## REVIEW

# Subtypes of functional *a*1-adrenoceptor

James R. Docherty

Received: 24 July 2009/Revised: 11 September 2009/Accepted: 5 October 2009/Published online: 28 October 2009 © Birkhäuser Verlag, Basel/Switzerland 2009

**Abstract** In this review, subtypes of functional  $\alpha$ 1-adrenoceptor are discussed. These are cell membrane receptors, belonging to the seven-transmembrane-spanning G-protein-linked family of receptors, which respond to the physiological agonist noradrenaline. al-Adrenoceptors can be divided into  $\alpha$ 1A-,  $\alpha$ 1B- and  $\alpha$ 1D-adrenoceptors, all of which mediate contractile responses involving Gq/11 and inositol phosphate turnover. A fourth  $\alpha$ 1-adrenoceptor, the  $\alpha$ 1L-, represents a functional phenotype of the  $\alpha$ 1A-adrenoceptor. al-Adrenoceptor subtype knock-out mice have refined our knowledge of the functions of  $\alpha$ -adrenoceptor subtypes, particuarly as subtype-selective agonists and antagonists are not available for all subtypes. al-Adrenoceptors function as stimulatory receptors involved particularly in smooth muscle contraction, especially contraction of vascular smooth muscle, both in local vasoconstriction and in the control of blood pressure and temperature, and contraction of the prostate and bladder neck. Central actions are now being elucidated.

Keywords  $\alpha$ 1-Adrenoceptors  $\cdot \alpha$ 1A-Adrenoceptors  $\cdot \alpha$ 1B-Adrenoceptors  $\cdot \alpha$ 1D-Adrenoceptors  $\cdot$  Blood pressure  $\cdot$ Smooth muscle contraction  $\cdot$  Vascular smooth muscle  $\cdot$ Benign prostatic hypertrophy

# Introduction

Adrenoceptors, or adrenergic receptors, are cell membrane receptors belonging to the seven-transmembrane-spanning

J. R. Docherty (🖂)

G-protein-linked superfamily of receptors. They respond to the sympathetic neurotransmitter noradrenaline and to the hormone adrenaline (and to various exogenous agonists) by producing a response within the cell involving a second messenger or ion channel. Adrenoceptors are classically the receptors involved in the "fight or flight" reaction, the mobilisation of resources caused by activation of the sympathetic nervous system that prepares the body for bouts of severe activity. Sympathetic activation will cause α1-adrenoceptor-mediated vasoconstriction in less vital vascular beds, particularly splanchnic and skin (although the skin vasculature may dilate later to dissipate heat), to divert blood to skeletal muscle in exercise. Sympathetic activation also mobilises blood from the reservoir in the large veins (the capacitance vessels) by veniconstriction, again largely involving  $\alpha 1$ - (and  $\alpha 2$ -) adrenoceptors.

Historically, employing a series of agonists, Ahlquist [1] described two types of adrenoceptor based on the rank order of potency of these agonists. The receptor termed  $\alpha$ was mainly excitatory, except in the intestine, and the receptor termed  $\beta$  was mainly inhibitory, except in the heart. In Ahlquist's classification, *a*-adrenoceptors were receptors present on smooth muscle, i.e. postjunctional receptors. These were later classified as postjunctional α1-adrenoceptors, when evidence accumulated for prejunctional  $\alpha$ 2-adrenoceptors [2]. Later, when evidence accumulated for  $\alpha$ 2-adrenoceptors located postjunctionally, this purely anatomical classification was refined into a pharmacological subclassification, independent of location [3]. Further advances in our understanding of  $\alpha$ 1-adrenoceptors have come from the development of new pharmacological methodologies for the study of receptors. The first of these was the technique of the radioligand binding assay: al-adrenoceptors were initially subdivided into  $\alpha 1A$  and  $\alpha 1B$ -subtypes, based on the affinities of a

Department of Physiology, Royal College of Surgeons in Ireland, 123, St. Stephen's Green, Dublin 2, Ireland e-mail: docherty@rcsi.ie

series of ligands, especially WB 4101 and prazosin [4], and based on the ability of the alkylating agent chloroethylclonidine to inactivate the  $\alpha$ 1B but not the  $\alpha$ 1A subtype [5]. Under this classification, functional receptors mediating contractions of rat vas deferens were  $\alpha$ 1A-, and those of rat spleen were  $\alpha$ 1B-adrenoceptors [5] (see Fig. 1).

The study of  $\alpha$ -adrenoceptors was revolutionised by molecular biology: cloning techniques revealed initially four subtypes of  $\alpha$ 1-adrenoceptor. The  $\alpha$ 1b-adrenceptor subtype (the lower case subscript being used for recombinant receptors and upper case subscript for pharmacologically defined receptor subtypes) was the first to be cloned, from the hamster [6], and this clone expressed a protein with the radioligand binding properties of the  $\alpha$ 1B-adrenoceptor. Other clones were the rat  $\alpha$ 1a- [7], the bovine  $\alpha 1c$ - [8] and rat  $\alpha 1d$ -adrenoceptor [9]. However, the  $\alpha$ 1a and  $\alpha$ 1d clones showed 99.8% homogeneity and appeared to represent the same subtype. It is now clear that the  $\alpha la/\alpha ld$  clone represents a novel subtype of  $\alpha l$ -adrenoceptor ( $\alpha$ 1D), whereas the  $\alpha$ 1c is now identified with the  $\alpha$ 1A-ligand binding site. These clones have now been renamed to match the functional receptors:  $\alpha$ 1A (formerly  $\alpha$ 1c),  $\alpha$ 1B (formerly  $\alpha$ 1b) and  $\alpha$ 1D (formerly  $\alpha$ 1a/ $\alpha$ 1d) (see Fig. 1). Hence, three genes for  $\alpha$ 1-adrenoceptors have now been identified ( $\alpha$ 1A,  $\alpha$ 1B,  $\alpha$ 1D) [see 10–12].

Figure 1 shows how the subclassification of  $\alpha$ 1-adrenoceptors has developed since 1948. The  $\alpha$ 1L-adrenoceptor is dependent on the  $\alpha$ 1A-adrenoceptor gene and is a phenotype of the  $\alpha$ 1A-adrenoceptor (see below).  $\alpha$ 1-Adrenoceptors are predominantly linked to the G-protein Gq/11 and activation of phospholipase C (PLC) (see Table 1). No adrenoceptor belongs to the class of ionotropic receptors, those with an intrinsic ion channel, unlike the situation with another monoamine, 5-hydroxytryptamine.



Fig. 1 The historical development of the subclassification of  $\alpha$ 1-adrenoceptors. For details, see text

The object of this review is to look at functional subtypes of  $\alpha$ 1-adrenoceptors and their physiological roles.

# Function of *a*1-adrenoceptors

 $\alpha$ 1-Adrenoceptors function as stimulatory receptors and are the classical adrenoceptors mediating smooth muscle contraction, and in the vascular system have a major role in the control of blood pressure. A fall in blood pressure due to causes such as haemorrhage will activate the baroreceptor reflex and cause sympathetic activation to vasconstrict less vital vascular beds, especially splanchnic and skin.  $\alpha$ 1-Adrenoceptor antagonists lower blood pressure in hypertension, but are not widely employed. al-Adrenoceptor agonist-mediated vasoconstriction can be used to treat hypotension, and these agonists are widely used as over the counter nasal decongestants, acting by reducing blood flow to the nasal musosa. Pseudoephedrine, when used as a nasal decongestant, shows some selectivity for local over cardiovascular actions: it is reported to have nasal actions at 60 mg [13], no cardiovascular actions at up to 120 mg [14] or cardiovascular actions at 120–180 mg [15–17]. The reason for this selectivity is unclear, but pseudoephedrine may show some slight selectivity for  $\alpha$ 1A-adrenoceptors [18] and moderate potency as a beta-2 adrenoceptor agonist [19].

Ocular effects involve  $\alpha$ 1-adrenoceptor-mediated dilatation of the pupil by contracting the dilator pupillae muscle, increasing the amount of light reaching the retina.  $\alpha$ 1-Adrenoceptor agonists also have actions to reduce intraocular pressure, presumably by restricting blood flow. Other actions include bronchoconstriction, constriction of sphincters in the gastrointestinal tract and salivary secretions.  $\alpha$ 1-Adrenoceptors may be important in the regulation of human lipid metabolism [20] and in the uptake of glucose into adipocytes [21].

Genitourinary actions are also important, and  $\alpha$ 1-adrenoceptors are involved in contraction of the vas deferens and in contracting the neck of the bladder, and are involved in prostate function.  $\alpha$ 1-Adrenoceptors mediate inhibition of micturition by constriction of the bladder neck, and this may involve mainly  $\alpha$ 1D-adrenoceptors [22], but the density of alpha1-adrenoceptors in the neck of the bladder is greater in males, suggesting an additional sexual function to prevent retrograde ejaculation into the bladder.  $\alpha$ 1-Adrenoceptors mediate contraction of the vas deferens and seminal vesicles, and this has an important role in ejaculation.  $\alpha$ 1A-KO mice, lacking the  $\alpha$ 1A-adrenoceptor, have a 50% reduction in pregnancy rate, with further reduction with knockout of all three alpha1-adrenoceptors, and this is mainly due to decreased ejaculatory function because of diminished contractile response of the vas

Table 1 Summary of *a*1-adrenoceptor subtype characteristics

Receptor subtype	α1A	α1B	α1D
Functional responses	Control of blood pressure; vasoconstriction; smooth muscle contraction	Regulatory; minor contractile role	Control of blood pressure; vasoconstriction; smooth muscle contraction
Location (relative to innervation)	Junctional and non-junctional		Junctional (mainly?)
Functional response (model systems)	Rat vas deferens contraction	Rat spleen contraction	Rat aorta contraction
Ligand-binding assay (other than transfected)	Rat submandibular gland	Rat spleen	(None)
Noradrenaline potency	Moderate	Moderate	High
Selective agonists	A61603	/	/
Selective antagonists	RS 100329	/	BMY 7378
Sensitivity to CEC <sup>a</sup>	+	++	+
Second messengers systems	Gq/11, PI turnover	Gq/11, PI turnover	Gq/11, PI turnover

<sup>a</sup> CEC affects all subtypes

A 61603, (*N*-[5-(4,5-dihydro-1H-imidazol-2yl)-2-hydroxy-5,6,7,8-tetrahydronaphthalen-1-yl]-methanesulfonamide); BMY 7378, (8-[2-(4-(2-methoxyphenyl) piperazin-1-yl)ethyl]-8-azaspiro[4,5]decane-7,9-dione); CEC, chloroethylclonidine; Gq/11, G-protein; PI, phosphoinositol

deferens [23]. The same is true of mice lacking the purinergic P2X1 receptor [24], suggesting that both  $\alpha$ 1A- and purinergic responses are required for normal vas deferens function and ejaculation.

