Review

Rubinstein-Taybi syndrome: molecular findings and therapeutic approaches to improve cognitive dysfunction

T. M. Hallam and R. Bourtchouladze*

Helicon Therapeutics, Inc., One Bioscience Park Drive, Farmingdale, New York 11735 (USA), Fax: +1 631 370 8846, e-mail: rb@helicontherapeutics.com

Received 18 November 2005; received after revision 22 February 2006; accepted 18 April 2006 Online First 19 June 2006

Abstract. Rubinstein-Taybi syndrome (RTS) is a rare human genetic disorder characterized by mental retardation and physical abnormalities. Many RTS patients have a genetic mutation which has been mapped to chromosome 16p13.3, a genomic region encoding cyclic AMP (cAMP) response element binding protein (CREB) binding protein (CBP). CBP is a transcriptional co-activator that binds to CREB when the latter is phosphorylated and promotes gene transcription. CREB-dependent gene transcription has been shown to underlie long-term memory formation.

In this review we will focus on recent findings regarding the biology of CBP and its role in memory formation and cognitive dysfunction in RTS. We will also review the role of CBP in other neurological disorders, including Alzheimer's disease, Huntington's disease and amyotrophic lateral sclerosis. Finally, we will discuss novel therapeutic approaches targeted to CBP/CREB function for treating the cognitive dysfunction of RTS and other neurological disorders.

Keywords. Rubinstein-Taybi Syndrome (RTS), cAMP-responsive element binding protein (CREB), CREB-binding protein (CBP), phosphodiesterase inhibitor 4 (PDE4), long-term memory, histone acetyltransferase (HAT), histone deacetylase inhibitor (HDAC).

Rubinstein-Taybi syndrome: What is it?

Rubinstein-Taybi syndrome (RTS) is a human genetic disorder characterized by mental retardation and physical abnormalities including broad thumbs, big and broad toes, short stature and craniofacial anomalies such as beaked nose, down-slanting palpebral fissures and hypoplastic maxilla [1–3]. RTS was first described by Rubinstein and Taybi in 1963. The first published case of RTS, however, was reported by Michail et al. (1957) from Athens, who described a 7-year-old boy with a disorder that retrospectively looks like RTS [4].

Compared with other genetic disorders with cognitive dysfunction (e.g. Down syndrome), RTS is a relatively rare syndrome. It occurs in about 1 in 125,000 to 1 in

720,000 births and accounts for as many as 1 in 300 cases of institutionalized mentally retarded patients. RTS affects males and females equally. The exact etiology of RTS is not established, and as of yet there is no indication for environmental causes, although they cannot be excluded [5]. The existing familial data indicating rare recurrence in siblings suggests that most mutations occur de novo. Such data are difficult to accumulate, however, because most RTS patients do not reproduce. In this regard, the recent report by Hennekam et al. of RTS in a mother and son provides important support for dominant inheritance of this disorder [5].

Although the exact molecular-genetic etiology of RTS is not clearly understood, several groups of researchers established that RTS is associated with breakpoints, mutations and microdeletions of chromosome 16p13.3 [6–13]. In 1995, Petrij et al. first reported that mutations in the

^{*} Corresponding author.

gene encoding the cyclic AMP (cAMP)-response element binding protein (CREB) binding protein (CBP), which is located on chromosome 16p13.3, could be causative of RTS [10]. Furthermore, these authors showed that RTS results not only from gross chromosomal rearrangement(s) of chromosome 16p, but also from heterozygous point mutations in the CBP gene itself, suggesting that the loss of one functional copy of the CBP gene underlies the developmental abnormalities and mental retardation in RTS patients [10]. A follow-up study of 194 patients with clinical features of RTS showed that microdeletions and truncating mutations in CBP account for approximately 20% of the mutations in individuals with RTS [11]. However, subsequent DNA sequencing of the CBP gene revealed pathogenic mutations in 56% of patients with RTS, a higher rate than previously detected [12]. More recently, Roelfsema et al. sequenced the CBP and EP300 genes (the latter shares some functional and sequence homology with CBP) in 92 patients with RTS and found 3 patients with inactivating mutations in EP300 [13]. This is the first gene other than CBP which has been linked to RTS. Taken together these data support the concept that RTS is a genetically heterogeneous disorder which may be caused by mutations in CBP as well as mutations in other genes in up to 50% of the cases.

Although CBP loss of function is a common feature in RTS patients, they nonetheless display a wide variation in the severity of mental retardation and other clinical manifestations. The intelligence level of RTS patients is usually very low, with an average IQ of 34 by aged 25 [2]. However, some individuals have higher scores and less profound developmental defects. These differences could be due to the heterogeneity of mutations in the CBP gene observed in RTS patients or to heterogeneity in genetic background. To date, two models of how CBP mutations may result in RTS have been proposed: (i) haploinsufficiency and (ii) dominant negative effects. For haploinsufficiency, two functional copies of the gene are required to produce sufficient protein for normal development and function, while for a dominant-negative mechanism, abnormal product derived from the mutant allele inhibits the wild-type product. To date, evidence supporting both mechanisms has been reported. For example, the haploinsufficiency mechanism is supported by observations that both truncated mutants and null mutants can cause human RTS [11, 12]. Alternatively, the findings that microinjection of the CBP-CREB binding domain into fibroblasts blocks transcriptional activation of a CRE-lacZ reporter gene support the dominant negative inhibiting mechanism of RTS [14, 15]. Taken together with evidence from studies of genetically engineered mouse models of RTS (see below), it seems likely that both mechanisms of CBP dysfunction can play a role in the development of RTS phenotypes.

Biology of CBP and cognition

CBP was first described in 1993 by Chrivia et al. as a nuclear transcription coactivator that binds specifically to CREB when it is phosphorylated by protein kinase A (PKA) [16]. CBP is a large nuclear phosphoprotein consisting of 2442 amino acids with a molecular mass of approximately 250 kDa that comprises several different domains: three cysteine-histidine-rich domains, a bromodomain, a PKA phosphorylation site, two zinc finger motifs, an N-terminal nuclear receptor binding domain, a C-terminal TAD and a histone acetyltransferase (HAT) domain [9]. Having many identified protein-interacting domains, CBP can function as a link and a signal integrator between the basal transcription machinery and certain DNA-binding factors, namely CREB, Jun, Fos and NFkB (for reviews see [17–20]). As a binding partner, CBP is a focal point for many molecular pathways. As such, CBP plays an important role in the complex biological processes that affect cell growth, transformation, development and neuronal plasticity (for reviews see [17, 19, 21-24]).

What are the biological mechanisms whereby dysfunction in CBP could cause the cognitive deficits in RTS? Biologically, this can be accomplished through disruption of either of the two main functions of CBP. First, CBP acts as a nuclear transcriptional co-activator that binds to the activated form of the CREB protein [16, 25, 26], and second, CBP acts as a HAT [19, 20, 27–30] (Fig. 1). In neurons, CBP can be activated by several kinases including (PKA), calcium/calmodulin-dependent kinase IV (CaMKIV) and mitogen-activated protein kinases (MAPK), suggesting that it is an important target of multiple signaling pathways [31–36]. Importantly, these signaling pathways (PKA, MAPK and CaMKIV) play a dominant role in activation of CREB in various physiological processes, including learning and memory [37– 49]. CREB phosphorylation enables its interaction with CBP and links kinase pathways driven by neuronal activity to transcription of genes that are important for learning and memory. Consequently, acting as a co-activator, CBP enhances the ability of phosphorylated CREB to activate transcription of cAMP-responsive genes. CREB and CRE-dependent genes have been established to underlie long-term memory formation in several invertebrate and vertebrate species [39, 50–60], raising the possibility that cognitive deficits in RTS patients may derive from impaired CBP function during long-term memory formation [61, 62].

Another important function of CBP is its HAT activity [27, 28]. Histone acetylation relates to the transcriptional availability of chromatin [63]. Specifically, hyperacetylated histones accumulate within transcriptionally active regions of the chromatin, and hypoacetylated histones accumulate within transcriptionally silent regions. His-

1727

tone acetylation disrupts repressive chromatin structure and allows for transcription apparently via three mechanisms: (i) it promotes transcription factor access to the DNA; (ii) it weakens inter-nucleasomal interactions, destabilizing higher-order chromatin structure; and (iii) it promotes the progression of RNA polymerase during transcription. In essence, when histones are acetylated, the DNA becomes 'unwound' from the histones and facilitates the binding of transcription complexes to DNA [63] (Fig. 1).

