

Article **One-Stage Pathway from Hollongdione to C17-Alkyne and Vinyl Chloride Following Mannich Bases and Carboxylic Acid**

Zarema Galimova ¹ , Irina Smirnova 1,* [,](https://orcid.org/0000-0001-7176-505X) Alexander Lobov ¹ [,](https://orcid.org/0000-0002-9223-508X) Dmitriy Polovyanenko ² , Tatyana Rybalova [2](https://orcid.org/0000-0003-2398-5271) and Oxana Kazakova [1](https://orcid.org/0000-0002-5606-1588)

- Ufa Institute of Chemistry of the Ufa Federal Research Centre of the Russian Academy of Sciences, 71, pr. Oktyabrya, 450054 Ufa, Russia; chemizara@gmail.com (Z.G.); lobovan@anrb.ru (A.L.); obf@anrb.ru (O.K.)
- ² N. N. Vorozhtsov Novosibirsk Institute of Organic Chemistry SB RAS, 630090 Novosibirsk, Russia; dpolo@nioch.nsc.ru (D.P.); rybalova@nioch.nsc.ru (T.R.)
- ***** Correspondence: si8081@yandex.ru

Abstract: Hollongdione is the first recorded example of the occurrence of a dammarane hexanortriterpene in nature possessing antiviral and cytotoxic activity. Its simple one-stage transformation into compounds with terminal alkyne and vinyl chloride fragments via the interaction with phosphorus halides is reported. The copper(I)-catalyzed Mannich reaction of 3-oxo-22,23,24,25,26,27 hexanor-dammar-20(21)-in **3** led to a series of aminomethylated products, while 17-carboxylic acid was obtained by ozone oxidation of 3-oxo-22,23,24,25,26,27-hexanor-dammar-20-chloro-20(21)-en **4**; the following direct amidation of the latter has been developed. The structures of all new molecules were established by spectroscopic studies that included 2D NMR correlation methods; the molecular structures of compounds **2**–**5** were determined by X-ray analysis.

Keywords: dammarane triterpenoids; hollongdione; alkyne; vinyl chloride; Mannich base; hollongdionoic acid

Citation: Galimova, Z.; Smirnova, I.; Lobov, A.; Polovyanenko, D.; Rybalova, T.; Kazakova, O. One-Stage Pathway from Hollongdione to C17-Alkyne and Vinyl Chloride Following Mannich Bases and Carboxylic Acid. *Int. J. Mol. Sci.* **2024**, *25*, 8356. [https://doi.org/10.3390/](https://doi.org/10.3390/ijms25158356) [ijms25158356](https://doi.org/10.3390/ijms25158356)

Academic Editor: Stefano Iotti

Received: 30 June 2024 Revised: 18 July 2024 Accepted: 26 July 2024 Published: 30 July 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

1. Introduction

The unique and complex structures of natural products make them important sources for exploring novel areas of the chemical space [\[1,](#page-14-0)[2\]](#page-14-1). The introduction of highly effective reaction centers in such molecules allows expanding the library of semi-synthetic compounds, with the aim of their future application as units in biochemical science.

Pentacyclic triterpenoids comprise a diverse group that is widely present in plants and has stable skeleton and reactive sites suitable for structural transformations. Triterpenoids usually possess a secondary hydroxyl group or a carbon atom at position *C*3, and a primary hydroxyl group or carboxyl group at position *C*28 (the general structures of triterpenoids with the key sites for modification are presented in Figure [1\)](#page-1-0). The most frequent modifications of the listed fragments include an acylation reaction resulting in various esters [\[3–](#page-14-2)[5\]](#page-14-3) and glycosides [\[6,](#page-14-4)[7\]](#page-14-5), since the addition of hydrosoluble moieties such as sugar substituents at the *C*3 and/or *C*28 positions can enhance both the water solubility and the pharmacological activity [\[8,](#page-14-6)[9\]](#page-14-7).

On the other hand, there is an oxygen function bonded to the ring *E* or *D*, a number of transformations of which have also been proposed in the literature. The most unique examples are Baeyer–Villiger oxidation of 20-oxo-30-nortaraxasteryl acetate that lead to *ε*-lactone [\[10\]](#page-14-8); 30-oxobetulinic acid conversion into a set of substituted dienes by the Wittig reaction with the use of triphenylphosphonium salts [\[11\]](#page-14-9); and an epimerization at *C*19 and a condensation reaction of several platanic carboxamides with the synthesis of novel *E*-ring *δ*−lactams [\[12\]](#page-14-10). Another attractive method of carbonyl group modification is a reaction of dehydration, which is a route to terminal acetylenes, which are key precursors in drug discovery [\[2](#page-14-1)[,13\]](#page-14-11). One widely used method of accessing alkynes is an interaction of methyl ketones with phosphorous-based reagents. This approach was found to be a short-stage,

Figure 1. The general structures of lupane, oleanane, and dammarane type triterpenoids. The cycles in the structures of triterpenoids are marked in blue. such as the general succession repair, because, and damination type interpendicis. The cycles

In this work, we decided to use dammarane-type triterpenoid hollongdione (22,23,24, 25,26,27-hexanordammar-3,20-dione) 1 with a C17 acetyl group as an initial scaffold [\[20\]](#page-14-18). Hollongdione (the structure of which is presented in Figure 2) can be obtained by onereactor synthesis from dipterocarpol, the main metabolite of Dipterocarpus alatus oleoresin or isolated from Dipterocarpus pilosus, Gardenia aubryi, Chisocheton penduliflorus, and leaves of Euphorbia leucocephala, which exhibits potent anti-influenza A virus activity [\[21](#page-14-19)-23]. Moreover, it is a crucial intermediate in the synthesis of the potent immunosuppressive agent 17α-23-(E)-dammara-20,23-diene-3β,25-diol and moderately active against small-cell lung cancer cells (N[CI-H](#page-15-0)[18](#page-15-1)7) [24,25]. Arylidene derivatives of hollongdione, described before, induce antiproliferative activity against melanoma and breast cancer through proapoptotic and antiangiogenic [mech](#page-15-2)anisms [26]. It is recognized as a hybrid molecule that contains cycle A , similar to triterpene, and a C17-acetyl group with a β -configuration attached to the D cycle, similar to the pregnane steroids.

