
1 **Title Page:**

2 **Comparative Analysis of Human-Chimpanzee Divergence in Brain**

3 **Connectivity and its Genetic Correlates**

4 **Short title: Chimpanzee-Human Brain Connectivity Divergence and its Genetic Correlates**

5 Yufan Wang^{1,2#}, Luqi Cheng^{3,4#}, Deying Li^{1,2}, Yuheng Lu^{1,2}, Changshuo Wang^{1,5}, Yaping Wang^{1,5},
6 Chaohong Gao^{1,5}, Haiyan Wang^{1,6}, Wim Vanduffel^{6,7,8,9}, William D. Hopkins¹⁰, Chet C. Sherwood¹¹,
7 Tianzi Jiang^{1,2,4}, Congying Chu^{1,2*}, Lingzhong Fan^{1,2,5,12*}

8 ¹ Brainnetome Center, Institute of Automation, Chinese Academy of Sciences, Beijing 100190,
9 China

10 ² School of Artificial Intelligence, University of Chinese Academy of Sciences, Beijing 100190,
11 China

12 ³ School of Life and Environmental Sciences, Guilin University of Electronic Technology, Guilin
13 541004, China

14 ⁴ Research Center for Augmented Intelligence, Zhejiang Lab, Hangzhou 311100, China

15 ⁵ Sino-Danish College, University of Chinese Academy of Sciences, Beijing 100190, China

16 ⁶ Department of Neurosciences, Laboratory of Neuro- and Psychophysiology, KU Leuven Medical
17 School, 3000 Leuven, Belgium

18 ⁷ Leuven Brain Institute, KU Leuven, 3000 Leuven, Belgium

19 ⁸ Athinoula A. Martinos Center for Biomedical Imaging, Massachusetts General Hospital,
20 Charlestown, MA 02129, USA

21 ⁹ Department of Radiology, Harvard Medical School, Boston, MA 02144, USA

22 ¹⁰ Department of Comparative Medicine, University of Texas MD Anderson Cancer Center, Bastrop,
23 TX 78602, USA

24 ¹¹ Department of Anthropology and Center for the Advanced Study of Human Paleobiology, The
25 George Washington University, Washington, DC 20052, USA

26 ¹² School of Health and Life Sciences, University of Health and Rehabilitation Sciences, Qingdao
27 266000, China

28

29 **#These authors contributed equally**

30 ***Corresponding Authors:**

31 Lingzhong Fan, Institute of Automation, Chinese Academy of Sciences, Beijing 100190, China.

32 Email: lingzhong.fan@ia.ac.cn, Phone: 010 - 8254 4523

33 or Congying Chu, Institute of Automation, Chinese Academy of Sciences, Beijing 100190, China.

34 Email: congying.chu@ia.ac.cn, Phone: 010 - 8254 4770

35

36 **Abstract**

37 Chimpanzees (*Pan troglodytes*) are humans' closest living relatives, making them the most directly
38 relevant comparison point for understanding human brain evolution. Zeroing in on the differences
39 in brain connectivity between humans and chimpanzees can provide key insights into the specific
40 evolutionary changes that might have occurred along the human lineage. However, conducting
41 comparisons of brain connectivity between humans and chimpanzees remains challenging, as cross-
42 species brain atlases established within the same framework are currently lacking. Without the
43 availability of cross-species brain atlases, the region-wise connectivity patterns between humans
44 and chimpanzees cannot be directly compared. To address this gap, we built the first Chimpanzee
45 Brainnetome Atlas (ChimpBNA) by following a well-established connectivity-based parcellation
46 framework. Leveraging this new resource, we found substantial divergence in connectivity patterns
47 across most association cortices, notably in the lateral temporal and dorsolateral prefrontal cortex
48 between the two species. Intriguingly, these patterns significantly deviate from the patterns of
49 cortical expansion observed in humans compared to chimpanzees. Additionally, we identified
50 regions displaying connectional asymmetries that differed between species, likely resulting from
51 evolutionary divergence. Genes associated with these divergent connectivities were found to be
52 enriched in cell types crucial for cortical projection circuits and synapse formation. These genes
53 exhibited more pronounced differences in expression patterns in regions with higher connectivity
54 divergence, suggesting a potential foundation for brain connectivity evolution. Therefore, our study
55 not only provides a fine-scale brain atlas of chimpanzees but also highlights the connectivity
56 divergence between humans and chimpanzees in a more rigorous and comparative manner and
57 suggests potential genetic correlates for the observed divergence in brain connectivity patterns
58 between the two species. This can help us better understand the origins and development of uniquely
59 human cognitive capabilities.

60

61 Introduction

62 Chimpanzees (*Pan troglodytes*) are among humans' (*Homo sapiens*) closest living primate relatives,
63 with a shared ancestor dating back approximately 6-8 million years ago ¹. Although chimpanzees
64 have brains that are approximately one-third the brain size of humans ^{2,3}, they demonstrate
65 similarities in neuroanatomical structure ^{4,5} and cognitive functions ⁶⁻⁹, including social behavior,
66 working memory, and tool use. Given their greater genetic proximity and more comparable
67 neurobiology to humans than other primates ¹⁰, elucidating chimpanzee brain organization is critical
68 as a comparative reference for understanding human evolution. Neuroimaging has enabled
69 quantitative comparisons of brain structure between chimpanzees and other primates ^{5,11-13}.
70 However, morphological changes in cortical areas alone cannot fully explain evolutionary adaptations,
71 especially with regard to the association cortices ^{14,15}.

72 Evolutionary changes in wiring space, especially in the white matter tracts beneath the cortex,
73 significantly influence anatomical and functional differences between species ^{14,16,17}. These
74 anatomical connections characterize brain regions and support flexible cognitive functions ^{18,19}.
75 Recent studies have mapped chimpanzee brain connections using diffusion MRI and revealed
76 substantial interspecies differences with humans ^{14,16,17,20-24}. Moreover, understanding brain
77 evolution requires a genetic perspective, as genetic factors strongly influence neural connectivity
78 and uncover molecular mechanisms driving interspecies differences in brain connections, revealing
79 insights into cognitive diversity and adaptation among primates ^{25,26}. However, comprehensive
80 whole-brain connectional analyses and comparisons between chimpanzees and humans, coupled
81 with genetic investigations into species differences in brain connections, are still lacking. Another
82 critical consideration is the neuroanatomical and functional asymmetries observed in both human
83 and non-human primates ^{27,28}. These asymmetries have been linked to faculties that require higher-
84 order cognitive processing, such as language and complex tool use ^{29,30}. Previous comparative
85 asymmetry studies mainly examined local structural features ^{31,32}, and limited research on
86 hemispheric asymmetries of connections to date only focused on few localized regions ^{33,34}.
87 Therefore, mapping brain-wide connectional asymmetries could illuminate lateralized human-
88 specific cognitive functions.

89 However, a major challenge in cross-species neuroscience is the lack of a common brain
90 reference system that facilitates comparison across different species. Therefore, the development of
91 brain atlases capable of facilitating such comparisons is essential for revealing the (dis)similarities
92 between species at the level of biologically meaningful subregions³⁵. Previous comparative
93 analyses defined homologous brain regions between species using cytoarchitecture,
94 myeloarchitecture, macroanatomy, connectivity patterns, functional activation, or a combination of
95 these features, leading to a variety of brain atlases for humans³⁶⁻³⁹ and chimpanzees^{12,21,40,41}.
96 However, inconsistencies in the modalities and scales used to construct these atlases render cross-
97 species comparisons challenging. Recent connectivity-based parcellation successfully delineated
98 distinct brain areas in humans, macaques, and marmosets using anatomical connections^{37,42,43}. This
99 demonstrates the feasibility of parcellating the chimpanzee brain based on diffusion MRI. Such a
100 standardized atlas approach also enables meaningful cross-species comparisons³³. Meanwhile,
101 generating homologous white matter tracts across species within a connectivity blueprint framework
102 has been used to predict homologous areas between species, even when their relative locations have
103 changed. This approach also helps identify unique aspects of brain organization, offering new
104 opportunities for investigating evolutionary changes in brain wiring^{44,45}.

105 To comprehensively investigate chimpanzee-human connective divergence, in the present
106 study, we first created the most refined atlas of the chimpanzee brain, i.e., the Chimpanzee
107 Brainnetome Atlas (ChimpBNA), following a well-established connectivity-based parcellation
108 framework³⁷ (Figure 1A). We then reconstructed homologous white matter tracts and built
109 connectivity blueprints for both humans and chimpanzees^{44,45}. Leveraging these blueprints, we
110 investigated cross-species connectivity divergence at the subregion level and their associated white
111 matter tracts (Figure 1B). Next, we examined the lateralization of connectivity patterns across the
112 entire brain in each species and aligned it to a common space¹⁴ to explore the relationship to
113 connectivity divergence between species (Figure 1C). Finally, we identified the genes and their
114 expression patterns associated with connectivity divergence between the species (Figure 1D). This
115 fine-grained chimpanzee parcellation and comprehensive cross-species connective analysis
116 provide new insights into human brain evolution.

117

118 Results

119 Connectivity-based Parcellation of the Chimpanzee Brain

120 Following the connectivity-based parcellation framework modified from our previous study (Figure
121 S1)³⁷, we first delineated 26 initial seed masks (19 cortical and 7 subcortical masks) on the
122 chimpanzee brain template and registered them to individual brains. We calculated the whole-brain
123 connectivity of each region and subdivided it into several clusters following the validation of indices
124 based on the reproducibility of the subjects' interhemispheric consistency in the topological
125 relationships. Using the connectivity derived from the probabilistic tractography of 46 chimpanzees
126 *in vivo* diffusion MRI data, we subdivided the brain into 200 cortical (Figure 2A) and 44 subcortical
127 regions (Figure S2), thereby building ChimpBNA, the most refined atlas of the chimpanzee brain
128 to date. The ChimpBNA parcellation is interactively accessible via the web viewer (Figure S3,
129 https://molicaca.github.io/atlas/chimp_atlas.html).

130 Considering the homology of sulci and gyri between chimpanzees and humans, the definition
131 and naming of initial regions of the chimpanzee brain followed that of the human brain when
132 constructing the Human Brainnetome Atlas (HumanBNA)³⁷, where 20 initial cortical seeds were
133 delineated. Here, we merged one seed, i.e., the posterior superior temporal sulcus (pSTS), into other
134 initial regions in the temporal lobe due to its uncertain definition in chimpanzees, thus obtaining 19
135 cortical seeds (Table S1). The boundaries between two large gyri were manually edited at the mid-
136 point of the sulcus. Due to the scarcity of chimpanzee brain atlases that could be employed for
137 referencing the labeling schemes, we named the subregions according to their topological positions.
138 The terminology of the parcellation is provided in Table S2. Although detailed whole-brain
139 cytoarchitectonic data from chimpanzee brains are unavailable for alignment, we utilized the
140 cytoarchitectonic maps of three well-described regions for validation^{21,40,46-49} and found good
141 consistency between histology and connectivity-base parcellation (inferior parietal lobule, Figure
142 S4A; inferior frontal gyrus, Figure S4B; superior temporal gyrus, Figure S4C). The details of the
143 parcellation results for each initial region are shown in Figure S5-S30.

144 To evaluate the validity of the ChimpBNA, distance-controlled boundary coefficient (DCBC)
145 was used to measure the homogeneity and heterogeneity of the parcellations using structural

146 connectivity⁵⁰. DCBC is an unbiased criterion that can compare within-parcel to between-parcel
147 similarity considering the spatial autocorrelation of the brain data and the scale of the parcellations.
148 A higher DCBC indicated that the parcellation provided a brain topography with higher within-
149 parcel similarity and higher between-parcel dissimilarity. We compared the DCBC of ChimpBNA
150 using the structural connectivity of each vertex/voxel with other parcellations of chimpanzees,
151 including Bailey and Bonin's parcellation (BB38)⁴⁰, and Davi130 parcellation¹². The results
152 demonstrated that ChimpBNA achieved the best performance for the delineation of the chimpanzee
153 brain based on structural connectivity (paired t-test, all $p < .001$, Bonferroni corrected; Figure S31).

154 To reveal the connectivity patterns of the subregions of ChimpBNA, we first obtained a region-
155 to-region connectivity matrix (Figure S32). Each term represented the structural connectivity
156 between subregions and was the count of the connected streamlines of probabilistic tractography.
157 More details of the intra- and inter-hemispheric connections between regions are shown in Figure
158 S32A-C. Taking the left SFG.r as an example, this area had the majority of its connections with
159 frontal subregions, including the SFG.ri, MFG.r, IFG.r, OrG.m, CG.r, and other subcortical regions
160 in the ipsilateral hemisphere and with the SPL.c, INS.rd, INS.cd, and CG.vc in the contralateral
161 hemisphere through the corpus callosum (Figure S32D).

162 We further assessed the structural hemispheric asymmetry of the chimpanzee brain for each
163 ChimpBNA subregion. The majority of subregions with greater gray matter volume in the left
164 hemisphere after adjusting for Bonferroni correction were located in the rostral inferior parietal
165 lobule (IPL), insula, rostral fusiform gyrus (FuG), and posterior middle frontal gyrus (MFG), while
166 gray matter volumes in the right were caudal IPL, anterior MFG, middle temporal gyrus (MTG),
167 and part of lateral occipital gyrus (Figure S33A). Leftward surface area asymmetry patterns were
168 found in rostral IPL, anterior temporal lobe, and medial occipital gyrus, while rightward
169 asymmetries were found in posterior IPL, anterior cingulate, and MTG (Figure S33B). The
170 hemispheric asymmetric results showed consistent patterns with previous studies^{12,32,33}.

171 We then reconstructed 45 homologous white matter tracts of the chimpanzee brain following
172 the protocols in a previous study⁴⁴ and performed probabilistic tractography from each vertex of
173 the white/gray matter surface to the whole brain. The regional connectivity blueprint was then
174 derived by multiplying the unwrapped white matter tracts matrix by the whole-brain connectivity

175 matrix ⁴⁵, with the middle cerebellar peduncle (MCP) disregarded because of its absence of
176 projection to the cortex. The rows showed the region-to-tract connectivity pattern of each subregion.
177 Taking the left SFG.r as an example again, the subregion was mainly connected with the forceps
178 minor (FMI), left inferior fronto-occipital fascicle (IFOF), and anterior thalamic radiations (ATR)
179 (Figure 2B).

180 **Connectivity Divergence between Species**

181 Leveraging connectivity blueprints built for chimpanzees and humans, we explored the connectivity
182 divergence between these two species. A modified dissimilarity measure used to quantify the
183 difference between two probability distributions, the symmetric Kullback-Leibler (KL) divergence
184 ⁴⁵, was calculated to measure the dissimilarity of the regional connectivity blueprints of the two
185 species between each subregion in the ChimpBNA and the HumanBNA. The minimum divergence
186 of each subregion of the HumanBNA was denoted as the connectivity divergence of that region,
187 resulting in a connectivity divergence map (Figure 3A, S34). Higher values in the divergence map
188 indicated that the region in humans had a connectivity pattern that was more dissimilar to the regions
189 in chimpanzees, i.e., might not be represented in this close phylogenetic relative ⁴⁵. One should note
190 that cladistic inferences about the direction of evolutionary change in trait values cannot be made
191 because the brain connectivity of the last common ancestor is not known. The minimum divergence
192 of each subregion of the ChimpBNA was also calculated (Figure S35).

193 As shown in Figure 3A, regions with the most different connectivity patterns between species
194 were located at the middle and posterior temporal lobe, especially in the anterior superior temporal
195 sulcus (aSTS) and both the rostral and caudal portion of the posterior superior temporal sulcus
196 (rpSTS and cpSTS), the caudal part of the IPL (corresponding to A39rv or PGa ⁵¹), the anterior part
197 of the precuneus (Pcun), the insula, and the inferior frontal gyrus (IFG), especially dorsal area 44
198 (A44d). In contrast, the occipital and sensorimotor cortices showed lower divergence. Notably, we
199 found that the connectivity divergence map showed a very low correlation with the map of cortical
200 expansion ($r = 0.057$, $p_{\text{spin}} = .579$; Figure 3C, S36) ¹³, although several regions in the prefrontal
201 cortex showed both morphological and connectional divergence (Figure 3C, right). This indicated
202 that the connectional changes reflect a unique aspect of brain reorganization.

