependent antiviral state. *Antimicrob Agents Chemother*. 2016;60(8):4552-4562.
- 153. Mullen NJ, Shukla SK, Thakur R, et al. DHODH inhibition enhances the efficacy of immune checkpoint blockade by increasing cancer cell antigen presentation. *bioRxiv* [Preprint]. 2024;13:2023.04.03.535399. doi:10.1101/2023.04.03.535399.
- 154. Nikolaou M, Pavlopoulou A, Georgakilas AG, Kyrodimos E. The challenge of drug resistance in cancer treatment: a current overview. *Clin Exp Metastasis*. 2018;35(4):309-318.
- Zitvogel L, Apetoh L, Ghiringhelli F, Kroemer G. Immunological aspects of cancer chemotherapy. *Nat Rev Immunol.* 2008;8(1): 59-73.
- 156. Oh DY, Lee KH, Lee DW, et al. Gemcitabine and cisplatin plus durvalumab with or without tremelimumab in chemotherapynaive patients with advanced biliary tract cancer: an open-label, single-centre, phase 2 study. *Lancet Gastroenterol Hepatol*. 2022;7(6):522-532.
- 157. Chen X, Wu X, Wu H, et al. Camrelizumab plus gemcitabine and oxaliplatin (GEMOX). *J Immunother Cancer*. 2020;8(2):e001240.
- 158. Padrón LJ, Maurer DM, O'Hara MH, et al. Sotigalimab and/ or nivolumab with chemotherapy in first-line metastatic pancreatic cancer: clinical and immunologic analyses from the randomized phase 2 PRINCE trial. *Nat Med.* 2022;28(6): 1167-1177.
- 159. Renouf DJ, Loree JM, Knox JJ, et al. The CCTG PA.7 phase II trial of gemcitabine and nab-paclitaxel with or without durvalumab

and tremelimumab as initial therapy in metastatic pancreatic ductal adenocarcinoma. *Nat Commun.* 2022;13(1):5020.

- 160. Sahai V, Griffith KA, Beg MS, et al. A randomized phase 2 trial of nivolumab, gemcitabine, and cisplatin or nivolumab and ipilimumab in previously untreated advanced biliary cancer: BilT-01. *Cancer*. 2022;128(19):3523-3530.
- 161. Kamdar M, Solomon ASR, Arnason J, et al. Lisocabtagene maraleucel versus standard of care with salvage chemotherapy followed by autologous stem cell transplantation as secondline treatment in patients with relapsed or refractory large B-cell lymphoma (TRANSFORM): results from an interim analysis of an open-label, randomised, phase 3 trial. *Lancet*. 2022;399(10343):2294-2308.
- 162. Kamath SD, Kalyan A, Kircher S, et al. Ipilimumab and gemcitabine for advanced pancreatic cancer: a phase Ib study. Oncologist. 2020;25(5):e808-e815.
- 163. Wang-Gillam A, Lim KH, McWilliams R, et al. Defactinib, pembrolizumab, and gemcitabine in patients with advanced treatment refractory pancreatic cancer: a phase I dose escalation and expansion study. *Clin Cancer Res.* 2022;28(24):5254-5262.
- 164. Wang-Gillam A, Lim KH, McWilliams R, et al. Erratum-Defactinib, Pembrolizumab, and gemcitabine in patients with advanced treatment refractory pancreatic cancer: a phase I dose escalation and expansion study. *Clin Cancer Res.* 2022;28(24):5254-5262.
- 165. Huang X, Liu Q, Zhong G, et al. Neoadjuvant toripalimab combined with gemcitabine and cisplatin in resectable locally advanced head and neck squamous cell carcinoma (NeoTGP01): an open label, single-arm, phase Ib clinical trial. *J Exp Clin Cancer Res.* 2022;41(1):300.
- 166. Feng K, Liu Y, Zhao Y, et al. Efficacy and biomarker analysis of nivolumab plus gemcitabine and cisplatin in patients with unresectable or metastatic biliary tract cancers: results from a phase II study. *J Immunother Cancer*. 2020;8(1):e000367.
- 167. Hewitt DB, Nissen N, Hatoum H, et al. A phase 3 randomized clinical trial of chemotherapy with or without algenpantucel-L (HyperAcute-pancreas) immunotherapy in subjects with borderline resectable or locally advanced unresectable pancreatic cancer. Ann Surg. 2022;275(1):45-53.
- 168. Mei Q, Zhang W, Liu Y, et al. Camrelizumab plus gemcitabine, vinorelbine, and pegylated liposomal doxorubicin in relapsed/refractory primary mediastinal B-cell lymphoma: a single-arm, open-label, phase II trial. *Clin Cancer Res.* 2020;26(17):4521-4530.
- 169. Palmer DH, Valle JW, Ma YT, et al. TG01/GM-CSF and adjuvant gemcitabine in patients with resected RAS-mutant adenocarcinoma of the pancreas (CT TG01-01): a single-arm, phase 1/2 trial. *Br J Cancer*. 2020;122(7):971-977.
- 170. Jiang H, Zheng Y, Qian J, et al. Efficacy and safety of sintilimab in combination with chemotherapy in previously untreated advanced or metastatic nonsquamous or squamous NSCLC: two cohorts of an open-label, phase 1b study. *Cancer Immunol Immunother*. 2021;70(3):857-868.
- 171. Cibula D, Rob L, Mallmann P, et al. Dendritic cell-based immunotherapy (DCVAC/OvCa) combined with second-line chemotherapy in platinum-sensitive ovarian cancer (SOV02): a randomized, open-label, phase 2 trial. *Gynecol Oncol.* 2021;162(3):652-660.
- 172. Roviello G, Ramello M, Catalano M, et al. Association between neutropenia and survival to nab-paclitaxel and

