The Clinical Translation of α-humulene – A Scoping Review

OPEN ACCESS

Authors
Nishaanth Dalavaye[1]*, Martha Nicholas[1]*, Manaswini Pillai[1]*, Simon Erridge[1,2], Mikael H. Sodergren[1,3]

Affiliations
1 Medical Cannabis Research Group, Department of Surgery and Cancer, Imperial College London, UK
2 Curaleaf Clinic, London, UK
3 Curaleaf International, London, UK

Correspondence
Mr. Mikael H. Sodergren FRCS PhD
Division of Surgery, Department of Surgery and Cancer,
Imperial College London, Academic Surgical Unit, 10th Floor
QEQM, St. Mary's Hospital
South Wharf Road, W2 1NY London, United Kingdom
Phone: + 44 20 33 12 66 66, Fax: + 44 20 33 12 63 09
m.sodergren@imperial.ac.uk

ABSTRACT

α-humulene, a sesquiterpene found in essential oils of various plant species, has garnered interest due to its potential therapeutic applications. This scoping review aims to consolidate α-humulene's evidence base, informing clinical translation, and guiding future research directions. A scoping review was conducted of EMBASE, MEDLINE, and PubMed databases up to 14th July 2023. All studies describing original research on α-humulene extraction, as well as pre-clinical and clinical research, were included for review. Three hundred and forty articles were analysed. α-humulene yields ranged from negligible to 60.90% across plant species. *In vitro* experiments demonstrated cytotoxicity against adenocarcinomas (such as colorectal, pulmonary, breast, prostatic, lung, and ovarian), with varying responses in other cell models. Mechanistic insights revealed its involvement in mitochondrial dysfunction, diminished intracellular glutathione levels, and the induction of oxidative stress. In rodent studies, oral administration of α-humulene at 50 mg/kg reduced inflammation markers in paw oedema and ovalbumin-induced airway inflammation. Intraperitoneal administration of α-humulene (50–200 mg/kg) exhibited cannabimimetic properties through cannabinoid 1 and adenosine A2a receptors. α-humulene also exhibited a multitude of properties with potential scope for therapeutic utilisation. However, there is a paucity of studies that have successfully translated this research into clinical populations with the associated disease. Potential barriers to clinical translation were identified, including yield variability, limited isolation studies, and challenges associated with terpene bioavailability. Consequently, rigorous pharmacokinetic studies and further mechanistic investigations are warranted to effectively uncover the potential of α-humulene.

Introduction

Terpenoids are a vast and diverse group encompassing several classes of secondary metabolites from plants, each being investigated for biomedical properties [1]. Notably, they have been described as having anti-inflammatory, analgesic, antimicrobial, antioxidant, and estrogenic properties [2, 3]. Sesquiterpenes are a class of terpene to which α-humulene (also known as α-caryophyllene) and its isomer β-caryophyllene belong. These sesquiterpenes share a three-isoprene unit structure, formed from the precursor farnesyl diphosphate, leading to the formation of cyclic or multi-ring compounds that contribute to their distinctive aroma [1].

* Nishaanth Dalavaye, Martha Nicholas, and Manaswini Pillai are to be considered as joint first authors
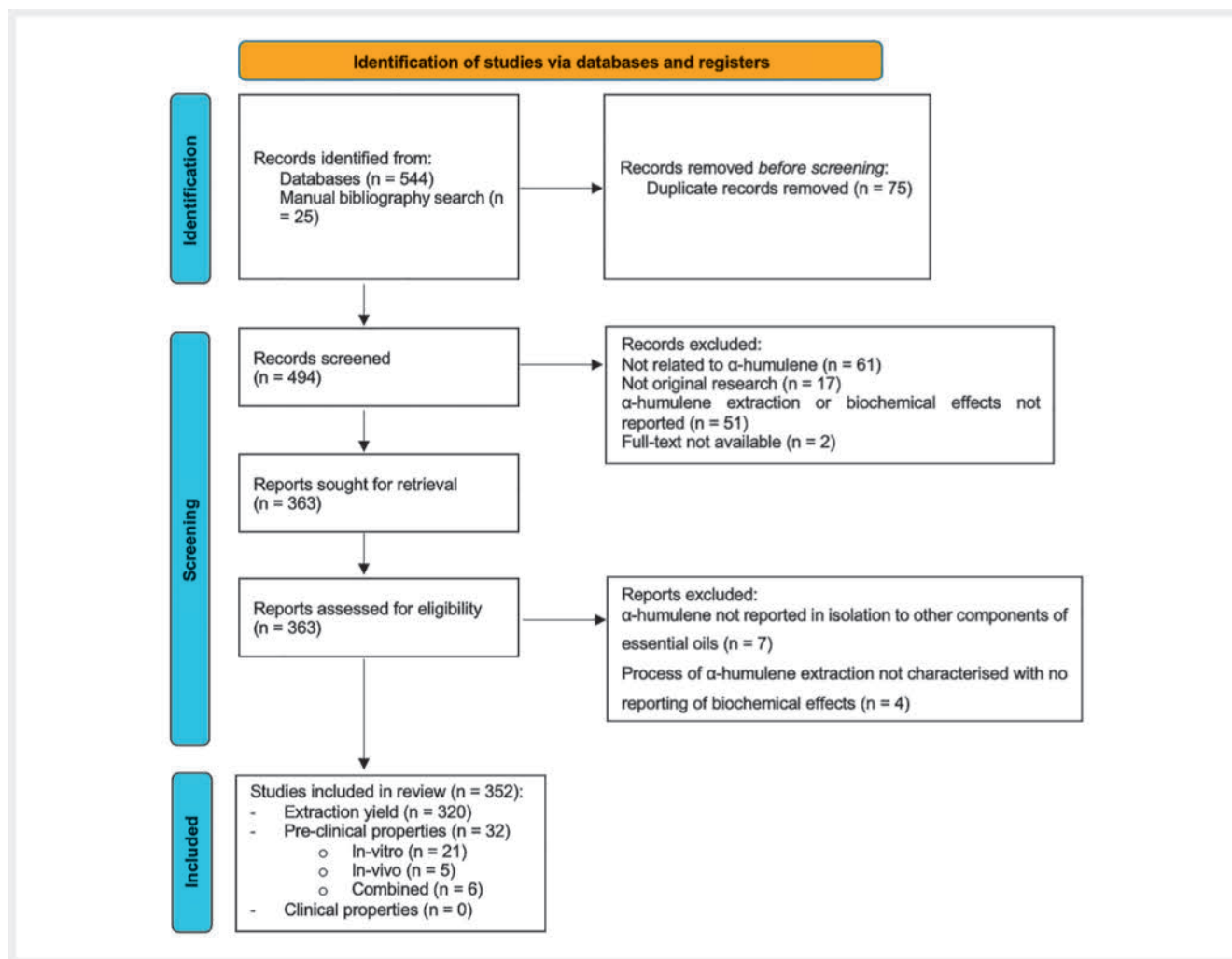
▶ Fig. 1 PRISMA flow chart showing the process of inclusion and exclusion of patients for analysis in this scoping review.

α-humulene and β-caryophyllene, though structurally similar, are differentiated by the open-ring structure present in α-humulene [4]. Historically, α-humulene was first identified as a major component in the essential oils of *Humulus lupulus* L., Cannabaceae, the common hop plant, from which it derived its name. Its structural elucidation was achieved through nuclear magnetic resonance spectroscopy. Furthermore, α-humulene has not only been sourced from *Humulus lupulus* but also from *Cannabis Indica* L., Cannabaceae, signifying its prevalence in botanical species for which there are already well-established agricultural processes [5]. The isolation and extraction of α-humulene from its botanical sources have been refined over time. Modern techniques, such as steam distillation, are employed to capture the volatile essential oils containing this sesquiterpene. Its abundance in various botanical sources makes it a subject of interest for both traditional and modern medicinal applications.

