DOI: 10.1002/epi4.12824

ORIGINAL ARTICLE

Accepted: 9 September 2023

First report of Tunisian patients with *CDKL5*-related encephalopathy

Chahnez Charfi Triki¹ | Salma Zouari Mallouli¹ | Marwa Ben Jdila² | Mariem Ben Said³ | Fatma Kamoun Feki¹ | Sarah Weckhuysen^{4,5,6} | Sabeur Masmoudi³ | Faiza Fakhfakh²

¹Child Neurology Department, Hedi Chaker Sfax University Hospital, and Research Laboratory LR19ES15, University of Sfax, Sfax, Tunisia

²Laboratory of Molecular and Functional Genetics, Faculty of Science of Sfax, University of Sfax, Sfax, Tunisia

³Laboratory of Molecular and Cellular Screening Processes (LPCMC), Center of Biotechnology of Sfax, University of Sfax, Sfax, Tunisia

⁴Applied & Translational Neurogenomics Group, VIB Center for Molecular Neurology, VIB, University of Antwerp, Antwerp, Belgium

⁵Department of Neurology, Antwerp University Hospital, Antwerp, Belgium

⁶Translational Neurosciences, Faculty of Medicine and Health Science, University of Antwerp, Antwerp, Belgium

Correspondence

Chahnez Charfi Triki, Child Neurology Department, Hedi Chaker Sfax University Hospital and Research Laboratory LR19ES15, University of Sfax, Sfax, Tunisia. Email: chahnezct@gmail.com

Abstract

Objective: Mutations in the cyclin-dependent kinase-like 5 gene (CDKL5) are associated with a wide spectrum of clinical presentations. Early-onset epileptic encephalopathy (EOEE) is the most recognized phenotype. Here we describe phenotypic features in eight Tunisian patients with CDKL5-related encephalopathy. **Methods:** We included all cases with clinical features consistent with CDKL5-related encephalopathy: infantile epileptic spasm, acquired microcephaly, movement disorders and visual impairment. We collected data about seizure types, electroencephalogram, magnetic resonance imaging, and metabolic analysis. The diagnosis of CDKL5 mutation was made thanks to Sanger sequencing with an ABI PRISM 3100-Avant automated DNA sequencer using a Big Dye Terminator Cycle Sequencing Reaction Kit v1.1. and Next Generation Sequencing (NGS) since the development of a gene panel responsible for DEE within the framework of "Strengthening the Sfax University Expertise for diagnosis and management of epileptic encephalopathies".

Results: We collected four boys and four girls aged meanly 6 years old with confirmed mutation on CDKL5 gene. Overall, we identified five de novo CDKL5 mutations including three Frame-shift mutations, one missense mutation, and a splicing variant. The mean age at first seizure onset was 4 months. The first seizure type was infantile epileptic spasm (4/8) followed by tonic (2/8) and myoclonic seizures (2/8). Out of eight cases, four exhibited two stages epileptic course while epilepsy in three other patients progressed on three stages. Regarding development, most cases (6/8) had psychomotor retardation from the start whilst the two others showed psychomotor regression with the onset of seizures. Additional clinical features included visual impairment (7/8), tone abnormalities (7/8), stereotypies (7/8), and acquired microcephaly (6/8).

Significance: Our present report delineates an unusual phenotype of CDKL5related encephalopathy with male gender predominance and delayed onset

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

epilepsy. It interestingly described new phenotypic features and uncommon benign developmental profiles in boys, different patterns of CDKL5-epilepsy, neuroimaging findings, and CDKL5 mutational spectrum.

K E Y W O R D S

CDKL5 mutation, clinical features, developmental and epileptic encephalopathy, epileptic spasm

1 INTRODUCTION

The cyclin-dependent kinase-like 5 (CDKL5)-also known as STK9-belongs to a small family of five distinct Ser/ Thr protein kinases, which contribute to signaling pathways basic for cell biology. The inactivation of the X-linked CDKL5 gene-or its mutations-was responsible for a wide spectrum of clinical phenotypes grouped under the umbrella of CDKL5 deficiency disorder (CDD).¹ The most recognized phenotype of CDKL5-related developmental and epileptic encephalopathy (DEE) is characterized by a 3-stage evolution of epilepsy with the onset of epilepsy before 3 months of age (stage 1), later progression into infantile epileptic spasm syndrome (IESS) (stage 2) and resulting ultimately in multifocal refractory epilepsy (stage 3).²⁻⁴ Epilepsy occurs in the setting of delayed psychomotor development and associates Rett-like features such as hand stereotypies and deceleration of head growth.² Despite the association of CDKL5 mutations with these well-known syndromic clusters, recent publications continue to provide phenotypic data different from previous descriptions, leading to a broadened spectrum of CDKL5-related disorders.²⁻⁶ Through this paper, we aim to depict the phenotypic specificities of Tunisian CDKL5 cases.

2 MATERIALS AND METHODS

2.1 | Patients and data collection

Files of patients with a follow-up at the Child neurology department of Hedi Chaker Sfax university hospital for Epileptic Encephalopathy (EE) beginning before the age of 1 year with neurodevelopmental impairment preceding or following the seizure onset, were retrospectively reviewed. Electro-clinical phenotyping was based on several EEGs coupled with video during the follow-up period, and the patient assignment was made according to the 2022 ILAE classification for seizures and epileptic syndromes.^{7–9} All patients had brain imaging and metabolic screening before their enrollment for genetic analysis. Patients with onset EE before the age of 1 year, and clinical features suggestive of *CDKL5* mutation, including acquired microcephaly and involuntary movement, were proposed for targeted gene

Key points

- *CDKL5* gene should be considered as fundamental for a gene panel designed to assess DDE as *CDKL5* mutation was frequently associated with such phenotype.
- *CDKL5*-related DEE affected both girls and boys justifying considering CDKL5 mutation in boys with DEE, too.
- Understanding epilepsy course associated with *CDKL5*-related DEE may help the development of a personalized management.

analysis using Sanger Sequencing. Since the development of a gene panel for DEE genetic assessment—including the *CDKL5* gene—at Sfax University within the framework of "Strengthening the Sfax University Expertise for diagnosis and management of epileptic encephalopathies" (SEED project GA n°856592-H202), CDKL5 gene sequencing was continued via Next Generation Sequencing (NGS) and SEED panel. Those whose neuroimaging showed major structural or signal abnormalities enough to explain the patient clinical picture or metabolic screening allowed the diagnosis of alternate inborn error of metabolism did not take part in genetic screening.

2.2 | Molecular methods

Blood samples were obtained from the children and their parents after informed consent. Genomic DNA was extracted from blood leukocytes by applying phenol-chloroform standard procedures. Polymerase Chain Reaction (PCR) was completed with amplification of the CDKL5 gene including 20 exons (2–21) and their exon-intron boundaries in all patients. PCR products were purified for mutation analysis. CDKL5 gene sequencing was performed by Sanger sequencing with an ABI PRISM 3100-Avant automated DNA sequencer using a Big Dye Terminator Cycle Sequencing Reaction Kit v1.1. The SEED panel was designed using

908 Epilepsia Open[®]

TABLE 1 Clinical, neuroimaging and genetic characteristics of patients with CDKL5 mutation.

