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# **Review**

# **Novel approaches to the treatment of small-cell lung cancer**

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Abstract. Small-cell lung cancer (SCLC) is character- antibodies and immunoconjugates as well as growth ized by its initial responsiveness to chemotherapy and factor antagonists and inhibitors. In addition, recent the appearance of early metastases. Although combina- advances in understanding the biology of SCLC have tion chemotherapy, in some instances together with stimulated new investigations searching to counter the radiation, has improved the prognosis of this disease, in molecular basis underlying the increased proliferation most patients SCLC ultimately recurs in a drug-resis- and the apoptosis deficiency of SCLC cells. This can tant form. Several new strategies for the eradication of be achieved using antisense oligodeoxynucleotides that SCLC are being explored at the preclinical level. The repress the expression of growth factor receptors and identification of selective target molecules on the surface anti-apoptosis genes, or by gene replacement to comof SCLC cells, together with the progress made in pensate for the loss or inactivation of tumor suppresantibody engineering, have provided new generations of sor genes.

**Key words.** Antisense; antibodies; growth factor inhibitors; chemotherapy; gene therapy; radiation therapy; small-cell lung cancer.

Lung cancer is the leading cause of cancer-related deaths, and its incidence continues to rise worldwide [1]. This cancer can be classified in two major histological types. Roughly 80% are non-small-cell lung cancers (NSCLCs), comprising adenocarcinoma, large-cell carcinoma, and squamous cell carcinoma. Most of the remaining cases are classified as small-cell lung cancer (SCLC), which has a different biological behavior, history, and treatment. SCLC is a very aggressive disease, characterized by rapid growth and early dissemination. In the seventies, SCLC was shown to be responsive to chemotherapy. While the median survival before the introduction of chemotherapy was  $2-4$  months [2], combination chemotherapy has increased survival about fourfold and resulted in long-term survival or cure in a small proportion of patients [3]. In most patients, however, the tumor relapses in a form resistant to any further cytotoxic treatment.

During the past decade, numerous studies have addressed the molecular and genetic mechanisms underlying the development and progression of malignant tumors. In the case of SCLC, many abnormalities have been identified, which allow the development of approaches that specifically target the molecular basis of this disease. Together with the improvements made in the field of drug development and delivery, this has opened new avenues for more effective treatment of SCLC, and may help to improve the treatment outcome of this hitherto incurable disease. This review will summarize the most prominent advances in the treatment of SCLC provided by preclinical and clinical studies dur- \* Corresponding author. ing the last decade, which deserve further investigation:

(i) recent improvements in chemo- and radiation therapy; (ii) approaches to target cell surface antigens, autocrine growth factors and their receptors, and modulators of drug resistance; (iii) approaches to target the molecular basis of increased proliferation and apoptosis deficiency of tumor cells, such as the use of bcl-2 antisense oligodeoxynucleotides or the replacement of inactivated or lost tumor suppressor genes. A summary of these issues is shown in figure 1.

## **Combination chemotherapy**

Back in the sixties it was shown that local therapy, either surgery or radiation alone, had no significant impact on the survival of SCLC patients [4, 5]. In the seventies, several chemotherapeutic drugs with activity against SCLC were identified, including nitrogen mustard, cyclophosphamide, methothrexate, doxorubicin, etoposide, procarbacine, and vincristine. It was also shown that a combination of up to three drugs yields better results than individual drugs used alone or sequentially. In the eighties, one new active drug, cisplatin, was identified, and several analogs of older agents, with similar modes of action but somewhat different toxicity profiles, were reported, including ifosfamide, epirubicin, teneposide, vindesin, and carboplatin. Based on many clinical studies, combination therapy with cisplatin and etoposide, or cyclophosphamide, doxorubicin and vincristine, both combina-



Figure 1. Overview of the various treatment approaches discussed in this review.

tions given for four to six cycles of treatment at 3 weekly intervals, has become the standard therapy of this disease [6].

Advances in chemotherapy reached a plateau in the eighties, despite many clinical trials testing further strategies to improve the results. Among these strategies were the intensification of drug delivery by weekly chemotherapy [7], the use of higher than conventional doses of standard chemotherapy, with or without hematopoietic growth factors [8, 9], the use of the anticoagulant warfarin together with chemotherapy [10], interferon maintenance [11], and the use of megestrol acetate to increase weight gain together with chemotherapy [12].

Currently, several new agents have been identified as having good activity in SCLC, including taxanes [13], gemcitabine [14], vinorelbine [15], and topotecan [16]. However, how these new agents should best be integrated in combination regimens and whether their inclusion will improve the outcome of SCLC have not yet been determined.

## **Radiation therapy**

The role of thoracic radiotherapy for limited-disease SCLC, i.e., SCLC confined to one hemithorax, has long been controversial. A meta-analysis of 13 randomized trials has settled this issue by demonstrating a moderate 5% survival benefit at 3 years for the use of combined chemotherapy and radiation therapy [17]. A recent publication from a large study showed that hyperfractionation compared to conventional fractionation gave a 5-year survival benefit from 16 to 26% [18]. For patients responding to therapy, relapses in the brain may occur frequently. To decrease the rate of brain relapses, many investigators suggested the use of prophylactic cranial irradiation. While it has been undisputed that such prophylactic cranial irradiation does reduce the occurrence of brain metastasis, the potential for neuropsychiatric side effects and lack of proof of a survival benefit left many physicians doubtful about the standard use of this treatment. Recent studies, however, failed to demonstrate an impairment of cognitive function and a meta-analysis demonstrated a significant survival benefit in complete responders from 15 to 20% at 3 years, suggesting prophylactic cranial irradiation should be offered to patients in complete remission  $[19-21]$ .

#### **Targeting cell surface antigens**

Antibodies recognizing surface antigens selectively expressed on tumor cells are potential therapeutic agents with high specificity. Recently, the efficacy of the antibody approach to cancer therapy has been revolutionized by such advances as the identification of novel antigens which are homogeneously expressed in the tumor but have low expression in normal vital tissues, and by genetic engineering techniques to produce tailormade targeting vehicles with reduced immunogenicity and improved effector functions. An impressive example is given by the HER-2/neu-specific humanized antibody Herceptin, which enhanced the chemosensitivity of tumor cells and lengthened remission times in patients with refractory metastatic breast cancer [22]. HER-2/neu is also over-expressed in a large proportion of NSCLCs, but is rarely associated with SCLC. In addition to the use of intact antibodies to recruit host effector cells and complement components or to inhibit growth factor signaling, rationally engineered full-size antibodies and single-chain Fv fragments have also been used in imaging and therapy studies to deliver radioisotopes, anti-tumor effector molecules, or immune response modifiers to solid tumor cells [23–26]. Single-chain Fv fragments are small molecules of about 25 – 30 kDa with improved tumor penetration ability. They can be genetically fused with effector proteins or chemically coupled with chemotherapeutic agents. Recent progress in phage display technology has expanded our ability to select human antibodies and construct an array of derivatives with desirable clinical properties. This technique also provides a powerful tool to search for novel target molecules selectively expressed on the surface of tumor cells.

The therapeutic efficacy of antibodies and antibody-effector conjugates not only depends on their tumortargeting potential, but also on the intrinsic properties of the antigen. For example, it is mandatory that the antigen is differentially expressed between tumors and normal tissues, that it is uniformly expressed by the tumor cell population, and that it is not shed from the cell surface or down-modulated upon antibody binding. Moreover, if the target site of a conjugate is located intracellularly, the antigen-antibody complex must be internalized from the surface by receptor-mediated endocytosis. In an attempt to develop novel antibodybased treatment approaches, there has been a collaborative effort between individual laboratories. The Second and Third International Workshops on Lung Tumor Antigens addressed the characterization and definition of surface molecules expressed on lung cancer cells and recognized by monoclonal antibodies [27, 28]. As a result, the nature and biological function of many of these antigens have been characterized. Many of the antibodies that recognize cell surface antigens associated with SCLC have also been investigated in vivo, e.g., in models of SCLC xenografts. These models are useful tools to assess the tumor localization and anti-tumor activity of antibodies and antibody-

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Table 1. Potential targets on SCLC cells for antibody-based diagnosis and therapy.	
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based immunoconjugates under complex physiological conditions. Since most of the antibodies tested do not cross-react with murine tissues, however, their clinical usefulness cannot be fully assessed in animal models. For example, antibodies recognizing CD24 and CD56 (neural cell adhesion molecule, NCAM), antigens also expressed on human hematopoietic cells but not their murine counterparts, efficiently localized to SCLC xenografts in mice [29, 30]. The antibodies, however, failed to detect tumors in patients, where they mainly localized to recticuloendothelial tissues [31, 32]. Based on these observations and the finding that most of the tumor-associated antigens are also expressed in normal vital tissues, only a few antigens remain that may serve as potential targets for antibody-based diagnosis and therapy of SCLC (table 1). The preclinical and clinical studies describing the use of these candidate antigens are reviewed below.

