

*Review*



# **Coffee Bean and Its Chemical Constituent Caffeine and Chlorogenic Acid as Promising Chemoprevention Agents: Updated Biological Studies against Cancer Cells**

**Mohamed Aborziza <sup>1</sup> , Riezki Amalia [2](https://orcid.org/0000-0002-8445-7045) , Ade Zuhrotun <sup>3</sup> [,](https://orcid.org/0000-0002-7451-9118) Nur Kusaira Khairul Ikram <sup>4</sup> [,](https://orcid.org/0000-0003-4953-927X) Dhania Novitasari <sup>1</sup> and Muchtaridi Muchtaridi 1,5,[\\*](https://orcid.org/0000-0002-6156-8025)**

- <sup>1</sup> Department of Pharmaceutical Analysis and Medicinal Chemistry, Faculty of Pharmacy, Universitas Padjadjaran, Sumedang 45363, Indonesia; mohamed22002@mail.unpad.ac.id (M.A.); dhania@unpad.ac.id (D.N.)
- <sup>2</sup> Department of Pharmacology and Clinical Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Sumedang 45363, Indonesia; riezki.amalia@unpad.ac.id
- <sup>3</sup> Department of Biology Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Sumedang 45363, Indonesia; ade.zuhrotun@unpad.ac.id
- 4 Institute of Biological Sciences, Faculty of Science, Universiti Malaya, Kuala Lumpur 50603, Malaysia; nkusaira@um.edu.my
- <sup>5</sup> Research Collaboration Centre for Radiopharmaceuticals Theranostic, National Research and Innovation Agency (BRIN), Jln. Raya Bandung Sumedang Km. 21, Sumedang 45363, Indonesia
- **\*** Correspondence: muchtaridi@unpad.ac.id

**Abstract:** Cancer is a complicated and ever-evolving disease that remains a significant global cause of disease and mortality. Its complexity, which is evident at the genetic and phenotypic levels, contributes to its diversity and resistance to treatment. Numerous scientific investigations on human and animal models demonstrate the potential of phytochemicals in cancer prevention. Coffee has been shown to possess potent anti-carcinogenic properties, and studies have documented the consumption of coffee as a beverage reduces the risk of cancer occurrence. The major secondary metabolites of coffee, named caffeine and chlorogenic acid, have been linked to anti-inflammatory and antineoplastic effects through various signaling. In light of this, this review article provides a comprehensive analysis based on studies in anticancer effects of coffee, chlorogenic acid, and caffeine published between 2010 and 2023, sourced from Scopus, Pubmed, and Google Scholar databases. We summarize recent advances and scientific evidence on the association of phytochemicals found in coffee with a special emphasis on their biological activities against cancer and their molecular mechanism deemed potential to be used as a novel therapeutic target for cancer prevention and therapy.

**Keywords:** coffee; caffeine; chlorogenic acid; chemopreventive; autophagy; tumor cells

# **1. Introduction**

The development of novel and effective cancer medicines is critical, given the global expanding incidence of malignant diseases. The multifaceted nature of cancer, as evidenced at both the molecular and clinical levels, highlights its diversity and treatment resistance [\[1\]](#page-8-0). Despite the obstacle in developing cancer chemopreventive agents based on natural sources, there are still several promising pieces of evidence that support the evaluation of potential active natural products with regards to reducing or reversing the premalignant tissues [\[2\]](#page-8-1). Furthermore, the notion of cancer chemoprevention surfaced from anecdotal experience with nutritious meals, as well as epidemiological studies, with most of that focused on cancer treatment. For instance, people who consume plant-based foods are thought to have a lesser risk of cancer, showing increased interest in dietary phytochemical studies [\[3\]](#page-8-2).

Coffee, being one of the most widely consumed beverages worldwide [\[4\]](#page-9-0), has been shown to contain potent natural chemopreventive and antineoplastic agents [\[5,](#page-9-1)[6\]](#page-9-2). Coffee



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is derived from the berries of the *Coffea* sp., but only two species were considered for production, including *Coffea arabica* (Arabica) and *Coffea canephora* (Robusta). Apart from its renowned effect as a stimulant due to the high amount of caffeine, the bioactive compounds in coffee have been increasingly explored for other biological activities, from antioxidant to associated activities, including antiinflammation and anticancer [\[7\]](#page-9-3). These occurrences have caught the attention of health experts, given that coffee consumption has rapidly expanded over the last few decades due to greater prosperity and economic interest [\[8](#page-9-4)[,9\]](#page-9-5).

The chemical constituents of coffee beverages are mostly determined by the processing procedures (pre-roasting and roasting) used to prepare green coffee beans. Furthermore, harvesting methods and industrial processes for green coffee, as well as consumer ways for preparing coffee beverages, all contribute to variations in the concentration of particular substances in the final product  $[10]$ . Various factors such as coffee species, growing circumstances, harvesting methods, and processing procedures (such as high-temperature roasting) affect the amount of bioactive chemicals in coffee, such as antioxidants and biogenic amines [\[11,](#page-9-7)[12\]](#page-9-8). Coffee beans comprise an abundance of xanthine-based caf-feine, polyphenol chlorogenic acids, and tannins [\[10\]](#page-9-6), followed by other polyphenols and flavonoids which possess the antioxidant properties [\[13\]](#page-9-9). Numerous epidemiological studies have demonstrated that coffee consumption has been associated with potential health advantages due to its anti-inflammatory and chemopreventive activities. It has been proposed that the antioxidative properties of several coffee ingredients are responsible for the decrease in inflammation when coffee is administered [\[14\]](#page-9-10). We therefore sought to review the recent advances and knowledge in the association of major phytochemicals present in coffee (caffeine and chlorogenic acid), with their preventive or therapeutic effects targeted at the cellular and molecular mechanisms that lead to cancer progression.

#### **2. Biochemistry and Metabolism of Caffeine from Coffee Beans**

Around 1.67% of dried green coffee contains caffeine (1,3,7-trimethylxanthine) regardless of the different geographical origins that would affect the amount of caffeine [\[15\]](#page-9-11). Concerning oral consumption of caffeine in beverages, the caffeine is mostly absorbed in the gastrointestinal tract and small intestine, with unnoticeable significant first pass effect. Following absorption, caffeine spreads swiftly throughout plasma-binding. It has been found to occur in bile, saliva, semen, breast milk, and umbilical cord blood. The caffeine-plasma concentration peaks between 15 and 120 min after oral consumption. Notably, after ingestion, caffeine may rapidly pass through cell membranes with detectable levels in the brain as early as 5 min [\[16\]](#page-9-12). The study by Lin et al. [\[17\]](#page-9-13) showed that daily caffeine intake affected higher concentrations of caffeine in gray matter and cerebral blood flow, indicating the accumulation of caffeine residual in the brain. The primary metabolism of caffeine occurs in the liver through phase-I oxidation by cytochrome P450 1A2 resulting in active paraxanthine as a major metabolite, followed by theobromine and theophylline [\[18](#page-9-14)[–22\]](#page-9-15) (Figure [1\)](#page-2-0). Prior reports have already discovered the connection between daily coffee consumption and caffeine metabolism through the polymorphism of CYP1A2 and CYP2A6 [\[23](#page-9-16)[,24\]](#page-9-17). The second phase conjugated-metabolism produces a mixture of di- and tri-methylated xanthine, uric acid, and acetylated uracil derivatives, all being excreted through urine [\[25\]](#page-9-18). Previous studies have established that the biological effects of caffeine are tightly associated at three primary modulatory points: an antagonistic action on adenosine receptors, calcium mobilization, and phosphodiesterases inhibition [\[26](#page-9-19)[,27\]](#page-9-20).

The capacity of caffeine (and metabolite paraxanthine) to inhibit adenosine receptors due to their similar purine structure, shows its significant effect regarding cellular energy and inflammatory response [\[28](#page-9-21)[,29\]](#page-9-22). Furthermore, caffeine induces intracellular activity on calcium and the cyclic adenosine monophosphate phosphodiesterase (cAMP) pathway by inhibiting phosphodiesterase in adipose tissue and skeletal muscle [\[30\]](#page-9-23), resulting in cardiostimulatory and antiasthmatic actions [\[31\]](#page-9-24). Adenosine receptor stimulation leads to an increase in cAMP production, which may reduce the inflammatory response in a variety of pathophysiological circumstances. Despite caffeine not being a selective adenosine receptor antagonist, its modulatory effects on adenosine receptors may worsen the acute inflammatory response which depends on its concentration [\[32](#page-9-25)[,33\]](#page-10-0). Additionally, caffeine inflammatory response which depends on its concentration [32,33]. Additionally, caffeine stimulates calcium release by activating ryanodine receptors in skeletal muscles, raising stimulates calcium release by activating ryanodine receptors in skeletal muscles, raising intracellular calcium and speeding up the excitation–contraction coupling process, thus intracellular calcium and speeding up the excitation–contraction coupling process, thus playing a crucial role in the neurotransmitters released by neurons [\[34,](#page-10-1)35]. Recent studies playing a crucial role in the neurotransmitters released by neurons [3[4,3](#page-10-2)5]. Recent studies of caffeine also documented several mechanisms that involve systemic metabolism and of caffeine also documented several mechanisms that involve systemic metabolism and oxidative-inflammatory signaling, indicating that caffeine also affects peripheral signaling oxidative-inflammatory signaling, indicating that caffeine also affects peripheral signaling and may have beneficial effects on the human body regarding the aging process [\[16\]](#page-9-12). and may have beneficial effects on the human body regarding the aging process [16].

