

*Review*



# **Metallothionein: A Comprehensive Review of Its Classification, Structure, Biological Functions, and Applications**

**Ruoqiu Yang <sup>1</sup> , Dumila Roshani <sup>2</sup> , Boya Gao <sup>1</sup> , Pinglan Li 1,\* and Nan Shang 2,[\\*](https://orcid.org/0000-0002-8620-4631)**

- <sup>1</sup> Key Laboratory of Precision Nutrition and Food Quality, College of Food Science and Nutritional Engineering, China Agricultural University, No, 17 Qinghua East Road, Haidian District, Beijing 100083, China; yangruoqiu@cau.edu.cn (R.Y.); s20213061034@cau.edu.cn (B.G.)
- <sup>2</sup> College of Engineering, China Agricultural University, No, 17 Qinghua East Road, Haidian District, Beijing 100083, China; dumila@cau.edu.cn
- **\*** Correspondence: lipinglan@cau.edu.cn (P.L.); nshang@cau.edu.cn (N.S.)

**Abstract:** Metallothionein is a cysteine-rich protein with a high metal content that is widely found in nature. In addition to heavy metal detoxification, metallothionein is well known as a potent antioxidant. The high sulfhydryl content of metallothionein confers excellent antioxidant activity, enabling it to effectively scavenge free radicals and mitigate oxidative stress damage. In addition, metallothionein can play a neuroprotective role by alleviating oxidative damage in nerve cells, have an anticancer effect by enhancing the ability of normal cells to resist unfavorable conditions through its antioxidant function, and reduce inflammation by scavenging reactive oxygen species. Due to its diverse biological functions, metallothionein has a broad potential for application in alleviating environmental heavy metal pollution, predicting and diagnosing diseases, and developing skin care products and health foods. This review summarizes the recent advances in the classification, structure, biological functions, and applications of metallothionein, focusing on its powerful antioxidant effects and related functions.

check for updates

**Citation:** Yang, R.; Roshani, D.; Gao, B.; Li, P.; Shang, N. Metallothionein: A Comprehensive Review of Its Classification, Structure, Biological Functions, and Applications. *Antioxidants* **2024**, *13*, 825. [https://doi.org/10.3390/](https://doi.org/10.3390/antiox13070825) [antiox13070825](https://doi.org/10.3390/antiox13070825)

Academic Editor: Simone Ciofi-Baffoni

Received: 27 May 2024 Revised: 24 June 2024 Accepted: 26 June 2024 Published: 9 July 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

**Keywords:** metallothionein; antioxidant; heavy metal detoxification; neuroprotective; anticancer; anti-inflammatory

# **1. Introduction**

Metal ions play an important role in cellular energy transfer, signaling, and other life activities; however, excessive concentrations can be toxic to cells. Cells have evolved two metal resistance mechanisms: metal membrane transport and intracellular chelation, and metallothionein is a chelator that responds to stimuli.

Metallothionein is a class of low-molecular-weight, cysteine-rich, high-metal-content proteins that was first discovered and isolated from equine kidneys by Margoshes and Valle in 1957 [\[1](#page-13-0)[,2\]](#page-13-1). Metallothionein is widely distributed in nature and has been found in vertebrates, invertebrates, plants, and microbes [\[3\]](#page-13-2). Since the day of its discovery, metallothionein's distinct structure has drawn attention. Its structure gives it specificity, stability, and dynamic variability while also greatly enhancing its antioxidant and metal ion binding capacities [\[4\]](#page-13-3). Metallothionein is capable of chelating metal ions via sulfhydryl groups to exert metal detoxification [\[5\]](#page-13-4). The high sulfhydryl content of metallothionein enables it to effectively scavenge various free radicals, such as hydroxyl radicals and oxygen radicals, and the reaction rate constant with hydroxyl radicals reaches 300 times that of glutathione [\[6\]](#page-13-5). Metallothionein, by virtue of its metal ion binding and antioxidant capacity, is able to reduce the damage caused by metal ions and oxidative stress on nerve cells [\[7](#page-13-6)[,8\]](#page-13-7). In addition, as an antioxidant, it can protect normal cells from external damage and reduce the risk of cancer [\[9\]](#page-13-8). When an inflammatory response occurs, metallothionein is able to scavenge reactive oxygen species to exert an anti-inflammatory effect and can also promote tissue repair and regeneration to a certain extent [\[10\]](#page-13-9). In recent years, many additional

studies on the biological functions of metallothionein have appeared, but there has been no recent comprehensive summary of its functions. Metallothionein, as a highly effective antioxidant, can play an important role in a variety of physiological and biochemical processes. A comprehensive understanding of the function of metallothionein is important for its maximum exploitation. The aim of this review is to introduce the classification, structure, biological function, and current application of metallothionein, with a focus on summarizing the various biological functions of metallothionein with a view to providing reference for its further development and utilization.

### **2. The Classification and Structure of Metallothionein**

#### *2.1. Classification*

Binz et al. proposed a new classification system to better distinguish between different species of metallothionein. They defined metallothionein as a superfamily, under which it is divided into various families based on evolutionary correlations. There are 15 families in the metallothionein superfamily, including vertebrate MTs, mollusc MTs, crustacean MTs, prokaryotic MTs, plant MTs, etc. (Figure [1\)](#page-1-0) [\[11\]](#page-13-10). The human MT family is divided into four classes, MT1 to MT4, encoded by 11 active genes (*MT1A*, *MT1B*, *MT1E*, *MT1F*, *MT1G*, *MT1H*, *MT1M*, *MT1X*, *MT2A*, *MT3*, and *MT4*) located in the chromosome 16 cluster [\[12\]](#page-13-11). MT1 and MT2 are expressed in most organs and tissues; MT3 is mainly expressed in the brain, whereas MT4 is expressed in stratified squamous epithelial cells [\[13–](#page-13-12)[15\]](#page-13-13).

<span id="page-1-0"></span>

**Figure 1.** Classification of metallothionein. This figure was drawn by Figdraw [\(www.figdraw.com](www.figdraw.com) accessed on 27 June 2024).

## *2.2. Structure*

Apoproteins do not form any typical secondary structures when they do not bind to metal ions [\[16\]](#page-13-14). Metallothionein acquires its structure only after binding to metals, and its tertiary structure depends on the nature and quantity of the metal ions [\[17\]](#page-13-15). Metallothionein is linked to metals mainly through the thiol group in cysteine residues, resulting in the formation of metallothionein clusters with a strong binding capacity for metal ions such as  $Cu^{2+}$  and  $Zn^{2+}$  [\[18](#page-13-16)[,19\]](#page-13-17). Mammalian metallothionein has a relatively unique 3D structure, containing two independent structural domains that combine to form a dumbbell-like structure [\[20\]](#page-13-18). The N-terminus is the  $\beta$  structural domain, and the C-terminus is the  $\alpha$ structural domain [\[21\]](#page-13-19). This structure is also regarded as the model for metallothionein (Figure [2\)](#page-2-0) [\[22\]](#page-13-20).

<span id="page-2-0"></span>

**Figure 2.** Biological functions of metallothionein. The NMR structure of the α-domain and β-domain of rat MT-2. The models were generated with UCSF ChimeraX [\(https://www.rbvi.ucsf.edu/chimerax](https://www.rbvi.ucsf.edu/chimerax) accessed on 27 June 2024) using RCSB PDB [\(https://www.rcsb.org/](https://www.rcsb.org/) accessed on 27 June 2024) coordinates of 1MRT and 2MRT. The α-domain has four metal ion binding sites, and the β-domain has three metal ion binding sites.

## **3. Biological Functions of Metallothionein**

Because of its high sulfhydryl content and potent metal ion binding capacity, metallothionein is beneficial in a variety of biological processes [\[4\]](#page-13-3). As was already mentioned, several studies have demonstrated that metallothionein has different functions, including the detoxification of heavy metals, antioxidant activity, neuroprotection, anticancer activity, and anti-inflammatory effects (Figure [3\)](#page-2-1).

<span id="page-2-1"></span>

**Figure 3.** Biological functions of metallothionein. This figure was drawn by Figdraw [\(www.figdraw.](www.figdraw.com) [com](www.figdraw.com) accessed on 27 June 2024).

## *3.1. Detoxification of Heavy Metals*

The ability to chelate metal ions is attributed to the abundance of cysteine residues with sulfhydryls present in metallothionein [\[5\]](#page-13-4). Table [1](#page-3-0) lists studies related to the detoxification of different metal ions by metallothionein. The chelation of metallothionein with metal ions is one of the most crucial pathways for metal detoxification in living cells [\[23\]](#page-14-0). Its degradation occurs in the liver after binding to metal ions [\[24\]](#page-14-1). When ZnMT is degraded, zinc is rapidly released from it, which continues to induce the production of new metallothionein. CuMT, on the other hand, undergoes oxidation to form insoluble polymers that accumulate in lysosomes and eventually enter the bile [\[25\]](#page-14-2).

Cadmium poisoning can cause damage to organs such as the liver and kidneys, trigger osteoporosis, increase the risk of cancer, affect the immune system, and trigger metabolic disorders [\[26\]](#page-14-3). Metallothionein was initially discovered in cadmium-rich environments and found to be bound to cadmium [\[1\]](#page-13-0). It has been shown that metallothionein plays an important role in cadmium detoxification. Studies have demonstrated that *E. coli* cells expressing metallothionein exhibit improved growth compared with those without metallothionein expression when cultured under different cadmium concentrations, indicating the significance of metallothionein cadmium detoxification [\[27\]](#page-14-4). The introduction of surface-engineered metallothionein bacteria into rice-growing soil can improve plant height and spike length, lower the cadmium concentration in cadmium-contaminated rice hulls, roots, shoots, and other parts of the rice, and significantly reduce cadmium toxicity [\[28\]](#page-14-5). Expression of the *PtMT2b* gene from *Populus trichocarpa* using *S. cerevisiae* effectively increased the yeast's tolerance to  $Cd^{2+}$  until the  $Cd^{2+}$  concentration reached 50  $\mu$ M, at which point its growth was completely inhibited [\[29\]](#page-14-6). Expression of IaMT in *E. coli* and determination of cell growth and  $Cd^{2+}$  accumulation under a  $Cd^{2+}$  environment revealed that IaMT was able to both enhance the tolerance of *E. coli* to  $Cd^{2+}$  and also increase the amount of intracellular  $Cd^{2+}$ accumulation and improve *E. coli's* tolerance to  $Cd^{2+}$  [\[30\]](#page-14-7). A sub-acute concentration of CdCl<sub>2</sub> solution was injected into grass carp, and MT was administered as an antidote four days later. It was discovered that the grass carp's blood and kidney Cd levels significantly decreased after receiving an MT injection. Simultaneously, MT significantly decreased blood apoptosis, mitigated the hemoglobin decrease brought on by a Cd injection, and assisted in lessening the immune system's reaction to Cd [\[31\]](#page-14-8).

