REVIEW

NF‑κ**B pathways in hematological malignancies**

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Abstract The nuclear factor κB or NF-κB transcription factor family plays a key role in several cellular functions, i.e. inflammation, apoptosis, cell survival, proliferation, angiogenesis, and innate and acquired immunity. The constitutive activation of NF-κB is typical of most malignancies and plays a major role in tumorigenesis. In this review, we describe NF-κB and its two pathways: the canonical pathway (RelA/p50) and the non-canonical pathway (RelB/ p50 or RelB/p52). We then consider the role of the NF-κB subunits in the development and functional activity of B cells, T cells, macrophages and dendritic cells, which are the targets of hematological malignancies. The relevance of the two pathways is described in normal B and T cells and in hematological malignancies, acute and chronic leukemias (ALL, AML, CLL, CML), B lymphomas (DLBCLs, Hodgkin's lymphoma), T lymphomas (ATLL, ALCL) and multiple myeloma. We describe the interaction of NF-κB with the apoptotic pathways induced by TRAIL and the transcription factor p53. Finally, we discuss therapeutic anti-tumoral approaches as mono-therapies or combination therapies aimed to block NF-κB activity and to induce apoptosis (PARAs and Nutlin-3).

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Introduction

The hallmarks of cancer are the distinctive and complementary capabilities of malignant cells to enable tumor growth and metastasis. They include sustenance of proliferative signaling, evasion of growth suppression, resistance to cell death, ability of replicative immortality, induction of angiogenesis, activation of invasion and metastasis, reprogramming of energy metabolism and evasion of immune destruction [[1\]](#page-14-0). As well, the inflammatory process has proved to be an emerging enabling hallmark for cancer progression [[2\]](#page-15-0). Very interestingly, the nuclear factor-κB (NF-κB) is able to induce most of these cellular alterations. There are several mechanisms that uncouple NF-κB from its normal regulatory processes and lead to its constitutive activation. Viral oncoproteins, i.e. Tax or v-Rel, have been shown to activate NF-κB and induce rapidly progressing lymphomas and leukemias. Cancer-related chromosomal translocations, deletion or mutations can disrupt genes encoding for the components of NF-κB signaling, i.e. MALT1, Bcl10, CARD11, TRAF3, NF-κB2, resulting in constitutive NF-κB activation. Constitutively activated NF-κB have been associated with several aspects of tumorigenesis, including promoting cancer cell proliferation, preventing apoptosis, increasing angiogenesis, cancer-related inflammation and metastatic potential [[3,](#page-15-1) [4\]](#page-15-2).

Before considering the role of the NF-κB pathways in hematological malignancies, we will introduce the pathways and the functions relevant to NF-κB activity in normal cells, with a particular attention to leukocytes, which represent the cellular targets of hematological malignancies.

Fig. 1 The canonical and non-canonical pathways of NF-κB and their interaction with the apoptotic pathways. The canonical and noncanonical NF-κB signaling pathways are represented in the *center* and to the *right* of the figure. The canonical pathway is activated through the TNF receptors' family, TLR4 and the antigen receptors BCR and TCR, while the non-canonical pathway is activated through a limited number of receptors, such as BAFF-R, CD40, RANK, CD30, and LTβ-R. The apoptotic pathways are summarized to the *left* of the figure in the extrinsic pathway (mediated by TRAIL) and the intrinsic

The two NF‑κ**B pathways**

The nuclear factor κB or NF-κB transcription factor family counts five subunits: RelA or p65, RelB, c-Rel or Rel, p50 (that with its precursor p105 is collectively known as NF- κ B1) and p52 (that with its precursor p100 is collectively known as NF-κB2). NF-κB subunits act as dimers to allow a functional NF-κB activation. All subunits share an ~300 residues long homologous element, referred to as the Rel homology region (RHR), which is responsible for dimerization, inhibitor binding, nuclear localization and DNA binding. These NF-κB proteins can form homodimeric and heterodimeric combinations, with the exception of RelB,

pathway (mediated by p53). *Arrows* indicate activating steps, and *bars* indicate inhibitory steps. *Numbers in circles* indicate the steps of the apoptotic pathways inhibited by molecules that are induced by NF-κB activation (c-FLIP, Bcl-2 family's molecules, IAPs). This is the case of ① procaspase 8 and 10, whose activation is inhibited by cFLIP, ② the pro-apoptotic molecules Bak/Bax inhibited by the prosurvival Bcl2, Bcl-X_L, Mcl-1, \circledcirc the effector caspases 3, 6, 7 targeted by the inhibitors of apoptosis (*IAPs*) family proteins, *④* the transcription factor p53 inhibited directly or indirectly by NF-κB

which does not share the common functional properties of the NF-κB family and seems not to form a stable, detectable homodimer. RelB has evolved into a specialized NFκB subunit with unique amino acids optimized for selective formation of heterodimers with p50 and p52 [\[5](#page-15-3), [6](#page-15-4)].

NF-κB signaling has been rationalized in two independent, yet interlinked, signaling pathways (Fig. [1](#page-1-0)).

The canonical or classical pathway, which is the most widely known NF-κB pathway, is mediated by the action of the RelA/p50 subunits (Fig. [1](#page-1-0), center). In resting conditions, RelA/p50 dimers reside in the cytoplasm complexed with the physiological inhibitor of NF-κB α (I κ B α), but upon cell activation, I κ B α is degraded by

ubiquitin-mediated proteasome activity. The complex RelA/p50 can translocate into the nucleus, and, in its active form, starts the transcription of target genes. The canonical pathway is activated by the $I \kappa B$ -kinase (IKK) complex, which is composed by IKKα or IKK1, IKKβ or IKK2, and IKK γ or NEMO, and that, in turn, phosphorylates IkB α and causes its degradation.

The non-canonical or alternative pathway is dependent on the activation of the RelB subunit associated with p50 or p52 (Fig. [1](#page-1-0), right). Differently from the canonical one, this pathway is activated by a more restricted number of ligands, such as the B cell-activating factor belonging to the TNF family (BAFF), CD40L, lymphotoxin β (LTβ), receptor activator nuclear factor-κB ligand (RANKL) or CD30L through the NF-κB-inducing kinase (NIK), which activates IKKα and, in turn, causes the degradation of p100, an IκBlike molecule which is the precursor of the p52 subunit. When cells are at rest, RelB is retained in the cytoplasm by p100 in the complexes RelB/p100 or RelB/p50/p100 and, upon p100 degradation, the dimers RelB/p50 and RelB/p52 [\[7](#page-15-5)] are released, translocate into the nucleus and start the transcription of target genes.

TNF-R-associated factors (TRAFs) are key intermediates in both the canonical and non-canonical NF-κB signaling pathways and act upstream of IKK or NIK activation, respectively. TRAF2 and TRAF3, in particular, seem to have distinct signaling functions and activities. While the recruitment of TRAF2 and TRAF6 is required for the activation of the canonical NF-κB pathway, TRAF3 does not seem to directly contribute to it, but rather to act as a negative regulator of TRAF2 and TRAF6 action, and seems to have a major role in the non-canonical pathway [[8\]](#page-15-6).

NF-κB subunits: functional roles by KO mice analysis

NF-κB was first discovered in 1986 in activated B cells for its property of interacting with the enhancer sequence of the immunoglobulin κ light-chain gene [\[9](#page-15-7)], stressing its relevance in B lymphocyte development. During the 1990s, the new technology of transgenic mice overexpressing exogenous copies of NF-κB or IκB genes, and of mice homozygous for null mutations [knock-out (KO) mice] in NF-κB, IκB or IKK genes, has provided invaluable models for elucidating the physiological functions and the regulation of the components of the two NF- κ B pathways [[10\]](#page-15-8) (see Table [1\)](#page-3-0). These approaches have shown that the different NF-κB subunits exert a relevant role in the development or functionality of mature lymphoid or myeloid cells. It was very soon evident that p50 and RelA are ubiquitously expressed in all cells, while the expression of RelB and p52 is restricted mostly to selected areas of hematopoietic organs and lymph nodes, i.e. thymic medulla, germinal centers (GCs), and marginal zone (MZ) of the

spleen. Moreover, RelB expression seems to be relevant in dendritic cells (DC) and B cells $[11-13]$ $[11-13]$. c-Rel expression is restricted to lymphocytes, monocytic, granulocytic, and erythroid cells in mouse fetal and adult hematopoietic organs [\[14](#page-15-11), [15](#page-15-12)].

Functionally, KO mice for RelA are characterized by embryonic lethality due to fetal hepatocyte apoptosis, while double KO mice for RelA and p50 show an earlier onset of embryonic death [\[16](#page-15-13)] (Table [1\)](#page-3-0). These findings indicate that the dimer RelA/p50 has an important antiapoptotic/prosurvival role, since its absence causes rapid cell death [\[17](#page-15-14)]. RelA acquires importance upon cell activation, since cell proliferation and functionality is impaired in RelA^{$-/-$} cells [\[18](#page-15-15)]. Moreover, RelA/p50 seems to be important in dendritic cell development from bone marrow precursors [[19\]](#page-15-16).

The overall picture for RelB KO mice is clearly different. Up to 50 % of mice die by 3 months from birth for multiple pathological lesions and defects in acquired and innate immunity, while double KO mice for RelB and p50 show a markedly increased inflammatory phenotype in both severity and extent of organ involvement, leading to premature death within 3–4 weeks after birth [\[20](#page-15-17)]. If RelA seems to have a general anti-apoptotic and proliferative role, RelB seems to mediate mostly anti-inflammatory responses and to affect the functionality of the cells involved in the immune response. In B cells, RelB is necessary for the normal production of Ag-specific IgG in response to T cell-dependent and T cell-independent stimulation [\[21](#page-15-18)], and in natural killer (NK) and T cells, RelB is required for the development of innate and adaptive responses [[22\]](#page-15-19) (Table [1\)](#page-3-0). Moreover, in RelB^{-/-} mice, T cell functions are impaired with T cell inflammatory infiltrates in various organs, and T cell-dependent myeloid hyperplasia and splenomegaly due to extramedullary hematopoiesis [\[23](#page-15-20), [24](#page-15-21)]. The absence of certain thymic and splenic dendritic cell (DC) populations [\[25](#page-15-22)] and the defective T cell-macrophage interaction in RelB^{-/−} mice [[21\]](#page-15-18) seem to indicate that the immune defects could be due to the functional impairment in DC. In double KO mice for RelB and p50, the functionality of myeloid and lymphoid cells is further impaired if compared to $\text{RelB}^{-/-}$ mice: B cells are absent from inflammatory infiltrates and their development and number are reduced, while myeloid hyperplasia increase and the thymus is atrophic [\[20](#page-15-17)].

c-Rel subunit is essential for a variety of functions in hematopoietic cells, although it is dispensable for the differentiation of hematopoietic precursors. Upon cell activation, c-Rel is required for T cell and B cell proliferation, isotype switching, cytokine and cell-surface activation markers production and for regulating gene expression and immune effector functions in different mature macrophage populations [\[14](#page-15-11), [15\]](#page-15-12). Recently, c-Rel has been found to be

Data are gathered from single and double KO mice for NF-κB subunits or components of the NF-κB pathways. Cells with a gray background indicate that the NF-κB signaling components are required for cell differentiation

Germinal centers (*GC*), marginal zones (*MZ*), Peyer's patches (*PP*), natural regulatory T cells (*nTreg*)

important for generating normal numbers of nTreg precursors and for the efficient induction, but not maintenance, of forkhead box P3 (Foxp3) + nTregs $[19, 26-28]$ $[19, 26-28]$ $[19, 26-28]$ $[19, 26-28]$.

Mice lacking the p50 subunit show multiple defects of immunity, since their B cells fail to proliferate in response to mitogens and elevated cell death [\[29](#page-15-25)] and are defective in basal and specific antibody production [[30\]](#page-15-26). On the contrary, mice lacking the p52 subunit show an abnormal architecture of spleen and lymph nodes since peri-follicular marginal zones and B follicular areas are absent and the defect seems to be related to an altered functionality of dendritic cells and macrophages rather than B and T cells' [\[11](#page-15-9), [12\]](#page-15-27). Double KO mice for p50 and p52 exhibit, soon after birth, growth retardation and craniofacial abnormalities, since they fail to generate mature osteoclasts and B cells, which are blocked at T1 stage [[31\]](#page-15-28).

KO mice for IκBα, the negative regulator of the RelA/ p50 complex, show, as expected, sustained NF-κB cellular activation and die 7–10 days postnatally, affected by a severe widespread inflammatory phenotype, indicating a major irreplaceable function of $I \kappa B\alpha$ in the termination of the NF-κB response [\[32,](#page-16-0) [33](#page-16-1)]. KO mice for the kinase IKKβ are characterized by embryonic death due to liver degeneration and apoptosis, a phenotype remarkably similar to RelA and p50 double KO mice [[34,](#page-16-2) [35\]](#page-16-3) (Table [1](#page-3-0)). IKK β is indeed the kinase of the IKK complex that is required for the activation of the canonical RelA/ p50 pathway.

Much of the knowledge gathered about the non-canonical pathway comes from studies on alymphoplasia (aly) mice, a natural strain with a mutant NIK gene [\[36\]](#page-16-4), or on mice genetically modified for IKKα, the IKKα KO mice or the IKK α^{AA} knock-in mice, in which the two serines in the activating phosphorylation sites of IKKα are replaced by alanines. IKKα KO mice die postnatally and show severe impairment of multiple morphogenetic events, including limb and skeletal patterning, and proliferation and differen-tiation of epidermal keratinocytes [[37](#page-16-5), [38\]](#page-16-6), while IKK α^{AA} mice are viable, but show defects in lymphoid organogenesis and GC formation $[39]$ $[39]$. IKK α is required for B cell maturation and the formation of secondary lymphoid organs, and induces the production of homeostatic chemokines, i.e. SLC, BLC, ELC, SDF1, and cytokines, i.e. BAFF, relevant in lymphoid organogenesis and adaptive immune reactions [[40,](#page-16-8) [41\]](#page-16-9).

Further in vivo and in vitro studies clarified that the two NF-κB pathways regulate several different functions that depend on the cell type and the stimulus applied [\[42](#page-16-10)]. The RelA/p50 canonical pathway was shown to promote inflammation, cell proliferation, cell survival through the production of several inhibitors of the apoptotic signaling, and to contribute to angiogenesis, tumor promotion and metastasis. The non-canonical pathway, instead, regulates lymphoid development and organization, adaptive immunity and has an anti-inflammatory activity [\[43](#page-16-11), [44](#page-16-12)].

Receptors of the non-canonical pathway

Both pathways play an important role in the functionality and development of myeloid and lymphoid cells, but the privileged localization of the RelB and p52 subunits in the lymphoid organs, where lymphocytes, macrophages, and dendritic cells reside and interact, seems to indicate an important involvement of the non-canonical pathway in lymphoid functions. Moreover, several members of the TNF receptors superfamily, such as LTβ-R, BAFF-R, CD40, RANK, have a major role in the activation of the non-canonical pathway, and mediate relevant functions in lymphocyte development and activation that stress the importance of this pathway in such activities. LTβ-R is expressed in lymphoid stromal and epithelial cells, and binds to two different ligands, lymphotoxin and LIGHT, both being primarily expressed in lymphocytes. The major function of LTβR is to mediate the development and maintenance of peripheral lymphoid organs. BAFF-R is predominantly expressed in B cells, and plays an important role in mediating the survival and maturation of peripheral B cells.