There is continual demand to identify novel compounds that possess anticancer, anti-inflammatory, and antimicrobial properties, in light of a persisting cancer burden, rising incidence of inflammatory conditions, and emergence of antimicrobial resis-

tance [6–8]. Despite promising preclinical evidence supporting the multimodal therapeutic potential of **α**-humulene, there are several barriers to its clinical translation. Whilst certain plant species are known to be rich sources of **α**-humulene, plant yields are often reported as being low, and so far, researchers have not reached a consensus on a named plant species that consistently yields high concentrations of α-humulene. At present, biosynthesis pathways are therefore being explored as an avenue to create synthetic α-humulene to overcome inherent challenges with the manufacturing of compounds that are reliant upon favourable agricultural conditions [9]. In addition, much of the research conducted to date has utilised a combination of organic compounds contained within the plant essential oil, rather than assessing the properties of α-humulene in isolation, resulting in a paucity of evidence summarising α-humulene's individual properties. It is therefore important to evaluate studies that report specifically the properties of α-humulene and identify species with acceptable yields in order to advance this scientific field.

A systematic review by Leite et al. [10] summarised the preclinical properties of sesquiterpene compounds, including α-humu-

▶ **Table 1** Summarising the species with the top five highest reported yields of α-humulene.

Species	Chemovar	Extraction	Isolation	Yield	Reference
Aframomum melegueta [alligator pepper]	Nigeria	Hydrodistillation for 3 h	Fractionation	60.90%	Ajaiyeoba E et al. 1999 [77]
Leptospermum sp. [Mt Maroon A. R. Bean 6665]	Australia	Hydrodistillation with incubation	GC-MS	44.00–51.00%	Brophy J et al. 2000 [78]
Humulus lupulus. [Chinook variety]	Brazil	Hydrodistillation using a Clevenger-type apparatus	GC-MS	31.50–34.62%	Duarte et al. 2023 [79]
Camponotus japonicus [insect]	Japan	Macerated in 10 mL of pentane	GC-MS	35.80%	Sakurai K et al. 2020 [80]
Zingiber nimmonii	India	Hydrodistillation using a Clevenger-type apparatus for 8 h	GC-MS	19.60%	Govindarajan et al. 2016 [81]
GC-MS – gas chromatography–mass spectrometry					

lene and β-caryophyllene. Whilst preclinical evidence has been promising regarding the properties of **α**-humulene, there has been minimal progress into clinical translation of this research. Hence, this review aimed to provide a synthesised evidence base for the prioritisation of future research, including optimisation of agriculture and manufacturing, alongside identification of the most promising biomedical applications.

Results

Search results

The database and manual bibliography search initially returned 544 studies (▶ **Fig. 1**). Four hundred and twelve full-text articles were assessed for eligibility, with 340 articles included for qualitative synthesis. Three hundred and seven studies included reported the extraction yields of α-humulene (Supplementary Material A). Thirty-two studies [11–42] were included for evaluation of the pre-clinical properties of α-humulene. These included investigations conducted *in vitro* [11–30], *in vivo* [31–36], and combined *in vitro* and *in vivo* experiments [37–42]. Notably, no studies were found to assess the clinical properties of α-humulene.

Yield of α-humulene from extraction

Yields of α-humulene were reported from 462 different plant and animal species (Supplementary Material B). Reported yields varied from nil to 60.90%. ▶ **Table 1** highlights the five species that exhibited the highest reported yields among the included studies. The most common method of α-humulene extraction was hydrodistillation in a Clevenger-type apparatus. Concurrently, isolation most frequently relied on gas chromatography–mass spectrometry (GC-MS). Among the species analysed, *Lantana camara* L., Verbenaceae; *Origanum majorana* L., Lamiaceae; *Cordia verbenacea* DC., Boraginaceae; *Cannabis sativa;* and *Daucus carota* L., Apiaceae were prominent contributors, with the greatest number of studies reporting α-humulene extraction data.

Specific properties of α-humulene
Antiproliferative properties

Thirteen studies evaluated the effects of α-humulene in cancer models (▶ **Table 2**) [11–20, 31, 37, 38]. Across these studies, α-humulene consistently demonstrated cytotoxic activity against tumour cells, with one exception by Loizzo et al. (2008) [20] involving human amelanotic melanoma (C32) and renal cell adenocarcinoma (ACHN) at a concentration of 9.3×10^{-7} to 1.2×10^{-4}. α-humulene, sourced from *Myrica rubra* Siebold & Zucc., Myricaceae, has demonstrated substantial anti-proliferative effects on colorectal cancer cell lines *in vitro*, marked by mitochondrial membrane potential disruption and enhanced efficacy when combined with conventional anticancer drugs [11]. In hepatocellular carcinoma (HCC), α-humulene from *Eupatorium odoratum* L., Asteraceae exhibited selective inhibition of HCC cell proliferation primarily *in vitro*, associated with the suppression of protein kinase B signalling [37]. Notably, α-humulene demonstrated dose-dependent inhibition of ovarian and lymphoblast cancer cell proliferation *in vitro* and synergistic effects with doxorubicin [12]. Its preferential cytotoxicity towards tumour cells, sparing non-tumour cells, indicates potential selectivity for actively dividing cancer cells. This anti-proliferative activity has also been related to apoptosis induction and modulation of reactive oxygen species (ROS) production [37]. In an *in vivo* study on clove terpenes, α-humulene induced significant glutathione S-transferase activity in mouse liver and small intestine tissues, suggesting a role in detoxification processes [31]. Fukuoka et al. (2004) [21] showed α-humulene's anti-proliferative properties in rat arterial smooth muscle cells, utilising a cell assay that induced proliferation with heat shock protein. The study reported an IC50 value of 0.122 μM, showcasing dose-dependent effects and superior potency compared to its analogue zerumbone. Inhibitory effects were demonstrated even at a concentration of 4.89×10^{-6} mol/L.

▶ **Table 2** Summary of studies investigating the anticancer properties of isolated α-humulene.