<span id="page-2-0"></span>

**Figure 1.** The major metabolism of caffeine in humans, adopted from [31]. **Figure 1.** The major metabolism of caffeine in humans, adopted from [\[31\]](#page-9-24).

# 3. Biochemistry and Metabolism of Chlorogenic Acid from Coffee Beans

There is a greater amount of chlorogenic acid (CGA) in green coffee bean than caffeine  $(5.43%)$ , and this would be lost during roasting  $[15,36]$  $[15,36]$ . Most of the biotransformation of chlorogenic acids in humans occurs in the colon, followed by the liver [37]. Dietary of chlorogenic acids in humans occurs in the colon, followed by the liver [\[37\]](#page-10-4). Dietary chlorogenic acids are absorbed in the small intestine, next they are hydrolyzed with es-terases from gut mucosa into quinic acid and caffeic acid (Figure [2\)](#page-3-0), and then they pass into the bloodstream. A substantial amount of the unaltered chlorogenic acid enters the colon, where it is metabolized by esterases produced by colon microflora. The colon plays cial role in transforming both caffeic and ferulic acid into dihydroferulic acid and facilitating their absorption through the intestine. Caffeic acid, e.g., 3, 4-dihydroxycinnamic a crucial role in transforming both caffeic and ferulic acid into dihydroferulic acid and acid, is converted by the enzyme catechol-O-methyltransferase into another phenolic acid, ferulic acid [\[38\]](#page-10-5). Both compounds have the ability to form an ester bond with quinic acid,

resulting in the formation of various isomers within the chlorogenic acid family. Most of the metabolized products from chlorogenic acid result from reaction with transferase, and are excreted as another form of benzoic acid called hippuric acid [\[39,](#page-10-6)[40\]](#page-10-7). genic acid family. Most of the metabolized products from chlorogenic acid result from resulting in the formation of various isomers within the chlorogenic acid ramily. Most of

<span id="page-3-0"></span>

**Figure 2.** The major metabolism of chlorogenic acid in humans, adopted from [37,40]. **Figure 2.** The major metabolism of chlorogenic acid in humans, adopted from [\[37](#page-10-4)[,40\]](#page-10-7).

#### **4. The Role of Coffee in Chemoprevention Activities on Carcinogenesis**

The targeted molecular pathways for developing and accessing future cancer-management techniques are carcinogenesis and chemoprevention. Chemoprevention refers to the use of pharmaceutical methods to stop or reverse the development of cancer before invasion and/or metastasis take place. According to epidemiological research, coffee consumption may be associated with a lower risk of cancer. The potential role of coffee in cancer chemoprevention has been supported by a number of experimental models, including human [\[41](#page-10-8)[,42\]](#page-10-9). The scientific literature has hypothesized a variety of coffee-dependent mechanisms, including the suppression of oxidative stress and damage, the activation of including the substrate in the determination processes of carentogens, and modulation of the inflammatory response. Furthermore, specific coffee ingredients and metadation of the inhamilatory response. I difference, specific correct ingredients<br>have been shown to affect tumor cell apoptosis, proliferation, and metastasis, and to exhibit anti-angiogenic properties [\[43,](#page-10-10)[44\]](#page-10-11). Interestingly, a higher intake of decaffeinated coffee significantly reduced the risk of colorectal cancer, but this effect was not observed with caffeinated coffee. However, it is known that caffeinated coffee can lower the risk of rectal tumor [\[45\]](#page-10-12). In another cohort study, both caffeinated and decaffeinated coffee consumption improved overall survival (OS) and progression free-survival (PFS) in patients with metastatic colorectal cancer [46]. Furthermore, frequent consumption of all coffee types lowered the chance of liver disease and carcinoma [\[47,](#page-10-14)48], while daily coffee intake reduced tumor size in invasive breast tumor with positive estrogen receptor (ER) more effectively than in triple-negative tumor [\[49\]](#page-10-16). These findings suggest that while drinking coffee with or without caffeine provides equivalent health benefits, caffeine may still play a role in some coffee-induced effects which also likely depend on the subsite of the tumor [\[50\]](#page-10-17). of metabolizing liver enzymes involved in the detoxification processes of carcinogens,

In addition to the chemopreventive activities demonstrated by caffeine and chlorogenic acid, studies have indicated that coffee extracts and kahweol also possess anticarcinogenesis properties in a number of cancer cell lines. Kahweol inhibits cancer growth in macrophage cells of mice via activating the NF-κB pathway [\[44,](#page-10-11)[51,](#page-10-18)[52\]](#page-10-19). Moreover, cotreatment with kahweol and cafestol has demonstrated anti-carcinogenic effects in male F344 rats [\[53\]](#page-10-20). Additionally, kahweol and cafestol together have been observed to provide

chemoprevention against malignancies caused by heterocyclic amines. Given that coffee constituents have the potential to exhibit antioxidant, cytotoxic, anti-mutagenic, and carcinogenic properties, they are therefore being studied for the treatment of different types of cancer, with particular attention to cafestol and kahweol [\[54](#page-10-21)[,55\]](#page-10-22), as these compounds may serve as valuable supplements to cancer prevention or therapy.

## **5. The Antitumor Activities of Coffee and Its Chemical Constituent**

A substansial report related to coffee extract and its metabolite constituents in cancer cells is summarized in Table [1.](#page-6-0) Caffeine directly inhibits the cyclin D/CDK 4/6 complex which causes G1 arrest independently of p53 [\[49\]](#page-10-16), and several reports also revealed that caffeine overrides the G2 phase arrest caused by DNA-damaging chemicals, propelling the cells into a deadly mitosis. Caffeine's capacity to restart Cdc25C and Cdc2 activity contributes to averting G2 arrest [\[56,](#page-10-23)[57\]](#page-10-24). Interestingly, due to its planar xanthine structure, caffeine is hypothetically a formed  $\pi$ - $\pi$  complex with nucleobases in DNA [\[58\]](#page-11-0), which is similar to conventional anticancer drugs [\[59\]](#page-11-1). In addition to triggering DNA intercalation, a report by Moura et al. [\[58\]](#page-11-0) found that caffeine had two possible roles: to protect DNA against DNA-damaging agents and to modulate intercalating drugs used in chemotherapy treatments.

Previous studies have documented that in melanoma cells, caffeine has a modulatory effect on the signaling cascades of AMP-activated protein kinase (AMPK), PI3K/Akt, and the mammalian target of rapamycin (mTOR) [\[60\]](#page-11-2). Moreover, caffeine downregulates the expression of several proteins, including retinoblastoma protein (Rb), extracellular signal-regulated kinases (ERK) 1/2, GSK3β, pyruvate dehydrogenase kinase 1 (PDK1), cyclin D1, cyclin E, c-Myc, Akt, and mTOR in various cancer cell lines [\[61–](#page-11-3)[63\]](#page-11-4). In another study, caffeine upregulates p300 expression in glioma cells [\[64\]](#page-11-5). Caffeine has been observed to reduce the phosphorylation of ERK induced by NF-κB in osteoclasts. A similar phenomenon also occurred in macrophage RAW 264.7 to suppress pro-inflammatory genes following lipopolysaccharide (LPS)-induced inflammation [\[62,](#page-11-6)[65\]](#page-11-7). In addition, coffee also demonstrated antitumor activity in vivo [\[66,](#page-11-8)[67\]](#page-11-9), and several studies have been conducted in humans to assess the correlation of coffee consumption and the risk of cancer [\[68\]](#page-11-10).

Chlorogenic acid in coffee has demonstrated antitumor action against cancer cell lines by reducing cell survival and suppressing reactive oxygen species (ROS) [\[69\]](#page-11-11). Additionally, it has been observed to suppress the production of cell adhesion molecules in human endothelial cells that are triggered by TNF-α16 [\[70\]](#page-11-12). Cafestol possesses anti-angiogenesis action in human umbilical vein endothelial cells, as it inhibits the proliferation, migration, and tube-formation ability of the cells [\[71](#page-11-13)[,72\]](#page-11-14). Ferulic acid also inhibits angiogenesis via targeting FGFR1 and activating the PI3K/Akt signaling pathways, limiting cell proliferation via cell cycle arrest and death in addition to reducing invasion, migration, and colony formation [\[73](#page-11-15)[,74\]](#page-11-16). Kahweol in green coffee bean has been shown to have an anti-angiogenic impact in zebrafish and chicken chorioallantoic membranes, in addition to exhibiting other significant activities including cell cycle arrest, anti-angiogenesis/proliferative, and associated phenomena [\[75\]](#page-11-17).

