

Review **Sex and Gender Differences in Alzheimer's Disease: Genetic, Hormonal, and Inflammation Impacts**

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Abstract: Two-thirds of Americans with Alzheimer's disease are women, indicating a profound variance between the sexes. Variances exist between the sexes in the age and intensity of the presentation, cognitive deficits, neuroinflammatory factors, structural and functional brain changes, as well as psychosocial and cultural circumstances. Herein, we summarize the existing evidence for sexual dimorphism and present the available evidence for these distinctions. Understanding these complexities is critical to developing personalized interventions for the prevention, care, and treatment of Alzheimer's disease.

Keywords: sex differences; Alzheimer's disease; risk factors; personalized medicine; neuroinflammation

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1. Introduction

Current estimates indicate that 6.7 million individuals in the United States (US) are afflicted with Alzheimer's disease (AD) [\[1,](#page-12-0)[2\]](#page-12-1); two-thirds of the patients with AD are women [\[3\]](#page-12-2). As the population of the US ages, AD and other forms of dementia will become increasingly prevalent. If medical advancements are not produced to prevent, abate, or cure this devastating illness, projection estimates predict that 13.8 million individuals will be diagnosed with AD by 2060 [\[2\]](#page-12-1). In 2019, 121,499 deaths in the US were attributed to AD [\[2\]](#page-12-1). Among people aged 65 or older in the US, twelve percent of women and nine percent of men have AD [\[4\]](#page-12-3). This statistic comprises approximately 7 million people, consisting of 4.1 million women and 2.6 million men [\[3\]](#page-12-2). Clearly, sex influences the frequency of AD with greater prevalence in women than men. In this review, we summarize evidence detailing the profound differences in AD between females and males. We describe epidemiological, genetic, neuropathological, hormonal, intestinal (gut microbiota), preclinical, and sociocultural evidence with an emphasis on the role of neuroinflammation in sex differences in AD.

2. Epidemiological Evidence for Sexual Dimorphism in Alzheimer's Disease

The frequency of AD is greater in women than men in the US [\[3\]](#page-12-2). However, there are equivocal reports comparing the incidence and prevalence of AD between females and males when controlled for age [\[5](#page-12-4)[,6\]](#page-12-5). While the explanations for these variations are not clear, they suggest that differences in the incidence of AD between the sexes may be geographically and temporally dependent [\[7\]](#page-12-6).

One European study reported differential incidence rates of AD by sex in those older than 85; in males, incidence decreased, but in females, incidence increased [\[8\]](#page-12-7). Two Asian studies reported a similar, albeit non-significant, trend [\[9](#page-12-8)[,10\]](#page-12-9). In contrast, the Cognitive Function and Aging Study conducted in the United Kingdom (UK) found that males had a higher incidence rate [\[11\]](#page-12-10).

Alternatively, several American studies showed that the incidence of AD increased similarly with age in men and women [\[12](#page-12-11)[–15\]](#page-12-12). In addition, a global comprehensive metaanalysis found that the incidence and prevalence of AD was not statistically different between men and women beyond 60 years of age [\[16\]](#page-12-13).

In epidemiological studies, estimates of prevalence usually derive from cross-sectional surveys conducted in community, institutional, or clinical settings and are vulnerable to survivor and selection biases [\[17\]](#page-12-14). For example, the main risk factor for developing AD is age, and, on average, females live longer than males [\[18,](#page-12-15)[19\]](#page-12-16). Reporting a higher prevalence of AD in women may result from survival bias rather than sex-specific variables that increase the disease risk [\[13\]](#page-12-17). Also, this survival bias should be considered when assessing the incidence rate in men. For instance, males tend to have a higher mortality rate from cardiovascular disease in middle age compared to females [\[19\]](#page-12-16). Yet, males who survive beyond 65 have a healthier cardiovascular risk profile, subsequently reducing their risk of developing dementia [\[19\]](#page-12-16). Thus, the comparison of the AD risk is between exceptionally robust males and females of average health and longevity [\[20\]](#page-12-18). Nonetheless, although survival bias may be a factor, it does not fully explain the sex differences in AD prevalence and incidence [\[21\]](#page-13-0). Though it is important to take longevity into account, new research indicates that the higher incidence of AD in women may be due to other components as well [\[22\]](#page-13-1). Therefore, biological, social, and behavioral elements all have a role in the variations in brain alterations, AD development, and symptom presentation between the sexes [\[7,](#page-12-6)[23,](#page-13-2)[24\]](#page-13-3). The longevity concept ignores certain significant factors. First of all, there is now a difference of less than five years in the average life expectancy in the United States between males and females [\[25\]](#page-13-4). Research conducted in Europe indicates that by 2030, the difference in longevity will be less than two years due to the steady increase in male survival rates [\[26\]](#page-13-5). Second, statistical models have demonstrated that, even after controlling for gender-dependent mortality rates, age at death, and variations in longevity, women still have a twofold greater incidence and lifetime risk of AD [\[27,](#page-13-6)[28\]](#page-13-7). Furthermore, differences in the structure, function, and age-related changes to the brain in men and women are well-documented [\[27\]](#page-13-6). According to recent findings, women tend to accrue more tangle load than men do with the same brain $Aβ$ amounts, although there is no difference in the lifetime risk of AD [\[29\]](#page-13-8). This finding suggests that the pathogenesis of AD may start sooner in women.

3. Genetic Risk Factors Associated with Sexual Dimorphism in Alzheimer's Disease

Many differences in gene expression have been observed between aging men and women. Genes involved in energy generation are often down-regulated in men, whereas immune response genes are typically up-regulated in women [\[30\]](#page-13-9). The apolipoprotein ε 4 (APOEε4) allele is the most powerful genetic risk factor for Late Age Onset Alzheimer's Disease (LOAD) [\[31](#page-13-10)[,32\]](#page-13-11), while the APOEε2 allele is protective [\[32\]](#page-13-11). In mild cognitive impairment (MCI), the impact of APOE ε 4 on the risk of AD is sex- and zygosity-specific [\[33\]](#page-13-12). In patients aged 65–69 who are homozygous for APOEε4 alleles, females have worse memory in comparison to males, and at ages 70–74, worse global cognition [\[33\]](#page-13-12). The APOE ε 4 allele impairs brain structure and metabolism more in females than in males [\[34\]](#page-13-13). With respect to the female AD-related risk genes, bridging integrator 1 (BIN1) has gained recognition as the second most significant susceptibility gene associated with sporadic AD [\[35](#page-13-14)[,36\]](#page-13-15), and a greater risk of developing AD in females compared to males [\[37\]](#page-13-16). BIN1 affects AD pathogenesis through the tau pathway and is overexpressed in the brains of AD patients [\[38\]](#page-13-17).

Differential Gene Expression and Pathway Enrichment in Male and Female AD

To demonstrate the differences between male and female subjects with AD, we reanalyzed the reposited Mayo Clinic Alzheimer's Disease Genetics Studies (MCADGS) dataset. The raw count matrix for RNA-seq of the temporal cortex was acquired from the Synapse web portal, specifically from Project SynID: syn2580853. In total, the analyzed dataset comprised 119 subjects and was split into two subsets based on biological sex information: female (N_{CTL} = 36, N_{AD} = 28) and male (N_{CTL} = 39, N_{AD} = 16). Notably, there was a similar percentage of two allele APOε4 (about 13%) and one allele APOε4 (about 50%) in male versus female subjects in this database. Subsequent analyses were conducted separately for each subset: female AD vs. female CTL and male AD vs. male CTL. Missing patient medical information (PMI) records in the metadata were imputed with a fixed value (-1) , followed by conducting a variance partitioning analysis using the variancePartition R package v1.30.2 [\[39\]](#page-13-18) with the formula ∼ *Age* + *RNA integrity* (*RIN*) + *postmorteminterval* (*PMI*) + *Diagnosis*. The analysis aimed to identify and filter out genes that do not explain the variability between the phenotypes. A differential analysis was conducted using the DESeq2 R package v1.40.2 [\[40\]](#page-13-19), employing the same formula.

Figure [1](#page-3-0) showcases the top differentially expressed genes in the female and male datasets, along with a summary of the covariates. A pathway enrichment analysis was performed using GSEA (Gene Set Enrichment Analysis) with the fGSEA R package v1.26.0 [\[41\]](#page-13-20). A rank value of [−*Log*10(*p* − *value*) × *L*2*FC*] was calculated based on the p-value and L2FC (i.e.*, Log*2 $\left\{Fold\ Change{CTL\over AD}\right\})$ output from the differential analysis. A curated list of pathways was obtained from the Biological Systems Lab website [\(https://download.](https://download.baderlab.org/EM_Genesets/current_release/Human/symbol/) [baderlab.org/EM_Genesets/current_release/Human/symbol/](https://download.baderlab.org/EM_Genesets/current_release/Human/symbol/) (accessed on 31 July 2024, ID: Human_AllPathways). The analysis of the female dataset yielded 168 significantly (q < 0.05) up-regulated and 36 significantly down-regulated pathways. Among them, 128 up-regulated and 14 down-regulated pathways were uniquely enriched in the female dataset. In the male dataset, 46 up-regulated pathways and 25 down-regulated pathways were significantly enriched, with six and three pathways respectively exclusive to this comparison. Table [1](#page-4-0) shows the statistically significant pathways that are distinct between the two comparisons.

Next, we determined the level of association with AD for each pathway in the PubMed database. Utilizing the Bio.Entrez v 1.78 package for Python 3.10 (Python Software Foundation, Washington, DE, USA), the PubMed database was queried for the number of times that any given pathway was mentioned with Alzheimer's disease, and that numerator was divided by the number of times that the pathway was mentioned in any publication. This result was used as a proxy for how related the pathway was to Alzheimer's disease. We then quantified the pathway's relative impact, calculating the Z-score of each pathway against a null distribution of 1000 random pathways that were sourced from the Bader Lab Gene Ontology database with the same Alzheimer's-relatedness calculation being performed as above. Those Z-scores are represented in the provided table. Higher positive Z-scores indicate a strong association with the AD literature, while Z-scores near zero suggest that the pathway has little prior association with the published literature in the PubMed database.

Notably, most of the top differentially expressed genes, as well as the significantly upregulated and down-regulated pathways are different in male versus female subjects with AD (Figure [1](#page-3-0) and Table [1\)](#page-4-0). This striking contrast suggests markedly different molecular perturbations in male vs. female subjects, which is consistent with prior epidemiological, animal model, and pathophysiological work in AD. Some of the findings highlighted in this exploratory analysis included pathways that were previously associated with AD, including the complement system [\(https://doi.org/10.1016/j.neuron.2018.10.031\)](https://doi.org/10.1016/j.neuron.2018.10.031), which was up-regulated in male and female subjects, as well as interleukin-10 and interferon signaling (IL-10), which were only up-regulated in female AD patients (cite: [https://](https://doi.org/10.1016/S0531-5565(00)00176-5) [doi.org/10.1016/S0531-5565\(00\)00176-5](https://doi.org/10.1016/S0531-5565(00)00176-5) and [https://doi.org/10.3389/fncel.2022.949340\)](https://doi.org/10.3389/fncel.2022.949340)

(Table 1). Interestingly, some of the pathways were less associated with AD based on our sematic association with the published literature. Interleukin-4 signaling was only up-regulated in male AD, while syndecan-1 signaling was up-regulated only in female subjects (Table 1). Female, but not male, AD subjects had down-regulation of glutamatergic pathways, suggesting a more severe course with a greater loss of excitatory synapses in female AD patients (Table 1). Taken together, this exploratory bioinformatics assessment of male versus female RNAseq profiles from human AD subjects highlights differences that may only be appreciated when male and female subjects are considered independently.

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The left panel shows the female subjects' results (female AD vs. female CTL), and the male subjects are represented on the right. (A,B) show the volcano plots of $[-Log_{10} (BH adj. p-value)]$ vs. $[Log_2 Fold$ *Change* (L2FC)] for the DGE output. The significance thresholds are set to 0.05 for the adj. p-value *Change* (*L2FC*)] for the DGE output. The significance thresholds are set to 0.05 for the adj. p-value and ± 1 for the L2FC. Green-colored genes are significantly down-regulated, and violet represents significant up-regulation. The top 20 genes—regardless of directionality—based on the value of rank = [-Log₁₀ (adj. *p*-value) \times L2FC] are labelled. (C,D) list the top ten down- and up-regulated **Figure 1.** Differential gene expression (DGE) analysis of the sex-based stratified data is presented. genes. Lighter-colored symbols are shared between the male and female lists. (**E**,**F**) summarize the demographic data for the cohorts under study. * CEA16: CEACAM16-AS1 & SD: Subjects Demographics.

Table 1. RNAseq-derived gender-specific enriched pathways in temporal cortex from female and male AD subjects.

An analysis of the differential gene expression was conducted using the Mayo Clinic Alzheimer's Disease Genetics Studies (MCADGS) dataset. The raw count matrix for RNA-seq of the temporal cortex was acquired from the Synapse web portal, specifically from Project SynID: syn2580853. N = 119 total subjects: female (N_{CTL} = 36, N_{AD} = 28); male (N_{CTL} = 39, N_{AD} = 16). Subsequent analyses were conducted separately for each subset: female AD vs. female CTL and male AD vs. male CTL. We determined the level of association with AD for each pathway in the PubMed database. Utilizing the Bio.Entrez package for Python, the PubMed database was queried for the number of times that any given pathway was mentioned with Alzheimer's disease, and that numerator was divided by the number of times the pathway was mentioned in any publication. This result was used as a proxy for how related the pathway was to Alzheimer's disease. We then quantified the pathway's relative impact, calculating the Z-score of each pathway against a null distribution of 1000 random pathways that were sourced from the Bader Lab Gene Ontology database with the same Alzheimer's-relatedness calculation being performed as above. Higher positive Z-scores indicate a strong association with the AD literature, while Z-scores near zero suggest that the specified pathway has less prior association with the published AD literature in the PubMed database.

4. Differences in Neuropathology between Human Males and Females with AD

Females and males have different regional frequencies of neurofibrillary tangles (NFTs), particularly when age intervals are included [\[42\]](#page-13-21). Female APOEε4 carriers are more likely than male carriers to develop amyloid plaques and NFTs in the early stages of the disease [\[43\]](#page-13-22). Additionally, compared to males, females exhibited higher NFT counts in the

hippocampus. As such, "hippocampal-sparing-AD" was more frequently found in males, whereas "limbic-dominating-AD" was more frequently found in females [\[42\]](#page-13-21).

