



# *Review* **An Update on the Entomology, Virology, Pathogenesis, and Epidemiology Status of West Nile and Dengue Viruses in Europe (2018–2023)**

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**Abstract:** In recent decades, increases in temperature and tropical rainfall have facilitated the spread of mosquito species into temperate zones. Mosquitoes are vectors for many viruses, including West Nile virus (WNV) and dengue virus (DENV), and pose a serious threat to public health. This review covers most of the current knowledge on the mosquito species associated with the transmission of WNV and DENV and their geographical distribution and discusses the main vertebrate hosts involved in the cycles of WNV or DENV. It also describes virological and pathogenic aspects of WNV or DENV infection, including emerging concepts linking WNV and DENV to the reproductive system. Furthermore, it provides an epidemiological analysis of the human cases of WNV and DENV reported in Europe, from 1 January 2018 to 31 December 2023, with a particular focus on Italy. The first autochthonous cases of DENV infection, with the most likely vector being *Aedes albopictus*, have been observed in several European countries in recent years, with a high incidence in Italy in 2023. The lack of treatments and effective vaccines is a serious challenge. Currently, the primary strategy to prevent the spread of WNV and DENV infections in humans remains to limit the spread of mosquitoes.

**Keywords:** WNV; DENV; mosquito; pathogenesis; epidemiology; Europe

# **1. Introduction**

In the past few decades, the changing climate is thought to have affected ecosystems. This has led to changes in the natural habitats of many animal species [\[1\]](#page-15-0). As temperatures rise, the metabolism of insects increases and their life cycle shortens. In this context, the activity of mosquitoes is greater and, consequently, the feeding, due to the effect of the bite, triggers a greater transmission of several pathogens, including those of the *Orthoflavivirus* genus, which cause diseases in humans and other animals, wild or domestic [\[2\]](#page-15-1). As a result, among orthoflavivirus infection, the number of human cases of West Nile virus (WNV) infections has increased not only in the United States of America (USA) [\[3\]](#page-15-2) but also in Europe, particularly Central and Mediterranean Europe [\[4,](#page-15-3)[5\]](#page-15-4). Data from the Centers for Disease Control and Prevention's (CDC) National Arbovirus Surveillance System (ArbonET) collected for 2023 showed 2566 US WNV cases [\[5\]](#page-15-4), more than double the number reported in 2022 [\[6\]](#page-15-5). In parallel, the European Centre for Disease Prevention and



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Control (ECDC) has reported nearly 700 human cases, with the highest frequency of WNV infections in Italy (i.e., 336 human cases of WNV infections) [\[7\]](#page-15-6).

Population growth, urbanization, and travel, as well as resistance to insecticides, whether in the form of larvicides or adulticides, have been the main cause of dengue virus (DENV) human infection worldwide over the last 50 years [\[8\]](#page-15-7). In 2023, more than 6 million cases and more than 6000 dengue-related deaths were reported in 92 countries worldwide [\[9\]](#page-15-8). Since the beginning of 2024, more than 10 million DENV cases and more than 5000 dengue-related deaths have been reported globally [\[10\]](#page-15-9). In particular, Brazil is a country with approximately 3 million laboratory-confirmed cases of dengue according to the World Health Organization (WHO) [\[11\]](#page-15-10). In general, DENV is recognized as endemic in tropical and sub-tropical countries, with the majority of cases reported in Brazil, Paraguay, Argentina, and Bolivia [\[9\]](#page-15-8). However, an increasing number of autochthonous/non-travelassociated DENV cases have been described in several European countries in recent years. In particular, Italy reported the highest number of confirmed cases  $(n = 82)$  in 2023 [\[9\]](#page-15-8). As far as 2024 was concerned, a total of 283 DENV cases have been reported in Italy, all associated with travel abroad [\[12\]](#page-15-11). No deaths have been reported [\[12\]](#page-15-11).

Albeit in a small percentage of infections, WNV and DENV can be the cause of severe disease such as West Nile neuroinvasive disease (WNND) (i.e., meningitis, encephalitis, and/or acute flaccid paralysis) and severe dengue illnesses [i.e., dengue hemorrhagic fever (DHF) and dengue shock syndrome (DSS)] [\[13,](#page-15-12)[14\]](#page-15-13). The other clinical illnesses caused by WNV and DENV [such as those related to WNV fever and dengue fever (DF)] are equally significant public health concerns due to their impact on human health, the direct costs of treatment, and the indirect costs of sick leave. In addition to the clinical manifestations associated with DENV infection, other viruses and their associated diseases are transmitted by DENV vectors, such as Zika and Chikungunya [\[15\]](#page-15-14).

To date, specific antiviral treatments for human use against WNV and DENV infections have not been approved [\[16\]](#page-15-15). Several human clinical studies have been conducted with vaccines against WNV [\[17\]](#page-15-16). However, the live attenuated vaccine ChimeriVax-WN02 is the only candidate against WNV due to its high immunogenicity, safety, and tolerability observed in all-age individuals [\[18](#page-15-17)[,19\]](#page-15-18). However, vaccine efficacy and cost effectiveness are significant concerns that limit WNV vaccination programs. The only licensed vaccines to prevent DENV infections are CYD-TDV (known as Dengvaxia) and TAK-003 (known as QDenga). Specifically, the first was approved in Europe in 2019 and can be administered to individuals aged between 6 and 45 years with a history of DENV infection. TAK-003, licensed in Europe in 2022, is approved for individuals aged 4 years and older, regardless of any previous exposure to DENV. However, vaccine efficacy was demonstrated against DENV-1 and DENV-2, but not against DENV-3, while the low incidence of DENV-4 limits the evaluation against this serotype [\[20,](#page-16-0)[21\]](#page-16-1).

This review aims to summarize the entomological literature on the mosquito vectors of WNV and DENV and to assess the role of animal hosts in their transmission. Furthermore, this review aims to provide new knowledge on the virological and pathogenic aspects of DENV and WNV infections. Finally, the epidemiology of the circulation of WNV and DENV in Europe from 1 January 2018 to 31 December 2023 is described, with a particular focus on the spread of these viruses in Italy.

## **2. West Nile Virus and Dengue Virus: Insights into the Vector Competence, Host Range, Transmission, Classification, Genome Organization, Virus Genetics, and Pathogenesis** *2.1. Mosquito Vectors of West Nile Virus*

Competent vectors can transmit WNV. Mosquito anatomical barriers regulating midgut infection, midgut escape, salivary gland infection, and transmission influence WNV competence [\[22,](#page-16-2)[23\]](#page-16-3). Temperature, an extrinsic factor, significantly influences WNV vector competence. Higher temperatures enhance virus replication in mosquitoes, speeding up dissemination and shortening incubation periods [\[24\]](#page-16-4). The main mosquito genus involved in the transmission of WNV is *Culex. Culex pipiens* (Linneaus, 1823) mosquitoes

are the primary vectors of WNV due to their seasonal abundance, vector competence, and high infection rates [\[25\]](#page-16-5). They serve as amplifying vectors in the bird-to-bird enzootic cycle, as bridging vectors in the bird-to-mammalian epizootic cycle, and as reservoirs [\[26\]](#page-16-6). The vector competence of *Cx. pipiens* is similar in northern and southern Europe. Its transmission rate increases between 18  $°C$  and 28  $°C$ , suggesting a temperature-dependent vector competence [\[26\]](#page-16-6). In addition to *Cx. pipiens* and *Culex Quinquefasciatus* (Say, 1823), other mosquito vectors of WNV are *Culex univittatus* (Theobald, 1901), *Culex theileri* (Theobald, 1903), *Culex perexiguus* (Theobald, 1903), *Culex modestus* (Ficaldi, 1890), and *Culex neavei* (Theobald, 1906). Climate change is expected to spread WNV to new areas by creating favorable conditions for mosquito vectors. Extreme weather conditions such as droughts, heat waves, and floods affect the life cycle and virus-carrying capacity of mosquitoes, increasing WNV replication and transmission to humans [\[27\]](#page-16-7). Table [1](#page-3-0) shows the distribution of the main *Cx*. mosquitoes involved in the transmission of WNV.

#### *2.2. Animal Hosts and Secondary Modes of Transmission of West Nile Virus*

WNV is maintained in an enzootic transmission cycle, with mosquitoes as vectors and birds acting as amplifying hosts. Birds are considered amplifying hosts for WNV because of the high levels of viremia found in their blood [\[28\]](#page-16-8). However, WNV can infect incidental hosts, such as humans and horses [\[29\]](#page-16-9), which are generally considered dead-end hosts because viral replication does not produce significant viremia to allow transmission to feeding mosquitoes (Figure [1A](#page-3-1)) [\[28](#page-16-8)[–30\]](#page-16-10). Moreover, WNV can sporadically infect other animal species, such as dogs, cats [\[31\]](#page-16-11), wild boars [\[32\]](#page-16-12), sheep [\[33\]](#page-16-13), alpacas, llamas, and wolves [\[30\]](#page-16-10). Similarly to humans and horses, in these dead-end hosts, the viremia generated is low and not sufficient to initiate a new cycle of transmission [\[30\]](#page-16-10). Alligators have been described as susceptible to WNV infection and able to develop a viremia of sufficient magnitude to predict at least low competence to infect feeding mosquitoes [\[34](#page-16-14)[,35\]](#page-16-15). In addition to the primary mode of transmission of WNV to hosts via competent mosquito vectors, WNV can also be acquired through blood transfusion [\[36\]](#page-16-16), organ transplantation [\[37\]](#page-16-17), perinatally, and breastfeeding [\[38\]](#page-16-18). WNV, like other orthoflaviviruses, can also be maintained within the mosquito population by vertical transmission from an infected female mosquito to her offspring, even if the rate of this transmission mode is low [\[39](#page-16-19)[–42\]](#page-16-20).

#### *2.3. Mosquito Vectors of Dengue Virus*

*Aedes aegypti* (Linnaeus, 1762) is the main vector for DENV infection [\[43\]](#page-16-21). However, even with low efficiency, *Aedes albopictus* (Skuse, 1895) can become infected and transmit DENV [\[44\]](#page-16-22). While humans are a dead-end host for WNV infection, as previously shown, they are considered to be an amplifying host for DENV infection (Figure [1B](#page-3-1)) [\[45\]](#page-16-23).

The distribution of *Ae. aegypti* and *Ae. Albopictus* is shown in Table [1.](#page-3-0) At the beginning of the 20th century, *Ae. aegypti* was recorded throughout the Mediterranean basin [\[46,](#page-16-24)[47\]](#page-16-25), with high abundance in France [\[48\]](#page-16-26), Greece [\[49\]](#page-16-27), Italy [\[50\]](#page-16-28), Russia [\[51\]](#page-16-29), Portugal [\[52\]](#page-17-0), and Spain [\[53\]](#page-17-1). Although the mosquito was eradicated from the Mediterranean region, it survived along the Black Sea coast [\[54](#page-17-2)[–56\]](#page-17-3). In addition, *Ae. aegypti*, has been established also in Cyprus since 2022 and could continue to spread to other European countries in the near future [\[57\]](#page-17-4). *Ae. albopictus* may be responsible for recent autochthonous cases of DENV in Europe [\[58\]](#page-17-5). This phenomenon can be explained by suitable climatic conditions (i.e., higher temperatures and increased rainfall) for the survival of the vector, for the development of the virus in the vector, and for an adequate pool of viremic individuals. In this context, DENV has been detected in *Ae. albopictus*, and the adaptation and spread of this mosquito species in temperate regions could be the cause of recently reported autochthonous DENV cases in Europe [\[58\]](#page-17-5).

#### *2.4. Animal Hosts and Secondary Modes of Transmission of Dengue Virus*

DENV is maintained by a sylvatic cycle involving non-human primates [\[59\]](#page-17-6) (Figure [1B](#page-3-1)). In rural areas, the spread of DENV to humans can occur when human populations come

into contact with the sylvatic cycle [\[60\]](#page-17-7). The RNA of the DENV genome has been found in various non-human primates [\[61](#page-17-8)[,62\]](#page-17-9), i.e., birds [\[63\]](#page-17-10), bats [\[62\]](#page-17-9), rodents [\[64\]](#page-17-11), dogs [\[65\]](#page-17-12), pigs [\[66\]](#page-17-13), cattle [\[67\]](#page-17-14), horses [\[68\]](#page-17-15), and marsupials [\[69\]](#page-17-16). Although DENV is maintained primarily by cross-species transmission between mosquito vectors and vertebrate hosts, vertical transmission of this virus within the mosquito vector population has been proposed as a maintenance mechanism [\[70\]](#page-17-17), supporting the persistence of DENV in the absence of a recognized host or under unfavorable conditions for mosquito activity [\[70\]](#page-17-17). In particular, DENV is transmitted vertically by transovarial transmission, where the virus infects the gonadal tissues of female mosquitoes, and by trans-egg transmission, where the virus infects the eggs during oviposition [\[71,](#page-17-18)[72\]](#page-17-19). DENV has a significantly higher effective vertical transmission rate in *Ae. albopictus* than in *Ae. aegypti* mosquitoes [\[70\]](#page-17-17). Specifically, DENV-1, one of the four DENV serotypes (i.e., DENV-1, DENV-2, DENV-3, and DENV-4) distinguished by genome sequence variation is better able to transmit vertically than the other DENV serotypes, but the explanation for this serotype-specific phenomenon is still unknown [\[70\]](#page-17-17).

<span id="page-3-0"></span>**Table 1.** Mosquito vector species for West Nile virus and dengue virus and their geographical distribution.



The main mosquito vector species for West Nile virus (WNV) and dengue virus (DENV) infections are shown, together with their geographical distributions. \* Currently, *Ae. aegypti* is present on the Black See Coast [\[56\]](#page-17-3) and since 2022 has been established in Cyprus, indicating a potential future spread of this mosquito across Europe [\[57\]](#page-17-4).  $\mathcal{L}_{1}$  virtual magnitude to predict at least low competence to infect feeding  $\mathcal{L}_{2}$ 

<span id="page-3-1"></span>

Figure 1. Life cycles of West Nile virus and dengue virus. (A) The life cycle of West Nile virus (WNV) is shown. The amplifying host is represented by a bird in which, after viral replication, the viral load in the blood is high enough to infect *Culex* mosquitoes. Humans and horses are considered the main hosts of WNV. They are known as dead-end hosts because their blood viremia is not high enough to infect other mosquitoes. (**B**) The sylvatic and urban cycles of dengue virus (DENV) are shown. In the sylvatic cycle, DENV is maintained mainly by *Aedes* mosquitoes and monkeys. The transition from the sylvatic to the urban cycle occurs in two ways: (i) DENV-infected mosquitoes move directly from the sylvatic to the urban area and infect humans; (ii) DENV-infected mosquitoes bite humans living in rural areas, who then move to urban areas. DENV is maintained in the urban cycle by *Ae. aegypti* mosquitoes and humans. The above image was created with <https://www.biorender.com/> (accessed on 15 July 2024).

#### *2.5. Classification*

The *Flaviviridae* family consists of enveloped viruses characterized by a positive singlestranded (ss)RNA genome and an icosahedral nucleocapsid. Four genera have been identified within this family: *Orthoflavivirus*, *Pestivirus*, *Hepacivirus*, and *Pegivirus*. *The Flavivirus* genus, known for its many arthropod-borne viruses, was renamed orthoflavivirus by the International Committee on Taxonomy of Viruses (ICTV) in 2023 to distinguish it from other members of the *Flaviviridae* family [\[82\]](#page-18-3).

Orthoflavivirus comprises more than 70 viruses, most of which are vector-borne pathogens, and are grouped into serocomplexes based on serological characteristics, including DENV, WNV, Japanese encephalitis virus (JEV), yellow fever virus (YFV), Zika virus (ZIKV), and USUTU virus (USUV) [\[83\]](#page-18-4).

USUV is considered an emerging orthoflavivirus and its amplifying host is the bird. A meta-analysis study conducted in Modena (northeast Italy) revealed the presence of USUV RNA in the cerebrospinal fluid of a high proportion of people, suggesting that USUV cases in humans may not be a sporadic event [\[84\]](#page-18-5).

Tick-borne encephalitis virus (TBEV), another orthoflavivirus, has caused epidemics across various Asian and African countries. Cases of TBEV infection have also been reported in several European countries, including Austria, Germany, Switzerland, France, Finland, Norway, Denmark, Slovenia, Romania, Hungary, Poland, Ukraine, and Russia [\[85\]](#page-18-6).

In addition, recent studies have identified novel orthoflaviviruses such as Aripo virus in Trinidad [\[86\]](#page-18-7), Menghai orthoflavivirus in China [\[87\]](#page-18-8), and Hanko virus in northern Europe [\[88\]](#page-18-9). Further details on the virological characteristics and genetic variability of notable orthoflaviviruses, such as WNV and DENV, are reviewed in the following sections.

