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REVIEW ARTICLE OPEN Pathology of pain and its implications for therapeutic interventions

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Pain is estimated to affect more than 20% of the global population, imposing incalculable health and economic burdens. Effective pain management is crucial for individuals suffering from pain. However, the current methods for pain assessment and treatment fall short of clinical needs. Benefiting from advances in neuroscience and biotechnology, the neuronal circuits and molecular mechanisms critically involved in pain modulation have been elucidated. These research achievements have incited progress in identifying new diagnostic and therapeutic targets. In this review, we first introduce fundamental knowledge about pain, setting gene mutation, epigenetic modification, posttranslational modification, inflammasome, signaling pathways and microbiota. To better present a comprehensive view of pain research, two prominent issues, sexual dimorphism and pain comorbidities, are discussed in detail based on current findings. The status quo of pain evaluation and manipulation is summarized. A series of improved and innovative pain management strategies, such as gene therapy, monoclonal antibody, brain-computer interface and microbial intervention, are making strides towards clinical application. We highlight existing limitations and future directions for enhancing the quality of preclinical and clinical research. Efforts to decipher the complexities of pain pathology will be instrumental in translating scientific discoveries into clinical practice, thereby improving pain management from bench to bedside.

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INTRODUCTION

Pain is defined as an unpleasant sensory and emotional experience associated with, or resembling that associated with, actual or potential tissue damage.¹ It is considered the most primitive and widespread human experience. Owing to its subjective nature, the interplay of nociceptive, cognitive, emotional and social components collectively shapes the pain experience.² Acute pain acts as a defense mechanism against noxious stimuli, infection, homeostasis dysfunction and secondary insults.^{3,4} Patients suffering from congenital insensitivity lack the ability to avoid damage, potentially leading to a predisposition toward self-mutilation.⁵ In contrast, chronic pain is inherently distressing and often the primary reason for patients to seek medical care. It poses a vast socioeconomic burden globally,⁶ with prevalence rates ranging from 10% to 40% and a relatively low recovery rate of only 5%.^{7–9} Pain relief has been a requisite and an important index for clinical treatment.

Pain serves as a crucial nexus between primary diseases and secondary outcomes. It can trigger a dynamic and detrimental interplay among biological, social and psychological factors, leading to disability and poor prognosis for patients. Pain-related psychiatric disorders, such as insomnia, depression, anxiety and impaired social interaction, can exacerbate the progression of primary diseases. These pathological deteriorations also negatively impact social relationships and self-esteem as evidenced by notable increases in divorce, substance abuse, and suicide rates.^{10–12} Chronic pain also undermines the survival benefits of

cancer treatment.¹³ Notably, pain is not equal to suffering. The outcomes of pain are affected by various factors unique to an individual. For instance, massage can elicit pleasant sensations despite transient pain, and an individual in a positive emotional state may exhibit greater pain tolerance. These examples underscore that pain extends beyond a mere biological event and is intricately processed by the brain.

Analgesic drugs are the mainstay of acute and chronic pain management. Despite their short-term effectiveness, significant concerns regarding drug dependence, addiction and other side effects have been raised.^{14,15} The misuse of analgesics has also garnered international attention. New insights into the mechanisms underlying pain sensitivity and recovery are gradually being reported. The development of new therapeutic modalities, drug delivery systems and nonpharmaceutical adjuvant therapies has potential value in pain management. However, these varied interventions still fall short of fully addressing the needs of an individual's quality of life.

This review will introduce the basic knowledge concerning pain research and then discuss current advances in understanding the pathology of pain perception and modulation. Two hot topics, sexual dimorphism and pain comorbidity, will also be discussed. Management approaches for pain will be summarized and remarked for fully displaying the status quo of pain research. Finally, we will discuss the existing limitations and propose future directions for enhancing the research and clinical practice of pain.

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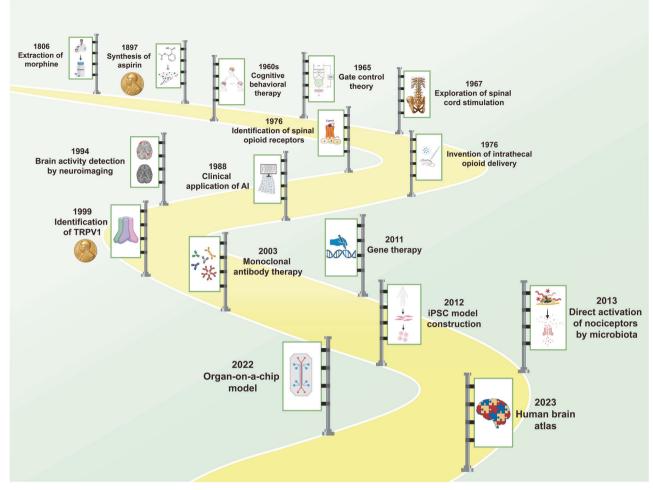


Fig. 1 The brief timeline of historic milestones in the field of pain therapy. Morphine was first extracted in 1806, which opened the chapter in fighting with pain using the fruits of modern medicine. Since then, many intervention methods for pain management were discovered and came into clinical application, such as CBT, spinal cord stimulation, monoclonal antibody therapy and gene therapy. The progress in the research on pain mechanisms and interdisciplinary collaboration boosted advances in pain therapy. In recent years, the wide application of high-throughput biotechnologies has further deepened the understanding in pain pathology and has contributed to the development of individualized pain management. Key milestones of pain therapy are chronologically illustrated in the figure. The achievements awarded by the Nobel prizes are marked with the medals

HISTORICAL MILESTONES OF INVESTIGATIONS INTO PAIN THERAPY

The history of human development is intertwined with the struggles against pain (Fig. 1). Opioid alkaloids, derived from the opium poppy, have been used for analgesia and euphoria for thousands of years. In 1806, Friedrich W. Sertürner pioneered the extraction of pure opioids. This event opened a new chapter in fighting with pain using modern medicine. Another representative drug, acetylsalicylic acid, also called aspirin, was synthesized by Felix Hoffman in 1897. Since then, non-steroid anti-inflammatory drugs (NSAIDs) have gradually become a mainstay in pain management. The discovery of their mechanisms was awarded the Nobel Prize in 1982. With the growing understanding of psychological factors of pain, psychologist Aaron Beck summarized the achievements and proposed cognitive behavioral therapy (CBT) in 1960s. The efficacy of CBT in treating mental disorders, including pain, has been substantiated by numerous cases. This finding underscores the tight link between pain and psychological factors. Advances in computers and algorithms have enabled rapid processing of complex data. In 1965, Melzack and Wall proposed the Gate Control theory. This theory depicted the important functions of spinal dorsal horn in modulating pain signals, offering novel insights into pain pathology and

approaches to clinical pain management.¹⁶ A clinical trial explored the analgesic effects of spinal cord stimulation to treat eight painful patients after 2 years. Half of the patients obtained longstanding pain relief within 2 min, which first proved the superiority of spinal cord stimulation.¹⁷ In 1976, the opioid receptor was identified in the primate spinal cord.¹⁸ In the same year, Yaksh and Rudy conducted intrathecal opioid delivery of narcotics in rats. It effectively exerted potent analgesia only at the spinal level. This exploring experiment laid the foundations for the development of spinal cord stimulation therapy.¹⁹ In 1988, artificial intelligence (AI) was first applied in a clinical trial focusing on the pain diagnosis. The results demonstrated that AI outperformed clinicians in differential diagnosis, highlighting its potency in the pain field.²⁰ Six years later, Edelman et al. utilized magnetic resonance imaging (MRI) to detect brain region activities, laying the groundwork for exploring regions involved in pain perception.²¹ At the end of 20th century, David Julius and colleagues identified the ion channel TRPV1, which is responsive to heat and then produce pain signals. This finding paved the way for discovering other temperature sensors. David Julius was honored with the Nobel Prize in 2021 for this breakthrough.

Entering new century, advanced technologies have been employed in basic research and pain management. The first clinical trial on a monoclonal antibody in neuropathic pain was reported in 2003.²² The effectiveness and safety of gene therapy were proved by a phase I clinical trial in 2011.²³ A year later, the technique for converting pluripotent stem cells into nociceptors was established. This progress has provided a better in-vitro model for pain research.²⁴ The associations between microbiota and pain have been revealed long before. However, it was commonly believed that microbiota activated nociceptors only through inducing inflammatory responses or secreting specific metabolites. A basic study in 2013 showed that gut microbiota could directly stimulate nociceptor neurons and induce pain sensation.²⁵ The revelation shifted previous perceptions in this field and marked a milestone in microbiota and pain research. Over the last decade, research breakthroughs have continued to emerge. The organ-on-a-chip technique was applied to create a spinal microphysiological system for investigating pain and opioid-induced tolerance.²⁶ It represents another significant advancement in experimental pain research tools. The latest milestone is the brain cell atlas, described using multi-omics by the BRAIN Initiative Cell Census Network project, which was reported in the special column of Science journals. This pioneering work parses brain structures at the single-cell level, providing valuable data for elucidating pain mechanisms.

CATEGORIES OF PAIN

Pain can be classified as nociceptive, neuropathic or nociplastic pain according to its etiology. One pain event tends to involve multiple categories. For instance, in a serious car accident, acute pain induced by open wounds can cause nociceptive pain. Spinal cord injury caused by a car crash may bring about perennial neuropathic pain. Posttraumatic stress disorder (PTSD) may also be triggered by this life-challenging event, resulting in somatic nociplastic pain. The etiology of cancer pain is more complicated, involving nerve invasion, organ damage, immune dysregulation and other unknown factors. Therefore, the clarification of pain categories is conducive to the development of pain research.

Nociceptive pain

Nociceptive pain refers to pain induced by a physiological protective system that protects against noxious stimuli,²⁷ which is the most frequent type of pain. It is by nature a transient response to actually or potentially harmful factors, triggering evasive action and protective behaviors. Inflammatory pain is one of the most representative subtypes of nociceptive pain. Somatic nociceptive pain is usually perceived in the dermis layer and is often described as lancinating, sharp or burning pain. In contrast, the sensation of visceral nociceptive pain is blurry and diffuse. The pain generated by cutting, burn and corrosion injuries can be classified as nociceptive pain.

Neuropathic pain

Neuropathic pain is defined as pain arising as a direct consequence of a lesion or disease affecting the somatosensory system, including central neurons and peripheral fibers (AB, AS, and C fibers). According to epidemiological investigations, 7-10% of the general population experiences neuropathic pain, accounting for 20–25% of patients suffering from chronic pain.^{28,29} The prevalence of neuropathic pain is dramatically increased in individuals with specific chronic diseases due to its mechanistic particularity. Diabetic polyneuropathy, cancer, herpes zoster, multiple sclerosis and spinal cord injury are important diseases with secondary involvement in neuropathic pain. Patients with neuropathic pain typically experience a series of manifestations, such as burning and electrical-shock sensations. Persistence and poor responses to analgesics create enormous health burdens for patients, usually accompanied by psychiatric disorders, such as depression, anxiety and insomnia.

Nociplastic pain

Some patients with explicit pain phenotypes fail to present with organic lesions and therefore cannot be classified as either of aforementioned types. In 2016, the concept of nociplastic pain was proposed and defined as a mechanistic descriptor for chronic pain states not characterized by clear activation of nociceptors or neuropathy but exhibiting clinical and psychophysical findings suggestive of altered nociceptive function. Its prevalence in the general population ranges from 5% to 15%, and there is a significant female preference.³⁰ Nociplastic pain is divided into five categories: chronic widespread pain, chronic primary musculoskeletal pain, chronic primary visceral pain, chronic primary headache pain and complex regional pain syndrome.³¹ Genetic, psychosocial, and environmental factors jointly contribute to the progression of nociplastic pain.³²

ANIMAL MODELS APPLIED FOR CURRENT RESEARCH ON PAIN

Experimental animal models are indispensable tools for basic and preclinical investigations into occurrence, diagnosis and treatment of pain. As pain is a multimodal event, an ideal pain model should encompass both biological and psychological factors. A diverse array of model preparation methods has been developed, including physical damages, chemical and biological irritants and psychosocial stressors (Fig. 2). Regrettably, standardized modeling approach that perfectly replicates pain development is still lacking. Most current models fail to accurately represent the mechanisms of specific pain types, potentially compromising the validity of basic research findings. In this section, we summarize the commonly employed methods of pain model generation to provide the swift access to pain research field for readers.

Physical damages

Surgery is a common method for generating nociceptive and neuropathic pain models. Chronic constriction injury (CCI), spared nerve injury (SNI) and spinal nerve ligation (SNL) are classical approaches for inducing neuropathic pain. CCI is produced by placing loosely constrictive ligatures around the common sciatic nerve. SNI entails the incision of tibial and common peroneal nerves, sparing the sural nerve. Therefore, a key advantage of SNI is better observation of impacts of injured and non-injured nerves. Following these procedures, the metapedes of both models typically develop hyperalgesia, and the mechanical withdrawal threshold decreases. Hyperalgesia usually peaks after 7 days of surgery and persists over two months. The spontaneous ongoing pain also becomes detectable after 7 days. The spinal nerve, due to its accessible anatomical position and significant physiological functions, is another idea target. Commonly, the L5 spinal nerve, located near the dorsal root ganglion (DRG) is selected for SNL modeling.³³ Pain perception typically develops within in 1–3 days, sooner than in CCI and SNI models. The mechanical and heat hyperalgesia can sustain 10 and 3 weeks, respectively. The spontaneous pain phenotype develops after one month of SNL.³⁴ The significant advantage of SNL is better investigations into the impacts on DRG. It is noteworthy that neonatal subjects may not experience mechanical allodynia or undergo delayedonset pain sensitivity following SNI, CCI, and SNL modeling, suggesting their unsuitability for early-life neuropathic pain studies. Although these three methods simulate physical nerve injury, it still remains unclear whether they can recapitulate the common diseases of neuropathic pain, such as diabetic neuropathy, neuropathic low back pain and radiculopathy.³⁷ Therefore, it should be cautious to draw conclusions concerning associations between etiological factors and clinical neuropathic pain based on these models.

Given the organ and tissues specificity of innervation, some studies exploring topical pain-associated diseases involve surgical damage to specific nerves to induce hyperalgesia at targeted sites. 4

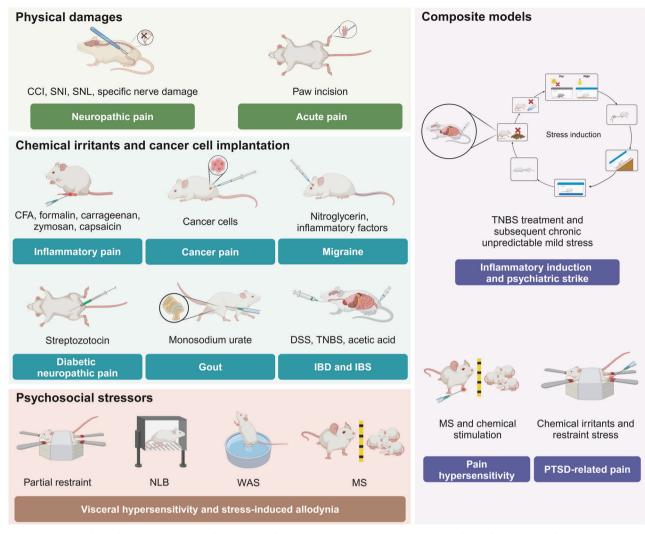


Fig. 2 Current animal models in pain research. Physical damages, chemical irritants, cancer cell implantation and psychosocial stressors constitute the three primary methods for preparing pain models. Furthermore, composite regimens that combine several of the aforementioned methods have been employed as pain is a multifactorial event. CCI chronic constriction injury, CFA complete Freund's adjuvant, DSS dextran sulfate sodium, IBD inflammatory bowel disease, IBS irritable bowel syndrome, MS maternal separation, NLB neonatal limited bedding, PTSD posttraumatic stress disorder, SNI spared nerve injury, SNL spinal nerve ligation, TNBS 2,4,6-trinitrobenzene sulfonic acid, WAS water avoidance stress

For example, T9 laminectomy combined with radical contusion damage is used to simulate spinal cord injury.³⁸ Trigeminal nerve root compression in inferior orbital fissure or inferior alveolar nerve is performed to generate animal models of trigeminal neuropathic pain.^{39,40} Furthermore, paw incision is an effective approach to imitate the status of postoperative pain or acute pain, which is extensively applied due to the simplicity and reproducibility.⁴¹

However, there are two significant limitations of physical damage models. First, despite precise intervention, inflammatory pain following operations, particularly in the acute phase, is inevitable. Consequently, research conclusions should be interpreted cautiously and comprehensively. Second, most methods are "all or nothing". They lack the capability to control the extent of damage, rendering them unsuitable for studies investigating the effects of varying degrees of nerve damage, with partially different underlying mechanisms. Electrocautery tends to progress into persistent allodynia,⁴² making it more suitable for the research on pain chronicity. Additionally, electrical stimulation is also employed to trigger pain sensations. Its non-invasive nature is noteworthy. Potential inflammatory responses following invasive

operations can be significantly reduced. Furthermore, some studies have verified the antalgic role of electrical stimulation.⁴³ Differentiating its pain-inducing and pain-relieving effects requires further investigation.

Chemical irritants and cancer cell implantation

Complete Freund's adjuvant (CFA) is a water-in-oil solvent composed of mineral oil, dead *Mycobacterium tuberculosis* and an antigen salt solution. It is extensively used in preparing topically inflammatory pain or arthritis models by injection into the paw or arthrosis, respectively. Paw injection of CFA can induce pain hypersensitivity and non-evoked ongoing pain after 24 h and it will last for 1–2 weeks. Joint pain occurs after 7 days of intra-articular injection. High-dose CFA is one of few approaches to generate models at the chronic phase of pain. Furthermore, CFA elicits synovitis and bone resorption without cartilage alteration, thus it has been evaluated as a robust model for the research on rheumatic arthritis.⁴⁴

Formalin is a protein coagulant commonly employed for tissue and cell fixation. Subcutaneous injection of formalin diluent into animal hind paws can generate local pain. Formalininduced evoked pain and spontaneous ongoing pain are characterized by a two-phase response. The first phase (0-5 min) results from the activation of peripheral nociceptors, whereas the second phase (10-40 min) reflects the development of inflammation and central sensitization.⁴⁵ Low-dose formalin directly activates nociceptors, while injection of high-dose formalin can exert additional tissue damage and inflammatory stimuli.⁴⁶ Hence, the evidences indicate a significant time and dose-dependent manner of formalin-induced pain. It is usually employed for investigations into pain mechanisms. Additionally, topical injection of carrageenan is mainly used for preparing transient joint inflammation. The hyperalgesia and spontaneous nociceptive behaviors occur within 3-5 h and lasts for 24 h. Zymosan is a typical agent for acute inflammation research. It can induce thermal and mechanical hyperalgesia after 30 min in a dose-dependent manner. Spontaneous pain can be observed after 24 h of high-dose injection of zymosan.⁴⁷ Capsaicin is commonly used for construction of skin inflammation and inflammatory bowel disease (IBD), as well as examination of analgesic drug efficacy. It promptly triggers evoked pain perception and fades within 1 h. Spontaneous ongoing pain occurs primarily within 5 min. Compared to the sustained and biphasic pain induced by formalin, it exhibits shorter lasting and monophasic duration.⁴⁸ Intriguingly, high-dose or continuous treatment reversely lead to neuronal desensitization and analgesic effects. The modeling regimens should be carefully investigated before generating pain modeling using capsaicin.

Notably, chemical pain inducers play a crucial role in generating models of gastrointestinal disorder-associated pain. Intrarectal administration of dextran sulfate sodium (DSS) and oral treatment with 2,4,6-trinitrobenzene sulfonic acid (TNBS) are classical methods for inducing IBD. The symptoms of visceral hypersensitivity are detectable within several weeks. The pathology induced by DSS shares more features of ulcerative colitis, while the immunological and histopathological mechanisms underlying Crohn's disease progression are following TNBS treatment.49 Researchers should choose proper chemical irritants according to disease types. For the research on irritable bowel syndrome (IBS), intracolonic injection of zymosan or acetic acid is commonly used, whereas with different treatment periods. Zymosan-induced visceral hypersensitivity can be detected only after 3 days.⁵⁰ Acetic acid requires 2-week continuous treatment.

Specific chemical agents are utilized for pain models based on disease-specific etiologies. For instance, diabetic neuropathy is one of the important causes of chronic pain. The streptDSSozotocin-induced diabetes model is used to investigate the mechanisms underlying neuropathic pain. The baseline of blood glucose is significantly elevated after intraperitoneal or intravenous injection of 3 days. Thermal and mechanical allodynia will be detectable after 2 weeks.⁵² The spontaneous ongoing pain lags behind evoked pain, which occurs after ~4 weeks.⁵³ Vasoconstriction dysfunction and inflammatory mediators are the etiological factors of migraine. Therefore, nitroglycerin or inflammatory factors are available for the generation of migraine models. Intravenous, subcutaneous or intraperitoneal injection of nitroglycerin induces acute evoked and spontaneous pain for 3-5 h. Repeated treatment can lead to the progression of chronic pain.⁵⁴ Topical administration of inflammatory factors, such as 5-hydroxytryptamine (5-HT), prostaglandin, histamine and bradykinin, relies on dural cannulation. Briefly, the rodents undergo craniotomy and cannula is inserted into the dura. The inflammatory factor solution is then pumped into the dura through cannula. This method can greatly avoid the effects of systemic delivery on other organs by nitroglycerin treatment, increasing the reliability of migraine models, however with a significant increase in operation complexity. Infusion numbers are positively Pathology of pain and its implications for therapeutic interventions Cao et al.

correlated with pain hypersensitivity and duration. Three-time infusion-induced hypersensitivity sustains ~1.5 h and eight-time infusion contributes to the prolongation into more than 5 h. However, this method cannot initiate long-period spontaneous pain.⁵⁵ Intraarticular injection of monosodium urate is a standard method for gout models. The swelling, mechanical hyperalgesia and ongoing pain can be observed after 2 h of injection. The symptoms reach the peak at 24 h and persist until 48 h.^{56,57}

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The mechanisms and treatment of cancer pain are hot topics of basic studies. Schwei et al. reported a bone cancer pain model.⁵⁸ This research has sparked an upsurge in research on pain associated with various types of cancer. This technique involves local transplantation of cancer cells, with pain hypersensitivity and spontaneous pain detectable within 1–2 weeks.⁵⁹ The concrete time varies based on cancer types, tumor cell loads and locations. Moreover, certain chemotherapeutic drugs for cancer treatment have significant neurotoxicity and easily cause neuropathic pain, such as oxaliplatin and paclitaxel. Administration of chemotherapeutic drugs can lead to phenotypes of hyperalgesia and spontaneous pain within 1 week.⁶⁰

Psychosocial stressors

Psychological and social factors are integral in pain perception, leading to the creation of animal models through psychosocial impairment techniques. These methods are employed to simulate clinically stress-associated diseases, such as IBS, gastric hyperalgesia and IC/BPS. For instance, partial restraint, a nontraumatic stress model, refers to wrapping the upper part of the animal's trunk in paper tape for several hours daily. The subjects develop spontaneous pain within a couple of weeks. However, this method is unsuitable for neonatal investigations into early-life stress. Neonatal limited bedding (NLB), which refers to subjects housed in a parochial cage, serves as an earlylife stressor. Subjects will endure persistent somatic hyperalgesia.⁶¹ Water avoidance stress (WAS) is another method to achieve movement restraint. Mice are placed on a small platform inadequate for standing by all fours and surrounded by water, which forces the subjects to remain continuously vigilant, resulting in a strong stress response. Repeated WAS can induce typical visceral hypersensitivity. Nevertheless, there have been no studies examining the effects of NLB and WAS on spontaneous ongoing pain.

The social relationship damage is also employed in pain model generation. Maternal separation (MS) is the severance between juvenile individuals and their dependent subjects, which affects nervous system development and increases the risks of adult psychiatric disorders.⁶² Therefore, MS has also been conducted in basic research on nociplastic pain. Current studies using this method have focused on visceral hypersensitivity. Animals undergoing early-life MS will suffer from pain hypersensitivity at the adult phase. Separation time is a critical factor in the effects of MS. Brief separation, more parallel to mother scavenging for food, has a relatively mild impact on juvenile subjects, whereas severe anxiety behaviors are observed following long-term separation. Likewise, the studies focusing on MS and spontaneous ongoing pain is still lacking.

Composite models

Despite the apparent pain-inducing effects of the above singlefactor models, their limitations on disease reducibility are obvious due to the etiological complexities of pain in patients. Some studies tried to simultaneously use several methods to corroborate each other.^{63,64} Furthermore, the comprehensive modeling strategies based on existing approaches can maximize the simulation capabilities of pain models. For example, the TC-IBS method includes trinitro-benzene-sulfonic acid treatment and

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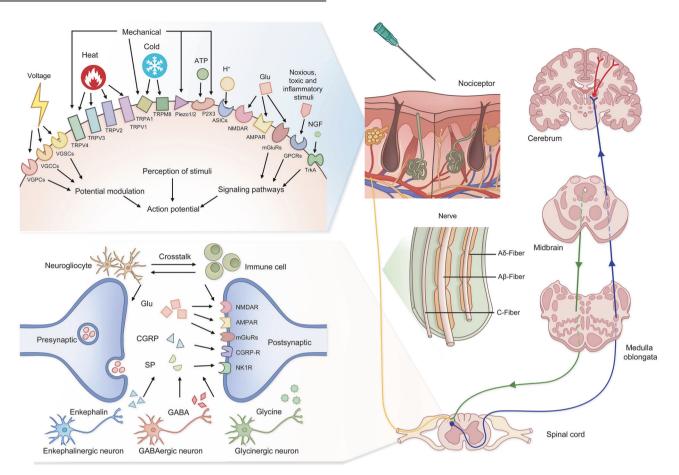


Fig. 3 Schematic illustration of pain sensation pathways. The exposure to pain-inducing events changes activity of specific receptors and activates action potential of peripheral nociceptors. The signals are then transmitted from DRG located to the spinal cord via afferent nerves. The nerves are categorized into $A\beta$, $A\delta$ and C fibers. During neuronal transmission, the presynaptic membrane releases various neurotransmitters into the subsynaptic membrane, inducing potential alterations in the subsequent neuron. The figure shows some representative neurotransmitters in during pain perception. Additionally, neurogliocytes, immune cells and other types of neurons collaboratively modulate pain signals. The DRG, as the relay station, is responsible for ascending transmission to the corresponding sensory cortex, which modulates the ultimate pain sensation. The descending regulatory pathways also play a role in pain modulation. ASIC acid-sensing ion channel, AMPAR α -amino-3-hydroxy-5- methylisoxazole-4-propionate receptor, CGRP calcitonin gene-related peptide, GABA gamma-aminobutyric acid, GPCR G protein-coupled receptor, mGluR metabotropic glutamate receptor, NGF nerve growth factor, NMDAR N-methyl-D-aspartate receptor potential melastatin 8, TRPV1 transient receptor potential vanilloid 1, TRPV2 transient receptor potential vanilloid 2, TRPV3 transient receptor potential vanilloid 3, TRPV4 transient receptor potential vanilloid 4, VGCC voltage-gated calcium channel, VGPC voltage-gated potassium channel, VGSC voltage-gated sodium channel

subsequent chronic unpredictable mild stress, with properties of both inflammatory induction and psychiatric strike.⁶⁵ Similarly, MS and chemical stimulation have been assembled to prepare a pain hypersensitivity model.⁶⁶ In the research on PTSD-related pain, chemical irritants and restraint stress are used simultaneously. Nervous system homeostasis and neurologic functions are typically disrupted, akin to PTSD symptoms.⁶⁷ Although these models are based on the superposition of different factors, they offer valuable insights for developing more scientific models for pain research.

BASIC CIRCUITS OF PAIN

Pain perception is a complex physiological process involving both the central nervous system (CNS) and peripheral nervous system (PNS). Numerous nervous structures, cells and molecules collectively underlie the transduction, transmission, modulation and perception of pain signals (Fig. 3). This section provides an overview of the basic mechanisms of pain perception for readers to better understand the subsequent contents in this review.

Peripheral transmission of pain signals

Nociceptors, peripheral transducers of pain signals, are located in the skin, mucosa, muscles, surface and interior of tendons, periosteum, vasculature, and internal organs. They are morphologically free or undifferentiated nerve endings, the cell bodies of which reside in the DRG and trigeminal ganglion. According to the received noxious stimuli, they can be divided into thermosensitive, mechanical-sensitive and injury signal-sensitive types. Compared to other sensors, the activation thresholds of nociceptors are relatively higher, ensuring that human body perceives normal tactile information without pain. Nociceptors are regarded as the gatekeepers and initiators of pain sensation.

The peripheral terminals of nociceptors have many types of ion channels, which can perceive external stimuli, code signals and generate membrane excitability. Ion channels produce electrical signals through regulating the ion current across membranes. The adjacent voltage-sensitive channels are forced open in a chain reaction. According to the precipitating factors of channel opening, they can be generally divided into two categories, voltage-gated ion channels and ligand-gated ion channels.

Voltage-gated ion channels refer to a kind of transmembrane proteins whose conformation is determined by membrane potentials. They play a crucial role in converting receptor potentials into a series of action potentials. Voltage-gated sodium channel (VGSC) family comprises 9 members, including Nav1.1 to Nav1.9. VGSCs rapidly adopt open conformations following cell membrane depolarization, allowing sodium to flow into cells down a concentration gradient. This process initiates action potentials and produces pain signals at nerve endings. VGSCs have typical differences in species, spatial and temporal distributions, as well as electrophysiological characteristics.^{68,69} Nav1.7, in particular, has garnered significant attention. Mutations in the Nav1.7 encoding gene Scn9a are associated with various pain disorders, such as inherited erythromelalgia, paroxysmal extreme pain disorder and small-fiber neuropathy.⁷⁰ Inhibiting Nav1.7 functions effectively mitigates neuropathic pain and stimulates the production of endogenous opioids.⁷¹ The role of Nav1.7 varies with different types of pain. For instance, it contributes to the development of neuropathic pain, whereas bone cancer pain and oxaliplatin-induced pain do not depend on Nav1.7-postive nociceptors.⁷² Other VGSCs, like Nav1.1, Nav1.6, and Nav1.8, also play important roles in pain modulation.^{68,73,7}

Voltage-gated calcium channels (VGCCs) are distributed in all types of excitable cells. They are composed of four subunits: $\alpha 1$, $\beta 1$ -4, $\alpha 2\delta 1$ -4, and $\gamma 1$ -8. Each VGCC type has a unique subunit composition, with $\alpha 2\delta$ being a crucial component. $\alpha 2\delta$ interacts with $\alpha 1$ and β subunits, enhancing peak potentials and rates of channel activation and inactivation. Noxious stimuli can upregulate $\alpha 2\delta$ expression in both the CNS and PNS, subsequently augmenting pain signals.^{75,76} The functions of calcium channels in sensory neurons are finely tuned by various factors, like adiponectin, neuromedin B and non-coding RNAs.⁷⁷⁻⁷⁹

In contrast to VGSCs and VGCCs, voltage-gated potassium channels (VGPCs) primarily facilitate potassium outflow from neurons, inducing membrane hyperpolarization and neuronal excitability attenuation. Noxious stimuli, such as mechanical force, heat and algogens, can downregulate potassium channel expression and inhibit their activity,⁸⁰ leading to ectopic spontaneous discharges in nociceptors.⁸¹

Transient receptor potential (TRP) channels, the most representative ligand-gated ion channels, are extensively distributed in both the CNS and PNS. TRP family members act as molecular sensors of pain and itch, responding to physical and chemical stimuli. Currently, 28 TRP members have been identified, with well-documented biological functions for TRPV1, TRPV2, TRPV3, TRPV4, TRPA1, and TRPM8.⁸² The mechanistic associations of TRPV1 and TRPA1 with pain modulation have been largely investigated. Their activation states and expression levels are positively associated with pain sensation.^{83,84} Intriguingly, variouNMDARs natural biotoxins induce pain perception just through targeting TRPV1 and TRPA1,^{85,86} demonstrating the ingeniousness of interspecies evolution. The TRP channel antagonists, like V116517 and BCTC, have shown significant potential in pain management.^{87,88}

N-methyl-D-aspartate receptors (NMDARs), consisting of various GluN subunits, are particularly sensitive to mechanical stimulation. Calcium influx through NMDARs is a critical inducer of electrical signal activation.⁸⁹ NMDARs interact with calcium channel subunit $a2\delta$, tonically activating primary afferent neurons.⁹⁰ The ion-specific permeability is controlled by Mg²⁺, and neuronal depolarization contributes to the activation of NMDARs. Both presynaptic and postsynaptic NMDARs modulate excitatory synaptic transmission and CNS synaptic plasticity, facilitating hyperalgesia.^{91,92} Neuropathic pain persistently activates NMDARs under continuous endogenous glutamate stimulation.^{93,94} This vicious cycle further exacerbates pain chronicity .

A variety of other channels distributed in nociceptors mediate the transformation of noxious stimuli into electrical signals. The calcium-permeable purinergic receptor (P2X) channels, another class of receptors for pain sensitization, exhibit hyperactivity dependent on extracellular ATP released from damaged cells.⁹⁵ Acid-sensing ion channels (ASICs), members of the epithelial sodium channel/degenerin family, detect alterations in extracellular pH and mediate hyperalgesia during the progression of inflammation, ischemia and cancer metastasis.⁹⁶ ASIC3 unilaterally suppresses P2X3 receptor currents through forming the protein complex due to their similar molecular structures and cellular colocalization,⁹⁷ illustrating the molecular mechanisms underlying pain harmonization. The identification of other receptors, such as Piezo 1/2, bradykinin and nicotinic acetylcholine receptors, has further expanded the understanding in pain generation mechanisms.⁹⁸

In addition to ion channels, signaling transduction receptors likewise receive stimuli and modulate pain-related signaling pathways. G protein-coupled receptors (GPCRs), the largest family of transmembrane proteins, mediate the physiological control of nociceptive transmission. GPCRs are widely expressed in neurons and other pain-associated cells, like glial and immune cells. Various pain-related ligands, such as opioids, glutamate, bradykinin, gamma-aminobutyric acid (GABA), cannabinoids, 5-HT, prostaglandins and histamine, have been identified. Activation of GPCRs leads to conformational changes in the α subunit, which regulates the activity of adenylate cyclase and phospholipase C, influencing the production of secondary messengers.⁹⁹ The mechanisms exert global effects on pain events, involving neuronal excitability, inflammatory response, intercellular com-munication and neurotransmitter release.¹⁰⁰ The functions of some ion channels are under the rigid control of GPCR-mediated signaling.¹⁰¹ Additionally, tropomyosin receptor kinases are receptors for neurotrophic factors such as nerve growth factor (NGF) and brain-derived neurotrophic factor (BDNF), responsible for neuronal survival and growth, immune homeostasis and neurotransmitter selection.¹

The electrical signals produced by nociceptors are subsequently transmitted to spinal cord via nerve afferent fibers, which are essentially axons of sensory neurons. There are three main categories of primary afferent fibers according to anatomical characteristics and functions, including AB, AS and C fibers. Spatial transcriptomics data further link the sensor subtypes with these nerve fibers.¹⁰³ Nociceptors pertain to Aδ and C fibers; effective stimulation to Aδ fibers induces sharp, needle-like fast pain, while C fiber activation leads to burning, blunt or inaccurately localized pain. A-type nerve fibers are wrapped with the myelin sheath composed of Schwann cells, accelerating transmission of action potentials. Recent studies have uncovered that Schwann cells respond to noxious stimuli and elicit intracellular oxidative stress reactions. The paracrine release of reactive oxygen species, like 4-hydroxynonenal and H₂O₂, significantly stimulates TRPA1, sustaining pain percep-^{4,104} Additionally, Nav1.7 is required for the initiation of tion.² action potentials in C fibers.⁷¹ Injury and inflammation can cause nerve fiber sensitization, decreasing thresholds and inducing persistent pain experiences.^{105,106}

Ascending pain transduction tracts

A single noxious event can activate multiple neurons in the DRG. Spinal projection neurons receive, integrate and transmit the complex signals into ascending pathways, thereby completing the conversion from the PNS to the CNS.

The spinothalamic tract and the trigeminothalamic tract are two fundamental pathways of somatalgia transduction. Specifically, the spinothalamic tract is shunted into lateral and anterior branches at the anterior white commissure. The lateral spinothalamic tract is responsible for transmitting pain information from the torso and limbs, while the trigeminothalamic tract consists of fibers from the spinal trigeminal nucleus and most pontine 7

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Table 1.	The summary	of neurotransmitters	involved in pai	n sensation
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Neurotransmitter	Туре	Receptor	Distribution	Function
Glutamate	Amino acid	NMDAR, AMPAR, mGluR	CNS, PNS	Excitatory
Aspartate	Amino acid	NMDAR, AMPAR, Kainate receptor	CNS	Excitatory
Histamine	Monoamine	H1	CNS, PNS	Excitatory
CGRP	Peptide	Heterotrimers of CALCRL, RAMP1 and RCP	CNS, PNS	Excitatory
Substance P	Peptide	NK1, NK2, NK3	CNS, PNS	Excitatory
GABA	Amino acid	GABA _A , GABA _B	CNS, PNS	Inhibitory
Glycine	Amino acid	GlyR	CNS	Inhibitory
Endogenous opioids	Peptide	μ -, κ- and δ-receptors	CNS, PNS	Inhibitory
Cannabinoids	Lipid	CB1, CB2	CNS, PNS	Inhibitory
5-HT	Indole derivative	5-HT1-4, 7	CNS, PNS	Dependent on 5-HT receptors
Norepinephrine	Monoamine	α 1, α 2, β adrenergic receptors	CNS, PNS	Excitatory (α 1, β), inhibitory (α 2
NO	Gasotransmitter	Diffusion across membrane	CNS, PNS	Excitatory or inhibitory

5-HT 5-hydroxytryptamine, AMPAR α-amino-3-hydroxy-5- methylisoxazole-4-propionate receptor, CALCRL calcitonin-receptor-like receptor, CB cannabinoid receptor, CGRP calcitonin gene-related peptide, CNS central nerve system, GABA gamma-aminobutyric acid, GlyR glycine receptor, mGluR metabotropic glutamate receptor, NK neurokinin, NMDAR N-methyl-D-aspartate receptor, NO nitric oxide, PNS peripheral nervous system, RAMP1 receptor activity modifying protein 1, RCP CGRP-receptor component protein

trigeminal nuclei, conveying pain sensations from the head and face. Additionally, there are other ascending pathways for somatalgia transduction, such as the spinomesencephalic tract, spinoreticular tract, spinohypothalamic tract and dorsal column postsynaptic fiber bundles.

In contrast, pathways for visceral pain transduction are more decentralized and do not conform to classical neural circuits. The cell bodies of afferent visceral sensory neurons through sympathetic nerves are located in the T1-L3 spinal ganglion. Those via parasympathetic nerves are distributed in the sensory ganglia of the glossopharyngeal nerve and vagus, as well as in the S2-4 spinal ganglion. The axons of afferent visceral sensory neurons are distributed throughout internal organs and their blood vessels.

Action potentials induce the release of various excitatory neurotransmitters and neuromodulators derived from neurons with different functions. They collectively bind to postsynaptic receptors, activating the excitability of next-level neurons. Glutamate, a typical excitatory neurotransmitter, is produced by excitatory neurons. Its release directly induces an excitatory postsynaptic potential. Conversely, inhibitory neurons secrete specific neurotransmitters, such as GABA, glycine and opioids, to prevent neuronal overexcitation. The identified neurotransmitters are listed in Table 1. Neuromodulators do not directly trigger alterations in postsynaptic biological effects. They mainly regulate neurotransmitter release and neuron excitability mainly through binding to GPCRs and rearranging intracellular signaling pathways. For instance, BDNF, has dual functions in pain signal modulation in both the PNS and CNS.^{107,108} Calcitonin gene-related peptide (CGRP) has properties of the enhancement in presynaptic glutamate transmission and responses to substance P.^{109,110}

Inflammatory cells (mastocyte, neutrophil, microglia, etc.) and mediators (TNF-α, interleukins, prostaglandin E2, etc.) act on neurons and affiliated cells within the nervous system. The activation of inflammatory cells and ectopic release of proinflammatory factors sensitize nociceptors and reduce pain threshold, facilitating responses to noxious stimuli and pain sensation caused by innocuous stimuli, like tickling.^{111–113} Long-term inflammation alters nervous system plasticity and promotes pain chronicity.¹¹⁴ The effects of neutrophils are time-phase dependent. Acute activation of neutrophil accelerates pain resolution. Early treatment with NSAIDs to inhibit inflammatory responses conversely prolongs the course of pain perception.¹¹⁵ Long-term infiltration of neutrophils leads to the exaggeration of nociceptive responses.¹¹⁶

Recent studies have demonstrated the impact of interactions between inflammatory cells and the nervous system on pain perception. For example, astrocytes can directly enhance microglia activity and promote pain perception.^{117,118} Lipocalin-2, an important secretory factor derived from activated astrocytes, also elevates TRPV4 expression and further promotes microglia activation.¹¹³ Macrophages secrete proinflammatory factors like IL-33 and recruit neutrophils. The inflammation-inducing effects of neutrophils can be promoted by IL-33, facilitating DRG excitability via TRPV1 channel activation.⁵⁶

Pain perception and modulation in the brain

The cerebrum coordinates the crosstalk among afferent axons, interneurons and projection neurons. A great number of clinical neuroimaging studies and basic research have identified a "pain matrix" in the brain, including a variety of brain nuclei like the periaqueductal gray (PAG), thalamus, hypothalamus, parabrachial nuclei (PB), nucleus tractus solitarius, amygdala, insular cortex (IC), somatosensory cortex, anterior cingulate cortex (ACC), prefrontal cortex (PFC). These areas appear to collaboratively modulate pain signals. The primary somatosensory cortex (S1) is a central hub of noxious sensation, receiving thalamocortical input from the ventral posterolateral thalamus.¹¹⁹ The output signals from S1 are transmitted to thalamic nuclei and several subcortical targets.¹²⁰ The neuronal activity and connectivity in S1 are dramatically enhanced following pain signal transmission, with neurons in layer 6 amplifying thalamocortical signaling while inhibiting innate antinociceptive mechanisms.¹²¹ Furthermore, the lateral PB and thalamus are key areas in receiving nociceptive projections from the spinal cord and integrating competitive signals that modulate pain. Chronic pain promotes significant changes in sensory circuit reorganization and metabolic patterns of these brain regions,^{122,123} further facilitating central sensitization. Along with PB, the central amygdala (CeA) also mediates pathophysiologic effects and behavioral responses to noxious stimuli through neural circuits connecting with brainstem and hypothalamus.^{124,12}

Pain perception depends not only on the damage degree but also on emotional, social and environmental factors. The degree of attention paid to pain, cognitive appraisal of threats and individual character are all associated with pain perception. Depressive states may aggravate or even directly elicit pain.¹²⁶ Pain memory can also reproduce forepassed pain feelings and cause hyperalcesia.¹²⁷ In terms of brain regions involved in emotion modulation, areas like S1, IC, ACC and PFC process afferent signals to generate pain perception and affection.¹²⁸ Particularly, the medial PFC (mPFC) plays a critical role in the development of chronic pain.¹²⁹ A subgroup of specific neuronal ensembles in the dorsomedial PFC processes nociceptive information and regulates pain chronicity.¹²⁹ Two implicated clusters of PFC neurons project to limbic regions, including the hypothalamus, nucleus accumbens (NAc) and amygdala, underpinning the negative emotional and physiological impacts on chronic pain.¹³⁰ Dopaminergic pathways in the ventral tegmental area and the substantia nigra compacta-NAc projection are responsible for pain aversion and pain-relief reward modulation.¹³¹ In summary, many important neural circuits contribute to the development of pain comorbidity.

Apart from the traditional pain circuits, many other areas are implicated in the perception and modulation of pain. For example, the entorhinal cortex and medial septum to hippocampus circuit, typically recognized for their roles in learning, memory or emotion regulation, have been found to modulate pathological pain.^{132–134} A contribution from adult hippocampal neurogenesis underlies pain chronicity, as well as the alleviative effects of environmental enrichment and exercise on chronic pain.^{135,136}

In addition to the ascending pain transduction tracts, many brain areas in the descending pain modulation pathways are involved, such as the PAG, rostral ventromedial medulla (RVM), locus coeruleus, lateral reticular nucleus, nucleus raphes magnus, nucleus reticularis paragiagantocellularis. These pathways are classified as descending inhibitory and facilitatory systems. They collectively keep a subtle balance under normal conditions. The development of pain hypersensitivity can disrupt this balance, favoring the descending facilitatory system.