The HAT domain of CBP resides in the central region of the protein, and both the N- and the C-terminal regions can activate transcription. This modular organization allows CBP to provide a physical bridge for assembly of transcription co-activator complexes. Taken together, it is plausible that CBP plays a dual role in transcription activation: (i) as a bridging protein interacting directly with CREB and other transcription factors; and (ii) as a HAT that contributes to transcription by acetylating histones, thereby disrupting repressive chromatin structure [19] (Fig. 1). Hence, it is likely that CBP dysfunction in RTS may interfere with the transcriptional machinery and activation of downstream genes by reducing the functional

availability of CREB and/or by disrupting HAT-induced chromatin remodeling.

While considering the role of CBP in the cognitive dysfunction of RTS, it is important to emphasize its critical role in development. Evolutionary CBP is a highly conserved protein sharing about 95% similarity between human and mouse. CBP orthologs have been found in a number of different organisms, including Drosophila melanogaster, Caenorhabditis elegans and Arabidopsis thaliana. Numerous studies have documented the involvement of CBP in embryogenesis and developmental processes, particularly in model systems such as Drosophila and mouse (for excellent reviews, see [21, 22]). At almost every stage of development CBP takes on different binding partners and expression patterns that affect cellular patterning and differentiation. During development many molecular pathways in vertebrates and invertebrates interact directly with CBP/CREB and/or are regulated by CBP/CREB. Thus far, however, it has been difficult to clearly link a specific defect in CBP during development to mental retardation observed in adult RTS patients. To date, we can only speculate that some of the developmental defects in CBP function con-

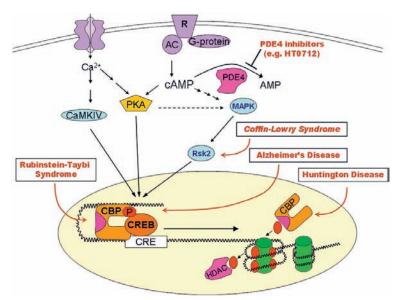


Figure 1. Biology of CBP and heritable forms of cognitive disorders. Molecular genetic mutations have been identified for various forms of heritable cognitive disorders. Interestingly, most of these mutations correspond to biochemical components of memory formation. Memory formation begins with the influx of Ca²⁺ through NMDA receptors and/or the activation of adenylyl cyclase via neuromodulators (e.g. dopamine). In turn, this causes a rise in cAMP levels that activates PKA and other kinases (e.g. MAPK and CaMKIV). The cAMP-signaling cascade reaches the nucleus, where its components (e.g. PKA, RSK2) phosphorylate transcription factors such as CREB. When CREB is phosphorylated, CBP binds to it, thereby promoting changes in gene expression important for long-term memory formation. Newly synthesized proteins result in long-term changes in cell function, such as growth and maturation of synaptic connections. To date, several molecular mutations have been identified for various inherited cognitive disorders, including Rubinstein-Taybi syndrome, Coffin-Lowry syndrome, Huntington's disease and Alzheimer's disease, all of which are consistent with disruptions of the biochemistry of brain plasticity and memory formation (also see, Weeber and Sweatt, 2002; Tully et al., 2003) [58, 62]. Recent evidence suggests that pharmacological treatments that enhance signaling in the CREB/CBP pathway (e.g. PDE4 inhibitors) can rescue memory deficits in animal models of RTS and AD [59, 72, 113]. (AC, adenylyl cyclase; AMP, adenosine monophosphate; CaMKIV, calcium-calmodulin-dependent protein kinase IV; CBP, cAMP-response-element-binding protein (CREB)-binding protein; HDAC, histone deacetylase; MAPK, mitogen-activated protein kinase; NMDA, N-methyl-D-aspartate; PDE4, phosphodiesterase 4; PKA, protein kinase A; RSK2, ribosomal S6 kinase).

tribute to the mental retardation in RTS patients (for our hypothesis see below).

Animal models of RTS: rescuing long-term memory deficits

The evidence that RTS is a consequence of reduced levels of CBP is based on RTS cases resulting from heterozygous defects of CBP. Indeed, many RTS patients are heterozygous for CBP mutations which yield truncations of the CBP C-terminus, suggesting that a dominant-negative mechanism may contribute to the clinical symptoms. As we described above, CBP is a transcriptional co-activator that binds to CREB when the latter is phosphorylated and facilitates gene transcription. Truncated CBP would bind to phospho-CREB but would not be able to activate transcription. CREB-dependent gene expression has been shown to underlie long-term memory formation in multiple species, leading to the intriguing speculation that the mental retardation in RTS patients may derive from reduced CBP function during long-term memory formation [61]. To this end, Oike et al. [15] generated a C-terminal truncation mutation in mouse CBP, which appears to act in a dominant-negative

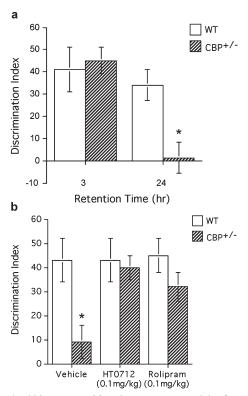


Figure 2. Object recognition in a mouse model of RTS. (*a*) Long-term (24 h) but not short-term (3 h) memory is impaired in CBP+/- mutant mice. (*b*) PDE4 inhibitors (HT-0712 and rolipram) ameliorate the long-term memory defect of CBP+/- mutant mice (* = p < 0.05 CBP+/- vs. wild-type mice) (adapted from [59]).

fashion to recapitulate many of the clinical features observed in RTS patients. Homozygous CBP-/- mutants are embryonic lethal, while heterozygous CBP+/- mice show reduced viability and growth retardation. In their initial study, Oike and colleagues showed that CBP+/- mice have normal learning and short-term memory but defective long-term memory for two passive avoidance tasks. These findings provided the first behavioral evidence of cognitive dysfunction in a mouse model of RTS, substantiating the notion that normal CBP function is required for memory formation.

We extended the observations of Oike et al. to a novel object recognition task, an ethologically relevant and nonaversive task that relies on a mouse's natural exploratory behavior [59]. Normal mice can remember having explored an object and show a preference for exploring novel objects. During training for this task, mice are presented with two identical objects, which they explore for some time by orienting toward, sniffing and crawling over. To test for memory of this experience, mice are presented at a later time with two different objects, one of which was previously explored and thus is 'familiar', and the other of which is novel. If the mouse remembers the familiar object, it will spend more time exploring the novel object.

We found that CBP*/- mutant mice have impaired long-term memory, but normal short-term memory for novel object recognition (Fig. 2a). These findings corroborate the observations of Oike et al. and establish that mutations in CBP can yield specific defects in long-term memory formation [15, 59].

Because CBP/CREB function was reduced but not eliminated in these mice, the possibility existed to improve long-term memory formation by enhancing upstream signaling onto CBP/CREB. To this end, we examined two phosphodiesterase 4 (PDE4) inhibitors, the prototypical PDE4 inhibitor rolipram and a novel one, HT0712, for their ability to abolish memory deficits in CBP+/- mice [59]. Indeed, as predicted, we found that a single dose of either rolipram or HT0712 delivered shortly before training was able to restore long-term memory to normal levels in CBP+/- mice (Fig. 2b). To address whether the drug's effects were specific to the molecular lesion in CBP, we reduced the duration of training so that wild-type mice showed almost no memory. We then gave increasing doses of the HT0712 until we achieved maximal long-term memory. We reasoned that because CBP+/- mice had less functional CBP than wild-type mice, they would require higher concentrations of the drug to respond comparably to wild-type mice. Indeed, we discovered that CBP+/- mice had a rightward shift in the dose response curve compared with wild-type mice. These findings provide strong support for the molecular specificity of the drug action [59] (Fig. 3).