CD40 is a TNFR member that is expressed on various cell types, including B cells, DC, monocytes, endothelial cells, and neurons. A major function of CD40 signaling is to regulate B cell activation and differentiation events, including proliferation and survival of activated B cells, germinal center (GC) formation, and antibody isotype switching. Another major function of CD40 is to mediate DC maturation and antigen presentation. CD30 expression on normal cells is restricted to activated T and B lymphocytes and eosinophils, and its cross-linking activates pleiotropic signals depending on the cell type and agents used. In malignant cells, CD30 is highly expressed in Hodgkin's lymphoma (HL) and anaplastic large cell lymphoma (ALCL), making it a relatively specific target for immunotherapy [\[45](#page-16-13)]. RANK is expressed in osteoclast precursors, DC and activated B cells, and primarily regulates osteoclastogenesis. It also has a role in DC survival and lymphoid organogenesis [[46\]](#page-16-14). TNF-R2 is expressed by almost all cells, but differently from the TNF-R1, which is the main pro-inflammatory TNF receptor, its functions are not well known. In T cells, TNF-R2, but not TNF-R1, has been shown to induce the activation of the non-canonical pathway $[47]$ $[47]$. Since the two NF- κ B pathways play a very important role in the development, survival and functional activation of lymphocytes, DC and macrophages, it is not unexpected that these pathways, when defective or over-activated, are the culprits of several hematological malignancies.

Antigen receptors and NF‑κ**B**

The B cell receptor (BCR) is the organizing principle of B cell biology, playing an obligatory role in B cell development, survival, antigen-driven clonal selection, and humoral immunity. The active, antigen-dependent BCR signaling pathway activates NF-κB, and is started by BCRs' antigen binding and clustering on the cell membrane [\[48](#page-16-16)].

In this pathway, the BCR consists of the antigen-binding IgH and IgL chains that are non-covalently coupled to the CD79A (Ig- $α$) and CD79B (Ig- $β$) subunits, which regulate BCR surface expression, internalization, and trafficking (Fig. [1,](#page-1-0) center). Upon antigen encounter and BCR clustering, CD79A and CD79B activate a variety of downstream signaling pathways. Each of these subunits has an immunoreceptor tyrosine-based activation motif (ITAM) consisting of two tyrosines in a conserved amino acid setting. BCR aggregation promotes tyrosine phosphorylation of these ITAMs by Src-family kinases, principally LYN, FYN, and BLK. Dually phosphorylated ITAMs recruit the kinase SYK, whose activation initiates a signaling cascade that engages the signaling pathways NF-κB, phosphoinositide 3-kinase (PI3-K), nuclear factor of activated T cells (NF-AT) and mitogen-activated protein kinase (MAP-K), ultimately leading to cell survival and proliferation. In the signaling cascade that finally triggers the NF-κB pathways, SYK interacts with the Bruton tyrosine kinase (BTK) that, in turn, activates the phospholipase $C\gamma$ 2 (PLC γ 2). PLC γ 2 then catalyzes the hydrolysis of phosphatidylinositol-4,5-bisphosphate [PI(4,5)P2] into diacylglycerol (DAG) and inositol triphosphate (IP3), resulting in increased intracellular calcium levels. The combination of DAG and increased intracellular calcium activates protein kinase Cβ (PKCβ), which in turn phosphorylates many substrates, including caspase recruitment domain-containing protein 11 (CARD11), which is a domain of the adaptor protein CARMA1. Once activated, CARD11 move to the plasma membrane and binds BCL10 and the mucosa-associated lymphoid tissue lymphoma translocation protein 1 (MALT1) to form the complex termed the CBM complex, which, together with the transforming growth factor (TGF) β-activated kinase 1 (TAK1), is finally responsible for the activation of the IKK complex and the canonical NF-κB pathway. This pathway is characteristic of GC B cells and initiates the germinal center response towards selection and terminal differentiation of GC B cells. On the contrary, there is a 'tonic' BCR signaling typical of mature B cells that relies solely on PI3-K signaling, does not need BCR aggregation and is required for B cell survival. Such tonic signaling is likely to be antigen-independent as it affects all B cells, regardless of their IgH and IgL V regions.

In T lymphocytes, the primary activating signal is the recognition of suitable peptide–MHC ligands on the

surface of antigen presenting cells (APC). The binding of MHC/antigen complexes by the T cell receptor (TCR)/CD3 orchestrates the intracellular events that depend on protein tyrosine phosphorylation. The signaling downstream of the TCR is similar to that of the BCR. Following the activation of the TCR, the Src family kinase LCK is recruited to the TCR complex via its association with CD4 or CD8 coreceptors, and phosphorylates the conserved ITAMs of the TCR, leading to the recruitment of ZAP-70, a 70 kDa protein tyrosine kinase of the SYK family. In T cells, the protein kinase Cθ (PKCθ), and not PKCβ, acts on CARD11 and allows the recruitment of the CBM complex to the immunological synapse (IS), and the subsequent activation of the IKK complex and of the canonical NF-κB pathway. Signals from other cell surface receptors, in particular CD28, cooperate with TCR signaling to produce full T cell activation [[49\]](#page-16-17).

Both BCR and TCR activate the canonical NF-κB pathway and the functions that are modulated by this pathway. Other receptors are required for the development or the functionality of B and T cells, and often activate both the canonical pathway and the non-canonical pathway. This is the case for BAFF-R, CD40, RANK, CD30 and LTβ-R. While BAFF-R and LTβ-R mediate primarily the non-canonical NF-κB pathway, CD40 and RANK have a role in the canonical NF-κB pathway, too. This unique feature of BAFFR is primarily due to its possession of an atypical TRAF-binding sequence, which interacts with TRAF3 and allows the triggering of the non-canonical NF- κ B signaling [\[50](#page-16-18)].

NF‑κ**B in lymphoid and myeloid development**

Hematological malignancies are characterized by altered signaling of the molecules belonging to the intracellular signaling pathways activated by the antigen receptors on B cells (BCR) or T cells (TCR), several growth factor receptors, or by receptors accessory to immune functions. These malignancies derive from a precursor cell that can be at different stages of development and can rely more or less strongly on NF- κ B signaling [[51,](#page-16-19) [52\]](#page-16-20).

B cells

Prepro-B cells are the earliest recognized B-cell precursors in the bone marrow, and become pro-B cells as soon as they begin to rearrange genes that encode the variable regions of the immunoglobulin μ (Ig μ) heavy chain. When they begin to also recombine the variable regions of the light chains [V(D)J recombination] and start to assemble the BCR, B cells become small pre-B cells. This process is mediated by the recombination activating gene 1 (RAG1) and recombination activating gene 2 (RAG2). The canonical NF-κB is

more active in pre-B cells if compared to pro-B cells, promotes the transition to small pre-B cells [\[53](#page-16-21)], and together with the non-canonical pathway, controls the generation of λ-chain B cells [[54\]](#page-16-22). These cells, called immature B cells, strongly interact with self-antigens and are either eliminated by apoptosis (negative selection), or undergo further rearrangement at the Igκ or Igλ loci to modify their specificity [\[55](#page-16-23)]. When they move to the spleen, immature B cells die by apoptosis in response to BCR crosslinking, as a checkout for potential self-reactivity [[56\]](#page-16-24). During the transition from immature to mature B cells, the response to BCR interaction changes from cell death to cell proliferation, since B cells become responsive to the survival cytokine BAFF.

Immature B cells become transitional B cells through stages T1 and T2, and develop either into mature follicular (FO) B cells, which can leave and enter the circulation, and are the major source of T-dependent immunity and B cell memory, or into non-circulating marginal zone (MZ) B cells that can mount T-dependent and T-independent immune responses. At this stage, BCR cross-linking (1) induces the activation of the classical pathway of NF-κB and of c-Rel [\[57](#page-16-25)] with production of anti-apoptotic proteins, (2) upregulates the expression of *Nfkb2* gene resulting in the production of the p100 protein, and (3) induces the survival signaling mediated by BAFF [[58](#page-16-26)], which, in turn, mediates the activation of the non-canonical NF-κB pathway.

NF-κB activity seems to have a major role in MZ B cells, since the number of these cells is reduced in several of NF-κB subunit's single gene deficiencies, including *Nfkb1* and *RelB*, but not *c*-*Rel* [\[59](#page-16-27)]. Mature B cells, when stimulated by a non-self (foreign) antigen and costimulated by antigen specific T helper cells, migrate into the B cell follicles of secondary lymphoid organs, such as lymph nodes, spleen and Peyer's patches, and start a productive immune response. At this stage, a new microenvironmental niche, known as the germinal center (GC) of the secondary lymphoid tissues, is formed and comprises B cells, now developed into germinal center B cells (GC B cells), T follicular helper (TfH) cells and follicular dendritic cells (fDC). GC B cells undergo affinity maturation and clonal expansion, are large, highly dividing cells, and the IgH of their BCR switches to IgG, IgA or IgE [\[60](#page-16-28)]. The antigen trapped on fDC is seized by the GC B cells and presented to the TfH cells, producing an immunological synapse that activates the B cell. During the germinal center reaction, at least two distinct DNA modifications—(1) somatic hypermutation (SHM) and (2) class switch recombination (CSR)—occur that are mediated by the B cell specific enzyme activationinduced cytidine deaminase (AID). GC B cells that recognize, through their BCRs, the antigen with high affinity, are positively selected to differentiate terminally into antibodysecreting plasma cells or memory B cells.

We can conclude that both the canonical and non-canonical NF-κB pathway subunits are required for the functional activation of leukocytes, while the non-canonical pathway, in the form of the subunits p50, RelB/p50 and the kinase IKKα, has a privileged role in B cell generation and maintenance (see Table [1](#page-3-0)). The receptors BCR and BAFF-R are essential for mature B cell survival, since mice that carry genetic mutations in either one of these genes lack mature B cells [\[57](#page-16-25)].

T cells and myeloid cells

T cell development takes place in the thymus, where CD4−CD8− double negative (DN) thymocytes progress towards $CD4+CD8+$ double positive (DP) thymocytes, and finally to the mature $CD4^+$ or $CD8^+$ single positive T cells, which exit the thymus and enter the circulation. All thymocytes have some constitutive NF-κB activity, which is particularly high in the DN phase, when the pre-T cell receptor (pre-TCR) is assembled. Still, it is not clear whether NF-κB is essential for DN thymocytes' survival or only contributes to it. In DP thymocytes, TCR seems to activate NF-κB to pro-apoptotic functions in negative selection and to antiapoptotic/pro-survival activities in positive selection [\[61](#page-16-29)]. As for B cells, in mature peripheral $CD4^+$ or $CD8^+$ T cells, TCR activates NF-κB to pro-survival signaling, most probably via the canonical pathway, given the involvement of IKKβ and I κ B α . NF- κ B seems to have a role in the generation and suppressive functionality of thymic T regulatory cells [\[62](#page-16-30)]. In particular, the NF-κB subunit c-Rel binds to the conserved non-coding sequence 3 (CNS3) of the Foxp3 locus and plays a major role in thymic and peripheral FoxP3 expression [[63\]](#page-16-31). Moreover, in vitro studies have shown that NF-κB, in the form of c-Rel, RelA or p50 subunits, also plays a role in the development of effector T cells [\[64](#page-16-32)].

Granulocyte-macrophage colony-stimulating factor (GM-CSF) and macrophage colony stimulating factor (M-CSF) are among the growth factors responsible for the differentiation of a myeloid progenitor into granulocytes and/or macrophages. The NF-κB subunits c-Rel and RelA have been shown to be involved in the development of macrophages, differently from granulocytes [[27\]](#page-15-29). NF-κB subunits p50, RelA and RelB seem to contribute to the differentiation of DC from progenitors [\[19](#page-15-16), [25](#page-15-22)] (Table [1\)](#page-3-0), while the NF-κB canonical pathway [[65\]](#page-16-33), but not the non-canonical RelB/p50 [\[66](#page-17-0)], seems to drive DC functional activation.

Hematological malignancies

Hematological malignancies comprise a collection of heterogeneous diseases, all originating from cells of the bone

marrow and the lymphatic system. The overall prevalence of hematological cancers is increasing, and they represent the fifth most common cancer group. Hematological malignancies include leukemias, lymphomas, plasma cell neoplasms, myeloproliferative neoplasms, histiocytic tumor, and dendritic cell neoplasms.

Hematological malignancies are classified according to several parameters, such as cell morphology and immunephenotype, together with genetic, molecular and clinical features [\[67](#page-17-1)]. More recently, genomic studies including candidate gene sequencing, copy number profiling and whole-genome/exome sequencing have led to the identification of recurrent mutations and amplifications/deletions in a spectrum of hematologic malignancies that have been shown to be prognostic of the clinical outcome. The disease entities are stratified according to their cell lineage (myeloid, lymphoid, and histiocytic/dendritic neoplasms) [\[68](#page-17-2), [69\]](#page-17-3) and their derivation from precursor to mature lymphoid cells, as for leukemias, lymphomas or malignancies of plasma cells [[70,](#page-17-4) [71\]](#page-17-5).

Leukemias

Leukemias are generally characterized by the accumulation of immature myeloid or lymphoid cells in the bone marrow, accompanied by the presence of large numbers of abnormal or immature leukocytes in the peripheral blood. While acute leukemias are clinically defined by a more rapid disease timing, which progress over weeks to months, ultimately culminating in bone marrow failure, chronic leukemias have a less aggressive presentation. Depending on the originating cell lineage, acute and chronic leukemia can be divided into acute lymphoid or myeloid leukemia (ALL or AML) and chronic lymphoid or myeloid leukemia (CLL or CML).

ALL is the predominant pediatric leukemia, while AML is the most predominant adult acute leukemia. AML is characterized by the accumulation of immature myeloid cells (erythrocytes, granulocytes, monocytes, platelets) in the bone marrow. The major subgroups of ALL include the precursor B and T lymphoblastic leukemias (pre B-ALL and pre T-ALL) with about 85 % of ALLs being pre-B cell tumors that typically manifest as childhood acute leukemias with extensive bone marrow and variable peripheral blood involvement [\[72](#page-17-6)].

Among chronic leukemias, CML is a rare disease and was the first malignancy for which a recurrent chromosomal translocation was identified, due to the rearrangement of the ABL1 proto-oncogene with the BCR gene. Whereas normal ABL is a weak tyrosine kinase, the BCR–ABL1 fusion gene encodes a chimeric protein with markedly enhanced tyrosine kinase (TK) activity. Moreover, chronic leukemias include chronic myeloproliferative neoplasms or disorders (MPDs), which are clonal disorders of haemopoiesis characterized by excessive production of terminally differentiated myeloid cells (i.e. the granulocytic, erythroid and megakaryocytic series). Myelodysplastic syndromes (MDSs) show ineffective myelopoiesis, with both excessive and at the same time ineffective cell proliferation [[73\]](#page-17-7).