Neuroimaging studies also report that females show greater neuropathology and cognitive decline than males. Females show significantly greater hippocampal atrophy [\[44](#page-14-0)[–46\]](#page-14-1) and a lower brain volume compared to males [\[7\]](#page-12-6). Hence, it follows that those females with MCI or AD experience a more rapid decline in memory and functional capacity as well as hippocampal atrophy [\[45,](#page-14-2)[47\]](#page-14-3). Females exhibit a twofold decline in overall cognitive function relative to males [\[48\]](#page-14-4). This supports a strong association between the extent of the neuropathological burden and cognitive performance in AD, and females are more susceptible than males.

Differences between males and females suggest the involvement of sex hormones. Indeed, the rapid decline in blood levels of ovarian hormones in the mid-life of females can have a profound effect on cognition. It is widely acknowledged that higher levels of estradiol are linked to an improved cognitive performance [\[49\]](#page-14-5). In regard to AD, estrogens regulate Aβ and tau [\[50,](#page-14-6)[51\]](#page-14-7) and decrease neuronal susceptibility to apoptosis when exposed to $Aβ$, particularly in the hippocampus [\[52\]](#page-14-8).

5. The Role of Sex Hormones in the Development of AD

5.1. Ovarian Hormones in the Development of AD

The biochemical and cellular mechanisms of AD start decades prior to the development of clinical signs and symptoms, resulting in a 15- to 20-year prodromal, silent stage that starts during mid-life in both sexes [\[53\]](#page-14-9). Coincidentally, menopause in women begins at approximately the same stage of life as the prodromal stage of AD, around the age of 50. Approximately 20 years later is the average age of AD diagnosis, 70 [\[54](#page-14-10)[,55\]](#page-14-11). Neuroimaging studies associated the menopausal transition (MT) with the beginning of AD pathology in mid-life, suggesting that the MT is involved in the development of AD [\[24](#page-13-3)[,56,](#page-14-12)[57\]](#page-14-13). Elevated Aβ deposition, decreased glucose metabolism, and the loss of white and gray matter volume have been observed in postmenopausal and perimenopausal women, indicating that they have a higher AD burden than premenopausal women and age-matched males [\[58\]](#page-14-14). Further evidence for a hormonal role in the increased susceptibility to AD of women comes from a study that reported a 70 percent greater risk of developing AD after ovarian resection and consequent estrogen deprivation [\[59](#page-14-15)[–61\]](#page-14-16). These results suggest that changes in the circulating levels of ovarian hormones, estrogens in particular play an important role in making women more susceptible than men to AD.

Estrogen receptors have been found in many brain areas, and estrogens regulate a number of physiological mechanisms in the brain, including synaptic plasticity, neuroinflammation, brain macronutrient utilization, blood–brain barrier (BBB) integrity, and docosahexaenoic acid (DHA) metabolism [\[62–](#page-14-17)[65\]](#page-14-18). Possibly, women are more prone to AD because of the profound metabolic changes that take place after menopause. The association between the abrupt decline in estradiol and increased oxidative stress in the brain at the time of menopause may be the consequential factors resulting in initiation of the prodromal phase of AD [\[66](#page-14-19)[,67\]](#page-14-20). Female patients with AD, in comparison to age-matched controls, have reduced levels of circulating 17β-estradiol [\[68\]](#page-14-21).

Women who are peri- or postmenopausal may be prescribed estradiol, progesterone, or a combination, broadly known as hormone replacement therapy (HRT) to boost endogenous ovarian hormone levels to treat unpleasant vasomotor and menopausal symptoms like irritability, depression, perspiration, hot flashes, and urinary incontinence [\[69\]](#page-14-22). Epidemiological studies on women's health conducted in the 1990s revealed that women who received HRT experienced a lower incidence of AD than untreated women [\[70](#page-14-23)[–72\]](#page-15-0). However, some clinical studies found no beneficial effect of HRT on the development or treatment of AD [\[73](#page-15-1)[–75\]](#page-15-2). A recent meta-analysis showed negative effects of HRT on cognitive function in women above 60 years of age. Among the included studies, just two investigated women who were under 60 years old: in one of these, no effect of oral estrogens on cognition was reported, and in the other study, conjugated equine estrogen (CEE) and medroxyprogesterone acetate (MPA) had positive benefits on cognition [\[76\]](#page-15-3). Another study investigating the modulating effect of age and APOEε4 on the response to HRT [\[77\]](#page-15-4) found that HRT was associated with better delayed memory and greater entorhinal and amygdala volumes only in APOEε4 carriers [\[77\]](#page-15-4).

5.2. Testicular Hormones as a Risk Factor for Developing AD

Testosterone levels are reduced in male AD patients compared to age-matched healthy males [\[78](#page-15-5)[,79\]](#page-15-6), and lower levels of testosterone are associated with a greater risk of developing AD and poorer cognitive function in elderly males [\[80\]](#page-15-7). Another study showed that free testosterone may decrease the early pathogenic deposition of Aβ in women, as well as slowing the development of AD-specific hippocampal neurodegeneration in men [\[81\]](#page-15-8). Testosterone is aromatized into 17β-estradiol in the brain, or into the stronger androgen dihydrotestosterone (DHT) by the enzyme 5-reductase [\[82\]](#page-15-9). The actions of testosterone in the brain are amplified to an extent by 17β -estradiol, which activates estrogen receptors (ERs) [\[83\]](#page-15-10).

Notably, only men are susceptible to prostate cancer, and over 50 percent of prostate cancer patients are now being treated with androgen-deprivation therapy (ADT) following diagnosis [\[84\]](#page-15-11). However, one study [\[85\]](#page-15-12), contrary to others [\[86\]](#page-15-13), indicates that ADT may increase the risk of developing dementia and cognitive dysfunction.

6. Sex-Related Differences in the Microbiota–Gut–Brain Axis

The human gut is home to billions of microbes. A strong correlation between the gut microbiota and the onset and progression of numerous neurological conditions exists, functionally representing the microbiota–gut–brain axis [\[87](#page-15-14)[,88\]](#page-15-15). Gut dysbiosis is associated with neurodegenerative diseases such as Alzheimer's disease, Huntington's disease, and Parkinson's disease [\[89](#page-15-16)[–92\]](#page-15-17). Moreover, gut microbiota play a role in regulating cognitive functions like memory and learning [\[93\]](#page-15-18).

Alterations in the intestinal microbes also contribute to the underlying causes of AD, given that the composition and activity of intestinal microorganisms may impact the pathological processes of dementia- and age-related cognitive decline [\[94–](#page-15-19)[97\]](#page-16-0). Sex disparities in the composition of gut microbiota have been reported, mostly in animal research [\[98,](#page-16-1)[99\]](#page-16-2). Some human studies also indicate a strong sex difference in gut microbiota [\[100](#page-16-3)[–102\]](#page-16-4), while others have not observed any significant difference [\[103,](#page-16-5)[104\]](#page-16-6). Long term antibiotic administration in mouse models of AD, including APPSWE/PS1∆E9 and APPPS1-21, has resulted in a decrease in A β deposition only in male mice [\[92](#page-15-17)[,105,](#page-16-7)[106\]](#page-16-8). Treatment with antibiotics in the early stages of life causes sex-specific microbiome changes that are associated with decreased extracellular Aβ deposition, decreased levels of Aβ peptides that are insoluble in formic acid (FA), changes in the morphology of plaque-associated microglia, and $A\beta$ neurodegenerative features. These effects were particular to male APPPS1-21 mice [\[107\]](#page-16-9). Additionally, the fecal microbiota transplantation of APPPS1-21 male mice that were previously treated with antibiotics in their lifespan led to partial restoration of $A\beta$ accumulation, hence confirming causality [\[92\]](#page-15-17). Another study [\[108\]](#page-16-10) examined the sex difference in the impact of the oral administration of probiotics, antibiotics, and synbiotics in the App^{NL-G-F} mouse model of AD. Their results demonstrated different effects of the alterations to gut microbiota between males and females. Among the male and female App^{NL-G-F} models of AD that were given probiotics, only female subjects experienced a decrease in Aβ plaques, microgliosis, and brain TNF- α , as well as improved memory, as compared to control groups that did not receive any treatment [\[108\]](#page-16-10).

Explaining the contribution of gut microbiota to AD pathology, bacteria-derived amyloids may seep from the intestinal lumen and build up in the brain and systemic circulation [\[109\]](#page-16-11). Reactive oxygen species quantities may rise as a result, and nuclear factorκB (NF-κB) signaling may be activated, up-regulating the proinflammatory microRNA-34a (miRNA-34a). Consequently, miRNA-34a would suppress TREM2 expression (triggering receptors that are expressed in microglial/myeloid cells-2), impairing phagocytosis and

causing $\text{A}β 42$ peptide buildup [\[109](#page-16-11)[,110\]](#page-16-12). The leakiness of the gut can be further aggravated by bacterially produced lipopolysaccharide and amyloids, which can continue to raise the levels of cytokines and proinflammatory molecules that are strongly associated with AD, such as IL-17A and IL-22 [\[111](#page-16-13)[,112\]](#page-16-14). These cytokines can penetrate the gastrointestinal tract and then the blood–brain barrier to enter the brain. The cytokines may proceed to initiate immunogenic responses, the release of reactive oxygen species, and the signaling of NF-κB, CD14, and toll-like receptor 2/1, all of which contribute to neurodegeneration [\[111](#page-16-13)[,113\]](#page-16-15).

7. Mood, Cultural, and Social Factors Involved in Sex Differences in AD

Depression is a risk factor for AD dementia in both men and women [\[114\]](#page-16-16). Recent estimates suggest that a midlife depression diagnosis may lead to a 70 percent increased risk of developing AD [\[115\]](#page-16-17). Females are twice as likely as males to experience depression [\[116\]](#page-16-18). Because mood and memory are associated with the same brain regions, depression may impact late-life cognitive function. In the Women's Health Initiative Memory Study (WHIMS), clinically significant depressive symptoms were associated with a nearly twofold increased risk of MCI and dementia [\[117\]](#page-16-19). Given that females have a higher lifelong prevalence of depression, this might be associated with the increased AD frequency [\[6\]](#page-12-5). Women exhibited higher mean scores on the NeuroPsychiatric Inventory (NPI) for depression, anxiety, and overall total neuropsychiatric symptoms (NPS) in a study of individuals with newly diagnosed AD dementia who were not receiving treatment for AD or neuropsychiatric symptoms [\[118\]](#page-16-20). According to several additional research articles, women who have been diagnosed with AD dementia exhibit more depression symptoms than males. In contrast, agitation is more prevalent in men with AD dementia than in women [\[119\]](#page-16-21).

It has been reported that a lower level of education is associated with higher risk of dementia in men and women [\[120,](#page-17-0)[121\]](#page-17-1). Moreover, engaging in cognitive activities lowers an older person's chance of dementia [\[122](#page-17-2)[,123\]](#page-17-3). Over 10% of the variation in an individual's cognitive performance may be explained by their intellectual lifestyle, which includes their educational background, occupations, and level of daily cognitive engagement [\[124\]](#page-17-4).

Thus, females born in the first half of the 20th century may have a higher risk of developing dementia due to their lower educational attainment compared to males [\[7\]](#page-12-6). Aside from the direct impacts of education on the risk of dementia-related diseases that some studies have documented, women's historically lower level of education may also indirectly raise the risk of dementia-related diseases through elevated levels of distress and mental health issues [\[125\]](#page-17-5). Furthermore, a cohort study reported that sex is not associated with the incidence rate of AD when controlling for the educational level [\[126\]](#page-17-6).

Gender norms have shaped career chances historically, with a higher proportion of males than women in the workforce [\[127\]](#page-17-7). Furthermore, there has long been a gender gap in occupations, with women being more likely to work in unpaid labor activities like childcare and less likely to hold professional or management positions [\[128\]](#page-17-8). Professional or management employment has been associated with a 22% decrease in cognitive decline as well as a 44% reduction in MCI, according to a meta-analysis of nine prospective studies [\[129\]](#page-17-9).

Males tend to suffer from a more severe course of the disease, leading to early mortality [\[130](#page-17-10)[,131\]](#page-17-11). In fact, the male sex is a significant predictor of both an aggressive disease course and progression to death following an AD diagnosis [\[132\]](#page-17-12). While male mortality is related to disease factors, such as dementia severity and delirium frequency, female mortality is not. Instead, female mortality is associated with measures of disability, the inability to perform daily tasks, comorbidities, and the presence of pressure sores and malnutrition [\[130\]](#page-17-10). One proposed reason for these conditions is that females show resilience to tau pathology, possibly due to their increased immune response compared to males [\[20\]](#page-12-18).

8. Animal Studies Evaluating Sex Differences in AD

8.1. Sex Differences in Animal Models of AD

Thus far, we have summarized the evidence in human AD research related to sex differences in the susceptibility to, progression of, and outcome of AD. Sex differences in AD pathology have also been reported in animal models. In summary, female mice exhibit increased neuropathological markers of AD and poorer cognitive outcomes compared to male mice (Table [2\)](#page-11-0).

Several transgenic mouse models are designed to increase Aβ production and deposition [\[133\]](#page-17-13). Mutant forms of amyloid precursor protein (APP), presenilin-1 (PS-1), and PS-2 genes are commonly used in these mouse models to induce familial AD [\[133](#page-17-13)[–135\]](#page-17-14). Studies on APP/PS-1 mice reported higher Aβ levels [\[133](#page-17-13)[,136–](#page-17-15)[139\]](#page-17-16), poorer learning and memory [\[137](#page-17-17)[,140\]](#page-17-18), greater neurodegeneration [\[136\]](#page-17-15), and a more severe inflammatory microenvironment [\[136\]](#page-17-15) in female compared to male APP/PS-1 mice. Conversely, Li X. et al. observed more severely impaired glucose and insulin tolerance and higher cholesterol and triglyceride levels in male compared to female APP/PS-1 mice [\[137\]](#page-17-17). Meanwhile, Davis et al. studied the effects of X chromosomes on AD pathology in an APP mouse model and found that the second X chromosome reduces neurological deficits and mortality without affecting the levels of Aβ or other protein markers [\[141\]](#page-17-19). Furthermore, another AD mouse model, Tg2576, with Swedish mutant human βAPP, expresses excessive amounts of hβAPP and starts to form A β plaques at the age of 8- to 10-months [\[142](#page-17-20)[,143\]](#page-17-21). Female Tg2576 mice have higher Aβ levels, increased Aβ plaques, and poorer cognitive function than males [\[144](#page-17-22)[,145\]](#page-17-23).