Currently, numerous cytotoxic medicines are utilized clinically for the treatment of various cancer types, despite their substantial side effects, low rate of cure, and development of resistance. Coffee, owing to its widespread availability, low cost, and racial compatibility, may hold promise as a significant anti-cancer treatment option [\[50\]](#page-10-17). In addition, combining coffee constituents (caffeine or chlorogenic acid) with existing chemotherapeutic drugs in cancer therapy has been evaluated. The combination of caffeine with doxorubicin prevented the efflux effect of doxorubicin from cancer cells and enhanced the cytotoxic activity [\[76\]](#page-11-18). A similar result was demonstrated in the synergistic effect of caffeine in cisplatin-treated sarcoma tumors [\[77\]](#page-11-19). Other antitumor drugs have also been summarized in a review by Ialongo et al. [\[78\]](#page-11-20). Clinical trials for caffeine have been reported in several publications, mainly combined with DNA-intercalating agents including cisplatin, doxorubicin, and the tyrosine kinase inhibitor dovitinib [\[79](#page-11-21)[–81\]](#page-11-22).



**Table 1.** The cytotoxic activities of chemical constituents of coffee bean against cancer cells.



<span id="page-6-0"></span>**Table 1.** *Cont.*

# **6. The Role of Coffee in Inducing Apoptosis toward Cancer Cells**

Coffee induces apoptosis by altering a number of the apoptotic response's constituent parts (Figure [3\)](#page-7-0). Different coffee compounds may target different apoptotic signaling mechanisms, such as increased cleavage of poly ADP ribose polymerase, downregulation of the signal transducer and activator of transcription 3 (STAT3) signaling pathway, and upregulation of the cyclic AMP-dependent transcription factor ATF3 [\[96,](#page-12-14)[98\]](#page-12-16). Caffeic acid has been shown to produce apoptotic cell death and dramatically decrease Akt signaling in PC-3 human prostate cancer cells, TW2.6, and HCT 15 colon cancer cell lines. Additionally, it has been proposed to decrease congenic survival and apoptotic cell death in SCC25, CAL27, and FaDu cell lines [\[99\]](#page-12-17). Numerous studies suggest that chemical constituents in coffee may possess apoptotic potential. The antioxidant function of these substances is also influenced by their environment. Some mechanisms of action include the inhibition of ROS generation and pro-survival gene expression, conformational changes in pro-apoptotic

<span id="page-7-0"></span>

proteins, loss of the mitochondrial membrane that activates caspases, and transcription<br>factor Sp1 [\[100\]](#page-12-18).

**Figure 3.** The hypothesized mechanism of coffee and its chemical compounds caffeine and chloro-**Figure 3.** The hypothesized mechanism of coffee and its chemical compounds caffeine and chlorogenic<br> acid of mediating apoptosis in cancer cells.

Despite its activity with respect to triggering apoptosis, it was later found that caffeine intake should be avoided in colorectal tumors treated with cell cycle modifying agents such as paclitaxel [\[101\]](#page-12-19). This was confirmed by Xu et al. who described that caffeine interferes with the anticancer effect of the antimitotic drug paclitaxel by preventing  $\alpha$ -tubulin acetyla-tion, which could enhance the progression of lung and cervical tumors [\[102\]](#page-12-20). It is important to note that the effect of caffeine in preventing the cytotoxicity of chemotherapy can be associated with cancer type, as caffeine enhanced the apoptosis in paclitaxel-induced breast cancer cells [\[103\]](#page-12-21). Nevertheless, these reports suggest that patients receiving antimitotic  $\epsilon$  cells  $\epsilon$  and  $\epsilon$  is the same way, more various manipulator as also been in the same  $\epsilon$  and  $\epsilon$  is the same way, mediator  $\epsilon$  and  $\epsilon$  is the same way, mediator  $\epsilon$  and  $\epsilon$  is the same  $\epsilon$  and  $\epsilon$  is the sa drugs as part of their cancer therapy regimen should avoid consuming foods or beverages<br>contributes of their normal relationship between and oxidative stress may also containing caffeine.

# **7. The Role of Coffee in Autophagy Process in Cancer Cells**  $\mathcal{L}$  treatments have proposed that induced that induced autophagy is an a

A double-membrane autophagosome is formed as part of the intracellular breakdown process known as autophagy. This mechanism facilitates the removal of inclusion bodies and misfolded cytotoxic proteins more effectively than apoptosis [\[104,](#page-12-22)[105\]](#page-12-23). Apart from programmed cell death, autophagy-induced cancer may also involve phosphatidylinositol 3-kinase (PI3K) pathways and the endoplasmic reticulum (ER) stress response. The dysregulation in this pathway has been linked to the development of cancer and resistance to cancer treatment, and it may have an impact on the level of autophagy in tumor cells [\[106,](#page-12-24)[107\]](#page-13-0). In the same way, mTOR has also been identified as an autophagy mediator that contributes to cell growth, survival, and proliferation [\[108,](#page-13-1)109]. Furthermore, the abnormal relationship between autophagy, inflammation, and oxidative stress may aid in the development of innovative pharmacotherapeutic approaches for the management and treatment of cancer. Recent developments have proposed that induced autophagy is a novel target for cancer  $t$ reatment [\[110](#page-13-3)[,111\]](#page-13-4).

It has been acknowledged that caffeine is able to suppress mTORC1 in both mice and in vitro models, to promote autophagosome generation in HepG2 cells, leading to the reduc-tion of intracellular fats, to enhance β-oxidation, and to control hepatosteatosis [\[112,](#page-13-5)[113\]](#page-13-6). It is noteworthy that caffeine in coffee has also exhibited cytoprotective effects in transformed

skin cells, preventing cellular senescence and suppressing ROS generation by inducing SIRT3/AMPK-mediated autophagy [\[114\]](#page-13-7). Despite its initial approach in normal tissues, many studies have proven that inducing autophagy in cancer cells can be beneficial for chemotherapy agents with respect to eliminating cancer cells. A study by Erzurumlu et al. [\[115\]](#page-13-8) showed that the addition of caffeine in docetaxel-treated breast cancer cells activated the unfolded protein response (UPR)-associated pathway and accelerated autophagy signaling due to increased Beclin-1 protein; this led to apoptosis in cancer cells as detected by the cleaved effector caspase-3. Also, methylxanthines derivatives (theophylline and caffeine) activated autophagy signaling through PTEN activation, followed by mTOR suppression in gastric tumor cells [\[116\]](#page-13-9). These findings open up the new challenge in caffeine development of inducing autophagy to initiate apoptosis in tumor cells, necessitating further experimental and clinical studies [\[117\]](#page-13-10).

#### **8. Conclusions and Recommendations**

Coffee has been shown to exhibit anticancer activities through several mechanisms and shows its potential to prevent carcinogenesis. Among these mechanisms, coffee and its major content caffeine have promising activities regarding autophagy, which serves as a potential therapeutic target due to its strong association with other mechanisms such as cellular senescence and ROS production. Further investigations are warranted to answer several interesting questions regarding the specific mechanisms by which chemical compounds in coffee induce autophagy. Additionally, the consumption of coffee as a daily beverage also shows reduced risk of cancer occurrence, thus supporting further exploration of the supplementation of coffee as a chemopreventive agent for cancer. One thing to note is that the physiological characteristics of whole coffee will probably vary since coffee is a complex, non-standardized beverage, despite its remarkable results from in vitro and in vivo investigations utilizing certain coffee secondary metabolite constituents, which have shown a variety of biological activities. As a result, the bioactivity of coffee in mixtures can be possibly affected by matrix, synergistic, and/or antagonist effects in preventing tumor progression. Also, only a small proportion of the substances consumed can pass through the circulatory system and enter the tissues, and very little of the absorbed content may retain the original structure of the phytochemical from coffee. For these reasons, it is noteworthy that the prevention of many diseases prompted by coffee use is usually the result of the combined action of numerous components, and in some cases, the synergistic effect of multiple types of compounds is substantially superior to the activity of single compounds.