A major stimulus to the development of new  $\alpha$ 1-adrenoceptor antagonist drugs has been drug therapy of benign prostatic hypertrophy, which affects an increasing proportion of men as they age, causing problems with micturition due to outflow obstruction. Outflow obstruction consists of a static component due to compression of the urethra by the enlarged prostate, and a dynamic component due to  $\alpha$ 1-adrenoceptor-mediated contraction of the bladder neck, prostate and urethra. The dynamic component may contribute nearly 50% of the total urethral obstruction [25], leading to the use of initially non-selective  $\alpha$ -adrenoceptor antagonists [26]. New  $\alpha$ 1-adrenoceptor antagonists were developed for effects in the lower urinary tract (see [27]), and the receptors involved were identifed as alA-adrenoceptors. Recent evidence suggests that, in addition to a1Aadrenoceptors, alD-adrenoceptors are also present to a significant extent in human prostate [28]. However, some antagonists that were selective for  $\alpha$ 1A-adrenoceptors in ligand-binding studies had low potencies in functional studies of the lower urinary tract, e.g. RS 17053 [29]. These studies brought the study of  $\alpha$ 1A-adrenoceptors into contact with parallel studies of  $\alpha$ 1L-adrenoceptors (see below).

## Selective ligands for alpha1-adrenoceptor subtypes

RS 100329 is a selective  $\alpha$ 1A-adrenoceptor antagonist [30], and A61603 is an  $\alpha$ 1A-adrenoceptor selective agonist,

reported to be 200 times more potent than noradrenaline at causing contractions of rat vas deferens [31].

Risperidone, AH11110A and cyclazosin have been proposed as selective  $\alpha$ 1B-adrenoceptor antagonists [32–34], but these selectivities have been questioned in functional studies [35–37]. Chloroethylclonidine has been used to identify subtypes of  $\alpha$ 1-adrenoceptor because of its reported actions to selectively alkylate  $\alpha$ 1B-adrenoceptors, but chloroethylclonidine interacts with all subtypes of  $\alpha$ 1-adrenoceptors [38, 40]. Overall there is currently no useful antagonist for the study of functional  $\alpha$ 1B-adrenoceptors.

BMY 7378 is a selective  $\alpha$ 1D-adrenoceptor antagonist [41], but also shows potency as an  $\alpha$ 2C-adrenoceptor antagonist [42] and is an antagonist/partial agonst at 5-HT1A receptors [43]. Despite this, it had proved to be a very useful selective antagonist in functional studies.

Silodosin (KMD-3213) is reported to be a selective  $\alpha$ 1A- and  $\alpha$ 1L-adrenoceptor antagonist, but is marketed as an  $\alpha$ 1A-adrenoceptor antagonist [44].

In the present author's opinion, the most reliable and widely studied selective antagonists are RS 100329 ( $\alpha$ 1A) and BMY 7378 ( $\alpha$ 1D) when used at appropriate concentrations and taking cogniscence of the pitfalls (see Table 1).

## Responses mediated by alA-adrenoceptors

Contractions are reported to be mediated at least partly by  $\alpha$ 1A-adrenoceptors in a number of tissues including rat vas deferens [45–48], rat renal artery (also  $\alpha$ 1D: [49]), rat tail artery [50, 51], rat right atrium (positive inotropic actions

[52]), rabbit ear artery [53], pig internal anal sphincter (also  $\alpha$ 1L: [54]), human vas deferens [55, 56] and human prostate ([57]; also  $\alpha$ 1B: [58],  $\alpha$ 1D: [28]; but see below for  $\alpha$ 1L). In rat vas deferens,  $\alpha$ 1A-adrenoceptors mediate two types of response: phasic, probably due to release of Ca<sup>2+</sup> from ryanodine sensitive stores, and tonic via protein kinase C involving diacylglycerol and influx of Ca<sup>2+</sup> via nifedipine-sensitive L-type channels [47] and possibly also T-type channels. Rat submandibular gland has been employed as a model of  $\alpha$ 1A-adrenoceptor ligand-binding sites (see [59]), but may contain both  $\alpha$ 1A- and  $\alpha$ 1B-adrenoceptors [60]. Positive inotropic actions of phenyl-ephrine in mouse involve alpha1A-adrenoceptors [61]. Contractions to noradrenaline were minimal in prostate from the  $\alpha$ 1A-adrenoceptor KO mice [62].

 $\alpha$ 1A-Adrenoceptor overexpression increases beta-adrenoceptor-mediated contractility in the heart and improves outcome from myocardiac infarction [63].

#### α1L-Adrenoceptors: α1A-adrenoceptors

One of the earliest functional subclassifications of  $\alpha$ 1-adrenoceptors was  $\alpha$ 1H and  $\alpha$ 1L, with high and low affinity for prazosin (see [64]), although prazosin has a wide range of affinities for  $\alpha$ 1-adrenoceptors in functional studies [11, 65]. Muramatsu and coworkers [66] subdivided  $\alpha$ 1-adrenoceptors into three subtypes,  $\alpha$ 1H,  $\alpha$ 1N and  $\alpha$ 1L, based on their affinities especially for prazosin.  $\alpha$ 1H-Adrenoceptors had high affinity for prazosin and appeared to match the  $\alpha$ 1A,  $\alpha$ 1B,  $\alpha$ 1D classification [67], whereas  $\alpha$ 1L (the  $\alpha$ 1N designation was dropped) had low affinity for prazosin and did not seem to match molecular cloningbased classifications. Under this classification and based on the low potency of prazosin,  $\alpha$ 1L-adrenoceptors were present in rabbit aorta, mesenteric and carotid arteries [66], guinea-pig aorta [67], rat anococcygeus mucle [68] and rat vas deferens (in addition to  $\alpha 1A$ : [69]; in longitudinal but not circular muscle: [70]). However, other authors have found that contractions of rat vas deferens to exogenous agonists are mediated by alA-adrenoceptors as demonstrated by the very significant correlation with  $\alpha$ 1Aadrenoceptor ligand binding sites [71].  $\alpha$ 1L-Adrenoceptors have also been reported in rabbit cutaneous resistance arteries (predominant adrenoceptor is  $\alpha 1B$ : [72]), rat small mesenteric artery [73], pig prostatic small arteries [74], guinea-pig aorta [75], rabbit iris [76], pig internal anal sphincter [54], rabbit bladder neck [77], in human, rat and dog urethra, dog and mouse prostate, but not in human prostate [58, 78, 79]. In contrast, Muramatsu et al. [67] reported alL-adrenoceptors in human prostate. alL-Adrenoceptor-mediated responses in prostate were abolished in  $\alpha$ 1A-adrenoceptor KO mice [62].

 $\alpha$ 1-Adrenoceptors displayed properties of the  $\alpha$ 1Aadrenoceptor in ligand bind studies, but properties of the  $\alpha$ 1L-adrenoceptor in functional studies [80, 81] or in intact tissue segments [82]. In studies of mRNA levels,  $\alpha$ 1Ladrenoceptors correlated with tissues expressing predominantly alpha1A-adrenoceptors [83]. Genetic polymorphism of  $\alpha$ 1A-adrenoceptors does not explain  $\alpha$ 1L-adrenoceptors, since human  $\alpha$ 1A-adrenoceptor splice variants [84] and homo- and heterodimers of human  $\alpha$ 1A variants [85, 86] have been found to have similar pharmacological characteristics. It can be concluded that  $\alpha$ 1L-adrenoceptors are a functional phenotype of the  $\alpha$ 1A-adrenoceptor, although as yet it is not clear under what circumstances  $\alpha$ 1L-adrenoceptor pharmacology is exhibited.

### Responses mediated by *α*1B-adrenoceptors

Studies of  $\alpha$ 1B-adrenoceptor-mediated function have been hampered by lack of a truly selective antagonist. Contractions are reported to be mediated at least partly by  $\alpha$ 1Badrenoceptors in a number of tissues including rat spleen (in addition to  $\alpha$ 2-adrenoceptors) [45, 46, 48], mouse spleen [87], rat right atrium (positive inotropic, also  $\alpha$ 1A: [52]), rabbit corpus cavernosum [48], rabbit cutaneous resistance arteries (also  $\alpha$ 1L: [72]) and human prostate (also  $\alpha$ 1A: [58], but see also  $\alpha$ 1L). Rat spleen is employed as a model of  $\alpha$ 1B-adrenoceptor ligand-binding sites (see [59]). Rat submandibular gland is reported to contain both  $\alpha$ 1A and  $\alpha$ 1B-adrenoceptors, but the secretion of saliva may mainly involve  $\alpha$ 1B-adrenoceptors [60].