A triple-transgenic familial mouse model, $3xTgAD$, carrying tau p_{301L} , PS1M146V, and APPSwe transgenes, was developed to explore the interplay between $A\beta$ and tau and their impact on synaptic function [\[146](#page-18-0)[,147\]](#page-18-1). This mouse model generates an ADlike pathology with $\text{A} \beta$ and tau accumulations. In human AD patients, $\text{A} \beta$ deposition starts in the cortical parts of the brain and subsequently spreads to the hippocampus. Contrastingly, tangle development often starts in the limbic brain region and later spreads to the cortical areas [\[148\]](#page-18-2). This is exactly the pattern of development seen in 3xTgAD mice [\[146\]](#page-18-0). Compared to male $3xTgAD$ mice, females showed increased $A\beta$ deposition and greater cognitive deficits in several studies [\[149](#page-18-3)[–153\]](#page-18-4). However, one study found comparable cognitive deficits in male and female 3xTgAD mice [\[154\]](#page-18-5).

It is widely accepted that the APOEε4 allele increases the chance of developing AD [\[31,](#page-13-10)[155\]](#page-18-6). Having the potential to be utilized as a possible treatment target for AD, many studies have investigated the connections between sex and APOEε4. Preclinical work suggests that the APOE ε 4 effects on cognition impairment are modulated by sex and age and are increased in aged female mice [\[156\]](#page-18-7). When studying mice that were carrying both genes of familial (3xTg) and sporadic (ApoE4) AD, hippocampal histology showed that female ApoE4/3xTg mice had elevated levels of Aβ proteins, β-site APP cleavage enzyme $(BACE1)$, and Sp1 (BACE1 transcription factor) in comparison to male ApoE4/3xTg, female 3xTg, and nonTg mice [\[157\]](#page-18-8). In addition, female ApoE4/3xTg mice showed a more severe AD pathology in the hippocampus and the earlier onset of spatial and memory impairment than male ApoE4/3xTg mice [\[157\]](#page-18-8).

Studies on EFAD mice, a mouse model of AD homozygousness for knock in ApoE gene and 5xFAD, also showed a sex-dependent effect of the ApoE4 allele [\[158](#page-18-9)[,159\]](#page-18-10). Female EFAD mice had a more severe AD pathology in terms of an increased plaque number and decreased plaque compaction due to lowered microglial interactions with Aβ deposits, an elevated level of soluble Aβ, and increased cerebral microbleeds and amyloid angiopathy [\[158](#page-18-9)[,159\]](#page-18-10). The ApoE4 allele altered lipid and amino acid metabolism throughout the brain in a sex-dependent manner [\[160\]](#page-18-11) and likely contributes to AD neuropathology through the impairment of energy, lipid, glucose, and amino acid metabolism in the synaptosomes' mitochondria [\[161\]](#page-18-12). In contrast, another study investigated ApoE4 and E3 male and female mice behaviors and found that only ApoE4/4 females recognized a new object in the novel object recognition (NOR) test [\[162\]](#page-18-13).

Genetic variations such as P301L and P301S have been found among AD patients with familial tauopathy, which may facilitate tau aggregation to produce paired helical filaments and NFTs [\[163](#page-18-14)[,164\]](#page-18-15). In comparison to male P301L-tg mice, female mice showed more body weight loss and a worse survival rate, strongly correlating with the accumulation of tau and p-tau in the regions of the brain which are most impacted by tauopathy [\[165\]](#page-18-16). Female mice with mutant APP and P301L had more NFT in all of the brain areas except the pons, compared to their male counterparts [\[166\]](#page-18-17), yet studies on P301S mutant mouse models indicated a more severe pathology in male mice compared to females [\[167,](#page-18-18)[168\]](#page-19-0).

Investigating sex differences in a mouse model of FTLD called the TAU58/2 line, which expresses the human 0N4R tau isoform via the P301S mutation, found that male TAU58/2 mice had more NFTs, higher soluble tau levels, and higher quantities of insoluble tau in hippocampal samples than female TAU58/2 mice [\[167\]](#page-18-18). Another recent study of the P301S mice model showed that male P301S animals, compared to female transgenic mice, lose weight more quickly and experience more severe dyskinesia and memory impairment. Male P301S mice showed distinct variations in a number of plasma variables, including as MIG, TNF-, IL-13, and IL-10, in comparison to female P301S mice, which was reasonable given the sex disparities in behavior and neuropathology [\[168\]](#page-19-0).

The 5XFAD mouse model was created in 2006 and expressed excess amounts of human PSEN1 protein with two Familial AD (FAD) mutations (M146L and L286V), as well as human APP with three FAD mutations: the Swedish (K670N, M671L), London (V7171), and Florida (I716V) mutations [\[169\]](#page-19-1). The mouse Thy1 promoter contains neuralspecific elements that control the expression of both of these transgenes, directing their overproduction only in brain neurons [\[170\]](#page-19-2). A recent study focusing on the genotype and sex difference in the 5XFAD mouse model [\[171\]](#page-19-3) found that females showed greater amounts of human APP and amyloid-β in addition to increased inflammation when compared to their male counterparts. Highlighting that these markers were associated with the hyperactivity that was seen in both sexes, female 5XFAD mice showed minor abnormalities in object and social exploration.

Regardless of their genotype, male mice expressed stress markers and neurotrophic factors more strongly than females, and these traits were associated with an improved cognitive function [\[171\]](#page-19-3). Other studies found poorer cognitive performance [\[172\]](#page-19-4), greater amyloid- β accumulation [\[173\]](#page-19-5), and a higher plaque burden [173] in female 5XFAD mice. On the other hand, Roddick et al. assessed olfactory-delayed matching-to-sample tasks in 5XFAD mice and observed that female mice performed better, indicating a better working memory [\[174\]](#page-19-6).

8.2. Sex Differences in Neuroinflammation Contributing to AD

Emerging evidence suggests that neuroinflammation has a pivotal role in AD onset and progression [\[175\]](#page-19-7). Currently, neuroinflammation is recognized as a main characteristic of AD. There is widespread agreement that inflammation starts at the very beginning of the AD pathology during the prodromal phase, and has both beneficial and detrimental effects [\[176\]](#page-19-8). Microglia, as the most prevalent of the phagocytic cells in the brain, can either enhance or inhibit AD progression. Early microglia activation is neuroprotective because the microglia remove soluble Λ β via phagocytosis, micropinocytosis, and Λ β-degrading enzyme-mediated proteolytic degradation [\[177–](#page-19-9)[179\]](#page-19-10). Nonetheless, the neuroinflammatory pathways in AD involve the activation and expansion of the microglia, which triggers the release of a range of inflammatory mediators [\[179\]](#page-19-10). In addition to directly damaging neurons, microglia-mediated neuroinflammation also promotes protein aggregation, which is one of the most notable characteristics of neurodegenerative diseases and plays a role in AD pathogenesis [\[178\]](#page-19-11). When A β , tau oligomers, and subsequent secretions activate the microglia, they draw in nearby microglia to speed up the active removal of misfolded protein aggregates and degenerated neuronal bodies. As the disease advances, the expression of many components linked to $A\beta$ clearance is down-regulated by the pro-inflammatory

cytokines that were generated in response to $A\beta$ aggregation, which encourages even more accumulation of Aβ and neurodegeneration [\[178\]](#page-19-11).

Most data about sex differences in the microglia come from rodent studies. Early in development, there are variations in the microglial density and shape associated with sex, and although these characteristics vary over the lifespan, some of the differences persist in the adult brain. It has been demonstrated that the amount of microglia in the fetal brains of male and female rats does not change at time points immediately before parturition [\[180\]](#page-19-12). But postnatal sex variations in the quantity and shape (and gene expression) of microglia start to show up, and these start to organize the rodent brain shortly after birth. At postnatal day 4 (P4), male rodents have a larger number of microglia than females in the parietal cortex, the amygdala, the CA1, CA3, and dentate gyrus (DG) areas of the hippocampus [\[180\]](#page-19-12). Although similar results have been observed in other studies [\[181\]](#page-19-13), there are studies that report a great number of microglia in the amygdala, hippocampus, and cortex of male rodents [\[182](#page-19-14)[–184\]](#page-19-15). In addition to the number of microglia, one study showed that there is a significant difference in brain aging in aged female mice compared to age-matched males in terms of microglial activation [\[185\]](#page-19-16). This sex difference was associated with an enhanced inflammatory environment in female mice compared to males through the increased expression of inflammatory genes, mostly related to microglia-specific transcripts, especially those involved in the complement system [\[185\]](#page-19-16). Moreover, microglia exhibit sex-specific migration rates, [\[184\]](#page-19-15) and interferon (IFN) γ regulates microglial mobility following an injury (microbleed) in only male rodents and not females [\[186\]](#page-19-17).

An investigation into the impact of sex and the APOE4 allele on cytokine production by mice' astrocytes showed that compared to APOE3, mixed-sex APOE4 primary mice' astrocytes have an elevated basal expression of many pro-inflammatory cytokines, such as IL-6, MCP-1, MIP-1α, TNF-α, IL-1β, and NOS2 [\[187\]](#page-19-18). In sex-specific cultures, an APOE4 female primary mouse's astrocytes had 1.5–2.5-fold higher levels of IL-6, IL-1β, and NOS2 than an APOE4 male's, and both were higher than an APOE3 primary mouse's astrocytes [\[187\]](#page-19-18).

Positron-emission tomography (PET) using the 18 kDa translocator protein (TSPO) has become popular over the past ten years as a method of evaluating microglial activity [\[188\]](#page-19-19). Studying female and male patients with AD showed a sex difference in cortical TSPO-PET signals, with a stronger increase in the TSPO-PET signal being observed in prodromal AD females as opposed to prodromal AD males [\[189\]](#page-19-20). Although this signal was not associated with $\text{A}β$ plaques, it could be related to tau accumulations [\[189\]](#page-19-20). Male brains' microglia in post-mortem tissue from AD patients had a uniformly ramified appearance, while the microglia in female brains showed a greatly varied morphology [\[190\]](#page-19-21). In addition, men showed a higher microglial density than women [\[190\]](#page-19-21).

Research conducted in vitro has demonstrated that the executive activities of activated microglia, such as phagocytosis [\[191\]](#page-19-22), and the release of inflammatory and toxic molecules, including nitric oxide (NO) and the tumor necrosis factor- α (TNF- α), are dependent on Ca^{2+} signaling [\[192\]](#page-19-23). Moreover, in vivo models showed that intracellular Ca^{2+} signaling pathways are particularly impaired in microglia that are associated with Amyloid β plaques [\[193\]](#page-19-24). Microglia in young brains rarely exhibit $Ca²⁺$ transients while at rest, yet they consistently react with fast Ca^{2+} signals when a nearby single neuron is damaged [\[194\]](#page-20-0). It has been shown that somatic Ca^{2+} transients become more common in microglia as they age and during the amyloid deposition process [\[193\]](#page-19-24). Another in vivo study showed that the percentage of active microglia in male mice altered very little through aging, in contrast to female mice, whose percentage of active cells nearly doubled throughout the same period [\[195\]](#page-20-1). These functional data indicate the "faster aging" of female microglia and are in line with recent single-cell transcriptome investigations into microglia [\[196\]](#page-20-2).

Table 2. Pre-clinical studies examining sex differences in murine models of Alzheimer's disease: ↓ means decreased, ↑ means increased, NOR: novel object recognition, BACE1: beta-site amyloid precursor protein cleaving enzyme, MWM: morris water maze, WT: wild-type, PnMS: pre-natal maternal stress, TTR: transthyretin, eIF2α: eukarytoic inititation factor 2 alpha, and NFT: neurofibrillary tangles.

9. Conclusions

While the underlying mechanisms of AD are not fully understood, differences between the sexes, genetic predispositions, hormones, and environmental factors may influence the onset and severity. Thus, it is essential to implement sex-specific approaches in AD research. Historically, the inclusion of females in preclinical studies and sex-specific analyses in clinical studies was limited, and females still experience exclusion from participating in studies in present times, thus causing a delay in the ability to address sex-specific issues. Future studies must consider the sex-specific mechanisms and outcomes of AD in both

males and females to approach their treatment and diagnosis more effectively. Such studies may then aid in bridging current knowledge gaps and could lead to improvements in AD diagnosis and treatments that benefit both sexes.