Furthermore, metallothionein exhibits detoxifying effects against various other heavy metal elements. Metallothionein can lessen the harmful effects of  $As<sup>3+</sup>$  on cells, as well as the disruption of phospholipid metabolism and damaged cell membranes caused by  $As^{3+}$ . Metallothionein chelates  $As^{3+}$  by forming a complex with  $As^{3+}$  and scavenges ROS, thus alleviating the toxic effects of  $As^{3+}$  [\[32\]](#page-14-9). ShMT, a metallothionein in freshwater crabs expressed in *E. coli*, has a strong binding ability to Zn, Cu, and Cd ions, with the order of affinity being  $Cu > Cd > Zn$  [\[33\]](#page-14-10). The chelating action of metallothionein on metal ions led to a significant increase in Zn<sup>2+</sup> and Cd<sup>2+</sup> accumulation in tobacco when the human metallothionein *HsMT1L* gene was expressed in tobacco, improving tolerance to Zn<sup>2+</sup> and Cd<sup>2+</sup>. *HsMT1L* is capable of removing heavy metal ions and plays a role in protecting plants from heavy metal toxicity [\[34\]](#page-14-11).

<b>Metallothionein Source</b>	<b>Target of Action</b>	Type of Metal Ion	Effect	Reference
Anabaena PCC 7120 NmtA	E. coli cells	$Cd^{2+}$	NmtA-expressing E. coli exhibits better growth at certain cadmium concentrations	$[27]$
Metallothionein expressed by Alishewanella sp. WH16-1-MT	Rice	$Cd^{2+}$	Increased plant height, spike length, and thousand-grain weight of rice, resulting in a significant reduction in Cd <sup>2+</sup> content in brown rice, rice husk, roots, and shoots	[28]
S. cerevisiae expresses the PtMT2b gene from Populus trichocarpa	S. cerevisiae	$Cd^{2+}$	Enhanced $Cd^{2+}$ tolerance in S. cerevisiae	[29]
Ipomoea aquatica metallothionein IaMT expressed in E. coli	E. coli	$Cd^{2+}$	Increased tolerance to and accumulation of $Cd^{2+}$ in E. coli	$[30]$
Rabbit liver MT-2	Grass carp	$Cd^{2+}$	Reduced cadmium levels in kidneys and blood, attenuating organ damage	$[31]$

<span id="page-3-0"></span>**Table 1.** Studies on the role of metallothionein in heavy metal detoxification.



## **Table 1.** *Cont.*

## *3.2. Antioxidant Effect*

Free radicals are produced during normal human metabolic processes and are also induced by exposure to radiation, ozone, and air pollutants [\[37\]](#page-14-14). When there is an imbalance between the production of free radicals and antioxidant defenses, it can lead to oxidative damage to nucleic acids, proteins, and lipids in the body, resulting in oxidative stress, which has an impact on the function of the body's antioxidant system [\[38](#page-14-15)[,39\]](#page-14-16). Oxidative stress triggers a variety of diseases, including inflammatory diseases, neurological disorders, cardiovascular diseases, cancer, and more [\[37\]](#page-14-14). Sulfhydryl groups serve as efficient scavengers of free radicals, making them preferred targets for their neutralization [\[40\]](#page-14-17). Metallothionein, as one of the major sources of thiols in cells, contains cysteine residues that can reduce oxidative damage by both preventing the generation of ROS and helping to quench it in vivo [\[40–](#page-14-17)[42\]](#page-14-18). Metallothionein can also provide metal cofactors for some antioxidant enzymes to exert antioxidant effects [\[43\]](#page-14-19). Metallothionein exhibits potent antioxidant properties, effectively scavenging various free radicals, such as hydroxyl and superoxide radicals. Its reaction rate constant with hydroxyl radicals is approximately 300 times higher than that of glutathione [\[6\]](#page-13-5). In addition, metallothionein has about 50-fold higher antioxidant activity against oxidative DNA damage and about 10-fold higher antioxidant activity against lipid peroxidation than glutathione [\[44\]](#page-14-20). MTF-1 and Nrf2 would regulate metallothionein expression by activating the ARE in the promoter region, while Nrf2 and its downstream antioxidant genes may also be regulated by metallothionein [\[45\]](#page-14-21). Several studies have confirmed the antioxidant effects of metallothionein (Table [2\)](#page-4-0).

<span id="page-4-0"></span>**Table 2.** Studies on the antioxidant effect of metallothionein.





**Table 2.** *Cont.*

The ability of *E. coli* to express GST-AmMT2 increased the bacteria's tolerance to  $H_2O_2$ , which may be explained by the fact that GST-AmMT2 increases CAT activity [\[46\]](#page-14-22). When metallothionein knockout mice and normal mice were subjected to intermittent hypoxic conditions, the former showed reduced lung structure damage and a shorter duration of oxidative stress [\[45\]](#page-14-21). Furthermore, another experiment using a knocked-out mouse metallothionein gene showed that when PPE was injected into the trachea of mice, almost no ROS was produced in the lungs of normal mice, but a higher amount was produced in the lungs of mice with the metallothionein gene knocked out [\[47\]](#page-14-23). The addition of rh-MT-III to oxidatively damaged *Caenorhabditis elegans* nematodes revealed restoration of motility, reduction of malondialdehyde and reactive oxygen species levels, enhancement of the antioxidant defense system, and alleviation of the extent of oxidative damage [\[48\]](#page-14-24). A certain degree of improvement in the tolerance of yeast to oxidative stress was observed when the *PdMT2A* gene was expressed, leading to an increase in the tolerance of yeast to H2O2. Expression of the *PdMT2A* gene in transgenic *Arabidopsis* seedlings produced seedlings that were considerably more SOD-active and more resistant to oxidative stress than normal plants [\[42\]](#page-14-18). The effect of MT3 on ROS production was examined when osteoblasts were differentiated in C2C12 cells. The findings demonstrated that when MT3 was overexpressed, ROS production was suppressed, and when MT3 was silenced, ROS production increased [\[49\]](#page-15-0). In HT1376 cells, MT2A knockdown raised endogenous ROS levels. In contrast, overexpression of MT2A inhibited  $H_2O_2$ -induced ROS production in the cells [\[50\]](#page-15-1). ROS levels are elevated during the early stages of adipose differentiation. Overexpression of MT3 reduces ROS levels. When MT3 expression was knocked down, the level of ROS was significantly increased [\[51\]](#page-15-2). LPS induces oxidative damage in the heart, mainly by increasing the production of superoxide anion radicals and reducing glutathione levels. The presence of metallothionein attenuates the degree of oxidative stress induced by LPS [\[52\]](#page-15-3). Overexpression of *GmMT-II* resulted in a significant increase in SOD, CAT, and POD activities in all transgenic *Arabidopsis* lines under high temperature and humidity stress [\[53\]](#page-15-4). When *LcMT3* was expressed in *Arabidopsis*, the amount of MDA and ROS decreased and the expression of SOD, CAT, and POD increased [\[54\]](#page-15-5).

#### *3.3. Neuroprotective Effect*

Alzheimer's disease is a type of dementia that is prevalent in the elderly and is characterized by the deposition of amyloid-β peptide  $(Aβ)$ , forming localized amyloidosis [\[55\]](#page-15-6). Aβ plaques are rich in metal ions, such as zinc and copper, which may induce the aggregation of Aβ peptides, leading to Alzheimer's disease [\[56\]](#page-15-7). The main pathological hallmark of Parkinson's disease is the accumulation of Lewy bodies, whose main protein component is α-synuclein (α-syn) [\[7\]](#page-13-6). Increased levels of metal ions trigger the misfolding of α-syn, which causes the formation of Lewy bodies and Parkinson's disease [\[57\]](#page-15-8). Metallothionein has a strong metal ion binding capacity, is able to reduce metal ion-induced neurotoxicity, and thus may play an important role in the treatment of diseases such as Alzheimer's disease and Parkinson's disease [\[7,](#page-13-6)[55\]](#page-15-6).

Prolonged metallothionein action resulted in decreased toxicity of A $\beta$  and  $\alpha$ -syn in aged transgenic Caenorhabditis elegans [\[58\]](#page-15-9).  $A\beta_{1-42}$  leads to increased  $Zn^{2+}$  content in dentate granule cells and impairs hippocampus-dependent memory. Dexamethasone injection increases metallothionein expression and maintains  $Zn^{2+}$  homeostasis [\[59\]](#page-15-10). The formation of α-syn-Cu (II) complexes catalyzes toxic reactions and accelerates neuronal death. The addition of  $Zn<sub>7</sub>MT-3$  removes Cu (II) and effectively inhibits the toxic effects of  $\alpha$ -syn-Cu (II) [\[60\]](#page-15-11).

Numerous other studies have demonstrated the neuroprotective properties of metallothionein in addition to these two disease-related investigations. Several studies on the neuroprotective effects of metallothionein have been listed in Table [3.](#page-6-0) Paraquat causes brain damage in zebrafish. Treatment with hMT2 has been shown to mitigate the negative effects of paraquat, including decreased lipid peroxidation and dopaminergic neurons [\[61\]](#page-15-12). Overexpression of MT3 in astrocytes resulted in lower levels of glutamate in the culture medium compared with control cells, demonstrating that MT3 expression enhances the buffering capacity of astrocytes against glutamate, thereby reducing the neurotoxicity caused by glutamate [\[62\]](#page-15-13). Rat spinal motor neurons were shown to benefit neuroprotectively from the addition of zonisamide, which was able to increase the expression of astrocyte metallothionein 2A and lessen oxidative stress-induced astrocyte damage [\[8\]](#page-13-7). Wild mice chronically exposed to an active volcano environment had high levels of heavy metal elements in their brains and were detected to have higher levels of MT-2A than normal mice. By chelating heavy metal ions, metallothionein mitigates the damage that volcanic pollutants cause to the central nervous system [\[63\]](#page-15-14). Injection of isoproterenol into mice resulted in increased synthesis of metallothionein, which helped to reduce  $A\beta_{1-42}$ -induced toxicity [\[64\]](#page-15-15).

<span id="page-6-0"></span>**Table 3.** Studies on the neuroprotective effect of metallothionein.



#### *3.4. Anticancer Effect*

As mentioned above, metallothionein has the functions of heavy metal detoxification and reduction of oxidative stress damage, which can enhance the ability of normal cells to resist external unfavorable conditions, thus having an anti-cancer effect [\[9\]](#page-13-8). At the same time, metallothionein can inhibit cancer cell growth, migration, and invasion and can induce cell cycle arrest in cancer cells, leading to apoptosis [\[65\]](#page-15-16). Studies on the anticancer effects of metallothionein have been listed in Table [4.](#page-7-0)

Furthermore, metallothionein has the effect of detoxifying heavy metals and reducing oxidative stress damage, which can enhance the adaptability of cells to anticancer drugs and alter cancer cells' resistance to chemotherapy, both of which have an anticancer effect.