	Patient 1 (P1) ^a	Patient 2 (P2) ^a	Patient 3 (P3) ^a	Patient 4 (P4)
Gender	M	М	М	F
Age at first visit (months)	16	8	5	2
Age at last visit (months)	208	76	149	28
Follow-up period (months)	192	68	145	26
Consanguinity	-	-	-	+
Family history of epilepsy	+	-	+	-
Clinical findings				
Acquired microcephaly	_	+	_	+
Abnormal movements	Hand stereotypies	Hand stereotypies Chorea	Hand stereotypies Chorea	Hand stereotypies
Development course				
Initial cognitive and motor development	Normal, with regression at onset of seizures	Normal, with regression at onset of seizures	Delayed	Delayed
Language	2 words (7 years)	Babbling (4 years)	-	Single words (2years)
Independent walking	7 years	-	8 years	-
Epilepsy				
Onset of epilepsy	6 months	8 months	5 months	40 days
First type of seizure/epilepsy syndrome	ES/IESS	ES/IESS	ES/IESS	Myoclonic seizures
Seizure-free period	12 months	32 months	7 months	16 months
Age at onset second seizure type	18 month	76 months	36 months	18 months
Type of seizures/epilepsy syndrome during evolution	FMTS	Multifocal seizures (FMTS and FMCS)	TS	Tonic spasm/IESS
Age at onset third seizure type	-	-	-	-
Type of seizures/epilepsy syndrome during evolution	-	-	-	-
Epilepsy course	2 stages	2 stages	2 stages	2 stages
Response to ASMs (Seizure-free)	+	-	+	No data
EEG findings				
At onset During follow-up	 Hypsarrhythmia Normal (16 months) 	 Hypsarrhythmia Diffuse epileptiform abnormalities (58 months) 	 Hypsarrhythmia Focal epileptiform abnormalities (48 months) 	 Ictal EEG: Diffuse polyspikes synchronous to myoclonus/Interictal: normal Interictal: multifocal epileptiform abnormalities (18 months)
Brain MRI	9 years: Normal	14 months: Fronto- temporal cortex atrophy + thin corpus callosum	5 years: Normal	2 years 3 months: Normal
CDKL5 mutation	De novo c.616 G>A (E	29) (p.Asp206Asn) at a soma	atic mosaic state	De novo c.149delA (p. Asn50MetfsTer26) at a heterozygote state
ACMG classification of the mutation	VUS			LP
Frequencies (gnomAD)	VNF			VNF

Abbreviations: (–), no; (+), yes; ASMs, anti-seizures medications; E, exon; ES, epileptic spasm; F, female; FMCS, focal motor clonic seizures; FMTS, focal motor tonic seizures; IESS, infantile epileptic spasm syndrome; LGS, Lennox Gastaut syndrome; LP, Likely pathogenic; M, male; S, stage; TS, tonic seizures; VNF, variant not found; VUS, variant of uncertain significance.

^aThe patient has been previously reported in a separate publication.¹⁵

^bThe patient has been previously reported in a separate publication.¹⁴

^cData according to CT scan.

