



Review article

The dynamic relationship between skin microbiomes and personal care products: A comprehensive review

Mahjabin Ferdaous Mim^a, Mahmudul Hasan Sikder^b,
Md. Zahid Hasan Chowdhury^a, Ashkar-Ul-Alam Bhuiyan^a, Nayeematul Zinan^a,
Shah Mohammad Naimul Islam^{a,*}

^a Institute of Biotechnology and Genetic Engineering, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, 1706, Bangladesh

^b Department of Pharmacology, Bangladesh Agricultural University, Mymensingh, 2202, Bangladesh

ARTICLE INFO

Keywords:

Active ingredients
Metagenomic
Microbiome
Microbiota changes
Skincare products

ABSTRACT

Healthy skin reflects a healthy microbiome and vice versa. The contemporary society, marked by a sharp increase in skin irritation cases, has compelled researchers, dermatologists, and the cosmetics industry to investigate the correlation between skin microbiomes and the use of skincare products. Different cosmetics can change skin's normal flora to a varying degree—some changes can be detrimental, there are also instances where these alterations aid in restoring the skin microbiome. Previous studies using artificial skin models, metagenomic analysis, and culture-based approaches have suggested that skincare products play an important role in skin microbial alteration. This article assessed current knowledge on microbial shifts from daily use of various personal and skincare products. We have also introduced a readily applicable framework, synthesized from various observations, which can be employed to identify the normal skin microbiome and evaluate the impact of personal care and skincare products on it. We also discussed how lifestyle choice remake skin microbial makeup. Future studies are warranted to examine the effect of personal and skincare product usage on skin microbiome across various age groups, genders, and body sites with a multi-study approach.

1. Introduction

Beauty is a cherished aspect for everyone, recognizing inner intelligence and external beauty propels toward motivation and refinement. Opting for cosmetics or skincare products regularly—daily, weekly, or monthly—instead of pills or supplements is a convenient, hassle-free way to enhance our appearance. This preference contributes to the exponential growth of the global cosmetics market, expected to reach \$463.5 billion by 2027 with a 5.3 % CAGR (compound annual growth rate) [1]. So, skin, the most versatile interface organ, comprising an average 30 m² surface area in adults, is the center of interest here [2]. Along with diversified chemicals and structural regimes, one thing that boosts skin's anisotropic heterogeneous nature is the composition of skin-dwelling microbiota. This is an absolute habitat for a diverse group of microorganisms, including bacteria, fungi, micro-eukaryotes (dust mites, Demodex mites), archaea, viruses, and phages [3]. A newborn baby gets colonized hugely after birth [4]. Gradually, due to topographical and site-specific diversity, the skin microbiota turns into a highly variable composition of an extremely versatile community depending on

* Corresponding author.

E-mail address: naimul@bsmrau.edu.bd (S.M.N. Islam).

the areas of the body, between individuals, and over time. Contrary to a popular myth, not all skin microorganisms are inherently dangerous; pathogenicity only happens when the ecosystem's equilibrium is interrupted and variety is diminished. This issue of disruption has stirred the curiosity of health-conscious individuals.

Over the past 5–10 years, there has been a significant rise in skin damage rates. While there could be multiple environmental factors contributing to this issue, researchers have identified the increased usage of synthetic chemical components in modern-day cosmetics as the primary culprit posing a significant threat [5]. Skin care products are highly appreciated until they attract some uninvited guests. Within the personal care industry, the skin microbiota represents a promising yet often overlooked area, as evidenced by the increasing use of formulations that may impact the balance of the skin microbiota. Numerous instances of skin diseases have been linked to this dysbiosis or perturbation of the skin microbiome [6]. The imbalance can often be traced back to excessive product usage. Understanding the intricate relationship between skin chemistry, microbes, and their variations due to cosmetic application is essential. Gaining a better understanding of the correlation between microorganisms and skin care routines could significantly expand our knowledge of the relationship between the skin microbiome, skin diseases, and overall skin health. This, in turn, might pave the way for a more sustainable and healthier human existence. This review aims to highlight the latest findings on how different skin care products affect the skin's microbiome, as well as the potential consequences associated with these alterations. It also provides insights into various methods for assessing the skin microbiome and screening the effects of skin care products on skin inhabitants.

2. Conducting literature review

The literature search strategy entailed a comprehensive review of scholarly articles published up to November 2023, focusing on the contemporary discussion surrounding microbial shifts arising from the habitual use of various personal and skincare products. Here we reviewed more than 100 research articles to compile regarding the whole knowledge. We searched online databases including Web of Science, Science Direct, Scopus, and Google Scholar. Keyword selection was guided by the thematic structures and content of the review that included skin microbiome, cosmetics impacts, microbial dysbiosis, dermatological research, microbial alteration, active ingredients of skin care products and so on. Moreover, the search criteria were strictly limited to English-language publications.

3. Skin microbiome: composition and diversity

Skin microbiota, also known as skin flora, refers to microorganisms that live on the skin. It is the second-largest microbiota of the human body in mass [7]. Skin provides vital nutrients for the development of this microbiota through sweat or stratum corneum, which is composed of 75–80 % proteins (primarily keratins and membrane proteins), 5–15 % lipids (ceramides, cholesterol mainly), 5–10 % unknown compounds, and water (15–20 % of the total tissue dry weight) [8]. Within 2- and 3-mm thickness of the skin, microbiome has the most variation over time than the gut's and the mouth's microbiomes [9]. The skin harbors a vast array of bacterial species, with over 1200 identified. Among them, more than 90 % belong to four phyla: Actinobacteria (52 %), including *Micrococcus*, *Propionibacteria*, and *Corynebacteria* genera; Firmicutes (24 %), comprising *Staphylococcus*, *Lactobacillus*, and *Streptococcus* genera; Proteobacteria (16 %), which includes *Paracoccus*, *Haematobacter*, and *Sphingomonas* genera; and Bacteroidetes (6 %), represented by the genera *Porphyromonas*, *Prevotella*, and *Flavobacterium* [10,11]. Estimates suggest that *Cutibacterium*, *Staphylococcus*, and *Corynebacterium* genera, found throughout various sites on the skin, constitute approximately 45–80 % of the skin microbiome [12]. Additionally, up to 4.2 % of the prokaryotic skin microbiome is composed of archaea, with the dominant group being Thaumarchaeota phyla [13].

Various fungi have been notably found on different body regions, including *Malassezia* spp., *Aspergillus*, *Cladosporium*, *Epicoccum*, *Phoma*, *Cryptococcus*, *Rhodotorula*, *Microsporum*, *Trichophyton*, *Saccharomyces*, *Candida*, and *Epidermophyton* [14].

Regarding viruses or phages, the skin can harbor Molluscum contagiosum virus, Merkel cell polyomavirus (MCPyV), Alphapapillomavirus, Simian virus, Acheta domestica densovirus (AddNV), Actinomyces phage, Propionibacterium phage, Polyomavirus HPyV7, Streptococcus phage, Stenotrophomonas phage, Enterobacteria phage, and more [7].

The resident microflora plays an advantageous role in modulating the immune system, occupying niches, and preventing the entry of harmful and contagious microorganisms into the skin. While skin microorganisms have been broadly identified and categorized [15], quantitative measurements are still necessary to compare investigations carried out by different researchers using various methodologies to consolidate their findings.

4. Interplay between cosmetics and skin microbiome

A vast array of skin care products, including body washes, gels, lotions, exfoliants, moisturizers, toners, and sunscreens, is available to address various beauty conditions, with a primary focus on nourishing the skin from within. However, researchers must acknowledge that cosmetic components can have both positive and negative impacts on the skin's microbiota [16]. Additionally, under specific physiochemical conditions, cosmetic compounds can serve as nutrient sources, promoting the growth of opportunistic pathogenic microorganisms that may lead to significant infections and diseases [17].

Certain research points to “microbiome resilience,” suggesting that there may be no negative effects on microorganisms due to the use of skin wash products. According to these studies, the concentrations of microorganisms are dramatically reduced by 5–10 times during product application [18]. However, the durability of these products varies depending on the application site, and their effects can last for weeks with highly individual reactions. These reactions may include changes in steroid and pheromone levels, as well as alterations in the structure and dynamics of bacterial and archaeal ecosystems, leading to a modified scenario [18].

As richness in microbial diversity has been proven to be linked with healthy skin [19], any kind of effect upon them destines changes leading to threats. But due to the broader range, studying their variations over skin has always been a challenge. These effects are not only occupied in the applied sites, but also other parts of the body [20]. For launching cosmetics, in depth studies are needed in context of microbiome. It is a fact of optimism that such practices are being encouraged nowadays in modern cosmetics. The first “Microbiome-friendly” certification for a cosmetic product was premiered in 2019 and was established by ‘MyMicrobiome’. Here the product’s purity, the targeted area’s specific bacteria’s safety, the preservation of the microbiome diversity, and the preservation of the skin’s natural balance are all confirmed (neither through the induction of harmful microorganisms nor by the repression of commensals) [21].