Evaluation of metallothionein expression in feline injection site fibrosarcoma revealed that there was a negative correlation between the tumor grade and the level of inflammation. Cells are protected from oxidative stress damage by metallothionein, and its down-regulation may raise the risk of DNA damage and, consequently, cancer risk [\[9\]](#page-13-8). In esophageal squamous cell carcinoma cells, overexpression of MT1M induces apoptosis, reduces cell viability, and inhibits epithelial-mesenchymal transition, thus acting as a tumor suppressor [\[66\]](#page-15-17). Cannabidiol was able to exert a therapeutic effect on colorectal cancer, and overexpression of MT1G and MT2A increased the number of dead cells and synergistically enhanced the anticancer effect of cannabidiol [\[67\]](#page-15-18). Overexpression of MT1E in hepatocellular carcinoma cells resulted in a significant decrease in both cell viability and cell number. Following MT1E knockdown, there was a notable decrease in the quantity of apoptotic cells [\[68\]](#page-15-19). Overexpression of MT1M inhibited the proliferation, migration, and invasion of gastric cancer cells and promoted apoptosis of gastric cancer cells. Meanwhile, MT1M may also play an anti-cancer role by decreasing the stemness of gastric cancer cells and increasing their sensitivity to 5-fluorouracil [\[69\]](#page-15-20). MT1G inhibited the proliferation, migration, and invasion of hepatocellular carcinoma cells in both in vivo and in vitro experiments. When overexpressed, it also demonstrated synergistic inhibition with sorafenib. When the expression of MT1G was disrupted, the effect of sorafenib was attenuated, indicating that MT1G was involved in the process of sorafenib action [\[70\]](#page-15-21). Overexpression of MT-1 can delay the progression of hepatocellular carcinoma. Decreased expression of MT-1 can also assist in the diagnosis of hepatocellular carcinoma to a certain extent [\[71\]](#page-15-22). Overexpression of MT2A inhibits proliferation and migration of colorectal cancer cells as well as growth and metastasis of cancer cells [\[72\]](#page-16-0).

Metallothionein can also play a role in the treatment of cancer by influencing chemotherapy resistance. There is a problem of resistance to gemcitabine in the treatment of pancreatic ductal adenocarcinoma, and MT1G is able to limit the secretion of activin A and inhibit pancreatic ductal adenocarcinoma cell stemness, thus overcoming the resistance to gemcitabine [\[73\]](#page-16-1).

In addition, metallothionein has a tendency to chelate with chemotherapy medications, lowering the medications' toxicity to cancer cells [\[74\]](#page-16-2). Malignant pleural mesothelioma can be successfully treated with platinum-based chemotherapy; however, metallothionein has the potential to bind to cisplatin and make it inactive. Knockdown of MT2A expression in malignant pleural mesothelioma cell lines resulted in a significant increase in apoptosis in response to cisplatin. Thus, inhibition of MT2A expression can significantly improve the therapeutic effect of cisplatin [\[75\]](#page-16-3). Chemotherapy resistance is a major challenge in the treatment of osteosarcoma. Four chosen osteosarcoma cell lines exhibit increased sensitivity to several chemotherapeutic agents when MT2A is silenced, which promotes the effectiveness of chemotherapeutic agents [\[76\]](#page-16-4). Carbon monoxide reduces levels of metallothionein, which reduces drug resistance in ovarian cancer cells. The therapeutic effect of cisplatin can be enhanced by decreasing the expression of metallothionein [\[77\]](#page-16-5).

<span id="page-7-0"></span>**Table 4.** Studies on the anticancer effect of metallothionein.





# **Table 4.** *Cont.*

# *3.5. Anti-Inflammatory Effect*

Any response that damages the body results in inflammation. Although the inflammatory process is normally self-limiting, intervention is needed to control acute inflammation and return the immune system to homeostasis [\[78\]](#page-16-6). The primary mechanism by which metallothionein exerts its anti-inflammatory properties is through its capacity to scavenge reactive oxygen species. Furthermore, it functions as a zinc chaperone, thereby triggering matrix metalloproteinases that facilitate tissue repair and regeneration during inflammatory conditions [\[10\]](#page-13-9). Inflammation can activate metallothionein expression through a variety of pathways, such as stimulation of antioxidant-responsive elements in promoter regions, specific metal-responsive elements, activation of the second messenger protein kinase pathway, and so on [\[79\]](#page-16-7). Several studies have reported the anti-inflammatory effects of metallothionein (Table [5\)](#page-8-0).

<span id="page-8-0"></span>**Table 5.** Studies on the anti-inflammatory effect of metallothionein.



# **Table 5.** *Cont.*



Studies on synovial cells isolated from patients with osteoarthritis revealed that MT-1 significantly reduced the expression of inflammatory cytokines such as TNF- $\alpha$  and IL-6 $\beta$ and exerted anti-inflammatory effects [\[80\]](#page-16-8). Overexpression of MT1 in the mouse liver in a CDAHFD mouse model resulted in the downregulation of genes related to inflammation and fibrosis, such as timp-1, coll1, ten- $\alpha$ , and mcp-1. It was demonstrated that MT1 could alleviate liver fibrosis and improve non-alcoholic steatohepatitis to a certain extent [\[81\]](#page-16-9). Naringenin was able to reduce the expression of pro-inflammatory cytokines by inducing MT1G expression and exerting an inhibitory effect on NF-κB activation. Proinflammatory cytokine expression rises after MT1G is knocked down [\[78\]](#page-16-6). Zinc supplementation improved the treatment of colitis in a mouse model of the disease by upregulating the expression of MT1 and MT2, regulating intestinal inflammation in terms of intestinal epithelial integrity, the immune system, metabolic function, and the defense against ox-idative stress [\[82\]](#page-16-10). When  $As^{3+}$  causes an inflammatory response in carp gills,  $Zn^{2+}$  can cause metallothionein to be expressed, prevent the NF-κB signaling pathway from being activated, decrease the number of inflammatory factors secreted, and ultimately stop the inflammatory response [\[83\]](#page-16-11). MT1 expression was similarly up-regulated during CuL5's anti-inflammatory effect on microglia, suggesting a potential correlation between MT1

and CuL5's anti-inflammatory effect [\[79\]](#page-16-7). During clearance of gram-negative bacterial infections, the non-canonical inflammasome of mouse caspase-11 is activated, but this process also causes severe inflammatory injury. MT3 expression increases  $Zn^{2+}$  levels, inhibits caspase-11 signaling through the TRIF-IRF3-STAT1 axis, and controls inflammation development [\[84\]](#page-16-12). Polysaccharides extracted from *Plantago asiatica* L. seeds were able to protect against inflammatory liver injury in mice to a certain extent. This resulted in an increase in metallothionein levels, which may have an anti-inflammatory effect by scavenging excess ROS generated during LPS-induced liver injury [\[85\]](#page-16-13). MT-1 levels were elevated in patients with ankylosing spondylitis and positively correlated with ankylosing spondylitis activity and inflammatory response, suggesting that MT-1 may be involved in defense systems against inflammatory processes [\[86\]](#page-16-14). An inflammatory response occurs during pre-eclampsia, and supplementation of zinc gluconate to pre-eclamptic rats increased MT levels, which significantly reduced pro-inflammatory cytokine levels and helped to alleviate the inflammatory response [\[87\]](#page-16-15). Exosomes from mesenchymal stromal cells were able to up-regulate the expression of MT-2, elevate the transcriptional level of I $\kappa$ B $\alpha$  in macrophages of mice with colitis, and inhibit the activation of NF- $\kappa$ B. This increased macrophage resistance to inflammation, which in turn facilitated the treatment of colitis [\[88\]](#page-16-16). Significantly elevated levels of Mt1 and Mt2 in alcoholic hepatitis mice reduce levels of lipid peroxides such as 4-hydroxynonenal and malondialdehyde and decrease the activation of stress kinases [\[89\]](#page-16-17). MT-1 and MT-2 inhibit the formation of osteoclasts and prevent osteoporosis and other damage caused by rheumatoid arthritis [\[90\]](#page-16-18). The ethyl acetate fraction of *Amomum villosum* var. *xanthioides* alleviates non-alcoholic steatohepatitis by enhancing antioxidant capacity and improving oxidative status through increased MT1 expression [\[91\]](#page-16-19). In addition, MT-1 is able to play an immunomodulatory role by regulating the proliferation and differentiation of immune cells and acting as a chemoattractant to regulate cellular infiltration, thus effectively alleviating the inflammatory response [\[92\]](#page-16-20).

# **4. Applications of Metallothionein**

In view of the multiple biological functions of metallothionein, it has a broad application prospect in environment, medicine, food and so on (Figure [4\)](#page-10-0).

<span id="page-10-0"></span>

**Figure 4.** Applications of metallothionein. This figure was drawn by Figdraw [\(www.figdraw.com](www.figdraw.com) accessed on 27 June 2024).

#### *4.1. Detection and Removal of Heavy Metal Ions from the Environment*

Metallothionein can be used as a biomarker for aquatic organisms exposed to heavy metal ions because its gene expression is stimulated by various heavy metal ions and can react at an early stage of environmental pollution [\[93\]](#page-16-21). Preparation of fish metallothioneinspecific polyclonal antibodies enables the detection of changes in metallothionein levels in the livers of a wide range of fish species, delivering early warning signs before heavy metal ion pollution of aquatic ecosystems occurs [\[93\]](#page-16-21). Metallothionein in plankton is also able to reflect the degree of contamination of organisms with heavy metal ions, which can be used in the assessment of environmental quality [\[94\]](#page-17-0). Fish levels of metallothionein increased significantly as a result of heavy metal ion pollution in the waters; hence, changes in fish levels of metallothionein can be used to track heavy metal ion pollution in the waters [\[95\]](#page-17-1).

Metallothionein demonstrates a good ability to bind heavy metal ions. When used in combination with other materials, it can remove heavy metal ions from the environment, thereby reducing the pollution associated with heavy metal ions [\[96\]](#page-17-2). Novel biosorbents were constructed using cellulose, metallothionein, and carbohydrate-binding modules, which can effectively adsorb  $Pb^{2+}$  and  $Zn^{2+}$  from polluted water and play a great role in removing toxic trace elements from water [\[97\]](#page-17-3). The synthesized metallothionein SmtAmodified selenium nanoparticles were able to efficiently adsorb and stably remove  $Cd^{2+}$ and  $Pb^{2+}$  and reduce cadmium and lead residues in wastewater to meet the national wastewater discharge standards [\[96\]](#page-17-2). After being genetically modified and co-assembled with magnetic nanoparticles, the metallothionein gene was introduced into E. coli and successfully removed  $Pb^{2+}$  and  $Cd^{2+}$ , with a removal efficiency of >80% [\[98\]](#page-17-4). Cloning of MT2A and MT3 into *E. coli* was effective in removing Cr<sup>6+</sup> from an aqueous solution, and the main functioning groups were the hydroxyl, phosphoryl, and carbonyl groups [\[99\]](#page-17-5). Encapsulation of MT3-expressing *E. coli* in the form of calcium alginate bio-beads was able to effectively remove three heavy metals, copper, zinc, and cadmium, from water, with the most significant removal effect on copper [\[100\]](#page-17-6). Dissolved metal ions in mine wastewater are difficult to remove. *Escherichia coli* overexpressing MTA were able to efficiently absorb nickel from wastewater, with a seven-fold increase in the ability to bioaccumulate nickel compared with the control [\[101\]](#page-17-7).