203 We next grouped the cortical subregions into seven intrinsic functional networks, i.e., the visual
204 (VIS), somatomotor (SMN), limbic (LN), dorsal-attention (DAN), ventral-attention (VAN),
205 frontoparietal (FPN), and default mode network (DMN) (Figure S37B, left) ⁵². Higher-order
206 cognitive networks displayed greater divergence than the VIS/SMN (Mann-Whitney U test, p
207 = .0045; Figure S37B, right), with the FPN having the greatest divergence.

208 Taking the precuneus as an example, the anterior (A5m) and the dorsal-middle (A7m)
209 subregions showed greater connectivity divergence than the ventral-middle (A31) and posterior
210 (dmPOS) subregions, highlighting the connectivity dissimilarity between chimpanzees and humans
211 in this heterogeneous region (Figure 3B, a). Different connectional patterns of the superior
212 longitudinal fasciculus I (SLF1), IFOF, and acoustic radiation (AR) contributed to the divergence
213 between chimpanzees and humans (Figure S38A, B). As for the IPL, the anterior (A40rv) and
214 posterior (A39rv) subregions showed greater divergence than the other subregions in the IPL (Figure
215 3B, b; Figure S38C, D), and in the insular cortex, the anterior subregions presented a greater
216 difference than the posterior subregions (Figure 3B, c; Figure S38E).

217 We used NeuroSynth to perform a meta-analysis to investigate the relationship between the
218 divergence maps and various cognitive functions examined in human subjects. This analysis showed
219 that regions with high divergence between humans and chimpanzees were characterized by higher-
220 order functions, such as "memories," "strategic," and "shape." In contrast, the lower end of the
221 divergence map was related to more basic sensory and motor processing, including "arm,"
222 "somatosensory," and "foot" (Figure 3D).

223 We further compared atlases of chimpanzees and humans using connectivity blueprints as
224 homologous features. For visualization, we projected regional connectivity blueprints of the two
225 species to a low-dimensional space using t-SNE and visualized each region in a 2-dimensional space
226 (Figure 3E). Regions with similar connectivity profiles were grouped together in the resulting space,
227 and regions belonging to one of the cortical systems (frontal, sensorimotor, temporal, parietal,
228 insular, cingulate, and occipital) were labeled with distinct colors. The sensorimotor, cingulate, and
229 occipital regions tended to form a group, but the other regions were more scattered, especially the
230 insular subregions, which were scattered among different cortical systems (Figure 3E). The median
231 coordinates of each cortical system were also calculated (Figure 3E, inset). The primary cortex,

232 including the sensorimotor and occipital regions, was closer between chimpanzees and humans,
233 while the frontal and parietal regions were farther apart, with the largest dissimilarity in the temporal
234 regions.

235 **Whole-brain level Connectional Lateralization**

236 To investigate the connectional lateralization of subregions of the chimpanzee brain, we split the
237 connectivity blueprint into two hemispheres, CB_L and CB_R , which only contained the respective left-
238 or right-hemispheric tracts and the commissural tracts (21 unilateral tracts and 2 commissural tracts).
239 We calculated the KL divergence between the homotopic subregions of the hemispheres of the
240 chimpanzees, indicating the inter-hemispheric differences in connectivity patterns. As shown in top
241 left panel of Figure 4A, the regions with highly asymmetric connectivity patterns included the
242 posterior temporal lobe, IPL, medial occipital cortex, and MFG. Inter-hemispheric differences in
243 human brain regions were also calculated the same as in the chimpanzee data, and the patterns were
244 largely consistent with previous studies (Figure 4A, bottom right) ⁵³.

245 Since the data for the two species were not in common space and could not be compared
246 directly, we used an alignment technique based on myelin data to transform the asymmetric
247 connectivity pattern between chimpanzees and humans ^{14,54}. The cross-species alignment was based
248 on a multimodal surface matching algorithm (MSM) ⁵⁵, using the distinction of areas in myelin
249 maps as anchor points, and derived a cortical registration between species. This allowed us to
250 compare species-shared and species-specific regions with asymmetric connectivity patterns in a
251 common space.

252 In the common human space, we focused on the human-specific regions that had asymmetric
253 connectivity patterns by calculating a variant of the exclusive *OR* maps between the chimpanzee
254 and human brains (Figure 4B) ⁵⁶. As shown in the bottom left panel of Figure 4B, the human maps
255 showed unique regions with asymmetric connectivity in the dorsal part of the IPL, anterior part of
256 the insular cortex, middle cingulate cortex (MCC), posterior part of the orbitofrontal cortex (OFC),
257 and most of the lateral prefrontal cortex (PFC). Weighted local correlation maps between the human-
258 specific asymmetry pattern and the connectivity divergence map provide a visualization of the
259 reorganization of the brain connectivity between the species. Regions with high values in the human

260 brain indicate marked connectivity differences between chimpanzees and asymmetry between
261 hemispheres. The posterior inferior parietal lobule, temporal lobe, posterior orbitofrontal cortex,
262 and dorsolateral prefrontal cortex showed the greatest differences (Figure 4B, bottom right).

263 Similarly, we found the chimpanzees showed unique connectional asymmetries in the several
264 posterior temporal regions and medial occipital cortex (Figure 4B, top left), and showed marked
265 connectional changes in the caudoventral part of the temporal lobe (Figure 4B, top right), which
266 could be considered chimpanzee-unique connectional features distinct from human.

267 We next investigated the tract contribution to the asymmetric pattern for several example
268 subregions. For the rostradorsal area 39 (A39rd) in humans, which showed human-specific
269 asymmetric connectivity pattern, the asymmetric tract IFOF, middle longitudinal fasciculus (MdLF)
270 and superior longitudinal fasciculus II (SLF2) showed their contribution (Figure 4C, left), while for
271 the IPL.v, which also showed an inter-hemispheric difference in chimpanzee, the asymmetry was
272 mainly driven by the SLF2 (Figure 4C, middle). This is consistent with the finding that the C2
273 subregion in the inferior parietal lobule of chimpanzees showed significant rightward asymmetric
274 connections with the SLF2³³. The SLF2 also contributed to the asymmetry observed in MFG
275 (Figure S39A). For the highly asymmetric MTG.cv in chimpanzees, the inferior longitudinal
276 fascicle (ILF) and vertical occipital fascicle (VOF) showed distinct lateralized connections (Figure
277 4C, right). The connectional asymmetries in MVOcC.rd and MVOcC.cv were driven by the
278 significant symmetric connections with optic radiation (Figure S39B).

279 **Gene Associations with Connectivity Divergence between Species**

280 We next investigated the association between the connectivity divergence map and gene expressions
281 using the Allen Human Brain Atlas (AHBA)⁵⁷. Note that the gene analysis was limited to left
282 hemisphere because few samples were obtained from the right hemisphere in AHBA. We used
283 partial least squares regression (PLSR) on the AHBA data and generated the first component (PLS1
284 score). The PLS1 score significantly correlated with the divergence map after a 10,000 times spin
285 test ($r = 0.39$, $p_{\text{spin}} < .011$; Figure 5A). 1939 genes with a Z -score greater than 3 were filtered using
286 10,000 times bootstrapping, including *EFCAB1*, *NUDT11*, *C2CD4C*, and *SYT17* which showed
287 higher Z -score (Table S3). These genes were enriched in excitatory neurons that showed the highest

288 level of significance, followed by astrocytes (Figure 5B). Specifically, the L6 and L2-3
289 intratelencephalic excitatory neurons were the most strongly enriched. The genes with a Z-score
290 higher than 3 were related to neuronal formation, neuron projection, and synapses (Figure 5C).

291 In addition, 71 of these genes significantly overlapped with human-accelerated genes related
292 to brain processes (HAR-BRAIN genes¹³) ($p < .005$; Figure 5D, Table S4). We also quantified the
293 evolutionary rate, i.e., dN/dS ⁵⁸, of these genes to examine whether these genes experienced positive
294 selection ($dN/dS > 1$), neutral selection ($dN/dS = 1$), or negative selection ($dN/dS < 1$) since the last
295 common ancestor of humans and chimpanzees. Macaque (*Macaca mulatta*) data was also utilized
296 for comparison with chimpanzee-human results. Fifty-six of 1939 genes were found to have $dN/dS >$
297 1 in the chimpanzee-human clade, whereas only 14 genes with positive selection were found for the
298 macaque data (Welch's t-test, $p < .0001$; Figure 5E, Table S5).

299 We further examined whether these filtered genes show differential expression between
300 humans and chimpanzees. We used comparative gene expression data from the PsychENCODE
301 database²⁶, which contains the expression levels of 16463 genes at 16 homologous brain locations
302 in humans ($n = 6$), chimpanzees ($n = 5$), and macaques ($n = 5$). Of the 1939 genes with Z-scores
303 above 3, 1473 overlapped with the PsychENCODE data and were used for further study (Table S6).
304 Gene expression data were normalized across the cortical areas to obtain Z scores and averaged to
305 a group-level gene expression for humans and chimpanzees. We selected three regions of interest
306 that showed distinct connectivity divergence in our study, i.e., the superior temporal cortex (STC),
307 dorsolateral frontal cortex (DFC), and primary visual cortex (V1C). We first assessed the two
308 species' differences using a paired t-test for each region. We found significant differences in first
309 two regions (STC: $t = 16.26$, $p < .001$, Bonferroni corrected; DFC: $t = 8.16$, $p < .001$, Bonferroni
310 corrected; V1C: $t = -1.88$, $p = .0599$) and more significant effect size in the STC and DFC than in
311 the V1C (STC: Cohen's $d = 0.42$; DFC: Cohen's $d = 0.21$; V1C: Cohen's $d = 0.05$). The differentially-
312 expressed genes between species for each region were called at a FDR of 0.01 (122 genes in STC,
313 92 genes in DFC, 118 genes in V1C; Figure S40) and were not attributable to variations in the ratio
314 of major cell types between species (Figure S41). This indicates a potential association between
315 gene expression differences and connectivity divergence between species.

316

317 **Discussion**

318 This study systematically compared the connectivity profiles of chimpanzees and humans, revealing
319 divergent connectivity patterns in wiring space across species, the potential genes involved, and the
320 asymmetric connectivity pattern at the whole-brain level. To enable cross-species comparisons, we
321 first developed a fine-grained atlas of the chimpanzee brain, ChimpBNA, based on anatomical
322 connectivity information and following our previous framework developed for the human brain. We
323 subdivided the chimpanzee brain into 122 subregions (100 cortical and 22 subcortical regions) per
324 hemisphere and mapped the structural connectivity pattern for each subregion. We used these
325 connectivity blueprints to examine connectivity divergence between chimpanzees and humans.
326 Specifically, we found that the two species showed profound dissimilarities in the posterior temporal
327 lobe, IPL, Pcu, insula, and IFG. We next explored the lateralization of the connectivity patterns for
328 each species and showed human-unique asymmetries in the insula, OFC, and lateral PFC, and
329 chimpanzee-specific asymmetries in the caudoventral temporal regions and medial occipital. The
330 genes filtered according to connectivity divergence were especially enriched in cells involved in the
331 construction of cortical projection circuits and the formation of synapses. The genes showed
332 different evolutionary rates and expression patterns between the species, suggesting a potential link
333 between gene expression and connectivity.

334 **Fine-grained Chimpanzee Brain Parcellation using Anatomical** 335 **Connectivity**

336 Atlases are essential for investigating the structural and functional characteristics of the brain by
337 providing a map in a common space and allowing comparison of results across studies⁵⁹. A series
338 of comparable brain atlases with a uniform parcellation scheme from different species could
339 facilitate efficient comparative studies and provide vital clues about how the human brain evolved
340^{60,61}. However, mapping non-human primate brains is incomplete and relatively preliminary, with
341 previous atlases having been constructed using different modalities and at different scales. This
342 hinders the translation of results across species and the understanding of human brain structure and
343 function³⁵. Although a few atlases have been developed for chimpanzee brains^{12,21,40,41}, a fine-

344 grained parcellation utilizing a uniform scheme comparable to humans has been lacking. To address
345 this deficiency, we constructed the ChimpBNA using anatomical connectivity profiles, which will
346 improve our understanding of the anatomical brain organization of chimpanzees and enable
347 comparative analysis across species.

348 The earliest chimpanzee whole brain atlas dates back to the 1950s when Bailey and his
349 colleagues delineated cortical regions based on histological sections⁴⁰. Although later mapped to
350 MRI slices, the information available is limited due to its coarseness^{21,41}. A more recent parcellation,
351 Davi130¹², was manually segmented based on macroanatomical landmarks but lacks connectivity
352 information on its subregions. While acknowledging that there is no consensus as to which type of
353 information reflects the true anatomical organization of the brain^{62,63}, the ChimpBNA provides a
354 whole-brain parcellation of the chimpanzee brain into robust and biologically plausible subregions,
355 together with detailed characterizations of structural connectivity patterns for each area. Anatomical
356 connections confirmed the boundaries previously detected by cytoarchitecture and identified several
357 subdivisions not previously reported in brain mapping of human and other non-human primates^{37,64}.
358 For example, the IPL contained 5 subregions in our atlas (Figure S4A, Figure S17) and showed
359 consistent rostral-caudal topological pattern across various parcellation schemes, except for a
360 ventral subregion located in the parietal operculum, which has been reported both by anatomical
361 connection-based and cytoarchitectonic parcellation^{33,46,65}. For another example, the chimpanzee's
362 IFG has received extensive attention because of its potential association with the evolution of
363 communication and language^{66,67}. Previous studies demonstrated that a homolog of Broca's area is
364 present in chimpanzee brains, but interindividual variability in the boundaries of area 44 and area
365 45 does not align with morphological sulcal anatomy⁶⁸. Here, we segregated the IFG into 6 more
366 refined subregions: two dorsal and a ventral portion of putative area 44, the rostral and caudal parts
367 of putative area 45, and one most rostral cluster approaching the frontal pole (Figure S4B, Figure
368 S7), providing a potential reference for further analysis of this area. Meanwhile, another language-
369 related homolog in the chimpanzee brain, the area Tpt, a component of Wernicke's area⁴⁸, was also
370 present in the posterior STG in the ChimpBNA (Figure S4C, Figure S11).

371 **Connectivity Divergence between Species**

372 Comparing evolutionary brain changes is fundamental for understanding anatomical and functional
373 similarities and differences across species⁶⁹. Although the expansion of brain areas and resulting
374 reorganization between chimpanzees and humans has been investigated¹³, the divergence in
375 connectivity patterns between humans and their closest primate relatives in the evolutionary tree
376 has yet to be comprehensively analyzed. The connectivity blueprints framework provides a common
377 space to enable comparisons of cortical organization and white matter connectivity across species,
378 quantitatively identifying not only common principles but also unique specializations throughout
379 evolution⁴⁵.

380 The current study presents a connectivity divergence map showing cortical regions such as the
381 middle and posterior temporal lobe, inferior parietal, precuneus, insular, and inferior frontal gyrus
382 with particularly higher connectivity divergence between humans and chimpanzees. Functional
383 decoding analysis using NeuroSynth data indicated that these regions were related to "memories,"
384 "shape," "strategic," and "self"⁷⁰⁻⁷² when assessed in humans. That said, it is important to recognize
385 that this analysis is derived from human data alone. The functional role that these pathways have in
386 chimpanzee cognition has not been studied, so it is difficult to make direct comparisons between
387 species. Further, humans and chimpanzees are different in many other anatomical and physiological
388 features (i.e., bipedalism, vocal control, etc.) that might be related to evolutionary changes in
389 connectivity that are not captured in the Neurosynth meta-analytic approach we adopted in this paper.
390 These limitations aside, given the evidence that evolutionary changes in anatomical connections
391 may contribute to functional differences in neural systems across species^{16,17,20,21}, the overall results
392 point to divergences in neuroanatomy that may potentially support unique species-specific cognitive
393 and motor specializations⁷³.

394 The precuneus is a heterogeneous region that is implicated in complex cognitive functions. The
395 anterior (A5m) and middle (A7m) subregions, which are involved in sensorimotor and cognitive
396 processes, had a greater divergence than the posterior subregion (dmPOS), which is associated with
397 visual processing⁷⁴. Chimpanzees generally have stronger and more flexible limbs to maneuver in
398 their environment, which could cause the difference in the anterior subregions, while humans

399 outperform apes in various cognitive tasks ⁷⁵.