5. Assessing skin microbiome and screening the response of cosmetics

There is a need to explore effective approaches for comparing the skin microbiome after using skin care products to determine any deviations from normal skin conditions. This initial step is crucial in establishing a connection between the amount of synthetic ingredients in a product and its impact on skin health. In the 1950s, American dermatologist Albert Kligman pioneered human microbiota research in dermatology, employing advanced cell culture methods [22,23]. Today, modern research techniques allow us to identify non-culturable, newer forms of microbes residing in the surface and deeper layers of human skin. Overcoming the limitations of culture-dependent studies, molecular methods, such as multi-omics approach integration [20], 18S and 16S rRNA gene sequencing, DNA barcoding, PCR-denaturing gradient gel electrophoresis sequencing [24], enable the detection of microbes present in low quantities. Additionally, studying the chemicals produced by microbes through mass spectrometry visualization and molecular networking provides a wide range of microbial studies [16]. To identify the microbes, various methods like swabbing, biopsy, or tape stripping [25] are used, with the selection depending on the categories of microbes to be identified [26]. For assessing the ultimate outcomes resulting from the application of different products, the use of artificial 3D skin models and leather skin models has become popular [27,28]. Other methods for screening include the duel culture method, Mass spectrophotometry, FTIR spectro microscopy (Fourier-transform infrared spectroscopy), or magnetic resonance imaging [29,30]. Incorporating bioinformatics studies can also shed light on the effects of chemicals and bioactive products from microbes. Fig. 1 illustrates a workflow for identifying skin microbes and outlines possible approaches for screening cosmetics in relation to changes in the skin microbiome. Molecular approaches are commendable, but they do raise some counter-thoughts, such as their inability to differentiate between the genes of living and dead organisms, which can lead to confusing results. Therefore, to gain a comprehensive understanding of skin microbes, it is essential to combine whole genome sequencing with skin models and cultural approaches.

6. Effect of cosmetics on the skin’s microbiome

6.1. Effect of common cosmetic preservatives on skin microbe dynamics

In cosmetics, ingredients such as water, oils, peptides, and carbohydrates create an environment conducive to the growth of microorganisms. To prevent contamination, preservatives are commonly added, which remain active on the skin upon application.

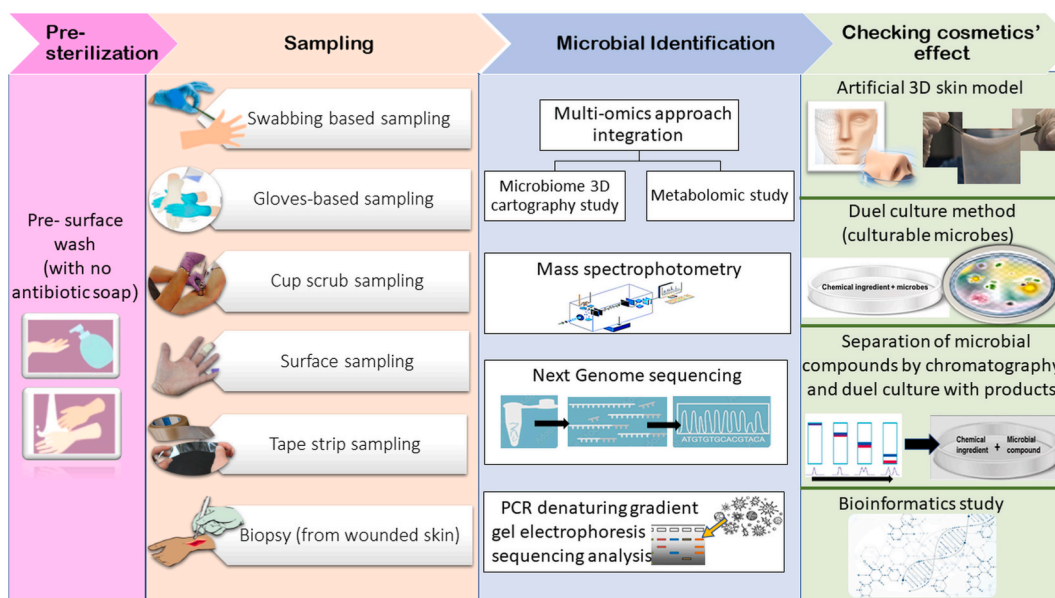


Fig. 1. A workflow of assessing skin microbe and cosmetics' response.

However, their blind use solely for preservation with antimicrobial properties might not serve their intended purpose [31].

A study conducted by Pinto, Ciardiello [32] revealed that various tested preservatives had an impact on the dynamics of targeted bacteria, including *Cutibacterium acnes* (previously known as *Propionibacterium acnes*), *Staphylococcus epidermidis*, and *Staphylococcus aureus*. While the combination of different preservatives in one bacterial strain (*S. aureus*) showed somewhat positive results, it proved to be both beneficial and harmful for the other two bacteria.

The most effective mixtures for reestablishing a pre-existing dysbiosis were found to contain hydroxyacetophenone, phenylpropanol, propanediol, caprylyl glycol, tocopherol, and tetrasodium glutamate diacetate because they act to moderately inhibit *C. acnes* and strongly inhibit *S. aureus* without simultaneously inhibiting *S. aureus* growth. The finest combinations to use in topical solutions for the skin and scalp where it is vital to maintain the eubiosis of the microbiota were proposed to be C1 (Sodium benzoate phenoxyethanol, ethylhexylglycerin), C4 (Sodium anisate, 1,2-hexanediol, ammonium acryloyldimethyltaurate/vp copolymer), C6 (Hydroxyacetophenone, phenylpropanol, propanediol, caprylyl glycol, tocopherol, disodium EDTA), and C7 (Benzyl alcohol, benzoic acid, dehydroacetic acid, ammonium acryloyldimethyltaurate/vp copolymer).

Toner, emulsion, cream, and baby cream contain common preservatives like parabens, 1, 2-hexanediol, phenoxyethanol. They exhibited potent antibacterial effects against *S. aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* as well as other skin-resident bacteria such as *S. epidermidis*, *Shigella flexneri*, *Enterobacter aerogenes* and so on that are not the target at all [33]. Paraben, used in common cream had shown to have robust effects against skin filamentous fungi and yeasts like *Candida* spp. and *Malassezia* spp. [34]. *C. acnes* and *S. epidermis* were significantly inhibited by the short-term usage of 0.5–5 % methylparaben in cosmetics [35]. Methylisothiazolinone was seen to inhibit *S. epidermidis* [36]. The use of phenoxyethanol disturbed the skin microbiota conferring the change both at the phylum level (Proteobacteria increased) and at species level (*Propionibacterium humerusii*, *S. epidermidis* decreased) [33,36]. The influence of preservatives may go beyond the parameters of the product composition and may have a potentially negative impact on the skin microbiota, according to a theory put up in light of the expanding understanding of the significance of the human microbiome. The specified and some common microbes changed by basic ingredients that are used in skin care products are illustrated in Fig. 2.

6.2. Changes in hand and palm microbiome by cosmetics

Hand and palm microbiome seems quite interesting with 83 % difference between the left and right hands of same individual, 87 % differences in difference in individuals and a greater overall diversity in female hands than males [37]. The amount of exposure to environmental elements, how often hands are washed all likely play a role in the variation in the skin microbiota of the hands. It is obviously affirmative to have a health care setting strategies for cleaning hands in but chronic washing is potential for dysbiosis in some individuals [38]. Healthcare personnel who routinely washed their hands have more pathogenic bacteria on them than those who didn't [39] indicating less microbial diversity led to the potentiality to increase pathogenic species including *S. aureus*,

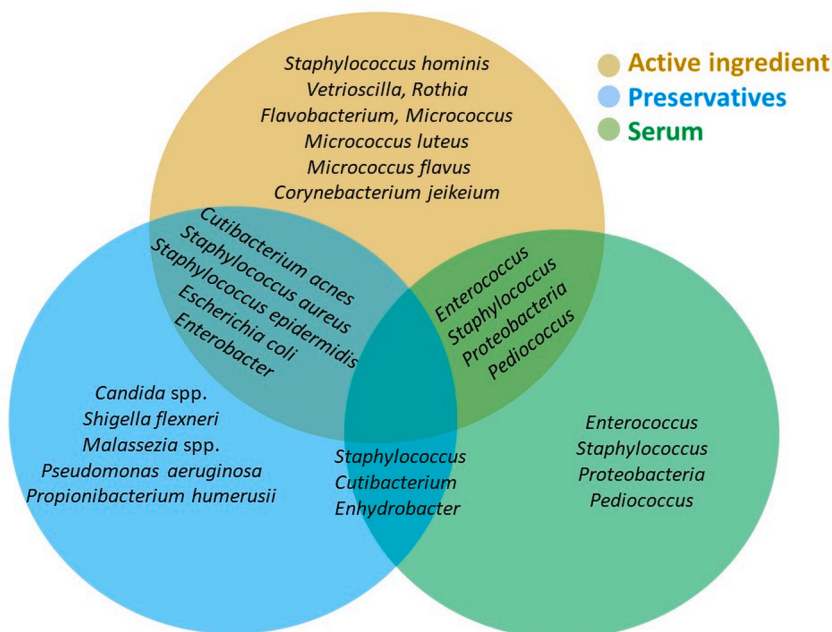


Fig. 2. A Venn diagram showing the specified and some common microbes changed by basic ingredients that are used in skin care products. Different color code represents different groups (Bronze- Active ingredient (eg.Kojic Acid, Isotretinoin etc.); Blue- Preservative (eg.Parabens, Methylparaben, Ethylparaben, Phenoxyethanol, Methylisothiazolinone etc.); Green- Serum (Galacto-oligosaccharide, Hyaluronic Acid etc.).

Table 1

The Alteration in skin microbes caused by the active ingredients used in cosmetics.