gemcitabine in patients with metastatic pancreatic cancer. *Sci Rep.* 2020;10:19281.

- 173. Blomstrand H, Green H, Fredrikson M, Gransmark E, Bjornsson B, Elander NO. Clinical characteristics and blood/serum bound prognostic biomarkers in advanced pancreatic cancer treated with gemcitabine and nab-paclitaxel. *BMC Cancer*. 2020;20:950.
- 174. Bruchard M, Mignot G, Derangère V, et al. Chemotherapytriggered cathepsin B release in myeloid-derived suppressor cells activates the Nlrp3 inflammasome and promotes tumor growth. *Nat Med.* 2013;19(1):57-64.
- 175. Konstantinopoulos PA, Costa AABA, Gulhan D, et al. A replication stress biomarker is associated with response to gemcitabine versus combined gemcitabine and ATR inhibitor therapy in ovarian cancer. *Nat Commun.* 2021;12(1):5574.
- 176. Wei CH, Gorgan TR, Elashoff DA, Hines OJ, Farrell JJ, Donahue TR. A meta-analysis of gemcitabine biomarkers in patients with pancreaticobiliary cancers. *Pancreas*. 2013;42(8):1303-1310.
- 177. Yu P, Luo S, Cai J, Li J, Peng C. ERAP2 as a potential biomarker for predicting gemcitabine response in patients with pancreatic cancer. *Aging*. 2022;14(19):7941-7958.

- 178. Leone P, Shin EC, Perosa F, Vacca A, Dammacco F, Racanelli V. MHC class I antigen processing and presenting machinery: organization, function, and defects in tumor cells. *J Natl Cancer Inst.* 2013;105(16):1172-1187.
- 179. Bauer C, Sterzik A, Bauernfeind F, et al. Concomitant gemcitabine therapy negatively affects DC vaccine-induced CD8+ T-cell and B-cell responses but improves clinical efficacy in a murine pancreatic carcinoma model. *Cancer Immunol Immunother*. 2014;63:321-333.

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