_Epilepsia Open®

FMFFF66549161272913012761212011201120111010111111111111111111111111111111111	Patient 5 (P5)	Patient 6 (P6) ^b	Patient 7 (P7)	Patient 8 (P8)
66 5 4 9 186 132 72 21 120 127 68 22 + + - - - - +(DE) - - - - +(DE) - - - + Hand stereotypies + + - - - - - - + - - - - - + -<	F	М	F	F
186 132 72 12 120 127 68 12 + - - - - +(DEF) - - + - - - - + - - - - - + - - - - - - + - - - - - - - -	66	5	4	9
1201276812 $+$ $+$ $ +$ (DFD) $ +$ $+$ (DFD) $ +$ $+$ $+$ $+$ $+$ Hand stereotypies $ +$ $ +$ $ -$ <tr< td=""><td>186</td><td>132</td><td>72</td><td>21</td></tr<>	186	132	72	21
+ - $+$ (DEP) $ -$ $ +$ Hand stereotypies $ +$ $ +$ Hand stereotypies $ +$ Hand stereotypies $-$	120	127	68	12
-+ (DER)*++Hand stereotypies Chorea Non-epilpatic myocionus*DelayedDelayedDelayedDelayedDelayedDelayedDelayedDelayed	+	+	-	-
Hand stereotypiesHand stereotypies Chorea Non-epilepite myoclonusHand stereotypies Chorea Non-epilepite myoclonusDelayedDelayedDelayedNonothsNaveks Myoclonus eszuresImonths Months Nonoths Steres SteresImonths Months MonthsImonths Months MonthsImonths Months Months Steres SteresImonths Months Months MonthsImonths Months MonthsImonths Months Months12 MonthsStages StagesImonths MonthsImonths Months MonthsImonths MonthsImonths Months2 Morea to foroto polyspikes (stabearges (186 months) StagesImonths StagesImonths MonthsImonths Months MonthsImonths Months2 Nota wallable (stabearges (186 months) (stabearges (186 months) (stabearges (186 months) MonthsImonths MonthsImonths MonthsImonths Months Months2 Nota wallable (stabearges (186 months) (stabearges (186 months) (stabearges (186 months) (stabearges (18	-	+ (DEE)	-	-
+ Hand stereotypies+ -+ Hand stereotypies+ Hand stereotypiesDelayedDelayedDelayedDelayedDelayedDelayedDelayedDelayedDelayedDelayed				
Hand stereotypies-Rand stereotypies choree Non-epleptir myoclonusHand stereotypies choree choree choree choree choreeHand stereotypies choree ch	+	+	+	+
Delayed Delayed <t< td=""><td>Hand stereotypies</td><td>_</td><td>Hand stereotypies</td><td>Hand stereotypies</td></t<>	Hand stereotypies	_	Hand stereotypies	Hand stereotypies
Non-epilepite myocionusDelayedDelayedDelayedDelayedDelayedDelayed </td <td></td> <td></td> <td>Chorea</td> <td></td>			Chorea	
Delayed <td></td> <td></td> <td>Non-epileptic myoclonus</td> <td></td>			Non-epileptic myoclonus	
DelayedDelayedDelayedDelayedDelayed3months3weeks1month8monthsTSMyoelonic seizuresTSES/IESS9months4months3months-12months5months4monthsNAAbsence seizuresES/IESSES/IESSES/IESS108 months5monthsES/IESSES/IESS108 months15months18months-17S, FMCS/LGS3stages3 stages-3 stages3 stages3 stages-1. Not available1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarthythmia2. Ictal EGG: FMCS synchronous ulischarges (186 months)1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarthythmia2. Ictal EGG: FMCS synchronous ulischarges (186 months)1. Focal paroxysm on well-organized background activity1. Not available (29 months)1. Hypsarthythmia (14 months) a. multifical epileptiform abnormalities (12 months)1. Hypsarthythmia (12 months)9years 3 months: NormalNomoths/11 years: Fronto-temporal (29 months)1. Smonths': Fronto-temporal cortex atrophy+thin corpus callosum maltimes (12 months)8 months: Fronto-temporal cortex atrophy+thelayed myelinationPe novo c.616 G>A (E9) (p.Ap206Asn) at a heterozygote statePe novo c.7288inG (E19) p.Glu930GlyfSTer9De novo c.910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote state a heteroz				
<	Delayed	Delayed	Delayed	Delayed
<td></td> <td></td> <td></td> <td></td>				
·····Smonths TSSweeks Myclonic seizuresImonth TSSmonths ES/IESSSmonths ES/IESSSmonths ES/IESSNamoths LamonthsMonths Mandhs ES/IESSSmonths ES/IESSSmonths ES/IESSSmonths ES/IESSSmonths ES/IESSNB months PMTS, FMCS/LGSSmonths Absence, TS/LGSBamonths Multifocal seizures3 stages t for to root opolar polyspikes discharges (186 months) * Diffuse irregular spike and weith background activity * Dysparrhythmia (1700nths) * Diffuse irregular spike and weith spike attrophythetia (2000mths) * Diffuse irregular spike and weith epilepifform abnormalities (2000mths) * Diffuse irregular spike and weith epilepifform abnormalities (2000mths)Smonths * Diffuse irregular spike and weith epilepifform abnormalities (2000mths) * Diffuse irregular spike and weith epilepifform abnormalities (2000mths)Smonths * Diffuse irregular spike and weith epilepifform abnormalities	-	_	_	-
Amounts TSAweeks Myoclonic seizuresImonth TSMonoths ES/IESS9months 12months4months months3months Manet ES/IESS-12months 12months4months monthsManet Manet ES/IESSManet ES/IESS108months PMTS, FMCS/LGS15months Absence, TS/LGSBanonths Multifocal seizures-13stages + 23stages - -3stages - 1. Not available 2. Inctail EEG: fonto-polyspikes discharges (166 months)1. Not available 2. Users atmosthy (166 months)2. Not available 2. Inctail EEG: (160 months)2. Not available 2. Inctail EEG: background activity (166 months)3. Not available 2. Inforto-polyspikes discharges (166 months)3. Thoot available 2. Inforto-polyspikes discharges (166 months)3. Proot c. Attrophy9 wears 3months: Normal Manet9 wears 4months: Normal Manet heterozygie state7 Months/11 years: Fronto-temporal discharges (166 months)9 monto-c.161 GSAN (219) (Apa206Ash) at a heterozygie state7 Months/11 years: Fronto-temporal discharges (166 months)7 Months/11 years: Fronto-temporal oper monot-clift scatteropy7 Months/11 years: Fronto-temporal heterozygie state <td>-</td> <td>_</td> <td>_</td> <td>-</td>	-	_	_	-
Smonths TSSweeks Mycolonic seizuresImonth TSSmonths ES/IESS9Months Smonths Absence seizuresAmonths Smonths ES/IESSSmonths Honths ES/IESSNA ES/IESS108 months FMTS, FMCS/LGSIS monts Absence, TS/LGSI8 months Multifocal seizures.3 stages +3 stages1. Not available 2. Ictal EEG: FMCS synchronour to fronto-polar polyspikes (96 months)1. Not available 2. Ictal EEG: FMCS synchronour to fronto-polar polyspikes (96 months)9 years 3 months: NormalToomths/11 years: Fronto-temporal (96 months)9 years 3 months: Normal Netrozypot stataDenove c.278 kinsG (E19) polu30G kirSrentVUSLPLPLPLPVUSLPLPLPVUSLPLPLPVUSLPLPVUSLPLPVUSLPNFVUSLPVUSLPVUSLP <t< td=""><td></td><td></td><td></td><td></td></t<>				
TSMyclonic seizuresTSES/IES39 months4 months3 months-12 months5 months4 monthsMAAbsence seizuresES/IES3ES/IES3ES/IES3108 months15 months18 months-108 months15 months18 months-108 months15 months18 months-108 months15 months3 stages-3 stages3 stages3 stages-12. lotal EEG: FMCS synchronousbackground activity 2. Hypsarrhythmia (17 months) 3. Diffuse irregular spike and wave (06 months)Not available 2. Hypsarrhythmia (14 months) 3. Diffuse irregular spike and wave (06 months)1. Mot available 2. Hypsarrhythmia (12 months) 3. Diffuse irregular spike and wave (06 months)1. Sonoths ⁻¹ , Fronto-temporal cortex atroph + thin corpus callosum8 months: remoral atnoph + thin corpus callosum9 gears 3 months: Normal heterozygote statePen novo c.1910_1911 ins 1910_(E12) p.Glu930GlyfsTer9Pen novo c.1910_1911 ins 1910_(E12) p.Glu930GlyfsTer92.153-1GS-A (E15) at a heterozygote stateVUSIPIPIPIPIPVNFVMFVMFVMFIP	3 months	3 weeks	1 month	8 months
9 months4 months3 months-12 months5 months4 monthsMAAbsence seizuresES/IESSES/IESSES/IESS108 months15 months18 months-FMTS, FMCS/LGS3 stages3 stages-3 stages3 stages3 stages-+1. Not available1. Focal paroxysm on well-organized background activity1. Not available 2. Hypsarrhythmia (14 months) 3. Diffuse irregular spike and wave (96 months)1. Not available 2. Hypsarrhythmia (29 months)1. Hypsarrhythmia 2. Interictal: multifocal epileptiform abnormalities (29 months)1. Hypsarrhythmia 2. Interictal: multifocal epileptiform abnormalities (20 months)1. Smonths: Fronto- temporal cortex atrophy + thin corpus callosum8 months: Fronto- temporal cortex atrophy + Delayed myelination0 pa novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote state2.153-IG>A (E15) at a heterozygote stateVUSLPLPLPLPLPVNFVNFVNFVNFVNF	TS	Myoclonic seizures	TS	ES/IESS
9 months4 months3 months- · · · · · · · · · · · · · · · · · · ·		5		
12 monthsSmonthsAmonthsNAAbsence seizuresES/IESSES/IESSES/IESS108 months15 monthsBanoths-FMCS,FMCS/LGS3 stagesBanoths-3 stages3 stages1 Not available2 Not available	9 months	4 months	3 months	-
Absence seizuresES/IESSES/IESSES/IESS108 months15 months16 months18 months1FMTS, FMCS/LGS3 baence, TS/LGS18 months113 stages3 stages3 stages11111 Not available1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarthythmia (14 months)1. Hypsarthythmia 2. Hypsarthythmia (29 months)1. Hypsarthythmia (29 months)1. Hypsarthythmia (29 months)9 parts 3 months: Normal1. Sonths/11 years: Fronto-temporal (96 months)3 months': Fronto-temporal cortex atrophy+thin corpus callosum altorphy-thelayed multifocal epilepilofism abnormalities (29 months)8 months: Fronto-temporal cortex atrophy+thin corpus callosum altorphy-thelayed altorphy-thelayed mul	12 months	5 months	4 months	NA
108 months FMTS, FMCS/LGS15 months Absence, TS/LGS18 months Multifocal seizures-3 stages +3 stages3 stages-3 rages +1. Not available +2. Ictal EEG: FMCS synchronous background activity + bysgarrhythmia (17 months) + bysgarrhythmia (14 months) + bysgarrhythmia (14 months) + bysgarrhythmia (14 months) + bysgarrhythmia (12 months)Bysgarrhythmia (12 months) + bysgarrhythmia (12 months) + bysgarrhythmia (12 months) + bysgarrhythmia (12 months)Bysgarrhythmia (12 months) + bysgarrhythmia (12 months) + bysgarrhythmia (12 months)9 ward Particle Par	Absence seizures	ES/IESS	ES/IESS	ES/IESS
108 months15 months18 months18 months				
FMTS, FMCS/LGSAbsence, TS/LGSMultifocal seizures-3 stages3 stages3 stages-41. Not available1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarrhythmia2. Ictal EEG: FMCS synchronous discharges (18 months)1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarrhythmia2. Ictal EEG: FMCS synchronous discharges (18 months)1. Fypsarrhythmia (7 months) abityms rirgular spike and wave (96 months)1. Not available 2. Hypsarrhythmia (12 months) (29 months)1. Hypsarrhythmia (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ⁶ : Fronto-temporal cortex atrophy+thin corpus callosum8 months: Fronto- temporal cortex atrophy+Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPLPVNFVNFVNFVNF	108 months	15 months	18 months	-
3 stages3 stages3 stages3 stages-3 stages	FMTS, FMCS/LGS	Absence, TS/LGS	Multifocal seizures	-
3 stages3 stages3 stages+1. Not available1. Focal paroxysm on well-organized background activity1. Not available1. Not available2. Ictal EEG: FMCS synchronous discharges (186 months)1. Focal paroxysm on well-organized background activity1. Not available1. Hypsarrhythmia3. Diffuse irregular spike and wave (96 months)1. Not available2. Hypsarrhythmia (14 months)2. Interictal: multifocal epileptiform abnormalities (29 months)3. multifocal epileptiform abnormalities (29 months)2. Interictal: multifocal epileptiform abnormalities (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ⁶ : Fronto-temporal cortex atrophy+ thin corpus callosum myelination8 months: Fronto- temporal cortex atrophy+ Delayed myelination0 p. novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote state2.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPLPVNFVNFVNFVNF				
+1. Not available 2. Ictal EEG: FMCS synchronous to fronto-polar polyspikes discharges (186 months)1. Focal paroxysm on well-organized background activity 2. Hypsarrhythmia (7 months) 3. Diffuse irregular spike and wave (96 months)1. Not available 2. Hypsarrhythmia (14 months) 3. multifocal epileptiform abnormalities (29 months)1. Hypsarrhythmia 2. Interictal: multifocal epileptiform abnormalities (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ⁶ : Fronto-temporal cortex atrophy+thin corpus callosum multifocal epileptiform abnormalities (12 months)8 months: Fronto- temporal cortex atrophy+Delayed myelination0 povo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF	3 stages	3 stages	3 stages	-
1. Not available 2. Ictal EEG: FMCS synchronous 2. Ictal EEG: FMCS synchronous discharges (186 months)1. Focal paroxysm on well-organized background activity 2. Hypsarrhythmia (7 months) 3. Diffuse irregular spike and wave (96 months)1. Not available 2. Hypsarrhythmia (14 months) 3. multifocal epileptiform abnormalities (29 months) (29 months)1. Hypsarrhythmia 2. Interictal: multifocal epileptiform abnormalities (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy+thin corpus callosum8 months: Fronto- temporal cortex atrophy+ Delayed myelination0 povo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPLPVNFVNFVNFVNF	+	-	-	-
1. Not available1. Focal paroxysm on weil-organized background activity1. Not available1. Not available1. Hypsarrhytnina2. Ictal EEG: FMCS synchronous to fronto-polar polyspikes discharges (186 months)2. Hypsarrhythmia (7 months) 3. Diffuse irregular spike and wave (96 months)2. Hypsarrhythmia (14 months) 3. multifocal epileptiform abnormalities (29 months)2. Interictal: multifocal epileptiform abnormalities (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy+ thin corpus callosum8 months: Fronto- temporal cortex atrophy+ Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPLPVNFVNFVNFVNF	1. N. 6	1	1. Makana lahla	1. II
2. Note EDG Finite SynchronousDetersponde uctivity2. Hypsamilytimina (Trunontis)2. Hypsamilytimina (Trunontis)2. Hypsarrhythmia (7 months)3. Diffuse irregular spike and wave (96 months)3. multifocal epileptiform abnormalities (29 months)epileptiform abnormalities (12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy + thin corpus callosum8 months: Fronto- temporal cortex atrophy + Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNF	1. Not available 2. Ictal EEG: EMCS synchronous	1. Focal paroxysm on well-organized	 Not available Hypsarrhythmia (14 months) 	 Hypsarrnytnmia Interictal: multifocal
discharges (186 months)3. Diffuse irregular spike and wave (96 months)(29 months)11 month9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy + thin corpus callosum8 months: Fronto- 	to fronto-polar polyspikes	2. Hypsarrhythmia (7 months)	3. multifocal epileptiform abnormalities	epileptiform
(96 months)(12 months)9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy+thin corpus callosum8 months: Fronto- temporal cortex atrophy+Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNF	discharges (186 months)	3. Diffuse irregular spike and wave	(29 months)	abnormalities
9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy+thin corpus callosum8 months: Fronto- temporal cortex atrophy+Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF		(96 months)		(12 months)
9 years 3 months: Normal7 months/11 years: Fronto-temporal cortex atrophy13 months ^c : Fronto-temporal cortex atrophy + thin corpus callosum8 months: Fronto- temporal cortex atrophy + Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote stateC2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF				
Sygers 3 months: Normal/ months/ Hyears: Fronto-temporal cortex atrophyIs months: Fronto-temporal cortex atrophy + thin corpus callosum8 months: Fronto- temporal cortex atrophy + Delayed myelinationDe novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNF	Oregona 2 month as Normal	7 m outbo/11 money Fronts town and	12 months ⁶ . Enoute tomorousl contour	0 m on the Energie
De novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF	9 years 3 months: Normai	/ months/11 years: Fronto-temporal	atrophy + thin corpus callosum	temporal cortex
De novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF		contex autophy	atophy i thin corpus canosani	atrophy + Delayed
De novo c.616 G>A (E9) (p.Asp206Asn) at a heterozygote stateDe novo c.2788insG (E19) p.Glu930GlyfsTer9De novo c.1910_1911 ins 19nt (E12) (p.Leu644Ter) at a heterozygote statec.2153-1G>A (E15) at a heterozygote stateVUSLPLPLPVNFVNFVNFVNF				myelination
(p.Asp206Asn) at a heterozygote statep.Glu930GlyfsTer9(p.Leu644Ter) at a heterozygote statea heterozygote stateVUSLPLPLPVNFVNFVNFVNF	De novo c.616 G>A (E9)	De novo c.2788insG (E19)	De novo c.1910_1911 ins 19nt (E12)	c.2153-1G>A (E15) at
heterozygote stateVUSLPVNFVNFVNFVNF	(p.Asp206Asn) at a	p.Glu930GlyfsTer9	(p.Leu644Ter) at a heterozygote state	a heterozygote state
VUSLPLPLPVNFVNFVNFVNF	heterozygote state			
VNF VNF VNF VNF	VUS		LP	LP
	VNF	VNF	VNF	VNF