Active ingredients/A mixture of active ingredients	Alteration in skin microbes	References		
Maltodextrin, <i>Zymomonas ferment</i> extract, honey extract, aqua	<i>Corynebacterium jeikeium</i> ↑	[58]		
	<i>Micrococcus luteus</i> ↑			
	<i>Staphylococcus aureus</i> ↑			
	<i>Micrococci</i> ↓			
	<i>Staphylococcus epidermidis</i> ↓			
	<i>Staphylococcus hominis</i> ↓			
	<i>Micrococcus flavus</i> ↓			
	<i>Cutibacterium avidum</i> ↓			
	Glycerin, <i>Laminaria digitata</i> extract, <i>Chlorella vulgaris</i> extract, saccharide isomerate, phenoxyethanol, ethylhexylglycerin, aqua, seawater		<i>Staphylococcus aureus</i> ↑	[58]
			<i>Staphylococcus hominis</i> <i>Micrococcus luteus</i> ↑	
<i>Cutibacterium avidum</i> ↑				
<i>S. epidermidis</i> ↓				
<i>Micrococcus flavus</i> ↓				
Fermented oil		<i>Staphylococcus</i> ↑	[59]	
	<i>Proteobacteria</i> ↓			
	ExpoZenfi by GREENTECH	Bacterial diversity ↑		[60]
<i>Staphylococcus epidermidis</i> ↑				
Isotretinoin	<i>Rothia</i> ↑	[61]		
	<i>Flavobacterium</i>			
	<i>Enterobacter</i>			
	<i>Micrococcus</i>			
	<i>Cutibacterium acnes</i> ↓			
<i>Halymenia durvillei</i> (red algae) extract	<i>Proteobacteria</i> phyla ↑	[60]		

(continued on next page)

Table 1 (continued)

Active ingredients/A mixture of active ingredients	Alteration in skin microbes	References
	Bacteroidetes phyla	
	↑	
	Actinobacteria phyla	
	↓	
	Firmicutes phyla	
	↓	
	<i>Corynebacterium kroppendenstedtii</i>	
	↓	

↑ indicates increase, ↓ indicates decrease.

Enterococcus spp., *Candida albicans* [40]. The alteration is most in the prominent skin taxa than others in antiseptic treatments [41]. The alcohol-based hand sanitizer and ethanol showed a reduction of the levels of viable aerobic-anaerobic bacteria [42]. Antimicrobial soap had impacts on epidermal antibacterial defense system and upon skin microbes alike *Streptococcus* [43]. Antibacterial wipes resulted in a decrease in the number of *S. aureus* [44].

Many cleansers use surfactants that not only take away the key defensive components of our protective skin barrier exposing it to outside aggressors, but also misbalances the pH of skin [42]. Skin pH is fundamental to maintaining a healthy microbial community. Higher concentration of NaOH in cleansers, soaps, makeup, creams or lotions leads to higher pH level, causing *Candida albicans*, a commensal microbe turning into a fungal infection [45]. Extreme amount of citric acid and lactic acid in cleanser or exfoliants lowered normal skin pH causing decreased *C. acnes*. Similarly, with the increase in pH, the activity and population had been seen to be boosted for *C. acnes* and *S. aureus* [46].

6.3. Microbiome alterations induced by the active ingredients in cosmetics

Active ingredients in skin care products are specifically formulated for various purposes such as brightening, addressing dryness, combating aging, treating sunburn, and managing acne. In addition to different extracts and chemicals, pre-pro-biotics are also included in this list. Collectively, all these groups can have both positive and negative impacts on the skin's microbiome. Plant extracts, fruit extracts, and seaweed extracts used in cosmetics have shown incredible potential in modifying *C. acnes* populations, effectively combating skin pathogenesis like acne vulgaris [47,48]. However, certain ingredients, like kojic acid found in skin lightening creams and lotions, have been found to be capable of traveling into the bloodstream [49]. Moreover, pure kojic acid, when exposed to UV light, has the potential to induce gene mutations in *E. coli* strains [50]. As *E. coli* strains naturally occur on the skin in small amounts, such mutations might lead to harmful consequences [51]. Table 1 provides an overview of research findings highlighting the changes in skin microbes caused by the active ingredients of cosmetics and Fig. 2 shows microbial changes caused by active ingredients along with preservatives and serum. In contrast to synthetic substances, which humans have only encountered in the last 60 years of their 200 000-year history, natural components, in the quantities found in nature, are not perceived as "foreign" to the skin's natural condition [52]. This is why branding products as "natural or organic" easily captures attention. Consumers often prefer using products labelled as "natural cosmetics" due to concerns about their skin. However, it's essential to recognize that natural products may not be as pure as they seem in reality. For instance, Methylisothiazolinone (MI), a synthetic compound used in some 'natural' labelled products, has been associated with potential harm, including neurotoxicity [53], allergic reactions [54], and changes in microorganisms [55]. Alarmingly, this MI preservative is not limited to adult products like makeup, eyeliners, makeup removers, blush, face powder, hair care products, nail and waxing products, moisturizing creams, and sunscreen; it is also used in baby wipes and bath products. Even in toddlers, their diapers, skin pH, and dermatitis are linked with *Candida albicans* and *S. aureus* [56]. The research conducted by Wallen-Russell [57] investigated the microbial community shifts caused by using three different cosmetic conditions: a 100 % truly natural product (JooMo's face wash), a product labelled as natural but containing 70 % synthetic ingredients, and a synthetic product with 75 % synthetic components. Microbiome sampling was done before product use (T1), after two weeks (T2), and after four weeks (T3) of product use. During the transition from T1 to T2 and T1 to T3, JooMo showed the fastest rise in Chao1 diversity and species richness. This suggests that cosmetics companies should conduct studies on skin microbiota when launching new active ingredients to ensure their products maintain, improve, or restore a healthy skin-microbiome balance, especially in cases of a disrupted microbiome.

6.4. Alternation of skin microbiome by different types of cosmetics

The topical application of personal hygiene products can alter the skin's lipid film, affecting microbial diversity and the overall richness of bacterial species on the skin. For instance, active body wash products have demonstrated potential advantages, as they can eliminate harmful bacteria such as *Brevibacterium casei* and *Rhodotorula mucilaginosa*, while promoting the growth of beneficial bacteria like *C. acnes* [62].

Furthermore, a basic skin care routine consisting of skin softener, lotion, essence, and cream with moisturizing compounds was found to impact the abundance of two common skin phyla: Actinobacteria (decreased) and Proteobacteria (increased) on facial cheeks [63]. The cause of these changes could be attributed to other skin bacteria growing and utilizing ingredients from the basic cosmetics,

Table 2
Some common skin care products causing microbial alteration in skin.

Product category	Alteration in skin microbes	References
Lotion application in lower foot (against xerosis, extreme dryness)	<p><i>Staphylococcus epidermidis</i> ↑</p> <p><i>Xanthomonas campestris</i> ↑</p> <p><i>Xanthomonas</i> spp.</p>	[64]
Short chain fructo-oligosaccharides (A prebiotic used in powder etc.)	<p>↑ <i>Staphylococcus epidermidis</i> at</p> <p>↑ lower concentration (0.5–5 %) <i>Staphylococcus aureus</i></p> <p>↓s <i>Staphylococcus epidermidis</i> at</p> <p>↓ higher concentration (10–15 %) <i>C. acnes</i> (complete halt)</p>	[65]
Spermidine used in lotion, cream	<p>↓ <i>Staphylococcus pneumonia</i> ↑</p> <p><i>Staphylococcus infantis</i> ↑</p> <p><i>Staphylococcus thermophiles</i> ↓</p>	[66]
Ceramides in moisturizers	<p><i>Streptococcus</i> spp.</p>	[67]
Serum cosmetics containing galacto-oligosaccharides	<p>↓ <i>Burkholderia</i> ↑</p> <p><i>Bifidobacteria</i> ↑</p> <p><i>Lactobacilli</i> ↑</p> <p><i>Lactococcus</i> ↑</p> <p><i>Sphingomonas</i> ↑</p> <p><i>Thermoanaerobacterium</i> ↑</p> <p><i>Staphylococcus aureus</i> ↓</p> <p><i>Staphylococcus</i> ↓</p> <p><i>Proteobacteria</i> ↓</p> <p><i>Cutibacterium</i> ↓</p>	[68]

(continued on next page)

Table 2 (continued)

Product category	Alteration in skin microbes	References
Foot powder use	<i>Pediococcus</i> ↓	[20]
	<i>Enhydrobacter</i> ↓	
	Enterobacteriaceae family ↓	
	<i>Micrococcus</i> ↑	
	<i>Anaerococcus</i> ↑	
	<i>Streptococcus</i> ↑	
	<i>Brevibacterium Acinetobacter</i> ↑	
Selenium in lotion, sunscreen, creams	Moraxellaceae family ↑	[69,70]
	<i>Staphylococcus aureus</i> ↓	
Moisturizers containing lipids	<i>Staphylococcus</i> ↑	[16,71,72]
	<i>Propionibacterium</i> ↑	

↑ indicates increase, ↓ indicates decrease.

or the cosmetics may be hindering the growth of normal skin bacterial groups, or even altering the skin environment itself. The following Table 2 provides an overview of product-based variations that influence skin microbes.

Sebocytes create sebum, a natural moisturizer that is essential for maintaining healthy skin. Excessive sebum production can lead to issues like acne, greasy skin, and clogged pores. To prevent such problems, many individuals use different skin care products, such as lotions [73], which have proven to be effective in reducing sebum production. However, decreased sebum can also result in a reduced food source for microbes, leading to a decline in the number of *Staphylococcus* bacteria and causing an imbalance in the skin microbiome. A comparison between makeup users and non-users revealed significantly higher microbial diversity on the forehead skin of makeup users, including an increase in the genera *Selenomonas*, *Aggregatibacter*, and *Aquicella* [74].

The changes in skin microbes are intricate and interconnected. Fig. 3A–F depicts the range of products (lotion, cream and moisture; powder, antiperspirant and deodorant; body, hand and face wash) that commonly influence bacterial or fungal populations, detailing those that have individual and combined effects on these species as evidenced by multiple research studies.

6.5. Microplastic

Microplastics and nanoplastics are ubiquitous constituents of human daily consumption, serving as pervasive pollutants that appear in many forms such as microbeads and fibers. The impact of microplastic load is terrifying for flora-fauna, terrestrial, ocean organisms [75,76] and soil microbiota [77], covering all sort of environmental microbiomes [78]. They can adsorb organic and inorganic contaminants on their surface [79]. In addition, biofilms can form on their surface and act as carriers of pathogenic vectors and antimicrobial resistance [80], career for pollutants [81], microorganisms and resistance genes [82]. Their effects on different animals' microbiome had been notified.