Lymphomas and plasma cell neoplasms

Lymphomas are a heterogeneous group of disorders characterized by the malignant proliferation of mature B or T cells, and generally present as solid tumors of the lymphoid system, particularly localized in lymph nodes, spleen, thymus and bone marrow. A few lymphomas can be localized at extranodal sites, such as skin, stomach, intestine or brain. The most common form of lymphoma is the diffuse large B cell lymphoma (DLBCL), which accounts for 30–40 % of newly diagnosed lymphomas. The molecular characterization of these tumors allowed DLBCL to be divided into three histologically indistinguishable molecular subtypes that differ in the expression of thousands of genes and apparently arise from B cells that are at separate stages of differentiation and activation. Germinal center B cell like (GCB) DLBCLs express genes typical of normal germinal center B cells, while activated B cell like (ABC) DLBCLs are characterized by genes found in B cells activated via the BCR. Primary mediastinal large B cell lymphomas typically express genes of mature B cells [\[74](#page-17-8)]. This classification has a prognostic relevance, since ABC DLBCL is associated to a less favorable outcome as compared to GCB DLBCL.

Other B lymphomas include Hodgkin's lymphomas, Burkitt's lymphoma, follicular lymphoma and marginal zone B cell lymphomas (MZL), which include the extranodal marginal zone B cell lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma), nodal MZL and primary splenic MZL. MALT lymphoma represents the most common extra-nodal lymphoma. Many cases of MALT lymphoma arise at sites affected by chronic inflammation and can be successfully treated by eradicating the source of inflammation. However, the concomitant insurgence of particular chromosomal abnormalities can promote disease progression.

Plasma cell or terminally differentiated B cell neoplasms have in common the expansion of a single clone of Agsecreting plasma cells and a resultant increase in serum level of a single homogeneous Ig or its fragments. Among these, multiple myeloma (MM) is the most important and the most common (10 % of hematological malignancies), and is characterized by multiple tumorous masses of neoplastic plasma cells scattered throughout the skeletal system [\[75](#page-17-9), [76\]](#page-17-10). It is noteworthy that, together with the clonal proliferation of malignant plasma cells, the bone marrow microenvironment of tumor cells has a pivotal role in the pathogenesis of multiple myeloma.

Mature T cell and natural killer (NK) cell neoplasms are less numerous than mature B cell neoplasms, and among them we can consider adult T cell leukemia/ lymphoma (ATLL), anaplastic large cell lymphomas (ALCL) or extranodal lymphomas, i.e. the enteropathyassociated T cell lymphoma, primary cutaneous T cell lymphomas [[77](#page-17-11)].

BCR and the pathogenesis of hematological malignancies

BCR activation is utilized not only by normal B cells, but also by neoplastic B cells for survival and expansion. This knowledge comes from gene expression profiling and functional analyses of BCR activation and signaling, as well as from the clinical efficacy of kinase inhibitors that interfere with BCR signaling.

The processes of V(D)J recombination in pre-B cells or of somatic hypermutation (SHM) and class switch recombination (CSR) in the germinal center reaction represent critical steps that might predispose to malignancy. For example, during V(D)J recombination, translocations $t(14;18)$ or $t(11;14)$ can arise and cause the dysregulation of the genes BCL2 or cyclin D1, which are juxtaposed to the IgH locus, and are characteristic of follicular lymphoma or mantle cell lymphoma, respectively [[78\]](#page-17-12). Activationinduced cytidine deaminase (AID), which operates during SHM and CSR, can mutate other genes in addition to Ig genes, as is the case of BCL6 or c-MYC. Deregulation of BCL6, which is a master regulator of germinal center B differentiation, characterizes most of DLBCLs, while translocation of the c-MYC gene is found in Burkitt's lymphoma. MALT lymphomas are associated with $t(1;14)$ and $t(11;18)$, where the $t(1;14)$ moves the IgG promoter upstream of the gene that encodes BCL10, and results in overexpression of a truncated BCL10 protein and constitutive NF-κB activation [\[79](#page-17-13), [80](#page-17-14)].

Genomic investigations of ABC DLBCL have provided clear genetic evidence that BCR signaling is central to the pathogenesis of human lymphomas [[48\]](#page-16-16) and depends on the activation of the NF-κB pathway. The B cell receptors in ABC DLBCL, similarly to BCRs in antigen-stimulated normal B cells, form prominent clusters in the plasma membrane. For these properties, the term 'chronic active BCR signaling' is used to describe the constitutive BCR activity in ABC DLBCL and several other lymphomas. On the contrary, Burkitt's lymphoma is characterized by the tonic BCR signaling and mainly depends on the activation of the PI3-K signaling pathway, and, together with GCB DLBCL, has been demonstrated to be scarcely dependent on NF-κB activity.

NF‑κ**B pathways in hematological malignancies**

The two NF-κB pathways seem to exert a specific role in the development of B and T cells or myeloid cells, with a major involvement of the non-canonical pathway in B cell development. Once matured, B cells and T cells exert their functional activities, probably through the combined activation of the two pathways, to allow the optimal cellular performance; on one side, the non-canonical pathway, which is more immune centered, and on the other side, the canonical pathway, which drives pro-survival and inflammatory activities.

The notion that the constitutive activation of NF-κB in cancer cells regulates the survival and pathogenesis of several malignancies is well acknowledged [[81,](#page-17-15) [82](#page-17-16)]. This can happen since NF-κB regulates the expression of proinflammatory cytokines, chemokines, growth factors, cell cycle regulators, anti-apoptotic molecules, adhesion molecules and matrix proteases. On the contrary, the individual contribution of the two NF-κB pathways in tumorigenesis is not so well appreciated. Here, we present an update of the current knowledge on the specific involvement of the canonical and non-canonical pathways in hematological malignancies (see Table [2](#page-9-0)).

AML

The myelodysplastic syndrome (MDS) comprises a group of hematopoietic stem cell disorders characterized by ineffective hematopoiesis leading to blood cytopenias and by a high risk of progression to acute myeloid leukemia (AML). In samples from 57 patients with MDS, activation of RelA was associated with blast counts, apoptosis suppression, and disease progression. Inhibition of the canonical pathway via siRNA, anti-RelA or Bortezomib, a drug that inhibits the proteasomal degradation of $I \kappa B\alpha$ and p100 (see ["Therapeutic approaches"](#page-13-0) for more details), induced rapid apoptosis of bone marrow cells [\[83](#page-17-17)]. A similar study on AML patients confirmed that the canonical pathway of NF-κB is constitutively active in AML cells in the form of RelA/p50 and p50/p50 complexes [[84\]](#page-17-18). Death-associated protein kinase 1 (DAPK1) is a tumor suppressor and its expression is epigenetically suppressed in several tumors. In AML, the FMS-like tyrosine kinase 3/internal tandem duplication (Flt3/ITD) initiates the activation of a noncanonical NF-κB signaling pathway mediated by RelB/ p52. It has been demonstrated that RelB/p52 is able to inhibit the tumor suppressor DAPK1 and promotes tumor development [\[85](#page-17-19)].

In murine erythroleukemia cells, the dimer RelB/p50, and not RelB or p50 alone, is induced by cAMP-dependent protein kinases and causes the upregulation of the transcription factor c-MYB, which is necessary for the proliferation

The triggers of the NFKB subunits' constitutive activation (if known) and the action mediated by the activated subunits are reported for several malignancies. Cells in gray indicate the few malignancies in which either the canonical or non-canonical pathway induces anti-tumorigenic activities. Data are presented in the RelB column when there is no mention in the paper of the subunit interacting with RelB. Data are from cell lines (*c.l.*), patients' material or mouse models. Constitutively active (c.a.)

of hematopoietic precursor cells, and in turn, inhibits cell differentiation [[86,](#page-17-20) [87\]](#page-17-21).

CML

In CML, the deregulated BCR-ABL tyrosine kinase has been demonstrated to induce NF-κB activation. In particular, constitutive activation of RelA-containing dimers has been measured in peripheral blood blasts from CML patients, indicating a role of the canonical NF-κB pathway in this malignancy [\[88](#page-17-22)]. We cannot exclude a contribution of the non-canonical pathway, since its activation has not yet been verified.

ALL

In a large population of patients with either B-ALL and T-ALL, the vast majority presents a constitutive activation of the canonical NF-κB pathway in the form of the RelA/p50 and p50/p50 complexes. It seems therefore that this constitutive activation of NF-κB serves as an important switch that ensures survival of ALL cells either by blocking apoptosis or by enhancing their proliferation [[89\]](#page-17-23). T-ALL is characterized by activating mutations of Notch1. When constitutively active, Notch1 positively regulates the expression of the NF-κB subunits RelB and NF-κB2, and activates the IKK complex, causing the activation of the non-canonical and canonical NF-κB pathways. Target genes overexpressed by NF-κB activation are BCL2A1, CD83, CCR7, CCL5, and TRAF1, which are regulated by both the canonical and the noncanonical pathways [\[90\]](#page-17-24).

CLL

In chronic lymphocytic leukemia (CLL), NF-κB has been found to be activated, to a variable degree, regardless of the disease stage or treatment status. In particular, RelAbinding complexes have been demonstrated to be constitutively active in peripheral blood samples of CLL patients, and their activation seems to be dependent on the action of the transcription activator STAT3 (signal transducer and activator of transcription-3) [[91\]](#page-17-25). More recently, a role for RelB has been determined, together with RelA, by studying B cells isolated from bone marrow aspirates of CLL patients. RelB activity seems not only to sustain tumor cell survival, but also to enhance cell sensitivity to proteasome inhibitor [[92\]](#page-17-26). In another study, RelB activation appeared to suppress apoptosis in CLL cell lines by inducing the anti-apoptotic gene Survivin [[93\]](#page-17-27). More generally, the canonical and non-canonical pathways seem to cooperate to CLL progression.

ATLL

Human T cell leukemia virus type I (HTLV-I) is an oncogenic retrovirus that is etiologically associated with adult T cell leukemia/lymphoma (ATLL). NF-κB is constitutively activated in this cancer in the form of RelA/p50 and p50/ p50 complexes [[94\]](#page-17-28) and can be activated by Tax, the transforming protein of HTLV-1, that directly interacts with the IKK complex or with IKKα and stimulates the processing of p100 to p52. Tax is thought therefore to activate both the canonical and the non-canonical pathways [\[95](#page-17-29)]. There are, anyway, alternative mechanisms independent of Tax that lead to NF-κB activation in this neoplasm. As for other lymphoid malignancies, in ATLL, a mutated form of the NF-κB2 gene has been described, and seems to interact with RelA and RelB and sustain the constitutive activation of NF-κB, playing a role in ATLL pathogenesis [\[96](#page-17-30)]. Another study performed on Tax-expressing mice demonstrated that NF-κB constitutive activation is dependent on RelB/p50 complex and follows the inactivation of the proapoptotic p53, indicating that decline in p53 function could be the first stage in the onset of ATLL [\[97](#page-17-31)].

DLBCL

Most genetic abnormalities in the diffuse large B cell lymphomas, except for the GCB DLBCL, result in the activation of the classical NF-κB pathway, with only two examples of mutations that are predictive of the activation of the non-canonical NF-κB pathway: (1) the structural alterations of the NF-κB2 gene and (2) the inactivation of TRAF3. By the way, it has not been demonstrated that their dysfunction will lead to the constitutive activation of the non-canonical pathway, rather than the canonical one [[98\]](#page-18-0).

ABC DLBCL, the most aggressive subtype of DLBCL, is strongly associated with the constitutive activation of the canonical NF-κB pathway. A recent paper has described a negative regulatory role of RelB in the activation of ABC DLBCL cell lines. RelB would be cleaved by the adapter Malt1 of the CBM complex and then undergo proteasomedependent degradation within a time frame that correlates with persistent nuclear translocation of NF-κB. In this way, RelB has been demonstrated to negatively regulate the activation of the canonical NF-κB, measured as RelA and c-Rel subunits, and therefore, its degradation by Malt1 could induce enhanced activation of the canonical pathway [\[99](#page-18-1)]. Other key features of this lymphoma are the genetic alterations that interfere with terminal B cell differentiation, as the inactivating mutations of the B lymphocyte induced maturation protein 1 (BLIMP1), a key regulator of plasma cell differentiation. It seems indeed that the canonical NFκB pathway, as an oncogenic event, and the BLIMP1, as

a tumor suppressor, interplay in the pathogenesis of ABC DLBCL [[100\]](#page-18-2).

Hodgkin's lymphoma and ALCL

Hodgkin's lymphoma can be divided into several subtypes, including the classical Hodgkin lymphoma (cHL), which is characterized by a low frequency of malignant Hodgkin and Reed–Sternberg (HRS) cells that derive from mature B cells embedded in a reactive background of non-neoplastic cells, with the majority of the tumor composed of various inflammatory cell types, including B cells, T cells, macrophages and granulocytes. This unique inflammatory milieu provides abundant cytokines that can activate NFκB. Anaplastic large cell lymphomas (ALCL) are unique lymphomas originating from cytotoxic T cells. They are defined by the proliferation of predominantly large lymphoid cells, and the cytotoxic molecules perforin and granzyme B.

ALCLs share a number of molecular aberrations with Hodgkin's lymphoma. Interestingly, the neoplastic HRS cells of cHL show a global downregulation of the B cellspecific gene expression program, whereas the T cellderived ALCLs have lost most of the expression of T cellspecific genes, supporting the view of a biologic analogy between both entities. Both lymphomas are characterized by the constitutive activation of the canonical and noncanonical NF-κB pathway and by the overexpression of the Notch1 and CD30 receptors, the last being overexpressed in multiple myeloma (MM), and adult T cell leukemia (ATLL), too.

In HL, it has been found that the maintenance of the HL phenotype strongly depends on the non-canonical NF-κB pathway, since the subunit RelB and the activating kinase NIK play a unique role in HL survival, differently from several DLBCLs [[101\]](#page-18-3). HL is often associated with Epstein–Barr virus (EBV), and the EBV-encoded latent membrane protein-1 (LMP-1) has been demonstrated to activate the canonical and non-canonical NFκB signaling pathways. Very interestingly, NF-κB constitutive activation has been observed in EBV-negative HL cell lines, and CD30 could be responsible for its activation [[102](#page-18-4)]. CD30 seems to activate the non-canonical pathway through the action of TRAF1, which is typically overexpressed in this malignancy [\[103\]](#page-18-5). Notch1, too, seems to play a role in the activation of the non-canonical pathway in HL, since it induces the processing of the NFκB2 gene product p100 into its active form p52. Moreover, Notch1 inhibition has been demonstrated to reduce NF-κB target genes and to impair the survival of HRS cells [\[104\]](#page-18-6).