909

Open Access

Design Studio to create a custom target enrichment library design. The design was based on GRCh37/hg19 reference sequences, with target sources obtained from the RefSeq database. In this custom design, all coding exons were targeted involving 25bp of the flanking intronic sequence of 116 genes. Genes were selected according to their relevance in DEE based on previously published genetic studies. The MiSeq Reporter software settings (Illumina) were adjusted to generate VCF files for index reads. VarAFT was used for variants annotation and filtering.¹⁰ In addition, we examined the pathogenicity of the different variants following the ACMG (American College of Medical Genetics and Genomics) standards and guidelines.¹¹ Moreover, the Allele Frequency in the Genome Aggregation Database (gnomAD) (https://gnomad.broadinstitute.org/)¹² was invested to assess the variant's frequency. Furthermore, we used a Human splicing finder to detect the impact of splicing variants on splicing signals.¹³ This study was approved by the ethics committee of the Sfax Region.

3 RESULTS

3.1 Demographic findings

Overall, the genetic analysis allowed the diagnosis of *CDKL5*-related DEE in 8 out of 44 analyzed patients (18.8%). Sanger sequencing, the technique used at the beginning of the study in 20 patients, identified the pathogenic variant of *CDKL5* mutation in six patients (30%) (respectively patients 1, 2, 3, 5, 6, and 7). Later, NGS using the developed panel was applied to locate *CDKL5* mutation in two other patients (Patients 4, 8) among the remaining 24 patients (8.3%) (Table 1). There were four boys and four girls (SR male/female=1) with a mean age at the last visit of 9 years old (range 1.75–17.33 years). The mean age at the first visit to the child neurology department was 14.37 months (range 2–66 months) with a median delay to the first visit of about 20 days (ranging from 0 to 69 months). Our patients have a mean follow-up period of 7.8 years.

Three patients had consanguineous parents (P4, P5, and P6) and a family history of epilepsy (P1, P3, and P6). One patient (P6), whose sister also suffered from DEE, displayed severe epilepsy and developmental delay. Unfortunately, the sister died before getting a blood sample for genetic screening.