Model	Concentration/Dose	Results	Reference
In vitro colorectal adenocarcinoma epithelial cells [CaCo-2 and SW-620]	5×10^{-5}, 1×10^{-4}, and 1.5×10^{-4} M	α-humulene exhibited antiproliferative activity in combination with oxaliplatin and 5-fluorouracil at 100 and 150 µmol/L due to decreased mitochondrial membrane potenzial.	Ambroz et al. 2019 [11]
In vitro hepatocellular carcinoma cells [huh7, SMMC-7721, HepG2 and Hep3B] and *in vivo* HepG2-bearing nude mice	*In vitro*: 6.1×10^{-6} – 2.4×10^{-4} M *In vivo*: 10 and 20 mg/kg	*In vitro*, α-humulene inhibited proliferation of all hepatocellular carcinoma cell lines at 15 µmol/L, inducing cytotoxicity via intrinsic apoptotic pathways. Similar findings were reported *in vivo* at 10 mg/kg.	Chen et al. 2019 [37]
In vitro ovarian cancer cells [A2780 and SKOV3 and lymphoblasts CCRF/CEM and CEM/ADR]	20, 40, 100, and 200 µM	α-humulene showed antiproliferative activity against certain ovarian cancer cell lines [A2780 at 40 µM, SKOV3 at 200 µM] and lymphoblast cell lines [CCRF/CEM at 200 µM, no effect on CEM/ADR at 200 µM].	Ambroz et al. 2017 [12]
In vitro colon adenocarcinoma [CaCo-2] and non-cancer cells [rat hepatocytes]	$0–2.4 \times 10^{-4}$ M	α-humulene demonstrated antiproliferative activity against cancer cells at 4.9×10^{-5} mol/L, with an IC50 of 24.4 ± 2.4. Additionally, α-humulene potentiated doxorubicin's anticancer properties in cancer cells, while showing no effect on non-cancer cell viability.	Ambroz et al. 2015 [13]
In vitro mice melanoma, human hepatocellular carcinoma, chronic human myelocytic leukaemia, and human promyelocytic leukaemia	9.3×10^{-7} – 1.2×10^{-4} M	No significant anticancer activity of α-humulene was identified for any concentration tested.	Costa et al. 2015 [38]
In vitro: colon human cancer [HT-29], human hepatocellular carcinoma [J5], and human pulmonary adenocarcinoma [A549]	$0–9.8 \times 10^{-4}$ M	α-humulene exhibited significant cytotoxicity against all cell lines, with IC50 values of 5.2×10^{-5}, 1.8×10^{-4}, and 1.3×10^{-4} mol/L for HT-29, J5, and A549, respectively.	Su et al. 2015 [14]
In vitro: human colorectal. adenocarcinoma [HCT-116], human breast cancer [MCF-7] and murine macrophages [RAW264.7]	7.6×10^{-6} – 4.9×10^{-4} M	α-humulene demonstrated cytotoxic potenzial by inhibiting cancer cell growth, with IC50 values of 3.1×10^{-4}, 4.2×10^{-4}, and 1.9×10^{-4} mol/L for HCT-116, MCF-7, and RAW264.7 cell lines, respectively.	Hadri et al. 2010 [15]
Murine small bowel mucosa and liver	9.8×10^{-5} M	α-humulene showed potenzial inhibitory action against carcinogenesis by increasing glutathione S-transferase [GST] activity. The enzyme activity increased by 99% in the liver and 152% in the small bowel.	Zheng et al. 1992 [31]
Cell lines of human breast adenocarcinoma [MCF-7], prostatic adenocarcinoma [PC-3], lung carcinoma [A-549], colon adenocarcinoma and fibroblasts [DLD-1 e L-929]	2.4×10^{-4} and 9.8×10^{-4} M	α-humulene caused dose-dependent glutathione depletion of 38% and 71% at 50 and 200 µM, respectively, along with increased production of reactive oxygen species by 163% and 278% after 1 and 4 hours. Normal human fibroblasts showed lower cytotoxic effects.	Legault et al. 2003 [16]
Cell lines of human breast adenocarcinoma [MCF-7], colon adenocarcinoma [DLD-1: ATCC # CCL-221], murine fibroblasts [L-929 ATCC # CCL-1]	7.8×10^{-5} – 3.1×10^{-4} M	α-humulene showed cytotoxicity at 1.6×10^{-4} and 3.1×10^{-4} mol/L. Cell growth inhibition by α-humulene was significantly increased from 50 ± 6% alone to 75 ± 6% by co-administration of non-cytotoxic levels [10 µg/mL] of caryophyllene, potenzially due to altered membrane permeability.	Legault et al. 2010 [17]

▶ **Table 2** *Continued*

Model	Concentration/Dose	Results	Reference
Cell lines of human breast adenocarcinoma [MCF-7 and MDA-MB-468], human malignant melanoma: [UACC-257]	Concentration not reported	α-humulene from *Eugenia zuchowskiae* inhibited all cell lines, with similar cytotoxicity against MCF-7 line as doxorubicin [LC50 of 1.1×10^{-4} and 1.4×10^{-4} mol/L, respectively].	Cole et al. 2007 [18]
Cell lines of human cervical carcinoma [HeLa], human colon adenocarcinoma [HT-29], monkey kidney [Vero]	$9.8 \times 10^{-7} - 9.8 \times 10^{-4}$ M	α-humulene demonstrated cytotoxicity against all cell lines. Tumour cell lines were more sensitive to cytotoxic activity than non-tumour Vero cells and murine macrophages.	Silva et al. 2008 [19]
Cell lines of human amelanotic melanoma [C32], renal cell adenocarcinoma [ACHN]	up to 4.9×10^{-4} M	α-humulene did not demonstrate cytotoxicity with IC50 > 4.9×10^{-4} mol/L against both C32 and ACHN lines. However, β-caryophyllene showed cytotoxic activity against both.	Loizzo et al. 2008 [20]

IC50 – half maximal inhibitory concentration; LC50 – half maximal lethal concentration

Anti-inflammatory properties

Exploration into the *in vivo* anti-inflammatory properties of α-humulene, isolated from *Cordia verbenacea*, has yielded significant insights as well. Passos et al. (2007) [39] showed the potent anti-inflammatory attributes of α-humulene by demonstrating its ability to significantly inhibit carrageenan-induced paw oedema in murine models and by a notable reduction in tumour necrosis factor (TNF)-α levels in response to carrageenan. Fernandes et al. (2007) [32] conducted a similar evaluation using oral administration of α-humulene against several experimental murine and rat models. Notably, administration of α-humulene at 50 mg/kg demonstrated a dose-dependent reduction in paw oedema, indicating its efficacy in mitigating the acute phase of inflammation. Additionally, it exhibited a sustained anti-inflammatory effect by inhibiting the late phase of carrageenan-induced oedema. Mechanistically, α-humulene interfered with multiple pathways involved in inflammation, including the inhibition of bradykinin, a platelet-activating factor, and histamine-induced oedema. Basting et al. (2019) [33] also observed a reduction in carrageenan-induced paw oedema. Despite not significantly affecting neutrophil migration, α-humulene suppressed the release of TNF-α and interleukin (IL)-1β and inhibited prostaglandin E2 production. Similar findings were observed by Medeiros et al. (2007) [34] in lipopolysaccharide-induced rat paw oedema. Key observations included a reduction in pro-inflammatory cytokines, inhibition of kinin B1 receptor upregulation, and suppression of neutrophil recruitment by targeting nuclear factor-kappa B (NF-κB) activation. Notably, α-humulene's efficacy surpassed that of trans-caryophyllene.

In a murine model of airway allergic inflammation, female BALB/c mice challenged with ovalbumin experienced a significant reduction in eosinophil recruitment to bronchoalveolar lavage fluid and lung tissue when administered α-humulene preventively or therapeutically [35]. α-humulene exhibited modulation of critical asthma-related mediators, including IL-5, C-C motif chemokine11, and leukotriene B4, along with the inhibition of P-selectin expression, a crucial factor in eosinophil migration. Additionally, α-humulene showed inhibitory effects on NF-κB and activator protein-1. Histological analysis indicated a decrease in mucus hypersecretion, suggesting a potential role in balancing T-helper cell responses.

Contrary to the widely positive findings reported regarding the anti-inflammatory activity of α-humulene, Viveiro et al. (2022) [22] investigated pterygium fibroblasts through *in vitro* exposure experiments. Third-passage pterygium fibroblasts were subjected to α-humulene concentrations (0.25, 2.5, and 25 µmol/L), and the subsequent cell viability assay revealed no significant cytotoxicity and minimal variation in inflammatory markers. This highlights the importance of considering cell-type-specific responses and experimental conditions in evaluating potential therapeutic benefits.

Antimicrobial properties

Early exploration into the *in vitro* antimicrobial potential of α-humulene was undertaken by Pichette et al. (2006) [23] who observed antibacterial activity against *Staphylococcus aureus* at a mean inhibitory concentration (MIC) of 1.3×10^{-5} mol/L. Subsequent investigations by Azizan et al. (2017) [24] expanded on this by demonstrating the dose-dependent bacteriostatic and bactericidal effects of α-humulene. Employing the broth microdilution method and α-humulene sourced from *Orthosiphon stamineus* and *Ficus deltoidei*, the study showed moderate to strong inhibition across a range of bacteria. Notably, oral Gram-negative species (*Porphyromonas gingivalis*, *Fusobacterium nucleatum*, and *Aggregatibacter actinomycetemcomitans*) exhibited greater susceptibility compared to Gram-positive bacteria (*Enterococcus faecalis*, *Streptococcus mutans*, *Streptococcus mitis*, and *Streptococcus salivarius*). Mechanistically, electron microscopy revealed morphological alterations, indicating that α-humulene interfered with the membrane structure or cell wall of oral bacteria. This effect was ascribed to the substantial electronegativity resulting from the carbon double bond configurations in its molecular structure.