## **Neural cell adhesion molecule**

NCAM, also known as CD56 or NKH01, was designated as a cluster-1 antigen on lung tumor cells [28]. NCAM is a homophilic cell adhesion molecule of 145 kDa abundantly expressed in virtually all SCLC cell lines and cells derived from tumor specimens [33, 34]. In normal tissues, NCAM expression is prominent in nerves, endocrine cells, natural killer (NK) cells and striated and cardiac muscle [35]. Monoclonal antibodies with specificity for NCAM were successfully used for imaging and therapy of SCLC xenografts growing in athymic mice [30, 36]. In a recent phase I study, 21 patients with relapsed or refractory SCLC were treated with the monoclonal antibody N901 which was conjugated with the protein toxin blocked ricin (bR) [37]. Patients received the immunotoxin N901-bR daily for 7 days by continuous infusion. Dose-limiting toxicity was vascular leak syndrome, whereas neuropathy was not observed. Most patients developed either human antimouse antibodies (HAMAs) or antibodies against the ricin moiety of the immunotoxin (HARA). One patient achieved a partial response which persisted for 3 months, 6 patients had stable disease. Although this result is encouraging, in view of the finding of Ornadel et al. [32] that NCAM antibodies do not efficiently localize to tumors in patients, the mechanisms underlying the clinical efficacy of N901-bR remain unclear.

## **Epithelial glycoprotein-2 (EGP-2)**

EGP-2, synonymously named Ep-CAM, EGP40, KSA, or GA733-2, was designated as a cluster-2 antigen on lung tumor cells [28]. EGP-2 was first cloned from a lung adenocarcinoma cell line by Strnad et al. [38]. It is a homophilic cell adhesion molecule of 40 kDa abundantly expressed on a variety of epithelial tumor cells, including a major fraction of SCLC [39]. Expression of EGP-2 is dynamically regulated during cell growth and differentiation, and recent investigation has identified its role in tissue invasion and its up-regulation in cells under growth-promoting conditions [40, 41]. The latter finding might explain the different levels of EGP-2 expression, which is high in malignant and low in normal epithelial cells. In a 7-year follow-up study in patients with advanced colorectal cancer, the mouse monoclonal antibody 17-1A (Panorex) produced significant anti-tumor effects [42], and a humanized recombinant variant is currently under clinical investigation in patients with refractory prostate, lung, or colon cancer. The mouse monoclonal antibody MOC31 was used for imaging studies in patients with SCLC [43], and is currently being re-engineered by our group in a collaborative effort to produce humanized scFv variants with high antigen-binding affinity [44].

#### **Lewis Y antigen**

A blood-group-related carbohydrate, the Y difucosylated hapten (Lewis Y antigen), also designated as a cluster-6 antigen on lung tumor cells [28], is associated with  $60 - 90\%$  of human tumors of epithelial origin, including breast, colon, and gastric cancer, and SCLC [45 – 48]. Its level of expression correlates with survival in patients with lung cancer [49]. The Lewis Y antigen is expressed in a stage-specific manner during embryonic development of the human lung, and its emergence on lung tumor cells appears to be the result of retrodifferentiation of cells to the stages of embryonic lung cells [50]. Despite high lytic activity in the serum of patients with SCLC, the mouse monoclonal antibody ABL364 could not produce clinical responses [51]. Cytotoxic immunoconjugates targeting the Lewis Y antigen have also been tested in preclinical and clinical studies [52, 53]. The immunotoxin LMB-1, composed of the monoclonal antibody B3 chemically linked to a genetically engineered form of *Pseudomonas* Exotoxin A, was used in a phase I study of 38 patients with solid tumors who failed conventional therapy [53]. Objective anti-tumor activity was observed in 5 patients, 18 had stable disease, 15 progressed. A complete remission was observed in a patient with metastatic breast cancer. A greater than 75% tumor reduction and resolution of all clinical symptoms lasting for more than 6 months was observed in a colon cancer patient with extensive retroperitoneal and cervical metastasis. Results from clinical trials with LMB-7, an immunotoxin which employs a humanized scFv version of B3 [54], and with LMB-9, a scFv stabilized by an interchain disulfide instead of a peptide linker, are awaited. These studies will reveal whether the impressive anti-tumor activity of improved second-generation immunotoxins in vitro and in animal models translates into therapeutic efficacy in patients.

#### **Gangliosides**

Gangliosides are glycosphingolipids, composed of a ceramide portion that anchors it into the plasma membrane and a carbohydrate moiety oriented to the extracellular space. Due to differences in the carbohydrate residues, many distinct gangliosides have high levels of expression particularly in neuroendocrine tumors such as melanoma and SCLC, but low levels of expression in normal tissues. Gangliosides are reported to be involved in the attachment of tumor cells to extracellular matrix proteins [55]. Based on their differential expression in tumors, the gangliosides GD2, GD3, and GM2 represent the most promising targets of antibody therapy [56-58]. A mouse-human chimeric antibody targeting GM2 was reported to induce effector-cell-mediated lysis of SCLC cells in vitro [58], and antibodies targeting GD2 demonstrated tumor-localizing and therapeutic potential in patients with melanoma and neuroblastoma [59, 60]. In a recent pilot study, the mouse monoclonal antibody 3F8 was used to target residual disease in SCLC patients [61]. Twelve patients were enrolled in this trial and 10 received intravenous 3F8 labeled with 131I. Five patients had recurrent or progressive disease after chemotherapy, 7 patients were subjected to diagnosis prior to the initiation of chemotherapy. Radionuclide scans demonstrated localization to all known sites of disease, apart from small brain metastases in 1 patient.

Anti-GD3 antibodies were also used to raise anti-idiotypic antibodies against GD3 in mice. In recent clinical trials, the anti-idiotypic antibody BEC2 was used to immunize patients with melanoma and SCLC [62]. Fifteen patients, 8 of whom presented with extensive disease, were enrolled in the study. BEC2 was administered intradermaly together with bacille Calmette-Guérin as an immune adjuvant. All patients developed anti-BEC2 antibodies. Five of 13 patients evaluable for serologic response developed anti-GD3 IgG antibodies. For these patients, survival from the time of initial diagnosis was significantly improved compared to the median survival of historical control groups. In view of these encouraging results, further clinical studies have been initiated to evaluate the potential of anti-ganglioside and anti-idiotypic ganglioside therapy in SCLC.

# **Targeting growth factors and growth factor receptors**

An autocrine growth loop in cancer was first described as a potential growth pathway for tumor cells more than 17 years ago [63]. A variety of neuroendocrine peptides such as the bombesin-like peptide gastrin-releasing peptide (GRP), and insulin-like growth factor-I (IGF-1) fit the definition of autocrine growth factors for SCLC. With the aim of inhibiting the growth of SCLC cells, these growth factors and their receptors have been evaluated as potential targets for various antagonists including antibodies and for antisense therapeutics.

## **Neuroendocrine growth factors**

SCLC is a neuroendocrine tumor which, according to the degree of neuroendocrine differentiation, is divided into a classic and a variant phenotype [64]. Classic SCLC accounts for 70% of all SCLCs. These tumor cells are characterized by dense-core neurosecretory granules and the production of high levels of neuroendocrine growth factors and peptide hormones that act as autocrine growth factors, such as dopa decarboxylase, neuron-specific enolase, and bombesin-like peptides. The bombesin-like GRP was initially cloned from lung carcinoid tumors and SCLC cell lines. It is a 27-amino-acid peptide produced from a large precursor protein of 145 amino acids that binds to its receptor with high affinity in the nanomolar range [65, 66] stimulating the growth of SCLC cells by elevating cytosolic  $Ca^{2+}$  and activating protein kinase C [67-69]. Synthetic GRP receptor antagonists and antibodies neutralizing secreted GRP have been used in a variety of preclinical studies to interrupt the autocrine and paracrine growth factor loops, and to inhibit the proliferation and clonogenicity of SCLC cells in vitro and in vivo. The broad-spectrum receptor antagonist substance P was identified as a potent analog in inhibiting bombesin-like peptide receptor binding in vitro [70]. The results of a phase II trial with the GRP-neutralizing monoclonal antibody 2A11 were published recently [71]. Patients with relapsed or refractory SCLC received 12 doses of 2A11 over a 4-week period. Twelve of 13 patients were eligible: 1 had a complete response to treatment, 4 achieved stable disease, and 7 showed progression. The responding patient received another cycle of treatment with 2A11 and remained without evidence of progression for 5 months. Since the growth of SCLC cells depends on multiple growth factors, more studies are necessary to determine the therapeutic potential of bombesin-like peptide inhibitors in patients with SCLC.

# **Insulin-like growth-factor-I**

IGF-I is a 70-amino-acid growth factor found at elevated levels in more than 90% of SCLC tumors [72-74]. It is involved in mitogenic signaling by binding to tyrosine kinase receptors causing phosphorylation of protein substrates and cell growth [75]. Specifically, the apoptosis-inhibiting pathway of IGF-I seems to route through Ras, PI-3 kinase, and the serine-threonine kinase Akt/protein kinase B [76, 77], ultimately impacting on Bad, a key modulator of Bcl-2 [78]. The IGF-I receptor is expressed in many human tissues and tumors including NSCLC and SCLC [74]. It is regulated by p53 and functions as an autocrine growth factor implicated in the apoptotic response of cells [79]. Analogously to the GRP autocrine growth loop, the IGF-I pathway can also be inhibited using receptor antagonists or ligands neutralizing the growth factor. Antibodies that block the specific binding of IGF-I to its receptor, such as the monoclonal antibody  $\alpha$ IR-3, are able to inhibit the growth of SCLC and NSCLC cells in vitro and in vivo [80].