Figure 2. The structure of hollongdione **1**. scribed before, induce antiproliferative activity against melanoma and breast cancer

The choice of the initial structure can be justified by the fact that there are no attempts of acetylene fragment formation as well as its further modification in the family of dammarane triterpenoids or pregnane steroids, especially among the closest analogues like panaxadiol and protopanaxadiol. Just in a few cases, changes were made based on the protopanaxadiol and protopanaxadiol. panaxadiol and protopanaxadiol. Just in a few cases, changes were made based on the ketones; different kinds of functionalities including hydroxy, fluoride, aldehyde, carboxy, ester, and enol ether were introduced for diversity of ring *D* [\[27](#page-15-3)[,28\]](#page-15-4).

Taking into account the above, we report herein a one-stage method of hollongdione modification into compounds with terminal alkyne and vinyl chloride fragments via the interaction with phosphorus halides. The obtained unsaturated hollongdiones were transformed to aminomethylated products and carboxamide.

2. Results

Procedure of compound **2**

A solution of 3*β*-hydroxy-dipterocarpol (0.440 g; 1.0 mmol) in AcOH (30 mL) was refluxed for 2 h; the reaction mixture was cooled to 0° C, and ozone was passed through it until the starting compound disappeared (TLC control). The mixture was then poured into H_2O (40 mL), removed by filtration, and washed with H_2O . The crude product was crystallized from benzene.

3*β*-Acetoxy-22,23,24,25,26,27-hexanor-dammar-20-one (**2**)

White solid, yield: 96%, mp: 126–127 °C, $[\alpha]^{20}$ _D = –60 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.59 (C18); 15.88 (C30); 16.31 (C19); 16.50 (C29); 18.15 (C6); 21.26 (C11); 21.31 (C2′); 23.71 (C2); 25.59 (C12); 25.98 (C16); 27.98 (C28); 30.06 (C21); 31.57 (C15); 35.49 (C7); 37.14 (C10); 37.92 (C4); 38.81 (C1); 40.53 (C8); 45.14 (C13); 50.07 (C14); 50.65 (C9); 54.27 (C17); 55.98 (C5); 80.87 (C3); 171.00 (C1'); 212.32 (C20). ¹H NMR (CDCl₃, δ ppm, *J* Hz): 0.83 (dd, 1H, 3 J_{5-6ax} = 11.7, 3 J_{5-6eq} = 2.6, H-5); 0.85 (s, 3H, H-29); 0.86 (s, 3H, H-28); 0.87 (s, 3H, H-30); 0.88 (s, 3H, H-19); 0.98 (s, 3H, H-18); 1.05 (td, 1H, ²J = 12.2, ³J_{1ax-2ax} = 12.2, $^3J_{1ax\text{-}2ax}$ = 5.6, H_{ax}-1); 1.16 (m, 1H, H_α-15); 1.20 (m, 1H, H_{ax}-12); 1.26 (m, 1H, H_{ax}-11); 1.29 (m, 1H, H_{eq}-7); 1.32 (dd, 1H, ³J_{9-11ax} = 11.8, ³J_{9-11eq} = 3.3, H-9); 1.47 (m, 1H, H_{ax}-6); 1.52 (m, 1H, H_{eq} -11); 1.53 (m, 1H, H_{eq} -6); 1.57 (m, 1H, H_{ax} -7); 1.63 (m, 1H, H_{ax} -2); 1.64 (m, 1H, Heq-12); 1.64 (m, 1H, Heq-2); 1.66 (m, 1H, Hβ-15); 1.70 (m, 1H, Heq-1); 1.71 (m, 1H, H_β-16); 1.92 (m, 1H, H-13); 1.93 (m, 1H, H_α-16); 2.04 (s, 3H, H-2'); 2.13 (s, 3H, H-21); 2.59 (td, 1H, ${}^{3}J_{17\text{-}13} = 10.9, {}^{3}J_{17\text{-}16\alpha} = 10.9, {}^{3}J_{17\text{-}16\beta} = 6.2$, H-17); 4.48 (dd, 1H, ${}^{3}J_{3\text{-}2\alpha\chi} = 10.7$, ${}^{3}J_{3\text{-2eq}} = 5.6, \text{H-3}.$

General procedure of compounds **3–8**

(a) Phosphoryl oxychloride (1 mL) was added to a solution of compound **1** or **2** (0.359 g or 0.403 g; 1.0 mmol) in 10 mL of anhydrous pyridine. The mixture was heated for 8 h under reflux, poured onto water, and extracted with chloroform $(3 \times 100 \text{ mL})$. The extract was washed with water $(3 \times 100 \text{ mL})$, dried over anhydrous calcium chloride, and evaporated under reduced pressure (water-jet pump). The residue was subjected to column chromatography on silica gel using eluent-petroleum ether-ethyl acetate (70:1→40:1). Compounds **6**, **7** were isolated in a mixture of products, with an overall yield of 87%.