Jang et al. (2020) [25] evaluated the *in vitro* antibacterial and antibiofilm effects of α-humulene extracted from *Bacteroides fragilis*. The study determined the MIC for cell growth and biofilm formation to be 9.8×10^{-6} mol/L. Through qRT-PCR analysis, concentration-dependent reductions in the expression of *bmeB1* and

bmeB3 genes were observed in various *Bacteroides fragilis* strains. This indicated increased antibiofilm action given these genes are implicated in the development of the biofilm matrix and antibiotic resistance. Notably, there was a marked reduction in cellular metabolic activity at concentrations of 3.9×10^{-5} to 7.8×10^{-5} mol/L. Moreover, confocal laser scanning microscopy revealed that α-humulene not only diminished cell density and thickness but also effectively reduced protein, carbohydrate, and nucleic acid levels. Rossato et al. (2022) [26] evaluated α-humulene's antibacterial potential against *Staphylococcus aureus* and *Enterococcus faecalis* using experimental light-cured periodontal dressing formulations and the modified direct contact model. Formulations with 10% and 20% α-humulene reduced bacterial growth after 1 and 24 hours of incubation compared to the control group, indicating sustained antibacterial activity.

Xing et al. (2018) [40] focused on evaluating the antifungal properties of humulene. Findings revealed a dose-dependent impediment of *Peronophythora litchii* growth, with scanning and transmission electron microscopy uncovering discernible morphological and ultrastructural changes. *In vivo* evaluations on litchi foliage and fruits demonstrated a notable reduction in disease severity. α-humulene exhibited weak inhibitory effects against *Peronophythora litchi* at high concentrations (8.7×10^{-4} to 4.4×10^{-3} mol/L) [40].

Antiallergic properties

The antiallergic potential of α-humulene was demonstrated by Tanaka et al. (1996) [36] in the context of treatment for atopic conditions. Using a sensitised murine model of passive cutaneous anaphylaxis, α-humulene administration prior to antigen challenge demonstrated dose-dependent inhibition of allergic reactions at 20, 40, and 80 mg/kg, with approximately four times the potency of the reference drug tranilast. However, the observed effects were less potent than the antiallergy effects triggered by β-caryophyllene. The study suggested the bicyclic ring structure inherent in β-caryophyllene may have contributed to the enhanced antiallergy activity of the compound. Additionally, Fernandes et al. (2007) [32] demonstrated α-humulene reduced paw oedema in sensitised mice challenged with ovoalbumin, suggesting its anti-inflammatory properties in alleviating allergic responses.

Antiparasitic properties

De Oliveria et al. (2017) [27] evaluated the antischistosomal effects of α-humulene against *Schistosoma mansoni* following *in vitro* exposure. At concentrations of 1 mol/L, α-humulene exhibited notable efficacy, causing mortality rates of 60% for female worms and 80% for male worms after a 72-hour incubation period. The sesquiterpene also induced a substantial reduction in motor activity and oviposition across all concentrations, highlighting its potential as a promising antiparasitic agent. Additionally, there were significant inhibitory effects of α-humulene on the excretory system of male *Schistosoma mansoni* adult worms. However, this inhibitory activity was not observed in female worms. The mechanism underlying this was attributed to the inhibition of the expression of P-glycoprotein, a product of the multidrug resistance 2 gene, within the excretory system of male *Schistosoma mansoni* worms. The study further employed Hoechst probe and scanning electron microscopy to assess the impact of α-humulene on the membrane integrity of *Schistosoma mansoni*. This highlighted the substantial damage caused to the tegument by α-humulene exposure.

Gastroprotective properties

Lemos et al. (2015) [41] investigated the potential gastroprotective effects of α-humulene through their study involving murine gastric ulcer models The researchers administered an oral dose of 30 mg/kg of isolated α-humulene, equivalent to omeprazole dosing. This dose led to a substantial reduction of 76.20% in gastric lesions induced by 0.2 ml of an ethanol/hydrogen chloride solution (60%/0.3 M). This investigation revealed two significant mechanisms contributing to the gastroprotective effects: a reduction in gastric acid secretion and an increase in mucus production.

Larvicidal properties

The larvicidal potential of α-humulene was examined by Govindarajan et al. (2016) [28] in an *in vivo* study encompassing three vector species: *Anopheles subpictus* Grassi (Culicidae)*; Aedes albopictus* Skuse (Culicidae); and *Culex tritaeniorhynchus* Giles (Culicidae). The researchers determined the lethal concentration 50 (LC50) values as 3.0×10^{-5}, 3.4×10^{-5}, and 3.6×10^{-5} mol/L for the respective species. The impact on non-target species was notably less, with a significantly lower LC50 of 5.0×10^{-3} mol/L observed in *Gambusia affinis* fish. Furthermore, there were no adverse effects on fish survival or swimming activity following the administration of α-humulene concentrations approaching the calculated larvae LC90.

Hung et al. (2021) [29] also evaluated the larvicidal effects of α-humulene from the essential oil of *Lantana camara*. It showed promising larvicidal activities against important mosquito vectors, with 48-hour LC50 values of 1.9×10^{-4} mol/L for *Anopheles aegypti* L. (Culicidae), 1.9×10^{-4} mol/L for *Aedes albopictus*, and 4.3×10^{-4} mol/L for *Culex quinquefasciatus* Say (Culicidae). Additionally, α-humulene exhibited notable mosquito larvicidal effects with an inhibitory concentration (IC50) value of 7.9×10^{-4} against electric eel acetylcholinesterase. Furthermore, *in silico* studies have demonstrated α-humulene exhibits significant binding energy in docking studies targeting sterol carrier protein-2, indicating its potential as an effective antilarvicidal agent [30].

Molluscicidal properties

In the context of molluscs acting as intermediate hosts for several helminths, α-humulene's potential molluscicidal properties have been subjected to scrutiny [29]. Notably, its LC50 values at 24 hours have been reported to be 9.3×10^{-5} mol/L, 9.3×10^{-5} mol/L, and 9.1×10^{-5} mol/L for *Pomacea caniculate* (Lam.), Ampullariidae; *Gyraulus convexiusculus* (Hutton), Planorbidae; and *Tarebia granifera* (Lam.), Thiaridae, respectively.

Cannabimimetic properties

The cannabimimetic properties of α-humulene were demonstrated by LaVigne et al. (2021) [42] through *in vivo* and *in vitro* experiments. Using various behavioural assays in mice, α-humulene manifested notable antinociceptive effects. The study identified specific receptor targets influenced by α-humulene, revealing in-

teractions with cannabinoid type 1 (CB1) and type 2 (CB2) receptors, as well as adenosine receptor A2a, through *in vitro* experiments. Furthermore, *in vivo* experiments demonstrated a synergistic interaction between α-humulene and the synthetic cannabinoid agonist WIN55,212–2, leading to enhanced antinociceptive effects. There were selective effects of α-humulene on various behaviours associated with the tetrad, emphasising its complex interplay with multiple receptor systems. Notably, the *in vitro* studies showed the CB1-dependent nature of α-humulene activation, requiring relatively high concentrations for receptor activation, a phenomenon reversible by the CB1 antagonist rimonabant.