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References

- 1. 2020 Alzheimer's disease facts and figures. *Alzheimers Dement.* **2020**, *16*, 391–460. [\[CrossRef\]](https://doi.org/10.1002/alz.12068) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32157811)
- 2. 2023 Alzheimer's disease facts and figures. *Alzheimers Dement.* **2023**, *19*, 1598–1695. [\[CrossRef\]](https://doi.org/10.1002/alz.13016)
- 3. Rajan, K.B.; Weuve, J.; Barnes, L.L.; McAninch, E.A.; Wilson, R.S.; Evans, D.A. Population estimate of people with clinical Alzheimer's disease and mild cognitive impairment in the United States (2020–2060). *Alzheimers Dement.* **2021**, *17*, 1966–1975. [\[CrossRef\]](https://doi.org/10.1002/alz.12362) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34043283)
- 4. 2014 National Population Projections Datasets. Available online: [https://www.census.gov/data/datasets/2014/demo/popproj/](https://www.census.gov/data/datasets/2014/demo/popproj/2014-popproj.html) [2014-popproj.html](https://www.census.gov/data/datasets/2014/demo/popproj/2014-popproj.html) (accessed on 5 May 2023).
- 5. Rocca, W.A. Time, Sex, Gender, History, and Dementia. *Alzheimer Dis. Assoc. Disord.* **2017**, *31*, 76–79. [\[CrossRef\]](https://doi.org/10.1097/WAD.0000000000000187) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28169841)
- 6. Mielke, M.M. Sex and Gender Differences in Alzheimer's Disease Dementia. *Psychiatr. Times* **2018**, *35*, 14–17. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30820070)
- 7. Mielke, M.M.; Vemuri, P.; Rocca, W.A. Clinical epidemiology of Alzheimer's disease: Assessing sex and gender differences. *Clin. Epidemiol.* **2014**, *6*, 37–48. [\[CrossRef\]](https://doi.org/10.2147/CLEP.S37929) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24470773)
- 8. Fratiglioni, L.; Launer, L.J.; Andersen, K.; Breteler, M.M.; Copeland, J.R.; Dartigues, J.F.; Lobo, A.; Martinez-Lage, J.; Soininen, H.; Hofman, A. Incidence of dementia and major subtypes in Europe: A collaborative study of population-based cohorts. Neurologic Diseases in the Elderly Research Group. *Neurology* **2000**, *54*, S10–S15. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10854355)
- 9. Yoshitake, T.; Kiyohara, Y.; Kato, I.; Ohmura, T.; Iwamoto, H.; Nakayama, K.; Ohmori, S.; Nomiyama, K.; Kawano, H.; Ueda, K.; et al. Incidence and risk factors of vascular dementia and Alzheimer's disease in a defined elderly Japanese population: The Hisayama Study. *Neurology* **1995**, *45*, 1161–1168. [\[CrossRef\]](https://doi.org/10.1212/WNL.45.6.1161)
- 10. Liu, C.K.; Lai, C.L.; Tai, C.T.; Lin, R.T.; Yen, Y.Y.; Howng, S.L. Incidence and subtypes of dementia in southern Taiwan: Impact of socio-demographic factors. *Neurology* **1998**, *50*, 1572–1579. [\[CrossRef\]](https://doi.org/10.1212/WNL.50.6.1572)
- 11. Matthews, F.E.; Stephan, B.C.; Robinson, L.; Jagger, C.; Barnes, L.E.; Arthur, A.; Brayne, C. A two decade dementia incidence comparison from the Cognitive Function and Ageing Studies I and II. *Nat. Commun.* **2016**, *7*, 11398. [\[CrossRef\]](https://doi.org/10.1038/ncomms11398)
- 12. Kawas, C.; Gray, S.; Brookmeyer, R.; Fozard, J.; Zonderman, A. Age-specific incidence rates of Alzheimer's disease: The Baltimore Longitudinal Study of Aging. *Neurology* **2000**, *54*, 2072–2077. [\[CrossRef\]](https://doi.org/10.1212/WNL.54.11.2072) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10851365)
- 13. Hebert, L.E.; Scherr, P.A.; McCann, J.J.; Beckett, L.A.; Evans, D.A. Is the risk of developing Alzheimer's disease greater for women than for men? *Am. J. Epidemiol.* **2001**, *153*, 132–136. [\[CrossRef\]](https://doi.org/10.1093/aje/153.2.132) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11159157)
- 14. Fillenbaum, G.G.; Heyman, A.; Huber, M.S.; Woodbury, M.A.; Leiss, J.; Schmader, K.E.; Bohannon, A.; Trapp-Moen, B. The prevalence and 3-year incidence of dementia in older Black and White community residents. *J. Clin. Epidemiol.* **1998**, *51*, 587–595. [\[CrossRef\]](https://doi.org/10.1016/S0895-4356(98)00024-9)
- 15. Edland, S.D.; Rocca, W.A.; Petersen, R.C.; Cha, R.H.; Kokmen, E. Dementia and Alzheimer disease incidence rates do not vary by sex in Rochester, Minn. *Arch. Neurol.* **2002**, *59*, 1589–1593. [\[CrossRef\]](https://doi.org/10.1001/archneur.59.10.1589) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12374497)
- 16. Fiest, K.M.; Roberts, J.I.; Maxwell, C.J.; Hogan, D.B.; Smith, E.E.; Frolkis, A.; Cohen, A.; Kirk, A.; Pearson, D.; Pringsheim, T.; et al. The Prevalence and Incidence of Dementia Due to Alzheimer's Disease: A Systematic Review and Meta-Analysis. *Can. J. Neurol. Sci.* **2016**, *43* (Suppl. S1), S51–S82. [\[CrossRef\]](https://doi.org/10.1017/cjn.2016.36)
- 17. Brayne, C.; Gill, C.; Huppert, F.A.; Barkley, C.; Gehlhaar, E.; Girling, D.M.; O'Connor, D.W.; Paykel, E.S. Incidence of clinically diagnosed subtypes of dementia in an elderly population. Cambridge Project for Later Life. *Br. J. Psychiatry* **1995**, *167*, 255–262. [\[CrossRef\]](https://doi.org/10.1192/bjp.167.2.255)
- 18. Chêne, G.; Beiser, A.; Au, R.; Preis, S.R.; Wolf, P.A.; Dufouil, C.; Seshadri, S. Gender and incidence of dementia in the Framingham Heart Study from mid-adult life. *Alzheimers Dement.* **2015**, *11*, 310–320. [\[CrossRef\]](https://doi.org/10.1016/j.jalz.2013.10.005)
- 19. Seshadri, S.; Wolf, P.A.; Beiser, A.; Au, R.; McNulty, K.; White, R.; D'Agostino, R.B. Lifetime risk of dementia and Alzheimer's disease. The impact of mortality on risk estimates in the Framingham Study. *Neurology* **1997**, *49*, 1498–1504. [\[CrossRef\]](https://doi.org/10.1212/WNL.49.6.1498) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9409336)
- 20. Dubal, D.B. Sex difference in Alzheimer's disease: An updated, balanced and emerging perspective on differing vulnerabilities. *Handb. Clin. Neurol.* **2020**, *175*, 261–273. [\[CrossRef\]](https://doi.org/10.1016/b978-0-444-64123-6.00018-7)
- 21. Shaw, C.; Hayes-Larson, E.; Glymour, M.M.; Dufouil, C.; Hohman, T.J.; Whitmer, R.A.; Kobayashi, L.C.; Brookmeyer, R.; Mayeda, E.R. Evaluation of Selective Survival and Sex/Gender Differences in Dementia Incidence Using a Simulation Model. *JAMA Netw. Open* **2021**, *4*, e211001. [\[CrossRef\]](https://doi.org/10.1001/jamanetworkopen.2021.1001)
- 22. Rahman, A.; Jackson, H.; Hristov, H.; Isaacson, R.S.; Saif, N.; Shetty, T.; Etingin, O.; Henchcliffe, C.; Brinton, R.D.; Mosconi, L. Sex and Gender Driven Modifiers of Alzheimer's: The Role for Estrogenic Control Across Age, Race, Medical, and Lifestyle Risks. *Front. Aging Neurosci.* **2019**, *11*, 315. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2019.00315) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31803046)
- 23. Ferretti, M.T.; Iulita, M.F.; Cavedo, E.; Chiesa, P.A.; Schumacher Dimech, A.; Santuccione Chadha, A.; Baracchi, F.; Girouard, H.; Misoch, S.; Giacobini, E.; et al. Sex differences in Alzheimer disease—The gateway to precision medicine. *Nat. Rev. Neurol.* **2018**, *14*, 457–469. [\[CrossRef\]](https://doi.org/10.1038/s41582-018-0032-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29985474)
- 24. Scheyer, O.; Rahman, A.; Hristov, H.; Berkowitz, C.; Isaacson, R.S.; Diaz Brinton, R.; Mosconi, L. Female Sex and Alzheimer's Risk: The Menopause Connection. *J. Prev. Alzheimers Dis.* **2018**, *5*, 225–230. [\[CrossRef\]](https://doi.org/10.14283/jpad.2018.34) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30298180)
- 25. Riedel, B.C.; Thompson, P.M.; Brinton, R.D. Age, APOE and sex: Triad of risk of Alzheimer's disease. *J. Steroid Biochem. Mol. Biol.* **2016**, *160*, 134–147. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2016.03.012) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26969397)
- 26. Bennett, J.E.; Li, G.; Foreman, K.; Best, N.; Kontis, V.; Pearson, C.; Hambly, P.; Ezzati, M. The future of life expectancy and life expectancy inequalities in England and Wales: Bayesian spatiotemporal forecasting. *Lancet* **2015**, *386*, 163–170. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(15)60296-3)
- 27. Carter, C.L.; Resnick, E.M.; Mallampalli, M.; Kalbarczyk, A. Sex and Gender Differences in Alzheimer's Disease: Recommendations for Future Research. *J. Women's Health* **2012**, *21*, 1018–1023. [\[CrossRef\]](https://doi.org/10.1089/jwh.2012.3789) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22917473)
- 28. Vina, J.; Lloret, A. Why women have more Alzheimer's disease than men: Gender and mitochondrial toxicity of amyloid-β peptide. *J. Alzheimer's Dis.* **2010**, *20*, S527–S533. [\[CrossRef\]](https://doi.org/10.3233/JAD-2010-100501) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20442496)
- 29. Buckley, R.F.; Mormino, E.C.; Amariglio, R.E.; Properzi, M.J.; Rabin, J.S.; Lim, Y.Y.; Papp, K.V.; Jacobs, H.I.; Burnham, S.; Hanseeuw, B.J. Sex, amyloid, and APOE ε 4 and risk of cognitive decline in preclinical Alzheimer's disease: Findings from three well-characterized cohorts. *Alzheimers Dement.* **2018**, *14*, 1193–1203. [\[CrossRef\]](https://doi.org/10.1016/j.jalz.2018.04.010)
- 30. Berchtold, N.C.; Cribbs, D.H.; Coleman, P.D.; Rogers, J.; Head, E.; Kim, R.; Beach, T.; Miller, C.; Troncoso, J.; Trojanowski, J.Q.; et al. Gene expression changes in the course of normal brain aging are sexually dimorphic. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 15605–15610. [\[CrossRef\]](https://doi.org/10.1073/pnas.0806883105) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18832152)
- 31. Strittmatter, W.J.; Saunders, A.M.; Schmechel, D.; Pericak-Vance, M.; Enghild, J.; Salvesen, G.S.; Roses, A.D. Apolipoprotein E: High-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 1977–1981. [\[CrossRef\]](https://doi.org/10.1073/pnas.90.5.1977)
- 32. Corder, E.H.; Saunders, A.M.; Strittmatter, W.J.; Schmechel, D.E.; Gaskell, P.C.; Small, G.W.; Roses, A.D.; Haines, J.L.; Pericak-Vance, M.A. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **1993**, *261*, 921–923. [\[CrossRef\]](https://doi.org/10.1126/science.8346443) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8346443)
- 33. Hobel, Z.; Isenberg, A.L.; Raghupathy, D.; Mack, W.; Pa, J. APOEE4 Gene Dose and Sex Effects on Alzheimer's Disease MRI Biomarkers in Older Adults with Mild Cognitive Impairment. *J. Alzheimers Dis.* **2019**, *71*, 647–658. [\[CrossRef\]](https://doi.org/10.3233/JAD-180859)
- 34. Sampedro, F.; Vilaplana, E.; de Leon, M.J.; Alcolea, D.; Pegueroles, J.; Montal, V.; Carmona-Iragui, M.; Sala, I.; Sánchez-Saudinos, M.B.; Antón-Aguirre, S.; et al. APOE-by-sex interactions on brain structure and metabolism in healthy elderly controls. *Oncotarget* **2015**, *6*, 26663–26674. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.5185)
- 35. Lambert, J.C.; Zelenika, D.; Hiltunen, M.; Chouraki, V.; Combarros, O.; Bullido, M.J.; Tognoni, G.; Fiévet, N.; Boland, A.; Arosio, B.; et al. Evidence of the association of BIN1 and PICALM with the AD risk in contrasting European populations. *Neurobiol. Aging* **2011**, *32*, e711–e755. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2010.11.022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21220176)
- 36. Seshadri, S.; Fitzpatrick, A.L.; Ikram, M.A.; DeStefano, A.L.; Gudnason, V.; Boada, M.; Bis, J.C.; Smith, A.V.; Carassquillo, M.M.; Lambert, J.C.; et al. Genome-wide analysis of genetic loci associated with Alzheimer disease. *JAMA* **2010**, *303*, 1832–1840. [\[CrossRef\]](https://doi.org/10.1001/jama.2010.574) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20460622)
- 37. Fan, C.C.; Banks, S.J.; Thompson, W.K.; Chen, C.H.; McEvoy, L.K.; Tan, C.H.; Kukull, W.; Bennett, D.A.; Farrer, L.A.; Mayeux, R.; et al. Sex-dependent autosomal effects on clinical progression of Alzheimer's disease. *Brain* **2020**, *143*, 2272–2280. [\[CrossRef\]](https://doi.org/10.1093/brain/awaa164)
- 38. Chapuis, J.; Hansmannel, F.; Gistelinck, M.; Mounier, A.; Van Cauwenberghe, C.; Kolen, K.V.; Geller, F.; Sottejeau, Y.; Harold, D.; Dourlen, P.; et al. Increased expression of BIN1 mediates Alzheimer genetic risk by modulating tau pathology. *Mol. Psychiatry* **2013**, *18*, 1225–1234. [\[CrossRef\]](https://doi.org/10.1038/mp.2013.1)
- 39. Hoffman, G.E.; Schadt, E.E. variancePartition: Interpreting drivers of variation in complex gene expression studies. *BMC Bioinform.* **2016**, *17*, 483. [\[CrossRef\]](https://doi.org/10.1186/s12859-016-1323-z)
- 40. Love, M.I.; Huber, W.; Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.* **2014**, *15*, 550. [\[CrossRef\]](https://doi.org/10.1186/s13059-014-0550-8)
- 41. Korotkevich, G.; Sukhov, V.; Budin, N.; Shpak, B.; Artyomov, M.N.; Sergushichev, A. Fast gene set enrichment analysis. *bioRxiv* **2021**, 060012. [\[CrossRef\]](https://doi.org/10.1101/060012)