IGF-I signaling in cells is inhibited by antisense oligodeoxynucleotides (ODNs) that repress expression of the IGF-I receptor. This has been demonstrated by Resnicoff et al. [81] who used antisense ODNs to reduce the number of IGF-I receptors on melanoma cells to inhibit their tumorigenesis in vivo. The antisense approach to inhibition of cancer-related gene expression has become particularly interesting since many of the initial limitations associated with antisense ODNs, such as limited stability, poor specificity, unpredictable targeting, and undesired non-antisense effects, have now been solved by chemical modifications [82, 83]. In addition, improvements in the manufacturing process have dramatically reduced the overall costs of ODN synthesis [84]. Antisense ODNs are fast, simple, and cost-effective tools available for validating new therapeutic targets, commonly the first step in modern drug development. The ability of these compounds to inhibit the expression of genes involved in cell growth and apoptosis regulation in a sequence-specific manner has been demonstrated in various preclinical and clinical studies [85 – 87].

## **Targeting the expression of drug transport proteins**

The issue of drug resistance in SCLC has been addressed extensively and genetic abnormalities that might contribute to the resistance phenotype, such as over-expression of genes coding for drug efflux pumps located in the plasma membrane, have been identified [88, 89]. The role of drug transporters in clinical drug resistance, however, remains unclear. Whereas the absence of MDR1 gene expression during chemotherapy of SCLC indicated a favorable prognosis in one study [90], in another study, no correlation between MDR1 gene expression in SCLC and NSCLC cells and chemosensitivity and clinical response to therapy was detected [91]. Recently, novel second-generation antisense ODNs targeting the MDR1 mRNA were used to down-regulate P-glycoprotein expression and increase drug uptake into cells in vitro [92]. To what extent this approach can help overcome clinical drug resistance in SCLC and other refractory tumors remains to be determined.

# **Targeting the expression of oncogenes and apoptosis inhibitors**

Considerable evidence has accumulated that cancer has a genetic origin based on the occurrence of mutations in families of genes implicated in DNA repair, growth control, and apoptosis [93-95]. This provides researchers with the opportunity to specifically target genetic lesions and abnormal gene products by therapeutic interventions. The most extensively investigated and advanced approaches to gene therapy of cancer are: (i) the use of antisense ODNs to inhibit the expression of oncogenes; (ii) replacement of inactivated or lost tumor suppressor genes; (iii) ex vivo transfection of autologous tumor cells with cytokine genes and their use as vaccines to stimulate an anti-tumor immune response.

#### **The bcl-2 family of apoptosis inhibitors**

Current research has provided evidence that the inability of tumor cells to undergo programmed cell death or apoptosis in response to cytotoxic damage is a critical determinant of drug resistance [96, 97]. The search for gene products involved in the regulation of cell growth and apoptosis has identified numerous proteins, and the list continues to grow. In particular, much interest has been devoted to members of the bcl-2 gene family of apoptosis regulators, because of their contribution to oncogenesis and drug resistance. This family of structurally related proteins includes members that exert opposing functions in apoptosis control. The Bcl-2 protein functions in normal and neoplastic cells to inhibit or delay apoptosis induced by a variety of endogenous and exogenous stimuli [98, 99]. The related Bcl-xL protein also acts as a negative regulator of apoptosis and protects cells from cell death where Bcl-2 is ineffective [100]. Over-expression of bcl-2 has been found in  $70-90\%$ of SCLC cell lines and tumor specimens [101 – 103]. We tested a number of first-generation phosphorothioate ODNs hybridizing to different regions of the bcl-2 mRNA [85]. These experiments revealed a sequence in the coding region as the most effective target site for antisense-mediated down-regulation of bcl-2 expression and induction of apoptosis in SCLC cells. Moreover, repression of the bcl-2 gene using this antisense ODN (2009) was shown to sensitize SCLC cells to chemotherapy, resulting in a synergistic cytotoxic effect [104]. In a recent phase I trial, the phosphorothioate antisense ODN G3139, which targets the translation initiation site of the bcl-2 mRNA, was administered as a 14-day continuous subcutaneous infusion in nine patients with relapsing non-Hodgkin lymphoma[105]. A reduction in tumor size was achieved in two patients, and in another two patients, the number of circulating lymphoma cells decreased during treatment. The dose-limiting toxicity of G3139 was related to the unspecific protein-binding activity of the phosphorothioate backbone [106]. The use of secondgeneration bcl-2 antisense ODNs with improved biological and pharmacological properties is under consideration in patients with refractory tumors. In addition to bcl-2, the anti-apoptotic bcl-xL gene is also expressed in a number of human tumors of different

histological origin including SCLC [73, 107, 108]. Since expression of bcl-xL also substantially contributes to resistance of tumor cells to a broad range of anti-cancer agents, it represents a further promising target for antisense therapeutics. That bcl-xL antisense ODNs can indeed induce apoptosis in SCLC cells is shown by the electron micrographs in figure 2. The role of bcl-xL antisense ODNs in cancer therapy and other proliferative diseases is currently under preclinical evaluation by several investigators.

## **The myc family of oncogenes**

In addition to the bcl-2 family members of apoptosis inhibitors, the myc family of oncogenes, which is transcriptionally deregulated or amplified in many human tumor cells including a significant proportion of SCLC cells [109 – 111] also represents an attractive target for gene repression approaches. The Myc phosphoproteins are transcription factors of the bHLH-zip family and there is substantial evidence for their role in the control of cell proliferation. In certain circumstances, however, Myc also promotes apoptosis  $[112-114]$ . Although the precise mechanism for this function remains obscure, Myc-induced apoptosis is efficiently suppressed by Bcl-2, which acts as a downstream inhibitor [115], and by survival signals, such as IGF-I or interleukin (IL)-3 [76, 116]. The higher level of Myc found in pretreated patients compared to patients prior to chemotherapy [117, 118] implies that it might be a negative regulator of apoptosis in SCLC cells. This finding is consistent with studies demonstrating poor response to chemotherapy and worse survival in patients with myc gene over-expression [119]. Antisense ODNs targeting the c-myc mRNA were used to inhibit the growth of NSCLC and to induce apoptosis in leukemia and melanoma cells in vitro and in vivo [120 – 122].

#### **Cyclins**

The passage of a cell through the cell cycle is controlled by cytoplasmic proteins, the most critical being cyclins and cyclin-dependent kinases (cdks), and alterations in cyclin expression provide the earliest examples of cell cycle regulators acting as oncogenes [123]. Cyclin D expression begins in early  $G<sub>1</sub>$ , when quiescent cells are stimulated to re-enter the cell cycle, and cyclin D expression remains at high levels as long as mitogens and proliferative signals are present. Cyclin D1 is the product of the bcl-1 gene, located near the translocation breakpoint (11;14) in B cell lymphomas. Cyclin D1 together with its cdk partner is responsible for transition to the S (DNA synthesis) phase by phosphorylating the product of the retinoblastoma susceptibility gene (Rb), which then releases transcription factors important in the initiation of DNA replication [124]. Inhibition of cyclin D1 blocks the cell cycle in  $G<sub>1</sub>$ , demonstrating the necessity of cyclin D for cell cycle control [125, 126]. Furthermore, reinforced expression of cyclin D1 can mimic the effects of growth factors in activating the cyclin E-Cdk2 complexes and promoting S phase entry [127]. These results suggest that cyclin D1 might be important for tumorigenesis. Further evidence for cyclin D1 as an oncogene was provided by experiments in which cyclin D1 over-expression transformed BRK cells with a defective adenovirus E1A protein [128] or rat fibroblasts with an activated Ha-ras gene [129]. Cyclin D1 over-expression may, however, be an early and purely proliferative event that eventually becomes oncogenic as further genetic aberrations emerge. Over-expression of cyclin D1 has also been identified as a frequent and early event during the development of NSCLC, where it is associated with a poorly differentiated histology and, surprisingly, a reduction in local relapse rate [130, 131]. Although overexpression of cyclin D1 is a key abnormality in lung carcinogenesis, it seems to be aless frequent event in SCLC, where its prognostic and therapeutic significance have not yet been addressed. Based on its frequent alteration and role in cell cycle control, cyclin D1 may serve as a potential target for gene inhibition approaches in early lung cancer. As shown by Schrump et al. [132], downregulation of cyclin D1 expression using an antisense approach could reduce the proliferation and tumorigenicity of murine lung cancer cells.

## **Prostate tumor-inducing gene 1 (PTI-1)**

Expression cloning and differential RNA display experiments revealed a novel potential oncogene of 2123 bp, PTI-1, which is expressed in prostate, breast, colon, and SCLC cells [133, 134]. In a recent study, Su et al. [135] demonstrated that stable expression of the nearly fulllength 1.9-kb PTI-1 gene in fibroblasts induced an aggressive phenotype with increased tumorigenicity in athymic mice. Blocking of PTI-1 expression with an antisense gene construct could revert the transformation of the cells and reduced their tumorigenic potential. Further studies are necessary to reveal whether PTI-1 indeed represents a potential target for antisense therapeutics in SCLC and other cancer diseases.