3-Oxo-22,23,24,25,26,27-hexanor-dammar-20(21)-in (**3**)

White solid, yield: 73%, mp: 161–162 °C, $[\alpha]^{20}$ _D = +12 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.33 (C30); 15.34 (C18); 16.04 (C19); 19.62 (C6); 21.03 (C29); 21.77 (C11); 24.77 (C12); 26.75 (C28); 29.93 (C16); 31.09 (C15); 31.55 (C17); 34.08 (C2); 34.71 (C7); 36.92 (C10); 39.95 (C1); 40.25 (C8); 47.39 (C4); 49.24 (C13); 49.26 (C14); 50.11 (C9); 55.34 (C5); 67.92 (C21); 89.04 (C20); 217.87 (C3). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.81 (s, 3H, H-30); 0.95 (s, 3H, H-19); 1.01 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.08 (s, 3H, H-28); 1.15 (m, 1H, H_α-15); 1.18 (qd, 1H, ²J = 12.4, ³J_{12ax-13} = 12.4, ³J_{12ax-11ax} = 12.4, ³J_{12ax-11eq} = 4.6, H_{ax}-12); 1.32 (m, 1H, Heq-7); 1.35 (m, 1H, Hax-11); 1.37 (dd, 1H, ³ *J*5-6ax = 11.7, ³ *J*5-6eq = 2.0, H-5); 1.39 (dd, 1H, $^3J_{9\text{-11ax}} = 12.5, \frac{3}{J_{9\text{-11eq}}} = 2.6, \text{ H-9}; \, 1.46 \text{ (ddd, 1H, }^2J = 13.2, \frac{3}{J_{1ax\text{-}2ax}} = 9.5, \frac{3}{J_{1ax\text{-}2eq}} = 7.8, \text{H}_{ax}$ 1); 1.48 (m, 1H, H_{eq}-6); 1.57 (m, 1H, H_{ax}-6); 1.58 (m, 1H, H_{eq}-11); 1.59 (m, 1H, H_{ax}-7); 1.69 (m, 1H, H_β-16); 1.71 (m, 1H, H_β-15); 1.74 (ddd, 1H, ³J_{13-12ax} = 12.4, ³J₁₃₋₁₇ = 12.2, ³J_{13-12eq} = 4.1, H-13); 1.87 (m, 1H, Heq-12); 1.95 (ddd, 1H, ² *J* = 13.2, ³ *J*1eq-2ax = 7.6, ³ *J*1eq-2eq = 4.7, Heq-1); 2.05 (d, 1H, 4 *J*₂₁₋₁₇ = 2.4, H-21); 2.11 (m, 1H, H_α-16); 2.34 (dddd, 1H, 3 *J*₁₇₋₁₃ = 12.2, 3 *J*_{17-16α} = 9.0, ${}^{3}J_{17\text{-}16\beta} = 6.4, {}^{4}J_{17\text{-}21} = 2.4, H$ -17); 2.43 (ddd, 1H, ²J = 15.6, ${}^{3}J_{2eq\text{-}1ax} = 7.8, {}^{3}J_{2eq\text{-}1eq} = 4.7,$ H_{eq} -2); 2.51 (ddd, 1H, ²J = 15.6, ³J_{2ax-1ax} = 9.5, ³J_{2ax-1eq} = 7.6, H_{ax}-2).

3-Oxo-22,23,24,25,26,27-hexanor-dammar-20-chloro-20(21)-ene (**4**)

White solid, yield: 7%, mp: 148–149 °C, $[\alpha]^{20}$ _D = -30 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.30 (C18); 15.96 (C30); 16.08 (C19); 19.66 (C6); 21.04 (C29); 21.80 (C11); 24.46 (C12); 26.75 (C28); 27.48 (C16); 31.28 (C15); 34.11 (C2); 34.79 (C7); 36.95 (C10); 39.99 (C1); 40.44 (C8); 45.14 (C13); 47.43 (C4); 49.40 (C14); 49.79 (C17); 50.28 (C9); 55.41 (C5); 111.59 (C21); 146.83 (C20); 218.00 (C3). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.88 (s, 3H, H-30); 0.96 (s, 3H, H-19); 1.03 (s, 3H, H-18); 1.05 (s, 3H, H-29); 1.09 (s, 3H, H-28); 1.13 (m, 1H, H_{ax}-12); 1.15 (m, 1H, H_α-15); 1.31 (m, 1H, H_{ax}-11); 1.33 (m, 1H, H_{eq}-7); 1.38 (dd, 1H,

 ${}^{3}J_{5-6ax} = 11.7, {}^{3}J_{5-6eq} = 2.0, H-5$); 1.40 (dd, 1H, ${}^{3}J_{9-11ax} = 12.6, {}^{3}J_{9-11eq} = 2.6, H-9$); 1.46 (ddd, $1H$, $2J = 13.2$, $3J_{1ax-2ax} = 9.5$, $3J_{1ax-2eq} = 7.8$, H_{ax} -1); 1.47 (m, 1H, H_{eq} -6); 1.55 (m, 1H, H_{eq} -11); 1.56 (m, 1H, H_{ax}-6); 1.60 (m, 1H, H_{ax}-7); 1.63 (m, 1H, H_{eq}-12); 1.67 (m, 1H, H_β-15); 1.74 (m, 1H, H_β-16); 1.88 (ddd, 1H, ³J_{13-12ax} = 12.4, ³J₁₃₋₁₇ = 10.6, ³J_{13-12eq} = 3.7, H-13); 1.89 (m, 1H, H_{α} -16); 1.94 (ddd, 1H, ²J = 13.2, ³J_{1eq-2ax} = 7.6, ³J_{1eq-2eq} = 4.7, H_{eq}-1); 2.43 (ddd, 1H, ²J = 15.6, ${}^{3}J_{2eq\text{-}1ax} = 7.8, {}^{3}J_{2eq\text{-}1eq} = 4.7, \text{H}_{eq}$ -2); 2.49 (td, 1H, ${}^{3}J_{17\text{-}13} = 10.6, {}^{3}J_{17\text{-}16\alpha} = 10.6, {}^{3}J_{17\text{-}16\beta} = 6.2,$ H-17); 2.51 (ddd, 1H, ² *J* = 15.6, ³ *J*2ax-1ax = 9.5, ³ *J*2ax-1eq = 7.6, Hax-2); 5.10 (d, 1H, ² *J* = 1.2, H_A -21); 5.11 (d, 1H, ²J = 1.2, H_B-21).