3.2 | Molecular findings

Five pathogenic CDKL5 variants (NM_003159.2) were identified in the eight patients in our cohort: the previously reported missense variant c.616 G>A in exon 9 (c.616 G>A; p.Asp206Asn), the frame-shift reported variant in exon 19 (c.2788insG; p.Glu930GlyfsTer9).^{14,15} A novel non-sense variant in exon 12 (c.1910_1911ins19nt; p.Leu644Ter), a novel frame-shift variant (c.149delA; p.Asn50MetfsTer26) in exon 4, and a novel splicing variant (c.2153-1G>A) in exon 15 (Table 1). All the variants detected were de novo in the patients since they were absent in their parents. The missense variant c.616 G>A in exon 9, was identified in a mosaic state in three boys (P1, P2, and P3) who were previously described,¹⁵ and in a heterozygous state in one girl (P5). The variant substitutes the highly conserved aspartate residue (Asp) with an asparagine residue (Asn) in position 206 (p.Asp206Asn) leading to conformational changes in different regions of the protein catalytic domain.¹⁵ The c.2788insG variant in exon 19 identified in patient 6 resulted in a frame-shift mutation with a consequent premature stop codon in the C-terminal domain (p.Glu930GlyfsTer9). These changes alter protein functioning and disturb the dynamic regulation of isoform levels especially hCDKL5 5 and hCDKL5 1 during pre and postnatal neurodevelopment.¹⁴ Variant c.1910 1911 ins 19nt in exon 12 (p.Leu644Ter) is also caused by the insertion of 19 nucleotides in exon 12 giving rise to a truncated protein in the C-terminal domain. The frameshift variant c.149delA (p.Asn50MetfsTer26) leads to a premature stop codon and the production of a truncated non-functional protein devoid of its catalytic domain. The last one is a splicing variant affecting a canonical region (c.2153-1G> A) predicted to alter a wild-type acceptor site according to Human Splicing Finder prediction tool. The five variants described in this study are absent from control population databases (gnomAD). In addition, the four truncating variants (c.149delA; c.2788insG; c.1910_1911 ins 19nt; c.2153-1G> A) are classified as "Likely Pathogenic" according to the ACMG classification (Table 1).

3.3 | Clinical findings

At presentation, six cases (two boys and four girls) (P3, P4, P5, P6, P7, and P8) had a global developmental delay from the start while the two remaining boys (P1 and P2) regressed with the onset of epilepsy. The main findings on examination at the first visit were poor eye contact (7/8) despite normal ophthalmologic exam, and severe tone abnormalities (7/8). Acquired microcephaly and dysmorphism were found in 6/8 and 4/8 of our patients, respectively. Common observed dysmorphic features in our patients included: a narrow forehead, low hairline, hypertelorism, deep-set eyes, ogival palate, and micrognathia. Two patients (P5 and P6) had mild hearing loss

due to retro-cochlear auditory pathways impairment, for which no genetic screening was necessary. During follow-up, four patients (three boys and one girl) improved their gross motor milestones with independent sitting position at a mean age of 30.5 months (P2 and P4) and independent walking at a mean age of 7.5 years (P1 and P3). All of them (P1, P2, P3, and P4) remained deeply impaired in cognitive skills. Other symptoms like abnormal involuntary movements, including chorea in three patients (P2, P3, and P7) and non-epileptic myoclonus in one patient (P7), as well as hand stereotypies (P1, P2, P3, P4, P5, and P7), were noted on follow-up examination with a mean delay of 15 and 33 months, respectively.

3.4 | Epilepsy and EEG findings

The first epileptic seizure occurred between 3 weeks and 8 months of age with a mean age at onset of 4 months. Age at onset of epilepsy was slightly more delayed in boys (4.9 months) than in girls (3.32 months). In the beginning, seizure types included epileptic spasms (ES) in three boys and one girl (P1, P2, P3, and P8), tonic seizures (TS) in two girls (P5 and P7), and myoclonic seizures in two cases, one boy and one girl (P4 and P6). Myoclonic seizures seemed to have earlier onset mean age (1 month) compared to TS (2months) and ES (6.75months). Awake and sleep EEG recordings in patients with ES at onset found interictal hypsarrhythmia in all four cases (P1, P2, P3, and P8) defining Infantile Epileptic Spasm Syndrome (IESS). During evolution, our patients showed two different epilepsy courses with the first made of a two-stage course in four patients (P1, P2, P3, and P4), and the second made of a three-stage course in three patients (P5, P6, and P7). The outcome of patient 8 (P8)-who had a short follow-up period limited to 12 months-needs further electro-clinical monitoring.

The critical time point determining the shift from one stage to another corresponded to the moment of marked changes in electro-clinical characteristics with the appearance of a new seizure type or significant increase in EEG interictal activity under optimal and well-conducted antiepileptic treatment. The different stages of seizures storm inconstantly alternated with periods with good seizure control, also called seizure-free periods.

Three children (P1, P2, and P3) among those who initially presented with IESS and one child (P4) out of the two patients with a myoclonic seizure at onset exhibited a two-stage epilepsy course. They displayed a different seizure type after a mean seizure-free period of 16.75 months (ranging from 4 to 32 months) and a mean time from epilepsy onset of 32 months (ranging from 12 to 68 months). Both patients 1 (P1) and 3 (P3) progressed to tonic seizures that responded well to ASMs, while patient 2 (P2) developed refractory multifocal epilepsy. Patient 4 (P4) evolved to IESS without hypsarrhythmia.

The two patients 5 (P5) and 7 (P7) who initially presented with TS, and one child (P6) with a myoclonic seizure at onset displayed a three-stage epilepsy course. Patient 5 (P5) had absence seizures as a second seizure type for a long time before he additionally presented focal motor tonic and clonic seizures defining a Lennox Gastaut Syndrome (LGS), which was responsive to ASMs. Patient 7 (P7) developed IESS with hypsarrhythmia, then refractory multifocal epilepsy. Patient 6 (P6) experienced IESS as a second epilepsy course, and subsequently, she evolved to her third stage in the form of intractable LGS.

In patients with a three-stage epilepsy course, the second seizure type occurred after a mean seizure-free period of 5.3 months (ranging from 3 to 9 months). The mean delay between the second (intermediate) and the third (ultimate) epilepsy course stage was 40 months (ranging from 10 to 96 months). This group included more girls (P5 and P7) than boys (P6), all of whom showed a poor developmental outcome. Further details regarding electroclinical data are available in Table 1.

3.5 | Brain MRI findings

Brain MRI was normal in four cases (P1, P3, P4, and P5) and demonstrated minor structural changes in the remaining patients (P2, P6, P7, and P8) with fronto-temporal cortex atrophy in four cases (P2, P6, P7, and P8) associated with thin corpus callosum in two cases (P2 and P7), and delayed myelination in patient 8 (P8) (Figure 1). Neuroimaging data are summarized in Table 1.

4 DISCUSSION

The present study categorized CDKL5 as a prominent DEE-related gene whose mutation was identified in nearly a fifth of our whole population. It identified five de novo CDKL5 gene mutations. Among these, three new variants constituted a unique finding of the present study. In addition, this paper drew epilepsy courses and key clinical features associated with CDKL5–related DEE. Indeed, CDKL5 mutation was unexpectedly found in four boys, and showed delayed onset epilepsy.^{14,15} Until 2018, only 25 mutations in *CDKL5* were reported in boys, as compared with 131 reported in females.¹⁶ These data gave evidence for female predominance. CDD is linked to the X chromosome and affects the female rather than the male gender (SR: 4/1) whatever the



FIGURE 1 The figure shows delayed myelination on Brain MRI of patients 8 performed at the age of 8 months. Axial T2 sequences (A and B) demonstrated a slight hyper-signal through the median thalamus and rostral lenticular nuclei as well as subcortical and deep white matter, While these stated brain area did not show any signal changes on axial Tl sequences (C and D).

phenotype. Indeed, males with germline variants are devoid of normal CDKL5 gene and run a lethal risk during fetal life^{5,17}; while survivors exhibit more severe disease courses. In contrast, phenotype severity in females was related to X chromosome inactivation at birth.¹⁶⁻¹⁸ In our cohort, we explain the relatively milder epilepsy phenotype in three boys by the presence of the mutation at a mosaic state in three cases.¹⁵ At the time of their first evaluation, our cases' mean age was 14.37 months (ranging from 2 to 66 months); this greatly exceeds what has been reported in a previous case series study about eight boys with CDKL5-related DEE whose ages at first examination ranged from 2 to 168 months.⁴ According to another report, the median age at the first visit was more delayed rising up to 4.7 years in females and 5.2 years in boys but the author included patients with DEE as well as those with Rett phenotype.¹⁹

Eighty percent of patients with CDD present with developmental delay, this is almost what was depicted in our series where six out of eight cases (75%) had a global developmental delay from the start. On the other hand, regression is reported in rare cases with frequent seizures or epileptic encephalopathy as it happened in two of our reports.^{2–4,18–25} In terms of clinical findings, our data met those commonly described in the literature.