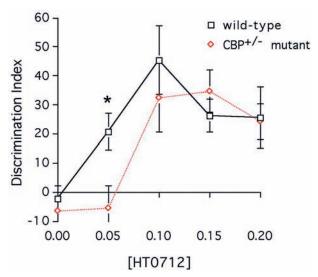


Figure 3. Pharmacological rescue of long-term memory deficit in CBP+/- mutants is dose-dependent. HT0712 dose sensitivity is decreased in CBP+/- mutant mice (* = p < 0.05 CBP+/- mice vs. wild-type controls) (adapted from [59]).

Our findings provided the first demonstration that the memory impairments of CBP +/- mutants can be ameliorated by inhibitors of PDE4. These PDE inhibitors likely enhance signaling to CREB during memory formation by increasing the magnitude and/or duration of cAMP levels in response to experience-dependent changes in neural activity [43, 64–68]. This effect is specific molecularly, because CBP+/- mutants require a higher dose of HT0712 than wild-type mice to produce an equivalent amount of enhanced memory. This result was expected since the genetic lesion of CBP+/- mice likely reduces the levels CBP/CREB available for functional activation via cAMP signaling. Hence, greater upstream enhancement is required for a comparable effect on downstream targets [59]. Given that CBP interacts with several transcription factors [17–20, 69, 70], our results strongly suggest CBP/ CREB to be the relevant interaction for long-term memory formation. Because of the observed molecular and pathological similarities between these CBP+/- mice and patients with Rubinstein-Taybi syndrome, we hypothesize that the mental retardation of RTS patients likely results, at least in part, from functional defects in longterm memory formation. Further, PDE4 inhibitors may ameliorate this biochemical block on memory formation, thereby rendering RTS patient capable of benefiting from training and experience.

Another model for RTS, a null allele mouse modeling the haploinsufficiency mechanism of RTS, was generated by Tanaka and colleagues [71]. Similar to the mice we studied, homozygous CBP mutants are embryonic lethal. Unlike our mice, however, fewer of those mutants (7 out of 21 mice studied) exhibit phenotypes resembling the clinical features of RTS [71], making these heterozygous

mice overall a less favorable model for RTS. In their original study Tanaka and colleagues did not perform behavioral analysis of these CBP+/- mutant mice. However, recently Alarcon and colleagues tested these mice in the similar fear based and object recognition tasks as we and Oike et al. did previously [59, 72]. In agreement with our findings, Alarcon et al. found that CBP+/- mutant mice have normal short-term memory but deficient long-term memory in the object recognition and contextual memory tasks. In addition to their behavioral findings, Alarcon et al. also showed that long-term potentiation (LTP), a form of synaptic plasticity that is thought to underlie the storage of some types of memory in the hippocampus, was deficient in CBP+/- mutant mice. This deficiency in LTP was abolished with the PDE4 inhibitor rolipram or by overexpressing a constitutively-active form of CREB [72]. These findings confirmed and strengthen our original hypothesis that PDE4 inhibitors would improve longterm memory formation by enhancing upstream signaling onto CBP/CREB.

Alarcon and colleagues also found reduced HAT activity in their mutant mice which raised the possibility that such molecular mechanism may also contribute to the observed memory deficits in these mice. To determine whether this was really the case, they treated animals with histone deacetylase (HDAC) inhibitor SAHA prior to training in contextual fear conditioning. The HDAC inhibitor reversed the memory deficit observed in contextual conditioning [72]. These findings showed that enhancing CREB function and/or increasing the HAT activity of CBP may be efficacious for treating RTS.

More recently, two additional groups of researchers generated mouse models of RTS in which the CBP trangene expression was restricted to the forebrain [73, 74]. The molecular techniques used by these authors allow for a mutation to be spatially restricted to neurons within the hippocampus, striatum, amygdala and cortex, as well as for a transgene to be temporally expressed postnatally. Both groups reported that CBP mutations restricted to the forebrain disrupt formation of long-term memory. These studies confirmed our and Alarcon's et al. findings and provide further support for the role of CBP in hippocampal dependent memory formation in the adult organism.

In summary, the present findings demonstrate that CBP/CREB likely function together as a molecular switch during long-term memory formation, and mutations in CBP can be detrimental to this process. The current evidence indicates that it is possible to overcome the deficits in long-term memory by enhancing CREB/CBP signaling with PDE4 inhibitors or with HDAC inhibitors. These findings suggest that PDE4 and HDAC inhibitors may prove to be novel therapeutics for treating cognitive dysfunction in RTS patients.

CBP in neurological diseases

In the past few years considerable progress has been made in revealing the role of CBP/CREB function not only in RTS, but also in the molecular etiology of other human neurological disorders, such as Huntington's disease, Alzheimer's disease and amyotrophic lateral sclerosis.

Polyglutamine diseases

Huntington's disease (HD) belongs to a family of neurodegenerative diseases including (i) spinal bulbar muscular atrophy (SBMA), (ii) (iii) dentatorubral and pallidoluysian atrophy (DRPLA) and (iv) spino-cerebellar ataxia (SCA) that are characterized by expanding CAG repeats coding for polyglutamine (polyQ) (for reviews see [75, 76]). Although there is no unambiguous single proteinprotein interaction that is responsible for the pathology of these diseases, there are two common factors connecting them. First, all are characterized by selective neuronal degeneration in specific regions of the CNS. Second, all these diseases contain expanding CAG repeats encoding for multiple copies of glutamine (polyglutamine [polyQ]) in the genome. Converging evidence from Drosophila [77-84], mice [85-87] and humans [87] suggests that polyQ aggregates cause neurotoxicity depending on where the polyQ is expressed. Here we focus on HD as a representative model on how polyQ can lead to CBP/ CREB dysfunction and neuronal death.

HD is an autosomal-dominant neurodegenerative disease caused by CAG trinucleotide repeats in exon 1 of the huntingtin (htt) gene. Normally, the htt protein is found predominately in the cytoplasm. In the mutant, however, htt is localized mainly in the nucleus where it forms aggregates of the mutant protein. These polyQ aggregates have been shown to bind to a number of transcription factors and co-factors, resulting in their functional impairment (reviewed in [88]). In particular, much of the attention has been on the functional disruption of CBP by the binding of polyQ to the HAT domain [82, 89, 90]. This decreases soluble levels of CBP [91]. The loss of accessible CBP leads to a transcriptional dysfunction of CBP/CREB-mediated gene expression [87, 88, 92-96] and a subsequent decline in neuronal survival [90]. In addition, the binding of polyQ to CBP results in a decrease in histone acetylation, which further restricts CBP/CREB transcription. Combined, these effects result in a lack of pro-survival signals, leading to neuronal death [87, 91, 94, 95, 97]. Converging evidence from recent studies suggests that biochemical interventions which facilitate CBP/CREB function can ameliorate the effects of mutant htt protein. For example, Nucifora et al. demonstrated that CBP was depleted from its normal nuclear location and was present in polyQ aggregates in a neuronal cell culture model of HD, a transgenic mouse model of HD and Huntington's disease in humans [87]. Expanded polyQ repeats inter-

fere with CBP-activated gene transcription. Importantly, Nucifora et al. found that by overexpressing CBP they could rescue CBP reporter gene expression and ameliorate polyQ-mediated toxicity [87]. In a second study, Wyttenbach et al. demonstrated a decrease in CRE-mediated transcription in polyQ transfected cells. By amplifying CREB/CBP signaling with cAMP or forskolin (which constitutively activates adenylyl cyclase to produce cAMP), they could partially rescue the CRE-mediated transcription [94]. Finally, Sugars et al. reported that overexpression of a transcriptionally active form of CREB rescued CRE-mediated transcription, reduced polyQ aggregation and protected against polyQ-induced cell death [95]. Additional evidence supporting CBP dysfunction comes from the correlation of hypoacetylation with neuronal degeneration in polyQ diseases [82, 84, 98]. For example, reversal of hypoacetylation by either CBP overexpression or HDAC inhibition rescues cell loss [82, 84] and extends animal survival in a dose-dependent manner in vivo [98, 99]. Taken together these findings indicate that modifying CBP/CREB function may provide a potential therapeutic approach for treating polyQ diseases such as HD.