3*β*-Acetoxy-22,23,24,25,26,27-hexanor-dammar-20-chloro-20(21)-ene (**6**)

White solid, yield: 80%, mp: 138–139 °C, $[\alpha]^{20}$ _D = -110 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.57 (C18); 16.02 (C30); 16.30 (C19); 16.50 (C29); 18.16 (C6); 21.28 (C11); 21.30 (C2′); 23.70 (C2); 24.42 (C12); 27.51 (C16); 27.98 (C28); 31.29 (C15); 35.38 (C7); 37.16 (C10); 37.90 (C4); 38.80 (C1); 40.54 (C8); 45.02 (C13); 49.41 (C14); 49.84 (C17); 50.83 (C9); 55.99 (C5); 80.86 (C3); 111.46 (C21); 146.93 (C20); 170.95 (C1'). ¹H NMR (CDCl₃, $δ$ ppm, *J* Hz): 0.84 (dd, 1H, 3 *J*_{5-6ax} = 11.5, 3 *J*_{5-6eq} = 1.8, H-5); 0.85 (s, 3H, H-29); 0.86 (s, 3H, H-28); 0.86 (s, 3H, H-30); 0.88 (s, 3H, H-19); 0.99 (s, 3H, H-18); 1.05 (ddd, 1H, $^2J = 11.8, \frac{3}{J_{1ax\text{-}2ax}} = 11.6, \frac{3}{J_{1ax\text{-}2eq}} = 4.7, \text{ H}_{ax}$ -1); 1.09 (qd, 1H, $^2J = 12.4, \frac{3}{J_{12ax\text{-}13}} = 12.4,$ ${}^{3}J_{12ax-11ax} = 12.4, {}^{3}J_{12ax-11eq} = 4.6, H_{ax}-12$; 1.12 (m, 1H, H_α-15); 1.23 (qd, 1H, ²J = 12.7, $^3J_{11ax\cdot9}$ = 12.6, $^3J_{11ax\cdot12ax}$ = 12.6, $^3J_{12ax\cdot11eq}$ = 4.6, H_{ax}-11); 1.29 (m, 1H, H_{eq}-7); 1.32 (dd, 1H, $^{3}J_{9\text{-11ax}}$ = 12.6, $^{3}J_{9\text{-11eq}}$ = 2.6, H-9); 1.46 (m, 1H, H_{ax}-6); 1.53 (m, 1H, H_{eq}-6); 1.54 (m, 1H, H_{eq} -11); 1.58 (m, 1H, H_{ax} -7); 1.60 (m, 1H, H_{eq} -12); 1.61 (m, 1H, H_{ax} -2); 1.64 (m, 1H, H_{eq} -2); 1.65 (m, 1H, H_β-15); 1.71 (m, 1H, H_{eq}-1); 1.74 (m, 1H, H_β-16); 1.84 (ddd, 1H, ³J_{13-12ax} = 12.4, $^3J_{13\text{-}17}$ = 10.6, $^3J_{13\text{-}12\text{eq}}$ = 3.7, H-13); 1.88 (m, 1H, H_α-16); 2.04 (s, 3H, H-2'); 2.47 (td, 1H, ${}^{3}J_{17\text{-}13} = 10.6, {}^{3}J_{17\text{-}16\alpha} = 10.6, {}^{3}J_{17\text{-}16\beta} = 6.2, \text{H-17}$; 4.48 (dd, 1H, ${}^{3}J_{3\text{-}2\alpha x} = 10.7, {}^{3}J_{3\text{-}2\text{eq}} = 5.8$, H-3); 5.08 (d, 1H, ²J = 1.2, H_A-21); 5.10 (d, 1H, ²J = 1.2, H_B-21).

3*β*-Acetoxy-22,23,24,25,26,27-hexanor-dammar-20(21)-in (**7**)

White solid, yield: 7%, mp: 149–150 °C, $[\alpha]^{20}$ _D = -61 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.41 (C30); 15.65 (C18); 16.29 (C19); 16.51 (C29); 18.15 (C6); 21.27 (C11); 21.31 (C2′); 23.72 (C2); 24.75 (C12); 28.00 (C28); 29.98 (C16); 31.13 (C15); 31.61 (C17); 35.32 (C7); 37.16 (C10); 37.92 (C4); 38.82 (C1); 40.39 (C8); 49.17 (C13); 49.32 (C14); 50.71 (C9); 55.97 (C5); 67.80 (C21); 80.86 (C3); 89.24 (C20); 170.98 (C1[']). ¹H NMR (CDCl₃, δ ppm, *J* Hz): 0.79 (s, 3H, H-30); 0.83 (dd, 1H, ³J_{5-6ax} = 11.5, ³J_{5-6eq} = 1.8, H-5); 0.85 (s, 3H, H-29); 0.85 (s, 3H, H-28); 0.88 (s, 3H, H-19); 0.97 (s, 3H, H-18); 1.05 (ddd, 1H, ²J = 11.8, ³J_{1ax-2ax} = 11.6, $^3J_{1ax\text{-}2eq}$ = 4.7, H_{ax}-1); 1.11 (m, 1H, H_{eq}-15); 1.16 (m, 1H, H_{ax}-12); 1.27 (m, 1H, H_{eq}-7); 1.29 (m, 1H, H_{ax}-11); 1.31 (dd, 1H, ³J_{9-11ax} = 12.6, ³J_{9-11eq} = 2.6, H-9); 1.46 (m, 1H, H_{ax}-6); 1.53 (m, 1H, H_{eq}-6); 1.56 (m, 1H, H_{ax}-7); 1.58 (m, 1H, H_{eq}-11); 1.61 (m, 1H, H_{ax}-2); 1.64 (m, 1H, H_{eq}-2); 1.68 (m, 1H, H_{eq}-16); 1.70 (m, 1H, H_{ax}-15); 1.71 (m, 1H, H_{eq}-1); 1.73 (m, 1H, H-13); 1.85 (m, 1H, H_{eq}-12); 2.03 (d, 1H, ⁴J₂₁₋₁₇ = 2.4, H-21); 2.04 (s, 3H, H-2[']); 2.10 (m, 1H, H_{ax}-16); 2.32 (tdd, 1H, $3\bar{J}_{17\text{-}13} = 10.6$, $3\bar{J}_{17\text{-}16\text{ax}} = 10.6$, $3\bar{J}_{17\text{-}16\text{eq}} = 6.2$, $4\bar{J}_{17\text{-}21} = 2.4$, H-17); 4.48 (dd, 1H, ${}^{3}J_{3\text{-2ax}} = 10.7, {}^{3}J_{3\text{-2eq}} = 5.8, H$ -3).