#### *2.6. Genome and Replicative Cycle of West Nile Virus and Dengue Virus*

The genome of WNV and DENV is a positive (ss)RNA of approximately 11 kilobases (Kb) containing a single open reading frame (ORF) encoding a polyprotein [\[89\]](#page-18-10) and two non-coding regions (NCR) located at 5′ and 3′ [\[89\]](#page-18-10). The polyprotein is cleaved into 10 viral proteins by viral and host proteases [\[90\]](#page-18-11). The 5′ terminal NCR encompasses RNA secondary structures, necessary for the translation initiation, the stem loop A (SLA), essential for genome replication, and the internal ribosome entry site (IRES) [\[91\]](#page-18-12). After binding to a cellular receptor, viral particles enter the cells by endocytosis and release the nucleocapsid into the cytosol. Once uncoated, the genome is immediately translated into a large polyprotein. Replication of the genome and the formation of new viral particles occur in the endoplasmic reticulum. Lastly, the virions are released from the cell by exocytosis (Figure [2\)](#page-5-0) [\[92\]](#page-18-13).

ssRNA+

<span id="page-5-0"></span>

**Figure 2. Orthoflavivirus replicative cycle.** There is an interaction between the viral envelope **Figure 2. Orthoflavivirus replicative cycle.** There is an interaction between the viral envelope part and of West Nile and depittative virus receptors and various receptors expressed on the cell membrane virus proteins of West Nile and dengue viruses and various receptors expressed on the cell membrane [e.g., dendritic cell-specific ICAM-3 grabbing non-integrin (DC-SIGN), mannose receptor, and glycosaminoglycans (GAGs)] that mediates endosome formation. Once the endosome is formed, fusion between membranes occurs, resulting in the release of the positive single-stranded (ss)RNA viral genome into the cytosol. The viral genome contains a 5′ internal ribosome entry site (IRES) secondary RNA structure that allows immediate translation into a large polyprotein by the host ribosomes in the endoplasmic reticulum (ER). The polyprotein is then cleaved by proteases into 3 structural proteins and 7 non-structural proteins, including RNA-dependent RNA polymerase, which is responsible for genome replication using a negative (ss)RNA template in the ER. In the ER, virions are assembled and acquire an envelope, but remain immature. The immature virions then move from the ER to the Golgi, where viral maturation takes place. Specifically, in the Golgi, immature virions are processed by the cellular Furin protease, which cleaves the enveloped proteins, resulting in mature, infectious virions. Finally, the viral particles are released by exocytosis. The above figure was created with <https://www.biorender.com/> (accessed on 15 July 2024).

#### *2.7. Genetic Variability of West Nile Virus*

WNV can be distinguished in nine different lineages, but only lineages 1 (WNV-1) and 2 (WNV-2) are of public health interest due to their widespread distribution across continents and the association with human neuroinvasive disease [\[93\]](#page-18-14). Specifically, three clades, namely, in alphabetic order, A, B, and C, can be considered for WNV-1 that are distributed in different geographical regions, as shown in Figure [2](#page-5-0) [\[94\]](#page-18-15). Originally discovered in Uganda in 1937, WNV-2 remained limited to sub-Saharan Africa and Madagascar until the early 2000s (Figure [3\)](#page-6-0) [\[95\]](#page-18-16). However, following its outbreak in Hungary and southern Russia [\[94](#page-18-15)[,96\]](#page-18-17), WNV-2 spread in Europe, causing outbreaks in northern Greece (2010) and in northeastern Italy (2011) [\[97–](#page-18-18)[99\]](#page-18-19). The migration of birds from African countries to Europe during this period is thought to have caused these outbreaks [\[99,](#page-18-19)[100\]](#page-18-20). Other WNV lineages

Golgi apparatus

include WNV-3, WNV-4, WNV-5, and WNV-6. The former was isolated on the border between the Czech Republic and Austria from *Cx. pipiens* and *Aedes rossicus* (Dolbeskin, Gorickaja, and Mitrofanova, 1930) and is named Rabensburg virus (RABV) [\[101\]](#page-18-21). WNV-4 is a unique virus that was isolated from ticks in the northwestern Caucasus Mountains of Russia in 1998 [\[102\]](#page-18-22). WNV-5, referred to as the 1c clade, was isolated in India [\[103\]](#page-18-23), while WNV-6 is putative and has been found in Spain [\[104\]](#page-19-0). WNV-7, known as Koutango virus (WN-KOUTV), has been isolated from ticks and rodents and is confined to Africa [\[100\]](#page-18-20). In addition, putative WNV lineages (WNV-8 and WNV-9) have been identified in *Culex perfuscus* (Theobald, 1903) mosquitoes in Kedoungou and *Uranotaenia unguiculata* (Edwards, 1913) mosquitoes in Austria, respectively [\[105\]](#page-19-1). Among WNV lineages, there is more than 85% amino acid sequence identity in the polypeptide, and the NS region is highly conserved *Trop. Med. Infect. Dis.* **2024**, *9*, 166 8 of 26 (Figure [3A](#page-6-0)).

<span id="page-6-0"></span>

**Figure 3. Taxonomy of West Nile and dengue viruses.** (A). The lineages of West Nile virus (WNV) are shown. The first two lineages are found in several regions of the world, while other lineages consist of a few strains that are restricted to specific areas and are not associated with severe disease. (B). The serotypes and genotypes of dengue virus (DENV) are shown. DENV is classified into 4 serotypes, each of which can be further subdivided into genotypes (with a maximum of 6% genetic *2.8. Genetic Variability of Dengue Virus*  (accessed on 15 July 2024).  $T_{\rm eff}$ diversity within the same genotype). The above figure was created with <https://www.biorender.com/>

# [106], are subdivided into several genotypes on the basis of the genome analysis of *2.8. Genetic Variability of Dengue Virus*

envelope region sequencing. Alongside the four distinct serotypes of DENV, an additional The four serotypes of DENV, which are DENV-1, DENV-2, DENV-3, and DENV-4 [\[106\]](#page-19-2), are subdivided into several genotypes on the basis of the genome analysis of envelope region sequencing. Alongside the four distinct serotypes of DENV, an additional DENV-5 serotype has been detected in a Malaysian man in 2013 [\[107\]](#page-19-3). However, the DENV-5 serotype circulates among non-human primates and survives in a sylvatic spreading cycle, suggesting a possible DENV spillover into humans [\[107\]](#page-19-3).

Concerning DENV-1, it includes five genotypes  $[108]$  (I–V): (i) genotype I is found in southeast Asia, China, and the Middle East; (ii) genotype II is widespread in Thailand; (iii) genotype III is characterized by sylvatic strains from Malesia; (iv) genotype IV refers

to strains from Pacific Rim countries, Australia, and Western Pacific Islands; (v) genotype V has been isolated from Asia, West Africa, and the Americas. As far as DENV-2 is concerned [\[109\]](#page-19-5), it is clustered in (i) the Asian genotype, subdivided into Asian 1 (Thailand and Malesia) and Asian 2 (Philippines, Sri Lanka, Vietnam, Taiwan, and China); (ii) the cosmopolitan genotype of Australia, East and West Africa, the Pacific and Indian Ocean islands, the Indian subcontinent, and the Middle East; (iii) the American genotype, isolated in Latin America and the Caribbean; (iv) the southeast Asian/American genotype of Vietnam, Thailand, and several tribes in the Americas; (v) the sylvatic genotype, widespread in West Africa and southeast Asia. Concerning DENV-3, it comprises four genotypes [\[110\]](#page-19-6) (I–V): (i) genotype I, widespread in the South Pacific islands, Malaysia, Indonesia, and the Philippines; (ii) genotype II strains, found in Thailand, Vietnam, and Bangladesh; (iii) genotype III strains, circulating in Sri Lanka, India, Africa, and Samoa; (iv) genotype IV strains, isolated in Puerto Rico, Latin and Central America, and the 1965 Tahiti strain; (v) genotype V strains, detected in Brazil and Colombia. The last DENV-4 strains can be grouped into five genotypes [\[111\]](#page-19-7) (I–V): (i) genotype I strains, found in Japan, the Philippines, Thailand, and Sri Lanka; (ii) genotype II strains, found in the Americas, the Caribbean, Indonesia, Malaysia, and Tahiti; (iii) genotype III strains, found in Thailand; (iv) genotype IV strains, found in Malaysia; (v) genotype V, found in India (Figure [3B](#page-6-0)).

#### *2.9. Insight into the Tropism and Pathogenesis of West Nile Virus*

Pathogenesis of WNV is shown in Figure [4.](#page-8-0) Specifically, WNV efficiently establishes its replication on cells that generally express the attachment factors, the adhesion molecule of dendritic cells (DC-SIGN), and the dendritic cell-specific ICAM-3 grabbing nonintegrinrelated (DC-SIGNR) [\[112,](#page-19-8)[113\]](#page-19-9). One week after virus inoculation, WNV is cleared from the blood and peripheral organs and migrates to the central nervous system (CNS). The mechanism by which WNV crosses the blood–brain barrier (BBB) is not fully understood. It has been shown that higher plasma viremia correlates positively with viral entry into the brain [\[114\]](#page-19-10). Several pro-inflammatory cytokines, produced during peripheral immune responses, are involved in the modulation of the BBB. Toll-like receptor 3 (TLR3)-deficient mice, which produce reduced levels of tumor necrosis factor-alpha (TNF-alpha), had less development of neuroinvasive forms compared to wild-type mice [\[115\]](#page-19-11). Alongside the compromission of the BBB, WNV can enter the CNS thought alternative mechanisms (Figure [4\)](#page-8-0). In the CNS, neurons, astrocytes, and microglial cells represent target cells for WNV infection, as observed in in vitro studies and in autopsied neural tissues of WNV patients with encephalomyelitis [\[116](#page-19-12)[–120\]](#page-19-13). However, differences can be observed between neuronal and glial WNV-infected cells. While rapid WNV replication kinetics and consequent cell death, predominantly apoptosis [\[121\]](#page-19-14), characterize neuronal cells, slow infection and continuous production of infectious particles are typical of WNV-infected astrocytes [\[122\]](#page-19-15). The death of neurons after WNV infection may be also due to neuronophagia, a mechanism by which inflammatory cells, predominantly microglia, phagocytize dying WNV neurons' infected cells [\[123\]](#page-19-16). In general, WNV infection in humans is characterized by a low viral load [<100 plaque-forming unit (PFU)/mL] and a short duration of viremia, occurring one to three days after infection and lasting up to 11 days [\[124\]](#page-19-17). However, cases of prolonged infection have been recorded in immunosuppressed patients infected with WNV, where detectable viremia persisted for more than 60 days [\[125\]](#page-19-18). Clinical manifestations associated with WNV infection are shown in Figure [5](#page-9-0) [\[126,](#page-19-19)[127\]](#page-19-20). In addition to the known cycle of WNV between blood-feeding mosquitoes and vertebrate hosts, the potential of this virus to be sexually transmitted has been recognized (Table [2\)](#page-10-0).

#### *2.10. Insight into the Tropism and Pathogenesis of Dengue Virus*

Insights into the pathogenesis of DENV are shown in Figure [4.](#page-8-0) DENV is able to bind an extensive panel of molecules, in agreement with the wide range of cell types that are susceptible to infection. A variety of host receptor candidates has been proposed, including glycosaminoglycans (GAGs, i.e., heparan sulfate and lectins), the DC-SIGN, mineralo-

corticoid receptor (MR) of macrophage, and the lipopolysaccharide (LPS) receptor CD14. Moreover, heath shock protein (HSP) 70 and 90 and T-cell/transmembrane, immunoglobulin, and mucin (TIM)-Tyro3, AXL, and MerTK (TAM) are other cellular receptors that DENV uses to promote infection [\[128,](#page-19-21)[129\]](#page-19-22). This non-specific targeting allows the virus to disseminate widely throughout the host, leading to the broad range of disease manifestations seen in DENV-infected patients [\[130\]](#page-19-23). The pathogenesis of DENV infection encompasses a wide spectrum of dengue disease severity, ranging from mild DF to severe DHF and DSS. DENV viremia is significantly higher in patients with DHF than in those with dengue at one week after symptom onset, suggesting a strong association with disease severity [\[131\]](#page-20-0). Severe manifestations of DENV infection are likely to result from a variety of immunopathogenic mechanisms involving viral and host factors. Specifically, hemorrhagic fever is characterized by elevated levels of pro-inflammatory cytokines, referred to as a "cytokine storm",<br>in and host factors. Specifically, here is considered by the specifically specifically in and the specifically similar to what has been shown for coronavirus disease 19 (COVID-19) [\[132](#page-20-1)[,133\]](#page-20-2). The cytokine storm is characterized by increased plasma levels of TNF-alpha, IL-6, IL-8, IL-10, IL-12, and matrix metalloproteases that are produced by T cells, monocytes, macrophages, and mast cells, which could increase vascular permeability and contribute to hemor-rhagic fever [134-[137\]](#page-20-4). One of the risk factors for the development of severe forms is pre-existing antibodies to DENV. This phenomenon, namely, antibody-dependent enhancement (ADE), is shown in Figure [6.](#page-10-1) Alongside the primary transmission mode of DENV via (ADE), is shown in Figure 6. Alongside the primary transmission mode of DENV via mosquito vectors, there are striking recent data concerning the sexual transmissibility of this mosquito vectors, there are striking recent data concerning the sexual transmissibility of viru[s \(](#page-10-0)Table 2). antibodies to DENV. This phenomenon, namely, antibody-dependent enhancement

<span id="page-8-0"></span>

**Figure 4. Pathogenesis of West Nile virus (WNV) and dengue virus (DENV).** When WNV and **Figure 4. Pathogenesis of West Nile virus (WNV) and dengue virus (DENV).** When WNV and  $\overline{\mathbf{V}}$  are in our lated into the skin, both viruses primarily infect Langerhans dendritic DENV are inoculated into the skin, both viruses primarily infect Langerhans dendritic cells and

keratinocytes [\[138,](#page-20-5)[139\]](#page-20-6). Regarding WNV infection, these cells migrate to the lymph nodes and cause primary viremia. Similarly, during DENV infection, Langerhans dendritic cells and keratinocytes migrate in the local draining lymph nodes, where monocytes and macrophages are also recruited and become targets of infection [\[138,](#page-20-5)[139\]](#page-20-6). Primary viremia facilitates widespread virus infection across multiple organs. WNV can replicate in various anatomical sites, including the spleen, liver, and kidney, prior to invading the CNS [\[140\]](#page-20-7). Concerning DENV, its presence was found in spleen, kidney, peripheral blood, bone marrow, and liver [141]. After peripheral dissemination, WNV crosses kidney, peripheral blood, bone marrow, and l[iver](#page-20-8) [141]. After peripheral dissemination, WNV the blood-brain barrier (BBB) and causes damage to the CNS. The proposed routes of entry into the CNS of WNV are by passive transport through the endothelium or choroid plex, transport through olfactory neurons, transport through immune-infected cells (Trojan horse), and axonal through olfactory neurons, transport through immune-infected cells (Trojan horse), and axonal retrograde transport from infected peripheral neurons [\[142\]](#page-20-9). The development of severe DENV retrograde transport from infected peripheral neurons [142]. The development of severe DENV disease depends on viral virulence factors and an adverse host immune response that collectively disease depends on viral virulence factors and an adverse host immune response that collectively results in abnormal hemostasis and increased vascular permeability [\[139\]](#page-20-6). The above figure was created with <https://www.biorender.com/> (accessed on 15 July 2024). resease depends on viral viralence factors and an adverse host minimity response that concentrely

<span id="page-9-0"></span>

**Figure 5. Clinical manifestation of West Nile virus (WNV) and dengue virus (DENV) infection**. **Figure 5. Clinical manifestation of West Nile virus (WNV) and dengue virus (DENV) infection**. The The majority of WNV infections (80%) are asymptomatic, while the remaining of infections (20%) majority of WNV infections (80%) are asymptomatic, while the remaining of infections (20%) present present with flu-like symptoms (i.e., headache, myalgia, fever, maculopapular rash, and with flu-like symptoms (i.e., headache, myalgia, fever, maculopapular rash, and gastrointestinal with he me symptoms (i.e., neadache, myangia, revel, macatopapatar rash, and gastromestinal symptoms) [\[143,](#page-20-10)[144\]](#page-20-11). Less than 1% of WNV-infected patients develop neuroinvasive diseases such as meningitis and meningoencephalitis and, among them,  $10\%$  had fatal outcomes [ $144,145$ ]. People who are at a greater risk of developing severe WNV infection are the elderly and immune-compromised individuals [146]. Generally, DENV infections are asymptoma[tic. H](#page-20-13)owever, DENV-infected patients may develop symptoms ranging from dengue fever (DF, characterized by mild flu-like syndrome) to dengue hemorrhagic fever (DHF, marked by decreased circulating plasma volume) and dengue shock syndrome (DSS, distinguished by multi-organ failure) [\[147\]](#page-20-14). A severe outcome related to DENV infection depends on previous exposure to the virus: heterotypic infections with a different DENV infection, depends on previous or posses to die virus necessity risk for the above in above in a  $\sim$ serotype, in comparison to the primary infection, pose the risk for individuals to develop severe illnesses [\[148\]](#page-20-15). The above image was created with <https://www.biorender.com/> (accessed on 15 July 2024).

