

Dominance between self-incompatibility alleles determines the mating system of *Capsella* allopolyploids

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Abstract

The shift from outcrossing to self-fertilization is one of the main evolutionary transitions in plants and has broad effects on evolutionary trajectories. In Brassicaceae, the ability to inhibit self-fertilization is controlled by 2 genes, SCR and SRK, tightly linked within the S-locus. A series of small non-coding RNAs also encoded within the S-locus regulates the transcriptional activity of SCR alleles, resulting in a linear dominance hierarchy between them. In Brassicaceae, natural allopolyploid species are often self-compatible (SC) even when one of the progenitor species is self-incompatible, but the reason why polyploid lineages tend to lose self-incompatibility (SI) and the timing of the loss of SI (immediately after ancestral hybridization between the progenitor species, or at a later stage after the formation of allopolyploid lineages) have generally remained elusive. We used a series of synthetic diploid and tetraploid hybrids obtained between self-fertilizing *Capsella orientalis* and outcrossing *Capsella grandiflora* to test whether the breakdown of SI could be observed immediately after hybridization, and whether the occurrence of SC phenotypes could be explained by the dominance interactions between S-haplotypes inherited from the parental lineages. We used RNA-sequencing data from young inflorescences to measure allele-specific expression of the SCR gene and infer dominance interactions in the synthetic hybrids. We then evaluated the seed set from autonomous self-pollination in the synthetic hybrids. Our results demonstrate that self-compatibility of the hybrids depends on the relative dominance between S-alleles inherited from the parental species, confirming that SI can be lost instantaneously upon formation of the ancestral allopolyploid lineage. They also confirm that the epigenetic regulation that controls dominance interactions between S-alleles can function between subgenomes in allopolyploids. Together, our results illustrate how a detailed knowledge of the mechanisms controlling SI can illuminate our understanding of the patterns of co-variation between the mating system and changes in ploidy.

Keywords: self-incompatibility, polyploidy, SRK, SCR, genetic dominance, *Capsella*

Lay Summary

Polyploidy is the inheritable condition of carrying more than two sets of chromosomes. It can result from within-species genome duplication (autopolyploidy), or from the merging of sets of chromosomes from different species following hybridization (allopolyploidy). Because sexual reproduction between individuals of different levels of ploidy is generally not successful, self-fertilization has been considered a key component of the establishment success of polyploid lineages. However, the reasons why the mating system of polyploids may differ from that of their parental species remains mysterious. In plants of the Brassicaceae family, several allopolyploid species arose from hybridization between an outcrossing and a self-fertilizing species, and in most cases, the resulting lineages are self-fertilizing. It has been proposed that the mating system of these allopolyploids depends on the dominance relationships between the functional and non-functional self-incompatibility alleles inherited from the parental species. Here, we tested this prediction by characterizing at the transcriptional (RNA-seq) and phenotypic levels (estimation of autonomous seed production) a series of synthetic *Capsella* diploid and tetraploid hybrids. We found that the predicted dominance relationships closely matched the observed expression of self-incompatibility alleles, as well as the self-compatibility phenotypes. Hence, the mating system of newly formed *Capsella* allotetraploids depends on the dominance relationship between self-incompatibility alleles inherited from the parents. Overall, our results improve our understanding of the mechanisms by which changes in ploidy can alter the system of mating over the course of evolution.

Introduction

Mating systems have far-reaching effects on plant evolution (Wright et al., 2013). For instance, shifts from outcrossing to self-fertilization are expected to reduce the effective rate of recombination and genetic polymorphism (Glémin et al., 2006),

while at the same time benefiting from a transmission advantage (Fisher, 1941) and providing reproductive assurance when mates are scarce (Jain, 1976). The establishment of polyploid populations is an iconic example of these effects. Whole-genome duplication (WGD) is prevalent in plants (Soltis et al., 2015), and polyploid

Received April 19, 2023; revisions received January 11, 2024; accepted March 4, 2024

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species are overrepresented in the Arctic flora (Brochmann et al., 2004) as well as in invasive (Pandit et al., 2011) and domesticated plants (Salman-Minkov et al., 2016). Moreover, ancient WGD events on phylogenies seem to be associated with drastic environmental changes (Vanneste et al., 2014). Therefore, WGD has often been hypothesized to allow faster adaptation and niche differentiation in changing environments (Baniaga et al., 2020; Selmecki et al., 2015). However, besides these potential long-term advantages, newly formed polyploid genotypes are also expected to suffer from the immediate lack of gametes of the same cytotype and from the lower fitness of interploidy hybrids. This phenomenon, known as “minority cytotype exclusion” (Husband, 2000; Levin, 1975), is expected to drastically hinder the success of newly formed polyploid lineages.

Self-fertilization should greatly increase the establishment success of polyploid populations by allowing them to avoid minority cytotype exclusion (Fowler & Levin, 2016). However, empirical surveys on the association between polyploidy and self-fertilization either confirmed the positive association (Barringer, 2007; Robertson et al., 2011) or found no association (Mable, 2004). This suggests that the current models fail to incorporate important details of the interaction between polyploidy and self-fertilization, such as genetic mechanisms controlling the mating system or the confounding effects of hybridization. An intriguing observation is that allopolyploid lineages (in which WGD occurred in association with hybridization) often exhibit low outcrossing rates, whereas autopolyploid lineages (where “only” WGD occurred) often exhibit predominant outcrossing or mixed mating systems (Husband et al., 2008), a pattern that was already suggested by Stebbins (1957). It is unknown whether these contrasted outcomes are due to the immediate effect of WGD on factors controlling the mating system, or to population genomic differences between auto- and allopolyploids that could influence the evolution of selfing at a later stage within the neopolyploid lineages.

The mating system of Brassicaceae species is controlled by a sporophytic self-incompatibility system, in which self-pollen is recognized by the allele-specific interaction between a pollen coat ligand protein (encoded by the SCR gene) and a stigma transmembrane receptor kinase (encoded by the SRK gene, Takayama et al., 2001). The two genes are tightly linked within a small genomic region called the S-locus, where a large number of S-alleles (also called S-haplotypes) typically segregate in self-incompatible species. S-haplotypes form a complex dominance hierarchy in anthers, whereby small non-coding RNA (sRNA) generated by dominant S-haplotypes transcriptionally silence the SCR gene of recessive S-haplotypes (Durand et al., 2014; Tarutani et al., 2010). Hence, while a large fraction of individuals are heterozygous at the S-locus, in most cases transcripts from only one of the two SCR alleles are present (Burghgraeve et al., 2020; Kakizaki et al., 2003), resulting in phenotypic dominance. Diversification of S-haplotypes in Brassicaceae is very ancient, as indicated by the very high level of nucleotide divergence among S-haplotype sequences and extensive trans-specific and even trans-generic sharing among related taxa (Castric & Vekemans, 2004). In *Arabidopsis* and *Capsella*, S-haplotypes are classified into four main dominance classes, related to their phylogenetic relationships with class I being the most recessive and class IV being the most dominant (Bachmann et al., 2019; Durand et al., 2014; Prigoda et al., 2005).

Several allopolyploid species of the Brassicaceae family originated from the hybridization between a self-incompatible (SI) and a self-compatible (SC) parental species, including *Arabidopsis suecica* (with *Arabidopsis thaliana* as SC parent and *Arabidopsis*

arenosa as SI parent; Novikova et al., 2017), *Arabidopsis kamchatica* (with *Arabidopsis lyrata* as SC parent and *Arabidopsis halleri* as SI parent; Kolesnikova et al., 2023; Shimizu-Inatsugi et al., 2009), and *Capsella bursa-pastoris* (with *Capsella orientalis* as SC parent and *Capsella grandiflora* as SI parent; Douglas et al., 2015). These three species have a recent allopolyploid origin, and all share the common feature of being self-compatible. The reason why these allopolyploid species originating from SI × SC hybridization are SC rather than SI is intriguing. An interesting possibility could be that the dominance interactions between S-haplotypes could have caused the instantaneous breakdown of SI in these species if the (non-functional) S-haplotype contributed by the SC species had retained the ability to suppress the expression of the (functional) SCR alleles contributed by the SI species in hybrid offspring (reviewed in Novikova et al., 2023). Consistent with this hypothesis, the allotetraploid *A. suecica*, *C. bursa-pastoris*, and some accessions of *A. kamchatica* all share the same nonfunctional S-allele as that of their respective SC parental species (Bachmann et al., 2019, 2021; Kolesnikova et al., 2023; Novikova et al., 2017). In addition, at least some resynthesized *A. suecica*- or *C. bursa-pastoris*-like allotetraploids are immediately SC after hybridization (Bachmann et al., 2021; Duan et al., 2023; Novikova et al., 2017), also supporting that the loss of SI could be an instant outcome of possessing one (relatively dominant) non-functional S-haplotype. A key prediction from this scenario is that the self-incompatibility of the resulting hybrid should vary according to the dominance of the S-haplotype contributed by the SI parent relative to that of the SC parent. However, formal proof of this hypothetical process, i.e., the establishment of a direct causal link between the relative dominance of the S-haplotypes, the expression of SCR alleles in anthers, and the loss of SI in allopolyploids, is still lacking. Evidence for the effect of dominance between functional and non-functional SCR alleles on the SC phenotype of transgenic lines of the allopolyploid *A. kamchatica* has been recently demonstrated (Yew et al., 2023), but the origin of the non-functional mutations occurring *in natura* (i.e., whether inherited from one parental species or appearing *de novo* within the neopolyploid lineage) remains unknown.