22,23,24,25,26,27-Hexanor-dammar-3,20-dichloro-20(21), 2(3)-diene (**8**)

White solid, yield: 8%, mp: 106–107 °C, $[\alpha]^{20}$ _D = –28 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.14 (C18); 15.98 (C30); 16.35 (C19); 20.01 (C29); 20.33 (C6); 21.71 (C11); 24.48 (C12); 27.54 (C16); 29.06 (C28); 31.30 (C15); 34.77 (C7); 36.42 (C10); 40.22 (C4); 40.35 (C8); 42.87 (C1); 45.17 (C13); 49.38 (C14); 49.44 (C9); 49.80 (C17); 54.37 (C5); 111.51 (C21); 122.07 (C2); 141.55 (C3); 146.94 (C20). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.87 (s, 3H, H-30); 0.92 $(s, 3H, H-19)$; 1.02 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.09 (qd, 1H, ²J = 12.4, ³J_{12ax-13} = 12.4, $^3J_{12ax\text{-}11ax}$ = 12.4, $^3J_{12ax\text{-}11eq}$ = 4.6, H_{ax}-12); 1.14 (s, 3H, H-28); 1.15 (m, 1H, H_α-15); 1.26 (dd, $1H$, $3J_{5-6ax} = 11.0$, $3J_{5-6eq} = 1.8$, H-5); 1.29 (qd, 1H, $^2J = 12.7$, $^3J_{11ax-9} = 12.6$, $^3J_{11ax-12ax} = 12.6$, ${}^{3}J_{12ax\text{-}11eq} = 4.6$, H_{ax}-11); 1.33 (m, 1H, H_{eq}-7); 1.36 (dd, 1H, ${}^{3}J_{9\text{-}11ax} = 12.6$, ${}^{3}J_{9\text{-}11eq} = 2.6$, H-9); 1.50 (m, 1H, H_{eq}-11); 1.55 (m, 1H, H_{ax}-6); 1.58 (m, 1H, H_{ax}-7); 1.58 (m, 1H, H_{eq}-6); 1.62 (m, 1H, H_{eq}-12); 1.65 (m, 1H, H_β-15); 1.71 (dd, 1H, ²J = 16.7, ³J_{1α-2} = 2.2, H_α-1); 1.75 (m, 1H, H_β-16); 1.87 (ddd, 1H, ${}^{3}J_{13\text{-}12ax} = 12.6$, ${}^{3}J_{13\text{-}17} = 10.9$, ${}^{3}J_{13\text{-}12eq} = 3.7$, H-13); 1.89

 $(m, 1H, H_{\alpha}-16)$; 2.09 (dd, 1H, ²J = 16.7, ³J_{1β-2} = 6.8, H_β-1); 2.49 (ddd, 1H, ³J₁₇₋₁₃ = 10.9, ${}^{3}J_{17\text{-}16\alpha} = 10.1, {}^{3}J_{17\text{-}16\beta} = 6.3, H-17$; 5.10 (d, 1H, ²J = 1.2, H_A-21); 5.11 (d, 1H, ²J = 1.2, H_B-21); 5.63 (dd, 1H, ${}^{3}J_{2-1\beta} = 6.8, {}^{3}J_{2-1\alpha} = 2.2, H-2$).

(b) Phosphoryl pentachloride (1.04 g; 5.0 mmol) was added to a solution of compound **1** or **2** (0.359 g or 0.403 g; 1.0 mmol) in 10 mL of anhydrous chloroform. The mixture was heated for 8 h under reflux, poured onto water, and extracted with chloroform $(3 \times 100 \text{ mL})$. The extract was washed with water $(3 \times 100 \text{ mL})$, dried over anhydrous calcium chloride, and evaporated under reduced pressure (water-jet pump). The residue was subjected to column chromatography on silica gel using eluent-petroleum ether-ethyl acetate (70:1→40:1). Compounds **6**, **7** were isolated in a mixture of products, with an overall yield of 77%.

3-Oxo-22,23,24,25,26,27-hexanor-dammar-20(21)-in (**3**)

White solid, yield: 14%, mp: 161–162 °C, $[\alpha]^{20}$ _D = +12 (c 0.05, CHCl₃).

3-Oxo-22,23,24,25,26,27-hexanor-dammar-20-chloro-20(21)-ene (**4**)

White solid, yield: 71%, mp: 148–149 °C, $[\alpha]_{\text{D}}^{20}$ = -30 (c 0.05, CHCl₃).