# **Gene therapy**

#### **Replacement of tumor suppressor genes**

Especially in tumors with a high propensity to metastatic spread such as SCLC, there are severe limitations in the efficiency of gene transfer by currently available vectors and gene transfer systems. Preclinical and clinical studies in other tumor types including NSCLC, however, have suggested that current delivery strategies may have therapeutic potential in defined clinical settings.

In SCLC cells, a number of genetic lesions have been reported, including the well-characterized tumor suppressor gene p53 and Rb, which are mutated or lost in 90% and 70% of cases, respectively. Wild-type p53 is a sequence-specific transcription factor whose expression



Figure 2. Electron mircographs of SW2 SCLC cells 96 h after treatment with bcl-2 or bcl-xL antisense oligodeoxynucleotides (ODNs): untreated cells (*A*), cells treated with a bcl-2 antisense ODN (*B*), cells treated with a mismatch control ODN (*C*), cells treated with a bcl-xL antisense ODN (*D*). As shown by the extensive membrane blebbing, cell death after treatment with the antisense ODNs was apoptotic by nature. (Prof. P. Grosscurth, Institute of Anatomy, University of Zürich, with permission).

can suppress the progression of cells from  $G_1$  into S phase [95, 136]. It is also required for the  $G_1$  arrest of cells in response to DNA damage, presumably to allow cells to repair damaged templates prior to replication [137, 138]. Substantial evidence suggests that a major part of p53 mediated growth arrest evolves through induction of the cdk inhibitor p21WAF1 (p21) [139]. In contrast, the mechanism by which p53 promotes apoptosis is less clear. Examples of p53-regulated targets are proteins involved in the apoptotic response, such as the Bcl-2 antagonist Bax [140, 141] and the IGF-I receptor [79], and proteins regulating angiogenesis [142, 143].

In its hypophosphorylated state, the Rb protein causes cell cycle arrest in the  $G_0/G_1$  phase, in part by inhibiting the function of transcription factors such as E2F [144, 145]. Inappropriate passage of cells through the  $G_1$ checkpoint may cause them to replicate damaged DNA in the S phase before DNA repair can be completed in  $G<sub>1</sub>$ , and to ignore signals inducing differentiation or cell death [146].

In view of these findings, p53 and Rb gene replacement might be of therapeutic benefit for many tumor types. Gene replacement with retroviral or adenoviral vectors harboring wt-p53 was shown to restore the apoptotic response of NSCLC cells in preclinical studies. In a recent phase I study with 21 patients presenting with NSCLC, local injection of the p53 adenoviral vector alone or in combination with cisplatin lengthened the time to disease progression and produced a bystander effect, possibly due to the anti-angiogenic effect of p53 [147]. The p53 tumor suppressor gene transactivates the cdk inhibitor gene p21, which is an important mediator of cell proliferation and differentiation [148]. In cell cycle regulation, p21 is a dual inhibitor of cdks and proliferating cell nuclear antigen, both of which are required for entering and progressing through the S phase of the cell cycle [149, 150]. Cells over-expressing p21 accumulate in the  $G_1$ phase and p21 null cells are impaired in their ability to undergo  $G_1$  arrest following DNA damage [149, 151]. In a recent preclinical study, p21 gene replacement using an adenoviral vector inhibited the growth of NSCLC cells in vitro and in a xenograft model by inducing  $G_0/G_1$ arrest [152]. Since the p53 and p21 tumor suppressor genes are frequently mutated or lost in SCLC cells, gene replacement approaches might be interesting treatment options for patients with limited-stage SCLC. Replacement of the Rb gene using adenoviral vectors inhibits the proliferation of and induces  $G_1$  arrest in Rb-altered human tumor cell lines including SCLC in vitro and in vivo [153, 154].

## **Vaccination gene therapy and immune activation**

Engineering of tumor cells with cytokine genes to enhance their immunogenicity has been the subject of extensive investigation in recent years [155, 156]. IL-2 has been the most commonly used cytokine in these studies, which mediates its immune stimulatory effect by binding to aspecific receptor constitutively expressed on T and NK cells [157, 158]. A clinical study suggested a possible effect of exogenous IL-2 in patients with SCLC [159], and an approach to genetically engineer autologous SCLC cells from patients with limited-stage SCLC to produce IL-2 has been proposed [160]. As shown by Meazza et al. [161], transfection of the IL-2 gene into SCLC cells reduced their tumorigenicity in nude mice, due to the recruitment and activation of macrophages, granulocytes, and NK cells at the tumor site. The idea underlying this approach is to prevent anergy of immune competent cells and to activate a systemic immune response by creating an environment of high doses of interleukins at the tumor site. A variety of epithelial tumor cells, including SCLC and NSCLC cells, however, up-regulate expression of the Fas-ligand (CD95L) on their surface, thereby creating an immuneprivileged environment almost inaccessible for cytotoxic T cells [162]. Further studies are needed to assess whether the rationale of the cytokine gene therapy approach will translate into clinical efficacy.

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- 1 Boring C. C., Squires T. S., Tong T. and Montgomery S. (1994) Cancer statistics, 1994. CA Cancer J. Clin. **44:** 7 – 26
- 2 Zelen M. (1973) Keynote address on biostatistics and data retrieval. Cancer Chemother. Rep. **4:** 31 – 42
- 3 Souhami R. L. and Law K. (1990) Longevity in small cell lung cancer: a report to the Lung Cancer Subcommittee of the United Kingdom Coordinating Committee for Cancer Research. Br. J. Cancer **61:** 584 – 589
- 4 Miller A. B., Fox W. and Tall R. (1969) Five-year follow-up of the Medical Research Council comparative trial of surgery and radiotherapy for the primary treatment of smallcelled or oat-celled carcinoma of the bronchus. Lancet **ii:** 501–505
- 5 Fox W. and Scadding J. G. (1973) Medical Research Council comparative trial of surgery and radiotherapy for primary treatment of small-celled or oat-celled carcinoma of bronchus: ten-year follow-up. Lancet **ii:** 63 – 65
- 6 Ihde D. C. (1992) Chemotherapy of lung cancer. N. Engl. J. Med. **327:** 1434–1441
- 7 Souhami R. L., Rudd R., Ruiz-de E. M., James L., Gower N., Harper P. G. et al. (1994) Randomized trial comparing weekly versus 3-week chemotherapy in small-cell lung cancer: a Cancer Research Campaign trial. J. Clin. Oncol. **12:** 1806–1813
- 8 Ihde D. C., Mulshine J. L., Kramer B. S., Steinberg S. M., Linnoila R. I., Gazdar A. F. et al. (1994) Prospective randomized comparison of high-dose and standard-dose etoposide and cisplatin chemotherapy in patients with extensive-stage small-cell lung cancer. J. Clin. Oncol. **12:** 2022 – 2034
- 9 Pujol J. L., Douillard J. Y., Riviere A., Quoix E., Lagrange J. L., Berthaud P. et al. (1997) Dose-intensity of a four-drug chemotherapy regimen with or without recombinant human

granulocyte-macrophage colony-stimulating factor in extensive-stage small-cell lung cancer: a multicenter randomized phase III study. J. Clin. Oncol. **15:** 2082 – 2089