The function of  $\alpha$ 1B-adrenoceptors has been clarified by the use of knockout technology. In aorta from a1B-KO mice there was a small reduction in the potency of noradrenaline or phenylephrine as compared to WT [88], or no significant change in potency [89]. Combined  $\alpha 1B/\alpha 1D$ -KO abolished contractions to noradrenaline and phenylephrine in aorta, having more effect than a1D-KO alone [89]. Daly et al. [90] demonstrated a minor contractile role of alB-adrenoceptors in mouse arteries, including the aorta and tail artery, using knockout technology. Although  $\alpha 1B$ -KO mice show some differences in vascular responsiveness, it has been pointed out that if the  $\alpha$ 1B-adrenoceptor has a regulatory or trophic role or is required for cell surface expression of other subtypes (see [86]), its absence affect vascular responses involving might other  $\alpha$ 1-adrenoceptors even though it was not directly involved in contraction [90]. Hence, results in studies of KO animals must be considered in the light of information from wildtype animals. Contractions in rat tail artery develop more slowly in  $\alpha$ 1B-adrenoceptor knock-out mice [91], so that subtle differences can be revealed following receptor knock-out.

 $\alpha$ 1B-Adrenoceptor overexpression decreases beta-adrenoceptor-mediated contractility in the heart [63] and results in hypertrophy of the cardiac muscle and hypotension [92], and predisposes to heart failure [63]. Pressor responses to phenylephrine in vivo and contractions in the isolated mesenteric artery were unchanged by  $\alpha$ 1B-adrenoceptor overexpression [92]. Overexpression of  $\alpha$ 1B-adrenoceptors blunts the positive inotropic actions on phenylephrine in mouse isolated heart because of a reduction in  $\alpha$ 1A-adrenoceptors, suggesting a regulatory rather than contractile role for this receptor [61].

## Responses mediated by a1D-adrenoceptors

Contractions are reported to be mediated at least partly by α1D-adrenoceptors in a number of tissues including rat aorta, mesenteric artery, iliac artery and pulmonary artery [93, 94], rat renal artery (also  $\alpha 1A$ : [49]), rat carotid artery, mesenteric artery, aorta [50], rabbit aorta (also possibly  $\alpha$ 1A: [53]) and rabbit ventricle (also other subtypes: [95]). In contrast,  $\alpha$ 1D-adrenoceptors are reported not to be involved in caudal, mesenteric or renal arteries [93]. In studies of mouse carotid artery from WT and alpha1D-KO, there was evidence for predominantly a1D-adrenoceptormediated contractions with some regulatory role for the alB-adrenoceptor [96]. Mouse aortic contractions to noradrenaline and phenylephrine were unaffected by alB-KO [89], markedly reduced by  $\alpha$ 1D-KO [89, 97], but abolished by combination of  $\alpha$ 1B/1D-KO [89], suggesting a regulatory or co-operative role for  $\alpha$ 1B-adrenoceptors. However, overexpression of a1B-adrenoceptors did not affect the  $\alpha$ 1D-adrenoceptor response of mouse aorta [98] or mesenteric artery [92]. In mouse mesenteric artery,  $\alpha 1B$ - had no role,  $\alpha$ 1D-adrenoceptors had a large role in contractions [99], and  $\alpha$ 1D-adrenoceptors can be revealed in femoral arteries using KO mice [100].

In addition to mediating contractions of vascular smooth muscle,  $\alpha$ 1-adrenoceptors may induce endotheliumdependent relaxations. It is reported that endotheliumdependent relaxations occur to phenylephrine in the rat mesenteric vascular bed due to  $\alpha$ 1D-adrenoceptor stimulaton [101], and  $\alpha$ 1D-adrenoceptor activation has trophic effects on endothelial cells [102].

Adrenoceptors mediating contractions to nerve stimulation are predominantly  $\alpha$ 1D in both rat [71] and mouse vas deferens (evidence from  $\alpha$ 1D-KO mice, [103]), although contractions of exogenous noradrenaline are predominantly  $\alpha$ 1A-adrenoceptor mediated [45]. In rat femoral arteries, contractions to exogenous noradrenaline were mediated by  $\alpha$ 1A-adrenoceptors, but responses to nerve released noradrenaline involved  $\alpha$ 1A- and  $\alpha$ 1Dadrenoceptors [100]. In addition to  $\alpha$ 1A-adrenoceptors,  $\alpha$ 1D-adrenoceptors are also expressed to a significant extent in human prostate [28], although their location has not been established.

Sympathectomy has been shown to alter the balance of  $\alpha$ 1-adrenoceptor subtypes in rat vas deferens. Although ligand-binding studies of normal rat vas deferens demonstrate a single population of  $\alpha$ 1A-adrenoceptors, tissues from rats sympathectomised with 6-hydroxydopamine demonstrate both  $\alpha$ 1A- and  $\alpha$ 1D-adrenoceptors [71]. Results obtained from sympathectomised rats suggests that phasic contractions are mainly  $\alpha$ 1D-adrenoceptor mediated, whereas tonic contractions are mainly a1Aadrenoceptor mediated, based on the effects of BMY 7378 and the alA-adrenoceptor antagonist RS 100329. Likewise, it has been reported that  $\alpha$ 1D-adrenoceptors are involved in reserpine-induced supersensitivity of rat tail artery [104]. These studies suggest that  $\alpha$ 1D-adrenoceptors are restricted to the junctional region by nerve activity, but if nerves are lost, these receptors spread from the junctional region along the smooth muscle. As a corollary, the rat aorta, which lacks a functional innervation, contains mainly a1D-adrenoceptors on the smooth muscle. How widespread are neuronal  $\alpha$ 1D-adrenoceptors? Clearly, contractions in a number of tissues are mediated by more than one subtype of  $\alpha$ 1-adrenoceptor, and currently available subtype-selective antagonists (particularly for  $\alpha 1B$ adrenoceptor) are often not selective enough to tease out clearly which receptors are present, requiring the continued use of  $\alpha$ 1-adrenoceptor KO mice.

#### **Inverse agonists**

It has become clear in recent years that antagonists may act as inverse agonists at  $\alpha$ 1-adrenoceptors. This means that they not only block the actions of agonists at the receptor, they also reduce the constitutive baseline activity of the G-protein coupled receptor in the absence of agonist. Pure antagonists, or neutral antagonists, do not affect baseline activity of the G-protein coupled receptor.

A number of studies have investigated the ability of calcium re-addition to produce contractions in the absence of an  $\alpha$ 1-adrenoceptor agonist following depletion of calcium stores, particularly in rat aorta, and the ability of  $\alpha$ 1-adrenoceptor antagonists to inhibit this contraction [105]. This phenomenon occurred in aorta [105] and iliac and proximal mesenteric arteries [106], but not tail artery (see [107]), and was blocked by benoxathian, WB 41001, prazosin, BMY 73778 and 5-methylurapidil [105, 107]. Furthermore, increased potency of BMY 7378 in aorta from SHR suggested an increase in this phenomenon in hypertension (see [107]). It was concluded by these authors that the phenomenon of contraction to calcium re-addition

occurred only for  $\alpha$ 1D-adrenoceptors, suggesting that these are constitutively active. Studies of human aortic smooth muscle cells have confirmed that the  $\alpha$ 1D-adrenoceptor is coupled to increases in intracellular calcium [108], and other studies of native receptors suggest the  $\alpha$ 1D is constitutively active [109].

In studies of constitutively active mutations of  $\alpha$ 1a and  $\alpha$ 1b adrenoceptors, a number of antagonists exhibited inverse agonism with marked inhibition: 5-methylurapidil, RS 17053 and tamsulosin at the alpha1a, and 5-methylurapidil at the 1b, but prazosin had only minor actions [110].

#### **Receptor dimers and oligomers**

G-protein-coupled receptors can also exist as dimers, or oligomers, both homologous and heterologous [111, 112]. Co-expression of the  $\alpha$ 1D- with  $\alpha$ 1B [86] or beta2-adrenoceptors [113] is reported to increase the cell surface expression of  $\alpha$ 1D-adrenoceptors, suggesting that  $\alpha$ 1D-adrenoceptor expression and function may involve heterodimerization with these other adrenoceptors. How this relates to expression of  $\alpha$ 1D-adrenoceptors in various smooth muscles is as yet unclear, as  $\alpha$ 1D-adrenoceptor-mediated actions can be easily investigated in functional studies (see above). Studies of other G-protein coupled receptors have found that the serotonin 5-HT2A and the glutamate mGlu2 receptor form functional dimers with distinct signalling [114].

#### α1-Adrenoceptor-mediated second messenger systems

 $\alpha$ 1-Adrenoceptor agonists can induce smooth muscle contraction and other responses by depolarisation-mediated calcium entry through L-type or T-type calcium channels, by directly activating Ca<sup>2+</sup> channels to cause calcium entry, by releasing Ca<sup>2+</sup> from intracellular stores or by sensitising the contractile apparatus to Ca<sup>2+</sup> [115] (see Fig. 2).

 $\alpha$ 1-Adrenoceptors are coupled to a wide variety of second messenger systems via G-proteins, predominatly by pertussis toxin-insensitive G-proteins of the Gq/11 family to phospholipase C [116, 117]. Activation of all  $\alpha$ 1-adrenoceptor subtypes results via phospholipase C in formation of inositol triphosphate and diacylglycerol. Diacylglycerol stimulates protein kinase C, and inositol triphosphate acts on the inositol triphosphate receptor in endoplasmic reticulum to release stored calcium: the net result is increased entry of extracellular Ca<sup>2+</sup> and/or release from Ca<sup>2+</sup> stores [116, 117] (see Fig. 2).  $\alpha$ 1-Adrenoceptor activation causes phospholipase A2 stimulation and arachidonic acid release in the mammalian COS cell line [118], possibly through Gi/Go [119], causes arachidonic acid release by



Fig. 2 A simplified diagram illustrating the possible ways in which  $\alpha$ 1-adrenoceptor activation can result in contraction, by Ca<sup>2+</sup> mobilisation or Ca<sup>2+</sup> sensitisation. *alpha1* alpha1-adrenoceptor, *SR* sarcoplasmic reticulum, *PLC* phospholipase C, *IP3* inositol triphosphate, *DAG* diacylgylcerol, *PKC* protein kinase C, *MLCK* myosin light chain kinase, *MLCP* myosin light chain phosphatase, *stim* stimulate

phospholipase D activation in rat tail artery [120] and can lead to cAMP production [118, 121]. The positive inotropic actions of alpha1-adrenoceptor agonists in rat heart involve Gs and stimulation of cAMP production leading to inhibition of potassium efflux [122].