Discussion

The scoping review undertaken in this study unveils the landscape of α-humulene's pharmacological potential, revealing a diverse spectrum of documented properties across various studies. These encompass anti-inflammatory, antimicrobial, antiproliferative, antiallergic, gastroprotective, and even cannabimimetic effects. α-humulene interacts with diverse biological pathways, suggesting its potential for addressing various health conditions.

The review further emphasises the pivotal role played by specific species that yield substantial amounts of α-humulene, carrying profound implications for pharmaceutical applications. Noteworthy among these is *Aframomum melegueta*, a West African spice renowned for its historical medicinal uses and significant α-humulene content, rendering it an enticing candidate for therapeutic extraction [43]. Likewise, several *Leptospermum* species, known for their potent antimicrobial properties, exhibit notable levels of α-humulene [44]. Additionally, *Humulus lupulus*, commonly known as hops, has a high α-humulene content. Given its well-documented applications and extensively studied properties, hops offer a versatile avenue for the development of α-humulene-based therapeutics [45]. Another plant of significance is *Cannabis sativa*, in which α-humulene is already utilised in full-spectrum cannabis-based medicinal products. The cannabimimetic effects of α-humulene may give rise to potential additive or synergistic effects when administered alongside cannabinoids and other active pharmaceutical ingredients, broadly referred to as 'the entourage effect' [46]. Collectively, these species enrich the available sources of α-humulene, highlighting its prevalence across a diverse range of botanicals. These species hold promise as potential sources for pharmaceutical extraction due to their abundant α-humulene content. By harnessing extracts derived from these species, either in combination with other compounds or as standalone treatments, further exploration of potential solutions for various health conditions becomes viable.

The translation of promising preclinical findings to clinical practice encounters barriers. Variability in α-humulene yield across different botanical sources poses logistical challenges for large-scale extraction [47]. The limited exploration of isolated α-humulene outside of whole essential oils highlights the importance of comprehensive investigations into isolated properties [48]. Addressing this inconsistency requires the identification of further plant species with high α-humulene yields or increased investigation of biosynthesis pathways for synthetic production.

The potential of α-humulene as an anticancer agent is particularly promising. Studies have established wide-ranging effects on various cancer cell lines, revealing nuanced interactions with distinct tumour types. This is coherent with preclinical studies on other terpenes, which similarly find anticancer potential [49]. The mechanism of action of α-humulene appears multifactorial, including increasing the production of reactive oxidative species and induction of apoptosis [16, 37]. Moreover, α-humulene was associated with glutathione depletion, which makes cancer cells more sensitive to stress caused by reactive oxidative species [16]. α-humulene was associated with an increase in GST activity, which is also seen with other terpenes [50]. This feature, however, is typically associated with improved cancer cell survival and resistance to certain chemotherapeutics due to the associated efflux of anticancer agents from the cell [51]. Putra et al. investigated α-humulene's interaction with the overexpressed HER-2 protein using docking methods and shed light on its potential as an anti-breast-cancer agent. The *in silico* molecular docking simulations reveal a binding energy of − 7.50 kcal/mol, affirming its efficacy against breast cancer [52]. As such, its effects within human studies are eagerly awaited, especially as preclinical studies show that α-humulene may have synergistic effects with doxorubicin and other chemotherapeutics [13]. This is particularly important as present studies indicate that α-humulene would not be a suitable chemotherapeutic agent in isolation and would otherwise be best used alongside traditional chemotherapeutics [53]. Its lower toxicity profile also supports this as a potential future use, provided efficacy can be determined [54]. Further mechanistic studies, including investigations into synergistic interactions with established chemotherapeutics, are ultimately necessary to fully leverage α-humulene's potential in cancer biology [55].

Beyond its cancer-related properties, α-humulene is a compelling anti-inflammatory agent. Its modulation of the NF-κB pathway and subsequent suppression of key inflammatory mediators demonstrates its potential in various inflammatory conditions [56]. Insights gained from murine models of asthma highlight its immunomodulatory potential, positioning α-humulene as a contender for treating inflammatory and atopic conditions [35]. Additionally, its analgesic potential and observed gastroprotective effects hold significance [57]. In contrast to traditional non-steroidal anti-inflammatory drugs, notorious for causing peptic ulcer disease and adverse renal effects, α-humulene offers a potentially safer alternative for managing inflammation-associated conditions [58].

The pharmacodynamic profile of α-humulene indicates its capability of addressing various aspects of health and disease. Its interactions with different molecular pathways suggest complex biochemical dialogues within cells and tissues. This complexity is particularly relevant for multifactorial conditions like cancer and chronic inflammatory diseases, where several dysregulated pathways contribute to aetiology and pathogenesis [59, 60]. By targeting these pathways, α-humulene introduces a novel therapeutic approach distinct from the traditional 'one-target-one-drug' paradigm [61, 62].

The antimicrobial properties of α-humulene enrich its pharmacological profile, spanning antibacterial, antiparasitic, and antifungal effects. Its efficacy in restraining biofilm formation and

curtailing gene expression associated with biofilm matrix development and antibiotic resistance is particularly relevant given the growing appreciation for the role of biofilm in antibiotic resistance [63,64]. This enhances the potential to address antibiotic-resistant infections, a pressing global health concern [65].

The paucity of clinical studies involving α-humulene necessitates thorough evaluation in the clinical setting to validate its efficacy, safety, and optimal dosage regimens in human subjects before widespread use [66]. In addition, it is crucial to emphasise the necessity of pharmacokinetic studies, particularly for terpenes like α-humulene. Due to their lipophilic nature, terpenes often exhibit poor water solubility and are susceptible to rapid metabolism and elimination, leading to low oral bioavailability [2,67]. Therefore, rigorous pharmacokinetic studies in animals and humans are essential to optimise dosing strategies to understand α-humulene's therapeutic potential [68]. Strategies to enhance α-humulene's bioavailability, such as formulations that improve solubility and stability, could significantly enhance its clinical utility [69]. There is one pilot study currently underway seeking to explore the effects of α-humulene on stress when combined with forest bathing, for which the results will be eagerly awaited [70].

Acknowledging the limitations of this scoping review is vital. The heterogeneity of study methodologies, including variations in cell lines, experimental conditions, and assessment methods, poses a challenge in directly comparing the results. This heterogeneity limits the ability to perform quantitative meta-analyses and emphasises the need for cautious interpretation. Variations in study design, quality, sample size, and reporting practices could impact the overall strength of evidence. The limited number of *in vivo* studies and the absence of clinical trials restrict the ability to directly extrapolate findings to human populations to provide clinical validation [71]. Preclinical studies often involve isolated cells or animal models, which may not fully replicate human physiological responses [72]. Furthermore, it is important to acknowledge the limitations associated with the compilation of α-humulene yield data from various locations, with a focus solely on the highest reported yields. This approach might not account for potential variations in cultivation practices, environmental factors, and genetic influences that can significantly affect yield outcomes. Relying solely on the highest reported yields could lead to an incomplete understanding of the compound's availability and potentially skew the representation of humulene yields. Moreover, studies often failed to specify whether the α-humulene yield was from the whole plant, flower, or another plant component. This lack of clarity restricts the interpretation, as there can be substantial variation in terpene content across various flower structures [73].

Overall, this systematic review provides valuable insights into α-humulene's potential therapeutic properties. However, addressing limitations through standardised methodologies, clinical trials, and consistent reporting practices is crucial for an accurate understanding of its multifaceted effects and clinical applications. The future of α-humulene's clinical translation hinges on collaborative efforts, pharmacokinetic evaluations, rigorous clinical trials, innovative formulation strategies, and partnerships across disciplines. Through these efforts, α-humulene's clinical translation

can be accelerated in light of its many promising therapeutic properties.