Here, we used an experimental approach based on a series of synthetic allopolyploid individuals obtained between the selfer *C. orientalis* and the outcrosser *C. grandiflora* (Duan et al., 2023) to test whether the breakdown of SI observed in allopolyploid species in Brassicaceae, such as *C. bursa-pastoris*, could be explained by the dominance interaction between S-haplotypes in anthers. First, we used published genomic and transcriptomic resequencing data to establish a methodology to infer S-locus genotypes and pollen S-locus phenotypes in *Capsella*. Then we used RNA-sequencing (RNA-seq) data from young inflorescences to measure allele-specific expression of the SCR and SRK genes in synthetic diploid and tetraploid *C. orientalis* × *C. grandiflora* hybrids, as an approximation of the early stages of natural *C. bursa-pastoris*. Those patterns of expression were used to infer patterns of allele dominance in anthers, and in particular those between the non-functional allele of *C. orientalis* and the alleles inherited from *C. grandiflora*. Finally, we compared the observed expression of SCR alleles with the seed set from autonomous self-pollination in the synthetic hybrids. Altogether, our results demonstrate that the relative dominance of S-alleles inherited from the parental species is a key determinant of the ability to self-fertilize in nascent allopolyploid lineages providing one potential explanation for the higher occurrence of selfing in allopolyploids than in autopolyploids in families with sporophytic SI.

Results

A methodology to determine S-locus genotypes and phenotypes in *Capsella*

First, we produced a comprehensive set of S-allele reference sequences in *Capsella*. For this, we genotyped individuals at the SRK gene using the NGSgenotyp pipeline (Genete et al., 2020) on publicly available short-read resequencing data of 180 *C. grandiflora* individuals from Monodendri, Greece (the Cg-9 population in Josephs et al., 2015). We started from a database of SRK sequences from *A. lyrata* and *A. halleri* that we complemented with 62 partial *C. grandiflora* SRK sequences previously obtained by Sanger sequencing by Jesper Bechsgaard and Mikkel Schierup (Guo et al., 2009; Neuffer et al., 2023; see Supplementary Information). We obtained a fully resolved S-locus genotype for 177 individuals (Supplementary Table S1), and identified 74 different S-alleles, including 25 previously unknown *C. grandiflora* S-alleles (Supplementary Table S2). For most of them, we were able to obtain full sequences of the exon 1 of SRK (available at <https://www.doi.org/10.6084/m9.figshare.22567558.v2>). Interestingly, one of those new S-alleles, noted H4047 (see Supplementary Information for description of the S-alleles notation), shared 99% identity with the SRK pseudogene (for which the coding sequence is interrupted at position 949 of exon 1) found at the S-locus in subgenome A of *C. bursa-pastoris* by Bachmann et al. (2021). A more complete description of this set of S-alleles is given in the Supplementary Information. Beside these 74 SRK alleles, we also identified five sequences clustering with SRK alleles (H0002, H0003, H0011, H0012, and H0013 in Supplementary Table S1, named CgrSRK01, CgrSRK06, CgrSRK09, CgrSRK51, and CgrSRK63 in Paetsch et al., 2006; and Neuffer et al., 2023, see Supplementary Table S2) that we considered as paralogous sequences unlinked to the S-locus, as already documented in Schierup et al. (2001) and Prigoda et al. (2005, see Supplementary Information).

Second, we validated the use of transcriptomic data to determine S-locus genotypes using the NGSgenotyp pipeline. We applied the approach described above to genotype the S-locus of four *C. grandiflora*, four *C. orientalis*, and 16 *C. bursa-pastoris* individuals, using published genome resequencing data as well as RNA-seq data obtained separately from leaf, root, or flower bud tissues from the exact same individuals (Kryvokhyzha et al., 2019). Strictly identical S-locus genotypes were inferred based on SRK sequences detected from genomic DNA and RNA-seq data from flower bud tissues (Supplementary Table S3), as expected by the codominant expression of SRK in pistils reported in Brassicaceae (Burghgraeve et al., 2020; Hatakeyama et al., 2001). These results validate the use of RNA-seq data from flower buds to reliably genotype the S-locus in *Capsella*. We note that all four individuals of *C. grandiflora* were heterozygous at the S-locus, and all *C. orientalis* individuals were homozygous for the allele called H4004n (we use the notation “n” to indicate that this allele is non-functional), in agreement with Bachmann et al. (2019, 2021; note that these authors refer to this allele as CoS12). In agreement with Bachmann et al. (2021), most allotetraploid *C. bursa-pastoris* individuals had two copies of the H4004n allele derived from the non-functional *C. orientalis* parental allele and all individuals had two copies of the non-functional H4047n allele (with an SRK sequence interrupted at position 949, see above), derived from the *C. grandiflora* parental allele H4047 (Supplementary Table S3). However, we found that a second allele, H2002, already known in *C. grandiflora* (Supplementary Fig. S2), segregates at the S-locus in the *C. orientalis* subgenome (present in two copies in accession DUB-RUS9 and in one copy, together with H4004n, in

LAB-RUS-4). As expected, we generally observed no or very low expression of SRK in leaves or roots (Supplementary Table S3).

Third, we evaluated whether S-locus phenotypes in pollen could be assessed based on RNA-seq data. We obtained RNA-seq data from flower buds for seven diploid *C. grandiflora* individuals used as parents in the production of synthetic polyploids and of hybrids with *C. orientalis* (see below), as well as for six synthetic autotetraploids (respectively, Cg2 and Cg4 in Supplementary Fig. S1). We obtained the SRK genotypes of these individuals using the NGSgenotyp pipeline, as described above. Five *C. grandiflora* S-alleles were found to segregate in this experimental material (Figure 1A and B; Supplementary Fig. S2), again with approximately balanced transcript levels between both SRK alleles in heterozygotes (Supplementary Tables S4 and S6). Then, we obtained full SCR transcript sequences for each of the five S-alleles (available at <https://www.doi.org/10.6084/m9.figshare.22567558.v2>) by applying the de novo assembly module of the NGSgenotyp pipeline, based on a reference database of known SCR sequences from *A. halleri* and *A. lyrata* (see Supplementary Material). We used these new reference sequences to compare patterns of allele-specific expression for SRK and SCR in the 13 *C. grandiflora* individuals (Supplementary Table S4). In agreement with the results of Burghgraeve et al. (2020) in *A. halleri*, allele-specific expression was much more asymmetric between SCR alleles than between SRK alleles. Indeed, in heterozygous individuals, one of the two alleles contributed over 99% of the total SCR transcript levels, with only three exceptions (individuals Cg2-12-3, Cg4-1-3, and Cg4-7-3; Supplementary Table S4). This very strong allelic imbalance was also found in tetraploid individuals, suggesting that a dominant allele is capable of repressing several co-occurring alleles at once. The identity of the predominantly expressed allele was fully concordant with expectations based on the predicted classes of dominance between S-alleles (Figure 1A; Supplementary Table S4). Even in cases where two S-alleles of the same dominance class co-occurred (e.g., H2008 and H2022 in individual Cg4-6-4, or H4015 and H4035 in Cg4-1-3), the asymmetry of the transcript levels remained very strong, hence enabling us to determine the putative dominance hierarchy among the five alleles, as follows: H4035 > H4015 > H2022 > H2008 > H1001. We also analyzed one diploid and one tetraploid *C. orientalis* individual (Supplementary Table S4) and observed a single S-allele in pistil and pollen, H4004n, whose SCR sequence was fully identical to that reported by Bachmann et al. (2019). These results also confirm that both SRK and SCR are still expressed in *C. orientalis*, as reported by Bachmann et al. (2019), even though they were shown to be non-functional based on crosses with *C. grandiflora* individuals carrying the H4004 allele.

The S-locus genotypes and phenotypes of diploid and synthetic tetraploid hybrids

Diploid and tetraploid hybrids were produced between *C. grandiflora* and *C. orientalis* (Supplementary Fig. S1; Duan et al., 2023), using seeds from two wild individuals of *C. grandiflora* and one inbred line of *C. orientalis*. Diploid hybrids were generated by crossing *C. orientalis* with *C. grandiflora*, while allotetraploids were created either by inducing genome doubling in diploid hybrids with colchicine treatment (“Sh” allotetraploids), or by crossing colchicine-induced autotetraploid *C. orientalis* with autotetraploid *C. grandiflora* (“Sd” allotetraploids). In all crosses, diploid or tetraploid *C. orientalis* was used as the maternal plant, mimicking the formation of natural *C. bursa-pastoris* (Hurka et al., 2012).