CBP in Alzheimer's

Alzheimer's disease (AD) is an age-associated neurodegenerative disease characterized by mild cognitive impairment at its onset. During the later stages, the disease progresses to severe deficits in multiple memory systems accompanied by changes in personality and cognitive decline. Current evidence suggests that AD begins as a disorder of synaptic function and plasticity (for reviews [100–103]). Over time AD progresses to mass neuronal cell death and irreversible brain damage. The initial synaptic dysfunction is associated with the deposition of the amyloid β -peptide (A β), particularly its 42-aa form $(A\beta_{1-42})$ [104–110]. $A\beta_{1-42}$ can disrupt synaptic function [107] and CREB transcription at very low (non-toxic) concentrations [111]. A β is a protein fragment cleaved from a larger protein called amyloid precursor protein (APP) during metabolism and is implicated in CBP/ CREB dysfunction. The final step in the generation of A β from APP is proteolysis by gamma-secretase. The most common cause of familial early onset AD is the result of mutations in the genes encoding APP and presenilins 1 and 2 (PS-1 and -2), the latter of which alters gammasecretase activity to increase $A\beta$ production. Although the precise mechanisms by which APP and PS mutations increase $A\beta$ deposition and contribute to the pathology of the disease remain to be determined, recent studies have demonstrated that mutations in APP and PS1 affect CREB/CBP function [66, 110–114].

Although there is eventual synaptic loss in mouse models of AD, the deficits in memory formation and LTP precede

synaptic depletion [106]. This suggests that biochemical changes occur before synapse and neuronal loss. Recently Vitolo and colleagues described an important link between $A\beta$ deposition and CREB dysfunction in cultured hippocampal neurons [66]. They observed a significant decrease in PKA activity in hippocampal neurons following $A\beta_{1-42}$ administration. They also found that pretreatment with $A\beta_{1-42}$ blocked the increase in CREB activation following glutamate stimulation. This $A\beta_{1-42}$ -induced decrease in PKA/CREB signaling also coincides with a decrease in LTP. Importantly, Vitolo and colleagues were able to rescue CREB activity in hippocampal cultures and LTP in hippocampal slices with the administration of the PDE4 inhibitor rolipram [66].

In addition to evidence of CREB/CBP dysfunction in vitro, recent studies on APP and PS1 double-transgenic mice also have pointed to dysfunction of CREB/CBP in vivo [113, 114]. APP and PS1 double-transgenic mice display progressive memory loss and A β deposition [115–118]. Recently, Gong et al. used rolipram to magnify cAMP/CREB signaling in their double transgenic animals [113]. They found that rolipram ameliorated both LTP and memory deficits in this mouse model of AD [113]. Importantly, this effect was long-lasting: the transgenic mice treated with rolipram at 3 months of age had normal contextual memory and normal levels of CREB at 7-8 months of age, while vehicle-treated mice did not [113]. In another study, Saura et al. evaluated PS1/2 conditional knockout mice in which PS1/2 is selectively eliminated in excitatory neurons in the forebrain [114]. The conditional knockout mice had LTP and memory deficits that were amplified with age. These mice had decreased levels of CBP leading to reduced transcription of CBP/CREB genes, such as c-fos and BDNF [114]. Taken together, these studies implicate CREB/CBP in AD and suggest the potential for therapeutic strategies targeted to CBP/CREB in age-related memory impairment.

CBP in amyotrophic lateral sclerosis

ALS, also called Lou Gherig's disease, is a neurodegenerative disease that is characterized by progressive degeneration of motor neurons in the brain and spinal cord. The loss of motor neurons results in muscle weakening and eventually paralysis and death. ALS has both sporadic and familial forms. Familial ALS is linked with a genetic mutation in the Cu²⁺/Zn²⁺ superoxide dismutase (SOD-1) gene and accounts for approximately 5–10% of all ALS cases (for reviews [119, 120]). The symptoms, pathology and progression of familial ALS are similar to the sporadic form, which suggests similar mechanisms of neurodegeneration. Several mouse strains with mutations in the SOD-1 gene have been generated as a mouse model of ALS (reviewed in [121]). These mouse models of ALS display motor neuron degeneration which is accompanied

by motor deficits and eventual death. The progression of the disease in the mouse model closely follows that of the human disease [122]. Recently, Rouaux et al. found reduced CBP levels in protein extracts prepared from lumbar spinal cord of transgenic SOD-1G86R mice [123]. In addition, histone acetylation was decreased in motor neuron nuclei in the spinal cord of SOD-1G86R mice compared with wild-type littermates [123]. Together, these studies provide support that CBP/CREB signaling is an important component of neuronal function and survival in ALS [24, 123].

Summary

Rubinstein-Taybi syndrome, Huntington's disease, Alzheimer's disease and amyotrophic lateral sclerosis all share impaired CBP/CREB function. Taken together, a pattern emerges that suggests that CBP/CREB dysfunction may be an important factor in multiple neuronal diseases. It is still unknown whether impaired CBP/CREB function is the cause of these neuronal diseases or a convergent downstream effector of the initiating diseases. However, because multiple signaling mechanisms converge onto the CREB/CBP molecular switch, the upstream components of these signaling pathways are emerging as therapeutic targets for drug intervention. Further research on molecular lesions associated with neuronal disorders will elucidate the molecular mechanisms of their pathologies. Such work promises to yield important new insights not only into the neurobiology of brain function but also to novel therapeutics for treating patients.

Acknowledgements. We would like to thank Marco Peters, Kellie McCormick-Hallam, Tim Tully and John Tallman for their comments on this manuscript.

- Rubinstein, J. H. and Taybi, H. (1963) Broad thumbs and toes and facial abnormalities. A possible mental retardation syndrome. Am. J. Dis. Child. 105, 588–608.
- 2 Hennekam, R. C., Baselier, A. C., Beyaert, E., Bos, A., Blok, J. B., Jansma, H. B., Thorbecke-Nilsen, V. V. and Veerman, H. (1992) Psychological and speech studies in Rubinstein-Taybi syndrome. Am. J. Ment. Retard. 96, 645–660.
- 3 Cantani, A. and Gagliesi, D. (1998) Rubinstein-Taybi syndrome. Review of 732 cases and analysis of the typical traits. Eur. Rev. Med. Pharmacol. Sci. 2, 81–87.
- 4 Michail, J., Matsoukas, J. and Theodorou, S. (1957) Pouce bot argue en forte abduction-extension et autres symptomes concomitants. Revue de chirurgie orthopédique et réparatrice de l'appareil locomoteur 43, 142–146.
- 5 Hennekam, R. C., Stevens, C. A. and Van de Kamp, J. J. (1990) Etiology and recurrence risk in Rubinstein-Taybi syndrome. Am. J. Med. Genet. Suppl. 6, 56–64.
- 6 Lacombe, D., Saura, R., Taine, L. and Battin, J. (1992) Confirmation of assignment of a locus for Rubinstein-Taybi syndrome gene to 16p13.3. Am. J. Med. Genet. 44, 126–128.
- 7 Tommerup, N., van der Hagen, C. B. and Heiberg, A. (1992) Tentative assignment of a locus for Rubinstein-Taybi syndrome to 16p13.3 by a de novo reciprocal translocation, t(7;16)(q34;p13.3). Am. J. Med. Genet. 44, 237–241.