#### *4.2. Disease Prediction and Diagnosis in Medicine*

In the field of medicine, it is possible to predict and diagnose disease by detecting changes in the content of metallothionein. Metallothionein has various functions, including antioxidants, heavy metal detoxification, and maintenance of metal ion homeostasis. Its content tends to change during the occurrence of diseases to counteract the adverse state of the organism [\[102\]](#page-17-8).

Genes like MT1E and MT1F can be used as biomarkers to predict the recurrence of hepatocellular carcinoma. Reduced expression of metallothionein may raise the risk of cancer, as evidenced by the down-regulation of these genes in patient samples with hepatocellular carcinoma [\[103\]](#page-17-9). MT-1 is able to predict the development of schizophrenia to a certain extent. The risk of schizophrenia is elevated when the level of MT-1 is reduced, probably due to the lack of metallothionein, which leads to an increased risk of oxidative damage in the body, thus raising the risk of schizophrenia [\[102\]](#page-17-8). Wilson disease is an inherited disorder of copper metabolism in which the liver and brain are affected by copper toxicity. Since metallothionein can detoxify copper, the expression of metallothionein is increased in the tissues of patients with Wilson disease. Positive metallothionein immunostaining has been demonstrated to be a useful tool in the diagnosis of Wilson disease [\[104\]](#page-17-10). The currently available prognostic indicators of hepatocellular carcinoma, such as tumor number and cell differentiation, have many limitations in the treatment of therapy and prognosis. To increase prognostic accuracy, multi-indicator prediction using oxidative stress biomarkers, like MT-3, can be used [\[105\]](#page-17-11). When colorectal cancer occurs, the levels of MT1B, MT1F, and MT1G genes are down-regulated, and MT genes may be able to serve as effective markers for the diagnosis of the disease. In addition, high expression levels of the MT1B, MT1H,

and MT1L genes are associated with a good prognosis in colorectal cancer patients and can predict the prognosis of the disease [\[106\]](#page-17-12). The greater the MT1X expression in patients with clear cell renal cell carcinoma, the greater the likelihood of both highly graded tumors and metastasizing tumors. Therefore, MT1X can predict tumorigenesis to some extent and predict the prognosis of clear cell renal cell carcinoma [\[107\]](#page-17-13). Patients with gastric cancer had down-regulated levels of metallothionein expression, and there was a significant positive correlation between MT mRNA and overall survival (OS), first-progression survival (FP), and post-progression survival (PPS). These results have the potential to predict the prognostic status of gastric cancer patients [\[108\]](#page-17-14). MT1JP expression was down-regulated in glioma patients. Glioma patients with high MTIJP expression had a longer survival time relative to those with low expression. Thus, the expression of MTIJP can provide some reference for the diagnosis of glioma [\[109\]](#page-17-15).

#### *4.3. Development of Products with Skincare Functions*

Metallothionein has many functions, such as anti-inflammatory, oxidative damage relief, anti-ultraviolet radiation, heavy metal detoxification, post-sunburn repair, accelerated wound healing, etc. The development of skincare products or functional foods with metallothionein as the active ingredient will be widely used in the field of skincare.

The expression of metallothionein genes was significantly elevated in dermatitis patients, and MT1X expression was able to reduce the inflammatory response triggered by allergens. At the same time, MTF1 expression can prevent allergen-induced oxidative stress, thus providing a protective effect on the skin [\[110\]](#page-17-16). By binding zinc in the cytoplasm, metallothionein can control zinc homeostasis and delay the onset of the hand-foot skin reaction. Zinc ions also protect the skin from oxidative damage by inducing the synthesis of metallothionein, a preferred target of oxidant attack [\[111\]](#page-17-17). UV radiation causes damage to the DNA of living organisms and also produces reactive oxygen species, leading to apoptosis and senescence. By stimulating the synthesis of metallothionein, 1,25-dihydroxyvitamin D3 can decrease the formation of sun-damaged cells and improve the skin's resistance to UV light [\[112\]](#page-17-18). After irradiating the skin with sun-simulated UV radiation and applying skin care products containing isoflavonoids, epidermal cell MT expression increased. Metallothionein exerts photoprotective effects mainly by scavenging free radicals [\[113\]](#page-17-19). When the back skin of mice lacking metallothionein was exposed to UVB radiation, the number of sun-damaged cells increased significantly, and the skin thickness was significantly higher than in control mice [\[114\]](#page-17-20). The skin was repaired after exposure to the sun, and the expression of MT1A was elevated. Metallothionein has a strong antioxidant capacity, and the elevated expression may be related to its involvement in skin cell repair after exposure to the sun [\[115\]](#page-17-21). When substances containing cadmium are applied to the skin, cadmium is readily absorbed into the skin and poses a health hazard. Cadmium causes apoptosis in keratin-forming cells, and curcumin is able to protect cells from cadmium toxicity damage by up-regulating the expression of MT2A and binding to cadmium [\[116\]](#page-17-22). After skin damage, the newly growing epidermis and skin cells at the wound edge express more metallothionein than does the surrounding normal skin. The increased expression of metallothionein may contribute to the enhancement of the anti-inflammatory activity of zinc and the enhancement of MMP-1 expression to promote the migration of keratinocytes, thus contributing to the healing of skin wounds [\[117\]](#page-17-23).

## **5. Conclusions**

Metallothionein plays a very important role in heavy metal detoxification, antioxidants, neuroprotection, anticancer, and anti-inflammatory activities due to its high metal ion binding capacity and high sulfhydryl content. The diverse biological functions of metallothionein also give it a broad application prospect in the fields of environmental protection, disease diagnosis and treatment, and skin care. However, the structure of metallothionein varies greatly among different species, and the metal ion binding state of metallothionein in vivo cannot be identified, resulting in the function of a particular metallothionein and

the mechanism of its functioning being unclear, which needs to be further investigated in the future.

**Author Contributions:** Conceptualization, R.Y., P.L. and N.S.; investigation, R.Y., B.G., P.L. and N.S.; writing—original draft preparation, R.Y. and D.R.; writing—review and editing, P.L. and N.S.; supervision, B.G., P.L. and N.S.; funding acquisition, P.L. and N.S. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Key R&D Program of China, grant number 2020YFD1000300 and Beijing Fishery Innovation Team, grant number BAIC07-2023-13.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## **References**