400 As for the lateral parietal cortex, previous studies have shown that the parietal operculum of
401 chimpanzees exhibited asymmetry and was involved with handedness in the use of tools ⁷⁶.
402 Although non-human primates share many tool capabilities, humans uniquely show greater manual
403 dexterity and conceptual knowledge of tool use ⁷⁰. Considerable divergence was found in the tool
404 processing network nodes ^{70,77}, the inferior frontal gyrus, and the most rostroventral inferior parietal
405 lobule (A40rv). These two regions were connected by the IFOF and SLF3, which have been linked
406 with the human tool-use circuit ⁷⁸⁻⁸⁰ and showed a different connectivity pattern between
407 chimpanzees and humans. While the rostroventral group of areas of the IPL deals with tool use and
408 sound, the middle and caudal IPL are implicated in nonspatial attention processes and semantic
409 processing, respectively ⁵¹. This is reflected by greater divergence in the A39rv (PGa) than in other
410 subregions of the IPL.

411 Of the cortical regions that showed the greatest connectivity divergence, the temporal lobe
412 merits special attention due to the significant changes that have been described in this region over
413 primate evolution ^{20,81-84}. Although the temporal cortex does not show as much cortical expansion
414 as the frontal cortex, there was a significant connectivity difference between chimpanzees and
415 humans in the temporal cortex. The posterior temporal lobe is involved with several human-unique
416 cognitive abilities, including language comprehension. The arcuate fasciculus, which is known to
417 be associated with language processing ⁸⁵, has been described to extend further anteriorly and
418 inferiorly in the temporal lobe in humans than in chimpanzees, resulting in the alternation of
419 connectivity patterns in this area, which might support the evolutionary specialization of this area
420 ^{14,44}. These findings suggest that these regions may have been expanded and reorganized in human
421 brains and resulted in the emergence of more complex linguistic abilities, notably speech processing.

422 We also compared the atlases of the two species by projecting the connectivity blueprints into
423 a low-dimensional space, linking the human and chimpanzee brains into a common connectivity
424 space. Subregions from the same cortical systems tended to group together. However, some regions
425 belonging to a system, e.g., the insular subregions, were more scattered. The posterior insular cortex
426 connected reciprocally with the secondary somatosensory cortex, while the anterior insular cortex
427 interconnected with regions in the temporal and ventral frontal cortex ⁸⁶, indicating posterior-

428 anterior trends of connectivity divergence in the insula. Subregions of the insular cortex were
429 scattered near the frontal, temporal, and sensorimotor systems in the low-dimensional space,
430 indicating the widely different functions related to the insular cortex, from multisensory information
431 processing to the engagement of social emotions and flexible behaviors⁸⁷. In addition, the primary
432 sensory and motor cortical areas, which showed less connectivity divergence, were closer between
433 chimpanzees and humans than the frontal and parietal regions, which showed a greater distance,
434 with the greatest dissimilarity in the temporal regions. Our results suggest that the additional
435 evolutionary adaptations for the connectivity of the temporal cortex may play a unique role in human
436 specialization^{20,84}.

437 **Asymmetric Connectivity Differences between Chimpanzees and** 438 **Humans**

439 Recent neuroimaging studies have shown that hemispheric asymmetry in structural characteristics
440³², connectivity patterns^{33,34}, and functional activity^{88,89} are not confined to humans but are actually
441 widespread among non-human primates. Here, we built whole-brain connectivity blueprints, which
442 represent the anatomical connectivity pattern of each subregion using homologous white matter
443 tracts across species, to investigate the inter-hemispheric difference for each species. We found that
444 chimpanzees showed highly asymmetric connectivity patterns in the posterior lateral temporal lobe,
445 inferior parietal, medial occipital cortex, and middle frontal gyrus. In contrast, humans showed
446 highly asymmetric connectivity patterns in the posterior temporal lobe, the superior and inferior
447 parietal cortex, the anterior insular cortex, the posterior orbitofrontal cortex, and most of the lateral
448 prefrontal cortex.

449 Previous studies reported that chimpanzees showed few leftward asymmetric connections
450 between the IPL and the temporal cortex³³, including the planum temporale, anterior superior
451 temporal gyrus, and anterior superior temporal sulcus, all of which have been implicated in human
452 language comprehension⁴⁸ as well as semantic and phonologic processing^{20,90}. Tract analysis
453 showed that the IPL_{ri} displayed significant rightward asymmetric connections with the SLF2,
454 which roughly corresponded to the conclusion of the C2 cluster of the IPL in a recent study³³ and

455 was implicated in auditory perception, which is one aspect of language processing ⁹¹. The SLF2
456 connects the angular gyrus and middle frontal gyrus of the chimpanzee brain ⁴⁴. The posterior
457 portion of the middle frontal gyrus, including the MFG.vi, MFG.dc, and MFG.vc, which likely
458 correspond to PB and PC in Bailey et al. ⁴⁰, also showed asymmetric connections in the SLF2
459 (Figure S39A). These regions were reported to be more activated by observing an action than by
460 producing grasping both transitively and intransitively ⁹². Asymmetric connectivity in the MTG.cv
461 was supported by the ILF, whose left volumetric asymmetry below the temporal cortex might
462 increase the left connectivity more than the right ⁹³. The MVOcC.rd and MVOcC.cv, which were
463 situated in the ventral part of the cuneus cortex and the calcarine sulcus, showed significant
464 asymmetric connections with optic radiation (Figure S39B). The asymmetric connection of the
465 subregions might be based on the asymmetric white matter volume underlying the occipital gyrus
466 ⁹³. We note here that this study consisted of 37 right-handed and 3 left-handed human subjects, while
467 no attempt was made to match chimpanzees on the basis of handedness. Thus, one might argue
468 differences in the distribution of right- and left-handed humans and chimpanzees might explain
469 some of the findings on divergence in asymmetry between the species. Recent studies in humans
470 have reported that individual differences in handedness are weakly associated with structural
471 asymmetries ⁹⁴. Therefore, though we cannot rule out this explanation, we do not believe that this
472 factor has a significant impact on the divergence patterns reported here.

473 Given the diversity of the brain in many aspects across species, a common space is required to
474 address these considerations. Based on myelin data, we aligned the chimpanzee brain with the
475 human brain into a common space and explored species-specific regions with asymmetric
476 connectivity patterns. Human uniqueness in asymmetric connectivity was found in the dorsal part
477 of the IPL, anterior insular cortex, MCC, posterior OFC, and most of the lateral PFC. In humans,
478 these regions are involved in tool use ⁹⁵, empathy ⁹⁶, planning ⁹⁷, and abstract reasoning ⁹⁸. Though
479 chimpanzees have been reported to exhibit each of these abilities, the available evidence suggests
480 that their aptitudes in these domains of function are more limited compared to humans ⁹⁹. The
481 posterior inferior parietal lobule, the temporal lobe, and the posterior orbitofrontal cortex all showed
482 distinct asymmetry between hemispheres and connectivity divergence between species, suggesting
483 a possible starting point for explaining the functions executed by these regions, with the emergence

484 of hemispheric specializations in human evolution. However, it is important to note that the
485 observed distinct species-specific patterns based on this comparative analysis cannot resolve the
486 direction of evolutionary changes on the human or chimpanzee lineage due to uncertainty in
487 determining brain connectivity of their last common ancestor; therefore, caution should be exercised
488 when interpreting these results in a phylogenetic context.

489 **Genetic Factors Associated with Connectivity Divergence between** 490 **Species**

491 The connectivity divergence and its underlying genetic associations were further investigated. The
492 filtered genes identified by PLSR using the AHBA dataset are implicated in synapse and axon-
493 related processes and neuronal projections, which are at the basis of macroscale anatomical
494 connections^{25,100}. Several genes, including *EFCAB1*, *C2CD4C*, and *SYT17*, were predicted to
495 enable calcium ion binding^{101,102}, which plays an important role in signal transduction and other
496 cellular processes. *SYT17* is also involved in positively regulating dendrite extension and enabling
497 syntaxin binding activity¹⁰³. *TMSB10* is related to actin monomer binding activity, thus regulating
498 cell migration¹⁰⁴⁻¹⁰⁶. Other genes that had a greater weight were related to metabolism or regulation
499 of transcription factor activity.

500 The filtered genes were enriched in excitatory neurons in the upper (L2/3) and deeper (L6)
501 layers as well as being enriched in astrocytes. The excitatory pyramidal neurons contribute to the
502 construction of major cortical projection circuits¹⁰⁷, and astrocytes are involved in regulating
503 synaptic transmission and plasticity¹⁰⁸. Our findings suggest that potential interactions between
504 excitatory neurons from different cortical layers and astrocytes contribute to divergent connectivity
505 between species at the cellular level.

506 From an evolutionary perspective, the genes shows signatures of positive selection (i.e., $dN/dS >$
507 1) in the chimpanzee-human phylogenetic group (i.e., the Hominini clade) compared with macaques
508 and markedly overlapped with the HAR-BRAIN genes, which are enriched in human-evolved
509 elements that are associated with brain development, cortical expansion, and formation of
510 connections^{13,109}. Chimpanzees share a more recent common ancestor with humans than macaques,

511 so they display more similar genetic changes due to their phylogenetic proximity. They are more
512 likely to have shared genetic adaptations and positive selection events, especially those related to
513 hominoid traits ^{110,111}. These genes were expressed unequally in the cortex, with more significant
514 differences in regions with a greater connectivity divergence. This suggests that the genes behind
515 the connectivity divergence played an essential role in modern human evolution and may reflect
516 increased cognitive specialization across species. However, we note that the difference in detected
517 gene transcript abundance levels may be due to differences in cellular composition between the two
518 species. With the development of single-cell RNA sequencing technology applied to a greater range
519 of brain regions across species ^{112,113}, these confounding effects can be distinguished in future
520 studies, and more accurate biological inferences could be made.

521 **Methodological Considerations**

522 This work has some technical and methodological limitations to acknowledge. The first concern is
523 the false positives produced by tractography ¹¹⁴. However, diffusion MRI tractography has been
524 irreplaceable for *in vivo* and non-invasively investigating the organization of humans and other non-
525 human primates, such as chimpanzees. A recent study mapped both deep and superficial white
526 matter in the chimpanzee brain, providing an important understanding of this species ¹¹⁵. Together
527 with *ex vivo* neuroanatomical data, the joint analysis may mitigate these technical issues and provide
528 even more insights ^{24,116}. Second, given the different brain sizes across species, the incidence of
529 potentially more acute curvatures in some tracts should also be considered. Because of the relatively
530 low-resolution diffusion images of the chimpanzee brain, only large subcortical nuclei were
531 parcellated based on anatomical connectivity. Other smaller nuclei and the cerebellum were not
532 considered in this study. Recent studies have analyzed these non-neocortical areas ^{117,118}, which
533 deserve greater attention. Finally, regarding parcellation reliability, it should be acknowledged that
534 the *a priori* macro-anatomical boundaries may not be related to the actual differentiation of cortical
535 areas based on pure connectivity profiles ³⁷. Future work needs to quantitatively examine the
536 correspondence between the ChimpBNA and microstructural parcellations in a greater range of
537 brain regions. However, it is difficult to define the number of subregions when parcellating starting
538 from the whole cortex, but the sulci were reported to be reliable determinants of the anatomo-

539 functional organization of brains ^{119,120}. Combined with existing primate parcellations ^{42,43},
540 ChimpBNA enables efficient cross-species brain comparisons. However, accurate homologous
541 regional correspondence across species remains lacking despite uniform atlas-building schemes.
542 Establishing more accurate homology mapping across species should be a priority for future work
543 on developing comparable cross-species brain atlases.

544 **Conclusion**

545 In summary, we constructed the fine-grained ChimpBNA based on anatomical connectivity and
546 performed a systematic comparative analysis of chimpanzee and human connectivity profiles. This
547 revealed divergent patterns across species, associated genetic factors, and whole-brain connectional
548 asymmetry. Distinct connectivity divergences between species were observed in the cortical
549 association areas, including the posterior temporal, inferior parietal, precuneus, insula, and inferior
550 frontal gyri, with related genes showing differential evolution and expression. We also identified
551 shared asymmetric areas constrained to the posterior temporal, inferior parietal, and mid-cingulate
552 cortices, while unique human asymmetry was seen in the insula, orbitofrontal cortex, and lateral
553 prefrontal cortex. The genes filtered according to the connectivity divergence were enriched into
554 cell types related to the construction of cortical projection circuits and involved with the formation
555 of synapses and showed different evolutionary rates and expression patterns across species. Our
556 findings indicate that connectivity divergence in chimpanzees and humans likely reflect functional
557 specialization in evolution and provide key steps toward elucidating the diversity of primate brains.
558

559 **Methods**

560 **Data Acquisitions and Preprocessing**

561 **Human data**

562 Data from 40 healthy human adults (*Homo sapiens*, 17 males) were selected as the same subjects as
563 in the construction of the Human Brainnetome Atlas (HumanBNA) ³⁷ from the S1200 subjects
564 release of the Human Connectome Project (HCP) database ¹²¹ (<http://www.humanconnectome.org/>).
565 All the scans and data from the individuals included in the study had passed the HCP quality control
566 and assurance standards.

567 The scanning procedures and acquisition parameters were detailed in previous publications ¹²².
568 In brief, T1w images were acquired with a 3D MPRAGE sequence on a Siemens 3T Skyra scanner
569 equipped with a 32-channel head coil with the following parameters: TR = 2400 ms, TE = 2.14 ms,
570 flip angle = 8°, FOV = 224×320 mm², voxel size = 0.7 mm isotropic. DWI images were acquired
571 using single-shot 2D spin-echo multiband echo planar imaging on a Siemens 3 Tesla Skyra system
572 (TR = 5520 ms, TE = 89.5 ms, flip angle = 78°, FOV = 210×180 mm). These consisted of three
573 shells (b-values = 1000, 2000, and 3000 s/mm²), with 90 diffusion directions isotropically
574 distributed among each shell and six b = 0 acquisitions within each shell, with a spatial resolution
575 of 1.25 mm isotropic voxels.

576 **Chimpanzee data**

577 Data from 46 adult chimpanzees (*Pan troglodytes*, 18 males) were available from the National
578 Chimpanzee Brain Resource (NCBR, <http://www.chimpanzeebrain.org>). The data, including T1w
579 and DWI, were acquired using previously described procedures at the Emory National Primate
580 Research Center (ENPRC) on a 3T MRI scanner under propofol anesthesia (10 mg/kg/h) ¹²³. All
581 procedures followed protocols approved by ENPRC and the Emory University Institutional Animal
582 Care and Use Committee (IACUC, approval no. YER-2001206). All data were obtained before the
583 2015 implementation of U.S. Fish and Wildlife Service and National Institutes of Health regulations
584 governing research with chimpanzees. All chimpanzee scans were completed by the end of 2012;
585 no new data were acquired for this study.

586 T1w images were collected at a 0.7×0.7×1 mm resolution. DWI images were acquired using a

587 single-shot spin-echo echo-planar sequence for 60 diffusion directions ($b = 1000 \text{ s/mm}^2$, $TR = 5900$
588 ms; $TE = 86 \text{ ms}$; 41 slices; 1.8 mm isotropic resolution). DWI images with phase-encoding
589 directions (left–right) of opposite polarity were acquired to correct susceptibility distortion. Five b
590 $= 0 \text{ s/mm}^2$ images were also acquired with matching imaging parameters for each repeat of a set of
591 DWI images.

592 **Data preprocessing**

593 The human T1w structural data had been preprocessed following the HCP's minimal preprocessing
594 pipeline ¹²², while the chimpanzee and macaque T1w structural data were preprocessed following
595 the HCP-NHP pipelines described in previous studies ^{11,124}. In brief, the processing pipeline included
596 imaging alignment to standard volume space using FSL, automatic anatomical surface
597 reconstruction using FreeSurfer ¹²⁵, and registration to a group average surface template space using
598 the multimodal surface matching (MSM) algorithm ⁵⁵. Human volume data were registered to
599 Montreal Neurological Institute (MNI) standard space, and surface data were transformed into
600 surface template space (fs_LR). Chimpanzee volume and surface data were registered to the
601 Yerkes29 chimpanzee template ¹¹. All subjects included in this study passed automatic FreeSurfer
602 quality control and visual inspection.

603 Preprocessing of the diffusion-weighted images was performed in a similar way in the human
604 and chimpanzee using FSL ¹²⁶. FSL's DTIFIT was used to fit a diffusion tensor model. Following
605 preprocessing, voxel-wise estimates of the fiber orientation distribution were calculated using
606 bedpostx, allowing for three fiber orientations for the human dataset and two fiber orientations for
607 the chimpanzee dataset due to the b-value in the diffusion data.