3-Chloro-22,23,24,25,26,27-hexanor-dammar-20-chloro-2(3),20(21)-diene (**5**)

White solid, yield: 8%, mp: 99–100 °C, $[\alpha]^{20}$ _D = +18 (c 0.05, CHCl₃). ¹H NMR (CDCl₃, δ ppm, *J* Hz): 0.87 (s, 3H, H-30); 0.92 (s, 3H, H-19); 1.02 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.09 (qd, 1H, 2 J = 12.4, 3 J_{12ax-13} = 12.4, 3 J_{12ax-11ax} = 12.4, 3 J_{12ax-11eq} = 4.6, H_{ax}-12); 1.14 (s, 3H, H-28); 1.15 (m, 1H, H_{eq}-15); 1.26 (dd, 1H, ³J_{5-6ax} = 11.0, ³J_{5-6eq} = 1.8, H-5); 1.29 (qd, $1H$, $2J = 12.7$, $3J_{11ax-9} = 12.6$, $3J_{11ax-12ax} = 12.6$, $3J_{12ax-11eq} = 4.6$, H_{ax} -11); 1.33 (m, 1H, H_{eq} -7); 1.36 (dd, 1H, ³J_{9-11ax} = 12.6, ³J_{9-11eq} = 2.6, H-9); 1.50 (m, 1H, H_{eq}-11); 1.55 (m, 1H, H_{ax}-6); 1.58 (m, 1H, H_{ax}-7); 1.58 (m, 1H, \dot{H}_{eq} -6); 1.62 (m, 1H, H_{eq}-12); 1.65 (m, 1H, H_{ax}-15); 1.71 $(dd, 1H, ²J = 16.7, ³J_{1α-2} = 2.2, H_α-1); 1.75 (m, 1H, H_{eq}-16); 1.87 (ddd, 1H, ³J_{13-12ax} = 12.6,$ ${}^{3}J_{13\text{-}17} = 10.9$, ${}^{3}J_{13\text{-}12\text{eq}} = 3.7$, H-13); 1.89 (m, 1H, H_{ax}-16); 2.09 (dd, 1H, ²J = 16.7, ${}^{3}J_{1\beta\text{-}2} = 6.8$, H_{β} -1); 2.49 (ddd, 1H, ³J₁₇₋₁₃ = 10.9, ³J_{17-16ax} = 10.1, ³J_{17-16eq} = 6.3, H-17); 5.10 (d, 1H, ²J = 1.2, $\rm H_A$ -21); 5.11 (d, 1H, ²J = 1.2, $\rm H_B$ -21); 5.63 (dd, 1H, $\rm{^{3}J_{2\text{-}1\beta}} = 6.8, \rm{^{3}J_{2\text{-}1\alpha}} = 2.2, \rm H$ -2). ¹³C NMR (CDCl3, δ ppm): 15.14 (C18); 15.98 (C30); 16.35 (C19); 20.01 (C29); 20.33 (C6); 21.71 (C11); 24.48 (C12); 27.54 (C16); 29.06 (C28); 31.30 (C15); 34.77 (C7); 36.42 (C10); 40.22 (C4); 40.35 (C8); 42.87 (C1); 45.17 (C13); 49.38 (C14); 49.44 (C9); 49.80 (C17); 54.37 (C5); 111.51 (C21); 122.07 (C2); 141.55 (C3); 146.94 (C20).

(c) A mixture of $2(0.403 \text{ g}; 1.0 \text{ mmol})$, $PCl₅(0.210 \text{ g}; 1.0 \text{ mmol})$, and a catalytic amount of DMAP in dry pyridine (5 mL) was refluxed for 2 h. After completion of the reaction, the mixture was poured onto water. The precipitate that formed was collected by filtration and washed with water. The residue was purified by $SiO₂$ column chromatography (eluent petroleum ether-ethyl acetate $(70:1 \rightarrow 40:1)$.

3*β*-Acetoxy-22,23,24,25,26,27-hexanor-dammar-20(21)-in (**7**)

White solid, yield: 89%, mp: 149–150 °C, $[\alpha]^{20}$ _D = -61 (c 0.05, CHCl₃).

General procedure of compounds **9a–c**

To a solution of compound **3** (0.204 g; 0.6 mmol) in 12 mL of dry dioxane was added paraformaldehyde (0.180 g, 6 mmol), 0.72 mmol of corresponding amine (diethylamine, *N*-methylpiperazine, or morpholine), NaOAc (0.250 g; 3 mmol), and CuI (6 mg; 0.03 mmol). The reaction mixture was stirred under argon for 10 h at 60 $°C$. After the reaction was completed by TLC, the mixture was diluted with water and extracted with CHCl₃ (3×20 mL), and the combined organic layer was washed with water, dried over $CaCl₂$, and evaporated under reduced pressure. The residue was purified by $SiO₂$ column chromatography (eluent petroleum ether-ethyl acetate $(40:1 \rightarrow 1:1)$, chloroform).

17-[3-(Diethylamino)-prop-1-yn-1-yl]-22,23,24,25,26,27-hexanor-dammar-3-one (**9a**)

White solid, yield: 78%, mp: 98–99 °C, $[\alpha]^{20}$ _D = -15 (c 0.05, CHCl₃). ¹³C NMR (CDCl₃, δ ppm): 12.21 (C3"); 15.39 (C18); 15.39 (C30); 16.08 (C19); 19.64 (C6); 21.03 (C29); 21.85 (C11); 24.92 (C12); 26.77 (C28); 30.19 (C16); 31.06 (C15); 31.90 (C17); 34.10 (C2); 34.70 (C7); 36.94 (C10); 39.98 (C1); 40.26 (C8); 41.07 (C1'); 47.14 (C2"); 47.41 (C4); 49.19 (C14); 49.42 (C13); 50.17 (C9); 55.35 (C5); 73.42 (C21); 89.88 (C20); 217.94 (C3).1H NMR (CDCl3, δ ppm, *J* Hz): 0.80 (s, 3H, H-30); 0.95 (s, 3H, H-19); 1.01 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.08 (s, 3H, H-28); 1.11 (t, 6H, 3] = 7.1, H-3''); 1.12 (m, 1H, H_α-15); 1.16 (m, 1H, H_{ax}-12); 1.32 (m, 1H, H_{ax}-11);