Also in ALCL, CD30 is among the receptors responsible for NF-κB activation and mediates several functions, i.e.

cell proliferation, but also cell cycle arrest and apoptosis. It is very interesting that only in ALCL, and not in HL, CD30 is able to mediate anti-proliferative effects. In particular, a prolonged stimulation of CD30 has been demonstrated to induce cell cycle arrest through the activation of the canonical pathway (RelA) [\[105](#page-18-7)], depending on specific intracellular domains of CD30 [[45\]](#page-16-13).

Finally, in both ALCL and HL, the dimer p50/p50 seems to be constitutively active and would be induced by the overexpression of Bcl3, a protein that shares structural features with IκBα proteins. Bcl3 has an anti-apoptotic effect in B and T lymphocytes and is tumorigenic in several malignancies [[106\]](#page-18-8).

Burkytt's lymphoma

Human Burkitt's lymphoma (BL) is a highly aggressive lymphoma, with the transcriptional deregulation of MYC due to chromosomal translocation being the pivotal event in lymphomagenesis. This lymphoma is characterized by a very low basal NF-κB activity consisting of RelB-containing heterodimers. Upon IKKβ activation, NF-κB is induced in the form of RelA and p50-containing complexes, and a reduced cell proliferation is observed together with enhanced apoptosis. MYC is able to induce the activation of the p53 pathway, and therefore apoptosis as a safeguard mechanism against malignant transformation. It seems that MYC overexpression sensitizes cells to NF-κB-induced apoptosis, and that persistent inactivity of NF-κB signaling is a prerequisite for MYC-mediated tumorigenesis [\[107](#page-18-9)].

Multiple myeloma

The activation of both the canonical and non-canonical NFκB pathway is a clear hallmark of multiple myeloma and the driving force for its pathogenesis. The genetic screening of 155 MM samples reported mutations in ten genes involved in NF-κB signaling that cause the inactivation of TRAF2, TRAF3, cylindromatosis (CYLD), cellular inhibitor of apoptosis (cIAP)1/cIAP2 and the activation of NFκB1, NF-κB2, CD40, LTBR, transmembrane activator and calcium-modulator, and cytophilin ligand interactor (TACI) [\[82](#page-17-16), [108](#page-18-10), [109\]](#page-18-11). More recently, it has been described that NF-κB2 mutation selectively activates the non-canonical NF-κB pathway, whereas CYLD, NF-κB1, and TACI mutations selectively activate the canonical pathway. Moreover, mutations affecting NIK expression, such as mutations in NIK, TRAF2, TRAF3, cIAP1, cIAP2, and CD40 genes, seem to activate primarily the non-canonical pathway. NIK overexpression and TRAF3 inactivation are two very common mechanisms in MM pathogenesis, even if their preferential involvement in the canonical or non-canonical NFκB is still debated [[108,](#page-18-10) [110\]](#page-18-12).

Recently, Baud et al. described further the important role of the non-canonical pathway in the pathogenesis of MM. RelB was found to be constitutively active in nearly 40 % of the newly diagnosed MM patients studied, to induce the expression of the anti-apoptotic genes cIAP2, Bcl- X_L and Bcl-2, and to promote cell survival [[111\]](#page-18-13).

The tumor microenvironment is extremely relevant to MM pathogenesis, growth and drug resistance. Adhesion of MM cells to fibronectin induces the activation of the complex RelB/p50 of NF-κB and expression of the antiapoptotic cIAP2, indicating non cell-autonomous pro-tumoral activities from the microenvironment cells [\[112](#page-18-14)].

Microenvironment relevance in tumorigenesis

The tumor microenvironment is constituted by a varied society of cells (fibroblasts, endothelial cells, smooth muscle cells, granulocytes, B, T and NK lymphocytes, mast cells, macrophages and dendritic cells), and by the extracellular matrix (ECM) that surrounds cancer cells. Noncell-autonomous, or micro-environmental, factors play an important role in tumor development, and therefore have been considered as important therapeutic targets for cancer treatment. Moreover, it has been demonstrated that gene expression profiling of the microenvironment is a good predictive tool of the clinical course of the malignancy [\[113](#page-18-15)].

There are several examples of non-autonomous relevance in tumorigenesis. In Hodgkin's lymphoma, the scarce HRS cells are embedded in a majority of inflammatory cells, and by expressing high level of several TNF receptor family members, can be chronically stimulated by the pro-inflammatory cytokines produced by the microenvironment. A recent work on an in vivo model of human T-ALL showed the unique role of the non-canonical NF-κB pathway (RelB subunit), which was activated in the stromal compartment and exerted a non-autonomous tumorigenic effect on leukemic cells [[114\]](#page-18-16). CLL is characterized by the accumulation of mature, monoclonal B lymphocytes in peripheral blood, bone marrow, lymph nodes and other secondary lymphoid organs. It has been demonstrated that the lymph node microenvironment promotes B cell receptor signaling, NF-κB activation, and tumor proliferation [\[115](#page-18-17)]. Another study demonstrated that, by co-culturing CCL bone marrow-derived B cells and bone marrow-derived stromal cells, the presence of high RelB activity, together with RelA activity, conferred survival advantages to CLL BM B cells [\[92](#page-17-26)]. Finally, in multiple myeloma, the activation of NF-κB in bone marrow stromal cells (BMSCs) also contributes to MM pathogenesis, since several NF-κBdependent factors produced by BMSCs lead to MM growth and prevent cell apoptosis [[82\]](#page-17-16).

From this overview on the canonical and non-canonical pathways in hematological malignancies, we can

summarize that in most neoplasms, from B and T leukemias to B and T lymphomas and multiple myeloma, both pathways are activated. The canonical pathway has generally a major role in tumorigenesis, as for ABC DLBCL, ALL, CML, AML, and MM (Table [2](#page-9-0)), but its relevance is clearly tumor and context-dependent. There are some exceptions to be considered: in ALCL (Table [2\)](#page-9-0), the canonical pathway has the ability to induce growth arrest and apoptosis depending on CD30 stimulation, and in Burkitt's lymphoma, a cancer with a low NF-κB activity, the canonical pathway, when activated, seems to mediate apoptosis and to counteract the oncogenic action of MYC.

Several evidences point to the relevant role of the noncanonical pathway in the pathogenesis of Hodgkin's lymphoma, probably due to its peculiar features, a deregulated BCR signaling, the overexpression of the receptors able to activate the non-canonical pathway, and the great majority of inflammatory cells surrounding the HRS cells that can chronically stimulate cancer cells. Very interestingly, in ABC DLBCL, RelB has been shown to counteract the activity of the canonical pathway and to induce a final antitumorigenic and pro-apoptotic activity. An inhibitory action of RelB on the canonical pathway or on genes expressed by the canonical pathway has been described in several cell types [\[116](#page-18-18), [117](#page-18-19)], and recently, RelB has been shown to inhibit cell proliferation and tumor growth through the activation of the transcription factor p53 [[118\]](#page-18-20). In some malignancies, RelB activation could be a mechanism to control excessive canonical pathway activation. Whether RelB is a tumor suppressor or tumor promoter depends most likely on several factors, such as the tumor type and the surrounding microenvironment.

NF‑κ**B signaling and apoptotic pathways**

The cellular commitment to survival or cell death is fundamental to the benefit of the organism, and errors in this process can lead to autoimmunity or cancer. One of the main functions of NF-κB is the blockade of cell death through the transcriptional induction of genes encoding anti-apoptotic and antioxidant proteins. The apoptotic pathway is controlled by two distinct, yet interlinked, signaling pathways. Extrinsic apoptosis is induced by the interaction of the pro-apoptotic ligands—tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), TNF or Fas ligand—to their specific receptors on infected or damaged cells. Intrinsic apoptosis can be triggered by several intracellular stress conditions, including DNA damage and oxidative stress, which can activate the transcription factor p53. NF-κB has been demonstrated to counteract apoptosis by regulating critical steps along the apoptotic pathways [\[119](#page-18-21)] (see Fig. [1](#page-1-0), left).

TRAIL activates the extrinsic apoptotic signaling pathway through TRAIL-R1 or TRAIL-R2, leading to the activation of caspase-8 and -9 and the assembly of the deathinducing signaling complex (DISC). Then, DISC activates Bid, which interacts with Bax and Bak and starts the intrinsic apoptotic pathway, or directly activates caspase-3, -6 and -7, inducing apoptosis. NF-κB activation induces the expression of several anti-apoptotic components of TRAIL signaling. Examples are (1) c-FLIP, which inhibits the activation of the caspases $8/10$ (Fig. [1,](#page-1-0) 0); (2) the pro-survival Bcl-2 proteins (Bcl-2, Bcl- X_L , Mcl-1), which inhibit the action of the pro-apoptotic (Bax, Bak) components (Fig. [1,](#page-1-0) ②); (3) inhibitor of apoptosis (IAP) proteins (c-IAP1, c-IAP2, X-IAP, survivin), which inhibit the caspases involved in the effector phase of apoptosis (Fig. [1](#page-1-0), ③).

The p53 tumor suppressor and transcription factor is one of the first lines of defense against the effects of genotoxic damage, oncogene activation, metabolic changes and hypoxia [\[120](#page-18-22)]. p53 induces the activation of the intrinsic apoptotic pathway, as previously described [\[121\]](#page-18-23). Briefly, p53 upregulates NOXA and PUMA, two proteins of the Bcl-2 family that interact with Bax and Bak at the mitochondrial site and start the apoptotic cascade with the formation of the signaling complex apoptosome, and the final activation of the effector caspases-3, -6 and -7 that induce cell apoptosis. The amount of p53 protein in cells is regulated by HDM2 (known as MDM2 in mice), which functions as an E3 ubiquitin ligase and induces p53 proteolysis with the final inhibition of the apoptotic pathway. Expression of HDM2 and the consequent downregulation of p53 can be induced by IKK and by NF-κB activation. NF-κB interacts with $p53$ in different ways, and it seems to act mainly as an antagonist to p53 transactivation (Fig. [1,](#page-1-0) ④). The canonical pathway seems to have a major role: IKKβ was shown to induce NF-κB activation, HDM2 expression and the subsequent p53 downregulation [\[122](#page-18-24)], while RelA was able to antagonize p53 activation through the sequestration of the p300 and CREB binding protein (CBP) co-activators [[123](#page-18-25)]. Also, the non-canonical pathway has been demonstrated to have an antagonistic role to p53, since $IKK\alpha$, through the phosphorylation of CBP, switches the binding preference of CBP from p53 to NF-κB [\[124](#page-18-26)]. Recent evidence reports that the canonical and non-canonical NF-κB pathways can, under some circumstances, mediate the pro-apoptotic effect, growth arrest and the inhibition of cancer development. In particular, RelA was able to stabilize p53 and cooperate with the pro-apoptotic signaling (via TRAIL-R2 induction) [\[125](#page-19-0), [126](#page-19-1)], and RelB could inhibit cell proliferation and tumor growth through p53 transactivation [\[127](#page-19-2)]. NF-κB2 is aberrantly expressed in many tumor types and has been implicated as a regulator of cell proliferation. In particular, p52 was shown to cooperate with p53 to regulate p53 target genes such as PUMA, TRAIL-R2 by associating directly to the p53-regulated promoters [[128\]](#page-19-3). Very interestingly, p53 has a non-autonomous effect, since it has a pro-tumorigenic role when its pathway is deregulated in stromal cells [[129](#page-19-4)], or, when expressed in stromal fibroblasts, p53 is able to attenuate cancer cell migration and invasion [\[130](#page-19-5)].

Therapeutic approaches

Given their relevance in cancer pathogenesis and progression, NF-κB and its signaling components are considered a good therapeutic target for the treatment of malignancies [[131](#page-19-6)].

To this purpose, numerous small inhibitory peptides have been designed to target specific patterns of the molecules most relevant to NF-κB activation, i.e. IKKs kinases, the proteasome 26S, or, more recently, the NF-κB subunits, and block their activity. Bortezomib is an inhibitor of the proteasome 26S, which is responsible for the degradation of IκBα and the processing of p100. Bortezomib has been approved for the treatment of newly diagnosed multiple myelomas, relapsed/refractory multiple myelomas, and mantle cell lymphomas. The anti-cancer mechanisms of Bortezomib, described in preclinical studies, include the upregulation of pro-apoptotic proteins (i.e. Noxa, IκBα), the inhibition of NF-κB and the suppression of several anti-apoptotic proteins (Bcl- X_L , Bcl-2). Other compounds selectively target and inhibit the catalytic subunits of the IKKs, preventing IκBα from degradation and the canonical NF-κB subunits from nuclear translocation. The main problem with the current targeting of kinase activity or proteasome degradation is specificity: the blocked mechanisms share their activity with other signaling pathways, and therefore, an effect on a pathway different from NF-κB can be observed. By using innovative technologies with an increased specificity to the target, such as siRNA, aptamers or combination of the two [[132\]](#page-19-7), potential toxic effects could be avoided. As we recently described, the optimization of these drugs with increased specificity towards NFκB subunits is ongoing [[42\]](#page-16-10).

TRAIL has emerged as a promising candidate for cancer therapy by virtue of its ability to trigger extrinsic apoptosis in various types of cancer cells without significant toxicity toward normal cells. The description and rationale for TRAIL-based therapies have been extensively treated in a recent review [[133\]](#page-19-8), and have shown to be relevant for the treatment of hematological malignancies, too [[134,](#page-19-9) [135](#page-19-10)]. Recombinant TRAIL molecules or agonistic antibodies targeting TRAIL receptors, termed pro-apoptotic receptor agonists (PARAs), have been shown to activate the apoptotic pathway that is often inhibited in cancer cells, due, among other factors, to p53 inactivation. A major limitation to the clinical benefit derived from cancer drug therapies, and from TRAIL-based therapies as well, is drug resistance. Mechanisms of TRAIL resistance have been extensively studied in the last decade, and it seems that NFκB activation, which is generally deregulated in cancer, has a major role in it.

Another important anti-neoplastic strategy relies on the possibility to activate the p53 pathway by disrupting the protein interaction between p53 and MDM2. The Nutlins were identified as the first potent and selective small-molecule MDM2 antagonists able to inhibit the p53/MDM2 interaction, leading to stabilization of p53 and activation of the p53 pathway [[136](#page-19-11)]. Nutlins are able to induce apoptotic cell death when added to primary leukemic cell cultures, but also to promote leukemic maturation [[137\]](#page-19-12), and paradoxically activate anti-apoptotic genes [\[138\]](#page-19-13). This suggests a preferential use of Nutlins in synergistic effect when used in combination with the chemotherapeutic drugs for the treatment of hematological malignancies. Nutlins also display noncell autonomous biological activities, such as inhibition of osteoprotegerin [[139](#page-19-14)], a soluble inhibitor of TRAIL [[140](#page-19-15)], and stromal derived factor-1 (SDF-1) expression by primary endothelial cells or stromal fibroblasts, respectively [\[130\]](#page-19-5), or induction of T cell proliferation by dendritic cells [\[141](#page-19-16)].