608 **Connectivity-based Parcellation**

609 **Initial seed mask definition**

610 Following the protocols we used in constructing the HumanBNA ³⁷, we identified the initial seeds
611 of the chimpanzee brain for the subsequent parcellation. Specifically, we started from the Desikan-
612 Killiany-Tourville (DKT) atlas ¹²⁷, which served as a foundational reference for identifying gyri and
613 sulci that was used as *a priori* information to guide the fine-scale subdivision process. ROIs
614 representing subdivisions of a larger gyrus, as well as distinct anatomical landmarks that the DKT

615 atlas delineates, were combined, and the boundaries between two gyri were manually edited at the
616 mid-point of the sulcus by two authors (W.Y. and C.L.). The resulting 19 cortical seeds were
617 delineated on the Yerkes29 mid-thickness surface. In addition, 7 subcortical regions were identified
618 using FreeSurfer and were included in the analysis. The full names and abbreviations of the initial
619 cortical and subcortical seed masks are listed in Table S1.

620 **Connectivity-based parcellation**

621 We used the connectivity-based parcellation procedure modified from our previous study^{33,37}.
622 Taking an initial region as a seed mask, probabilistic tractography was applied to the native
623 gray/white matter interface using probtrackx^{128,129}. Each vertex was sampled 10,000 times based
624 on the orientation probability model for each voxel, with a curvature threshold of 0.2, a step length
625 of 0.5 mm, and a number of steps of 3200. The pial surfaces were set as stop masks to prevent
626 streamlines from crossing sulci. The connectivity of voxels that were only reached by no more than
627 4/10,000 streamlines was removed. This process resulted in a whole-brain connectivity profile, a
628 matrix ($M \times N$) between all the vertices in the ROI (M) and the whole-brain voxels (N).

629 A cross-correlation matrix ($M \times M$) was calculated to quantify the similarity between the
630 connectivity profile of vertices in the ROI. Then, the cross-correlation matrix across all subjects was
631 averaged to construct a group matrix. Spectral clustering was used to define the distinct subregions
632³⁷. Since the number of clusters must be predefined, we explored 2-12 parcellations. After relabeling
633 across subjects using a group-level labelling scheme as the reference¹³⁰, we used cross-validation
634 to determine the optimal solution that yielded the greatest consistency across the subjects
635 considering three clustering indices: Cramer's V (CV), Dice coefficient (Dice), and topological
636 distance (TpD). The details of the description of these indices can be found in a previous study¹³¹.
637 The local peak of Cramer's V and Dice indicates better reproducibility than the surrounding
638 solutions, and a TpD closer to 0 indicates a more similar topological arrangement between
639 hemispheres. The pairwise strategy was used to evaluate the reproducibility of the parcellation. All
640 pairs of subjects were evaluated, and the average consistency was computed. The optimal number
641 of clusters was selected based on searching for the local peak of Cramer's V or Dice primarily, as
642 well as smaller TpD.

643 The subcortical regions were parcellated similarly but were conducted in volumetric space. For

644 a broader application, the parcellation of the cortex was also mapped to the volumetric space by
645 assigning each gray matter voxel to the label of its closest vertex.

646 **Comparisons with existing parcellations**

647 Previous parcellations of chimpanzee brains were used to compare with ChimpBNA, including (1)
648 The initial seed parcellation of ChimpBNA (Init); (2) Bailey and Bonin's brain atlas of chimpanzees,
649 which has a digital 3D version (BB38)⁴⁰; (3) Davi130 parcellation (Davi130)¹².

650 We compared the parcellations using a homogeneous metric, the distance-controlled boundary
651 coefficient (DCBC)⁵⁰. The basic idea underlying DCBC is that any two vertices/voxels within the
652 same region should have more similarity between their structural connectivity profiles than those
653 belonging to distinct regions. Taking the spatial smoothness of brain data into consideration, DCBC
654 binned each vertex/voxel pair (bin step = 1 mm) based on their geometric distance ranging from 0
655 to 100 mm and then compared the similarity between the within- and between-parcel within each
656 bin. A higher DCBC indicates that the parcellation reflected a more homogeneous topography based
657 on structural connectivity. The DCBC of ChimpBNA was compared with that of other parcellations
658 across subjects using paired t-test and adjusted using Bonferroni correction¹³².

659 **Mapping anatomical connectivity patterns**

660 To compute the structural connectome between all subregions, we performed probabilistic
661 tractography sampling 10,000 times for each voxel of each subregion and counted the connected
662 streamlines as the weight of the edge between the subregions of each subject. The connectome of
663 subjects was normalized and averaged to generate a group connectome for subsequent analysis.

664 **Hemispheric asymmetry analysis**

665 The surface area and gray matter volume of each subregion in ChimpBNA were computed from
666 FreeSurfer. The hemispheric asymmetry of subregions was determined using the formula $Asym =$
667 $(L - R) / (L + R) * 0.5$ ¹², where the L and R represent the structural index for each subregion in the
668 left and right hemispheres, respectively. Positive $Asym$ values indicate a leftward asymmetry, and
669 negative values indicate a rightward asymmetry. One-sample t-test were conducted for each
670 subregion, and significant leftward or rightward asymmetry was determined with a $p < .05$ after
671 correcting for multiple comparisons using Bonferroni correction¹³².

672 **Connectivity Analysis between Species**

673 **Construction of connectivity blueprints**

674 Using XTRACT⁵³, we reconstructed 45 homologous white matter tracts⁴⁴ for each human and
675 chimpanzee subject. We performed probabilistic tractography using FSL's probtrackx2^{128,129} to map
676 the whole-brain connectivity pattern. Specifically, the white surface was set as a seed region tracking
677 to the rest of the brain with the ventricles removed. The pial surface was used as a stop mask to
678 prevent streamlines from crossing sulci. Each vertex was sampled 10,000 times (10,000 trackings)
679 based on the orientation probability model for each voxel, with a curvature threshold of 0.2, a step
680 length of 0.5 mm, and a number of steps of 3200. This resulted in a (*whole-surface vertices*) ×
681 (*whole-brain voxels*) matrix for further analysis.

682 Tractography of the white matter tracts was vectorized to a (*tracts*) × (*whole-brain voxels*)
683 matrix and was multiplied by the whole-brain connectivity matrix, deriving a (*tracts*) × (*whole-*
684 *surface vertices*) matrix, i.e., connectivity blueprint⁴⁵. We averaged the connectivity profiles of the
685 vertices within each subregion of the HumanBNA and ChimpBNA to form the regional connectivity
686 blueprints across subjects. The matrix columns showed each subregion's connectivity distribution
687 pattern, while the rows provided the cortical termination patterns of the tracts. Note that the middle
688 cerebellar peduncle (MCP) does not project to the cortex. We disregarded the tract when
689 constructing blueprints. So the final *tracts* = 44.

690 **Connectivity divergence between species**

691 We used connectivity blueprints built for each species to explore the connectivity divergence across
692 species⁴⁵. We calculated the symmetric Kullback Leibler (KL) divergence for each pair of
693 subregions in ChimpBNA and HumanBNA and searched for the minimal value for each subregion
694 in the HumanBNA. Specifically, for an example subregion in HumanBNA (Figure S34B, a), a
695 connectivity blueprint (Figure S34B, b) represents how it was connected to each white matter tract.
696 Using the KL divergence measure, this connectivity profile was compared with that of all
697 chimpanzee subregions (Figure S34B, c). The subregion with the connectivity blueprint most
698 similar to the human one, i.e., with the smallest KL divergence, was picked, and this KL divergence
699 value was assigned to the human subregion (Figure S34B, d). A higher value on this cross-species

700 divergence map means that the subregion has a more dissimilar connectivity profile absent in
701 chimpanzees.

702 **Correlation with expansion maps of surface areas**

703 The expansion map of surface areas between chimpanzees and humans was obtained from a
704 previous study ¹³. We calculated the Spearman correlation between the cortical expansion and
705 connectivity divergence map. Spin tests (10,000 times) were used to assess the correspondence
706 between brain maps ¹³³.

707 We also investigate the local correlation between these two maps. The correlation was
708 computed using a sliding window around every vertex on the sphere, with a search kernel of 40°
709 that corresponds to a circular window with a radius of approximately 7cm ¹⁴. A weighted mask
710 derived from the multiplication of two maps was applied to the correlation map for magnification.
711 Higher values on the local correlation map indicate marked cortical expansion and connectivity
712 differences between chimpanzees and humans.

713 **Connectivity divergence in functional networks**

714 Resting-state functional networks were obtained from the Yeo 7-network atlas ⁵². We assigned each
715 cortical subregion in the HumanBNA to one of the 7 networks. For each subregion in the atlas, we
716 computed the ratio of the vertices belonging to each of the 7 networks and finally decided on the
717 label using a majority vote. Connectivity divergence among the 7 networks was examined, and the
718 divergence between the VIS/SMN networks and higher-order networks (the other five networks)
719 was compared using the Mann-Whitney U test.

720 **NeuroSynth term-based meta-analysis**

721 To investigate the association between functional decoding and the resultant cross-species
722 divergence map, we projected the meta-analytical task-based activation along the divergence map.
723 We used 590 terms related to cognitive processes that had been selected in previous publications
724 (the full list of terms can be found in a previous study ¹³⁴). The activation maps of the cognitive
725 terms were downloaded from the NeuroSynth database (<https://neurosynth.org/>). We generated 20
726 binarized masks at five-percentile increments of the divergence map, which ranged from 0-5% to
727 95%-100%, and input them into the subsequent meta-analysis. Each function term had a mean
728 activation z-score per bin. Terms included in the visualization of meta-analysis results were those

729 with the highest three z-scores in each bin. A significance threshold of $z > 0.5$ was added as a
730 visualization constraint.

731 **Connectivity embedding**

732 We used connectivity blueprints as homologous features in a common space to compare atlases
733 across species. We projected the regional connectivity blueprints of humans and chimpanzees to a
734 low-dimensional space using t-SNE. Thus, subregions with similar connectivity profiles were
735 grouped together in the resulting space. We also assigned subregions with labels of distinct cortical
736 landmarks and calculated the median coordinates of cortical systems to compare across species.

737 **Whole-brain level connectional lateralization**

738 We investigated the lateralization of the connectivity patterns at the whole-brain level. Specifically,
739 we obtained two hemispheric connectivity blueprints, CB_L and CB_R , which only contain left- or
740 right-hemispheric and commissural tracts. We calculated the symmetric KL divergence between
741 each pair of (CB_L , CB_R) rows^{45,53}, i.e., homotopic subregions between hemispheres. We calculated
742 the asymmetric connectivity pattern for humans and chimpanzees separately.

743 **Aligning chimpanzee and human brains into a common space**

744 Since the species were not in a common space and could not be compared directly, we used an
745 alignment technique based on myelin data to transform the connectivity pattern of the chimpanzee
746 to the human cortex^{14,54}. This method implemented the registration using multimodal surface
747 matching algorithm⁵⁵, initialized first by aligning a set of homologous ROIs between species,
748 followed by aligning the whole-hemisphere myelin maps using the ROIs-registration step as
749 initialization. The method had been shown to align the myelin maps well across species in the
750 previous study¹⁴, and the resulting transformations could account for relocations of other modalities.

751 After the alignment, we could explore the species-shared and species-specific regions with a
752 asymmetric connectivity patterns in a common human space. Here, we first focused on the human-
753 specific asymmetry map, which was calculated as the asymmetric connectivity pattern map of
754 humans minus the intersection between humans and chimpanzees⁵⁶, i.e., $human-(human*chimp)$,
755 with high values corresponding to regions whose connectivity patterns were asymmetric in humans
756 but not in chimpanzees. Similarly, for the chimpanzee-specific asymmetry, we calculated it as
757 $chimp-(chimp*human)$, for the species-shared asymmetry, we calculated it as $human*chimp$.

758 To investigate the relationship between the species-specific asymmetry map and the
759 connectivity divergence map we generated in the previous section, we computed a local correlation
760 map with a weighted mask to up-weight the brain areas that both showed high values. The weighted
761 mask was generated by multiplying the species-specific asymmetry map by the connectivity
762 divergence map.

763 **Gene Analysis across Species**

764 **AHBA data and preprocessing**

765 Regional microarray gene expression data were obtained from 6 postmortem brains (1 female; ages
766 24-57; $n = 4$ left-hemisphere only, $n = 2$ both left and right hemispheres) provided by the Allen
767 Human Brain Atlas (AHBA, <https://human.brain-map.org/>)⁵⁷. We only used data from the left
768 hemisphere because few samples in the AHBA were obtained from the right hemisphere. The data
769 were processed using the abagen toolbox (version 0.1.1; <https://github.com/rmarkello/abagen>)¹³⁵.
770 First, the microarray probes were reannotated to remove those that did not match a valid Entrez ID
771¹³⁶. Then, the probes were filtered based on their expression intensity relative to background noise,
772 and the probes with the maximum summed adjacency when representing the corresponding gene
773 expression were kept, yielding 15633 genes corresponding to more than one probe. Last, we
774 resampled the output gene expression map in fsaverage5 space to fs_LR32k space for subsequent
775 study and then compiled the regional expression levels for each gene using HumanBNA parcellation
776 scheme (105 subregions in the left cerebral hemisphere)³⁷ to form a 105×15633 regional
777 transcription matrix.

778 **Gene enrichment analysis**

779 We used partial least squares regression (PLSR) to characterize the genetic mechanisms underlying
780 the divergence map. We generated 10,000 surrogate maps using BrainSMASH¹³³ to ensure that the
781 explained variance of the first PLS component (PLS1) was more significant than chance. 10,000x
782 bootstrapping was used to assess the error of each gene's PLS1 weight, and Z-scores were calculated
783 as the ratio of the weight of each gene to its standard error¹³⁷. We obtained the genes that had Z-
784 scores higher than 3 and submitted the gene sets for gene enrichment analysis. TopGene
785 (<https://toppgene.cchmc.org/>)¹³⁸, which contains the whole list of AHBA genes as the background

786 gene set, was used to conduct the analysis. The following term categories were assessed: GO:
787 Molecular Function, GO: Biological Process, GO: Cellular Component, Pathway, and Disease.

788 **Cell-type enrichment analysis**

789 We further performed cell-type enrichment analysis on the filtered genes using CellGO
790 (<http://www.cellgo.world>)¹³⁹, a deep learning-based tool for cell type-specific gene function
791 interpretation. We used the single-cell datasets accessible from a recent study¹¹³. The single-nucleus
792 RNA sequencing (snRNA-seq) data from postmortem brain samples from the dorsolateral PFC of 4
793 human adults (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE207334>). Six major
794 classes of cells were provided in CellGO, including excitatory neurons (ExN), inhibitory neurons
795 (InN), oligodendrocytes (Oligo), oligodendrocyte precursor cells (OPC), astrocytes (Astro), and
796 microglia cells (Microglia). Subtypes of neurons were also available, including six excitatory
797 neurons (L2/3IT, L3/5IT, L5/6NP, L6CT, L6IT, L6b) and five inhibitory neurons (LAMP5, PVALB,
798 SST, VIP, SNCG). The enrichment P-values of the cell types resulting from the submitted genes
799 were based on the Kolmogorov-Smirnov (K-S) test in CellGO.

800 **Evolution-related analysis of genes**

801 We first explored whether the filtered genes overlapped with human-accelerated genes related to
802 brain processes (HAR-BRAIN genes)¹³ and tested the significance using the hypergeometric test.
803 Then, we quantified the evolutionary rate of these genes. The evolutionary rate, i.e., dN/dS , is
804 defined as the ratio between the nonsynonymous substitution rate and the synonymous substitution
805 rate⁵⁸. Values of $dN/dS < 1$, $= 1$, and > 1 indicate negative purifying selection, neutral evolution,
806 and positive selection, respectively. We obtained the homologous genes between chimpanzees and
807 humans in BioMart¹⁴⁰ and calculated the dN/dS as the chimpanzee-human evolutionary rate.
808 Macaque data was also selected for comparison with chimpanzee-human results. Welch's t-test was
809 used to test the statistical significance.