1.32 (m, 1H, H_{eq}-7); 1.37 (dd, 1H, ³J_{5-6ax} = 11.7, ³J_{5-6eq} = 2.6, H-5); 1.39 (dd, 1H, ³J_{9-11ax} = 12.6, $^3J_{9\text{-}11\text{eq}} = 2.6$, H-9); 1.46 (ddd, 1H, ²J = 13.2, $^3J_{1ax\text{-}2ax} = 9.6$, $^3J_{1ax\text{-}2\text{eq}} = 7.8$, H_{ax}-1); 1.48 (m, 1H, H_{eq}-6); 1.56 (m, 1H, H_{ax}-6); 1.57 (m, 1H, H_{eq}-11); 1.59 (m, 1H, H_{ax}-7); 1.63 (m, 1H, H_β-16); 1.68 (m, 1H, H_β-15); 1.69 (ddd, 1H, ³J_{13-12ax} = 11.9, ³J₁₃₋₁₇ = 11.4, ³J_{13-12eq} = 3.7, H-13); 1.83 (m, 1H, H_{eq}-12); 1.94 (ddd, 1H, ²J = 13.2, ³J_{1eq-2ax} = 7.6, ³J_{1eq-2eq} = 4.7, H_{eq}-1); 2.07 (m, 1H, H_α-16); 2.35 (ddd, 1H, ${}^{3}J_{17\text{-}13} = 11.4$, ${}^{3}J_{17\text{-}16\alpha} = 10.5$, ${}^{3}J_{17\text{-}16\beta} = 6.4$, H-17); 2.43 (ddd, $1H$, $2J = 15.6$, $3J_{2eq-1ax} = 7.8$, $3J_{2eq-1eq} = 4.7$, H_{eq} -2); 2.51 (ddd, $1H$, $2J = 15.6$, $3J_{2ax-1ax} = 9.6$, ${}^{3}J_{2ax\text{-}1eq}$ = 7.6, H_{ax}-2); 2.60 (q, 4H, ³J = 7.1, H-2''); 3.45 (s, 2H, H-1'). ¹⁵N NMR (CDCl₃, δ ppm): 46.35 (N1′′).

17-[3-(Pyrrolidin-1-yl)-prop-1-yn-1-yl]-22,23,24,25,26,27-hexanor-dammar-3-one (**9b**)

White solid, yield: 87%, mp: 100–102 °C, $[\alpha]^{20}$ _D = +67 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.36 (C30); 15.39 (C18); 16.07 (C19); 19.63 (C6); 21.03 (C29); 21.79 (C11); 24.09 (C3′′(4′′)); 24.94 (C12); 26.75 (C28); 30.02 (C16); 31.09 (C15); 31.86 (C17); 34.08 (C2); 34.71 (C7); 36.94 (C10); 39.97 (C1); 40.29 (C8); 43.45 (C1′); 47.41 (C4); 49.27 (C13); 49.29 (C14); 50.12 (C9); 52.14 (C2"(5")); 55.36 (C5); 73.53 (C21); 89.50 (C20); 217.83 (C3). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.81 (s, 3H, H-30); 0.95 (s, 3H, H-19); 1.01 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.08 (s, 3H, H-28); 1.15 (m, 1H, H_{α} -15); 1.19 (m, 1H, H_{ax} -12); 1.32 (m, 1H, H_{ax}-11); 1.32 (m, 1H, H_{eq}-7); 1.38 (dd, 1H, ³J_{5-6ax} = 11.8, ³J_{5-6eq} = 2.7, H-5); 1.39 (dd, 1H, $^{3}J_{9\text{-11ax}}$ = 12.7, $^{3}J_{9\text{-11eq}}$ = 2.7, H-9); 1.47 (m, 1H, H_{ax}-1); 1.48 (m, 1H, H_{eq}-6); 1.56 (m, 1H, H_{ax}-6); 1.57 (m, 1H, H_{eq}-11); 1.59 (m, 1H, H_{ax}-7); 1.65 (m, 1H, H_β-16); 1.71 (m, 1H, H_β-15); 1.74 (ddd, 1H, ³ *J*13-12ax = 11.9, ³ *J*13-17 = 11.4, ³ *J*13-12eq = 3.7, H-13); 1.82 (m, 1H, Heq-12); 1.94 $(\text{ddd}, 1\text{H}, ^2\text{J} = 13.2, ^3\text{J}_{1\text{eq-2ax}} = 7.6, ^3\text{J}_{1\text{eq-2eq}} = 4.7, \text{H}_{\text{eq}}-1)$; 1.96 (m, 4H, H-3"(4")); 2.08 (m, 1H, H_α-16); 2.36 (ddd, 1H, ${}^{3}J_{17\text{-}13} = 11.4$, ${}^{3}J_{17\text{-}16\alpha} = 10.5$, ${}^{3}J_{17\text{-}16\beta} = 6.4$, H-17); 2.43 (ddd, $1H$, $2J = 15.6$, $3J_{2eq-1ax} = 7.8$, $3J_{2eq-1eq} = 4.7$, H_{eq} -2); 2.51 (ddd, $1H$, $2J = 15.6$, $3J_{2ax-1ax} = 9.6$, ${}^{3}J_{2ax\text{-}1eq}$ = 7.6, H_{ax}-2); 2.97 (m, 4H, H-2"(5")); 3.63 (br.s, 2H, H-1').