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#### **References**

- <span id="page-8-0"></span>1. Schneider, K.A. All About Breast Cancer. In *Counseling about Cancer*; Wiley: Hoboken, NJ, USA, 2011; pp. 151–185.
- <span id="page-8-1"></span>2. Ma, L.; Zhang, M.; Zhao, R.; Wang, D.; Ma, Y.; Li, A. Plant Natural Products: Promising Resources for Cancer Chemoprevention. *Molecules* **2021**, *26*, 933. [\[CrossRef\]](https://doi.org/10.3390/molecules26040933) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33578780)
- <span id="page-8-2"></span>3. Kotecha, R.; Takami, A.; Espinoza, J.L. Dietary phytochemicals and cancer chemoprevention: A review of the clinical evidence. *Oncotarget* **2016**, *7*, 52517–52529. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.9593) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27232756)
- <span id="page-9-0"></span>4. Freedman, N.D.; Park, Y.; Abnet, C.C.; Hollenbeck, A.R.; Sinha, R. Association of coffee drinking with total and cause-specific mortality. *N. Engl. J. Med.* **2012**, *366*, 1891–1904. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1112010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22591295)
- <span id="page-9-1"></span>5. Mishra, M.; Panta, R.; Miyares, M. Influence of coffee and its components on breast cancer: A review. *Asian Pac. J. Trop. Dis.* **2016**, *6*, 827–831. [\[CrossRef\]](https://doi.org/10.1016/S2222-1808(16)61140-4)
- <span id="page-9-2"></span>6. Sado, J.; Kitamura, T.; Kitamura, Y.; Sobue, T.; Nishino, Y.; Tanaka, H.; Nakayama, T.; Tsuji, I.; Ito, H.; Suzuki, T.; et al. Association between coffee consumption and all-sites cancer incidence and mortality. *Cancer Sci.* **2017**, *108*, 2079–2087. [\[CrossRef\]](https://doi.org/10.1111/cas.13328) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28746796)
- <span id="page-9-3"></span>7. Andersen, L.F.; Jacobs, D.R., Jr.; Carlsen, M.H.; Blomhoff, R. Consumption of coffee is associated with reduced risk of death attributed to inflammatory and cardiovascular diseases in the Iowa Women's Health Study. *Am. J. Clin. Nutr.* **2006**, *83*, 1039–1046. [\[CrossRef\]](https://doi.org/10.1093/ajcn/83.5.1039) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16685044)
- <span id="page-9-4"></span>8. Sachs, J.D.; Cordes, K.Y.; Rising, J.; Toledano, P.; Maennling, N. *Ensuring Economic Viability and Sustainability of Coffee Production*; Columbia Center on Sustainable Investment: New York, NY, USA, 2019.
- <span id="page-9-5"></span>9. Gobbi, L.; Maddaloni, L.; Prencipe, S.A.; Vinci, G. Bioactive Compounds in Different Coffee Beverages for Quality and Sustainability Assessment. *Beverages* **2023**, *9*, 3. [\[CrossRef\]](https://doi.org/10.3390/beverages9010003)
- <span id="page-9-6"></span>10. de Melo Pereira, G.V.; de Carvalho Neto, D.P.; Júnior, A.I.M.; do Prado, F.G.; Pagnoncelli, M.G.B.; Karp, S.G.; Soccol, C.R. Chemical composition and health properties of coffee and coffee by-products. *Adv. Food Nutr. Res.* **2020**, *91*, 65–96. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32035601)
- <span id="page-9-7"></span>11. Cortés-Macías, E.T.; López, C.F.; Gentile, P.; Girón-Hernández, J.; López, A.F. Impact of post-harvest treatments on physicochemical and sensory characteristics of coffee beans in Huila, Colombia. *Postharvest Biol. Technol.* **2022**, *187*, 111852. [\[CrossRef\]](https://doi.org/10.1016/j.postharvbio.2022.111852)
- <span id="page-9-8"></span>12. Jeszka-Skowron, M.; Sentkowska, A.; Pyrzyńska, K.; De Peña, M.P. Chlorogenic acids, caffeine content and antioxidant properties of green coffee extracts: Influence of green coffee bean preparation. *Eur. Food Res. Technol.* **2016**, *242*, 1403–1409. [\[CrossRef\]](https://doi.org/10.1007/s00217-016-2643-y)
- <span id="page-9-9"></span>13. Carvalho Neto, D.P.d.; Gonot-Schoupinsky, X.P.; Gonot-Schoupinsky, F.N. Coffee as a naturally beneficial and sustainable ingredient in personal care products: A systematic scoping review of the evidence. *Front. Sustain.* **2021**, *2*, 697092. [\[CrossRef\]](https://doi.org/10.3389/frsus.2021.697092)
- <span id="page-9-10"></span>14. Karpinska, J.; Swisłocka, R.; Lewandowski, W. A mystery of a cup of coffee; an insight look by chemist. ´ *BioFactors* **2017**, *43*, 621–632. [\[CrossRef\]](https://doi.org/10.1002/biof.1371) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28613019)
- <span id="page-9-11"></span>15. Awwad, S.; Issa, R.; Alnsour, L.; Albals, D.; Al-Momani, I. Quantification of Caffeine and Chlorogenic Acid in Green and Roasted Coffee Samples Using HPLC-DAD and Evaluation of the Effect of Degree of Roasting on Their Levels. *Molecules* **2021**, *26*, 7502. [\[CrossRef\]](https://doi.org/10.3390/molecules26247502) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34946584)
- <span id="page-9-12"></span>16. Barcelos, R.P.; Lima, F.D.; Carvalho, N.R.; Bresciani, G.; Royes, L.F. Caffeine effects on systemic metabolism, oxidativeinflammatory pathways, and exercise performance. *Nutr. Res.* **2020**, *80*, 1–17. [\[CrossRef\]](https://doi.org/10.1016/j.nutres.2020.05.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32589582)
- <span id="page-9-13"></span>17. Lin, Y.-S.; Weibel, J.; Landolt, H.-P.; Santini, F.; Garbazza, C.; Kistler, J.; Rehm, S.; Rentsch, K.; Borgwardt, S.; Cajochen, C.; et al. Time to Recover From Daily Caffeine Intake. *Front. Nutr.* **2022**, *8*, 787225. [\[CrossRef\]](https://doi.org/10.3389/fnut.2021.787225) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35187019)
- <span id="page-9-14"></span>18. Jandova, Z.; Gill, S.C.; Lim, N.M.; Mobley, D.L.; Oostenbrink, C. Binding Modes and Metabolism of Caffeine. *Chem. Res. Toxicol.* **2019**, *32*, 1374–1383. [\[CrossRef\]](https://doi.org/10.1021/acs.chemrestox.9b00030) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31132250)
- 19. Lelo, A.; Kjellen, G.; Birkett, D.J.; Miners, J.O. Paraxanthine metabolism in humans: Determination of metabolic partial clearances and effects of allopurinol and cimetidine. *J. Pharmacol. Exp. Ther.* **1989**, *248*, 315–319. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2913277)
- 20. Nehlig, A. Interindividual Differences in Caffeine Metabolism and Factors Driving Caffeine Consumption. *Pharmacol. Rev.* **2018**, *70*, 384–411. [\[CrossRef\]](https://doi.org/10.1124/pr.117.014407)
- 21. Tarka, S.M., Jr.; Arnaud, M.J.; Dvorchik, B.H.; Vesell, E.S. Theobromine kinetics and metabolic disposition. *Clin. Pharmacol. Ther.* **1983**, *34*, 546–555. [\[CrossRef\]](https://doi.org/10.1038/clpt.1983.212)
- <span id="page-9-15"></span>22. Labedzki, A.; Buters, J.; Jabrane, W.; Fuhr, U. Differences in caffeine and paraxanthine metabolism between human and murine CYP1A2. *Biochem. Pharmacol.* **2002**, *63*, 2159–2167. [\[CrossRef\]](https://doi.org/10.1016/S0006-2952(02)01019-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12110375)
- <span id="page-9-16"></span>23. Josse, A.R.; Da Costa, L.A.; Campos, H.; El-Sohemy, A. Associations between polymorphisms in the AHR and CYP1A1-CYP1A2 gene regions and habitual caffeine consumption. *Am. J. Clin. Nutr.* **2012**, *96*, 665–671. [\[CrossRef\]](https://doi.org/10.3945/ajcn.112.038794) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22854411)
- <span id="page-9-17"></span>24. Cornelis, M.C.; Kacprowski, T.; Menni, C.; Gustafsson, S.; Pivin, E.; Adamski, J.; Artati, A.; Eap, C.B.; Ehret, G.; Friedrich, N.; et al. Genome-wide association study of caffeine metabolites provides new insights to caffeine metabolism and dietary caffeine-consumption behavior. *Hum. Mol. Genet.* **2016**, *25*, 5472–5482. [\[CrossRef\]](https://doi.org/10.1093/hmg/ddw334) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27702941)
- <span id="page-9-18"></span>25. Arnaud, M.J. Components of coffee. *Caffeine Coffee Health* **1993**, *43*, 43–95.
- <span id="page-9-19"></span>26. Sardão, V.A.; Oliveira, P.J.; Moreno, A.J.M. Caffeine Enhances the Calcium-Dependent Cardiac Mitochondrial Permeability Transition: Relevance for Caffeine Toxicity. *Toxicol. Appl. Pharmacol.* **2002**, *179*, 50–56. [\[CrossRef\]](https://doi.org/10.1006/taap.2001.9334) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11884236)