Polystyrene reduced of the relative abundance of phyla Firmicutes, Bacteroidetes and Verrucomicrobia, increased Actinobacteria with 0.5 μm particles size in the gastrointestinal tract of mice [83]. Polystyrene microplastics fibres and fragments reduced Actinobacteria; fibers affected specific bacteria genera while fragments caused a decrease in *Pseudomonas* and *Aeromonas* genus and all three shapes increased the relative abundance of *Gordonia* [84]. It's 5 μm increased *Staphylococcus* and 200 μm boosted the genera *Vibrio*, *Acinetobacter*, *Porphyromonas*, *Haemophilus*, *Neisseria* and *Lactococcus* in zebra fish [85]. Phyla Cyanobacteria, Chloroflexi, Fusobacteria and Proteobacteria were increased, while Nitrospirae, Bacteroidetes, and Firmicutes were reduced in freshwater crabs [86] via

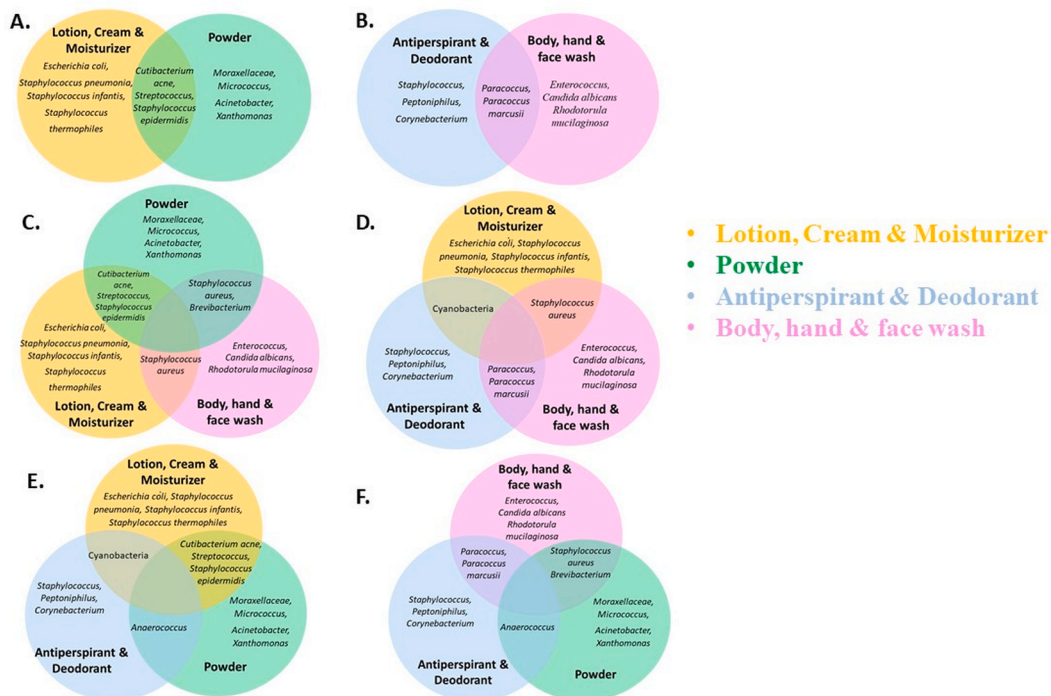


Fig. 3. Venn diagrams illustrating the specific and common microbial changes influenced by skincare products. A: Alteration of microbes by Lotion, Cream & Moisturizer + Powder. B: Alteration of microbes by Antiperspirant & Deodorant + Body, hand & face wash. C: Alteration of microbes by Powder + Lotion, Cream & Moisturizer + Body, hand & face wash. D: Alteration of microbes by Lotion, Cream & Moisturizer + Antiperspirant & Deodorant + Body, hand & face wash. E: Alteration of microbes by Lotion, Cream & Moisturizer + Antiperspirant & Deodorant + Powder. F: Alteration of microbes by Body, hand & face wash + Antiperspirant & Deodorant + Powder.

microplastic intervention. In Human, exposure to Polystyrene MPs induced gut microbial shifts increasing α -diversity and abundance of potentially harmful pathobionts, such as Dethiosulfovibrionaceae and Enterobacteriaceae family [87]. Polyethylene alters Firmicutes, Bacteroidetes, Proteobacteria, and Verrucomicrobia phyla [88]. Reductions in *Bifidobacterium* spp., *Clostridium* spp., *Staphylococcus* spp., total aerobic and anaerobic bacteria, and increase in *Escherichia* or *Shigella*, *Cloacibacillus*, *Bilophila*, *Eisenbergiella*, *Megasphaera* and *Oscillibacter* genus including rise in Firmicutes phylum [89] had been shown in different chambers of gut. Polypropylene microplastics caused significant changes in fish gut microbiome, reducing beneficial lactic acid bacteria and increasing potential pathogenic microorganism Proteobacteria and Vibrionales [90].

Regarding human microbiomes, mammal microbiomes are more similar to that of humans than non-mammal microbiomes. Microbead is the primary source of Microplastics used as cleansing or exfoliating agents in a diversity of personal care and cosmetic products such as shower gels, toothpaste, nail polishes, or eye shadows, amongst many others. From the above-mentioned studies, we can predict there is highest possible chance of those microplastic altering our normal skin microbiome. In a research conducted in Macao showed 100 skin care products (facial skin, body skin, cosmetics) out of 144 are with microplastic [91]. A significant quantity of personal care and cosmetics products are found to include microplastics such as polyethylene, polyethylene terephthalate, polypropylene, and other similar substances. The aforementioned alterations in the environment and gut microbiota may lead us to speculate that they could potentially impact the composition of the skin microbiome. Unfortunately, there is a lack of relevant data regarding this particular matter. A comprehensive investigation is necessary to ascertain the impact of microplastic exposure, either alone or in conjunction with skin-care products, on the host microbiome.

7. Consequences of altered microbiome

In contrast to individual variances, it is a well-known fact that most adult human microbiomes remain stable without intervention [9,92,93]. However, shifts can occur due to both intrinsic and extrinsic factors, leading to the opposite phenomenon. Thus far, it has been established that the use of various skincare products can trigger microbial alterations. Whether this shifting will be named after positive or negative, is dependent upon the species of microbes being changed. Depending upon the types of products use, the consequences might vary. For sure, many of the skin care products offer us a variety of favors in skin commensals. One of the consequences that terrifies all is microbes misbalance causes their shifting from friend to foe for instance *C. acnes* [94]. The same statement is justified as commensals *S. epidermidis* would be turned into pathogen if its number is increased creating serious skin irritation by boosting atopic dermatitis [95]. But in normal skin, *S. epidermidis* aids in skin homeostasis and lowers the pathogenic inflammation triggered by

C. acnes as it drops off the TLR2 protein (Toll like receptor 2 protein) production, which reasons skin inflammation [96]. Dysbiosis and altered microbial biodiversity of the skin microbiome has been linked with many diseases [97]. Acne was modulated by Proteobacteria, Firmicutes and Actinobacteria [98] and also by many species like *C. acnes* [99], *Streptococcus* [100], *S. epidermidis* [101], *Malassezia* species [102,103] etc. Atopic dermatitis, a chronic skin inflammatory disease impacts roughly 15%–20% of children and 1%–3% of adults worldwide and has skyrocketed 2- to 3-fold [104]. It had seen to have relation with significant skin microbial changes such as *S. aureus*, *S. epidermidis* and *Staphylococcus haemolyticus* [105,106], *Propionibacterium*, *Corynebacterium* and *Streptococcus* [105,107]. Some common skin dwellers *Staphylococcus*, *Corynebacterium*, *Streptococcus* and *Propionibacterium* were related to skin diseases like psoriatic lesion [108]. *Malassezia* fungal species linking with folliculitis [109], *Staphylococcus* with diabetic skin [110], *S. aureus* with allergies [111], *Streptococcus*, *Staphylococcus*, *Fusobacterium* with leishmaniasis [112], *Acinetobacter lwoffii* with allergic sensitization and inflammation [113] had been reported. Apart from the common one (*Malassezia* fungi) [114] some low abundant genera like *Gordonia* and *Geobacillus* also had seen linking with skin disease-rosacea [115]. Regarding bodies defense mechanism, some ardently serious issues are involved. Shifting of normal microbiome of *Staphylococcus*, and in particular *S. epidermidis* results in the disruption of skin immune system causing the natural antimicrobial bacteriocins production hampered [116]. Changes in *Porphyromonas*, *Streptococcus*, *Peptostreptococcus*, *Sphingomonas*, *Stenotrophomonas*, *Anaerococcus*, *Staphylococcus*, *Corynebacterium* etc. were related to wound healing process [117]. So even minor shift may result some catastrophe. These findings hold significance from an immunological perspective, as they imply a direct communication link between host health and microbial cells in the skin.

While individual microorganisms alone may or may not directly cause cancer, the overall composition and diversity of the skin microbiome can have an impact. Cosmetics can alter the skin microbiome by reducing its diversity and balance, resulting in microbial imbalance (dysbiosis). This, in turn, can lead to inflammation, immune suppression, oxidative stress, and infections, all of which can contribute to the promotion of skin cancer [118]. The studies mentioned above indicate varying levels of microbial increase and decrease in the skin microbial community. Some of these common microbes have been shown to be linked to cancer. For instance, *S. aureus* is associated with the carcinogenic progression from actinic keratosis to squamous cell carcinoma (SCC), a type of skin cancer that develops in the flat squamous cells located in the outer layer of the skin.” [119,120]. Moreover, it can influence the expression of Human beta-defensin-2, thereby causing SCC proliferation [121]. **Cutaneous T cell lymphoma, another type of skin cancer had been linked with *S. aureus*** [122]. Some research showed that *Corynebacterium* species may influence the development of malignant melanoma, the deadliest form of skin cancer, accounting for 75% of all skin cancer-related deaths [123,124]. Skin cancers might be boosted as a result of skin microbiome alteration demonstrating skin barrier disruption [125] for *Staphylococcus epidermidis* [126].