- 42. Liesinger, A.M.; Graff-Radford, N.R.; Duara, R.; Carter, R.E.; Hanna Al-Shaikh, F.S.; Koga, S.; Hinkle, K.M.; DiLello, S.K.; Johnson, M.F.; Aziz, A.; et al. Sex and age interact to determine clinicopathologic differences in Alzheimer's disease. *Acta Neuropathol.* **2018**, *136*, 873–885. [\[CrossRef\]](https://doi.org/10.1007/s00401-018-1908-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30219939)
- 43. Corder, E.H.; Ghebremedhin, E.; Taylor, M.G.; Thal, D.R.; Ohm, T.G.; Braak, H. The biphasic relationship between regional brain senile plaque and neurofibrillary tangle distributions: Modification by age, sex, and APOE polymorphism. *Ann. N. Y. Acad. Sci.* **2004**, *1019*, 24–28. [\[CrossRef\]](https://doi.org/10.1196/annals.1297.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15246987)
- 44. Ardekani, B.A.; Convit, A.; Bachman, A.H. Analysis of the MIRIAD Data Shows Sex Differences in Hippocampal Atrophy Progression. *J. Alzheimers Dis.* **2016**, *50*, 847–857. [\[CrossRef\]](https://doi.org/10.3233/JAD-150780)
- 45. Holland, D.; Desikan, R.S.; Dale, A.M.; McEvoy, L.K. Higher rates of decline for women and apolipoprotein E epsilon4 carriers. *AJNR Am. J. Neuroradiol.* **2013**, *34*, 2287–2293. [\[CrossRef\]](https://doi.org/10.3174/ajnr.A3601)
- 46. Hua, X.; Hibar, D.P.; Lee, S.; Toga, A.W.; Jack, C.R., Jr.; Weiner, M.W.; Thompson, P.M. Sex and age differences in atrophic rates: An ADNI study with n=1368 MRI scans. *Neurobiol. Aging* **2010**, *31*, 1463–1480. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2010.04.033) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20620666)
- 47. Koran, M.E.I.; Wagener, M.; Hohman, T.J. Sex differences in the association between AD biomarkers and cognitive decline. *Brain Imaging Behav.* **2017**, *11*, 205–213. [\[CrossRef\]](https://doi.org/10.1007/s11682-016-9523-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26843008)
- 48. Barnes, L.L.; Wilson, R.S.; Bienias, J.L.; Schneider, J.A.; Evans, D.A.; Bennett, D.A. Sex Differences in the Clinical Manifestations of Alzheimer Disease Pathology. *Arch. Gen. Psychiatry* **2005**, *62*, 685–691. [\[CrossRef\]](https://doi.org/10.1001/archpsyc.62.6.685) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15939846)
- 49. Luine, V.N. Estradiol and cognitive function: Past, present and future. *Horm. Behav.* **2014**, *66*, 602–618. [\[CrossRef\]](https://doi.org/10.1016/j.yhbeh.2014.08.011)
- 50. Anastasio, T.J. Exploring the contribution of estrogen to amyloid-Beta regulation: A novel multifactorial computational modeling approach. *Front. Pharmacol.* **2013**, *4*, 16. [\[CrossRef\]](https://doi.org/10.3389/fphar.2013.00016)
- 51. Alvarez-de-la-Rosa, M.; Silva, I.; Nilsen, J.; Pérez, M.M.; García-Segura, L.M.; Avila, J.; Naftolin, F. Estradiol prevents neural tau hyperphosphorylation characteristic of Alzheimer's disease. *Ann. N. Y. Acad. Sci.* **2005**, *1052*, 210–224. [\[CrossRef\]](https://doi.org/10.1196/annals.1347.016)
- 52. Nilsen, J.; Chen, S.; Irwin, R.W.; Iwamoto, S.; Brinton, R.D. Estrogen protects neuronal cells from amyloid beta-induced apoptosis via regulation of mitochondrial proteins and function. *BMC Neurosci.* **2006**, *7*, 74. [\[CrossRef\]](https://doi.org/10.1186/1471-2202-7-74) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17083736)
- 53. Sperling, R.; Mormino, E.; Johnson, K. The evolution of preclinical Alzheimer's disease: Implications for prevention trials. *Neuron* **2014**, *84*, 608–622. [\[CrossRef\]](https://doi.org/10.1016/j.neuron.2014.10.038)
- 54. Mosconi, L.; Brinton, R.D. How would we combat menopause as an Alzheimer's risk factor? *Expert Rev. Neurother.* **2018**, *18*, 689–691. [\[CrossRef\]](https://doi.org/10.1080/14737175.2018.1510320) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30091648)
- 55. Sperling, R.A.; Karlawish, J.; Johnson, K.A. Preclinical Alzheimer disease-the challenges ahead. *Nat. Rev. Neurol.* **2013**, *9*, 54–58. [\[CrossRef\]](https://doi.org/10.1038/nrneurol.2012.241)
- 56. Mosconi, L.; Berti, V.; Quinn, C.; McHugh, P.; Petrongolo, G.; Osorio, R.S.; Connaughty, C.; Pupi, A.; Vallabhajosula, S.; Isaacson, R.S.; et al. Perimenopause and emergence of an Alzheimer's bioenergetic phenotype in brain and periphery. *PLoS ONE* **2017**, *12*, e0185926. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0185926)
- 57. Mosconi, L.; Rahman, A.; Diaz, I.; Wu, X.; Scheyer, O.; Hristov, H.W.; Vallabhajosula, S.; Isaacson, R.S.; de Leon, M.J.; Brinton, R.D. Increased Alzheimer's risk during the menopause transition: A 3-year longitudinal brain imaging study. *PLoS ONE* **2018**, *13*, e0207885. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0207885) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30540774)
- 58. Mosconi, L.; Berti, V.; Quinn, C.; McHugh, P.; Petrongolo, G.; Varsavsky, I.; Osorio, R.S.; Pupi, A.; Vallabhajosula, S.; Isaacson, R.S.; et al. Sex differences in Alzheimer risk: Brain imaging of endocrine vs chronologic aging. *Neurology* **2017**, *89*, 1382–1390. [\[CrossRef\]](https://doi.org/10.1212/WNL.0000000000004425)
- 59. Rocca, W.A.; Bower, J.H.; Maraganore, D.M.; Ahlskog, J.E.; Grossardt, B.R.; de Andrade, M.; Melton, L.J., 3rd. Increased risk of cognitive impairment or dementia in women who underwent oophorectomy before menopause. *Neurology* **2007**, *69*, 1074–1083. [\[CrossRef\]](https://doi.org/10.1212/01.wnl.0000276984.19542.e6)
- 60. Rocca, W.A.; Grossardt, B.R.; Shuster, L.T. Oophorectomy, estrogen, and dementia: A 2014 update. *Mol. Cell. Endocrinol.* **2014**, *389*, 7–12. [\[CrossRef\]](https://doi.org/10.1016/j.mce.2014.01.020)
- 61. Phung, T.K.; Waltoft, B.L.; Laursen, T.M.; Settnes, A.; Kessing, L.V.; Mortensen, P.B.; Waldemar, G. Hysterectomy, oophorectomy and risk of dementia: A nationwide historical cohort study. *Dement. Geriatr. Cogn. Disord.* **2010**, *30*, 43–50. [\[CrossRef\]](https://doi.org/10.1159/000314681)
- 62. McCarthy, M.; Raval, A.P. The peri-menopause in a woman's life: A systemic inflammatory phase that enables later neurodegenerative disease. *J. Neuroinflammation* **2020**, *17*, 317. [\[CrossRef\]](https://doi.org/10.1186/s12974-020-01998-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33097048)
- 63. Zandi, P.P.; Carlson, M.C.; Plassman, B.L.; Welsh-Bohmer, K.A.; Mayer, L.S.; Steffens, D.C.; Breitner, J.C. Hormone replacement therapy and incidence of Alzheimer disease in older women: The Cache County Study. *JAMA* **2002**, *288*, 2123–2129. [\[CrossRef\]](https://doi.org/10.1001/jama.288.17.2123) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12413371)
- 64. Cui, J.; Shen, Y.; Li, R. Estrogen synthesis and signaling pathways during aging: From periphery to brain. *Trends Mol. Med.* **2013**, *19*, 197–209. [\[CrossRef\]](https://doi.org/10.1016/j.molmed.2012.12.007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23348042)
- 65. Henderson, V.W.; St John, J.A.; Hodis, H.N.; McCleary, C.A.; Stanczyk, F.Z.; Shoupe, D.; Kono, N.; Dustin, L.; Allayee, H.; Mack, W.J. Cognitive effects of estradiol after menopause: A randomized trial of the timing hypothesis. *Neurology* **2016**, *87*, 699–708. [\[CrossRef\]](https://doi.org/10.1212/WNL.0000000000002980) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27421538)
- 66. Li, R.; Singh, M. Sex differences in cognitive impairment and Alzheimer's disease. *Front. Neuroendocr.* **2014**, *35*, 385–403. [\[CrossRef\]](https://doi.org/10.1016/j.yfrne.2014.01.002)
- 67. Demetrius, L.A.; Eckert, A.; Grimm, A. Sex differences in Alzheimer's disease: Metabolic reprogramming and therapeutic intervention. *Trends Endocrinol. Metab.* **2021**, *32*, 963–979. [\[CrossRef\]](https://doi.org/10.1016/j.tem.2021.09.004)
- 68. Manly, J.J.; Merchant, C.A.; Jacobs, D.M.; Small, S.A.; Bell, K.; Ferin, M.; Mayeux, R. Endogenous estrogen levels and Alzheimer's disease among postmenopausal women. *Neurology* **2000**, *54*, 833–837. [\[CrossRef\]](https://doi.org/10.1212/WNL.54.4.833) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10690972)
- 69. Johnson, S.R. Menopause and hormone replacement therapy. *Med. Clin. N. Am.* **1998**, *82*, 297–320. [\[CrossRef\]](https://doi.org/10.1016/S0025-7125(05)70608-8)
- 70. Henderson, V.W.; Paganini-Hill, A.; Emanuel, C.K.; Dunn, M.E.; Buckwalter, J.G. Estrogen replacement therapy in older women. Comparisons between Alzheimer's disease cases and nondemented control subjects. *Arch. Neurol.* **1994**, *51*, 896–900. [\[CrossRef\]](https://doi.org/10.1001/archneur.1994.00540210068014)
- 71. Mortel, K.F.; Meyer, J.S. Lack of postmenopausal estrogen replacement therapy and the risk of dementia. *J. Neuropsychiatry Clin. Neurosci.* **1995**, *7*, 334–337. [\[CrossRef\]](https://doi.org/10.1176/jnp.7.3.334)
- 72. Kawas, C.; Resnick, S.; Morrison, A.; Brookmeyer, R.; Corrada, M.; Zonderman, A.; Bacal, C.; Lingle, D.D.; Metter, E. A prospective study of estrogen replacement therapy and the risk of developing Alzheimer's disease: The Baltimore Longitudinal Study of Aging. *Neurology* **1997**, *48*, 1517–1521. [\[CrossRef\]](https://doi.org/10.1212/WNL.48.6.1517)
- 73. Mulnard, R.A.; Cotman, C.W.; Kawas, C.; van Dyck, C.H.; Sano, M.; Doody, R.; Koss, E.; Pfeiffer, E.; Jin, S.; Gamst, A.; et al. Estrogen replacement therapy for treatment of mild to moderate Alzheimer disease: A randomized controlled trial. Alzheimer's Disease Cooperative Study. *JAMA* **2000**, *283*, 1007–1015. [\[CrossRef\]](https://doi.org/10.1001/jama.283.8.1007)
- 74. Grady, D.; Yaffe, K.; Kristof, M.; Lin, F.; Richards, C.; Barrett-Connor, E. Effect of postmenopausal hormone therapy on cognitive function: The Heart and Estrogen/progestin Replacement Study. *Am. J. Med.* **2002**, *113*, 543–548. [\[CrossRef\]](https://doi.org/10.1016/S0002-9343(02)01270-6)
- 75. Shumaker, S.A.; Legault, C.; Rapp, S.R.; Thal, L.; Wallace, R.B.; Ockene, J.K.; Hendrix, S.L.; Jones, B.N., 3rd; Assaf, A.R.; Jackson, R.D.; et al. Estrogen plus progestin and the incidence of dementia and mild cognitive impairment in postmenopausal women: The Women's Health Initiative Memory Study: A randomized controlled trial. *JAMA* **2003**, *289*, 2651–2662. [\[CrossRef\]](https://doi.org/10.1001/jama.289.20.2651)
- 76. Zhou, H.-H.; Yu, Z.; Luo, L.; Xie, F.; Wang, Y.; Wan, Z. The effect of hormone replacement therapy on cognitive function in healthy postmenopausal women: A meta-analysis of 23 randomized controlled trials. *Psychogeriatrics* **2021**, *21*, 926–938. [\[CrossRef\]](https://doi.org/10.1111/psyg.12768) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34622524)
- 77. Saleh, R.N.M.; Hornberger, M.; Ritchie, C.W.; Minihane, A.M. Hormone replacement therapy is associated with improved cognition and larger brain volumes in at-risk APOE4 women: Results from the European Prevention of Alzheimer's Disease (EPAD) cohort. *Alzheimer's Res. Ther.* **2023**, *15*, 10. [\[CrossRef\]](https://doi.org/10.1186/s13195-022-01121-5)
- 78. Hogervorst, E.; Bandelow, S.; Combrinck, M.; Smith, A.D. Low free testosterone is an independent risk factor for Alzheimer's disease. *Exp. Gerontol.* **2004**, *39*, 1633–1639. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2004.06.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15582279)
- 79. Moffat, S.D.; Zonderman, A.B.; Metter, E.J.; Kawas, C.; Blackman, M.R.; Harman, S.M.; Resnick, S.M. Free testosterone and risk for Alzheimer disease in older men. *Neurology* **2004**, *62*, 188–193. [\[CrossRef\]](https://doi.org/10.1212/WNL.62.2.188) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14745052)
- 80. Lv, W.; Du, N.; Liu, Y.; Fan, X.; Wang, Y.; Jia, X.; Hou, X.; Wang, B. Low Testosterone Level and Risk of Alzheimer's Disease in the Elderly Men: A Systematic Review and Meta-Analysis. *Mol. Neurobiol.* **2016**, *53*, 2679–2684. [\[CrossRef\]](https://doi.org/10.1007/s12035-015-9315-y)
- 81. Lee, J.H.; Byun, M.S.; Yi, D.; Choe, Y.M.; Choi, H.J.; Baek, H.; Sohn, B.K.; Lee, J.Y.; Kim, H.J.; Kim, J.W.; et al. Sex-specific association of sex hormones and gonadotropins, with brain amyloid and hippocampal neurodegeneration. *Neurobiol. Aging* **2017**, *58*, 34–40. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2017.06.005)
- 82. Ishikawa, T.; Glidewell-Kenney, C.; Jameson, J.L. Aromatase-independent testosterone conversion into estrogenic steroids is inhibited by a 5 alpha-reductase inhibitor. *J. Steroid Biochem. Mol. Biol.* **2006**, *98*, 133–138. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2005.09.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16386416)
- 83. Bianchi, V.E. Impact of Testosterone on Alzheimer's Disease. *World J. Mens. Health* **2022**, *40*, 243–256. [\[CrossRef\]](https://doi.org/10.5534/wjmh.210175) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35021306)