- <span id="page-13-0"></span>1. Margoshes, M.; Vallee, B.L. A Cadmium Protein from Equine Kidney Cortex. *J. Am. Chem. Soc.* **1957**, *79*, 4813–4814. [\[CrossRef\]](https://doi.org/10.1021/ja01574a064)
- <span id="page-13-1"></span>2. Jamrozik, D.; Dutczak, R.; Machowicz, J.; Wojtyniak, A.; Smedowski, A.; Pietrucha-Dutczak, M. Metallothioneins, a Part of the Retinal Endogenous Protective System in Various Ocular Diseases. *Antioxidants* **2023**, *12*, 1251. [\[CrossRef\]](https://doi.org/10.3390/antiox12061251) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37371981)
- <span id="page-13-2"></span>3. Krizkova, S.; Kepinska, M.; Emri, G.; Eckschlager, T.; Stiborova, M.; Pokorna, P.; Heger, Z.; Adam, V. An insight into the complex roles of metallothioneins in malignant diseases with emphasis on (sub)isoforms/isoforms and epigenetics phenomena. *Pharmacol. Ther.* **2018**, *183*, 90–117. [\[CrossRef\]](https://doi.org/10.1016/j.pharmthera.2017.10.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28987322)
- <span id="page-13-3"></span>4. Coyle, P.; Philcox, J.C.; Carey, L.C.; Rofe, A.M. Metallothionein: The multipurpose protein. *Cell Mol. Life Sci.* **2002**, *59*, 627–647. [\[CrossRef\]](https://doi.org/10.1007/s00018-002-8454-2)
- <span id="page-13-4"></span>5. Liu, Y.; Wu, Z.; Guo, K.; Zhou, Y.; Xing, K.; Zheng, J.; Sun, Y.; Zhang, J. Metallothionein-1 gene from *Exopalaemon carinicauda* and its response to heavy metal ions challenge. *Mar. Pollut. Bull.* **2022**, *175*, 113324. [\[CrossRef\]](https://doi.org/10.1016/j.marpolbul.2022.113324) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35051848)
- <span id="page-13-5"></span>6. Miyazaki, I.; Asanuma, M. Multifunctional Metallothioneins as a Target for Neuroprotection in Parkinson's Disease. *Antioxidants* **2023**, *12*, 894. [\[CrossRef\]](https://doi.org/10.3390/antiox12040894)
- <span id="page-13-6"></span>7. McLeary, F.A.; Rcom-H'cheo-Gauthier, A.N.; Goulding, M.; Radford, R.A.W.; Okita, Y.; Faller, P.; Chung, R.S.; Pountney, D.L. Switching on Endogenous Metal Binding Proteins in Parkinson's Disease. *Cells* **2019**, *8*, 179. [\[CrossRef\]](https://doi.org/10.3390/cells8020179) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30791479)
- <span id="page-13-7"></span>8. Kanbara, S.; Ohkawara, B.; Nakashima, H.; Ohta, K.; Koshimizu, H.; Inoue, T.; Tomita, H.; Ito, M.; Masuda, A.; Ishiguro, N.; et al. Zonisamide ameliorates progression of cervical spondylotic myelopathy in a rat model. *Sci. Rep.* **2020**, *10*, 13138. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-70068-0)
- <span id="page-13-8"></span>9. Mikiewicz, M.; Pazdzior-Czapula, K.; Fiedorowicz, J.; Gesek, M.; Otrocka-Domagala, I. Metallothionein expression in feline injection site fibrosarcomas. *BMC Vet. Res.* **2023**, *19*, 42. [\[CrossRef\]](https://doi.org/10.1186/s12917-023-03604-5)
- <span id="page-13-9"></span>10. Jose, D.; Allen, A.L.; Blakley, B.; Al-Dissi, A. Evaluation of metallothionein and Ki-67 expression in chronic cholangiohepatitis in cats. *Can. J. Vet. Res.* **2021**, *85*, 36–44.
- <span id="page-13-10"></span>11. Binz, P.A.; Kägi, J.H.R. Metallothionein: Molecular evolution and classification. In *Metallothionein IV*; Springer: Basel, Switzerland, 1999; pp. 7–13.
- <span id="page-13-11"></span>12. Alvarez-Barrios, A.; Alvarez, L.; Garcia, M.; Artime, E.; Pereiro, R.; Gonzalez-Iglesias, H. Antioxidant Defenses in the Human Eye: A Focus on Metallothioneins. *Antioxidants* **2021**, *10*, 89. [\[CrossRef\]](https://doi.org/10.3390/antiox10010089) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33440661)
- <span id="page-13-12"></span>13. Thirumoorthy, N.; Manisenthil Kumar, K.T.; Shyam Sundar, A.; Panayappan, L.; Chatterjee, M. Metallothionein: An overview. *World J. Gastroenterol.* **2007**, *13*, 993–996. [\[CrossRef\]](https://doi.org/10.3748/wjg.v13.i7.993) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17373731)
- 14. Moffatt, P.; Seguin, C. Expression of the gene encoding metallothionein-3 in organs of the reproductive system. *DNA Cell Biol.* **1998**, *17*, 501–510. [\[CrossRef\]](https://doi.org/10.1089/dna.1998.17.501) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9655243)
- <span id="page-13-13"></span>15. Quaife, C.J.; Findley, S.D.; Erickson, J.C.; Froelick, G.J.; Kelly, E.J.; Zambrowicz, B.P.; Palmiter, R.D. Induction of a new metallothionein isoform (MT-IV) occurs during differentiation of stratified squamous epithelia. *Biochemistry* **1994**, *33*, 7250–7259. [\[CrossRef\]](https://doi.org/10.1021/bi00189a029) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8003488)
- <span id="page-13-14"></span>16. Sutherland, D.E.; Stillman, M.J. Challenging conventional wisdom: Single domain metallothioneins. *Metallomics* **2014**, *6*, 702–728. [\[CrossRef\]](https://doi.org/10.1039/C3MT00216K) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24469686)
- <span id="page-13-15"></span>17. Ziller, A.; Fraissinet-Tachet, L. Metallothionein diversity and distribution in the tree of life: A multifunctional protein. *Metallomics* **2018**, *10*, 1549–1559. [\[CrossRef\]](https://doi.org/10.1039/C8MT00165K) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30229264)
- <span id="page-13-16"></span>18. Stankovic, R.K.; Chung, R.S.; Penkowa, M. Metallothioneins I and II: Neuroprotective significance during CNS pathology. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 484–489. [\[CrossRef\]](https://doi.org/10.1016/j.biocel.2006.09.010)
- <span id="page-13-17"></span>19. Sakulsak, N. Metallothionein: An Overview on its Metal Homeostatic Regulation in Mammals. *Int. J. Morphol.* **2012**, *30*, 1007–1012. [\[CrossRef\]](https://doi.org/10.4067/S0717-95022012000300039)
- <span id="page-13-18"></span>20. Braun, W.; Vasak, M.; Robbins, A.H.; Stout, C.D.; Wagner, G.; Kagi, J.H.; Wuthrich, K. Comparison of the NMR solution structure and the x-ray crystal structure of rat metallothionein-2. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 10124–10128. [\[CrossRef\]](https://doi.org/10.1073/pnas.89.21.10124)
- <span id="page-13-19"></span>21. Juarez-Rebollar, D.; Rios, C.; Nava-Ruiz, C.; Mendez-Armenta, M. Metallothionein in Brain Disorders. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 5828056. [\[CrossRef\]](https://doi.org/10.1155/2017/5828056)
- <span id="page-13-20"></span>22. Chan, J. Studies of metal binding reactions in metallothioneins by spectroscopic, molecular biology, and molecular modeling techniques. *Coord. Chem. Rev.* **2002**, *233–234*, 319–339. [\[CrossRef\]](https://doi.org/10.1016/S0010-8545(02)00176-5)
- <span id="page-14-0"></span>23. Vijver, M.G.; Van Gestel, C.A.; Lanno, R.P.; Van Straalen, N.M.; Peijnenburg, W.J. Internal metal sequestration and its ecotoxicological relevance: A review. *Environ. Sci. Technol.* **2004**, *38*, 4705–4712. [\[CrossRef\]](https://doi.org/10.1021/es040354g) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15487776)
- <span id="page-14-1"></span>24. Bremner, I. Nutritional and physiological significance of metallothionein. *Exp. Suppl.* **1987**, *52*, 81–107. [\[CrossRef\]](https://doi.org/10.1007/978-3-0348-6784-9_5)
- <span id="page-14-2"></span>25. Richards, M.P. Recent developments in trace element metabolism and function: Role of metallothionein in copper and zinc metabolism. *J. Nutr.* **1989**, *119*, 1062–1070. [\[CrossRef\]](https://doi.org/10.1093/jn/119.7.1062)
- <span id="page-14-3"></span>26. Järup, L.; Åkesson, A. Current status of cadmium as an environmental health problem. *Toxicol. Appl. Pharm.* **2009**, *238*, 201–208. [\[CrossRef\]](https://doi.org/10.1016/j.taap.2009.04.020)
- <span id="page-14-4"></span>27. T, V.D.; Chandwadkar, P.; Acharya, C. NmtA, a novel metallothionein of *Anabaena* sp. strain PCC 7120 imparts protection against cadmium stress but not oxidative stress. *Aquat. Toxicol.* **2018**, *199*, 152–161. [\[CrossRef\]](https://doi.org/10.1016/j.aquatox.2018.03.035)
- <span id="page-14-5"></span>28. Yu, Y.; Shi, K.; Li, X.; Luo, X.; Wang, M.; Li, L.; Wang, G.; Li, M. Reducing cadmium in rice using metallothionein surfaceengineered bacteria WH16-1-MT. *Environ. Res.* **2022**, *203*, 111801. [\[CrossRef\]](https://doi.org/10.1016/j.envres.2021.111801) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34339701)
- <span id="page-14-6"></span>29. De Oliveira, V.H.; Ullah, I.; Dunwell, J.M.; Tibbett, M. Bioremediation potential of Cd by transgenic yeast expressing a metallothionein gene from *Populus trichocarpa*. *Ecotoxicol. Environ. Saf.* **2020**, *202*, 110917. [\[CrossRef\]](https://doi.org/10.1016/j.ecoenv.2020.110917)
- <span id="page-14-7"></span>30. Huang, Y.-Y.; Gong, F.-Y.; Shen, C.; He, C.-T.; Fu, H.-L.; Wang, X.-S.; Tan, X.; Xu, P.-L.; Yang, Z.-Y. Cloning, characterization and expression analysis of metallothioneins from Ipomoea aquatica and their cultivar-dependent roles in Cd accumulation and detoxification. *Ecotoxicol. Environ. Saf.* **2018**, *165*, 450–458. [\[CrossRef\]](https://doi.org/10.1016/j.ecoenv.2018.08.089) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30218968)
- <span id="page-14-8"></span>31. Huang, X.; Xiong, G.; Feng, Y.; Fan, W.; Yang, S.; Duan, J.; Duan, Y.; Wang, K.; Ou, Y.; Rehman, T.; et al. Protective effects of metallothionein and vitamin E in the trunk kidney and blood of cadmium poisoned *Ctenopharyngodon idellus*. *Fish. Physiol. Biochem.* **2020**, *46*, 1053–1061. [\[CrossRef\]](https://doi.org/10.1007/s10695-020-00771-2)
- <span id="page-14-9"></span>32. Qi, Z.; Wang, Q.; Wang, H.; Tan, M. Metallothionein Attenuated Arsenic-Induced Cytotoxicity: The Underlying Mechanism Reflected by Metabolomics and Lipidomics. *J. Agric. Food Chem.* **2021**, *69*, 5372–5380. [\[CrossRef\]](https://doi.org/10.1021/acs.jafc.1c00724) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33939412)
- <span id="page-14-10"></span>33. He, Y.; Wang, L.; Ma, W.; Lu, X.; Li, Y.; Liu, J. Secretory expression, immunoaffinity purification and metal-binding ability of recombinant metallothionein (ShMT) from freshwater crab Sinopotamon henanense. *Ecotoxicol. Environ. Saf.* **2019**, *169*, 457–463. [\[CrossRef\]](https://doi.org/10.1016/j.ecoenv.2018.11.065) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30472469)