In addition to signalling through heterotrimeric G-proteins,  $\alpha$ 1-adrenoceptors may mediate responses through other mechanisms. In rat tail artery,  $\alpha$ 1-adrenoceptormediated calcium sensitisation is due mainly to the activation, via the small GTP binding protein RhoA, of Rho kinase [123], which phosphorylates and so inhibits myosin light-chain phosphatase (see [124]) (see Fig. 2).

#### Control of blood pressure

 $\alpha$ 1-Adrenoceptors in the vascular system have a major role in the control of blood pressure and in the baroreflex response to falls in blood pressure. Piascik et al. [125] reported that the  $\alpha$ 1A-adrenoceptor subtype played a role in the tonic maintenance of blood pressure in the conscious rat, whereas the  $\alpha$ 1B-adrenoceptor (or perhaps more correctly non-alpha1A-adrenoceptor) subtype participates in the response to exogenous agonists. In the pithed rat, both pressor nerve responses and responses to exogenous noradrenaline are reported to involve both  $\alpha$ 1A- and  $\alpha$ 1Dadrenoceptors [126], and in the pithed mouse the pressor response to noradrenaline is largely  $\alpha$ 1A-adrenoceptor mediated [127].

Studies of knock-out mice have given insights into the role of the various subtypes of alpha1-adrenoceptor in blood pressure control. In knock-out mice lacking the  $\alpha$ 1Aadrenoceptor, there was a significant fall in blood pressure, both in tail cuff meaurement and invasive recording, but the pressor response to phenylephrine was largely unchanged [128] (Table 2). In knock-out mice lacking the  $\alpha$ 1B-adrenoceptor subtype, there was no effect on basal blood pressure [88, 89], but the pressor responses to phenylephrine were significantly blunted [88] or unchanged [89]. However, in  $\alpha 1A/\alpha 1B$  double knockout mice, there was no significant fall in blood pressure [129]. In mice lacking the alpha1D-adrenoceptor, or both  $\alpha$ 1D and  $\alpha$ 1B, there was a significant fall in resting blood pressure both by tail cuff and invasive recording, and a small fall in the pressor response both to phenylephrine and noradrenaline [89, 97]. Of note in Table 2 are several facts. Firstly, tail cuff SBP seems a poor guide to invasive MAP. Secondly, MAP in the WT varied markedly among studies (varying between 116.5 and 138 mmHg). This can be explained partly by differing genetic backgrounds, but perhaps also by surgical preparation. Although all studies were in conscious animals, the animals were allowed to recover for 3-24 h, so that animals would not have fully recovered from the surgical trauma.

It was found that the pressor responses to the  $\alpha$ -adrenoceptor agonists noradrenaline and/or phenylephrine were almost unchanged in  $\alpha$ 1A-KO mice, but reduced in both  $\alpha$ 1D- and  $\alpha$ 1B-KO mice. This is perhaps surprising as one might have expected, from the wealth of published studies, that the  $\alpha$ 1A and  $\alpha$ 1D-adrenoceptors would be most important for control of blood pressure. However, there may have been compensatory mechanisms for the loss of  $\alpha$ 1A-adrenoceptors in particular, and the  $\alpha$ 1B-adrenoceptor may have a modulatory rather than contractile role (see above). Hence, both  $\alpha$ 1A- and  $\alpha$ 1D-adrenoceptors are involved in acute blood pressure control, which would agree with the findings with antagonist drugs.

#### **Temperature control**

Another important role of vascular  $\alpha$ 1-adrenoceptors is temperature control, as vasoconstriction of superficial blood vessels is an important mechanism to conserve heat. Methylenedioxymethamphetamine (MDMA) is a widely used recreational drug of abuse, and toxic effects include a life-threatening hyperthermia that can occur particularly when the drug is used in a "rave" environment. In animal studies, MDMA disrupts thermoregulation, often causing hypothermia at low ambient temperatures and hyperthermia at high ambient temperatures [130, 131]. In the presence of  $\alpha$ 1-adrenoceptor antagonists, the monophasic hyperthermic response produced by MDMA in mouse became a biphasic response: hypothermia followed by hyperthermia, and this probably involves both  $\alpha$ 1A- and  $\alpha$ 1D-adrenoceptors [132].  $\beta$ 3-Adrenoceptors have also been implicated in thermogenesis [133], but  $\beta$ 3-adrenoceptor ligands have been associated with  $\alpha$ 1-adrenoceptor antagonism, and some of the actions may involve alpha1adrenoceptors [59, 134].

Parameter	WT	α1Α-ΚΟ	α1B-KO	α1D-KO	Notes	Ref.
Conscious tail cuff SBP (mmHg)	114	104*	/	/		[128]
	99	/	99	93*	1B/1D double KO: 92*	[ <mark>89</mark> ]
	108.7	/	/	99.1*	1A/1B double KO: 112	[ <mark>97</mark> ]
	111	/	/	/		[129]
Conscious invasive MAP (mmHg)	138	121*	/	/	24 h post surgery	[128]
	119.3	/	118.5	/	3 h post surgery	[ <mark>88</mark> ]
	118	/	111	109*	1B/1D double KO: 103*; 24 h post surgery	[89]
	116.5	/	/	106.9*	24 h post surgery	[ <mark>97</mark> ]
Phe pressor		No change [128]	Decrease [88]	Small decrease [97]		
			No change [89]	No change [89]	1B/1D double KO: decrease [89]	
NA pressor			Small decrease [88]	Small decrease [97]		
			Small decrease [89]	Decrease [89]	1B/1D double KO: decrease [89]	

**Table 2** Blood pressure responses in WT and  $\alpha$ 1-adrenoceptor subtype KO mice

Results taken from [88, 89, 97, 128, 129]

SBP systolic blood pressure (recorded by tail cuff), HR heart rate, MAP mean arterial pressure (recorded by invasive cannula), phe pressor pressor response to phenylephrine relative to WT, NA pressor pressor response to noradrenaline relative to WT

\*Significant difference from respective WT

Peripheral effects of MDMA at  $\alpha$ 1-adrenoceptors can explain a component of the hyperthermia: cutaneous vasoconstriction by MDMA prevents an early hypothermic response to the drug. At low ambient temperatures, cutaneous vasoconstriction is already marked so that MDMA produces little further vasoconstriction and the, presumed central, hypothermic actions of MDMA may predominate. At high ambient temperatures cutaneous dilatation has occurred, allowing a marked vasoconstrictor component to the actions of MDMA, and hyperthermia predominates. Hence, peripheral  $\alpha$ 1-adrenoceptor-mediated vasoconstrictor actions of MDMA modulate central hypo- and hyperthermic components.

## Neuronal *a*1-adrenoceptors

 $\alpha$ 1A-Adrenoceptors [135] and  $\alpha$ 1B-adrenoceptors [136] are involved in a number of actions in neurones and glial cells in the CNS.  $\alpha$ 1D-Adrenoceptors are also present in the CNS as demonstrated by a significant fall in alpha1binding in  $\alpha$ 1D-KO mice [97].  $\alpha$ 1B-Adrenoceptor overexpression resulted in apoptotic neurodegeneration with a corresponding multiple system atrophy including a Parkinson-like syndrome and grand mal seizures [137].

## α1-Adrenoceptor-mediated inhibition

Although the concept of prejunctional inhibition mediated by  $\alpha$ 2-adrenoceptors is well established (see [2]), some studies support the contention that inhibitory prejunctional  $\alpha$ 1-adrenoceptors exist in pithed rat, rat ventricle, rat vas deferens, rat kidney, dog heart, rat atria, rat tail artery, guinea-pig atria and on the cholinergic nerves of rat gastric fundus (for references, see [11]). Prejunctional inhibition in the CNS involving  $\alpha$ 1-adrenoceptors is also reported in, for instance, the paraventricular nucleus [138]. Other studies suggest transsynaptic inhibition by prostaglandins or purproduced postjunctionally by  $\alpha$ 1-adrenoceptor ines stimulation [139, 140], and the possibility of  $\alpha$ 2-adrenoceptor-mediated actions of some  $\alpha$ 1-adrenoceptor antagonists must be considered.

## α1-Adrenoceptor-mediated facilitation

 $\alpha$ 1-Adrenoceptor agonists have been reported to facilitate release of acetylcholine in rat heart [141] and cat [142] and rat bladder [143, 144]. These  $\alpha$ 1-adrenoceptors may be on the soma of bladder parasympathetic neurones and mediate a slow postsynaptic depolarisation [143]. In the CNS, facilitation of rat spinal motoneuron activity [145] and of vasopressin release [146] are reported to be mediated by  $\alpha$ 1-adrenoceptors.  $\alpha$ 1-Adrenoceptor activation stimulates

inhibitory GABAergic neurotransmission in rat spinal cord [147], rat cerebellum [148], mouse accessory olfactory bulb [149] and mouse hypothalamus [150]. The  $\alpha$ 1-adrenoceptor-mediated facilitation may involve protein kinase C and increases in intracellular calcium [146, 148, 151].