Indeed, cortical visual impairment evidenced in most children as poor eye contact regardless of normal ophthalmologic exam stands for at least 75% of individuals among numerous series.^{3,22–29} Accordingly, severe tone abnormalities present in most of our patients are a wellrecognized feature of CDD.^{5,23,30} Auditory pathways impairment seen in two of our eight reports have not yet been reported to the best of our knowledge. Nonetheless, further genetic analysis assessing deafness causing gene mutations was not required. In fact, our patients displayed isolated retro-cochlear damage on Brain Evoked Response, which was an uncommon finding for hearing impairment of a genetic origin.³¹ Rett-like features including hand stereotypies occurred in seven out of eight patients after a median follow-up period of 33 months. This is consistent with the observed rate of around 80% in the literature where hand stereotypies appear since the first year of life and become more obvious over time.^{2,5,17,32,33} Other movement disorders such as chorea noted in three cases in association with myoclonus in one case among our patient cohort were previously reported.^{34–36}

According to the literature, limitation in milestones attainment across different developmental areas concerned mainly communications and fine motor skills

compared to achievements in gross motor skills. Moreover, when attained, they were significantly delayed.^{2,17,28} In a recent publication assessing girls, independent walking was attained by 22%-23% by 4 years and a half whilst only 16% of subjects could speak single words by 7 years of age.²⁸ In our study, among patients with sufficient follow-up period, just one girl and one boy acquired sitting positions at 13 and 48 months, respectively. Two boys (25%) were able to walk independently at, respectively, 7 and 8 years old while others (one boy and two girls) remained bedridden. Only two cases (one girl and one boy) were able to talk single words at 2 and 7 years, respectively. Although males are known to have a severe neurodevelopmental profile, males achieved more developmental milestones than wait in the present study. A previous study also reported four boys who acquired independent sitting positions and walking with limited communication skills. The author thus suggested a variability relative to developmental milestones attainment in males with the CDKL5. He then explained disagreement with the literature data by the limited size of male samples reported and the selection of severely affected boys for CDKL5 mutation screening.²⁸

Dysmorphic features described in four cases in our cohort (50%) were observed in a small number of patients (5.7%) of previous reports.^{5,17,18} Common observed facial features in our patients included: a narrow forehead, low hairline, hypertelorism, deep-set eyes, ogival palate, and micrognathia. Our findings were different from those describing subtle dysmorphic features including mostly broad or high forehead, deep-set eyes, deep philtrum, prominent lips, puffy phalanges, and tapered fingers.^{5,18,19,22,23,25,37} Deceleration of head growth seen in six of our eight cases (75%) exceeded the rate of 10% reported in the literature.^{3,5,19,21–25,34} We explained this finding by the fact that acquired microcephaly was one of the selection criteria for Sanger Sequencing our study cohort.

Consistent with the findings of previous studies, ES was the leading seizure type (4/8) at onset epilepsy in our cohort and appeared in later disease stages in three other cases. Other seizure types included tonic (2/8)and myoclonic (2/8) seizures, too. The estimated frequency of ES at the onset of the disease is about 23% and occurred in about 81% at some point of the disease course.^{17,35,36} According to the *CDKL5* Centers of Excellence (COEs), median epileptic spasm onset was 4 months of age ranging from 2 weeks to 36 months but the author did not specify if he considered all cases with spasms whatever the time of their occurrence.⁵ Overall, the median age of epilepsy onset is 6 weeks with 90% onset by 3 months.^{5,19,32} By contrast, our reports showed more delayed ES onset between 5 and 8 months while early-onset seizures from 3 weeks to 3 months were

Epilepsia Open[®]

more often tonic and myoclonic seizures with a mean age at onset epilepsy of 4 months (ranging from 3 weeks to 8 months) considering all seizure types. EEG at onset ES as the first seizure type demonstrated hypsarrhythmia in all cases. Among those who developed ES as a second seizure type, only two out of three displayed hypsarrhythmia on EEG. In fact, ES associated with CDD may occur without hypsarrhythmia with an EEG showing only a rare interictal epileptiform activity, or even normal.^{5,27} During the follow-up period, our patients showed two models of epileptic seizure evolution thus leading to dividing them into two groups according to their epilepsy course. The second group (P5, P6, P7) displayed the characteristic three-stage electro-clinical epilepsy with onset at the latest 3 months of age. This typical progression was first described by Bahi-Buisson et al who paid attention to three stages with the onset of epilepsy before 3 months of age (stage 1), followed by epileptic spasms (stage 2), and later multifocal refractory epilepsy (stage 3).^{2,5,6,17,26,38-40} Next, a subsequent two-stage epilepsy pattern was reported with a second stage made of hyper motor-tonic-spasms sequence occurring between 3.5 and 13 months (median 7.5 months), after a period of 1.5-11 months from onset epilepsy (stage 1) but the author did not precise first seizure type.^{2,5,6,17,26,38–40} In the present study, the twostage epilepsy course was also noted in P1, P2, P3, and P4. These latter presented with delayed ES between 5 and 8 months at onset in three boys (P1, P2, and P3) and myoclonic seizure at 40 days in one girl (P4) (stage 1) and ultimately developed polymorphous seizures (stage 2) including tonic (P1 and P3), multifocal seizures (P2) and tonic spasm (P4). These heterogeneous data arising from different clinical observations indicate the need for further studies with a greater sample size to improve our understanding of CDKL5-related epilepsy.

According to the COE cohort, seizure-free periods varied between 1 and 12 months for 32% of families while only 13% of families estimated this period to be more than 12 months. They specified this honeymoon period typically occurs before 2 years of age, yet some can experience seizure-free periods later in childhood or into their teenage years.⁵ These data are consistent with our results where patients had a median free-seizure period of around 11 months.

Regarding neuroimaging data, some researchers found no abnormalities on brain MRI²⁶ while others noticed mild frontal lobe atrophy in almost all patients.²³ This ascertainment was also seen in four of our eight patients in association with thin corpus callosum in two cases, hypomyelination, and basal ganglia signal abnormalities carrying out additional characteristics for patients with CDD. Our study added three novel pathogenic variants to the n Access

TABLE 2 The table shows clinical phenotype, epilepsy course, neuroimaging features in patient with CDKL5 mutation as reported in the present and previous study.