The PAG, RVM and a portion of the pontine dorsolateral reticular formation participate in the descending inhibitory system. These brain areas exert inhibitory modulation on nociceptive information via the descending spinal dorsolateral tract. Multiple nuclei have their own descending pathways towards spinal dorsal horn. The PAG is a hub of descending inhibitory system, with the dorsomedial PFC, amygdala and hypothalamus regulating its activation.^{137–139} The projection from the PAG to RVM and locus coeruleus attenuates neuropathic pain and accompanying emotional dysregulation.^{140,141} The RVM receives input signals from the PAG, nucleus tractus solitaries, parabrachial nucleus and other supraspinal sites processing nociceptive information. Two types of functional neurons, Oncell and Off-cell, have been identified in the RVM, executing descending facilitation and inhibition, and collaboratively determining the adjusting strengths of the RVM.¹⁴²

The descending facilitatory system enhances responses to noxious stimuli by decreasing pain thresholds. It is not typically active during normal pain perception. Pain hypersensitivity and catastrophizing can suppress the activity of the inhibitory system while arousing the facilitatory system.^{143,144} The RVM, ACC, PAG, parabrachial nucleus are major components of this system.¹⁴⁵ Despite distinct regulatory functions, it is not difficult to find that there is considerable overlap in anatomical regions. The bidirectional functions of the RVM have been greatly manifested, possibly attributable to different subtypes of neurons in the nuclei. Descending facilitation from this region is the critical factor of neuropathic pain development.¹⁴⁶ Furthermore, physiological behaviors are partially governed by the limbic system, which modulates activities of the brain regions in descending facilitatory system.¹⁴⁷ This is an essential mechanism by which pain perception is affected by individual emotion, experience and memory.

Default mode network (DMN) is a collection of brain regions that are actively engaged when an individual is at rest or not

actively engaged in a task that requires external attention. DMN mainly includes the precuneus, posterior cingulate cortex, mPFC, medial temporal lobe and angular gyrus. These regions are interconnected and work together to support internally focused cognition, including self-reflection, episodic memory retrieval, and mind-wandering. DMN is implicated in the regulation of pain sensation. Pain competes for cognitive resources with other kinds of attention-demanding stimuli, which is closely associated with DMN dysfunctions. The functional connectivity between brain regions within DMN is enhanced and its amplification is correlated with pain severity.¹⁴⁸ Specifically, mPFC, a critical hub of DMN, can present increased high frequency oscillations. Its connectivity to the posterior constituents of the DMN is meanwhile impaired.¹⁴⁹ In the patients with persistent post-traumatic headache, PAGseeded functional connectivity is reduced, accompanied by the structural reconstruction of the ACC and posterior cingulate cortex.^{150,151} In addition to internal networks, the abnormality of DNM connectivity with other brain regions has been extensively discovered, involving the insula, ventral lateral/posterolateral nucleus and postcentral gyrus.¹⁵²⁻¹⁵⁴ The advances in DMN research provide strong proof for identifying mechanisms underlying emotional changes that affect pain perception. For instance, mind wandering restores the ectopic connectivity between PAG and DMN, redirecting spontaneous attention away from pain.¹⁵⁵ The thalamic-DMN decoupling has been proved as an important mechanism of mindfulness meditation.¹⁵⁶ On the other hand, negative mood promotes pain hypersensitivity through influencing DMN functional connectivity during the progression of chronic pain.¹⁵⁷ Notably, despite close associations between DMN and chronic pain shown by most studies, acute pain can likewise induce alterations in oscillatory activity and functional connectivity of DMN, which underpins attentional processes in the presence of pain.15

MOLECULAR MECHANISMS OF PAIN MODULATION

In addition to the basic circuits and corresponding molecules as introduced above, a series of molecular mechanisms underlie pain perception under intricate but well-regulated control. With the development of high-quality preclinical research, the scattered advancements are gradually converging into the systemic body of knowledge, contributing to the identification of numerous promising therapeutic targets. Herein, we summarize current achievements in related molecular mechanisms to present a more complete network of pain modulation (Fig. 4).

Gene mutation

Most gene mutations are neutral, but a small minority may cause diseases, including pain disorders. Various mutations can lead to totally different clinical outcomes, ranging from pain insensitivity to extreme pain sensation. Erythromelalgia, familial episodic pain syndrome, congenital insensitivity to pain with anhidrosis and Fabry disease are the representative inherited diseases with specific gene mutations. Due to the individual differences, mutation patterns associated with pain disorders are sporadic and most data have been presented as case reports. Mutations in ion channel-encoding genes account for a large portion of existing investigations.

Mutations of multiple sites of *Scn9a* gene cause truncation or function loss of Nav1.7, leading to congenital insensitivity to pain. Some cases are complicated with anosmia, while other patients have normal olfactory sensation,^{159–161} suggesting that mechanisms by which Nav1.7 modulates pain and olfaction partially overlap. Common missense mutants of *Scn9a* are correlated with pain severity of clinical patients with symptomatic disc herniation.¹⁶² Mutations in introns, which do not directly encode Nav1.7 protein, can also affect pain sensitivity. A novel homozygous substitution in *Scn9a* intron 3 interferes with mRNA splicing and leads to Nav1.7

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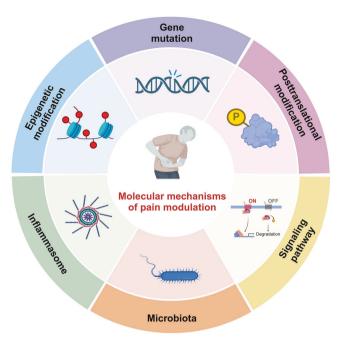


Fig. 4 The schematic illustration of molecular mechanisms underlying pain modulation. The molecular mechanisms are generally categorized into six aspects, including gene mutation, epigenetic modification, posttranslational modification, inflammasome, signaling pathways and microbiota. They orchestrate pain perception and modulation

inactivation. Furthermore, mutations are not confined to channel function deficiency. A1632E is a type of gain-of-function mutation. Nav1.7/A1632E mutants can form dimers and maintain persistent currents, exempt from the effects of inactivation particles targeting VGSCs.¹⁶³ Such non-canonical mechanisms greatly expand the understanding in gene mutation functions.

Mutations in other VGSC-encoding genes also contribute to the dysregulation of pain perception. Two missense mutations in Scn11a (c.673 C > T and c.2423 C > G) facilitate channel activity and promote hyperexcitability of Nav1.9 in DRG sensory neurons, which is a critical reason for familial episodic pain syndrome.¹⁶ The mutation at the R222S site of Scn11a has also been identified in patients with mechanical hyperalgesia sensitive to cold exposure.¹⁶⁵ Conversely, a heterozygous nonsynonymous mutation in exon 15 of Scn11a causes excessive activation at resting potential and sustained depolarization of nociceptors in individuals with the congenital inability to experience pain. The resultant action potential and excitatory transmission are impaired, leading to a loss of pain perception. This mechanism of overactivation-induced inactivation is similar to pain relief by capsaicin.¹⁶⁶ Additionally, the Nav1.1 channel with L263V missense mutation enhances spike activity induced by P2X3 and 5-HT3 receptors, increasing the excitability of peripheral trigeminal neurons and contributing to migraine pain.

One mutation pattern of the Cav3.1 channel in trigeminal neuralgia has been recently identified. The missense mutation of *Cacna1g* gene, encoding a1 subunit of Cav3.1, leads to the replacement of arginine with glutamine at position 706. Current density is enhanced and neuron excitability is significantly elevated.¹⁶⁸ Intriguingly, an a261 mutant with arginine at position 217 does not change pain sensitivity but blocks the analgesic efficacy of pregabalin for neuropathic pain.¹⁶⁹ This finding clearly demonstrates that the analgesic action of pregabalin relies on a261 subunit blockade.

In addition to VGSC and VGCC mutations, different VGPC variants have distinct impacts on pain sensitivity. A frameshift

mutation in *Kcnk18* gene, encoding the two-pore potassium channel, causes its loss of functions. Neuronal excitability is significantly increased, exaggerating mechanical and thermal hypersensitivity during migraine progression.¹⁷⁰ A recent study focusing on gene mutations in women requiring no analgesia during childbirth has identified *Kcng4* with excessive heterozygotes carrying the rare allele of SNP rs140124801. The product, Kv6.4 mutant, loses the capability of regulating Kv2.1 activity. The potassium outflux and sensory neuron hyperpolarization in uterus are promoted, attenuating childbirth pain.¹⁷¹

The roles of mutations in representative members of the TRP family in pain modulation have been unveiled. TRPV1 with N331K mutation directly causes functional deficiency.¹⁷² The G564S mutant is a gain-of-function variant. Nevertheless, the overactivation-induced inactivation is also observed in this mutation pattern. This membrane transport of G564S mutant is simultaneously inhibited.¹⁷³ Notably, in addition to natural mutation, Trpv1 gene can be chemically edited by an alkylating agent to produce a loss-of-function product.¹⁷⁴ For the research on TRPA1, the N855S mutant exhibits a fivefold increase in inward current in activated nociceptors, resulting in the development of familial episodic pain syndrome.^{175,176} A nonsense mutation in Trpa1 gene causes TRPV1 protein truncation, which can further assemble with wildtype TRPA1. The complex lowers energetic barriers and alters pore architecture, leading to neuronal hyperactivation.¹⁷

Additionally, mutations in genes regulating neuron development and axon outgrowth have been found to modulate pain sensation loss or sensitivity, including transcription factors, structural proteins, membrane channels and receptors.¹⁷⁸ Taken together, a great number of genes and mutated sites have been identified to have associations and causalities with pain. However, we have to acknowledge current research limitations: i) The concrete mechanisms by which these mutated proteins gain or lose functions are largely unknown. High-resolution structures and interactions may be promising research directions. ii) The typical individual differences in gene mutations mean that current achievements have lower universality, limiting their further clinical translation. iii) Few studies have investigated potential drugs targeting the mutants, leading to the dreadful scarcity of clinical therapies against congenital pain disorders. Robert et al. found a peptide with properties of blocking P2X7 receptor mutants without restraining normal channels, which is associated with nerve injury and inflammatory allodynia.¹⁷

Epigenetic modification

Despite differences in hereditary information, there are extremely high similarities in gene sequences between individuals with significantly different characteristics. Environment, behavior and age can produce apparent and persistent influences on humans. These phenomena cannot be forcefully explained by inherent genetic information alone. Epigenetics refers to alterations in gene expression not rooted in DNA sequences. Rapid advancements in epigenetics knowledge have unveiled novel mechanisms underlying physiological and pathological processes. It primarily includes DNA methylation, histone modification and non-coding RNAs. These three molecular mechanisms play essential roles in pain modulation (Fig. 5).

DNA methylation pertains to forms of DNA chemical modification. Catalysis of DNA methyltransferases (DNMTs) can transfer methyl groups derived from S-adenosylmethionine to specific bases. Most DNA methylation sites exhibit aggregated distributions, known as CpG islands. DNA methylation changes chromatin structure, DNA conformation, DNA stability and interactions with proteins, precisely regulating gene expression without editing base sequences. Studies have shown close associations between DNA methylation and pain perception.

Pathology of pain and its implications for therapeutic interventions Cao et al.

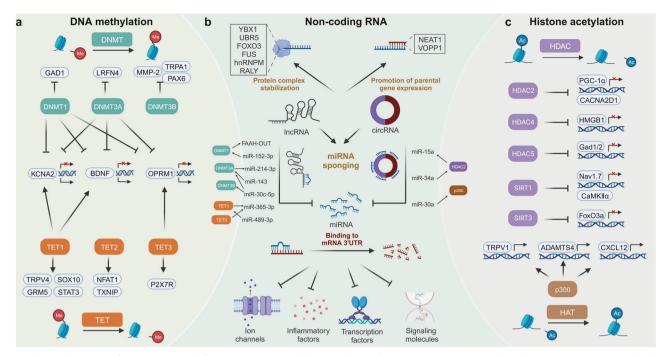


Fig. 5 The mechanisms of epigenetic modification in pain modulation. The mechanisms are categorized into three aspects: DNA methylation, non-coding RNA and histone acetylation. **a** For DNA methylation, DNMTs and TETs are responsible for DNA methylation and demethylation, respectively. They regulate expression of various genes associated with pain perception. The expression of KCNA2, BDNF and OPRM1 are simultaneously under the control of DNMTs and TETs. **b** Non-coding RNAs, comprising miRNAs, IncRNAs and circRNAs, play various roles. miRNAs can bind to 3'UTR of mRNAs associated with pain, negatively regulating their expression. Some IncRNAs and circRNAs act as miRNA sponges to counteract the functions of downstream targets. Certain IncRNAs and circRNAs directly interact with proteins to enhance their stabilization, thereby affecting pain sensitivity. Several non-coding RNAs, like IncRNA NEAT1 and circVOPP1, have been shown to stabilize the mRNAs of their parental genes related to pain to promote their expression. **c** HDACs and HATs collaboratively maintain the balance in histone acetylation. Specific HDACs, including HDAC2, HDAC4, HDAC5, SIRT1 and SIRT3, along with HAT p300, regulate expression of genes involved in pain modulation. Notably, non-coding RNAs regulate expression of enzymes associated with DNA methylation and histone acetylation. The expression of non-coding RNAs are, in turn, regulated by the other two mechanisms. circRNA microRNA, TET ten-eleven-translocation protein, UTR untranslated region

Patients suffering from chronic pain universally undergo significant changes in DNA methylation states, particularly in promoter regions.^{184,185} Global methylation data have been used to investigate pain-associated mechanisms with the support of bioinformatic analysis, such as G-protein coupled cholinergic signaling, neuron development and immunomodulation.^{184,186,187} DNA methylation has quick responses to pain. Its alterations can be detected at the early phase of neuropathic pain and persist chronically.¹⁸⁸ DNA methylation has disease and organ specificities. For example, there are huge differences between DNA methylation induced by diabetes neuropathy, nerve injury and chemotherapy, although they are all typical neuropathic pain. The CpG sites present prevailing hypomethylation in DRG, whereas the CNS, such as spinal cord and PFC, gains more DNA methyla-The methylation levels of genes encoding classical tion. positive regulators of neuropathic and nociplastic pain sensation, like TRPA1, CGRP, and BDNF, are significantly altered in patients with pain disorders. The methylation levels negatively regulate their expression, potentially causing hyperalgesia or pain insensitivity.^{190–193}

The DNMT family mainly consists of three enzymes with catalytic activity, including DNMT1, DNMT3A, and DNMT3B, responsible for adding methyl to specific gene regions. They generally present hypomethylation and participate in neuropathic pain modulation in both the CNS and PNS.¹⁹⁴ DNMT1 and DNMT3A upregulate methylation of promoter and 5'-untranslated region of *Kcna2* gene, decreasing membrane densities of VGPCs and Kv current, leading to central sensitization and neuropathic

pain.^{194,195} They also methylate promoters of genes encoding non-coding RNAs, with dysfunctions in these downstream noncoding RNAs contributing to various pain disorders, ranging from pain hypersensitivity to insensitivity.^{196,197} Systemic inhibition of DNMT activity results in alleviation of neuropathic pain.¹⁸⁵ Therefore, it can be concluded that despite manifold targets of DNA methylation, its overall effects are pain hypersensitivity.

Ten-eleven-translocation proteins (TETs) mediate DNA demethylation, dramatically maintaining DNA methylation stability and shaping epigenome landscape along with the DNMT family. TET1, TET2, and TET3 are the main members. The doublesided nature of TET1 has been revealed. On the one hand, it can remove restrictions on gene expression induced by DNA methylation during the progression of nociceptive and neuropathic pain, involving membrane receptors (mGluR5), ion channels (TRPV4), transcription factors (SOX10), and signal transduction factors (STAT3 and BDNF).¹⁹⁸⁻²⁰¹ On the other hand, some reports have shown the analgesic properties of TET1. It can rescue suppression of VGPC functions by regulating methylation of Kcna2 and K2p1.1 promoters in the neuropathic pain models.^{202,203} Restoration of PROX1 levels following TET1 overexpression attenuates depression comorbidity through neurogenesis enhancement.²⁰⁴ Some studies have claimed the opposite roles of TET1 in the same therapy.^{198,204} More strangely, the contradictory data are based on investigations into the similar pain types and model generation methods, reflecting the complexity of epigenetic modification in pain sensation. Some factors not easily perceived, such as pain inducer doses, disease courses and

experimental environments, may affect DNA methylation and require more attention in subsequent research.

The *Oprm1* gene encodes µ-opioid receptor and its hypermethylation positively correlates with pain severity and opioid tolerance. Long-term exposure to opioids further enhances *Oprm1* methylation levels.^{205,206} These vicious cycles via epigenomics are critical mechanisms underlying the opioid tolerance development. Moreover, molecules with properties of neuropathic and nociplastic pain modulation, such as stress-related protein FKBP5, peptide hormone leptin, CDK5 regulatory subunit-associated protein CDK5RAP1 are under strict control of DNA methylation.²⁰⁷⁻²⁰⁹

Histone is a key component of chromatin, with five types of core histones, including H1, H2A, H2B, H3, and H4. Histone acetylation, primarily occurring at lysine sites of H3 and H4, is an essential mechanism controlling histone activity. Unlike DNA methylation, acute pain has no evident impact on histone acetylation, which only responds to pain chronicity.⁴ Global alterations in histone acetylation are identified in both CNS and PNS.^{39,210,211} During nociceptive and neuropathic pain development, H3 and H4 acetylation is upregulated in DRG and spinal dorsal horn.²¹² Key brain regions, such as the CeA, PFC and hippocampus, exhibit significant changes in histone acetylation, which are involved in visceral hypersensitivity, neuropathic pain sensation and its comorbidities.^{213–215} In the descending pain modulation pathways, persistent enhancement of H3 acetylation occurs in the RVM, while this molecular event is short-lived, fading after long-term stress in the locus coeruleus.^{211,216} These findings suggest distinct regulatory effects of histone acetylation in different brain regions. Inflammatory mediators like IL-6 and TNF-α promote hyperacetylation of H3 and H4, enhancing neuron excitability in neuropathic models.215,21

The dynamic balance of histone acetylation is maintained by histone acetyltransferases (HATs) and histone deacetylases (HDACs). HDACs have 18 kinds of members, some of which are closely associated with pain perception. The reductions in HDAC1 and HDAC2 expression lead to the abnormal synaptic transmission, followed by somatic and visceral hypersensitivity.75,213,218 However, nuclear recruitment of HDAC2 driven by transcription factor Sp1 conversely aggravates neuronal dysregulation and microglial inflammation,²¹⁹ suggesting that the cellular distribution of epigenetic regulators is another factor in pain modulation. Existing negative results concerning HDAC3 indicate its weak associations with pain modulation.^{75,213} Inhibition of HDAC4 translocation into the cytoplasm epigenetically decreases HMGB1 expression and functions as an analgesic approach for neuropathic pain.²²⁰ Accumulation of HDAC5 in the nucleus inhibits H3 acetylation of Gad1 and Gad2 promoters, impairing GABAergic neuron activity and contributing to aberrant activation of astrocytes through direct interaction with STAT3. These mechanisms can lead to the development of peripheral neuropathic pain.^{221,222} The analgesic properties of SIRT1 and SIRT3, class III of HDACs, have also been revealed. Restoring their expression downregulation in nervous lesions mitigates ectopic discharge ²²⁴ alleviating of sensory neurons and excessive oxidative stress,²² emotional vulnerability of neuropathic pain.214

p300 is a representative molecule for pain modulation among HATs. Neuropathy following chemotherapy, stress and diabetes results in the upregulation of p300 expression or enhancement in p300 activity. It epigenetically modifies the hypothalamic–pituitary–adrenal (HPA) axis and promotes responses to norepinephrine.^{225–227} p300 is also involved in inflammatory pain through activating macrophages and elevating expression of TNF- α , IL-1 β , CCL2, and CXCL10.²²⁸ Regretfully, other HATs' roles in modulating pain are rarely investigated. Future research should pay attention to this shortcoming.

EZH2 is a histone methyltransferase catalyzing histone H3 methylation on K27 site.²²⁹ In the rodent models suffering from

nerve injury and cancer pain, the expression of EZH2 can be significantly upregulated in the CNS. The microglia are subsequently activated, accompanied with the abrupt release of proinflammatory factors. These mechanisms contribute to the development of mechanical and thermal hyperalgesia. Downregulation of EZH2 expression or topical injection of EZH2 inhibitors have been found to alleviate neuropathic and cancer pain.^{230–232} Although several investigations have verified the paininduced role of EZH2, its regulatory network of molecular mechanisms is still largely unclear. One study shows that mTOR signaling pathway-mediated autophagy may be a functional target of EZH2.²³³ The expression and activity of EZH2 are also under rigorous control of non-coding RNAs, including Incenc1, miR-124-3p, and miR-378.^{234–236} Moreover, EZH2 has been selected as a biomarker of evaluating efficacy of analgesic methods for neuropathic pain.²¹

Non-coding RNAs are multiple kinds of RNAs mostly incapable of encoding proteins, but their functions are not secondary to proteins. MicroRNAs (miRNAs), long non-coding RNAs (IncRNAs) and circular RNAs (circRNAs) are key molecules. Their remarkable mechanisms in pain modulation have been demonstrated by numerous studies.

The primary function of miRNAs is binding to the 3' untranslated regions of mRNA, blocking mRNA translation and promoting mRNA degradation. Patients with pain experience have evidently altered miRNA profiles, consequently dysregulating the expression of downstream targets.²³⁸ A large number of target genes have been identified, including but not limited to ion channels, inflammatory mediators, signaling molecules and transcription factors.^{239–243} Importantly, the miRNA regulatory network on pain is intricate, although most studies focused on their one-to-one relationships with target genes. One miRNA can regulate many downstream mRNAs. miR-183 cluster controls expression of over 80% of recognized pain-regulated genes.²⁴⁴ One target gene is likewise under regulation by multiple miRNAs, such as TRPV1.^{245,246}

LncRNAs and circRNAs can suppress miRNAs through complementary base pairing, described as miRNA sponges. This mechanism is the main research direction of current studies on pain modulation.^{247,248} Additionally, some lncRNAs and circRNAs interact with transcription factors, changing their activity and nuclear localization. These molecular events lead to alterations in neuropathic pain-related gene expression, such as KCNN1, G9A, and VEGFB.^{249–251} Certain lncRNAs and circRNAs have capabilities in regulating parental gene expression and modulating neuropathic pain, like lncRNA NEAT1 and circVOPP1.^{252,253}

Non-coding RNAs as exosomal cargos play important roles in intercellular communications. Hyperactivated neurons release exosomes loaded with non-coding RNAs with immunomodulation properties, like miR-21-5p. The exosomes are phagocytosed by macrophages, initiating the proinflammatory phenotype.²⁵⁴ This evidence demonstrates that sensory neurons are not only victims, but also accomplices in the progression of hyperalgesia triggered by neuroinflammation. The interactions among astrocytes, microglia and macrophages via exosomal non-coding RNAs exquisitely regulate inflammatory pain degrees.^{255,256} Altogether, the ectopic levels of non-coding RNAs have great potential in pain evaluation.²⁵⁷ Correction of abnormal non-coding RNA networks using gene editing and chemical treatment has achieved favorable outcomes for nociceptive, nociplastic and neuropathic pain as shown in preclinical research.²⁵⁸⁻²⁶⁰ Future clinical trials are eagerly required to promote the translational application of noncoding RNAs.

Notably, there exists crosstalk between the above three aspects of epigenetic modification. For example, DNA methylation and histone acetylation jointly regulate the expression of neuropathic pain-related genes.^{225,261} miRNAs directly suppress the expression of key enzymes of the other epigenetic aspects.²³⁹ Noncoding

RNA expression is under control of DNA methylation and histone acetylation.^{225,243,262} Overall, investigations into epigenetic modification have unveiled a new landscape of mechanisms underlying pain modulation. The achievements may lay the foundations for progress in pain management.

Posttranslational modification (PTM)

The activity, structure, cellular localization and interactions of proteins are critically regulated by PTMs. PTMs refer to the chemical modifications involving the addition or removal of specific groups in amino acid residues. To date, more than 600 kinds of PTMs have been identified. Common PTMs include phosphorylation, ubiquitination, glycosylation, methylation, etc. Histone acetylation, as mentioned in the previous section of epigenetic modification, also belong to PTMs. Novel PTMs, such as crotonylation, succinylation and lactylation, are continuously being discovered with advancements in biotechnology.²⁶³ The uncovered mechanisms concerning PTMs in pain modulation are concentrated on several PTMs.

The associations between phosphorylation and pain have received the most attention among PTMs. Fyn, a member of the Src family protein kinases, phosphorylates downstream targets. Its regulatory functions on pain perception have been extensively revealed. In responses to nerve injury and inflammation, IL-33 and BDNF enhance phosphorylation and catalytic action of Fyn in a PKA-dependent manner.²⁶⁴ GluN2B, a subunit of NMDAR, at Tyr1472 is phosphorylated by Fyn. This molecular event inhibits GluN2B endocytosis, increasing its membrane densities and synaptic currents mediated by NMDAR.264,265 The molecular functions of SHP-1 are opposite to Fyn, mediating target protein dephosphorylation. The DRG produces PD-L1 in response to acute and chronic pain. It further phosphorylates SHP-1, downregulating expression and phosphorylation of TRPV1.^{266,267} The attenuation of sodium channels and potentiation of potassium channels mediated by SHP-1 are observed, inhibiting the excitability of sensory neurons^{267,268} However, this mechanism can also be exploited by melanoma cells secreting PD-L1, leading to pain relief in the early stages of cancer development.²⁶⁷ The inconspicuous symptoms may contribute to delayed treatment and poor prognosis in cancer patients.

From the perspective of pain-related molecules regulated by phosphorylation, ion channels and hyperalgesia-related signaling pathways are the main targets. Nav1.7 phosphorylation by PKC and Fyn,^{269,270} TRPA1 phosphorylation by CDK5²⁷¹ and constitu-tive phosphorylation of TRPM8²⁷² are the key regulators of pain perception. Most signaling pathways rely on phosphorylation regulation. The activities of these pathways sustain functions of neurons and non-neuronal cells. For example, the NF-KB signaling pathway, closely associated with inflammation, is activated via p65 subunit phosphorylation in neurons, astrocytes and microglia, facilitating neuropathic and endometriosis-associated pain progression.^{273,274} The MAPK signaling mediates signal transduction from the membrane to the nucleus. p38 MAPK phosphorylation promotes the development of neuropathic and postoperative pain sensation through affecting downstream molecules and crosstalk with other signaling pathways.^{275,276} MAPK phosphorylation in immune cells is crucial for regulating the release of proinflammatory factors.²⁷⁷ Moreover, STAT3, Wnt, AMPK, mTOR, JNK signaling pathways involved in pain modulation are also under control of phosphorylation.278,22

Ubiquitination is another modification associated with pain modulation. The classical mechanism of ubiquitination involves the degradation of endogenous proteins catalyzed by the concerted action of ubiquitin-activating enzyme (E1), ubiquitinconjugating enzymes (E2) and ubiquitin-protein ligases (E3). NEDD4, an E3 ubiquitin ligase, promotes the degradation of multiple target proteins enhancing neuropathic pain hypersensitivity, including Nav1.7, TRPA1, and NMDAR subunit GluN2D.^{280–282} Nerve injury-induced NEDD4 downregulation is an important molecular event in pain development, regulated by histone acetylation.²⁸¹ Restoration of NEDD4 dysregulation effectively mitigates allodynia phenotypes.²⁸³ Cbl-b, another E3 ubiquitin ligase, processes nociceptive information in the spinal cord. Noxious stimuli induce Cbl-b dephosphorylation.²⁸⁴ The ubiquitination levels of GluN2B and TrkA are subsequently decreased, leading to inflammatory pain hypersensitivity.^{284,285} However, some studies have reported the neuropathic paininducing functions of Cbl-b based on its direct ubiquitination effects on K⁺-Cl⁻ co-transporter 2 and IL-2.^{286,287} The paradoxical conclusions suggest that roles of ubiquitination enzymes may depend on pain types and progression stages.

VGCC activity is susceptible to ubiquitination. TRPV1, IL-1 β and HMGB1 activate functions of the deubiquitinating enzyme USP5.^{288–290} USP5 directly interacts with Cav3.2 and potentiates whole-cell currents, promoting neuropathic and inflammatory pain development.²⁹¹ Many follow-up studies have verified the functions of the USP5/Cav3.2 axis and explored therapeutic approaches targeting this ubiquitinating event.^{292,293} For Cav2.2, its elements in the proximal C terminus are liable to ubiquitination,²⁹⁴ which has been found to contribute to neuropathic allodynia.²⁹⁵ The roles of other ubiquitination enzymes, like HUWE1 and UBR5, in nociceptive and neuropathic pain modulation have been partially revealed.^{296,297} In the present context, all the ubiquitination enzymes linked to pain modulation pertain to E3 ubiquitin ligases.

Small ubiquitin-related modifier (SUMOylation) has similar modification processes with ubiquitination, but its mechanisms and functions have considerable differences. SUMOvlation relies on covalent conjugation of SUMO with target proteins and regulates their localization, interaction and stability.²⁹⁸ Despite its late discovery of SUMOylation, its associations with hyperalgesia have garnered much attention. Dynamic alterations in SUMOylation can be detected in response to neuronal activity within a few minutes, reconfiguring ionic current densities.299 The robust impact of SUMOylation is ascribed to wide-range regulation of ion channels. CRMP2, coordinated by neuronal development and synaptic plasticity, is SUMOylated with SUMOylation E2 enzyme Ubc9, facilitating membrane localization and current density of Nav1.7.300,301 CRMP2 SUMOylation is a potential biomarker of persistent neuropathic pain.³⁰² CRMP2 SUMOylation is also regulated by multiple PTMs. SUMO-specific protease SENP1 induces CRMP2 deSUMOylation, eliciting antinociceptive effects.³⁰³ CRMP2 phosphorylation by CDK5 enhances its SUMOylation, whereas Fyn-induced phosphorylation downregulates SUMOylation.³⁰⁴ Antagonists targeting CRMP2/Ubc9 axis have shown desirable performances in trigeminal neuropathic pain relief.³⁰⁵ Importantly, CRMP2 SUMOylation occurs in chronic neuropathic pain, rather than physiological pain.³⁰¹ Therapies against this mechanism may have fewer side effects of losing sensations of normal acute pain. Besides CRMP2, other mechanisms by which SUMOylation affects ion channels have been identified, including direct promotion of Kir7.1 and HCN, suppression of USP5/Cav3.2 axis and enhancement in PKCɛ functions to phosphorylate TRPV1.³⁰⁶

Other PTMs likewise participate in pain development. Abnormal glycosylation is a crucial factor in diabetic complications due to dysglycemia. N-glycosylation modification enhances pore opening, channel permeability and membrane expression of ion channels like Cav3.2 and TRPV1.^{310–312} Nitrosylation, carbonylation, palmitoylation and succinylation have also been proved to modulate nociceptive and neuropathic pain perception, as well as opioid tolerance.^{313–316}

In summary, PTMs constitute regulatory networks of pain modulation, exerting milder and reversible effects on protein functions compared to other mechanisms. Furthermore, PTMs can respond more quickly to noxious stimuli probably because PTMs

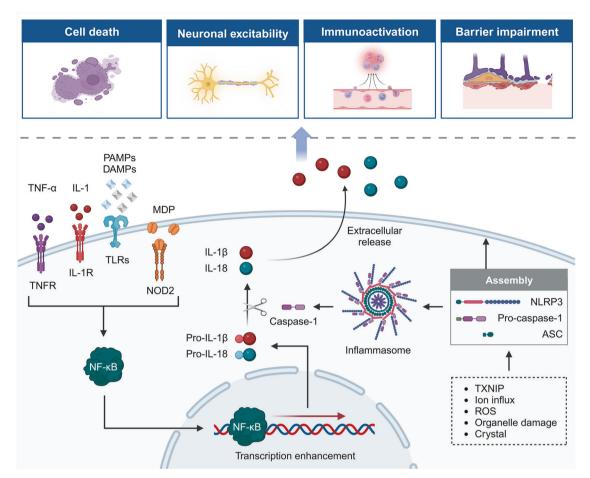


Fig. 6 The mechanisms underlying NLRP3 inflammasome-medicated hyperalgesia. Certain ligands, including TNF-α, IL-1, PAMPs, DAMPs and MDP, bind to the corresponding receptors, activating NF-kB signaling. The activated NF-kB is transported into the nucleus and promote the expression of pro-IL-1β and pro-IL-18. Various molecular and cellular events regulate the assembly of inflammasomes. Caspase-1, derived from inflammasomes, facilitates the maturation of IL-1 β and IL-18. These cytokines are subsequently released, leading to cell death, neuronal hyperexcitability, immune activation and impairment of brain barriers. These processes collectively contribute to hyperalgesia. ASC apoptosisassociated speck-like protein containing a CARD, DAMP damage-associated molecular pattern, IL-1 interleukin-1, IL-1R interleukin-1 receptor, IL-18 interleukin-18, MDP muramyl dipeptide, NF-xB nuclear factor-kappa B, NOD2 nucleotide-binding oligomerization domain 2, PAMP pathogen associated molecular pattern, ROS reactive oxygen species, TLR Toll-like receptor, TNF-α tumor necrosis factor-α, TNFR tumor necrosis factor receptor, TXNIP thioredoxin-interacting protein

function on the basis of mature proteins and do not require their de novo synthesis. These features underscore the prominence of PTMs in pain evaluation and treatment. Although our current understanding represents just the tip of the iceberg, the immense research and clinical value of PTMs has already emerged.

Inflammasome

Inflammasomes are the complexes composed of NOD-like receptors, apoptosis-associated speck-like protein containing CARD and pro-Caspase-1. According to assembled components, they are mainly classified as NLRP1, NLRP3, NLRC4, IPAF, and AIM2. Inflammasomes are recognized as multiprotein signaling platforms orchestrating inflammatory responses and host defense against microbial invasion. Inflammasomes were initially recognized as mediators of danger signals and pathogens.³¹⁷ As the research moves along, their causalities with pain perception, especially inflammatory pain, have been increasingly revealed (Fig. 6).

Nerve injury induces inflammasome activation within several days.⁴⁰ There are significant sex differences in inflammasome activation patterns. In males, high concentration of NLRP1 and NLRP3 inflammasomes can be detected. NLRP3 inflammasome regulates pain perception in both sensory neurons and nonneuronal cells. By contrast, NLRP3 and AIM2 inflammasomes are

more abundant in females, with pain signal modulation controlled only by sensory neurons, independent of non-neuronal cells.^{318,31} Activation of inflammasomes promotes the conversion from pro-Caspase-1 to its active form, Caspase-1, which cleaves precursors of IL-1β and IL-18 to produce mature inflammatory factors.³²⁰ They induce a series of pathological processes associated with pain, including cell death, neuronal excitability, immune overactivation. The brain barrier is subsequently impaired, further aggravating central neuroinflammation.

Most basic studies focus on the upstream regulatory pathways of NLRP3 inflammasome. TXNIP, a critical protein for regulating oxidative stress and inflammatory responses, interacts with NLRP3 inflammasome to promote inflammation activation. TXNIP functions on NLRP3 inflammasome are under strict control of epigenetic modification. For instance, HDAC2 promotes deacetylation of histone H4, decreasing miR-183 expression and antagonizing its inhibitory effects on TXNIP.³²¹ TET2 upregulates TXNIP expression at the transcriptional level and facilitates progression of diabetic neuropathic pain.³²² Activation of TRP family members, TRPV1, TRPA1 and TRPV4, enhances NLRP3 inflammasome activity.^{47,323,324} Moreover, pathological events, like organelle damage and exposure to crystal, also promote NLRP3 inflammasome activity.

The wealth of therapeutic approaches for pain relief are linked to the inhibition of inflammasomes. Inflammasome-related markers are widely employed to reflect the therapeutic efficacy.^{325,326} While research in this field is flourishing, potential side effects of therapies targeting inflammasome activity should be emphasized. The inflammasomes have the important physiological functions. At the early stage of nerve injury, Schwann cells and macrophages rapidly activate inflammasomes, facilitating macrophage phagocytosis of axonal debris. The regulatory factors derived from macrophages can drive Schwann cells to repair damaged nerves.^{327,328} The inflammasome product IL-1 β is also critical for the host defense against bacterial invasion.³²⁹ Therefore, impertinent suppression of inflammasome activity for pain relief may lead to delayed nerve regeneration and immunosuppression, necessitating careful validation and resolution of related adverse events.

Signaling pathways

Signaling pathways refer to a series of biochemical reactions within cells triggered by specific signals. They collectively regulate nearly all physiological and pathological processes in all aspects of gene expression and functions. The mechanistic associations between pain perception and signaling network abnormality have been gradually uncovered. A large number of preclinical and clinical studies have further demonstrated the potential value of restoration of signaling pathway dysfunctions in alleviating pain. This section will summarize several classical signaling pathways associated with pain aiming to present more integral landscape of molecular mechanisms underlying pain development.

Wnt/β-catenin signaling is one of the most critical pathways, responsible for morphogenesis, carcinogenesis and other physiological processes. Activation of Wnt blocks the formation of β-catenin destruction complex and promotes its transportation to the nucleus, regulating the expression of specific genes. In the animal models with neuropathic pain, Wnt3a, Wnt5a and Wnt10a are significantly upregulated, accompanied with increased nuclear translocation of β-catenin.^{330,331} Some molecular regulators, such as SFRP1, LPAR3, miR-26a-5p and LncCRNDE, jointly control Wnt/β-catenin signaling-mediated neuropathic pain.^{331–334} A recent study demonstrated that GPR177 promotes the secretion of Wnt5a from A-fiber DRG neurons, which further enhances rapid currents of TRPV1+ nociceptive DRG neurons.³³⁵ This finding has strongly revealed the functions of intercellular communications via Wnt/β-catenin signaling during neuropathic pain progression.

The downstream mechanisms by which Wnt/β-catenin signaling promotes pain hypersensitivity are complicated. Activation of Wnt facilitates trafficking of classical receptors P2X3 and NR2B to the membrane in Ca2+-dependent manners in primary sensory neurons, leading to peripheral sensitization and bone cancer pain progression.^{336,337} VGLUT2 is a critical component of glutamate neurotransmitter system, associated with various kinds of pain. Wnt1 signaling can upregulate VGLUT2 expression and potentiate neuropathic pain development.³³⁸ Furthermore, Wnt/β-catenin signaling promotes the biosynthesis and release of substance P, TNF- α and IL-18.^{339,340} For the research on pain management, some natural compounds with analgesic properties, like isoquer-citrin, persicae semen and resveratrol.^{341–343} Inhibition of Wnt/ β -catenin signaling has also shown the potential in treating nociceptive and nociplastic pain.^{344,345} Amazingly, Wnt pathway inhibitors have been applied to treat osteoarthritis-related pain by clinical trials and showed promising performance.³⁴

MAPK signaling is another important pathway within eukaryotic signaling networks. There are many branches of MAPK signaling, including ERK, JNK, p38 MAPK and ERK5 pathways. They all participate in the regulation of growth, differentiation, stress and inflammation though with differences in concrete functions. MAPK signaling has been proved as a pivot of $TNF-\alpha$ -mediated pain hypersensitivity. During the progression of acute nociceptive pain,

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TNF-α activates p38 MAPK and increases the ser110 site of Nav1.7. Hyperphosphorylation of Nav1.7 contributes to its insertion into the somatic membrane of neurons.³⁴⁷ The expression of Nav1.8 and Nav1.9 is meanwhile upregulated.³⁴⁸ MAPK-ERK-CREB signaling has also been found to increase Nav1.6 expression in the oxaliplatin-induced neuropathic pain model.³⁴⁹ Therefore, MAPK signaling has a broad promotion effect on VGSCs, facilitate the formation of pain hypersensitivity. In addition to TNF-α, MAPK signaling is responsible for the pain-inducing effects of other secretory factors. For instance, inhibition of macrophage- and Schwann cell-derived nerve growth factor mitigates neuropathic pain through inhibiting TAK1-MAPK pathway in the CCI model.³⁵⁰ It also activates p38 MAPK signaling and upregulates ASIC3 expression in DRG cells, facilitating mechanical allodynia development.³⁵¹ Substance P released from trigeminal ganglion neurons activates satellite glial cells through ERK1/2 and p38 pathways. The hyperactivated glial cells produce more IL-1ß and 52 As TNF- α and play an accessory role in inflammatory pain.³ stepping into the stage of pain hypersensitivity maintenance, JNK signaling gradually play a leading role in the astrocytes in response to TNF- α .²⁷⁷ However, one study showed that glial glutamate transporter can be upregulated by p38 MAPK. This mechanism prevents long-lasting ongoing spontaneous pain. These results suggest that the branches of MAPK signaling may have opposite effects.³⁵

There have been various regimens concerning MAPK signaling interference in preclinical research. Antisense oligonucleotides targeting p38 have been synthesized. They effectively inhibit microglia and astrocyte activation through suppressing MAPK signaling, thereby functioning as an analgesic method for inflammatory and neuropathic pain relief.^{354,355} Some clinically applied drugs, such as tetrahydropalmatine, lidocaine and opioids, have been demonstrated to achieve analgesic effects for nociceptive and neuropathic pain, at least partially, through blocking MAPK signaling.^{356–358} A large number of natural compounds may serve as MAPK signaling inhibitors for treating neuropathic pain.³⁵⁹

PI3K/Akt/mTOR signaling functions as a regulator of cell survival, proliferation, angiogenesis, metabolism, autophagy, etc. Different with previous introduced pathways, PI3K/Akt/mTOR signaling seems to play a two-sided role in pain development and management. On the one hand, overactivation of PI3K/Akt/ mTOR signaling has been detected in models with nociceptive, neuropathic pain and opioid tolerance.^{360,361} Mechanistically, Akt phosphorylates ASIC1a at the Ser25 site, which promotes its forward trafficking and membrane expression.³⁶² mTOR activation has been proved to facilitate reconstruction of nociceptive terminals following inflammation, diminishment of ACC synaptic protein involved in neuropathic pain perception. These mechanisms collectively underpin the pain hypersensitivity progression.^{360,363} Furthermore, NALP1 inflammasome activation can be elicited by PI3K/Akt signaling, accelerating the formation of opioid tolerance.³⁶⁴ Topical injection or systemic administration of PI3K/ Akt/mTOR signaling inhibitors have shown good performance in reversing hyperalgesia and opioid tolerance.³

On the other hand, however, activation of this signaling may exert analgesic effects. Neurotrophic factor derived from bone marrow mesenchymal stem cells can enhance PI3K/Akt signaling activity, transforming destructive M1 phenotype into regenerative M2 phenotype of microglia. The autophagy is meanwhile enhanced.³⁶⁷ These mechanisms restore the abnormal discharging C-fiber neurons.³⁶⁸ During the progression of chronic postoperative pain, microglia can downregulate activity of PI3K/Akt signaling in astrocytes, which induces astrocyte transformation into A1 phenotype and promotes the chronicity of pain.³⁶⁹ Additionally, PI3K/Akt signaling has been proved to participate in nerve regeneration and alleviate neuropathic pain.³⁷⁰ The above evidence has suggested that the functions of PI3K/Akt/mTOR

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signaling may depend on cell types, pain types and development stages. Crude inhibition or activation of this signaling may be ineffective and bring about potential side effects. It is eagerly required to explore more precise targeted therapies for PI3K/Akt/ mTOR signaling interference.

AMPK is a hub regulator of biological energy metabolism. The dysregulation of AMPK signaling contributes to various metabolism-related diseases. AMPK signaling-mediated metabolic disorders have been proved as an important factor of hyperalgesia. For instance, HSP22, a kind of heat shock proteins, is downregulated in the spinal cord neurons of models with nerve injury. Restoration of HSP22 expression improves mitochondrial biogenesis and reduces oxidative stress through activating AMPK/ PGC-1 α pathway, attenuating neuropathic pain.³⁷¹ Meanwhile, this mechanism underpins osteoarthritis pain relief caused by Sestrin2 overexpression.³⁷² AMPK signaling serves as a sensor of intracellular glucose concentrations. AMPK signaling hyperactivity can rapidly reduce TRPA1 membrane expression and its channel activity. High-glucose exposure significantly inhibits AMPK signaling in DRG neurons and potentiates TRPA1-mediated hyperalgesia, which is a critical mechanism underlying painful diabetic neuropathy.³⁷³ Suppressing NLRP3 inflammasome is another identified mechanism of AMPK-induced analgesia.³⁷²

AMPK signaling exerts huge impact on non-neuronal cells. Its activation promotes M2-type polarization of microglia and reduced the release of proinflammatory factors.³⁷⁵ AMPK signaling has also been found to participate in the functions of endocannabinoid-induced analgesia through reprogramming of the phosphoproteome and bioenergetics of macrophages.³⁷⁶ The autophagic flux of Schwann cells is also enhanced by AMPK hyperphosphorylation, attenuating peripheral neuropathic pain.³⁷⁷ Taken together, existing studies have shown that AMPK signaling may play a protective role in pain perception. Some drugs and therapeutic regimens as AMPK agonists, such as metformin and caloric restriction diet, have been proved to effectively contribute to pain relief.