- 10 Maurer L. H., Herndon J. E. II, Hollis D. R., Aisner J., Carey R. W., Skarin A. T. et al. (1997) Randomized trial of chemotherapy and radiation therapy with or without warfarin for limited-stage small-cell lung cancer: a Cancer and LeukemiaGroup B study. J. Clin. Oncol. **15:** 3378– 3387
- 11 Kelly K., Crowley J. J., Bunn-PA J., Hazuka M. B., Beasley K., Upchurch C. et al. (1995) Role of recombinant interferon alfa-2a maintenance in patients with limited-stage small-cell lung cancer responding to concurrent chemoradiation: a Southwest Oncology Group study. J. Clin. Oncol. 13: 2924 – 2930
- 12 Rowland K. M. J., Loprinzi C. L., Shaw E. G., Maksymiuk A. W., Kuross S. A., Jung S. H. et al. (1996) Randomized double-blind placebo-controlled trial of cisplatin and etoposide plus megestrol acetate/placebo in extensive-stage small-cell lung cancer: a North Central Cancer Treatment Group study. J. Clin. Oncol. **14:** 135–141
- 13 Ettinger D. S., Finkelstein D. M., Sarma R. P. and Johnson D. H. (1995) Phase II study of paclitaxel in patients with extensive-disease small-cell lung cancer: an Eastern Cooperative Oncology Group study. J. Clin. Oncol. **13:** 1430–1435
- 14 Cormier Y., Eisenhauer E., Muldal A., Gregg R., Ayoub J., Goss G. et al. (1994) Gemcitabine is an active new agent in previously untreated extensive small cell lung cancer (SCLC): a study of the National Cancer Institute of Canada Clinical Trials Group. Ann. Oncol. **5:** 283 – 285
- 15 Furuse K., Kubota K., Kawahara M., Takada M., Kimura I., Fujii M. et al. (1996) Phase II study of vinorelbine in heavily previously treated small cell lung cancer: Japan Lung Cancer Vinorelbine Study Group. Oncology **53:** 169–172
- 16 Schiller J. H., Kim K., Hutson P., DeVore R., Glick J., Stewart J. et al. (1996) Phase II study of topotecan in patients with extensive-stage small-cell carcinoma of the lung: an Eastern Cooperative Oncology Group trial. J. Clin. Oncol. **14:** 2345 – 2352
- 17 Pignon J. P., Arriagada R., Ihde D. C., Johnson D. H., Perry M. C., Souhami R. L. et al. (1992) A meta-analysis of thoracic radiotherapy for small-cell lung cancer. N. Engl. J. Med. **327:** 1618 – 1624
- 18 Turrisi A., Kyungmann K., Blum R., Sause W. T., Livingston R. B., Komaki R. et al. (1999) Twice-daily compared with once-daily thoracic radiotherapy in limited small-cell lung cancer treated concurrently with cisplatin and etoposide. N. Engl. J. Med. **340:** 265 – 271
- 19 Arriagada R., Le C. T., Borie F., Riviere A., Chomy P., Monnet I. et al. (1995) Prophylactic cranial irradiation for patients with small-cell lung cancer in complete remission. J. Natl. Cancer Inst. **87:** 183–190
- 20 Arriagada R., Auperin A., Pignon J. P., Gregor A., Stephens R., Kristjansen P. E. G. et al. (1998) Prophylactic cranial irradiation overview (PCIO) in patients with small cell lung cancer (SCLC) in complete remission (CR) (Abstract no. 1758) Proc. Am. Soc. Clin. Oncol. **17**.
- 21 Gregor A., Cull A., Stephens R. J., Kirkpatrick J. A., Yarnold J. R., Girling D. J. et al. (1997) Prophylactic cranial irradiation is indicated following complete response to induction therapy in small cell lung cancer: results of a multicentre randomised trial. Eur. J. Cancer **33:** 1752– 1758
- 22 Pegram M. D., Lipton A., Hayes D. F., Weber B. L., Baselga J. M., Tripathy D. et al. (1998) Phase II study of receptor-enhanced chemosensitivity using recombinant humanized anti-p185HER2/neu monoclonal antibody plus cisplatin in patients with HER2/neu-overexpressing metastatic breast cancer refractory to chemotherapy treatment. J. Clin. Oncol. **16:** 2659 – 2671
- 23 Yang D. J., Kuan C. T., Payne J., Kihara A., Murray A., Wang L. M. et al. (1998) Recombinant heregulin *Pseudomonas* exotoxin fusion proteins: interactions with the heregulin receptors and antitumor activity in vivo. Clin. Cancer Res. **4:** 993–1004
- 24 Colcher D., Bird R., Roselli M., Hardman K. D., Johnson S., Pope S. et al. (1990) In vivo tumor targeting of a recombinant single-chain antigen-binding protein. J. Natl. Cancer Inst. **82:** 1191– 1197
- 25 Begent R. H., Verhaar M. J., Chester K. A., Casey J. L., Green A. J., Napier M. P. et al. (1996) Clinical evidence of efficient tumor targeting based on single-chain Fv antibody selected from a combinatorial library. Nat. Med. **2:** 979 – 984
- 26 Dolman C. S., Mueller B. M., Lode H. N., Xiang R., Gillies S. D. and Reisfeld R. A. (1998) Suppression of human prostate carcinoma metastases in severe combined immunodeficient mice by interleukin 2 immunocytokine therapy. Clin. Cancer Res. **4:** 2551–2557
- Souhami R. L., Beverley P. C., Bobrow L. G. and Ledermann J. A. (1991) Antigens of lung cancer: results of the second international workshop on lung cancer antigens. J. Natl. Cancer Inst. **83:** 609 –612
- Stahel R. A., Gilks W. R. and Schenker T. (1994) Antigens of lung cancer: results of the Third International Workshop on Lung Tumor and Differentiation Antigens. J. Natl. Cancer Inst. **86:** 669– 672
- Zangemeister W. U., Lehmann H. P., Waibel R., Wawrzynczak E. J. and Stahel R. A. (1993) Action of a CD24-specific deglycosylated ricin-A-chain immunotoxin in conventional and novel models of small-cell-lung-cancer xenograft. Int. J. Cancer **53:** 521 – 528
- 30 Waibel R., Mannhart M., O'Hara C. J., Brocklehurst C., Zangemeister W. U., Schenker T. et al. (1993) Monoclonal antibody SEN7 recognizes a new epitope on the neural cell adhesion molecule present on small cell lung cancer but not on lymphocytes. Cancer Res. **53:** 2840–2845
- 31 Ledermann J. A., Marston N. J., Stahel R. A., Waibel R., Buscombe J. R. and Ell P. J. (1993) Biodistribution and tumour localisation of 131I SWA11 recognising the cluster w4 antigen in patients with small cell lung cancer. Br. J. Cancer **68:** 119-121
- 32 Ornadel D., Ledermann J. A., Eagle K., Pedley R. B., Boxer G., Ward S. E. et al. (1998) Biodistribution of a radiolabelled monoclonal antibody NY3D11 recognizing the neural cell adhesion molecule in tumour xenografts and patients with small cell lung cancer. Br. J. Cancer **77:** 103–109
- 33 Doria M. I. J., Montag A. G. and Franklin W. A. (1988) Immunophenotype of small cell lung carcinoma: expression of NKH-1 and transferrin receptor and absence of most myeloid antigens. Cancer **62:** 1939 –1945
- 34 Skarin A. T. (1993) Analysis of long-term survivors with small-cell lung cancer. Chest **103:** 440S–444S
- 35 Mechtersheimer G., Staudter M. and Moller P. (1991) Expression of the natural killer cell-associated antigens CD56 and CD57 in human neural and striated muscle cells and in their tumors. Cancer Res. **51:** 1300–1307
- 36 Hosono M. N., Hosono M., Endo K., Ueda R. and OnoyamaY. (1994) Effect of hyperthermiaon tumor uptake of radiolabeled anti-neural cell adhesion molecule antibody in small-cell lung cancer xenografts. J. Nuclear Med. **35:** 504 –509
- 37 Lynch T. J. L., Lambert J. M., Coral F., Shefner J., Wen P., Blattler W. A. et al. (1997) Immunotoxin therapy of smallcell lung cancer: a phase I study of N901-blocked ricin. J. Clin. Oncol. **15:** 723 –734
- Strand J., Hamilton A. E., Beavers L. S., Gamboa G. C., Apelgren L. D., Taber L. D. et al. (1989) Molecular cloning and characterization of a human adenocarcinoma/epithelial cell surface antigen complementary DNA. Cancer Res. **49:** 314– 317
- 39 Zimmermann S., Wels W., Froesch B. A., Gerstmayer B., Stahel R. A. and Zangemeister Wittke U. (1997) A novel immunotoxin recognising the epithelial glycoprotein-2 has potent antitumoural activity on chemotherapy-resistant lung cancer. Cancer Immunol. Immunother. **44:** 1–9