Study	Present study	Liang et al (2011) ²¹	Zhao et al (2014) ³⁶	Siri et al (2021) ⁴³
Number	8	12	10	50
Age (Average/months)				
First visit	14.37 (2–66)	26 (6-55)	No data	No data
Last visit	108 (21–208)	No data	31.5 (5-60)	71 (5–252)
Gender (SR: Female/Male)	4F/4M: 1	9 F/3 M: 3	9 F/1 M: 9	5 new boys and review of 45 previously reported males
Physical examination				
Acquired microcephaly	75%	16.6%	100%	20%
Dysmorphism	50%	No data	No data	20%
Neurologic features				
Poor eye contact	87.5%	No data	No data	64%
Tone abnormalities	87.5%			
Axial hypotonia	87.5%	83%	100%	68%
Limb spasticity	75%	No data	No data	26%
Autistic features				
Stereotyped movements	87.5%	41.6%	90%	8%
Movement disorders	37.5%	No data	No data	46%
Development				
Speech	12.5% (single word)	0%	10% (single word)	No data
Limited hand skills	25%	No data	100%	2%
Sitting	37.5%	16.6%	70% (12.4 months)	6%
Walking	25%	0%	10% (few steps)	4%
Epilepsy				
Age at onset (Average/ months)	4 (3 weeks–8 months)	1.5 (4 days–6 months)	1.5 (10 days–3.3 months)	5 (4 days–132 months)
First seizure type	ES (50%); TS (25%); Myoclonic (25%)	ES (75%); Focal (25%); GTC (25%); Myoclonic (8.3%)	Focal (100%); ES (10%)	ES (38%); Focal and GT
Second seizure type	ES (62.5%); Focal motor seizure tonic (37.5%) clonic (25%); TS (25%) Absence (25%)	No data	ES (80%); GT (20%); Myoclonic (20%)	Myoclonic (36%)
Disease course	Two (50%)/Three-stage course (37.5%)	No data	Two-stage course	Three-stage course
Refractory EE to ASM	50%	91.6%	100%	96%
Seizure-free period	11 (3–32 months)	No data	No data	8% (2 weeks-24 months)
EEG				
Hypsarrhythmia at onset	50%	No data	50%	16%
Hypsarrhythmia during follow-up	25%	No data	No data	22%
Brain MRI				
Fronto-temporal cortex atrophy	50%	100%	0%	28%
Thin corpus callosum	25%	0%	0%	10%
Delayed myelination	12.5%	0%	0%	0%
Vermian hypoplasia	0%	0%	0%	2%
Normal	50%	0%	100%	NA

TABLE 2 (Continued)

n	1	5
2	T	J

Study	Present study	Liang et al (2011) ²¹	Zhao et al (2014) ³⁶	Siri et al (2021) ⁴³
Genetic				
Novel/recurrent mutation	4/0	6/2	9/1	46/4
De novo	4	58.3%	100%	96%
Mutation type	Frame-shift (3) Missense (1)	Nonsense (3) Frame-shift (3) Missense (2)	Microdeletion (3) Insertion; Splicing (2) Nonsense; Missense (1)	Missense (2/5) Frame-shift; in-frame deletion; splice site (1/5)
Genotype–phenotype correlation	Seizure control in three cases with MS mutation	Seizure control in one patient with MS mutation	Seizure control and in one patient with splicing mutation	Late onset epilepsy in one patient with MS mutation

previously set of published mutations and thus gave a continuation for earlier reviews. In that issue, most reported *CDKL5* mutations were also sporadic and considered as de novo with a very small number of recurrent variants.²¹ More data about *CDKL5* mutation-related disorder according to literature are available in Table 2.

Although the phenotypes of our patients overlap at some points with those reported in the literature, we noted a few such as milder phenotypes in boys, hearing loss, and dysmorphic traits. Furthermore, our cohort showed phenotypic heterogeneity with inconsistent disease outcome and severity between patients even in those carrying the same mutation indicating probably epigenetic factors implication in the determinism of gene expression. This hypothesis may explain our failure and that of previous studies to establish any genotype–phenotype correlation.^{5,27,29} Although some authors thought that frameshift mutations in the C-terminal region cause more severe phenotypes as was the case in our sixth (P6) and seventh (P7) reports; the relationship between genotype and phenotype is still ambiguous.^{36,41,42}

5 | CONCLUSION

Our data as well as those of the literature well demonstrated CDD is responsible for DEE. The present study interestingly described new phenotypic features such as hearing loss and peculiar dysmorphic traits. We also depicted more reports relative to unusual benign developmental profiles in boys carrying CDKL5 mutation, different patterns of CDKL5-epilepsy, neuroimaging findings, and CDKL5 mutational spectrum. However, we lacked information about CDKL5-related comorbidities such as sleep disorders, and respiratory and feeding problems. Consequently, a screen for CDKL5 mutations is mandatory in the setting of early-onset epilepsy mainly IS and associated Rett-like features independently of associated unusual phenotypic findings. However, the fact that previous studies described CDKL5 in boys,⁴³ and half of our sample individuals with CDKL5 mutations were male warrants the necessity of considering the analysis of CDKL5 in boys with DEE, too.

AUTHOR CONTRIBUTIONS

Chahnez Charfi Triki involved in conceptualization, editing, and supervision. Salma Zouari Mallouli involved in data curation, first draft redaction, and editing. Fatma Kamoun Feki and Sarah weckhuysen involved in editing and supervision. Marwa Ben Jdila, Faiza Fakhfakh, Mariem Ben Said, and Sabeur Masmoudi involved in genetic analysis and editing.

CONFLICT OF INTEREST STATEMENT

Neither of the authors has any conflict of interest to disclose. We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ETHICS STATEMENT

The consent form used in the present study is approved by the regional ethics committee.

PATIENT CONSENT STATEMENT

Informed consent of children's caregivers was obtained.

ORCID

Chahnez Charfi Triki [®] https://orcid. org/0000-0003-2918-3819 Salma Zouari Mallouli [®] https://orcid. org/0000-0002-2298-3613 Mariem Ben Said [®] https://orcid. org/0000-0003-4098-4537 Fatma Kamoun Feki [®] https://orcid. org/0000-0002-5058-7693

REFERENCES

916

- Eyers PA. A new consensus for evaluating CDKL5/STK9dependent signalling mechanisms. EMBO J. 2018;37:e100848. https://doi.org/10.15252/embj.2018100848
- Bahi-Buisson N, Bienvenu T. CDKL5-related disorders: from clinical description to molecular genetics. Mol Syndromol. 2012;2:137–52.
- Bahi-Buisson N, Nectoux J, Rosas-Vargas H, Milh M, Boddaert N, Girard B, et al. Key clinical features to identify girls with CDKL5 mutations. Brain. 2008;131:2647–61.
- Mirzaa GM, Paciorkowski AR, Marsh ED, Berry-Kravis EM, Medne L, Alkhateeb A, et al. CDKL5 and ARX mutations in males with early-onset epilepsy. Pediatr Neurol. 2014;48:367–77.
- Olson HE, Demarest ST, Pestana-Knight EM, Swanson LC, Iqbal S, Lal D, et al. Cyclin-dependent kinase-like 5 deficiency disorder: clinical review. Pediatr Neurol. 2019;97:18–25.
- 6. Sartori S, di Rosa G, Polli R, Bettella E, Tricomi G, Tortorella G, et al. A novel CDKL5 mutation in a 47,XXY boy with the early-onset seizure variant of Rett syndrome. Am J Med Genet A. 2009;149A:232–6.
- Zuberi SM, Wirrell E, Yozawitz E, Wilmshurst JM, Specchio N, Riney K, et al. ILAE classification and definition of epilepsy syndromes with onset in neonates and infants: position statement by the ILAE task force on nosology and definitions. Epilepsia. 2022;63:1349–97.
- Fisher RS, Cross JH, French JA, Higurashi N, Hirsch E, Jansen FE, et al. Operational classification of seizure types by the International League Against Epilepsy: position paper of the ILAE Commission for Classification and Terminology. Epilepsia. 2017;58:522–30.
- Scheffer IE, Berkovic S, Capovilla G, Connolly MB, French J, Guilhoto L, et al. ILAE classification of the epilepsies: position paper of the ILAE Commission for Classification and Terminology. Epilepsia. 2017;58:512–21.
- Desvignes J-P, Bartoli M, Delague V, Krahn M, Miltgen M, Béroud C, et al. VarAFT: a variant annotation and filtration system for human next generation sequencing data. Nucleic Acids Res. 2018;46:W545–53.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17:405–24.
- 12. gnomAD. Genome Aggregation. Available from: https://gnomad.broadinstitute.org/
- Desmet F-O, Hamroun D, Lalande M, Collod-Béroud G, Claustres M, Béroud C. Human splicing finder: an online bioinformatics tool to predict splicing signals. Nucleic Acids Res. 2009;37:e67.
- 14. Jdila MB, Triki C, Rhouma BB, Jomaa RB, Issa AB, Ammar-Keskes L, et al. A novel C-terminal truncated mutation in hCDKL5 protein causing a severe West syndrome: comparison with previous truncated mutations and genotype/phenotype correlation. Int J Dev Neurosci. 2019;72:22–30.
- Jdila MB, Issa AB, Khabou B, Rhouma BB, Kamoun F, Ammar-Keskes L, et al. Novel mutations in the CDKL5 gene in complex genotypes associated with West syndrome with variable phenotype: first description of somatic mosaic state. Clin Chim Acta. 2017;473:51–9.