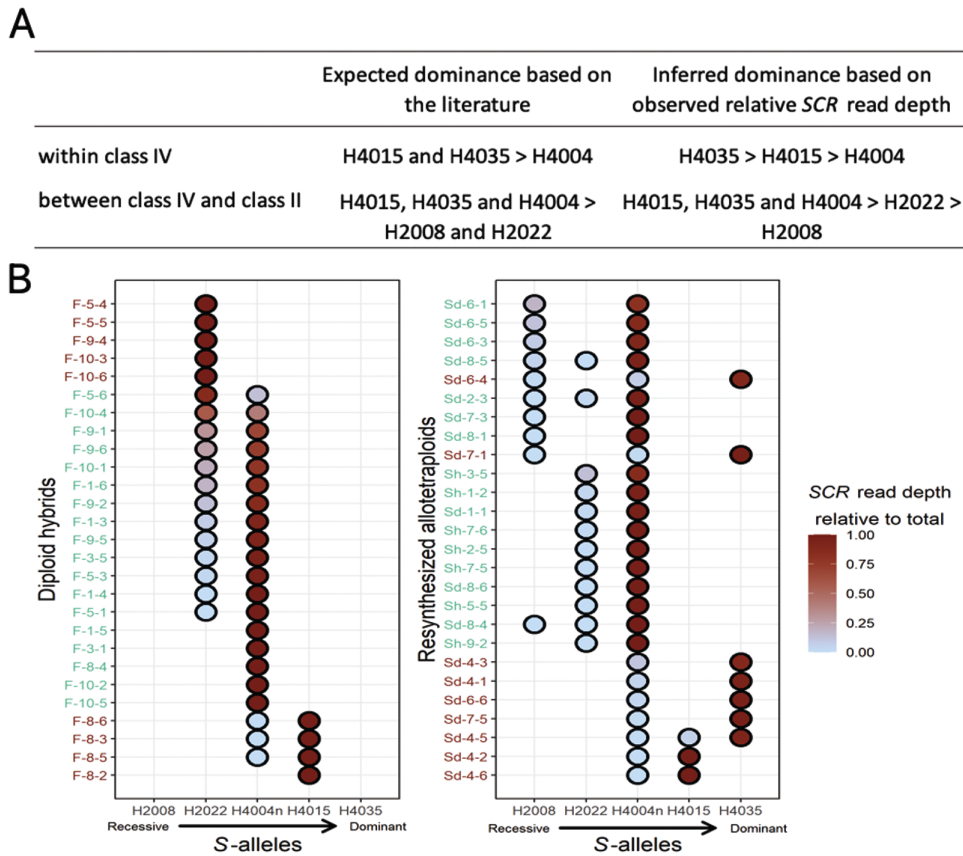


Figure 1. Predicted and observed dominance relationships between *C. grandiflora* S-alleles and the *C. orientalis* H4004n allele in the pollen and prediction of self-incompatibility phenotypes of hybrid individuals. A. Predicted dominance relationships based on SRK genotypes and previous studies in *Arabidopsis halleri* (Burghraeve et al., 2020; Durand et al., 2014; Llaurens et al., 2008; Yew et al., 2023) and *A. lyrata* (Prigoda et al., 2005), and inferred dominance based on observed relative SCR read depth from RNAseq data (Supplementary Tables S4 and S5). B. Observed relative SCR read depth of H4004n relative to other alleles in diploid (F) and tetraploid (Sd, Sh) individuals, and predicted self-incompatibility phenotype: green individual label, self-compatible (SC) because H4004n is dominant over *C. grandiflora* allele(s); brown individual label, self-incompatible (SI) because H4004n is recessive to *C. grandiflora* allele(s).

Then we analyzed RNA-seq data from flower buds of 27 diploid hybrids (F) and 26 tetraploid hybrids (Sh and Sd). By mapping RNA-seq raw reads on SRK reference sequences we found that five different S-alleles were segregating among these 53 individuals (Figure 1A and B; Supplementary Table S5): the non-functional *C. orientalis* H4004n allele along with four *C. grandiflora* alleles (all alleles described above except H1001). We could determine full genotypes for all diploid hybrids, whereas for tetraploids we could determine only the identity of the S-alleles present in each individual, but not their relative copy numbers, leaving some uncertainties in the exact genotypes (Supplementary Table S5). We checked relative patterns of expression of SCR in order to test the hypothesis of Bachmann et al. (2021) that H4004n may have retained the ability to transcriptionally repress S-alleles of a lower dominance class. In agreement with this hypothesis, we found that in 11 of the 13 diploid hybrids and in all 12 tetraploid hybrids possessing both alleles H4004n and H2022, the former was expressed majoritarily (Figure 1B; Supplementary Table S6). Also in all 8 tetraploid hybrids possessing both alleles H4004n and H2008, the former was expressed majoritarily. Hence, SCR expression of both H2022 and H2008 is repressed in the presence of H4004n when H4015 and H4035 are absent, in agreement with previous results showing dominance of class IV alleles over class II alleles (Burghraeve et al., 2020; Durand et al., 2014; Llaurens et al., 2008; Prigoda et al., 2005). In contrast, in the three diploid

and two tetraploid hybrids sharing only alleles H4004n and H4015, it was the latter that was expressed majoritarily (Figure 1B), indicating dominance of H4015 over H4004n. Similarly, dominance of H4035 over H4004n was confirmed by patterns of SCR expression of the seven tetraploid hybrids sharing these two S-alleles. These results are in agreement with previous results showing recessivity of H4004 with respect to all other class IV alleles tested to date (Durand et al., 2014; Llaurens et al., 2008; Yew et al., 2023). We then used two different approaches to predict the SI phenotype of hybrids: (1) based on the S-locus genotype inferred from SRK data, a hybrid individual was predicted to be SC if it carried the *C. orientalis* H4004n allele and none of the S-alleles derived from *C. grandiflora* that are predicted to be more dominant than H4004n (i.e., H4035 and H4015), and to be SI otherwise (i.e., carrying H2008 and/or H2022) (Figure 1A); (2) based on the SCR relative expression data, a hybrid individual was predicted to be SC if the relative SCR read depth of allele H4004n was higher than 0.5, and to be SI otherwise (Figure 1B). As shown in Supplementary Fig. S3, predictions of the SI phenotypes based on the two methods were highly congruent. Overall, 18 of the 27 homoploid hybrids (F individuals) and 17 of the 26 tetraploid hybrids (Sd and Sh individuals) were predicted to be functionally self-compatible based on approach (1), while the remaining individuals were predicted to be self-incompatible (Figure 1B).

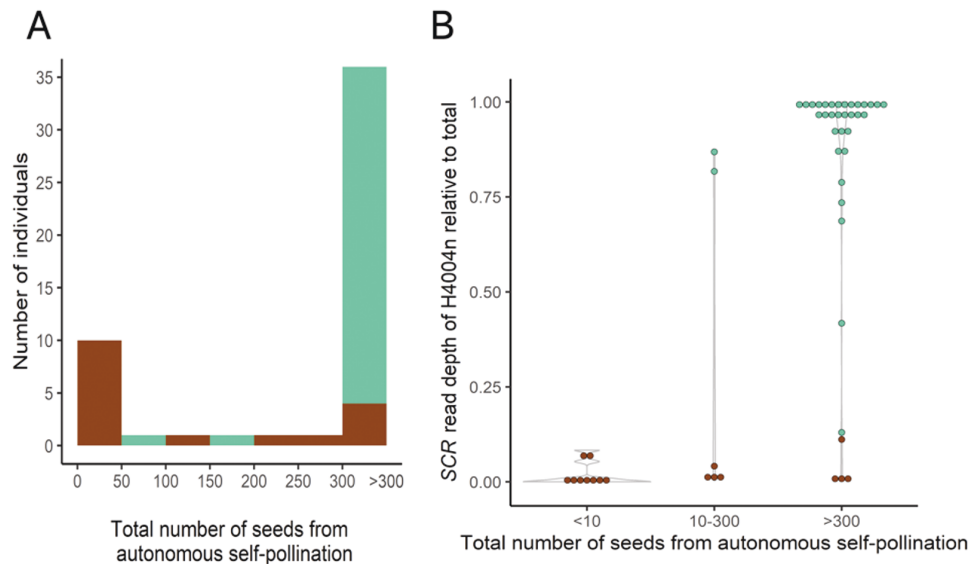


Figure 2. Seed production under autonomous self-pollination in the diploid and tetraploid *C. orientalis* × *C. grandiflora* hybrids in relation to expected self-incompatibility phenotype (SRK-based prediction) and relative expression of the non-functional H4004n SCR allele. (A) Distribution of seed production for expected self-compatible (SC, green) and self-incompatible (SI, brown) phenotypes. (B) Relationships of the SRK-predicted dominance of the H4004n SCR allele (green, SC; brown, SI), the observed relative expression level of the H4004n SCR allele (green, SC; brown, SI), the observed relative expression level of the H4004n SCR allele and seed production.

Inferred S-locus phenotypes predict the ability to self-fertilize in hybrids

We then compared the autonomous seed set of diploid and tetraploid hybrids to test whether the S-locus genotypes and the observed transcriptional dominance of SCR alleles can explain which *C. orientalis* × *C. grandiflora* hybrids are SC and which are SI. Seed production under autonomous selfing was used as an indicator of self-compatibility and showed a clear bimodal distribution with most individuals either in the 0–50 seeds or in the > 300 seeds categories (Figure 2A).