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- 40 Basak S., Speicher D., Eck S., Wunner W., Maul G., Simmons M. S. et al. (1998) Colorectal carcinoma invasion inhibition by CO17-1A/GA733 antigen and its murine homologue. J. Natl. Cancer Inst. **90:** 691 – 697
- 41 Cirulli V., Crisa L., Beattie G. M., Mally M. I., Lopez A. D., Fannon A. et al. (1998) KSA antigen Ep-CAM mediates cell-cell adhesion of pancreatic epithelial cells: morphoregulatory roles in pancreatic islet development. J. Cell Biol. **140:** 1519 – 1534
- 42 Riethmuller G., Holz E., Schlimok G., Schmiegel W., Raab R., Hoffken K. et al. (1998) Monoclonal antibody therapy for resected Dukes' C colorectal cancer: seven-year outcome of amulticenter randomized trial. J. Clin. Oncol. **16:** 1788– 1794
- 43 Kosterink J. G., Jonge M. W. de, Smit E. F., Piers D. A., Kengen R. A., Postmus P. E. et al. (1995) Pharmacokinetics and scintigraphy of indium-111-DTPA-MOC-31 in small-cell lung carcinoma. J. Nuclear Med. **36:** 2356–2362
- 44 Willuda J., Honegger A., Waibel R., Stahel R., Zangemeister-Wittke U. and Plückthun A. (1999) Rational engineering for high stability is required for tumor targeting of a highaffinity scFv fragment specific for the panepithelial glycoprotein EGP-2 (EP-CAM). Proc. Am. Assoc. Cancer Res. **40:** 354
- 45 UraY., Dion A. S., Williams C. J., Olsen B. D., Redfield E. S., Ishida M. et al. (1992) Quantitative dot blot analyses of blood-group-related antigens in paired normal and malignant human breast tissues. Int. J. Cancer **50:** 57 – 63
- 46 Cooper H. S., Malecha M. J., Bass C., Fagel P. L. and Steplewski Z. (1991) Expression of blood group antigens H-2, Le(y), and sialylated-Le(a) in human colorectal carcinoma: an immunohistochemical study using double-labeling techniques. Am. J. Pathol. **138:** 103 – 110
- 47 Murata K., Egami H., Shibata Y., Sakamoto K., Misumi A. and Ogawa M. (1992) Expression of blood group-related antigens, ABH, Lewis(a), Lewis(b), Lewis(x), Lewis(y), CA19-9, and CSLEX1 in early cancer, intestinal metaplasia, and uninvolved mucosa of the stomach. Am. J. Clin. Pathol. **98:** 67 – 75
- 48 Leoni F., Colnaghi M. I., Canevari S., Menard S., Colzani E., Facheris P. et al. (1992) Glycolipids carrying Le(y) are preferentially expressed on small-cell lung cancer cells as detected by the monoclonal antibody MLuC1. Int. J. Cancer **51:** 225 – 231
- 49 Miyake M., Taki T., Hitomi S. and Hakomori S. (1992) Correlation of expression of H/Le(y)/Le(b) antigens with survival in patients with carcinoma of the lung. N. Engl. J. Med. **327:** 14 – 18
- 50 Miyake M., Zenita K., Tanaka O., Okada Y. and Kannagi R. (1988) Stage-specific expression of SSEA-1-related antigens in the developing lung of human embryos and its relation to the distribution of these antigens in lung cancers. Cancer Res. **48:** 7150 – 7158
- 51 Stahel R. A., Lacroix H., Sculier J. P., Morant R., Richner J., Janzek E. et al. (1992) Phase I/II study of monoclonal antibody against Lewis Y hapten in relapsed small-cell lung cancer. Ann. Oncol. **3:** 319 – 320
- 52 Siegall C. B. (1995) Targeted therapy of carcinomas using BR96 sFv-PE40, a single-chain immunotoxin that binds to the Le(y) antigen. Semin. Cancer Biol. **6:** 289 –295
- 53 Pai L. H., Wittes R., Setser A., Willingham M. C. and Pastan I. (1996) Treatment of advanced solid tumors with immunotoxin LMB-1: an antibody linked to *Pseudomonas* exotoxin. Nat. Med. **2:** 350 – 353
- 54 Pastan I. H., Pai L. H., Brinkmann U. and Fitzgerald D. J. (1995) Recombinant toxins: new therapeutic agents for cancer. Ann. NY Acad. Sci. **758:** 345 – 354
- 55 Cheresh D. A., Pierschbacher M. D., Herzig M. A. and Mujoo K. (1986) Disialogangliosides GD2 and GD3 are involved in the attachment of human melanoma and neuroblastoma cells to extracellular matrix proteins. J. Cell Biol. **102:** 688 – 696
- 56 Cheresh D. A., Honsik C. J., Staffileno L. K., Jung G. and Reisfeld R. A. (1985) Disialoganglioside GD3 on human melanoma serves as a relevant target antigen for monoclonal antibody-mediated tumor cytolysis. Proc. Natl. Acad. Sci. USA **82:** 5155 –5159
- 57 Fuentes R., Allman R. and Mason M. D. (1997) Ganglioside expression in lung cancer cell lines. Lung Cancer **18:** 21 – 33
- 58 Hanibuchi M., Yano S., Nishioka Y., Yanagawa H. and Sone S. (1996) Anti-ganglioside GM2 monoclonal antibodydependent killing of human lung cancer cells by lymphocytes and monocytes. Jpn. J. Cancer Res. **87:** 497–504
- 59 Houghton A. N., Mintzer D., Cordon C. C., Welt S., Fliegel B., Vadhan S. et al. (1985) Mouse monoclonal IgG3 antibody detecting GD3 ganglioside: a phase I trial in patients with malignant melanoma. Proc. Natl. Acad. Sci. USA **82:** 1242–1246
- 60 Cheung N. K. and Miraldi F. D. (1988) Iodine 131 labeled GD2 monoclonal antibody in the diagnosis and therapy of human neuroblastoma. Prog. Clin. Biol. Res. **271:** 595 – 604
- 61 Grant S. C., Kostakoglu L., Kris M. G., Yeh S. D., Larson S. M., Finn R. D. et al. (1996) Targeting of small-cell lung cancer using the anti-GD2 ganglioside monoclonal antibody 3F8: a pilot trial. Eur. J. Nuclear Med. **23:** 145– 149
- 62 Kris M. G. and Miller V. (1997) Long-term survival in 15 patients with small cell lung cancer (SCLC) immunized with BEC2 plus BCG after initial therapy: an update. Proc. Am. Soc. Clin. Oncol. **16:** 1630
- 63 Sporn M. B. and Todaro G. J. (1980) Autocrine secretion and malignant transformation of cells. N. Engl. J. Med. **303:** 878–880
- 64 Carney D. N., Gazdar A. F., Bepler G., Guccion J. G., Marangos P. J., Moody T. W. et al. (1985) Establishment and identification of small cell lung cancer cell lines having classic and variant features. Cancer Res. **45:** 2913 – 2923
- 65 Sausville E. A., Lebacq V. A., Spindel E. R., Cuttitta F., Gazdar A. F. and Battey J. F. (1986) Expression of the gastrin-releasing peptide gene in human small cell lung cancer: evidence for alternative processing resulting in three distinct mRNAs. J. Biol. Chem. **261:** 2451–2457
- Spindel E. R., Zilberberg M. D. and Chin W. W. (1987) Analysis of the gene and multiple messenger ribonucleic acids (mRNAs) encoding human gastrin-releasing peptide: alternate RNA splicing occurs in neural and endocrine tissue. Mol. Endocrinol. **1:** 224– 232
- Heikkila R., Trepel J. B., Cuttitta F., Neckers L. M. and Sausville E. A. (1987) Bombesin-related peptides induce calcium mobilization in a subset of human small cell lung cancer cell lines. J. Biol. Chem. **262:** 16456–16460
- 68 Moody T. W., Murphy A., Mahmoud S. and Fiskum G. (1987) Bombesin-like peptides elevate cytosolic calcium in small cell lung cancer cells. Biochem. Biophys. Res. Commun. **147:** 189 –195
- 69 Jones C. L., Beck L. K., BroznaJ. P., Holley M., Dempsey E. J. and Kane M. A. (1995) Properties of classic protein kinase C in human small cell lung carcinoma NCI-H345 cells. Cell Growth Differ. **6:** 1627 –1634
- 70 Bepler G., Zeymer U., Mahmoud S., Fiskum G., Palaszynski E., Rotsch M. et al. (1988) Substance P analogues function as bombesin receptor antagonists and inhibit small cell lung cancer clonal growth. Peptides **9:** 1367– 1372
- 71 Kelley M. J., LinnoilaR. I., Avis I. L., Georgiadis M. S., CuttittaF., Mulshine J. L. et al. (1997) Antitumor activity of a monoclonal antibody directed against gastrin-releasing peptide in patients with small cell lung cancer. Chest **112:**  $256 - 261$
- 72 Nakanishi Y., Mulshine J. L., Kasprzyk P. G., Natale R. B., Maneckjee R., Avis I. et al. (1988) Insulin-like growth factor-I can mediate autocrine proliferation of human small cell lung cancer cell lines in vitro. J. Clin. Invest. **82:** 354 – 359