<span id="page-10-1"></span>

Antibody-dependent enhancement (ADE)

**Figure 6. Antibody-dependent enhancement (ADE).** This phenomenon can be explained by a **Figure 6. Antibody-dependent enhancement (ADE).** This phenomenon can be explained by a subset of IgG targeting viral proteins of one dengue virus (DENV) serotype cross-reacting with those of other serotypes and resulting in a poor neutralizing function [\[149](#page-20-16)[–153\]](#page-20-17). The antibody–DENV complex binds to the Fcγ receptors (FcγR) on circulating phagocytes (i.e., monocytes), facilitating the infection [\[154\]](#page-20-18). This process enhances viral replication and triggers excessive cytokine production. Moreover, ADE initiates an immune cascade that leads to severe dengue disease.

<span id="page-10-0"></span>**Table 2.** Detection of West Nile and dengue virus in the reproductive tract.



Studies on WNV and DENV localization in the reproductive tract are shown.

# **3. A Six-Year Epidemiological Report on West Nile Virus and Dengue Virus in Europe (2018–2023)**

Epidemiological data on the circulation of WNV and DENV in Europe were obtained from the ECDC. Specifically, data for WNV were obtained from the "Annual Epidemiological Reports (AERs) for 2018 and 2019" [\[168,](#page-21-9)[169\]](#page-21-10), "Epidemiological update: West Nile virus transmission season in Europe for 2020, 2021 and 2022" [\[170–](#page-21-11)[172\]](#page-21-12) and "Weekly updates: 2023 West Nile virus transmission season for 2023" [\[173\]](#page-21-13). Data on DENV circulation in Europe were obtained from the "Surveillance Report Dengue annual epidemiological report for 2018, 2019, 2020, 2021 and 2022" [\[174–](#page-21-14)[178\]](#page-21-15) and from the "Autochthonous vectorial transmission of dengue in Europe Union/European Economic Area (EU/EEA)" [\[179\]](#page-21-16) section of the ECDC website for 2023. For Italy, the number of DENV infections was collected from the Istituto Superiore di Sanità reports until 2023 ("Bollettini periodici arbovirosi") [\[180\]](#page-21-17).

# *3.1. The Number of West Nile Virus Cases in Europe from 2018 to 2023*

All WNV cases in the European Union (EU) are shown in Figure [7.](#page-11-0) Considering all EU cases from 2018 to 2023, the number of WNV cases decreased from 2018 to 2021, while a new peak of cases was observed in 2022 [\[168–](#page-21-9)[172\]](#page-21-12). Between 2018 and 2023, 406 deaths were reported among cases of WNV infection, with a peak registered in 2018, which corresponds to the highest number of WNV human cases reported in that year (Figure [7\)](#page-11-0) [\[168](#page-21-9)[–173\]](#page-21-13). In European countries where WNV is endemic, COVID-19 restrictions had little or no

effect. This is consistent with what has been observed in avian surveillance, where it coincides with an epidemic in these animals [\[181\]](#page-22-0). Over the years, most of the cases have been observed in Mediterranean EU countries, including Italy and Greece, where summer temperatures are usually high, allowing mosquitoes to spread. Interestingly, WNV was not detected in both birds and humans in Germany before 2018, while the first WNV cases were observed in birds but not in humans in 2018 [\[182\]](#page-22-1). In addition, two outbreaks of WNV in horses were observed in September and October 2018 [\[183\]](#page-22-2). Insecticide resistance in *Cx. pipiens* mosquitoes has also been reported in Europe, posing a significant threat to the spread of mosquito-borne disease vectors. Recent data examining the evolution and spread of WNV in Europe suggest that, in addition to ecological conditions favoring the presence of birds and mosquitoes, the intensity of agricultural activity may be an important driver of WNV emergence and spread [\[184\]](#page-22-3).

<span id="page-11-0"></span>

**Figure 7. The number of cases and deaths due to West Nile virus (WNV) infection in European Figure 7. The number of cases and deaths due to West Nile virus (WNV) infection in European Union countries from 2018 to 2023.** WNV cases and deaths for 2018 (A), 2019 (B), 2020 (C), 2021 (D), 2022 (**E**) and 2023 (**F**) are shown. Data were collected from the European Centre for Disease 2022 (**E**) and 2023 (**F**) are shown. Data were collected from the European Centre for Disease Prevention  $\sum_{i=1}^{\infty}$  reports up december 2023 ( $\sum_{i=1}^{\infty}$   $\sum_{i=1}^{\infty}$ and Control (ECDC) reports updated on 31 December 2023 ("West Nile virus infection—Annual epidemiological report for 2018", 2019; "West Nile virus infection—Annual epidemiological report 2020", 2021; "Epidemiological update: West Nile virus transmission season in Europe, 2021", 2022; for 2019", 2020; "Epidemiological update: West Nile virus transmission season in Europe, 2020", 2021;  $\mathbf{E} = \mathbf{E} \cdot \mathbf{W} \cdot \mathbf{W} \cdot \mathbf{W}$ "Epidemiological update: West Nile virus transmission season in Europe, 2021", 2022; "Epidemiologi-*3.2. The Number of Dengue Virus Cases in Europe from 2018 to 2023: The Emergence of Dengue*  Nile virus transmission season, 2023"). cal update: West Nile virus transmission season in Europe, 2022", 2023; "Weekly updates: 2023 West

### 3.2. The Number of Dengue Virus Cases in Europe from 2018 to 2023: The Emergence of Dengue traveled abroad (Figure 8). Most of the DENV cases in the EU were observed in 2019; in *Virus Autochthonous Cases*

DENV cases in Europe are mainly imported cases, due to infected people who have  $y_{\text{old}}$  abroad (Figure 8), Most of the DENN cases in the FU was absorped traveled abroad (Figure [8\)](#page-12-0). Most of the DENV cases in the EU were observed in 2019;

in parallel, DENV epidemics were reported in many countries around the world in the same year, including Bangladesh [\[185\]](#page-22-4), Nepal [\[186\]](#page-22-5), and Honduras [\[187\]](#page-22-6). Besides the recent impact of the spread of SARS-CoV-2 on the circulation of DENV, autochthonous cases have been reported in Europe over the past decade, probably due to the estab-*Trop. Med. Infect. Dis.* **2024**, *9*, 166 14 of 26 lishment of *Ae. albopictus* on the continent. The first autochthonous cases of DENV infection in the EU were described in 2010, in Croatia and France (15 and 2 cases, respec-tively) [\[188,](#page-22-7)[189\]](#page-22-8). Several human cases of autochthonous DENV infection were then reported in France until 2015 (1 case in 2013, 4 cases in 2014, and 8 cases in 2015) [\[190](#page-22-9)[,191\]](#page-22-10). In 2018, 6 autochthonous cases of DENV were reported in France [\[192\]](#page-22-11), and it was also the first year in which autochthonous DENV cases were reported in Spain [\[193\]](#page-22-12); in particular, 3 cases ne mathematic and EDIV was contract person in Span (1920), at particular, could be expected in August (Autonomous Communities of Murcia and Andalusia) (none of the DENV-infected individuals had traveled to endemic areas in the previous 2 weeks before symptom onset) and the other 3 cases in October, geographically close to the first cases (Autonomous Community of Murcia). Sequencing showed that DENV-1 caused the outbreak in Spain. However*,* no *Ae. albopictus* mosquitoes were positive for DENV-RNA detection. One novel DENV human infection was reported in Germany, but this was linked to an infection in a virology laboratory [\[175\]](#page-21-18). In 2022, 66 autochthonous vector transmissions of DENV were observed in mainland EU/EEA, including 60 and 6 cases in France and Spain, respectively [\[178\]](#page-21-15). In 2023, a slight decrease of DENV in non-travel-associated infections was recorded, with 45 cases in France and 3 cases in Spain [\[179\]](#page-21-16). autochthonous Communities of Murcha and Amadriaship (none  $\alpha$  as a recorded, which  $45 \text{ cases in frame and 5 cases in Spin (177)}.$ 

Italy reported its first autochthonous cases in 2020 (see further section below). In Italy reported its first autochthonous cases in 2020 (see further section below). In 2021, there was a general decrease in human DENV infections, probably due to COVID-2021, there was a general decrease in human DENV infections, probably due to COVID-19 travel restrictions, but 2 new autochthonous cases were identified in France [\[194\]](#page-22-13). In 19 travel restrictions, but 2 new autochthonous cases were identified in France [194]. In contrast to 2022, when no autochthonous cases were reported in Italy, a new peak of 82 non-travel-associated DENV infections was recorded in 2023 [195–198]. non-travel-associated DENV infections was recorded [in 20](#page-22-14)[23 \[1](#page-22-15)95–198].

<span id="page-12-0"></span>

**Figure 8. Total and autochthonous dengue virus (DENV) cases in European Union (EU) countries**. **Figure 8. Total and autochthonous dengue virus (DENV) cases in European Union (EU) countries**. Total and autochthonous DENV cases for 2018 (A), 2019 (B), 2020 (C), 2021 (D), 2022 (E) and 2023 (F)

are shown. All DENV cases, including autochthonous and imported cases, found in Europe between 2018 and 2022 were collected from the European Centre for Disease Prevention and Control (ECDC) reports ("Surveillance Report Dengue annual epidemiological report for 2018", 2019; "Surveillance Report Dengue annual epidemiological report for 2019", 2021; "Surveillance Report Dengue annual epidemiological report for 2020", 2022; "Surveillance Report Dengue annual epidemiological report Dengue annual epidemiological report for 2020", 2022; "Surveillance Report Dengue annual for 2021", 2023; "Surveillance Report Dengue annual epidemiological report for 2022", 2024). Annual ECDC reports on the total number of cases in Europe, the sum of travelers returning from dengueendemic areas, and autochthonous cases for 2023 are not yet available. The only available data on imported cases of DENV infections for 2023 are for France and Italy, respectively, from the "Santè publique France" and "Istituto Superiore di Sanità" websites [\[197,](#page-22-16)[199\]](#page-22-17). In parallel, data on autochthonous DENV infections in Europe in 2023 are summarized on the ECDC website in the section "Autochthonous vectorial transmission of dengue in EU/EEA" [\[179\]](#page-21-16). returning from dengths of the total funder or cases in Europe, the sum or thevelets featuring from dength

#### *3.3. West Nile Virus Circulation in Italy 3.3. West Nile Virus Circulation in Italy*  Human WNV infections from 2018 to 2023 are shown in Figure 9. Analyses of *Cx.*

Human WNV infections from 2018 to 2023 are shown in Figure [9.](#page-13-0) Analyses of *Cx. pipiens* mosquitoes, the vector for WNV in Italy, collected from 2018 to 2020 showed the circulation of WNV-2 in Italy during these years. The first circulations in Italy of both WNV-1 and WNV-2 were detected in human WNV infections in 2021. In particular, WNV-1 human infections were reported in 2021 in the provinces of Padua (n = 2) and Vicenza  $(n = 1)$  [\[200\]](#page-22-18). In the same year, WNV-2 was detected in a patient with neuroinvasive disease and in a blood donor  $[200]$ . In 2022, the presence of WNV-1 was confirmed in  $\sim$ 2 human cases (i.e., a blood donor and a patient with encephalitis) in the province of Padua. With regard to WNV-2, in 2022, one WNV nucleic acid test (NAT)-positive blood regard to WNV-2, in 2022, one WNV nucleic acid test (NAT)-positive blood donor was donor was identified in the province of Venice. Regarding the circulation of WNV lineages in 2023, veterinary surveillance conducted on horses, mosquitoes, and birds confirmed in 2023, veterinary surveillance conducted on horses, mosquitoes, and birds confirmed the presence of WNV-2 in northern Italy (Emilia Romagna, Friuli Venezia Giulia, Liguria, presence of WNV-2 in northern Italy (Emilia Romagna, Friuli Venezia Giulia, Liguria, Marche, Veneto, Lombardy, and Piedmont), as well as in Apulia, Sardinia, and Sicily [\[201\]](#page-22-19). Marche, Veneto, Lombardy, and Piedmont), as well as in Apulia, Sardinia, and Sicily [201]. At the same time, the circulation of WNV-1 was restricted to Emilia-Romagna, Sicily, Veneto, At the same time, the circulation of WNV-1 was restricted to Emilia-Romagna, Sicily, and Campania [\[201\]](#page-22-19). *pipiens* most most most most vector from 2018 to 2023 are shown in Figure 9. Analyses of C  $\mu$  identified in the province of Venice. Regarding the circulation of WNV lineages in 2023,  $\mu$ 

<span id="page-13-0"></span>

**Figure 9.** West Nile virus (WNV) cases in Italy from 2018 to 2023 according to clinical manifestations **Figure 9.** West Nile virus (WNV) cases in Italy from 2018 to 2023 according to clinical manifestations (fever, neuroinvasive disease, and cases in blood donors). All data were collected from reports of (fever, neuroinvasive disease, and cases in blood donors). All data were collected from reports of the "Istituto Superiore di Sanità" ("Sorveglianza integrata del West Nile e Usutu virus 2018", 2018; the "Istituto Superiore di Sanità" ("Sorveglianza integrata del West Nile e Usutu virus 2018", 2018; "Sorveglianza integrata del West Nile e Usutu virus 2019", 2019; "Sorveglianza integrata del West "Sorveglianza integrata del West Nile e Usutu virus 2019", 2019; "Sorveglianza integrata del West Nile e Usutu virus 2020", 2020; "Sorveglianza integrata del West Nile e Usutu virus 2021", 2021; Nile e Usutu virus 2020", 2020; "Sorveglianza integrata del West Nile e Usutu virus 2021", 2021; "Sorveglianza integrata del West Nile e Usutu virus 2022", 2022; "Sorveglianza integrata del West Nile e Usutu virus 2023", 2023). \* There were 42 deaths reported in 2018; ● There were 5 deaths "Sorveglianza integrata del West Nile e Usutu virus 2022", 2022; "Sorveglianza integrata del West Nile e Usutu virus 2023", 2023).  $*$  There were 42 deaths reported in 2018;  $*$  There were 5 deaths reported in 2019; ▲ There were 37 deaths reported in 2022.

# *3.4. Dengue Virus Circulation in Italy 3.4. Dengue Virus Circulation in Italy*

Data on DENV infections in Italy from 2018 to 2023 are shown in Figure 10. DENV Data on DENV infections in Italy from 2018 to 2023 are shown in Figure [10.](#page-14-0) DENV infections in Italy are mainly imported as the mosquito vector of DENV (Ae. aegypti) is present in the country. Notably, the first autochthonous cases in Italy, likely due to the not present in the country. Notably, the first autochthonous cases in Italy, likely due to the presence of *Ae. albopictus* in Italy, were identified in August 2020. In particular, a family presence of *Ae. albopictus* in Italy, were identified in August 2020. In particular, a family cluster of DENV infections was reported in the province of Vicenza (northeastern Italy), cluster of DENV infections was reported in the province of Vicenza (northeastern Italy),



<span id="page-14-0"></span>starting from a traveler returning from Indonesia with DENV symptomatic infection. One month after the onset of symptoms, six additional household members became infected with DENV-1, as did other individuals living in the vicinity of the primary case [\[202\]](#page-23-0).

#### **Figure 10. The number of dengue virus (DENV) cases, including autochthonous cases, in Italy Figure 10. The number of dengue virus (DENV) cases, including autochthonous cases, in Italy from 2018 to 2023.** Dengue endemic countries were reported for imported cases. All data were **from 2018 to 2023.** Dengue endemic countries were reported for imported cases. All data were collected from reports of the Istituto Superiore di Sanità ("Arbovirosi in Italia. Dengue. Anno 2018", collected from reports of the Istituto Superiore di Sanità ("Arbovirosi in Italia. Dengue. Anno 2018", 2018; "Arbovirosi in Italia. Dengue. Anno 2019", 2019; "Arbovirosi in Italia. Dengue. Anno 2020", 2018; "Arbovirosi in Italia. Dengue. Anno 2019", 2019; "Arbovirosi in Italia. Dengue. Anno 2020",  $2020$ ;  $\mu$ <sub>b</sub> in Italia. Dengue. Anno 2021", 2021; "Arborita. Italia. Dengue. Anno 2022" 2022; "Arbovirosi in Italia. Dengue. Anno 2023", 2023). 2020; "Arbovirosi in Italia. Dengue. Anno 2021", 2021; "Arbovirosi in Italia. Dengue. Anno 2022", 2022; "Arbovirosi in Italia. Dengue. Anno 2023", 2023).