- 8 Breuning, M. H., Dauwerse, H. G., Fugazza, G., Saris, J. J., Spruit, L., Wijnen, H., Tommerup, N., van der Hagen, C. B., Imaizumi, K., Kuroki, Y. et al. (1993) Rubinstein-Taybi syndrome caused by submicroscopic deletions within 16p13.3. Am. J. Hum. Genet. 52, 249–254.
- 9 Giles, R. H., Petrij, F., Dauwerse, H. G., den Hollander, A. I., Lushnikova, T., van Ommen, G. J., Goodman, R. H., Deaven, L. L., Doggett, N. A., Peters, D. J. and Breuning, M. H. (1997) Construction of a 1.2-Mb contig surrounding, and molecular analysis of, the human CREB-binding protein (CBP/CREBBP) gene on chromosome 16p13.3. Genomics 42, 96–114.
- 10 Petrij, F., Giles, R. H., Dauwerse, H. G., Saris, J. J., Hennekam, R. C., Masuno, M., Tommerup, N., van Ommen, G. J., Goodman, R. H., Peters, D. J. et al. (1995) Rubinstein-Taybi syndrome caused by mutations in the transcriptional co-activator CBP. Nature 376, 348–351.
- 11 Petrij, F., Dauwerse, H. G., Blough, R. I., Giles, R. H., van der Smagt, J. J., Wallerstein, R., Maaswinkel-Mooy, P. D., van Karnebeek, C. D., van Ommen, G. J., van Haeringen, A., Rubinstein, J. H., Saal, H. M., Hennekam, R. C., Peters, D. J. and Breuning, M. H. (2000) Diagnostic analysis of the Rubinstein-Taybi syndrome: five cosmids should be used for microdeletion detection and low number of protein truncating mutations. J. Med. Genet. 37, 168–176.
- 12 Bartsch, O., Schmidt, S., Richter, M., Morlot, S., Seemanova, E., Wiebe, G. and Rasi, S. (2005) DNA sequencing of CREBBP demonstrates mutations in 56% of patients with Rubinstein-Taybi syndrome (RSTS) and in another patient with incomplete RSTS. Hum. Genet. 117, 485–493.
- 13 Roelfsema, J. H., White, S. J., Ariyurek, Y., Bartholdi, D., Niedrist, D., Papadia, F., Bacino, C. A., den Dunnen, J. T., van Ommen, G. J., Breuning, M. H., Hennekam, R. C. and Peters, D. J. (2005) Genetic heterogeneity in Rubinstein-Taybi syndrome: mutations in both the CBP and EP300 genes cause disease. Am. J. Hum. Genet. 76, 572–580.
- 14 Parker, D., Ferreri, K., Nakajima, T., LaMorte, V. J., Evans, R., Koerber, S. C., Hoeger, C. and Montminy, M. R. (1996) Phosphorylation of CREB at Ser-133 induces complex formation with CREB-binding protein via a direct mechanism. Mol. Cell. Biol. 16, 694–703.
- 15 Oike, Y., Hata, A., Mamiya, T., Kaname, T., Noda, Y., Suzuki, M., Yasue, H., Nabeshima, T., Araki, K. and Yamamura, K. (1999) Truncated CBP protein leads to classical Rubinstein-Taybi syndrome phenotypes in mice: implications for a dominant-negative mechanism. Hum. Mol. Genet. 8, 387–396.
- 16 Chrivia, J. C., Kwok, R. P., Lamb, N., Hagiwara, M., Montminy, M. R. and Goodman, R. H. (1993) Phosphorylated CREB binds specifically to the nuclear protein CBP. Nature 365, 855–859.
- 17 Janknecht, R. and Hunter, T. (1996) Versatile molecular glue. Transcriptional control. Curr. Biol. 6, 951–954.
- 18 Giles, R. H., Peters, D. J. and Breuning, M. H. (1998) Conjunction dysfunction: CBP/p300 in human disease. Trends Genet. 14, 178–183.
- 19 Chan, H. M. and La Thangue, N. B. (2001) p300/CBP proteins: HATs for transcriptional bridges and scaffolds. J. Cell. Sci. 114, 2363–2373.
- 20 Kalkhoven, E. (2004) CBP and p300: HATs for different occasions. Biochem. Pharmacol. 68, 1145–1155.
- 21 Goodman, R. H. and Smolik, S. (2000) CBP/p300 in cell growth, transformation, and development. Genes Dev. 14, 1553–1577.
- 22 McManus, K. J. and Hendzel, M. J. (2001) CBP, a transcriptional coactivator and acetyltransferase. Biochem. Cell. Biol. 79, 253–266.
- 23 Janknecht, R. (2002) The versatile functions of the transcriptional coactivators p300 and CBP and their roles in disease. Histol. Histopathol. 17, 657–668.

- 24 Rouaux, C., Loeffler, J. P. and Boutillier, A. L. (2004) Targeting CREB-binding protein (CBP) loss of function as a therapeutic strategy in neurological disorders. Biochem. Pharmacol. 68, 1157–1164.
- 25 Kwok, R. P., Lundblad, J. R., Chrivia, J. C., Richards, J. P., Bachinger, H. P., Brennan, R. G., Roberts, S. G., Green, M. R. and Goodman, R. H. (1994) Nuclear protein CBP is a coactivator for the transcription factor CREB. Nature 370, 223–226.
- 26 Cardinaux, J. R., Notis, J. C., Zhang, Q., Vo, N., Craig, J. C., Fass, D. M., Brennan, R. G. and Goodman, R. H. (2000) Recruitment of CREB binding protein is sufficient for CREB-mediated gene activation. Mol. Cell. Biol. 20, 1546–1552.
- 27 Bannister, A. J. and Kouzarides, T. (1996) The CBP co-activator is a histone acetyltransferase. Nature 384, 641–643.
- 28 Ogryzko, V. V., Schiltz, R. L., Russanova, V., Howard, B. H. and Nakatani, Y. (1996) The transcriptional coactivators p300 and CBP are histone acetyltransferases. Cell 87, 953–959.
- 29 Korzus, E., Torchia, J., Rose, D. W., Xu, L., Kurokawa, R., McInerney, E. M., Mullen, T. M., Glass, C. K. and Rosenfeld, M. G. (1998) Transcription factor-specific requirements for coactivators and their acetyltransferase functions. Science 279, 703–707.
- 30 Kouzarides, T. (2000) Acetylation: a regulatory modification to rival phosphorylation? EMBO J. 19, 1176–1179.
- 31 Janknecht, R. and Nordheim, A. (1996) MAP kinase-dependent transcriptional coactivation by Elk-1 and its cofactor CBP. Biochem. Biophys. Res. Commun. 228, 831–837.
- 32 Swope, D. L., Mueller, C. L. and Chrivia, J. C. (1996) CREB-binding protein activates transcription through multiple domains. J. Biol. Chem. 271, 28138–28145.
- 33 Chawla, S., Hardingham, G. E., Quinn, D. R. and Bading, H. (1998) CBP: a signal-regulated transcriptional coactivator controlled by nuclear calcium and CaM kinase IV. Science 281, 1505–1509.
- 34 Liu, Y. Z., Chrivia, J. C. and Latchman, D. S. (1998) Nerve growth factor up-regulates the transcriptional activity of CBP through activation of the p42/p44(MAPK) cascade. J. Biol. Chem. 273, 32400–32407.
- 35 Ait-Si-Ali, S., Carlisi, D., Ramirez, S., Upegui-Gonzalez, L. C., Duquet, A., Robin, P., Rudkin, B., Harel-Bellan, A. and Trouche, D. (1999) Phosphorylation by p44 MAP Kinase/ERK1 stimulates CBP histone acetyl transferase activity in vitro. Biochem. Biophys. Res. Commun. 262, 157–162.
- 36 Impey, S., Fong, A. L., Wang, Y., Cardinaux, J. R., Fass, D. M., Obrietan, K., Wayman, G. A., Storm, D. R., Soderling, T. R. and Goodman, R. H. (2002) Phosphorylation of CBP mediates transcriptional activation by neural activity and CaM kinase IV. Neuron 34, 235–244.
- 37 Huang, Y. Y., Kandel, E. R., Varshavsky, L., Brandon, E. P., Qi, M., Idzerda, R. L., McKnight, G. S. and Bourtchouladze, R. (1995) A genetic test of the effects of mutations in PKA on mossy fiber LTP and its relation to spatial and contextual learning. Cell 83, 1211–1222.
- 38 Bito, H., Deisseroth, K. and Tsien, R. W. (1996) CREB phosphorylation and dephosphorylation: a Ca(2+)- and stimulus duration-dependent switch for hippocampal gene expression. Cell 87, 1203–1214.
- 39 Impey, S., Mark, M., Villacres, E. C., Poser, S., Chavkin, C. and Storm, D. R. (1996) Induction of CRE-mediated gene expression by stimuli that generate long-lasting LTP in area CA1 of the hippocampus. Neuron 16, 973–982.
- 40 Abel, T., Nguyen, P. V., Barad, M., Deuel, T. A., Kandel, E. R. and Bourtchouladze, R. (1997) Genetic demonstration of a role for PKA in the late phase of LTP and in hippocampus-based long-term memory. Cell 88, 615–626.
- 41 Atkins, C. M., Selcher, J. C., Petraitis, J. J., Trzaskos, J. M. and Sweatt, J. D. (1998) The MAPK cascade is required for mammalian associative learning. Nat. Neurosci. 1, 602–609.