We found that both the predicted dominance of the H4004n allele and the relative expression level of its SCR allele are strong predictors of the ability of autonomous seed production, with only a few exceptions (Figure 2A and B). The SRK-based prediction of self-compatibility was strongly associated with seed production categories (Fisher's exact test, p -value < .001), with most individuals predicted to be self-incompatible producing no or few seeds under autonomous pollination (<300 seeds). In contrast, individuals who were predicted to be self-compatible usually produced more than 300 seeds. Similarly, the relative expression level of the H4004n SCR allele significantly differed among the three seed production categories (Kruskal–Wallis H test, p -value < 0.001). Individuals with a higher proportion of the H4004n SCR allele expression (e.g., larger than 0.25) usually had autonomous seed production above 300, and individuals with a lower proportion of H4004n expression usually had no or few seeds.

Discussion

Assessing dominance relationships between S-alleles using RNA-seq data

At the S-locus in Brassicaceae, the genotype-to-phenotype map is complicated by the widespread existence of dominance/recessivity interactions between S-alleles. Determination of these dominance relationships ultimately relies on phenotypic assays based on controlled pollinations. Following Shiba et al. (2002), Burghgraeve et al. (2020) recently demonstrated that phenotypic

dominance in pollen can be predicted with high accuracy from the simple comparison of transcript abundances, using quantitative RT-PCR of SCR transcripts. However, allele-specific qPCR primers need to be designed and optimized for every single S-haplotype whose expression is to be quantified, which is not practical when large numbers of S-haplotypes segregate. Here, we show that RNA-seq data from flower buds can be used to reliably infer dominance relationships between S-alleles in pollen. These data are relatively simpler to obtain, as they do not require specific optimization steps beyond a generic RNA-seq library construction and can thus be generalized more readily than the qRT-PCR approach of Burghgraeve et al. (2020), or the labor-intensive phenotypic assessment of dominance by controlled pollination assays. A limitation of this new approach, however, is that it can only quantify transcripts of SCR alleles whose nucleotide sequence is known a priori, which is only the case for a subset of the numerous S-haplotypes typically found in SI species, including *C. grandiflora*. A potential caveat to this new approach is that it relies on comparing mapping densities of (Illumina) sequencing reads on the nucleotide sequence of SCR alleles, which have relatively short coding sequences, thus making accurate mapping a potential challenge. The high levels of nucleotide divergence among SCR alleles are expected to (at least partially) compensate for this limitation, and accordingly, we found that for the five S-alleles considered in our crossing design, cross-mapping of individual sequencing reads among alleles was negligible, making the method highly reliable.

When applying the method to diploid *C. grandiflora* individuals, we found that the putatively dominant S-haplotype represented > 99% of the global level of SCR transcripts in six out of seven individuals (Supplementary Table S4), corresponding to nearly complete dominance at the transcriptional level, in line with Burghgraeve et al. (2020). The relative dominance levels we inferred among the *Capsella* S-haplotypes we studied here were also fully concordant with the dominance interactions previously measured by controlled crosses for the trans-specifically shared S-haplotypes in *A. lyrata* and *A. halleri* (Durand et al., 2014; Laurens et al., 2008; Prigoda et al., 2005). Specifically, we confirmed

that both S-haplotypes from class IV were dominant over both S-haplotypes from class II, which were themselves dominant over the single class I S-haplotype (Figure 1A; Supplementary Table S4). Another original result from our analysis is that a single SCR allele was predominantly expressed in each tetraploid *C. grandiflora* individual, suggesting that the transcriptional silencing machinery controlling dominance remains effective in an autotetraploid context, in line with the phenotypic patterns of dominance between S-haplotypes observed in tetraploid individuals of *A. lyrata* (Mable et al., 2004). Although we could determine without ambiguity the identity of all S-alleles present in tetraploid individuals, we note that the exact number of gene copies of each allele remains uncertain because S-locus genotypes were determined using RNA-seq data instead of genomic resequencing (Genete et al., 2020). Hence, precise genotyping of tetraploid individuals based on genomic DNA will be needed to quantify the extent of this phenomenon. Our approach also demonstrated that the silencing machinery was functional in an allotetraploid context, with functional interactions occurring between genetic determinants belonging to different parental subgenomes, i.e., a dominant allele within the *C. orientalis* subgenome was capable of silencing recessive alleles from the *C. grandiflora* subgenome, and vice-versa. This is consistent with recent results from Yew et al. (2023) and Dou et al. (2023) who used a different approach based on genetic transformation in the allotetraploid *A. kamchatica* and *Brassica napus*, respectively, to show that the sRNA precursor from a dominant non-functional allele was capable of silencing a functional S-allele from the other subgenome.

Three potential caveats could have blurred the link between the dominance relationship of SCR alleles, the occurrence of self-compatibility and seed production. First, the effect of dominance among S-alleles was tested by measuring seed production under autonomous selfing rather than by directly observing the SI reaction by controlled self-pollination. Second, the newly formed interspecific hybrids are expected to have lower fitness due to interactions between divergent genomes (Fishman & Sweigart, 2018), therefore individuals with no seed or few seeds could also result from hybrid incompatibility rather than SI. Third, the individuals were not strictly separated in the growth chamber during flowering time, so for individuals that generated a small number of seeds, we cannot rule out the possibility of pollen contamination from other plants. The fact that we still observed a strong association between the relative expression level of the non-functional H4004n SCR allele and autonomous seed production, in spite of these potential limitations, provides strong evidence that the machinery controlling dominance relationships between SCR alleles is a major determinant of the self-compatibility phenotype of the hybrids we obtained.

S-haplotypes dominance mediates the effect of WGD on the breakdown of SI in *Capsella* allopolyploids

In *Capsella*, a genus with a sporophytic SI system, our results formally establish that the ability to self-fertilize immediately upon hybridization between the SC and SI parental species depends on the relative dominance of the non-functional allele inherited from the selfer *C. orientalis* as compared to that of the functional S-allele(s) inherited from the outcrosser *C. grandiflora*, in line with the model proposed by Novikova et al. (2023). This observation raises several intriguing questions. First, the non-functional S-allele needs to still retain the capacity to remain dominant (Fujimoto et al., 2006). This is the consequence of the particular genetic architecture of dominance between S-haplotypes, where dominance modifiers (small non-coding RNAs) are distinct from the gene they regulate (Billiard

& Castric, 2011). This particular genetic architecture of dominance might be less rare than it was long thought to be (Billiard et al., 2021), but remains to be investigated in other families with sporophytic SI where dominance relationships have been demonstrated such as Asteraceae (Samaha & Boyle, 1989) and Convolvulaceae (Kowayama et al., 1994). For Convolvulaceae, the occurrence of such a mechanism would explain the observation by Kakeda et al. (2000) that a non-functional S-allele in *Ipomoea trifida* was found to be dominant over a functional allele and could enforce self-compatibility in heterozygotes. Second, the variation we observed relies on the fact that the non-functional S-haplotype that was fixed in *C. orientalis* has an intermediate level of dominance. If it had been the most recessive, then all allotetraploid individuals would by definition have inherited a more dominant S-haplotype from *C. grandiflora*, and would thus have remained SI. In contrast, if *C. orientalis* had fixed the most dominant S-haplotype, then all hybrids would have turned SC. Similar cases of SC parental species involved in allopolyploidy events that had previously fixed dominant non-functional S-haplotypes, e.g., *A. thaliana* and SC populations of *A. lyrata*, have been reviewed by Novikova et al. (2023). If there is a general trend that S-haplotypes at high levels of dominance are more likely to be fixed in SC taxa, then the dominance interaction itself can contribute to the association between allopolyploids and self-compatibility. Some factors affecting the fixation probability of SC mutations have been studied by Tsuchimatsu and Shimizu (2013), but to the best of our knowledge, the effect of dominance on the fixation probability of SC mutations in a sporophytic SI system remains to be investigated formally. A third interesting question is why most known examples of recent allotetraploids in Brassicaceae involve hybridization between a selfer and an outcrosser (Novikova et al., 2023). A more general survey of recent allopolyploids would be needed to determine the generality of this pattern, but one tempting hypothesis is that such scenario would provide for a genetic mechanism introducing instantaneous self-compatibility of the allopolyploid individuals, hence facilitating the establishment of the neopolyploid populations (Novikova et al., 2023). Determining whether differences in the intensity of genetic conflicts between the outcrosser and the selfer genomes (in particular over development of the endosperm, Rebernik et al., 2015) can oppose this selective advantage would be an interesting next step. Finally, a parallel can be drawn with the more general process of Haldane's sieve, in which advantageous alleles tend to be fixed more readily in natural populations when they are dominant because they are directly exposed to natural selection (Haldane, 1924). Here, in contrast, the selective advantage would go to hybrid lineages that have inherited from their SI parent an S-haplotype more recessive than the S-allele that was fixed in the selfing lineage.