Coffee aqueous extract is being used in many skins care formula that had demonstrated links with Colorectal cancer [127]. Tryptophan, a microbial metabolite detected on skin, also used in cosmetics [128] has effect on aryl hydrocarbon receptor. Imbalanced regulation of aryl hydrocarbon receptor expression or activity promotes cancer development alike breast cancer [129], lung cancer [130], oesophageal cancer [131] etc. *Malassezia* sp. affects skin cancer [132]. It increased aryl hydrocarbon receptor activity conferring increased tumorigenesis [133]. *Candida* is related with an increased risk of numerous malignancies, including hematologic, head and neck, pancreatic, skin, and thyroid cancer [134]. Viruses are also included in this list [135]. More study is needed to better understand how various cosmetics products promoter different forms of cancer and how they can be managed to prevent or treat cancer.

8. Conclusion

A diverse and thriving skin microbiome is essential for human health, making it a novel objective for skin care products to focus on maintaining or restoring the skin’s microbiota. The application of additional skin care products can significantly impact the microbial diversity on the skin, resulting in both positive and negative differences. Each type of product used on the skin has its own individual and combined effects. Common skin microbes like *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Corynebacterium*, *Cutibacterium acnes*, and *Malassezia* spp. are frequently altered by diversified arrays of such products application. As microbial alteration is linked with lots of diseases ranging from skin irritation to cancers not only in skin but also of other organs, precise studies are needed. A combination of sampling methods, culture-based strategies, and modernized research tools, such as artificial skin models, spectrophotometry, bioinformatics, and metagenomic analysis, is necessary. To gain a comprehensive understanding of the skin’s microbiome and its complex interactions when using various cosmetics, further investigations considering factors like age, location, and specific body sites are essential. Ultimately, these studies aim to ensure a healthy human skin microbiome. No definitive conclusion has been reached to universally categorize any product as entirely positive or negative. The question arises whether the negative impacts outweigh the positive ones.

Data availability statement

No data was used for the research described in the article.

CRedit authorship contribution statement

Mahjabin Ferdaous Mim: Writing – original draft, Visualization, Data curation, Conceptualization. **Mahmudul Hasan Sikder:** Writing – review & editing. **Md. Zahid Hasan Chowdhury:** Visualization, Data curation. **Ashkar-Ul-Alam Bhuiyan:** Visualization. **Nayeematul Zinan:** Data curation. **Shah Mohammad Naimul Islam:** Writing – review & editing, Conceptualization.

Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: Shah Mohammad Naimul Islam reports administrative support was provided by Bangabandhu Sheikh Mujibur Rahman Agricultural University.

References

- [1] Shankar, Cosmetics market by category, gender and distribution channel: opportunity analysis and industry forecast, 2021–2027, Allied market research (2021). <https://www.alliedmarketresearch.com/press-release/cosmetics-market.html>.
- [2] R.L. Gallo, Human skin is the largest epithelial surface for interaction with microbes, *J. Invest. Dermatol.* 137 (2017) 1213–1214.
- [3] S. Verbanic, C.Y. Kim, J.M. Deacon, I.A. Chen, Improved single-swab sample preparation for recovering bacterial and phage DNA from human skin and wound microbiomes, *BMC Microbiol.* 19 (2019) 1–13.
- [4] J. Oh, A.L. Byrd, M. Park, H.H. Kong, J.A. Segre, Temporal stability of the human skin microbiome, *Cell* 165 (2016) 854–866.
- [5] C. Wallen-Russell, S. Wallen-Russell, Meta analysis of skin microbiome: new link between skin microbiota diversity and skin health with proposal to use this as a future mechanism to determine whether cosmetic products damage the skin, *Cosmet. Toilet.* 4 (2017) 14.
- [6] I.A. Myles, K.W. Williams, J.D. Reckhow, M.L. Jammeh, N.B. Pincus, I. Sastalla, D. Saleem, K.D. Stone, S.K. Datta, Transplantation of human skin microbiota in models of atopic dermatitis, *JCI Insight* 1 (2016).
- [7] A.L. Byrd, Y. Belkaid, J.A. Segre, The human skin microbiome, *Nat. Rev. Microbiol.* 16 (2018) 143–155.
- [8] J.S. Barbieri, K. Wanat, J. Seykora, *Skin: Basic Structure and Function*, 2014.
- [9] E.K. Costello, C.L. Lauber, M. Hamady, N. Fierer, J.I. Gordon, R. Knight, Bacterial community variation in human body habitats across space and time, *Science* 326 (2009) 1694–1697.
- [10] E.A. Grice, H.H. Kong, G. Renaud, A.C. Young, NISC Comparative Sequencing Program, G.G. Bouffard, R.W. Blakesley, T.G. Wolfsberg, M.L. Turner, J.A. Segre, A diversity profile of the human skin microbiota, *Genome Res.* 18 (2008) 1043–1050.
- [11] E.A. Grice, H.H. Kong, S. Conlan, C.B. Deming, J. Davis, A.C. Young, NISC Comparative Sequencing Program, G.G. Bouffard, R.W. Blakesley, P.R. Murray, Topographical and temporal diversity of the human skin microbiome, *Science* 324 (2009) 1190–1192.
- [12] S. Samaras, M. Hoptroff, *The Microbiome of Healthy Skin*, *Skin Microbiome Handbook: from Basic Research to Product Development*, 2020, pp. 1–32.
- [13] A.J. Probst, A.K. Auerbach, C. Moissl-Eichinger, Archaea on human skin, *PLoS One* 8 (2013) e65388.
- [14] H. Li, B.N. Goh, W.K. Teh, Z. Jiang, J.P.Z. Goh, A. Goh, G. Wu, S.S. Hoon, M. Raida, A. Camattari, Skin commensal *Malassezia globosa* secreted protease attenuates *Staphylococcus aureus* biofilm formation, *J. Invest. Dermatol.* 138 (2018) 1137–1145.
- [15] R. Sfriso, M. Egert, M. Gempeler, R. Voegeli, R. Campiche, Revealing the secret life of skin-with the microbiome you never walk alone, *Int. J. Cosmet. Sci.* 42 (2020) 116–126.
- [16] A. Bouslimani, C. Porto, C.M. Rath, M. Wang, Y. Guo, A. Gonzalez, D. Berg-Lyon, G. Ackermann, G.J. Moeller Christensen, T. Nakatsuji, *Molecular Cartography of the Human Skin Surface in 3D*, vol. 112, *Proceedings of the National Academy of Sciences*, 2015, pp. E2120–E2129.
- [17] E. Neza, M. Centini, Microbiologically contaminated and over-preserved cosmetic products according Rapex 2008–2014, *Cosmet. Toilet.* 3 (2016) 3.
- [18] B. Murphy, M. Hoptroff, D. Arnold, R. Eccles, S. Campbell-Lee, In-vivo impact of common cosmetic preservative systems in full formulation on the skin microbiome, *PLoS One* 16 (2021) e0254172.
- [19] B.B. Finlay, M.-C. Arrietta, *Let Them Eat Dirt*, Greystone Books, 2016.
- [20] A. Bouslimani, R. da Silva, T. Kosciulek, S. Janssen, C. Callewaert, A. Amir, K. Dorrestein, A.V. Melnik, L.S. Zaramela, J.-N. Kim, The impact of skin care products on skin chemistry and microbiome dynamics, *BMC Biol.* 17 (2019) 1–20.
- [21] MyMicrobiome. Retrieved from, <https://microbiome-friendly.com/en/>, February 3, 2023.
- [22] B. Dréno, E. Araviiskaia, E. Berardesca, G. Gontijo, M. Sanchez Viera, L.F. Xiang, R. Martin, T. Bieber, Microbiome in healthy skin, update for dermatologists, *J. Eur. Acad. Dermatol. Venereol.* 30 (2016) 2038–2047.
- [23] D.M. Pillsbury, *Manual OF dermatology*, *AJN The American Journal of Nursing* 43 (1943) 791.
- [24] W. Li, L. Han, P. Yu, C. Ma, X. Wu, J. Xu, Nested PCR-denaturing gradient gel electrophoresis analysis of human skin microbial diversity with age, *Microbiol. Res.* 169 (2014) 686–692.
- [25] H.H. Kong, B. Andersson, T. Clavel, J.E. Common, S.A. Jackson, N.D. Olson, J.A. Segre, C. Traidl-Hoffmann, Performing skin microbiome research: a method to the madness, *J. Invest. Dermatol.* 137 (2017) 561–568.
- [26] S. Verbanic, C.Y. Kim, J.M. Deacon, I.A. Chen, Improved single-swab sample preparation for recovering bacterial and phage DNA from human skin and wound microbiomes, *BMC Microbiol.* 19 (2019) 1–13.
- [27] H. Niehues, J.A. Bouwstra, A. El Ghalbzouri, J.M. Brandner, P.L. Zeeuwen, E.H. Van Den Bogaard, 3D skin models for 3R research: the potential of 3D reconstructed skin models to study skin barrier function, *Exp. Dermatol.* 27 (2018) 501–511.
- [28] D. Pinto, T. Ciardiello, M. Franzoni, F. Pasini, G. Giuliani, F. Rinaldi, Effect of commonly used cosmetic preservatives on skin resident microflora dynamics, *Sci. Rep.* 11 (2021) 8695.
- [29] T. Coenye, H.J. Nelis, In vitro and in vivo model systems to study microbial biofilm formation, *J. Microbiol. Methods* 83 (2010) 89–105.
- [30] A.V. Gannesen, E.L. Zdorovenko, E.A. Botchkova, J. Hardouin, S. Massier, D.S. Kopsitsyn, M.V. Gorbachevskii, A.A. Kadykova, A.S. Shashkov, M.V. Zhurina, Composition of the biofilm matrix of *Cutibacterium acnes* acneic strain RT5, *Front. Microbiol.* 10 (2019) 1284.
- [31] M.C. Cao, T.T. Feng, X.C. Zhang, Investigation on preservatives use in commercial cosmetics, *J Environ Hyg* 7 (2017) 296–300.
- [32] D. Pinto, T. Ciardiello, M. Franzoni, F. Pasini, G. Giuliani, F. Rinaldi, Effect of commonly used cosmetic preservatives on skin resident microflora dynamics, *Sci. Rep.* 11 (2021) 8695.
- [33] J.-J. Jeong, D.-H. Kim, Effects of cosmetics and their preservatives on the growth and composition of human skin microbiota, *Journal of the Society of Cosmetic Scientists of Korea* 41 (2015) 127–134.
- [34] S.A. Nasrollahi, M. Fattahi, A. Khamesipour, F. Amiri, M. Ahmadi, M.S. Kavkani, E. Lotfali, A. Ayatollahi, S.E. Skandari, A. Firooz, Effects of cosmetic preservatives on healthy facial skin microflora, *J. Clin. Aesthet. Dermatol.* 15 (2022) 34.
- [35] N. Halla, I.P. Fernandes, S.A. Heleno, P. Costa, Z. Boucherit-Otmani, K. Boucherit, A.E. Rodrigues, I.C. Ferreira, M.F. Barreiro, Cosmetics preservation: a review on present strategies, *Molecules* 23 (2018) 1571.
- [36] Q. Wang, S. Cui, L. Zhou, K. He, L. Song, H. Liang, C. He, Effect of cosmetic chemical preservatives on resident flora isolated from healthy facial skin, *J. Cosmet. Dermatol.* 18 (2019) 652–658.
- [37] N. Fierer, M. Hamady, C.L. Lauber, R. Knight, The Influence of Sex, Handedness, and Washing on the Diversity of Hand Surface Bacteria, vol. 105, *Proceedings of the National Academy of Sciences*, 2008, pp. 17994–17999.
- [38] A.A. Nash, R.G. Dalziel, J.R. Fitzgerald, Attachment to and entry of microorganisms into the body. *Mims' Pathogenesis of Infectious Disease*, Elsevier, Amsterdam, The Netherlands, 2015, pp. 9–49.
- [39] E. Larson, Skin hygiene and infection prevention: more of the same or different approaches? *Clin. Infect. Dis.* 29 (1999) 1287–1294.
- [40] M. Rosenthal, A. Aiello, E. Larson, C. Chenoweth, B. Foxman, Healthcare workers' hand microbiome may mediate carriage of hospital pathogens, *Pathogens* 3 (2014) 1–13.
- [41] A.J. SanMiguel, J.S. Meisel, J. Horwinski, Q. Zheng, C.W. Bradley, E.A. Grice, Antiseptic agents elicit short-term, personalized, and body site-specific shifts in resident skin bacterial communities, *J. Invest. Dermatol.* 138 (2018) 2234–2243.