- 73 Reeve J. G., Xiong J., Morgan J. and Bleehen N. M. (1996) Expression of apoptosis-regulatory genes in lung tumour cell lines: relationship to p53 expression and relevance to acquired drug resistance. Br. J. Cancer **73:** 1193 –1200
- 74 Minuto F., Del M. P., Barreca A., Alama A., Cariola G. and Giordano G. (1988) Evidence for autocrine mitogenic stimulation by somatomedin-C/insulin-like growth factor I on an established human lung cancer cell line. Cancer Res. **48:** 3716 – 3719
- 75 Rechler M. M. and Nissley S. P. (1985) The nature and regulation of the receptors for insulin-like growth factors. Annu. Rev. Physiol. **47:** 425–442
- 76 Kauffmann Z. A., Rodriguez V. P., Ulrich E., Gilbert C., Coffer P., Downward J. et al. (1997) Suppression of c-Mycinduced apoptosis by Ras signalling through PI(3)K and PKB. Nature **385:** 544–548
- 77 Dudek H., Datta S. R., Franke T. F., Birnbaum M. J., Yao R., Cooper G. M. et al. (1997) Regulation of neuronal survival by the serine-threonine protein kinase Akt. Science **275:** 661 – 665
- 78 Minshall C., Arkins S., Straza J., Conners J., Dantzer R., Freund G. G. et al. (1997) IL-4 and insulin-like growth factor-I inhibit the decline in Bcl-2 and promote the survival of IL-3-deprived myeloid progenitors. J. Immunol. **159:** 1225 – 1232
- 79 Baserga R., Hongo A., Rubini M., Prisco M. and Valentinis B. (1997) The IGF-I receptor in cell growth, transformation and apoptosis. Biochim. Biophys. Acta **1332:** F105–F126
- 80 Zia F., Jacobs S., Kull F., Cuttitta F., Mulshine J. L. and Moody T. W. (1996) Monoclonal antibody alpha IR-3 inhibits non-small cell lung cancer growth in vitro and in vivo. J. Cell Biochem. Suppl. **24:** 269 – 275
- 81 Resnicoff M., Coppola D., Sell C., Rubin R., Ferrone S. and Baserga R. (1994) Growth inhibition of human melanoma cells in nude mice by antisense strategies to the type 1 insulin-like growth factor receptor. Cancer Res. **54:** 4848 – 4850
- 82 Crooke S. T. (1998) An overview of progress in antisense therapeutics. Antisense Nucleic Acid Drug Dev. **8:** 115 – 122
- 83 Crooke S. T. (1998) Basic principles of antisense therapeutics. Antisense Res. Appl. **131:** 1 – 50
- 84 Bennett C. F. (1998) Antisense oligonucleotides: is the glass half full or half empty? Biochem. Pharmacol. **55:** 9-19
- 85 Ziegler A., Luedke G. H., Fabbro D., Altmann K. H. and Zangemeister-Wittke U. (1997) Induction of apoptosis in small-cell lung cancer cells by an antisense oligodeoxynucleotide targeting the Bcl-2 coding sequence. J. Natl. Cancer Inst. **89:** 1027 – 1036
- 86 MoniaB. P., Sasmor H., Johnston J. F., Freier S. M., Lesnik E. A., Muller M. et al. (1996) Sequence-specific antitumor activity of a phosphorothioate oligodeoxyribonucleotide targeted to human C-raf kinase supports an antisense mechanism of action in vivo. Proc. Natl. Acad. Sci. USA **93:** 15481 – 15484
- Skorski T., Nieborowska Skorska M., Wlodarski P., Perrotti D., Hoser G., Kawiak J. et al. (1997) Treatment of Philadelphia leukemia in severe combined immunodeficient mice by combination of cyclophosphamide and bcr/abl antisense oligodeoxynucleotides. J. Natl. Cancer Inst. **89:** 124 –133
- 88 Zaman G. J., Versantvoort C. H., Smit J. J., Eijdems E. W., Haas M. de, Smith A. J. et al. (1993) Analysis of the expression of MRP, the gene for a new putative transmembrane drug transporter, in human multidrug resistant lung cancer cell lines. Cancer Res. **53:** 1747 –1750
- 89 Narasaki F., Matsuo I., Ikuno N., Fukuda M., Soda H. and Oka M. (1996) Multidrug resistance-associated protein (MRP) gene expression in human lung cancer. Anticancer Res. **16:** 2079 – 2082
- 90 Poupon M. F., Arvelo F., Goguel A. F., Bourgeois Y., Jacrot M., Hanania N. et al. (1993) Response of small-cell lung cancer xenografts to chemotherapy: multidrug resistance and direct clinical correlates. J. Natl. Cancer Inst. **85:** 2023 – 2029
- 91 Lai S. L., Goldstein L. J., Gottesman M. M., Pastan I., Tsai C. M., Johnson B. E. et al. (1989) MDR1 gene expression in lung cancer. J. Natl. Cancer Inst. **81:** 1144–1150
- 92 Alahari S. K., DeLong R., Fisher M. H., Dean N. M., Viliet P. and Juliano R. L. (1998) Novel chemically modified oligonucleotides provide potent inhibition of P-glycoprotein expression. J. Pharmacol. Exp. Ther. **286:** 419–428
- Bishop J. M. (1991) Molecular themes in oncogenesis. Cell **64:** 235 –248
- 94 Weinberg R. A. (1991) Tumor suppressor genes. Science **254:** 1138–1146
- 95 Vogelstein B. and Kinzler K. W. (1992) p53 function and dysfunction. Cell **70:** 523–526
- 96 Fisher D. E. (1994) Apoptosis in cancer therapy: crossing the threshold. Cell **78:** 539– 542
- Smets L. A. (1994) Programmed cell death (apoptosis) and response to anti-cancer drugs. Anticancer Drugs **5:** 3–9
- 98 Reed J. C. (1997) Double identity for proteins of the Bcl-2 family. Nature **387:** 773 – 776
- 99 Adams J. M. and Cory S. (1998) The Bcl-2 protein family: arbiters of cell survival. Science **281:** 1322–1326
- 100 Simonian P. L., Grillot D. A. and Nunez G. (1997) Bcl-2 and Bcl-XL can differentially block chemotherapy-induced cell death. Blood **90:** 1208–1216
- 101 Ikegaki N., Katsumata M., Minna J. D. and Tsujimoto Y. (1994) Expression of *bcl*-<sup>2</sup> in small cell lung carcinoma cells. Cancer Res. **54:** 6–8
- 102 Jiang S. X., Sato Y., Kuwao S. and Kameya T. (1995) Expression of *bcl*-<sup>2</sup> oncogene protein is prevalent in small cell lung carcinomas. J. Pathol. **177:** 135–138
- 103 Ben Ezra J. M., Kornstein M. J., Grimes M. M. and Krystal G. (1994) Small cell carcinomas of the lung express the Bcl-2 protein. Am. J. Pathol. **145:** 1036 –1040
- 104 Zangemeister-Wittke U., Schenker T., Luedke G. H. and Stahel R. A. (1998) Synergistic cytotoxicity of bcl-2 antisense oligodeoxynucleotides and etoposide, doxorubicin and cisplatin on small cell lung cancer cell lines. Br. J. Cancer **78:**  $1035 - 1042$
- 105 Webb A., Cunningham D., Cotter F., Clarke P. A., Distefano F., Ross P. et al. (1997) Bcl-2 antisense therapy in patients with non-Hodgkin-lymphoma. Lancet **49:** 1137 – 1141
- 106 Guvakova M. A., Yakubov L. A., Vlodavsky I., Tonkinson J. L. and Stein C. A. (1995) Phosphorothioate oligodeoxynucleotides bind to basic fibroblast growth factor, inhibit its binding to cell surface receptors, and remove it from low affinity binding sites on extracellular matrix. J. Biol. Chem. **270:** 2620– 2627
- 107 Tuscano J. M., Druey K. M., RivaA., PenaJ., Thompson C. B. and Kehrl J. H. (1996) Bcl-x rather than Bcl-2 mediates CD40-dependent centrocyte survival in the germinal center. Blood **88:** 1359 –1364
- 108 Dole M. G., Jasty R., Cooper M. J., Thompson C. B., Nunez G. and Castle V. P. (1995) Bcl-xL is expressed in neuroblastoma cells and modulates chemotherapy-induced apoptosis. Cancer Res. **55:** 2576–2582
- 109 Wong A. J., Ruppert J. M., Eggleston J., Hamilton S. R., Baylin S. B. and Vogelstein B. (1986) Gene amplification of c-myc and N-myc in small cell carcinoma of the lung. Science **233:** 461 –464
- 110 Prins J., De V. E. and Mulder N. H. (1993) The myc family of oncogenes and their presence and importance in small-cell lung carcinoma and other tumour types. Anticancer Res. **13:** 1373– 1385
- 111 Little C. D., Nau M. M., Carney D. N., Gazdar A. F. and Minna J. D. (1983) Amplification and expression of the c-myc oncogene in human lung cancer cell lines. Nature **306:** 194– 196
- 112 Evan G. I., Wyllie A. H., Gilbert C. S., Littlewood T. D., Land H., Brooks M. et al. (1992) Induction of apoptosis in fibroblasts by c-myc protein. Cell **69:** 119–128