A similar mechanism was proposed by Bachmann et al. (2021) for the evolution of selfing in the natural allotetraploid *C. bursa-pastoris* from *C. orientalis* and *C. grandiflora* parents. Intriguingly, while *C. bursa-pastoris* is a strong selfer, the S-haplotype it inherited from *C. grandiflora* (H4047) belongs to class IV, and thus would a priori be expected to be at least as dominant as the S-haplotype it inherited from *C. orientalis* (H4004n). Also, we identified one *C. bursa-pastoris* individual lacking the H4004n allele but carrying a (more recessive) class II allele (H2002, Supplementary Table S3), presumably inherited from *C. orientalis*. Hence, other mechanisms might be needed to explain the loss of SI in the early development of the *C. bursa-pastoris* lineage. It should be noted, however, that dominance interactions between the class IV S-alleles have been firmly established at the phenotypic level for a small number of S-alleles only, so the possibility remains that some class IV S-alleles (in this particular case, H4047) could actually be more

recessive than H4004n. This is suggested by the observation of a putative target of the small non-coding RNA produced by the *C. orientalis* S-haplotype in close proximity to the H4047n SCR pseudogene within the A subgenome of *C. bursa-pastoris* (Bachmann et al., 2021). As we found that both alleles (H4004 and H4047) are segregating in the Cg-9 population, it would be interesting to obtain living material carrying those alleles and perform controlled crosses to establish their relative dominance.

Predictions on the effect of WGD on mating system evolution need to take into account the genetic determination of mating systems, the mating systems of parental species, and the type of polyploidy

Associations between WGD and an autogamous mating system have been largely debated in plant biology, but a general consensus is still lacking (Barringer, 2007; Husband et al., 2008; Mable, 2004; Robertson et al., 2011). Part of the uncertainty stems from the fact that broad-scale studies focused on taxa comprising a mixture of different SI systems (e.g., self-recognition-based sporophytic SI, self-recognition or non-self-recognition-based gametophytic SI), and different types of polyploidy (autopolyploidy, allopolyploidy, or a combination of both, i.e., segmental allopolyploidy). These SI systems and types of polyploidy strikingly differ with respect to the mechanistic effect of WGD on maintaining a fully functional SI response in neopolyploids (Table 1) and are also expected to differ in the conditions allowing evolution of *de novo* mutations altering the mating system in neopolyploid lineages (Husband et al., 2008). Regarding the former effect, in some systems, polyploidy will immediately generate a mechanical breakdown of SI (i.e., gametophytic SI systems with non-self-pollen/pistil recognition or sporophytic SI systems with one SC parent carrying a dominant non-functional allele, as demonstrated in this work), while in other SI systems no such effect is expected (Table 1). The type of polyploidy, i.e., auto- vs allopolyploidy, will also have an impact, as autopolyploids with non-self-pollen/pistil recognition systems will systematically be SC, while SI would be maintained in other systems. A detailed meta-analysis taking these factors into account would be an interesting next step.

Material and methods

Methodological approach to type S-alleles in *Capsella* experimental material based on RNA-seq data

To build an extended dataset of reference sequences of SRK from the self-incompatible species *Capsella grandiflora*, we genotyped

180 individuals of the Cg-9 population of *C. grandiflora* from Monodendri, Greece (Josephs et al., 2015) at the SRK gene with the NGSgenotyp pipeline (Genete et al., 2020) using raw Illumina reads available from Sequence Read Archive (SRA, Supplementary Table S1). For the SRK reference database, we used available sequences of SRK from *A. lyrata* and *A. halleri* (Genete et al., 2020; Takou et al., 2021), and 62 partial sequences from *Capsella grandiflora* (Guo et al., 2009; Neuffer et al., 2023; see Supplementary Table S2). Briefly, this pipeline filters raw reads with a dictionary of k-mers extracted from the reference database, then uses Bowtie2 to align filtered reads against each reference sequence from the database and produces summary statistics with Samtools (v1.4; Danecek et al., 2021) allowing it to identify S-alleles present in each individual. The pipeline NGSgenotyp also contains a *de novo* assembly approach module which produces full sequences of the S-domain of SRK for alleles present as partial sequences in the database as well as for newly identified S-alleles.

We then compared the results of the S-alleles genotyping approach obtained from either genomic DNA or RNA-seq data from flower buds, leaf, and root tissues. For this, we used published data from Kryvokhyzha et al. (2019) on four individuals each of *C. grandiflora* and *C. orientalis*, and on 16 individuals of *C. bursa-pastoris*. We applied the NGSgenotyp pipeline separately on each dataset, using the SRK reference database expanded with the *Capsella* S-allele sequences obtained above. Our analysis showed that S-allele typing based on RNA-seq data from flower buds gave identical results than those obtained from genomic DNA, so in the rest of the analyses we only used flower buds RNA-seq data.

Creating and sequencing the transcriptome of synthetic hybrids and polyploids and assessing their mating system

To test the dominance relationship among SI alleles and its phenotypic consequences we used diploid and tetraploid hybrids of *C. orientalis* × *C. grandiflora*, using the synthetic hybrids generated by Duan et al. (2023; Supplementary Fig. S1). We measured fruit set under autonomous selfing to test whether self-compatibility of these hybrids can be predicted by the dominance of SCR alleles, by comparing the SCR alleles identified in transcriptomes of inflorescences with fruit-set from spontaneous self-pollination in 27 diploid and 26 tetraploid hybrids.

In short, the diploid and tetraploid hybrids were generated from one inbred line of *C. orientalis* (URAL-RUSS), and seeds that were collected from two wild *C. grandiflora* individuals of the same population (85.1 and 85.24). Specifically, all the synthetic hybrids were descendants of three *C. grandiflora* individuals (85.1-5, 85.24-1, and 85.24-5), two of which (85.24-1 and 85.24-5) had

Table 1. Expected instantaneous effect of polyploidy on the self-compatibility phenotype of neopolyploids depending on the type of SI system of the parental species (SI, self-incompatible; SC, self-compatible) and on the type of polyploidy (autopolyploidy vs allopolyploidy).

Type of SI system	Type of polyploidy		
	Autopolyploidy	Allopolyploidy (SI × SI parental species)	Allopolyploidy (SC × SI parental species)
Gametophytic SI with non-self-recognition	SC ¹	SC ¹	Should depend on the type of SC mutation
Gametophytic SI with self-recognition	SI ²	SI ³	Should depend on the type of SC mutation
Sporophytic SI	SI ⁴	SI	Depends on relative dominance level of SI and SC alleles ⁵

¹Automatic breakdown of SI in diploid heteroallelic pollen carrying two different S-alleles (Entani et al., 1999; Kubo et al., 2010; Luu et al., 2001; Tsukamoto et al., 2005).

²Hauck et al., 2006.

³Vieira et al., 2008.

⁴Mable et al., 2004.

⁵This study.

the same maternal plant. Diploid hybrids (F) were obtained by crossing *C. orientalis* with *C. grandiflora*. Tetraploid hybrids (allotetraploids) were generated in two ways: in the first case the two diploid species were first crossed, and WGD was induced on the first generation of diploid hybrids with colchicine solution, resulting in “hybridization-first” synthetic allotetraploids (Sh); in the second case, WGD was induced in both diploid species, then the synthetic autotetraploids were crossed to obtain “WGD-first” allotetraploids (Sd). In addition, several diploid hybrids were suspected to have spontaneous WGD without colchicine treatment based on observations of organ size and the shape of trichomes, including individual F-3-5 which was used in the present study. In all interspecific crosses, diploid or tetraploid *C. orientalis* served as the maternal plant. The second generation of diploid hybrids, Sh-allotetraploids and Sd-allotetraploids as well as the diploid and tetraploid parental species were then grown together in a growth chamber. Each of the three hybrid groups was represented by six lines (independent hybridization events), and each line was represented by six individuals. The six individuals of the same line were full siblings from self-fertilization. An overview of the mating scheme used to create the different resynthesized hybrids and polyploids is given in [Supplementary Fig. S1](#).

The ability to generate seeds with only autonomous selfing was recorded for all hybrid individuals as a categorical factor. The hybrid individuals were classified into three categories based on the total number of seeds: (1) having almost no seeds (<10 seeds), (2) having few seeds (10–300 seeds), and (3) having plenty of seeds (>300 seeds). As the hybrid individuals were not strictly separated in the growth chamber during flowering, the cutoff of 10 seeds was applied to reduce noise from occasional pollen contamination from other *Capsella* plants. The seed set data of two individuals were removed from the dataset because they were severely affected by disease during flowering time.

The first group of RNA-seq data was from [Duan et al. \(2023\)](#). Total RNA was extracted from the inflorescence of 6 diploid hybrids and 14 allotetraploids, using a cetyl-trimethylammonium-bromide-based method. Sequencing libraries were prepared with Illumina TruSeq Stranded mRNA (poly-A selection) kit, and sequenced on three NovaSeq 6000 S4 lanes with 150-bp paired-end reads (SNP&SEQ Technology Platform in Uppsala). One sequencing library was prepared and sequenced for each diploid sample, and two libraries were prepared for each tetraploid sample. On average 38.6 and 77.3 million read pairs were generated for the diploid and tetraploid samples, respectively.

To obtain a larger sample size, we performed a second group of RNA-seq on inflorescences of 33 additional hybrid individuals, including 20 diploid hybrids and 13 allotetraploids. Inflorescence samples of this second group were from the same experiment as those from the first group, and were collected at the same time, and stored at -80°C before sequencing. Total RNA was extracted using an RNeasy Plant Mini Kit (Qiagen). The library preparation and sequencing platform were the same as the first group, except that one library was prepared for each individual, regardless of ploidy level. Sequencing of the second group of inflorescence samples yielded an average library size of 97.4 million RNA-seq reads.