- 84. Barry, M.J.; Delorenzo, M.A.; Walker-Corkery, E.S.; Lucas, F.L.; Wennberg, D.C. The rising prevalence of androgen deprivation among older American men since the advent of prostate-specific antigen testing: A population-based cohort study. *BJU Int.* **2006**, *98*, 973–978. [\[CrossRef\]](https://doi.org/10.1111/j.1464-410X.2006.06416.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16879443)
- 85. Gonzalez, B.D.; Jim, H.S.; Booth-Jones, M.; Small, B.J.; Sutton, S.K.; Lin, H.Y.; Park, J.Y.; Spiess, P.E.; Fishman, M.N.; Jacobsen, P.B. Course and Predictors of Cognitive Function in Patients With Prostate Cancer Receiving Androgen-Deprivation Therapy: A Controlled Comparison. *J. Clin. Oncol.* **2015**, *33*, 2021–2027. [\[CrossRef\]](https://doi.org/10.1200/JCO.2014.60.1963) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25964245)
- 86. Alibhai, S.M.; Timilshina, N.; Duff-Canning, S.; Breunis, H.; Tannock, I.F.; Naglie, G.; Fleshner, N.E.; Krahn, M.D.; Warde, P.; Marzouk, S.; et al. Effects of long-term androgen deprivation therapy on cognitive function over 36 months in men with prostate cancer. *Cancer* **2017**, *123*, 237–244. [\[CrossRef\]](https://doi.org/10.1002/cncr.30320) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27583806)
- 87. Cryan, J.F.; O'Riordan, K.J.; Cowan, C.S.M.; Sandhu, K.V.; Bastiaanssen, T.F.S.; Boehme, M.; Codagnone, M.G.; Cussotto, S.; Fulling, C.; Golubeva, A.V.; et al. The Microbiota-Gut-Brain Axis. *Physiol. Rev.* **2019**, *99*, 1877–2013. [\[CrossRef\]](https://doi.org/10.1152/physrev.00018.2018) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31460832)
- 88. Sasmita, A.O. Modification of the gut microbiome to combat neurodegeneration. *Rev. Neurosci.* **2019**, *30*, 795–805. [\[CrossRef\]](https://doi.org/10.1515/revneuro-2019-0005)
- 89. Scheperjans, F.; Aho, V.; Pereira, P.A.; Koskinen, K.; Paulin, L.; Pekkonen, E.; Haapaniemi, E.; Kaakkola, S.; Eerola-Rautio, J.; Pohja, M.; et al. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov. Disord.* **2015**, *30*, 350–358. [\[CrossRef\]](https://doi.org/10.1002/mds.26069)
- 90. Vizcarra, J.A.; Wilson-Perez, H.E.; Espay, A.J. The power in numbers: Gut microbiota in Parkinson's disease. *Mov. Disord.* **2015**, *30*, 296–298. [\[CrossRef\]](https://doi.org/10.1002/mds.26116)
- 91. Kong, G.; Cao, K.L.; Judd, L.M.; Li, S.; Renoir, T.; Hannan, A.J. Microbiome profiling reveals gut dysbiosis in a transgenic mouse model of Huntington's disease. *Neurobiol. Dis.* **2020**, *135*, 104268. [\[CrossRef\]](https://doi.org/10.1016/j.nbd.2018.09.001)
- 92. Dodiya, H.B.; Kuntz, T.; Shaik, S.M.; Baufeld, C.; Leibowitz, J.; Zhang, X.; Gottel, N.; Zhang, X.; Butovsky, O.; Gilbert, J.A.; et al. Sex-specific effects of microbiome perturbations on cerebral Aβ amyloidosis and microglia phenotypes. *J. Exp. Med.* **2019**, *216*, 1542–1560. [\[CrossRef\]](https://doi.org/10.1084/jem.20182386) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31097468)
- 93. Chu, C.; Murdock, M.H.; Jing, D.; Won, T.H.; Chung, H.; Kressel, A.M.; Tsaava, T.; Addorisio, M.E.; Putzel, G.G.; Zhou, L.; et al. The microbiota regulate neuronal function and fear extinction learning. *Nature* **2019**, *574*, 543–548. [\[CrossRef\]](https://doi.org/10.1038/s41586-019-1644-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31645720)
- 94. Shen, L.; Liu, L.; Ji, H.F. Alzheimer's Disease Histological and Behavioral Manifestations in Transgenic Mice Correlate with Specific Gut Microbiome State. *J. Alzheimers Dis.* **2017**, *56*, 385–390. [\[CrossRef\]](https://doi.org/10.3233/JAD-160884) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27911317)
- 95. Bäuerl, C.; Collado, M.C.; Diaz Cuevas, A.; Viña, J.; Pérez Martínez, G. Shifts in gut microbiota composition in an APP/PSS1 transgenic mouse model of Alzheimer's disease during lifespan. *Lett. Appl. Microbiol.* **2018**, *66*, 464–471. [\[CrossRef\]](https://doi.org/10.1111/lam.12882) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29575030)
- 96. Kong, Y.; Jiang, B.; Luo, X. Gut microbiota influences Alzheimer's disease pathogenesis by regulating acetate in Drosophila model. *Future Microbiol.* **2018**, *13*, 1117–1128. [\[CrossRef\]](https://doi.org/10.2217/fmb-2018-0185)
- 97. Shen, L.; Ji, H.F. Associations Between Gut Microbiota and Alzheimer's Disease: Current Evidences and Future Therapeutic and Diagnostic Perspectives. *J. Alzheimers Dis.* **2019**, *68*, 25–31. [\[CrossRef\]](https://doi.org/10.3233/JAD-181143)
- 98. Fransen, F.; van Beek, A.A.; Borghuis, T.; Meijer, B.; Hugenholtz, F.; van der Gaast-de Jongh, C.; Savelkoul, H.F.; de Jonge, M.I.; Faas, M.M.; Boekschoten, M.V.; et al. The Impact of Gut Microbiota on Gender-Specific Differences in Immunity. *Front. Immunol.* **2017**, *8*, 754. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2017.00754) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28713378)
- 99. Elderman, M.; Hugenholtz, F.; Belzer, C.; Boekschoten, M.; van Beek, A.; de Haan, B.; Savelkoul, H.; de Vos, P.; Faas, M. Sex and strain dependent differences in mucosal immunology and microbiota composition in mice. *Biol. Sex Differ.* **2018**, *9*, 26. [\[CrossRef\]](https://doi.org/10.1186/s13293-018-0186-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29914546)
- 100. Sinha, T.; Vich Vila, A.; Garmaeva, S.; Jankipersadsing, S.A.; Imhann, F.; Collij, V.; Bonder, M.J.; Jiang, X.; Gurry, T.; Alm, E.J.; et al. Analysis of 1135 gut metagenomes identifies sex-specific resistome profiles. *Gut Microbes* **2019**, *10*, 358–366. [\[CrossRef\]](https://doi.org/10.1080/19490976.2018.1528822)
- 101. Singh, P.; Manning, S.D. Impact of age and sex on the composition and abundance of the intestinal microbiota in individuals with and without enteric infections. *Ann. Epidemiol.* **2016**, *26*, 380–385. [\[CrossRef\]](https://doi.org/10.1016/j.annepidem.2016.03.007)
- 102. Dominianni, C.; Sinha, R.; Goedert, J.J.; Pei, Z.; Yang, L.; Hayes, R.B.; Ahn, J. Sex, body mass index, and dietary fiber intake influence the human gut microbiome. *PLoS ONE* **2015**, *10*, e0124599. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0124599) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25874569)
- 103. Haro, C.; Rangel-Zúñiga, O.A.; Alcalá-Díaz, J.F.; Gómez-Delgado, F.; Pérez-Martínez, P.; Delgado-Lista, J.; Quintana-Navarro, G.M.; Landa, B.B.; Navas-Cortés, J.A.; Tena-Sempere, M.; et al. Intestinal Microbiota Is Influenced by Gender and Body Mass Index. *PLoS ONE* **2016**, *11*, e0154090. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0154090) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27228093)
- 104. Takagi, T.; Naito, Y.; Inoue, R.; Kashiwagi, S.; Uchiyama, K.; Mizushima, K.; Tsuchiya, S.; Dohi, O.; Yoshida, N.; Kamada, K.; et al. Differences in gut microbiota associated with age, sex, and stool consistency in healthy Japanese subjects. *J. Gastroenterol.* **2019**, *54*, 53–63. [\[CrossRef\]](https://doi.org/10.1007/s00535-018-1488-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29926167)
- 105. Minter, M.R.; Hinterleitner, R.; Meisel, M.; Zhang, C.; Leone, V.; Zhang, X.; Oyler-Castrillo, P.; Zhang, X.; Musch, M.W.; Shen, X.; et al. Antibiotic-induced perturbations in microbial diversity during post-natal development alters amyloid pathology in an aged APP(SWE)/PS1(∆E9) murine model of Alzheimer's disease. *Sci. Rep.* **2017**, *7*, 10411. [\[CrossRef\]](https://doi.org/10.1038/s41598-017-11047-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28874832)
- 106. Minter, M.R.; Zhang, C.; Leone, V.; Ringus, D.L.; Zhang, X.; Oyler-Castrillo, P.; Musch, M.W.; Liao, F.; Ward, J.F.; Holtzman, D.M.; et al. Antibiotic-induced perturbations in gut microbial diversity influences neuro-inflammation and amyloidosis in a murine model of Alzheimer's disease. *Sci. Rep.* **2016**, *6*, 30028. [\[CrossRef\]](https://doi.org/10.1038/srep30028)
- 107. Dodiya, H.B.; Lutz, H.L.; Weigle, I.Q.; Patel, P.; Michalkiewicz, J.; Roman-Santiago, C.J.; Zhang, C.M.; Liang, Y.; Srinath, A.; Zhang, X.; et al. Gut microbiota-driven brain Aβ amyloidosis in mice requires microglia. *J. Exp. Med.* **2022**, *219*, e20200895. [\[CrossRef\]](https://doi.org/10.1084/jem.20200895) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34854884)
- 108. Kaur, H.; Nookala, S.; Singh, S.; Mukundan, S.; Nagamoto-Combs, K.; Combs, C.K. Sex-Dependent Effects of Intestinal Microbiome Manipulation in a Mouse Model of Alzheimer's Disease. *Cells* **2021**, *10*, 2370. [\[CrossRef\]](https://doi.org/10.3390/cells10092370) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34572019)
- 109. Zhao, Y.; Lukiw, W.J. Microbiome-generated amyloid and potential impact on amyloidogenesis in Alzheimer's disease (AD). *J. Nat. Sci.* **2015**, *1*, e138.
- 110. Zhao, Y.; Lukiw, W.J. TREM2 signaling, miRNA-34a and the extinction of phagocytosis. *Front. Cell. Neurosci.* **2013**, *7*, 131. [\[CrossRef\]](https://doi.org/10.3389/fncel.2013.00131)
- 111. Hill, J.M.; Lukiw, W.J. Microbial-generated amyloids and Alzheimer's disease (AD). *Front. Aging Neurosci.* **2015**, *7*, 9. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2015.00009)
- 112. Zhang, J.; Ke, K.F.; Liu, Z.; Qiu, Y.H.; Peng, Y.P. Th17 cell-mediated neuroinflammation is involved in neurodegeneration of aβ1-42-induced Alzheimer's disease model rats. *PLoS ONE* **2013**, *8*, e75786. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0075786) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24124514)
- 113. Friedland, R.P. Mechanisms of molecular mimicry involving the microbiota in neurodegeneration. *J. Alzheimers Dis.* **2015**, *45*, 349–362. [\[CrossRef\]](https://doi.org/10.3233/JAD-142841) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25589730)
- 114. Goveas, J.S.; Espeland, M.A.; Woods, N.F.; Wassertheil-Smoller, S.; Kotchen, J.M. Depressive symptoms and incidence of mild cognitive impairment and probable dementia in elderly women: The Women's Health Initiative Memory Study. *J. Am. Geriatr. Soc.* **2011**, *59*, 57–66. [\[CrossRef\]](https://doi.org/10.1111/j.1532-5415.2010.03233.x)
- 115. Ownby, R.L.; Crocco, E.; Acevedo, A.; John, V.; Loewenstein, D. Depression and risk for Alzheimer disease: Systematic review, meta-analysis, and metaregression analysis. *Arch. Gen. Psychiatry* **2006**, *63*, 530–538. [\[CrossRef\]](https://doi.org/10.1001/archpsyc.63.5.530) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16651510)
- 116. Kessler, R.C.; McGonagle, K.A.; Swartz, M.; Blazer, D.G.; Nelson, C.B. Sex and depression in the National Comorbidity Survey. I: Lifetime prevalence, chronicity and recurrence. *J. Affect. Disord.* **1993**, *29*, 85–96. [\[CrossRef\]](https://doi.org/10.1016/0165-0327(93)90026-G) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8300981)
- 117. Nebel, R.A.; Aggarwal, N.T.; Barnes, L.L.; Gallagher, A.; Goldstein, J.M.; Kantarci, K.; Mallampalli, M.P.; Mormino, E.C.; Scott, L.; Yu, W.H.; et al. Understanding the impact of sex and gender in Alzheimer's disease: A call to action. *Alzheimers Dement.* **2018**, *14*, 1171–1183. [\[CrossRef\]](https://doi.org/10.1016/j.jalz.2018.04.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29907423)
- 118. Spalletta, G.; Musicco, M.; Padovani, A.; Rozzini, L.; Perri, R.; Fadda, L.; Canonico, V.; Trequattrini, A.; Pettenati, C.; Caltagirone, C.; et al. Neuropsychiatric symptoms and syndromes in a large cohort of newly diagnosed, untreated patients with Alzheimer disease. *Am. J. Geriatr. Psychiatry* **2010**, *18*, 1026–1035. [\[CrossRef\]](https://doi.org/10.1097/JGP.0b013e3181d6b68d)
- 119. Lee, J.; Lee, K.J.; Kim, H. Gender differences in behavioral and psychological symptoms of patients with Alzheimer's disease. *Asian J. Psychiatr.* **2017**, *26*, 124–128. [\[CrossRef\]](https://doi.org/10.1016/j.ajp.2017.01.027) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28483073)
- 120. Maccora, J.; Peters, R.; Anstey, K.J. What does (low) education mean in terms of dementia risk? A systematic review and meta-analysis highlighting inconsistency in measuring and operationalising education. *SSM Popul. Health* **2020**, *12*, 100654. [\[CrossRef\]](https://doi.org/10.1016/j.ssmph.2020.100654)
- 121. Xu, W.; Tan, L.; Wang, H.F.; Tan, M.S.; Tan, L.; Li, J.Q.; Zhao, Q.F.; Yu, J.T. Education and Risk of Dementia: Dose-Response Meta-Analysis of Prospective Cohort Studies. *Mol. Neurobiol.* **2016**, *53*, 3113–3123. [\[CrossRef\]](https://doi.org/10.1007/s12035-015-9211-5)
- 122. Crowe, M.; Andel, R.; Pedersen, N.L.; Johansson, B.; Gatz, M. Does participation in leisure activities lead to reduced risk of Alzheimer's disease? A prospective study of Swedish twins. *J. Gerontol. B Psychol. Sci. Soc. Sci.* **2003**, *58*, P249–P255. [\[CrossRef\]](https://doi.org/10.1093/geronb/58.5.P249)