During the COVID-19 pandemic, there was a reduction of imported DENV cases from 185 in 2019 to 34 (−80.6% compared to the previous year) in 2020 and 11 (−75.6% compared to the previous year) in 2021 [175,177]. By contrast, an increased rate of DENV infections was described in 2022, with a total of 117 new DENV cases, as SARS-CoV-2 restrictions were almost completely removed. Imported DENV cases in Italy derived mainly from Thailand, the Maldives, and Cuba, where DENV is considered endemic [203-205]. Among EU countries, France had the highest number of autochthonous DENV cases, with a peak of DENV infections (65 confirmed cases) in 2022 [190]. It was considered the most likely country responsible for the imported DENV cases described in 2022 from Italy [179].

Data on DENV infections in Italy in 2023 reported 295 reported cases in travelers returning from dengue-endemic regions (mainly Mexico, Thailand, India, Cuba, and Indonesia); 1 of these DENV cases resulted in death [\[195](#page-22-14)[,197\]](#page-22-16). In parallel, 82 new autochthonous cases were registered in the province of Lodi, Lombardy (41 cases of DENV-1 infection)  $[204,206]$  $[204,206]$ , in the province of Latina (2 cases), in Rome (38 cases), and in Anzio, Lazio region (1 case)  $[12,198]$  $[12,198]$ .

#### **4. Conclusions**

The latitudinal range of mosquitoes and other arthropods is expected to expand, allowing these disease-carrying insects to spread from the tropics to temperate regions where they were previously absent. With rising temperatures and the effects of global warming, vector capacity has increased and will continue to increase. As a result, there will be an increase in (i) the number of daily bites; (ii) the likelihood of disease transmission from the vector to humans; (iii) the likelihood of the vector becoming infected; (iv) the long incubation period of the pathogen; (v) the time between the moment the pathogen is ingested and the moment a mosquito can transmit the disease; (vi) the mortality and reproduction rate of the vector. Peaks in WNV infections were reported in Italy in 2018 and 2022. These events were partly attributed to rising temperatures followed by rainfall, which facilitates the spread of *Cx. pipiens*. As for DENV infection, it remains the most commonly identified cause of fever in sick international travelers, as confirmed by numerous cases of DENV in Europe. However, the recent outbreak of autochthonous cases of dengue virus

(DENV) in Europe has become an urgent concern. A notable increase in autochthonous cases occurred in Italy in 2023, highlighting the urgent need to strengthen DENV surveillance in the country and the potential for tropical disease outbreaks in non-endemic regions where competent vectors may be present.