- <span id="page-14-11"></span>34. Zheng, Y.; Cui, M.; Ni, L.; Qin, Y.; Li, J.; Pan, Y.; Zhang, X. Heterologous Expression of Human Metallothionein Gene *HsMT1L* Can Enhance the Tolerance of Tobacco (*Nicotiana nudicaulis* Watson) to Zinc and Cadmium. *Genes* **2022**, *13*, 2413. [\[CrossRef\]](https://doi.org/10.3390/genes13122413)
- <span id="page-14-12"></span>35. Zou, C.; Chen, Y.; Li, H.; Li, W.; Wei, J.; Li, Z.; Wang, X.; Chen, T.; Huang, H. Engineered Bacteria EcN-MT Alleviate Liver Injury in Cadmium-Exposed Mice via its Probiotics Characteristics and Expressing of Metallothionein. *Front. Pharmacol.* **2022**, *13*, 857869. [\[CrossRef\]](https://doi.org/10.3389/fphar.2022.857869) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35281910)
- <span id="page-14-13"></span>36. Tsyganov, V.E.; Tsyganova, A.V.; Gorshkov, A.P.; Seliverstova, E.V.; Kim, V.E.; Chizhevskaya, E.P.; Belimov, A.A.; Serova, T.A.; Ivanova, K.A.; Kulaeva, O.A.; et al. Efficacy of a Plant-Microbe System: *Pisum sativum* (L.) Cadmium-Tolerant Mutant and Rhizobium leguminosarum Strains, Expressing Pea Metallothionein Genes PsMT1 and PsMT2, for Cadmium Phytoremediation. *Front. Microbiol.* **2020**, *11*, 15. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2020.00015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32063892)
- <span id="page-14-14"></span>37. Lobo, V.; Patil, A.; Phatak, A.; Chandra, N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn. Rev.* **2010**, *4*, 118–126. [\[CrossRef\]](https://doi.org/10.4103/0973-7847.70902)
- <span id="page-14-15"></span>38. Paithankar, J.G.; Saini, S.; Dwivedi, S.; Sharma, A.; Chowdhuri, D.K. Heavy metal associated health hazards: An interplay of oxidative stress and signal transduction. *Chemosphere* **2021**, *262*, 128350. [\[CrossRef\]](https://doi.org/10.1016/j.chemosphere.2020.128350) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33182141)
- <span id="page-14-16"></span>39. McCord, J.M. The evolution of free radicals and oxidative stress. *Am. J. Med.* **2000**, *108*, 652–659. [\[CrossRef\]](https://doi.org/10.1016/S0002-9343(00)00412-5)
- <span id="page-14-17"></span>40. Thornalley, P.J.; Vasak, M. Possible role for metallothionein in protection against radiation-induced oxidative stress. Kinetics and mechanism of its reaction with superoxide and hydroxyl radicals. *Biochim. Biophys. Acta* **1985**, *827*, 36–44. [\[CrossRef\]](https://doi.org/10.1016/0167-4838(85)90098-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2981555)
- 41. Eibl, J.K.; Abdallah, Z.; Ross, G.M. Zinc-metallothionein: A potential mediator of antioxidant defence mechanisms in response to dopamine-induced stress. *Can. J. Physiol. Pharmacol.* **2010**, *88*, 305–312. [\[CrossRef\]](https://doi.org/10.1139/Y10-022)
- <span id="page-14-18"></span>42. Patankar, H.V.; Al-Harrasi, I.; Al Kharusi, L.; Jana, G.A.; Al-Yahyai, R.; Sunkar, R.; Yaish, M.W. Overexpression of a Metallothionein 2A Gene from Date Palm Confers Abiotic Stress Tolerance to Yeast and *Arabidopsis thaliana*. *Int. J. Mol. Sci.* **2019**, *20*, 2871. [\[CrossRef\]](https://doi.org/10.3390/ijms20122871) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31212812)
- <span id="page-14-19"></span>43. Lazo, J.S.; Pitt, B.R. Metallothioneins and cell death by anticancer drugs. *Annu. Rev. Pharmacol. Toxicol.* **1995**, *35*, 635–653. [\[CrossRef\]](https://doi.org/10.1146/annurev.pa.35.040195.003223)
- <span id="page-14-20"></span>44. Bensellam, M.; Laybutt, D.R.; Jonas, J.C. Emerging Roles of Metallothioneins in Beta Cell Pathophysiology: Beyond and Above Metal Homeostasis and Antioxidant Response. *Biology* **2021**, *10*, 176. [\[CrossRef\]](https://doi.org/10.3390/biology10030176) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33652748)
- <span id="page-14-21"></span>45. Lin, X.; Jagadapillai, R.; Cai, J.; Cai, L.; Shao, G.; Gozal, E. Metallothionein induction attenuates the progression of lung injury in mice exposed to long-term intermittent hypoxia. *Inflamm. Res.* **2020**, *69*, 15–26. [\[CrossRef\]](https://doi.org/10.1007/s00011-019-01287-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31707449)
- <span id="page-14-22"></span>46. Babaei-Bondarti, Z.; Shahpiri, A. A metallothionein type 2 from Avicennia marina binds to iron and mediates hydrogen peroxide balance by activation of enzyme catalase. *Phytochemistry* **2020**, *176*, 112396. [\[CrossRef\]](https://doi.org/10.1016/j.phytochem.2020.112396) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32353553)
- <span id="page-14-23"></span>47. Tanaka, K.I.; Shiota, S.; Sakakibara, O.; Shimoda, M.; Takafuji, A.; Takabatake, M.; Kadota, Y.; Kawakami, T.; Suzuki, S.; Kawahara, M. Exacerbation of Elastase-Induced Emphysema via Increased Oxidative Stress in Metallothionein-Knockout Mice. *Biomolecules* **2022**, *12*, 583. [\[CrossRef\]](https://doi.org/10.3390/biom12040583) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35454172)
- <span id="page-14-24"></span>48. Sun, Z.; Qin, J.; Yuan, H.; Guo, M.; Shang, M.; Niu, S.; Li, Y.; Li, Q.; Xue, Y. Recombinant human metallothionein-III alleviates oxidative damage induced by copper and cadmium in *Caenorhabditis elegans*. *J. Appl. Toxicol.* **2023**, *43*, 1242–1252. [\[CrossRef\]](https://doi.org/10.1002/jat.4460) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36918407)
- <span id="page-15-0"></span>49. Li, S.; Kim, M.J.; Lee, S.H.; Jin, L.; Cong, W.; Jeong, H.G.; Lee, K.Y. Metallothionein 3 Promotes Osteoblast Differentiation in C2C12 Cells via Reduction of Oxidative Stress. *Int. J. Mol. Sci.* **2021**, *22*, 4312. [\[CrossRef\]](https://doi.org/10.3390/ijms22094312)
- <span id="page-15-1"></span>50. Sung, H.C.; Chang, K.S.; Chen, Y.T.; Hsu, S.Y.; Lin, Y.H.; Hou, C.P.; Feng, S.H.; Tsui, K.H.; Juang, H.H. Metallothionein 2A with Antioxidant and Antitumor Activity Is Upregulated by Caffeic Acid Phenethyl Ester in Human Bladder Carcinoma Cells. *Antioxidants* **2022**, *11*, 1509. [\[CrossRef\]](https://doi.org/10.3390/antiox11081509) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36009228)
- <span id="page-15-2"></span>51. Li, Y.; Lee, S.H.; Piao, M.; Kim, H.S.; Lee, K.Y. Metallothionein 3 Inhibits 3T3-L1 Adipocyte Differentiation via Reduction of Reactive Oxygen Species. *Antioxidants* **2023**, *12*, 640. [\[CrossRef\]](https://doi.org/10.3390/antiox12030640) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36978888)
- <span id="page-15-3"></span>52. Chen, Y.; Zhao, J.; Ye, H.; Ceylan-Isik, A.F.; Zhang, B.; Liu, Q.; Yang, Y.; Dong, M.; Luo, B.; Ren, J. Beneficial impact of cardiac heavy metal scavenger metallothionein in sepsis-provoked cardiac anomalies dependent upon regulation of endoplasmic reticulum stress and ferroptosis but not autophagy. *Life Sci.* **2024**, *336*, 122291. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2023.122291) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38030060)
- <span id="page-15-4"></span>53. Liu, S.; Liu, Y.; Liu, C.; Li, Y.; Zhang, F.; Ma, H. Isolation and Characterization of the GmMT-II Gene and Its Role in Response to High Temperature and Humidity Stress in *Glycine max*. *Plants* **2022**, *11*, 1503. [\[CrossRef\]](https://doi.org/10.3390/plants11111503) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35684276)
- <span id="page-15-5"></span>54. Chen, Y.; Zhu, W.; Deng, H.; Pei, X.; Zhang, J.; Liu, J.; Ma, P. Heterologous expression of the Leymus chinensis metallothionein gene LcMT3 confers enhanced tolerance to salt stress in *Escherichia coli*, yeast, and *Arabidopsis thaliana*. *J. Plant Physiol.* **2023**, *287*, 154022. [\[CrossRef\]](https://doi.org/10.1016/j.jplph.2023.154022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37301036)
- <span id="page-15-6"></span>55. Zar˛eba, N.; Kepinska, M. The Function of Transthyretin Complexes with Metallothionein in Alzheimer's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 9003. [\[CrossRef\]](https://doi.org/10.3390/ijms21239003)
- <span id="page-15-7"></span>56. Smith, D.P.; Ciccotosto, G.D.; Tew, D.J.; Fodero-Tavoletti, M.T.; Johanssen, T.; Masters, C.L.; Barnham, K.J.; Cappai, R. Concentration dependent  $Cu^{2+}$  induced aggregation and dityrosine formation of the Alzheimer's disease amyloid-beta peptide. *Biochemistry* **2007**, *46*, 2881–2891. [\[CrossRef\]](https://doi.org/10.1021/bi0620961)
- <span id="page-15-8"></span>57. Santner, A.; Uversky, V.N. Metalloproteomics and metal toxicology of alpha-synuclein. *Metallomics* **2010**, *2*, 378–392. [\[CrossRef\]](https://doi.org/10.1039/b926659c) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21072383)
- <span id="page-15-9"></span>58. Pretsch, D.; Rollinger, J.M.; Schmid, A.; Genov, M.; Wohrer, T.; Krenn, L.; Moloney, M.; Kasture, A.; Hummel, T.; Pretsch, A. Prolongation of metallothionein induction combats Ass and alpha-synuclein toxicity in aged transgenic *Caenorhabditis elegans*. *Sci. Rep.* **2020**, *10*, 11707. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-68561-7)
- <span id="page-15-10"></span>59. Tamano, H.; Suzuki, H.; Murakami, T.; Fujii, H.; Adlard, P.A.; Bush, A.I.; Takeda, A. Amyloid beta(1-42)-Induced Rapid Zn(2+) Influx into Dentate Granule Cells Attenuates Maintained LTP Followed by Retrograde Amnesia. *Mol. Neurobiol.* **2019**, *56*, 5041–5050. [\[CrossRef\]](https://doi.org/10.1007/s12035-018-1429-6)
- <span id="page-15-11"></span>60. Calvo, J.S.; Mulpuri, N.V.; Dao, A.; Qazi, N.K.; Meloni, G. Membrane insertion exacerbates the alpha-Synuclein-Cu(II) dopamine oxidase activity: Metallothionein-3 targets and silences all alpha-synuclein-Cu(II) complexes. *Free Radic. Biol. Med.* **2020**, *158*, 149–161. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2020.07.006)
- <span id="page-15-12"></span>61. Mohamad Najib, N.H.; Yahaya, M.F.; Das, S.; Teoh, S.L. The effects of metallothionein in paraquat-induced Parkinson disease model of zebrafish. *Int. J. Neurosci.* **2023**, *133*, 822–833. [\[CrossRef\]](https://doi.org/10.1080/00207454.2021.1990916) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34623211)
- <span id="page-15-13"></span>62. Paryani, F.; Kwon, J.S.; Ng, C.W.; Madden, N.; Ofori, K.; Tang, A.; Lu, H.; Li, J.; Mahajan, A.; Davidson, S.M.; et al. Multi-OMIC analysis of Huntington disease reveals a neuroprotective astrocyte state. *bioRxiv* **2023**. [\[CrossRef\]](https://doi.org/10.1101/2023.09.08.556867)