- <span id="page-9-20"></span>27. Ribeiro, J.A.; Sebastiao, A.M. Caffeine and adenosine. *J. Alzheimer's Dis.* **2010**, *20*, S3–S15. [\[CrossRef\]](https://doi.org/10.3233/JAD-2010-1379) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20164566)
- <span id="page-9-21"></span>28. Gliottoni, R.C.; Meyers, J.R.; Arngrímsson, S.Á.; Broglio, S.P.; Motl, R.W. Effect of caffeine on quadriceps muscle pain during acute cycling exercise in low versus high caffeine consumers. *Int. J. Sport Nutr. Exerc. Metab.* **2009**, *19*, 150–161. [\[CrossRef\]](https://doi.org/10.1123/ijsnem.19.2.150) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19478340)
- <span id="page-9-22"></span>29. Monteiro, J.P.; Alves, M.G.; Oliveira, P.F.; Silva, B.M. Structure-bioactivity relationships of methylxanthines: Trying to make sense of all the promises and the drawbacks. *Molecules* **2016**, *21*, 974. [\[CrossRef\]](https://doi.org/10.3390/molecules21080974) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27472311)
- <span id="page-9-23"></span>30. Orbán, C.; Vásárhelyi, Z.; Bajnok, A.; Sava, F.; Toldi, G. Effects of caffeine and phosphodiesterase inhibitors on activation of neonatal T lymphocytes. *Immunobiology* **2018**, *223*, 627–633. [\[CrossRef\]](https://doi.org/10.1016/j.imbio.2018.07.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30177027)
- <span id="page-9-24"></span>31. Institute of Medicine (US) Committee on Military Nutrition Research. *Caffeine for the Sustainment of Mental Task Performance: Formulations for Military Operations*; National Academies Press: Washington, DC, USA, 2001.
- <span id="page-9-25"></span>32. Meeusen, R.; Roelands, B.; Spriet, L.L. Caffeine, exercise and the brain. In *Limits of Human Endurance*; Karger Publishers: Basel, Switzerland, 2013; Volume 76, pp. 1–12.
- <span id="page-10-0"></span>33. Ohta, A.; Lukashev, D.; Jackson, E.K.; Fredholm, B.B.; Sitkovsky, M. 1,3,7-trimethylxanthine (caffeine) may exacerbate acute inflammatory liver injury by weakening the physiological immunosuppressive mechanism. *J. Immunol.* **2007**, *179*, 7431–7438. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.179.11.7431) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18025187)
- <span id="page-10-1"></span>34. Kong, H.; Jones, P.P.; Koop, A.; Zhang, L.; Duff, H.J.; Chen, S.R. Caffeine induces  $Ca^{2+}$  release by reducing the threshold for luminal Ca2+ activation of the ryanodine receptor. *Biochem. J.* **2008**, *414*, 441–452. [\[CrossRef\]](https://doi.org/10.1042/BJ20080489) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18518861)
- <span id="page-10-2"></span>35. Alasmari, F. Caffeine induces neurobehavioral effects through modulating neurotransmitters. *Saudi Pharm. J.* **2020**, *28*, 445–451. [\[CrossRef\]](https://doi.org/10.1016/j.jsps.2020.02.005)
- <span id="page-10-3"></span>36. Van Cuong, T.; Ling, L.H.; Quan, G.K.; Tiep, T.D.; Nan, X.; Qing, C.X.; Le Linh, T. Effect of roasting conditions on several chemical constituents of Vietnam Robusta coffee. *Ann. Univ. Dunarea Jos Galati Fascicle VI-Food Technol.* **2014**, *38*, 43–56.
- <span id="page-10-4"></span>37. Olthof, M.R.; Hollman, P.C.; Katan, M.B. Chlorogenic acid and caffeic acid are absorbed in humans. *J. Nutr.* **2001**, *131*, 66–71. [\[CrossRef\]](https://doi.org/10.1093/jn/131.1.66)
- <span id="page-10-5"></span>38. Zhao, Y.; Wang, J.; Ballevre, O.; Luo, H.; Zhang, W. Antihypertensive effects and mechanisms of chlorogenic acids. *Hypertens. Res.* **2012**, *35*, 370–374. [\[CrossRef\]](https://doi.org/10.1038/hr.2011.195) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22072103)
- <span id="page-10-6"></span>39. Nabavi, S.F.; Tejada, S.; Setzer, W.N.; Gortzi, O.; Sureda, A.; Braidy, N.; Daglia, M.; Manayi, A.; Nabavi, S.M. Chlorogenic Acid and Mental Diseases: From Chemistry to Medicine. *Curr. Neuropharmacol.* **2017**, *15*, 471–479. [\[CrossRef\]](https://doi.org/10.2174/1570159X14666160325120625) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27012954)
- <span id="page-10-7"></span>40. Olthof, M.R.; Hollman, P.C.H.; Buijsman, M.N.C.P.; van Amelsvoort, J.M.M.; Katan, M.B. Chlorogenic Acid, Quercetin-3- Rutinoside and Black Tea Phenols Are Extensively Metabolized in Humans. *J. Nutr.* **2003**, *133*, 1806–1814. [\[CrossRef\]](https://doi.org/10.1093/jn/133.6.1806) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12771321)
- <span id="page-10-8"></span>41. Lee, H.N.; Kim, J.K.; Kim, J.H.; Lee, S.J.; Ahn, E.K.; Oh, J.S.; Seo, D.W. A mechanistic study on the anti-cancer activity of ethyl caffeate in human ovarian cancer SKOV-3 cells. *Chem. Biol. Interact.* **2014**, *219*, 151–158. [\[CrossRef\]](https://doi.org/10.1016/j.cbi.2014.05.017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24892518)
- <span id="page-10-9"></span>42. Siegel, R.L.; Miller, K.D.; Wagle, N.S.; Jemal, A. Cancer statistics, 2023. *CA Cancer J Clin* **2023**, *73*, 17–48. [\[CrossRef\]](https://doi.org/10.3322/caac.21763)
- <span id="page-10-10"></span>43. Lima, C.S.; Spindola, D.G.; Bechara, A.; Garcia, D.M.; Palmeira-dos-Santos, C.; Peixoto-da-Silva, J.; Erustes, A.G.; Michelin, L.F.G.; Pereira, G.J.S.; Smaili, S.S.; et al. Cafestol, a diterpene molecule found in coffee, induces leukemia cell death. *Biomed. Pharmacother.* **2017**, *92*, 1045–1054. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2017.05.109) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28618649)
- <span id="page-10-11"></span>44. Makino, T.; Izumi, K.; Hiratsuka, K.; Kano, H.; Shimada, T.; Nakano, T.; Kadomoto, S.; Naito, R.; Iwamoto, H.; Yaegashi, H.; et al. Anti-proliferative and anti-migratory properties of coffee diterpenes kahweol acetate and cafestol in human renal cancer cells. *Sci. Rep.* **2021**, *11*, 675. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-80302-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33436830)
- <span id="page-10-12"></span>45. Um, C.Y.; McCullough, M.L.; Guinter, M.A.; Campbell, P.T.; Jacobs, E.J.; Gapstur, S.M. Coffee consumption and risk of colorectal cancer in the Cancer Prevention Study-II Nutrition Cohort. *Cancer Epidemiol.* **2020**, *67*, 101730. [\[CrossRef\]](https://doi.org/10.1016/j.canep.2020.101730) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32526644)
- <span id="page-10-13"></span>46. Mackintosh, C.; Yuan, C.; Ou, F.-S.; Zhang, S.; Niedzwiecki, D.; Chang, I.-W.; O'Neil, B.H.; Mullen, B.C.; Lenz, H.-J.; Blanke, C.D.; et al. Association of Coffee Intake With Survival in Patients With Advanced or Metastatic Colorectal Cancer. *JAMA Oncol.* **2020**, *6*, 1713–1721. [\[CrossRef\]](https://doi.org/10.1001/jamaoncol.2020.3938) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32940631)
- <span id="page-10-14"></span>47. Tran, K.T.; Coleman, H.G.; McMenamin, Ú.C.; Cardwell, C.R. Coffee consumption by type and risk of digestive cancer: A large prospective cohort study. *Br. J. Cancer* **2019**, *120*, 1059–1066. [\[CrossRef\]](https://doi.org/10.1038/s41416-019-0465-y)
- <span id="page-10-15"></span>48. Tanaka, K.; Tamakoshi, A.; Sugawara, Y.; Mizoue, T.; Inoue, M.; Sawada, N.; Matsuo, K.; Ito, H.; Naito, M.; Nagata, C. Coffee, green tea and liver cancer risk: An evaluation based on a systematic review of epidemiologic evidence among the Japanese population. *Jpn. J. Clin. Oncol.* **2019**, *49*, 972–984. [\[CrossRef\]](https://doi.org/10.1093/jjco/hyz097) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31790152)
- <span id="page-10-16"></span>49. Rosendahl, A.H.; Perks, C.M.; Zeng, L.; Markkula, A.; Simonsson, M.; Rose, C.; Ingvar, C.; Holly, J.M.P.; Jernström, H. Caffeine and Caffeic Acid Inhibit Growth and Modify Estrogen Receptor and Insulin-like Growth Factor I Receptor Levels in Human Breast Cancer. *Clin. Cancer Res.* **2015**, *21*, 1877–1887. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-14-1748) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25691730)