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#### **References**

- <span id="page-15-0"></span>1. Abbass, K.; Qasim, M.Z.; Song, H.; Murshed, M.; Mahmood, H.; Younis, I. A Review of the Global Climate Change Impacts, Adaptation, and Sustainable Mitigation Measures. *Environ. Sci. Pollut. Res.* **2022**, *29*, 42539–42559. [\[CrossRef\]](https://doi.org/10.1007/s11356-022-19718-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35378646)
- <span id="page-15-1"></span>2. Bellone, R.; Failloux, A.-B. The Role of Temperature in Shaping Mosquito-Borne Viruses Transmission. *Front. Microbiol.* **2020**, *11*, 584846. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2020.584846) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33101259)
- <span id="page-15-2"></span>3. Hadfield, J.; Brito, A.F.; Swetnam, D.M.; Vogels, C.B.F.; Tokarz, R.E.; Andersen, K.G.; Smith, R.C.; Bedford, T.; Grubaugh, N.D. Twenty Years of West Nile Virus Spread and Evolution in the Americas Visualized by Nextstrain. *PLOS Pathog.* **2019**, *15*, e1008042. [\[CrossRef\]](https://doi.org/10.1371/journal.ppat.1008042) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31671157)
- <span id="page-15-3"></span>4. Bakonyi, T.; Haussig, J.M. West Nile Virus Keeps on Moving up in Europe. *Eurosurveillance* **2020**, *25*, 2001938. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2020.25.46.2001938) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33213684)
- <span id="page-15-4"></span>5. Current Year Data (2024)|West Nile Virus|CDC. Available online: [https://www.cdc.gov/west-nile-virus/data-maps/](https://www.cdc.gov/west-nile-virus/data-maps/current-year-data.html?CDC_AAref_Val=https://www.cdc.gov/westnile/statsmaps/current-season-data.html) [current-year-data.html?CDC\\_AAref\\_Val=https://www.cdc.gov/westnile/statsmaps/current-season-data.html](https://www.cdc.gov/west-nile-virus/data-maps/current-year-data.html?CDC_AAref_Val=https://www.cdc.gov/westnile/statsmaps/current-season-data.html) (accessed on 16 July 2024).
- <span id="page-15-5"></span>6. Historic Data (1999–2023)|West Nile Virus|CDC. Available online: [https://www.cdc.gov/west-nile-virus/data-maps/historic](https://www.cdc.gov/west-nile-virus/data-maps/historic-data.html?CDC_AAref_Val=https://www.cdc.gov/westnile/statsmaps/historic-data.html)[data.html?CDC\\_AAref\\_Val=https://www.cdc.gov/westnile/statsmaps/historic-data.html](https://www.cdc.gov/west-nile-virus/data-maps/historic-data.html?CDC_AAref_Val=https://www.cdc.gov/westnile/statsmaps/historic-data.html) (accessed on 16 July 2024).
- <span id="page-15-6"></span>7. Epidemiological Update: West Nile Virus Transmission Season in Europe. 2023. Available online: [https://www.ecdc.europa.eu/](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2023-0#:~:text=During%20the%202023%20West%20Nile) [en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2023-0#:~:text=During%20the%202023](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2023-0#:~:text=During%20the%202023%20West%20Nile) [%20West%20Nile](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2023-0#:~:text=During%20the%202023%20West%20Nile) (accessed on 23 May 2024).
- <span id="page-15-7"></span>8. Gubler, D.J. Dengue, Urbanization and Globalization: The Unholy Trinity of the 21st Century. *Trop. Med. Health* **2011**, *39*, 3–11. [\[CrossRef\]](https://doi.org/10.2149/tmh.2011-S05)
- <span id="page-15-8"></span>9. Dengue Cases January–December 2023. Available online: [https://www.ecdc.europa.eu/en/publications-data/dengue-cases](https://www.ecdc.europa.eu/en/publications-data/dengue-cases-january-december-2023)[january-december-2023](https://www.ecdc.europa.eu/en/publications-data/dengue-cases-january-december-2023) (accessed on 16 July 2024).
- <span id="page-15-9"></span>10. European Centre for Disease Prevention and Control Dengue Worldwide Overview. Available online: [https://www.ecdc.europa.](https://www.ecdc.europa.eu/en/dengue-monthly) [eu/en/dengue-monthly](https://www.ecdc.europa.eu/en/dengue-monthly) (accessed on 16 July 2024).
- <span id="page-15-10"></span>11. Dengue-Global Situation. Available online: [https://www.who.int/emergencies/disease-outbreak-news/item/2024-DON518#:~:](https://www.who.int/emergencies/disease-outbreak-news/item/2024-DON518#:~:text=In%202024,%20Brazil%20is%20the) [text=In%202024,%20Brazil%20is%20the](https://www.who.int/emergencies/disease-outbreak-news/item/2024-DON518#:~:text=In%202024,%20Brazil%20is%20the) (accessed on 16 July 2024).
- <span id="page-15-11"></span>12. EpiCentro Febbre Dengue-News. Available online: <https://www.epicentro.iss.it/febbre-dengue/aggiornamenti> (accessed on 16 July 2024).
- <span id="page-15-12"></span>13. Weatherhead, J.E.; Miller, V.E.; Garcia, M.N.; Hasbun, R.; Salazar, L.; Dimachkie, M.M.; Murray, K.O. Long-Term Neurological Outcomes in West Nile Virus–Infected Patients: An Observational Study. *Am. J. Trop. Med. Hyg.* **2015**, *92*, 1006–1012. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.14-0616) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25802426)
- <span id="page-15-13"></span>14. Kalayanarooj, S. Clinical Manifestations and Management of Dengue/DHF/DSS. *Trop. Med. Health* **2011**, *39*, 83–87. [\[CrossRef\]](https://doi.org/10.2149/tmh.2011-S10) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22500140)
- <span id="page-15-14"></span>15. Paixão, E.S.; Teixeira, M.G.; Rodrigues, L.C. Zika, chikungunya and dengue: The causes and threats of new and re-emerging arboviral diseases. *BMJ Glob Health* **2018**, *3* (Suppl. S1), e000530, Erratum in *BMJ Glob Health* **2018**, *3*, e000530corr1. [\[CrossRef\]](https://doi.org/10.1136/bmjgh-2017-000530) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29435366) [\[PubMed Central\]](https://www.ncbi.nlm.nih.gov/pmc/PMC5759716)
- <span id="page-15-15"></span>16. Sinigaglia, A.; Peta, E.; Riccetti, S.; Barzon, L. New Avenues for Therapeutic Discovery against West Nile Virus. *Expert Opin. Drug Discov.* **2020**, *15*, 333–348. [\[CrossRef\]](https://doi.org/10.1080/17460441.2020.1714586)
- <span id="page-15-16"></span>17. Kaiser, J.A.; Barrett, A.D.T. Twenty Years of Progress toward West Nile Virus Vaccine Development. *Viruses* **2019**, *11*, 823. [\[CrossRef\]](https://doi.org/10.3390/v11090823)
- <span id="page-15-17"></span>18. da Silveira, L.T.C.; Tura, B.; Santos, M. Systematic Review of Dengue Vaccine Efficacy. *BMC Infect. Dis.* **2019**, *19*, 750. [\[CrossRef\]](https://doi.org/10.1186/s12879-019-4369-5)
- <span id="page-15-18"></span>19. Arroyo, J.; Miller, C.; Catalan, J.; Myers, G.A.; Ratterree, M.S.; Trent, D.W.; Monath, T.P. ChimeriVax-West Nile Virus Live-Attenuated Vaccine: Preclinical Evaluation of Safety, Immunogenicity, and Efficacy. *J. Virol.* **2004**, *78*, 12497–12507. [\[CrossRef\]](https://doi.org/10.1128/JVI.78.22.12497-12507.2004)
- <span id="page-16-0"></span>20. Tricou, V.; Yu, D.; Reynales, H.; Biswal, S.; Saez-Llorens, X.; Sirivichayakul, C.; Lopez, P.; Borja-Tabora, C.; Bravo, L.; Kosalaraksa, P.; et al. Long-Term Efficacy and Safety of a Tetravalent Dengue Vaccine (TAK-003): 4·5-Year Results from a Phase 3, Randomised, Double-Blind, Placebo-Controlled Trial. *Lancet Glob. Health* **2024**, *12*, e257–e270. [\[CrossRef\]](https://doi.org/10.1016/S2214-109X(23)00522-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38245116)
- <span id="page-16-1"></span>21. Thomas, S.J. Is New Dengue Vaccine Efficacy Data a Relief or Cause for Concern? *npj Vaccines* **2023**, *8*, 1265. [\[CrossRef\]](https://doi.org/10.1038/s41541-023-00658-2)
- <span id="page-16-2"></span>22. Girard, Y.A.; Klingler, K.A.; Higgs, S. West Nile virus dissemination and tissue tropisms in orally infected *Culex pipiens quinquefasciatus*. *Vector Borne Zoonotic Dis.* **2004**, *4*, 109–122. [\[CrossRef\]](https://doi.org/10.1089/1530366041210729)
- <span id="page-16-3"></span>23. Lewis, J.; Gallichotte, E.N.; Randall, J.; Glass, A.; Foy, B.D.; Ebel, G.D.; Kading, R.C. Intrinsic Factors Driving Mosquito Vector Competence and Viral Evolution: A Review. *Front. Cell. Infect. Microbiol.* **2023**, *13*, 1330600. [\[CrossRef\]](https://doi.org/10.3389/fcimb.2023.1330600)
- <span id="page-16-4"></span>24. Ciota, A.T. West Nile virus and its vectors. *Curr. Opin. Insect Sci.* **2017**, *22*, 28–36. [\[CrossRef\]](https://doi.org/10.1016/j.cois.2017.05.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28805636)
- <span id="page-16-5"></span>25. Andreadis, T.G. The contribution of *Culex pipiens* complex mosquitoes to transmission and persistence of West Nile virus in North America. *J. Am. Mosq. Control Assoc.* **2012**, *28* (Suppl. S4), 137–151. [\[CrossRef\]](https://doi.org/10.2987/8756-971X-28.4s.137)
- <span id="page-16-6"></span>26. Vogels, C.B.; Göertz, G.P.; Pijlman, G.P.; Koenraadt, C.J. Vector competence of European mosquitoes for West Nile virus. *Emerg. Microbes Infect.* **2017**, *6*, e96. [\[CrossRef\]](https://doi.org/10.1038/emi.2017.82) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29116220)
- <span id="page-16-7"></span>27. Paz, S. Effects of Climate Change on Vector-Borne Diseases: An Updated Focus on West Nile Virus in Humans. *Emerg. Top. Life Sci.* **2019**, *3*, 143–152. [\[CrossRef\]](https://doi.org/10.1042/etls20180124)
- <span id="page-16-8"></span>28. Campbell, G.L.; Marfin, A.A.; Lanciotti, R.S.; Gubler, D.J. West Nile Virus. *Lancet Infect. Dis.* **2002**, *2*, 519–529. [\[CrossRef\]](https://doi.org/10.1016/S1473-3099(02)00368-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12206968)
- <span id="page-16-9"></span>29. Bunning, M.L.; Bowen, R.A.; Cropp, B.C.; Sullivan, K.G.; Davis, B.S.; Komar, N.; Godsey, M.; Baker, D.; Hettler, D.L.; Holmes, D.A.; et al. Experimental Infection of Horses with West Nile Virus. *Emerg. Infect. Dis.* **2002**, *8*, 380–386. [\[CrossRef\]](https://doi.org/10.3201/eid0804.010239) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11971771)
- <span id="page-16-10"></span>30. Martín-Acebes, M.A. West Nile Virus: A Re-Emerging Pathogen Revisited. *World J. Virol.* **2012**, *1*, 51. [\[CrossRef\]](https://doi.org/10.5501/wjv.v1.i2.51) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24175211)
- <span id="page-16-11"></span>31. Austgen, L.E.; Bowen, R.A.; Bunning, M.L.; Davis, B.S.; Mitchell, C.J.; Chang, G.J.J. Experimental Infection of Cats and Dogs with West Nile Virus. *Emerg. Infect. Dis.* **2004**, *10*, 82–88. [\[CrossRef\]](https://doi.org/10.3201/eid1001.020616) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15078601)
- <span id="page-16-12"></span>32. Escribano-Romero, E.; Lupulović, D.; Merino-Ramos, T.; Blázquez, A.-B.; Lazić, G.; Lazić, S.; Saiz, J.-C.; Petrović, T. West Nile Virus Serosurveillance in Pigs, Wild Boars, and Roe Deer in Serbia. *Vet. Microbiol.* **2015**, *176*, 365–369. [\[CrossRef\]](https://doi.org/10.1016/j.vetmic.2015.02.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25724332)
- <span id="page-16-13"></span>33. Rimoldi, G.; Mete, A.; Adaska, J.M.; Anderson, M.L.; Symmes, K.P.; Diab, S. West Nile Virus Infection in Sheep. *Vet. Pathol.* **2016**, *54*, 155–158. [\[CrossRef\]](https://doi.org/10.1177/0300985816653796) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27312367)
- <span id="page-16-14"></span>34. Bowen, R.A.; Nemeth, N.M. Experimental Infections with West Nile Virus. *Curr. Opin. Infect. Dis.* **2007**, *20*, 293–297. [\[CrossRef\]](https://doi.org/10.1097/QCO.0b013e32816b5cad) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17471040)
- <span id="page-16-15"></span>35. Klenk, K.; Snow, J.; Morgan, K.; Bowen, R.; Stephens, M.; Foster, F.; Gordy, P.; Beckett, S.; Komar, N.; Gubler, D.; et al. Alligators as West Nile Virus Amplifiers. *Emerg. Infect. Dis.* **2004**, *10*, 2150–2155. [\[CrossRef\]](https://doi.org/10.3201/eid1012.040264) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15663852)
- <span id="page-16-16"></span>36. Pealer, L.N.; Marfin, A.A.; Petersen, L.R.; Lanciotti, R.S.; Page, P.L.; Stramer, S.L.; Stobierski, M.G.; Signs, K.; Newman, B.; Kapoor, H.; et al. Transmission of West Nile Virus through Blood Transfusion in the United States in 2002. *N. Engl. J. Med.* **2003**, *349*, 1236–1245. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa030969)
- <span id="page-16-17"></span>37. Iwamoto, M.; Jernigan, D.B.; Guasch, A.; Trepka, M.J.; Blackmore, C.G.; Hellinger, W.C.; Pham, S.M.; Zaki, S.; Lanciotti, R.S.; Lance-Parker, S.E.; et al. Transmission of West Nile Virus from an Organ Donor to Four Transplant Recipients. *N. Engl. J. Med.* **2003**, *348*, 2196–2203. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa022987)
- <span id="page-16-18"></span>38. Centers for Disease Control and Prevention. Possible West Nile Virus Transmission to an Infant through Breast-Feeding— Michigan, 2002. *JAMA J. Am. Med. Assoc.* **2002**, *288*, 1976–1977. [\[CrossRef\]](https://doi.org/10.1001/jama.288.16.1976)
- <span id="page-16-19"></span>39. Anderson, J.F.; Main, A.J.; Delroux, K.; Fikrig, E. Extrinsic Incubation Periods for Horizontal and Vertical Transmission of West Nile Virus by *Culex Pipiens Pipiens* (Diptera: Culicidae). *J. Med. Entomol.* **2008**, *45*, 445–451. [\[CrossRef\]](https://doi.org/10.1093/jmedent/45.3.445) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18533438)
- 40. Baqar, S.; Murphy, J.R.; Watts, D.M.; Hayes, C.G. Vertical Transmission of West Nile Virus by *Culex* and *Aedes* Species Mosquitoes. *Am. J. Trop. Med. Hyg.* **1993**, *48*, 757–762. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.1993.48.757)
- 41. Dohm, D.J.; Sardelis, M.R.; Turell, M.J. Experimental Vertical Transmission of West Nile Virus by *Culex Pipiens* (Diptera: Culicidae): Table 1. *J. Med. Entomol.* **2002**, *39*, 640–644. [\[CrossRef\]](https://doi.org/10.1603/0022-2585-39.4.640)
- <span id="page-16-20"></span>42. Nelms, B.M.; Fechter-Leggett, E.; Carroll, B.D.; Macedo, P.; Kluh, S.; Reisen, W.K. Experimental and Natural Vertical Transmission of West Nile Virus by California *Culex* (Diptera: Culicidae) Mosquitoes. *J. Med. Entomol.* **2013**, *50*, 371–378. [\[CrossRef\]](https://doi.org/10.1603/ME12264) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23540126)
- <span id="page-16-21"></span>43. Novelo, M.; Dutra, H.L.C.; Metz, H.C.; Jones, M.; Sigle, L.T.; Frentiu, F.D.; Allen, S.L.; Chenoweth, S.F.; McGraw, E.A. Dengue and Chikungunya Virus Loads in the Mosquito *Aedes* Are Determined by Distinct Genetic Architectures. *PLOS Pathog.* **2023**, *19*, e1011307. [\[CrossRef\]](https://doi.org/10.1371/journal.ppat.1011307) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37043515)
- <span id="page-16-22"></span>44. Rezza, G. *Aedes Albopictus* and the Reemergence of Dengue. *BMC Public Health* **2012**, *12*, 72. [\[CrossRef\]](https://doi.org/10.1186/1471-2458-12-72) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22272602)
- <span id="page-16-23"></span>45. Weaver, S.C.; Barrett, A.D.T. Transmission Cycles, Host Range, Evolution and Emergence of Arboviral Disease. *Nat. Rev. Microbiol.* **2004**, *2*, 789–801. [\[CrossRef\]](https://doi.org/10.1038/nrmicro1006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15378043)
- <span id="page-16-24"></span>46. Briscoe, M.S. *Aedes Aegypti* the Yellow Fever Mosquito, Its Life History, Bionomics and Structure. *J. Natl. Med. Assoc.* **1962**, *54*, 132.
- <span id="page-16-25"></span>47. Schaffner, F.; Mathis, A. Dengue and Dengue Vectors in the WHO European Region: Past, Present, and Scenarios for the Future. *Lancet Infect. Dis.* **2014**, *14*, 1271–1280. [\[CrossRef\]](https://doi.org/10.1016/S1473-3099(14)70834-5)
- <span id="page-16-26"></span>48. Blanchard, R. Le danger du paludisme et de la fièvre jaune en France; moyens de l'éviter. *Bull. Acad. Med.* **1917**, *77*, 657–669.
- <span id="page-16-27"></span>49. Blanc, G.; Caminopetros, J. Comment les fait épidémiologiques, en Grèce, montrent le rôle exclusif joué par le *Stegomyia fasciata* (*Aedes aegypti*) dans la transmission de la dengue. *Arch. Inst. Pasteur Hell.* **1930**, *2*, 277–294.
- <span id="page-16-28"></span>50. La Face, L.; Raffaele, G. Sulla presenza della *Stegomyia fasciata* nell'Italia meridionale e in Sicilia. *Il Policlin.* **1928**, *35*, 2095.
- <span id="page-16-29"></span>51. Marzinowsky, E.I. De l'existence de *Stegomyia fasciata* (*St. calopus*) en Russie. *Bull. Soc. Pathol. Exot.* **1914**, *7*, 590–593.
- <span id="page-17-0"></span>52. Sarmento, M.; França, C. Sur quelques culicides portugais. *C R Hebdo Séances Mém. Soc. Biol.* **1902**, *54*, 152–153.
- <span id="page-17-1"></span>53. Collado, G. Datos Actuales Sobre La Distribución Geográfica de Los Culicidos Españoles. *Eos Rev. Española Entomol.* **1930**, *6*, 329–347.
- <span id="page-17-2"></span>54. IuV, I.; Riabova, T.E.; NIa, M.; Bezzhonova, O.V.; Ganushkina, L.A.; Semenov, V.B.; Tarkhov, G.A.; Vasilenko, L.E.; Guzeeva, T.M.; Shevereva, T.V.; et al. First evidence for breeding *Aedes aegypti* L in the area of Greater Sochi and in some towns of Abkhasia. *Med. Parazitol.* **2008**, *3*, 40–43. (In Russian)
- 55. Wint, W.; Jones, P.; Kraemer, M.; Alexander, N.; Schaffner, F. Past, Present and Future Distribution of the Yellow Fever Mosquito *Aedes Aegypti*: The European Paradox. *Sci. Total Environ.* **2022**, *847*, 157566. [\[CrossRef\]](https://doi.org/10.1016/j.scitotenv.2022.157566)
- <span id="page-17-3"></span>56. Trájer, A.J. *Aedes Aegypti* in the Mediterranean Container Ports at the Time of Climate Change: A Time Bomb on the Mosquito Vector Map of Europe. *Heliyon* **2021**, *7*, e07981. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2021.e07981)