- <span id="page-15-14"></span>63. Navarro-Sempere, A.; Martinez-Peinado, P.; Rodrigues, A.S.; Garcia, P.V.; Camarinho, R.; Grindlay, G.; Gras, L.; Garcia, M.; Segovia, Y. Metallothionein expression in the central nervous system in response to chronic heavy metal exposure: Possible neuroprotective mechanism. *Environ. Geochem. Health* **2023**, *45*, 8257–8269. [\[CrossRef\]](https://doi.org/10.1007/s10653-023-01722-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37580456)
- <span id="page-15-15"></span>64. Kawano, Y.; Tamura, K.; Egawa, M.; Tamano, H.; Takeda, A. Isoproterenol, an adrenergic beta receptor agonist, induces metallothionein synthesis followed by canceling amyloid beta(1-42)-induced neurodegeneration. *Biometals* **2022**, *35*, 303–312. [\[CrossRef\]](https://doi.org/10.1007/s10534-022-00365-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35064350)
- <span id="page-15-16"></span>65. Kwon, I.S.; Hwang, Y.N.; Park, J.H.; Na, H.H.; Kwon, T.H.; Park, J.S.; Kim, K.C. Metallothionein Family Proteins as Regulators of Zinc Ions Synergistically Enhance the Anticancer Effect of Cannabidiol in Human Colorectal Cancer Cells. *Int. J. Mol. Sci.* **2023**, *24*, 16621. [\[CrossRef\]](https://doi.org/10.3390/ijms242316621) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38068944)
- <span id="page-15-17"></span>66. Li, D.; Peng, W.; Wu, B.; Liu, H.; Zhang, R.; Zhou, R.; Yao, L.; Ye, L. Metallothionein MT1M Suppresses Carcinogenesis of Esophageal Carcinoma Cells through Inhibition of the Epithelial-Mesenchymal Transition and the SOD1/PI3K Axis. *Mol. Cells* **2021**, *44*, 267–278. [\[CrossRef\]](https://doi.org/10.14348/molcells.2021.2179)
- <span id="page-15-18"></span>67. Zhou, L.; Deng, X.; Xiao, X.; Liao, Y.; Chen, W.; Dai, Q. Kruppel-like factor 9 inhibits growth and metastasis of cholangiocarcinoma cells by targeted regulation of metallothionein 1 M transcription. *Tissue Cell* **2022**, *79*, 101962. [\[CrossRef\]](https://doi.org/10.1016/j.tice.2022.101962)
- <span id="page-15-19"></span>68. Liu, Q.; Lu, F.; Chen, Z. Identification of MT1E as a novel tumor suppressor in hepatocellular carcinoma. *Pathol. Res. Pr.* **2020**, *216*, 153213. [\[CrossRef\]](https://doi.org/10.1016/j.prp.2020.153213)
- <span id="page-15-20"></span>69. Li, K.; Sun, S.; Lu, Y.; Liang, W.; Xu, X.; Zhang, H.; Chang, Z.; Wang, C.; Gao, Y.; Chen, L. MT1M regulates gastric cancer progression and stemness by modulating the Hedgehog pathway protein GLI1. *Biochem. Biophys. Res. Commun.* **2023**, *670*, 63–72. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2023.05.121)
- <span id="page-15-21"></span>70. Wei, T.; Lin, R.; Fu, X.; Lu, Y.; Zhang, W.; Li, Z.; Zhang, J.; Wang, H. Epigenetic regulation of the DNMT1/MT1G/KLF4/CA9 axis synergises the anticancer effects of sorafenib in hepatocellular carcinoma. *Pharmacol. Res.* **2022**, *180*, 106244. [\[CrossRef\]](https://doi.org/10.1016/j.phrs.2022.106244) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35550167)
- <span id="page-15-22"></span>71. Wu, Y.J.; Ko, B.S.; Liang, S.M.; Lu, Y.J.; Jan, Y.J.; Jiang, S.S.; Shyue, S.K.; Chen, L.; Liou, J.Y. ZNF479 downregulates metallothionein-1 expression by regulating ASH2L and DNMT1 in hepatocellular carcinoma. *Cell Death Dis.* **2019**, *10*, 408. [\[CrossRef\]](https://doi.org/10.1038/s41419-019-1651-9)
- <span id="page-16-0"></span>72. Liu, X.; Quan, J.; Shen, Z.; Zhang, Z.; Chen, Z.; Li, L.; Li, X.; Hu, G.; Deng, X. Metallothionein 2A (MT2A) controls cell proliferation and liver metastasis by controlling the MST1/LATS2/YAP1 signaling pathway in colorectal cancer. *Cancer Cell Int.* **2022**, *22*, 205. [\[CrossRef\]](https://doi.org/10.1186/s12935-022-02623-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35642057)
- <span id="page-16-1"></span>73. Li, K.; Zhang, Z.; Mei, Y.; Yang, Q.; Qiao, S.; Ni, C.; Yao, Y.; Li, X.; Li, M.; Wei, D.; et al. Metallothionein-1G suppresses pancreatic cancer cell stemness by limiting activin A secretion via NF-kappaB inhibition. *Theranostics* **2021**, *11*, 3196–3212. [\[CrossRef\]](https://doi.org/10.7150/thno.51976) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33537082)
- <span id="page-16-2"></span>74. Merlos Rodrigo, M.A.; Jimenez Jimemez, A.M.; Haddad, Y.; Bodoor, K.; Adam, P.; Krizkova, S.; Heger, Z.; Adam, V. Metallothionein isoforms as double agents—Their roles in carcinogenesis, cancer progression and chemoresistance. *Drug Resist. Updat.* **2020**, *52*, 100691. [\[CrossRef\]](https://doi.org/10.1016/j.drup.2020.100691) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32615524)
- <span id="page-16-3"></span>75. Borchert, S.; Suckrau, P.M.; Walter, R.F.H.; Wessolly, M.; Mairinger, E.; Steinborn, J.; Hegedus, B.; Hager, T.; Herold, T.; Eberhardt, W.E.E.; et al. Impact of metallothionein-knockdown on cisplatin resistance in malignant pleural mesothelioma. *Sci. Rep.* **2020**, *10*, 18677. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-75807-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33122816)
- <span id="page-16-4"></span>76. Mangelinck, A.; da Costa, M.E.M.; Stefanovska, B.; Bawa, O.; Polrot, M.; Gaspar, N.; Fromigue, O. MT2A is an early predictive biomarker of response to chemotherapy and a potential therapeutic target in osteosarcoma. *Sci. Rep.* **2019**, *9*, 12301. [\[CrossRef\]](https://doi.org/10.1038/s41598-019-48846-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31444479)
- <span id="page-16-5"></span>77. Kawahara, B.; Ramadoss, S.; Chaudhuri, G.; Janzen, C.; Sen, S.; Mascharak, P.K. Carbon monoxide sensitizes cisplatin-resistant ovarian cancer cell lines toward cisplatin via attenuation of levels of glutathione and nuclear metallothionein. *J. Inorg. Biochem.* **2019**, *191*, 29–39. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2018.11.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30458366)
- <span id="page-16-6"></span>78. Yang, J.; Liu, L.; Li, M.; Huang, X.; Yang, H.; Li, K. Naringenin inhibits pro-inflammatory cytokine production in macrophages through inducing MT1G to suppress the activation of NF-kappaB. *Mol. Immunol.* **2021**, *137*, 155–162. [\[CrossRef\]](https://doi.org/10.1016/j.molimm.2021.07.003)
- <span id="page-16-7"></span>79. Choo, X.Y.; McInnes, L.E.; Grubman, A.; Wasielewska, J.M.; Belaya, I.; Burrows, E.; Quek, H.; Martin, J.C.; Loppi, S.; Sorvari, A.; et al. Novel Anti-Neuroinflammatory Properties of a Thiosemicarbazone-Pyridylhydrazone Copper(II) Complex. *Int. J. Mol. Sci.* **2022**, *23*, 722. [\[CrossRef\]](https://doi.org/10.3390/ijms231810722) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36142627)
- <span id="page-16-8"></span>80. Wang, C.; Gong, Z.; Hu, S.; Zhang, G. Metallothionein-1 is associated with osteoarthritis disease activity and suppresses proinflammatory cytokines production in synovial cells. *Int. Immunopharmacol.* **2019**, *75*, 105815. [\[CrossRef\]](https://doi.org/10.1016/j.intimp.2019.105815) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31465913)
- <span id="page-16-9"></span>81. Li, X.; Zhong, S.; Sun, Y.; Huang, X.; Li, Y.; Wang, L.; Wu, Y.; Yang, M.; Yuan, H.X.; Liu, J.; et al. Integration analysis identifies the role of metallothionein in the progression from hepatic steatosis to steatohepatitis. *Front. Endocrinol.* **2022**, *13*, 951093. [\[CrossRef\]](https://doi.org/10.3389/fendo.2022.951093)
- <span id="page-16-10"></span>82. Foligne, B.; George, F.; Standaert, A.; Garat, A.; Poiret, S.; Peucelle, V.; Ferreira, S.; Sobry, H.; Muharram, G.; Lucau-Danila, A.; et al. High-dose dietary supplementation with zinc prevents gut inflammation: Investigation of the role of metallothioneins and beyond by transcriptomic and metagenomic studies. *FASEB J.* **2020**, *34*, 12615–12633. [\[CrossRef\]](https://doi.org/10.1096/fj.202000562RR) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32729971)
- <span id="page-16-11"></span>83. Lu, H.; Zhao, H.; Wang, Y.; Guo, M.; Mu, M.; Liu, Y.; Nie, X.; Huang, P.; Xing, M. Arsenic (III) induces oxidative stress and inflammation in the gills of common carp, which is ameliorated by zinc (II). *J. Inorg. Biochem.* **2021**, *225*, 111617. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2021.111617) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34571403)
- <span id="page-16-12"></span>84. Chowdhury, D.; Gardner, J.C.; Satpati, A.; Nookala, S.; Mukundan, S.; Porollo, A.; Landero Figueroa, J.A.; Subramanian Vignesh, K. Metallothionein 3-Zinc Axis Suppresses Caspase-11 Inflammasome Activation and Impairs Antibacterial Immunity. *Front. Immunol.* **2021**, *12*, 755961. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2021.755961) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34867993)
- <span id="page-16-13"></span>85. Li, F.; Huang, D.; Nie, S.; Xie, M. Polysaccharide from the Seeds of *Plantago asiatica* L. Protect Against Lipopolysaccharide-Induced Liver Injury. *J. Med. Food* **2019**, *22*, 1058–1066. [\[CrossRef\]](https://doi.org/10.1089/jmf.2018.4394) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31560611)
- <span id="page-16-14"></span>86. Ma, Y.; Du, J.; Yin, Z.; Dai, H.; Wei, Y.; Xia, Y.; Li, L.; Ye, Z.; Huang, Z. Metallothionein-1 is Positively Correlated with Inflammation and Ankylosing Spondylitis Activity. *J. Inflamm. Res.* **2022**, *15*, 5935–5944. [\[CrossRef\]](https://doi.org/10.2147/JIR.S382827) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36274830)
- <span id="page-16-15"></span>87. Lang, Q.; Wei, J.; Tian, M.; Wei, S.; Yu, X.; Zhao, C.; Zhang, J.; Huang, B. Attenuated effect of zinc gluconate on oxidative stress, inflammation, and angiogenic imbalance in pre-eclampsia rats. *Life Sci.* **2022**, *310*, 121055. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2022.121055) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36228770)
- <span id="page-16-16"></span>88. Liu, H.; Liang, Z.; Wang, F.; Zhou, C.; Zheng, X.; Hu, T.; He, X.; Wu, X.; Lan, P. Exosomes from mesenchymal stromal cells reduce murine colonic inflammation via a macrophage-dependent mechanism. *JCI Insight* **2019**, *4*, e131273. [\[CrossRef\]](https://doi.org/10.1172/jci.insight.131273)