In addition to the abovementioned classical signaling pathways, NF- κ B, JAK/STAT, TGF- β , Notch, Hippo/YAP1, Hedgehog, cGAS-STING signaling have been proved to be associated with pain modulation. Notably, these signaling pathways have compact crosstalk. One molecular target or pain phenotype may be under control of various pathways. The interlaced signaling network underpins hyperalgesia and pain relief. Likewise, interfering with pivots within signaling pathways may bring about unexpectable side effects, which may contribute to slow paces of clinical translation of agonists or inhibitors targeting signaling pathways. Further studies should strictly inspect their effectiveness and safety.

Microbiota

The human microbiome, comprising $\sim 10^{14}$ microbes, is a symbiotic superorganism.³⁷⁸ Microbiota reside in the gastrointestinal tract, skin, respiratory tract and reproductive system. The interactions between eukaryotic systems and microorganisms play a significant role in physiological and pathological modulation.³⁷⁹ The mechanisms underlying pain perception, particularly gastrointestinal dysfunctions, regulated by microbiota are gradually being uncovered, which can be generally divided into indirect and direct pathways.

For the indirect pathways, microbiota dysbiosis can be recognized by immune systems. Pathogen-associated molecular patterns (PAMPs) refer to the highly conserved structural components of microbiota that are recognized by host cells. PAMPs essentially constitute the molecular immunogenic signatures of pathogens, which include lipopolysaccharide (LPS), peptidoglycan, teichoic acid, and mannose. The recognition of PAMPs contributes to the activation of immune cells, producing proinflammatory cytokines, chemokines and neuropeptides. Their topical and plasma levels are sharply increased, and the crosstalk between immune cells promotes nociceptive pain progression.^{277,380–382} Particularly, the abrupt release of TNF- α caused by microbiota dysbiosis exacerbates nociceptive pain hypersensitivity in the PNS.^{277,383}

The direct pathways are more complicated, which are schematically illustrated in Fig. 7. Microbiota-derived compounds are critically involved in the direct pathways. Histamine pertains to one type of amine compound originating from histidine decarboxylation. It plays important roles in perceiving nociceptive information. Gram-positive bacteria utilize pyruvoyl-dependent histidine decarboxylase and gram-negative bacteria employ pyridoxal phosphate-dependent histidine decarboxylase to produce histamine. Gut histamine concentration is correlated with the degree of abdominal pain.^{384,385} A recent study showed that *Klebsiella aerogenes* is a main producer of histamine, and its ectopic histamine metabolism is the culprit of IBSinduced abdominal pain.³⁸⁶ Activation of the H1 receptor induces excitation of TRPV1 signaling transduction and sensitization of nociceptive nerves.^{387,388}

5-HT is an important metabolite product of gut microbiota, such as *Corynebacterium*, *Streptococcus* and *Enterococcus*.³⁸⁹ Plasma 5-HT is significantly increased during the progression of IBS.⁶⁵ A combined network analysis based on IBS patients indicated that visceral hypersensitivity is associated with gut microbiota related to the local 5-HT system.³⁹⁰ Long-term exposure to 5-HT enhances its interactions of 5-HT3 and 5-HT4 receptors with cannabinoid signaling, deteriorating the progression of visceral hypersensitivity.³⁹¹ Gut homeostasis modifiers alleviate abdominal pain by restoring normal levels of 5-HT.³⁹²

Moreover, classical excitatory neurotransmitters glutamate and aspartic acid, together with inhibitory neurotransmitter GABA, are also important products of microbiota and participate in pain modulation. Glutamate and aspartic acid are synthesized by Bacillus bacteria in the guts. The absorbable products facilitate excitability of sensory neurons.^{393,394} Many bacteria produce GABA, including *Lactobacillus, Bifidobacterium* and *Bacteroides*.^{395,396} Studies have proved that supplementation with specific gut probiotics effectively increases GABA levels and restores visceral hypersensitivity.³⁹⁷ Of note, fungi are also able to produce GABA, ³⁹⁸ implying the importance of nonbacterial species in pain modulation.

Direct interactions of microbiota with neurons are another direct mechanism of pain induction, independent of immune activation or secreted metabolite regulation. The antigenicity of microbiota constitutive elements endows sensory neurons with cognition capability. Toll-like receptors (TLRs) act as detectors of microbiota invasion, initiating innate immune responses by recognizing PAMPs and activating intracellular signaling pathways. There are 13 functional categories of TLRs, and they monitor different types of PAMPs derived from microbiota.

TLR4 is a critical receptor mediating chronic pain development. Its expression can be detected in the afferent fibers of trigeminal nerves and DRG. Peripheral nerve injury and inflammation trigger significant increases in TLR4 expression in the spinal cord.³⁹⁹ LPS is a classical activator of TLR4. TLR4 activated by LPS increases the firing frequencies of TRPV1⁺ neurons, leading to nociceptive, nociplastic and neuropathic pain under bacterial dysbio-00,401 sis 380,4 Gut microbiota may interplay with TLR4 receptors and induce oxaliplatin-induced mechanical hyperalgesia.⁴ During the progression of IBS, LPS regulates the TLR4-MyD88 receptor signaling pathway to inhibit the synthesis of resolvin D1 in colonic tuft cells, inducing visceral pain hypersensitivity and colonic inflammation.⁴⁰⁴ Apart from peripheral TLR4, upregulation of TLR4 expression in the PFC and hippocampus is observed during abdominal pain progression, providing strong evidence for the crosstalk of the gut-brain axis.

Bacterial flagellin is a typical activator of TLR5. A positive association between TLR5 expression and the abundance of flagellated bacteria has been observed in rats with chronic

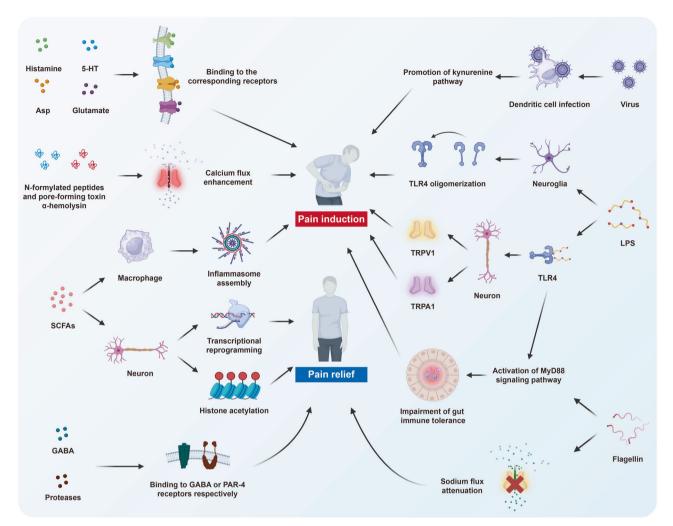


Fig. 7 The direct mechanisms underlying regulation of pain perception by microbiota. The microbiota-derived metabolites, including histamine, 5-HT, Asp and glutamate, bind to their corresponding receptors to enhance pain sensation. In contrast, GABA and proteases can respectively bind to GABA and PAR-4 receptors to alleviate pain. N-formylated peptides and the pore-forming toxin α -hemolysin produced by microbiota, enhance the calcium flux in nociceptors and induce pain perception. SCFAs exhibit dual effects. They interact with macrophages, inducing inflammasome assembly and pain perception. On the other hand, SCFAs promote transcriptional reprogramming and histone acetylation in neurons, reducing pain hypersensitivity phenotypes. Viral infection of dendritic cells facilitates the kynurenine pathway, contributing to pain development. LPS possesses the typical properties of pain induction in a TLR4-dependent manner. TLR4 oligomerization in neuroglia, activation of TRPV1 and TRPA1 in neurons and immune tolerance impairment in intestinal epithelial cells through TLR4/ MyD88 signaling pathway are mechanisms underlying LPS-induced pain sensation. Another bacterial component, flagellin, likewise attenuates gut immune tolerance through regulating MyD88 signaling pathway. However, it can decrease sodium flux in nociceptors, inversely alleviating pain perception. 5-HT 5-hydroxytryptamine, Asp aspartic acid, SCFA short-chain fatty acid, γ -aminobutyric acid, PAR-4 receptor, LPS lipopolysaccharide, TLR4 toll-like receptor 4, TRPV1 transient receptor potential vanilloid 1, TRPA1 transient receptor potential ankyrin 1

abdominal pain.⁴⁰⁶ HMGB1 is released by macrophages or necrotic cells under infection or inflammation. HMGB1 acts as a ligand of TLR5 and activates the NF- κ B signaling pathway, leading to the development of allodynia.⁴⁰⁷ Nevertheless, one study revealed the analgesic effects of TLR5. Flagellin induces TLR5dependent blockade of sodium currents of A β fibers. The combined use of flagellin and the membrane-impermeable lidocaine derivative QX-314 effectively alleviates mechanical allodynia induced by chemotherapy, nerve injury and diabetic neuropathy.⁴⁰⁸

Several critical but sporadic research studies on direct interactions between microbiota and neurons provide new perspectives on microbiota regulation in pain. LPS can activate TRPV1, TRPM3, and TRPM8 channels located in the DRG.^{409,410} N-formylated peptides and the pore-forming toxin α -hemolysin produced by *Staphylococcus aureus* directly increase calcium flux and action

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potentials of nociceptors.²⁵ The excitability of DRG neurons is downregulated by gastrointestinal commensal bacteria, such as *Faecalibacterium prausnitzii*, through blockade of PAR-4.⁴¹¹ Furthermore, acute treatment with the lysate of *Escherichia coli*, rather than purified LPS, dramatically facilitates afferent discharge of colonic mesenteric nerves. Deletion of TLR signaling effects cannot rescue pain hypersensitivity,⁴¹² implying that there are undiscovered mechanisms and targets by which microbiota directly stimulate nociceptor hyperactivity.

Taken together, the microbiota plays an essential role in pain perception, especially for visceral hypersensitivity. Abnormal microbiota signatures associated with pain have been depicted. The mechanisms underlying bidirectional interactions between microbiota and the nervous system have been gradually revealed, which spawn many future directions for pain diagnosis and treatment. 17

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MECHANISMS OF PAIN SEXUAL DIMORPHISM

Pain seemingly exhibits a disparity between the sexes. Epidemiological data demonstrate a significant predisposition for pain in females.^{413,414} More basic experiments using female animals, particularly since the beginning of this decade, have advanced our understanding of the characteristics and mechanisms of sexual dimorphism in pain.⁴¹⁵ This phenomenon is typically characterized by four aspects: higher morbidity susceptibility, stronger pain sensation, more pronounced negative effects and poorer responses to analgesic drugs.^{416–418} Beyond the impact of congenital physiological factors, such as menstruation, the intrinsic mechanisms of pain modulation have been shown to contribute to sexual dimorphism. Significant sex differences in the structures and functions of the CNS and PNS during pain perception have been observed.^{419,420} High-throughput data further reveal global differences in gene polymorphism and expression.^{103,421,422} Some therapeutic regimens have been proposed to address inefficacy of analgesic drugs due to sexual dimorphism.⁴²³ We will give an overview of the mechanisms by which pain has typical sex preferences, illustrating the sophisticated modulation of pain (Fig. 8).

Hormone

Sex hormones, crucial determinants of sexual characteristics, affect pain sensitivity through their periodic fluctuation.⁴²⁴ Nerve injury, in turn, induces the upregulation of estrogen receptor expression in the DRG, microglia and spinal dorsal horn. 17 β -estradiol, a primary component of estrogen, binds to estrogen receptors,

regulating interactions between inositol 1,4,5-triphosphate and ryanodine receptors. It subsequently enhances calcium transients and neuronal excitability in response to noxious stimuli. Concurrently, the inflammasome activity and the release of proinflammatory factors are promoted, accompanied with a significant decrease in membrane expression of GABA receptors.^{425–427} Additionally, 17β-estradiol is involved in the functional modulation of specific brain regions, such as the hippocampus, parabrachial and amygdala.^{428,429} Other estrogen metabolites, including 2- and 4-HEMs and 2- or 4-OHE1, directly simulate TRPV1 and TRPA1, contributing to the dramatical reduction in uterine pain threshold.⁴³⁰ Conversely, testosterone has been proved to have protective effects against nociceptive pain.⁴³¹

Prolactin is responsible for mammary gland growth and the stimulation and maintenance of lactation. The prolactin receptor (PRLR) in sensory neurons is imperative for initiating pain sensation in females, but unnecessary in males.⁴²⁷ The PRLR protein is detectable only in sensory neurons of females, although its mRNA is found in both sexes.⁴³² The effects of PRLR on nociceptive pain modulation rely on mutual antagonism between its isoforms PRLR-L and PRLR-S. PRLR-S enhances neuronal excitability, while PRLR-L upregulation selectively rescues pain sensitivity.⁴³³ Activation of κ -opioid receptor signaling in the hypothalamus induces dysregulation of PRLR isoforms and prolactin production in sensory neurons, accompanied by the release of CGRP.⁴³⁴ The neuronal hyperreactivity to stimuli during migraine progression is subsequently enhanced.⁴³⁵

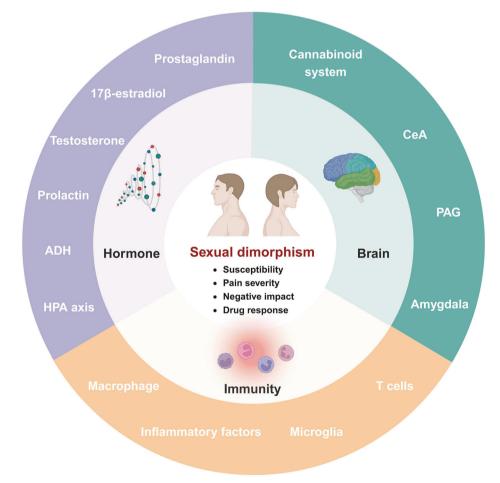


Fig. 8 The mechanisms underlying sexual dimorphism of pain. The sexual dimorphism in pain is characterized as four aspects: susceptibility, pain severity, negative impact of pain and responses to analgesic drug. These differences are attributed to variations in hormones, brain function, and immunity. The specific factors in each mechanism, as identified by existing studies, are displayed in the corresponding outer ring. ADH antidiuretic hormone, CeA central amygdala, HPA hypothalamic-pituitary-adrenal, PAG periaqueductal gray

Sex differences in the neuroendocrine stress axis are preliminarily revealed. Corticotropin-releasing factor expression in the CeA is elevated in response to stress-induced visceral pain.⁴³⁶ Knockdown of β 2-adrenergic receptors in nociceptors effectively alleviates hyperalgesia in females, but is inefficacious in males. Blockade of glucocorticoid receptors attenuates pain following paclitaxel-induced peripheral neuropathy only in males.⁴³⁷ These findings suggest that sexual dimorphism in pain may be associated with two independent neuroendocrine stress axes. Additionally, prostaglandin and antidiuretic hormone also have close associations with pain susceptivity in females.^{438,439} However, there is still a lack of mechanistic investigations into non-sex hormones.

Brain

The differences in brain region functions and circuits constitute another important factor in sexual dimorphism in pain. Interestingly, females exhibit more pronounced empathy for pain, which is associated with sex differences in brain region activities.⁴²⁰ Most studies focus on the amygdala and PAG. The amygdala, a hub for emotional processing, shows apparent differences in responses and projections between healthy men and women. During the progression of visceral pain, the amygdala in males is more connected with the anterior cingulate subregions and insula, while in females, it shows stronger functional connectivity with prefrontal modulatory regions.⁴⁴⁰ This feature may underlie sexual dimorphism in pain. One study using MRI data indicated that some amygdala subnuclei, including the lateral and basal nuclei in the left hemisphere, exhibit distinct neuronal excitability. These regions act as modulators of the interplay between negative emotions and pain, with changes in their volumes serving as complementary mediators in sexual dimorphism.⁴⁴¹

The ascending pain transduction tracts are more engaged in preventing pain habituation in males.⁴⁴² The regulatory signals from the CeA project to the ventrolateral PAG (vPAG). Inflammatory pain can induce overactivation of the CeA-vPAG pathway in females, resulting in more severe chronic pain sensations.445 In addition to differences in the upstream regulatory pathways, PAG functions also exhibit sexual dimorphism. Critical mediators in the endocannabinoid system, 2-arachidonoylglycerol (2-AG) and anandamide, are inhibited in the PAG of females,⁴⁴⁴ indicating the relative ineffectiveness of endocannabinoid-associated analgesia. Furthermore, fear-conditioned analgesia, which refers to pain relief caused by re-exposure to a condition previously associated with aversive stimuli, is coordinated by the PAG. Males exhibit more robust fear-conditioned analgesia, possibly due to the higher presence of GABA in the PAG of males.445 Moreover, congenital structural differences in brain regions, such as thicker posterior insula and precuneus cortices in female migraine patients, may have associations with sexual dimorphism.⁴⁴⁶ The aforementioned mechanisms collectively support the evidence that differences in brain regions contribute to sexual dimorphism in pain sensation.

Immunity

Males suffering from pain present more pronounced profiles of immune activation, despite females experiencing more intense pain. Among all the immune regulators, microglia have shown the most salient sex-dependent functions. Microglia in males are more sensitive to nociception, as evidenced by increased cell hyperactivity and the release of more pro-inflammatory factors.⁴⁴⁷ Overactivation of microglia leads to higher required doses of opioid analgesia in females.⁴⁴⁸ Peripheral macrophages can also exhibit distinct polarization and motility when stimulated by proinflammatory factors in both sexes.⁴⁴⁹ Several key factors have been identified as causes of sexual dimorphism in microglia activity, such as HMGB1, SETD7, and P2X4R. They all activate microglia by enhancing inflammation-associated signaling 19

pathways.^{447,450,451} More importantly, inhibiting microglia activation or antagonizing stimulating factors effectively attenuates hyperalgesia only in males, not in females.^{452,453} These results imply that male pain hypersensitivity is largely dependent on microglia, whereas the link with female hyperalgesia is less clear. In other words, the progress in pain management research through targeting immune responses in males is therefore more promising, particularly with the recent insights into microglia mechanisms. Additionally, dysregulation of proinflammatory factors, including IL-6, IL-23 and TNF- α , is an important reason of poor sensation of nociceptive, neuropathic and nociplastic pain, as well as the drug responses in females.^{452,454,455}

In summary, sex differences of hormones, brain regions and immune responses collectively mediate the sexual dimorphism in pain. Nevertheless, this is emphatically not the complete picture, and sexual dimorphism does not simply mean that females are all along at more disadvantaged positions. Women may gain greater adaptation and habituation to persistent heat pain.⁴ Voluntarv exercise, like running, is effective in restraining nociceptive stimuli in females by inhibiting inflammatory responses and DRG excitability. Conversely, it enhances the release of proinflammatory factors in males.^{457,458} Furthermore, cognitive disorders following chronic neuropathic pain occur only in males.⁴⁵⁹ Some studies have reported the negative results or even opposite conclusions regarding pain differences,460,461 highlighting the complexity of sexual dimorphism in pain modulation. Future research is eagerly required to delve into its deep-seated reasons.

MECHANISMS OF PAIN COMORBIDITIES

Pain is not an isolated event. The development of pain sensation is usually predicated on disease progression. Pain, in turn, is likely to cause comorbid physiological disorders, such as depression, anxiety, sleep disturbance and cognitive impairment. These comorbidities greatly reduce the quality of life and increase medical costs. A large number of neural circuits and molecular regulators have been found to promote pain comorbidities. This section will summarize the mechanistic achievements, categorized according to the types of comorbidities.

Depression

The prevalence of comorbid depression is alarmingly high, approximately ranging from 40% to 60%. There is a significant positive correlation between the severities of pain and depression.^{462,463} The ACC is critically involved in comorbid depression associated with chronic pain. It drives the enhancement in firing and bursting activity of the ACC.⁴⁶⁴ Microglia and glutamatergic neurons are concurrently activated.⁴⁶⁵ The synaptic structures undergo reorganization, facilitating pain-induced depression.^{92,4} The molecular mechanisms underlying these pathological changes are scattered so far. For instance, histone acetylation and phosphorylation of the transcription factor ATF/CREB contribute to the upregulation of MAPK phosphatase-1 expression in the ACC.⁴⁶⁷ Tiam1 coordinates cytoskeleton reorganization and membrane NMDAR stability.92 The imbalance of TREM-1 and TREM-2, two receptors expressed on microglia, mediates inflammation outbreaks in the ACC. These molecular events collectively underpin ACC hyperactivity and depressive-like behaviors.⁴

Certain neural circuits, especially those involving the amygdala, play a role in the formation of depression comorbidity. The ACCamygdala pathway is the core of depression comorbidity in pain. Activation of this pathway in rodents is sufficient to replicate brain transcriptome signatures of clinical patients. The activity of ACCamygdala pathway is dependent on signal factor Sema4A⁴⁶⁸ The nucleus of the solitary tract and the dorsal raphe nucleus are two brain regions regulating depression. They respond to pain signals and project to CeA.^{469,470} Subsequently, DNMT1 expression in GABAergic neurons is upregulated, partially due to the positive 20

functions of upstream IncRNA XR_351665.⁴⁷¹ The DNMT1mediated DNA methylation network suppresses GABAergic neuron activity, inducing comorbid depressive symptoms.⁴⁷² Additionally, some studies reveal the side effects of active protection from neuropathic pain. Neurons of the mPFC and locus coeruleus are activated at the acute phase, which partially block the outburst of pain sensation through enhancing inhibitory signals. It can be regarded as negative feedback for neuropathic pain relief. However, this restorative analgesia induces dysfunctions of the endocannabinoid system in the mPFC and adrenoreceptors in the ACC as pain chronicity develops, resulting in neuropathic pain-induced depression.^{473,474} These findings highlight the complexity of depression comorbidity. It should be noted that single therapy against depression comorbidity may have risks in aggravating hyperalgesia.

The kynurenine pathway accounts for approximately degradation of 95% dietary tryptophan. Tryptophan 2,3-dioxygenase in liver or indoleamine 2,3-dioxygenase (IDO) in other organs is the rate-limiting enzyme of the kynurenine pathway.475 Evidence has shown the critical roles of the kynurenine pathway in regulating visceral and inflammatory pain perception.476,477 The enhancement in the kynurenine pathway, characterized by an elevated kynurenine/tryptophan ratio, correlates positively with pain sensation.478 Kynurenine is then metabolized into a pronociceptive metabolite 3-hydroxykynurenine catalyzed by kynurenine 3-monooxygenase (KMO). After a series of enzymatic reactions in the kynurenine pathway, the final product, quinolinic acid, also participates in neuropathic pain modulation.⁴⁷⁹ For the depression comorbidity, pain induces upregulation of IDO expression both in the hippocampus and liver, accompanied by alterations in metabolites in the kynurenine pathway and increased expression of KMO. Ablation of IDO and KMO functions effectively mitigates depression-like behaviors following chronic pain, providing novel therapeutic targets for attenuating depression comorbidity.

Anxiety

The core feature of anxiety is excessive worry, manifested as concerns about possible future catastrophes or unhappy events that are disproportionate to reality, or not clearly understood in terms of their objects or contents. Pain dramatically increases incidence rates of anxiety comorbidity.480 Some neural circuits and related molecular mechanisms are critically implicated in the development of anxiety comorbidity. The paraventricular thalamic nucleus generates excitatory projections in response to chronic pain. This neuronal input triggers activation of specific neurons expressing nitric oxide synthase (NOS) in the ventromedial PFC. The enhanced synthesis and release of NO promote nitrosylation modification and trafficking.⁴⁸¹ This molecular event initiates network excitability and anxiety-like behaviors.482 The prelimbic cortex (PrL) is a hub of orchestrating nociception and emotion. TNF-a-induced projections from the PrL activate the basolateral amygdala and meanwhile suppress activity of the vPAG, respectively mediating anxiety and neuropathic pain.⁴⁸³ Although the vPAG is not involved in anxiety, the front section of PAG can project to the lateral habenula, leading to neuronal hyperactivity in the lateral habenula. This pathway promotes neuropathic pain and anxiety comorbidity.⁴⁸⁴ Moreover, GABAergic neurons in the lateral septum and CART-positive neurons in the bed nucleus of the stria terminalis are activated by chronic pain, projecting to the lateral hypothalamus. These two inhibitory inputs decrease the lateral hypothalamus activity and cause maladaptive anxiety in models with the CFA or SNI-induced model.⁴

Sleep disturbance

Pain-induced insomnia is essentially a defense mechanism under normal conditions. The prominent symptoms are difficulty in falling asleep and frequent arousals. It increases vigilance under potential threats or at vulnerability status for improving survival possibilities of individuals. However, the catastrophic amplification of this action brings significant health burdens for patients with pain. One large-population study has shown nine shared functional connectivity mediating the vicious interactions between sleep disturbance and pain. Notably, the causality from chronic pain to sleep disturbance comorbidity may be stronger than the effects of sleep loss on pain worsening.⁴⁸⁷

The NAc is an important region in regulating reward, addiction and disgust. A specific subset of neurons in the NAc is activated in response to noxious stimuli or upon awakening. This subset exerts inhibitory inputs divergently to the ventral tegmental and preoptic areas. Downregulation of ventral tegmental area activity causes neuropathic pain hypersensitivity, and the projection to the preoptic area leads to sleep disturbance.⁴⁸⁸ The dual functions of the NAc may provide a target for therapies against pain and sleep disturbance comorbidity. Despite the lack of mechanistic research on frequent arousals, an observational study indicated that the arousal infraslow can appear at the interval of 50 s during non-rapid eye movement sleep. Autonomic arousals and local somatosensory sensitivity are significantly enhanced.489 This phenomenon underpins the sleep disturbance induced by frequent arousals.

Furthermore, poor sleep, in turn, aggravates pain experience. Long-term sleep disturbance suppresses the functions of the endocannabinoid system in the thalamic reticular nucleus, which promotes projections to the ventroposterior region of the thalamus. This projection is critically involved in hyperalgesia.⁴⁹⁰ Nerve injury causes a significant increase in signal input from the parabrachial nucleus, facilitating the overactivity of cholinergic neurons in the anterior nucleus basalis during non-rapid eye movement sleep. These neurons stimulate pyramidal neurons and hyperalgesia through directly acting on the S1 region.⁴⁹¹

There are a few investigations on comorbidities of other mental diseases, such as anhedonia, memory impairment and spatial awareness dysfunctions.^{136,492,493} Notably, some neuronal circuits mediate more than one comorbidity of pain. The overlaps between depression and anxiety have attracted considerable attention. Despite distinct clinical symptoms, many brain regions simultaneously regulate depression and anxiety, including the CeA, ACC, locus coeruleus and amygdala.^{494–496} In general, the research on pain comorbidities unveils close associations of pain with other psychiatric disorders. The importance of investigations in this field is not only derived from huge burdens of pain comorbidities. The achievements may provide new diagnostic biomarkers and therapeutic targets. However, the risks of novel therapies in exacerbating mental diseases should not be neglected, given the crosstalk between comorbidities.

APPROACHES FOR PAIN ASSESSMENT AND DIAGNOSIS

Pain, often challenging to describe and quantify in clinical practice, encompasses various factors for assessment and diagnosis, including its site, degree, type, frequency, duration and pathological phenotypes. In the past for a long time, pain assessment has heavily relied on scales, which is highly subjective and lacks universality and accuracy. Misinterpretation of patient-reported pain may cause medication overprescription or poor responses. The emergence of various novel achievements has provided improved options. This section will focus on the methods that have been applied or showing potential for pain assessment and diagnosis.

Pain questionaries

Pain questionaries are the commonly used tools for pain screening and evaluation, especially for neuropathic pain. The singledimensional pain scales are the simplest tool, like VAS, FPS-R and NRS. They rely on the single type of visualized information, including numbers, faces and languages. The subjects can finish examination tasks within 1 min. Due to its simplicity and understandability, whereas compromised with accuracy, they are widely used for pain screening. For the multidimensional scales, NPQ, PD-Q, LANSS and DN4 serves as the representative tools. They include more indicators involving psychology, sleep and emotion. Patients suffering from neuropathic pain can be screened out based on symptom description and self-perception. Screening questionnaires enable rapid initial judgments but may compromise assessment accuracy, like NPQ with only 64% sensitivity and 74% specificity in distinguishing neuropathic pain from nociceptive pain.⁴⁹⁷ PD-Q questionnaire overcomes the weakness of yesor-no options in previous questionnaires and uses quantitative scoring options instead. It considerably improves the evaluation accuracy.⁴⁹⁸ Until now, PD-Q has been widely employed in screening of chronic neuropathic pain.⁴⁹⁹

Evaluation questionnaires are designed to assess the patient's symptoms and evaluate the effectiveness of clinical therapy. NPSI, is a self-administered questionnaire developed to describe the clinical symptoms of pain. Unlike the screening questionnaire, NPSI focuses on assessing the effectiveness of pain management and delineation of pain subtypes.⁵⁰⁰ BPI, with advantages of rapid test and wide-range application, is mainly used to evaluate the pain severity and the impact on quality of life. It has been proved as a reliable evaluation tool for diabetic neuropathy pain.⁵⁰¹ OASIS is a questionnaire used to assess different pain dimensions in osteoarthritis patients with OA. Its most obvious feature is the assessing changes in pain characteristics over time, suggesting that it could be used in the future for drug studies to assess patient responsiveness to treatment.⁵⁰²

As a rapid evaluation method of pain, questionnaires have prominent characteristics. They are able to offer efficient and costeffective pain assessment, promoting accessibility for patients and interpretability for doctors. The subjective sensations are the most important indicators of pain, since pain severity determines the clinical outcomes of individuals. However, pain questionnaires cannot be used for diagnosis and not encouraged to serve as the sole evaluation tool. More methods should be conducted alongside to consolidate the clinical results.

Quantitative sensory testing (QST) and neurophysiological tests QST is a psychophysical detection method to assess the perception of touch, vibration, proprioception and sensitivity to various stimuli like pinprick, blunt pressure and temperature in experimental settings. It is commonly applied in mechanistic investigations for providing personalized medicine. As an important neuroelectrophysiological technique, QST is valued for simple operation, non-invasive nature and robust repeatability.⁵⁰³ It stimulates different fibers to produce sensations. Temperature sensation is used to examine the functions of A\delta and C fibers, while vibration sensation serves as a marker of AB nerve fiber activity.^{504,505} QST typically categorizes pain disorders into sensory loss, thermal hyperalgesia, and mechanical hyperalgesia. Quantitative thermal testing (QTT), a kind of QST, is the most widely used sensory testing. QTT contains four main indicators, including cold detection, warm detection, cold pain threshold and heat pain threshold. They collectively reflect the nerve fiber integrity, especially small fibers.^{506,507} QTT is suitable for patients with lumbar disc herniation who have inconsistent imaging and clinical manifestations or contraindications to MRI examination.⁵⁰⁸ It also shows effectiveness in trigeminal neuralgia-associated pain assessment according to alterations in temperature perception thresholds.

Although QTT effectively compensates for the questionnaire shortcomings, it still has obvious limitations. As a method causing additional pain experience and psychological stress, QTT is commonly applied in experimental research, far from clinical use. Similar to other psychophysical testing methods, QTT requires active engagement of patients, and test results are susceptible to

the psychological state.⁵¹⁰ The lengthy examination can cause fatigue and boredom, affecting the accuracy of pain assessment.⁵¹¹ Administration of analgesic drugs, like opioids, may also interfere with detection results.⁵¹² Therefore, the potential of QTT in pain evaluation needs more in-depth exploration to figure out its optimal application conditions and reference value.

Unlike QST, which is used to evaluate small-fiber neuropathy, neurophysiological tests are primarily used to evaluate large noninvasive afferent fibers. These tests include nerve conduction, trigeminal reflexes (including the blinking reflex), and somatosensory evoked potentials.⁵¹³ Some tests achieved 94% specificity and 87% sensitivity in diagnosing certain neuropathologic pain.⁵¹⁴ A comprehensive clinical examination, pain questionnaire, QST and neurophysiological tests are helpful for the clinical assessment of patients with neuropathic pain.

Brain imaging

The activities of specific brain regions are associated with pain severity. Based on these findings, brain imaging techniques have been developed, including structural MRI (sMRI), functional MRI (fMRI) and electroencephalograph (EEG). Brain imaging has emerged as a potential diagnostic biomarker for each type of pain. fMRI is an imaging technique developed on the basis of traditional MRI. It analyzes functional brain network connectivity, detecting neural activity through blood oxygen level-dependent signals. High signal intensities in fMRI are correlated with increased pain perception processing in specific brain regions. The data derived from fMRI can distinguish patients with acute or chronic nociplastic pain from healthy controls with over 63% accuracy.^{515,516} sMRI, engine by AI, utilizes gray matter images for diagnosis, achieving over 70% accuracy based on machine learning models.^{517,518} A prediction model constructed by squares-discriminant analysis likewise accurately distinguishes patients with chronic abdominal pain from the healthy controls.⁵ EEG is employed to detect brain oscillatory activities and functional connectivity changes at different frequencies. It has presented good performances in pain diagnosis due to its properties of ease of use and cost-effectiveness.519,520 More importantly, EEG serves as a cross-site pain diagnostic biomarker to differentiate patients with different types and severities of pain.52

However, constrained by the limitations of cross-sectional studies, these investigations have not delved into other pain biomarkers like monitoring and prognostic markers. This underscores the need for longitudinal studies focused on identifying dynamic detection and prognostic biomarkers. Long-term observation of subacute patients has shown that changes in NAc volume often correlate with the transition to chronic pain,^{522,523} indicating the potential of brain imaging as a prognostic tool. fMRI in identifying prognostic biomarkers for pain has been utilized.⁵²⁴ The functional connectivity between the NAc and PFC, which strengthens with persistent pain, can predict chronic back pain with an area under the ROC curve of 0.81.⁵²⁵ This connectivity's effectiveness in predicting short-term pain reduction in chronic pelvic pain syndrome has been validated with 73.1% accuracy.⁵²⁶

Despite the potential of these models in pain prognosis prediction, generalizing them to all pain types is challenging due to sample size limitations and the practicality of longitudinal studies. Recently, several pain-focused databases like OpenfMRI, OpenPain, and the Pain and Interoception Imaging Network repository have been developed, which should facilitate the construction of larger-scale, more reliable prediction models.

In summary, brain imaging's non-invasive nature and reproducibility make it an objective tool for pain mechanism research, drug action site identification, and intervention therapy development. However, there is a current shortage of longitudinal brain imaging biomarker studies. Future research should focus on largescale studies utilizing shared data to develop composite biomarkers. Most importantly, individualized pain diagnosis and treatment based on patient classification is a critical future research direction.

Molecular biomarkers

Given the close correlations, associations, and causal links between pain and molecular mechanisms, certain key molecules hold potential in clinical diagnosis (Table 2). This research area is an increasingly mainstream. The genetic variants have been linked to neuropathic pain, such as *Hla*, *Comt*, *Oprm1*, and *Gch1*.⁵²⁷ Large cohort studies have made initial forays. An investigation using the data from Genome Wide Association Study identified 171 differentiated mutant phenotypes in neuropathic ocular pain.²⁰⁶ Many targets, however, lack universality due to varied mutation sites.^{528,529} In the future, machine-learning algorithms may enable the combined application of multiple genetic variants.

Besides gene mutations, gene expression products exhibit more promising potential in pain diagnosis, though lacking direct and robust evidence. The transcriptome analysis has shown that GTF2H2, KLHL5, LRRC37A4P, PRR24 and MRPL23 are highly expressed in the serum of patients with neuropathic pain. The diagnostic model constructed by this gene expression signature can reach an area under curve (AUC) of 0.83.530 Combinational use of anti-inflammatory and pro-inflammatory cytokines in serum, such as IL-4, IL-6, and TNF-a, has been proved as potential biomarkers of neuropathic pain or visceral hypersensitivity, with AUC at least 0.66.^{531–533} Metabolomics has led to the emergence of serum metabolites as pain biomarkers. Patients with chronic pain often have abnormal serum levels of kynurenine pathway metabolites,⁵³⁴ although their diagnostic efficacy remains unvalidated. Urine-based models using multiple metabolite indicators provide better predictive value, with an AUC of 0.749.53

Molecular probes combined with imaging are increasingly being used for diseases requiring biopsy, like tumors.⁵³⁶ They can significantly enhance diagnostic efficiency, particularly for small lesions.⁵³⁷ This progress can provide enlightenment for pain diagnosis. The molecular probes can be designed to interact with pain biomarkers. Magnetic Resonance Spectroscopy, a new MRI technology, could potentially detect molecular pain biomarkers, improving diagnosis and continuous condition monitoring. Signal intensity changes could also monitor drug effects, aiding in dynamic treatment adjustment. Importantly, molecular probes have potential in informing individualized intervention regimens.

The other techniques like skin biopsy for peripheral neuropathy and gut microbiome analysis for pain severity correlation offer potential diagnostic avenues.^{538,539} Skin biopsy is an examination method to investigate nociceptive fibers in the human epidermis and dermis. It is used to investigate peripheral neuropathy, including diabetic neuropathy and infectious neuropathy, with sensitivity and specificity approaching 90%.540 The primary advantage of nerve ultrasound lies in its non-invasive and convenient nature, enabling the assessment of peripheral neuropathy through the use of a high-resolution ultrasound probe, as opposed to skin biopsy.⁵⁴¹ However, the positive findings of both skin biopsy and neuroultrasound cannot be directly linked to the diagnosis of pain.542 The primary challenge in pain diagnosis is the absence of objective standards and effective assessment approaches. The progress in pain diagnosis seriously lags behind the research on pain mechanisms and therapies. Integrating objective diagnosis with psychosocial research to develop comprehensive clinical diagnostic approaches remains a complex task requiring further investigation and validation.

INTERVENTION METHODS FOR PAIN RELIEF

Human has been grappling with pain throughout the long history. The complexity of pain's etiologies, including biological,

psychological and social factors, necessitates multidisciplinary treatment approaches. Primary disease therapies are crucial. However, for the incurable diseases and pain chronicity, specialized analgesic methods are meanwhile required. The surge in preclinical research contributes to the emergence of many novel therapies and intervention targets. Identification of underlying mechanisms provides additional insights into the efficacy of widely used therapies. The multimodal approach mainly includes three aspects according to the development stages: the traditional, rejuvenating and emerging therapies (Fig. 9). The traditional therapies refer to the widely applied approaches with mostly identified mechanisms underlying pain relief. By contrast, although the rejuvenating therapies have been used for pain management for a long history, their mechanisms are largely unknown and gradually being revealed. Some novel strategies hail from the old and plain therapies by virtue of modern biotechnology. The emerging therapies are completely the products of biotechnological advances, which require further explorations and validations. This section aims to summarize current achievements, offering a clear and comprehensive overview for researchers and clinicians.

Non-opioid pharmaceuticals

Non-opioid drugs are the most commonly used for pain management by virtue of their high dependability and safety. They mainly include NSAIDs, antidepressants, gabapentinoids, antiepileptics and cannabinoids. The non-opioid pharmaceuticals are applicable to different types of pain. For example, NSAIDs are evaluated as the first-line treatment for inflammatory pain, while antidepressants and antiepileptics are preferred for neuropathic pain caused by nerve injury and neuropathy.

NSAIDs function as cyclooxygenase (COX) inhibitors, reducing prostaglandin production. COX-2, inducibly expressed in immune cells under specific stress events like infection and inflammation, is targeted by NSAIDs to suppress immune cell activation and encourage oligodendrocyte differentiation.⁵⁴³ This leads to a decrease in proinflammatory factors and restoration of nociceptor sensitivity. In addition to the peripheral effects, NSAIDs also impact activities of brain regions, such as the PAG, CeA and spinal cord, ^{544,545} which serve as important central mechanisms of analgesic effects of NSAIDs. They have been recommended as first-line or primary analgesic drugs by many authoritative guidelines, such as PROSPECT for postoperative pain, ACP/AAFP guideline for musculoskeletal acute pain, ESCEO/OARSI guidelines for osteoarthritis and ACP guideline for gout.^{546–549}

Tricyclic antidepressants (TCAs) inhibit the reuptake of norepinephrine and 5-HT, thereby increasing monoamine neurotransmitter concentrations in synaptic clefts. According to the evidence of basic research, 5-HT enhances the inhibitory effects of serotoninergic system on the descending spinal bulb, together with the influences on β 2-adrenergic receptor-mediated antiallodynic action.⁵⁵⁰ The depression-independent mechanisms have also been identified. TCAs modulate opioid and endocannabinoid systems.^{551,552} The densities of membrane ion channels and glutamate transporters are downregulated.^{553,554} They likewise exhibit the anti-inflammatory properties.⁵⁵⁵ TCAs are recommended as the first-choice medications for neuropathic pain according to the IASP guideline.⁵⁵⁶

Selective serotonin reuptake inhibitors (SSRIs), such as fluoxetine, paroxetine and sertraline, are another kind of antidepressants. They have pharmacological properties of inhibiting 5-HT reuptake, increasing synaptic 5-HT concentrations. The regulatory effects of 5-HT depend on subtypes of 5-HT receptors, which have various and even opposite functions on pain modulation. Overall, SSRIs act as pain suppressors, but a literature review of related clinical trials has shown that the efficacy of SSRIs is less effective than that of TCAs, possibly due to canceling various receptors out.⁵⁵⁷ Their wide-range pharmacological actions further increases

Ref. 753 754 755 756 757 530 758 532 533 759 535 762 AUC area under curve, CLBP chronic low back pain, CRPS complex regional pain syndrome, FPI foundation pain index, ICBPS interstitial cystitis/bladder pain syndrome, NLR neutrophil-to-lymphocyte ratio, UAP 231 760 761 763 Serum NLR differentiates patients with IC/BPS from CASP5, and differentiate UAP patients from healthy individuals. OPG, as a serum marker, can effectively distinguish DNAJC18 and CNTN1 can predict pain severity in This gene signature can differentiate neuropathic differentiate endometriosis patients from controls. Serum TNF- α level can predict the progression of requiring different doses of opioids for analgesia. Jrinary ATP has predictive value in the diagnosis distinguishing chronic neuropathic pain patients Serum IL-6 level combined with clinical imaging A FPI model based on 11 metabolites effectively differentiates chronic pain patients from healthy WLS expression serves as a biomarker panel for occurrence of neuropathic pain in patients with slCAM-1 can be used to evaluate CLBP patients predictive value for the Serum IL-4, peritoneal IL-2 and IL-4 are able to the required postoperative dosages of sufentanil for gastric neuropathy in patients with spinal cord injury. Gal-9 effectively differentiates endometriosis The panel effectively distinguishes patients and patient subjective feeling can effectivel to evaluate the severity of The combination of MYC, STAT1, TLR4, CRPS patients from healthy controls. pain patients from healthy controls. Preoperative serum F13B predicts and evaluation of IC/BPS severity. men and women, respectively. patients from healthy controls with different pain intensities Vitamin D shows good osteoarthritis patients. from healthy controls. rheumatoid arthritis. nealthy controls cancer patients. sPIICP is used Performance controls. 0.647/0.706/ 0.588 Specificity 0.938 0.790 0.855 0.643 0.760 0.794 0.674 0.921 0.636/0.727/ 0.697 Sensitivity 0.830 0.940 0.740 0.760 0.736 0.962 0.484 0.937 0.660/0.710/ 0.700 0.780/0.630 Accuracy 0.710 0.806 0.890 0.859 0.973 0.830 0.852 0.800 0.840 0.980 0.749 0.765 AUC/ 0.800 0.811 participants Number of 1038 212 213 135 227 140 487 106 240 80 93 50 54 68 58 2 Chronic neuropathic Rheumatoid arthritis Psychiatric patients Postoperative pain Neuropathic pain Neuropathic pain The summary of molecular biomarkers in pain diagnosis Endometriosis Endometriosis Osteoarthritis Chronic pain Chronic pain with pain Disease IC/BPS IC/BPS CRPS CLBP UAP pain peritoneal fluid Serum and Urinary Urinary Sample Serum Skin MYC, STAT1, TLR4, CASP5WLS GTF2H2, KLHL5, LRRC37A4P, PRR24, MRPL23 Serum IL-4, peritoneal fluid OPRM1, OPRK1, OPRD1, urgent abdominal pair 11 metabolites (e.g. methylmalonic acid, xanthurenic acid) DNAJC18, CNTN1 Vitamin D Biomarker **SIGMAR1** IL-2, IL-4 Table 2. sICAM-1 sPIICP TNF-α Gal-9 F13B Ddo NLR IL-6 ATP

Pathology of pain and its implications for therapeutic interventions Cao et al.