- <span id="page-17-4"></span>57. Increasing Risk of Mosquito-Borne Diseases in EU/EEA Following Spread of Aedes Species. Available online: [https://www.ecdc.](https://www.ecdc.europa.eu/en/news-events/increasing-risk-mosquito-borne-diseases-eueea-following-spread-aedes-species) [europa.eu/en/news-events/increasing-risk-mosquito-borne-diseases-eueea-following-spread-aedes-species](https://www.ecdc.europa.eu/en/news-events/increasing-risk-mosquito-borne-diseases-eueea-following-spread-aedes-species) (accessed on 16 July 2024).
- <span id="page-17-5"></span>58. Lühken, R.; Brattig, N.W.; Becker, N. Introduction of Invasive Mosquito Species into Europe and Prospects for Arbovirus Transmission and Vector Control in an Era of Globalization. *Infect. Dis. Poverty* **2023**, *12*, 109. [\[CrossRef\]](https://doi.org/10.1186/s40249-023-01167-z)
- <span id="page-17-6"></span>59. Valentine, M.J.; Murdock, C.C.; Kelly, P.J. Sylvatic Cycles of Arboviruses in Non-Human Primates. *Parasites Vectors* **2019**, *12*, 463. [\[CrossRef\]](https://doi.org/10.1186/s13071-019-3732-0)
- <span id="page-17-7"></span>60. Vasilakis, N.; Cardosa, J.; Hanley, K.A.; Holmes, E.C.; Weaver, S.C. Fever from the Forest: Prospects for the Continued Emergence of Sylvatic Dengue Virus and Its Impact on Public Health. Nature Reviews. *Microbiology* **2011**, *9*, 532–541. [\[CrossRef\]](https://doi.org/10.1038/nrmicro2595) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21666708)
- <span id="page-17-8"></span>61. Althouse, B.M.; Durbin, A.P.; Hanley, K.A.; Halstead, S.B.; Weaver, S.C.; Cummings, D.A.T. Viral Kinetics of Primary Dengue Virus Infection in Non-Human Primates: A Systematic Review and Individual Pooled Analysis. *Virology* **2014**, *452–453*, 237–246. [\[CrossRef\]](https://doi.org/10.1016/j.virol.2014.01.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24606701)
- <span id="page-17-9"></span>62. Gwee, S.X.W.; St John, A.L.; Gray, G.C.; Pang, J. Animals as Potential Reservoirs for Dengue Transmission: A Systematic Review. *One Health* **2021**, *12*, 100216. [\[CrossRef\]](https://doi.org/10.1016/j.onehlt.2021.100216) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33598525)
- <span id="page-17-10"></span>63. Kaul, H.K.; Venkateshan, C.N.; Mishra, A.C.; Modi, G.B.; Ghosh, S.N. Serological Evidence of Arbovirus Activity in Birds and Small Mammals in Japanese Encephalitis Affected Areas of Bankura District, West Bengal. *Indian J. Med. Res.* **1976**, *64*, 1535–1539.
- <span id="page-17-11"></span>64. Cigarroa-Toledo, N.; Talavera-Aguilar, L.G.; Baak-Baak, C.M.; García-Rejón, J.E.; Hernandez-Betancourt, S.; Blitvich, B.J.; Machain-Williams, C. Serologic Evidence of Flavivirus Infections in Peridomestic Rodents in Merida, Mexico. *J. Wildl. Dis.* **2016**, *52*, 168–172. [\[CrossRef\]](https://doi.org/10.7589/2015-05-116) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26540177)
- <span id="page-17-12"></span>65. Thongyuan, S.; Kittayapong, P. First Evidence of Dengue Infection in Domestic Dogs Living in Different Ecological Settings in Thailand. *PLoS ONE* **2017**, *12*, e0180013. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0180013)
- <span id="page-17-13"></span>66. Rudnick, A. Studies of the Ecology of Dengue in Malaysia: A Preliminary Report. *J. Med. Entomol.* **1965**, *2*, 203–208. [\[CrossRef\]](https://doi.org/10.1093/jmedent/2.2.203) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/5827577)
- <span id="page-17-14"></span>67. Albanese, M.; Di Cuonzo, G.; Randazzo, G.; Srihongse, S.; Tringali, G. Survey for Arbovirus Antibodies in Domestic Animals of Western Sicily. *Ann. Sclavo* **1971**, *13*, 641–647.
- <span id="page-17-15"></span>68. Beck, C.; Leparc-Goffart, I.; Desoutter, D.; Debergé, E.; Bichet, H.; Lowenski, S.; Dumarest, M.; Gonzalez, G.; Migné, C.; Vanhomwegen, J.; et al. Serological Evidence of Infection with Dengue and Zika Viruses in Horses on French Pacific Islands. *PLoS Neglected Trop. Dis.* **2019**, *13*, e0007162. [\[CrossRef\]](https://doi.org/10.1371/journal.pntd.0007162)
- <span id="page-17-16"></span>69. de Thoisy, B.; Lacoste, V.; Germain, A.; Muñoz-Jordán, J.; Colón, C.; Mauffrey, J.-F.; Delaval, M.; Catzeflis, F.; Kazanji, M.; Matheus, S.; et al. Dengue Infection in Neotropical Forest Mammals. *Vector Borne Zoonotic Dis.* **2009**, *9*, 157–170. [\[CrossRef\]](https://doi.org/10.1089/vbz.2007.0280) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18945183)
- <span id="page-17-17"></span>70. Lequime, S.; Paul, R.E.; Lambrechts, L. Determinants of Arbovirus Vertical Transmission in Mosquitoes. *PLoS Pathog.* **2016**, *12*, e1005548. [\[CrossRef\]](https://doi.org/10.1371/journal.ppat.1005548) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27171170)
- <span id="page-17-18"></span>71. Rosen, L. Further Observations on the Mechanism of Vertical Transmission of Flaviviruses by *Aedes* Mosquitoes. *Am. J. Trop. Med. Hyg.* **1988**, *39*, 123–126. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.1988.39.123) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2840833)
- <span id="page-17-19"></span>72. Joshi, V.; Mourya, D.T.; Sharma, R.C. Persistence of Dengue-3 Virus through Transovarial Transmission Passage in Successive Generations of *Aedes Aegypti* Mosquitoes. *Am. J. Trop. Med. Hyg.* **2002**, *67*, 158–161. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.2002.67.158)
- <span id="page-17-20"></span>73. Harbach, R.E. *Culex Pipiens*: Species versus Species Complex–Taxonomic History and Perspective. *J. Am. Mosq. Control Assoc.* **2012**, *28*, 10–23. [\[CrossRef\]](https://doi.org/10.2987/8756-971X-28.4.10) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23401941)
- <span id="page-17-21"></span>74. Samy, A.M.; Elaagip, A.H.; Kenawy, M.A.; Ayres, C.F.J.; Peterson, A.T.; Soliman, D.E. Climate Change Influences on the Global Potential Distribution of the Mosquito *Culex Quinquefasciatus*, Vector of West Nile Virus and Lymphatic Filariasis. *PLoS ONE* **2016**, *11*, e0163863. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0163863) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27695107)
- <span id="page-17-22"></span>75. Am, G.; el Said, S.; Soliman, B.A.; An, H.; Shoukry, A. Distribution and Bionomics of Egyptian *Culex Univittatus* (Theobald). *J. Egypt. Soc. Parasitol.* **1987**, *17*, 17–31.
- <span id="page-17-23"></span>76. Harbach, R.E. The Mosquitoes of the Subgenus *Culex* in Southwestern Asia and Egypt (Diptera: Culicidae). *Contrib. Am. Entomol. Inst.* **1988**, *24*, 1.
- <span id="page-17-24"></span>77. Wilkerson, R.C.; Linton, Y.M.; Strickman, D. *Mosquitoes of the World*; JHU Press: Baltimore, MD, USA, 2021.
- <span id="page-17-25"></span>78. Kline, D.L.; Darsie, R.F.; Ward, R.A. Identification and Geographical Distribution of the Mosquitoes of North America, North of Mexico. *J. Med. Entomol.* **2006**, *43*, 124–125. [\[CrossRef\]](https://doi.org/10.1603/0022-2585(2006)043[0124:IAGDOT]2.0.CO;2)
- <span id="page-18-0"></span>79. European Centre for Disease Prevention and Control Aedes Aegypti—Factsheet for Experts. Available online: [https://www.ecdc.](https://www.ecdc.europa.eu/en/disease-vectors/facts/mosquito-factsheets/aedes-aegypti) [europa.eu/en/disease-vectors/facts/mosquito-factsheets/aedes-aegypti](https://www.ecdc.europa.eu/en/disease-vectors/facts/mosquito-factsheets/aedes-aegypti) (accessed on 16 July 2024).
- <span id="page-18-1"></span>80. Aedes Albopictus—Factsheet for Experts. Available online: [https://www.ecdc.europa.eu/en/disease-vectors/facts/mosquito](https://www.ecdc.europa.eu/en/disease-vectors/facts/mosquito-factsheets/aedes-albopictus#:~:text=Aedes%20albopictus%20is%20considered%20to)[factsheets/aedes-albopictus#:~:text=Aedes%20albopictus%20is%20considered%20to](https://www.ecdc.europa.eu/en/disease-vectors/facts/mosquito-factsheets/aedes-albopictus#:~:text=Aedes%20albopictus%20is%20considered%20to) (accessed on 16 July 2024).
- <span id="page-18-2"></span>81. Benedict, M.Q.; Levine, R.S.; Hawley, W.A.; Lounibos, L.P. Spread of the Tiger: Global Risk of Invasion by the Mosquito *Aedes Albopictus*. *Vector-Borne Zoonotic Dis.* **2007**, *7*, 76–85. [\[CrossRef\]](https://doi.org/10.1089/vbz.2006.0562)
- <span id="page-18-3"></span>82. Postler, T.S.; Beer, M.; Blitvich, B.J.; Bukh, J.; De Lamballerie, X.; Drexler, J.F.; Imrie, A.; Kapoor, A.; Karganova, G.G.; Lemey, P.; et al. Renaming of the Genus *Flavivirus* to *Orthoflavivirus* and Extension of Binomial Species Names within the Family *Flaviviridae*. *Arch. Virol.* **2023**, *168*, 224. [\[CrossRef\]](https://doi.org/10.1007/s00705-023-05835-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37561168)
- <span id="page-18-4"></span>83. Pierson, T.C.; Diamond, M.S. The Continued Threat of Emerging Flaviviruses. *Nat. Microbiol.* **2020**, *5*, 796–812. [\[CrossRef\]](https://doi.org/10.1038/s41564-020-0714-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32367055)
- <span id="page-18-5"></span>84. Grottola, A.; Marcacci, M.; Tagliazucchi, S.; Gennari, W.; Di Gennaro, A.; Orsini, M.; Monaco, F.; Marchegiano, P.; Marini, V.; Meacci, M.; et al. Usutu Virus Infections in Humans: A Retrospective Analysis in the Municipality of Modena, Italy. *Clin. Microbiol. Infect.* **2017**, *23*, 33–37. [\[CrossRef\]](https://doi.org/10.1016/j.cmi.2016.09.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27677699)
- <span id="page-18-6"></span>85. Amicizia, D.; Domnich, A.; Panatto, D.; Lai, P.L.; Cristina, M.L.; Avio, U.; Gasparini, R. Epidemiology of Tick-Borne Encephalitis (TBE) in Europe and Its Prevention by Available Vaccines. *Hum. Vaccines Immunother.* **2013**, *9*, 1163–1171. [\[CrossRef\]](https://doi.org/10.4161/hv.23802) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23377671)
- <span id="page-18-7"></span>86. Auguste, A.J.; Langsjoen, R.M.; Porier, D.L.; Erasmus, J.H.; Bergren, N.A.; Bolling, B.G.; Luo, H.; Singh, A.; Guzman, H.; Popov, V.L.; et al. Isolation of a Novel Insect-Specific Flavivirus with Immunomodulatory Effects in Vertebrate Systems. *Virology* **2021**, *562*, 50–62. [\[CrossRef\]](https://doi.org/10.1016/j.virol.2021.07.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34256244)
- <span id="page-18-8"></span>87. Zhang, X.; Guo, X.; Fan, H.; Zhao, Q.; Zuo, S.; Sun, Q.; Pei, G.; Cheng, S.; An, X.; Wang, Y.; et al. Complete Genome Sequence of Menghai Flavivirus, a Novel Insect-Specific *flavivirus* from China. *Arch. Virol.* **2017**, *162*, 1435–1439. [\[CrossRef\]](https://doi.org/10.1007/s00705-017-3232-5)
- <span id="page-18-9"></span>88. Huhtamo, E.; Moureau, G.; Cook, S.; Julkunen, O.; Putkuri, N.; Kurkela, S.; Uzcátegui, N.Y.; Harbach, R.E.; Gould, E.A.; Vapalahti, O.; et al. Novel Insect-Specific Flavivirus Isolated from Northern Europe. *Virology* **2012**, *433*, 471–478. [\[CrossRef\]](https://doi.org/10.1016/j.virol.2012.08.038)
- <span id="page-18-10"></span>89. Markoff, L. 5'- and 3'-Noncoding Regions in Flavivirus RNA. *Adv. Virus Res.* **2003**, *59*, 177–228. [\[CrossRef\]](https://doi.org/10.1016/s0065-3527(03)59006-6)
- <span id="page-18-11"></span>90. Shiryaev, S.A.; Strongin, A.Y. Structural and Functional Parameters of the Flaviviral Protease: A Promising Antiviral Drug Target. *Future Virol.* **2010**, *5*, 593–606. [\[CrossRef\]](https://doi.org/10.2217/fvl.10.39)
- <span id="page-18-12"></span>91. Lee, E.; Bujalowski, P.J.; Teramoto, T.; Gottipati, K.; Scott, S.D.; Padmanabhan, R.; Choi, K.H. Structures of Flavivirus RNA Promoters Suggest Two Binding Modes with NS5 Polymerase. *Nat. Commun.* **2021**, *12*, 2530. [\[CrossRef\]](https://doi.org/10.1038/s41467-021-22846-1)
- <span id="page-18-13"></span>92. Renner, M.; Dejnirattisai, W.; Carrique, L.; Martin, I.S.; Karia, D.; Ilca, S.L.; Ho, S.F.; Kotecha, A.; Keown, J.R.; Mongkolsapaya, J.; et al. Flavivirus Maturation Leads to the Formation of an Occupied Lipid Pocket in the Surface Glycoproteins. *Nat. Commun.* **2021**, *12*, 1238. [\[CrossRef\]](https://doi.org/10.1038/s41467-021-21505-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33623019)
- <span id="page-18-14"></span>93. Mackenzie, J.S.; Williams, D.T. The Zoonotic Flaviviruses of Southern, South-Eastern and Eastern Asia, and Australasia: The Potential for Emergent Viruses. *Zoonoses Public Health* **2009**, *56*, 338–356. [\[CrossRef\]](https://doi.org/10.1111/j.1863-2378.2008.01208.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19486319)
- <span id="page-18-15"></span>94. Bakonyi, T.; Ivanics, É.; Erdélyi, K.; Ursu, K.; Ferenczi, E.; Weissenböck, H.; Nowotny, N. Lineage 1 and 2 Strains of Encephalitic West Nile Virus. *Emerg. Infect. Dis.* **2006**, *12*, 618. [\[CrossRef\]](https://doi.org/10.3201/eid1204.051379) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16704810)
- <span id="page-18-16"></span>95. Mencattelli, G.; Ndione, M.H.D.; Rosà, R.; Marini, G.; Diagne, C.T.; Diagne, M.M.; Fall, G.; Faye, O.; Diallo, M.; Faye, O.; et al. Epidemiology of West Nile Virus in Africa: An Underestimated Threat. *PLoS Neglected Trop. Dis.* **2022**, *16*, e0010075. [\[CrossRef\]](https://doi.org/10.1371/journal.pntd.0010075) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35007285)
- <span id="page-18-17"></span>96. Platonov, A.E.; Karan', L.S.; Shopenskaia, T.A.; Fedorova, M.V.; Koliasnikova, N.M.; Rusakova, N.M.; Shishkina, L.V.; Arshba, T.E.; Zhuravlev, V.I.; Govorukhina, M.V.; et al. Genotyping of West Nile Fever Virus Strains Circulating in Southern Russia as an Epidemiological Investigation Method: Principles and Results. *Zhurnal Mikrobiol. Epidemiol. I Immunobiol.* **2011**, *2*, 29–37.
- <span id="page-18-18"></span>97. Kolodziejek, J.; Seidel, B.; Jungbauer, C.; Dimmel, K.; Kolodziejek, M.; Rudolf, I.; Hubálek, Z.; Allerberger, F.; Nowotny, N. West Nile Virus Positive Blood Donation and Subsequent Entomological Investigation, Austria, 2014. *PLoS ONE* **2015**, *10*, e0126381. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0126381)
- 98. Wodak, E.; Richter, S.; Zoltán, B.; Revilla-Fernández, S.; Weissenböck, H.; Nowotny, N.; Winter, P. Detection and Molecular Analysis of West Nile Virus Infections in Birds of Prey in the Eastern Part of Austria in 2008 and 2009. *Vet. Microbiol.* **2011**, *149*, 358–366. [\[CrossRef\]](https://doi.org/10.1016/j.vetmic.2010.12.012) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21276665)
- <span id="page-18-19"></span>99. Hernández-Triana, L.M.; Jeffries, C.L.; Mansfield, K.L.; Carnell, G.; Fooks, A.R.; Johnson, N. Emergence of West Nile Virus Lineage 2 in Europe: A Review on the Introduction and Spread of a Mosquito-Borne Disease. *Front. Public Health* **2014**, *2*, 271. [\[CrossRef\]](https://doi.org/10.3389/fpubh.2014.00271) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25538937)
- <span id="page-18-20"></span>100. Fall, G.; Diallo, D.; Soumaila, H.; Ndiaye, E.H.; Lagare, A.; Sadio, B.D.; Ndione, M.H.D.; Wiley, M.; Dia, M.; Diop, M.; et al. First Detection of the West Nile Virus Koutango Lineage in Sandflies in Niger. *Pathogens* **2021**, *10*, 257. [\[CrossRef\]](https://doi.org/10.3390/pathogens10030257)
- <span id="page-18-21"></span>101. Aliota, M.T.; Jones, S.; Dupuis, A.P.; Ciota, A.T.; Hubálek, Z.; Kramer, L.D. Characterization of Rabensburg Virus, a Flavivirus Closely Related to West Nile Virus of the Japanese Encephalitis Antigenic Group. *PLoS ONE* **2012**, *7*, e39387. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0039387)
- <span id="page-18-22"></span>102. Prilipov, A.G.; Kinney, R.M.; Samokhvalov, E.I.; Savage, H.M.; Al'khovskiĭ, S.V.; Tsuchiya, K.R.; Gromashevskiĭ, V.L.; Sadykova, G.K.; Shatalov, A.G.; Vyshemirskiĭ, O.I.; et al. Analysis of New Variants of West Nile Fever Virus. *Vopr. Virusol.* 2002, 47, 36-41. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12271724)
- <span id="page-18-23"></span>103. Bondre, V.P.; Jadi, R.S.; Mishra, A.C.; Yergolkar, P.N.; Arankalle, V.A. West Nile Virus Isolates from India: Evidence for a Distinct Genetic Lineage. *J. Gen. Virol.* **2007**, *88*, 875–884. [\[CrossRef\]](https://doi.org/10.1099/vir.0.82403-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17325360)
- <span id="page-19-0"></span>104. Vázquez, A.; Sánchez-Seco, M.P.; Ruiz, S.; Molero, F.; Hernández, L.; Moreno, J.; Magallanes, A.; Tejedor, C.G.; Tenorio, A. Putative New Lineage of West Nile Virus, Spain. *Emerg. Infect. Dis.* **2010**, *16*, 549–552. [\[CrossRef\]](https://doi.org/10.3201/eid1603.091033) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20202444)
- <span id="page-19-1"></span>105. Rajaiah, P.; Mayilsamy, M.; Kumar, A. West Nile Virus in India: An Update on Its Genetic Lineages. *J. Vector Borne Dis.* **2023**, *60*, 225–237. [\[CrossRef\]](https://doi.org/10.4103/0972-9062.374039)
- <span id="page-19-2"></span>106. Monath, T.P. *The Arboviruses*; National Institutes of Health: Boca Raton, FL, USA, 2021.
- <span id="page-19-3"></span>107. Mustafa, M.S.; Rasotgi, V.; Jain, S.; Gupta, V. Discovery of Fifth Serotype of Dengue Virus (DENV-5): A New Public Health Dilemma in Dengue Control. *Med. J. Armed Forces India* **2015**, *71*, 67–70. [\[CrossRef\]](https://doi.org/10.1016/j.mjafi.2014.09.011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25609867)
- <span id="page-19-4"></span>108. Rico-Hesse, R. Molecular Evolution and Distribution of Dengue Viruses Type 1 and 2 in Nature. *Virology* **1990**, *174*, 479–493. [\[CrossRef\]](https://doi.org/10.1016/0042-6822(90)90102-W) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2129562)