Combination therapies

In hematological malignancies, there are several evidences of the antagonism between the apoptotic pathway driven by TRAIL or p53, and the pro-survival NF-κB pathway. Whatever the nature of the anti-tumoral drug considered, the activation of the NF-κB pathway can confer resistance to the treatment.

In follicular lymphoma, resistance to TRAIL-based therapies is promoted by CD40L through the activation of NFκB and the subsequent upregulation of c-FLIP and Bcl- X_L [\[142](#page-19-17)]. In CLL, constitutively activated Notch 1 signaling is involved in the activation of NF-κB and in the increased expression of c-IAP2 and X-IAP, which eventually cause survival and apoptosis resistance [\[143](#page-19-18)]. Drug resistance can also be driven in a non-autonomous fashion by stromal cells, as in CLL, where stromal cells of the bone marrow express CD40L and show NF- κ B activity [\[144](#page-19-19), [145\]](#page-19-20); or in MM, due to increased synthesis of c-FLIP by the NF-κBactivated stromal microenvironment [[146\]](#page-19-21).

Combined regimens of therapeutics inhibiting NF-κB activation on one side and specifically activating the proapoptotic pathway have shown promising results, both in preclinical models and in clinical trials. This is the case of hematological malignancies, too. Nutlin-3, by stabilizing p53, enhances Bortezomib-mediated mitochondrial apoptosis in p53-mutated mantle cell lymphomas [\[147](#page-19-22)], or potentiates the cytotoxic effects of Bortezomib in multiple myelomas [\[148](#page-19-23)], even overcoming non-autonomous protumorigenic effects of surrounding stromal cells [\[149](#page-19-24)].

Bortezomib, on the other side, cooperatively enhances TRAIL-mediated apoptosis in AML [[150\]](#page-19-25), CLL, MM [\[146](#page-19-21)], and Burkitt's lymphoma via upregulation of TRAIL and its death receptors [\[151](#page-19-26)]. Failure in overcoming TRAIL resistance in TRAIL-based therapy could be due to the fact that, along with the apoptotic extrinsic pathway, TRAIL is able to induce non-apoptotic functions on several malig-nancies [\[152](#page-19-27)]; a fact that has been ascribed to the activation of NF-κB and PI3-K/AKT [\[153](#page-19-28)]. Therefore, the final choice of the combination therapy to consider strongly depends on the particular malignancy that is targeted.

Conclusions

The anti-apoptotic role of the canonical NF-κB pathway is a common feature of all malignancies, independent of the cell of origin. Hematological malignancies can be considered differently from other malignancies in terms of NF-κB pathways, since (1) the components of the non-canonical pathway are preferentially expressed in lymphoid organs where the development and activation of lymphoid and myeloid cells take place; and (2) the receptors that mediate the non-canonical pathway are strongly involved in the processes of immune response and lymphocyte survival. For these reasons, hematological malignancies appear to be a preferential site for the action of the non-canonical NFκB signaling. In this regard, it is interesting that in Hodgkin's lymphoma, characterized by unpaired BCR activity and enhanced expression of "non-canonical receptors", the non-canonical pathway seems to have a relevant role, while in ABC DLBCL, which is strongly dependent on the BCRdependent activation of the canonical pathway, RelB activity antagonizes the activation of the canonical pathway.

More research is needed to better define the relative role of the two NF-κB pathways, and in particular, in what cancers the non-canonical pathway cooperate or inhibit the activation of the canonical pathway. In this way, new therapies targeting specifically either or both NF-κB pathways could be developed. The combined therapeutic approach, obtained by blocking NF-κB and inducing the apoptotic response, will possibly obtain a most favorable outcome.

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Conflict of interest The authors declare no conflicts of interest.

References

1. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144(5):646–674. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.cell.2011.02.013) [cell.2011.02.013](http://dx.doi.org/10.1016/j.cell.2011.02.013)

- 2. Mantovani A, Allavena P, Sica A, Balkwill F (2008) Cancerrelated inflammation. Nature 454(7203):436–444. doi[:10.1038/](http://dx.doi.org/10.1038/nature07205) [nature07205](http://dx.doi.org/10.1038/nature07205)
- 3. Karin M, Cao Y, Greten FR, Li ZW (2002) NF-kappaB in cancer: from innocent bystander to major culprit. Nat Rev Cancer 2(4):301–310. doi[:10.1038/nrc780](http://dx.doi.org/10.1038/nrc780)
- 4. Ben-Neriah Y, Karin M (2011) Inflammation meets cancer, with NF-kappaB as the matchmaker. Nat Immunol 12(8):715–723. doi:[10.1038/ni.2060](http://dx.doi.org/10.1038/ni.2060)
- 5. Huang DB, Vu D, Ghosh G (2005) NF-kappaB RelB forms an intertwined homodimer. Structure 13(9):1365–1373. doi:[10.1016/j.str.2005.06.018](http://dx.doi.org/10.1016/j.str.2005.06.018)
- 6. Vu D, Huang DB, Vemu A, Ghosh G (2013) A structural basis for selective dimerization by NF-kappaB RelB. J Mol Biol 425(11):1934–1945. doi:[10.1016/j.jmb.2013.02.020](http://dx.doi.org/10.1016/j.jmb.2013.02.020)
- 7. Derudder E, Dejardin E, Pritchard LL, Green DR, Korner M, Baud V (2003) RelB/p50 dimers are differentially regulated by tumor necrosis factor-alpha and lymphotoxin-beta receptor activation: critical roles for p100. J Biol Chem 278(26):23278– 23284. doi[:10.1074/jbc.M300106200](http://dx.doi.org/10.1074/jbc.M300106200)
- 8. Vallabhapurapu S, Matsuzawa A, Zhang W, Tseng PH, Keats JJ, Wang H, Vignali DA, Bergsagel PL, Karin M (2008) Nonredundant and complementary functions of TRAF2 and TRAF3 in a ubiquitination cascade that activates NIK-dependent alternative NF-kappaB signaling. Nat Immunol 9(12):1364–1370. doi:[10.1038/ni.1678](http://dx.doi.org/10.1038/ni.1678)
- 9. Sen R, Baltimore D (1986) Inducibility of kappa immunoglobulin enhancer-binding protein Nf-kappa B by a posttranslational mechanism. Cell 47(6):921–928. doi:[10.1016/0092-8674\(86\)90807-X](http://dx.doi.org/10.1016/0092-8674(86)90807-X)
- 10. Gerondakis S, Grossmann M, Nakamura Y, Pohl T, Grumont R (1999) Genetic approaches in mice to understand Rel/NF-kappaB and IkappaB function: transgenics and knockouts. Oncogene 18(49):6888–6895. doi[:10.1038/sj.onc.1203236](http://dx.doi.org/10.1038/sj.onc.1203236)
- 11. Caamano JH, Rizzo CA, Durham SK, Barton DS, Raventos-Suarez C, Snapper CM, Bravo R (1998) Nuclear factor (NF) kappa B2 (p100/p52) is required for normal splenic microarchitecture and B cell-mediated immune responses. J Exp Med 187(2):185–196. doi:[10.1084/jem.187.2.185](http://dx.doi.org/10.1084/jem.187.2.185)
- 12. Franzoso G, Carlson L, Poljak L, Shores EW, Epstein S, Leonardi A, Grinberg A, Tran T, Scharton-Kersten T, Anver M, Love P, Brown K, Siebenlist U (1998) Mice deficient in nuclear factor (NF)-kappa B/p52 present with defects in humoral responses, germinal center reactions, and splenic microarchitecture. J Exp Med 187(2):147–159. doi[:10.1084/jem.187.2.147](http://dx.doi.org/10.1084/jem.187.2.147)
- 13. Weih DS, Yilmaz ZB, Weih F (2001) Essential role of RelB in germinal center and marginal zone formation and proper expression of homing chemokines. J Immunol 167(4):1909–1919
- 14. Grigoriadis G, Zhan Y, Grumont RJ, Metcalf D, Handman E, Cheers C, Gerondakis S (1996) The Rel subunit of NF-kappaBlike transcription factors is a positive and negative regulator of macrophage gene expression: distinct roles for Rel in different macrophage populations. EMBO J 15(24):7099–7107
- 15. Gerondakis S, Strasser A, Metcalf D, Grigoriadis G, Scheerlinck JY, Grumont RJ (1996) Rel-deficient T cells exhibit defects in production of interleukin 3 and granulocyte-macrophage colony-stimulating factor. Proc Natl Acad Sci USA 93(8):3405–3409. doi[:10.1073/pnas.93.8.3405](http://dx.doi.org/10.1073/pnas.93.8.3405)
- 16. Horwitz BH, Scott ML, Cherry SR, Bronson RT, Baltimore D (1997) Failure of lymphopoiesis after adoptive transfer of NF-kappaB-deficient fetal liver cells. Immunity 6(6):765–772. doi:[10.1016/S1074-7613\(00\)80451-3](http://dx.doi.org/10.1016/S1074-7613(00)80451-3)
- 17. Beg AA, Baltimore D (1996) An essential role for NF-kappaB in preventing TNF-alpha-induced cell death. Science 274(5288):782–784. doi:[10.1126/science.274.5288.782](http://dx.doi.org/10.1126/science.274.5288.782)
- 18. Doi TS, Takahashi T, Taguchi O, Azuma T, Obata Y (1997) NFkappa B RelA-deficient lymphocytes: normal development of T cells and B cells, impaired production of IgA and IgG1 and reduced proliferative responses. J Exp Med 185((5):953–961. doi:[10.1084/jem.185.5.953](http://dx.doi.org/10.1084/jem.185.5.953)
- 19. Ouaaz F, Arron J, Zheng Y, Choi Y, Beg AA (2002) Dendritic cell development and survival require distinct NFkappaB subunits. Immunity 16(2):257–270. doi[:10.1016/](http://dx.doi.org/10.1016/S1074-7613(02)00272-8) [S1074-7613\(02\)00272-8](http://dx.doi.org/10.1016/S1074-7613(02)00272-8)
- 20. Weih F, Durham SK, Barton DS, Sha WC, Baltimore D, Bravo R (1997) p50-NF-kappaB complexes partially compensate for the absence of RelB: severely increased pathology in p50(−/−)relB(−/−) double-knockout mice. J Exp Med 185(7):1359–1370. doi[:10.1084/jem.185.7.1359](http://dx.doi.org/10.1084/jem.185.7.1359)
- 21. Weih F, Warr G, Yang H, Bravo R (1997) Multifocal defects in immune responses in RelB-deficient mice. J Immunol 158(11):5211–5218
- 22. Caamano J, Alexander J, Craig L, Bravo R, Hunter CA (1999) The NF-kappa B family member RelB is required for innate and adaptive immunity to *Toxoplasma gondii*. J Immunol 163(8):4453–4461
- 23. Burkly L, Hession C, Ogata L, Reilly C, Marconi LA, Olson D, Tizard R, Cate R, Lo D (1995) Expression of relB is required for the development of thymic medulla and dendritic cells. Nature 373(6514):531–536. doi:[10.1038/373531a0](http://dx.doi.org/10.1038/373531a0)
- 24. Weih F, Carrasco D, Durham SK, Barton DS, Rizzo CA, Ryseck RP, Lira SA, Bravo R (1995) Multiorgan inflammation and hematopoietic abnormalities in mice with a targeted disruption of RelB, a member of the NF-kappa B/Rel family. Cell 80(2):331–340. doi:[10.1016/0092-8674\(95\)90416-6](http://dx.doi.org/10.1016/0092-8674(95)90416-6)
- 25. Wu L, D'Amico A, Winkel KD, Suter M, Lo D, Shortman K (1998) RelB is essential for the development of myeloid-related CD8alpha-dendritic cells but not of lymphoid-related CD8alpha+ dendritic cells. Immunity 9(6):839–847. doi[:10.1016/](http://dx.doi.org/10.1016/S1074-7613(00)80649-4) [S1074-7613\(00\)80649-4](http://dx.doi.org/10.1016/S1074-7613(00)80649-4)
- 26. Grigoriadis G, Vasanthakumar A, Banerjee A, Grumont R, Overall S, Gleeson P, Shannon F, Gerondakis S (2011) c-Rel controls multiple discrete steps in the thymic development of Foxp3+ CD4 regulatory T cells. PLoS One 6(10):e26851. doi:[10.1371/journal.pone.0026851](http://dx.doi.org/10.1371/journal.pone.0026851)
- 27. Grossmann M, Metcalf D, Merryfull J, Beg A, Baltimore D, Gerondakis S (1999) The combined absence of the transcription factors Rel and RelA leads to multiple hemopoietic cell defects. Proc Natl Acad Sci USA 96(21):11848–11853. doi[:10.1073/p](http://dx.doi.org/10.1073/pnas.96.21.11848) [nas.96.21.11848](http://dx.doi.org/10.1073/pnas.96.21.11848)
- 28. Grossmann M, O'Reilly LA, Gugasyan R, Strasser A, Adams JM, Gerondakis S (2000) The anti-apoptotic activities of Rel and RelA required during B-cell maturation involve the regulation of Bcl-2 expression. EMBO J 19(23):6351–6360. doi[:10.10](http://dx.doi.org/10.1093/emboj/19.23.6351) [93/emboj/19.23.6351](http://dx.doi.org/10.1093/emboj/19.23.6351)
- 29. Grumont RJ, Rourke IJ, O'Reilly LA, Strasser A, Miyake K, Sha W, Gerondakis S (1998) B lymphocytes differentially use the Rel and nuclear factor kappaB1 (NF-kappaB1) transcription factors to regulate cell cycle progression and apoptosis in quiescent and mitogen-activated cells. J Exp Med 187(5):663–674. doi:[10.1084/jem.187.5.663](http://dx.doi.org/10.1084/jem.187.5.663)
- 30. Sha WC, Liou HC, Tuomanen EI, Baltimore D (1995) Targeted disruption of the p50 subunit of NF-kappa B leads to multifocal defects in immune responses. Cell 80(2):321–330. doi:[10.1016/0092-8674\(95\)90415-8](http://dx.doi.org/10.1016/0092-8674(95)90415-8)
- 31. Franzoso G, Carlson L, Xing L, Poljak L, Shores EW, Brown KD, Leonardi A, Tran T, Boyce BF, Siebenlist U (1997) Requirement for NF-kappaB in osteoclast and B-cell development. Genes Dev 11(24):3482–3496. doi[:10.1101/](http://dx.doi.org/10.1101/gad.11.24.3482) [gad.11.24.3482](http://dx.doi.org/10.1101/gad.11.24.3482)
- 32. Beg AA, Sha WC, Bronson RT, Baltimore D (1995) Constitutive NF-kappa B activation, enhanced granulopoiesis, and neonatal lethality in I kappa B alpha-deficient mice. Genes Dev 9(22):2736–2746. doi[:10.1101/gad.9.22.2736](http://dx.doi.org/10.1101/gad.9.22.2736)
- 33. Klement JF, Rice NR, Car BD, Abbondanzo SJ, Powers GD, Bhatt PH, Chen CH, Rosen CA, Stewart CL (1996) Ikappa-Balpha deficiency results in a sustained NF-kappaB response and severe widespread dermatitis in mice. Mol Cell Biol 16(5):2341–2349
- 34. Li ZW, Chu W, Hu Y, Delhase M, Deerinck T, Ellisman M, Johnson R, Karin M (1999) The IKKbeta subunit of IkappaB kinase (IKK) is essential for nuclear factor kappaB activation and prevention of apoptosis. J Exp Med 189(11):1839–1845. doi:[10.1084/jem.189.11.1839](http://dx.doi.org/10.1084/jem.189.11.1839)
- 35. Tanaka M, Fuentes ME, Yamaguchi K, Durnin MH, Dalrymple SA, Hardy KL, Goeddel DV (1999) Embryonic lethality, liver degeneration, and impaired NF-kappa B activation in IKK-betadeficient mice. Immunity 10(4):421–429. doi[:10.1016/](http://dx.doi.org/10.1016/S1074-7613(00)80042-4) [S1074-7613\(00\)80042-4](http://dx.doi.org/10.1016/S1074-7613(00)80042-4)
- 36. Yamada T, Mitani T, Yorita K, Uchida D, Matsushima A, Iwamasa K, Fujita S, Matsumoto M (2000) Abnormal immune function of hemopoietic cells from alymphoplasia (aly) mice, a natural strain with mutant NF-kappa B-inducing kinase. J Immunol 165(2):804–812
- 37. Hu Y, Baud V, Delhase M, Zhang P, Deerinck T, Ellisman M, Johnson R, Karin M (1999) Abnormal morphogenesis but intact IKK activation in mice lacking the IKKalpha subunit of IkappaB kinase. Science 284(5412):316–320. doi[:10.1126/](http://dx.doi.org/10.1126/science.284.5412.316) [science.284.5412.316](http://dx.doi.org/10.1126/science.284.5412.316)
- 38. Takeda K, Takeuchi O, Tsujimura T, Itami S, Adachi O, Kawai T, Sanjo H, Yoshikawa K, Terada N, Akira S (1999) Limb and skin abnormalities in mice lacking IKKalpha. Science 284(5412):313–316. doi:[10.1126/science.284.5412.313](http://dx.doi.org/10.1126/science.284.5412.313)
- 39. Senftleben U, Cao Y, Xiao G, Greten FR, Krahn G, Bonizzi G, Chen Y, Hu Y, Fong A, Sun SC, Karin M (2001) Activation by IKKalpha of a second, evolutionary conserved. NF-kappa B signaling pathway. Science 293(5534):1495–1499. doi[:10.1126/](http://dx.doi.org/10.1126/science.1062677) [science.1062677](http://dx.doi.org/10.1126/science.1062677)
- 40. Dejardin E, Droin NM, Delhase M, Haas E, Cao Y, Makris C, Li ZW, Karin M, Ware CF, Green DR (2002) The lymphotoxin-beta receptor induces different patterns of gene expression via two NF-kappaB pathways. Immunity 17(4):525–535. doi:[10.1016/S1074-7613\(02\)00423-5](http://dx.doi.org/10.1016/S1074-7613(02)00423-5)
- 41. Bonizzi G, Bebien M, Otero DC, Johnson-Vroom KE, Cao Y, Vu D, Jegga AG, Aronow BJ, Ghosh G, Rickert RC, Karin M (2004) Activation of IKKalpha target genes depends on recognition of specific kappaB binding sites by RelB:p52 dimers. EMBO J 23(21):4202–4210. doi[:10.1038/sj.emboj.7600391](http://dx.doi.org/10.1038/sj.emboj.7600391)
- 42. Gasparini C, Feldmann M (2012) NF-kappaB as a target for modulating inflammatory responses. Curr Pharm Des 18(35):5735–5745. doi[:10.2174/138161212803530763](http://dx.doi.org/10.2174/138161212803530763)
- 43. Pomerantz JL, Baltimore D (2002) Two pathways to NF-kappaB. Mol Cell 10(4):693–695. doi:[10.1016/S1097-2765\(02\)00697-4](http://dx.doi.org/10.1016/S1097-2765(02)00697-4)
- 44. Bonizzi G, Karin M (2004) The two NF-kappaB activation pathways and their role in innate and adaptive immunity. Trends Immunol 25(6):280–288. doi:[10.1016/j.it.2004.03.008](http://dx.doi.org/10.1016/j.it.2004.03.008)
- 45. Buchan SL, Al-Shamkhani A (2012) Distinct motifs in the intracellular domain of human CD30 differentially activate canonical and alternative transcription factor NF-kappaB signaling. PLoS One 7(9):e45244. doi:[10.1371/journal.pone.0045244](http://dx.doi.org/10.1371/journal.pone.0045244)
- 46. Theill LE, Boyle WJ, Penninger JM (2002) RANK-L and RANK: T cells, bone loss, and mammalian evolution. Annu Rev Immunol 20:795–823. doi:[10.1146/annurev.immunol.20.10030](http://dx.doi.org/10.1146/annurev.immunol.20.100301.064753) [1.064753](http://dx.doi.org/10.1146/annurev.immunol.20.100301.064753)
- 47. Rauert H, Wicovsky A, Muller N, Siegmund D, Spindler V, Waschke J, Kneitz C, Wajant H (2010) Membrane tumor