810 **Gene expression differences using PsychENCODE**

811 We used comparative gene expression data from the PsychENCODE database
812 (<https://evolution.psyencode.org/>)²⁶. The PsychENCODE database contains 16463 gene
813 expressions at 16 homologous brain locations (10 cortical, 5 subcortical, and 1 limbic) in three
814 species (human, $n = 6$; chimpanzee, $n = 5$; macaque, $n = 5$). The gene expression levels were

815 quantified by RPKM (reads per kilobase of exon model per million mapped reads) and corrected
816 for batch effects using the R package ComBat for normalization (v3.52.0)¹⁴¹. We further normalized
817 the gene expression data across cortical areas to *Z* scores within each individual¹³⁶ and averaged
818 across individuals to obtain a group-level gene expression for each species. Only human and
819 chimpanzee data was used in this study.

820 Due to the low spatial sampling of the cortical regions, we only selected three regions of
821 interest that showed distinct connectivity divergence for further study, i.e., the superior temporal
822 cortex (STC), dorsolateral frontal cortex (DFC), and primary visual cortex (V1C). The difference
823 between the gene expression of the two species for each region was assessed using paired t-test, and
824 the significance was determined with a $p < .001$ after correcting for multiple comparisons using
825 Bonferroni correction¹³². For each region, the differentially expressed genes between species were
826 computed with the R package DESeq (v1.44.0)¹⁴². The potential effects by cell-type ratio between
827 species was investigated following the method used in a previous publication²⁶. We first selected
828 the top 100 genes enriched in each cell type of sorted-cell of human neocortex¹⁴³. Genes enriched
829 in cell type *A* were selected by ranking the genes based on the difference between mean expression
830 in *A* and the maximum mean expression in cell types except *A*. The adjusted *p*-values of cell-type
831 enriched genes in differential gene expression analysis between species were then examined.
832

833 **Data availability**

834 The surface and volumetric representation files of the ChimpBNA are available at Github repo
835 (<https://github.com/FANLabCASIA/ChimpBNA>). The ChimpBNA parcellation is also interactively
836 accessible and downloadable via the web viewer (https://molicaca.github.io/atlas/chimp_atlas.html).
837 The chimpanzee data are available at the National Chimpanzee Brain Resource
838 (<http://www.chimpanzeebrain.org/>). The human data are available from the Human Connectome
839 Project (<https://db.humanconnectome.org/>). The Yeo 7-network atlas can be downloaded at
840 [https://github.com/ThomasYeoLab/CBIG/tree/master/stable_projects/brain_parcellation/Yeo2011_](https://github.com/ThomasYeoLab/CBIG/tree/master/stable_projects/brain_parcellation/Yeo2011_fcMRI_clustering/1000subjects_reference/Yeo_JNeurophysiol11_SplitLabels/fs_LR32k)
841 [fcMRI_clustering/1000subjects_reference/Yeo_JNeurophysiol11_SplitLabels/fs_LR32k](https://github.com/ThomasYeoLab/CBIG/tree/master/stable_projects/brain_parcellation/Yeo2011_fcMRI_clustering/1000subjects_reference/Yeo_JNeurophysiol11_SplitLabels/fs_LR32k). The
842 human gene expression data are available in the Allen Brain Atlas ([https://human.brain-](https://human.brain-map.org/static/download)
843 [map.org/static/download](https://human.brain-map.org/static/download)). The single-nucleus RNA sequencing data from the human adults are
844 available at <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE207334>. The gene
845 expression data for humans and chimpanzees are available at BioMart
846 (<https://www.ensembl.org/info/data/biomart/index.html>) and in the PsychENCODE dataset
847 (<https://evolution.psychencode.org/>). The cell-type data is available at <http://www.brainrnaseq.org/>.
848 The source data underlying Figures and Supplementary Figures are provided as Source Data at
849 Github repo (<https://github.com/FANLabCASIA/ChimpBNA>).

850

851 **Code availability**

852 The HCP-Pipeline can be found at <https://github.com/Washington-University/HCPpipelines>, and
853 the NHP-HCP-Pipeline can be found at <https://github.com/Washington-University/NHPPipelines>.
854 The neuroimaging preprocessing software used for the other datasets is freely available including
855 FreeSurfer v6.0 (<http://surfer.nmr.mgh.harvard.edu/>), and FSL v6.0.5
856 (<https://fsl.fmrib.ox.ac.uk/fsl/fslwiki>). The gene processing pipeline is available (abagen,
857 <https://github.com/rmarkello/abagen>), and the gene enrichment analysis process was conducted at
858 <https://toppgene.cchmc.org/>. The cell-type enrichment analysis process was conducted at
859 <http://www.cellgo.world>. The brain maps were presented using Workbench v1.5.0
860 (<https://www.humanconnectome.org/software/connectome-workbench>). The tracts were visualized

861 using ITK-SNAP 4.0.1 (<http://www.itksnap.org/>) and Paraview 5.11.0 (<https://www.paraview.org/>).

862

863 **Acknowledgments**

864 This work was partially supported by STI2030-Major Projects (Grant No. 2021ZD0200203), the
865 Natural Science Foundation of China (Grant Nos. 82072099, 82202253, 62250058), and the China
866 Postdoctoral Science Foundation (2022M722915). Data were provided in part by the National
867 Chimpanzee Brain Resource (supported by NIH NS092988, NIH HG011641, NIH AG067419, NSF
868 EF-2021785, and NSF DRL-2219759), Human Connectome Project, WU-Minn Consortium
869 (Principal Investigators: David Van Essen and Kamil Ugurbil; 1U54MH091657) funded by the 16
870 NIH Institutes and Centers that support the NIH Blueprint for Neuroscience Research; and by the
871 McDonnell Center for Systems Neuroscience at Washington University. The authors appreciate the
872 English language and editing assistance of Rhoda E. and Edmund F. Perozzi, PhDs.

873

874 References

- 875 1 Staes, N. *et al.* Evolutionary divergence of neuroanatomical organization and related genes in
876 chimpanzees and bonobos. *Cortex* **118**, 154-164, doi:10.1016/j.cortex.2018.09.016 (2019).
- 877 2 Van Essen, D. C. *et al.* Cerebral cortical folding, parcellation, and connectivity in humans,
878 nonhuman primates, and mice. *Proc Natl Acad Sci U S A* **116**, 26173-26180,
879 doi:10.1073/pnas.1902299116 (2019).
- 880 3 Herculano-Houzel, S. The remarkable, yet not extraordinary, human brain as a scaled-up
881 primate brain and its associated cost. *Proc Natl Acad Sci U S A* **109 Suppl 1**, 10661-10668,
882 doi:10.1073/pnas.1201895109 (2012).
- 883 4 Hopkins, W. D. *et al.* Evolution of the central sulcus morphology in primates. *Brain Behav*
884 *Evol* **84**, 19-30, doi:10.1159/000362431 (2014).
- 885 5 Hopkins, W. D., Li, X., Crow, T. & Roberts, N. Vertex- and atlas-based comparisons in
886 measures of cortical thickness, gyrification and white matter volume between humans and
887 chimpanzees. *Brain Struct Funct* **222**, 229-245, doi:10.1007/s00429-016-1213-1 (2017).
- 888 6 Call, J. Chimpanzee social cognition. *Trends Cogn Sci* **5**, 388-393, doi:10.1016/s1364-
889 6613(00)01728-9 (2001).
- 890 7 Escribano, D. *et al.* Chimpanzees organize their social relationships like humans. *Sci Rep* **12**,
891 16641, doi:10.1038/s41598-022-20672-z (2022).
- 892 8 Inoue, S. & Matsuzawa, T. Working memory of numerals in chimpanzees. *Curr Biol* **17**,
893 R1004-1005, doi:10.1016/j.cub.2007.10.027 (2007).
- 894 9 Inoue-Nakamura, N. & Matsuzawa, T. Development of stone tool use by wild chimpanzees
895 (Pan troglodytes). *J Comp Psychol* **111**, 159-173, doi:10.1037/0735-7036.111.2.159 (1997).
- 896 10 Varki, A. & Altheide, T. K. Comparing the human and chimpanzee genomes: searching for
897 needles in a haystack. *Genome Res* **15**, 1746-1758, doi:10.1101/gr.3737405 (2005).
- 898 11 Donahue, C. J., Glasser, M. F., Preuss, T. M., Rilling, J. K. & Van Essen, D. C. Quantitative
899 assessment of prefrontal cortex in humans relative to nonhuman primates. *Proc Natl Acad Sci U S*
900 *A* **115**, E5183-E5192, doi:10.1073/pnas.1721653115 (2018).
- 901 12 Vickery, S. *et al.* Chimpanzee brain morphometry utilizing standardized MRI preprocessing

-
- 902 and macroanatomical annotations. *Elife* **9**, doi:10.7554/eLife.60136 (2020).
- 903 13 Wei, Y. *et al.* Genetic mapping and evolutionary analysis of human-expanded cognitive
904 networks. *Nat Commun* **10**, 4839, doi:10.1038/s41467-019-12764-8 (2019).
- 905 14 Eichert, N. *et al.* Cross-species cortical alignment identifies different types of anatomical
906 reorganization in the primate temporal lobe. *Elife* **9**, doi:10.7554/eLife.53232 (2020).
- 907 15 Mars, R., Passingham, R., Neubert, F., Verhagen, L. & Sallet, J. *Evolutionary specializations*
908 *of human association cortex.* (2016).
- 909 16 Rilling, J. K. *et al.* The evolution of the arcuate fasciculus revealed with comparative DTI.
910 *Nat Neurosci* **11**, 426-428, doi:10.1038/nn2072 (2008).
- 911 17 Balezeau, F. *et al.* Primate auditory prototype in the evolution of the arcuate fasciculus. *Nat*
912 *Neurosci* **23**, 611-614, doi:10.1038/s41593-020-0623-9 (2020).
- 913 18 Passingham, R. E., Stephan, K. E. & Kotter, R. The anatomical basis of functional
914 localization in the cortex. *Nat Rev Neurosci* **3**, 606-616, doi:10.1038/nrn893 (2002).
- 915 19 Thiebaut de Schotten, M. & Forkel, S. J. The emergent properties of the connected brain.
916 *Science* **378**, 505-510, doi:10.1126/science.abq2591 (2022).
- 917 20 Sierpowska, J. *et al.* Comparing human and chimpanzee temporal lobe neuroanatomy reveals
918 modifications to human language hubs beyond the frontotemporal arcuate fasciculus. *Proc Natl*
919 *Acad Sci U S A* **119**, e2118295119, doi:10.1073/pnas.2118295119 (2022).
- 920 21 Ardesch, D. J. *et al.* Evolutionary expansion of connectivity between multimodal association
921 areas in the human brain compared with chimpanzees. *Proc Natl Acad Sci U S A* **116**, 7101-7106,
922 doi:10.1073/pnas.1818512116 (2019).
- 923 22 Roumazeilles, L. *et al.* Longitudinal connections and the organization of the temporal cortex
924 in macaques, great apes, and humans. *PLoS Biol* **18**, e3000810, doi:10.1371/journal.pbio.3000810
925 (2020).
- 926 23 Bryant, K. L. *et al.* Organization of extrastriate and temporal cortex in chimpanzees
927 compared to humans and macaques. *Cortex* **118**, 223-243, doi:10.1016/j.cortex.2019.02.010
928 (2019).
- 929 24 Eichner, C. *et al.* Detailed mapping of the complex fiber structure and white matter pathways
930 of the chimpanzee brain. *Nat Methods*, doi:10.1038/s41592-024-02270-1 (2024).

-
- 931 25 Li, D. *et al.* Converging Topographic Axes of Connectivity and Genetic Architectures
932 Scaffold Cortical Patterning. *bioRxiv*, 2023.2009. 2006.556618 (2023).
- 933 26 Sousa, A. M. M. *et al.* Molecular and cellular reorganization of neural circuits in the human
934 lineage. *Science* **358**, 1027-1032, doi:10.1126/science.aan3456 (2017).
- 935 27 Corballis, M. C. Evolution of cerebral asymmetry. *Prog Brain Res* **250**, 153-178,
936 doi:10.1016/bs.pbr.2019.04.041 (2019).
- 937 28 Hopkins, W. D., Misiura, M., Pope, S. M. & Latash, E. M. Behavioral and brain asymmetries
938 in primates: a preliminary evaluation of two evolutionary hypotheses. *Ann N Y Acad Sci* **1359**, 65-
939 83, doi:10.1111/nyas.12936 (2015).
- 940 29 Bishop, D. V. Cerebral asymmetry and language development: cause, correlate, or
941 consequence? *Science* **340**, 1230531, doi:10.1126/science.1230531 (2013).
- 942 30 Hopkins, W. D. *et al.* Motor skill for tool-use is associated with asymmetries in Broca's area
943 and the motor hand area of the precentral gyrus in chimpanzees (*Pan troglodytes*). *Behav Brain*
944 *Res* **318**, 71-81, doi:10.1016/j.bbr.2016.10.048 (2017).
- 945 31 Hopkins, W. D. Neuroanatomical asymmetries in nonhuman primates in the homologs to
946 Broca's and Wernicke's areas: a mini-review. *Emerg Top Life Sci* **6**, 271-284,
947 doi:10.1042/ETLS20210279 (2022).
- 948 32 Xiang, L., Crow, T. J., Hopkins, W. D. & Roberts, N. Comparison of Surface Area and
949 Cortical Thickness Asymmetry in the Human and Chimpanzee Brain. *Cereb Cortex*,
950 doi:10.1093/cercor/bhaa202 (2020).
- 951 33 Cheng, L. *et al.* Connectional asymmetry of the inferior parietal lobule shapes hemispheric
952 specialization in humans, chimpanzees, and rhesus macaques. *Elife* **10**, doi:10.7554/eLife.67600
953 (2021).
- 954 34 Xia, X., Gao, F. & Yuan, Z. Species and individual differences and connectional asymmetry
955 of Broca's area in humans and macaques. *Neuroimage* **244**, 118583,
956 doi:10.1016/j.neuroimage.2021.118583 (2021).
- 957 35 Fan, L. Mapping the Human Brain: What Is the Next Frontier? *Innovation (Camb)* **2**, 100073,
958 doi:10.1016/j.xinn.2020.100073 (2021).
- 959 36 Brodmann, K. *Vergleichende Lokalisationslehre der Grosshirnrinde in ihren Prinzipien*

-
- 960 *dargestellt auf Grund des Zellenbaues.* (Barth, 1909).
- 961 37 Fan, L. *et al.* The Human Brainnetome Atlas: A New Brain Atlas Based on Connectional
962 Architecture. *Cereb Cortex* **26**, 3508-3526, doi:10.1093/cercor/bhw157 (2016).
- 963 38 Glasser, M. F. *et al.* A multi-modal parcellation of human cerebral cortex. *Nature* **536**, 171-
964 178, doi:10.1038/nature18933 (2016).
- 965 39 Schaefer, A. *et al.* Local-Global Parcellation of the Human Cerebral Cortex from Intrinsic
966 Functional Connectivity MRI. *Cereb Cortex* **28**, 3095-3114, doi:10.1093/cercor/bhx179 (2018).
- 967 40 Bailey, P., Bonin, G. V. & McCulloch, W. S. *The isocortex of the chimpanzee.* (1950).
- 968 41 van den Heuvel, M. P. *et al.* Evolutionary modifications in human brain connectivity
969 associated with schizophrenia. *Brain* **142**, 3991-4002, doi:10.1093/brain/awz330 (2019).
- 970 42 Liu, C. *et al.* A digital 3D atlas of the marmoset brain based on multi-modal MRI.
971 *Neuroimage* **169**, 106-116, doi:10.1016/j.neuroimage.2017.12.004 (2018).
- 972 43 Lu, Y. *et al.* Macaque Brainnetome Atlas: A multifaceted brain map with parcellation,
973 connection, and histology. *Sci Bull (Beijing)*, doi:10.1016/j.scib.2024.03.031 (2024).
- 974 44 Bryant, K. L., Li, L., Eichert, N. & Mars, R. B. A comprehensive atlas of white matter tracts
975 in the chimpanzee. *PLoS Biol* **18**, e3000971, doi:10.1371/journal.pbio.3000971 (2020).
- 976 45 Mars, R. B. *et al.* Whole brain comparative anatomy using connectivity blueprints. *Elife* **7**,
977 doi:10.7554/eLife.35237 (2018).
- 978 46 Reyes, L. D. *et al.* Cytoarchitecture, myeloarchitecture, and parcellation of the chimpanzee
979 inferior parietal lobe. *Brain Struct Funct* **228**, 63-82, doi:10.1007/s00429-022-02514-w (2023).
- 980 47 Schenker, N. M. *et al.* Broca's area homologue in chimpanzees (Pan troglodytes):
981 probabilistic mapping, asymmetry, and comparison to humans. *Cereb Cortex* **20**, 730-742,
982 doi:10.1093/cercor/bhp138 (2010).
- 983 48 Spocter, M. A. *et al.* Wernicke's area homologue in chimpanzees (Pan troglodytes) and its
984 relation to the appearance of modern human language. *Proc Biol Sci* **277**, 2165-2174,
985 doi:10.1098/rspb.2010.0011 (2010).
- 986 49 Gallardo, G. *et al.* Morphological evolution of language-relevant brain areas. *PLoS Biol* **21**,
987 e3002266, doi:10.1371/journal.pbio.3002266 (2023).
- 988 50 Zhi, D., King, M., Hernandez-Castillo, C. R. & Diedrichsen, J. Evaluating brain parcellations