- <span id="page-19-5"></span>109. Vasilakis, N.; Holmes, E.C.; Fokam, E.B.; Faye, O.; Diallo, M.; Sall, A.A.; Weaver, S.C. Evolutionary Processes among Sylvatic Dengue Type 2 Viruses. *J. Virol.* **2007**, *81*, 9591–9595. [\[CrossRef\]](https://doi.org/10.1128/JVI.02776-06) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17553878)
- <span id="page-19-6"></span>110. Wittke, V.; Robb, T.E.; Thu, H.M.; Nisalak, A.; Nimmannitya, S.; Kalayanrooj, S.; Vaughn, D.W.; Endy, T.P.; Holmes, E.C.; Aaskov, J.G. Extinction and Rapid Emergence of Strains of Dengue 3 Virus during an Interepidemic Period. *Virology* **2002**, *301*, 148–156. [\[CrossRef\]](https://doi.org/10.1006/viro.2002.1549) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12359455)
- <span id="page-19-7"></span>111. Chen, R.; Han, G.Z. Dengue in China: Comprehensive Phylogenetic Evaluation Reveals Evidence of Endemicity and Complex Genetic Diversity. *Am. J. Trop. Med. Hyg.* **2016**, *94*, 198–202. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.15-0546)
- <span id="page-19-8"></span>112. Davis, C.W.; Nguyen, H.Y.; Hanna, S.L.; Sanchez, M.D.; Doms, R.W.; Pierson, T.C. West Nile Virus Discriminates between DC-SIGN and DC-SIGNR for Cellular Attachment and Infection. *J. Virol.* **2006**, *80*, 1290–1301. [\[CrossRef\]](https://doi.org/10.1128/JVI.80.3.1290-1301.2006)
- <span id="page-19-9"></span>113. Martina, B.E.E.; Koraka, P.; van den Doel, P.; Rimmelzwaan, G.F.; Haagmans, B.L.; Osterhaus, A.D.M.E. DC-SIGN Enhances Infection of Cells with Glycosylated West Nile Virus in Vitro and Virus Replication in Human Dendritic Cells Induces Production of IFN-α and TNF-α. *Virus Res.* **2008**, *135*, 64–71. [\[CrossRef\]](https://doi.org/10.1016/j.virusres.2008.02.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18405996)
- <span id="page-19-10"></span>114. Samuel, M.A.; Diamond, M.S. Alpha/Beta Interferon Protects against Lethal West Nile Virus Infection by Restricting Cellular Tropism and Enhancing Neuronal Survival. *J. Virol.* **2005**, *79*, 13350–13361. [\[CrossRef\]](https://doi.org/10.1128/JVI.79.21.13350-13361.2005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16227257)
- <span id="page-19-11"></span>115. Wang, T.; Town, T.; Alexopoulou, L.; Anderson, J.F.; Fikrig, E.; Flavell, R.A. Toll-like Receptor 3 Mediates West Nile Virus Entry into the Brain Causing Lethal Encephalitis. *Nat. Med.* **2004**, *10*, 1366–1373. [\[CrossRef\]](https://doi.org/10.1038/nm1140) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15558055)
- <span id="page-19-12"></span>116. Quick, E.D.; Leser, J.S.; Clarke, P.; Tyler, K.L. Activation of Intrinsic Immune Responses and Microglial Phagocytosis in an Ex Vivo Spinal Cord Slice Culture Model of West Nile Virus Infection. *J. Virol.* **2014**, *88*, 13005–13014. [\[CrossRef\]](https://doi.org/10.1128/JVI.01994-14) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25165111)
- 117. Shrestha, B.; Gottlieb, D.; Diamond, M.P. Infection and Injury of Neurons by West Nile Encephalitis Virus. *J. Virol.* **2003**, *77*, 13203–13213. [\[CrossRef\]](https://doi.org/10.1128/JVI.77.24.13203-13213.2003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14645577)
- 118. Cheeran, M.C.J.; Hu, S.; Sheng, W.S.; Rashid, A.; Peterson, P.K.; Lokensgard, J.R. Differential Responses of Human Brain Cells to West Nile Virus Infection. *J. Neurovirol.* **2005**, *11*, 512–524. [\[CrossRef\]](https://doi.org/10.1080/13550280500384982) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16338745)
- 119. Hunsperger, E.; Roehrig, J. Characterization of West Nile Viral Replication and Maturation in Peripheral Neurons in Culture. *J. Neurovirol.* **2005**, *11*, 11–22. [\[CrossRef\]](https://doi.org/10.1080/13550280590900454) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15804955)
- <span id="page-19-13"></span>120. Wang, P.; Arjona, A.; Zhang, Y.; Sultana, H.; Dai, J.; Yang, L.; LeBlanc, P.M.; Doiron, K.; Saleh, M.; Fikrig, E. Caspase-12 Controls West Nile Virus Infection via the Viral RNA Receptor RIG-I. *Nat. Immunol.* **2010**, *11*, 912–919. [\[CrossRef\]](https://doi.org/10.1038/ni.1933) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20818395)
- <span id="page-19-14"></span>121. Chu, J.J.H. The Mechanism of Cell Death during West Nile Virus Infection Is Dependent on Initial Infectious Dose. *J. Gen. Virol.* **2003**, *84*, 3305–3314. [\[CrossRef\]](https://doi.org/10.1099/vir.0.19447-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14645911)
- <span id="page-19-15"></span>122. Diniz, J.A.P.; Da Rosa, A.P.A.T.; Guzman, H.; Xu, F.; Xiao, S.Y.; Popov, V.L.; Vasconcelos, P.F.C.; Tesh, R.B. West Nile Virus Infection of Primary Mouse Neuronal and Neuroglial Cells: The Role of Astrocytes in Chronic Infection. *Am. J. Trop. Med. Hyg.* **2006**, *75*, 691–696. [\[CrossRef\]](https://doi.org/10.4269/ajtmh.2006.75.691)
- <span id="page-19-16"></span>123. Peng, B.H.; Wang, T. West Nile Virus Induced Cell Death in the Central Nervous System. *Pathogens* **2019**, *8*, 215. [\[CrossRef\]](https://doi.org/10.3390/pathogens8040215) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31683807)
- <span id="page-19-17"></span>124. Pupella, S.; Pisani, G.; Cristiano, K.; Catalano, L.; Grazzini, G. West Nile Virus in the Transfusion Setting with a Special Focus on Italian Preventive Measures Adopted in 2008–2012 and Their Impact on Blood Safety. *Blood Transfus.* **2013**, *11*, 463–564. [\[CrossRef\]](https://doi.org/10.2450/2013.0077-13)
- <span id="page-19-18"></span>125. Brenner, W.; Storch, G.; Buller, R.; Vij, R.; Devine, S.; DiPersio, J. West Nile Virus Encephalopathy in an Allogeneic Stem Cell Transplant Recipient: Use of Quantitative PCR for Diagnosis and Assessment of Viral Clearance. *Bone Marrow Transplant.* **2005**, *36*, 369–370. [\[CrossRef\]](https://doi.org/10.1038/sj.bmt.1705059) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15968282)
- <span id="page-19-19"></span>126. DeBiasi, R.L.; Tyler, K.L. West Nile Virus Meningoencephalitis. *Nat. Clin. Pract. Neurol.* **2006**, *2*, 264–275. [\[CrossRef\]](https://doi.org/10.1038/ncpneuro0176)
- <span id="page-19-20"></span>127. Bai, F.; Thompson, E.A.; Vig, P.J.S.; Leis, A.A. Current Understanding of West Nile Virus Clinical Manifestations, Immune Responses, Neuroinvasion, and Immunotherapeutic Implications. *Pathogens* **2019**, *8*, 193. [\[CrossRef\]](https://doi.org/10.3390/pathogens8040193) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31623175)
- <span id="page-19-21"></span>128. Anwar, M.N.; Akhtar, R.; Abid, M.; Khan, S.A.; Rehman, Z.U.; Tayyub, M.; Malik, M.I.; Shahzad, M.K.; Mubeen, H.; Qadir, M.S.; et al. The Interactions of Flaviviruses with Cellular Receptors: Implications for Virus Entry. *Virology* **2022**, *568*, 77–85. [\[CrossRef\]](https://doi.org/10.1016/j.virol.2022.02.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35149346)
- <span id="page-19-22"></span>129. Taguwa, S.; Maringer, K.; Li, X.; Bernal-Rubio, D.; Rauch, J.N.; Gestwicki, J.E.; Andino, R.; Fernandez-Sesma, A.; Frydman, J. Defining Hsp70 Subnetworks in Dengue Virus Replication Reveals Key Vulnerability in Flavivirus Infection. *Cell* **2015**, *163*, 1108–1123. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2015.10.046) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26582131)
- <span id="page-19-23"></span>130. Cruz-Oliveira, C.; Freire, J.M.; Conceição, T.M.; Higa, L.M.; Castanho, M.A.R.B.; Da Poian, A.T. Receptors and Routes of Dengue Virus Entry into the Host Cells. *FEMS Microbiol. Rev.* **2015**, *39*, 155–170. [\[CrossRef\]](https://doi.org/10.1093/femsre/fuu004)
- <span id="page-20-0"></span>131. Morsy, S.; Hashan, M.R.; Hieu, T.H.; Mohammed, A.A.; Elawady, S.S.; Ghosh, P.; Elgendy, M.A.; Le, H.-H.; Attiah, M.; Iqtadar, S.; et al. The Association between Dengue Viremia Kinetics and Dengue Severity: A Systemic Review and Meta-Analysis. *Rev. Med. Virol.* **2020**, *30*, 1–10. [\[CrossRef\]](https://doi.org/10.1002/rmv.2121)
- <span id="page-20-1"></span>132. Chaturvedi, U.C.; Agarwal, R.; Elbishbishi, E.A.; Mustafa, A.S. Cytokine Cascade in Dengue Hemorrhagic Fever: Implications for Pathogenesis. *FEMS Immunol. Med. Microbiol.* **2000**, *28*, 183–188. [\[CrossRef\]](https://doi.org/10.1111/j.1574-695X.2000.tb01474.x)
- <span id="page-20-2"></span>133. Malavige, G.N.; Jeewandara, C.; Ogg, G.S. Dengue and COVID-19: Two Sides of the Same Coin. *J. Biomed. Sci.* **2022**, *29*, 48. [\[CrossRef\]](https://doi.org/10.1186/s12929-022-00833-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35786403)
- <span id="page-20-3"></span>134. Rothman, A.L. Immunity to Dengue Virus: A Tale of Original Antigenic Sin and Tropical Cytokine Storms. *Nat. Rev. Immunol.* **2011**, *11*, 532–543. [\[CrossRef\]](https://doi.org/10.1038/nri3014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21760609)
- 135. Luplerdlop, N.; Missé, D.; Bray, D.; Deleuze, V.; Gonzalez, J.-P.; Leardkamolkarn, V.; Yssel, H.; Veas, F. Dengue-Virus-Infected Dendritic Cells Trigger Vascular Leakage through Metalloproteinase Overproduction. *EMBO Rep.* **2006**, *7*, 1176–1181. [\[CrossRef\]](https://doi.org/10.1038/sj.embor.7400814) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17028575)
- 136. Puerta-Guardo, H.; Raya-Sandino, A.; González-Mariscal, L.; Rosales, V.H.; Ayala-Davila, J.; Chávez-Mungía, B.; Martinez-Fong, D.; Medina, F.; Ludert, J.E.; del Angel, R.M. Rosa The Cytokine Response of U937-Derived Macrophages Infected through Antibody-Dependent Enhancement of Dengue Virus Disrupts Cell Apical-Junction Complexes and Increases Vascular Permeability. *J. Virol.* **2013**, *87*, 7486–7501. [\[CrossRef\]](https://doi.org/10.1128/JVI.00085-13) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23616663)
- <span id="page-20-4"></span>137. St John, A.L.; Rathore, A.P.S.; Raghavan, B.; Ng, M.-L.; Abraham, S.N. Contributions of Mast Cells and Vasoactive Products, Leukotrienes and Chymase, to Dengue Virus-Induced Vascular Leakage. *eLife* **2013**, *2*, e00481. [\[CrossRef\]](https://doi.org/10.7554/eLife.00481) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23638300)
- <span id="page-20-5"></span>138. Samuel, M.A.; Diamond, M.S. Pathogenesis of West Nile Virus Infection: A Balance between Virulence, Innate and Adaptive Immunity, and Viral Evasion. *J. Virol.* **2006**, *80*, 9349–9360. [\[CrossRef\]](https://doi.org/10.1128/JVI.01122-06) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16973541)
- <span id="page-20-6"></span>139. Martina, B.E.E.; Koraka, P.; Osterhaus, A.D.M.E. Dengue Virus Pathogenesis: An Integrated View. *Clin. Microbiol. Rev.* **2009**, *22*, 564–581. [\[CrossRef\]](https://doi.org/10.1128/CMR.00035-09) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19822889)
- <span id="page-20-7"></span>140. Steele, K.E.; Linn, M.J.; Schoepp, R.J.; Komar, N.; Geisbert, T.W.; Manduca, R.M.; Calle, P.P.; Raphael, B.L.; Clippinger, T.L.; Larsen, T.; et al. Pathology of Fatal West Nile Virus Infections in Native and Exotic Birds during the 1999 Outbreak in New York City, New York. *Vet. Pathol.* **2000**, *37*, 208–224. [\[CrossRef\]](https://doi.org/10.1354/vp.37-3-208)
- <span id="page-20-8"></span>141. Begum, F.; Das, S.; Mukherjee, D.; Mal, S.; Ray, U. Insight into the Tropism of Dengue Virus in Humans. *Viruses* **2019**, *11*, 1136. [\[CrossRef\]](https://doi.org/10.3390/v11121136)
- <span id="page-20-9"></span>142. Winkelmann, E.R.; Luo, H.; Wang, T. West Nile Virus Infection in the Central Nervous System. *F1000Research* **2016**, *5*, 105. [\[CrossRef\]](https://doi.org/10.12688/f1000research.7404.1)
- <span id="page-20-10"></span>143. Petersen, L.R.; Marfin, A.A. West Nile Virus: A Primer for the Clinician. *Ann. Intern. Med.* **2002**, *137*, 173. [\[CrossRef\]](https://doi.org/10.7326/0003-4819-137-3-200208060-00009)
- <span id="page-20-11"></span>144. Watson, J.T.; Pertel, P.E.; Jones, R.C.; Siston, A.M.; Paul, W.S.; Austin, C.C.; Gerber, S.I. Clinical Characteristics and Functional Outcomes of West Nile Fever. *Ann. Intern. Med.* **2004**, *141*, 360. [\[CrossRef\]](https://doi.org/10.7326/0003-4819-141-5-200409070-00010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15353427)
- <span id="page-20-12"></span>145. Roberts, J.A.; Kim, C.Y.; Dean, A.; Kulas, K.E.; St George, K.; Hoang, H.E.; Thakur, K.T. Clinical and Diagnostic Features of West Nile Virus Neuroinvasive Disease in New York City. *Pathogens* **2024**, *13*, 382. [\[CrossRef\]](https://doi.org/10.3390/pathogens13050382) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38787234)
- <span id="page-20-13"></span>146. Montgomery, R.R.; Murray, K.O. Risk Factors for West Nile Virus Infection and Disease in Populations and Individuals. *Expert Rev. Anti-Infect. Ther.* **2015**, *13*, 317–325. [\[CrossRef\]](https://doi.org/10.1586/14787210.2015.1007043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25637260)
- <span id="page-20-14"></span>147. Bilal Khan, M.; Yang, Z.S.; Lin, C.C.; Hsu, M.C.; Nayim Urbina, A.; Assavalapsakul, W.; Wang, W.H.; Chen, Y.H.; Wang, S.F. Dengue Overview: An Updated Systemic Review. *J. Infect. Public Health* **2023**, *16*, 1625–1642. [\[CrossRef\]](https://doi.org/10.1016/j.jiph.2023.08.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37595484)
- <span id="page-20-15"></span>148. Ripoll, D.R.; Wallqvist, A.; Chaudhury, S. Molecular Simulations Reveal the Role of Antibody Fine Specificity and Viral Maturation State on Antibody-Dependent Enhancement of Infection in Dengue Virus. *Front. Cell. Infect. Microbiol.* **2019**, *9*, 200. [\[CrossRef\]](https://doi.org/10.3389/fcimb.2019.00200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31275864)
- <span id="page-20-16"></span>149. Guzman, M.G.; Alvarez, M.; Halstead, S.B. Secondary Infection as a Risk Factor for Dengue Hemorrhagic Fever/Dengue Shock Syndrome: An Historical Perspective and Role of Antibody-Dependent Enhancement of Infection. *Arch. Virol.* **2013**, *158*, 1445–1459. [\[CrossRef\]](https://doi.org/10.1007/s00705-013-1645-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23471635)
- 150. Dutta, D.; Ghosh, A.C.; Dutta, C.; Soumi, S.; Biswas, S. Cross Reactivity of SARS-CoV-2 with Other Pathogens, Especially Dengue Virus: A Historical Perspective. *J. Med. Virol.* **2023**, *95*, e28557. [\[CrossRef\]](https://doi.org/10.1002/jmv.28557) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36755367)
- 151. Dejnirattisai, W.; Jumnainsong, A.; Onsirisakul, N.; Fitton, P.; Vasanawathana, S.; Limpitikul, W.; Puttikhunt, C.; Edwards, C.; Duangchinda, T.; Supasa, S.; et al. Cross-Reacting Antibodies Enhance Dengue Virus Infection in Humans. *Science* **2010**, *328*, 745–748. [\[CrossRef\]](https://doi.org/10.1126/science.1185181)
- 152. Halstead, S.B. Observations Related to Pathogensis of Dengue Hemorrhagic Fever. VI. Hypotheses and Discussion. *Yale J. Biol. Med.* **1970**, *42*, 350–362.
- <span id="page-20-17"></span>153. Halstead, S.B. Pathogenesis of Dengue: Dawn of a New Era. *F1000Research* **2015**, *4*, F100 Faculty Rev-1353. [\[CrossRef\]](https://doi.org/10.12688/f1000research.7024.1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26918141)
- <span id="page-20-18"></span>154. Teo, A.; Tan, H.D.; Loy, T.; Chia, P.Y.; Chua, C.L.L. Understanding Antibody-Dependent Enhancement in Dengue: Are Afucosylated IgG1s a Concern? *PLoS Pathog.* **2023**, *19*, e1011223. [\[CrossRef\]](https://doi.org/10.1371/journal.ppat.1011223)
- <span id="page-20-19"></span>155. Gorchakov, R.; Gulas-Wroblewski, B.E.; Ronca, S.E.; Ruff, J.C.; Nolan, M.S.; Berry, R.; Alvarado, R.E.; Gunter, S.M.; Murray, K.O. Optimizing PCR Detection of West Nile Virus from Body Fluid Specimens to Delineate Natural History in an Infected Human Cohort. *Int. J. Mol. Sci.* **2019**, *20*, 1934. [\[CrossRef\]](https://doi.org/10.3390/ijms20081934) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31010160)
- <span id="page-21-0"></span>156. Armah, H.B.; Wang, G.; Omalu, B.I.; Tesh, R.B.; Gyure, K.A.; Chute, D.J.; Smith, R.D.; Dulai, P.; Vinters, H.V.; Kleinschmidt-DeMasters, B.K.; et al. Systemic Distribution of West Nile Virus Infection: Postmortem Immunohistochemical Study of Six Cases. *Brain Pathol.* **2007**, *17*, 354–362. [\[CrossRef\]](https://doi.org/10.1111/j.1750-3639.2007.00080.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17610522)
- <span id="page-21-1"></span>157. Sandhu, T.; Sidhu, D.; Dhillon, M. Antigenic Distribution of West Nile Virus in Various Organs of Wildly Infected American Crows (*Corvus Brachyrhynchos*). *J. Glob. Infect. Dis.* **2011**, *3*, 138–142. [\[CrossRef\]](https://doi.org/10.4103/0974-777X.81690) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21731300)
- <span id="page-21-2"></span>158. Palmieri, C.; Franca, M.; Uzal, F.; Anderson, M.; Barr, B.; Woods, L.; Moore, J.; Woolcock, P.; Shivaprasad, H.L. Pathology and Immunohistochemical Findings of West Nile Virus Infection in Psittaciformes. *Vet. Pathol.* **2010**, *48*, 975–984. [\[CrossRef\]](https://doi.org/10.1177/0300985810391112) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21160024)
- <span id="page-21-3"></span>159. Kelley, R.E.; Berger, J.R.; Kelley, B.P. WEST NILE VIRUS MENINGO-ENCEPHALITIS: POSSIBLE SEXUAL TRANSMISSION. *J. La. State Med. Soc. Off. Organ La. State Med. Soc.* **2016**, *168*, 21–22.
- <span id="page-21-4"></span>160. Blitvich, B.J.; Magalhaes, T.; Laredo-Tiscareño, S.V.; Foy, B.D. Sexual Transmission of Arboviruses: A Systematic Review. *Viruses* **2020**, *12*, 933. [\[CrossRef\]](https://doi.org/10.3390/v12090933) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32854298)