- 42 Bourtchouladze, R., Abel, T., Berman, N., Gordon, R., Lapidus, K. and Kandel, E. R. (1998) Different training procedures recruit either one or two critical periods for contextual memory consolidation, each of which requires protein synthesis and PKA. Learn. Mem. 5, 365–374.
- 43 Barad, M., Bourtchouladze, R., Winder, D. G., Golan, H. and Kandel, E. (1998) Rolipram, a type IV-specific phosphodiesterase inhibitor, facilitates the establishment of long-lasting long-term potentiation and improves memory. Proc. Natl. Acad. Sci. USA 95, 15020–15025.
- 44 Impey, S., Obrietan, K., Wong, S. T., Poser, S., Yano, S., Wayman, G., Deloulme, J. C., Chan, G. and Storm, D. R. (1998) Cross talk between ERK and PKA is required for Ca2+ stimulation of CREB-dependent transcription and ERK nuclear translocation. Neuron 21, 869–883.
- 45 Wong, S. T., Athos, J., Figueroa, X. A., Pineda, V. V., Schaefer, M. L., Chavkin, C. C., Muglia, L. J. and Storm, D. R. (1999) Calcium-stimulated adenylyl cyclase activity is critical for hippocampus-dependent long-term memory and late phase LTP. Neuron 23, 787–798.
- 46 Soderling, T. R. (1999) The Ca-calmodulin-dependent protein kinase cascade. Trends Biochem. Sci. 24, 232–236.
- 47 Poser, S. and Storm, D. R. (2001) Role of Ca2+-stimulated adenylyl cyclases in LTP and memory formation. Int. J. Dev. Neurosci. 19, 387–394.
- 48 Kang, H., Sun, L. D., Atkins, C. M., Soderling, T. R., Wilson, M. A. and Tonegawa, S. (2001) An important role of neural activity-dependent CaMKIV signaling in the consolidation of long-term memory. Cell 106, 771–783.
- 49 Morozov, A., Muzzio, I. A., Bourtchouladze, R., Van-Strien, N., Lapidus, K., Yin, D., Winder, D. G., Adams, J. P., Sweatt, J. D. and Kandel, E. R. (2003) Rap1 couples cAMP signaling to a distinct pool of p42/44MAPK regulating excitability, synaptic plasticity, learning, and memory. Neuron 39, 309–325.
- 50 Bourtchuladze, R., Frenguelli, B., Blendy, J., Cioffi, D., Schutz, G. and Silva, A. J. (1994) Deficient long-term memory in mice with a targeted mutation of the cAMP-responsive element-binding protein. Cell 79, 59–68.
- 51 Yin, J. C., Wallach, J. S., Del Vecchio, M., Wilder, E. L., Zhou, H., Quinn, W. G. and Tully, T. (1994) Induction of a dominant negative CREB transgene specifically blocks longterm memory in *Drosophila*. Cell 79, 49–58.
- 52 Bartsch, D., Ghirardi, M., Skehel, P. A., Karl, K. A., Herder, S. P., Chen, M., Bailey, C. H. and Kandel, E. R. (1995) Aplysia CREB2 represses long-term facilitation: relief of repression converts transient facilitation into long-term functional and structural change. Cell 83, 979–992.
- 53 Guzowski, J. F. and McGaugh, J. L. (1997) Antisense oligo-deoxynucleotide-mediated disruption of hippocampal cAMP response element binding protein levels impairs consolidation of memory for water maze training. Proc. Natl. Acad. Sci. USA 94, 2693–2698.
- 54 Kogan, J. H., Frankland, P. W., Blendy, J. A., Coblentz, J., Marowitz, Z., Schutz, G. and Silva, A. J. (1997) Spaced training induces normal long-term memory in CREB mutant mice. Curr. Biol. 7, 1–11.
- 55 Dubnau, J. and Tully, T. (1998) Gene discovery in *Drosophila*: new insights for learning and memory. Annu. Rev. Neurosci. 21, 407–444.
- 56 Albright, T. D., Jessell, T. M., Kandel, E. R. and Posner, M. I. (2000) Neural science: a century of progress and the mysteries that remain. Neuron 25 Suppl. S1–S55.
- 57 Kida, S., Josselyn, S. A., de Ortiz, S. P., Kogan, J. H., Chevere, I., Masushige, S. and Silva, A. J. (2002) CREB required for the stability of new and reactivated fear memories. Nat. Neurosci. 5, 348–355.
- 58 Tully, T., Bourtchouladze, R., Scott, R. and Tallman, J. (2003) Targeting the CREB pathway for memory enhancers. Nat. Rev. Drug Discov. 2, 267–277.

- 59 Bourtchouladze, R., Lidge, R., Catapano, R., Stanley, J., Gossweiler, S., Romashko, D., Scott, R. and Tully, T. (2003) A mouse model of Rubinstein-Taybi syndrome: defective long-term memory is ameliorated by inhibitors of phosphodiesterase 4. Proc. Natl. Acad. Sci. USA 100, 10518–10522.
- 60 Josselyn, S. A., Kida, S. and Silva, A. J. (2004) Inducible repression of CREB function disrupts amygdala-dependent memory. Neurobiol. Learn. Mem. 82, 159–163.
- 61 D'Arcangelo, G. and Curran, T. (1995) Smart transcription factors. Nature 376, 292–293.
- 62 Weeber, E. J. and Sweatt, J. D. (2002) Molecular neurobiology of human cognition. Neuron 33, 845–848.
- 63 Eberharter, A. and Becker, P. B. (2002) Histone acetylation: a switch between repressive and permissive chromatin. Second in review series on chromatin dynamics. EMBO Rep. 3, 224–229.
- 64 Scott, R., Bourtchuladze, R., Gossweiler, S., Dubnau, J. and Tully, T. (2002) CREB and the discovery of cognitive enhancers. J. Mol. Neurosci. 19, 171–177.
- 65 Nagakura, A., Takagi, N. and Takeo, S. (2002) Impairment of cerebral cAMP-mediated signal transduction system and of spatial memory function after microsphere embolism in rats. Neuroscience 113, 519–528.
- 66 Vitolo, O. V., Sant'Angelo, A., Costanzo, V., Battaglia, F., Arancio, O. and Shelanski, M. (2002) Amyloid beta -peptide inhibition of the PKA/CREB pathway and long-term potentiation: reversibility by drugs that enhance cAMP signaling. Proc. Natl. Acad. Sci. USA 99, 13217–13221.
- 67 Zhang, H. T., Huang, Y., Suvarna, N. U., Deng, C., Crissman, A. M., Hopper, A. T., De Vivo, M., Rose, G. M. and O'Donnell, J. M. (2005) Effects of the novel PDE4 inhibitors MEM1018 and MEM1091 on memory in the radial-arm maze and inhibitory avoidance tests in rats. Psychopharmacology 179, 613–619.
- 68 Monti, B., Berteotti, C. and Contestabile, A. (2006) Subchronic rolipram delivery activates hippocampal CREB and Arc, enhances retention and slows down extinction of conditioned fear. Neuropsychopharmacology 31, 278–286.
- 69 Kamei, Y., Xu, L., Heinzel, T., Torchia, J., Kurokawa, R., Gloss, B., Lin, S. C., Heyman, R. A., Rose, D. W., Glass, C. K. and Rosenfeld, M. G. (1996) A CBP integrator complex mediates transcriptional activation and AP-1 inhibition by nuclear receptors. Cell 85, 403–414.
- 70 Torchia, J., Rose, D. W., Inostroza, J., Kamei, Y., Westin, S., Glass, C. K. and Rosenfeld, M. G. (1997) The transcriptional co-activator p/CIP binds CBP and mediates nuclear-receptor function. Nature 387, 677–684.
- 71 Tanaka, Y., Naruse, I., Maekawa, T., Masuya, H., Shiroishi, T. and Ishii, S. (1997) Abnormal skeletal patterning in embryos lacking a single Cbp allele: a partial similarity with Rubinstein-Taybi syndrome. Proc. Natl. Acad. Sci. USA 94, 10215–10220.
- 72 Alarcon, J. M., Malleret, G., Touzani, K., Vronskaya, S., Ishii, S., Kandel, E. R. and Barco, A. (2004) Chromatin acetylation, memory, and LTP are impaired in CBP+/– mice: a model for the cognitive deficit in Rubinstein-Taybi syndrome and its amelioration. Neuron 42, 947–959.
- 73 Korzus, E., Rosenfeld, M. G. and Mayford, M. (2004) CBP histone acetyltransferase activity is a critical component of memory consolidation. Neuron 42, 961–972.
- 74 Wood, M. A., Kaplan, M. P., Park, A., Blanchard, E. J., Oliveira, A. M., Lombardi, T. L. and Abel, T. (2005) Transgenic mice expressing a truncated form of CREB-binding protein (CBP) exhibit deficits in hippocampal synaptic plasticity and memory storage. Learn. Mem. 12, 111–119.
- 75 Zoghbi, H. Y. and Orr, H. T. (2000) Glutamine repeats and neurodegeneration. Annu. Rev. Neurosci. 23, 217–247.
- 76 Ross, C. A. (2002) Polyglutamine pathogenesis: emergence of unifying mechanisms for Huntington's disease and related disorders. Neuron 35, 819–822.