- [42] C. Zapka, J. Leff, J. Henley, J. Tittl, E. De Nardo, M. Butler, R. Griggs, N. Fierer, S. Edmonds-Wilson, Comparison of standard culture-based method to culture-independent method for evaluation of hygiene effects on the hand microbiome, *mBio* 8 (2017), <https://doi.org/10.1128/mbio.00093-17>.
- [43] A.M. Two, T. Nakatsuji, P.F. Kotel, E. Arvanitidou, L. Du-Thumm, T.R. Hata, R.L. Gallo, The cutaneous microbiome and aspects of skin antimicrobial defense system resist acute treatment with topical skin cleansers, *J. Invest. Dermatol.* 136 (2016) 1950–1954.
- [44] S. Ameri, S. Ahmad Nasrollahi, A. Samadi, F. Amiri, S. Ahmadvand, S. Yadangi, M. Fattahi, M. Ehsani, A. Firooz, Assessment of skin microbiota and biometric parameters: a comprehensive comparison of four types of hand cleansers, *Iranian Journal of Dermatology* 24 (2021) 306–314.
- [45] F. Ripplke, E. Berardesca, T.M. Weber, pH and microbial infections, pH of the Skin: Issues and Challenges, vol. 54, 2018, pp. 87–94.
- [46] H.C. Kortling, K. Hübner, K. Greiner, G. Hamm, O. Braun-Falco, Differences in the skin surface pH and bacterial microflora due to the long-term application of synthetic detergent preparations of pH 5.5 and pH 7.0. Results of a crossover trial in healthy volunteers, *Acta Derm. Venereol.* 70 (1990) 429–431.
- [47] S. Gervason, I. Metton, E. Gemrot, E. Ranouille, G. Skorski, M. Cabannes, J.-Y. Berthon, E. Filaire, *Rhodomyrtus tomentosa* fruit extract and skin microbiota: a focus on *C. acnes* phylotypes in acne subjects, *Cosmet. Toilet.* 7 (2020) 53.
- [48] J.-H. Lee, S.-H. Eom, E.-H. Lee, Y.-J. Jung, H.-J. Kim, M.-R. Jo, K.-T. Son, H.-J. Lee, J.H. Kim, M.-S. Lee, In vitro antibacterial and synergistic effect of phlorotannins isolated from edible brown seaweed *Eisenia bicyclis* against acne-related bacteria, *ALGAE* 29 (2014) 47–55.
- [49] H. Fukase, Percutaneous Absorption Study of Kojic Acid in Humans, CPC Clinic, Medical Facility, 2005. Kagoshima, Japan.
- [50] H.E. Wollny, *Salmonella typhimurium* and *Escherichia coli* Reverse Mutation Assay with Kojic Acid. RCC-CCR Project Number 612701, Unpublished Data, 1998.
- [51] Z. Petkovsek, K. Elersic, M. Gubina, D. Zgur-Bertok, M. Starcic Erjavec, Virulence potential of *Escherichia coli* isolates from skin and soft tissue infections, *J. Clin. Microbiol.* 47 (2009) 1811–1817.
- [52] M.J. Blaser, S. Falkow, What are the consequences of the disappearing human microbiota? *Nat. Rev. Microbiol.* 7 (2009) 887–894.
- [53] C.L. Burnett, W.F. Bergfeld, D.V. Belsito, C.D. Klaassen, J.G. Marks, R.C. Shank, T.J. Slaga, P.W. Snyder, F.A. Andersen, Final report of the safety assessment of methylisothiazolinone, *Int. J. Toxicol.* 29 (2010) 1875–2135.
- [54] M.A.R. Scherrer, V.B. Rocha, A.R.C. Andrade, Contact dermatitis to methylisothiazolinone, *An. Bras. Dermatol.* 90 (2015) 912–914.
- [55] M. Fournière, T. Latire, D. Souak, M.G.J. Feuilloley, G. Bedoux, *Staphylococcus epidermidis* and *Cutibacterium acnes*: two major sentinels of skin microbiota and the influence of cosmetics, *Microorganisms* 8 (2020) 1752.
- [56] M. Šikić Pogačar, U. Maver, N. Marcun Varda, D. Mičetić-Turk, Diagnosis and management of diaper dermatitis in infants with emphasis on skin microbiota in the diaper area, *Int. J. Dermatol.* 57 (2018) 265–275.
- [57] C. Wallen-Russell, The role of everyday cosmetics in altering the skin microbiome: a study using biodiversity, *Cosmet. Toilet.* 6 (2018) 2.
- [58] M. Rademacher, M.-K. Zinn, R. Beinho, D.P. Bockmühl, A new model to investigate the effects of cosmetics on skin microorganisms in vitro, *Cosmet. Toilet.* 9 (2022) 88.
- [59] T. Ciardiello, D. Pinto, L. Marotta, G. Giuliani, F. Rinaldi, Effects of fermented oils on alpha-biodiversity and relative abundance of cheek resident skin microbiota, *Cosmet. Toilet.* 7 (2020) 34.
- [60] E. Filaire, C. Vialleix, J.-P. Cadoret, S. Guénard, C. Muller, A. Dreux-Zigha, J.-Y. Berthon, Characterization of reactive and sensitive skin microbiota: effect of *Halymenia durvillei* (HD) extract treatment, *Cosmet. Toilet.* 6 (2019) 69.
- [61] W.H. McCoy, E. Otchere, B.A. Rosa, J. Martin, C.M. Mann, M. Mitreva, Skin ecology during sebaceous drought—how skin microbes respond to isotretinoin, *J. Invest. Dermatol.* 139 (2019) 732.
- [62] R. Friso, J. Claypool, Microbial reference frames reveal distinct shifts in the skin microbiota after cleansing, *Microorganisms* 8 (2020) 1634.
- [63] H.J. Lee, S.E. Jeong, S. Lee, S. Kim, H. Han, C.O. Jeon, Effects of cosmetics on the skin microbiome of facial cheeks with different hydration levels, *Microbiologyopen* 7 (2018) e00557.
- [64] B. Murphy, S. Grimshaw, M. Hoptroff, S. Paterson, D. Arnold, A. Cawley, S.E. Adams, F. Falciani, T. Dadd, R. Eccles, Alteration of barrier properties, stratum corneum ceramides and microbiome composition in response to lotion application on cosmetic dry skin, *Sci. Rep.* 12 (2022) 5223.
- [65] C. Le Bourgot, C. Meunier, E. Gao, V. Murat, M. Micheletto, E. Tedesco, F. Benetti, Effects of short chain fructo-oligosaccharides on selected skin bacteria, *Sci. Rep.* 12 (2022) 9702.
- [66] G. Kim, M. Kim, M. Kim, C. Park, Y. Yoon, D.-H. Lim, H. Yeo, S. Kang, Y.-G. Lee, N.-I. Beak, Spermidine-induced recovery of human dermal structure and barrier function by skin microbiome, *Commun. Biol.* 4 (2021) 231.
- [67] B. Howard, C.C. Bascom, P. Hu, R.L. Binder, G. Fadayel, T.G. Huggins, B.B. Jarrold, R. Osborne, H.L. Rocchetta, D. Swift, Aging-associated changes in the adult human skin microbiome and the host factors that affect skin microbiome composition, *J. Invest. Dermatol.* 142 (2022) 1934–1946. e21.
- [68] K.-B. Hong, Y.H. Hong, E.Y. Jung, K. Jo, H.J. Suh, Changes in the diversity of human skin microbiota to cosmetic serum containing prebiotics: results from a randomized controlled trial, *J. Personalized Med.* 10 (2020) 91.
- [69] K. Kaur, G. Rath, Formulation and evaluation of UV protective synbiotic skin care topical formulation, *J. Cosmet. Laser Ther.* 21 (2019) 332–342.
- [70] M. Stolzoff, S. Wang, T. Webster, Efficacy and Mechanism of Selenium Nanoparticles as Antibacterial Agents, 2016.
- [71] M. Unno, O. Cho, T. Sugita, Inhibition of Propionibacterium acnes lipase activity by the antifungal agent ketoconazole, *Microbiol. Immunol.* 61 (2017) 42–44.
- [72] C. Holland, T.N. Mak, U. Zimny-Arndt, M. Schmid, T.F. Meyer, P.R. Jungblut, H. Brüggemann, Proteomic identification of secreted proteins of *Propionibacterium acnes*, *BMC Microbiol.* 10 (2010) 1–11.
- [73] N. Weber, K. Schwabe, C.M. Schempp, U. Wölflle, Effect of a botanical cleansing lotion on skin sebum and erythema of the face: a randomized controlled blinded half-side comparison, *J. Cosmet. Dermatol.* 18 (2019) 821–826.
- [74] T. Staudinger, A. Pipal, B. Redl, Molecular analysis of the prevalent microbiota of human male and female forehead skin compared to forearm skin and the influence of make-up, *J. Appl. Microbiol.* 110 (2011) 1381–1389.
- [75] Y. Jin, J. Xia, Z. Pan, J. Yang, W. Wang, Z. Fu, Polystyrene microplastics induce microbiota dysbiosis and inflammation in the gut of adult zebrafish, *Environ. Pollut.* 235 (2018) 322–329.
- [76] L.W.D. van Raamsdonk, M. van der Zande, A.A. Koelmans, R.L. Hoogenboom, R.J.B. Peters, M.J. Groot, A.A. Peijnenburg, Y.J.A. Weesepoel, Current insights into monitoring, bioaccumulation, and potential health effects of microplastics present in the food chain, *Foods* 9 (2020) 72.
- [77] J.-J. Guo, X.-P. Huang, L. Xiang, Y.-Z. Wang, Y.-W. Li, H. Li, Q.-Y. Cai, C.-H. Mo, M.-H. Wong, Source, migration and toxicology of microplastics in soil, *Environ. Int.* 137 (2020) 105263.
- [78] H. Wei, L. Wu, Z. Liu, M. Saleem, X. Chen, J. Xie, J. Zhang, Meta-analysis reveals differential impacts of microplastics on soil biota, *Ecotoxicol. Environ. Saf.* 230 (2022) 113150.
- [79] P.D. Dissanayake, S. Kim, B. Sarkar, P. Oleszczuk, M.K. Sang, M.N. Haque, J.H. Ahn, M.S. Bank, Y.S. Ok, Effects of microplastics on the terrestrial environment: a critical review, *Environ. Res.* 209 (2022) 112734.
- [80] K. Kaur, S. Reddy, P. Barathe, U. Oak, V. Shriram, S.S. Kharat, M. Govarthanan, V. Kumar, Microplastic-associated pathogens and antimicrobial resistance in environment, *Chemosphere* 291 (2022) 133005.
- [81] T.S.M. Amelia, W.M.A.W.M. Khalik, M.C. Ong, Y.T. Shao, H.-J. Pan, K. Bhubalan, Marine microplastics as vectors of major ocean pollutants and its hazards to the marine ecosystem and humans, *Prog. Earth Planet. Sci.* 8 (2021) 1–26.
- [82] K.S. Stenger, O.G. Wikmark, C.C. Bezuidenhout, L.G. Molale-Tom, Microplastics pollution in the ocean: potential carrier of resistant bacteria and resistance genes, *Environ. Pollut.* 291 (2021) 118130.
- [83] L. Lu, Z. Wan, T. Luo, Z. Fu, Y. Jin, Polystyrene microplastics induce gut microbiota dysbiosis and hepatic lipid metabolism disorder in mice, *Sci. Total Environ.* 631 (2018) 449–458.
- [84] R. Qiao, Y. Deng, S. Zhang, M.B. Wolosker, Q. Zhu, H. Ren, Y. Zhang, Accumulation of different shapes of microplastics initiates intestinal injury and gut microbiota dysbiosis in the gut of zebrafish, *Chemosphere* 236 (2019) 124334.
- [85] W. Gu, S. Liu, L. Chen, Y. Liu, C. Gu, H.-Q. Ren, B. Wu, Single-cell RNA sequencing reveals size-dependent effects of polystyrene microplastics on immune and secretory cell populations from zebrafish intestines, *Environ. Sci. Technol.* 54 (2020) 3417–3427.