#### **Concluding remarks**

Pharmacological and receptor knockout techniques have greatly increased our understanding of  $\alpha$ 1-adrenoceptors in terms of location and function of the three subtypes. Areas of particular interest in the next few years will be investigation of the role of  $\alpha$ 1-adrenoceptor subtypes in the central nervous system, development of 'missing' subtype selective agonists and antagonists, further development of drugs for benign prostatic hypertrophy and elucidation of the role of the  $\alpha$ 1B-adrenoceptor.

#### References

- Ahlquist RP (1948) A study of the adrenotropic receptors. Am J Physiol 153:586–600
- Starke K (1977) Regulation of noradrenaline release by presynaptic receptor systems. Rev Physiol Biochem Pharmacol 77:1–124
- Starke K, Langer SZ (1979) A note on the terminology for presynaptic receptors. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 1–3
- Morrow AL, Creese I (1986) Characterization of alphaladrenergic receptor subtypes in rat brain: a re-evaluation of [3H]WB4101 and [3H]prazosin binding. Mol Pharmacol 29:321–330
- Han C, Abel PW, Minneman KP (1987) Alpha1-adrenoceptor subtypes linked to different mechanisms for increasing intracellular Ca<sup>2+</sup> in smooth muscle. Nature 329:333–335
- Cotecchia S, Schwinn DA, Randall RR, Lefkowitz RJ, Caron MG, Kobilka BK (1988) Molecular cloning and expression of the cDNA for the hamster alpha1-adrenergic receptor. Proc Natl Acad Sci USA 85:7159–7163
- Lomasney JW, Cotecchia S, Lorenz W, Leung W-Y, Schwinn DA, Yang-Feng TL, Brownstein M, Lefkowitz RJ, Caron M (1991) Molecular cloning and expression of the cDNA for the alpha1<sub>A</sub>-adrenergic receptor. J Biol Chem 266:6365–6369
- Schwinn DA, Lomasney JW, Lorenz W, Szklut PJ, Fremeau RT, Yang-Feng TL, Caron MG, Lefkowitz RJ, Cotecchia S (1990) Molecular cloning and expression of the cDNA for a novel alpha1-adrenergic receptor subtype. J Biol Chem 265:8183–8189
- Perez DM, Piascik MT, Graham RM (1991) Solution-phase library screening for the identification of rare clones: isolation of an alpha<sub>1D</sub>-adrenergic receptor cDNA. Mol Pharmacol 40:876– 883
- Hieble JP, Bylund DB, Clarke DE, Eikenbur DC, Langer SZ, Lefkowitz RJ, Minneman KP, Ruffolo RR (1995) International Union of Pharmacology. X. Recommendation for nomenclature of alpha1-adrenoceptors: Consensus update. Pharmacol Rev 47:267–270
- Docherty JR (1998) Subtypes of functional alphal- and alpha2adrenoceptors. Eur J Pharmacol 361:1–15

- Guimarães S, Moura D (2001) Vascular adrenoceptors: an update. Pharmacol Rev 53:319–356
- Hodges AN, Lynn BM, Bula JE, Donaldson MG, Dagenais MO, McKenzie DC (2003) Effects of pseudoephedrine on maximal cycling power and submaximal cycling efficiency. Med Sci Sports Exerc 35:1316–1319
- Swain RA, Harsha DM, Baenziger J, Saywell RM Jr (1997) Do pseudoephedrine or phenylpropanolamine improve maximum oxygen uptake and time to exhaustion? Clin J Sport Med 7:168– 173
- Drew CD, Knight GT, Hughes DT, Bush M (1978) Comparison of the effects of D-(-)-ephedrine and L-(+)-pseudoephedrine on the cardiovascular and respiratory systems in man. Br J Clin Pharmacol 6:221–225
- Empey DW, Young GA, Letley E, John GC, Smith P, Mc-Donnell KA, Bagg LR, Hughes DT (1980) Dose–response study of the nasal decongestant and cardiovascular effects of pseudoephedrine. Br J Clin Pharmacol 9:351–358
- Bright TP, Sandage BW Jr, Fletcher HP (1981) Selected cardiac and metabolic responses to pseudoephedrine with exercise. J Clin Pharmacol 21:488–492
- Ma G, Bavadekar SA, Davis YM, Lalchandani SG, Nagmani R, Schaneberg BT, Khan IA, Feller D (2007) Pharmacological effects of ephedrine alkaloids on human alpha(1)- and alpha(2)adrenergic receptor subtypes. J Pharmacol Exp Ther 322:214
- Vansal SS, Feller DR (1999) Direct effects of ephedrine isomers on human beta-adrenergic receptor subtypes. Biochem Pharmacol 58:807–810
- Ulahannan TJ, Karpe F, Humphreys SM, Matthews DR, Frayn KN (2002) Effects of acute administration of doxazosin on fasting and postprandial haemodynamics and lipid metabolism in healthy subjects. Horm Metab Res 34:499–503
- Flechtner-Mors M, Jenkinson CP, Alt A, Biesalski HK, Adler G, Ditschuneit HH (2004) Sympathetic regulation of glucose uptake by the alpha1-adrenoceptor in human obesity. Obes Res 12:612–620
- 22. Chen Q, Takahashi S, Zhong S, Hosoda C, Zheng HY, Ogushi T, Fujimura T, Ohta N, Tanoue A, Tsujimoto G, Kitamura T (2005) Function of the lower urinary tract in mice lacking alpha1d-adrenoceptor. J Urol 174:370–374
- Sanbe A, Tanaka Y, Fujiwara Y, Tsumura H, Yamauchi J, Cotecchia S, Koike K, Tsujimoto G, Tanoue A (2007) Alphaladrenoceptors are required for normal male sexual function. Br J Pharmacol 152:332–340
- Mulryan K, Gitterman DP, Lewis CJ, Vial C, Leckie BJ, Cobb AL, Brown JE, Conley EC, Buell G, Pritchard CA, Evans RJ (2000) Reduced vas deferens contraction and male infertility in mice lacking P2X1 receptors. Nature 403:86–89
- Furuya S, Kumamoto Y, Yokoyama E, Tsukamoto T, Izumi T, Abiko Y (1982) Alpha-adrenergic activity and urethral pressure in prostatic zone in benign prostatic hypertrophy. J Urol 128:836–839
- Caine M (1986) The present role of alpha-adrenergic blockers in the treatment of benign prostatic hypertrophy. J Urol 136:1–4
- Nickel JC, Sander S, Moon TD (2008) A meta-analysis of the vascular-related safety profile and efficacy of alpha-adrenergic blockers for symptoms related to benign prostatic hyperplasia. Int J Clin Pract 62:1547–1559
- Kojima Y, Sasaki S, Shinoura H, Hayashi Y, Tsujimoto G, Kohri K (2006) Quantification of alpha1-adrenoceptor subtypes by real-time RT-PCR and correlation with age and prostate volume in benign prostatic hyperplasia patients. Prostate 66:761–767
- Ford APDW, Arredondo NF, Blue DR, Bonhaus DW, Jasper J, Kava MS, Lesnick J, Pister JR, Shieh IM, Vimont RL, Williams TJ, McNea JE, Stamey TA, Clarke DE (1996) RS-17053, a

selective alpha<sub>1A</sub>-adrenoceptor antagonist, displays low affinity for functional alpha1-adrenoceptors in human prostate: implications for adrenoceptor classification. Mol Pharmacol 49:209–215

- 30. Williams TJ, Blue DR, Daniels DV, Davis B, Elworthy T, Gever JR, Kava MS, Morgans D, Padilla F, Tassa S, Vimont RL, Chapple CR, Chess-Williams R, Eglen RM, Clarke DE, Ford AP (1999) In vitro alpha1-adrenoceptor pharmacology of Ro 70-0004 and RS-100329, novel alpha1A-adrenoceptor selective antagonists. Br J Pharmacol 127:252–258
- Knepper SM, Buckner SA, Brune ME, DeBernardis JF, Meyer MD, Hancock AA (1995) A-61603, a potent alpha1-adrenergic receptor agonist, selective for the alpha<sub>1A</sub> receptor subtype. J Pharmacol Exp Ther 274:97–103
- Sleight AJ, Koek W, Bigg DC (1993) Binding of antipsychotic drugs at alpha1A- and alpha1B-adrenoceptors: risperidone is selective for the alpha 1B-adrenoceptors. Eur J Pharmacol 238:407–410
- 33. King HK, Goetz AS, Ward SDC, Saussy DL Jr (1994) AH11110A is selective for the  $\alpha$ 1B subtype of  $\alpha$ 1-adrenoceptors. Soc Neurosci Abstr 20 p 52
- 34. Marucci G, Angeli P, Buccioni M, Gulini U, Melchiorre C, Sagratini G, Testa R, Giardinà D (2005) (+)-Cyclazosin, a selective alpha1B-adrenoceptor antagonist: functional evaluation in rat and rabbit tissues. Eur J Pharmacol 522:100–107
- Eltze M (1996) In functional experiments, risperidone is selective, not for the B, but for the A subtype of alpha-1 adrenoceptor. Eur J Pharmacol 295:69–73
- 36. Stam WB, Van der Graaf PH, Saxena PR (1998) Functional characterisation of the pharmacological profile of the putative alpha1B-adrenoceptor antagonist, (+)-cyclazosin. Eur J Pharmacol 361:79–83
- 37. Eltze M, König H, Ullrich B, Grebe T (2001) Failure of AH11110A to functionally discriminate between alpha(1)-adrenoceptor subtypes A, B and D or between alpha(1)- and alpha(2)-adrenoceptors. Eur J Pharmacol 415:265–276
- Michel MC, Kerker J, Branchek TA, Forray C (1993) Selective irreversible binding of chloroethylclonidine at alpha1- and alpha2-adrenoceptor subtypes. Mol Pharmacol 44:1165–1170
- O'Rourke M, Kearns S, Docherty JR (1995) Investigations of the actions of chloroethylclonidine in rat aorta. Br J Pharmacol 115:1399–1406
- O'Rourke M, Gavin K, Docherty JR (1997) Further investigation of the alpha-adrenoceptor-mediated actions of chloroethylclonidine in rat aorta. Eur J Pharmacol 336:37–42
- 41. Goetz AS, King HK, Ward SDC, True TA, Rimele TJ, Saussy DL (1995) BMY 7378 is a selective antagonist of the D subtype of alpha1-adrenoceptors. Eur J Pharmacol 272:R5–R6
- 42. Cleary L, Murad K, Bexis S, Docherty JR (2005) The alpha (1D)-adrenoceptor antagonist BMY 7378 is also an alpha (2C)-adrenoceptor antagonist. Auton Autacoid Pharmacol 25:135–141
- 43. Chaput Y, de Montigny C (1988) Effects of the 5-hydroxytryptamine receptor antagonist, BMY 7378, on 5-hydroxytryptamine neurotransmission: electrophysiological studies in the rat central nervous system. J Pharmacol Exp Ther 246:359–370
- 44. Murata S, Taniguchi T, Takahashi M, Okada K, Akiyama K, Muramatsu I (2000) Tissue selectivity of KMD-3213, an alpha(1)-adrenoreceptor antagonist, in human prostate and vasculature. J Urol 164:578–583
- 45. Aboud RW, Shafii M, Docherty JR (1993) Investigations of the subtypes of alpha1-adrenoceptor mediating contractions of rat aorta, vas deferens and spleen. Br J Pharmacol 109:80–87
- 46. Burt RP, Chapple CR, Marshall I (1995) Evidence for a functional alpha<sub>1A</sub>- (alpha<sub>1C</sub>-) adrenoceptor mediating contraction of rat epididymal vas deferens and an alpha<sub>1B</sub>-adrenoceptor