- 123. Fabrigoule, C. Do leisure activities protect against Alzheimer's disease? *Lancet Neurol.* **2002**, *1*, 11. [\[CrossRef\]](https://doi.org/10.1016/S1474-4422(02)00010-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12849540)
- 124. Vemuri, P.; Lesnick, T.G.; Przybelski, S.A.; Knopman, D.S.; Roberts, R.O.; Lowe, V.J.; Kantarci, K.; Senjem, M.L.; Gunter, J.L.; Boeve, B.F.; et al. Effect of lifestyle activities on Alzheimer disease biomarkers and cognition. *Ann. Neurol.* **2012**, *72*, 730–738. [\[CrossRef\]](https://doi.org/10.1002/ana.23665) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23280791)
- 125. Hasselgren, C.; Ekbrand, H.; Halleröd, B.; Mellqvist Fässberg, M.; Zettergren, A.; Johansson, L.; Skoog, I.; Dellve, L. Sex differences in dementia: On the potentially mediating effects of educational attainment and experiences of psychological distress. *BMC Psychiatry* **2020**, *20*, 434. [\[CrossRef\]](https://doi.org/10.1186/s12888-020-02820-9)
- 126. Kukull, W.A.; Higdon, R.; Bowen, J.D.; McCormick, W.C.; Teri, L.; Schellenberg, G.D.; van Belle, G.; Jolley, L.; Larson, E.B. Dementia and Alzheimer disease incidence: A prospective cohort study. *Arch. Neurol.* **2002**, *59*, 1737–1746. [\[CrossRef\]](https://doi.org/10.1001/archneur.59.11.1737)
- 127. U.S. Bureau of Labor Statistics. *Women in the Labor Force: A Databook*; Retrieved; U.S. Bureau of Labor Statistics: Washington, DC, USA, 2015.
- 128. Roos, P.A.; Stevens, L.M. Integrating occupations: Changing occupational sex segregation in the United States from 2000 to 2014. *Demogr. Res.* **2018**, *38*, 127–154. [\[CrossRef\]](https://doi.org/10.4054/DemRes.2018.38.5)
- 129. Huang, L.Y.; Hu, H.Y.; Wang, Z.T.; Ma, Y.H.; Dong, Q.; Tan, L.; Yu, J.T. Association of Occupational Factors and Dementia or Cognitive Impairment: A Systematic Review and Meta-Analysis. *J. Alzheimers Dis.* **2020**, *78*, 217–227. [\[CrossRef\]](https://doi.org/10.3233/JAD-200605)
- 130. Lapane, K.L.; Gambassi, G.; Landi, F.; Sgadari, A.; Mor, V.; Bernabei, R. Gender differences in predictors of mortality in nursing home residents with AD. *Neurology* **2001**, *56*, 650–654. [\[CrossRef\]](https://doi.org/10.1212/WNL.56.5.650)
- 131. Geerlings, M.I.; Deeg, D.J.; Schmand, B.; Lindeboom, J.; Jonker, C. Increased risk of mortality in Alzheimer's disease patients with higher education? A replication study. *Neurology* **1997**, *49*, 798–802. [\[CrossRef\]](https://doi.org/10.1212/WNL.49.3.798)
- 132. Stern, Y.; Tang, M.X.; Albert, M.S.; Brandt, J.; Jacobs, D.M.; Bell, K.; Marder, K.; Sano, M.; Devanand, D.; Albert, S.M.; et al. Predicting time to nursing home care and death in individuals with Alzheimer disease. *JAMA* **1997**, *277*, 806–812. [\[CrossRef\]](https://doi.org/10.1001/jama.1997.03540340040030)
- 133. Wang, J.; Tanila, H.; Puolivali, J.; Kadish, I.; van Groen, T. Gender differences in the amount and deposition of amyloidbeta in APPswe and PS1 double transgenic mice. *Neurobiol. Dis.* **2003**, *14*, 318–327. [\[CrossRef\]](https://doi.org/10.1016/j.nbd.2003.08.009) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14678749)
- 134. Price, D.L.; Sisodia, S.S.; Gandy, S.E. Amyloid beta amyloidosis in Alzheimer's disease. *Curr. Opin. Neurol.* **1995**, *8*, 268–274. [\[CrossRef\]](https://doi.org/10.1097/00019052-199508000-00004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7582041)
- 135. Hardy, J. The Alzheimer family of diseases: Many etiologies, one pathogenesis? *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 2095–2097. [\[CrossRef\]](https://doi.org/10.1073/pnas.94.6.2095) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9122152)
- 136. Jiao, S.S.; Bu, X.L.; Liu, Y.H.; Zhu, C.; Wang, Q.H.; Shen, L.L.; Liu, C.H.; Wang, Y.R.; Yao, X.Q.; Wang, Y.J. Sex Dimorphism Profile of Alzheimer's Disease-Type Pathologies in an APP/PS1 Mouse Model. *Neurotox. Res.* **2016**, *29*, 256–266. [\[CrossRef\]](https://doi.org/10.1007/s12640-015-9589-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26707129)
- 137. Li, X.; Feng, Y.; Wu, W.; Zhao, J.; Fu, C.; Li, Y.; Ding, Y.; Wu, B.; Gong, Y.; Yang, G.; et al. Sex differences between APPswePS1dE9 mice in A-beta accumulation and pancreatic islet function during the development of Alzheimer's disease. *Lab. Anim.* **2016**, *50*, 275–285. [\[CrossRef\]](https://doi.org/10.1177/0023677215615269) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26519428)
- 138. Ordonez-Gutierrez, L.; Fernandez-Perez, I.; Herrera, J.L.; Anton, M.; Benito-Cuesta, I.; Wandosell, F. AbetaPP/PS1 Transgenic Mice Show Sex Differences in the Cerebellum Associated with Aging. *J. Alzheimers Dis.* **2016**, *54*, 645–656. [\[CrossRef\]](https://doi.org/10.3233/JAD-160572) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27567877)
- 139. Howlett, D.R.; Richardson, J.C.; Austin, A.; Parsons, A.A.; Bate, S.T.; Davies, D.C.; Gonzalez, M.I. Cognitive correlates of Aβ deposition in male and female mice bearing amyloid precursor protein and presenilin-1 mutant transgenes. *Brain Res.* **2004**, *1017*, 130–136. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2004.05.029) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15261108)
- 140. Pistell, P.J.; Zhu, M.; Ingram, D.K. Acquisition of conditioned taste aversion is impaired in the amyloid precursor protein/presenilin 1 mouse model of Alzheimer's disease. *Neuroscience* **2008**, *152*, 594–600. [\[CrossRef\]](https://doi.org/10.1016/j.neuroscience.2008.01.025)
- 141. Davis, E.J.; Broestl, L.; Abdulai-Saiku, S.; Worden, K.; Bonham, L.W.; Minones-Moyano, E.; Moreno, A.J.; Wang, D.; Chang, K.; Williams, G.; et al. A second X chromosome contributes to resilience in a mouse model of Alzheimer's disease. *Sci. Transl. Med.* **2020**, *12*, eaaz5677. [\[CrossRef\]](https://doi.org/10.1126/scitranslmed.aaz5677)
- 142. Games, D.; Adams, D.; Alessandrini, R.; Barbour, R.; Berthelette, P.; Blackwell, C.; Carr, T.; Clemens, J.; Donaldson, T.; Gillespie, F.; et al. Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. *Nature* **1995**, *373*, 523–527. [\[CrossRef\]](https://doi.org/10.1038/373523a0)
- 143. Hsiao, K.; Chapman, P.; Nilsen, S.; Eckman, C.; Harigaya, Y.; Younkin, S.; Yang, F.; Cole, G. Correlative memory deficits, Abeta elevation, and amyloid plaques in transgenic mice. *Science* **1996**, *274*, 99–102. [\[CrossRef\]](https://doi.org/10.1126/science.274.5284.99)
- 144. Callahan, M.J.; Lipinski, W.J.; Bian, F.; Durham, R.A.; Pack, A.; Walker, L.C. Augmented senile plaque load in aged female beta-amyloid precursor protein-transgenic mice. *Am. J. Pathol.* **2001**, *158*, 1173–1177. [\[CrossRef\]](https://doi.org/10.1016/S0002-9440(10)64064-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11238065)
- 145. Schmid, S.; Rammes, G.; Blobner, M.; Kellermann, K.; Bratke, S.; Fendl, D.; Kaichuan, Z.; Schneider, G.; Jungwirth, B. Cognitive decline in Tg2576 mice shows sex-specific differences and correlates with cerebral amyloid-beta. *Behav. Brain Res.* **2019**, *359*, 408–417. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2018.11.022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30458163)
- 146. Oddo, S.; Caccamo, A.; Shepherd, J.D.; Murphy, M.P.; Golde, T.E.; Kayed, R.; Metherate, R.; Mattson, M.P.; Akbari, Y.; LaFerla, F.M. Triple-transgenic model of Alzheimer's disease with plaques and tangles: Intracellular Abeta and synaptic dysfunction. *Neuron* **2003**, *39*, 409–421. [\[CrossRef\]](https://doi.org/10.1016/S0896-6273(03)00434-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12895417)
- 147. Oddo, S.; Caccamo, A.; Kitazawa, M.; Tseng, B.P.; LaFerla, F.M. Amyloid deposition precedes tangle formation in a triple transgenic model of Alzheimer's disease. *Neurobiol. Aging* **2003**, *24*, 1063–1070. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2003.08.012) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14643377)
- 148. Mesulam, M.M. Neuroplasticity failure in Alzheimer's disease: Bridging the gap between plaques and tangles. *Neuron* **1999**, *24*, 521–529. [\[CrossRef\]](https://doi.org/10.1016/S0896-6273(00)81109-5)
- 149. Blazquez, G.; Canete, T.; Tobena, A.; Gimenez-Llort, L.; Fernandez-Teruel, A. Cognitive and emotional profiles of aged Alzheimer's disease (3xTgAD) mice: Effects of environmental enrichment and sexual dimorphism. *Behav. Brain Res.* **2014**, *268*, 185–201. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2014.04.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24746486)
- 150. Carroll, J.C.; Rosario, E.R.; Kreimer, S.; Villamagna, A.; Gentzschein, E.; Stanczyk, F.Z.; Pike, C.J. Sex differences in beta-amyloid accumulation in 3xTg-AD mice: Role of neonatal sex steroid hormone exposure. *Brain Res.* **2010**, *1366*, 233–245. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2010.10.009) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20934413)
- 151. Clinton, L.K.; Billings, L.M.; Green, K.N.; Caccamo, A.; Ngo, J.; Oddo, S.; McGaugh, J.L.; LaFerla, F.M. Age-dependent sexual dimorphism in cognition and stress response in the 3xTg-AD mice. *Neurobiol. Dis.* **2007**, *28*, 76–82. [\[CrossRef\]](https://doi.org/10.1016/j.nbd.2007.06.013)
- 152. Hirata-Fukae, C.; Li, H.F.; Hoe, H.S.; Gray, A.J.; Minami, S.S.; Hamada, K.; Niikura, T.; Hua, F.; Tsukagoshi-Nagai, H.; Horikoshi-Sakuraba, Y.; et al. Females exhibit more extensive amyloid, but not tau, pathology in an Alzheimer transgenic model. *Brain Res.* **2008**, *1216*, 92–103. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2008.03.079)
- 153. Yang, J.T.; Wang, Z.J.; Cai, H.Y.; Yuan, L.; Hu, M.M.; Wu, M.N.; Qi, J.S. Sex Differences in Neuropathology and Cognitive Behavior in APP/PS1/tau Triple-Transgenic Mouse Model of Alzheimer's Disease. *Neurosci. Bull.* **2018**, *34*, 736–746. [\[CrossRef\]](https://doi.org/10.1007/s12264-018-0268-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30099679)
- 154. Giménez-Llort, L.; García, Y.; Buccieri, K.; Revilla, S.; Suñol, C.; Cristofol, R.; Sanfeliu, C. Gender-Specific Neuroimmunoendocrine Response to Treadmill Exercise in 3xTg-AD Mice. *Int. J. Alzheimers Dis.* **2010**, *2010*, 128354. [\[CrossRef\]](https://doi.org/10.4061/2010/128354) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20981262)
- 155. Roses, A.D. Apolipoprotein E alleles as risk factors in Alzheimer's disease. *Annu. Rev. Med.* **1996**, *47*, 387–400. [\[CrossRef\]](https://doi.org/10.1146/annurev.med.47.1.387) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8712790)
- 156. Raber, J.; Wong, D.; Buttini, M.; Orth, M.; Bellosta, S.; Pitas, R.E.; Mahley, R.W.; Mucke, L. Isoform-specific effects of human apolipoprotein E on brain function revealed in ApoE knockout mice: Increased susceptibility of females. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 10914–10919. [\[CrossRef\]](https://doi.org/10.1073/pnas.95.18.10914)
- 157. Hou, X.; Adeosun, S.O.; Zhang, Q.; Barlow, B.; Brents, M.; Zheng, B.; Wang, J. Differential contributions of ApoE4 and female sex to BACE1 activity and expression mediate Aβ deposition and learning and memory in mouse models of Alzheimer's disease. *Front. Aging Neurosci.* **2015**, *7*, 207. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2015.00207) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26582141)
- 158. Cacciottolo, M.; Christensen, A.; Moser, A.; Liu, J.; Pike, C.J.; Smith, C.; LaDu, M.J.; Sullivan, P.M.; Morgan, T.E.; Dolzhenko, E.; et al. The APOE4 allele shows opposite sex bias in microbleeds and Alzheimer's disease of humans and mice. *Neurobiol. Aging* **2016**, *37*, 47–57. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2015.10.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26686669)
- 159. Stephen, T.L.; Cacciottolo, M.; Balu, D.; Morgan, T.E.; LaDu, M.J.; Finch, C.E.; Pike, C.J. APOE genotype and sex affect microglial interactions with plaques in Alzheimer's disease mice. *Acta Neuropathol. Commun.* **2019**, *7*, 82. [\[CrossRef\]](https://doi.org/10.1186/s40478-019-0729-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31113487)
- 160. Shang, Y.; Mishra, A.; Wang, T.; Wang, Y.; Desai, M.; Chen, S.; Mao, Z.; Do, L.; Bernstein, A.S.; Trouard, T.P.; et al. Evidence in support of chromosomal sex influencing plasma based metabolome vs APOE genotype influencing brain metabolome profile in humanized APOE male and female mice. *PLoS ONE* **2020**, *15*, e0225392. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0225392)
- 161. Shi, L.; Du, X.; Zhou, H.; Tao, C.; Liu, Y.; Meng, F.; Wu, G.; Xiong, Y.; Xia, C.; Wang, Y.; et al. Cumulative effects of the ApoE genotype and gender on the synaptic proteome and oxidative stress in the mouse brain. *Int. J. Neuropsychopharmacol.* **2014**, *17*, 1863–1879. [\[CrossRef\]](https://doi.org/10.1017/S1461145714000601)