- 77 Warrick, J. M., Paulson, H. L., Gray-Board, G. L., Bui, Q. T., Fischbeck, K. H., Pittman, R. N. and Bonini, N. M. (1998) Expanded polyglutamine protein forms nuclear inclusions and causes neural degeneration in *Drosophila*. Cell 93, 939–949.
- 78 Marsh, J. L., Walker, H., Theisen, H., Zhu, Y. Z., Fielder, T., Purcell, J. and Thompson, L. M. (2000) Expanded polyglutamine peptides alone are intrinsically cytotoxic and cause neurodegeneration in *Drosophila*. Hum. Mol. Genet. 9, 13–25.
- 79 Jackson, G. R., Salecker, I., Dong, X., Yao, X., Arnheim, N., Faber, P. W., MacDonald, M. E. and Zipursky, S. L. (1998) Polyglutamine-expanded human huntingtin transgenes induce degeneration of *Drosophila* photoreceptor neurons. Neuron 21, 633–642.
- 80 Kazemi-Esfarjani, P. and Benzer, S. (2000) Genetic suppression of polyglutamine toxicity in *Drosophila*. Science 287, 1837–1840.
- 81 Fernandez-Funez, P., Nino-Rosales, M. L., de Gouyon, B., She, W. C., Luchak, J. M., Martinez, P., Turiegano, E., Benito, J., Capovilla, M., Skinner, P. J., McCall, A., Canal, I., Orr, H. T., Zoghbi, H. Y. and Botas, J. (2000) Identification of genes that modify ataxin-1-induced neurodegeneration. Nature 408, 101–106.
- 82 Steffan, J. S., Bodai, L., Pallos, J., Poelman, M., McCampbell, A., Apostol, B. L., Kazantsev, A., Schmidt, E., Zhu, Y. Z., Greenwald, M., Kurokawa, R., Housman, D. E., Jackson, G. R., Marsh, J. L. and Thompson, L. M. (2001) Histone deacetylase inhibitors arrest polyglutamine-dependent neurodegeneration in *Drosophila*. Nature 413, 739–743.
- 83 Takeyama, K., Ito, S., Yamamoto, A., Tanimoto, H., Furutani, T., Kanuka, H., Miura, M., Tabata, T. and Kato, S. (2002) Androgen-dependent neurodegeneration by polyglutamine-expanded human androgen receptor in *Drosophila*. Neuron 35, 855–864.
- 84 Taylor, J. P., Taye, A. A., Campbell, C., Kazemi-Esfarjani, P., Fischbeck, K. H. and Min, K. T. (2003) Aberrant histone acetylation, altered transcription, and retinal degeneration in a *Drosophila* model of polyglutamine disease are rescued by CREB-binding protein. Genes Dev. 17, 1463–1468.
- 85 Davies, S. W., Turmaine, M., Cozens, B. A., DiFiglia, M., Sharp, A. H., Ross, C. A., Scherzinger, E., Wanker, E. E., Mangiarini, L. and Bates, G. P. (1997) Formation of neuronal intranuclear inclusions underlies the neurological dysfunction in mice transgenic for the HD mutation. Cell 90, 537–548.
- 86 Turmaine, M., Raza, A., Mahal, A., Mangiarini, L., Bates, G. P. and Davies, S. W. (2000) Nonapoptotic neurodegeneration in a transgenic mouse model of Huntington's disease. Proc. Natl. Acad. Sci. USA 97, 8093–8097.
- 87 Nucifora, F. C. Jr, Sasaki, M., Peters, M. F., Huang, H., Cooper, J. K., Yamada, M., Takahashi, H., Tsuji, S., Troncoso, J., Dawson, V. L., Dawson, T. M. and Ross, C. A. (2001) Interference by huntingtin and atrophin-1 with cbp-mediated transcription leading to cellular toxicity. Science 291, 2423–2428.
- 88 Sugars, K. L. and Rubinsztein, D. C. (2003) Transcriptional abnormalities in Huntington disease. Trends Genet. 19, 233– 238.
- 89 Kazantsev, A., Preisinger, E., Dranovsky, A., Goldgaber, D. and Housman, D. (1999) Insoluble detergent-resistant aggregates form between pathological and nonpathological lengths of polyglutamine in mammalian cells. Proc. Natl. Acad. Sci. USA 96, 11404–11409.
- 90 McCampbell, A., Taye, A. A., Whitty, L., Penney, E., Steffan, J. S. and Fischbeck, K. H. (2001) Histone deacetylase inhibitors reduce polyglutamine toxicity. Proc. Natl. Acad. Sci. USA 98, 15179–15184.
- 91 McCampbell, A., Taylor, J. P., Taye, A. A., Robitschek, J., Li, M., Walcott, J., Merry, D., Chai, Y., Paulson, H., Sobue, G. and Fischbeck, K. H. (2000) CREB-binding protein sequestration by expanded polyglutamine. Hum. Mol. Genet. 9, 2197–2202.