mediating contraction of the rat spleen. Br J Pharmacol 115:467-475

- 47. Burt RP, Chapple CR, Marshall I (1998) Alpha<sub>1A</sub>-Adrenoceptor mediated contraction of rat prostatic vas deferens and the involvement of ryanodine stores and Ca<sup>2+</sup> influx stimulated by diacylglycerol and PKC. Br J Pharmacol 123:317–325
- 48. Noble AJ, Chess-Williams R, Couldwell C, Furukawa K, Uchyiuma T, Korstanje C, Chapple CR (1997) The effects of tamsulosin, a high affinity antagonist at functional alpha<sub>1A</sub>- and alpha<sub>1D</sub>-adrenoceptor subtypes. Br J Pharmacol 120:231–238
- Villalobos-Molina R, Lopez-Guerrero JJ, Ibarra M (1997) Alpha1D- and alpha1A-adrenoceptors mediate contraction in rat renal artery. Eur J Pharmacol 322:225–227
- Villalobos-Molina R, Ibarra M (1996) Alpha1-adrenoceptors mediating contraction in arteries of normotensive and spontaneously hypertensive rats are of the alpha1D or alpha 1A subtypes. Eur J Pharmacol 298:257–263
- 51. Lachnitt WG, Tran AM, Clarke DE, Ford APDW (1997) Pharmacological characterization of an alpha<sub>1A</sub>-adrenoceptor mediating contractile responses to noradrenaline in isolated caudal artery of rat. Br J Pharmacol 120:819–826
- 52. Yu G-S, Han C (1994) Role of alpha<sub>1A</sub>- and alpha<sub>1B</sub>-adrenoceptors in phenylephrine induced positive inotropic response in isolated rat left atrium. J Cardiovasc Pharmacol 24:745–752
- Fagura MS, Lyfdford SJ, Douggall IG (1997) Pharmacological classification of alpha1-adrenoceptors mediating contractions of rabbit isolated ear artery: comparison with rat isolated thoracic aorta. Br J Pharmacol 120:247–258
- Mills K, Hausman N, Chess-Williams R (2008) Characterization of the alpha1-adrenoceptor subtype mediating contractions of the pig internal anal sphincter. Br J Pharmacol 155:110–117
- 55. Furukawa K, Rosario DJ, Smith DJ, Chapple CR, Uchiyama T, Chess-Williams R (1995) Alpha<sub>1A</sub>-adrenoceptor-mediated contractile responses of the human vas deferens. Br J Pharmacol 116:1605–1610
- 56. Moriyama N, Nasu K, Takeuchi T, Akiyama K, Murata S, Mishimatsu H, Yano J, Tsujimoto G, Kawabe K (1997) Quantification and distribution of alpha1-adrenoceptor subtype mRNAs in human vas deferens: comparison with those of epididymal and pelvic portions. Br J Pharmacol 122:1009–1014
- Marshall I, Burt RP, Chapple CR (1995) Noradrenaline contractions of human prostate mediated by alpha<sub>1A</sub>- (alpha<sub>1c</sub>-) adrenoceptor subtype. Br J Pharmacol 115:781–786
- Teng C-M, Guh J-H, Ko F-N (1994) Functional identification of alpha1-adrenoceptor subtypes in human prostate: comparison with those in rat vas deferens and spleen. Eur J Pharmacol 265:61–66
- 59. Bexis S, Docherty JR (2009) Role of  $\alpha$ 1- and  $\beta$ 3-adrenoceptor subtypes in the modulation by SR59230A of the effects of MDMA on body temperature in the mouse. Br J Pharmacol 158:259–266
- 60. Bruchas MR, Toews ML, Bockman CS, Abel PW (2008) Characterization of the alpha1-adrenoceptor subtype activating extracellular signal-regulated kinase in submandibular gland acinar cells. Eur J Pharmacol 578:349–358
- 61. Ross SA, Rorabaugh BR, Chalothorn D, Yun J, Gonzalez-Cabrera PJ, McCune DF, Piascik MT, Perez DM (2003) The alpha(1B)adrenergic receptor decreases the inotropic response in the mouse Langendorff heart model. Cardiovasc Res 60:598–607
- 62. Gray K, Short J, Ventura S (2008) The alpha1A-adrenoceptor gene is required for the alpha1L-adrenoceptor-mediated response in isolated preparations of the mouse prostate. Br J Pharmacol 155:103–109
- Woodcock EA (2007) Roles of alpha1A- and alpha1B-adrenoceptors in heart: insights from studies of genetically modified mice. Clin Exp Pharmacol Physiol 34:884–888

- Flavahan NA, Vanhoutte PA (1986) Alpha1-adrenoceptor subclassification in vascular smooth muscle. Trends Pharmacol Sci 7:347–349
- 65. Docherty JR (1989) The pharmacology of alpha<sub>1</sub>- and alpha<sub>2</sub>adrenoceptors: evidence for and against a further subdivision. Pharmacol Ther 44:241–284
- 66. Muramatsu I, Ohmura T, Kigoshi S, Hashimoto S, Oshita M (1990) Pharmacological subclassification of alpha1-adrenoceptors in vascular smooth muscle. Br J Pharmacol 99:197–201
- Muramatsu I, Oshita M, Ohmura T, Kigoshi S, Akino H, Gobara M, Okada K (1995) Pharmacological characterization of alphaladrenoceptor subtypes in the human prostate: functional and binding studies. Br J Urol 74:572–578
- Ford APDW, Berge NV, Clarke DE (1993) Characterisation of alpha1-adrenoceptors in isolated anococcygeus muscle of rat. Br J Pharmacol 109:112P
- 69. Ohmura T, Oshita M, Kigoshi S, Muramatsu I (1992) Identification of alpha1-adrenoceptor subtypes in the rat vas deferens: binding and functional studies. Br J Pharmacol 107:698–704
- 70. Amobi NI, Guillebaud J, Kaisary AV, Turner E, Smith IC (2002) Discrimination by SZL49 between contractions evoked by noradrenaline in longitudinal and circular muscle of human vas deferens. Br J Pharmacol 136:127–135
- Cleary L, Slattery J, Bexis S, Docherty JR (2004) Sympathectomy reveals alpha1A- and alpha1D-adrenoceptor components to contractions to noradrenaline in rat vas deferens. Br J Pharmacol 143:745–752
- 72. Smith KM, MacMillan JB, McGrath JC (1997) Investigation of alpha1-adrenoceptor subtypes mediating vasoconstriction in rabbit cutaneous resistance arteries. Br J Pharmacol 122: 825–832
- 73. Van der Graaf PH, Shankley NP, Black JW (1996) Analysis of the effects of alpha1-adrenoceptor antagonists on noradrenalinemediate contraction of rat small mesenteric artery. Br J Pharmacol 118:1308–1316
- 74. Recio P, Orensanz LM, Martínez MP, Navarro-Dorado J, Bustamante S, García-Sacristán A, Prieto D, Hernández M (2008) Noradrenergic vasoconstriction of pig prostatic small arteries. Naunyn Schmiedebergs Arch Pharmacol 376:397–406
- Yamamoto Y, Koike K (1999) Alpha1-adrenoceptors in the guinea pig thoracic aorta. J Smooth Muscle Res 35:181–192
- 76. Nakamura S, Taniguchi T, Suzuki F, Akagi Y, Muramatsu I (1999) Evaluation of alpha1-adrenoceptors in the rabbit iris: pharmacological characterization and expression of mRNA. Br J Pharmacol 127:1367–1374
- 77. Kava MS, Blue DR, Vimont RL, Clarke DE, Ford APDW (1998) Alpha<sub>1L</sub>-adrenoceptor mediation of smooth muscle contraction in rabbit bladder neck: a model for lower urinary tract tissues of man. Br J Pharmacol 123:1359–1366
- 78. Fukasawa R, Taniguchi N, Moriyama N, Ukai Y, Yamazaki S, Ueki T, Kameyama S, Kimura K, Kawabe K (1998) The alpha1L-adrenoceptor subtype in the lower urinary tract: a comparison of human urethra and prostate. Br J Urol 82:733–737
- 79. Gray KT, Ventura S (2006) Alpha1L-adrenoceptors mediate contractions of the isolated mouse prostate. Eur J Pharmacol 540:155–161
- Ford APDW, Daniels D, Chang DJ, Gever JR, Jasper JR, Lesnick JD, Clarke DE (1997) Pharmacological pleiotropism of the human recombinant alpha<sub>1A</sub>-adrenoceptor: implications for alpha1-adrenoceptor classification. Br J Pharmacol 121: 137–1135
- Daniels DV, Gever JR, Jasper JR, Kava MS, Lesnick JD, Meloy TD, Stepan G, Williams TJ, Clarke DE, Chang DJ, Ford AP (1999) Human cloned alpha1A-adrenoceptor isoforms display alpha1L-adrenoceptor pharmacology in functional studies. Eur J Pharmacol 370:337–343