- <span id="page-10-17"></span>50. Safe, S.; Kothari, J.; Hailemariam, A.; Upadhyay, S.; Davidson, L.A.; Chapkin, R.S. Health Benefits of Coffee Consumption for Cancer and Other Diseases and Mechanisms of Action. *Int. J. Mol. Sci.* **2023**, *24*, 2706. [\[CrossRef\]](https://doi.org/10.3390/ijms24032706) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36769029)
- <span id="page-10-18"></span>51. Choi, D.W.; Lim, M.S.; Lee, J.W.; Chun, W.; Lee, S.H.; Nam, Y.H.; Park, J.M.; Choi, D.H.; Kang, C.D.; Lee, S.J.; et al. The Cytotoxicity of Kahweol in HT-29 Human Colorectal Cancer Cells Is Mediated by Apoptosis and Suppression of Heat Shock Protein 70 Expression. *Biomol. Ther.* **2015**, *23*, 128–133. [\[CrossRef\]](https://doi.org/10.4062/biomolther.2014.133)
- <span id="page-10-19"></span>52. Oh, S.H.; Hwang, Y.P.; Choi, J.H.; Jin, S.W.; Lee, G.H.; Han, E.H.; Chung, Y.H.; Chung, Y.C.; Jeong, H.G. Kahweol inhibits proliferation and induces apoptosis by suppressing fatty acid synthase in HER2-overexpressing cancer cells. *Food Chem. Toxicol.* **2018**, *121*, 326–335. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2018.09.008)
- <span id="page-10-20"></span>53. Bovell-Benjamin, A.C. Chapter 22—Bioactivity, Benefits and Safety of Traditional and Ethnic Foods. In *Ensuring Global Food Safety*; Boisrobert, C.E., Stjepanovic, A., Oh, S., Lelieveld, H.L.M., Eds.; Academic Press: San Diego, CA, USA, 2010; pp. 363–382.
- <span id="page-10-21"></span>54. Huber, W.W.; Rossmanith, W.; Grusch, M.; Haslinger, E.; Prustomersky, S.; Peter-Vörösmarty, B.; Parzefall, W.; Scharf, G.; Schulte-Hermann, R. Effects of coffee and its chemopreventive components kahweol and cafestol on cytochrome P450 and sulfotransferase in rat liver. *Food Chem. Toxicol.* **2008**, *46*, 1230–1238. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2007.09.094)
- <span id="page-10-22"></span>55. Ren, Y.; Wang, C.; Xu, J.; Wang, S. Cafestol and Kahweol: A Review on Their Bioactivities and Pharmacological Properties. *Int. J. Mol. Sci.* **2019**, *20*, 4238. [\[CrossRef\]](https://doi.org/10.3390/ijms20174238)
- <span id="page-10-23"></span>56. Tej, G.; Nayak, P.K. Mechanistic considerations in chemotherapeutic activity of caffeine. *Biomed. Pharmacother.* **2018**, *105*, 312–319. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2018.05.144) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29864619)
- <span id="page-10-24"></span>57. Meisaprow, P.; Aksorn, N.; Vinayanuwattikun, C.; Chanvorachote, P.; Sukprasansap, M. Caffeine Induces G0/G1 Cell Cycle Arrest and Inhibits Migration through Integrin αv, β3, and FAK/Akt/c-Myc Signaling Pathway. *Molecules* **2021**, *26*, 7659. [\[CrossRef\]](https://doi.org/10.3390/molecules26247659) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34946741)
- <span id="page-11-0"></span>58. Moura, T.A.; Junior, R.L.R.; Rocha, M.S. Caffeine modulates the intercalation of drugs on DNA: A study at the single molecule level. *Biophys. Chem.* **2021**, *277*, 106653. [\[CrossRef\]](https://doi.org/10.1016/j.bpc.2021.106653) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34217911)
- <span id="page-11-1"></span>59. Davies, D.B.; Veselkov, D.A.; Djimant, L.N.; Veselkov, A.N. Hetero-association of caffeine and aromatic drugs and their competitive binding with a DNA oligomer. *Eur. Biophys. J.* **2001**, *30*, 354–366. [\[CrossRef\]](https://doi.org/10.1007/s002490100150) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11592692)
- <span id="page-11-2"></span>60. Caini, S.; Masala, G.; Saieva, C.; Kvaskoff, M.; Savoye, I.; Sacerdote, C.; Hemmingsson, O.; Hammer Bech, B.; Overvad, K.; Tjønneland, A.; et al. Coffee, tea and melanoma risk: Findings from the European Prospective Investigation into Cancer and Nutrition. *Int. J. Cancer* **2017**, *140*, 2246–2255. [\[CrossRef\]](https://doi.org/10.1002/ijc.30659)
- <span id="page-11-3"></span>61. Gibbs, B.F.; Gonçalves Silva, I.; Prokhorov, A.; Abooali, M.; Yasinska, I.; Casely-Hayford, M.A.; Berger, S.M.; Fasler-Kan, E.; Sumbayev, V. Caffeine affects the biological responses of human hematopoietic cells of myeloid lineage via downregulation of the mTOR pathway and xanthine oxidase activity. *Oncotarget* **2015**, *6*, 28678. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.5212)
- <span id="page-11-6"></span>62. Miwa, S.; Sugimoto, N.; Yamamoto, N.; Shirai, T.; Nishida, H.; Hayashi, K.; Kimura, H.; Takeuchi, A.; Igarashi, K.; Yachie, A.; et al. Caffeine induces apoptosis of osteosarcoma cells by inhibiting AKT/mTOR/S6K, NF-κB and MAPK pathways. *Anticancer Res.* **2012**, *32*, 3643–3649.
- <span id="page-11-4"></span>63. Saiki, S.; Sasazawa, Y.; Imamichi, Y.; Kawajiri, S.; Fujimaki, T.; Tanida, I.; Kobayashi, H.; Sato, F.; Sato, S.; Ishikawa, K.; et al. Caffeine induces apoptosis by enhancement of autophagy via PI3K/Akt/mTOR/p70S6K inhibition. *Autophagy* **2011**, *7*, 176–187. [\[CrossRef\]](https://doi.org/10.4161/auto.7.2.14074)
- <span id="page-11-5"></span>64. Chen, J.C.; Hwang, J.H. Effects of caffeine on cell viability and activity of histone deacetylase 1 and histone acetyltransferase in glioma cells. *Ci Ji Yi Xue Za Zhi* **2016**, *28*, 103–108. [\[CrossRef\]](https://doi.org/10.1016/j.tcmj.2016.06.005)
- <span id="page-11-7"></span>65. Hwang, J.H.; Kim, K.J.; Ryu, S.J.; Lee, B.Y. Caffeine prevents LPS-induced inflammatory responses in RAW264.7 cells and zebrafish. *Chem. Biol. Interact.* **2016**, *248*, 1–7. [\[CrossRef\]](https://doi.org/10.1016/j.cbi.2016.01.020)
- <span id="page-11-8"></span>66. Tsikis, S.; Hoefer, L.; Bethimoutis, G.; Nicolaidou, E.; Paparizos, V.; Antoniou, C.; Chardalias, L.; Stavropoulos, G.E.; Sharma, S.; Long, B.C.; et al. Risk factors, prevalence, and site concordance of human papillomavirus in high-risk Greek men. *Eur. J. Cancer Prev.* **2018**, *27*, 514–520. [\[CrossRef\]](https://doi.org/10.1097/CEJ.0000000000000366) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28394804)
- <span id="page-11-9"></span>67. Gligor, O.; Clichici, S.; Moldovan, R.; Decea, N.; Vlase, A.M.; Fizesan, I.; Pop, A.; Virag, P.; Filip, G.A.; Vlase, L.; et al. An In Vitro and In Vivo Assessment of Antitumor Activity of Extracts Derived from Three Well-Known Plant Species. *Plants* **2023**, *12*, 1840. [\[CrossRef\]](https://doi.org/10.3390/plants12091840) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37176897)
- <span id="page-11-10"></span>68. Nigra, A.D.; Teodoro, A.J.; Gil, G.A. A Decade of Research on Coffee as an Anticarcinogenic Beverage. *Oxidative Med. Cell. Longev.* **2021**, *2021*, 4420479. [\[CrossRef\]](https://doi.org/10.1155/2021/4420479) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34567408)
- <span id="page-11-11"></span>69. Yamagata, K.; Izawa, Y.; Onodera, D.; Tagami, M. Chlorogenic acid regulates apoptosis and stem cell marker-related gene expression in A549 human lung cancer cells. *Mol. Cell Biochem.* **2018**, *441*, 9–19. [\[CrossRef\]](https://doi.org/10.1007/s11010-017-3171-1)
- <span id="page-11-12"></span>70. Chang, W.C.; Chen, C.H.; Lee, M.F.; Chang, T.; Yu, Y.M. Chlorogenic acid attenuates adhesion molecules upregulation in IL-1beta-treated endothelial cells. *Eur. J. Nutr.* **2010**, *49*, 267–275. [\[CrossRef\]](https://doi.org/10.1007/s00394-009-0083-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19937041)
- <span id="page-11-13"></span>71. Moeenfard, M.; Cortez, A.; Machado, V.; Costa, R.; Luís, C.; Coelho, P.; Soares, R.; Alves, A.; Borges, N.; Santos, A. Anti-Angiogenic Properties of Cafestol and Kahweol Palmitate Diterpene Esters. *J. Cell. Biochem.* **2016**, *117*, 2748–2756. [\[CrossRef\]](https://doi.org/10.1002/jcb.25573) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27129115)
- <span id="page-11-14"></span>72. Wang, S.; Yoon, Y.C.; Sung, M.J.; Hur, H.J.; Park, J.H. Antiangiogenic properties of cafestol, a coffee diterpene, in human umbilical vein endothelial cells. *Biochem. Biophys. Res. Commun.* **2012**, *421*, 567–571. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2012.04.046) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22525673)