-
- 989 using the distance-controlled boundary coefficient. *Hum Brain Mapp* **43**, 3706-3720,
990 doi:10.1002/hbm.25878 (2022).
- 991 51 Caspers, S. *et al.* Organization of the human inferior parietal lobule based on receptor
992 architectonics. *Cereb Cortex* **23**, 615-628, doi:10.1093/cercor/bhs048 (2013).
- 993 52 Yeo, B. T. *et al.* The organization of the human cerebral cortex estimated by intrinsic
994 functional connectivity. *J Neurophysiol* **106**, 1125-1165, doi:10.1152/jn.00338.2011 (2011).
- 995 53 Warrington, S. *et al.* XTRACT - Standardised protocols for automated tractography in the
996 human and macaque brain. *Neuroimage* **217**, 116923, doi:10.1016/j.neuroimage.2020.116923
997 (2020).
- 998 54 Mars, R. B., Jbabdi, S. & Rushworth, M. F. S. A Common Space Approach to Comparative
999 Neuroscience. *Annu Rev Neurosci* **44**, 69-86, doi:10.1146/annurev-neuro-100220-025942 (2021).
- 1000 55 Robinson, E. C. *et al.* MSM: a new flexible framework for Multimodal Surface Matching.
1001 *Neuroimage* **100**, 414-426, doi:10.1016/j.neuroimage.2014.05.069 (2014).
- 1002 56 Warrington, S. *et al.* Concurrent mapping of brain ontogeny and phylogeny within a common
1003 space: Standardized tractography and applications. *Sci Adv* **8**, eabq2022,
1004 doi:10.1126/sciadv.abq2022 (2022).
- 1005 57 Hawrylycz, M. J. *et al.* An anatomically comprehensive atlas of the adult human brain
1006 transcriptome. *Nature* **489**, 391-399, doi:10.1038/nature11405 (2012).
- 1007 58 Kryazhimskiy, S. & Plotkin, J. B. The population genetics of dN/dS. *PLoS Genet* **4**,
1008 e1000304, doi:10.1371/journal.pgen.1000304 (2008).
- 1009 59 Eickhoff, S. B., Thirion, B., Varoquaux, G. & Bzdok, D. Connectivity-based parcellation:
1010 Critique and implications. *Hum Brain Mapp* **36**, 4771-4792, doi:10.1002/hbm.22933 (2015).
- 1011 60 Striedter, G. F. *et al.* NSF workshop report: discovering general principles of nervous system
1012 organization by comparing brain maps across species. *Brain Behav Evol* **83**, 1-8,
1013 doi:10.1159/000360152 (2014).
- 1014 61 Balan, P. F. *et al.* MEBRAINS 1.0: A new population-based macaque atlas. *Imaging*
1015 *Neuroscience* **2**, 1-26, doi:10.1162/imag_a_00077 %J Imaging Neuroscience (2024).
- 1016 62 Eickhoff, S. B., Constable, R. T. & Yeo, B. T. T. Topographic organization of the cerebral
1017 cortex and brain cartography. *Neuroimage* **170**, 332-347, doi:10.1016/j.neuroimage.2017.02.018

-
- 1018 (2018).
- 1019 63 Eickhoff, S. B., Yeo, B. T. T. & Genon, S. Imaging-based parcellations of the human brain.
1020 *Nat Rev Neurosci* **19**, 672-686, doi:10.1038/s41583-018-0071-7 (2018).
- 1021 64 Van Essen, D. C., Donahue, C., Dierker, D. L. & Glasser, M. F. in *Micro-, Meso- and Macro-*
1022 *Connectomics of the Brain* (eds H. Kennedy, D. C. Van Essen, & Y. Christen) 89-106 (2016).
- 1023 65 Reyes, L. D. *Tool-use and the Chimpanzee Brain: An Investigation of Gray and White Matter,*
1024 *and a Focused Study of Inferior Parietal Microstructure*, The George Washington University,
1025 (2017).
- 1026 66 Schenker, N. M. *et al.* A comparative quantitative analysis of cytoarchitecture and
1027 minicolumnar organization in Broca's area in humans and great apes. *J Comp Neurol* **510**, 117-
1028 128, doi:10.1002/cne.21792 (2008).
- 1029 67 Keller, S. S., Roberts, N. & Hopkins, W. A comparative magnetic resonance imaging study of
1030 the anatomy, variability, and asymmetry of Broca's area in the human and chimpanzee brain. *J*
1031 *Neurosci* **29**, 14607-14616, doi:10.1523/JNEUROSCI.2892-09.2009 (2009).
- 1032 68 Hopkins, W. D. J. O. o. H. L. C. & Primates, D. w. N. *Motor and communicative correlates of*
1033 *the inferior frontal gyrus (Broca's Area) in chimpanzees*. Vol. 153 (2018).
- 1034 69 Friedrich, P. *et al.* Imaging evolution of the primate brain: the next frontier? *Neuroimage* **228**,
1035 117685, doi:10.1016/j.neuroimage.2020.117685 (2021).
- 1036 70 Lewis, J. W. Cortical networks related to human use of tools. *Neuroscientist* **12**, 211-231,
1037 doi:10.1177/1073858406288327 (2006).
- 1038 71 Pasupathy, A. Neural basis of shape representation in the primate brain. *Prog Brain Res* **154**,
1039 293-313, doi:10.1016/S0079-6123(06)54016-6 (2006).
- 1040 72 Patterson, K., Nestor, P. J. & Rogers, T. T. Where do you know what you know? The
1041 representation of semantic knowledge in the human brain. *Nat Rev Neurosci* **8**, 976-987,
1042 doi:10.1038/nrn2277 (2007).
- 1043 73 van den Heuvel, M. P. *et al.* Human and chimpanzee shared and divergent neurobiological
1044 systems for general and specific cognitive brain functions. *Proc Natl Acad Sci U S A* **120**,
1045 e2218565120, doi:10.1073/pnas.2218565120 (2023).
- 1046 74 Margulies, D. S. *et al.* Precuneus shares intrinsic functional architecture in humans and

-
- 1047 monkeys. *Proc Natl Acad Sci U S A* **106**, 20069-20074, doi:10.1073/pnas.0905314106 (2009).
- 1048 75 Bruner, E., Preuss, T. M., Chen, X. & Rilling, J. K. Evidence for expansion of the precuneus
1049 in human evolution. *Brain Struct Funct* **222**, 1053-1060, doi:10.1007/s00429-015-1172-y (2017).
- 1050 76 Gilissen, E. P. & Hopkins, W. D. Asymmetries of the parietal operculum in chimpanzees (Pan
1051 troglodytes) in relation to handedness for tool use. *Cereb Cortex* **23**, 411-422,
1052 doi:10.1093/cercor/bhs029 (2013).
- 1053 77 Wen, H., Xu, T., Wang, X., Yu, X. & Bi, Y. Brain intrinsic connection patterns underlying tool
1054 processing in human adults are present in neonates and not in macaques. *Neuroimage* **258**,
1055 119339, doi:10.1016/j.neuroimage.2022.119339 (2022).
- 1056 78 Bi, Y. *et al.* The white matter structural network underlying human tool use and tool
1057 understanding. *J Neurosci* **35**, 6822-6835, doi:10.1523/JNEUROSCI.3709-14.2015 (2015).
- 1058 79 Ramayya, A. G., Glasser, M. F. & Rilling, J. K. A DTI investigation of neural substrates
1059 supporting tool use. *Cereb Cortex* **20**, 507-516, doi:10.1093/cercor/bhp141 (2010).
- 1060 80 Peeters, R. *et al.* The representation of tool use in humans and monkeys: common and
1061 uniquely human features. *J Neurosci* **29**, 11523-11539, doi:10.1523/JNEUROSCI.2040-09.2009
1062 (2009).
- 1063 81 Bryant, K. L. & Preuss, T. M. J. D. E. F. s. t. b. *A comparative perspective on the human*
1064 *temporal lobe.* (2018).
- 1065 82 Rilling, J. K. & Seligman, R. A. A quantitative morphometric comparative analysis of the
1066 primate temporal lobe. *J Hum Evol* **42**, 505-533, doi:10.1006/jhev.2001.0537 (2002).
- 1067 83 Rilling, J. K. Human temporal lobes have been reorganized: A response to Pearson *et al.*,
1068 "updated imaging and phylogenetic comparative methods reassess relative temporal lobe size in
1069 anthropoids and modern humans". *Am J Biol Anthropol* **182**, 3-6, doi:10.1002/ajpa.24798 (2023).
- 1070 84 Braunsdorf, M. *et al.* Does the temporal cortex make us human? A review of structural and
1071 functional diversity of the primate temporal lobe. *Neurosci Biobehav Rev* **131**, 400-410,
1072 doi:10.1016/j.neubiorev.2021.08.032 (2021).
- 1073 85 Becker, Y., Loh, K. K., Coulon, O. & Meguerditchian, A. The Arcuate Fasciculus and
1074 language origins: Disentangling existing conceptions that influence evolutionary accounts.
1075 *Neurosci Biobehav Rev* **134**, 104490, doi:10.1016/j.neubiorev.2021.12.013 (2022).

-
- 1076 86 Jakab, A., Molnar, P. P., Bogner, P., Beres, M. & Berenyi, E. L. Connectivity-based
1077 parcellation reveals interhemispheric differences in the insula. *Brain Topogr* **25**, 264-271,
1078 doi:10.1007/s10548-011-0205-y (2012).
- 1079 87 Gogolla, N. The insular cortex. *Curr Biol* **27**, R580-R586, doi:10.1016/j.cub.2017.05.010
1080 (2017).
- 1081 88 Wey, H. Y. *et al.* Multi-region hemispheric specialization differentiates human from
1082 nonhuman primate brain function. *Brain Struct Funct* **219**, 2187-2194, doi:10.1007/s00429-013-
1083 0620-9 (2014).
- 1084 89 Lopez-Persem, A. *et al.* Differential functional connectivity underlying asymmetric reward-
1085 related activity in human and nonhuman primates. *Proc Natl Acad Sci U S A* **117**, 28452-28462,
1086 doi:10.1073/pnas.2000759117 (2020).
- 1087 90 Insausti, R. Comparative neuroanatomical parcellation of the human and nonhuman primate
1088 temporal pole. *J Comp Neurol* **521**, 4163-4176, doi:10.1002/cne.23431 (2013).
- 1089 91 Tagliatela, J. P., Russell, J. L., Schaeffer, J. A. & Hopkins, W. D. Visualizing vocal
1090 perception in the chimpanzee brain. *Cereb Cortex* **19**, 1151-1157, doi:10.1093/cercor/bhn157
1091 (2009).
- 1092 92 Hecht, E. E. *et al.* Differences in neural activation for object-directed grasping in
1093 chimpanzees and humans. *J Neurosci* **33**, 14117-14134, doi:10.1523/JNEUROSCI.2172-13.2013
1094 (2013).
- 1095 93 Hopkins, W. D., Tagliatela, J. P., Nir, T., Schenker, N. M. & Sherwood, C. C. A voxel-based
1096 morphometry analysis of white matter asymmetries in chimpanzees (*Pan troglodytes*). *Brain*
1097 *Behav Evol* **76**, 93-100, doi:10.1159/000319010 (2010).
- 1098 94 Sha, Z. *et al.* Handedness and its genetic influences are associated with structural
1099 asymmetries of the cerebral cortex in 31,864 individuals. *Proc Natl Acad Sci U S A* **118**,
1100 doi:10.1073/pnas.2113095118 (2021).
- 1101 95 Stout, D. & Chaminade, T. Stone tools, language and the brain in human evolution. *Philos*
1102 *Trans R Soc Lond B Biol Sci* **367**, 75-87, doi:10.1098/rstb.2011.0099 (2012).
- 1103 96 Panksepp, J. & Panksepp, J. B. Toward a cross-species understanding of empathy. *Trends*
1104 *Neurosci* **36**, 489-496, doi:10.1016/j.tins.2013.04.009 (2013).

-
- 1105 97 Mattar, M. G. & Lengyel, M. Planning in the brain. *Neuron* **110**, 914-934,
1106 doi:10.1016/j.neuron.2021.12.018 (2022).
- 1107 98 Kaufmann, A. & Cahen, A. Temporal representation and reasoning in non-human animals.
1108 *Behav Brain Sci* **42**, e257, doi:10.1017/S0140525X19000487 (2019).
- 1109 99 Schwartz, B. L. & Beran, M. J. *Primate cognitive studies*. (Cambridge University Press,
1110 2022).
- 1111 100 Axer, M. & Amunts, K. Scale matters: The nested human connectome. *Science* **378**, 500-504,
1112 doi:10.1126/science.abq2599 (2022).
- 1113 101 Sudhof, T. C. Calcium control of neurotransmitter release. *Cold Spring Harb Perspect Biol* **4**,
1114 a011353, doi:10.1101/cshperspect.a011353 (2012).
- 1115 102 Hjejij, R. *et al.* Pathogenic variants in CLXN encoding the outer dynein arm docking-
1116 associated calcium-binding protein calaxin cause primary ciliary dyskinesia. *Genet Med* **25**,
1117 100798, doi:10.1016/j.gim.2023.100798 (2023).
- 1118 103 Ruhl, D. A. *et al.* Synaptotagmin 17 controls neurite outgrowth and synaptic physiology via
1119 distinct cellular pathways. *Nat Commun* **10**, 3532, doi:10.1038/s41467-019-11459-4 (2019).
- 1120 104 Xiao, R. *et al.* TMSB10 promotes migration and invasion of cancer cells and is a novel
1121 prognostic marker for renal cell carcinoma. *Int J Clin Exp Pathol* **12**, 305-312 (2019).
- 1122 105 Driver, A. M., Shumrick, C. & Stottmann, R. W. Ttc21b Is Required in Bergmann Glia for
1123 Proper Granule Cell Radial Migration. *J Dev Biol* **5**, doi:10.3390/jdb5040018 (2017).
- 1124 106 Osorio, D. S. & Gomes, E. R. Connecting the nucleus to the cytoskeleton for nuclear
1125 positioning and cell migration. *Adv Exp Med Biol* **773**, 505-520, doi:10.1007/978-1-4899-8032-
1126 8_23 (2014).
- 1127 107 Spruston, N. Pyramidal neurons: dendritic structure and synaptic integration. *Nat Rev*
1128 *Neurosci* **9**, 206-221, doi:10.1038/nrn2286 (2008).
- 1129 108 Allen, N. J. & Eroglu, C. Cell Biology of Astrocyte-Synapse Interactions. *Neuron* **96**, 697-
1130 708, doi:10.1016/j.neuron.2017.09.056 (2017).
- 1131 109 Won, H., Huang, J., Opland, C. K., Hartl, C. L. & Geschwind, D. H. Human evolved
1132 regulatory elements modulate genes involved in cortical expansion and neurodevelopmental
1133 disease susceptibility. *Nat Commun* **10**, 2396, doi:10.1038/s41467-019-10248-3 (2019).