necrosis factor (TNF) induces p100 processing via TNF receptor-2 (TNFR2). J Biol Chem 285(10):7394–7404. doi:[10.1074/jbc.M109.037341](http://dx.doi.org/10.1074/jbc.M109.037341)

- 48. Young RM, Staudt LM (2013) Targeting pathological B cell receptor signalling in lymphoid malignancies. Nat Rev Drug Discov 12(3):229–243. doi:[10.1038/nrd3937](http://dx.doi.org/10.1038/nrd3937)
- 49. Weil R, Israel A (2006) Deciphering the pathway from the TCR to NF-kappaB. Cell Death Differ 13(5):826–833. doi:[10.1038/sj](http://dx.doi.org/10.1038/sj.cdd.4401856) [.cdd.4401856](http://dx.doi.org/10.1038/sj.cdd.4401856)
- 50. Sun SC (2011) Non-canonical NF-kappaB signaling pathway. Cell Res 21(1):71–85. doi[:10.1038/cr.2010.177](http://dx.doi.org/10.1038/cr.2010.177)
- 51. Gerondakis S, Banerjee A, Grigoriadis G, Vasanthakumar A, Gugasyan R, Sidwell T, Grumont RJ (2012) NF-kappaB subunit specificity in hemopoiesis. Immunol Rev 246(1):272–285. doi:[10.1111/j.1600-065X.2011.01090.x](http://dx.doi.org/10.1111/j.1600-065X.2011.01090.x)
- 52. Bottero V, Withoff S, Verma IM (2006) NF-kappaB and the regulation of hematopoiesis. Cell Death Differ 13(5):785–797. doi: [10.1038/sj.cdd.4401888](http://dx.doi.org/10.1038/sj.cdd.4401888)
- 53. Jimi E, Phillips RJ, Rincon M, Voll R, Karasuyama H, Flavell R, Ghosh S (2005) Activation of NF-kappaB promotes the transition of large, CD43+ pre-B cells to small, CD43-pre-B cells. Int Immunol 17(6):815–825. doi:[10.1093/intimm/dxh263](http://dx.doi.org/10.1093/intimm/dxh263)
- 54. Derudder E, Cadera EJ, Vahl JC, Wang J, Fox CJ, Zha S, van Loo G, Pasparakis M, Schlissel MS, Schmidt-Supprian M, Rajewsky K (2009) Development of immunoglobulin lambdachain-positive B cells, but not editing of immunoglobulin kappa-chain, depends on NF-kappaB signals. Nat Immunol 10(6):647–654. doi:[10.1038/ni.1732](http://dx.doi.org/10.1038/ni.1732)
- 55. Siebenlist U, Brown K, Claudio E (2005) Control of lymphocyte development by nuclear factor-kappaB. Nat Rev Immunol 5(6):435–445. doi[:10.1038/nri1629](http://dx.doi.org/10.1038/nri1629)
- 56. Kaileh M, Sen R (2012) NF-kappaB function in B lymphocytes. Immunol Rev 246(1):254–271. doi:[10.1111/j.1600-065X.2012.01106.x](http://dx.doi.org/10.1111/j.1600-065X.2012.01106.x)
- 57. Castro I, Wright JA, Damdinsuren B, Hoek KL, Carlesso G, Shinners NP, Gerstein RM, Woodland RT, Sen R, Khan WN (2009) B cell receptor-mediated sustained c-Rel activation facilitates late transitional B cell survival through control of B cell activating factor receptor and NF-kappaB2. J Immunol 182(12):7729–7737. doi:[10.4049/jimmunol.0803281](http://dx.doi.org/10.4049/jimmunol.0803281)
- 58. Smith SH, Cancro MP (2003) Cutting edge: B cell receptor signals regulate BLyS receptor levels in mature B cells and their immediate progenitors. J Immunol 170(12):5820–5823
- 59. Pillai S, Cariappa A (2009) The follicular versus marginal zone B lymphocyte cell fate decision. Nat Rev Immunol 9(11):767– 777. doi:[10.1038/nri2656](http://dx.doi.org/10.1038/nri2656)
- 60. Victora GD, Nussenzweig MC (2012) Germinal centers. Annu Rev Immunol 30:429–457. doi[:10.1146/](http://dx.doi.org/10.1146/annurev-immunol-020711-075032) [annurev-immunol-020711-075032](http://dx.doi.org/10.1146/annurev-immunol-020711-075032)
- 61. Mora AL, Stanley S, Armistead W, Chan AC, Boothby M (2001) Inefficient ZAP-70 phosphorylation and decreased thymic selection in vivo result from inhibition of NF-kappaB/Rel. J Immunol 167(10):5628–5635
- 62. Oh H, Ghosh S (2013) NF-kappaB: roles and regulation in different $CD4(+)$ T-cell subsets. Immunol Rev $252(1):41-51$. doi:[10.1111/imr.12033](http://dx.doi.org/10.1111/imr.12033)
- 63. Long M, Park SG, Strickland I, Hayden MS, Ghosh S (2009) Nuclear factor-kappaB modulates regulatory T cell development by directly regulating expression of Foxp3 transcription factor. Immunity 31(6):921–931. doi[:10.1016/j.immuni.2009.09.022](http://dx.doi.org/10.1016/j.immuni.2009.09.022)
- 64. Molinero LL, Cubre A, Mora-Solano C, Wang Y, Alegre ML (2012) T cell receptor/CARMA1/NF-kappaB signaling controls T-helper (Th) 17 differentiation. Proc Natl Acad Sci USA 109(45):18529–18534. doi[:10.1073/pnas.1204557109](http://dx.doi.org/10.1073/pnas.1204557109)
- 65. Yoshimura S, Bondeson J, Foxwell BM, Brennan FM, Feldmann M (2001) Effective antigen presentation by dendritic cells

is NF-kappaB dependent: coordinate regulation of MHC, costimulatory molecules and cytokines. Int Immunol 13(5):675– 683. doi:[10.1093/intimm/13.5.675](http://dx.doi.org/10.1093/intimm/13.5.675)