- 82. Morishima S, Suzuki F, Yoshiki H, Md Anisuzzaman AS, Sathi ZS, Tanaka T, Muramatsu I (2008) Identification of the alpha1L-adrenoceptor in rat cerebral cortex and possible relationship between alpha1L- and alpha1A-adrenoceptors. Br J Pharmacol 153:1485–1494
- 83. Martí D, Miquel R, Ziani K, Gisbert R, Ivorra MD, Anselmi E, Moreno L, Villagrasa V, Barettino D, D'Ocon P (2005) Correlation between mRNA levels and functional role of alphaladrenoceptor subtypes in arteries: evidence of alphalL as a functional isoform of the alphalA-adrenoceptor. Am J Physiol Heart Circ Physiol 289:H1923–H1932
- 84. Shibata K, Hirasawa A, Moriyama N, Kawabe K, Ogawa S, Tsujimoto G (1996) Alpha<sub>1a</sub>-adrenoceptor polymorphism: pharmacological characterization and association with benign prostatic hypertrophy. Br J Pharmacol 118:1403–1408
- 85. Ramsay D, Carr IC, Pediani J, Lopez-Gimenez JF, Thurlow R, Fidock M, Milligan G (2004) High-affinity interactions between human alpha1A-adrenoceptor C-terminal splice variants produce homo- and heterodimers but do not generate the alpha1Ladrenoceptor. Mol Pharmacol 66:228–239
- Uberti MA, Hall RA, Minneman KP (2003) Subtype-specific dimerization of alpha1-adrenoceptors: effects on receptor expression and pharmacological properties. Mol Pharmacol 64:1379–1390
- Eltze M (1996) Functional evidence for an alpha1B-adrenoceptor mediating contraction of the mouse spleen. Eur J Pharmacol 311:187–198
- Cavalli A, Lattion A, Hummler E, Nenninger M, Pedrazzini T (1997) Decreased blood pressure response in mice deficient of the alpha 1b-adrenergic receptor. Proc Nat Acad Sci USA 94:11589–11595
- 89. Hosoda C, Koshimizu TA, Tanoue A, Nasa Y, Oikawa R, Tomabechi T, Fukuda S, Shinoura H, Oshikawa S, Takeo S, Kitamura T, Cotecchia S, Tsujimoto G (2005) Two alphaladrenergic receptor subtypes regulating the vasopressor response have differential roles in blood pressure regulation. Mol Pharmacol 67:912–922
- Daly CJ, Deighan C, McGee A, Mennie D, Ali Z, McBride M, McGrath JC (2002) A knockout approach indicates a minor vasoconstrictor role for vascular alpha1B-adrenoceptors in mouse. Physiol Genomics 9:85–91
- 91. Daly CJ, Cotecchia S, McGrath JC (1998) Low frequency electrical field stimulation elicits responses in segments of mouse tail artery which are slower in alpha1B-knockout mice than in control mice. Naunyn Schmiedebergs Arch Pharmacol 358:R600
- 92. Zuscik MJ, Chalothorn D, Hellard D, Deighan C, McGee A, Daly CJ, Waugh DJ, Ross SA, Gaivin RJ, Morehead AJ, Thomas JD, Plow EF, McGrath JC, Piascik MT, Perez DM (2001) Hypotension, autonomic failure, and cardiac hypertrophy in transgenic mice overexpressing the alpha 1B-adrenergic receptor. J Biol Chem 276:13738–13743
- 93. Piascik MT, Guarino RD, Smith MS, Soltis EE, Saussy DL, Perez DM (1995) The specific contribution of the novel alpha-1D adrenoceptor to the contraction of vascular smooth muscle. J Pharmacol Exp Ther 275:1583–1589
- Hussain M, Marshall I (1997) Characterization of alpha1-adrenoceptor subtypes mediating contractions to phenylephrine in rat thoracic aorta, mesenteric artery and pulmonary artery. Br J Pharmacol 122:849–858
- 95. Yang H-T, Endoh M (1997) Pharmacological evidence for alpha<sub>1D</sub>-adrenoceptors in the rabbit ventricular myocardium: analysis with BMY 7378. Br J Pharmacol 122:1541–1550
- 96. Deighan C, Methven L, Naghadeh MM, Wokoma A, Macmillan J, Daly CJ, Tanoue A, Tsujimoto G, McGrath JC (2005) Insights into the functional roles of alpha(1)-adrenoceptor subtypes in

mouse carotid arteries using knockout mice. Br J Pharmacol 144:558-565

- 97. Tanoue A, Nasa Y, Koshimizu T, Shinoura H, Oshikawa S, Kawai T, Sunada S, Takeo S, Tsujimoto G (2002) The alpha(1D)-adrenergic receptor directly regulates arterial blood pressure via vasoconstriction. J Clin Invest 109:765–775
- 98. Chalothorn D, McCune DF, Edelmann SE, Tobita K, Keller BB, Lasley RD, Perez DM, Tanoue A, Tsujimoto G, Post GR, Piascik MT (2003) Differential cardiovascular regulatory activities of the alpha1B- and alpha1D-adrenoceptor subtypes. J Pharmacol Exp Ther 305:1045–1053
- 99. Hosoda C, Tanoue A, Shibano M, Tanaka Y, Hiroyama M, Koshimizu TA, Cotecchia S, Kitamura T, Tsujimoto G, Koike K (2005) Correlation between vasoconstrictor roles and mRNA expression of alpha1-adrenoceptor subtypes in blood vessels of genetically engineered mice. Br J Pharmacol 146:456–466
- 100. Zacharia J, Hillier C, Tanoue A, Tsujimoto G, Daly CJ, McGrath JC, MacDonald A (2005) Evidence for involvement of alpha1D-adrenoceptors in contraction of femoral resistance arteries using knockout mice. Br J Pharmacol 146:942–951
- 101. Filippi S, Parenti A, Donnini S, Granger HJ, Fazzini A, Ledda F (2001) Alpha(1D)-adrenoceptors cause endothelium-dependent vasodilatation in the rat mesenteric vascular bed. J Pharmacol Exp Ther 296:869–875
- 102. Vinci MC, Bellik L, Filippi S, Ledda F, Parenti A (2007) Trophic effects induced by alpha1D-adrenoceptors on endothelial cells are potentiated by hypoxia. Am J Physiol Heart Circ Physiol 293:H2140–H2147
- 103. Bexis S, Cleary L, McGrath JC, Tanoue A, Tsujimoto G, Docherty JR (2008) Alpha(1D)-adrenoceptors mediate nerve and agonist-evoked contractions in mouse vas deferens: evidence obtained from knockout technology. Auton Autacoid Pharmacol 28:81–85
- 104. Taki N, Tanaka T, Zhang L, Suzuki F, Israilova M, Taniguchi T, Hiraizumi-Hiraoka Y, Shinozuka K, Kunitomo M, Muramatsu I (2004) Alpha-1D adrenoceptors are involved in reserpineinduced supersensitivity of rat tail artery. Br J Pharmacol 142:647–656
- Noguera MA, Ivorra MD, D'Ocon P (1996) Functional evidence of inverse agonism in vascular smooth muscle. Br J Pharmacol 119:158–164
- 106. Ziani K, Gisbert R, Noguera MA, Ivorra MD, D'Ocon P (2002) Modulatory role of a constitutively active population of alpha(1D)-adrenoceptors in conductance arteries. Am J Physiol Heart Circ Physiol 282:H475–H481
- 107. Gisbert R, Pérez-Vizcaino F, Cogolludo AL, Noguera MA, Ivorra MD, Tamargo J, D'Ocon P (2003) Cytosolic Ca<sup>2+</sup> and phosphoinositide hydrolysis linked to constitutively active alpha1D-adrenoceptors in vascular smooth muscle. J Pharmacol Exp Ther 305:1006–1014
- 108. García-Cazarín ML, Smith JL, Olszewski KA, McCune DF, Simmerman LA, Hadley RW, Kraner SD, Michael T, Piascik MT (2008) The α1D-adrenergic receptor is expressed intracellularly and coupled to increases in intracellular calcium and reactive oxygen species in human aortic smooth muscle cells. J Mol Signal 3:6
- 109. McCune DF, Edelmann SE, Olges JR, Post GR, Waldrop BA, Waugh DJ, Perez DM, Piascik MT (2000) Regulation of the cellular localization and signaling properties of the alpha1B- and alpha1D-adrenoreceptors by agonists and inverse antagonists. Mol Pharmacol 57:659–666
- 110. Rossier O, Abuin L, Fanelli F, Leonardi A, Cotecchia S (1999) Inverse agonism and neutral antagonism at alpha1A- and alpha1B-adrenergic receptor subtypes. Mol Pharmacol 56: 858–866