- 162. McLean, J.W.; Bhattrai, A.; Vitali, F.; Raikes, A.C.; Wiegand, J.L.; Brinton, R.D. Contributions of sex and genotype to exploratory behavior differences in an aged humanized APOE mouse model of late-onset Alzheimer's disease. *Learn. Mem.* **2022**, *29*, 321–331. [\[CrossRef\]](https://doi.org/10.1101/lm.053588.122)
- 163. Lossos, A.; Reches, A.; Gal, A.; Newman, J.P.; Soffer, D.; Gomori, J.M.; Boher, M.; Ekstein, D.; Biran, I.; Meiner, Z.; et al. Frontotemporal dementia and parkinsonism with the P301S tau gene mutation in a Jewish family. *J. Neurol.* **2003**, *250*, 733–740. [\[CrossRef\]](https://doi.org/10.1007/s00415-003-1074-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12796837)
- 164. Alonso Adel, C.; Mederlyova, A.; Novak, M.; Grundke-Iqbal, I.; Iqbal, K. Promotion of hyperphosphorylation by frontotemporal dementia tau mutations. *J. Biol. Chem.* **2004**, *279*, 34873–34881. [\[CrossRef\]](https://doi.org/10.1074/jbc.M405131200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15190058)
- 165. Buccarello, L.; Grignaschi, G.; Castaldo, A.M.; Di Giancamillo, A.; Domeneghini, C.; Melcangi, R.C.; Borsello, T. Sex Impact on Tau-Aggregation and Postsynaptic Protein Levels in the P301L Mouse Model of Tauopathy. *J. Alzheimers Dis.* **2017**, *56*, 1279–1292. [\[CrossRef\]](https://doi.org/10.3233/JAD-161087) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28157099)
- 166. Lewis, J.; Dickson, D.W.; Lin, W.L.; Chisholm, L.; Corral, A.; Jones, G.; Yen, S.H.; Sahara, N.; Skipper, L.; Yager, D.; et al. Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science* **2001**, *293*, 1487–1491. [\[CrossRef\]](https://doi.org/10.1126/science.1058189) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11520987)
- 167. van Eersel, J.; Stevens, C.H.; Przybyla, M.; Gladbach, A.; Stefanoska, K.; Chan, C.K.; Ong, W.Y.; Hodges, J.R.; Sutherland, G.T.; Kril, J.J.; et al. Early-onset axonal pathology in a novel P301S-Tau transgenic mouse model of frontotemporal lobar degeneration. *Neuropathol. Appl. Neurobiol.* **2015**, *41*, 906–925. [\[CrossRef\]](https://doi.org/10.1111/nan.12233) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25763777)
- 168. Sun, Y.; Guo, Y.; Feng, X.; Jia, M.; Ai, N.; Dong, Y.; Zheng, Y.; Fu, L.; Yu, B.; Zhang, H.; et al. The behavioural and neuropathologic sexual dimorphism and absence of MIP-3alpha in tau P301S mouse model of Alzheimer's disease. *J. Neuroinflammation* **2020**, *17*, 72. [\[CrossRef\]](https://doi.org/10.1186/s12974-020-01749-w)
- 169. Oakley, H.; Cole, S.L.; Logan, S.; Maus, E.; Shao, P.; Craft, J.; Guillozet-Bongaarts, A.; Ohno, M.; Disterhoft, J.; Van Eldik, L.; et al. Intraneuronal beta-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: Potential factors in amyloid plaque formation. *J. Neurosci.* **2006**, *26*, 10129–10140. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.1202-06.2006)
- 170. Moechars, D.; Lorent, K.; De Strooper, B.; Dewachter, I.; Van Leuven, F. Expression in brain of amyloid precursor protein mutated in the alpha-secretase site causes disturbed behavior, neuronal degeneration and premature death in transgenic mice. *EMBO J.* **1996**, *15*, 1265–1274. [\[CrossRef\]](https://doi.org/10.1002/j.1460-2075.1996.tb00468.x)
- 171. Sil, A.; Erfani, A.; Lamb, N.; Copland, R.; Riedel, G.; Platt, B. Sex Differences in Behavior and Molecular Pathology in the 5XFAD Model. *J. Alzheimers Dis.* **2022**, *85*, 755–778. [\[CrossRef\]](https://doi.org/10.3233/JAD-210523)
- 172. O'Leary, T.P.; Brown, R.E. Visuo-spatial learning and memory impairments in the 5xFAD mouse model of Alzheimer's disease: Effects of age, sex, albinism, and motor impairments. *Genes. Brain Behav.* **2022**, *21*, e12794. [\[CrossRef\]](https://doi.org/10.1111/gbb.12794)
- 173. Devi, L.; Alldred, M.J.; Ginsberg, S.D.; Ohno, M. Sex- and brain region-specific acceleration of beta-amyloidogenesis following behavioral stress in a mouse model of Alzheimer's disease. *Mol. Brain* **2010**, *3*, 34. [\[CrossRef\]](https://doi.org/10.1186/1756-6606-3-34) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21059265)
- 174. Roddick, K.M.; Schellinck, H.M.; Brown, R.E. Olfactory delayed matching to sample performance in mice: Sex differences in the 5XFAD mouse model of Alzheimer's disease. *Behav. Brain Res.* **2014**, *270*, 165–170. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2014.04.038) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24786328)
- 175. Ghosh, P.; Singh, R.; Ganeshpurkar, A.; Pokle, A.V.; Singh, R.B.; Singh, S.K.; Kumar, A. Cellular and molecular influencers of neuroinflammation in Alzheimer's disease: Recent concepts & roles. *Neurochem. Int.* **2021**, *151*, 105212. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2021.105212) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34656693)
- 176. Scuderi, C.; Facchinetti, R.; Steardo, L.; Valenza, M. Neuroinflammation in Alzheimer's Disease: Friend or Foe? *FASEB J.* **2020**, *34*, 1. [\[CrossRef\]](https://doi.org/10.1096/fasebj.2020.34.s1.00381)
- 177. Webers, A.; Heneka, M.T.; Gleeson, P.A. The role of innate immune responses and neuroinflammation in amyloid accumulation and progression of Alzheimer's disease. *Immunol. Cell Biol.* **2020**, *98*, 28–41. [\[CrossRef\]](https://doi.org/10.1111/imcb.12301)
- 178. Dong, Y.; Li, X.; Cheng, J.; Hou, L. Drug Development for Alzheimer's Disease: Microglia Induced Neuroinflammation as a Target? *Int. J. Mol. Sci.* **2019**, *20*, 558. [\[CrossRef\]](https://doi.org/10.3390/ijms20030558) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30696107)
- 179. Simon, E.; Obst, J.; Gomez-Nicola, D. The Evolving Dialogue of Microglia and Neurons in Alzheimer's Disease: Microglia as Necessary Transducers of Pathology. *Neuroscience* **2019**, *405*, 24–34. [\[CrossRef\]](https://doi.org/10.1016/j.neuroscience.2018.01.059)
- 180. Schwarz, J.M.; Sholar, P.W.; Bilbo, S.D. Sex differences in microglial colonization of the developing rat brain. *J. Neurochem.* **2012**, *120*, 948–963. [\[CrossRef\]](https://doi.org/10.1111/j.1471-4159.2011.07630.x)
- 181. Mouton, P.R.; Long, J.M.; Lei, D.L.; Howard, V.; Jucker, M.; Calhoun, M.E.; Ingram, D.K. Age and gender effects on microglia and astrocyte numbers in brains of mice. *Brain Res.* **2002**, *956*, 30–35. [\[CrossRef\]](https://doi.org/10.1016/S0006-8993(02)03475-3)
- 182. Bollinger, J.L.; Salinas, I.; Fender, E.; Sengelaub, D.R.; Wellman, C.L. Gonadal hormones differentially regulate sex-specific stress effects on glia in the medial prefrontal cortex. *J. Neuroendocr.* **2019**, *31*, e12762. [\[CrossRef\]](https://doi.org/10.1111/jne.12762)
- 183. Manwani, B.; Liu, F.; Scranton, V.; Hammond, M.D.; Sansing, L.H.; McCullough, L.D. Differential effects of aging and sex on stroke induced inflammation across the lifespan. *Exp. Neurol.* **2013**, *249*, 120–131. [\[CrossRef\]](https://doi.org/10.1016/j.expneurol.2013.08.011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23994069)
- 184. Guneykaya, D.; Ivanov, A.; Hernandez, D.P.; Haage, V.; Wojtas, B.; Meyer, N.; Maricos, M.; Jordan, P.; Buonfiglioli, A.; Gielniewski, B.; et al. Transcriptional and Translational Differences of Microglia from Male and Female Brains. *Cell Rep.* **2018**, *24*, 2773–2783.e2776. [\[CrossRef\]](https://doi.org/10.1016/j.celrep.2018.08.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30184509)
- 185. Mangold, C.A.; Wronowski, B.; Du, M.; Masser, D.R.; Hadad, N.; Bixler, G.V.; Brucklacher, R.M.; Ford, M.M.; Sonntag, W.E.; Freeman, W.M. Sexually divergent induction of microglial-associated neuroinflammation with hippocampal aging. *J. Neuroinflammation* **2017**, *14*, 141. [\[CrossRef\]](https://doi.org/10.1186/s12974-017-0920-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28732515)
- 186. Boghozian, R.; Sharma, S.; Narayana, K.; Cheema, M.; Brown, C.E. Sex and interferon gamma signaling regulate microglia migration in the adult mouse cortex in vivo. *Proc. Natl. Acad. Sci. USA* **2023**, *120*, e2302892120. [\[CrossRef\]](https://doi.org/10.1073/pnas.2302892120) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37428916)
- 187. Mhatre-Winters, I.; Eid, A.; Han, Y.; Tieu, K.; Richardson, J.R. Sex and APOE Genotype Alter the Basal and Induced Inflammatory States of Primary Astrocytes from Humanized Targeted Replacement Mice. *ASN Neuro* **2023**, *15*, 17590914221144549. [\[CrossRef\]](https://doi.org/10.1177/17590914221144549) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36604975)
- 188. Stefaniak, J.; O'Brien, J. Imaging of neuroinflammation in dementia: A review. *J. Neurol. Neurosurg. Psychiatry* **2016**, *87*, 21–28. [\[CrossRef\]](https://doi.org/10.1136/jnnp-2015-311336) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26384512)
- 189. Biechele, G.; Rauchmann, B.S.; Janowitz, D.; Buerger, K.; Franzmeier, N.; Weidinger, E.; Guersel, S.; Schuster, S.; Finze, A.; Harris, S.; et al. Associations between sex, body mass index and the individual microglial response in Alzheimer's disease. *J. Neuroinflammation* **2024**, *21*, 30. [\[CrossRef\]](https://doi.org/10.1186/s12974-024-03020-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38263017)
- 190. Guillot-Sestier, M.-V.; Araiz, A.R.; Mela, V.; Gaban, A.S.; O'Neill, E.; Joshi, L.; Chouchani, E.T.; Mills, E.L.; Lynch, M.A. Microglial metabolism is a pivotal factor in sexual dimorphism in Alzheimer's disease. *Commun. Biol.* **2021**, *4*, 711. [\[CrossRef\]](https://doi.org/10.1038/s42003-021-02259-y)
- 191. Koizumi, S.; Shigemoto-Mogami, Y.; Nasu-Tada, K.; Shinozaki, Y.; Ohsawa, K.; Tsuda, M.; Joshi, B.V.; Jacobson, K.A.; Kohsaka, S.; Inoue, K. UDP acting at P2Y6 receptors is a mediator of microglial phagocytosis. *Nature* **2007**, *446*, 1091–1095. [\[CrossRef\]](https://doi.org/10.1038/nature05704)
- 192. Färber, K.; Kettenmann, H. Functional role of calcium signals for microglial function. *Glia* **2006**, *54*, 656–665. [\[CrossRef\]](https://doi.org/10.1002/glia.20412)
- 193. Brawek, B.; Schwendele, B.; Riester, K.; Kohsaka, S.; Lerdkrai, C.; Liang, Y.; Garaschuk, O. Impairment of in vivo calcium signaling in amyloid plaque-associated microglia. *Acta Neuropathol.* **2014**, *127*, 495–505. [\[CrossRef\]](https://doi.org/10.1007/s00401-013-1242-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24407428)
- 195. Olmedillas Del Moral, M.; Fröhlich, N.; Figarella, K.; Mojtahedi, N.; Garaschuk, O. Effect of Caloric Restriction on the in vivo Functional Properties of Aging Microglia. *Front. Immunol.* **2020**, *11*, 750. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2020.00750) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32411143)
- 196. Sala Frigerio, C.; Wolfs, L.; Fattorelli, N.; Thrupp, N.; Voytyuk, I.; Schmidt, I.; Mancuso, R.; Chen, W.T.; Woodbury, M.E.; Srivastava, G.; et al. The Major Risk Factors for Alzheimer's Disease: Age, Sex, and Genes Modulate the Microglia Response to Aβ Plaques. *Cell Rep.* **2019**, *27*, 1293–1306.e1296. [\[CrossRef\]](https://doi.org/10.1016/j.celrep.2019.03.099) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31018141)
- 197. Sierksma, A.S.; Vanmierlo, T.; De Vry, J.; Raijmakers, M.E.; Steinbusch, H.W.; van den Hove, D.L.; Prickaerts, J. Effects of prenatal stress exposure on soluble Abeta and brain-derived neurotrophic factor signaling in male and female APPswe/PS1dE9 mice. *Neurochem. Int.* **2012**, *61*, 697–701. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2012.06.022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22750275)
- 198. Ordonez-Gutierrez, L.; Anton, M.; Wandosell, F. Peripheral amyloid levels present gender differences associated with aging in AbetaPP/PS1 mice. *J. Alzheimers Dis.* **2015**, *44*, 1063–1068. [\[CrossRef\]](https://doi.org/10.3233/JAD-141158) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25408213)
- 199. Oliveira, S.M.; Ribeiro, C.A.; Cardoso, I.; Saraiva, M.J. Gender-dependent transthyretin modulation of brain amyloid-beta levels: Evidence from a mouse model of Alzheimer's disease. *J. Alzheimers Dis.* **2011**, *27*, 429–439. [\[CrossRef\]](https://doi.org/10.3233/JAD-2011-110488)
- 200. Bayer, T.A.; Schafer, S.; Simons, A.; Kemmling, A.; Kamer, T.; Tepest, R.; Eckert, A.; Schussel, K.; Eikenberg, O.; Sturchler-Pierrat, C.; et al. Dietary Cu stabilizes brain superoxide dismutase 1 activity and reduces amyloid Abeta production in APP23 transgenic mice. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14187–14192. [\[CrossRef\]](https://doi.org/10.1073/pnas.2332818100)
- 201. Granger, M.W.; Franko, B.; Taylor, M.W.; Messier, C.; George-Hyslop, P.S.; Bennett, S.A. A TgCRND8 Mouse Model of Alzheimer's Disease Exhibits Sexual Dimorphisms in Behavioral Indices of Cognitive Reserve. *J. Alzheimers Dis.* **2016**, *51*, 757–773. [\[CrossRef\]](https://doi.org/10.3233/JAD-150587)

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