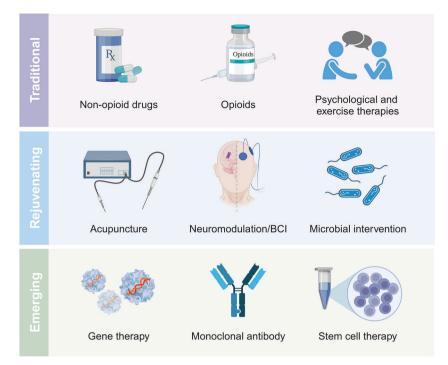


Fig. 9 The summary of currently developed therapies for pain management. The nine methods are categorized into three groups, including traditional, rejuvenating, and emerging therapies. BCI brain-computer interface

risks of side effects. SSRIs are not used as the first-line pharmaceuticals in clinical pain management.

Serotonin-norepinephrine reuptake inhibitors, like venlafaxine and duloxetine, show better performance in treating neuropathic pain because of the additional effects on noradrenaline. It serves as the first-line medication for neuropathic pain recommended by the IASP guideline.⁵⁵⁶ A multicenter clinical trial called OPTION-DM investigated the efficacy of sequential administration supplemented with pregabalin. This regimen effectively alleviated diabetic peripheral neuropathic pain but with a significant increase in nausea compared to monotherapy.⁵⁵⁸

Ketamine is known for rapid-acting antidepressant properties. Preclinical studies have shown that its analgesic effects are mostly dependent on NMDAR antagonism, blocking the wind-up phenomenon in the spinal dorsal horn.⁵⁵⁹ The ascending inhibitory pathway activity and antiinflammation are also promoted. The ketamine metabolite, 2R, 6R-hydroxynorketamine, activates mTOR signaling in PFC, which in turn enhances ketamine efficacy.⁵⁶⁰ This metabolite also downregulates CGRP release through inhibiting the activities of TRPV1 and TRPA1.⁵⁶¹ For clinical practice, ketamine is recommended to treat moderate-severe postoperative pain by the ASRA/AAPM/ASA consensus guideline. Patients with opioid tolerance or obstructive sleep apnea may gain benefits from ketamine for pain relief due to its properties of compensate opioid efficacy and decreasing opioid requirements.⁵⁶²

Gabapentinoids are the classical antiepileptics. Basic studies have shown that gabapentinoids bind to $\alpha 2\delta$ subunit, decreasing ion influx through neuronal VGCCs. Local administration of gabapentinoids inhibits responses to evoked stimuli.⁵⁶³ The effects of gabapentinoids partially rely on the opioid system, involving the rostral ACC and downstream mesolimbic reward circuits engaged in learned pain-motivated behaviors.⁵⁶⁴ Gabapentinoids have been recommended for painful diabetic polyneuropathy.⁵⁶⁵ Pregabalin has better efficacy for the treatment of post-total joint arthroplasty than gabapentin.⁵⁶⁶ However, gabapentinoids are not or weakly recommended for other types of postoperative pain according to the PROSPECT guidelines because of their high risk-benefit ratio.^{567,568} Ongoing clinical trials are trying to expand indications of gabapentinoids in pain management.^{558,569}

The molecular mechanisms of cannabinoids are complicated, dependent on cannabinoid receptor types, including cannabinoid receptor-1 (CB1) and cannabinoid receptor-2 (CB2). CB1 activation in the mPFC inhibits activities of dorsal horn and peripheral neurons, preventing neuropathic pain chronicity.⁵⁷⁰ The circuit from the zona incerta to posterior complex of the thalamus is critically involved in pain sensation. The hyperactivity of CB1 specifically expressed in axon terminals of this circuit mitigates nocifensive responses via presynaptic inhibition.⁵⁷¹ Intriguingly, CB1 counterintuitively sensitizes TRPV1 channel during prostaglandin E2-induced inflammation. The overactivation of TRPV1 inhibits action potential through dampening the depolarization rate in the capsaicin-induced model.⁵⁷²

CB2, enriched in immune cells, are linked to inflammation. CB2 activation decreases HDAC1 and IKBa expression, as well as promoting the release of anti-inflammatory cytokine IL-10 in microglia.⁵⁷³ The levels of proinflammatory factors are declined, such as IL-1 β and TNF- α , accompanied by NLRP3 inflammasome deactivation.^{574,575} CB2 effects are not limited to peripheral inflammation inhibition. 2-AG can bind to CB2 receptor in the ACC and promotes fear-conditioned analgesia.⁵⁷⁶ Additionally, CB1 and CB2 receptors have overlaps in mechanisms, like interactions with Nav1.7 and Nav1.8 to inhibit neuronal activity.^{577,578} However, the endocannabinoid system can activate GABAergic neurons in the mPFC. The excessive release of GABA may increase risks in depression comorbidity.⁴⁷³ Researchers should be cautious in cannabinoids-induced negative emotion, particularly given the increasing trend of healthy individuals overusing commercial cannabinoids, like cannabidiol (CBD) gummies or water.

A large number of clinical trials have explored the efficacy and appropriate administration regimens of cannabinoids. For instance, δ -9-tetrahydrocannabinol (THC) alleviates chronic neuropathic pain compared to placebo therapies. The reduction in functional connectivity between the ACC and sensorimotor cortex

is observed, suggesting that cognitive-emotional modulation is a critical mechanism underlying its analgesic effects of THC.⁵⁷⁹ A meta-analysis including 16 clinical trials and 1750 cases showed that cannabis-based medicines are significantly effective in chronic neuropathic pain relief.⁵⁸⁰ Furthermore, CBD can exert profound pharmacological interactions with THC.⁵⁸¹ A phase-II adaptive Bayesian trial indicated that CBD can restore cannabis use disorder.⁵⁸² However, many studies have reported the negative results of cannabinoids, involving acute nociceptive pain, abdominal pain, postoperative pain and inflammation-associated pain.⁵⁸³⁻⁵⁸⁶ A meta-analysis focusing showed that existing data do not support cannabinoids as an effective drug for both chronic non-cancer and cancer related pain.^{587,588} The effects of medical cannabinoids differ. Only dronabinol and nabiximol exhibit significant therapeutic effects on pain associated with a variety of diseases.⁵⁸⁹ The transient adverse side effects further impaired application value of cannabinoids.⁵⁸⁷ Taken together, cannabinoids are suitable for the treatment of neuropathic pain, rather than other kinds of pain. The drug types, administration dosage and regimens need to be extensively investigated by subsequent clinical trials.

There are several challenges of clinical trials on cannabinoids. Most current studies evaluated pain intensities based on single pain indicators. However, quality of life, sleep, motivational and cognitive dimensions are also have closely associated with patient pain experience. Studies should include more comprehensive indexes to assess analgesic effects of cannabinoids. The interactions with other drugs, like opioids and NSAIDs, are still lacking. The efficacy and potential risks of combinational use should be carefully clarified. The investigations into optimal therapeutic ratios of cannabinoids for different pain conditions are also required, which may provide proofs for developing strategies of separating analgesia from adverse effects. Furthermore, real-world data concerning cannabinoids are currently absent. The data can better illuminate benefits and harms of cannabinoids.

Opioids

Opioids are analgesic drugs usually used for treating many types of moderate and severe pain at all ages, based on extensive evidence-based medicine. They play a vital role in clinical symptomatic and palliative therapies. Emerging clinical trials have explored novel opioid regimens. For instance, an international, open-label trial gives a strong recommendation for two-step cancer pain management, which refers to bypassing weak opioids in the pathway from non-opioid therapies to strong opioids. This regimen can decrease the cost of two-step approach and achieve the comparable efficacy.⁵⁹⁰ The combinational use of CBD effectively improves patients' quality of life receiving opioid treatment.^{591,592} The sustained-release and topically administered forms have entered clinical trials for improving analgesic efficacy and reducing adverse event risks.^{593,594}

Opioids are agonists of μ , κ , and δ receptors, with diminishing effects across these subtypes. Opioid-induced analgesia involves multiple mechanisms. The canonical manner is the suppression of adenylyl cyclase and high-threshold VGCCs through G protein coupling pathways. The inwardly rectifying potassium channels are meanwhile activated, accompanied with inhibition of TRP family members, VGSCs and ASICs.^{595–598} These events jointly decrease neuronal excitability and excitatory neurotransmitter levels. In the brain's reward circuits, opioids mitigate GABA-driven inhibitory neurotransmission. The suppression on dopaminergic neurons in the striatum and PFC is reversed.^{599,600} The proinflammatory neuropeptide release is also downregulated, further promoting the analgesic effects of opioids.⁶⁰¹

The side effects of opioids are the vital reasons for a series of clinical and social problems, mainly including tolerance, hyperalgesia, respiratory depression and gastrointestinal reaction. They 25

are driven by complicated mechanisms, but some crucial cross nodes have been identified. β -arrestin 2 is a negative regulator of GPCR signaling, implicated in opioid tolerance, addiction and respiratory depression through coupling with intracellular and cytoplasmic regions of phosphorylated μ receptors.⁶⁰² Biased agonists with reduced β -arrestin 2 recruitment, like 2S-LP2 and EM-2, are being developed to alleviate these effects.^{603,604} However, there are dissenting opinions that β -arrestin 2 as a scaffolding protein is unlikely to be an ideal pharmacological target. Evidences have shown that severe side effects are not observed in mice with ablation of β -arrestin 2 functions,^{605,606} implying that unknown mechanisms independent of β -arrestin 2 may contribute to chronic opioid tolerance.⁶⁰⁷ Instead, targeting recruited molecules of β -arrestin 2, like vasopressin 1b receptor, may be a promising approach.⁶⁰⁸

Neuroglia cells are involved in opioid tolerance and hyperalgesia. MAPK/NF- κ B signaling activation in microglia promotes release of proinflammatory factors and upregulates expression of TLR4.⁶⁰⁹ Inhibitors of MAPK/NF- κ B signaling can attenuate opioidassociated side effects in rodent models.^{610–612} CR4056, an imidazoline I2 receptor ligand, suppresses microglia activation and enhances analgesic effects of morphine.⁶¹³ NMDAR in astrocytes is another potent target. The inhibitors targeting NMDAR effectively block intercellular communications between astrocytes and neurons, alleviating opioid tolerance.⁶¹⁴ Antagonizing IL-33-mediated crosstalk between astrocytes and oligodendrocytes also prolongs morphine's analgesic effects.⁶¹⁵ Therefore, neuroglia may serve as promising targets in mitigating opioid side effects.

Tramadol is a weak opioid agonist widely used for pain relief. Tramadol is primarily used for postoperative pain and chronic musculoskeletal pain management. It has also been recommended as the non-first-line drug for the treatment of neuropathic pain by CPS and EFNS guidelines.^{616,617} Additionally, it can be safely and effectively used for delivery analgesia without affecting the newborn's respiration. Recently, a chewable tablet has been invented and used for children, further proving its safety.⁶¹⁸ For the basic mechanistic research, tramadol exhibits a dual mechanism, primarily through the activation of opioid receptors in the CNS presynaptic membrane and the inhibition of 5-HT and norepinephrine reuptake in the presynaptic membrane of the descending inhibitory system of spinal cord. This dual mechanism allows tramadol to achieve analgesic intensity comparable to opioids at appropriate dosages.

Otherwise, nitric oxide, a gasotransmitter, activates presynaptic and postsynaptic guanylate cyclase, leading to the production of cGMP. This signaling further promotes opioid tolerance proved by basic studies. NOS inhibitors, like L-NAME and aminoguanidine, attenuate morphine tolerance. Repeated administration can further alleviate the withdrawal symptoms of opioids.^{619,620} Despite promising preclinical results, the translational speed has slowed markedly in recent years. The clinical application of NOS inhibitors remains cautious, pending further evidence.

Psychological and exercise therapies

Restoration of psychological and behavioral disorders has demonstrated significant value in pain management. CBT, which integrates behavior modification with psychotherapy, is a goldstandard approach for treating mental diseases. Its analgesic effects are applicable to adults at all ages with chronic pain, such as chronic low back pain, osteoarthritis and IBS.^{621,622} Notably, a recent meta-analysis based on 153 trials and 8713 participants has strongly recommended CBT for management of chronic pain associated with temporomandibular disorders.⁶²³ To further promote application of CBT, clinical trials began to investigate the efficacy of online CBT. The results showed its comparable competences with traditional psychotherapies at the dramatically lower cost,^{624,625} providing new directions of CBT development.

For the mechanistic investigations, CBT induces global alterations in brain region activities. Prevention of pain catastrophizing is an important mechanism of CBT, which relies on the regulation of the ventral posterior cingulate cortex, a hub of the DMN. CBT impairs the connectivity between the somatomotor and salience network regions in fibromyalgia patients.⁶²⁶ The connectivity strength involving the ventral posterior cingulate cortex is negatively correlated with CBT efficacy.⁶²⁷ The mPFC is another key node in the DMN. CBT facilitates new long-term potentiation connections in the mPFC with other critical regions, like the amygdala and insula, underpinning significant correction of chronic nociplastic pain.^{628,629} The enhanced crosstalk of the amygdala with the ACC, frontal and precentral gyrus is also related to the responsiveness to CBT.⁶³⁰ Although most evidences only demonstrate its correlations with brain region activities, these extensive and profound influences suggest the substantial potency of CBT.

Exercise is another cost-effective therapy against pain, which has been strongly recommended by recent guidelines for the clinical management of pain associated with motor system, including fibromyalgia, osteoarthritis, low back pain, chronic musculoskeletal pain and temporomandibular disor-ders.^{549,623,631-633} It has also been evaluated as an important component of therapies against cancer and neuropathic pain.^{634,635} Inflammation mitigation is a major mechanism of exercise, which is mentioned above in the section of pain sexual dimorphism.^{457,458} The descending regulatory pathway is an important target of exercise. Regular exercise increases the concentrations of endogenous opioids (β-endorphin and enkephalin) in the PAG and RVM.⁶³⁶ The activities of regions, such as the anterior insula, left dorsolateral PFC, locus coeruleus and midbrain reticular formation, are globally altered in patients with nociplastic pain.^{637,638} However, due to various methods and intensities of exercise therapy, the optimal prescription and delivery for specific diseases should be extensively discussed based on more high-level basic and clinical research.

Placebo effect refers to the phenomenon that symptoms are alleviated through psychological functions produced by patients' belief after receiving dummy treatment. It is especially common in the pain research. The forms of placebos are various, such as tablets, pseudostimulus and sham operation. Their efficacy rivals some classical modalities.⁶³⁹ Recent investigations using virtual reality technology have proved that physical entities are unnecessary for pain management.⁶⁴⁰ The adjuvant functions in synergistically enhancing other therapies have also been demonstrated.⁶⁴¹ On the other hand, the reliability of existing analgesic methods is questioned because it is hard to tell whether placebo effect is involved. It is great pleasure to see that project designers of clinical trials have recently been aware of this confounding factor, leading to recalibration of specific therapy effectiveness.

Activation of opioid and endocannabinoid systems is the dominant mechanism of placebo-induced analgesia, which hints that placebos may become alternative therapies of opioids and cannabinoids.⁶⁴² The PFC, insula and somatosensory cortex have been found to be engaged in this top-down effect, together with processing pain anticipation and perception by the thalamus and brainstem.^{643,644} Some researchers hold the view that functional connectivity may be more sensitive for manifesting placebo effect than isolated brain regions. The decreased connection between the left medial PFC and bilateral insula, responsible for cognition modulation, is correlated with placebo effect in patients with chronic back pain.⁶⁴⁵ The circuits from prefrontal cognitive to pain processing regions also serve as indicators of responsiveness to placebos.⁶⁴⁴ Furthermore,

nocebo has properties of hyperalgesia induction. Existing basic studies have revealed totally different neural networks subserving placebo and nocebo effects,⁶⁴⁶ further validating the complexity of pain perception.

In addition to the above methods, mindfulness, short-term dynamic psychotherapy and hypnosis are other psychotherapies for pain management. The research on psychological and exercise therapies has thrived. However, a disparity exists between the abundance of clinical trials and the scarcity of basic research, partly due to challenges in replicating psychological or voluntary exercise in animal models. Future studies should address this imbalance and develop more foundational experimental methods.

Acupuncture

Acupuncture originates in ancient China and its effectiveness has been confirmed through clinical practice. According to the guidelines, acupuncture can serve as an alternative and complementary therapy for pain management, especially for cancer, low back pain and postoperative and osteoarthritis pain.^{546,632,647,648} Additionally, clinical trials stimulation of pain-specific acupoints produces inhibitory effects of comorbidities of depression, anxiety and sleep disturbance.^{649,650} The underlying principles of acupuncture, based on traditional Chinese medicine, initially led to skepticism within modern medicine. In the last century, Jisheng Han et al. clarified the spatial and temporal events and related mechanisms of acupuncture-induced analgesia. This is a historic milestone of utilizing biomedical technologies to elucidate acupuncture-induced analgesia mechanisms.

Early preclinical studies indicated that acupuncture stimulates Aδ and C afferent nerves and promotes secretion of endogenous opioid peptide, as well as the reduction in activities of norepinephrine and serotonin systems. With the deepening of basic research on clinical patients, the nodes in default mode and frontoparietal networks have been identified.651 The connectivity between the amygdala, right middle cingulate cortex and temporal gyrus is enhanced in patients with nociplastic pain.⁶⁵² Activities of the thalamus, caudate, claustrum and lentiform are likewise modulated.⁶⁵³ Importantly, there are typical differences between acupuncture and sham control groups, allaying the concerns of the placebo effect.⁶⁵² Molecular mechanisms underlying acupuncture are considerably diverse, including epigenetic modification, PTMs, non-coding RNAs, inflammasome and microbiota.^{41,204,574,654,655} However, most studies merely reveal their changes following acupuncture, which may be accompanying effects, rather than action mechanisms. The causal studies using rescuing experiments are needed. Taken together, acupuncture is a promising therapy for pain management, but requires further exploration to develop individualized regimens.

Neuromodulation and brain-computer interface (BCI)

Neuromodulation refers to the approaches that directly or indirectly implant electrodes into innervation regions to improve pathological changes and clinical symptoms. Brain, spinal cord, vagus, sacral nerve, auditory nerve, etc. are all the interventional targets of invasive neuromodulation. Deep brain stimulation has garnered the most attention with the deepening understanding in brain region functions. The sensory thalamus, PAG, ACC and periventricular gray matter are main anatomic regions of deep brain stimulation.⁶⁵⁶ In contrast, transcranial alternating current stimulation represents noninvasive neuromodulation. Though less potent, its safety and convenience may propel it to the forefront of future research. In summary, neuromodulation has proven effective in clinical trials for chronic pain management.^{657,658}

Spinal cord stimulation refers to a pain management technique that involves implanting electrodes into the epidural

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space in the spinal cord for modulating neural electrophysiology. The non-nociceptive electrical signals can inhibit the transmission of nociceptive signals by stimulating the large diameter AB-fibers. The most common indications of spinal cord stimulation are complex regional pain syndrome, failed back surgery syndrome and peripheral neuropathy induced by ischemia, herpes zoster or diabetes. Many clinical trials and meta-analysis have verified the efficacy of spinal cord stimulation as a supplementary approach in attenuating neuropathic pain, especially for complex regional pain syndrome and failed back surgery syndrome.^{659,660} It has also been recommended as a third-line therapy for patients who have failed to respond to gabapentinoids and antidepressants.⁶⁶¹ In addition to the tonic stimulation, some novel stimulation waveforms have been proposed, including burst, high frequency and close-loop stimulation. These further enhance pain relief or reduce the risks associated with paresthesia perception.⁶

Although spinal cord stimulation was initiated based on Gate control theory, the extensive regulatory mechanisms have been gradually uncovered by many preclinical studies. Opioid system is involved in the effects of spinal cord stimulation and the stimulation at different frequencies rely on different endorphins and opioid receptors.⁶⁶³ The expression of CB receptors is upregulated following spinal cord stimulation treatment. Nociceptive-evoked activation of supraspinal areas, such as the locus coeruleus, RVM, reticular formation and PAG, can be inhibited by spinal cord stimulation.^{664,665} The descending inhibitory system is activated, leading to the release of 5-HT and attenuation of chronic neuropathic pain.⁶⁶⁶ The above mechanisms collectively contribute to the analgesic effects of spinal cord stimulation.

Despite significant achievements in spinal cord stimulation, there are difficulties in positioning specific pain regions, such as low back, knee and groin. The complex anatomy of spinal cord, shunting of electrical stimulation through cerebrospinal fluid and relative displacement of spinal cord in the canalis spinalis all impair the application of spinal cord stimulation.⁶⁶⁷ DRG stimulation can overcome these shortcomings. It simultaneously activates AB, AS, and C fibers. The cerebrospinal fluid around DRG forms a groove, attenuating the dispersion of electrical currents and avoiding the side effects of paresthesia within peripheral regions. Moreover, it can produce stable currents in the regions that spinal cord stimulation hardly achieves. According to the existing evidences from clinical trials, DRG stimulation has been selected as the primary treatment of lower limb type I or II complex regional pain syndrome.^{668,669} Patients suffering from chronic postsurgical inguinal pain, knee pain and types of chronic intractable pain can gain typical benefits from DRG stimulation.^{670–672} Some studies have found that stimulation frequencies are a determinant factor of DRG stimulation efficacy and 20 Hz might become the best choice.673,674 There are limitations in clinical investigations into either spinal cord stimulation or DRG stimulation. Although the effectiveness of electrical stimulation has been proved, the concrete regimens, such as stimulation frequencies, treatment interval, best indications, need further exploration. Their surgical characteristics make it difficult to set standard sham groups and adhere to blinding principles, affecting the reliability of current clinical data. More high-guality real-world studies should be conducted to compensate for these shortcominas.

The innovation of BCI has ushered neuromodulation into a new era. Preclinical and clinical research has shown its potential in alleviating neuropathic pain. Hence, in this section, we highlight this cutting-edge technique. Pain perception drives fluctuations in the brain network. Extracting features of this process may provide the sources of decoding pain perception with great accuracy.⁶⁷⁵ BCI can analyze the data supported by AI and produce real-time neuromodulation on multiple regions to

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mitigate pain. The S1, ACC, and PFC are the crucial targets of BCI. More importantly, the regulatory effects are not unilateral. Brain has been found to actively communicate with BCI and change its responses to pain, embodied by enhanced activities of the ACC and PAG and modulation of pain attention.⁶⁷⁶

The studies aiming at prompting more extensive use are ongoing. Patients with phantom limb pain have been trained with BCI to control a phantom hand.^{677,678} A three-day training session can alleviate pain perception for more than 1 week.⁶⁷⁹ In contrast, another study indicated that despite the enhanced discriminability for movement and prosthetic control, overconcentration on the phantom hand driven by BCI intensifies neuropathic pain. Dissociation between prosthetic and phantom hands is a more feasible way for analgesia.⁶⁸⁰ A multisensory intervention strategy consisting of BCI, virtual reality, and transcutaneous electrical nerve stimulation sharply increases the efficiency of decoding pain memory and attenuating neuropathic pain.⁶⁸¹ The invention of a memory and attenuating neuropathic pain.68 home-use, patient-managed BCI device has further accelerated the translation of BCI.⁶⁸² More importantly, another application of BCI is assisting movement for paralytic patients. Pain perception has been found to damage the performance of BCI on motor system control.⁶⁸³ These findings underscore the broader significance of these advancements, extending beyond mere pain relief.

Microbial intervention

The breadth of research on microbiota and pain has guided the development of related approaches to analgesics. Investigations into associations between microbiota dysbiosis and pain progression have promoted the translation of novel interventional regimens, mainly including probiotics supplementation and fecal microbiota transplantation (FMT). Notably, given the natural associations between microbiota and gastrointestinal tract, most studies focus on therapies against abdominal pain, particularly IBS.

Probiotic supplementation has exhibited translational value. In the preclinical research, administration of Saccharomyces boulardii reduces colonic TRPV1 expression and alleviates pain sensation in an IBS model.⁶⁸⁴ Lactobacillus paracasei and butyrate-producing Roseburia hominis can respectively attenuate visceral hypersensitivity through mitigating dysfunctions of gut homeostasis.685,686 Bifidobacterium dentium and Lactococcus lactis both have properties of enzymatic decarboxylation of glutamate. Their analgesic effectiveness by GABA production has been detected in visceral hypersensitivity models.^{397,687} The nociceptive perception induced by 5-HT is ameliorated by Lactobacillus plantarum through downregulating responses of the HPA axis.⁶⁸⁸ The efficacy of probiotic supplementation have also been confirmed by clinical trials.^{689,690} In addition to natural probiotics, researchers have attempted to exploit the advantages of bioengineering. Genetically engineered Lactococcus lactis delivering in-situ IL-10 and IL-22 have been successfully generated. They effectively alleviate visceral hypersensitivity.69

Probiotics have promising potentials in treating other diseases. For example, the achievements of probiotic application in migraine therapy are relatively mature. Multispecies probiotic strategies have stepped into the validation phase of clinical trials and have exhibited desirable therapeutic effects.⁶⁹³ The formation of dental plague biofilms is recognized as important contributors of oral diseases. A cocktail comprised of 5 bacteriophages targeting Salmonella typhimurium can rapidly inhibit biofilms produced by this pathogenic bacterium on the surfaces of teeth, alleviating caries-induced pain in vitro.⁶⁹⁴ In the model of chemotherapy-induced neuropathic pain, elimination of irinotecan-elicited microbiota communities using antibiotic administration effectively relieves hyperalgesia through inactivating TLR4-dependent mechanisms.40

Probiotic supplementation mainly influences specific bacterial species and hardly cause global effects on microbiome, which can

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be compensated by the whole microbiome transplants. In fact, oral consumption of fecal juice was employed to treat patients with food poisoning or diarrhea 1700 years ago. FMT from healthy controls effectively alleviates visceral hypersensitivity both in preclinical and clinical research.^{695,696} Repeated FMT treatment can clinically compensate for the unsustainable curative effects of a single intervention.⁶⁹⁷ The effectiveness and safety of FMT via different delivery routes have also been confirmed in clinical patients, including nasal feeding, enteroclysis and oral administration.^{697–699} The success rates of FMT are dependent on microbial diversity, stability and allocation between donors and recipients.^{696,697} Regretfully, sufficient evidence for establishing rigorous criteria for FMT matching, similar to those for clinical transfusion and organ transplantation, is lacking and eagerly required.

Gene therapy

The burgeoning advancements in bioengineering have significantly augmented the application of gene editing in both basic and clinical research. The identification of pivotal regulators lays a robust theoretical groundwork for gene therapies. Gene therapy methodologies can be categorized into two types based on delivery systems: viral infection and naked plasmids. Viral infection exploits the innate ability of viruses to transport exogenous genetic material into host cells. Viruses are engineered to carry gene editing systems, mainly including zincfinger nucleases, transcription activator-like effector nucleases and the Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) system. Undoubtedly, CRISPR/Cas9 technique has garnered considerable attention in current studies. Its recent endorsement for clinical application further underscores its substantial potential. Naked plasmids offer a viable alternative to circumvent the limitations of viral infection. The transfection of plasmids, encoding specific target genes, into cells, can modulate expression of particular RNAs or proteins, thereby influencing pain perception.

A single-arm clinical trial constructed a delivery system using herpes simplex virus to increase preproenkephalin levels in DRG. This methodology efficaciously mitigated pain perception in patients with intractable focal cancer pain.²³ This study inspires follow-up basic and clinical investigations into viral delivery systems. Some preclinical studies further expanded the potential of gene therapy. Ion channels are important targets in gene therapy. A study focusing on the genetic variations of TRPV1 conducted the CRISPR/Cas9-mediated introduction of a K710N TRPV1 missense variant, resulting in reduced calcium influx and dampened neuronal excitability. It decreased calcium influx and inhibited neuronal excitability, suppressing nociceptive and neuropathic pain.⁷⁰⁰ CBD3 is a peptide aptamer that antagonizes activity of Cav2.2, and delivery of CBD3-encoding gene via adenoassociated virus (AAV) alleviates neuropathic pain.⁷⁰¹ The integration of glutamate-gated chloride channel silences sensory neurons without affecting motor or proprioceptive functions, attesting to the precision and safety of AAV vectors.⁷

Intriguingly, basic studies have leveraged epigenetic and PTM mechanisms to modulate Nav1.7 expression. The CRISPR-dCas9 system guides inactivated Cas9 protein to *Scn9a* and circumvolutes the target gene instead of editing the sequences, thereby epigenetically blocking *Scn9a* transcription. It exerts long-lasting analgesia for inflammatory and neuropathy-induced pain.⁶⁹ Upregulation of SENP1 via CRISPR system reduces Nav1.7 expression through regulating CRMP2 deSUMOylation.³⁰³ Remolding phosphorylation sites on histone H3.1 by AAV raises the threshold for thermal nociception.⁷⁰³ The indirect regulatory functions of gene therapies may offer more rapid responses and enhanced safety based on the features of epigenetics and PTMs. Furthermore, introduction of the ASC-encoding gene effectively suppresses the schwannoma growth and mitigates associated

cancer pain perception.^{704,705} A combinational gene delivery strategy, aiming for simultaneous modulation of neuronal excitability and neuroinflammation, has shown promising results.⁷⁰⁶

Naked plasmids encoding hepatocyte growth factor (HGF) represent a potent approach for treating ischemic diseases. Their role in pain modulation has recently come to light. Topical administration of the naked HGF plasmids suppresses macrophage infiltration in DRG, concomitantly diminishing the release of proinflammatory factors, including IL-1 β , IL-6, and TNF- α .^{707,708} This pain relief strategy has passed phase III study on diabetic peripheral neuropathy,⁷⁰⁹ which is the gene therapy closet to clinical application to date. Similarly, IL-10 delivered by naked plasmids induces lasting neuropathic pain suppression independently of its endogenous form.⁷¹⁰

Taken together, the dual-track development of gene therapies has opened a new direction for upcoming studies. Some future prospects need to be highlighted. Although clinical trials have proven the short-term safety of gene therapies, long-term followup is essential to identify underlying risks, given the typical irreversibility of gene editing. Moreover, beyond Cas9, additional genetic scissors have been identified, offering superior applicability.⁷¹¹ Loading with the new systems may further improve the efficacy of pain management. Lastly, while current studies focus on hyperalgesia, the phenomenon of pain loss remains underinvestigated. Gene mutation is a critical cause of pain loss. Therefore, it is speculated that gene therapies could be effective in this domain.

Monoclonal antibody

Monoclonal antibody, akin to gene therapy, are part of novel strategies of targeted treatment. Monoclonal antibody inhibits functions of target proteins, which is widely applied in treating cancer and autoimmune diseases. In the pain research, CGRP monoclonal antibodies are undoubtedly the drugs with remarkable performances. Erenumab, fremanezumab, galcanezumab and eptinezumab, targeting CGRP or its receptors, have been approved by FDA to prevent migraine flares with applicability to a broad population. Current trials have been further expanding indications like post-traumatic headache and trigeminal neural-gia^{712,713} and exploring administration routes like oral and intranasal administration.^{714,715}

Antibodies targeting NGF have also been proved effective in relieving pain associated with various diseases based on clinical and preclinical research. A meta-analysis has shown that NGF antibodies effectively alleviate symptoms of hip and knee osteoarthritis. Whereas, the drugs may increase risks in therapy discontinuation due to side effects.⁷¹⁶ Basic studies have exhibited their additional potential in treating postoperative and diabetic neuropathy pain.^{717,718} Furthermore, Antibodies targeting proinflammatory factors, like TNF- α and IL-6, are clinically used for immune disorders, naturally attenuating concomitant pain. Neuropathic pain can also be mitigated by these antibodies,⁷¹⁹ suggesting there may be overlaps in mechanisms of inflammation and neuropathic pain. Other monoclonal antibodies being investigated are the candidates for clinical pain management, like antibodies against functions of HMGB1, TrkA and PAR2.⁷²⁰⁻⁷²² Modern bioengineering technologies spawn antibodies targeting more direct molecules, including Nav1.7 and TRPV1.^{723,724} The novel monoclonal antibodies proved by preclinical research are listed in Table 3. Despite insufficient evidence on their effectiveness and safety, development of new targets and mechanism-based approaches is an inexorable trend.

Stem cell therapy

Stem cell therapy is an important branch of regenerative medicine. It has been extensively applied in treating various

diseases. It is categorized into placenta and umbilical cord-, hematopoietic-, bone marrow- and adipose-derived types according to the origins. Their general mechanisms include the following three aspects. The direct way is rapidly dividing at lesions and preparing injured tissues. Stem cells also release a batch of trophic factors, like TGF- β , vascular endothelial growth factor (VEGF) and other cytokines. The paracrine function is regarded as more important than direct engraftment for chronic diseases, because the persistent efficacy can be observed after displacement and absorbance of stem cells.⁷²⁵ Stem cells are also able to exert global immunomodulation, such as inhibition of monocyte differentiation to dendritic cells, T cell development and NK cell regulation.

In the preclinical research, stem cells derived from bone marrow, adipose and peripheral nerve significantly attenuate neuropathic pain.^{726–728} The opioid tolerance is reversed by stem cell transplantation.⁷²⁹ VEGF is a potent regulator of counteracting complex neuropathy. Bioengineered stem cells expressing VEGF exhibit better efficacy in pain relief of patients with Parkinson's disease.⁷³⁰ Another notable finding is the significance of stem cell spheroids, which outperform the monolayer-cultured stem cells. The capability of neurotrophic factor secretion is dramatically enhanced, accompanied with impaired immune cell infiltration. The in-vivo survival time of spheroids is also extended.^{731,732} Development of stem cell spheroid treatment may be a promising direction. The effectiveness and short-term safety of stem cell therapy in pain induced by neuromuscular skeletal disorders and neuropathy have been validated by clinical trials.^{733,734} However, the long-term risks in carcinogenesis and autoimmune disease induction, need careful evaluation.

In conclusion, various therapeutic strategies have been developed and applied in clinical pain management. Besides the abovementioned modalities, there are other developing approaches, such as immunotherapy, nerve regeneration, diet management and surgical intervention. Another trend is integration with biomaterials. They are able to potentiate targeting abilities of loaded pharmaceuticals, consequently enhancing potencies for pain relief and avoiding undesirable side effects.⁷³⁵ However, clinical requirement for pain relief is far from satisfaction, again implying the complexity of pain sensation. Multidisciplinary cooperation, supported by high-quality preclinical and clinical research, may be a key solution to this dilemma.

SHORTCOMINGS OF THE CURRENT PAIN RESEARCH

The strength of evidence in existing research, particularly regarding brain regions, is generally weak. Studies often detected activity alterations in brain regions after pain induction or intervention, along with synergistic regional reactions. On the basis of these data, they inferred that some brain regions and functional connectivity might participate in pain modulation. Similar situations were also observed in the research on specific inflammatory mediators, neurotransmitters and molecular regulators. These findings are useful for developing novel diagnostic biomarkers and screening out potential targets. Nevertheless, they fall short in supporting concrete conclusions about pain etiology and treatment. Researchers should exercise caution in drawing conclusions based on assumptions from previous reports and established knowledge. Such weak evidence may obscure the true nature of pain modulation. For instance, the accepted view that TRPV1 activation serves as a marker of hypersensitivity, used in pain perception assessment. However, CB1-dependent TRPV1 overactivation is an important mechanism in dipyrone-induced analgesia,⁵⁷² overturning the stereotype regarding TRPV1. β-arrestin 2 has been recognized as a critical mediator of opioidinduced respiratory depression based on previous studies. Its role is now questioned, as β -arrestin 2 knockout does not affect respiratory rhythms.⁶⁰⁶ Otherwise, GABA is a negative regulator of hyperalgesia, but its depression comorbidity-inducing effects⁴⁷³ are often overlooked. Therefore, the lack of studies related to causal and mechanistic data increases difficulties in identifying more valuable targets.

The experimental methods require significant improvement. The reliance on mouse and rat models for human disease studies has been long debated due to substantial differences in nervous systems.⁷³⁶ This species gap may introduce biases in clinical translation. Additionally, gene editing and controlling certain substance levels in in-vivo models are relatively challenging compared to in-vitro models, slowing progress in exploring mechanisms underlying pain modulation. As mentioned previously, pain-related disorders are multifaceted. A single modeling approach cannot fully simulate human pain sensation, further affecting the credibility of current research. Fortunately, scientists are addressing this by developing composite modeling strategies. Moreover, commonly used pain indicators, such as c-fos expression, ion channel activity and hormone levels, may not accurately reflect pain severity in experimental animals. The representativeness of these indicators warrants scrutiny and validation. The above problems, of course, are universal flaws of basic experiments and are not unique to pain research.

The potential of multi-omics and high-resolution approaches remain underexploited. The authors only find that microbiome combined with metabolomics is widely used in the existing studies on microbiota and pain. While the findings have deepened our understanding and advanced pain diagnosis and treatment, it risks forming research stereotypes of the multiomics pattern of microbiome plus metabolomics. Furthermore, several recent studies analyzed basic structures and mechanisms underlying responses to nociception using single-cell transcriptome. However, other omics approaches, like transcriptomes, proteomics and spatial omics, are overlooked, especially the combinational application. The multi-omics approaches and high-resolution have led to significant discoveries in other nervous system diseases, such as Alzheimer's disease, depression and autism spectrum disorder.^{737–739} By contrast, few pioneer investigations have preliminarily shown the potential of transcriptomes and proteomics in exploring pain modulation,^{740,741} implying that pain research lags due to underutilization of multi-omics analysis. On the other hand, the efficiencies of omics data analysis integration are relatively low. The depth of omics data analysis is limited, especially for the basic research for investigating pain causality. In summary, the small data pool and rough analytical tools of omics both hamper the clinical progress in pain relief.

Data on pain modulation mechanisms and therapeutic regimens often present contradictions. The prime example is that the performances of specific therapeutic approaches are different across clinical trials, particularly in alternative and complementary therapies for pain management. Such discrepancies, which are influenced by numerous unpredictable confounding factors, are common in clinical research. However, similar inconsistencies are also observed in preclinical research. For instance, 17B-estradiol is reported as both a promoter of pain in females and a protective agent against hyperalgesia.⁷⁴² Moreover, the changes in expression of HDACs in response to pain modeling are controversial.^{213,743} These opposing conclusions without reasonable explanations create barriers to deeper studies. Thus, monism is unsuitable for estimating the roles of pain regulators, which may function distinctly under various conditions.

The progress in clinical trials for pain management is slow. The scarcity of large-sample, multicenter clinical trials hamper clinical translation of novel pain management approaches.

Table 3.	The preclinical researc	The preclinical research progress in novel monoclonal antib	tibodies for pain relief		
Target	Antibody	Disease	Experimental model	Effect	Ref.
Nav1.7	SVmab	Paclitaxel-induced neuropathic pain	In-vitro HEK293 and human DRG neurons	SVmab blocks Na^+ currents and neuronal excitability.	723
P2X3	12D4	Inflammation and visceral hypersensitivity	HEK-293 cell line and primary DRG neurons in vitro and female rats treated with formalin, CFA and TNBS in vivo	12D4 binds to inactivated state of P2X3 and significantly mitigates visceral pain.	764
P2X4R	scFv95	Trigeminal neuralgia	Constriction of the trigeminal infraorbital nerve in male and female BALB/c mice	scFv95 completely reverses the neuropathic pain only in the males.	765
BDNF	R3bH01	Peripheral nerve injury	SNI and tibial nerve transection in male SD rats	R3bH01 inhibits hyperactivity to heat and mechanical stimuli of sensory neurons in a dose-dependent manner.	766
HMGB1	anti-HMGB1 nAb	Trigeminal neuralgia	Constriction of distal infraorbital nerve in male ddY mice	Anti-HMGB1 nAb mitigates neuropathic pain through blocking activation of immune cells.	720
TrkA	MNAC13	Inflammation and nerve injury	Treatment with formalin and CCI in male CD1 mice	MNAC13 alleviates hyperalgesia and has synergistic effects in combination with opioids.	721
PAR-2	PAR650097	Migraine	Injection of CGRP or SLIGRL in female C57BL/6 J mice	PAR650097 prevents pain perception of both CGRP-dependent and independent migraine.	722
PD-L1	Nivolumab	Bone cancer pain	Inoculation of lung cancer cells into male and female C57BL/6 J mice	Nivolumab suppresses bone cancer pain by suppressing osteoclastogenesis.	767
RAGE	11E6	Inflammation and nerve injury	CFA stimulation or CCI in male C57BL/6.1 mice	11E6 decreases mechanical hypersensitivity in a dose-dependent manner.	768
CD11D	anti-CD11d mAb	Spinal cord injury	The laminectomy at T11 in male Wistar rats	No significant efficacies of anti-CD11d mAb are observed.	769
GlyR _α 3	FAbs 9A11, 19C8 and 14E3		HEK293 cell line and male SD rats	Selective inhibition of GlyR α 3 may have potentials in treating hyperalgesia and has good safety in vivo.	561
64WW	MMP9 mAb	Paclitaxel-induced pain	Administration of paclitaxel in male and female CD1 mice	MMP9 mAb alleviates paclitaxel-induced pain through inhibiting inflammatory responses to oxidative stress and loss of intraepidermal nerve fibers.	770
TRPM8	ACC-049 (polyclonal antibody)		Primary DRG neurons derived from male SD rats	The effects of ACC-049 on TRPM8 blockade are verified.	1.22
VEGFR-1	D16F7	Chemotherapy-induced pain (oxaliplatin, paclitaxel and vincristine)	Administration of chemotherapeutic drugs in male CD1 mice	D16F7 inhibits the development of chemotherapy-induced pain through interfering with astrocyte-neuron crosstalk.	772
RGMa	AE12-1 and AE12-1Y	Spinal cord injury	Extradural impact-compression in female Wistar rats	AE12-1 and AE12-1Y enhance neuronal survival capability, plasticity of descending serotonergic pathways and axonal regeneration. The activation of microglia and CGRP expression are also inhibited, alleviating pain perception.	773
OxPL	E06	Inflammation	CFA treatment in male Wistar rats and female Lewis rats	E06 suppresses hyperalgesia and CGRP release by blocking TRPV1 and TRPA1.	774

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CCI chronic constriction injury, CFA complete Freund's adjuvant, DRG dorsal root ganglion, SD Sprague-Dawley, TNBS trinitro-benzene-sulfonic acid

Limited patient inclusion reduces the practicality of subgroup analysis in identifying potential beneficiaries. Long-shot clinical investigations for high-level evidence-based support are lacking. Moreover, potential bias of publication, favoring studies with positive results, is a serious yet neglected problem. The clinical trials that display negative data or vigilance about the side effects have more difficulties in gaining extensive attention. Although this phenomenon has been improved to some extent recently, the follow-up impact following the previous phenomenon will persistently exist, probably causing more waste of basic and clinical resources.

Some mechanisms remain underexplored, especially in burgeoning research areas. For instance, current studies primarily focus on a few PTMs, including phosphorylation, ubiguitination, SUMOylation and glycosylation. This does not mean the unimportant roles of other PTMs. Instead, sporadic studies have reported their potentials in regulating nociceptor sensitivity and molecular activity, implying the unidentified PTM networks in pain modulation. Similarly, NLRP3 inflammasome is undisputedly a key mediator in hyperalgesia. This research trend may make other inflammasomes with properties in pain modulation, like NLRP2, ignorable. Otherwise, in the research on microbiota and pain, nonbacterial microbiota, like fungi, which regulate pain sensation, receive minimal attention. Basic knowledge about pain and biofilms, an important accessory structure of microbial community, is also limited. The sluggish paces with research frontiers may result in missing out on many diagnostic and therapeutic methods.

FUTURE PERSPECTIVES OF THE PAIN RESEARCH

Improvement in human-based in-vitro systems as research models. To surpass the inherent limitations of experimental animals as in-vivo models and in-vitro two-dimensional cells models (cell lines and induced pluripotent stem cells), the utilization of organoid and organs-on-a-chip technologies is essential in the follow-up studies. Organoids are the threedimensional culture systems derived from self-organizing stem cells. Organs-on-a-chip systems are in-vitro microfluidic devices containing the cell types of interest in close recapitulation of the original tissue structure, function, and physiology. The advent of these two in-vitro culture biotechnologies provides more opportunities for narrowing the gap in mechanistic insights into pain. Organoids and organs-on-a-chip systems possess both the maneuverability of in-vitro cell line models and integrity of in-vivo animal models. They have been widely adopted in various research areas, establishing a robust foundation for data production. Nevertheless, few studies on pain perception chose 31

them as experimental models, likely due to technological and financial barriers. The application of new biotechnologies always requires powerful supports of experimental technique and funding. To this end, just as developing trends of other biotechnologies, like high-throughput sequencing and antibody preparation, extensive commercialization of organoids and organs-on-a-chip systems may sharply reduce their application thresholds, fully unlocking research potential. Moreover, improvement in extracellular matrix structures and functions is important likewise. Three-dimensional bioprinting, microfluidics device and biomaterials should be further developed to generate organs with fine and complex structures incorporated with vasculature and innervation networks. Furthermore, crvoelectron microscopy (cryo-EM) is an emerging technique for analyzing molecular structures, providing in-depth evidences for investigating mechanisms and exploring new analgesic drugs. We are glad to see that cryo-EM has been extensively used in pain research (Table 4). More studies are required to illustrate functions of critical regulators in pain modulation.