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- 113 Sakamuro D., Eviner V., Elliott K. J., Showe L., White E. and Prendergast G. C. (1995) c-Myc induces apoptosis in epithelial cells by both p53-dependent and p53-independent mechanisms. Oncogene **11:** 2411– 2418
- 114 Thompson E. B. (1998) The many roles of c-Myc in apoptosis. Annu. Rev. Physiol. **60:** 575 – 600
- 115 Alarcon R. M., Rupnow B. A., Graeber T. G., Knox S. J. and Giaccia A. J. (1996) Modulation of c-Myc activity and apoptosis in vivo. Cancer Res. **56:** 4315 –4319
- 116 Askew D. S., Ashmun R. A., Simmons B. C. and Cleveland J. L. (1991) Constitutive c-myc expression in an IL-3-dependent myeloid cell line suppresses cell cycle arrest and accelerates apoptosis. Oncogene **6:** 1915– 1922
- 117 Johnson B. E., Russell E., Simmons A. M., Phelps R., Steinberg S. M., Ihde D. C. et al. (1996) MYC family DNA amplification in 126 tumor cell lines from patients with small cell lung cancer. J. Cell Biochem. Suppl. **24:** 210 – 217
- 118 Bergh J. C. (1990) Gene amplification in human lung cancer: the myc family genes and other proto-oncogenes and growth factor genes. Am. Rev. Respir. Dis. **142:** S20 –S26
- 119 Funa K., Steinholtz L., Nou E. and Bergh J. (1987) Increased expression of N-myc in human small cell lung cancer biopsies predicts lack of response to chemotherapy and poor prognosis. Am. J. Clin. Pathol. **88:** 216 – 220
- 120 Robinson L. A., Smith L. J., Fontaine M. P., Kay H. D., Mountjoy C. P. and Pirruccello S. J. (1995) c-myc antisense oligodeoxyribonucleotides inhibit proliferation of non-small cell lung cancer. Ann. Thorac. Surg. **60:** 1583 – 1591
- 121 Kimura S., Maekawa T., Hirakawa K., Murakami A. and Abe T. (1995) Alterations of c-myc expression by antisense oligodeoxynucleotides enhance the induction of apoptosis in HL-60 cells. Cancer Res. **55:** 1379 – 1384
- 122 Leonetti C., D'Agnano I., Lozupone F., Valentini A., Geiser T., Zon G. et al. (1996) Antitumor effect of c-myc antisense phosphorothioate oligodeoxynucleotides on human melanoma cells in vitro and in mice. J. Natl. Cancer Inst. **88:**  $419 - 429$
- 123 Hunter T. and Pines J. (1994) Cyclins and cancer. II. Cyclin D and CDK inhibitors come of age. Cell **79:** 573 –582
- 124 Donnellan R. and Chetty R. (1998) Cyclin D1 and human neoplasia. Mol. Pathol. **51:** 1–7
- 125 Baldin V., Lukas J., Marcote M. J., Pagano M. and Draetta G. (1993) Cyclin D1 is anuclear protein required for cell cycle progression in G1. Genes Dev. **7:** 812 – 821
- 126 Quelle D. E., Ashmun R. A., Shurtleff S. A., Kato J. Y., Bar S. D., Roussel M. F. et al. (1993) Overexpression of mouse D-type cyclins accelerates G1 phase in rodent fibroblasts. Genes Dev. **7:** 1559 – 1571
- 127 Prall O. W., Rogan E. M. and Sutherland R. L. (1998) Estrogen regulation of cell cycle progression in breast cancer cells. J. Steroid Biochem. Mol. Biol. **65:** 169–174
- 128 Hinds P. W., Dowdy S. F., Eaton E. N., Arnold A. and Weinberg R. A. (1994) Function of a human cyclin gene as an oncogene. Proc. Natl. Acad. Sci. USA **91:** 709–713
- 129 Lovec H., Sewing A., Lucibello F. C., Muller R. and Moroy T. (1994) Oncogenic activity of cyclin D1 revealed through cooperation with Ha-ras: link between cell cycle control and malignant transformation. Oncogene **9:** 323 – 326
- 130 Betticher D. C., Heighway J., Hasleton P. S., Altermatt H. J., Ryder W. D., Cerny T. et al. (1996) Prognostic significance of CCND1 (cyclin D1) overexpression in primary resected non-small-cell lung cancer. Br. J. Cancer **73:** 294 –300
- 131 Betticher D. C., White G. R., Vonlanthen S., Liu X., Kappeler A., Altermatt H. J. et al. (1997) G1 control gene status is frequently altered in resectable non-small cell lung cancer. Int. J. Cancer **74:** 556–562
- 132 Schrump D. S., Chen A. and Consoli U. (1996) Inhibition of lung cancer proliferation by antisense cyclin D. Cancer Gene Ther. **3:** 131 – 135
- 133 Shen R., Su Z. Z., Olsson C. A. and Fisher P. B. (1995) Identification of the human prostatic carcinoma oncogene PTI-1 by rapid expression cloning and differential RNA display. Proc. Natl. Acad. Sci. USA **92:** 6778 – 6782
- 134 Sun Y., Lin J., Katz A. E. and Fisher P. B. (1997) Human prostatic carcinoma oncogene PTI-1 is expressed in human tumor cell lines and prostate carcinoma patient blood samples. Cancer Res. **57:** 18 – 23
- 135 Su Z. Z., Goldstein N. I. and Fisher P. B. (1998) Antisense inhibition of the PTI-1 oncogene reverses cancer phenotypes. Proc. Natl. Acad. Sci. USA **95:** 1764–1769
- 136 Ullrich S. J., Anderson C. W., Mercer W. E. and Appella E. (1992) The p53 tumor suppressor protein, amodulator of cell proliferation. J. Biol. Chem. **267:** 15259 –15262
- 137 Kastan M. B., Onyekwere O., Sidransky D., Vogelstein B. and Craig R. W. (1991) Participation of p53 protein in the cellular response to DNA damage. Cancer Res. 51: 6304-6311
- 138 Kuerbitz S. J., Plunkett B. S., Walsh W. V. and Kastan M. B. (1992) Wild-type p53 is a cell cycle checkpoint determinant following irradiation. Proc. Natl. Acad. Sci. USA **89:** 7491 – 7495
- 139 Hansen R. and Oren M. (1997) p53: from inductive signal to cellular effect. Curr. Opin. Genet. Dev. **7:** 46 – 51
- 140 Yin C., Knudson C. M., Korsmeyer S. J. and Van D. T. (1997) Bax suppresses tumorigenesis and stimulates apoptosis in vivo. Nature **385:** 637 –640
- 141 Miyashita T., Krajewski S., Krajewska M., Wang H. G., Lin H. K., Liebermann D. A. et al. (1994) Tumor suppressor p53 is a regulator of bcl-2 and bax gene expression in vitro and in vivo. Oncogene **9:** 1799–1805
- 142 Dameron K. M., Volpert O. V., Tainsky M. A. and Bouck N. (1994) Control of angiogenesis in fibroblasts by p53 regulation of thrombospondin-1. Science **265:**1582 – 1584
- 143 Bouvet M., Ellis L. M., Nishizaki M., Fujiwara T., Liu W. B., Bucana C. D. et al. (1998) Adenovirus-mediated wild-type p53 gene transfer down-regulates vascular endothelial growth factor expression and inhibits angiogenesis in human colon cancer. Cancer Res. **58:** 2288 –2292
- 144 Goodrich D. W., Wang N. P., Qian Y. W., Lee E. Y. and Lee W. H. (1991) The retinoblastoma gene product regulates progression through the G1 phase of the cell cycle. Cell **67:** 293–302
- 145 Weintraub S. J., Prater C. A. and Dean D. C. (1992) Retinoblastoma protein switches the E2F site from positive to negative element. Nature **358:** 259–261
- 146 Wiman K. G. (1993) The retinoblastoma gene: role in cell cycle control and cell differentiation. FASEB J. **7:** 841 – 845
- 147 Roth J. A. (1998) Gene replacement strategies for lung cancer. Curr. Opin. Oncol. **10:** 127 – 132
- 148 Deiry W. S. el, Tokino T., Velculescu V. E., Levy D. B., Parsons R., Trent J. M. et al. (1993) WAF1, a potential mediator of p53 tumor suppression. Cell **75:** 817–825
- 149 Harper J. W., Adami G. R., Wei N., Keyomarsi K. and Elledge S. J. (1993) The p21 Cdk-interacting protein Cip1 is apotent inhibitor of G1 cyclin-dependent kinases. Cell **75:**  $805 - 816$
- 150 Waga S., Hannon G. J., Beach D. and Stillman B. (1994) The p21 inhibitor of cyclin-dependent kinases controls DNA replication by interaction with PCNA. Nature **369:** 574 –578
- 151 Deng C., Zhang P., Harper J. W., Elledge S. J. and Leder P. (1995) Mice lacking p21CIP1/WAF1 undergo normal development, but are defective in G1 checkpoint control. Cell **82:** 675 –684
- 152 Joshi U. S., Chen Y. Q., Kalemkerian G. P., Adil M. R., Kraut M. and Sarkar F. H. (1998) Inhibition of tumor cell growth by p21(WAF1) adenoviral gene transfer in lung cancer. Cancer Gene Ther. **5:** 183– 191
- 153 Ookawa K., Shiseki M., Takahashi R., Yoshida Y., Terada M. and Yokota J. (1993) Reconstitution of the RB gene suppresses the growth of small-cell lung carcinoma cells carrying multiple genetic alterations. Oncogene **8:**2175 – 2181
- 154 Demers G. W., Harris M. P., Wen S. F., Engler H., Nielsen L. L. and Maneval D. C. (1998) A recombinant adenoviral vector expressing full-length human retinoblastoma susceptibility gene inhibits human tumor cell growth. Cancer Gene Ther. **5:** 207 – 214
- 155 Colombo M. P. and Forni G. (1994) Cytokine gene transfer in tumor inhibition and tumor therapy: where are we now? Immunol. Today **15:** 48 – 51
- 156 Schmidt W. G. and Schmidt W. I. (1995) Cytokines and clinical gene therapy. Eur. J. Immunol. **25:** 1137– 1140
- 157 Taniguchi T. (1992) Structure and function of IL-2 and IL-2 receptors. Behring Inst. Mitt. 87 –95
- 158 Voss S. D., Sondel P. M. and Robb R. J. (1992) Characterization of the interleukin 2 receptors (IL-2R) expressed on human natural killer cells activated in vivo by IL-2: association of the p64 IL-2R gamma chain with the IL-2R beta chain in functional intermediate-affinity IL-2R. J. Exp. Med. **176:** 531–541
- 159 Clamon G., Herndon J., Perry M. C., Ozer H., Kreisman H., Maher T. et al. (1993) Interleukin-2 activity in patients with extensive small-cell lung cancer: a phase II trial of Cancer and Leukemia Group B. J. Natl. Cancer Inst. **85:** 316– 320
- 160 Cassileth P. A., Podack E., Sridhar K., Savaraj N. and Hanlon J. (1995) Phase I study of transfected cancer cells expressing the interleukin-2 gene product in limited stage small cell lung cancer. Hum. Gene Ther. **6:** 369–383
- 161 Meazza R., Marciano S., Sforzini S., Orengo A. M., Coppolecchia M., Musiani P. et al. (1996) Analysis of IL-2 receptor expression and of the biological effects of IL-2 gene transfection in small-cell lung cancer. Br. J. Cancer **74:** 788– 795
- 162 Niehans G. A., Brunner T., Frizelle S. P., Liston J. C., Salerno C. T., Knapp D. J. et al. (1997) Human lung carcinomas express Fas ligand. Cancer Res. **57:** 1007 – 1012