Determination of S-locus genotype and phenotype in the *Capsella* hybrids and synthetic polyploids and confrontation with self-compatibility phenotype assessments

In Brassicaceae, the self-incompatibility phenotype in pollen depends on complex dominance relationships among S-alleles,

achieved through modifier genetic elements consisting of small RNAs encoded by precursors lying at the S-locus of dominant alleles and targeting the SCR gene of recessive alleles ([Durand et al., 2014](#); [Tarutani et al., 2010](#)). Dominance in pollen is thus regulated at the transcriptional level and is associated with very strong inhibition of mRNA production of recessive alleles ([Burghgraeve et al., 2020](#)), which could potentially be revealed by analyzing RNA-seq data. Hence, we tested this approach by performing S-allele typing with the NGSgenotyp pipeline using separately a SRK reference database, to determine the S-locus genotype of individuals (because SRK alleles are always co-expressed in the style, [Hatakeyama et al., 2001](#)), and an SCR reference database, to determine which S-allele is majoritarily expressed in pollen (and thus putatively dominant). We applied this approach to RNA-seq data from eight diploid individuals (seven *C. grandiflora* + one *C. orientalis*) and seven tetraploids (six *C. grandiflora* + one *C. orientalis*) used as parents in the hybrid experiments ([Duan et al., 2023](#)), with the same SRK reference database as above enlarged with newly obtained *C. grandiflora* allele sequences, and for SCR with a reference database of sequences from *A. halleri* and *A. lyrata* (Genbank sequences). For SCR, because of the higher sequence divergence among S-alleles than for SRK, we modified the NGSgenotyp parameters by reducing the k-mer size used for filtering to a value of 15 (the default size used for SRK was 20). This allowed us, with the de novo assembly module of NGSgenotyp, to obtain full coding sequences of SCR for all *C. grandiflora* alleles present in the hybrids. In order to quantify the relative expression of SCR alleles, we used the genotyp module from NGSgenotyp to map individual RNA-seq reads data against each reference SCR exon 2 sequence (Bowtie2 v2.4.4; [Langmead & Salzberg, 2012](#)). As the SCR sequences from the database are small, the alignment mode was set to “local” to allow partial mapping of the reads (soft clipping) in a way that optimizes the alignment score. Then we used the mean read depth delivered by Samtools (v1.14; [Danecek et al., 2021](#)) to compute the ratio of the mean read depth of the predominantly expressed allele (i.e., the putative dominant allele) to the sum of the read depths of all alleles present. We applied the same approach in synthetic tetraploid *C. grandiflora* individuals, but for SRK data we could only report the number and identity of alleles present, and thus it was not possible to precisely genotype individuals, i.e., to determine the number of gene copies of any given allele identified (when the total number of alleles detected in an individual was lower than 4, which was the case for all tetraploid individuals).

Once the proposed approach was validated on *C. grandiflora* diploid and tetraploid individuals, we applied it to the experimental diploid and tetraploid hybrids. We identified two categories of hybrid individuals, in terms of pollen SI phenotype, depending on relative dominance levels of the inherited *C. grandiflora* and *C. orientalis* parental alleles: individuals with predominant expression of the *C. orientalis* allele, which is a non-functional S-allele; and individuals with predominant expression of one of the *C. grandiflora* alleles. The dominance of the non-functional *C. orientalis* allele is expected to cause a breakdown of self-incompatibility because it will impede recognition and rejection of self-pollen, and thus we assigned an expected self-compatible phenotype to those individuals, and an expected self-incompatible phenotype to individuals with a dominant SCR allele from *C. grandiflora*. Then we compared seed production data under autonomous selfing with the prediction of self-compatible/self-incompatible phenotype based on SCR dominance. The association between SRK-predicted self-compatibility and seed production categories was tested with Fisher’s exact test in R software environment version

3.6.3 (R Core Team, 2020). The relative expression level of the H4004n SCR allele (mean read depth of H4004n/sum of the mean read depth of all other S-alleles of the individual) in the three seed production categories were compared by the Kruskal–Wallis H test.

Supplementary material

Supplementary material is available online at *Evolution Letters*.

Data and code availability

The RNA-sequencing data of the additional *Capsella* hybrids generated by this article are available in the Sequence Read Archive (SRA) of the National Center for Biotechnology Information (NCBI), and can be accessed with BioProject number PRJNA946929. All new *Capsella* SRK and SCR sequences obtained by de novo assembly with the NGSgenotyp pipeline are posted at <https://www.doi.org/10.6084/m9.figshare.22567558.v2>.

Author contributions

Conception and coordination of the study: T.D., M.L., V.C., and X.V. Creation of experimental material: T.D. and A.S. RNAseq data: T.D. and Z.Z. Bioinformatics: T.D. and M.G. Data analysis: T.D., X.V., V.C., C.P., and M.G. Writing of the manuscript: T.D., X.V., V.C., and M.L. All authors reviewed the manuscript.

Funding

The work on SI in the Lille group is supported by the Agence Nationale de la Recherche (TE-MoMa project, grant number ANR-18-CE02-0020-01), the Région Hauts-de-France and the Ministère de l'Enseignement Supérieur et de la Recherche (CPER Climibio and CPER Ecrin grants), and the European Fund for Regional Economic Development. The study in Uppsala was supported by grant 2019-00806 from the Swedish Research Council to M.L. and Nilsson-Ehle research grant from the Royal Physiographic Society in Lund to T.D. M.L. also acknowledges support from the Erik Philip-Sörensen Foundation. T.D. was supported by the Sven and Lilly Lawski Foundation. The computation and data handling were provided by the Swedish National Infrastructure for Computing (SNIC) at Uppmax, partially funded by the Swedish Research Council through grant agreement no 2018-05973.

Conflict of interest: The authors declare no conflict of interest.

Acknowledgments

We thank Elsa Sundkvist for her help with seed counting, Jesper Besgaard for the early sharing of unpublished SRK sequences from *C. grandiflora*, and Tyler Kent and Stephen Wright for a preliminary analysis of S-alleles diversity in the Cg-9 population. Finally, we thank Barbara Mable and two anonymous reviewers for their critical comments on the manuscript.

References

Bachmann, J. A., Tedder, A., Fracassetti, M., Steige, K. A., Lafon-Placette, C., Köhler, C., & Slotte, T. (2021). On the origin of the widespread self-compatible allotetraploid *Capsella bursa-pastoris* (Brassicaceae). *Heredity*, 127(1), 124–134. <https://doi.org/10.1038/s41437-021-00434-9>

Bachmann, J. A., Tedder, A., Laenen, B., Fracassetti, M., Désamoré, A., Lafon-Placette, C., Steige, K. A., Callot, C., Marande, W., Neuffer, B., Bergès, H., Köhler, C., Castric, V., & Slotte, T. (2019). Genetic basis and timing of a major mating system shift in *Capsella*. *The New Phytologist*, 224(1), 505–517. <https://doi.org/10.1111/nph.16035>

Baniaga, A. E., Marx, H. E., Arrigo, N., & Barker, M. S. (2020). Polyploid plants have faster rates of multivariate niche differentiation than their diploid relatives. *Ecology Letters*, 23(1), 68–78. <https://doi.org/10.1111/ele.13402>

Barringer, B. C. (2007). Polyploidy and self-fertilization in flowering plants. *American Journal of Botany*, 94(9), 1527–1533. <https://doi.org/10.3732/ajb.94.9.1527>

Billiard, S., & Castric, V. (2011). Evidence for Fisher's dominance theory: How many "special cases?". *Trends in Genetics*, 27(11), 441–445. <https://doi.org/10.1016/j.tig.2011.06.005>

Billiard, S., Castric, V., & Llaurens, V. (2021). The integrative biology of genetic dominance. *Biological Reviews of the Cambridge Philosophical Society*, 96(6), 2925–2942. <https://doi.org/10.1111/brv.12786>

Brochmann, C., Brysting, A. K., Alsos, I. G., Borgen, L., Grundt, H. H., Scheen, A. C., & Elven, R. (2004). Polyploidy in arctic plants. *Biological Journal of the Linnean Society*, 82(4), 521–536. <https://doi.org/10.1111/j.1095-8312.2004.00337.x>

Burghgraeve, N., Simon, S., Barral, S., Fobis-Loisy, I., Holl, A. C., Ponitzki, C., Schmitt, E., Vekemans, X., & Castric, V. (2020). Base-pairing requirements for small RNA-mediated gene silencing of recessive self-incompatibility alleles in *Arabidopsis halleri*. *Genetics*, 215(3), 653–664. <https://doi.org/10.1534/genetics.120.303351>

Castric, V., & Vekemans, X. (2004). Plant self-incompatibility in natural populations: A critical assessment of recent theoretical and empirical advances. *Molecular Ecology*, 13(10), 2873–2889. <https://doi.org/10.1111/j.1365-294X.2004.02267.x>

Danecek, P., Bonfield, J. K., Liddle, J., Marshall, J., Ohan, V., Pollard, M. O., Whitwham, A., Keane, T., McCarthy, S. A., Davies, R. M., & Li, H. (2021). Twelve years of SAMtools and BCFtools. *GigaScience*, 10(2), giab008. <https://doi.org/10.1093/gigascience/giab008>