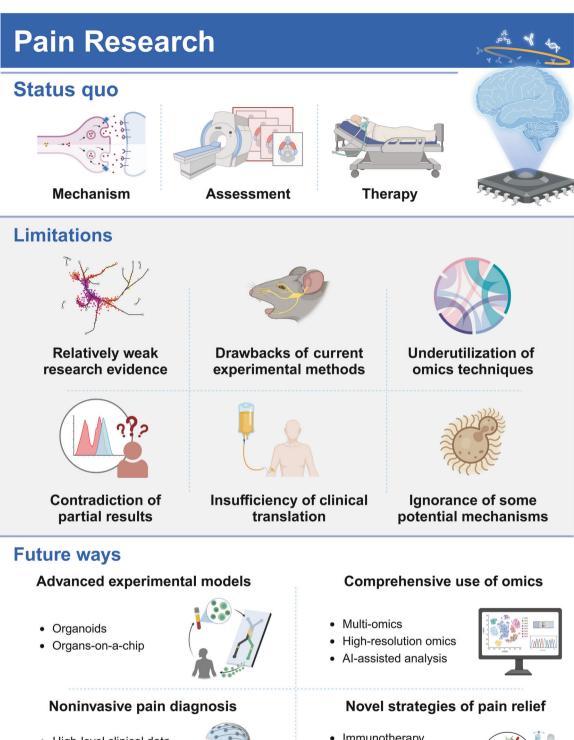
Comprehensive use of omics profiling. Single-cell and spatial omics technologies have become landmark achievements in the technological revolution. Pain research has begun employing single-cell and spatial omics to unravel new mechanisms, such as compiling DRG atlas, identifying mechanistic networks and discovering new cell subtypes.⁷⁴⁴ More focus is needed in areas like analgesic therapy responses, opioid tolerance and pain sensation variations among populations. Furthermore, the integration of multi-omics detection with these high-resolution technologies is absent but greatly needed. The joint analysis of microbiome and metabolomics has already laid a foundation. Expanding omics types and data volumes should be further encouraged. The rapid advancement of AI offers robust data analysis capabilities for multi-omics technologies.745 Future studies should try to widen the scope of multi-omics profiling and deepen omics data mining. Recently, the most comprehensive human brain cell map has been disclosed. This work set a benchmark for future omics studies. Although introduction of new omics approaches may disclose more unexplored zones and put more demand on researchers, it will tremendously broaden the horizons concerning pain modulation. The profits in better understanding mechanisms and identifying more related targets can boost the development of pain diagnosis and treatment.

Development of noninvasive methods for pain diagnosis. Due to the subjectivity and heterogeneity of pain perception, clinical pain evaluation typically depends on scales and assessment by doctors, which are not always reliable. Novel approaches like gene biomarkers and brain imaging have yet

Table 4.	The research progress in structure analysis of pain-associated	molecules using cryo-EM	
Molecule	Aim	Significance	Ref.
NaV1.3	To investigate the mechanisms by which bulleyaconitine and a selective inhibitor ICA121431 block Nav1.3.	Bulleyaconitine resides around domain I-II fenestration and blocks ion channels. ICA121431 stabilizes the activation state of Nav1.3 through preferentially binding to the domain IV voltage-sensor.	775
Nav1.7	To investigate the structures of Nav1.7 bound with riluzole and lamotrigine.	Riluzole binds to the central cavity of Nav1.7 pore, whereas lamotrigine has dual-pocket mechanisms. In addition to the above site, lamotrigine additionally resides in the beneath intracellular gate, synergistically inhibiting Nav1.7 activity.	776
Nav1.7	To discriminate the structural basis of various Nav1.7 antagonists.	Carbamazepine, bupivacaine and lacosamide reside in a binding site beneath the intracellular gate. Lacosamide closely binds to the selectivity filter. Vinpocetine plugs in the III-IV fenestrations, whereas vixotrigine can penetrate the IV-I fenestrations.	777

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Molecule	Aim	Significance	Re
Nav1.7	To investigate the structural basis of Nav1.7 mutants.	The first voltage-sensing domain exhibits inactivation conformation, which is attributable to transference of three gating charge residues into the cytosolic side, leading to intracellular gate rearrangement and contraction. The data provide proofs for understanding pain- related mutations.	778
Cav1.2	To identify Cav1.2 structures and the structural mechanisms of the calcium channel blockers.	Pinaverium bromide, an antispasmodic drug, is inserted into IFM- binding site and induces global alterations of Cav1.2, while calciseptine binds to the shoulder of the pore domain.	779
Cav2.2	To elucidate the inactivation state of Cav2.2.	S6 helices in the second voltage-sensing domain are essential for maintaining inactivation, and a W-helix from the domain II–III linker further stabilizes the close state of Cav2.2. This finding may contribute to the development of novel Cav2.2 blockers.	78
Cav2.3	To explore the structures of Cav2.3 in complex with $\alpha 2\delta$ -1 and $\beta 3$ subunits, together with the differences with a Cav2.3 mutant.	Cav2.3 has the high similarity in cellular structures with Cav2.2. CH2II helix is deleted in Cav2.3 mutant, which is critical for stabilizing the inactivated conformation.	78
Cav3.1	To show the structures of Cav3.1 alone and in complex with Z944, a selective blocker of Cav3.1.	The structural mechanisms of Z944 residing in the central cavity of the pore domain of Cav3.1 are revealed, deepening the understanding in distinct channel properties of VGCCs.	78
KCNQ4	To explore the structures of KCNQ4 and in complex with an opener retigabine or an inhibitor linopirdine.	Signaling factor PIP2 is inserted into the cavity of voltage-sensing domain for channel activity modulation. The fenestration and cytosolic cavity are the binding regions of retigabine and linopirdine, respectively. The data lay foundations for developing new drugs targeting KCNQ4.	78
TRPV3	To display the structure of TRPV3-dyclonine complex.	Dyclonine sticks into the channel pore and blocks the connection with membrane environments.	78
TRPV3	To identify the opening structure of TRPV3.	The structural changes in both the outer pore and the intracellular gate occur in channel opening. S6 helices in the gate are further elongated and rotated.	78
CB1, CB2	To identify the structures of CB1 and CB2 in complex with Gi.	The structures of Gi complexes with CB1 or CB2 are obtained, which provide proofs for clarifying their selectivity. Cholesterol is identified as an endogenous allosteric modulator of CB1.	78
CB2	To investigate the mechanisms underlying activation and signaling of CB2.	The structures of CB2- G_i complex and interactions with potent agonist WIN 55,212-2 are identified. Moreover, a unique motif in CB2 is found to underlie its specificity of G_i coupling.	78
GABA _A	To explore the structural basis of $\alpha1\beta3\gamma2L$, a major isoform of $GABA_A$ receptor.	Intracellular M3-M4 loops in $\alpha 1\beta 3\gamma 2L$ are vulnerable to physiological and pharmacological regulation, which is an important reason of heteromeric organization of the GABA _A receptor.	78
GABA _A	To explore the principles of pentamer formation of $GABA_A$ receptor.	The permutational landscape of GABA _A receptor is depicted based on the theory of differential assembly. GABA _A arrangements are found to meanwhile respond to GABA and histamine.	78
GABA _B	To screen the ligands pre-associating with the $GABA_{B}$ receptor.	Transmembrane domains are the key sites of ligands binding to the GABA _B receptor. A heterodimer interface between helices 3 and 5 is found to be essential for GABA _B inactivation.	79
GlyR	To clarify the mechanisms by which THC, the principal psychoactive component in cannabis sativa, modulates GlyR activity.	THC activates GlyR through directly regulating the opening states of the channel pore.	79
mGlu2	To elucidate the structures of mGlu2 homodimers and complexes with heterotrimeric ${\rm G}_{\rm i}.$	An asymmetric TM6-TM6 interface is critical for the extensive rearrangement of transmembrane domains. The mechanisms of family A and B GPCRs rely on the TM6-TM6 interface, while their family C G protein coupling functions in a TM6-independent manner.	79
MRGPRX1	To explore the mechanisms underlying the activation of MRGPRX1, a molecule mediating pain perception.	The three structures of MRGPRX1 in complex with opioid peptide fragment BAM8-22, BAM8-22 plus positive allosteric modulator ML382 and a synthetic agonist compound-16 are revealed. The data provide evidence for developing novel non-opioid drugs.	79
A ₁ R	To clarify the mechanisms underlying MIPS521, a positive allosteric modulator of A1R.	A_1R binds to a heterotrimer of adenosine, MIPS521 and G_{i2} via an extrahelical lipid-detergent-facing allosteric binding pocket involving helixes 1, 6 and 7. The data provide new evidence for developing allosteric drugs.	79
P2X4	To identify the structures of P2X4 in complex with specific antagonists BX430 and BAY-1797.	BX430 and BAY-1797 can both bind to the allosteric site at the subunit interface of the P2X4 extracellular domain.	79



- High-level clinical data
- · Rapid targeted diagnosis
- Streamlining markers
- · Multimodal diagnosis

- Immunotherapy
- **Biomaterials**
- Learn from nature
- Comprehensive strategy



Fig. 10 The status quo, limitations and future perspectives of the pain research. Most current preclinical and clinical studies in the pain field focused on its mechanisms, assessment and therapy. However, there are some limitations as follows. i) Current research evidence is relatively weak and the underlying mechanisms remain largely unknown. ii) The in-vitro and in-vivo experimental models cannot thoroughly mimic the clinical conditions of pain. iii) The research value of omics techniques is not fully exploited. iv) There are contradictions in the results from some studies. v) Current progress in clinical translation of pain research achievements is far from clinical requirements. vi) Some potential mechanisms underlying pain modulation should be emphasized, like the regulatory role of fungi. Herein, we propose four future perspectives for pain research, including development of advanced experimental models, comprehensive application of omics, emphasis on noninvasive pain diagnosis and optimization of strategies for pain relief

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to effectively address these limitations. Based on the fact, the future directions can be concluded into four aspects. i) Collection of more high-level evidence-based medical data. Since existing data have shown their optimal performance, trials with larger samples and subsequent commercialization processes should be advocated. This measure may allow patients to benefit from these achievements at an earlier time. The differences of pain perception in various subpopulations can be better excavated. ii) Rapid diagnosis based on molecular targets and easily accessible samples. Current approaches have difficulties in balancing efficiency and accuracy. Novel bioengineering techniques are favorable for overcoming this dilemma. Lin and colleagues set a good example. They develop a nanochip detecting saliva CGRP concentrations for migraine diagnosis. The results can be obtained within 10 min.746 More investigations into rapid diagnosis need to be encouraged. iii) Streamlining detected indicators. Some studies used ten more biomarkers or global high-throughput data to train pain models. Despite theoretically good performances, they may not conform to clinical reality due to their high cost and complexity. Herein, the authors call for investigations into applications of smaller-scale indicators. iv) Multimodal diagnostic methods. Although we encourage decreases in included indicators of one diagnostic methods, the advantages of multimodal approaches should be highlighted. Multimodal approaches can mutually compensate for shortcomings of single method, like BCI combined with skin conductance.⁶⁸¹ The integration of AI in multimodal approaches could offer the most efficient ways to meet realistic conditions.

Extension of novel strategies for pain management. i) Activation of immunoreaction against pain. Immune disorders significantly contribute to hyperalgesia, and most approaches pertain to the passively mitigation. The success of cancer immunotherapy hints that the immune system may be another key to pain relief. For instance, Sara et al. creatively employed microbiota antigens to activate specific immunity, obliterating visceral hypersensitivityassociated microbiota.747 This research is opening new possibilities in this field. On the basis of the reported advantages of immunotherapy, responders are likely to obtain more benefits from it, embodied by lasting medical effects, mild side effects and good tolerance.⁷⁴⁸ ii) Leveraging biomaterial superiority. Interdisciplinary investigation is a promising approach to promote translation, mainly including efficacy enhancement, potency prolongation, side effect mitigation and cost minimization. Despite its promise, collaborations involving biomaterials remain scarce. The need for more high-quality studies is evident. iii) Formation of comprehensive treatment strategies. The characteristics of pain, as a multifactorial disease, necessitates multifaceted interventions, including, analgesics, emotional management, social relations improvement, together with alternative and complementary therapies. Research is shifting towards examining the synergistic effects of combined therapies, such as analgesic 49,750 More drugs with CBT and cell stem therapy with probiotics.⁷ research investment should be conducted to make one plus one larger than two. iv) Learning from nature. Many breakthroughs in pain research are inspired by natural phenomena. Certain natural compounds are found to have analgesic efficacy, potentially more effective than synthetic drugs.^{751,752} The natural compounds and endogenous substances, such as melatonin, curcumin and peppermint, with a history of safe use, offer wide regulatory mechanisms. The advantages can effectively decrease risks associated with new drug development.

CONCLUSIONS

Pain, a kind of universal experience, brings about overwhelming physical and mental distress. Attenuating pain is a fundamental

right of patients. Extensive preclinical and clinical studies have delved into pain pathology and the molecular mechanisms of pain modulation. Key brain regions and critical molecules have been identified involved in pain perception. The research achievements regarding TRPV1, TRPM8 and Piezo even garner recognition of the Nobel prize. The significant progress has facilitated a shift from empirical management to personalized interventions. A range of promising diagnostic and therapeutic approaches have emerged. Some Chinese traditional therapies are rehabilitated by modern medical evidence. Multidisciplinary collaborations have further advanced the field and integrated cutting-edge technologies, such as virtual reality, biomaterials, and high-resolution omics technologies. However, current research limitations are impeding further advancements in pain management. This review suggests several potential directions for future research (Fig. 10). The authors believe that the unpleasure experience of pain will be eradicated as neuroscience continues to evolve.

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AUTHOR CONTRIBUTIONS

B.W., Y.W. and F.L. conceptualized and designed the idea. B.C., Q.X., Y.S., R.Z. and H.L. performed the literature search and finished the draft. B.C., Q.X., Y.S., R.Z., H.L. and J.Z. drew the figures. B.W., Y.W., F.L. and J.Z. revised the manuscript for intellectual contents. All authors have read and approved this review.

ADDITIONAL INFORMATION

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REFERENCES

- Raja, S. N. et al. The revised International Association for the Study of Pain definition of pain: concepts, challenges, and compromises. *Pain* 161, 1976–1982 (2020).
- Basbaum, A. I., Bautista, D. M., Scherrer, G. & Julius, D. Cellular and molecular mechanisms of pain. *Cell* 139, 267–284 (2009).
- Julius, D. & Basbaum, A. I. Molecular mechanisms of nociception. *Nature* 413, 203–210 (2001).
- Yang, D. et al. Nociceptor neurons direct goblet cells via a CGRP-RAMP1 axis to drive mucus production and gut barrier protection. *Cell* 185, 4190–4205.e4125 (2022).
- Goodwin, G. & McMahon, S. B. The physiological function of different voltagegated sodium channels in pain. *Nat. Rev. Neurosci.* 22, 263–274 (2021).
- Steglitz, J., Buscemi, J. & Ferguson, M. J. The future of pain research, education, and treatment: a summary of the IOM report "Relieving pain in America: a blueprint for transforming prevention, care, education, and research". *Transl. Behav. Med* 2, 6–8 (2012).
- Dahlhamer, J. et al. Prevalence of chronic pain and high-impact chronic pain among adults—United States, 2016. *MMWR Morb. Mortal. Wkly Rep.* 67, 1001–1006 (2018).
- Fayaz, A., Croft, P., Langford, R. M., Donaldson, L. J. & Jones, G. T. Prevalence of chronic pain in the UK: a systematic review and meta-analysis of population studies. *BMJ Open* 6, e010364 (2016).
- Goldberg, D. S. & McGee, S. J. Pain as a global public health priority. BMC Public Health 11, 770 (2011).
- Morasco, B. J. et al. Systematic review of prevalence, correlates, and treatment outcomes for chronic non-cancer pain in patients with comorbid substance use disorder. *Pain* **152**, 488–497 (2011).
- 11. Vieira, E. B. et al. Chronic pain, associated factors, and impact on daily life: are there differences between the sexes? *Cad. Saude Publica* **28**, 1459–1467 (2012).
- Tang, N. K. & Crane, C. Suicidality in chronic pain: a review of the prevalence, risk factors and psychological links. *Psychol. Med.* 36, 575–586 (2006).

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- Quinten, C. et al. Baseline quality of life as a prognostic indicator of survival: a meta-analysis of individual patient data from EORTC clinical trials. *Lancet Oncol.* 10, 865–871 (2009).
- Moore, R. A., Derry, S., Aldington, D. & Wiffen, P. J. Adverse events associated with single dose oral analgesics for acute postoperative pain in adults - an overview of Cochrane reviews. *Cochrane Database Syst. Rev.* 2015, Cd011407 (2015).
- Zafeiri, A., Mitchell, R. T., Hay, D. C. & Fowler, P. A. Over-the-counter analgesics during pregnancy: a comprehensive review of global prevalence and offspring safety. *Hum. Reprod. Update* 27, 67–95 (2021).
- Melzack, R. & Wall, P. D. Pain mechanisms: a new theory. *Science* **150**, 971–979 (1965).
- Wall, P. D. & Sweet, W. H. Temporary abolition of pain in man. *Science* 155, 108–109 (1967).
- Lamotte, C., Pert, C. B. & Snyder, S. H. Opiate receptor binding in primate spinal cord: distribution and changes after dorsal root section. *Brain Res.* **112**, 407–412 (1976).
- Yaksh, T. L. & Rudy, T. A. Analgesia mediated by a direct spinal action of narcotics. *Science* **192**, 1357–1358 (1976).
- Mathew, B., Norris, D., Hendry, D. & Waddell, G. Artificial intelligence in the diagnosis of low-back pain and sciatica. *Spine* 13, 168–172 (1988).
- Edelman, R. R. et al. Qualitative mapping of cerebral blood flow and functional localization with echo-planar MR imaging and signal targeting with alternating radio frequency. *Radiology* **192**, 513–520 (1994).
- Karppinen, J. et al. Tumor necrosis factor-alpha monoclonal antibody, infliximab, used to manage severe sciatica. *Spine* 28, 750–753 (2003).
- 23. Fink, D. J. et al. Gene therapy for pain: results of a phase I clinical trial. Ann. Neurol. **70**, 207–212 (2011).
- Chambers, S. M. et al. Combined small-molecule inhibition accelerates developmental timing and converts human pluripotent stem cells into nociceptors. *Nat. Biotechnol.* **30**, 715–720 (2012).
- Chiu, I. M. et al. Bacteria activate sensory neurons that modulate pain and inflammation. *Nature* 501, 52–57 (2013).
- Cai, H. et al. Engineering human spinal microphysiological systems to model opioid-induced tolerance. *Bioact. Mater.* 22, 482–490 (2023).
- Souza Monteiro de Araujo, D., Nassini, R., Geppetti, P. & De Logu, F. TRPA1 as a therapeutic target for nociceptive pain. *Expert Opin. Ther. Targets* 24, 997–1008 (2020).
- Torrance, N., Smith, B. H., Bennett, M. I. & Lee, A. J. The epidemiology of chronic pain of predominantly neuropathic origin. Results from a general population survey. J. Pain. 7, 281–289 (2006).
- Bouhassira, D., Lantéri-Minet, M., Attal, N., Laurent, B. & Touboul, C. Prevalence of chronic pain with neuropathic characteristics in the general population. *Pain* 136, 380–387 (2008).
- Dydyk, A. M. & Givler, A. Dydyk, A. M. & Givler, A. Central Pain Syndrome (StatPearls Publishing LLC., 2022).
- Treede, R. D. et al. Chronic pain as a symptom or a disease: the IASP classification of chronic pain for the International Classification of Diseases (ICD-11). *Pain* 160, 19–27 (2019).
- Hausteiner-Wiehle, C. & Henningsen, P. Nociplastic pain is functional pain. Lancet 399, 1603–1604 (2022).
- Li, Y., Dorsi, M. J., Meyer, R. A. & Belzberg, A. J. Mechanical hyperalgesia after an L5 spinal nerve lesion in the rat is not dependent on input from injured nerve fibers. *Pain* 85, 493–502 (2000).
- Minert, A., Gabay, E., Dominguez, C., Wiesenfeld-Hallin, Z. & Devor, M. Spontaneous pain following spinal nerve injury in mice. *Exp. Neurol.* 206, 220–230 (2007).
- Howard, R. F., Walker, S. M., Mota, M. P. & Fitzgerald, M. The ontogeny of neuropathic pain: postnatal onset of mechanical allodynia in rat spared nerve injury (SNI) and chronic constriction injury (CCI) models. *Pain* **115**, 382–389 (2005).
- 36. Liu, P. et al. The delayed-onset mechanical pain behavior induced by infant peripheral nerve injury is accompanied by sympathetic sprouting in the dorsal root ganglion. *Biomed. Res. Int.* **2020**, 9165475 (2020).
- Sadler, K. E., Mogil, J. S. & Stucky, C. L. Innovations and advances in modelling and measuring pain in animals. *Nat. Rev. Neurosci.* 23, 70–85 (2022).
- Gaudet, A. D., Fonken, L. K., Ayala, M. T., Maier, S. F. & Watkins, L. R. Aging and miR-155 in mice influence survival and neuropathic pain after spinal cord injury. *Brain Behav. Immun.* 97, 365–370 (2021).
- Lin, R. et al. Immunohistochemical analysis of histone H3 acetylation in the trigeminal root entry zone in an animal model of trigeminal neuralgia. J. Neurosurg. 131, 828–838 (2018).
- Sun, X. et al. The NLRP3-related inflammasome modulates pain behavior in a rat model of trigeminal neuropathic pain. *Life Sci.* 277, 119489 (2021).

- 41. Yang, L. et al. Electroacupuncture attenuates surgical pain-induced delirium-like behavior in mice via remodeling gut microbiota and dendritic spine. *Front Immunol.* **13**, 955581 (2022).
- Katsuda, Y. et al. Histone modification of pain-related gene expression in spinal cord neurons under a persistent postsurgical pain-like state by electrocautery. *Mol. Brain* 14, 146 (2021).
- Ilfeld, B. M. et al. Percutaneous peripheral nerve stimulation (Neuromodulation) for postoperative pain: a randomized, sham-controlled pilot study. *Anesthesiol*ogy 135, 95–110 (2021).
- Bas, D. B., Su, J., Wigerblad, G. & Svensson, C. I. Pain in rheumatoid arthritis: models and mechanisms. *Pain. Manag.* 6, 265–284 (2016).
- Araya, E. I. et al. Acute orofacial pain leads to prolonged changes in behavioral and affective pain components. *Pain* 161, 2830–2840 (2020).
- Pepino, L., Malapert, P., Saurin, A. J., Moqrich, A. & Reynders, A. Formalin-evoked pain triggers sex-specific behavior and spinal immune response. *Sci. Rep.* 13, 9515 (2023).
- Kodji, X. et al. Evidence that a TRPA1-mediated murine model of temporomandibular joint pain involves NLRP3 inflammasome activation. *Pharmacology* 14, 1073 (2021).
- Sawynok, J., Reid, A. & Meisner, J. Pain behaviors produced by capsaicin: influence of inflammatory mediators and nerve injury. J. Pain. 7, 134–141 (2006).
- Wirtz, S. et al. Chemically induced mouse models of acute and chronic intestinal inflammation. *Nat. Protoc.* 12, 1295–1309 (2017).
- Xie, Z. et al. Piezo2 channels expressed by colon-innervating TRPV1-lineage neurons mediate visceral mechanical hypersensitivity. *Neuron* 111, 526–538.e524 (2023).
- Fei, L. & Wang, Y. microRNA-495 reduces visceral sensitivity in mice with diarrhea-predominant irritable bowel syndrome through suppression of the PI3K/AKT signaling pathway via PKIB. *IUBMB Life* **72**, 1468–1480 (2020).
- Khan, A. et al. Anti-neuropathic pain activity of Ajugarin-I via activation of Nrf2 signaling and inhibition of TRPV1/TRPM8 nociceptors in STZ-induced diabetic neuropathy. *Pharm. Res.* 183, 106392 (2022).
- Agarwal, N. et al. Evoked hypoalgesia is accompanied by tonic pain and immune cell infiltration in the dorsal root ganglia at late stages of diabetic neuropathy in mice. *Mol. Pain.* 14, 1744806918817975 (2018).
- Chen, H. et al. IL-17 crosses the blood-brain barrier to trigger neuroinflammation: a novel mechanism in nitroglycerin-induced chronic migraine. *J. Headache Pain* 23, 1 (2022).
- Oshinsky, M. L. & Gomonchareonsiri, S. Episodic dural stimulation in awake rats: a model for recurrent headache. *Headache* 47, 1026–1036 (2007).
- Yin, C. et al. IL-33/ST2 induces neutrophil-dependent reactive oxygen species production and mediates gout pain. *Theranostics* 10, 12189–12203 (2020).
- Rossato, M. F. et al. Monosodium urate crystal interleukin-1β release is dependent on Toll-like receptor 4 and transient receptor potential V1 activation. *Rheumatology* 59, 233–242 (2020).
- Schwei, M. J. et al. Neurochemical and cellular reorganization of the spinal cord in a murine model of bone cancer pain. J. Neurosci. 19, 10886–10897 (1999).
- 59. Ding, Z. et al. Inhibition of spinal ferroptosis-like cell death alleviates hyperalgesia and spontaneous pain in a mouse model of bone cancer pain. *Redox Biol.* **62**, 102700 (2023).
- Wei, W. et al. A compound mitigates cancer pain and chemotherapy-induced neuropathic pain by dually targeting nNOS-PSD-95 interaction and GABA(A) receptor. *Neurotherapeutics* 18, 2436–2448 (2021).
- Green, P. G., Alvarez, P. & Levine, J. D. A role for gut microbiota in early-life stress-induced widespread muscle pain in the adult rat. *Mol. Pain* 17, 17448069211022952 (2021).
- Nishi, M. Effects of early-life stress on the brain and behaviors: implications of early maternal separation in rodents. *Int. J. Mol. Sci.* 21, 7212 (2020).
- Wang, Y. et al. TRPV1 SUMOylation regulates nociceptive signaling in models of inflammatory pain. *Nat. Commun.* 9, 1529 (2018).
- Courties, A. et al. Human-specific duplicate CHRFAM7A gene is associated with more severe osteoarthritis and amplifies pain behaviours. *Ann. Rheum. Dis.* 82, 710–718 (2023).
- Ma, J. et al. The comprehensive pathophysiological changes in a novel rat model of postinflammatory visceral hypersensitivity. FASEB J. 33, 13560–13571 (2019).
- Hasegawa, R., Saito-Nakaya, K., Gu, L., Kanazawa, M. & Fukudo, S. Maternal separation and TNBS-induced gut inflammation synergistically alter the sexually differentiated stress response in rats. *Biopsychosoc. Med.* 17, 7 (2023).
- 67. Kozlova, E. V. et al. Induction of distinct neuroinflammatory markers and gut dysbiosis by differential pyridostigmine bromide dosing in a chronic mouse model of GWI showing persistent exercise fatigue and cognitive impairment. *Life Sci.* 288, 120153 (2022).
- Touska, F. et al. Heat-resistant action potentials require TTX-resistant sodium channels Na(V)1.8 and Na(V)1.9. J. Gen. Physiol. 150, 1125–1144 (2018).

- 36
- Moreno, A. M. et al. Long-lasting analgesia via targeted in situ repression of Na(V)1.7 in mice. Sci. Transl. Med. 13, eaay9056 (2021).
- Waxman, S. G. Painful Na-channelopathies: an expanding universe. *Trends Mol. Med.* 19, 406–409 (2013).
- Deng, L. et al. Nav1.7 is essential for nociceptor action potentials in the mouse in a manner independent of endogenous opioids. *Neuron* **111**, 2642–2659.e2613 (2023).
- Minett, M. S. et al. Pain without nociceptors? Nav1.7-independent pain mechanisms. *Cell Rep.* 6, 301–312 (2014).
- Osteen, J. D. et al. Selective spider toxins reveal a role for the Nav1.1 channel in mechanical pain. *Nature* 534, 494–499 (2016).
- 74. Chen, L. et al. Sodium channel Nav1.6 in sensory neurons contributes to vincristine-induced allodynia. *Brain* **143**, 2421–2436 (2020).
- 75. Zhang, J. et al. HDAC2 in primary sensory neurons constitutively restrains chronic pain by repressing α2δ-1 expression and associated NMDA receptor activity. J. Neurosci. 42, 8918–8935 (2022).
- Davies, A. et al. Functional biology of the alpha(2)delta subunits of voltagegated calcium channels. *Trends Pharm. Sci.* 28, 220–228 (2007).
- Zhang, Y. et al. Adiponectin receptor 1-mediated stimulation of Cav3.2 channels in trigeminal ganglion neurons induces nociceptive behaviors in mice. J. Headache Pain 24, 117 (2023).
- 78. Zhang, Y. et al. Neuromedin B receptor stimulation of Cav3.2 T-type Ca(2+) channels in primary sensory neurons mediates peripheral pain hypersensitivity. *Theranostics* **11**, 9342–9357 (2021).
- Gandla, J., Lomada, S. K., Lu, J., Kuner, R. & Bali, K. K. miR-34c-5p functions as pronociceptive microRNA in cancer pain by targeting Cav2.3 containing calcium channels. *Pain* 158, 1765–1779 (2017).
- Dawes, J. M. et al. Immune or genetic-mediated disruption of CASPR2 causes pain hypersensitivity due to enhanced primary afferent excitability. *Neuron* 97, 806–822.e810 (2018).
- Higerd-Rusli, G. P. et al. Inflammation differentially controls transport of depolarizing Nav versus hyperpolarizing Kv channels to drive rat nociceptor activity. *Proc. Natl Acad. Sci. USA* **120**, e2215417120 (2023).
- Moran, M. M. TRP channels as potential drug targets. Annu. Rev. Pharm. Toxicol. 58, 309–330 (2018).
- Blake, K. J. et al. Staphylococcus aureus produces pain through pore-forming toxins and neuronal TRPV1 that is silenced by QX-314. *Nat. Commun.* 9, 37 (2018).
- De Logu, F. et al. Schwann cell endosome CGRP signals elicit periorbital mechanical allodynia in mice. *Nat. Commun.* 13, 646 (2022).
- 85. Yang, S. et al. A bimodal activation mechanism underlies scorpion toxin-induced pain. *Sci. Adv.* **3**, e1700810 (2017).
- Lin King, J. V. et al. A cell-penetrating scorpion toxin enables mode-specific modulation of TRPA1 and pain. *Cell* **178**, 1362–1374.e1316 (2019).
- Arendt-Nielsen, L. et al. A randomized, double-blind, positive-controlled, 3-way cross-over human experimental pain study of a TRPV1 antagonist (V116517) in healthy volunteers and comparison with preclinical profile. *Pain* **157**, 2057–2067 (2016).
- Heber, S. et al. TRPV1 antagonist BCTC inhibits pH 6.0-induced pain in human skin. *Pain* 161, 1532–1541 (2020).
- Lee, J. et al. Functional interactions between NMDA receptors and TRPV1 in trigeminal sensory neurons mediate mechanical hyperalgesia in the rat masseter muscle. *Pain* 153, 1514–1524 (2012).
- Deng, M., Chen, S. R., Chen, H. & Pan, H. L. α2δ-1-bound N-methyl-D-aspartate receptors mediate morphine-induced hyperalgesia and analgesic tolerance by potentiating glutamatergic input in rodents. *Anesthesiology* **130**, 804–819 (2019).
- Xie, R. G. et al. Presynaptic NMDARs on spinal nociceptor terminals statedependently modulate synaptic transmission and pain. *Nat. Commun.* 13, 728 (2022).
- Ru, Q. et al. TIAM1-mediated synaptic plasticity underlies comorbid depressionlike and ketamine antidepressant-like actions in chronic pain. J. Clin. Investig. 132, e158545 (2022).
- Deng, M. et al. Mitogen-activated protein kinase signaling mediates opioidinduced presynaptic NMDA receptor activation and analgesic tolerance. J. Neurochem. 148, 275–290 (2019).
- Dedek, A. et al. Loss of STEP61 couples disinhibition to N-methyl-d-aspartate receptor potentiation in rodent and human spinal pain processing. *Brain* 142, 1535–1546 (2019).
- Inoue, K. Nociceptive signaling of P2X receptors in chronic pain states. *Purinergic Signal* 17, 41–47 (2021).
- Abdelhamid, R. E. & Sluka, K. A. ASICs mediate pain and inflammation in musculoskeletal diseases. *Physiology* **30**, 449–459 (2015).
- Stephan, G. et al. The ASIC3/P2X3 cognate receptor is a pain-relevant and ligand-gated cationic channel. *Nat. Commun.* 9, 1354 (2018).

- Khan, A., Khan, S. & Kim, Y. S. Insight into pain modulation: nociceptors sensitization and therapeutic targets. *Curr. Drug Targets* 20, 775–788 (2019).
- 99. Wong, T. S. et al. G protein-coupled receptors in neurodegenerative diseases and psychiatric disorders. *Signal Transduct. Target Ther.* **8**, 177 (2023).
- Doyen, P. J., Beckers, P., Brook, G. A. & Hermans, E. Regulators of G protein signalling as pharmacological targets for the treatment of neuropathic pain. *Pharm. Res.* **160**, 105148 (2020).
- 101. Mayer, F., Gunawan, A. L., Tso, P. & Aponte, G. W. Glucagon-like peptide 1 and glucose-dependent insulinotropic polypeptide stimulate release of substance P from TRPV1- and TRPA1-expressing sensory nerves. Am. J. Physiol. Gastrointest. Liver Physiol. **319**, G23–G35 (2020).
- Giraud, F., Pereira, E., Anizon, F. & Moreau, P. Recent advances in pain management: relevant protein kinases and their inhibitors. *Molecules* 26, 2696 (2021).
- Tavares-Ferreira, D. et al. Spatial transcriptomics of dorsal root ganglia identifies molecular signatures of human nociceptors. *Sci. Transl. Med.* 14, eabj8186 (2022).
- De Logu, F. et al. Schwann cells expressing nociceptive channel TRPA1 orchestrate ethanol-evoked neuropathic pain in mice. J. Clin. Investig. 129, 5424–5441 (2019).
- McWilliams, K. & Fallon, M. Fast-acting fentanyl preparations and pain management. QJM 106, 887–890 (2013).
- Chen, L. & Michalsen, A. Management of chronic pain using complementary and integrative medicine. *BMJ* 357, j1284 (2017).
- 107. Sikandar, S. et al. Brain-derived neurotrophic factor derived from sensory neurons plays a critical role in chronic pain. *Brain* **141**, 1028–1039 (2018).
- Yue, L. et al. Brain-derived neurotrophic factor in the infralimbic cortex alleviates inflammatory pain. *Neurosci. Lett.* 655, 7–13 (2017).
- 109. Marvizón, J. C. et al. Calcitonin receptor-like receptor and receptor activity modifying protein 1 in the rat dorsal horn: localization in glutamatergic presynaptic terminals containing opioids and adrenergic alpha2C receptors. *Neuroscience* **148**, 250–265 (2007).
- Marquez de Prado, B., Hammond, D. L. & Russo, A. F. Genetic enhancement of calcitonin gene-related Peptide-induced central sensitization to mechanical stimuli in mice. J. Pain. 10, 992–1000 (2009).
- 111. Tanioku, T. et al. Tmem45b is essential for inflammation- and tissue injuryinduced mechanical pain hypersensitivity. *Proc. Natl Acad. Sci. USA* **119**, e2121989119 (2022).
- 112. Chu, J. et al. ATP-releasing SWELL1 channel in spinal microglia contributes to neuropathic pain. *Sci. Adv.* **9**, eade9931 (2023).
- 113. Hu, X. et al. A TRPV4-dependent neuroimmune axis in the spinal cord promotes neuropathic pain. J. Clin. Investig. **133**, e161507 (2023).
- 114. Caspi, Y. et al. Structural plasticity of axon initial segment in spinal cord neurons underlies inflammatory pain. *Pain* **164**, 1388–1401 (2023).
- Parisien, M. et al. Acute inflammatory response via neutrophil activation protects against the development of chronic pain. *Sci. Transl. Med.* 14, eabj9954 (2022).
- Caxaria, S. et al. Neutrophils infiltrate sensory ganglia and mediate chronic widespread pain in fibromyalgia. *Proc. Natl Acad. Sci. USA* **120**, e2211631120 (2023).
- Lim, H., Lee, H., Noh, K. & Lee, S. J. IKK/NF-kB-dependent satellite glia activation induces spinal cord microglia activation and neuropathic pain after nerve injury. *Pain* 158, 1666–1677 (2017).
- 118. Verkhratsky, A. et al. Astrocytes in human central nervous system diseases: a frontier for new therapies. *Signal Transduct. Target Ther.* **8**, 396 (2023).
- Prescott, S. A., Ma, Q. & De Koninck, Y. Normal and abnormal coding of somatosensory stimuli causing pain. *Nat. Neurosci.* 17, 183–191 (2014).
- Guo, K., Yamawaki, N., Barrett, J. M., Tapies, M. & Shepherd, G. M. G. Corticothalamo-cortical circuits of mouse forelimb S1 are organized primarily as recurrent loops. J. Neurosci. 40, 2849–2858 (2020).
- Ziegler, K. et al. Primary somatosensory cortex bidirectionally modulates sensory gain and nociceptive behavior in a layer-specific manner. *Nat. Commun.* 14, 2999 (2023).
- Hiraga, S. I. et al. Microglial depletion under thalamic hemorrhage ameliorates mechanical allodynia and suppresses aberrant axonal sprouting. *JCI Insight* 5, e131801 (2020).
- 123. Fanton, S. et al. Anti-satellite glia cell IgG antibodies in fibromyalgia patients are related to symptom severity and to metabolite concentrations in thalamus and rostral anterior cingulate cortex. *Brain Behav. Immun.* **114**, 371–382 (2023).
- 124. Corder, G. et al. An amygdalar neural ensemble that encodes the unpleasantness of pain. *Science* **363**, 276–281 (2019).
- Crook, R., Weston, M., Wilde, R. P. & Hartnell, G. G. Aortic valvoplasty: comparison of the techniques and results of transeptal and retrograde methods. *Clin. Radio.* 42, 110–113 (1990).
- 126. Zhou, Y. et al. Plasma inflammatory cytokines and treatment-resistant depression with comorbid pain: improvement by ketamine. *J. Neuroinflamm.* **18**, 200 (2021).

- 127. Taddio, A., Shah, V., Gilbert-MacLeod, C. & Katz, J. Conditioning and hyperalgesia in newborns exposed to repeated heel lances. *JAMA* **288**, 857–861 (2002).
- Mercer Lindsay, N., Chen, C., Gilam, G., Mackey, S. & Scherrer, G. Brain circuits for pain and its treatment. *Sci. Transl. Med.* 13, eabj7360 (2021).
- 129. Qi, X. et al. A nociceptive neuronal ensemble in the dorsomedial prefrontal cortex underlies pain chronicity. *Cell Rep.* **41**, 111833 (2022).
- Bhattacherjee, A. et al. Spatial transcriptomics reveals the distinct organization of mouse prefrontal cortex and neuronal subtypes regulating chronic pain. *Nat. Neurosci.* 26, 1880–1893 (2023).
- 131. Yang, H. et al. Pain modulates dopamine neurons via a spinal-parabrachialmesencephalic circuit. *Nat. Neurosci.* 24, 1402–1413 (2021).
- 132. Shao, S. et al. Ventral hippocampal CA1 modulates pain behaviors in mice with peripheral inflammation. *Cell Rep.* **42**, 112017 (2023).
- Jiang, Y. Y. et al. Cholinergic neurons in medial septum maintain anxiety-like behaviors induced by chronic inflammatory pain. *Neurosci. Lett.* 671, 7–12 (2018).
- Zhang, M. et al. Anxiolytic effects of hippocampal neurosteroids in normal and neuropathic rats with spared nerve injury. J. Neurochem. 141, 137–150 (2017).
- 135. Zheng, J. et al. Adult hippocampal neurogenesis along the dorsoventral axis contributes differentially to environmental enrichment combined with voluntary exercise in alleviating chronic inflammatory pain in mice. *J. Neurosci.* **37**, 4145–4157 (2017).
- Xia, S. H. et al. Chronic pain impairs memory formation via disruption of neurogenesis mediated by mesohippocampal brain-derived neurotrophic factor signaling. *Biol. Psychiatry* 88, 597–610 (2020).
- Yin, J. B. et al. dmPFC-vIPAG projection neurons contribute to pain threshold maintenance and antianxiety behaviors. J. Clin. Investig. 130, 6555–6570 (2020).
- 138. Huang, J. et al. A neuronal circuit for activating descending modulation of neuropathic pain. *Nat. Neurosci.* **22**, 1659–1668 (2019).
- 139. Christensen, J., MacPherson, N., Li, C., Yamakawa, G. R. & Mychasiuk, R. Repeat mild traumatic brain injuries (RmTBI) modify nociception and disrupt orexinergic connectivity within the descending pain pathway. J. Headache Pain 24, 72 (2023).
- Tobaldini, G., Sardi, N. F., Guilhen, V. A. & Fischer, L. Pain inhibits pain: an ascending-descending pain modulation pathway linking mesolimbic and classical descending mechanisms. *Mol. Neurobiol.* 56, 1000–1013 (2019).
- 141. Lopez-Alvarez, V. M., Puigdomenech, M., Navarro, X. & Cobianchi, S. Monoaminergic descending pathways contribute to modulation of neuropathic pain by increasing-intensity treadmill exercise after peripheral nerve injury. *Exp. Neurol.* 299, 42–55 (2018).
- 142. Jiao, Y. et al. Molecular identification of bulbospinal ON neurons by GPER, which drives pain and morphine tolerance. *J. Clin. Investig.* **133**, e154588 (2023).
- 143. Costa, A. R. et al. Neuropathic pain induced alterations in the opioidergic modulation of a descending pain facilitatory area of the brain. *Front. Cell Neurosci.* **13**, 287 (2019).
- 144. Suzuki, R., Morcuende, S., Webber, M., Hunt, S. P. & Dickenson, A. H. Superficial NK1-expressing neurons control spinal excitability through activation of descending pathways. *Nat. Neurosci.* 5, 1319–1326 (2002).
- Lockwood, S. & Dickenson, A. H. What goes up must come down: insights from studies on descending controls acting on spinal pain processing. *J. Neural Transm.* **127**, 541–549 (2020).
- Tillu, D. V., Gebhart, G. F. & Sluka, K. A. Descending facilitatory pathways from the RVM initiate and maintain bilateral hyperalgesia after muscle insult. *Pain* 136, 331–339 (2008).
- Liang, S. H. et al. A neural circuit from thalamic paraventricular nucleus to central amygdala for the facilitation of neuropathic pain. J. Neurosci. 40, 7837–7854 (2020).
- 148. Hemington, K. S. et al. Patients with chronic pain exhibit a complex relationship triad between pain, resilience, and within- and cross-network functional connectivity of the default mode network. *Pain* **159**, 1621–1630 (2018).
- 149. Baliki, M. N., Mansour, A. R., Baria, A. T. & Apkarian, A. V. Functional reorganization of the default mode network across chronic pain conditions. *PLoS One* 9, e106133 (2014).
- 150. Niu, X. et al. Mild traumatic brain injury is associated with effect of inflammation on structural changes of default mode network in those developing chronic pain. *J. Headache Pain* **21**, 135 (2020).
- 151. Niu, X. et al. Disruption of periaqueductal grey-default mode network functional connectivity predicts persistent post-traumatic headache in mild traumatic brain injury. J. Neurol. Neurosurg. Psychiatry **90**, 326–332 (2019).
- 152. Hsiao, F. J. et al. Altered insula-default mode network connectivity in fibromyalgia: a resting-state magnetoencephalographic study. J. Headache Pain 18, 89 (2017).
- 153. Tu, Y. et al. Distinct thalamocortical network dynamics are associated with the pathophysiology of chronic low back pain. *Nat. Commun.* **11**, 3948 (2020).
- 154. Mawla, I. et al. Large-scale momentary brain co-activation patterns are associated with hyperalgesia and mediate focal neurochemistry and cross-network functional connectivity in fibromyalgia. *Pain* **164**, 2737–2748 (2023).