- 92 Shimohata, T., Nakajima, T., Yamada, M., Uchida, C., Onodera, O., Naruse, S., Kimura, T., Koide, R., Nozaki, K., Sano, Y., Ishiguro, H., Sakoe, K., Ooshima, T., Sato, A., Ikeuchi, T., Oyake, M., Sato, T., Aoyagi, Y., Hozumi, I., Nagatsu, T., Takiyama, Y., Nishizawa, M., Goto, J., Kanazawa, I., Davidson, I., Tanese, N., Takahashi, H. and Tsuji, S. (2000) Expanded polyglutamine stretches interact with TAFII130, interfering with CREB-dependent transcription. Nat. Genet. 26, 29–36.
- 93 Steffan, J. S., Kazantsev, A., Spasic-Boskovic, O., Greenwald, M., Zhu, Y. Z., Gohler, H., Wanker, E. E., Bates, G. P., Housman, D. E. and Thompson, L. M. (2000) The Huntington's disease protein interacts with p53 and CREB-binding protein and represses transcription. Proc. Natl. Acad. Sci. USA 97, 6763–6768.
- 94 Wyttenbach, A., Swartz, J., Kita, H., Thykjaer, T., Carmichael, J., Bradley, J., Brown, R., Maxwell, M., Schapira, A., Orntoft, T. F., Kato, K. and Rubinsztein, D. C. (2001) Polyglutamine expansions cause decreased CRE-mediated transcription and early gene expression changes prior to cell death in an inducible cell model of Huntington's disease. Hum. Mol. Genet. 10, 1829–1845
- 95 Sugars, K. L., Brown, R., Cook, L. J., Swartz, J. and Rubinsztein, D. C. (2004) Decreased cAMP response element-mediated transcription: an early event in exon 1 and full-length cell models of Huntington's disease that contributes to polyglutamine pathogenesis. J. Biol. Chem. 279, 4988–4999.
- 96 Iijima-Ando, K., Wu, P., Drier, E. A., Iijima, K. and Yin, J. C. (2005) cAMP-response element-binding protein and heat-shock protein 70 additively suppress polyglutamine-mediated toxicity in *Drosophila*. Proc. Natl. Acad. Sci. USA 102, 10261–10266.
- 97 Jiang, H., Nucifora, F. C. Jr, Ross, C. A. and DeFranco, D. B. (2003) Cell death triggered by polyglutamine-expanded huntingtin in a neuronal cell line is associated with degradation of CREB-binding protein. Hum. Mol. Genet. 12, 1–12.
- 98 Ferrante, R. J., Kubilus, J. K., Lee, J., Ryu, H., Beesen, A., Zucker, B., Smith, K., Kowall, N. W., Ratan, R. R., Luthi-Carter, R. and Hersch, S. M. (2003) Histone deacetylase inhibition by sodium butyrate chemotherapy ameliorates the neurodegenerative phenotype in Huntington's disease mice. J. Neurosci. 23, 9418–9427.
- 99 Gardian, G., Browne, S. E., Choi, D. K., Klivenyi, P., Gregorio, J., Kubilus, J. K., Ryu, H., Langley, B., Ratan, R. R., Ferrante, R. J. and Beal, M. F. (2005) Neuroprotective effects of phenylbutyrate in the N171–82Q transgenic mouse model of Huntington's disease. J. Biol. Chem. 280, 556–563.
- 100 Masliah, E. (1995) Mechanisms of synaptic dysfunction in Alzheimer's disease. Histol. Histopathol. 10, 509–519.
- 101 Small, D. H., Mok, S. S. and Bornstein, J. C. (2001) Alzheimer's disease and Abeta toxicity: from top to bottom. Nat. Rev. Neurosci. 2, 595–598.
- 102 Walsh, D. M. and Selkoe, D. J. (2004) Deciphering the molecular basis of memory failure in Alzheimer's disease. Neuron 44, 181–193.
- 103 LaFerla, F. M. and Oddo, S. (2005) Alzheimer's disease: Abeta, tau and synaptic dysfunction. Trends Mol. Med. 11, 170–176.
- 104 Cullen, W. K., Suh, Y. H., Anwyl, R. and Rowan, M. J. (1997) Block of LTP in rat hippocampus *in vivo* by beta-amyloid precursor protein fragments. Neuroreport 8, 3213–3217.
- 105 Itoh, A., Akaike, T., Sokabe, M., Nitta, A., Iida, R., Olariu, A., Yamada, K. and Nabeshima, T. (1999) Impairments of long-term potentiation in hippocampal slices of beta-amyloid-infused rats. Eur. J. Pharmacol. 382, 167–175.
- 106 Chapman, P. F., White, G. L., Jones, M. W., Cooper-Blacketer, D., Marshall, V. J., Irizarry, M., Younkin, L., Good, M. A., Bliss, T. V., Hyman, B. T., Younkin, S. G. and Hsiao, K. K. (1999) Impaired synaptic plasticity and learning in aged

- amyloid precursor protein transgenic mice. Nat. Neurosci. 2, 271–276.
- 107 Walsh, D. M., Klyubin, I., Fadeeva, J. V., Cullen, W. K., Anwyl, R., Wolfe, M. S., Rowan, M. J. and Selkoe, D. J. (2002) Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation *in vivo*. Nature 416, 535–539.
- 108 Oddo, S., Caccamo, A., Shepherd, J. D., Murphy, M. P., Golde, T. E., Kayed, R., Metherate, R., Mattson, M. P., Akbari, Y. and LaFerla, F. M. (2003) Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction. Neuron 39, 409–421.
- 109 Brendza, R. P., Bacskai, B. J., Cirrito, J. R., Simmons, K. A., Skoch, J. M., Klunk, W. E., Mathis, C. A., Bales, K. R., Paul, S. M., Hyman, B. T. and Holtzman, D. M. (2005) Anti-Abeta antibody treatment promotes the rapid recovery of amyloidassociated neuritic dystrophy in PDAPP transgenic mice. J. Clin. Invest. 115, 428–433.
- 110 Puzzo, D., Vitolo, O., Trinchese, F., Jacob, J. P., Palmeri, A. and Arancio, O. (2005) Amyloid-beta peptide inhibits activation of the nitric oxide/cGMP/cAMP-responsive element-binding protein pathway during hippocampal synaptic plasticity. J. Neurosci. 25, 6887–6897.
- 111 Tong, L., Thornton, P. L., Balazs, R. and Cotman, C. W. (2001) Beta -amyloid-(1–42) impairs activity-dependent cAMP-response element-binding protein signaling in neurons at concentrations in which cell survival Is not compromised. J. Biol. Chem. 276, 17301–17306.
- 112 Marambaud, P., Wen, P. H., Dutt, A., Shioi, J., Takashima, A., Siman, R. and Robakis, N. K. (2003) A CBP binding transcriptional repressor produced by the PS1/epsilon-cleavage of N-cadherin is inhibited by PS1 FAD mutations. Cell 114, 635–645.
- 113 Gong, B., Vitolo, O. V., Trinchese, F., Liu, S., Shelanski, M. and Arancio, O. (2004) Persistent improvement in synaptic and cognitive functions in an Alzheimer mouse model after rolipram treatment. J. Clin. Invest. 114, 1624–1634.
- 114 Saura, C. A., Choi, S. Y., Beglopoulos, V., Malkani, S., Zhang, D., Shankaranarayana Rao, B. S., Chattarji, S., Kelleher, R. J. 3rd, Kandel, E. R., Duff, K., Kirkwood, A. and Shen, J. (2004) Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration. Neuron 42, 23–36.

- 115 Holcomb, L., Gordon, M. N., McGowan, E., Yu, X., Benkovic, S., Jantzen, P., Wright, K., Saad, I., Mueller, R., Morgan, D., Sanders, S., Zehr, C., O'Campo, K., Hardy, J., Prada, C. M., Eckman, C., Younkin, S., Hsiao, K. and Duff, K. (1998) Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes. Nat. Med. 4, 97–100.
- 116 Morgan, D., Diamond, D. M., Gottschall, P. E., Ugen, K. E., Dickey, C., Hardy, J., Duff, K., Jantzen, P., DiCarlo, G., Wilcock, D., Connor, K., Hatcher, J., Hope, C., Gordon, M. and Arendash, G. W. (2000) A beta peptide vaccination prevents memory loss in an animal model of Alzheimer's disease. Nature 408, 982–985.
- 117 Arendash, G. W., King, D. L., Gordon, M. N., Morgan, D., Hatcher, J. M., Hope, C. E. and Diamond, D. M. (2001) Progressive, age-related behavioral impairments in transgenic mice carrying both mutant amyloid precursor protein and presenilin-1 transgenes. Brain Res. 891, 42–53.
- 118 Gordon, M. N., King, D. L., Diamond, D. M., Jantzen, P. T., Boyett, K. V., Hope, C. E., Hatcher, J. M., DiCarlo, G., Gottschall, W. P., Morgan, D. and Arendash, G. W. (2001) Correlation between cognitive deficits and Abeta deposits in transgenic APP+PS1 mice. Neurobiol. Aging 22, 377– 385.
- 119 Bruijn, L. I., Miller, T. M. and Cleveland, D. W. (2004) Unraveling the mechanisms involved in motor neuron degeneration in ALS. Annu. Rev. Neurosci. 27, 723–749.
- 120 Bendotti, C. and Carri, M. T. (2004) Lessons from models of SOD1-linked familial ALS. Trends Mol. Med. 10, 393–400.
- 121 Shibata, N. (2001) Transgenic mouse model for familial amyotrophic lateral sclerosis with superoxide dismutase-1 mutation. Neuropathology 21, 82–92.
- 122 Hand, C. K., Khoris, J., Salachas, F., Gros-Louis, F., Lopes, A. A., Mayeux-Portas, V., Brewer, C. G., Brown, R. H. Jr, Meininger, V., Camu, W. and Rouleau, G. A. (2002) A novel locus for familial amyotrophic lateral sclerosis, on chromosome 18q. Am. J. Hum. Genet. 70, 251–256.
- 123 Rouaux, C., Jokic, N., Mbebi, C., Boutillier, S., Loeffler, J. P. and Boutillier, A. L. (2003) Critical loss of CBP/p300 histone acetylase activity by caspase-6 during neurodegeneration. EMBO J. 22, 6537–6549.