- <span id="page-11-15"></span>73. Muchtaridi, M.; Lestari, D.; Khairul Ikram, N.K.; Gazzali, A.M.; Hariono, M.; Wahab, H.A. Decaffeination and Neuraminidase Inhibitory Activity of Arabica Green Coffee (*Coffea arabica*) Beans: Chlorogenic Acid as a Potential Bioactive Compound. *Molecules* **2021**, *26*, 3402. [\[CrossRef\]](https://doi.org/10.3390/molecules26113402)
- <span id="page-11-16"></span>74. Seow, L.-J.; Shamlan, S.; Seow, E.-K. Influence of roasting degrees on the antioxidant and anti-angiogenic effects of Coffea liberica. *J. Food Meas. Charact.* **2021**, *15*, 4030–4036. [\[CrossRef\]](https://doi.org/10.1007/s11694-021-00987-7)
- <span id="page-11-17"></span>75. Dong, S.; Kong, J.; Kong, J.; Shen, Q.; Kong, F.; Sun, W.; Zheng, L. Low Concentration of Caffeine Inhibits the Progression of the Hepatocellular Carcinoma via Akt Signaling Pathway. *Anticancer Agents Med. Chem.* **2015**, *15*, 484–492. [\[CrossRef\]](https://doi.org/10.2174/1871520615666150209110832) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25666502)
- <span id="page-11-18"></span>76. Chung, F.-L.; Wang, M.; Rivenson, A.; Iatropoulos, M.J.; Reinhardt, J.C.; Pittman, B.; Ho, C.-T.; Amin, S.G. Inhibition of lung carcinogenesis by black tea in Fischer rats treated with a tobacco-specific carcinogen: Caffeine as an important constituent. *Cancer Res.* **1998**, *58*, 4096–4101. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9751618)
- <span id="page-11-19"></span>77. Tsuchiya, H.; Tomita, K.; Yamamoto, N.; Mori, Y.; Asada, N. Caffeine-potentiated chemotherapy and conservative surgery for high-grade soft-tissue sarcoma. *Anticance. Res.* **1998**, *18*, 3651–3656.
- <span id="page-11-20"></span>78. Ialongo, D.; Tudino, V.; Arpacioglu, M.; Messore, A.; Patacchini, E.; Costi, R.; Di Santo, R.; Madia, V.N. Synergistic Effects of Caffeine in Combination with Conventional Drugs: Perspectives of a Drug That Never Ages. *Pharmaceuticals* **2023**, *16*, 730. [\[CrossRef\]](https://doi.org/10.3390/ph16050730) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37242514)
- <span id="page-11-21"></span>79. Takeuchi, A.; Tsuchiya, H.; Yamamoto, N.; Hayashi, K.; Yamauchi, K.; Kawahara, M.; Miyamoto, K.; Tomita, K. Caffeinepotentiated chemotherapy for patients with high-grade soft tissue sarcoma: Long-term clinical outcome. *Anticancer Res.* **2007**, *27*, 3489–3495. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17972506)
- 80. de Weger, V.A.; Goel, S.; von Moos, R.; Schellens, J.H.M.; Mach, N.; Tan, E.; Anand, S.; Scott, J.W.; Lassen, U. A drug–drug interaction study to assess the effect of the CYP1A2 inhibitor fluvoxamine on the pharmacokinetics of dovitinib (TKI258) in patients with advanced solid tumors. *Cancer Chemother. Pharmacol.* **2018**, *81*, 73–80. [\[CrossRef\]](https://doi.org/10.1007/s00280-017-3469-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29101463)
- <span id="page-11-22"></span>81. Dougherty, J.B.; Kelsen, D.; Kemeny, N.; Magill, G.; Botet, J.; Niedzwiecki, D. Advanced Pancreatic Cancer: A Phase I–II Trial of Cisplatin, High-Dose Cytarabine, and Caffeine. *J. Natl. Cancer Inst.* **1989**, *81*, 1735–1738. [\[CrossRef\]](https://doi.org/10.1093/jnci/81.22.1735) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2681796)
- <span id="page-12-0"></span>82. Park, J.J.; Hwang, S.J.; Park, J.H.; Lee, H.J. Chlorogenic acid inhibits hypoxia-induced angiogenesis via down-regulation of the HIF-1α/AKT pathway. *Cell. Oncol.* **2015**, *38*, 111–118. [\[CrossRef\]](https://doi.org/10.1007/s13402-014-0216-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25561311)
- <span id="page-12-1"></span>83. Huang, S.; Wang, L.L.; Xue, N.N.; Li, C.; Guo, H.H.; Ren, T.K.; Zhan, Y.; Li, W.B.; Zhang, J.; Chen, X.G.; et al. Chlorogenic acid effectively treats cancers through induction of cancer cell differentiation. *Theranostics* **2019**, *9*, 6745–6763. [\[CrossRef\]](https://doi.org/10.7150/thno.34674) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31660066)
- <span id="page-12-2"></span>84. Burgos-Morón, E.; Calderón-Montaño, J.M.; Orta, M.L.; Pastor, N.; Pérez-Guerrero, C.; Austin, C.; Mateos, S.; López-Lázaro, M. The coffee constituent chlorogenic acid induces cellular DNA damage and formation of topoisomerase I- and II-DNA complexes in cells. *J. Agric. Food Chem.* **2012**, *60*, 7384–7391. [\[CrossRef\]](https://doi.org/10.1021/jf300999e)
- <span id="page-12-3"></span>85. Liu, H.; Hua, Y.; Zheng, X.; Shen, Z.; Luo, H.; Tao, X.; Wang, Z. Effect of coffee consumption on the risk of gastric cancer: A systematic review and meta-analysis of prospective cohort studies. *PLoS ONE* **2015**, *10*, e0128501. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0128501) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26023935)
- <span id="page-12-4"></span>86. Yan, Y.; Liu, N.; Hou, N.; Dong, L.; Li, J. Chlorogenic acid inhibits hepatocellular carcinoma in vitro and in vivo. *J. Nutr. Biochem.* **2017**, *46*, 68–73. [\[CrossRef\]](https://doi.org/10.1016/j.jnutbio.2017.04.007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28458139)
- <span id="page-12-5"></span>87. Rathod, M.A.; Patel, D.; Das, A.; Tipparaju, S.R.; Shinde, S.S.; Anderson, R.F. Inhibition of radical-induced DNA strand breaks by water-soluble constituents of coffee: Phenolics and caffeine metabolites. *Free Radic. Res.* **2013**, *47*, 480–487. [\[CrossRef\]](https://doi.org/10.3109/10715762.2013.788167) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23521605)
- <span id="page-12-6"></span>88. Pounis, G.; Tabolacci, C.; Costanzo, S.; Cordella, M.; Bonaccio, M.; Rago, L.; D'Arcangelo, D.; Di Castelnuovo, F.A.; de Gaetano, G.; Donati, M.B.; et al. Reduction by coffee consumption of prostate cancer risk: Evidence from the Moli-sani cohort and cellular models. *Int. J. Cancer* **2017**, *141*, 72–82. [\[CrossRef\]](https://doi.org/10.1002/ijc.30720) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28436066)
- <span id="page-12-7"></span>89. Shim, S.G.; Jun, D.W.; Kim, E.K.; Saeed, W.K.; Lee, K.N.; Lee, H.L.; Lee, O.Y.; Choi, H.S.; Yoon, B.C. Caffeine attenuates liver fibrosis via defective adhesion of hepatic stellate cells in cirrhotic model. *J. Gastroenterol. Hepatol.* **2013**, *28*, 1877–1884. [\[CrossRef\]](https://doi.org/10.1111/jgh.12317) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23808892)
- <span id="page-12-8"></span>90. Safa, M.; Bashash, D.; Hamidpoor, M. Induction of cell death and decreased cell proliferation in acute promyelocytic leukemia cells (NB4) by caffeine. *Sci. J. Iran. Blood Transfus Organ* **2016**, *12*, 331–339.
- <span id="page-12-9"></span>91. Wang, X.; Lim, L.-T. Chapter 27—Physicochemical Characteristics of Roasted Coffee. In *Coffee in Health and Disease Prevention*; Preedy, V.R., Ed.; Academic Press: San Diego, CA, USA, 2015; pp. 247–254.
- <span id="page-12-10"></span>92. Choi, M.J.; Park, E.J.; Oh, J.H.; Min, K.J.; Yang, E.S.; Kim, Y.H.; Lee, T.J.; Kim, S.H.; Choi, Y.H.; Park, J.W.; et al. Cafestol, a coffee-specific diterpene, induces apoptosis in renal carcinoma Caki cells through down-regulation of anti-apoptotic proteins and Akt phosphorylation. *Chem. Biol. Interact.* **2011**, *190*, 102–108. [\[CrossRef\]](https://doi.org/10.1016/j.cbi.2011.02.013)
- <span id="page-12-11"></span>93. Cárdenas, C.; Quesada, A.R.; Medina, M.A. Anti-angiogenic and anti-inflammatory properties of kahweol, a coffee diterpene. *PLoS ONE* **2011**, *6*, e23407. [\[CrossRef\]](https://doi.org/10.1371/annotation/38262cc6-07cc-4074-8ce7-2181d4d0fbdc)
- <span id="page-12-12"></span>94. Seo, H.Y.; Kim, M.K.; Lee, S.H.; Hwang, J.S.; Park, K.G.; Jang, B.K. Kahweol Ameliorates the Liver Inflammation through the Inhibition of NF-κB and STAT3 Activation in Primary Kupffer Cells and Primary Hepatocytes. *Nutrients* **2018**, *10*, 863. [\[CrossRef\]](https://doi.org/10.3390/nu10070863) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29973533)