-
- 1134 110 Bakewell, M. A., Shi, P. & Zhang, J. More genes underwent positive selection in chimpanzee
1135 evolution than in human evolution. *Proc Natl Acad Sci U S A* **104**, 7489-7494,
1136 doi:10.1073/pnas.0701705104 (2007).
- 1137 111 Arbiza, L., Dopazo, J. & Dopazo, H. Positive selection, relaxation, and acceleration in the
1138 evolution of the human and chimp genome. *PLoS Comput Biol* **2**, e38,
1139 doi:10.1371/journal.pcbi.0020038 (2006).
- 1140 112 Jorstad, N. L. *et al.* Comparative transcriptomics reveals human-specific cortical features.
1141 *Science* **382**, eade9516, doi:10.1126/science.ade9516 (2023).
- 1142 113 Ma, S. *et al.* Molecular and cellular evolution of the primate dorsolateral prefrontal cortex.
1143 *Science* **377**, eabo7257, doi:10.1126/science.abo7257 (2022).
- 1144 114 Maier-Hein, K. H. *et al.* The challenge of mapping the human connectome based on diffusion
1145 tractography. *Nat Commun* **8**, 1349, doi:10.1038/s41467-017-01285-x (2017).
- 1146 115 Chauvel, M. *et al.* In vivo mapping of the deep and superficial white matter connectivity in
1147 the chimpanzee brain. *Neuroimage* **282**, 120362, doi:10.1016/j.neuroimage.2023.120362 (2023).
- 1148 116 Howard, A. F. D. *et al.* An open resource combining multi-contrast MRI and microscopy in
1149 the macaque brain. *Nat Commun* **14**, 4320, doi:10.1038/s41467-023-39916-1 (2023).
- 1150 117 Blostein, N. Variation in subcortical anatomy: relating interspecies differences, heritability,
1151 and brain-behavior relationships. *bioRxiv* (2022).
- 1152 118 Parks, A. N., Bryant, K. L., Rankin, E. K., Smaers, J. B. & Hecht, E. E. Segmentation and
1153 morphometric MRI atlas of the chimpanzee cerebellum. *bioRxiv* (2022).
- 1154 119 Amiez, C. *et al.* A revised perspective on the evolution of the lateral frontal cortex in
1155 primates. *Sci Adv* **9**, eadf9445, doi:10.1126/sciadv.adf9445 (2023).
- 1156 120 Amiez, C. *et al.* Sulcal organization in the medial frontal cortex provides insights into
1157 primate brain evolution. *Nat Commun* **10**, 3437, doi:10.1038/s41467-019-11347-x (2019).
- 1158 121 Van Essen, D. C. *et al.* The WU-Minn Human Connectome Project: an overview.
1159 *Neuroimage* **80**, 62-79, doi:10.1016/j.neuroimage.2013.05.041 (2013).
- 1160 122 Glasser, M. F. *et al.* The minimal preprocessing pipelines for the Human Connectome
1161 Project. *Neuroimage* **80**, 105-124, doi:10.1016/j.neuroimage.2013.04.127 (2013).
- 1162 123 Chen, X. *et al.* Brain aging in humans, chimpanzees (*Pan troglodytes*), and rhesus macaques

-
- 1163 (Macaca mulatta): magnetic resonance imaging studies of macro- and microstructural changes.
1164 *Neurobiol Aging* **34**, 2248-2260, doi:10.1016/j.neurobiolaging.2013.03.028 (2013).
- 1165 124 Donahue, C. J. *et al.* Using Diffusion Tractography to Predict Cortical Connection Strength
1166 and Distance: A Quantitative Comparison with Tracers in the Monkey. *J Neurosci* **36**, 6758-6770,
1167 doi:10.1523/JNEUROSCI.0493-16.2016 (2016).
- 1168 125 Fischl, B. FreeSurfer. *Neuroimage* **62**, 774-781, doi:10.1016/j.neuroimage.2012.01.021
1169 (2012).
- 1170 126 Jenkinson, M., Beckmann, C. F., Behrens, T. E., Woolrich, M. W. & Smith, S. M. Fsl.
1171 *Neuroimage* **62**, 782-790, doi:10.1016/j.neuroimage.2011.09.015 (2012).
- 1172 127 Klein, A. & Tourville, J. 101 labeled brain images and a consistent human cortical labeling
1173 protocol. *Front Neurosci* **6**, 171, doi:10.3389/fnins.2012.00171 (2012).
- 1174 128 Behrens, T. E., Berg, H. J., Jbabdi, S., Rushworth, M. F. & Woolrich, M. W. Probabilistic
1175 diffusion tractography with multiple fibre orientations: What can we gain? *Neuroimage* **34**, 144-
1176 155, doi:10.1016/j.neuroimage.2006.09.018 (2007).
- 1177 129 Hernandez-Fernandez, M. *et al.* Using GPUs to accelerate computational diffusion MRI:
1178 From microstructure estimation to tractography and connectomes. *Neuroimage* **188**, 598-615,
1179 doi:10.1016/j.neuroimage.2018.12.015 (2019).
- 1180 130 Munkres, J. J. J. o. t. s. f. i. & mathematics, a. Algorithms for the assignment and
1181 transportation problems. *Journal of the society for industrial and applied mathematics* **5**, 32-38
1182 (1957).
- 1183 131 Li, H. *et al.* ATPP: A Pipeline for Automatic Tractography-Based Brain Parcellation. *Front*
1184 *Neuroinform* **11**, 35, doi:10.3389/fninf.2017.00035 (2017).
- 1185 132 Armstrong, R. A. When to use the Bonferroni correction. *Ophthalmic Physiol Opt* **34**, 502-
1186 508, doi:10.1111/opo.12131 (2014).
- 1187 133 Burt, J. B., Helmer, M., Shinn, M., Anticevic, A. & Murray, J. D. Generative modeling of
1188 brain maps with spatial autocorrelation. *Neuroimage* **220**, 117038,
1189 doi:10.1016/j.neuroimage.2020.117038 (2020).
- 1190 134 Karolis, V. R., Corbetta, M. & Thiebaut de Schotten, M. The architecture of functional
1191 lateralisation and its relationship to callosal connectivity in the human brain. *Nat Commun* **10**,

1192 1417, doi:10.1038/s41467-019-09344-1 (2019).

1193 135 Markello, R. D. *et al.* Standardizing workflows in imaging transcriptomics with the abagen
1194 toolbox. *Elife* **10**, doi:10.7554/eLife.72129 (2021).

1195 136 Arnatkeviciute, A., Fulcher, B. D. & Fornito, A. A practical guide to linking brain-wide gene
1196 expression and neuroimaging data. *Neuroimage* **189**, 353-367,
1197 doi:10.1016/j.neuroimage.2019.01.011 (2019).

1198 137 Morgan, S. E. *et al.* Cortical patterning of abnormal morphometric similarity in psychosis is
1199 associated with brain expression of schizophrenia-related genes. *Proc Natl Acad Sci U S A* **116**,
1200 9604-9609, doi:10.1073/pnas.1820754116 (2019).

1201 138 Chen, J., Bardes, E. E., Aronow, B. J. & Jegga, A. G. ToppGene Suite for gene list
1202 enrichment analysis and candidate gene prioritization. *Nucleic Acids Res* **37**, W305-311,
1203 doi:10.1093/nar/gkp427 (2009).

1204 139 Li, P., Wei, J. & Zhu, Y. CellGO: a novel deep learning-based framework and webserver for
1205 cell-type-specific gene function interpretation. *Brief Bioinform* **25**, doi:10.1093/bib/bbad417
1206 (2023).

1207 140 Durinck, S. *et al.* BioMart and Bioconductor: a powerful link between biological databases
1208 and microarray data analysis. *Bioinformatics* **21**, 3439-3440, doi:10.1093/bioinformatics/bti525
1209 (2005).

1210 141 Johnson, W. E., Li, C. & Rabinovic, A. Adjusting batch effects in microarray expression data
1211 using empirical Bayes methods. *Biostatistics* **8**, 118-127, doi:10.1093/biostatistics/kxj037 (2007).

1212 142 Anders, S. & Huber, W. Differential expression analysis for sequence count data. *Genome*
1213 *Biol* **11**, R106, doi:10.1186/gb-2010-11-10-r106 (2010).

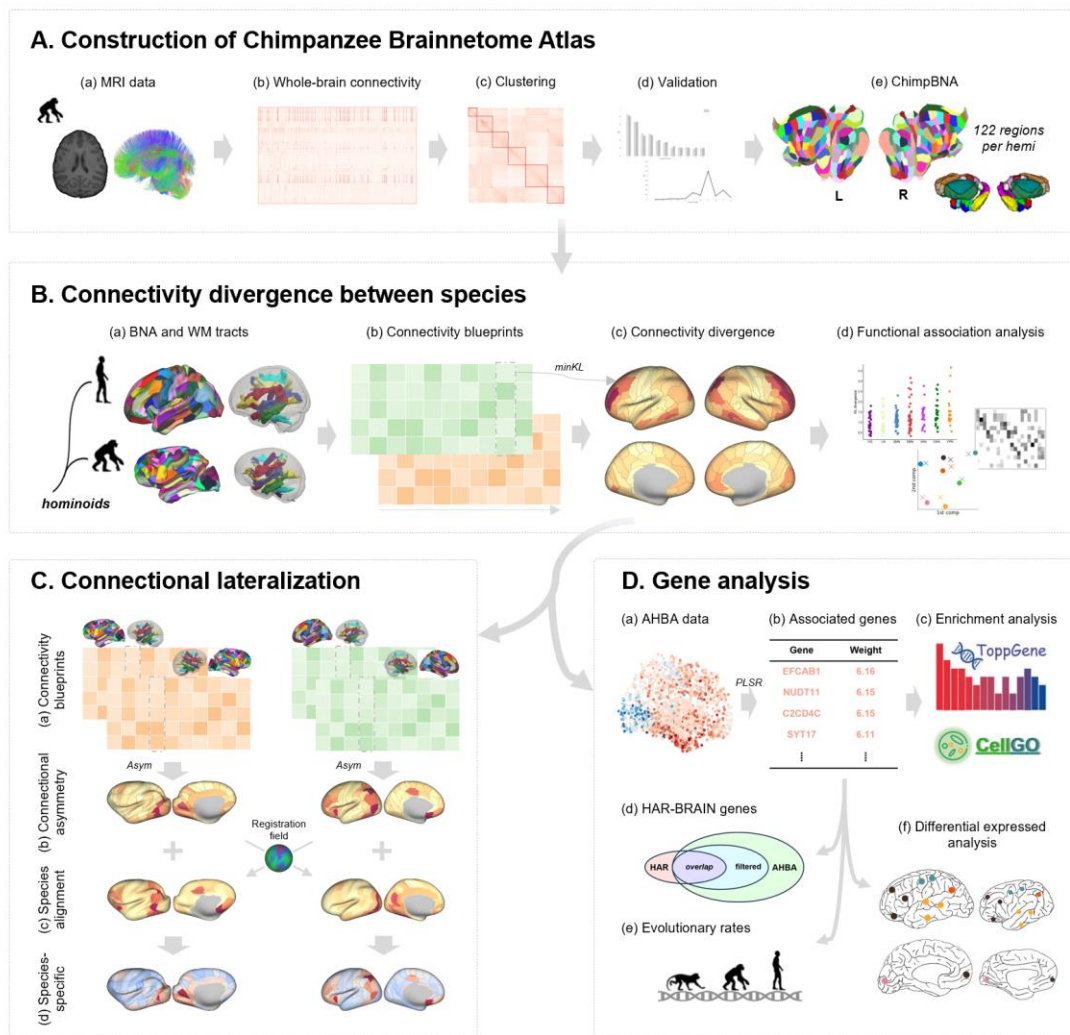
1214 143 Zhang, Y. *et al.* Purification and Characterization of Progenitor and Mature Human
1215 Astrocytes Reveals Transcriptional and Functional Differences with Mouse. *Neuron* **89**, 37-53,
1216 doi:10.1016/j.neuron.2015.11.013 (2016).

1217

1218

1219

Figures

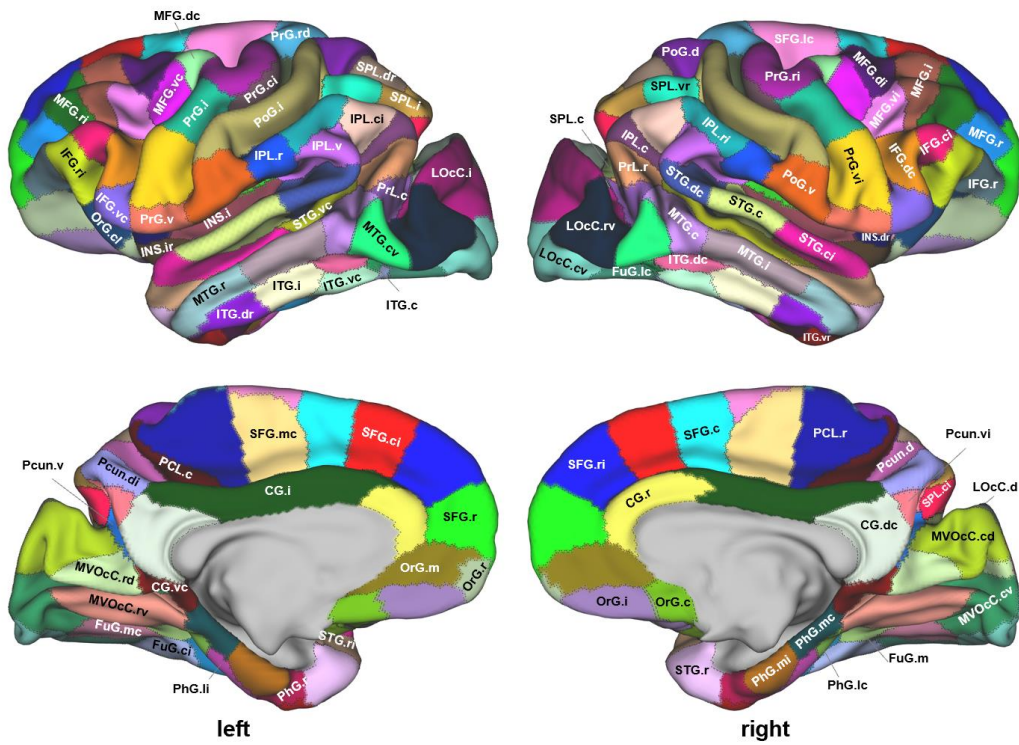


1220

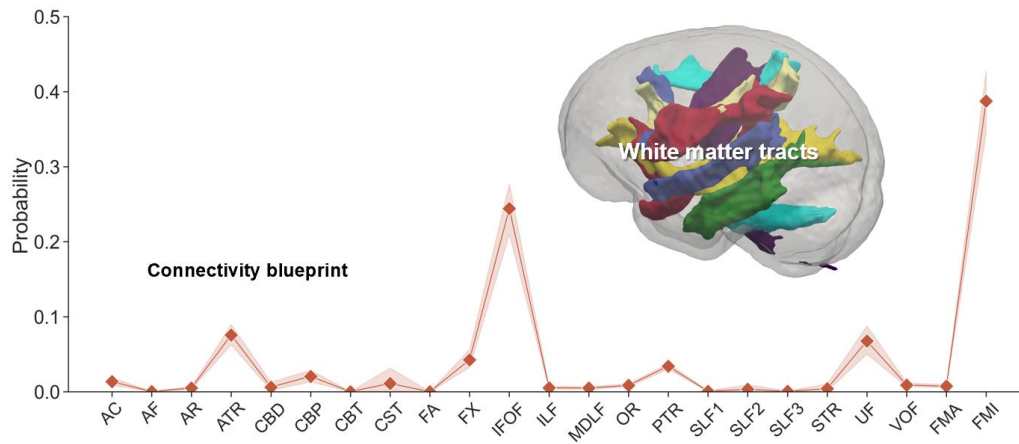
1221 **Figure 1. Analysis pipeline.** (A) Following a connectivity-based parcellation procedure, we used
 1222 MRI data from chimpanzee brains (a) to construct the Chimpanzee Brainnetome Atlas.
 1223 Tractography and similarity matrices were performed (b) for subsequent spectral clustering (c). The
 1224 clustering results were validated using several indices (d), and the final parcellation of the whole
 1225 brain was obtained (e). (B) We utilized the Brainnetome Atlas of humans and chimpanzees and
 1226 homologous white matter tracts (a), to build regional connectivity blueprints for each species (b).
 1227 The blueprints were used to explore the connectivity divergence between humans and chimpanzees
 1228 (c), followed by functional association analysis of this divergence (d). (C) We used the connectivity
 1229 blueprints from two hemispheres for each species (a) to investigate the asymmetric connectivity
 1230 pattern (b). Myelin-based registration was used to align the two species into a common space (c).