- 16. Liang J-S. Phenotypic manifestations between male and female children with CDKL5 mutations. Brain Dev. 2019;41:783–9.
- Jakimiec M, Paprocka J, Śmigiel R. CDKL5 deficiency disorder-a complex epileptic encephalopathy. Brain Sci. 2020;10:107.
- Archer HL, Evans J, Edwards S, Colley J, Newbury-Ecob R, O'Callaghan F, et al. CDKL5 mutations cause infantile spasms, early onset seizures, and severe mental retardation in female patients. J Med Genet. 2006;43:729–34.
- Fehr S, Wilson M, Downs J, Williams S, Murgia A, Sartori S, et al. The CDKL5 disorder is an independent clinical entity associated with early-onset encephalopathy. Eur J Hum Genet. 2013;21:266–73.
- Fehr S, Downs J, Ho G, de Klerk N, Forbes D, Christodoulou J, et al. Functional abilities in children and adults with the CDKL5 disorder. Am J Med Genet A. 2016;170:2860–9.
- 21. Liang J-S, Shimojima K, Takayama R, Natsume J, Shichiji M, Hirasawa K, et al. CDKL5 alterations lead to early epileptic encephalopathy in both genders. Epilepsia. 2011;52:1835–42.
- Mei D, Marini C, Novara F, Bernardina BD, Granata T, Fontana E, et al. Xp22.3 genomic deletions involving the CDKL5 gene in girls with early onset epileptic encephalopathy. Epilepsia. 2010;51:647–54.
- 23. Olson HE, Poduri A. CDKL5 mutations in early onset epilepsy: case report and review of the literature. J Pediatr Epilepsy. 2012;1:151–9.
- Russo S, Marchi M, Cogliati F, Bonati MT, Pintaudi M, Veneselli E, et al. Novel mutations in the CDKL5 gene, predicted effects and associated phenotypes. Neurogenetics. 2009;10:241–50.
- Sartori S, Polli R, Bettella E, Rossato S, Andreoli W, Vecchi M, et al. Pathogenic role of the X-linked cyclin-dependent kinase-like 5 and aristaless-related homeobox genes in epileptic encephalopathy of unknown etiology with onset in the first year of life. J Child Neurol. 2011;26:683–91.
- Artuso R, Mencarelli MA, Polli R, Sartori S, Ariani F, Pollazzon M, et al. Early-onset seizure variant of Rett syndrome: definition of the clinical diagnostic criteria. Brain Dev. 2010;32:17–24.
- 27. Mefford HC. Phenotype to genotype and Back again. Epilepsy Curr. 2020;20:88–9.
- 28. Fehr S, Leonard H, Ho G, Williams S, de Klerk N, Forbes D, et al. There is variability in the attainment of developmental milestones in the CDKL5 disorder. J Neurodev Disord. 2015;7:2.
- Demarest ST, Olson HE, Moss A, Pestana-Knight E, Zhang X, Parikh S, et al. CDKL5 deficiency disorder: relationship between genotype, epilepsy, cortical visual impairment and development. Epilepsia. 2020;60:1733–42.
- Fehr S, Wong K, Chin R, Williams S, de Klerk N, Forbes D, et al. Seizure variables and their relationship to genotype and functional abilities in the CDKL5 disorder. Neurology. 2016;87:2206–13.
- Michalski N, Petit C. Central auditory deficits associated with genetic forms of peripheral deafness. Hum Genet. 2022;141(3– 4):335–45. https://doi.org/10.1007/s00439-021-02339-3
- 32. Frullanti E, Papa FT, Grillo E, Clarke A, Ben-Zeev B, Pineda M, et al. Analysis of the phenotypes in the Rett networked database. Int J Genomics. 2019;2019:6956934.
- Saitsu H, Osaka H, Nishiyama K, Tsurusaki Y, Doi H, Miyake N, et al. A girl with early-onset epileptic encephalopathy

associated with microdeletion involving CDKL5. Brain Dev. 2012;34:364-7.

- 34. Nemos C, Lambert L, Giuliano F, Doray B, Roubertie A, Goldenberg A, et al. Mutational spectrum of CDKL5 in early-onset encephalopathies: a study of a large collection of French patients and review of the literature. Clin Genet. 2009;76:357–71.
- Wong VC-N, Kwong AK-Y. CDKL5 variant in a boy with infantile epileptic encephalopathy: case report. Brain Dev. 2015;37:446-8.
- 36. Zhao Y, Zhang X, Bao X, Zhang Q, Zhang J, Cao G, et al. Clinical features and gene mutational spectrum of CDKL5-related diseases in a cohort of Chinese patients. BMC Med Genet. 2014;15:24.
- Elia M, Falco M, Ferri R, Spalletta A, Bottitta M, Calabrese G, et al. CDKL5 mutations in boys with severe encephalopathy and early-onset intractable epilepsy. Neurology. 2008;71:997–9.
- Grosso S, Brogna A, Bazzotti S, Renieri A, Morgese G, Balestri P. Seizures and electroencephalographic findings in CDKL5 mutations: case report and review. Brain Dev. 2007;29:239–42.
- 39. Klein KM, Yendle SC, Harvey AS. A distinctive seizure type in patients with CDKL5 mutations: hypermotor-tonic-spasms sequence. Neurology. 2011;76:1436–8.
- 40. Melani F, Mei D, Pisano T, Savasta S, Franzoni E, Ferrari AR, et al. CDKL5 gene-related epileptic encephalopathy:

electroclinical findings in the first year of life. Dev Med Child Neurol. 2011;53:354–60.

- Bahi-Buisson N, Villeneuve N, Caietta E, Jacquette A, Maurey H, Matthijs G, et al. Recurrent mutations in the CDKL5 gene: genotype-phenotype relationships. Am J Med Genet A. 2012;158A:1612–9.
- 42. Kilstrup-Nielsen C, Rusconi L, La Montanara P, Ciceri D, Bergo A, Bedogni F, et al. What we know and would like to know about CDKL5 and its involvement in epileptic encephalopathy. Neural Plast. 2012;2012:728267.
- 43. Siri B, Varesio C, Freri E, Darra F, Gana S, Mei D, et al. CDKL5 deficiency disorder in males: five new variants and review of the literature. Eur J Paediatr Neurol. 2021;33:9–20.

How to cite this article: Charfi Triki C, Zouari Mallouli S, Ben Jdila M, Ben Said M, Kamoun Feki F, Weckhuysen S, et al. First report of Tunisian patients with *CDKL5*-related encephalopathy. Epilepsia Open. 2024;9:906–917. <u>https://doi.org/10.1002/</u> epi4.12824