Materials and Methods

A scoping review, utilising methods outlined by Arksey and O'Malley and Levac et al. [74,75], was conducted of the current literature on α-humulene.

Research question

This scoping review focused on identifying and clarifying key research aspects and characteristics of the available literature with regards to α-humulene. Given its potential therapeutic effects in the clinical setting, this review evaluated the evidence base for α-humulene in terms of its extraction and properties that may be translated for medicinal purposes. This review also aimed to identify any specific gaps in the evidence base that may inform the work of future researchers in this area of sesquiterpene research.

Data sources and search strategy

A broad search was conducted of MEDLINE, PubMed, and EMBASE databases in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines [76]. A search was conducted from 1946 to July 14, 2023, utilising the search terms 'humulene', 'alpha-humulene', and 'alpha-caryophyllene' with the Boolean operator 'OR' (**Table S1**, Supporting Information). The literature search was conducted by three independent researchers. For any discrepancies identified, a senior author planned to review these if necessary. Additional relevant articles were included through manual search of the bibliographies of included studies. Articles were screened in relation to the topic area and included if deemed to meet the inclusion criteria. The precise search strategies performed can be found in Supplementary Material C.

Study selection criteria

Inclusion criteria consisted of original articles related to α-humulene that included extraction, pre-clinical, and clinical research. Studies were excluded if they did not constitute original primary research or the outcomes of α-humulene were not reported in isolation from other essential oils extracted from plant species.

Data extraction and presentation

Data extraction was performed independently by three authors. If outcomes were not reported within the published article, but described within the methodology, corresponding authors were contacted for additional information. Concentrations are presented as percentage yield or micromolar concentrations (μM) with the standard deviation (S. D.), standard error (S. E.), or range if reported.

Supporting information

Supplementary Material A: References for studies of extraction yields for different species
Supplementary Material B: Reported yields of α-humulene
Supplementary Material C: Search strategy

Contributors' Statement

Data collection: N. Dalavaye, M. Nicholas, M. Pillai; Design of the study: N. Dalavaye, M. Nicholas, M. Pillai, S. Erridge, M.H.Sodergren; Statistical analysis: N. Dalavaye, M. Nicholas, M. Pillai, S. Erridge; analysis and interpretation of the data: N. Dalavaye, M. Nicholas, M. Pillai, S. Erridge, M.H.Sodergren; drafting the manuscript: N. Dalavaye, M. Nicholas, M. Pillai, S. Erridge; critical revision of the manuscript: N. Dalavaye, M. Nicholas, M. Pillai, S. Erridge, M.H.Sodergren

Conflict of Interest

SE is the Head of Research at Curaleaf Clinic. MHS is the Chief Medical Officer at Curaleaf International.

References

[1] Degenhardt J, Köllner TG, Gershenzon J. Monoterpene and sesquiterpene synthases and the origin of terpene skeletal diversity in plants. Phytochemistry 2009; 70: 1621–1637

[2] Masyita A, Mustika Sari R, Dwi Astuti A, Yasir B, Rahma Rumata N, Emran TB, Nainu F, Simal-Gandara J. Terpenes and terpenoids as main bioactive compounds of essential oils, their roles in human health and potential application as natural food preservatives. Food Chem X 2022; 13: 100217

[3] Yang W, Chen X, Li Y, Guo S, Wang Z, Yu X. Advances in pharmacological activities of terpenoids. Nat Prod Commun 2020; 15: doi: 10.1177/1934578X20903555

[4] Mander L, Hung-Wen L Comprehensive Natural Products II: Chemistry and Biology, Volumes 1–10. München: Elsevier Science; 2010

[5] Simonsen JL, Todd AR. 32. Cannabis indica. Part X. The essential oil from Egyptian hashish. J Chem Soc 1942; 188–188

[6] Ugai T, Sasamoto N, Lee HY, Ando M, Song M, Tamimi RM, Kawachi I, Campbell PT, Giovannucci EL, Weiderpass E, Rebbeck TR, Ogino S. Is early-onset cancer an emerging global epidemic? Current evidence and future implications. Nat Rev Clin Oncol 2022; 19: 656–673

[7] Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, Ferrucci L, Gilroy DW, Fasano A, Miller GW, Miller AH, Mantovani A, Weyand CM, Barzilai N, Goronzy JJ, Rando TA, Effros RB, Lucia A, Kleinstreuer N, Slavich GM. Chronic inflammation in the etiology of disease across the life span. Nat Med 2019; 25: 1822–1832

[8] Andersson DI, Balaban NQ, Baquero F, Courvalin P, Glaser P, Gophna U, Kishony R, Molin S, Tønjum T. Antibiotic resistance: turning evolutionary principles into clinical reality. FEMS Microbiol Rev 2020; 44: 171–188

[9] Harada H, Yu F, Okamoto S, Kuzuyama T, Utsumi R, Misawa N. Efficient synthesis of functional isoprenoids from acetoacetate through metabolic pathway-engineered Escherichia coli. Appl Microbiol Biotechnol 2009; 81: 915–925

[10] Leite GM, Barbosa MO, Lopes MJP, Delmondes GA, Bezerra DS, Araújo IM, Carvalho de Alencar CD, Coutinho HDM, Peixoto LR, Barbosa-Filho JM, Felipe CFB, Barbosa R, Alencar de Menezes IR, Kerntof MR. Pharmacological and toxicological activities of α-humulene and its isomers: A systematic review. Trends Food Sci Technol 2021; 115: 255–274

[11] Ambrož M, Šmatová M, Šadibolová M, Pospíšilová E, Hadravská P, Kašparová M, Hanušová Skarková V, Králová V, Skálová L. Sesquiterpenes α-humulene and β-caryophyllene oxide enhance the efficacy of 5-fluorouracil and oxaliplatin in colon cancer cells. Acta Pharm 2019; 69: 121–128

[12] Ambrož M, Matoušková P, Skarka A, Zajdlová M, Žáková K, Skálová L. The effects of selected sesquiterpenes from Myrica rubra essential oil on the efficacy of doxorubicin in sensitive and resistant cancer cell lines. Molecules 2017; 22: 1021–1028

[13] Ambrož M, Boušová I, Skarka A, Hanušová V, Králová V, Matoušková P, Szotáková B, Skálová L. The influence of sesquiterpenes from Myrica rubra on the antiproliferative and pro-oxidative effects of doxorubicin and its accumulation in cancer cells. Molecules 2015; 20: 15343–15358

[14] Su YC, Hsu KP, Wang EIC, Ho CL. Composition, in vitro cytotoxic, and antimicrobial activities of the flower essential oil of diospyros discolor from Taiwan. Nat Prod Commun 2015; 10: 1311–1314. doi: https://doi.org/10.1177/1934578X1501000744

[15] Hadri A, Gómez del Río MA, Sanz J, González Coloma A, Idaomar M, Ribas Ozonas B, Benedí González J, Sánchez Reus MI. Cytotoxic activity of alpha-humulene and trans-caryophyllene from Salvia officinalis in animal and human tumor cells. An Real Acad Farm 2010; 76: 343–356

[16] Legault J, Dahl W, Debiton E, Pichette A, Madelmont JC. Antitumor activity of balsam fir oil: Production of reactive oxygen species induced by alpha-humulene as possible mechanism of action. Planta Med 2003; 69: 402–407

[17] Legault J, Pichette A. Potentiating effect of β-caryophyllene on anticancer activity of α-humulene, isocaryophyllene and paclitaxel. J Pharm Pharmacol 2010; 59: 1643–1647

[18] Cole RA, Bansal A, Moriarity DM, Haber WA, Setzer WN. Chemical composition and cytotoxic activity of the leaf essential oil of Eugenia zuchowskiae from Monteverde, Costa Rica. J Nat Med 2007; 61: 414–417