- <span id="page-21-5"></span>161. Liew, C.H. The First Case of Sexual Transmission of Dengue in Spain. *J. Travel Med.* **2019**, *27*, taz087. [\[CrossRef\]](https://doi.org/10.1093/jtm/taz087)
- 162. Le Tortorec, A.; Matusali, G.; Mahé, D.; Aubry, F.; Mazaud-Guittot, S.; Houzet, L.; Dejucq-Rainsford, N. From Ancient to Emerging Infections: The Odyssey of Viruses in the Male Genital Tract. *Physiol. Rev.* **2020**, *100*, 1349–1414. [\[CrossRef\]](https://doi.org/10.1152/physrev.00021.2019)
- 163. Lalle, E.; Colavita, F.; Iannetta, M.; Gebremeskel Teklè, S.; Carletti, F.; Scorzolini, L.; Bordi, L.; Vincenti, D.; Castilletti, C.; Ippolito, G.; et al. Prolonged Detection of Dengue Virus RNA in the Semen of a Man Returning from Thailand to Italy, January 2018. *Eurosurveillance* **2018**, *23*, 18–00197. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2018.23.18.18-00197) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29741153)
- 164. Mons, J.; Mahé-Poiron, D.; Mansuy, J.-M.; Lheureux, H.; Nigon, D.; Moinard, N.; Hamdi, S.; Pasquier, C.; Dejucq-Rainsford, N.; Bujan, L. Effects of Acute Dengue Infection on Sperm and Virus Clearance in Body Fluids of Men. *Emerg. Infect. Dis.* **2022**, *28*, 1146. [\[CrossRef\]](https://doi.org/10.3201/eid2806.212317) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35608553)
- <span id="page-21-6"></span>165. Dengue Fever –Spain. Available online: <https://www.who.int/emergencies/disease-outbreak-news/item/2019-DON214> (accessed on 22 June 2024).
- <span id="page-21-7"></span>166. Iannetta, M.; Lalle, E.; Musso, M.; Carletti, F.; Scorzolini, L.; D'Abramo, A.; Chinello, P.; Castilletti, C.; Ippolito, G.; Capobianchi, M.R.; et al. Persistent Detection of Dengue Virus RNA in Vaginal Secretion of a Woman Returning from Sri Lanka to Italy, April 2017. *Eurosurveillance* **2017**, *22*, 30600. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2017.22.34.30600) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28857045)
- <span id="page-21-8"></span>167. Lee, C.; Lee, H. Probable Female to Male Sexual Transmission of Dengue Virus Infection. *Infect. Dis.* **2018**, *51*, 150–152. [\[CrossRef\]](https://doi.org/10.1080/23744235.2018.1521004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30318968)
- <span id="page-21-9"></span>168. European Centre for Disease Prevention and Control West Nile Virus Infection-Annual Epidemiological Report for 2018. Available online: <https://www.ecdc.europa.eu/en/publications-data/west-nile-virus-infection-annual-epidemiological-report-2018> (accessed on 16 July 2024).
- <span id="page-21-10"></span>169. European Centre for Disease Prevention and Control West Nile Virus Infection-Annual Epidemiological Report for 2019. Available online: <https://www.ecdc.europa.eu/en/publications-data/west-nile-virus-infection-annual-epidemiological-report-2019> (accessed on 16 July 2024).
- <span id="page-21-11"></span>170. European Centre for Disease Prevention and Control Epidemiological Update: West Nile Virus Transmission Season in Europe. 2020. Available online: [https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2020)[season-europe-2020](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2020) (accessed on 10 August 2023).
- 171. European Centre for Disease Prevention and Control Epidemiological Update: West Nile Virus Transmission Season in Europe. 2021. Available online: [https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2021)[season-europe-2021](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2021) (accessed on 16 July 2024).
- <span id="page-21-12"></span>172. Epidemiological Update: West Nile Virus Transmission Season in Europe. 2022. Available online: [https://www.ecdc.europa.eu/](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2022) [en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2022](https://www.ecdc.europa.eu/en/news-events/epidemiological-update-west-nile-virus-transmission-season-europe-2022) (accessed on 16 July 2024).
- <span id="page-21-13"></span>173. Weekly Updates: 2023 West Nile Virus Transmission Season. Available online: [https://www.ecdc.europa.eu/en/west-nile-fever/](https://www.ecdc.europa.eu/en/west-nile-fever/surveillance-and-disease-data/disease-data-ecdc) [surveillance-and-disease-data/disease-data-ecdc](https://www.ecdc.europa.eu/en/west-nile-fever/surveillance-and-disease-data/disease-data-ecdc) (accessed on 16 July 2024).
- <span id="page-21-14"></span>174. European Centre for Disease Prevention and Control Dengue-Annual Epidemiological Report for 2018. Available online: [https:](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2018) [//www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2018](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2018) (accessed on 30 December 2023).
- <span id="page-21-18"></span>175. European Centre for Disease Prevention and Control Dengue-Annual Epidemiological Report for 2019. Available online: [https:](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2019) [//www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2019](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2019) (accessed on 30 December 2023).
- 176. European Centre for Disease Prevention and Control Dengue-Annual Epidemiological Report for 2020. Available online: [https:](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2020) [//www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2020](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2020) (accessed on 30 December 2023).
- <span id="page-21-19"></span>177. European Centre for Disease Prevention and Control Dengue-Annual Epidemiological Report for 2021. Available online: [https:](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2021) [//www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2021](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2021) (accessed on 30 December 2023).
- <span id="page-21-15"></span>178. Dengue-Annual Epidemiological Report for 2022. Available online: [https://www.ecdc.europa.eu/en/publications-data/dengue](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2022)[annual-epidemiological-report-2022](https://www.ecdc.europa.eu/en/publications-data/dengue-annual-epidemiological-report-2022) (accessed on 12 July 2024).
- <span id="page-21-16"></span>179. Autochthonous Transmission of Dengue Virus in Mainland EU/EEA, 2010-Present. Available online: [https://www.ecdc.europa.](https://www.ecdc.europa.eu/en/all-topics-z/dengue/surveillance-and-disease-data/autochthonous-transmission-dengue-virus-eueea) [eu/en/all-topics-z/dengue/surveillance-and-disease-data/autochthonous-transmission-dengue-virus-eueea](https://www.ecdc.europa.eu/en/all-topics-z/dengue/surveillance-and-disease-data/autochthonous-transmission-dengue-virus-eueea) (accessed on 16 July 2024).
- <span id="page-21-17"></span>180. EpiCentro Arbovirosi • Bollettini Periodici Arbovirosi. Available online: <https://www.epicentro.iss.it/arbovirosi/bollettini> (accessed on 16 July 2024).
- <span id="page-22-0"></span>181. Michel, F.; Sieg, M.; Fischer, D.; Keller, M.; Eiden, M.; Reuschel, M.; Schmidt, V.; Schwehn, R.; Rinder, M.; Urbaniak, S.; et al. Evidence for West Nile Virus and Usutu Virus Infections in Wild and Resident Birds in Germany, 2017 and 2018. *Viruses* **2019**, *11*, 674. [\[CrossRef\]](https://doi.org/10.3390/v11070674) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31340516)
- <span id="page-22-1"></span>182. Ruscher, C.; Patzina-Mehling, C.; Melchert, J.; Graff, S.L.; McFarland, S.E.; Hieke, C.; Kopp, A.; Prasser, A.; Tonn, T.; Schmidt, M.; et al. Ecological and Clinical Evidence of the Establishment of West Nile Virus in a Large Urban Area in Europe, Berlin, Germany, 2021 to 2022. *Eurosurveillance* **2023**, *28*, 2300258. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2023.28.48.2300258) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38037727)
- <span id="page-22-2"></span>183. Department for Environment, Food and Rural Affairs Animal and Plant Health Agency Advice Services—International Disease Monitoring 1 Updated Outbreak Assessment West Nile Virus in Germany and Southern Europe Disease Report. 2018. Available online: <https://assets.publishing.service.gov.uk/media/61606cecd3bf7f55fe946bc2/300921-oa-wnv-germany.pdf> (accessed on 16 July 2024).
- <span id="page-22-3"></span>184. Lu, L.; Zhang, F.; Oude Munnink, B.B.; Munger, E.; Sikkema, R.S.; Pappa, S.; Tsioka, K.; Sinigaglia, A.; Dal Molin, E.; Shih, B.B.; et al. West Nile Virus Spread in Europe: Phylogeographic Pattern Analysis and Key Drivers. *PLoS Pathog.* **2024**, *20*, e1011880. [\[CrossRef\]](https://doi.org/10.1371/journal.ppat.1011880) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38271294)
- <span id="page-22-4"></span>185. Mamun, M.A.; Misti, J.M.; Griffiths, M.D.; Gozal, D. The Dengue Epidemic in Bangladesh: Risk Factors and Actionable Items. *Lancet* **2019**, *394*, 2149–2150. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(19)32524-3)
- <span id="page-22-5"></span>186. Pandey, B.D.; Costello, A. The Dengue Epidemic and Climate Change in Nepal. *Lancet* **2019**, *394*, 2150–2151. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(19)32689-3)
- <span id="page-22-6"></span>187. Zambrano, L.I.; Rodriguez, E.; Espinoza-Salvado, I.A.; Fuentes-Barahona, I.C.; Lyra de Oliveira, T.; Luciano da Veiga, G.; Cláudio da Silva, J.; Valle-Reconco, J.A.; Rodríguez-Morales, A.J. Spatial Distribution of Dengue in Honduras during 2016–2019 Using a Geographic Information Systems (GIS)–Dengue Epidemic Implications for Public Health and Travel Medicine. *Travel Med. Infect. Dis.* **2019**, *32*, 101517. [\[CrossRef\]](https://doi.org/10.1016/j.tmaid.2019.101517)
- <span id="page-22-7"></span>188. La Ruche, G.; Souarès, Y.; Armengaud, A.; Peloux-Petiot, F.; Delaunay, P.; Desprès, P.; Lenglet, A.; Jourdain, F.; Leparc-Goffart, I.; Charlet, F.; et al. First Two Autochthonous Dengue Virus Infections in Metropolitan France, September 2010. *Eurosurveillance* **2010**, *15*, 19676. [\[CrossRef\]](https://doi.org/10.2807/ese.15.39.19676-en)
- <span id="page-22-8"></span>189. Gjenero-Margan, I.; Aleraj, B.; Krajcar, D.; Lesnikar, V.; Klobučar, A.; Pem-Novosel, I.; Kurečić-Filipović, S.; Komparak, S.; Martić, R.; Đuričić, S.; et al. Autochthonous Dengue Fever in Croatia, August-September 2010. *Eurosurveillance* 2011, 16, 19805. [\[CrossRef\]](https://doi.org/10.2807/ese.16.09.19805-en) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21392489)
- <span id="page-22-9"></span>190. Franke, F.; Giron, S.; Cochet, A.; Jeannin, C.; Leparc-Goffart, I.; de Valk, H.; Jourdain, F.; de Lamballerie, X.; L'Ambert, G.; Paty, M.C. Autochthonous Chikungunya and Dengue Fever Outbreak in Mainland France, 2010-2018. *European Journal of Public Health* **2019**, *29*. [\[CrossRef\]](https://doi.org/10.1093/eurpub/ckz186.628)
- <span id="page-22-10"></span>191. Marchand, E.; Prat, C.; Jeannin, C.; Lafont, E.; Bergmann, T.; Flusin, O.; Rizzi, J.; Roux, N.; Busso, V.; Deniau, J.; et al. Autochthonous Case of Dengue in France, October 2013. *Eurosurveillance* **2013**, *18*, 20661. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES2013.18.50.20661)
- <span id="page-22-11"></span>192. European Centre for Disease Prevention and Control Main Conclusions and Options for Response Main Conclusions. 2018. Available online: <https://www.ecdc.europa.eu/sites/default/files/documents/08-10-2018-RRA-Dengue-France.pdf> (accessed on 16 July 2024).
- <span id="page-22-12"></span>193. Monge, S.; García-Ortúzar, V.; López Hernández, B.; Lopaz Pérez, M.; Delacour-Estrella, S.; Sánchez-Seco, M.; Fernández Martinez, B.; García San Miguel, L.; García-Fulgueiras, A.; Sierra Moros, M. Characterization of the First Autochthonous Dengue Outbreak in Spain (August–September 2018). *Acta Trop.* **2020**, *205*, 105402. [\[CrossRef\]](https://doi.org/10.1016/j.actatropica.2020.105402)
- <span id="page-22-13"></span>194. Cochet, A.; Calba, C.; Jourdain, F.; Grard, G.; Durand, G.A.; Guinard, A.; Noël, H.; Paty, M.-C.; Franke, F. Autochthonous Dengue in Mainland France, 2022: Geographical Extension and Incidence Increase. *Eurosurveillance* **2022**, *27*, 2200818. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2022.27.44.2200818) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36330819)
- <span id="page-22-14"></span>195. Virus Dengue, Identificato in Provincia Di Lodi Primo Caso Lombardo. Available online: [https://www.lombardianotizie.online/](https://www.lombardianotizie.online/virus-dengue-lodi/) [virus-dengue-lodi/](https://www.lombardianotizie.online/virus-dengue-lodi/) (accessed on 30 December 2023).
- 196. Virus Dengue, a Castiglione d'Adda (Lodi) Individuati Altri 2 Casi. Available online: [https://www.lombardianotizie.online/](https://www.lombardianotizie.online/virus-dengue/) [virus-dengue/](https://www.lombardianotizie.online/virus-dengue/) (accessed on 30 December 2023).
- <span id="page-22-16"></span>197. EpiCentro Arbovirosi • Riepilogo 2023. Available online: <https://www.epicentro.iss.it/arbovirosi/dashboard-2023> (accessed on 16 July 2024).
- <span id="page-22-15"></span>198. De Carli, G.; Carletti, F.; Spaziante, M.; Maria, E.; Rueca, M.; Spezia, P.G.; Vantaggio, V.; Barca, A.; De Liberato, C.; Romiti, F.; et al. Outbreaks of Autochthonous Dengue in Lazio Region, Italy, August to September 2023: Preliminary Investigation. *Eurosurveillance* **2023**, *28*, 2300552. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2023.28.44.2300552) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37917030)
- <span id="page-22-17"></span>199. Sante Publique France Chikungunya, Dengue et Zika-Chikungunya, Dengue et Zika-Données de La Surveillance Renforcée En France Métropolitaine En 2022. Available online: [https://www.santepubliquefrance.fr/maladies-et-traumatismes/maladies-a](https://www.santepubliquefrance.fr/maladies-et-traumatismes/maladies-a-transmission-vectorielle/chikungunya/articles/donnees-en-france-metropolitaine/chikungunya-dengue-et-zika-donnees-de-la-surveillance-renforcee-en-france-metropolitaine-en-2022#block-452632)[transmission-vectorielle/chikungunya/articles/donnees-en-france-metropolitaine/chikungunya-dengue-et-zika-donnees](https://www.santepubliquefrance.fr/maladies-et-traumatismes/maladies-a-transmission-vectorielle/chikungunya/articles/donnees-en-france-metropolitaine/chikungunya-dengue-et-zika-donnees-de-la-surveillance-renforcee-en-france-metropolitaine-en-2022#block-452632)[de-la-surveillance-renforcee-en-france-metropolitaine-en-2022#block-452632](https://www.santepubliquefrance.fr/maladies-et-traumatismes/maladies-a-transmission-vectorielle/chikungunya/articles/donnees-en-france-metropolitaine/chikungunya-dengue-et-zika-donnees-de-la-surveillance-renforcee-en-france-metropolitaine-en-2022#block-452632) (accessed on 16 July 2024).
- <span id="page-22-18"></span>200. Barzon, L.; Montarsi, F.; Quaranta, E.; Monne, I.; Pacenti, M.; Michelutti, A.; Toniolo, F.; Danesi, P.; Marchetti, G.; Gobbo, F.; et al. Early Start of Seasonal Transmission and Co-Circulation of West Nile Virus Lineage 2 and a Newly Introduced Lineage 1 Strain, Northern Italy, June 2022. *Eurosurveillance* **2022**, *27*, 2200548. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2022.27.29.2200548)
- <span id="page-22-19"></span>201. EpiCentro La Sorveglianza Dei Casi Umani Di Infezione Da West Nile Virus. Available online: [https://www.epicentro.iss.it/](https://www.epicentro.iss.it/westnile/bollettino) [westnile/bollettino](https://www.epicentro.iss.it/westnile/bollettino) (accessed on 30 December 2023).
- <span id="page-23-0"></span>202. Barzon, L.; Gobbi, F.; Capelli, G.; Montarsi, F.; Martini, S.; Riccetti, S.; Sinigaglia, A.; Pacenti, M.; Pavan, G.; Rassu, M.; et al. Autochthonous Dengue Outbreak in Italy 2020: Clinical, Virological and Entomological Findings. *J. Travel Med.* **2021**, *28*, taab130. [\[CrossRef\]](https://doi.org/10.1093/jtm/taab130) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34409443)
- <span id="page-23-1"></span>203. Gutiérrez-Bugallo, G.; Boullis, A.; Martinez, Y.; Hery, L.; Rodríguez, M.; Bisset, J.A.; Vega-Rúa, A. Vector Competence of *Aedes Aegypti* from Havana, Cuba, for Dengue Virus Type 1, Chikungunya, and Zika Viruses. *PLoS Neglected Trop. Dis.* **2020**, *14*, e0008941. [\[CrossRef\]](https://doi.org/10.1371/journal.pntd.0008941)
- <span id="page-23-3"></span>204. Mutoh, Y.; Moriya, A.; Yasui, Y.; Saito, N.; Takasaki, T.; Hiramatsu, S.; Izuchi, T.; Umemura, T.; Ichihara, T. Two Cases of Dengue Virus Type 2 (DENV-2) Infection in a Japanese Couple Returning from the Maldives during the 2018 Dengue Outbreak. *Jpn. J. Infect. Dis.* **2020**, *73*, 58–60. [\[CrossRef\]](https://doi.org/10.7883/yoken.JJID.2019.153) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31474701)
- <span id="page-23-2"></span>205. Thisyakorn, U.; Saokaew, S.; Gallagher, E.; Kastner, R.; Sruamsiri, R.; Oliver, L.; Hanley, R. Epidemiology and Costs of Dengue in Thailand: A Systematic Literature Review. *PLoS Neglected Trop. Dis.* **2022**, *16*, e0010966. [\[CrossRef\]](https://doi.org/10.1371/journal.pntd.0010966) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36534668)
- <span id="page-23-4"></span>206. Cassaniti, I.; Ferrari, G.; Senatore, S.; Rossetti, E.; Defilippo, F.; Maffeo, M.; Vezzosi, L.; Campanini, G.; Sarasini, A.; Paolucci, S.; et al. Preliminary Results on an Autochthonous Dengue Outbreak in Lombardy Region, Italy, August 2023. *Eurosurveillance* **2023**, *28*, 2300471. [\[CrossRef\]](https://doi.org/10.2807/1560-7917.ES.2023.28.37.2300471) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37707980)

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