- 155. Kucyi, A., Salomons, T. V. & Davis, K. D. Mind wandering away from pain dynamically engages antinociceptive and default mode brain networks. *Proc. Natl Acad. Sci. USA* **110**, 18692–18697 (2013).
- Riegner, G. et al. Disentangling self from pain: mindfulness meditation-induced pain relief is driven by thalamic-default mode network decoupling. *Pain* 164, 280–291 (2023).
- 157. Letzen, J. E. & Robinson, M. E. Negative mood influences default mode network functional connectivity in patients with chronic low back pain: implications for functional neuroimaging biomarkers. *Pain* **158**, 48–57 (2017).
- 158. Alshelh, Z. et al. Disruption of default mode network dynamics in acute and chronic pain states. *Neuroimage Clin.* **17**, 222–231 (2018).
- 159. Cox, J. J. et al. An SCN9A channelopathy causes congenital inability to experience pain. *Nature* **444**, 894–898 (2006).
- Ahmad, S. et al. A stop codon mutation in SCN9A causes lack of pain sensation. Hum. Mol. Genet. 16, 2114–2121 (2007).
- Yammine, T. et al. Novel SCN9A variant associated with congenital insensitivity to pain. *Mol. Biol. Rep.* 50, 6293–6298 (2023).
- 162. Kurzawski, M. et al. Common missense variant of SCN9A gene is associated with pain intensity in patients with chronic pain from disc herniation. *Pain. Med.* **19**, 1010–1014 (2018).
- Rühlmann, A. H. et al. Uncoupling sodium channel dimers restores the phenotype of a pain-linked Na(v) 1.7 channel mutation. Br. J. Pharm. 177, 4481–4496 (2020).
- Zhang, X. Y. et al. Gain-of-function mutations in SCN11A cause familial episodic pain. Am. J. Hum. Genet. 93, 957–966 (2013).
- 165. Matsubara, Y., Okuda, H., Harada, K. H., Youssefian, S. & Koizumi, A. Mechanical allodynia triggered by cold exposure in mice with the Scn11a p.R222S mutation: a novel model of drug therapy for neuropathic pain related to Na(V)1.9. Naunyn Schmiedebergs Arch. Pharm. **394**, 299–306 (2021).
- Leipold, E. et al. A de novo gain-of-function mutation in SCN11A causes loss of pain perception. *Nat. Genet.* 45, 1399–1404 (2013).
- 167. Suleimanova, A., Talanov, M., van den Maagdenberg, A. & Giniatullin, R. Deciphering in silico the role of mutated Na (V) 1.1 sodium channels in enhancing trigeminal nociception in familial hemiplegic migraine type 3. *Front. Cell Neurosci.* **15**, 644047 (2021).
- 168. Jeong, S., Shim, J. S., Sin, S. K., Park, K. S. & Lee, J. H. Phosphorylation states greatly regulate the activity and gating properties of Ca(v) 3.1 T-type Ca(2+) channels. J. Cell Physiol. 238, 210–226 (2023).
- 169. Field, M. J. et al. Identification of the alpha2-delta-1 subunit of voltagedependent calcium channels as a molecular target for pain mediating the analgesic actions of pregabalin. *Proc. Natl Acad. Sci. USA* **103**, 17537–17542 (2006).
- Pettingill, P. et al. A causal role for TRESK loss of function in migraine mechanisms. *Brain* **142**, 3852–3867 (2019).
- 171. Lee, M. C. et al. Human labor pain is influenced by the voltage-gated potassium channel K(V)6.4 subunit. *Cell Rep.* **32**, 107941 (2020).
- 172. Katz, B. et al. Nociception and pain in humans lacking a functional TRPV1 channel. J. Clin. Investig. 133, e153558 (2023).
- 173. Duo, L. et al. TRPV1 gain-of-function mutation impairs pain and itch sensations in mice. *Mol. Pain* **14**, 1744806918762031 (2018).
- Christoph, T., Kögel, B., Schiene, K., Peters, T. & Schröder, W. Investigation of TRPV1 loss-of-function phenotypes in TRPV1 Leu206Stop mice generated by N-ethyl-Nnitrosourea mutagenesis. *Biochem. Biophys. Res. Commun.* 500, 456–461 (2018).
- Zíma, V. et al. Structural modeling and patch-clamp analysis of pain-related mutation TRPA1-N855S reveal inter-subunit salt bridges stabilizing the channel open state. *Neuropharmacology* **93**, 294–307 (2015).
- Kremeyer, B. et al. A gain-of-function mutation in TRPA1 causes familial episodic pain syndrome. *Neuron* 66, 671–680 (2010).
- 177. Bali, A. et al. Molecular mechanism of hyperactivation conferred by a truncation of TRPA1. *Nat. Commun.* **14**, 2867 (2023).
- 178. Chen, Y. C. et al. Transcriptional regulator PRDM12 is essential for human pain perception. *Nat. Genet.* **47**, 803–808 (2015).
- Bhattacherjee, A. et al. Neuronal cytoskeletal gene dysregulation and mechanical hypersensitivity in a rat model of Rett syndrome. *Proc. Natl Acad. Sci. USA* 114, E6952–e6961 (2017).
- Nahorski, M. S. et al. A novel disorder reveals clathrin heavy chain-22 is essential for human pain and touch development. *Brain* 138, 2147–2160 (2015).
- Habib, A. M. et al. A novel human pain insensitivity disorder caused by a point mutation in ZFHX2. *Brain* 141, 365–376 (2018).
- Perini, I. et al. Mutation carriers with reduced c-afferent density reveal cortical dynamics of pain-action relationship during acute pain. *Cereb. Cortex* 30, 4858–4870 (2020).
- Sorge, R. E. et al. Genetically determined P2X7 receptor pore formation regulates variability in chronic pain sensitivity. *Nat. Med.* 18, 595–599 (2012).

- 38
- 184. Goodin, B. R. et al. Epigenome-wide DNA methylation profiling of conditioned pain modulation in individuals with non-specific chronic low back pain. *Clin. Epigenet.* **14**, 45 (2022).
- 185. Li, X. et al. Intraperitoneal 5-azacytidine alleviates nerve injury-induced pain in rats by modulating DNA methylation. *Mol. Neurobiol.* **60**, 2186–2199 (2023).
- 186. Aroke, E. N. et al. Identification of DNA methylation associated enrichment pathways in adults with non-specific chronic low back pain. *Mol. Pain* 16, 1744806920972889 (2020).
- Carlsen, L. N. et al. DNA-methylation and immunological response in medication overuse headache. *Cephalalgia* 43, 3331024221147482 (2023).
- Garriga, J. et al. Nerve injury-induced chronic pain is associated with persistent DNA methylation reprogramming in dorsal root ganglion. J. Neurosci. 38, 6090–6101 (2018).
- 189. Chen, W. et al. Whole genomic DNA methylation profiling of cpg sites in promoter regions of dorsal root ganglion in diabetic neuropathic pain mice. J. Mol. Neurosci. 71, 2558–2565 (2021).
- 190. Wan, D. et al. DNA methylation of RAMP1 gene in migraine: an exploratory analysis. *J. Headache Pain* **16**, 90 (2015).
- Bell, J. T. et al. Differential methylation of the TRPA1 promoter in pain sensitivity. Nat. Commun. 5, 2978 (2014).
- 192. Gombert, S. et al. Epigenetic divergence in the TRPA1 promoter correlates with pressure pain thresholds in healthy individuals. *Pain* **158**, 698–704 (2017).
- 193. Polli, A. et al. DNA methylation and brain-derived neurotrophic factor expression account for symptoms and widespread hyperalgesia in patients with chronic fatigue syndrome and comorbid fibromyalgia. *Arthritis Rheumatol.* **72**, 1936–1944 (2020).
- 194. Rodrigues, D., Monteiro, C., Cardoso-Cruz, H. & Galhardo, V. Altered brain expression of dna methylation and hydroxymethylation epigenetic enzymes in a rat model of neuropathic pain. *Int. J. Mol. Sci.* 24 (2023).
- Zhao, J. Y. et al. DNA methyltransferase DNMT3a contributes to neuropathic pain by repressing Kcna2 in primary afferent neurons. *Nat. Commun.* 8, 14712 (2017).
- 196. Liu, L. et al. Epigenetic reduction of miR-214-3p upregulates astrocytic colonystimulating factor-1 and contributes to neuropathic pain induced by nerve injury. *Pain* **161**, 96–108 (2020).
- 197. Mikaeili, H. et al. Molecular basis of FAAH-OUT-associated human pain insensitivity. *Brain* **146**, 3851–3865 (2023).
- 198. Hsieh, M. C. et al. Melatonin impedes Tet1-dependent mGluR5 promoter demethylation to relieve pain. J. Pineal. Res. 63 (2017).
- Deng, J. et al. Oxaliplatin-induced neuropathic pain involves HOXA6 via a TET1dependent demethylation of the SOX10 promoter. *Int. J. Cancer* 147, 2503–2514 (2020).
- Lai, C. Y. et al. MicroRNA-489-3p attenuates neuropathic allodynia by regulating oncoprotein DEK/TET1-dependent epigenetic modification in the dorsal horn. *Neuropharmacology* **210**, 109028 (2022).
- Xu, Z. H. et al. TET1-TRPV4 signaling contributes to bone cancer pain in Rats. Brain Sci. 13, 644 (2023).
- 202. Jia, S. et al. TET1 overexpression attenuates paclitaxel-induced neuropathic pain through rescuing K(2p)1.1 expression in primary sensory neurons of male rats. *Life Sci.* 297, 120486 (2022).
- 203. Wu, Q. et al. TET1 overexpression mitigates neuropathic pain through rescuing the expression of μ-opioid receptor and Kv1.2 in the primary sensory neurons. *Neurotherapeutics* **16**, 491–504 (2019).
- Li, Y. et al. Electroacupuncture ameliorates depression-like behaviors comorbid to chronic neuropathic pain via Tet1-mediated restoration of adult neurogenesis. Stem Cells 41, 384–399 (2023).
- Doehring, A., Oertel, B. G., Sittl, R. & Lötsch, J. Chronic opioid use is associated with increased DNA methylation correlating with increased clinical pain. *Pain* 154, 15–23 (2013).
- 206. Agulló, L. et al. Sex differences in opioid response linked to OPRM1 and COMT genes DNA methylation/genotypes changes in patients with chronic pain. J. *Clin. Med.* **12**, 3449 (2023).
- 207. Achenbach, J., Rhein, M., Glahn, A., Frieling, H. & Karst, M. Leptin promoter methylation in female patients with painful multisomatoform disorder and chronic widespread pain. *Clin. Epigenet.* **14**, 13 (2022).
- Zhang, C. et al. PiRNA-DQ541777 contributes to neuropathic pain via targeting Cdk5rap1. J. Neurosci. 39, 9028–9039 (2019).
- Maiarù, M. et al. A DNA methylation signature in the stress driver gene Fkbp5 indicates a neuropathic component in chronic pain. *Clin. Epigenet.* 15, 155 (2023).
- Louwies, T. & Greenwood-Van Meerveld, B. Sex differences in the epigenetic regulation of chronic visceral pain following unpredictable early life stress. *Neurogastroenterol. Motil.* 32, e13751 (2020).
- 211. Imbe, H. & Kimura, A. Increase of histone acetylation in the GABAergic neurons in the rostral ventromedial medulla associated with mechanical hypersensitivity after repeated restraint stress. *Brain Res. Bull.* **142**, 394–402 (2018).

- Liang, L. & Tao, Y. X. Expression of acetyl-histone H3 and acetyl-histone H4 in dorsal root ganglion and spinal dorsal horn in rat chronic pain models. *Life Sci.* 211, 182–188 (2018).
- Guan, L. et al. Contribution of amygdala histone acetylation in early life stressinduced visceral hypersensitivity and emotional comorbidity. *Front. Neurosci.* 16, 843396 (2022).
- 214. Zhou, C. et al. SIRT1 decreases emotional pain vulnerability with associated CaMKIIa deacetylation in central amygdala. *J. Neurosci.* **40**, 2332–2342 (2020).
- 215. Zhao, Y. T. et al. Adaptation of prelimbic cortex mediated by IL-6/STAT3/Acp5 pathway contributes to the comorbidity of neuropathic pain and depression in rats. *J. Neuroinflamm.* **19**, 144 (2022).
- 216. Imbe, H. & Kimura, A. Repeated forced swim stress affects the expression of pCREB and ΔFosB and the acetylation of histone H3 in the rostral ventromedial medulla and locus coeruleus. *Brain Res Bull.* **127**, 11–22 (2016).
- Ding, H. H. et al. TNF-a/STAT3 pathway epigenetically upregulates Nav1.6 expression in DRG and contributes to neuropathic pain induced by L5-VRT. J. Neuroinflamm. 16, 29 (2019).
- Li, Z. et al. SCFAs ameliorate chronic postsurgical pain-related cognition dysfunction via the ACSS2-HDAC2 axis in rats. *Mol. Neurobiol.* 59, 6211–6227 (2022).
- Miao, J., Chen, Z., Wu, Y., Hu, Q. & Ji, T. Sp1 inhibits PGC-1α via HDAC2-catalyzed histone deacetylation in chronic constriction injury-induced neuropathic pain. ACS Chem. Neurosci. 13, 3438–3452 (2022).
- Lin, T. B. et al. Melatonin relieves neuropathic allodynia through spinal MT2enhanced PP2Ac and downstream HDAC4 shuttling-dependent epigenetic modification of hmgb1 transcription. J. Pineal. Res. 60, 263–276 (2016).
- Gu, P. et al. Central endothelin-1 confers analgesia by triggering spinal neuronal histone deacetylase 5 (HDAC5) nuclear exclusion in peripheral neuropathic pain in mice. J. Pain 22, 454–471 (2021).
- 222. Fan, T. et al. Histone deacetylase 5-induced deficiency of signal transducer and activator of transcription-3 acetylation contributes to spinal astrocytes degeneration in painful diabetic neuropathy. *Glia* **71**, 1099–1119 (2023).
- Xu, L. J. et al. Reduction of SIRT1-mediated epigenetic upregulation of Nav1.7 contributes to oxaliplatin-induced neuropathic pain. *Pain Physician* 26, E213–E222 (2023).
- Zhou, C. et al. SIRT3 alleviates neuropathic pain by deacetylating FoxO3a in the spinal dorsal horn of diabetic model rats. *Reg. Anesth. Pain Med.* 46, 49–56 (2021).
- Hong, S., Zheng, G. & Wiley, J. W. Epigenetic regulation of genes that modulate chronic stress-induced visceral pain in the peripheral nervous system. *Gastroenterology* **148**, 148–157.e147 (2015).
- 226. Wang, C. et al. Epigenetic up-regulation of ADAMTS4 in sympathetic ganglia is involved in the maintenance of neuropathic pain following nerve injury. *Neurochem. Res.* **48**, 2350–2359 (2023).
- 227. Xu, T. et al. Epigenetic upregulation of CXCL12 expression mediates antitubulin chemotherapeutics-induced neuropathic pain. *Pain* **158**, 637–648 (2017).
- Zou, Y. et al. Involvement of histone lysine crotonylation in the regulation of nerve-injury-induced neuropathic pain. Front. Immunol. 13, 885685 (2022).
- Weng, H. R., Taing, K., Chen, L. & Penney, A. EZH2 Methyltransferase regulates neuroinflammation and neuropathic pain. *Cells* 12 (2023).
- Yadav, R. & Weng, H. R. EZH2 regulates spinal neuroinflammation in rats with neuropathic pain. *Neuroscience* 349, 106–117 (2017).
- 231. Chen, H. et al. Evidence of the involvement of spinal EZH2 in the development of bone cancer pain in rats. J. Pain Res. 14, 3593–3600 (2021).
- 232. An, Q. et al. Calcitonin gene-related peptide regulates spinal microglial activation through the histone H3 lysine 27 trimethylation via enhancer of zeste homolog-2 in rats with neuropathic pain. J. Neuroinflamm. 18, 117 (2021).
- Meng, X. L. et al. Increased EZH2 levels in anterior cingulate cortex microglia aggravate neuropathic pain by inhibiting autophagy following brachial plexus avulsion in rats. *Neurosci. Bull.* 36, 793–805 (2020).
- 234. Zhang, Z., Sun, X., Zhao, G., Ma, Y. & Zeng, G. LncRNA embryonic stem cells expressed 1 (Lncenc1) is identified as a novel regulator in neuropathic pain by interacting with EZH2 and downregulating the expression of Bai1 in mouse microglia. *Exp. Cell Res.* **399**, 112435 (2021).
- Zhang, Y. et al. miR-124-3p attenuates neuropathic pain induced by chronic sciatic nerve injury in rats via targeting EZH2. J. Cell Biochem. 120, 5747–5755 (2019).
- Gao, P. et al. Overexpression of miR-378 alleviates chronic sciatic nerve injury by targeting EZH2. *Neurochem. Res.* 46, 3213–3221 (2021).
- Chen, S. H. et al. Ultrahigh frequency transcutaneous electrical nerve stimulation for neuropathic pain alleviation and neuromodulation. *Neurotherapeutics* 21, e00336 (2024).
- Yang, X. et al. Transcriptome profiling of miRNA-mRNA interactions and associated mechanisms in chemotherapy-induced neuropathic pain. *Mol. Neurobiol.* 60, 5672–5690 (2023).
- Francés, R. et al. Identification of epigenetic interactions between MicroRNA-30c-5p and DNA methyltransferases in neuropathic pain. *Int. J. Mol. Sci.* 23, 13994 (2022).

- Yan, X. T. et al. MicroRNA-93 alleviates neuropathic pain through targeting signal transducer and activator of transcription 3. *Int. Immunopharmacol.* 46, 156–162 (2017).
- 241. Sabirzhanov, B. et al. Inhibition of NOX2 signaling limits pain-related behavior and improves motor function in male mice after spinal cord injury: Participation of IL-10/miR-155 pathways. *Brain Behav. Immun.* **80**, 73–87 (2019).
- Pan, Z. et al. miRNA-23a/CXCR4 regulates neuropathic pain via directly targeting TXNIP/NLRP3 inflammasome axis. J. Neuroinflamm. 15, 29 (2018).
- 243. Qi, R. et al. Histone methylation-mediated microRNA-32-5p down-regulation in sensory neurons regulates pain behaviors via targeting Cav3.2 channels. Proc. Natl Acad. Sci. USA 119, e2117209119 (2022).
- 244. Peng, C. et al. miR-183 cluster scales mechanical pain sensitivity by regulating basal and neuropathic pain genes. *Science* **356**, 1168–1171 (2017).
- Ma, Y. et al. TRPV1, targeted by miR-338-3p, induces neuropathic pain by interacting with NECAB2. J. Mol. Neurosci. 71, 55–65 (2021).
- 246. Zhou, Q. et al. Decreased miR-199 augments visceral pain in patients with IBS through translational upregulation of TRPV1. *Gut* **65**, 797–805 (2016).
- 247. Ni, H. et al. Upregulation of LncRNA71132 in the spinal cord regulates hypersensitivity in a rat model of bone cancer pain. *Pain* **164**, 180–196 (2023).
- Bai, X. et al. CircNf1-mediated CXCL12 expression in the spinal cord contributes to morphine analgesic tolerance. *Brain Behav. Immun.* **107**, 140–151 (2023).
- 249. Pan, Z. et al. Downregulation of a dorsal root ganglion-specifically enriched long noncoding rna is required for neuropathic pain by negatively regulating RALYtriggered Ehmt2 expression. Adv. Sci. 8, e2004515 (2021).
- Du, S. et al. A nerve injury-specific long noncoding RNA promotes neuropathic pain by increasing Ccl2 expression. J. Clin. Investig. 132, e153563 (2022).
- 251. Zhang, S. B. et al. CircAnks1a in the spinal cord regulates hypersensitivity in a rodent model of neuropathic pain. *Nat. Commun.* **10**, 4119 (2019).
- 252. Maruyama, M. et al. Neat1 IncRNA organizes the inflammatory gene expressions in the dorsal root ganglion in neuropathic pain caused by nerve injury. *Front. Immunol.* **14**, 1185322 (2023).
- 253. Noor, S. et al. Prenatal alcohol exposure dysregulates spinal and circulating immune cell circular RNA expression in adult female rats with chronic sciatic neuropathy. *Front. Neurosci.* **17**, 1180308 (2023).
- Simeoli, R. et al. Exosomal cargo including microRNA regulates sensory neuron to macrophage communication after nerve trauma. *Nat. Commun.* 8, 1778 (2017).
- Liao, K. et al. Morphine-mediated release of miR-138 in astrocyte-derived extracellular vesicles promotes microglial activation. *J. Extracell. Vesicles* 10, e12027 (2020).
- Jean-Toussaint, R. et al. Therapeutic and prophylactic effects of macrophagederived small extracellular vesicles in the attenuation of inflammatory pain. *Brain Behav. Immun.* 94, 210–224 (2021).
- Andelic, M. et al. Integrative miRNA-mRNA profiling of human epidermis: unique signature of SCN9A painful neuropathy. *Brain* 146, 3049–3062 (2023).
- Zhao, C. et al. CircFOXO3 protects against osteoarthritis by targeting its parental gene FOXO3 and activating PI3K/AKT-mediated autophagy. *Cell Death Dis.* 13, 932 (2022).
- Wang, B. et al. A sensory neuron-specific long non-coding RNA reduces neuropathic pain by rescuing KCNN1 expression. *Brain* 146, 3866–3884 (2023).
- Kong, E. et al. Microglial exosome miR-124-3p in hippocampus alleviates cognitive impairment induced by postoperative pain in elderly mice. J. Cell Mol. Med. 28, e18090 (2024).
- Sun, N. et al. MeCP2 epigenetic silencing of Oprm1 gene in primary sensory neurons under neuropathic pain conditions. *Front. Neurosci.* 15, 743207 (2021).
- 262. Pan, Z. et al. Hydroxymethylation of microRNA-365-3p regulates nociceptive behaviors via Kcnh2. J. Neurosci. 36, 2769–2781 (2016).
- 263. Wu, X. et al. Targeting protein modifications in metabolic diseases: molecular mechanisms and targeted therapies. *Signal Transduct. Target Ther.* 8, 220 (2023).
- 264. Yang, H. B. et al. cAMP-dependent protein kinase activated Fyn in spinal dorsal horn to regulate NMDA receptor function during inflammatory pain. J. Neurochem. 116, 93–104 (2011).
- 265. Kimura, Y. et al. IL-33 induces orofacial neuropathic pain through Fyndependent phosphorylation of GluN2B in the trigeminal spinal subnucleus caudalis. Brain Behav. Immun. 99, 266–280 (2022).
- Xiao, X. et al. Shp-1 dephosphorylates TRPV1 in dorsal root ganglion neurons and alleviates CFA-induced inflammatory pain in rats. *Pain* **156**, 597–608 (2015).
- Chen, G. et al. PD-L1 inhibits acute and chronic pain by suppressing nociceptive neuron activity via PD-1. *Nat. Neurosci.* 20, 917–926 (2017).
- Liu, B. L., Cao, Q. L., Zhao, X., Liu, H. Z. & Zhang, Y. Q. Inhibition of TRPV1 by SHP-1 in nociceptive primary sensory neurons is critical in PD-L1 analgesia. *JCl Insight* 5, e137386 (2020).
- 269. Li, Y. et al. Nav1.7 is phosphorylated by Fyn tyrosine kinase which modulates channel expression and gating in a cell type-dependent manner. *Mol. Pain* 14, 1744806918782229 (2018).

- 270. Kerth, C. M., Hautvast, P., Körner, J., Lampert, A. & Meents, J. E. Phosphorylation of a chronic pain mutation in the voltage-gated sodium channel Nav1.7 increases voltage sensitivity. J. Biol. Chem. 296, 100227 (2021).
- Sulak, M. A., Ghosh, M., Sinharoy, P., Andrei, S. R. & Damron, D. S. Modulation of TRPA1 channel activity by Cdk5 in sensory neurons. *Channels* 12, 65–75 (2018).
 Rivera, B. et al. Constitutive phosphorylation as a key regulator of TRPM8
- CHANNEL Function. J. Neurosci. **41**, 8475-8493 (2021).
- 273. He, L. et al. Toll-like receptor 7 contributes to neuropathic pain by activating NF-KB in primary sensory neurons. *Brain Behav. Immun.* **87**, 840–851 (2020).
- Su, W. et al. Suppression of TLR4-MyD88 signaling pathway attenuated chronic mechanical pain in a rat model of endometriosis. J. Neuroinflamm. 18, 65 (2021).
- 275. Wang, L. et al. Pellino1 regulates neuropathic pain as well as microglial activation through the regulation of MAPK/NF-κB signaling in the spinal cord. J. Neuroinflamm. 17, 83 (2020).
- 276. Luo, X. et al. Intrathecal administration of antisense oligonucleotide against p38α but not p38β MAP kinase isoform reduces neuropathic and postoperative pain and TLR4-induced pain in male mice. *Brain Behav. Immun.* **72**, 34–44 (2018).
- Zhang, R. et al. Spinal microglia-derived TNF promotes the astrocytic JNK/CXCL1 pathway activation in a mouse model of burn pain. *Brain Behav. Immun.* 102, 23–39 (2022).
- Cheng, K., Martin, L. F., Slepian, M. J., Patwardhan, A. M. & Ibrahim, M. M. Mechanisms and pathways of pain photobiomodulation: a narrative review. *J. Pain.* 22, 763–777 (2021).
- 279. Dubin, A. E. & Patapoutian, A. Nociceptors: the sensors of the pain pathway. J. Clin. Investig. **120**, 3760–3772 (2010).
- Gautam, V. et al. Nedd4 is a specific E3 ubiquitin ligase for the NMDA receptor subunit GluN2D. *Neuropharmacology* 74, 96–107 (2013).
- Wang, C., Chen, R., Zhu, X. & Zhang, X. Suberoylanilide hydroxamic acid ameliorates pain sensitization in central neuropathic pain after spinal cord injury via the HDAC5/NEDD4/SCN9A axis. *Neurochem Res* 48, 2436–2450 (2023).
- Wang, S. et al. The ubiquitin E3 ligase Nedd4-2 relieves mechanical allodynia through the ubiquitination of TRPA1 channel in db/db mice. *Eur. J. Neurosci.* 53, 1691–1704 (2021).
- Deftu, A. F. et al. The antidiabetic drug metformin regulates voltage-gated sodium channel Na(V)1.7 via the ubiquitin-ligase NEDD4-2. *eNeuro*. 9, ENEURO.0409-21.2022 (2022).
- Zhang, Z. Y. et al. Ubiquitination and functional modification of GluN2B subunitcontaining NMDA receptors by Cbl-b in the spinal cord dorsal horn. *Sci. Signal* 13, eaaw1519 (2020).
- Chen, H. K. et al. Cbl-b modulated TrkA ubiquitination and function in the dorsal root ganglion of mice. *Eur. J. Pharm.* **921**, 174876 (2022).
- Ma, J. J. et al. BDNF modulated KCC2 ubiquitylation in spinal cord dorsal horn of mice. *Eur. J. Pharm.* **906**, 174205 (2021).
- Jeong, J. S. et al. Increased expression of the Cbl family of E3 ubiquitin ligases decreases Interleukin-2 production in a rat model of peripheral neuropathy. *BMC Anesthesiol.* 18, 87 (2018).
- Stemkowski, P. et al. TRPV1 nociceptor activity initiates USP5/T-type channelmediated plasticity. *Cell Rep.* 17, 2901–2912 (2016).
- Stemkowski, P. L. et al. Identification of interleukin-1 beta as a key mediator in the upregulation of Cav3.2-USP5 interactions in the pain pathway. *Mol. Pain* 13, 1744806917724698 (2017).
- 290. Tomita, S. et al. Ca(v)3.2 overexpression in L4 dorsal root ganglion neurons after L5 spinal nerve cutting involves Egr-1, USP5 and HMGB1 in rats: an emerging signaling pathway for neuropathic pain. *Eur. J. Pharm.* 888, 173587 (2020).
- 291. García-Caballero, A. et al. The deubiquitinating enzyme USP5 modulates neuropathic and inflammatory pain by enhancing Cav3.2 channel activity. *Neuron* 83, 1144–1158 (2014).
- 292. Sun, L. et al. Targeted ubiquitination of sensory neuron calcium channels reduces the development of neuropathic pain. *Proc. Natl Acad. Sci. USA* **119**, e2118129119 (2022).
- 293. Tomita, S. et al. Critical role of Ca(v)3.2 T-type calcium channels in the peripheral neuropathy induced by bortezomib, a proteasome-inhibiting chemotherapeutic agent, in mice. *Toxicology* **413**, 33–39 (2019).
- 294. Marangoudakis, S. et al. Differential ubiquitination and proteasome regulation of Ca(V)2.2 N-type channel splice isoforms. J. Neurosci. **32**, 10365–10369 (2012).
- 295. Lai, C. Y. et al. Spinal Fbxo3-dependent Fbxl2 ubiquitination of active zone protein RIM1α mediates neuropathic allodynia through CaV2.2 activation. J. Neurosci. 36, 9722–9738 (2016).
- Liu, X. D. et al. DHX9/DNA-tandem repeat-dependent downregulation of ciRNA-Fmn1 in the dorsal horn is required for neuropathic pain. *Acta Pharm. Sin.* 44, 1748–1767 (2023).
- 297. Zhang, Z. Y. et al. Ubiquitination and inhibition of glycine receptor by HUWE1 in spinal cord dorsal horn. *Neuropharmacology* **148**, 358–365 (2019).
- 298. Vertegaal, A. C. O. Signalling mechanisms and cellular functions of SUMO. *Nat. Rev. Mol. Cell Biol.* **23**, 715–731 (2022).

- 40
- Parker, A. R., Forster, L. A. & Baro, D. J. Modulator-gated, SUMOylation-mediated, activity-dependent regulation of ionic current densities contributes to shortterm activity homeostasis. J. Neurosci. 39, 596–611 (2019).
- François-Moutal, L. et al. Inhibition of the Ubc9 E2 SUMO-conjugating enzyme-CRMP2 interaction decreases NaV1.7 currents and reverses experimental neuropathic pain. *Pain* **159**, 2115–2127 (2018).
- Moutal, A. et al. Studies on CRMP2 SUMOylation-deficient transgenic mice identify sex-specific Nav1.7 regulation in the pathogenesis of chronic neuropathic pain. *Pain* 161, 2629–2651 (2020).
- Moutal, A. et al. Blocking CRMP2 SUMOylation reverses neuropathic pain. *Mol. Psychiatry* 23, 2119–2121 (2018).
- 303. Gomez, K. et al. Targeted transcriptional upregulation of SENP1 by CRISPR activation enhances deSUMOylation pathways to elicit antinociception in the spinal nerve ligation model of neuropathic pain. *Pain* **165**, 866–883 (2023).
- Dustrude, E. T. et al. Hierarchical CRMP2 posttranslational modifications control NaV1.7 function. Proc. Natl Acad. Sci. USA 113, E8443–E8452 (2016).
- Loya-Lopez, S. I. et al. Intranasal CRMP2-Ubc9 inhibitor regulates NaV1.7 to alleviate trigeminal neuropathic pain. *Pain* 165, 573–588 (2023).
- Lv, Y. Y. et al. SUMOylation of Kir7.1 participates in neuropathic pain through regulating its membrane expression in spinal cord neurons. *CNS Neurosci. Ther.* 28, 1259–1267 (2022).
- 307. Forster, L. A., Jansen, L. R., Rubaharan, M., Murphy, A. Z. & Baro, D. J. Alterations in SUMOylation of the hyperpolarization-activated cyclic nucleotide-gated ion channel 2 during persistent inflammation. *Eur. J. Pain.* 24, 1517–1536 (2020).
- Garcia-Caballero, A. et al. SUMOylation regulates USP5-Cav3.2 calcium channel interactions. *Mol. Brain* 12, 73 (2019).
- Zhao, X., Xia, B., Cheng, J., Zhu, M. X. & Li, Y. PKCc SUMOylation is required for mediating the nociceptive signaling of inflammatory pain. *Cell Rep.* 33, 108191 (2020).
- Ondacova, K., Karmazinova, M., Lazniewska, J., Weiss, N. & Lacinova, L. Modulation of Cav3.2 T-type calcium channel permeability by asparagine-linked glycosylation. *Channels* **10**, 175–184 (2016).
- Joksimovic, S. L. et al. Glycosylation of Ca(V)3.2 channels contributes to the hyperalgesia in peripheral neuropathy of type 1 diabetes. *Front. Cell Neurosci.* 14, 605312 (2020).
- Veldhuis, N. A. et al. N-glycosylation determines ionic permeability and desensitization of the TRPV1 capsaicin receptor. J. Biol. Chem. 287, 21765–21772 (2012).
- 313. Janes, K. et al. A3 adenosine receptor agonist prevents the development of paclitaxel-induced neuropathic pain by modulating spinal glial-restricted redoxdependent signaling pathways. *Pain* **155**, 2560–2567 (2014).
- Ilari, S. et al. Antioxidant modulation of sirtuin 3 during acute inflammatory pain: the ROS control. *Pharm. Res.* **157**, 104851 (2020).
- Cao, Y., Wang, W., Zhan, X. & Zhang, Y. PRDX6: a protein bridging S-palmitoylation and diabetic neuropathy. Front. Endocrinol. 13, 992875 (2022).
- Huang, Y., Lu, S., Chen, Y., Feng, Y. & Lu, W. Morphine induces HADHA succinylation, while HADHA desuccinylation alleviates morphine tolerance by influencing autophagy. *Naunyn Schmiedebergs Arch. Pharm.* 397, 1589–1600 (2023).
- Broz, P. & Dixit, V. M. Inflammasomes: mechanism of assembly, regulation and signalling. *Nat. Rev. Immunol.* 16, 407–420 (2016).
- Cowie, A. M., Menzel, A. D., O'Hara, C., Lawlor, M. W. & Stucky, C. L. NOD-like receptor protein 3 inflammasome drives postoperative mechanical pain in a sex-dependent manner. *Pain* 160, 1794–1816 (2019).
- Green-Fulgham, S. M. et al. Interleukin-1beta and inflammasome expression in spinal cord following chronic constriction injury in male and female rats. *Brain Behav. Immun.* 115, 157–168 (2023).
- Filippone, A. et al. BAY-117082-driven NLRP3 inflammasome inhibition resolves nitro-glycerine (NTG) neuronal damage in in vivo model of migraine. *Biomed. Pharmacother.* 156, 113851 (2022).
- 321. Miao, J., Zhou, X., Ji, T. & Chen, G. NF-κB p65-dependent transcriptional regulation of histone deacetylase 2 contributes to the chronic constriction injuryinduced neuropathic pain via the microRNA-183/TXNIP/NLRP3 axis. J. Neuroinflamm. 17, 225 (2020).
- 322. Chen, W. et al. The upregulation of NLRP3 inflammasome in dorsal root ganglion by ten-eleven translocation methylcytosine dioxygenase 2 (TET2) contributed to diabetic neuropathic pain in mice. J. Neuroinflamm. 19, 302 (2022).
- 323. Ruan, Y. et al. Paeoniflorin alleviates CFA-induced inflammatory pain by inhibiting TRPV1 and succinate/SUCNR1-HIF-1α/NLPR3 pathway. *Int. Immuno-pharmacol.* **101**, 108364 (2021).
- 324. Lan, Z. et al. Mechanosensitive TRPV4 is required for crystal-induced inflammation. *Ann. Rheum. Dis.* **80**, 1604–1614 (2021).
- 325. Hsiao, H. T., Liu, Y. Y., Wang, J. C., Lin, Y. C. & Liu, Y. C. The analgesic effect of propofol associated with the inhibition of hypoxia inducible factor and inflammasome in complex regional pain syndrome. J. Biomed. Sci. 26, 74 (2019).
- Liu, Q. et al. Melatonin alleviates morphine analgesic tolerance in mice by decreasing NLRP3 inflammasome activation. *Redox Biol.* 34, 101560 (2020).

- 327. Brosius Lutz, A. et al. An RNA-sequencing transcriptome of the rodent Schwann cell response to peripheral nerve injury. J. Neuroinflamm. **19**, 105 (2022).
- Stratton, J. A. et al. Macrophages regulate Schwann cell maturation after nerve injury. Cell Rep. 24, 2561–2572.e2566 (2018).
- Wu, W. H. et al. Interleukin-1β secretion induced by mucosa-associated gut commensal bacteria promotes intestinal barrier repair. *Gut Microbes* 14, 2014772 (2022).
- 330. Kim, H. K. et al. Blockers of Wnt3a, Wnt10a, or β-catenin prevent chemotherapyinduced neuropathic pain in vivo. *Neurotherapeutics* 18, 601–614 (2021).
- 331. Wei, M., Li, L., Zhang, Y., Zhang, M. & Su, Z. Downregulated circular RNA zRANB1 mediates Wnt5a/β-Catenin signaling to promote neuropathic pain via miR-24-3p/LPAR3 axis in CCI rat models. *Gene* **761**, 145038 (2020).
- Tang, J. et al. Secreted frizzled-related protein 1 regulates the progression of neuropathic pain in mice following spinal nerve ligation. J. Cell Physiol. 233, 5815–5822 (2018).
- Lu, Y. et al. Human PMSCs-derived small extracellular vesicles alleviate neuropathic pain through miR-26a-5p/Wnt5a in SNI mice model. J. Neuroinflamm. 19, 221 (2022).
- Zhang, Q., Zhu, D. & Li, Q. LncRNA CRNDE exacerbates neuropathic pain in chronic constriction injury-induced(CCI) rats through regulating miR-146a-5p/ WNT5A pathway. *Bioengineered* 12, 7348–7359 (2021).
- Xie, Y. K. et al. GPR177 in A-fiber sensory neurons drives diabetic neuropathic pain via WNT-mediated TRPV1 activation. *Sci. Transl. Med.* 14, eabh2557 (2022).
- He, J. J. et al. Wht5b/Ryk-mediated membrane trafficking of P2X3 receptors contributes to bone cancer pain. *Exp. Neurol.* 334, 113482 (2020).
- 337. Zhang, Y. K. et al. WNT signaling underlies the pathogenesis of neuropathic pain in rodents. J. Clin. Investig. **123**, 2268–2286 (2013).
- Zhang, Z. L. et al. Wnt1/β-catenin signaling upregulates spinal VGLUT2 expression to control neuropathic pain in mice. *Neuropharmacology* 164, 107869 (2020).
- 339. Li, Y. S. et al. Up-regulation of the biosynthesis and release of substance P through Wnt/β-catenin signaling pathway in rat dorsal root ganglion cells. *PLoS* One **10**, e0129701 (2015).
- Bai, X. et al. Wnt3a/YTHDF1 regulated oxaliplatin-induced neuropathic pain via TNF-α/IL-18 expression in the spinal cord. *Cell Mol. Neurobiol.* 43, 1583–1594 (2023).
- Resham, K., Khare, P., Bishnoi, M. & Sharma, S. S. Neuroprotective effects of isoquercitrin in diabetic neuropathy via Wnt/β-catenin signaling pathway inhibition. *Biofactors* 46, 411–420 (2020).
- 342. Jun, J. Y. et al. Persicae semen promotes bone union in rat fractures by stimulating osteoblastogenesis through BMP-2 and Wnt signaling. *Int. J. Mol. Sci.* 24, 7388 (2023).
- 343. Genovese, T. et al. Resveratrol Inhibition of the WNT/β-catenin pathway following discogenic low back pain. *Int. J. Mol. Sci.* 23, 4092 (2022).
- 344. Deshmukh, V. et al. Modulation of the Wnt pathway through inhibition of CLK2 and DYRK1A by lorecivivint as a novel, potentially disease-modifying approach for knee osteoarthritis treatment. *Osteoarthr. Cartil.* 27, 1347–1360 (2019).
- 345. D'Amico, R. et al. Wnt/β-catenin pathway in experimental model of fibromyalgia: role of hidrox[®]. Biomedicines 9, 1683 (2021).
- 346. Yazici, Y. et al. Lorecivivint, a novel intraarticular CDC-like kinase 2 and dualspecificity tyrosine phosphorylation-regulated kinase 1A inhibitor and wnt pathway modulator for the treatment of knee osteoarthritis: a phase II randomized trial. Arthritis Rheumatol. 72, 1694–1706 (2020).
- 347. Tyagi, S. et al. Compartment-specific regulation of Na(V)1.7 in sensory neurons after acute exposure to TNF-α. *Cell Rep.* **43**, 113685 (2024).
- 348. de Lima, F. O. et al. Unveiling targets for treating postoperative pain: the role of the TNF-α/p38 MAPK/NF-κB/Nav1.8 and Nav1.9 pathways in the mouse model of incisional pain. Int. J. Mol. Sci. 23, 11630 (2022).
- Shao, J. et al. MAPK-ERK-CREB signaling pathway upregulates Nav1.6 in oxaliplatin-induced neuropathic pain in the rat. *Toxicol. Lett.* 384, 149–160 (2023).
- 350. Dai, W. L., Yan, B., Bao, Y. N., Fan, J. F. & Liu, J. H. Suppression of peripheral NGF attenuates neuropathic pain induced by chronic constriction injury through the TAK1-MAPK/NF-κB signaling pathways. *Cell Commun. Signal* **18**, 66 (2020).
- Chaumette, T. et al. c-Jun/p38MAPK/ASIC3 pathways specifically activated by nerve growth factor through TrkA are crucial for mechanical allodynia development. *Pain* 161, 1109–1123 (2020).
- 352. Zhang, Y. et al. Activation of mitogen-activated protein kinases in satellite glial cells of the trigeminal ganglion contributes to substance P-mediated inflammatory pain. *Int. J. Oral. Sci.* **11**, 24 (2019).
- 353. Reichl, S. et al. Activation of glial glutamate transporter via MAPK p38 prevents enhanced and long-lasting non-evoked resting pain after surgical incision in rats. *Neuropharmacology* **105**, 607–617 (2016).
- 354. Shin, J. et al. p38 siRNA-encapsulated PLGA nanoparticles alleviate neuropathic pain behavior in rats by inhibiting microglia activation. *Nanomedicine* 13, 1607–1621 (2018).

- 355. Ling, Y. et al. Antioxidant cascade nanoenzyme antagonize inflammatory pain by modulating MAPK/p-65 signaling pathway. *Adv. Sci.* **10**, e2206934 (2023).
- 356. Wu, D. et al. Levo-tetrahydropalmatine ameliorates neuropathic pain by inhibiting the activation of the Clec7a-MAPK/NF-κB-NLRP3 inflammasome axis. *Phytomedicine* **121**, 155075 (2023).
- Zhang, S., Li, Y. & Tu, Y. Lidocaine attenuates CFA-induced inflammatory pain in rats by regulating the MAPK/ERK/NF-κB signaling pathway. *Exp. Ther. Med.* 21, 211 (2021).
- Zhang, T. et al. Spinal endomorphins attenuate burn-injury pain in male mice by inhibiting p38 MAPK signaling pathway through the mu-opioid receptor. *Eur. J. Pharm.* **903**, 174139 (2021).
- 359. Khan, A. et al. Withametelin, a steroidal lactone, isolated from datura innoxa attenuates STZ-induced diabetic neuropathic pain in rats through inhibition of NF-kB/MAPK signaling. *Food Chem. Toxicol.* **175**, 113742 (2023).
- Wong, C. et al. mTORC2 mediates structural plasticity in distal nociceptive endings that contributes to pain hypersensitivity following inflammation. *J. Clin. Investig.* **132**, e152635 (2022).
- 361. Qiao, L., Tang, Q., An, Z. & Qi, J. Minocycline relieves neuropathic pain in rats with spinal cord injury via activation of autophagy and suppression of PI3K/Akt/ mTOR pathway. J. Pharm. Sci. 153, 12–21 (2023).
- Duan, B. et al. PI3-kinase/Akt pathway-regulated membrane insertion of acidsensing ion channel 1a underlies BDNF-induced pain hypersensitivity. J. Neurosci. 32, 6351–6363 (2012).
- Um, S. W., Kim, M. J., Leem, J. W., Bai, S. J. & Lee, B. H. Pain-relieving effects of mTOR inhibitor in the anterior cingulate cortex of neuropathic rats. *Mol. Neurobiol.* 56, 2482–2494 (2019).
- 364. Tian, Y. et al. Early single aspirin-triggered lipoxin blocked morphine antinociception tolerance through inhibiting NALP1 inflammasome: involvement of Pl3k/Akt signaling pathway. *Brain Behav. Immun.* **50**, 63–77 (2015).
- 365. Dai, W. L. et al. Selective blockade of spinal D2DR by levo-corydalmine attenuates morphine tolerance via suppressing PI3K/Akt-MAPK signaling in a MOR-dependent manner. *Exp. Mol. Med.* **50**, 1–12 (2018).
- 366. You, H. P. et al. Taselisib moderates neuropathic pain through PI3K/AKT signaling pathway in a rat model of chronic constriction injury. *Brain Res. Bull.* 199, 110671 (2023).
- 367. Bai, J. et al. Exercise facilitates the M1-to-M2 polarization of microglia by enhancing autophagy via the BDNF/AKT/mTOR pathway in neuropathic pain. *Pain Physician* 25, E1137–E1151 (2022).
- 368. Zhong, Z. et al. Bone marrow mesenchymal stem cells upregulate PI3K/AKT pathway and down-regulate NF-κB pathway by secreting glial cell-derived neurotrophic factors to regulate microglial polarization and alleviate deaf-ferentation pain in rats. *Neurobiol. Dis.* **143**, 104945 (2020).
- 369. Li, T. et al. Microglia induce the transformation of A1/A2 reactive astrocytes via the CXCR7/PI3K/Akt pathway in chronic post-surgical pain. J. Neuroinflamm. 17, 211 (2020).
- 370. Wei, J. et al. Maresin 1 promotes nerve regeneration and alleviates neuropathic pain after nerve injury. *J. Neuroinflamm.* **19**, 32 (2022).
- 371. Zhang, L. et al. Heat shock protein 22 attenuates nerve injury-induced neuropathic pain via improving mitochondrial biogenesis and reducing oxidative stress mediated by spinal AMPK/PGC-1α pathway in male rats. J. Neuroimmune Pharm. 19, 5 (2024).
- 372. Sun, J. et al. Sestrin2 overexpression attenuates osteoarthritis pain via induction of AMPK/PGC-1α-mediated mitochondrial biogenesis and suppression of neuroinflammation. *Brain Behav. Immun.* **102**, 53–70 (2022).
- 373. Wang, S. et al. Negative regulation of TRPA1 by AMPK in primary sensory neurons as a potential mechanism of painful diabetic neuropathy. *Diabetes* 67, 98–109 (2018).
- Zheng, T. et al. Salidroside alleviates diabetic neuropathic pain through regulation of the AMPK-NLRP3 inflammasome axis. *Toxicol. Appl Pharm.* 416, 115468 (2021).
- 375. Lu, G. et al. AMPK activation attenuates central sensitization in a recurrent nitroglycerin-induced chronic migraine mouse model by promoting microglial M2-type polarization. *J. Headache Pain.* **25**, 29 (2024).
- Chen, M. et al. Endocannabinoid biosynthetic enzymes regulate pain response via LKB1-AMPK signaling. Proc. Natl Acad. Sci. USA 120, e2304900120 (2023).
- 377. Xiao, P. Y. et al. UNC5B overexpression alleviates peripheral neuropathic pain by stimulating netrin-1-dependent autophagic flux in Schwann cells. *Mol. Neurobiol.* 59, 5041–5055 (2022).
- 378. Shoskes, D. A. et al. The urinary microbiome differs significantly between patients with chronic prostatitis/chronic pelvic pain syndrome and controls as well as between patients with different clinical phenotypes. *Urology* **92**, 26–32 (2016).
- 379. Hou, K. et al. Microbiota in health and diseases. *Signal Transduct. Target Ther.* **7**, 135 (2022).
- Zhou, S. Y. et al. FODMAP diet modulates visceral nociception by lipopolysaccharide-mediated intestinal inflammation and barrier dysfunction. J. Clin. Invest 128, 267–280 (2018).