1231 Thus the species-specific asymmetric connectivity pattern could be calculated (d). **(D)** AHBA data
1232 (a) was used to identify the genes associated with the connectivity divergence by PLSR (b), the
1233 filtered genes were input to gene enrichment analysis and cell-type enrichment analysis (c), as well
1234 as evolutionary investigation, including overlap with HAR-BRAIN genes (d), evolutionary rates (e),
1235 and differentially expressed analysis between the two species (f).
1236

A. Cortical regions of ChimpBNA



B. Region-to-tract connection



1237

1238 **Figure 2. The Chimpanzee Brainnetome Atlas and connections of the chimpanzee brain. (A)**

1239 Cortical regions of the Chimpanzee Brainnetome Atlas. 100 cortical subregions were identified per

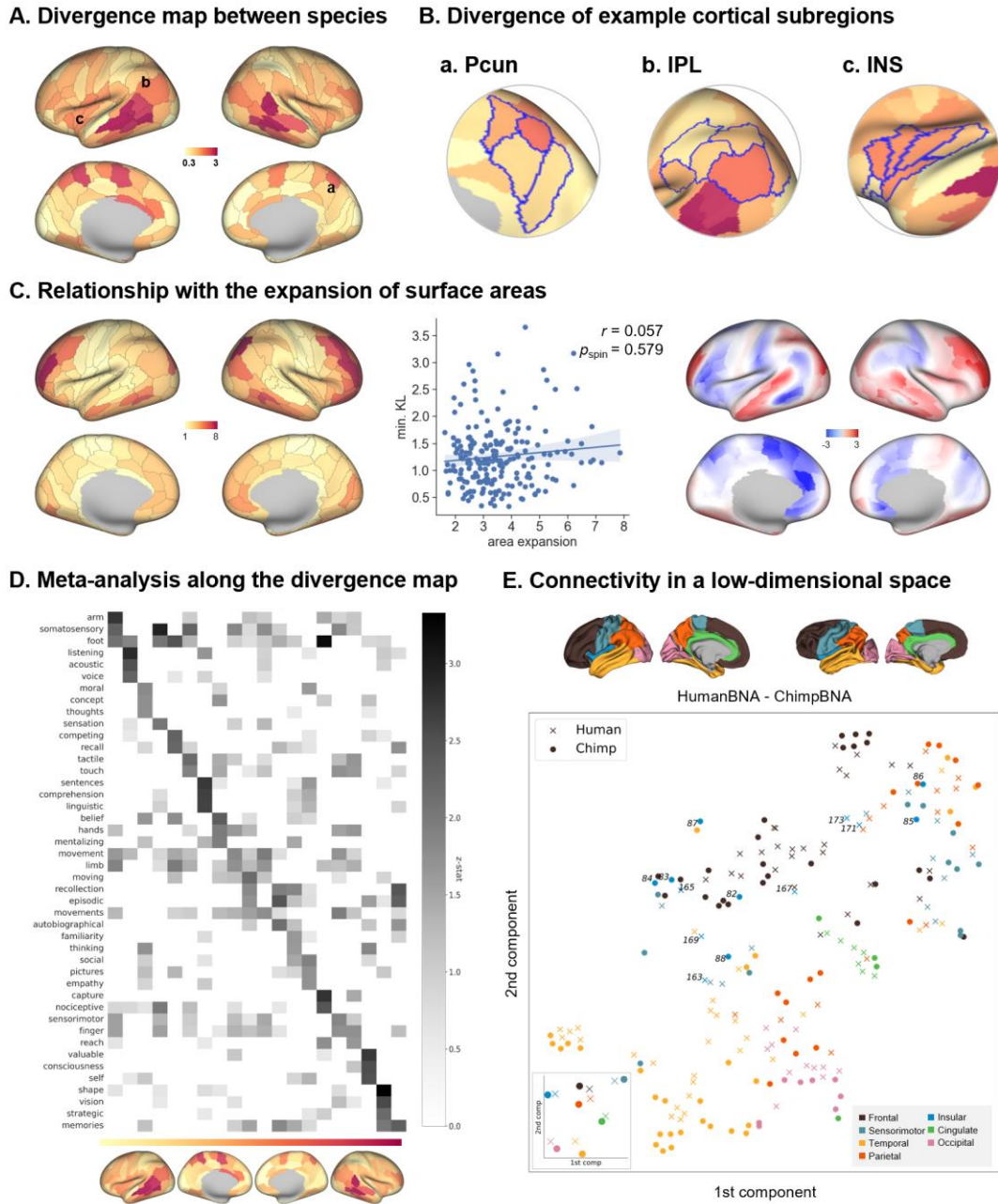
1240 hemisphere using anatomical connectivity profiles. **(B)** Region-to-tract connections of the

1241 chimpanzee brain. White matter tracts were reconstructed following protocols in the previous study

1242 ⁴⁴, and the connectivity blueprint of an exemplar region, SFG.r, is shown. SFG.r, superior frontal

1243 gyrus, rostral part.

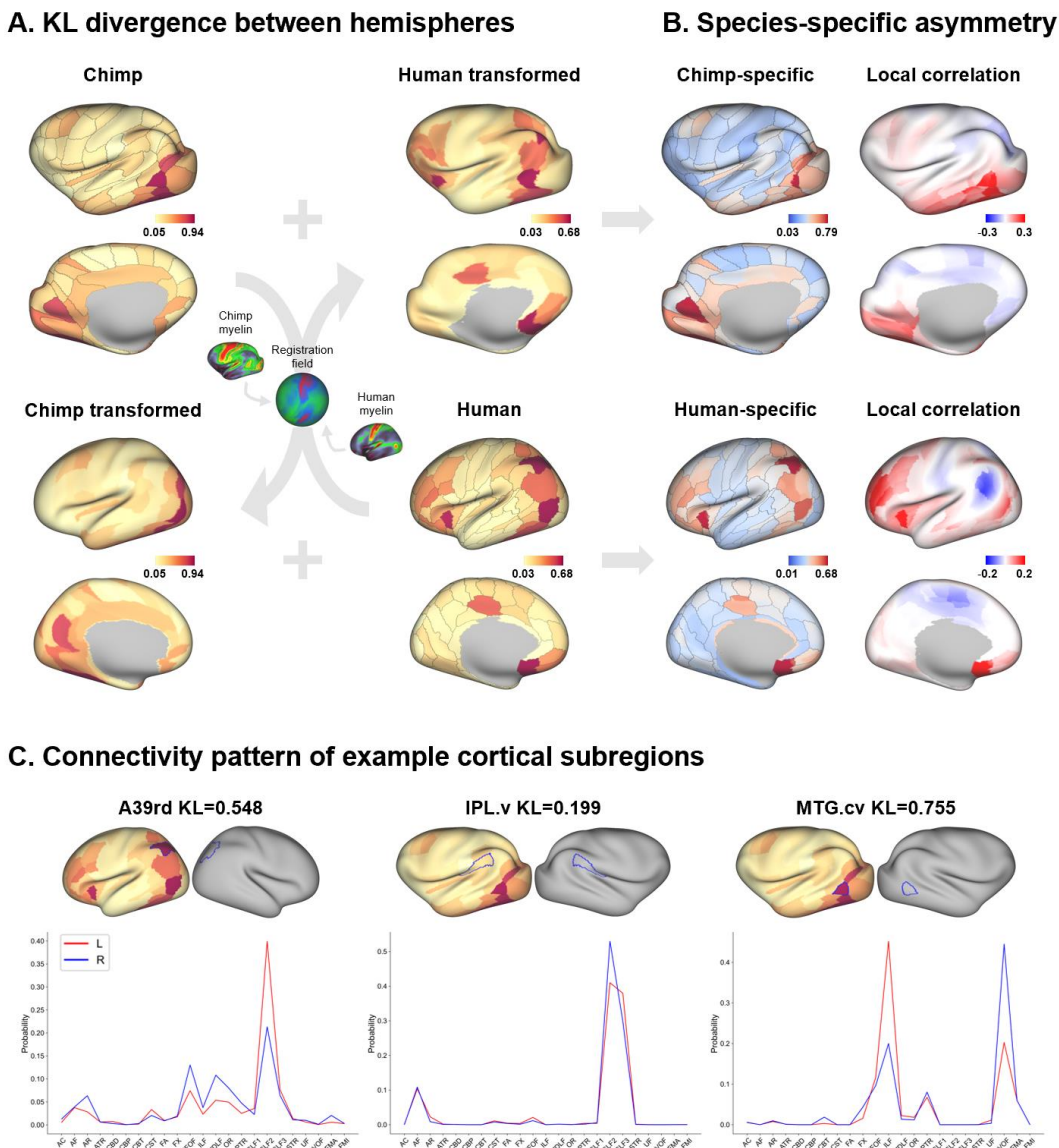
1244



1245

1246 **Figure 3. Connectivity divergence between species.** (A) Connectivity blueprints were used to
 1247 calculate the KL divergence between species to determine the extent of dissimilarity between their
 1248 connectivity profiles. Higher values in the divergence map indicated that the region in humans had
 1249 a connectivity pattern that was more dissimilar to the regions in chimpanzees. (B) Connectivity
 1250 divergence of several example ROIs, i.e., Pccn, IPL, and INS, were investigated at the subregion
 1251 level. (C) The connectivity divergence map showed a very low correlation with the map of cortical
 1252 expansion between chimpanzees and humans¹³. The right panel showed the weighted local

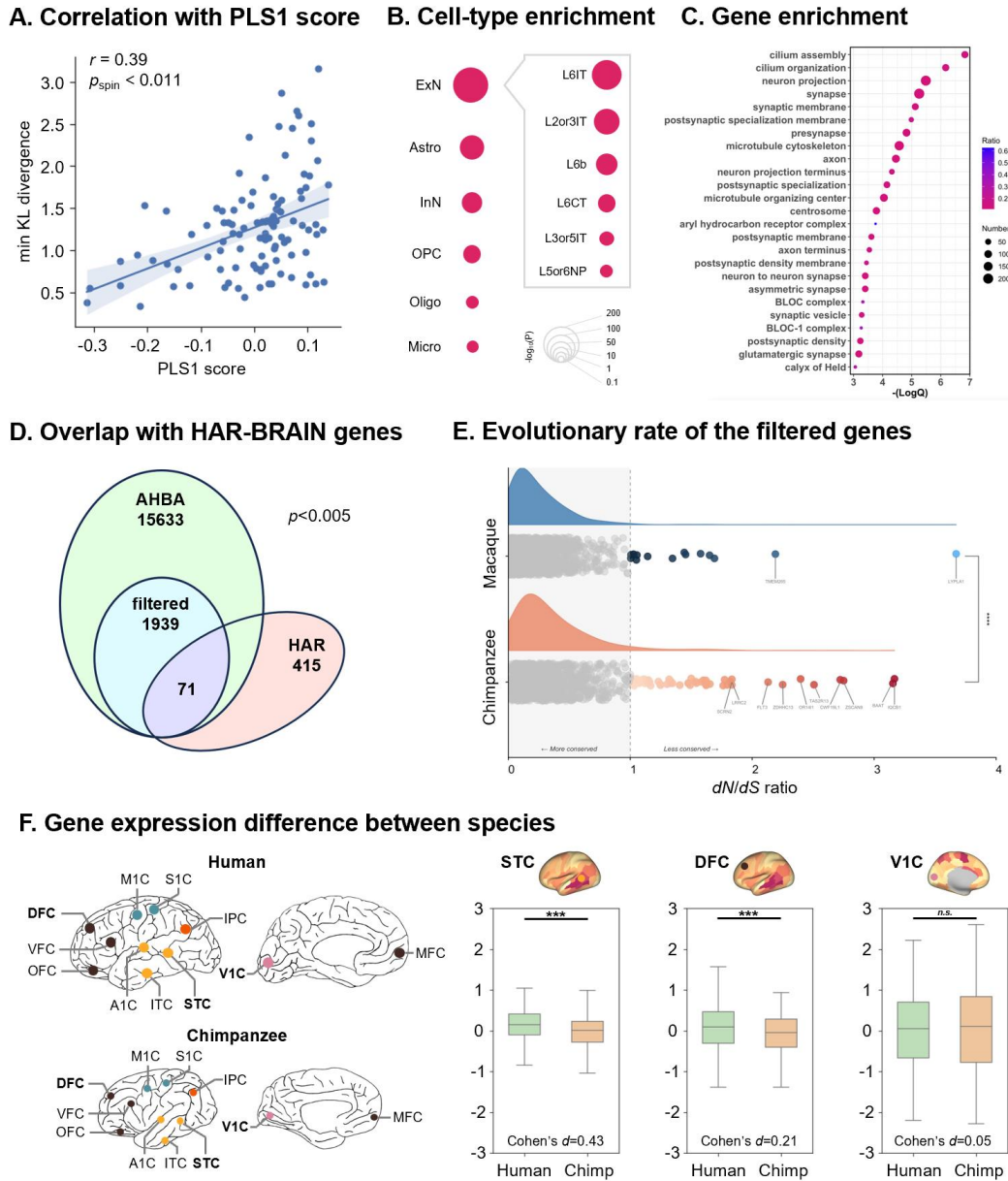
1253 correlation between the area expansion map and the connectivity divergence map. Regions with
1254 high correlation coefficients indicate marked both cortical expansion and connectivity differences
1255 between chimpanzees and humans. **(D)** The divergence map was input for functional decoding.
1256 NeuroSynth terms with the highest three z-scores for each binarized mask of the divergence map
1257 were visualized. **(E)** Subregions of chimpanzees and human left brains were projected into a low-
1258 dimensional space using their connectivity profile, with each color representing a cortical system.
1259 The figure inset indicates the center of each system. Pcun, precuneus; IPL, inferior parietal lobule;
1260 INS, insular cortex.
1261



1262

1263 **Figure 4. Species-specific whole-brain level connective lateralization.** (A) Connectivity
 1264 blueprints were used to calculate the KL divergence between homotopic subregions between
 1265 hemispheres. The divergence map of the connective lateralization of the chimpanzees and humans
 1266 was aligned and compared. (B) Species-specific asymmetric connectivity patterns were calculated,
 1267 and the weighted local correlation with the connectivity divergence map is shown, where a higher
 1268 value indicates both marked connectivity differences between species and asymmetry between
 1269 hemispheres. (C) Connection probability of tracts of several example ROIs. In A39rd in humans,
 1270 the inter-hemisphere differences were driven by IFOF, MdLF, and SLF2, while in IPL.v in
 1271 chimpanzees, the asymmetry was mainly driven by SLF2. MTG.cv in the chimpanzee brain, in

1272 which the KL divergence was greater, showed differences in their tract connections, mostly driven
1273 separately by the ILF and VOF. A39rd, rostradorsal area 39; IPL.v, inferior parietal lobule, ventral
1274 part; MTG.cv, middle temporal gyrus, caudoventral part; IFOF, inferior fronto-occipital fasciculus;
1275 MdLF, middle longitudinal fasciculus; SLF2, superior longitudinal fascicle II; ILF, inferior
1276 longitudinal fasciculus; VOF, vertical occipital fasciculus.
1277



1278

1279 **Figure 5. Gene association with connectivity divergence between species. (A)** The divergence
 1280 map shows a significant correlation with the PLS1 score of genes with the AHBA dataset (left brain).
 1281 **(B)** Cell-type enrichment analysis of genes identified using bootstrapping with the most positive or
 1282 negative weights ($|Z| > 3$) in PLSR. **(C)** These genes were used in an enrichment analysis and found
 1283 to be associated with neuronal projection and synapse formation processes. **(D)** 71 genes overlapped
 1284 with the HAR-BRAIN genes ($p < .005$). **(E)** 56 genes had a dN/dS ratio > 1 (greater than 1 means
 1285 less conserved in chimpanzees), but only 14 genes in the macaque against the human genome
 1286 (Welch's t-test $p < .0001$). **(F)** Differences in these filtered genes were compared using human and

1287 chimpanzee data from the PsychENCODE database. 1473 of 1939 genes that overlapped in the
1288 database were used for analysis. Three regions of interest exhibiting distinct connectivity divergence
1289 were considered, and a paired t-test was utilized to assess differences between the two species.
1290 Significant differences were found in first two regions, and the effect size was more significant in
1291 the STC and DFC than in VIC. STC, superior temporal cortex; DFC, dorsolateral frontal cortex;
1292 VIC, primary visual cortex. *** indicates $p < .001$.