- <span id="page-12-13"></span>95. Oh, J.H.; Lee, J.T.; Yang, E.S.; Chang, J.S.; Lee, D.S.; Kim, S.H.; Choi, Y.H.; Park, J.W.; Kwon, T.K. The coffee diterpene kahweol induces apoptosis in human leukemia U937 cells through down-regulation of Akt phosphorylation and activation of JNK. *Apoptosis* **2009**, *14*, 1378–1386. [\[CrossRef\]](https://doi.org/10.1007/s10495-009-0407-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19768546)
- <span id="page-12-14"></span>96. Park, G.H.; Song, H.M.; Jeong, J.B. Kahweol from Coffee Induces Apoptosis by Upregulating Activating Transcription Factor 3 in Human Colorectal Cancer Cells. *Biomol. Ther.* **2017**, *25*, 337–343. [\[CrossRef\]](https://doi.org/10.4062/biomolther.2016.114) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27871156)
- <span id="page-12-15"></span>97. Jeon, J.S.; Kim, H.T.; Jeong, I.H.; Hong, S.R.; Oh, M.S.; Park, K.H.; Shim, J.H.; Abd El-Aty, A.M. Determination of chlorogenic acids and caffeine in homemade brewed coffee prepared under various conditions. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **2017**, *1064*, 115–123. [\[CrossRef\]](https://doi.org/10.1016/j.jchromb.2017.08.041)
- <span id="page-12-16"></span>98. Park, J.B. Isolation and quantification of major chlorogenic acids in three major instant coffee brands and their potential effects on H2O<sup>2</sup> -induced mitochondrial membrane depolarization and apoptosis in PC-12 cells. *Food Funct.* **2013**, *4*, 1632–1638. [\[CrossRef\]](https://doi.org/10.1039/c3fo60138b) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24061869)
- <span id="page-12-17"></span>99. Kim, H.G.; Hwang, Y.P.; Han, E.H.; Choi, J.H.; Kwon, K.-i.; Chung, Y.C.; Jeong, M.H.; Jeong, T.C.; Kang, W.; Jeong, H.G. The coffee diterpene kahweol inhibits metastasis by modulating expressions of MMPs and VEGF via STAT3 inactivation. *Food Chem.* **2012**, *133*, 1521–1529. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2012.02.043)
- <span id="page-12-18"></span>100. Sen, A.; Papadimitriou, N.; Lagiou, P.; Perez-Cornago, A.; Travis, R.C.; Key, T.J.; Murphy, N.; Gunter, M.; Freisling, H.; Tzoulaki, I.; et al. Coffee and tea consumption and risk of prostate cancer in the European Prospective Investigation into Cancer and Nutrition. *Int. J. Cancer* **2019**, *144*, 240–250. [\[CrossRef\]](https://doi.org/10.1002/ijc.31634) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29943826)
- <span id="page-12-19"></span>101. Mhaidat, N.M.; Alzoubi, K.H.; Al-Azzam, S.I.; Alsaad, A.A. Caffeine inhibits paclitaxel-induced apoptosis in colorectal cancer cells through the upregulation of Mcl-1 levels. *Mol. Med. Rep.* **2014**, *9*, 243–248. [\[CrossRef\]](https://doi.org/10.3892/mmr.2013.1763) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24173825)
- <span id="page-12-20"></span>102. Xu, H.; Wang, L.; Shi, B.; Hu, L.; Gan, C.; Wang, Y.; Xiang, Z.; Wang, X.; Sheng, J. Caffeine inhibits the anticancer activity of paclitaxel via down-regulation of α-tubulin acetylation. *Biomed. Pharmacother.* **2020**, *129*, 110441. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2020.110441) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32580047)
- <span id="page-12-21"></span>103. Saunders, D.E.; Lawrence, W.D.; Christensen, C.; Wappler, N.L.; Ruan, H.; Deppe, G. Paclitaxel-induced apoptosis in MCF-7 breast-cancer cells. *Int. J. Cancer* **1997**, *70*, 214–220. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1097-0215(19970117)70:2%3C214::AID-IJC13%3E3.0.CO;2-I)
- <span id="page-12-22"></span>104. Mizushima, N. Autophagy: Process and function. *Genes. Dev.* **2007**, *21*, 2861–2873. [\[CrossRef\]](https://doi.org/10.1101/gad.1599207) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18006683)
- <span id="page-12-23"></span>105. Van Limbergen, J.; Stevens, C.; Nimmo, E.R.; Wilson, D.C.; Satsangi, J. Autophagy: From basic science to clinical application. *Mucosal Immunol.* **2009**, *2*, 315–330. [\[CrossRef\]](https://doi.org/10.1038/mi.2009.20)
- <span id="page-12-24"></span>106. Udristioiu, A.; Nica-Badea, D. Autophagy dysfunctions associated with cancer cells and their therapeutic implications. *Biomed. Pharmacother.* **2019**, *115*, 108892. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2019.108892) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31029889)
- <span id="page-13-0"></span>107. Yang, Z.J.; Chee, C.E.; Huang, S.; Sinicrope, F.A. The role of autophagy in cancer: Therapeutic implications. *Mol. Cancer Ther.* **2011**, *10*, 1533–1541. [\[CrossRef\]](https://doi.org/10.1158/1535-7163.MCT-11-0047) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21878654)
- <span id="page-13-1"></span>108. Kim, Y.C.; Guan, K.-L. mTOR: A pharmacologic target for autophagy regulation. *J. Clin. Investig.* **2015**, *125*, 25–32. [\[CrossRef\]](https://doi.org/10.1172/JCI73939) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25654547)
- <span id="page-13-2"></span>109. Rabanal-Ruiz, Y.; Otten, E.G.; Korolchuk, V.I. mTORC1 as the main gateway to autophagy. *Essays Biochem.* **2017**, *61*, 565–584. [\[CrossRef\]](https://doi.org/10.1042/ebc20170027)
- <span id="page-13-3"></span>110. Lin, Y.; Jiang, M.; Chen, W.; Zhao, T.; Wei, Y. Cancer and ER stress: Mutual crosstalk between autophagy, oxidative stress and inflammatory response. *Biomed. Pharmacother.* **2019**, *118*, 109249. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2019.109249) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31351428)
- <span id="page-13-4"></span>111. Rahman, M.A.; Ahmed, K.R.; Haque, F.; Park, M.N.; Kim, B. Recent Advances in Cellular Signaling Interplay between Redox Metabolism and Autophagy Modulation in Cancer: An Overview of Molecular Mechanisms and Therapeutic Interventions. *Antioxidants* **2023**, *12*, 428. [\[CrossRef\]](https://doi.org/10.3390/antiox12020428) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36829987)
- <span id="page-13-5"></span>112. Sinha, R.A.; Farah, B.L.; Singh, B.K.; Siddique, M.M.; Li, Y.; Wu, Y.; Ilkayeva, O.R.; Gooding, J.; Ching, J.; Zhou, J.; et al. Caffeine stimulates hepatic lipid metabolism by the autophagy-lysosomal pathway in mice. *Hepatology* **2014**, *59*, 1366–1380. [\[CrossRef\]](https://doi.org/10.1002/hep.26667)
- <span id="page-13-6"></span>113. Pietrocola, F.; Malik, S.A.; Mariño, G.; Vacchelli, E.; Senovilla, L.; Chaba, K.; Niso-Santano, M.; Maiuri, M.C.; Madeo, F.; Kroemer, G. Coffee induces autophagy in vivo. *Cell Cycle* **2014**, *13*, 1987–1994. [\[CrossRef\]](https://doi.org/10.4161/cc.28929) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24769862)
- <span id="page-13-7"></span>114. Li, Y.F.; Ouyang, S.H.; Tu, L.F.; Wang, X.; Yuan, W.L.; Wang, G.E.; Wu, Y.P.; Duan, W.J.; Yu, H.M.; Fang, Z.Z.; et al. Caffeine Protects Skin from Oxidative Stress-Induced Senescence through the Activation of Autophagy. *Theranostics* **2018**, *8*, 5713–5730. [\[CrossRef\]](https://doi.org/10.7150/thno.28778) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30555576)
- <span id="page-13-8"></span>115. Erzurumlu, Y.; Çataklı, D.; Doğan, H.K.; Aydoğdu, E. Caffeine May Improve the Chemotherapeutic Effect of Docetaxel by Inducing UPR and Autophagy in Breast Cancer Cells. *FABAD J. Pharm. Sci.* **2023**, *48*, 91–104. [\[CrossRef\]](https://doi.org/10.55262/fabadeczacilik.1164699)
- <span id="page-13-9"></span>116. Liu, H.; Song, J.; Zhou, Y.; Cao, L.; Gong, Y.; Wei, Y.; Yang, H.; Tang, L. Methylxanthine derivatives promote autophagy in gastric cancer cells targeting PTEN. *Anticancer Drugs* **2019**, *30*, 347–355. [\[CrossRef\]](https://doi.org/10.1097/CAD.0000000000000724)
- <span id="page-13-10"></span>117. Benvenuto, M.; Albonici, L.; Focaccetti, C.; Ciuffa, S.; Fazi, S.; Cifaldi, L.; Miele, M.T.; De Maio, F.; Tresoldi, I.; Manzari, V.; et al. Polyphenol-Mediated Autophagy in Cancer: Evidence of In Vitro and In Vivo Studies. *Int. J. Mol. Sci.* **2020**, *21*, 6635. [\[CrossRef\]](https://doi.org/10.3390/ijms21186635)

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