Dou, S., Zhang, T., Wang, L., Yang, C., Quan, C., Liang, X., Ma, C., & Dai, C. (2023). The self-compatibility is acquired after polyploidization: A case study of *Brassica napus* self-incompatible trilinear hybrid breeding system. *New Phytologist*, 241(4), 1690–1707. <https://doi.org/10.1111/nph.19451>

Douglas, G. M., Gos, G., Steige, K. A., Salcedo, A., Holm, K., Josephs, E. B., Arunkumar, R., Ågren, J. A., Hazzouri, K. M., Wang, W., Platts, A. E., Williamson, R. J., Neuffer, B., Lascoux, M., Slotte, T., & Wright, S. I. (2015). Hybrid origins and the earliest stages of diploidization in the highly successful recent polyploid *Capsella bursa-pastoris*. *Proceedings of the National Academy of Sciences of the United States of America*, 112(9), 2806–2811. <https://doi.org/10.1073/pnas.1412277112>

Duan, T., Sicard, A., Glémin, S., & Lascoux, M. (2023). Expression pattern of resynthesized allotetraploid *Capsella* is determined by hybridization, not whole-genome duplication. *The New Phytologist*, 237(1), 339–353. <https://doi.org/10.1111/nph.18542>

Durand, E., Méheust, R., Soucaze, M., Goubet, P. M., Gallina, S., Poux, C., Fobis-Loisy, I., Guillon, E., Gaude, T., Sarazin, A., Figeac, M., Prat, E., Marande, W., Bergès, H., Vekemans, X., Billiard, S., & Castric, V. (2014). Dominance hierarchy arising from the evolution of a complex small RNA regulatory network. *Science*, 346(6214), 1200–1205. <https://doi.org/10.1126/science.1259442>

Entani, T., Takayama, S., Iwano, M., Shiba, H., Che, F. -S., & Isogai, A. (1999). Relationship between polyploidy and pollen self-incompatibility phenotype in *Petunia hybrida* Vilm. *Bioscience*,

- Biotechnology, and Biochemistry, 63(11), 1882–1888. <https://doi.org/10.1271/bbb.63.1882>
- Fisher, R. A. (1941). Average excess and average effect of a gene substitution. *Annals of Eugenics*, 11(1), 53–63. <https://doi.org/10.1111/j.1469-1809.1941.tb02272.x>
- Fishman, L., & Sweigart, A. L. (2018). When two rights make a wrong: The evolutionary genetics of plant hybrid incompatibilities. *Annual Review of Plant Biology*, 69, 707–731. <https://doi.org/10.1146/annurev-arplant-042817-040113>
- Fowler, N. L., & Levin, D. A. (2016). Critical factors in the establishment of allopolyploids. *American Journal of Botany*, 103(7), 1236–1251. <https://doi.org/10.3732/ajb.1500407>
- Fujimoto, R., Sugimura, T., Fukai, E., & Nishio, T. (2006). Suppression of gene expression of a recessive SP11/SCR allele by an untranscribed SP11/SCR allele in Brassica self-incompatibility. *Plant Molecular Biology*, 61(4-5), 577–587. <https://doi.org/10.1007/s11103-006-0032-9>
- Genete, M., Castric, V., & Vekemans, X. (2020). Genotyping and de novo discovery of allelic variants at the Brassicaceae self-incompatibility locus from short-read sequencing data. *Molecular Biology and Evolution*, 37(4), 1193–1201. <https://doi.org/10.1093/molbev/msz258>
- Glémin, S., Bazin, E., & Charlesworth, D. (2006). Impact of mating systems on patterns of sequence polymorphism in flowering plants. *Proceedings of the Royal Society B: Biological Sciences*, 273(1604), 3011–3019. <https://doi.org/10.1098/rspb.2006.3657>
- Guo, Y. L., Bechsgaard, J. S., Slotte, T., Neuffer, B., Lascoux, M., Weigel, D., & Schierup, M. H. (2009). Recent speciation of *Capsella rubella* from *Capsella grandiflora*, associated with loss of self-incompatibility and an extreme bottleneck. *Proceedings of the National Academy of Sciences of the United States of America*, 106(13), 5246–5251. <https://doi.org/10.1073/pnas.0808012106>
- Haldane, J. B. S. (1924). A mathematical theory of natural and artificial selection, Part I. *Transactions of the Cambridge Philosophical Society*, 23, 19–41.
- Hatakeyama, K., Takasaki, T., Suzuki, G., Nishio, T., Watanabe, M., Isogai, A., & Hinata, K. (2001). The S receptor kinase gene determines dominance relationships in stigma expression of self-incompatibility in Brassica. *The Plant Journal: For Cell and Molecular Biology*, 26(1), 69–76. <https://doi.org/10.1046/j.1365-313x.2001.01009.x>
- Hauck, N. R., Yamane, H., Tao, R., & Iezzoni, A. F. (2006). Accumulation of non-functional S-haplotypes results in the breakdown of gametophytic self-incompatibility in tetraploid *Prunus*. *Genetics*, 172(2), 1191–1198. <https://doi.org/10.1534/genetics.105.049395>
- Hurka, H., Friesen, N., German, D. A., Franzke, A., & Neuffer, B. (2012). ‘Missing link’ species *Capsella orientalis* and *Capsella thracica* elucidate evolution of model plant genus *Capsella* (Brassicaceae). *Molecular Ecology*, 21(5), 1223–1238. <https://doi.org/10.1111/j.1365-294X.2012.05460.x>
- Husband, B. C. (2000). Constraints on polyploid evolution: A test of the minority cytotype exclusion principle. *Proceedings of the Royal Society B: Biological Sciences*, 267(1440), 217–223. <https://doi.org/10.1098/rspb.2000.0990>
- Husband, B. C., Ozimec, B., Martin, S. L., & Pollock, L. (2008). Mating consequences of polyploid evolution in flowering plants: Current trends and insights from synthetic polyploids. *International Journal of Plant Sciences*, 169(1), 195–206. <https://doi.org/10.1086/523367>
- Jain, S. (1976). The evolution of inbreeding in plants. *Annual Review of Ecology and Systematics*, 7, 469–495.
- Josephs, E. B., Lee, Y. W., Stinchcombe, J. R., & Wright, S. I. (2015). Association mapping reveals the role of purifying selection in the maintenance of genomic variation in gene expression. *Proceedings of the National Academy of Sciences of the United States of America*, 112(50), 15390–15395. <https://doi.org/10.1073/pnas.1503027112>
- Kakeda, K., Tsukada, H., & Koyama, Y. (2000). A self-compatible mutant S allele conferring a dominant negative effect on the functional S allele in *Ipomoea trifida*. *Sexual Plant Reproduction*, 13(3), 119–125. <https://doi.org/10.1007/s004970000048>
- Kakizaki, T., Takada, Y., Ito, A., Suzuki, G., Shiba, H., Takayama, S., Isogai, A., & Watanabe, M. (2003). Linear dominance relationship among four class-II S haplotypes in pollen is determined by the expression of SP11 in Brassica self-incompatibility. *Plant and Cell Physiology*, 44(1), 70–75. <https://doi.org/10.1093/pcp/pcg009>
- Kolesnikova, U. K., Scott, A. D., Van de Velde, J. D., Burns, R., Tikhomirov, N. P., Pfordt, U., Clarke, A. C., Yant, L., Seregin, A. P., Vekemans, X., Laurent, S., & Novikova, P. Y. (2023). Transition to self-compatibility associated with dominant s-allele in a diploid Siberian progenitor of allotetraploid *Arabidopsis kamchatica* Revealed by *Arabidopsis lyrata* Genomes. *Molecular Biology and Evolution*, 40(7), msad122. <https://doi.org/10.1093/molbev/msad122>
- Koyama, Y., Takashi, H., Muraoka, K., Tani, T., & Hara, K., & Shiotani, I. (1994). Number, frequency and dominance relationships of S-alleles in diploid *Ipomoea trifida*. *Heredity*, 73, 275–283. <https://doi.org/10.1038/hdy.1994.134>
- Kryvokhyzha, D., Milesi, P., Duan, T., Orsucci, M., Wright, S. I., Glémin, S., & Lascoux, M. (2019). Towards the new normal: Transcriptomic convergence and genomic legacy of the two subgenomes of an allopolyploid weed (*Capsella bursa-pastoris*). *PLoS Genetics*, 15(5), e1008131. <https://doi.org/10.1371/journal.pgen.1008131>
- Kubo, K. -I., Entani, T., Takara, A., Wang, N., Fields, A. M., Hua, Z., Toyoda, M., Kawashima, S. -I., Ando, T., Isogai, A., Kao, T. -hui, & Takayama, S. (2010). Collaborative non-self recognition system in S-RNase-based self-incompatibility. *Science*, 330(6005), 796–799. <https://doi.org/10.1126/science.1195243>
- Langmead, B., & Salzberg, S. L. (2012). Fast gapped-read alignment with Bowtie 2. *Nature Methods*, 9(4), 357–359. <https://doi.org/10.1038/nmeth.1923>
- Levin, D. A. (1975). Minority cytotype exclusion in local plant populations. *Taxon*, 24(1), 35–43. <https://doi.org/10.2307/1218997>
- Llaurens, V., Billiard, S., Leducq, J. B., Castric, V., Klein, E. K., & Vekemans, X. (2008). Does frequency-dependent selection with complex dominance interactions accurately predict allelic frequencies at the self-incompatibility locus in *Arabidopsis halleri*? *Evolution*, 62(10), 2545–2557. <https://doi.org/10.1111/j.1558-5646.2008.00469.x>
- Luu, D. T., Qin, X., Laublin, G., Yang, Q., & Morse, D., & Cappadocia, M. (2001). Rejection of S-heteroallelic pollen by a dual-specific S-RNase in *Solanum chacoense* predicts a multimeric SI pollen component. *Genetics*, 159, 329–335. <https://doi.org/10.1093/genetics/159.1.329>
- Mable, B. K. (2004). Polyploidy and self-compatibility: is there an association? *New Phytologist*, 162(3), 803–811. <https://doi.org/10.1111/j.1469-8137.2004.01055.x>
- Mable, B. K., Beland, J., & Di Berardo, C. (2004). Inheritance and dominance of self-incompatibility alleles in polyploid *Arabidopsis lyrata*. *Heredity*, 93(5), 476–486. <https://doi.org/10.1038/sj.hdy.6800526>
- Neuffer, B., Bechsgaard, J., Paetsch, M., Titel, C., Wesse, C., Bona, E., Schimpf, R., Žerdoner Čalasan, A., & Hurka, H. (2023). S-alleles and mating system in natural populations of *Capsella grandiflora* (Brassicaceae) and its congeneric relatives. *Flora*, 299, 152206. <https://doi.org/10.1016/j.flora.2022.152206>
- Novikova, P. Y., Kolesnikova, U. K., & Scott, A. D. (2023). Ancestral self-compatibility facilitates the establishment of allopolyploids