- 66. Gasparini C, Foxwell BM, Feldmann M (2009) RelB/p50 regulates CCL19 production, but fails to promote human DC maturation. Eur J Immunol 39(8):2215–2223. doi[:10.1002/](http://dx.doi.org/10.1002/eji.200939209) [eji.200939209](http://dx.doi.org/10.1002/eji.200939209)
- 67. Heel K, Tabone T, Rohrig KJ, Maslen PG, Meehan K, Grimwade LF, Erber WN (2013) Developments in the immunophenotypic analysis of haematological malignancies. Blood Rev 27(4):193–207. doi:[10.1016/j.blre.2013.06.005](http://dx.doi.org/10.1016/j.blre.2013.06.005)
- 68. Rodriguez-Abreu D, Bordoni A, Zucca E (2007) Epidemiology of hematological malignancies. Ann Oncol 18(Suppl 1):i3–i8. doi:[10.1093/annonc/mdl443](http://dx.doi.org/10.1093/annonc/mdl443)
- 69. Campo E, Swerdlow SH, Harris NL, Pileri S, Stein H, Jaffe ES (2011) The 2008 WHO classification of lymphoid neoplasms and beyond: evolving concepts and practical applications. Blood 117(19):5019–5032. doi[:10.1182/blood-2011-01-293050](http://dx.doi.org/10.1182/blood-2011-01-293050)
- 70. Braggio E, Egan JB, Fonseca R, Stewart AK (2013) Lessons from next-generation sequencing analysis in hematological malignancies. Blood Cancer J 3:e127. doi:[10.1038/bcj.2013.26](http://dx.doi.org/10.1038/bcj.2013.26)
- 71. Rampal R, Levine RL (2013) Leveraging cancer genome information in hematologic malignancies. J Clin Oncol 31(15):1885–1892. doi[:10.1200/JCO.2013.48.7447](http://dx.doi.org/10.1200/JCO.2013.48.7447)
- 72. Brown CM, Larsen SR, Iland HJ, Joshua DE, Gibson J (2012) Leukaemias into the 21st century: part 1: the acute leukaemias. Intern Med J 42(11):1179–1186. doi:[10.1111/j.1445-5994.2012.02938.x](http://dx.doi.org/10.1111/j.1445-5994.2012.02938.x)
- 73. Gibson J, Iland HJ, Larsen SR, Brown CM, Joshua DE (2013) Leukaemias into the 21st century. Part 2: the chronic leukaemias. Intern Med J 43(5):484–494. doi[:10.1111/imj.12135](http://dx.doi.org/10.1111/imj.12135)
- 74. Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, Boldrick JC, Sabet H, Tran T, Yu X, Powell JI, Yang L, Marti GE, Moore T, Hudson J Jr, Lu L, Lewis DB, Tibshirani R, Sherlock G, Chan WC, Greiner TC, Weisenburger DD, Armitage JO, Warnke R, Levy R, Wilson W, Grever MR, Byrd JC, Botstein D, Brown PO, Staudt LM (2000) Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. Nature 403(6769):503–511. doi:[10.1038/35000501](http://dx.doi.org/10.1038/35000501)
- 75. Rajkumar SV (2009) Multiple myeloma. Curr Probl Cancer 33(1):7–64. doi:[10.1016/j.currproblcancer.2009.01.001](http://dx.doi.org/10.1016/j.currproblcancer.2009.01.001)
- 76. Raab MS, Podar K, Breitkreutz I, Richardson PG, Anderson KC (2009) Multiple myeloma. Lancet 374(9686):324–339. doi:[10.1016/S0140-6736\(09\)60221-X](http://dx.doi.org/10.1016/S0140-6736(09)60221-X)
- 77. Jaffe ES (2009) The 2008 WHO classification of lymphomas: implications for clinical practice and translational research. Hematology (Am Soc Hematol Educ Program):523–531. doi:[10](http://dx.doi.org/10.1182/asheducation-2009.1.523) [.1182/asheducation-2009.1.523](http://dx.doi.org/10.1182/asheducation-2009.1.523)
- 78. Shaffer AL 3rd, Young RM, Staudt LM (2012) Pathogenesis of human B cell lymphomas. Annu Rev Immunol 30:565–610. doi:[10.1146/annurev-immunol-020711-075027](http://dx.doi.org/10.1146/annurev-immunol-020711-075027)
- 79. Nogai H, Dorken B, Lenz G (2011) Pathogenesis of non-Hodgkin's lymphoma. J Clin Oncol 29(14):1803–1811. doi[:10.1200/](http://dx.doi.org/10.1200/JCO.2010.33.3252) [JCO.2010.33.3252](http://dx.doi.org/10.1200/JCO.2010.33.3252)
- 80. Rui L, Schmitz R, Ceribelli M, Staudt LM (2011) Malignant pirates of the immune system. Nat Immunol 12(10):933–940. doi:[10.1038/ni.2094](http://dx.doi.org/10.1038/ni.2094)
- 81. Jost PJ, Ruland J (2007) Aberrant NF-kappaB signaling in lymphoma: mechanisms, consequences, and therapeutic implications. Blood 109(7):2700–2707. doi[:10.1182/](http://dx.doi.org/10.1182/blood-2006-07-025809) [blood-2006-07-025809](http://dx.doi.org/10.1182/blood-2006-07-025809)
- 82. Li ZW, Chen H, Campbell RA, Bonavida B, Berenson JR (2008) NF-kappaB in the pathogenesis and treatment of multiple myeloma. Curr Opin Hematol 15(4):391–399. doi[:10.1097/](http://dx.doi.org/10.1097/MOH.0b013e328302c7f4) [MOH.0b013e328302c7f4](http://dx.doi.org/10.1097/MOH.0b013e328302c7f4)
- 83. Braun T, Carvalho G, Coquelle A, Vozenin MC, Lepelley P, Hirsch F, Kiladjian JJ, Ribrag V, Fenaux P, Kroemer G (2006) NF-kappaB constitutes a potential therapeutic target in highrisk myelodysplastic syndrome. Blood 107(3):1156–1165. doi:[10.1182/blood-2005-05-1989](http://dx.doi.org/10.1182/blood-2005-05-1989)
- 84. Guzman ML, Neering SJ, Upchurch D, Grimes B, Howard DS, Rizzieri DA, Luger SM, Jordan CT (2001) Nuclear factor-kappaB is constitutively activated in primitive human acute myelogenous leukemia cells. Blood 98(8):2301–2307. doi[:10.1182/](http://dx.doi.org/10.1182/blood.V98.8.2301) [blood.V98.8.2301](http://dx.doi.org/10.1182/blood.V98.8.2301)
- 85. Shanmugam R, Gade P, Wilson-Weekes A, Sayar H, Suvannasankha A, Goswami C, Li L, Gupta S, Cardoso AA, Al Baghdadi T, Sargent KJ, Cripe LD, Kalvakolanu DV, Boswell HS (2012) A noncanonical Flt3ITD/NF-kappaB signaling pathway represses DAPK1 in acute myeloid leukemia. Clin Cancer Res 18(2):360–369. doi:[10.1158/1078-0432.CCR-10-3022](http://dx.doi.org/10.1158/1078-0432.CCR-10-3022)
- 86. Suhasini M, Reddy CD, Reddy EP, DiDonato JA, Pilz RB (1997) cAMP-induced NF-kappaB (p50/relB) binding to a c-myb intronic enhancer correlates with c-myb up-regulation and inhibition of erythroleukemia cell differentiation. Oncogene 15(15):1859–1870. doi[:10.1038/sj.onc.1201530](http://dx.doi.org/10.1038/sj.onc.1201530)
- 87. Suhasini M, Pilz RB (1999) Transcriptional elongation of c-myb is regulated by NF-kappaB (p50/RelB). Oncogene 18(51):7360–7369. doi[:10.1038/sj.onc.1203158](http://dx.doi.org/10.1038/sj.onc.1203158)
- 88. Kirchner D, Duyster J, Ottmann O, Schmid RM, Bergmann L, Munzert G (2003) Mechanisms of Bcr-Abl-mediated NF-kappaB/Rel activation. Exp Hematol 31(6):504–511. doi:[10.1016/S0301-472X\(03\)00069-9](http://dx.doi.org/10.1016/S0301-472X(03)00069-9)
- 89. Kordes U, Krappmann D, Heissmeyer V, Ludwig WD, Scheidereit C (2000) Transcription factor NF-kappaB is constitutively activated in acute lymphoblastic leukemia cells. Leukemia 14(3):399–402. doi:[10.1038/sj.leu.2401705](http://dx.doi.org/10.1038/sj.leu.2401705)
- 90. Vilimas T, Mascarenhas J, Palomero T, Mandal M, Buonamici S, Meng F, Thompson B, Spaulding C, Macaroun S, Alegre ML, Kee BL, Ferrando A, Miele L, Aifantis I (2007) Targeting the NF-kappaB signaling pathway in Notch1-induced T-cell leukemia. Nat Med 13(1):70–77. doi:[10.1038/nm1524](http://dx.doi.org/10.1038/nm1524)
- 91. Liu Z, Hazan-Halevy I, Harris DM, Li P, Ferrajoli A, Faderl S, Keating MJ, Estrov Z (2011) STAT-3 activates NF-kappaB in chronic lymphocytic leukemia cells. Mol Cancer Res 9(4):507– 515. doi:[10.1158/1541-7786.MCR-10-0559](http://dx.doi.org/10.1158/1541-7786.MCR-10-0559)
- 92. Xu J, Zhou P, Wang W, Sun A, Guo F (2013) RelB, together with RelA, sustains cell survival and confers proteasome inhibitor sensitivity of chronic lymphocytic leukemia cells from bone marrow. J Mol Med (Berl). doi:[10.1007/s00109-013-1081-6](http://dx.doi.org/10.1007/s00109-013-1081-6)
- 93. Mineva ND, Rothstein TL, Meyers JA, Lerner A, Sonenshein GE (2007) CD40 ligand-mediated activation of the de novo RelB NF-kappaB synthesis pathway in transformed B cells promotes rescue from apoptosis. J Biol Chem 282(24):17475– 17485. doi[:10.1074/jbc.M607313200](http://dx.doi.org/10.1074/jbc.M607313200)
- 94. Mori N, Fujii M, Ikeda S, Yamada Y, Tomonaga M, Ballard DW, Yamamoto N (1999) Constitutive activation of NF-kappaB in primary adult T-cell leukemia cells. Blood 93(7):2360–2368
- 95. Sun SC, Yamaoka S (2005) Activation of NF-kappaB by HTLV-I and implications for cell transformation. Oncogene 24(39):5952–5964. doi[:10.1038/sj.onc.1208969](http://dx.doi.org/10.1038/sj.onc.1208969)
- 96. Isogawa M, Higuchi M, Takahashi M, Oie M, Mori N, Tanaka Y, Aoyagi Y, Fujii M (2008) Rearranged NF-kappa B2 gene in an adult T-cell leukemia cell line. Cancer Sci 99(4):792–798. doi:[10.1111/j.1349-7006.2008.00750.x](http://dx.doi.org/10.1111/j.1349-7006.2008.00750.x)
- 97. Ohsugi T, Ishida T, Shimasaki T, Okada S, Umezawa K (2013) p53 dysfunction precedes the activation of nuclear factorkappaB during disease progression in mice expressing Tax, a human T-cell leukemia virus type 1 oncoprotein. Carcinogenesis 34(9):2129–2136. doi[:10.1093/carcin/bgt144](http://dx.doi.org/10.1093/carcin/bgt144)
- 98. Demchenko YN, Kuehl WM (2010) A critical role for the NFkB pathway in multiple myeloma. Oncotarget 1(1):59–68
- 99. Hailfinger S, Nogai H, Pelzer C, Jaworski M, Cabalzar K, Charton JE, Guzzardi M, Decaillet C, Grau M, Dorken B, Lenz P, Lenz G, Thome M (2011) Malt1-dependent RelB cleavage promotes canonical NF-kappaB activation in lymphocytes and lymphoma cell lines. Proc Natl Acad Sci USA 108(35):14596– 14601. doi[:10.1073/pnas.1105020108](http://dx.doi.org/10.1073/pnas.1105020108)
- 100. Calado DP, Zhang B, Srinivasan L, Sasaki Y, Seagal J, Unitt C, Rodig S, Kutok J, Tarakhovsky A, Schmidt-Supprian M, Rajewsky K (2010) Constitutive canonical NF-kappaB activation cooperates with disruption of BLIMP1 in the pathogenesis of activated B cell-like diffuse large cell lymphoma. Cancer Cell 18(6):580–589. doi:[10.1016/j.ccr.2010.11.024](http://dx.doi.org/10.1016/j.ccr.2010.11.024)
- 101. Ranuncolo SM, Pittaluga S, Evbuomwan MO, Jaffe ES, Lewis BA (2012) Hodgkin lymphoma requires stabilized NIK and constitutive RelB expression for survival. Blood 120(18):3756– 3763. doi[:10.1182/blood-2012-01-405951](http://dx.doi.org/10.1182/blood-2012-01-405951)
- 102. Nonaka M, Horie R, Itoh K, Watanabe T, Yamamoto N, Yamaoka S (2005) Aberrant NF-kappaB2/p52 expression in Hodgkin/ Reed–Sternberg cells and CD30-transformed rat fibroblasts. Oncogene 24(24):3976–3986. doi:[10.1038/sj.onc.1208564](http://dx.doi.org/10.1038/sj.onc.1208564)
- 103. Guo F, Sun A, Wang W, He J, Hou J, Zhou P, Chen Z (2009) TRAF1 is involved in the classical NF-kappaB activation and CD30-induced alternative activity in Hodgkin's lymphoma cells. Mol Immunol 46(13):2441–2448. doi:[10.1016/j.molimm.2009.05.178](http://dx.doi.org/10.1016/j.molimm.2009.05.178)
- 104. Schwarzer R, Dorken B, Jundt F (2012) Notch is an essential upstream regulator of NF-kappaB and is relevant for survival of Hodgkin and Reed–Sternberg cells. Leukemia 26(4):806–813. doi:[10.1038/leu.2011.265](http://dx.doi.org/10.1038/leu.2011.265)
- 105. Wright CW, Rumble JM, Duckett CS (2007) CD30 activates both the canonical and alternative NF-kappaB pathways in anaplastic large cell lymphoma cells. J Biol Chem 282(14):10252– 10262. doi[:10.1074/jbc.M608817200](http://dx.doi.org/10.1074/jbc.M608817200)
- 106. Mathas S, Johrens K, Joos S, Lietz A, Hummel F, Janz M, Jundt F, Anagnostopoulos I, Bommert K, Lichter P, Stein H, Scheidereit C, Dorken B (2005) Elevated NF-kappaB p50 complex formation and Bcl-3 expression in classical Hodgkin, anaplastic large-cell, and other peripheral T-cell lymphomas. Blood 106(13):4287–4293. doi:[10.1182/blood-2004-09-3620](http://dx.doi.org/10.1182/blood-2004-09-3620)
- 107. Klapproth K, Sander S, Marinkovic D, Baumann B, Wirth T (2009) The IKK2/NF-(kappa)B pathway suppresses MYCinduced lymphomagenesis. Blood 114(12):2448–2458. doi:[10.1182/blood-2008-09-181008](http://dx.doi.org/10.1182/blood-2008-09-181008)
- 108. Annunziata CM, Davis RE, Demchenko Y, Bellamy W, Gabrea A, Zhan F, Lenz G, Hanamura I, Wright G, Xiao W, Dave S, Hurt EM, Tan B, Zhao H, Stephens O, Santra M, Williams DR, Dang L, Barlogie B, Shaughnessy JD Jr, Kuehl WM, Staudt LM (2007) Frequent engagement of the classical and alternative NF-kappaB pathways by diverse genetic abnormalities in multiple myeloma. Cancer Cell 12(2):115–130. doi:[10.1016/j.ccr.2007.07.004](http://dx.doi.org/10.1016/j.ccr.2007.07.004)
- 109. Keats JJ, Fonseca R, Chesi M, Schop R, Baker A, Chng WJ, Van Wier S, Tiedemann R, Shi CX, Sebag M, Braggio E, Henry T, Zhu YX, Fogle H, Price-Troska T, Ahmann G, Mancini C, Brents LA, Kumar S, Greipp P, Dispenzieri A, Bryant B, Mulligan G, Bruhn L, Barrett M, Valdez R, Trent J, Stewart AK, Carpten J, Bergsagel PL (2007) Promiscuous mutations activate the noncanonical NF-kappaB pathway in multiple myeloma. Cancer Cell 12(2):131–144. doi[:10.1016/j.ccr.2007.07.003](http://dx.doi.org/10.1016/j.ccr.2007.07.003)
- 110. Demchenko YN, Glebov OK, Zingone A, Keats JJ, Bergsagel PL, Kuehl WM (2010) Classical and/or alternative NF-kappaB pathway activation in multiple myeloma. Blood 115(17):3541– 3552. doi:[10.1182/blood-2009-09-243535](http://dx.doi.org/10.1182/blood-2009-09-243535)