- 381. Dansereau, M. A. et al. Mechanistic insights into the role of the chemokine CCL2/ CCR2 axis in dorsal root ganglia to peripheral inflammation and pain hypersensitivity. *J. Neuroinflamm.* **18**, 79 (2021).
- 382. Tan, X. et al. Bone marrow mesenchymal stem cells alleviate stress-induced hyperalgesia via restoring gut microbiota and inhibiting neuroinflammation in the spinal cord by targeting the AMPK/NF-κB signaling pathway. *Life Sci.* **314**, 121318 (2023).
- 383. Lee, J., Lee, G., Ko, G. & Joong Lee, S. Nerve injury-induced gut dysbiosis contributes to spinal cord TNF-α expression and nociceptive sensitization. *Brain Behav. Immun.* **110**, 155–161 (2023).
- Lyons, D. E., Beery, J. T., Lyons, S. A. & Taylor, S. L. Cadaverine and aminoguanidine potentiate the uptake of histamine in vitro in perfused intestinal segments of rats. *Toxicol. Appl Pharm.* **70**, 445–458 (1983).
- Hoffmann, K. M., Gruber, E., Deutschmann, A., Jahnel, J. & Hauer, A. C. Histamine intolerance in children with chronic abdominal pain. *Arch. Dis. Child* 98, 832–833 (2013).
- De Palma, G. et al. Histamine production by the gut microbiota induces visceral hyperalgesia through histamine 4 receptor signaling in mice. *Sci. Transl. Med.* 14, eabj1895 (2022).
- Wouters, M. M. et al. Histamine receptor H1-mediated sensitization of TRPV1 mediates visceral hypersensitivity and symptoms in patients with irritable bowel syndrome. *Gastroenterology* 150, 875–887.e879 (2016).
- Rudick, C. N., Bryce, P. J., Guichelaar, L. A., Berry, R. E. & Klumpp, D. J. Mast cellderived histamine mediates cystitis pain. *PLoS One* 3, e2096 (2008).
- Roshchina, V. V. New trends and perspectives in the evolution of neurotransmitters in microbial, plant, and animal cells. Adv. Exp. Med. Biol. 874, 25–77 (2016).
- 390. Labus, J. S. et al. Evidence for an association of gut microbial Clostridia with brain functional connectivity and gastrointestinal sensorimotor function in patients with irritable bowel syndrome, based on tripartite network analysis. *Microbiome* 7, 45 (2019).
- Feng, C. C. et al. Vagal anandamide signaling via cannabinoid receptor 1 contributes to luminal 5-HT modulation of visceral nociception in rats. *Pain* 155, 1591–1604 (2014).
- 392. Ling, X. et al. Effects of Chang-Kang-Fang formula on the microbiota-gut-brain axis in rats with irritable bowel syndrome. *Front. Pharm.* **13**, 778032 (2022).
- 393. Cao, D. Y., Bai, G., Ji, Y. & Traub, R. J. Epigenetic upregulation of metabotropic glutamate receptor 2 in the spinal cord attenuates oestrogen-induced visceral hypersensitivity. *Gut* 64, 1913–1920 (2015).
- Clos-Garcia, M. et al. Gut microbiome and serum metabolome analyses identify molecular biomarkers and altered glutamate metabolism in fibromyalgia. *EBioMedicine* 46, 499–511 (2019).
- Strandwitz, P. et al. GABA-modulating bacteria of the human gut microbiota. Nat. Microbiol. 4, 396–403 (2019).
- Royo, F. et al. Orally administered bifidobacterium adolescentis diminishes serum glutamate concentration in mice. *Microbiol. Spectr.* 11, e0506322 (2023).
- 397. Laroute, V. et al. Lactococcus lactis NCDO2118 exerts visceral antinociceptive properties in rat via GABA production in the gastro-intestinal tract. *Elife* 11, e77100 (2022).
- Dhakal, R., Bajpai, V. K. & Baek, K. H. Production of gaba (γ-Aminobutyric acid) by microorganisms: a review. *Braz. J. Microbiol* 43, 1230–1241 (2012).
- 399. Tanga, F. Y., Nutile-McMenemy, N. & DeLeo, J. A. The CNS role of Toll-like receptor 4 in innate neuroimmunity and painful neuropathy. *Proc. Natl Acad. Sci.* USA 102, 5856–5861 (2005).
- 400. Diogenes, A., Ferraz, C. C., Akopian, A. N., Henry, M. A. & Hargreaves, K. M. LPS sensitizes TRPV1 via activation of TLR4 in trigeminal sensory neurons. *J. Dent. Res.* **90**, 759–764 (2011).
- Lian, N. et al. Drinking hydrogen-rich water alleviates chemotherapy-induced neuropathic pain through the regulation of gut microbiota. J. Pain Res. 14, 681–691 (2021).
- 402. Shen, S. et al. Gut microbiota is critical for the induction of chemotherapyinduced pain. *Nat. Neurosci.* **20**, 1213–1216 (2017).
- Wardill, H. R. et al. Irinotecan-induced gastrointestinal dysfunction and pain are mediated by common TLR4-dependent mechanisms. *Mol. Cancer Ther.* 15, 1376–1386 (2016).
- 404. Grabauskas, G. et al. Gut microbiota alter visceral pain sensation and inflammation via modulation of synthesis of resolvin D1 in colonic tuft cells. *Gastroenterology* **S0016-5085**, 00829–0 (2022).
- 405. Tramullas, M., Finger, B. C., Dinan, T. G. & Cryan, J. F. Obesity takes its toll on visceral pain: high-fat diet induces toll-like receptor 4-dependent visceral hypersensitivity. *PLoS One* **11**, e0155367 (2016).
- 406. Mallaret, G. et al. Involvement of toll-like receptor 5 in mouse model of colonic hypersensitivity induced by neonatal maternal separation. *World J. Gastroenterol.* 28, 3903–3916 (2022).
- Das, N. et al. HMGB1 activates proinflammatory signaling via TLR5 leading to allodynia. *Cell Rep.* 17, 1128–1140 (2016).

- 42
- Xu, Z. Z. et al. Inhibition of mechanical allodynia in neuropathic pain by TLR5mediated A-fiber blockade. *Nat. Med.* 21, 1326–1331 (2015).
- Boonen, B. et al. Differential effects of lipopolysaccharide on mouse sensory TRP channels. *Cell Calcium* 73, 72–81 (2018).
- Meseguer, V. et al. TRPA1 channels mediate acute neurogenic inflammation and pain produced by bacterial endotoxins. *Nat. Commun.* 5, 3125 (2014).
- Sessenwein, J. L. et al. Protease-mediated suppression of DRG neuron excitability by commensal bacteria. J. Neurosci. 37, 11758–11768 (2017).
- Ochoa-Cortes, F. et al. Bacterial cell products signal to mouse colonic nociceptive dorsal root ganglia neurons. *Am. J. Physiol. Gastrointest. Liver Physiol.* 299, G723–G732 (2010).
- Vetvik, K. G. & MacGregor, E. A. Sex differences in the epidemiology, clinical features, and pathophysiology of migraine. *Lancet Neurol.* 16, 76–87 (2017).
- 414. van der Horst-Bruinsma, I. E., Zack, D. J., Szumski, A. & Koenig, A. S. Female patients with ankylosing spondylitis: analysis of the impact of gender across treatment studies. *Ann. Rheum. Dis.* **72**, 1221–1224 (2013).
- 415. Mogil, J. S. Qualitative sex differences in pain processing: emerging evidence of a biased literature. *Nat. Rev. Neurosci.* **21**, 353–365 (2020).
- 416. Vacca, V. et al. Higher pain perception and lack of recovery from neuropathic pain in females: a behavioural, immunohistochemical, and proteomic investigation on sex-related differences in mice. *Pain* **155**, 388–402 (2014).
- 417. Navratilova, E., Fillingim, R. B. & Porreca, F. Sexual dimorphism in functional pain syndromes. *Sci. Transl. Med.* **13**, eabj7180 (2021).
- Serrano, N. E., Saputra, S. G., Íbias, J., Company, M. & Nazarian, A. Pain-induced impulsivity is sexually dimorphic and mu-opioid receptor sensitive in rats. *Psychopharmacology* 238, 3447–3462 (2021).
- 419. Jeon, S. M., Chang, D., Geske, A., Ginty, D. D. & Caterina, M. J. Sex-dependent reduction in mechanical allodynia in the sural-sparing nerve injury model in mice lacking merkel cells. *J. Neurosci.* **41**, 5595–5619 (2021).
- Christov-Moore, L. & lacoboni, M. Sex differences in somatomotor representations of others' pain: a permutation-based analysis. *Brain Struct. Funct.* 224, 937–947 (2019).
- 421. Ray, P. R. et al. RNA profiling of human dorsal root ganglia reveals sex differences in mechanisms promoting neuropathic pain. *Brain* **146**, 749–766 (2023).
- Johnston, K. J. A. et al. Sex-stratified genome-wide association study of multisite chronic pain in UK Biobank. *PLoS Genet.* 17, e1009428 (2021).
- Baggio, D. F. et al. Sex dimorphism in resolvin d5-induced analgesia in rat models of trigeminal pain. J. Pain. 24, 717–729 (2023).
- 424. Kaur, S. et al. Sex differences and estrous cycle effects of peripheral serotoninevoked rodent pain behaviors. *Neuroscience* **384**, 87–100 (2018).
- 425. Xu, Z. et al. Positive interaction between GPER and β-alanine in the dorsal root ganglion uncovers potential mechanisms: mediating continuous neuronal sensitization and neuroinflammation responses in neuropathic pain. J. Neuroinflamm. **19**, 164 (2022).
- 426. Lu, Y. C., Chen, C. W., Wang, S. Y. & Wu, F. S. 17Beta-estradiol mediates the sex difference in capsaicin-induced nociception in rats. J. Pharm. Exp. Ther. 331, 1104–1110 (2009).
- Paige, C. et al. Neuroendocrine mechanisms governing sex differences in hyperalgesic priming involve prolactin receptor sensory neuron signaling. J. Neurosci. 40, 7080–7090 (2020).
- 428. Wu, Y. W. et al. Hippocampal nerve growth factor potentiated by 17β-estradiol and involved in allodynia of inflamed TMJ in rat. *J. Pain* **13**, 555–563 (2012).
- 429. Hornung, R., Pritchard, A., Kinchington, P. R. & Kramer, P. R. Comparing gene expression in the parabrachial and amygdala of diestrus and proestrus female rats after orofacial varicella zoster injection. *Int. J. Mol. Sci.* **21**, 5749 (2020).
- 430. Xie, Z. et al. Estrogen metabolites increase nociceptor hyperactivity in a mouse model of uterine pain. *JCI Insight* **7**, e149107 (2022).
- 431. Butkevich, I. P., Barr, G. A. & Vershinina, E. A. Sex differences in formalin-induced pain in prenatally stressed infant rats. *Eur. J. Pain* **11**, 888–894 (2007).
- 432. Patil, M. et al. Prolactin regulates pain responses via a female-selective nociceptor-specific mechanism. *iScience* **20**, 449–465 (2019).
- Chen, Y. et al. The prolactin receptor long isoform regulates nociceptor sensitization and opioid-induced hyperalgesia selectively in females. *Sci. Transl. Med.* 12, eaay7550 (2020).
- Avona, A. et al. Meningeal CGRP-prolactin interaction evokes female-specific migraine behavior. Ann. Neurol. 89, 1129–1144 (2021).
- Watanabe, M. et al. Dysregulation of serum prolactin links the hypothalamus with female nociceptors to promote migraine. *Brain* 145, 2894–2909 (2022).
- 436. Prusator, D. K. & Greenwood-Van Meerveld, B. Amygdala-mediated mechanisms regulate visceral hypersensitivity in adult females following early life stress: importance of the glucocorticoid receptor and corticotropin-releasing factor. *Pain* **158**, 296–305 (2017).
- 437. Ferrari, L. F., Araldi, D., Green, P. G. & Levine, J. D. Marked sexual dimorphism in neuroendocrine mechanisms for the exacerbation of paclitaxel-induced painful peripheral neuropathy by stress. *Pain* **161**, 865–874 (2020).

- 438. Shen, B. Q., Sankaranarayanan, I., Price, T. J. & Tavares-Ferreira, D. Sex-differences in prostaglandin signaling: a semi-systematic review and characterization of PTGDS expression in human sensory neurons. *Sci. Rep.* **13**, 4670 (2023).
- Maddahi, A., Edvinsson, L. & Warfvinge, K. Expression of vasopressin and its receptors in migraine-related regions in CNS and the trigeminal system: influence of sex. J. Headache Pain 23, 152 (2022).
- Labus, J. S. et al. Sex differences in emotion-related cognitive processes in irritable bowel syndrome and healthy control subjects. *Pain* **154**, 2088–2099 (2013).
- 441. Zhang, H. et al. The role of negative emotions in sex differences in pain sensitivity. *Neuroimage* **245**, 118685 (2021).
- 442. Da Silva, J. T. et al. Sex differences in visceral sensitivity and brain activity in a rat model of comorbid pain: a longitudinal study. *Pain* **165**, 698–706 (2023).
- 443. Cantu, D. J., Kaur, S., Murphy, A. Z. & Averitt, D. L. Sex differences in the amygdaloid projections to the ventrolateral periaqueductal gray and their activation during inflammatory pain in the rat. J. Chem. Neuroanat. **124**, 102123 (2022).
- 444. Levine, A. et al. Sex differences in the expression of the endocannabinoid system within V1M cortex and PAG of Sprague Dawley rats. *Biol. Sex. Differ.* **12**, 60 (2021).
- 445. Llorente-Berzal, A. et al. Sexually dimorphic expression of fear-conditioned analgesia in rats and associated alterations in the endocannabinoid system in the periaqueductal grey. *Neuroscience* **480**, 117–130 (2022).
- 446. Maleki, N. et al. Her versus his migraine: multiple sex differences in brain function and structure. *Brain* **135**, 2546–2559 (2012).
- 447. Agalave, N. M. et al. Sex-dependent role of microglia in disulfide high mobility group box 1 protein-mediated mechanical hypersensitivity. *Pain* **162**, 446–458 (2021).
- Doyle, H. H., Eidson, L. N., Sinkiewicz, D. M. & Murphy, A. Z. Sex differences in microglia activity within the periaqueductal gray of the rat: a potential mechanism driving the dimorphic effects of morphine. *J. Neurosci.* 37, 3202–3214 (2017).
- 449. Friedman, T. N. et al. Sex differences in peripheral immune cell activation: Implications for pain and pain resolution. *Brain Behav. Immun.* **114**, 80–93 (2023).
- 450. Shen, Y. et al. SETD7 mediates spinal microgliosis and neuropathic pain in a rat model of peripheral nerve injury. *Brain Behav. Immun.* 82, 382–395 (2019).
- Mapplebeck, J. C. S. et al. Microglial P2X4R-evoked pain hypersensitivity is sexually dimorphic in rats. *Pain* 159, 1752–1763 (2018).
- 452. Del Rivero, T., Fischer, R., Yang, F., Swanson, K. A. & Bethea, J. R. Tumor necrosis factor receptor 1 inhibition is therapeutic for neuropathic pain in males but not in females. *Pain* **160**, 922–931 (2019).
- 453. Inyang, K. E. et al. The antidiabetic drug metformin prevents and reverses neuropathic pain and spinal cord microglial activation in male but not female mice. *Pharm. Res.* **139**, 1–16 (2019).
- 454. Ji, J. et al. IL-23 enhances C-fiber-mediated and blue light-induced spontaneous pain in female mice. *Front. Immunol.* **12**, 787565 (2021).
- 455. Mun, C. J. et al. Sex differences in interleukin-6 responses over time following laboratory pain testing among patients with knee osteoarthritis. J. Pain 21, 731–741 (2020).
- Hashmi, J. A. & Davis, K. D. Women experience greater heat pain adaptation and habituation than men. *Pain* 145, 350–357 (2009).
- 457. Mifflin, K. A. et al. Voluntary wheel running reveals sex-specific nociceptive factors in murine experimental autoimmune encephalomyelitis. *Pain* **160**, 870–881 (2019).
- 458. Lee, S. et al. Voluntary running attenuates behavioural signs of low back pain: dimorphic regulation of intervertebral disc inflammation in male and female SPARC-null mice. Osteoarthr. Cartil. **30**, 110–123 (2022).
- 459. Won, S., Park, K., Lim, H. & Lee, S. J. Sexual dimorphism in cognitive disorders in a murine model of neuropathic pain. *Behav. Brain Funct.* 16, 1 (2020).
- Morales-Medina, J. C., Flores, G., Vallelunga, A., Griffiths, N. H. & lannitti, T. Cerebrolysin improves peripheral inflammatory pain: Sex differences in two models of acute and chronic mechanical hypersensitivity. *Drug Dev. Res.* 80, 513–518 (2019).
- Liu, R. H., Xue, M., Li, X. H. & Zhuo, M. Sex difference in synaptic plasticity in the anterior cingulate cortex of adult mice. *Mol. Brain* 13, 41 (2020).
- 462. Frank, P. et al. Association between depression and physical conditions requiring hospitalization. JAMA Psychiatry 80, 690–699 (2023).
- 463. Maneeton, N., Maneeton, B. & Srisurapanont, M. Prevalence and predictors of pain in patients with major depressive disorder. *Asian J. Psychiatr.* 6, 288–291 (2013).
- 464. Sellmeijer, J. et al. Hyperactivity of anterior cingulate cortex areas 24a/24b drives chronic pain-induced anxiodepressive-like consequences. J. Neurosci. 38, 3102–3115 (2018).
- 465. Wu, K. et al. The microglial innate immune receptors TREM-1 and TREM-2 in the anterior cingulate cortex (ACC) drive visceral hypersensitivity and depressive-

like behaviors following DSS-induced colitis. *Brain Behav. Immun.* **112**, 96–117 (2023).

- 466. Albrecht, D. S. et al. The neuroinflammatory component of negative affect in patients with chronic pain. *Mol. Psychiatry* **26**, 864–874 (2021).
- Barthas, F. et al. Cingulate overexpression of mitogen-activated protein kinase phosphatase-1 as a key factor for depression. *Biol. Psychiatry* 82, 370–379 (2017).
- 468. Becker, L. J. et al. The basolateral amygdala-anterior cingulate pathway contributes to depression-like behaviors and comorbidity with chronic pain behaviors in male mice. *Nat. Commun.* **14**, 2198 (2023).
- He, S., Huang, X., Zheng, J., Zhang, Y. & Ruan, X. An NTS-CeA projection modulates depression-like behaviors in a mouse model of chronic pain. *Neurobiol. Dis.* **174**, 105893 (2022).
- Zhou, W. et al. A neural circuit for comorbid depressive symptoms in chronic pain. *Nat. Neurosci.* 22, 1649–1658 (2019).
- 471. Ding, X. et al. LncRNA XR_351665 contributes to chronic pain-induced depression by upregulating DNMT1 via sponging miR-152-3p. *J. Pain* **24**, 449–462 (2023).
- Ding, X. et al. DNMT1 mediates chronic pain-related depression by inhibiting GABAergic neuronal activation in the central amygdala. *Biol. Psychiatry* 94, 672–684 (2023).
- 473. Mecca, C. M. et al. Dynamic change of endocannabinoid signaling in the medial prefrontal cortex controls the development of depression after neuropathic pain. J. Neurosci. 41, 7492–7508 (2021).
- 474. Llorca-Torralba, M. et al. Pain and depression comorbidity causes asymmetric plasticity in the locus coeruleus neurons. *Brain* **145**, 154–167 (2022).
- Cervenka, I., Agudelo, L. Z. & Ruas, J. L. Kynurenines: tryptophan's metabolites in exercise, inflammation, and mental health. *Science* 357, eaaf9794 (2017).
- 476. Myint, K. et al. Psychological Stresses in children trigger cytokine—and kynurenine metabolite-mediated abdominal pain and proinflammatory changes. *Front. Immunol.* **12**, 702301 (2021).
- Binvignat, M. et al. Serum tryptophan metabolites are associated with erosive hand osteoarthritis and pain: results from the DIGICOD cohort. *Osteoarthr. Cartil.* 31, 1132–1143 (2023).
- Barjandi, G. et al. Plasma tryptophan and kynurenine in females with temporomandibular disorders and fibromyalgia-An exploratory pilot study. J. Oral. Rehabil. 47, 150–157 (2020).
- Maganin, A. G. et al. Meningeal dendritic cells drive neuropathic pain through elevation of the kynurenine metabolic pathway in mice. J. Clin. Investig. 132, e153805 (2022).
- Csupak, B., Sommer, J. L., Jacobsohn, E. & El-Gabalawy, R. A population-based examination of the co-occurrence and functional correlates of chronic pain and generalized anxiety disorder. J. Anxiety Disord. 56, 74–80 (2018).
- Liang, H. Y. et al. nNOS-expressing neurons in the vmPFC transform pPVTderived chronic pain signals into anxiety behaviors. *Nat. Commun.* 11, 2501 (2020).
- 482. Shao, S. et al. Sex-dependent expression of N-cadherin-GluA1 pathway-related molecules in the prefrontal cortex mediates anxiety-like behavior in male offspring following prenatal stress. Stress 24, 612–620 (2021).
- Gao, F. et al. Elevated prelimbic cortex-to-basolateral amygdala circuit activity mediates comorbid anxiety-like behaviors associated with chronic pain. J. Clin. Investig. 133, e166356 (2023).
- 484. Zhang, W. W. et al. Tachykinin receptor 3 in the lateral habenula alleviates pain and anxiety comorbidity in mice. *Front. Immunol.* **14**, 1049739 (2023).
- 485. Yamauchi, N. et al. Chronic pain-induced neuronal plasticity in the bed nucleus of the stria terminalis causes maladaptive anxiety. *Sci. Adv.* **8**, eabj5586 (2022).
- 486. Wang, D. et al. Lateral septum-lateral hypothalamus circuit dysfunction in comorbid pain and anxiety. *Mol. Psychiatry* **28**, 1090–1100 (2023).
- 487. Sun, J. et al. Polygenic evidence and overlapped brain functional connectivities for the association between chronic pain and sleep disturbance. *Transl. Psychiatry* **10**, 252 (2020).
- Sun, H. et al. A common neuronal ensemble in nucleus accumbens regulates pain-like behaviour and sleep. *Nat. Commun.* 14, 4700 (2023).
- Cardis, R. et al. Cortico-autonomic local arousals and heightened somatosensory arousability during NREMS of mice in neuropathic pain. *Elife* 10, e65835 (2021).
- Ding, W. et al. The endocannabinoid N-arachidonoyl dopamine is critical for hyperalgesia induced by chronic sleep disruption. *Nat. Commun.* 14, 6696 (2023).
- Zhou, H., Li, M., Zhao, R., Sun, L. & Yang, G. A sleep-active basalocortical pathway crucial for generation and maintenance of chronic pain. *Nat. Neurosci.* 26, 458–469 (2023).
- Wang, X. Y. et al. A glutamatergic DRN-VTA pathway modulates neuropathic pain and comorbid anhedonia-like behavior in mice. *Nat. Commun.* 14, 5124 (2023).
- 493. Yue, C. et al. The role of the gut-microbiota-brain axis via the subdiaphragmatic vagus nerve in chronic inflammatory pain and comorbid spatial working

memory impairment in complete Freund's adjuvant mice. J. Psychiatr. Res. 166, 61–73 (2023).

- 494. Jiang, H. et al. Contribution of AMPA receptor-mediated LTD in LA/BLA-CeA pathway to comorbid aversive and depressive symptoms in neuropathic pain. J. Neurosci. 41, 7278–7299 (2021).
- 495. Li, Z. Z. et al. Extracellular matrix protein laminin β1 regulates pain sensitivity and anxiodepression-like behaviors in mice. J. Clin. Investig. **131**, e146323 (2021).
- 496. Pereira-Silva, R. et al. Attenuation of the diffuse noxious inhibitory controls in chronic joint inflammatory pain is accompanied by anxiodepressive-like behaviors and impairment of the descending noradrenergic modulation. *Int. J. Mol. Sci.* **21**, 2973 (2020).
- Krause, S. J. & Backonja, M.-M. Development of a neuropathic pain questionnaire. *Clin. J. Pain* 19, 306–314 (2003).
- 498. Freynhagen, R., Baron, R., Gockel, U. & Tölle, T. R. painDETECT: a new screening questionnaire to identify neuropathic components in patients with back pain. *Curr. Med. Res. Opin.* 22, 1911–1920 (2006).
- 499. Gauffin, J., Hankama, T., Kautiainen, H., Hannonen, P. & Haanpää, M. Neuropathic pain and use of Pain DETECT in patients with fibromyalgia: a cohort study. *BMC Neurol.* **13**, 21 (2013).
- 500. Fishbain, D. A. et al. Can the neuropathic pain scale discriminate between nonneuropathic and neuropathic pain? *Pain. Med.* 9, 149–160 (2008).
- Zelman, D. C., Gore, M., Dukes, E., Tai, K.-S. & Brandenburg, N. Validation of a modified version of the brief pain inventory for painful diabetic peripheral neuropathy. *J. Pain. Symptom Manag.* **29**, 401–410 (2005).
- Perrot, S., Trouvin, A.-P. & Bouhassira, D. Three dimensions of pain in osteoarthritis: development and validation of the Osteoarthritis Symptom Inventory Scale. *Pain* 164, 1566–1577 (2023).
- 503. Rolke, R. et al. Quantitative sensory testing: a comprehensive protocol for clinical trials. *Eur. J. Pain* **10**, 77–88 (2006).
- Uddin, Z. & MacDermid, J. C. Quantitative sensory testing in chronic musculoskeletal pain. *Pain. Med.* 17, 1694–1703 (2016).
- 505. Siao, P. & Cros, D. P. Quantitative sensory testing. Phys. Med. Rehabil. Clin. North Am. 14, 261–286 (2003).
- Mücke, M. et al. Quantitative sensory testing (QST). English version. Schmerz 35, 153–160 (2021).
- 507. Magrinelli, F. et al. The association between serum cytokines and damage to large and small nerve fibers in diabetic peripheral neuropathy. J. Diabetes Res. 2015, 547834 (2015).
- Erbüyün, S. C., Ovalı, G. Y., Özyurt, B. & Yentür, E. A. Lumbar disc herniation, the association between quantitative sensorial test and magnetic resonance imaging findings. *Pain. Med.* **19**, 1373–1381 (2018).
- Flor, H. et al. Subtle sensory abnormalities detected by quantitative sensory testing in patients with trigeminal neuralgia. *Pain Physician* 19, 507–518 (2016).
- Zhou, P., Chen, Y., Zhang, J., Wang, K. & Svensson, P. Quantitative sensory testing for assessment of somatosensory function in human oral mucosa: a review. Acta Odontol. Scand. 76, 13–20 (2018).
- Krumova, E. K., Geber, C., Westermann, A. & Maier, C. Neuropathic pain: is quantitative sensory testing helpful? *Curr. Diabetes Rep.* 12, 393–402 (2012).
- 512. Katz, N. P., Paillard, F. C. & Edwards, R. R. Review of the performance of quantitative sensory testing methods to detect hyperalgesia in chronic pain patients on long-term opioids. *Anesthesiology* **122**, 677–685 (2015).
- Truini, A., Garcia-Larrea, L. & Cruccu, G. Reappraising neuropathic pain in humans-how symptoms help disclose mechanisms. *Nat. Rev. Neurol.* 9, 572–582 (2013).
- ST4. Cruccu, G. et al. AAN-EFNS guidelines on trigeminal neuralgia management. *Eur. J. Neurol.* 15, 1013–1028 (2008).
- 515. Mano, H. et al. Classification and characterisation of brain network changes in chronic back pain: a multicenter study. *Wellcome Open Res.* 3, 19 (2018).
- 516. López-Solà, M. et al. Towards a neurophysiological signature for fibromyalgia. *Pain* **158**, 34–47 (2017).
- Ung, H. et al. Multivariate classification of structural MRI data detects chronic low back pain. Cereb. Cortex 24, 1037–1044 (2014).
- Labus, J. S. et al. Multivariate morphological brain signatures predict patients with chronic abdominal pain from healthy control subjects. *Pain* 156, 1545–1554 (2015).
- Ta Dinh, S. et al. Brain dysfunction in chronic pain patients assessed by restingstate electroencephalography. *Pain* 160, 2751–2765 (2019).
- 520. Levitt, J. et al. Pain phenotypes classified by machine learning using electroencephalography features. *NeuroImage* **223**, 117256 (2020).
- 521. Zolezzi, D. M., Maria Alonso-Valerdi, L., Naal-Ruiz, N. E. & Ibarra-Zarate, D. I. Identification of neuropathic pain severity based on linear and non-linear EEG features. Annu. Int. Conf. IEEE Eng. Med. Biol. Soc. 2021, 169–173 (2021).
- 522. Makary, M. M. et al. Loss of nucleus accumbens low-frequency fluctuations is a signature of chronic pain. Proc. Natl Acad. Sci. USA 117, 10015–10023 (2020).

- 44
- Gohel, S. R. & Biswal, B. B. Functional integration between brain regions at rest occurs in multiple-frequency bands. *Brain Connect.* 5, 23–34 (2015).
- Bosma, R. L. et al. Brain dynamics and temporal summation of pain predicts neuropathic pain relief from ketamine infusion. *Anesthesiology* **129**, 1015–1024 (2018).
- 525. Baliki, M. N. et al. Corticostriatal functional connectivity predicts transition to chronic back pain. *Nat. Neurosci.* **15**, 1117–1119 (2012).
- Kutch, J. J. et al. Resting-state functional connectivity predicts longitudinal pain symptom change in urologic chronic pelvic pain syndrome: a MAPP network study. *Pain* **158**, 1069–1082 (2017).
- Veluchamy, A., Hébert, H. L., Meng, W., Palmer, C. N. A. & Smith, B. H. Systematic review and meta-analysis of genetic risk factors for neuropathic pain. *Pain* 159, 825–848 (2018).
- 528. Suri, P. et al. Genome-wide meta-analysis of 158,000 individuals of European ancestry identifies three loci associated with chronic back pain. *PLoS Genet.* **14**, e1007601 (2018).
- 529. Meloto, C. B. et al. Human pain genetics database: a resource dedicated to human pain genetics research. *Pain* **159**, 749–763 (2018).
- Hu, L., Yin, W., Ma, Y., Zhang, Q. & Xu, Q. Gene expression signature of human neuropathic pain identified through transcriptome analysis. *Front. Genet.* 14, 1127167 (2023).
- Xu, J. et al. Tumor necrosis factor-alpha is a potential diagnostic biomarker for chronic neuropathic pain after spinal cord injury. *Neurosci. Lett.* 595, 30–34 (2015).
- Breidthardt, T. et al. Inflammatory biomarkers and clinical judgment in the emergency diagnosis of urgent abdominal pain. *Clin. Chem.* 65, 302–312 (2019).
- Drosdzol-Cop, A., Skrzypulec-Plinta, V. & Stojko, R. Serum and peritoneal fluid immunological markers in adolescent girls with chronic pelvic pain. *Obstetr. Gynecol. Surv.* 67, 374–381 (2012).
- 534. Gunn, J., Hill, M. M., Cotten, B. M. & Deer, T. R. An analysis of biomarkers in patients with chronic pain. *Pain. Physician* **23**, E41–E49 (2020).
- 535. Amirdelfan, K. et al. Clinical validation of a multi-biomarker assay for the evaluation of chronic pain patients in a cross-sectional, observational study. *Pain. Ther.* 9, 511–529 (2020).
- Bodei, L., Herrmann, K., Schöder, H., Scott, A. M. & Lewis, J. S. Radiotheranostics in oncology: current challenges and emerging opportunities. *Nat. Rev. Clin. Oncol.* 19, 534–550 (2022).
- Vonk, J., de Wit, J. G., Voskuil, F. J. & Witjes, M. J. H. Improving oral cavity cancer diagnosis and treatment with fluorescence molecular imaging. *Oral. Dis.* 27, 21–26 (2021).
- 538. Devigili, G. et al. Diagnostic criteria for small fibre neuropathy in clinical practice and research. *Brain J. Neurol.* **142**, 3728–3736 (2019).
- 539. Truini, A. et al. Joint European Academy of Neurology-European Pain Federation-Neuropathic Pain Special Interest Group of the International Association for the Study of Pain guidelines on neuropathic pain assessment. *Eur. J. Neurol.* **30**, 2177–2196 (2023).
- 540. European Federation of Neurological Societies/Peripheral Nerve Society Guideline on the use of skin biopsy in the diagnosis of small fiber neuropathy. Report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society. J. Peripheral Nervous Syst. 15, 79–92 (2010).
- Pitarokoili, K., Gold, R. & Fisse, A. L. Nerve ultrasound for the diagnosis and follow-up of peripheral neuropathies. *Curr. Opin. Neurol.* 36, 373–381 (2023).
- 542. Truini, A. et al. Does the epidermal nerve fibre density measured by skin biopsy in patients with peripheral neuropathies correlate with neuropathic pain? *Pain* 155, 828–832 (2014).
- Preisner, A. et al. Non-steroidal anti-inflammatory drug indometacin enhances endogenous remyelination. Acta Neuropathol. 130, 247–261 (2015).
- Khasabova, I. A. et al. Sensitization of nociceptors by prostaglandin E(2)-glycerol contributes to hyperalgesia in mice with sickle cell disease. *Blood* 133, 1989–1998 (2019).
- 545. Wei, H. L. et al. Disrupted functional connectivity of the amygdala predicts the efficacy of non-steroidal anti-inflammatory drugs in migraineurs without aura. *Front. Mol. Neurosci.* **15**, 819507 (2022).
- Aldamluji, N., Burgess, A., Pogatzki-Zahn, E., Raeder, J. & Beloeil, H. PROSPECT guideline for tonsillectomy: systematic review and procedure-specific postoperative pain management recommendations. *Anaesthesia* 76, 947–961 (2021).
- Qaseem, A. et al. Management of acute and recurrent gout: a clinical practice guideline from the American College of Physicians. *Ann. Intern. Med.* 166, 58–68 (2017).
- 548. Qaseem, A. et al. Nonpharmacologic and pharmacologic management of acute pain from non-low back, musculoskeletal injuries in adults: a clinical guideline from the American College of Physicians and American Academy of Family Physicians. Ann. Intern. Med. **173**, 739–748 (2020).
- Arden, N. K. et al. Non-surgical management of knee osteoarthritis: comparison of ESCEO and OARSI 2019 guidelines. *Nat. Rev. Rheumatol.* 17, 59–66 (2021).

- Yalcin, I. et al. beta(2)-adrenoceptors are critical for antidepressant treatment of neuropathic pain. Ann. Neurol. 65, 218–225 (2009).
- 551. Benbouzid, M. et al. Delta-opioid receptors are critical for tricyclic antidepressant treatment of neuropathic allodynia. *Biol. Psychiatry* **63**, 633–636 (2008).
- 552. Deng, L., Lee, W. H., Xu, Z., Makriyannis, A. & Hohmann, A. G. Prophylactic treatment with the tricyclic antidepressant desipramine prevents development of paclitaxel-induced neuropathic pain through activation of endogenous analgesic systems. *Pharm. Res.* **114**, 75–89 (2016).
- 553. Barnet, C. S., Tse, J. Y. & Kohane, D. S. Site 1 sodium channel blockers prolong the duration of sciatic nerve blockade from tricyclic antidepressants. *Pain* **110**, 432–438 (2004).
- 554. Tai, Y. H. et al. Amitriptyline preserves morphine's antinociceptive effect by regulating the glutamate transporter GLAST and GLT-1 trafficking and excitatory amino acids concentration in morphine-tolerant rats. *Pain* **129**, 343–354 (2007).
- 555. Tai, Y. H. et al. Amitriptyline suppresses neuroinflammation and up-regulates glutamate transporters in morphine-tolerant rats. *Pain* **124**, 77–86 (2006).
- Scholz, J. et al. The IASP classification of chronic pain for ICD-11: chronic neuropathic pain. *Pain* 160, 53–59 (2019).
- 557. Ansari, A. The efficacy of newer antidepressants in the treatment of chronic pain: a review of current literature. *Harv. Rev. Psychiatry* 7, 257–277 (2000).
- 558. Tesfaye, S. et al. Comparison of amitriptyline supplemented with pregabalin, pregabalin supplemented with amitriptyline, and duloxetine supplemented with pregabalin for the treatment of diabetic peripheral neuropathic pain (OPTION-DM): a multicentre, double-blind, randomised crossover trial. *Lancet* **400**, 680–690 (2022).
- 559. Guirimand, F., Dupont, X., Brasseur, L., Chauvin, M. & Bouhassira, D. The effects of ketamine on the temporal summation (wind-up) of the R(III) nociceptive flexion reflex and pain in humans. *Anesth. Analg.* **90**, 408–414 (2000).
- 560. Paul, R. K. et al. (R,S)-Ketamine metabolites (R,S)-norketamine and (2S,6S)hydroxynorketamine increase the mammalian target of rapamycin function. *Anesthesiology* **121**, 149–159 (2014).
- 561. Liu, A. R. et al. The potent analgesia of intrathecal 2R, 6R-HNK via TRPA1 inhibition in LF-PENS-induced chronic primary pain model. J. Headache Pain. 24, 141 (2023).
- 562. Schwenk, E. S. et al. Consensus guidelines on the use of intravenous ketamine infusions for acute pain management from the american society of regional anesthesia and pain medicine, the american academy of pain medicine, and the American Society of Anesthesiologists. *Reg. Anesth. Pain Med* **43**, 456–466 (2018).
- 563. Zhang, X., Peng, L. & Liu, D. Pregabalin alleviates neuropathic pain via inhibition of the PKCε/TRPV1 pathway. *Neurosci. Lett.* **766**, 136348 (2022).
- 564. Bannister, K. et al. Multiple sites and actions of gabapentin-induced relief of ongoing experimental neuropathic pain. *Pain* **158**, 2386–2395 (2017).
- 565. Price, R. et al. Oral and topical treatment of painful diabetic polyneuropathy: practice guideline update summary: report of the AAN guideline subcommittee. *Neurology* **98**, 31–43 (2022).
- 566. Hannon, C. P. et al. Gabapentinoids in total joint arthroplasty: the clinical practice guidelines of the American Association of Hip and Knee Surgeons, American Society of Regional Anesthesia and Pain Medicine, American Academy of Orthopaedic Surgeons, Hip Society, and Knee Society. J. Arthroplast. 35, 2700–2703 (2020).
- 567. Anger, M. et al. PROSPECT guideline for total hip arthroplasty: a systematic review and procedure-specific postoperative pain management recommendations. *Anaesthesia* 76, 1082–1097 (2021).
- 568. Korwin-Kochanowska, K. et al. PROSPECT guideline for hallux valgus repair surgery: a systematic review and procedure-specific postoperative pain management recommendations. *Reg. Anesth. Pain. Med.* **45**, 702–708 (2020).
- 569. Hah, J. et al. Effect of perioperative gabapentin on postoperative pain resolution and opioid cessation in a mixed surgical cohort: a randomized clinical trial. JAMA Surg. 153, 303–311 (2018).
- 570. Tran, H. et al. Descending mechanism by which medial prefrontal cortex endocannabinoid signaling controls the development of neuropathic pain and neuronal activity of dorsal root ganglion. *Pain* **165**, 102–114 (2023).
- 571. Wang, H. et al. Incerta-thalamic circuit controls nocifensive behavior via cannabinoid type 1 receptors. *Neuron* **107**, 538–551.e537 (2020).
- 572. Goncalves Dos Santos, G. et al. CB(1) receptor-dependent desensitisation of TRPV1 channels contributes to the analgesic effect of dipyrone in sensitised primary sensory neurons. Br. J. Pharm. 177, 4615–4626 (2020).
- 573. Borgonetti, V. et al. Cannabidiol-rich non-psychotropic Cannabis sativa L. oils attenuate peripheral neuropathy symptoms by regulation of CB2-mediated microglial neuroinflammation. *Phytother. Res.* **37**, 1924–1937 (2023).
- 574. Gao, F. et al. Electroacupuncture inhibits NLRP3 inflammasome activation through CB2 receptors in inflammatory pain. *Brain Behav. Immun.* **67**, 91–100 (2018).
- 575. Carey, L. M. et al. Peripheral sensory neuron CB2 cannabinoid receptors are necessary for both CB2-mediated antinociceptive efficacy and sparing of

morphine tolerance in a mouse model of anti-retroviral toxic neuropathy. *Pharm. Res.* **187**, 106560 (2023).