- in Brassicaceae. *Plant Reproduction*, 36(1), 125–138. <https://doi.org/10.1007/s00497-022-00451-6>
- Novikova, P. Y., Tsuchimatsu, T., Simon, S., Nizhynska, V., Voronin, V., Burns, R., Fedorenko, O. M., Holm, S., Säll, T., Prat, E., Marande, W., Castric, V., & Nordborg, M. (2017). Genome sequencing reveals the origin of the allotetraploid *Arabidopsis suecica*. *Molecular Biology and Evolution*, 34(4), 957–968. <https://doi.org/10.1093/molbev/msw299>
- Paetsch, M., Mayland-Quellhorst, S., & Neuffer, B. (2006). Evolution of the self-incompatibility system in the Brassicaceae: Identification of S-locus receptor kinase (SRK) in self-incompatible *Capsella grandiflora*. *Heredity*, 97(4), 283–290. <https://doi.org/10.1038/sj.hdy.6800854>
- Pandit, M. K., Poccock, M. J. O., & Kunin, W. E. (2011). Ploidy influences rarity and invasiveness in plants. *Journal of Ecology*, 99(5), 1108–1115. <https://doi.org/10.1111/j.1365-2745.2011.01838.x>
- Prigoda, N. L., Nassuth, A., & Mable, B. K. (2005). Phenotypic and genotypic expression of self-incompatibility haplotypes in *Arabidopsis lyrata* suggests unique origin of alleles in different dominance classes. *Molecular Biology and Evolution*, 22(7), 1609–1620. <https://doi.org/10.1093/molbev/msi153>
- R Core Team. (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing.
- Rebernik, C. A., Lafon-Placette, C., Hatorangan, M. R., Slotte, T., & Köhler, C. (2015). Non-reciprocal interspecies hybridization barriers in the *Capsella* genus are established in the endosperm. *PLoS Genetics*, 11(6), e1005295. <https://doi.org/10.1371/journal.pgen.1005295>
- Robertson, K., Goldberg, E. E., & Igić, B. (2011). Comparative evidence for the correlated evolution of polyploidy and self-compatibility in Solanaceae. *Evolution*, 65(1), 139–155. <https://doi.org/10.1111/j.1558-5646.2010.01099.x>
- Salman-Minkov, A., Sabath, N., & Mayrose, I. (2016). Whole-genome duplication as a key factor in crop domestication. *Nature Plants*, 2, 16115. <https://doi.org/10.1038/nplants.2016.115>
- Samaha, R. R., & Boyle, T. H. (1989). Self-incompatibility of *Zinnia angustifolia* HBK (Compositae): 11. Genetics. *Journal of Heredity*, 80(5), 368–372. <https://doi.org/10.1093/oxfordjournals.jhered.a110876>
- Schierup, M. H., Mable, B. K., Awadalla, P., & Charlesworth, D. (2001). Identification and characterization of a polymorphic receptor kinase gene linked to the self-incompatibility locus of *Arabidopsis lyrata*. *Genetics*, 158(1), 387–399. <https://doi.org/10.1093/genetics/158.1.387>
- Selmecki, A. M., Maruvka, Y. E., Richmond, P. A., Guillet, M., Shores, N., Sorenson, A. L., De, S., Kishony, R., Michor, F., Dowell, R., & Pellman, D. (2015). Polyploidy can drive rapid adaptation in yeast. *Nature*, 519(7543), 349–352. <https://doi.org/10.1038/nature14187>
- Shiba, H., Iwano, M., Entani, T., Ishimoto, K., Che, F.-S., Satta, Y., Ito, A., Takada, Y., Watanabe, M., Isogai, A., & Takayama, S. (2002). The dominance of alleles controlling self-incompatibility in Brassica pollen is regulated at the RNA level. *Plant Cell*, 14, 491–504.
- Shimizu-Inatsugi, R., Lihová, J., Iwanaga, H., Kudoh, H., Marhold, K., Savolainen, O., Watanabe, K., Yakubov, V. V., & Shimizu, K. (2009). The allopolyploid *Arabidopsis kamchatica* originated from multiple individuals of *Arabidopsis lyrata* and *Arabidopsis halleri*. *Molecular Ecology*, 18(19), 4024–4048. <https://doi.org/10.1111/j.1365-294X.2009.04329.x>
- Soltis, P. S., Marchant, D. B., Peer, Y. Van de, & Soltis, D. E. (2015). Polyploidy and genome evolution in plants. *Current Opinion in Genetics & Development*, 35, 119–125.
- Stebbins, G. L. (1957). Self fertilization and population variability in the higher plants. *The American Naturalist*, 91(861), 337–354. <https://doi.org/10.1086/281999>
- Takayama, S., Shimosato, H., Shiba, H., Funato, M., Che, F. S., Watanabe, M., Iwano, M., & Isogai, A. (2001). Direct ligand–receptor complex interaction controls Brassica self-incompatibility. *Nature*, 413(6855), 534–538. <https://doi.org/10.1038/35097104>
- Takou, M., Hämälä, T., Koch, E. M., Steige, K. A., Dittberner, H., Yant, L., Genete, M., Sunyaev, S., Castric, V., Vekemans, X., Savolainen, O., & de Meaux, J. (2021). Maintenance of adaptive dynamics and no detectable load in a range-edge outcrossing plant population. *Molecular Biology and Evolution*, 38(5), 1820–1836. <https://doi.org/10.1093/molbev/msaa322>
- Tarutani, Y., Shiba, H., Iwano, M., Kakizaki, T., Suzuki, G., Watanabe, M., Isogai, A., & Takayama, S. (2010). Trans-acting small RNA determines dominance relationships in Brassica self-incompatibility. *Nature*, 466(7309), 983–986. <https://doi.org/10.1038/nature09308>
- Tsuchimatsu, T., & Shimizu, K. K. (2013). Effects of pollen availability and the mutation bias on the fixation of mutations disabling the male specificity of self-incompatibility. *Journal of Evolutionary Biology*, 26(10), 2221–2232. <https://doi.org/10.1111/jeb.12219>
- Tsukamoto, T., Ando, T., Watanabe, H., Marchesi, E., & Kao, T. -H. (2005). Duplication of the S-locus F-box gene is associated with breakdown of pollen function in an S-haplotype identified in a natural population of self-incompatible *Petunia axillaris*. *Plant Molecular Biology*, 57(1), 141–153. <https://doi.org/10.1007/s11103-004-6852-6>
- Vanneste, K., Baele, G., Maere, S., & Peer, Y. V. D. (2014). Analysis of 41 plant genomes supports a wave of successful genome duplications in association with the Cretaceous–Paleogene boundary. *Genome Research*, 24, 1334–1347.
- Vieira, J., Santos, R. A., Ferreira, S. M., & Vieira, C. P. (2008). Inferences on the number and frequency of S-pollen gene (SFB) specificities in the polyploid *Prunus spinosa*. *Heredity*, 101(4), 351–358. <https://doi.org/10.1038/hdy.2008.60>
- Wright, S. I., Kalisz, S., & Slotte, T. (2013). Evolutionary consequences of self-fertilization in plants. *Proceedings of the Royal Society B: Biological Sciences*, 280(1760), 20130133. <https://doi.org/10.1098/rspb.2013.0133>
- Yew, C. L., Tsuchimatsu, T., Shimizu-Inatsugi, R., Yasuda, S., Hatakeyama, M., Kakui, H., Ohta, T., Suwabe, K., Watanabe, M., Takayama, S., & Shimizu, K. K. (2023). Dominance in self-compatibility between subgenomes of allopolyploid *Arabidopsis kamchatica* shown by transgenic restoration of self-incompatibility. *Nature Communications*, 14(1), 1–11.