- 111. Minneman KP (2007) Heterodimerization and surface localization of G protein coupled receptors. Biochem Pharmacol 73:1043–1050
- 112. Dalrymple MB, Pfleger KD, Eidne KA (2008) G protein-coupled receptor dimers: functional consequences, disease states and drug targets. Pharmacol Ther 118:359–371
- 113. Uberti MA, Hague C, Oller H, Minneman KP, Hall RA (2005) Heterodimerization with beta2-adrenergic receptors promotes surface expression and functional activity of alpha1D-adrenergic receptors. J Pharmacol Exp Ther 313:16–23
- 114. González-Maeso J, Ang RL, Yuen T, Chan P, Weisstaub NV, López-Giménez JF, Zhou M, Okawa Y, Callado LF, Milligan G, Gingrich JA, Filizola M, Meana JJ, Sealfon SC (2008) Identification of a serotonin/glutamate receptor complex implicated in psychosis. Nature 452:9
- 115. Chen XL, Rembold CM (1995) Phenylephrine contracts rat tail artery by one electromechanical and three pharmacomechanical mechanisms. Am J Physiol 268:H74–H81
- 116. Minneman KP (1988) Alpha1-adrenergic receptor subtypes, inositol phosphates and sources of cell Ca<sup>2+</sup>. Pharmacol Rev 40:87–119
- 117. Wu D, Katz A, Lee C, Simon MI (1992) Activation of phospholipase C by alpha1-adrenergic receptors in mediated by the alpha subunits of Gq family. J Biol Chem 267: 25798–25802
- 118. Perez DM, DeYoung MP, Graham RM (1993) Coupling of expressed alpha<sub>1B</sub>- and alpha<sub>1D</sub>-adrenergic receptors to multiple signaling pathways is both G-protein and cell type specific. Mol Pharmacol 44:784–795
- 119. Exton JH (1994) Phosphatidylcholine breakdown and signal transduction. Biochim Biophys Acta 1212:26–42
- 120. Gu H, Trajkovic S, LaBelle EF (1992) Norepinephrine-induced phosphatidylcholine hydrolysis by phospholipases D and C in rat tail artery. Am J Physiol 262:C1376–C1383
- 121. Ruan Y, Kan H, Parmentier J-H, Fatima S, Allen LF, Malik KU (1998) Alpha-1A adrenergic receptor stimulation with phenylephrine promotes arachidonic acid release by activation of phospholipase D in rat-1 fibroblasts: inhibition by protein kinase A. J Pharmacol Exp Ther 284:575–585
- 122. Gallego M, Setién R, Puebla L, Boyano-Adánez Mdel C, Arilla E, Casis O (2005) Alpha1-adrenoceptors stimulate a Galphas protein and reduce the transient outward K<sup>+</sup> current via a cAMP/PKA-mediated pathway in the rat heart. Am J Physiol Cell Physiol 288:C577–C585
- 123. Mueed I, Bains P, Zhang L, MacLeod KM (2004) Differential participation of protein kinase C and Rho kinase in α1-adrenoceptor mediated contraction in rat arteries. Can J Physiol Pharmacol 82:895–902
- 124. Somlyo AP, Somlyo AV (2003) Ca<sup>2+</sup> sensitivity of smooth muscle and non muscle myosin II: modulated by G proteins, kinases, and myosin phosphatase. Physiol Rev 83:1325–1358
- 125. Piascik MT, Kusiak JW, Barron KW (1990) Alpha1-adrenoceptor subtypes and the regulation of peripheral hemodynamics in the conscious rat. Eur J Pharmacol 186:273–278
- 126. Castillo EF, López RM, Rodríguez-Silverio J, Bobadilla RA, Castillo C (1998) Alpha 1D-adrenoceptors contribute to the neurogenic vasopressor response in pithed rats. Fundam Clin Pharmacol 12:584–589
- 127. López-Guerrero JJ, Ibarra M, Villalobos-Molina R (2005) Postjunctional alpha1-adrenoceptors in the vasculature of the pithed mouse are of the alpha1A-subtype. Auton Autacoid Pharmacol 25:101–103
- 128. Rokosh DG, Simpson PC (2002) Knockout of the alpha 1A/Cadrenergic receptor subtype: the alpha 1A/C is expressed in resistance arteries and is required to maintain arterial blood pressure. Proc Natl Acad Sci USA 99:9474–9479

- 129. O'Connell TD, Ishizaka S, Nakamura A, Swigart PM, Rodrigo MC, Simpson GL, Cotecchia S, Rokosh DG, Grossman W, Foster E, Simpson PC (2003) The alpha(1A/C)- and alpha(1B)adrenergic receptors are required for physiological cardiac hypertrophy in the double-knockout mouse. J Clin Invest 111:1783–1791
- 130. Malberg JE, Seiden LS (1998) Small changes in ambient temperature cause large changes in 3, 4-methylenedioxymethamphetamine (MDMA)-induced serotonin neurotoxicity and core body temperature in the rat. J Neurosci 18:5086–5094
- 131. Green AR, O'Shea E, Saadat KS, Elliott JM, Colado MI (2005) Studies on the effect of MDMA ('ecstasy') on the body temperature of rats housed at different ambient room temperatures. Br J Pharmacol 146:306–312
- 132. Bexis S, Docherty JR (2008) Role of alpha(1)-adrenoceptor subtypes in the effects of methylenedioxy methamphetamine (MDMA) on body temperature in the mouse. Br J Pharmacol 153:591–597
- 133. Sprague JE, Brutcher RE, Mills EM, Caden D, Rusyniak DE (2004) Attenuation of 3, 4-methylenedioxymethamphatamine (MDMA, Ecstasy)-induced rhabdomyolysis with  $\alpha$ 1- plus  $\beta$ 3-adrenoceptor antagonists. Br J Pharmacol 142:667–670
- 134. Brahmadevara N, Shaw AM, MacDonald A (2004) Alphaladrenoceptor antagonist properties of CGP 12177A and other beta-adrenoceptor ligands: evidence against beta(3)- or atypical beta-adrenoceptors in rat aorta. Br J Pharmacol 142:781–787
- 135. Papay R, Gaivin R, Jha A, McCune DF, McGrath JC, Rodrigo MC, Simpson PC, Doze VA, Perez DM (2006) Localization of the mouse alpha1A-adrenergic receptor (AR) in the brain: alpha1AAR is expressed in neurons, GABAergic interneurons, and NG2 oligodendrocyte progenitors. J Comp Neurol 497:209–222
- 136. Papay R, Gaivin R, McCune DF, Rorabaugh BR, Macklin WB, McGrath JC, Perez DM (2004) Mouse alpha1B-adrenergic receptor is expressed in neurons and NG2 oligodendrocytes. J Comp Neurol 478:1–10
- 137. Zuscik MJ, Sands S, Ross SA, Waugh DJ, Gaivin RJ, Morilak D, Perez DM (2000) Overexpression of the alpha1B-adrenergic receptor causes apoptotic neurodegeneration: multiple system atrophy. Nat Med 6:1388–1394
- 138. Chen Q, Li DP, Pan HL (2006) Presynaptic alphal adrenergic receptors differentially regulate synaptic glutamate and GABA release to hypothalamic presympathetic neurons. J Pharmacol Exp Ther 316:733–742
- 139. Rump LC, Majewski H (1987) Modulation of noradrenaline release through alpha<sub>1</sub>- and alpha<sub>2</sub>-adrenoceptors in rat isolated kidney. J Cardiovasc Pharmacol 9:500–507
- 140. Shinozuka K, Kunitomo M, Bjur RA, Westfall DP, Hattori K (1995) Effect of methoxamine on noradrenaline release in the caudal artery of hypertensive rats. Clin Exp Pharmacol Physiol 22:S88–S90
- 141. Bognar TI, Baretti R, Fischer S, Veldet C, Fuder H (1990) Alpha-adrenoceptor mediated facilitation of acetylcholine release in rat perfused heart. J Pharmacol Exp Ther 254:702–710
- 142. Keast JR, Kawatani M, De Groat WC (1990) Sympathetic modulation of cholinergic transmission in cat vesical ganglia is mediated by alpha1- and alpha2-adrenoceptors. Am J Physiol 258:R44–R50
- 143. Yoshimura N, de Groat WC (1992) Patch clamp analysis of afferent and efferent neurons that innervate the urinary bladder of the rat. Soc Neurosci Abstr 18:126
- 144. Somogyi GT, Tanowitz M, de Groat WC (1995) Prejunctional facilitatory alpha-1 adrenoceptors in the rat urinary bladder. Br J Pharmacol 114:1710–1716
- 145. Wada T, Hasegawa Y, Ono H (1997) Characterization of alpha1-adrenoceptor subtypes in facilitation of rat spinal motoneuron activity. Eur J Pharmacol 340:45–52

- 146. Sladek CD, Song Z (2008) Regulation of vasopressin release by co-released neurotransmitters: mechanisms of purinergic and adrenergic synergism. Prog Brain Res 170:93–107
- 147. Yuan WX, Chen SR, Chen H, Pan HL (2009) Stimulation of alpha(1)-adrenoceptors reduces glutamatergic synaptic input from primary afferents through GABA(A) receptors and T-type Ca(2+) channels. Neuroscience 158:1616–1624
- 148. Herold S, Hecker C, Deitmer JW, Brockhaus J (2005) Alphaladrenergic modulation of synaptic input to Purkinje neurons in rat cerebellar brain slices. J Neurosci Res 82:571–579
- Araneda RC, Firestein S (2006) Adrenergic enhancement of inhibitory transmission in the accessory olfactory bulb. J Neurosci 26:3292–3298
- Li Y, van den Pol AN (2005) Direct and indirect inhibition by catecholamines of hypocretin/orexin neurons. J Neurosci 25:173–183
- 151. Somogyi GT, Tanowitz M, Zernova G, de Groat WC (1996)  $M_1$  muscarinic receptor-induced facilitation of ACh and noradrenaline release in the rat bladder is mediated by protein kinase C. J Physiol 496:245–254