- 576. Corcoran, L., Mattimoe, D., Roche, M. & Finn, D. P. Attenuation of fearconditioned analgesia in rats by monoacylglycerol lipase inhibition in the anterior cingulate cortex: potential role for CB(2) receptors. *Br. J. Pharm.* **177**, 2240–2255 (2020).
- 577. Zhang, H. B. & Bean, B. P. Cannabidiol inhibition of murine primary nociceptors: tight binding to slow inactivated states of Na(v)1.8 channels. J. Neurosci. 41, 6371–6387 (2021).
- 578. Ghovanloo, M. R. et al. Inhibition of sodium conductance by cannabigerol contributes to a reduction of dorsal root ganglion neuron excitability. *Br. J. Pharm.* **179**, 4010–4030 (2022).
- 579. Weizman, L. et al. Cannabis analgesia in chronic neuropathic pain is associated with altered brain connectivity. *Neurology* **91**, e1285–e1294 (2018).
- Mücke, M., Phillips, T., Radbruch, L., Petzke, F. & Häuser, W. Cannabis-based medicines for chronic neuropathic pain in adults. *Cochrane Database Syst. Rev.* 3, Cd012182 (2018).
- 581. van de Donk, T. et al. An experimental randomized study on the analgesic effects of pharmaceutical-grade cannabis in chronic pain patients with fibro-myalgia. *Pain* **160**, 860–869 (2019).
- 582. Freeman, T. P. et al. Cannabidiol for the treatment of cannabis use disorder: a phase 2a, double-blind, placebo-controlled, randomised, adaptive Bayesian trial. *Lancet Psychiatry* 7, 865–874 (2020).
- 583. Bebee, B. et al. The CANBACK trial: a randomised, controlled clinical trial of oral cannabidiol for people presenting to the emergency department with acute low back pain. *Med. J. Aust.* **214**, 370–375 (2021).
- 584. de Vries, M., van Rijckevorsel, D. C. M., Vissers, K. C. P., Wilder-Smith, O. H. G. & van Goor, H. Tetrahydrocannabinol does not reduce pain in patients with chronic abdominal pain in a phase 2 placebo-controlled study. *Clin. Gastroenterol. Hepatol.* **15**, 1079–1086.e1074 (2017).
- Narang, G. et al. Effect of cannabidiol oil on post-ureteroscopy pain for urinary calculi: a randomized, double-blind, placebo-controlled trial. *J. Urol.* 209, 726–733 (2023).
- Vela, J. et al. Cannabidiol treatment in hand osteoarthritis and psoriatic arthritis: a randomized, double-blind, placebo-controlled trial. *Pain* 163, 1206–1214 (2022).
- 587. Wang, L. et al. Medical cannabis or cannabinoids for chronic non-cancer and cancer related pain: a systematic review and meta-analysis of randomised clinical trials. *BMJ* 374, n1034 (2021).
- 588. Stockings, E. et al. Cannabis and cannabinoids for the treatment of people with chronic noncancer pain conditions: a systematic review and meta-analysis of controlled and observational studies. *Pain* 159, 1932–1954 (2018).
- Bilbao, A. & Spanagel, R. Medical cannabinoids: a pharmacology-based systematic review and meta-analysis for all relevant medical indications. *BMC Med.* 20, 259 (2022).
- 590. Fallon, M. et al. An international, open-label, randomised trial comparing a twostep approach versus the standard three-step approach of the WHO analgesic ladder in patients with cancer. Ann. Oncol. 33, 1296–1303 (2022).
- 591. Capano, A., Weaver, R. & Burkman, E. Evaluation of the effects of CBD hemp extract on opioid use and quality of life indicators in chronic pain patients: a prospective cohort study. *Postgrad. Med.* **132**, 56–61 (2020).
- 592. Zylla, D. M. et al. A randomized trial of medical cannabis in patients with stage IV cancers to assess feasibility, dose requirements, impact on pain and opioid use, safety, and overall patient satisfaction. *Support Care Cancer* 29, 7471–7478 (2021).
- 593. Blanken, P., Nuijten, M., van den Brink, W. & Hendriks, V. M. Clinical effects beyond cocaine use of sustained-release dexamphetamine for the treatment of cocaine dependent patients with comorbid opioid dependence: secondary analysis of a double-blind, placebo-controlled randomized trial. *Addiction* **115**, 917–923 (2020).
- 594. Cui, J. et al. Local infiltration of HYR-PB21, a sustained-release formulation of bupivacaine, provides analgesia and reduces opioid requirement after haemorrhoidectomy: a randomised controlled trial. Br. J. Anaesth. **129**, 970–976 (2022).
- 595. Dembla, S. et al. Anti-nociceptive action of peripheral mu-opioid receptors by Gbeta-gamma protein-mediated inhibition of TRPM3 channels. *Elife* 6, e26280 (2017).
- 596. Endres-Becker, J. et al. Mu-opioid receptor activation modulates transient receptor potential vanilloid 1 (TRPV1) currents in sensory neurons in a model of inflammatory pain. *Mol. Pharm.* **71**, 12–18 (2007).
- 597. MacDonald, D. I. et al. A central mechanism of analgesia in mice and humans lacking the sodium channel Na(V)1.7. Neuron 109, 1497–1512.e1496 (2021).
- 598. Borg, C. B. et al. Mechanism and site of action of big dynorphin on ASIC1a. Proc. Natl Acad. Sci. USA 117, 7447–7454 (2020).
- 599. Mitrano, D. A. et al. α-1 Adrenergic receptors are localized on presynaptic elements in the nucleus accumbens and regulate mesolimbic dopamine transmission. *Neuropsychopharmacology* **37**, 2161–2172 (2012).

- 600. Colvin, L. A., Bull, F. & Hales, T. G. Perioperative opioid analgesia-when is enough too much? A review of opioid-induced tolerance and hyperalgesia. *Lancet* **393**, 1558–1568 (2019).
- Basso, L. et al. TRPV1 promotes opioid analgesia during inflammation. *Sci. Signal.* 12 (2019).
- 602. Mafi, A., Kim, S. K. & Goddard, W. A. 3rd Mechanism of β-arrestin recruitment by the μ-opioid G protein-coupled receptor. *Proc. Natl Acad. Sci. USA* **117**, 16346–16355 (2020).
- Pasquinucci, L. et al. (25)-N-2-methoxy-2-phenylethyl-6,7-benzomorphan compound (2S-LP2): discovery of a biased mu/delta opioid receptor agonist. *Eur. J. Med. Chem.* **168**, 189–198 (2019).
- 604. Piekielna-Ciesielska, J. et al. Functional selectivity of EM-2 analogs at the muopioid receptor. *Front. Pharm.* **14**, 1133961 (2023).
- 605. Kliewer, A. et al. Phosphorylation-deficient G-protein-biased μ-opioid receptors improve analgesia and diminish tolerance but worsen opioid side effects. *Nat. Commun.* **10**, 367 (2019).
- Bachmutsky, I., Wei, X. P., Durand, A. & Yackle, K. ß-arrestin 2 germline knockout does not attenuate opioid respiratory depression. *Elife* 10, e62552 (2021).
- 607. Muchhala, K. H., Jacob, J. C., Dewey, W. L. & Akbarali, H. I. Role of β-arrestin-2 in short- and long-term opioid tolerance in the dorsal root ganglia. *Eur. J. Pharm.* 899, 174007 (2021).
- 608. Koshimizu, T. A. et al. Complex formation between the vasopressin 1b receptor, β -arrestin-2, and the μ -opioid receptor underlies morphine tolerance. *Nat. Neurosci.* **21**, 820–833 (2018).
- 609. Wang, X. et al. Sinomenine alleviates dorsal root ganglia inflammation to inhibit neuropathic pain via the p38 MAPK/CREB signalling pathway. *Eur. J. Pharm.* 897, 173945 (2021).
- 610. Pan, Y. et al. Metformin reduces morphine tolerance by inhibiting microglialmediated neuroinflammation. J. Neuroinflamm. **13**, 294 (2016).
- Zhang, Y. et al. Lidocaine alleviates morphine tolerance via AMPK-SOCS3dependent neuroinflammation suppression in the spinal cord. *J. Neuroinflamm.* 14, 211 (2017).
- 612. Cui, Y. et al. A novel role of minocycline: attenuating morphine antinociceptive tolerance by inhibition of p38 MAPK in the activated spinal microglia. *Brain Behav. Immun.* 22, 114–123 (2008).
- 613. Sala, E. et al. Improved efficacy, tolerance, safety, and abuse liability profile of the combination of CR4056 and morphine over morphine alone in rodent models. *Br. J. Pharm.* **177**, 3291–3308 (2020).
- 614. Li, Q. et al. Hydrogen enriched saline alleviates morphine tolerance via inhibiting neuroinflammation, GLT-1, GS nitration and NMDA receptor trafficking and functioning in the spinal cord of rats. *Neurosci. Lett.* **755**, 135847 (2021).
- 615. Gritsch, S. et al. Oligodendrocyte ablation triggers central pain independently of innate or adaptive immune responses in mice. *Nat. Commun.* **5**, 5472 (2014).
- Attal, N. et al. EFNS guidelines on the pharmacological treatment of neuropathic pain: 2010 revision. *Eur. J. Neurol.* 17, 1113–e1188 (2010).
- Moulin, D. et al. Pharmacological management of chronic neuropathic pain: revised consensus statement from the Canadian Pain Society. *Pain Res. Manag.* 19, 328–335 (2014).
- 618. Yoo, O. et al. A randomised controlled trial of a novel tramadol chewable tablet: pharmacokinetics and tolerability in children. *Anaesthesia* **77**, 438–448 (2022).
- Abdel-Zaher, A. O., Hamdy, M. M., Aly, S. A., Abdel-Hady, R. H. & Abdel-Rahman, S. Attenuation of morphine tolerance and dependence by aminoguanidine in mice. *Eur. J. Pharm.* **540**, 60–66 (2006).
- 620. Wolińska, R. et al. Nitric oxide modulates tapentadol antinociceptive tolerance and physical dependence. *Eur. J. Pharm.* **907**, 174245 (2021).
- 621. Korownyk, C. S. et al. PEER simplified chronic pain guideline: Management of chronic low back, osteoarthritic, and neuropathic pain in primary care. *Can. Fam. Physician* **68**, 179–190 (2022).
- 622. Colombel, J. F., Shin, A. & Gibson, P. R. AGA clinical practice update on functional gastrointestinal symptoms in patients with inflammatory bowel disease: expert review. *Clin. Gastroenterol. Hepatol.* **17**, 380–390.e381 (2019).
- 623. Busse, J. W. et al. Management of chronic pain associated with temporomandibular disorders: a clinical practice guideline. *Bmj* 383, e076227 (2023).
- 624. Mourad, G., Eriksson-Liebon, M., Karlström, P. & Johansson, P. The effect of internet-delivered cognitive behavioral therapy versus psychoeducation only on psychological distress in patients with noncardiac chest pain: randomized controlled trial. J. Med. Internet Res. 24, e31674 (2022).
- 625. Lalouni, M. et al. Clinical and cost effectiveness of online cognitive behavioral therapy in children with functional abdominal pain disorders. *Clin. Gastroenterol. Hepatol.* **17**, 2236–2244.e2211 (2019).
- 626. Lee, J. et al. A randomized controlled neuroimaging trial of cognitive behavioral therapy for fibromyalgia pain. *Arthritis Rheumatol.* **76**, 130–140 (2023).
- 627. Yoshino, A. et al. Changes in resting-state brain networks after cognitivebehavioral therapy for chronic pain. *Psychol. Med.* 48, 1148–1156 (2018).

- 46
- Greenwald, J. D. & Shafritz, K. M. An integrative neuroscience framework for the treatment of chronic pain: from cellular alterations to behavior. *Front. Integr. Neurosci.* 12, 18 (2018).
- 629. Nahman-Averbuch, H. et al. Alterations in brain function after cognitive behavioral therapy for migraine in children and adolescents. *Headache* **60**, 1165–1182 (2020).
- 630. Nahman-Averbuch, H. et al. Identification of neural and psychophysical predictors of headache reduction after cognitive behavioral therapy in adolescents with migraine. *Pain* **162**, 372–381 (2021).
- 631. Macfarlane, G. J. et al. EULAR revised recommendations for the management of fibromyalgia. *Ann. Rheum. Dis.* **76**, 318–328 (2017).
- 632. Qaseem, A. et al. Noninvasive treatments for acute, subacute, and chronic low back pain: a clinical practice guideline from the American College of Physicians. *Ann. Intern. Med.* **166**, 514–530 (2017).
- 633. Lin, I. et al. What does best practice care for musculoskeletal pain look like? Eleven consistent recommendations from high-quality clinical practice guidelines: systematic review. Br. J. Sports Med. 54, 79–86 (2020).
- 634. Nijs, J. et al. Towards precision pain medicine for pain after cancer: the Cancer Pain Phenotyping Network multidisciplinary international guidelines for pain phenotyping using nociplastic pain criteria. *Br. J. Anaesth.* **130**, 611–621 (2023).
- 635. Zhang, Y. H. et al. Exercise for neuropathic pain: a systematic review and expert consensus. *Front. Med.* **8**, 756940 (2021).
- Bush, N. J., Schneider, V., Sevel, L., Bishop, M. D. & Boissoneault, J. Associations of regional and network functional connectivity with exercise-induced low back pain. J. Pain 22, 1606–1616 (2021).
- Ellingson, L. D., Stegner, A. J., Schwabacher, I. J., Koltyn, K. F. & Cook, D. B. Exercise strengthens central nervous system modulation of pain in fibromyalgia. *Brain Sci.* 6, 8 (2016).
- 638. Baraniuk, J. N., Amar, A., Pepermitwala, H. & Washington, S. D. Differential effects of exercise on fMRI of the midbrain ascending arousal network nuclei in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) and Gulf War Illness (GWI) in a Model of Postexertional Malaise (PEM). *Brain Sci.* **12**, 78 (2022).
- 639. Kong, J. T. et al. Effect of electroacupuncture vs sham treatment on change in pain severity among adults with chronic low back pain: a randomized clinical trial. *JAMA Netw. Open* **3**, e2022787 (2020).
- 640. Ho, J. T., Krummenacher, P., Lesur, M. R., Saetta, G. & Lenggenhager, B. Real bodies not required? Placebo analgesia and pain perception in immersive virtual and augmented reality. *J. Pain* **23**, 625–640 (2022).
- 641. Davies, J. N., Colagiuri, B., Sharpe, L. & Day, M. A. Placebo effects contribute to brief online mindfulness interventions for chronic pain: results from an online randomized sham-controlled trial. *Pain* **164**, 2273–2284 (2023).
- Benedetti, F., Amanzio, M., Rosato, R. & Blanchard, C. Nonopioid placebo analgesia is mediated by CB1 cannabinoid receptors. *Nat. Med.* 17, 1228–1230 (2011).
- 643. Wagner, I. C., Rütgen, M., Hummer, A., Windischberger, C. & Lamm, C. Placeboinduced pain reduction is associated with negative coupling between brain networks at rest. *Neuroimage* **219**, 117024 (2020).
- 644. Tétreault, P. et al. Brain connectivity predicts placebo response across chronic pain clinical trials. *PLoS Biol.* **14**, e1002570 (2016).
- 645. Hashmi, J. A. et al. Brain networks predicting placebo analgesia in a clinical trial for chronic back pain. *Pain* **153**, 2393–2402 (2012).
- 646. Fu, J. et al. Distinct neural networks subserve placebo analgesia and nocebo hyperalgesia. *Neuroimage* **231**, 117833 (2021).
- 647. Mao, J. J. et al. Integrative medicine for pain management in oncology: society for integrative oncology-ASCO guideline. J. Clin. Oncol. 40, 3998–4024 (2022).
- Luo, X. et al. Acupuncture for treatment of knee osteoarthritis: a clinical practice guideline. J. Evid. Based Med. 16, 237–245 (2023).
- 649. Zhang, J. et al. Acupuncture for chemotherapy-associated insomnia in breast cancer patients: an assessor-participant blinded, randomized, sham-controlled trial. *Breast Cancer Res.* **25**, 49 (2023).
- 650. Liao, H. Y., Satyanarayanan, S. K., Lin, Y. W. & Su, K. P. Clinical efficacy and immune effects of acupuncture in patients with comorbid chronic pain and major depression disorder: A double-blinded, randomized controlled crossover study. *Brain Behav. Immun.* **110**, 339–347 (2023).
- 651. Yu, Z. et al. A coordinate-based meta-analysis of acupuncture for chronic pain: evidence from fMRI studies. *Front. Neurosci.* **16**, 1049887 (2022).
- 652. Liu, L. et al. Changes in brain connectivity linked to multisensory processing of pain modulation in migraine with acupuncture treatment. *Neuroimage Clin.* **36**, 103168 (2022).
- 653. Ha, G. et al. Coordinate-based (ALE) meta-analysis of acupuncture for musculoskeletal pain. *Front. Neurosci.* **16**, 906875 (2022).
- 654. Wang, W. et al. Phosphoproteomic profiling of rat's dorsal root ganglia reveals mTOR as a potential target in bone cancer pain and electro-acupuncture's analgesia. *Front Pharm.* **12**, 593043 (2021).

- 655. Yao, C. et al. Transcriptome profiling of microRNAs reveals potential mechanisms of manual therapy alleviating neuropathic pain through microRNA-547-3pmediated Map4k4/NF-κb signaling pathway. J. Neuroinflamm. **19**, 211 (2022).
- 656. Tan, H., Yamamoto, E. A., Elkholy, M. A. & Raslan, A. M. Treating chronic pain with deep brain stimulation. *Curr. Pain. Headache Rep.* 27, 11–17 (2023).
- 657. Galhardoni, R. et al. Insular and anterior cingulate cortex deep stimulation for central neuropathic pain: disassembling the percept of pain. *Neurology* 92, e2165–e2175 (2019).
- 658. Lempka, S. F. et al. Randomized clinical trial of deep brain stimulation for poststroke pain. *Ann. Neurol.* **81**, 653–663 (2017).
- 659. Duarte, R. V. et al. Systematic review and meta-analysis of placebo/sham controlled randomised trials of spinal cord stimulation for neuropathic pain. *Pain* 161, 24–35 (2020).
- 660. Mong, M. S. A., Lai, M. Y. C., Cheng, L. J. & Lau, Y. Novel spinal cord stimulation waveforms for treating back and leg pain: a systematic review and metaanalysis of randomized controlled trials. *Neuromodulation* **26**, 905–916 (2023).
- Moisset, X. et al. Pharmacological and non-pharmacological treatments for neuropathic pain: systematic review and French recommendations. *Rev. Neurol.* 176, 325–352 (2020).
- Fontaine, D. Spinal cord stimulation for neuropathic pain. *Rev. Neurol.* 177, 838–842 (2021).
- 663. Zhai, F. J. et al. Involvement of opioid peptides in the analgesic effect of spinal cord stimulation in a rat model of neuropathic pain. *Neurosci. Bull.* 38, 403–416 (2022).
- 664. Saadé, N. E., Tabet, M. S., Banna, N. R., Atweh, S. F. & Jabbur, S. J. Inhibition of nociceptive evoked activity in spinal neurons through a dorsal columnbrainstem-spinal loop. *Brain Res.* **339**, 115–118 (1985).
- 665. Carstens, E. & Watkins, L. R. Inhibition of the responses of neurons in the rat spinal cord to noxious skin heating by stimulation in midbrain periaqueductal gray or lateral reticular formation. *Brain Res.* **382**, 266–277 (1986).
- 666. Heijmans, L., Mons, M. R. & Joosten, E. A. A systematic review on descending serotonergic projections and modulation of spinal nociception in chronic neuropathic pain and after spinal cord stimulation. *Mol. Pain* **17**, 17448069211043965 (2021).
- 667. Graham, R. D., Sankarasubramanian, V. & Lempka, S. F. Dorsal root ganglion stimulation for chronic pain: hypothesized mechanisms of action. *J. Pain.* 23, 196–211 (2022).
- 668. Deer, T. R. et al. Dorsal root ganglion stimulation yielded higher treatment success rate for complex regional pain syndrome and causalgia at 3 and 12 months: a randomized comparative trial. *Pain* **158**, 669–681 (2017).
- 669. Levy, R. M. et al. Therapy habituation at 12 months: spinal cord stimulation versus dorsal root ganglion stimulation for complex regional pain syndrome type I and II. J. Pain 21, 399–408 (2020).
- 670. Kinfe, T. M. et al. Unilateral L4-dorsal root ganglion stimulation evokes pain relief in chronic neuropathic postsurgical knee pain and changes of inflammatory markers: part II whole transcriptome profiling. J. Transl. Med. 17, 205 (2019).
- 671. Mol, F. et al. Comparing the efficacy of dorsal root ganglion stimulation with conventional medical management in patients with chronic postsurgical inguinal pain: post hoc analyzed results of the SMASHING study. *Neuromodulation* 26, 1788–1794 (2023).
- 672. Deer, T. R., Grigsby, E., Weiner, R. L., Wilcosky, B. & Kramer, J. M. A prospective study of dorsal root ganglion stimulation for the relief of chronic pain. *Neuromodulation* **16**, 67–71 (2013).
- 673. Piedade, G. S., Gillner, S., McPhillips, P. S., Vesper, J. & Slotty, P. J. Effect of low-frequency dorsal root ganglion stimulation in the treatment of chronic pain. *Acta Neurochir.* **165**, 947–952 (2023).
- 674. Piedade, G. S., Gillner, S., McPhillips, P. S., Vesper, J. & Slotty, P. J. Frequency dependency of therapeutic efficacy in dorsal root ganglion stimulation for neuropathic pain. *Acta Neurochir.* **164**, 1193–1199 (2022).
- Xu, F. et al. One-dimensional local binary pattern and common spatial pattern feature fusion brain network for central neuropathic pain. *Int. J. Neural Syst.* 33, 2350030 (2023).
- 676. Zhang, S. et al. Pain control by co-adaptive learning in a brain-machine interface. *Curr. Biol.* **30**, 3935–3944.e3937 (2020).
- 677. Sun, G. et al. Closed-loop stimulation using a multiregion brain-machine interface has analgesic effects in rodents. *Sci. Transl. Med.* **14**, eabm5868 (2022).
- 678. Zhang, Q. et al. A prototype closed-loop brain-machine interface for the study and treatment of pain. *Nat. Biomed. Eng.* **7**, 533–545 (2023).
- 679. Yanagisawa, T. et al. BCI training to move a virtual hand reduces phantom limb pain: a randomized crossover trial. *Neurology* **95**, e417–e426 (2020).
- 680. Yanagisawa, T. et al. Induced sensorimotor brain plasticity controls pain in phantom limb patients. *Nat. Commun.* **7**, 13209 (2016).
- Aurucci, G. V., Preatoni, G., Damiani, A. & Raspopovic, S. Brain-computer interface to deliver individualized multisensory intervention for neuropathic pain. *Neurotherapeutics* 20, 1316–1329 (2023).

- 682. Al-Taleb, M. K. H., Purcell, M., Fraser, M., Petric-Gray, N. & Vuckovic, A. Home used, patient self-managed, brain-computer interface for the management of central neuropathic pain post spinal cord injury: usability study. *J. Neuroeng. Rehabil.* **16**, 128 (2019).
- 683. Vuckovic, A. et al. The influence of central neuropathic pain in paraplegic patients on performance of a motor imagery based brain computer interface. *Clin. Neurophysiol.* **126**, 2170–2180 (2015).
- 684. Constante, M. et al. Saccharomyces boulardii CNCM I-745 modulates the microbiota-gut-brain axis in a humanized mouse model of Irritable Bowel Syndrome. *Neurogastroenterol. Motil.* **33**, e13985 (2021).
- Verdú, E. F. et al. Specific probiotic therapy attenuates antibiotic induced visceral hypersensitivity in mice. *Gut* 55, 182–190 (2006).
- Zhang, J. et al. Beneficial effect of butyrate-producing Lachnospiraceae on stress-induced visceral hypersensitivity in rats. J. Gastroenterol. Hepatol. 34, 1368–1376 (2019).
- 687. Pokusaeva, K. et al. GABA-producing bifidobacterium dentium modulates visceral sensitivity in the intestine. *Neurogastroenterol. Motil.* **29**, e12904 (2017).
- Liu, Y. W. et al. Lactobacillus plantarum PS128 ameliorated visceral hypersensitivity in rats through the gut-brain axis. *Probiot. Antimicrob. Proteins* 12, 980–993 (2020).
- 689. Luo, M., Liu, Q., Xiao, L. & Xiong, L. S. Golden bifid might improve diarrheapredominant irritable bowel syndrome via microbiota modulation. *J. Health Popul. Nutr.* **41**, 21 (2022).
- 690. Kajander, K. et al. Clinical trial: multispecies probiotic supplementation alleviates the symptoms of irritable bowel syndrome and stabilizes intestinal microbiota. *Aliment. Pharm. Ther.* **27**, 48–57 (2008).
- 691. Martín, R. et al. Effects in the use of a genetically engineered strain of Lactococcus lactis delivering in situ IL-10 as a therapy to treat low-grade colon inflammation. *Hum. Vaccin. Immunother.* **10**, 1611–1621 (2014).
- Meynier, M. et al. AhR/IL-22 pathway as new target for the treatment of postinfectious irritable bowel syndrome symptoms. *Gut Microbes* 14, 2022997 (2022).
- 693. Martami, F. et al. The effects of a multispecies probiotic supplement on inflammatory markers and episodic and chronic migraine characteristics: a randomized double-blind controlled trial. *Cephalalgia* **39**, 841–853 (2019).
- 694. Var, I., AlMatar, M., Heshmati, B. & Albarri, O. Bacteriophage cocktail can effectively control salmonella biofilm on gallstone and tooth surfaces. *Curr. Drug Targets* 24, 613–625 (2023).
- 695. Bai, T. et al. Fecal microbiota transplantation is effective in relieving visceral hypersensitivity in a postinfectious model. *Biomed. Res. Int.* **2018**, 3860743 (2018).
- 696. Cruz-Aguliar, R. M. et al. An open-labeled study on fecal microbiota transfer in irritable bowel syndrome patients reveals improvement in abdominal pain associated with the relative abundance of akkermansia muciniphila. *Digestion* **100**, 127–138 (2019).
- 697. Holvoet, T. et al. Fecal microbiota transplantation reduces symptoms in some patients with irritable bowel syndrome with predominant abdominal bloating: short- and long-term results from a placebo-controlled randomized trial. *Gastroenterology* **160**, 145–157.e148 (2021).
- 698. Holster, S. et al. The effect of allogenic versus autologous fecal microbiota transfer on symptoms, visceral perception and fecal and mucosal microbiota in irritable bowel syndrome: a randomized controlled study. *Clin. Transl. Gastroenterol.* **10**, e00034 (2019).
- 699. Guo, Q. et al. Dynamic changes of intestinal flora in patients with irritable bowel syndrome combined with anxiety and depression after oral administration of enterobacteria capsules. *Bioengineered* **12**, 11885–11897 (2021).
- 700. He, S. et al. A human TRPV1 genetic variant within the channel gating domain regulates pain sensitivity in rodents. J. Clin. Investig. 133, e163735 (2023).
- Yu, H. et al. AAV-encoded Ca(V)2.2 peptide aptamer CBD3A6K for primary sensory neuron-targeted treatment of established neuropathic pain. *Gene Ther.* 26, 308–323 (2019).
- 702. Weir, G. A. et al. Using an engineered glutamate-gated chloride channel to silence sensory neurons and treat neuropathic pain at the source. *Brain* 140, 2570–2585 (2017).
- Mészár, Z. et al. CRISPR/Cas9-based mutagenesis of histone H3.1 in spinal dynorphinergic neurons attenuates thermal sensitivity in mice. *Int. J. Mol. Sci.* 23, 3178 (2022).
- 704. Ahmed, S. G. et al. Gene therapy with apoptosis-associated speck-like protein, a newly described schwannoma tumor suppressor, inhibits schwannoma growth in vivo. *Neuro Oncol.* **21**, 854–866 (2019).
- 705. Ahmed, S. G., Maguire, C. A., Cao, S. A. & Brenner, G. J. Schwannoma gene therapy via adeno-associated viral vector delivery of apoptosis-associated speck-like protein containing CARD (ASC): preclinical efficacy and safety. *Int. J. Mol. Sci.* 23, 819 (2022).

- Ji, H. et al. Combination gene delivery reduces spinal cord pathology in rats with peripheral neuropathic pain. J. Pain 24, 2211–2227 (2023).
- 707. Nho, B., Ko, K. R., Kim, S. & Lee, J. Intramuscular injection of a plasmid DNA vector expressing hepatocyte growth factor (HGF) ameliorated pain symptoms by controlling the expression of pro-inflammatory cytokines in the dorsal root ganglion. *Biochem Biophys. Res. Commun.* **607**, 60–66 (2022).
- 708. Hu, C. et al. Gene therapy for neuropathic pain induced by spared nerve injury with naked plasmid encoding hepatocyte growth factor. *J. Gene Med.* **19** (2017).
- 709. Kessler, J. A. et al. Gene therapy for diabetic peripheral neuropathy: a randomized, placebo-controlled phase III study of VM202, a plasmid DNA encoding human hepatocyte growth factor. *Clin. Transl. Sci.* 14, 1176–1184 (2021).
- 710. Clements, M. A. et al. Intrathecal non-viral interleukin-10 gene therapy ameliorates neuropathic pain as measured by both classical static allodynia and a novel supra-spinally mediated pain assay, the Two-Arm Rodent Somatosensory (TARS) task. *Brain Behav. Immun.* **111**, 177–185 (2023).
- 711. Altae-Tran, H. et al. Uncovering the functional diversity of rare CRISPR-Cas systems with deep terascale clustering. *Science* **382**, eadi1910 (2023).
- 712. Ashina, H. et al. Efficacy, tolerability, and safety of erenumab for the preventive treatment of persistent post-traumatic headache attributed to mild traumatic brain injury: an open-label study. J. Headache Pain 21, 62 (2020).
- 713. Schott Andersen, A. S. et al. Safety and efficacy of erenumab in patients with trigeminal neuralgia in Denmark: a double-blind, randomised, placebo-controlled, proof-of-concept study. *Lancet Neurol.* **21**, 994–1003 (2022).
- Lipton, R. B. et al. Safety, tolerability, and efficacy of zavegepant 10 mg nasal spray for the acute treatment of migraine in the USA: a phase 3, double-blind, randomised, placebo-controlled multicentre trial. *Lancet Neurol.* 22, 209–217 (2023).
- 715. Berman, G. et al. Safety of rimegepant, an Oral CGRP receptor antagonist, plus CGRP monoclonal antibodies for migraine. *Headache* **60**, 1734–1742 (2020).
- 716. Seah, K. T. M., Rammanohar, J., Sutton, J., To, K. & Khan, W. S. The effectiveness of anti-nerve growth factor monoclonal antibodies in the management of pain in osteoarthritis of the hip and knee: a PRISMA systematic review and metaanalysis. *Pain. Med.* 22, 1185–1204 (2021).
- 717. Majuta, L. A., Guedon, J. G., Mitchell, S. A. T., Ossipov, M. H. & Mantyh, P. W. Antinerve growth factor therapy increases spontaneous day/night activity in mice with orthopedic surgery-induced pain. *Pain* **158**, 605–617 (2017).
- Dong, X. et al. Analgesic effects of nerve growth factor-directed monoclonal antibody on diabetic neuralgia in an animal model. *FEBS Open Bio* 12, 1325–1335 (2022).
- 719. LaMacchia, Z. M. et al. Perispinal injection of a TNF blocker directed to the brain of rats alleviates the sensory and affective components of chronic constriction injury-induced neuropathic pain. *Brain Behav. Immun.* 82, 93–105 (2019).
- 720. Kochi, T. et al. Pretreatment with high mobility group box-1 monoclonal antibody prevents the onset of trigeminal neuropathy in mice with a distal infraorbital nerve chronic constriction injury. *Molecules* 26, 2035 (2021).
- Ugolini, G., Marinelli, S., Covaceuszach, S., Cattaneo, A. & Pavone, F. The function neutralizing anti-TrkA antibody MNAC13 reduces inflammatory and neuropathic pain. *Proc. Natl Acad. Sci. USA* **104**, 2985–2990 (2007).
- 722. Kopruszinski, C. M. et al. Characterization and preclinical evaluation of a protease activated receptor 2 (PAR2) monoclonal antibody as a preventive therapy for migraine. *Cephalalgia* **40**, 1535–1550 (2020).
- 723. Bang, S. et al. Differential inhibition of Nav1.7 and neuropathic pain by hybridoma-produced and recombinant monoclonal antibodies that target Nav1.7 : differential activities of Nav1.7-targeting monoclonal antibodies. *Neurosci. Bull.* 34, 22–41 (2018).
- 724. Trkulja, C. L. et al. Rational antibody design for undruggable targets using kinetically controlled biomolecular probes. *Sci. Adv.* **7**, eabe6397 (2021).
- 725. Padda, J. et al. Stem cell therapy and its significance in pain management. *Cureus* **13**, e17258 (2021).
- 726. Wang, Q., He, H., Xie, S., Wei, Q. & He, C. Mesenchymal stem cells transplantation for neuropathic pain induced by peripheral nerve injury in animal models: a systematic review. *Stem Cells Dev.* **29**, 1420–1428 (2020).
- 727. Zhang, Y. et al. Therapeutic effects of peripherally administrated neural crest stem cells on pain and spinal cord changes after sciatic nerve transection. *Stem Cell Res. Ther.* **12**, 180 (2021).
- Lin, C. H. et al. Autologous adipose-derived stem cells reduce burn-induced neuropathic pain in a rat model. *Int. J. Mol. Sci.* 19, 34 (2017).
- 729. Li, F., Liu, L., Cheng, K., Chen, Z. & Cheng, J. The use of stem cell therapy to reverse opioid tolerance. *Clin. Pharm. Ther.* **103**, 971–974 (2018).
- Li, M., Li, J., Chen, H. & Zhu, M. VEGF-expressing mesenchymal stem cell therapy for safe and effective treatment of pain in Parkinson's disease. *Cell Transpl.* 32, 9636897221149130 (2023).
- Lee, N. et al. Mesenchymal stem cell spheroids alleviate neuropathic pain by modulating chronic inflammatory response genes. *Front. Immunol.* 13, 940258 (2022).

- 48
- Lee, H. L. et al. Peripheral nerve-derived stem cell spheroids induce functional recovery and repair after spinal cord injury in rodents. *Int. J. Mol. Sci.* 22, 4141 (2021).
- 733. Shang, Z., Wang, M., Zhang, B., Wang, X. & Wanyan, P. Clinical translation of stem cell therapy for spinal cord injury still premature: results from a single-arm metaanalysis based on 62 clinical trials. *BMC Med.* **20**, 284 (2022).
- 734. Sun, Y., Zhao, J., Zhang, L., Li, Z. & Lei, S. Effectiveness and safety of stem cell therapy for diabetic foot: a meta-analysis update. *Stem Cell Res. Ther.* **13**, 416 (2022).
- Vijayavenkataraman, S. Nerve guide conduits for peripheral nerve injury repair: a review on design, materials and fabrication methods. *Acta Biomater.* **106**, 54–69 (2020).
- Fang, R. et al. Conservation and divergence of cortical cell organization in human and mouse revealed by MERFISH. *Science* 377, 56–62 (2022).
- 737. Wang, M. et al. Transformative network modeling of multi-omics data reveals detailed circuits, key regulators, and potential therapeutics for Alzheimer's disease. *Neuron* **109**, 257–272.e214 (2021).
- Li, Z. et al. Multi-omics analyses of serum metabolome, gut microbiome and brain function reveal dysregulated microbiota-gut-brain axis in bipolar depression. *Mol. Psychiatry* 27, 4123–4135 (2022).
- Jang, W. E. et al. Cntnap2-dependent molecular networks in autism spectrum disorder revealed through an integrative multi-omics analysis. *Mol. Psychiatry* 28, 810–821 (2023).
- 740. Wang, H. et al. The effect of tuina on ulcerative colitis model mice analyzed by gut microbiota and proteomics. *Front. Microbiol.* **13**, 976239 (2022).
- 741. Zeber-Lubecka, N. et al. Gene expression-based functional differences between the bladder body and trigonal urothelium in adolescent female patients with micturition dysfunction. *Biomedicines* **10**, 1435 (2022).
- 742. Vacca, V. et al. 17beta-estradiol counteracts neuropathic pain: a behavioural, immunohistochemical, and proteomic investigation on sex-related differences in mice. *Sci. Rep.* 6, 18980 (2016).
- Cherng, C. H. et al. Baicalin ameliorates neuropathic pain by suppressing HDAC1 expression in the spinal cord of spinal nerve ligation rats. *J. Formos. Med. Assoc.* 113, 513–520 (2014).
- 744. Chen, L. et al. Single-cell RNA sequencing in the context of neuropathic pain: progress, challenges, and prospects. *Transl. Res.* **251**, 96–103 (2023).
- 745. Allesøe, R. L. et al. Discovery of drug-omics associations in type 2 diabetes with generative deep-learning models. *Nat. Biotechnol.* **41**, 399–408 (2023).
- Lin, X. et al. One-droplet saliva detection on photonic crystal-based competitive immunoassay for precise diagnosis of migraine. *SmartMat.* e1252 (2023).
- 747. Botschuijver, S. et al. Intestinal fungal dysbiosis is associated with visceral hypersensitivity in patients with irritable bowel syndrome and rats. *Gastroenterology* **153**, 1026–1039 (2017).
- Lin, M. J. et al. Cancer vaccines: the next immunotherapy frontier. *Nat. Cancer* 3, 911–926 (2022).
- 749. Gould, H. M. et al. A randomized placebo-controlled trial of desipramine, cognitive behavioral therapy, and active placebo therapy for low back pain. *Pain* 161, 1341–1349 (2020).
- 750. Chen, K. H. et al. Synergic effect of early administration of probiotics and adipose-derived mesenchymal stem cells on alleviating inflammation-induced chronic neuropathic pain in rodents. *Int. J. Mol. Sci.* 23, 11974 (2022).
- 751. Castro, J. et al. α-Conotoxin Vc1.1 inhibits human dorsal root ganglion neuroexcitability and mouse colonic nociception via GABA(B) receptors. *Gut* 66, 1083–1094 (2017).
- 752. Yang, S. et al. Discovery of a selective NaV1.7 inhibitor from centipede venom with analgesic efficacy exceeding morphine in rodent pain models. *Proc. Natl Acad. Sci. USA* **110**, 17534–17539 (2013).
- 753. Kringel, D. et al. Emergent biomarker derived from next-generation sequencing to identify pain patients requiring uncommonly high opioid doses. *Pharmacogenom. J.* **17**, 419–426 (2017).
- Luchting, B. et al. Soluble intercellular adhesion molecule-1: a potential biomarker for pain intensity in chronic pain patients. *Biomark. Med.* 11, 265–276 (2017).
- Niculescu, A. B. et al. Towards precision medicine for pain: diagnostic biomarkers and repurposed drugs. *Mol. Psychiatry* 24, 501–522 (2019).
- Li, J. et al. Label-free serum proteomics for the identification of the putative biomarkers of postoperative pain in patients with gastric cancer. *Mol. Omics* 19, 351–361 (2023).
- 757. Brubel, R. et al. Serum galectin-9 as a noninvasive biomarker for the detection of endometriosis and pelvic pain or infertility-related gynecologic disorders. *Fertil. Steril.* **108**, 1016–1025.e2 (2017).
- Islam, B. et al. The identification of blood biomarkers of chronic neuropathic pain by comparative transcriptomics. *Neuromol. Med.* 24, 320–338 (2022).
- 759. Papaneophytou, C. et al. Associations between serum biomarkers of cartilage metabolism and serum hyaluronic acid, with risk factors, pain categories, and

disease severity in knee osteoarthritis: a pilot study. *BMC Musculoskelet. Disord.* **23**, 195 (2022).

- 760. Krämer, H. H. et al. Osteoprotegerin: a new biomarker for impaired bone metabolism in complex regional pain syndrome? *Pain* **155**, 889–895 (2014).
- Wu, Y., He, Y., Qi, J., Wang, S. & Wang, Z. Urinary ATP may be a biomarker of interstitial cystitis/bladder pain syndrome and its severity. *Biomol. Biomed.* 24, 170–175 (2023).
- 762. Yesil, H. et al. Association between serum vitamin D levels and neuropathic pain in rheumatoid arthritis patients: a cross-sectional study. *Int. J. Rheum. Dis.* 21, 431–439 (2018).
- 763. Ke, H., Zhu, L., Wang, Q. & Xu, K. Neutrophil-to-lymphocyte ratio as a promising non-invasive biomarker for symptom assessment and diagnosis of interstitial cystitis/bladder pain syndrome. *BMC Urol.* 23, 180 (2023).
- Shcherbatko, A. et al. Modulation of P2X3 and P2X2/3 receptors by monoclonal antibodies. J. Biol. Chem. 291, 12254–12270 (2016).
- Westlund, K. N. et al. Single-dose P2 X4R single-chain fragment variable antibody permanently reverses chronic pain in male mice. *Int. J. Mol. Sci.* 22, 13612 (2021).
- 766. Stack, E. et al. In vitro affinity optimization of an anti-BDNF monoclonal antibody translates to improved potency in targeting chronic pain states in vivo. *MAbs* 12, 1755000 (2020).
- 767. Wang, K. et al. PD-1 blockade inhibits osteoclast formation and murine bone cancer pain. J. Clin. Investig. **130**, 3603–3620 (2020).
- Brederson, J. D. et al. A monoclonal antibody against the receptor for advanced glycation end products attenuates inflammatory and neuropathic pain in the mouse. *Eur. J. Pain.* 20, 607–614 (2016).
- 769. Hurtado, A. et al. Anti-CD11d monoclonal antibody treatment for rat spinal cord compression injury. *Exp. Neurol.* **233**, 606–611 (2012).
- 770. Tonello, R., Lee, S. H. & Berta, T. Monoclonal antibody targeting the matrix metalloproteinase 9 prevents and reverses paclitaxel-induced peripheral neuropathy in mice. J. Pain. 20, 515–527 (2019).
- 771. Miller, S. et al. Antibodies to the extracellular pore loop of TRPM8 act as antagonists of channel activation. *PLoS One* **9**, e107151 (2014).
- 772. Micheli, L. et al. VEGF-A/VEGFR-1 signalling and chemotherapy-induced neuropathic pain: therapeutic potential of a novel anti-VEGFR-1 monoclonal antibody. J. Exp. Clin. Cancer Res. 40, 320 (2021).
- 773. Mothe, A. J. et al. RGMa inhibition with human monoclonal antibodies promotes regeneration, plasticity and repair, and attenuates neuropathic pain after spinal cord injury. *Sci. Rep.* 7, 10529 (2017).
- 774. Oehler, B. et al. Inflammatory pain control by blocking oxidized phospholipidmediated TRP channel activation. *Sci. Rep.* **7**, 5447 (2017).
- 775. Li, X. et al. Structural basis for modulation of human Na(V)1.3 by clinical drug and selective antagonist. *Nat. Commun.* **13**, 1286 (2022).
- 776. Huang, J., Fan, X., Jin, X., Teng, L. & Yan, N. Dual-pocket inhibition of Na(v) channels by the antiepileptic drug lamotrigine. *Proc. Natl Acad. Sci. USA* **120**, e2309773120 (2023).
- 777. Wu, Q. et al. Structural mapping of Na(v)1.7 antagonists. Nat. Commun. 14, 3224 (2023).
- Huang, G. et al. Unwinding and spiral sliding of S4 and domain rotation of VSD during the electromechanical coupling in Na(v)1.7. Proc. Natl Acad. Sci. USA 119, e2209164119 (2022).
- 779. Gao, S. et al. Structural basis for human Ca(v)1.2 inhibition by multiple drugs and the neurotoxin calciseptine. *Cell* **186**, 5363–5374.e5316 (2023).
- 780. Dong, Y. et al. Closed-state inactivation and pore-blocker modulation mechanisms of human Ca(V)2.2. *Cell Rep.* **37**, 109931 (2021).
- 781. Yao, X. et al. Structures of the R-type human Ca(v)2.3 channel reveal conformational crosstalk of the intracellular segments. *Nat. Commun.* 13, 7358 (2022).
- 782. Zhao, Y. et al. Cryo-EM structures of apo and antagonist-bound human Ca(v)3.1. *Nature* **576**, 492–497 (2019).
- Li, T. et al. Structural basis for the modulation of human KCNQ4 by smallmolecule drugs. *Mol. Cell* 81, 25–37.e24 (2021).
- Neuberger, A., Nadezhdin, K. D. & Sobolevsky, A. I. Structural mechanism of TRPV3 channel inhibition by the anesthetic dyclonine. *Nat. Commun.* 13, 2795 (2022).
- Singh, A. K., McGoldrick, L. L. & Sobolevsky, A. I. Structure and gating mechanism of the transient receptor potential channel TRPV3. *Nat. Struct. Mol. Biol.* 25, 805–813 (2018).
- 786. Hua, T. et al. Activation and signaling mechanism revealed by cannabinoid receptor-G(i) complex structures. *Cell* **180**, 655–665.e618 (2020).
- Xing, C. et al. Cryo-EM structure of the human cannabinoid receptor CB2-G(i) signaling complex. *Cell* 180, 645–654.e613 (2020).
- 788. Laverty, D. et al. Cryo-EM structure of the human α1β3γ2 GABA(A) receptor in a lipid bilayer. *Nature* 565, 516–520 (2019).
- Sente, A. et al. Differential assembly diversifies GABA(A) receptor structures and signalling. *Nature* 604, 190–194 (2022).

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- 790. Park, J. et al. Structure of human GABA(B) receptor in an inactive state. Nature 584, 304–309 (2020).
- 791. Kumar, A. et al. Structural basis for cannabinoid-induced potentiation of alpha1glycine receptors in lipid nanodiscs. *Nat. Commun.* **13**, 4862 (2022).
- 792. Seven, A. B. et al. G-protein activation by a metabotropic glutamate receptor. *Nature* **595**, 450–454 (2021).
- 793. Liu, Y. et al. Ligand recognition and allosteric modulation of the human MRGPRX1 receptor. *Nat. Chem. Biol.* **19**, 416–422 (2023).
- Draper-Joyce, C. J. et al. Positive allosteric mechanisms of adenosine A(1) receptor-mediated analgesia. *Nature* 597, 571–576 (2021).
- 795. Shen, C. et al. Structural insights into the allosteric inhibition of P2X4 receptors. *Nat. Commun.* **14**, 6437 (2023).

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