- [86] Z. Liu, P. Yu, M. Cai, D. Wu, M. Zhang, M. Chen, Y. Zhao, Effects of microplastics on the innate immunity and intestinal microflora of juvenile *Eriocheir sinensis*, *Sci. Total Environ.* 685 (2019) 836–846.
- [87] E. Fournier, J. Ratel, S. Denis, M. Leveque, P. Ruiz, C. Mazal, F. Amiard, M. Edely, V. Bezirard, E. Gaultier, Exposure to polyethylene microplastics alters immature gut microbiome in an infant in vitro gut model, *J. Hazard Mater.* 443 (2023) 130383.
- [88] Y. Zhao, Z. Qin, Z. Huang, Z. Bao, T. Luo, Y. Jin, Effects of polyethylene microplastics on the microbiome and metabolism in larval zebrafish, *Environ. Pollut.* 282 (2021) 117039.
- [89] A. Tamargo, N. Molinero, J.J. Reinoso, V. Alcolea-Rodríguez, R. Portela, M.A. Bañares, J.F. Fernández, M.V. Moreno-Arribas, PET microplastics affect human gut microbiota communities during simulated gastrointestinal digestion, first evidence of plausible polymer biodegradation during human digestion, *Sci. Rep.* 12 (2022) 528.
- [90] D. Montero, S. Rimoldi, S. Torrecillas, J. Rapp, F. Moroni, A. Herrera, M. Gómez, Á. Fernández-Montero, G. Terova, Impact of polypropylene microplastics and chemical pollutants on European sea bass (*Dicentrarchus labrax*) gut microbiota and health, *Sci. Total Environ.* 805 (2022) 150402.
- [91] S.M. Bashir, S. Kimiko, C.-W. Mak, J.K.-H. Fang, D. Gonçalves, Personal care and cosmetic products as a potential source of environmental contamination by microplastics in a densely populated Asian city, *Front. Mar. Sci.* 8 (2021) 683482.
- [92] G.E. Flores, J.G. Caporaso, J.B. Henley, J.R. Rideout, D. Domogala, J. Chase, J.W. Leff, Y. Vázquez-Baeza, A. Gonzalez, R. Knight, Temporal variability is a personalized feature of the human microbiome, *Genome Biol.* 15 (2014) 1–13.
- [93] J. Oh, A.L. Byrd, M. Park, H.H. Kong, J.A. Segre, Temporal stability of the human skin microbiome, *Cell* 165 (2016) 854–866.
- [94] C. Dessinoti, A.D. Katsambas, The role of Propionibacterium acnes in acne pathogenesis: facts and controversies, *Clin. Dermatol.* 28 (2010) 2–7.
- [95] M.M. Brown, A.R. Horswill, Staphylococcus epidermidis—skin friend or foe? *PLoS Pathog.* 16 (2020) e1009026.
- [96] J.-P. Claudel, N. Auffret, M.-T. Leccia, F. Poli, S. Corvec, B. Dréno, Staphylococcus epidermidis: a potential new player in the physiopathology of acne? *Dermatology* 235 (2019) 287–294.
- [97] D. McDonald, G. Ackermann, L. Khalilova, C. Baird, D. Heyland, R. Kozar, M. Lemieux, K. Derenski, J. King, C. Vis-Kampen, Extreme dysbiosis of the microbiome in critical illness, *mSphere* 1 (2016), <https://doi.org/10.1128/msphere.00199-16>.
- [98] B. Dreno, R. Martin, D. Moyal, J.B. Henley, A. Khammari, S. Seité, Skin microbiome and acne vulgaris: Staphylococcus, a new actor in acne, *Exp. Dermatol.* 26 (2017) 798–803.
- [99] A. Balato, S. Cacciapuoti, R. Di Caprio, C. Marasca, A. Masarà, A. Raimondo, G. Fabbrocini, Human microbiome: composition and role in inflammatory skin diseases, *Arch. Immunol. Ther. Exp.* 67 (2019) 1–18.
- [100] C.C. Coughlin, S.M. Swink, J. Horwinski, G. Sfyroera, J. Bugayev, E.A. Grice, A.C. Yan, The preadolescent acne microbiome: a prospective, randomized, pilot study investigating characterization and effects of acne therapy, *Pediatr. Dermatol.* 34 (2017) 661–664.
- [101] Y. Wang, S. Kuo, M. Shu, J. Yu, S. Huang, A. Dai, A. Two, R.L. Gallo, C.-M. Huang, Staphylococcus epidermidis in the human skin microbiome mediates fermentation to inhibit the growth of Propionibacterium acnes: implications of probiotics in acne vulgaris, *Appl. Microbiol. Biotechnol.* 98 (2014) 411–424.
- [102] K. Findley, J. Oh, J. Yang, S. Conlan, C. Deming, J.A. Meyer, D. Schoenfeld, E. Nomicos, M. Park, Topographic diversity of fungal and bacterial communities in human skin, *Nature* 498 (2013) 367–370.
- [103] E.A. Grice, The Skin Microbiome: Potential for Novel Diagnostic and Therapeutic Approaches to Cutaneous Disease, NIH Public Access, 2014.
- [104] K.H. Kim, Overview of atopic dermatitis, *Asia Pac. Allergy* 3 (2013) 79–87.
- [105] H.H. Kong, J. Oh, C. Deming, S. Conlan, E.A. Grice, M.A. Beatson, E. Nomicos, E.C. Polley, H.D. Komarow, P.R. Murray, Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis, *Genome Res.* 22 (2012) 850–859.
- [106] J.B. Henley, R.M.M. Ing, H. Zelenkova, Microbiome of affected and unaffected skin of patients with atopic dermatitis before and after emollient treatment, *J. Drugs Dermatol.* 13 (2014) 1365–1372.
- [107] M.E. Gonzalez, J.V. Schaffer, S.J. Orlow, Z. Gao, H. Li, A.V. Alekseyenko, M.J. Blaser, Cutaneous microbiome effects of fluticasone propionate cream and adjunctive bleach baths in childhood atopic dermatitis, *J. Am. Acad. Dermatol.* 75 (2016) 481–493. e8.
- [108] M. Yerushalmi, O. Elalouf, M. Anderson, V. Chandran, The skin microbiome in psoriatic disease: a systematic review and critical appraisal, *Journal of Translational Autoimmunity* 2 (2019) 100009.
- [109] A. Velegraki, C. Cafarchia, G. Gaitanis, R. Iatta, T. Boekhout, Malassezia infections in humans and animals: pathophysiology, detection, and treatment, *PLoS Pathog.* 11 (2015) e1004523.
- [110] M. Gardiner, M. Vicaretti, J. Sparks, S. Bansal, S. Bush, M. Liu, A. Darling, E. Harry, C.M. Burke, A longitudinal study of the diabetic skin and wound microbiome, *PeerJ* 5 (2017) e3543.
- [111] O. Tsilochristou, G. du Toit, P.H. Sayre, G. Roberts, K. Lawson, M.L. Sever, H.T. Bahnson, S. Radulovic, M. Basting, M. Plaut, Association of Staphylococcus aureus colonization with food allergy occurs independently of eczema severity, *J. Allergy Clin. Immunol.* 144 (2019) 494–503.
- [112] V.R. Salgado, A.T.L. de Queiroz, S.S. Sanabani, C.I. de Oliveira, E.M. Carvalho, J.M.L. Costa, M. Barral-Netto, A. Barral, The microbiological signature of human cutaneous leishmaniasis lesions exhibits restricted bacterial diversity compared to healthy skin, *Mem. Inst. Oswaldo Cruz* 111 (2016) 241–251.
- [113] N. Fyhrquist, L. Ruokolainen, A. Suomalainen, S. Lehtimäki, V. Veckman, J. Vendelin, P. Karisola, M. Lehto, T. Savinko, H. Jarva, Acinetobacter species in the skin microbiota protect against allergic sensitization and inflammation, *J. Allergy Clin. Immunol.* 134 (2014) 1301–1309. e11.
- [114] N.N. Schommer, R.L. Gallo, Structure and function of the human skin microbiome, *Trends Microbiol.* 21 (2013) 660–668.
- [115] A.K. Zaidi, K. Spaunhurst, D. Sprockett, Y. Thomason, M.W. Mann, P. Fu, C. Ammons, M. Gerstenblith, M.S. Tuttle, D.L. Popkin, Characterization of the facial microbiome in twins discordant for rosacea, *Exp. Dermatol.* 27 (2018) 295–298.
- [116] G.J.M. Christensen, H. Brüggemann, Bacterial skin commensals and their role as host guardians, *Benef. Microbes* 5 (2014) 201–215.
- [117] S.E. Gardner, S.L. Hillis, K. Heilmann, J.A. Segre, E.A. Grice, The neuropathic diabetic foot ulcer microbiome is associated with clinical factors, *Diabetes* 62 (2013) 923–930.
- [118] Y.R. Woo, S.H. Cho, J.D. Lee, H.S. Kim, The human microbiota and skin cancer, *Int. J. Mol. Sci.* 23 (2022) 1813.
- [119] J. Kullander, O. Forslund, J. Dillner, Staphylococcus aureus and squamous cell carcinoma of the skin, *Cancer Epidemiol. Biomarkers Prev.* 18 (2009) 472–478.
- [120] D.L.A. Wood, N. Lachner, J.-M. Tan, S. Tang, N. Angel, A. Laino, R. Linedale, K.-A. Lê Cao, M. Morrison, I.H. Frazer, A natural history of actinic keratosis and cutaneous squamous cell carcinoma microbiomes, *mBio* 9 (2018), <https://doi.org/10.1128/mbio.01432-18>.
- [121] N. Madhusudhan, M.R. Pausan, B. Halwachs, M. Durdević, M. Windisch, J. Kehrmann, V. Patra, P. Wolf, P. Boukamp, C. Moissl-Eichinger, Molecular profiling of keratinocyte skin tumors links Staphylococcus aureus overabundance and increased human β -defensin-2 expression to growth promotion of squamous cell carcinoma, *Cancers* 12 (2020) 541.
- [122] J.C.M. Jackow, J.C. Cather, V. Hearne, A.T. Asano, J.M. Musser, M. Duvic, Association of erythrodermic cutaneous T-cell lymphoma, superantigen-positive Staphylococcus aureus, and oligoclonal T-cell receptor V β gene expansion, *Blood, Am. J. Hematol.* 89 (1997) 32–40.
- [123] M. Glud, R. Gniadecki, MicroRNAs in the pathogenesis of malignant melanoma, *J. Eur. Acad. Dermatol. Venereol.* 27 (2013) 142–150.
- [124] S. Mizuhashi, I. Kajihara, S. Sawamura, H. Kanemura, K. Makino, J. Aoi, T. Makino, S. Masuguchi, S. Fukushima, H. Ihn, Skin microbiome in acral melanoma: Corynebacterium is associated with advanced melanoma, *J. Dermatol.* 48 (2021) e15–e16.
- [125] C. Zoschke, M. Ulrich, M. Sochorová, C. Wolff, K. Vávrová, N. Ma, C. Ulrich, J.M. Brandner, M. Schäfer-Korting, The barrier function of organotypic non-melanoma skin cancer models, *J. Contr. Release* 233 (2016) 10–18.
- [126] M.R. Williams, L. Cau, Y. Wang, D. Kaul, J.A. Sanford, L.S. Zaramela, S. Khalil, A.M. Butcher, K. Zengler, A.R. Horswill, Interplay of staphylococcal and host proteases promotes skin barrier disruption in Netherton syndrome, *Cell Rep.* 30 (2020) 2923–2933. e7.
- [127] R.S. Chapkin, L.A. Davidson, H. Park, U. Jin, Y. Fan, Y. Cheng, M.E. Hensel, K.K. Landrock, C. Allred, R. Menon, Role of the aryl hydrocarbon receptor (AhR) in mediating the effects of coffee in the colon, *Mol. Nutr. Food Res.* 65 (2021) 2100539.
- [128] P. Qiao, C. Zhang, J. Yu, S. Shao, J. Zhang, H. Fang, J. Chen, Y. Luo, D. Zhi, Q. Li, Quinolinic acid, a tryptophan metabolite of the skin microbiota, negatively regulates NLRP3 inflammasome through AhR in psoriasis, *J. Invest. Dermatol.* 142 (2022) 2184–2193. e6.