[19] da Silva SL, Chaar JS, Figueiredo PMS, Yano T. Cytotoxic evaluation of essential oil from Casearia sylvestris Sw on human cancer cells and erythrocytes. Acta Amazon 2008; 38: 107–112

[20] Loizzo MR, Tundis R, Saab AM, Statti GA, Menichini F. Antiproliferative effects of essential oils and their major constituents in human renal adenocarcinoma and amelanotic melanoma cells. Cell Prolif 2008; 41: 1002–1012

[21] Fukuoka K, Sawabe A, Sugimoto T, Koga M, Okuda H, Kitayama T, Shirai M, Komai K, Komemushi S, Matsuda K. Inhibitory actions of several natural products on proliferation of rat vascular smooth muscle cells induced by Hsp60 from Chlamydia pneumoniae J138. J Agric Food Chem 2004; 52: 6326–6329

[22] Viveiros MMH, Silva MG, da Costa JGM, de Oliveira AG, Rubio C, Padovani CR, Rainho CA, Schellini SA. Anti-inflammatory effects of α-humulene and β-caryophyllene on pterygium fibroblasts. Int J Ophthalmol 2022; 15: 1903–1907

[23] Pichette A, Larouche PL, Lebrun M, Legault J. Composition and antibacterial activity of Abies balsamea essential oil. Phytother Res 2006; 20: 371–373

[24] Azizan N, Mohd Said S, Zainal Abidin Z, Jantan I. Composition and antibacterial activity of the essential oils of Orthosiphon stamineus Benth and Ficus deltoidea Jack against pathogenic oral bacteria. Molecules 2017; 22: 2135

[25] Jang HI, Rhee KJ, Eom YB. Antibacterial and antibiofilm effects of alpha-humulene against Bacteroides fragilis. Can J Microbiol 2020; 66: 389–399

[26] Rossato TCA, Alves T, Cuevas-Suárez CE, Rosa WLO, Silva AF, Piva E, Zanchi CH, Lund RG. Effect of alpha-humulene incorporation on the properties of experimental light-cured periodontal dressings. Braz Oral Res 2022; 36: e091

[27] de Oliveira RN, Dos Santos KR, Mendes TMF, Garcia VL, Santos Oliveira AS, de Lourdes Sierpe Jeraldo V, Allegretti SM. Sesquiterpenes evaluation on Schistosoma mansoni: Survival, excretory system and membrane integrity. Biomed Pharmacother 2017; 90: 813–820

[28] Govindarajan M, Benelli G. α-Humulene and β-elemene from Syzygium zeylanicum (Myrtaceae) essential oil: Highly effective and eco-friendly larvicides against Anopheles subpictus, Aedes albopictus, and Culex tritaeniorhynchus (Diptera: Culicidae). Parasitol Res 2016; 115: 2771–2778

[29] Huy Hung N, Ngoc Dai D, Satyal P, Thi Huong L, Thi Chinh B, Quang Hung D, Anh Tai T, Setzer WN. Lantana camara essential oils from Vietnam:

Chemical composition, molluscicidal, and mosquito larvicidal activity. Chem Biodivers 2021; 18: e2100145

[30] Andrade-Ochoa S, Correa-Basurto J, Rodríguez-Valdez LM, Sánchez-Torres LE, Nogueda-Torres B, Nevárez-Moorillón GV. In vitro and in silico studies of terpenes, terpenoids and related compounds with larvicidal and pupaecidal activity against Culex quinquefasciatus Say (Diptera: Culicidae). Chem Cent J 2018; 12: 53

[31] Zheng GQ, Kenney PM, Lam LKT. Sesquiterpenes from clove (Eugenia caryophyllata) as potential anticarcinogenic agents. J Nat Prod 1992; 55: 999–1003

[32] Fernandes ES, Passos GF, Medeiros R, da Cunha FM, Ferreira J, Campos MM, Pianowski LF, Calixto JB. Anti-inflammatory effects of compounds alpha-humulene and (−)-trans-caryophyllene isolated from the essential oil of Cordia verbenacea. Eur J Pharmacol 2007; 569: 228–236

[33] Basting RT, Spindola HM, Sousa IMO, Queiroz NCA, Trigo JR, de Carvalho JE, Foglio MA. Pterodon pubescens and Cordia verbenacea association promotes a synergistic response in antinociceptive model and improves the anti-inflammatory results in animal models. Biomed Pharmacother 2019; 112: 108693

[34] Medeiros R, Passos GF, Vitor CE, Koepp J, Mazzuco TL, Pianowski LF, Campos MM, Calixto JB. Effect of two active compounds obtained from the essential oil of Cordia verbenacea on the acute inflammatory responses elicited by LPS in the rat paw. Br J Pharmacol 2007; 151: 618–627

[35] Rogerio AP, Andrade EL, Leite DFP, Figueiredo CP, Calixto JB. Preventive and therapeutic anti-inflammatory properties of the sesquiterpene α-humulene in experimental airways allergic inflammation. Br J Pharmacol 2009; 158: 1074–1087

[36] Tanaka S, Akimoto M, Tambe Y, Tabata M, Ikeshiro Y. Volatile antiallergic principles from a traditional herbal prescription of kampo medicine. Phytother Res 1996; 10: 238–241

[37] Chen H, Yuan J, Hao J, Wen Y, Lv Y, Chen L, Yang X. α-Humulene inhibits hepatocellular carcinoma cell proliferation and induces apoptosis through the inhibition of Akt signaling. Food Chem Toxicol 2019; 134: 110830

[38] Costa EV, Menezes LRA, Rocha SLA, Baliza IRS, Dias RB, Rocha CAG, Soares MBP, Bezerra DP. Antitumor properties of the leaf essential oil of Zornia brasiliensis. Planta Med 2015; 81: 563–567

[39] Passos GF, Fernandes ES, da Cunha FM, Ferreira J, Pianowski LF, Campos MM, Calixto JB. Anti-inflammatory and anti-allergic properties of the essential oil and active compounds from Cordia verbenacea. J Ethnopharmacol 2007; 110: 323–333

[40] Xing M, Zheng L, Deng Y, Xu D, Xi P, Li M, Kong G, Jiang Z. Antifungal activity of natural volatile organic compounds against litchi downy blight pathogen Peronophythora litchii. Molecules 2018; 23: 358

[41] Lemos M, Santin JR, Mizuno CS, Boeing T, De Sousa JPB, Nanayakkara D, Kenupp Bastos J, Faloni de Andrade S. Copaifera langsdorffii: Evaluation of potential gastroprotective of extract and isolated compounds obtained from leaves. Rev Bras Farmacogn 2015; 25: 238–245

[42] LaVigne JE, Hecksel R, Keresztes A, Streicher JM. Cannabis sativa terpenes are cannabimimetic and selectively enhance cannabinoid activity. Sci Rep 2021; 11: 8232

[43] Ilic N, Schmidt BM, Poulev A, Raskin I. Toxicological evaluation of grains of paradise (Aframomum melegueta) [Roscoe] K. Schum. J Ethnopharmacol 2010; 127: 352–356

[44] Caputo L, Smeriglio A, Trombetta D, Cornara L, Trevena G, Valussi M, Fratianni F, De Feo V, Nazzaro F. Chemical composition and biological activities of the essential oils of Leptospermum petersonii and Eucalyptus gunnii. Front Microbiol 2020; 11: 409

[45] Astray G, Gullón P, Gullón B, Munekata PES, Lorenzo JM. Humulus lupulus L. as a natural source of functional biomolecules. Appl Sci 2020; 10: 5074

[46] Russo EB. Taming THC: Potential cannabis synergy and phytocannabinoid-terpenoid entourage effects. Br J Pharmacol 2011; 163: 1344–1364