- 111. Cormier F, Monjanel H, Fabre C, Billot K, Sapharikas E, Chereau F, Bordereaux D, Molina TJ, Avet-Loiseau H, Baud V (2013) Frequent engagement of RelB activation is critical for cell survival in multiple myeloma. PLoS ONE 8(3):e59127. doi:[10.1371/journal.pone.0059127](http://dx.doi.org/10.1371/journal.pone.0059127)
- 112. Landowski TH, Olashaw NE, Agrawal D, Dalton WS (2003) Cell adhesion-mediated drug resistance (CAM-DR) is associated with activation of NF-kappa B (RelB/p50) in myeloma cells. Oncogene 22(16):2417–2421. doi[:10.1038/sj.](http://dx.doi.org/10.1038/sj.onc.1206315) [onc.1206315](http://dx.doi.org/10.1038/sj.onc.1206315)
- 113. Dave SS, Wright G, Tan B, Rosenwald A, Gascoyne RD, Chan WC, Fisher RI, Braziel RM, Rimsza LM, Grogan TM, Miller TP, LeBlanc M, Greiner TC, Weisenburger DD, Lynch JC, Vose J, Armitage JO, Smeland EB, Kvaloy S, Holte H, Delabie J, Connors JM, Lansdorp PM, Ouyang Q, Lister TA, Davies AJ, Norton AJ, Muller-Hermelink HK, Ott G, Campo E, Montserrat E, Wilson WH, Jaffe ES, Simon R, Yang L, Powell J, Zhao H, Goldschmidt N, Chiorazzi M, Staudt LM (2004) Prediction of survival in follicular lymphoma based on molecular features of tumor-infiltrating immune cells. N Engl J Med 351(21):2159– 2169. doi[:10.1056/NEJMoa041869](http://dx.doi.org/10.1056/NEJMoa041869)
- 114. dos Santos NR, Williame M, Gachet S, Cormier F, Janin A, Weih D, Weih F, Ghysdael J (2008) RelB-dependent stromal cells promote T-cell leukemogenesis. PLoS One 3(7):e2555. doi:[10.1371/journal.pone.0002555](http://dx.doi.org/10.1371/journal.pone.0002555)
- 115. Herishanu Y, Perez-Galan P, Liu D, Biancotto A, Pittaluga S, Vire B, Gibellini F, Njuguna N, Lee E, Stennett L, Raghavachari N, Liu P, McCoy JP, Raffeld M, Stetler-Stevenson M, Yuan C, Sherry R, Arthur DC, Maric I, White T, Marti GE, Munson P, Wilson WH, Wiestner A (2011) The lymph node microenvironment promotes B-cell receptor signaling, NF-kappaB activation, and tumor proliferation in chronic lymphocytic leukemia. Blood 117(2):563–574. doi:[10.1182/blood-2010-05-284984](http://dx.doi.org/10.1182/blood-2010-05-284984)
- 116. Marienfeld R, May MJ, Berberich I, Serfling E, Ghosh S, Neumann M (2003) RelB forms transcriptionally inactive complexes with RelA/p65. J Biol Chem 278(22):19852–19860. doi:[10.1074/jbc.M301945200](http://dx.doi.org/10.1074/jbc.M301945200)
- 117. Gasparini C, Foxwell BM, Feldmann M (2013) RelB/ p50 regulates TNF production in LPS-stimulated dendritic cells and macrophages. Cytokine 61(3):736–740. doi:[10.1016/j.cyto.2012.12.029](http://dx.doi.org/10.1016/j.cyto.2012.12.029)
- 118. Jacque E, Billot K, Authier H, Bordereaux D, Baud V (2012) RelB inhibits cell proliferation and tumor growth through p53 transcriptional activation. Oncogene. doi[:10.1038/onc.2012.282](http://dx.doi.org/10.1038/onc.2012.282)
- 119. Baldwin AS (2012) Regulation of cell death and autophagy by IKK and NF-kappaB: critical mechanisms in immune function and cancer. Immunol Rev 246(1):327–345. doi:[10.1111/j.1600-065X.2012.01095.x](http://dx.doi.org/10.1111/j.1600-065X.2012.01095.x)
- 120. Johnson RF, Perkins ND (2012) Nuclear factor-kappaB, p53, and mitochondria: regulation of cellular metabolism and the Warburg effect. Trends Biochem Sci 37(8):317–324. doi:[10.1016/j.tibs.2012.04.002](http://dx.doi.org/10.1016/j.tibs.2012.04.002)
- 121. Secchiero P, Bosco R, Celeghini C, Zauli G (2011) Recent advances in the therapeutic perspectives of Nutlin-3. Curr Pharm Des 17(6):569–577. doi[:10.2174/138161211795222586](http://dx.doi.org/10.2174/138161211795222586)
- 122. Tergaonkar V, Pando M, Vafa O, Wahl G, Verma I (2002) p53 stabilization is decreased upon NFkappaB activation: a role for NFkappaB in acquisition of resistance to chemotherapy. Cancer Cell 1(5):493–503. doi:[10.1016/S1535-6108\(02\)00068-5](http://dx.doi.org/10.1016/S1535-6108(02)00068-5)
- 123. Webster GA, Perkins ND (1999) Transcriptional cross talk between NF-kappaB and p53. Mol Cell Biol 19(5):3485–3495
- 124. Huang WC, Ju TK, Hung MC, Chen CC (2007) Phosphorylation of CBP by IKKalpha promotes cell growth by switching the binding preference of CBP from p53 to NF-kappaB. Mol Cell 26(1):75–87. doi[:10.1016/j.molcel.2007.02.019](http://dx.doi.org/10.1016/j.molcel.2007.02.019)
- 125. Fujioka S, Schmidt C, Sclabas GM, Li Z, Pelicano H, Peng B, Yao A, Niu J, Zhang W, Evans DB, Abbruzzese JL, Huang P, Chiao PJ (2004) Stabilization of p53 is a novel mechanism for proapoptotic function of NF-kappaB. J Biol Chem 279(26):27549–27559. doi[:10.1074/jbc.M313435200](http://dx.doi.org/10.1074/jbc.M313435200)
- 126. Shetty S, Graham BA, Brown JG, Hu X, Vegh-Yarema N, Harding G, Paul JT, Gibson SB (2005) Transcription factor NF-kappaB differentially regulates death receptor 5 expression involving histone deacetylase 1. Mol Cell Biol 25(13):5404–5416. doi :[10.1128/MCB.25.13.5404-](http://dx.doi.org/10.1128/MCB.25.13.5404-) 5416.2005
- 127. Jacque E, Billot K, Authier H, Bordereaux D, Baud V (2013) RelB inhibits cell proliferation and tumor growth through p53 transcriptional activation. Oncogene 32(21):2661–2669. doi:[10](http://dx.doi.org/10.1038/onc.2012.282) [.1038/onc.2012.282](http://dx.doi.org/10.1038/onc.2012.282)
- 128. Schumm K, Rocha S, Caamano J, Perkins ND (2006) Regulation of p53 tumour suppressor target gene expression by the p52 NF-kappaB subunit. EMBO J 25(20):4820–4832. doi:[10.1038/s](http://dx.doi.org/10.1038/sj.emboj.7601343) [j.emboj.7601343](http://dx.doi.org/10.1038/sj.emboj.7601343)
- 129. Kiaris H, Chatzistamou I, Trimis G, Frangou-Plemmenou M, Pafiti-Kondi A, Kalofoutis A (2005) Evidence for nonautonomous effect of p53 tumor suppressor in carcinogenesis. Cancer Res 65(5):1627–1630. doi:[10.1158/0008-5472.CAN-04-3791](http://dx.doi.org/10.1158/0008-5472.CAN-04-3791)
- 130. Moskovits N, Kalinkovich A, Bar J, Lapidot T, Oren M (2006) p53 attenuates cancer cell migration and invasion through repression of SDF-1/CXCL12 expression in stromal fibroblasts. Cancer Res 66(22):10671–10676. doi[:10.1158/0008-5472.](http://dx.doi.org/10.1158/0008-5472.CAN-06-2323) [CAN-06-2323](http://dx.doi.org/10.1158/0008-5472.CAN-06-2323)
- 131. Baud V, Karin M (2009) Is NF-kappaB a good target for cancer therapy? Hopes and pitfalls. Nat Rev Drug Discov 8(1):33–40. doi:[10.1038/nrd2781](http://dx.doi.org/10.1038/nrd2781)
- 132. Chu TC, Twu KY, Ellington AD, Levy M (2006) Aptamer mediated siRNA delivery. Nucleic Acids Res 34(10):e73. doi:[10.109](http://dx.doi.org/10.1093/nar/gkl388) [3/nar/gkl388](http://dx.doi.org/10.1093/nar/gkl388)
- 133. Gasparini C, Vecchi Brumatti L, Monasta L, Zauli G (2013) TRAIL-based therapeutic approaches for the treatment of pediatric malignancies. Curr Med Chem 20(17):2254–2271. doi:[10.2174/0929867311320170009](http://dx.doi.org/10.2174/0929867311320170009)
- 134. Secchiero P, Zauli G (2008) Tumor-necrosis-factor-related apoptosis-inducing ligand and the regulation of hematopoiesis. Curr Opin Hematol 15(1):42–48. doi[:10.1097/MOH.0b013e328](http://dx.doi.org/10.1097/MOH.0b013e3282f15fa6) [2f15fa6](http://dx.doi.org/10.1097/MOH.0b013e3282f15fa6)
- 135. Secchiero P, Melloni E, Corallini F, Beltrami AP, Alviano F, Milani D, D'Aurizio F, di Iasio MG, Cesselli D, Bagnara GP, Zauli G (2008) Tumor necrosis factor-related apoptosis-inducing ligand promotes migration of human bone marrow multipotent stromal cells. Stem Cells 26(11):2955–2963. doi[:10.1634/st](http://dx.doi.org/10.1634/stemcells.2008-0512) [emcells.2008-0512](http://dx.doi.org/10.1634/stemcells.2008-0512)
- 136. Secchiero P, di Iasio MG, Gonelli A, Zauli G (2008) The MDM2 inhibitor Nutlins as an innovative therapeutic tool for the treatment of haematological malignancies. Curr Pharm Des 14(21):2100–2110. doi[:10.2174/138161208785294663](http://dx.doi.org/10.2174/138161208785294663)
- 137. Secchiero P, Zerbinati C, Melloni E, Milani D, Campioni D, Fadda R, Tiribelli M, Zauli G (2007) The MDM-2 antagonist nutlin-3 promotes the maturation of acute myeloid leukemic blasts. Neoplasia 9(10):853–861. doi[:10.1593/neo.07523](http://dx.doi.org/10.1593/neo.07523)
- 138. Secchiero P, Melloni E, di Iasio MG, Tiribelli M, Rimondi E, Corallini F, Gattei V, Zauli G (2009) Nutlin-3 up-regulates the expression of Notch1 in both myeloid and lymphoid leukemic cells, as part of a negative feedback antiapoptotic mechanism. Blood 113(18):4300–4308. doi[:10.1182/blood-2008-11-187708](http://dx.doi.org/10.1182/blood-2008-11-187708)
- 139. Secchiero P, Corallini F, Rimondi E, Chiaruttini C, di Iasio MG, Rustighi A, Del Sal G, Zauli G (2008) Activation of the p53 pathway down-regulates the osteoprotegerin expression and release by vascular endothelial cells. Blood 111(3):1287–1294. doi:[10.1182/blood-2007-05-092031](http://dx.doi.org/10.1182/blood-2007-05-092031)
- 140. Zauli G, Melloni E, Capitani S, Secchiero P (2009) Role of fulllength osteoprotegerin in tumor cell biology. Cell Mol Life Sci 66(5):841–851. doi:[10.1007/s00018-008-8536-x](http://dx.doi.org/10.1007/s00018-008-8536-x)
- 141. Gasparini C, Tommasini A, Zauli G (2012) The MDM2 inhibitor Nutlin-3 modulates dendritic cell-induced T cell proliferation. Hum Immunol 73(4):342–345. doi:[10.1016/j.humimm.2012.01.018](http://dx.doi.org/10.1016/j.humimm.2012.01.018)
- 142. Travert M, Ame-Thomas P, Pangault C, Morizot A, Micheau O, Semana G, Lamy T, Fest T, Tarte K, Guillaudeux T (2008) CD40 ligand protects from TRAIL-induced apoptosis in follicular lymphomas through NF-kappaB activation and up-regulation of c-FLIP and Bcl-xL. J Immunol 181(2):1001–1011
- 143. Rosati E, Sabatini R, Rampino G, Tabilio A, Di Ianni M, Fettucciari K, Bartoli A, Coaccioli S, Screpanti I, Marconi P (2009) Constitutively activated Notch signaling is involved in survival and apoptosis resistance of B-CLL cells. Blood 113(4):856– 865. doi:[10.1182/blood-2008-02-139725](http://dx.doi.org/10.1182/blood-2008-02-139725)
- 144. Hertlein E, Byrd JC (2010) Signalling to drug resistance in CLL. Best Prac Res Clin Haematol 23(1):121–131. doi:[10.1016/j.beha.2010.01.007](http://dx.doi.org/10.1016/j.beha.2010.01.007)
- 145. Romano MF, Lamberti A, Tassone P, Alfinito F, Costantini S, Chiurazzi F, Defrance T, Bonelli P, Tuccillo F, Turco MC, Venuta S (1998) Triggering of CD40 antigen inhibits fludarabine-induced apoptosis in B chronic lymphocytic leukemia cells. Blood 92(3):990–995
- 146. Perez LE, Parquet N, Meads M, Anasetti C, Dalton W (2010) Bortezomib restores stroma-mediated APO2L/TRAIL apoptosis resistance in multiple myeloma. Eur J Haematol 84(3):212–222. doi:[10.1111/j.1600-0609.2009.01381.x](http://dx.doi.org/10.1111/j.1600-0609.2009.01381.x)
- 147. Jin L, Tabe Y, Kojima K, Zhou Y, Pittaluga S, Konopleva M, Miida T, Raffeld M (2010) MDM2 antagonist Nutlin-3 enhances bortezomib-mediated mitochondrial apoptosis in TP53-mutated mantle cell lymphoma. Cancer Lett 299(2):161– 170. doi:[10.1016/j.canlet.2010.08.015](http://dx.doi.org/10.1016/j.canlet.2010.08.015)
- 148. Saha MN, Jiang H, Jayakar J, Reece D, Branch DR, Chang H (2010) MDM2 antagonist nutlin plus proteasome inhibitor velcade combination displays a synergistic anti-myeloma activity. Cancer Biol Ther 9(11):936–944. doi:[10.4161/cbt.9.11.11882](http://dx.doi.org/10.4161/cbt.9.11.11882)
- 149. Ooi MG, Hayden PJ, Kotoula V, McMillin DW, Charalambous E, Daskalaki E, Raje NS, Munshi NC, Chauhan D, Hideshima T, Buon L, Clynes M, O'Gorman P, Richardson PG, Mitsiades CS, Anderson KC, Mitsiades N (2009) Interactions of the Hdm2/p53 and proteasome pathways may enhance the antitumor activity of bortezomib. Clin Cancer Res 15(23):7153–7160. doi:[10.1158/1078-0432.CCR-09-1071](http://dx.doi.org/10.1158/1078-0432.CCR-09-1071)
- 150. Conticello C, Adamo L, Vicari L, Giuffrida R, Iannolo G, Anastasi G, Caruso L, Moschetti G, Cupri A, Palumbo GA, Gulisano M, De Maria R, Giustolisi R, Di Raimondo F (2008) Antitumor activity of bortezomib alone and in combination with TRAIL in human acute myeloid leukemia. Acta Haematol 120(1):19–30. doi[:10.1159/000151511](http://dx.doi.org/10.1159/000151511)
- 151. Kabore AF, Sun J, Hu X, McCrea K, Johnston JB, Gibson SB (2006) The TRAIL apoptotic pathway mediates proteasome inhibitor induced apoptosis in primary chronic lymphocytic leukemia cells. Apoptosis 11(7):1175–1193. doi[:10.1007/](http://dx.doi.org/10.1007/s10495-006-8048-9) [s10495-006-8048-9](http://dx.doi.org/10.1007/s10495-006-8048-9)
- 152. Di Pietro R, Zauli G (2004) Emerging non-apoptotic functions of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)/Apo2L. J Cell Physiol 201(3):331–340. doi:[10.1002/jcp.20099](http://dx.doi.org/10.1002/jcp.20099)
- 153. Zauli G, Sancilio S, Cataldi A, Sabatini N, Bosco D, Di Pietro R (2005) PI-3K/Akt and NF-kappaB/IkappaBalpha pathways are activated in Jurkat T cells in response to TRAIL treatment. J Cell Physiol 202(3):900–911. doi:[10.1002/jcp.20202](http://dx.doi.org/10.1002/jcp.20202)