- [129] L.M. Gearhart-Serna, J.B. Davis, M.K. Jolly, N. Jayasundara, S.J. Sauer, R.T. Di Giulio, G.R. Devi, A polycyclic aromatic hydrocarbon-enriched environmental chemical mixture enhances AhR, antiapoptotic signaling and a proliferative phenotype in breast cancer cells, *Carcinogenesis* 41 (2020) 1648–1659.
- [130] S. Portal-Nuñez, U.T. Shankavaram, M. Rao, N. Datrice, S. Atay, M. Aparicio, K.A. Camphausen, P.M. Fernández-Salguero, H. Chang, P. Lin, Aryl hydrocarbon receptor-induced adrenomedullin mediates cigarette smoke carcinogenicity in humans and mice, *Cancer Res.* 72 (2012) 5790–5800.
- [131] M.J. Roth, W.-Q. Wei, J. Baer, C.C. Abnet, G.-Q. Wang, L.R. Sternberg, A.C. Warner, L.L. Johnson, N. Lu, C.A. Giffen, Aryl hydrocarbon receptor expression is associated with a family history of upper gastrointestinal tract cancer in a high-risk population exposed to aromatic hydrocarbons, *Cancer Epidemiol. Biomarkers Prev.* 18 (2009) 2391–2396.
- [132] G. Gaitanis, P. Magiatis, M. Hantschke, I.D. Bassukas, A. Velegraki, The *Malassezia* genus in skin and systemic diseases, *Clin. Microbiol. Rev.* 25 (2012) 106–141.
- [133] Y. Sato, T. Fujimura, K. Tanita, L. Chunbing, S. Matsushita, Y. Fujisawa, A. Otsuka, Y. Yamamoto, T. Hidaka, S. Aiba, *Malassezia*-derived aryl hydrocarbon receptor ligands enhance the CCL20/Th17/soluble CD163 pathogenic axis in extra-mammary Paget's disease, *Exp. Dermatol.* 28 (2019) 933–939.
- [134] L.-M. Chung, J.-A. Liang, C.-L. Lin, L.-M. Sun, C.-H. Kao, Cancer risk in patients with candidiasis: a nationwide population-based cohort study, *Oncotarget* 8 (2017) 63562.
- [135] M.E. Spurgeon, P.F. Lambert, Merkel cell polyomavirus: a newly discovered human virus with oncogenic potential, *Virology* 435 (2013) 118–130.