- <span id="page-16-17"></span>89. Ma, J.; Cao, H.; Rodrigues, R.M.; Xu, M.; Ren, T.; He, Y.; Hwang, S.; Feng, D.; Ren, R.; Yang, P.; et al. Chronic-plusbinge alcohol intake induces production of proinflammatory mtDNA-enriched extracellular vesicles and steatohepatitis via ASK1/p38MAPKalpha-dependent mechanisms. *JCI Insight* **2020**, *5*, e136496. [\[CrossRef\]](https://doi.org/10.1172/jci.insight.136496)
- <span id="page-16-18"></span>90. Royzman, D.; Andreev, D.; Stich, L.; Peckert-Maier, K.; Wild, A.B.; Zinser, E.; Muhl-Zurbes, P.; Jones, E.; Adam, S.; Frey, S.; et al. The soluble CD83 protein prevents bone destruction by inhibiting the formation of osteoclasts and inducing resolution of inflammation in arthritis. *Front. Immunol.* **2022**, *13*, 936995. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.936995)
- <span id="page-16-19"></span>91. Cho, J.H.; Lee, J.S.; Kim, H.G.; Lee, H.W.; Fang, Z.; Kwon, H.H.; Kim, D.W.; Lee, C.M.; Jeong, J.W. Ethyl Acetate Fraction of Amomum villosum var. xanthioides Attenuates Hepatic Endoplasmic Reticulum Stress-Induced Non-Alcoholic Steatohepatitis via Improvement of Antioxidant Capacities. *Antioxidants* **2021**, *10*, 998. [\[CrossRef\]](https://doi.org/10.3390/antiox10070998)
- <span id="page-16-20"></span>92. Subramanian Vignesh, K.; Deepe, G., Jr. Metallothioneins: Emerging Modulators in Immunity and Infection. *Int. J. Mol. Sci.* **2017**, *18*, 2197. [\[CrossRef\]](https://doi.org/10.3390/ijms18102197)
- <span id="page-16-21"></span>93. Nagamatsu, P.C.; Vargas, D.A.R.; Prodocimo, M.M.; Opuskevitch, I.; Ferreira, F.; Zanchin, N.; de Oliveira Ribeiro, C.A.; de Souza, C. Synthetic fish metallothionein design as a potential tool for monitoring toxic metals in water. *Environ. Sci. Pollut. Res. Int.* **2021**, *28*, 9517–9528. [\[CrossRef\]](https://doi.org/10.1007/s11356-020-11427-2)
- <span id="page-17-0"></span>94. Strogyloudi, E.; Paraskevopoulou, V.; Campillo, J.A.; Zervoudaki, S.; Bouga, V.; Catsiki, V.A.; Dassenakis, E.; Krasakopoulou, E. Metal and metallothionein levels in zooplankton in relation to environmental exposure: Spatial and temporal variability (Saronikos Gulf, Greece). *Environ. Sci. Pollut. Res. Int.* **2021**, *28*, 28640–28657. [\[CrossRef\]](https://doi.org/10.1007/s11356-021-12591-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33544340)
- <span id="page-17-1"></span>95. Freire, M.M.; Gomez, C.; Moreira, J.C.; Linde Arias, A.R. Multibiomarker approach in fish to assess a heavily polluted Brazilian estuary, Guanabara Bay. *Environ. Monit. Assess.* **2022**, *195*, 187. [\[CrossRef\]](https://doi.org/10.1007/s10661-022-10752-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36504393)
- <span id="page-17-2"></span>96. Zhou, Z.; Dong, Y.; Zhu, L.; Xia, X.; Li, S.; Wang, G.; Shi, K. Effective and stable adsorptive removal of Cadmium(II) and Lead(II) using selenium nanoparticles modified by microbial SmtA metallothionein. *Chemosphere* **2022**, *307*, 135818. [\[CrossRef\]](https://doi.org/10.1016/j.chemosphere.2022.135818) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35944684)
- <span id="page-17-3"></span>97. Mwandira, W.; Nakashima, K.; Togo, Y.; Sato, T.; Kawasaki, S. Cellulose-metallothionein biosorbent for removal of Pb(II) and Zn(II) from polluted water. *Chemosphere* **2020**, *246*, 125733. [\[CrossRef\]](https://doi.org/10.1016/j.chemosphere.2019.125733)
- <span id="page-17-4"></span>98. Zhu, N.; Zhang, B.; Yu, Q. Genetic Engineering-Facilitated Coassembly of Synthetic Bacterial Cells and Magnetic Nanoparticles for Efficient Heavy Metal Removal. *ACS Appl. Mater. Interfaces* **2020**, *12*, 22948–22957. [\[CrossRef\]](https://doi.org/10.1021/acsami.0c04512) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32338492)
- <span id="page-17-5"></span>99. Akkurt, S.; Oguz, M.; Alkan Uckun, A. Bioreduction and bioremoval of hexavalent chromium by genetically engineered strains (*Escherichia coli* MT2A and *Escherichia coli* MT3). *World J. Microbiol. Biotechnol.* **2022**, *38*, 45. [\[CrossRef\]](https://doi.org/10.1007/s11274-022-03235-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35075546)
- <span id="page-17-6"></span>100. Gupta, D.; Satpati, S.; Dixit, A.; Ranjan, R. Fabrication of biobeads expressing heavy metal-binding protein for removal of heavy metal from wastewater. *Appl. Microbiol. Biotechnol.* **2019**, *103*, 5411–5420. [\[CrossRef\]](https://doi.org/10.1007/s00253-019-09852-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31065755)
- <span id="page-17-7"></span>101. Diep, P.; Leo Shen, H.; Wiesner, J.A.; Mykytczuk, N.; Papangelakis, V.; Yakunin, A.F.; Mahadevan, R. Engineered nickel bioaccumulation in *Escherichia coli* by NikABCDE transporter and metallothionein overexpression. *Eng. Life Sci.* **2023**, *23*, 2200133. [\[CrossRef\]](https://doi.org/10.1002/elsc.202200133)
- <span id="page-17-8"></span>102. Yilmaz, S.; Kilic, N.; Kaya, S.; Tasci, G. A Potential Biomarker for Predicting Schizophrenia: Metallothionein-1. *Biomedicines* **2023**, *11*, 590. [\[CrossRef\]](https://doi.org/10.3390/biomedicines11020590)
- <span id="page-17-9"></span>103. Wang, S.; Gribskov, M. Transcriptome analysis identifies metallothionein as biomarkers to predict recurrence in hepatocellular cacinoma. *Mol. Genet. Genom. Med.* **2019**, *7*, e693. [\[CrossRef\]](https://doi.org/10.1002/mgg3.693)
- <span id="page-17-10"></span>104. Wiethoff, H.; Mohr, I.; Fichtner, A.; Merle, U.; Schirmacher, P.; Weiss, K.H.; Longerich, T. Metallothionein: A game changer in histopathological diagnosis of Wilson disease. *Histopathology* **2023**, *83*, 936–948. [\[CrossRef\]](https://doi.org/10.1111/his.15041)
- <span id="page-17-11"></span>105. Miao, C.; He, X.; Chen, G.; Kahlert, U.D.; Yao, C.; Shi, W.; Su, D.; Hu, L.; Zhang, Z. Seven oxidative stress-related genes predict the prognosis of hepatocellular carcinoma. *Aging* **2023**, *15*, 15050–15063. [\[CrossRef\]](https://doi.org/10.18632/aging.205330)
- <span id="page-17-12"></span>106. Hung, K.C.; Huang, T.C.; Cheng, C.H.; Cheng, Y.W.; Lin, D.Y.; Fan, J.J.; Lee, K.H. The Expression Profile and Prognostic Significance of Metallothionein Genes in Colorectal Cancer. *Int. J. Mol. Sci.* **2019**, *20*, 3849. [\[CrossRef\]](https://doi.org/10.3390/ijms20163849) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31394742)
- <span id="page-17-13"></span>107. Ding, Y.; Fang, J.; Chen, M.; Xu, Y.; Liu, N.; Fang, S.; Xiang, W.; Chen, R.; Wu, C.; Yu, H. MT1X is an oncogene and indicates prognosis in ccRCC. *Biosci. Rep.* **2022**, *42*, BSR20221128. [\[CrossRef\]](https://doi.org/10.1042/BSR20221128)
- <span id="page-17-14"></span>108. Tong, M.; Lu, W.; Liu, H.; Wu, J.; Jiang, M.; Wang, X.; Hao, J.; Fan, D. Evaluation of MT Family Isoforms as Potential Biomarker for Predicting Progression and Prognosis in Gastric Cancer. *Biomed. Res. Int.* **2019**, *2019*, 2957821. [\[CrossRef\]](https://doi.org/10.1155/2019/2957821) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31380415)
- <span id="page-17-15"></span>109. Chen, J.; Lou, J.; Yang, S.; Lou, J.; Liao, W.; Zhou, R.; Qiu, C.; Ding, G. MT1JP inhibits glioma progression via negative regulation of miR-24. *Oncol. Lett.* **2020**, *19*, 334–342. [\[CrossRef\]](https://doi.org/10.3892/ol.2019.11085)
- <span id="page-17-16"></span>110. Sirvent, S.; Vallejo, A.F.; Corden, E.; Teo, Y.; Davies, J.; Clayton, K.; Seaby, E.G.; Lai, C.; Ennis, S.; Alyami, R.; et al. Impaired expression of metallothioneins contributes to allergen-induced inflammation in patients with atopic dermatitis. *Nat. Commun.* **2023**, *14*, 2880. [\[CrossRef\]](https://doi.org/10.1038/s41467-023-38588-1)
- <span id="page-17-17"></span>111. Yeh, C.N.; Huang, W.K.; Lu, C.W.; Chen, C.P.; Lin, S.H.; Pan, Y.R.; Wu, C.E. A Potential Association of Zinc Deficiency and Tyrosine Kinase Inhibitor-Induced Hand-Foot Skin Reaction. *Biol. Trace Elem. Res.* **2023**, *201*, 5540–5545. [\[CrossRef\]](https://doi.org/10.1007/s12011-023-03618-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36892689)
- <span id="page-17-18"></span>112. Jagoda, S.V.; Dixon, K.M. Protective effects of 1,25 dihydroxyvitamin D(3) and its analogs on ultraviolet radiation-induced oxidative stress: A review. *Redox Rep.* **2020**, *25*, 11–16. [\[CrossRef\]](https://doi.org/10.1080/13510002.2020.1731261) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32093585)
- <span id="page-17-19"></span>113. Widyarini, S.; Allanson, M.; Gallagher, N.L.; Pedley, J.; Boyle, G.M.; Parsons, P.G.; Whiteman, D.C.; Walker, C.; Reeve, V.E. Isoflavonoid photoprotection in mouse and human skin is dependent on metallothionein. *J. Investig. Dermatol.* **2006**, *126*, 198–204. [\[CrossRef\]](https://doi.org/10.1038/sj.jid.5700013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16417237)
- <span id="page-17-20"></span>114. Wang, W.H.; Li, L.F.; Zhang, B.X.; Lu, X.Y. Metallothionein-null mice exhibit reduced tolerance to ultraviolet B injury in vivo. *Clin. Exp. Dermatol.* **2004**, *29*, 57–61. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2230.2004.01424.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14723724)
- <span id="page-17-21"></span>115. Tanaka, Y.; Parker, R.; Aganahi, A. Up-Regulated Expression of ICAM1, MT1A, PTGS2, LCE3D, PPARD, and GM-CSF2 Following Solar Skincare Protection and Repair Strategies in a 3-Dimensional Reconstructed Human Skin Model. *Clin. Cosmet. Investig. Dermatol.* **2023**, *16*, 2829–2839. [\[CrossRef\]](https://doi.org/10.2147/CCID.S428170) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37850108)
- <span id="page-17-22"></span>116. Wahyudi, L.D.; Yu, S.H.; Cho, M.K. The effect of curcumin on the cadmium-induced mitochondrial apoptosis pathway by metallothionein 2A regulation. *Life Sci.* **2022**, *310*, 121076. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2022.121076) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36243116)
- <span id="page-17-23"></span>117. Agren, M.S.; Chafranska, L.; Eriksen, J.O.; Forman, J.L.; Bjerrum, M.J.; Schjerling, P.; Larsen, H.F.; Cottarelli, E.; Jorgensen, L.N.; Gjerdrum, L.M.R. Spatial expression of metallothionein, matrix metalloproteinase-1 and Ki-67 in human epidermal wounds treated with zinc and determined by quantitative immunohistochemistry: A randomised double-blind trial. *Eur. J. Cell Biol.* **2021**, *100*, 151147. [\[CrossRef\]](https://doi.org/10.1016/j.ejcb.2020.151147)

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.