[47] Sharmeen J, Mahomoodally F, Zengin G, Maggi F. Essential oils as natural sources of fragrance compounds for cosmetics and cosmeceuticals. Molecules 2021; 26: 666

[48] Guzmán E, Lucia A. Essential oils and their individual components in cosmetic products. Cosmetics 2021; 8: 114

[49] Tomko AM, Whynot EG, Ellis LD, Dupré DJ. Anti-cancer potential of cannabinoids, terpenes, and flavonoids present in cannabis. Cancers (Basel) 2020; 12: 1985

[50] Elegbede JA, Maltzman TH, Elson CE, Gould MN. Effects of anticarcinogenic monoterpenes on phase II hepatic metabolizing enzymes. Carcinogenesis 1993; 14: 1221–1223

[51] Singh RR, Reindl KM. Glutathione S-transferases in cancer. Antioxidants 2021; 10: 701

[52] Putra IMH, Pratama IPAAC, Putra KDA, Pradnyaswari GAD, Laksmiani NPL. The potency of alpha-humulene as HER-2 inhibitor by molecular docking. Pharm Rep 2022; 2: 19

[53] Zhu S, Zhang T, Zheng L, Liu H, Song W, Liu D, Li Z, Pan CX. Combination strategies to maximize the benefits of cancer immunotherapy. J Hematol Oncol 2021; 14: 156

[54] Blowman K, Magalhães M, Lemos MFL, Cabral C, Pires IM. Anticancer properties of essential oils and other natural products. Evid-Based Complementary Altern Med 2018; 2018: 1–12

[55] Boshuizen J, Peeper DS. Rational cancer treatment combinations: An urgent clinical need. Mol Cell 2020; 78: 1002–1018

[56] Yu H, Lin L, Zhang Z, Zhang H, Hu H. Targeting NF-κB pathway for the therapy of diseases: Mechanism and clinical study. Signal Transduct Target Ther 2020; 5: 209

[57] Liktor-Busa E, Keresztes A, LaVigne J, Streicher JM, Largent-Milnes TM. Analgesic potential of terpenes derived from Cannabis sativa. Pharmacol Rev 2021; 73: 1269–1297

[58] Tai FWD, McAlindon ME. Non-steroidal anti-inflammatory drugs and the gastrointestinal tract. Clin Med (Northfield Il) 2021; 21: 131–134

[59] Wu S, Zhu W, Thompson P, Hannun YA. Evaluating intrinsic and non-intrinsic cancer risk factors. Nat Commun 2018; 9: 3490

[60] Zhao H, Wu L, Yan G, Chen Y, Zhou M, Wu Y, Li Y. Inflammation and tumor progression: Signaling pathways and targeted intervention. Signal Transduct Target Ther 2021; 6: 263

[61] Hashem S, Ali TA, Akhtar S, Nisar S, Sageena G, Ali S, Al-Mannai S, Therachiyil L, Mir R, Elfaki I, Mir MM, Jamal F, Masoodi T, Uddin S, Singh M, Haris M, Macha M, Bhat AA. Targeting cancer signaling pathways by natural products: Exploring promising anti-cancer agents. Biomed Pharmacother 2022; 150: 113054

[62] Stine ZE, Schug ZT, Salvino JM, Dang CV. Targeting cancer metabolism in the era of precision oncology. Nat Rev Drug Discov 2022; 21: 141–162

[63] Sharma D, Misba L, Khan AU. Antibiotics versus biofilm: An emerging battleground in microbial communities. Antimicrob Resist Infect Control 2019; 8: 76

[64] Bowler P, Murphy C, Wolcott R. Biofilm exacerbates antibiotic resistance: Is this a current oversight in antimicrobial stewardship? Antimicrob Resist Infect Control 2020; 9: 162

[65] Antimicrobial Resistance Collaborators. Global burden of bacterial antimicrobial resistance in 2019: A systematic analysis. The Lancet 2022; 399: 629–655

[66] Subbiah V. The next generation of evidence-based medicine. Nat Med 2023; 29: 49–58

[67] Prasanthi D, Lakshmi PK. Terpenes: Effect of lipophilicity in enhancing transdermal delivery of alfuzosin hydrochloride. J Adv Pharm Technol Res 2012; 3: 216–223

[68] Palmer ME, Andrews LJ, Abbey TC, Dahlquist AE, Wenzler E. The importance of pharmacokinetics and pharmacodynamics in antimicrobial drug development and their influence on the success of agents developed to combat resistant gram negative pathogens: A review. Front Pharmacol 2022; 13: 888079

[69] Palrasu M, Wright L, Patel M, Leech L, Branch S, Harrelson S, Khan S. Perspectives on challenges in cannabis drug delivery systems: Where are we? Med Cannabis Cannabinoids 2022; 5: 102–119

[70] University of Washington. Bratman G. Do terpenes play a role in the stress-reducing effects of a forest bathing intervention? ClinicalTrials. gov Identifier: NCT05316597 (2022 Nov 1). Accessed 2023-12-23 at: https://clinicaltrials.gov/ct2/show/NCT05316597

[71] Sun D, Gao W, Hu H, Zhou S. Why 90% of clinical drug development fails and how to improve it? Acta Pharm Sin B 2022; 12: 3049–3062

[72] Seyhan AA. Lost in translation: The valley of death across preclinical and clinical divide – identification of problems and overcoming obstacles. Transl Med Commun 2019; 4: 18

[73] Keefover-Ring K. The chemical biogeography of a widespread aromatic plant species shows both spatial and temporal variation. Ecol Evol 2022; 12: e9265

[74] Arksey H, O'Malley L. Scoping studies: Towards a methodological framework. Int J Soc Res Methodol 2005; 8: 19–32

[75] Levac D, Colquhoun H, O'Brien KK. Scoping studies: Advancing the methodology. Implement Sci 2010; 5: 69

[76] Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, Shamseer L, Tetzlaff JM, Akl EA, Brennan SE, Chou R, Glanville J, Grimshaw JM, Hróbjartsson A, Lalu MM, Li T, Loder EW, Mayo-Wilson E, McDonald S, McGuinness LA, Stewart LA, Thomas J, Tricco AC, Welch VA, Whiting P, Moher D. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. BMJ 2021; 372: 71

[77] Ajaiyeoba EO, Ekundayo O. Essential oil constituents of Aframomum melegueta (Roscoe) K. Schum. seeds (alligator pepper) from Nigeria. Flavour Fragr J 1999; 14: 109–111

[78] Brophy JJ, Goldsack RJ, Bean AR, Forster PI, Lepsch BJ. Leaf essential oils of the genus Leptospermum (Myrtaceae) in eastern Australia. Part 6. Leptospermum polygalifolium and allies. Flavour Fragr J 2000; 15: 271–277

[79] Duarte PF, do Nascimento LH, Fischer B, Lohmann AM, Bandiera VJ, Fernandes IA, Magro JD, Valduga E, Cansian RL, Paroul N, Junges A. Effect of extraction time on the yield, chemical composition, and antibacterial activity of hop essential oil against lactic acid bacteria (Lactobacillus brevis and Lactobacillus casei) beer spoilage. Curr Microbiol 2023; 80: 237

[80] Sakurai K, Tomiyama K, Yaguchi Y, Asakawa Y. The characteristic smell emitted from two scale insects, *Ceroplastes japonicus* and *Ceroplastes rubens*. Biosci Biotechnol Biochem 2020; 84: 1541–1545

[81] Govindarajan M, Rajeswary M, Arivoli S, Tennyson S, Benelli G. Larvicidal and repellent potential of Zingiber nimmonii (J. Graham) Dalzell (Zingiberaceae) essential oil: an eco-friendly tool against malaria, dengue, and lymphatic filariasis mosquito vectors? Parasitol Res 2016; 115: 1807–1816