17-[(3-Morpholino-prop-1-yn-1-yl]-22,23,24,25,26,27-hexanor-dammar-3-one (**9C**) White solid, yield: 82%, mp: 105–106 °C, $[\alpha]_{D}^{20} = -32$ (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.38 (C30); 15.40 (C18); 16.07 (C19); 19.64 (C6); 21.04 (C29); 21.80 (C11); 24.94 (C12); 26.77 (C28); 30.12 (C16); 31.08 (C15); 31.91 (C17); 34.10 (C2); 34.71 (C7); 36.95 (C10); 39.98 (C1); 40.29 (C8); 47.41 (C4); 47.74 (C1'); 49.23 (C13); 49.24 (C14); 50.14 (C9); 52.05 (C2"(4")); 55.37 (C5); 66.48 (C3"(5")); 74.83 (C21); 89.30 (C20); 217.87 (C3). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.81 (s, 3H, H-30); 0.95 (s, 3H, H-19); 1.00 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.08 (s, 3H, H-28); 1.13 (m, 1H, H_{α} -15); 1.17 (m, 1H, H_{ax} -12); 1.29 (m, 1H, Hax-11); 1.32 (m, 1H, Heq-7); 1.37 (dd, 1H, ³ *J*5-6ax = 11.7, ³ *J*5-6eq = 2.7, H-5); 1.39 (dd, 1H, $^{3}J_{9\text{-11ax}}$ = 12.7, $^{3}J_{9\text{-11eq}}$ = 2.7, H-9); 1.46 (m, 1H, H_{ax}-1); 1.48 (m, 1H, H_{eq}-6); 1.55 (m, 1H, H_{ax}-6); 1.57 (m, 1H, \dot{H}_{eq} -11); 1.59 (m, 1H, H_{ax}-7); 1.66 (m, 1H, H_β-16); 1.70 (m, 1H, H_β-15); 1.71 (ddd, 1H, ³ *J*13-12ax = 11.9, ³ *J*13-17 = 11.4, ³ *J*13-12eq = 3.7, H-13); 1.83 (m, 1H, Heq-12); 1.94 $(\text{ddd}, 1\text{H}, ^2\text{J} = 13.2, ^3\text{J}_{1\text{eq}-2\text{ax}} = 7.6, ^3\text{J}_{1\text{eq}-2\text{eq}} = 4.7, \text{H}_{\text{eq}}-1)$; 2.08 (m, 1H, H_α-16); 2.34 (ddd, 1H, ${}^{3}J_{17\text{-}13} = 11.4$, ${}^{3}J_{17\text{-}16\alpha} = 10.5$, ${}^{3}J_{17\text{-}16\beta} = 6.4$, H-17); 2.43 (ddd, 1H, ${}^{2}J = 15.6$, ${}^{3}J_{2eq\text{-}1ax} = 7.8$, ${}^{3}J_{2eq\text{-}1eq} = 4.7$, H_{eq}-2); 2.51 (ddd, 1H, ²J = 15.6, ${}^{3}J_{2ax\text{-}1ax} = 9.6$, ${}^{3}J_{2ax\text{-}1eq} = 7.6$, H_{ax}-2); 2.65 (m, 4H, H-2′′(6′′)); 3.36 (br.s, 2H, H-1′); 3.81 (m, 4H, H-3′′(5′′)).

General procedure of compounds **10** and **11**

Through a solution of compound **4** or **6** (0.377 g or 0.421 g; 1 mmol) in 50 mL of $(CH₃)₂CO:H₂O$ (20:1) solution, 2 eq. of ozone at room temperature was passed until the starting substance disappeared (TLC-control). The residue was subjected to column chromatography on SiO₂ using eluent-petroleum ether-ethyl acetate (10:1→1:1), chloroform.

3-Oxo-22,23,24,25,26,27-hexanor-dammar-17-carboxylic acid (**10**)

White solid, yield: 94%, mp: 135-137 °C, $[\alpha]^{20}$ _D = -48 (c 0.05, CHCl₃). ¹³C NMR (CDCl₃, δ ppm): 15.29 (C18); 15.55 (C30); 16.08 (C19); 19.64 (C6); 21.03 (C29); 21.71 (C11); 25.34 (C12); 26.26 (C16); 26.76 (C28); 31.53 (C15); 34.08 (C2); 34.87 (C7); 36.92 (C10); 39.97 (C1); 40.39 (C8); 45.56 (C17); 46.59 (C13); 47.42 (C4); 50.05 (C9); 50.06 (C14); 55.36 (C5); 181.26 (C20); 217.92 (C3). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.87 (s, 3H, H-30); 0.95 (s, 3H, H-19); 1.03 (s, 3H, H-18); 1.04 (s, 3H, H-29); 1.09 (s, 3H, H-28); 1.20 (ddd, 1H, ²J = 12.1, ³J_{15α-16β} = 8.6, ³J_{15α-16α} = 2.4,

 H_{α} -15); 1.25 (qd, 1H, ²J = 12.6, ³J_{12ax-13} = 12.6, ³J_{12ax-11ax} = 12.6, ³J_{12ax-11eq} = 4.6, H_{ax}-12); 1.32 (qd, 1H, $^{2}J = 12.2$, $^{3}J_{11ax-9} = 12.2$, $^{3}J_{11ax-12ax} = 12.2$, $^{3}J_{11ax-12eq} = 3.7$, H_{ax}-11); 1.35 (m, 1H, H_{eq}-7); 1.38 (dd, 1H, ³J_{5-6ax} = 11.7, ³J_{5-6eq} = 2.8, H-5); 1.41 (dd, 1H, ³J_{9-11ax} = 12.5, $^3J_{9\text{-}11\text{eq}} = 2.6$, H-9); 1.46 (ddd, 1H, ²J = 13.2, $^3J_{1ax\text{-}2ax} = 9.6$, $^3J_{1ax\text{-}2\text{eq}} = 7.8$, H_{ax}-1); 1.48 (m, 1H, H_{eq}^1 -6); 1.56 (m, 1H, H_{eq}-11); 1.57 (m, 1H, H_{ax}-6); 1.61 (m, 1H, H_{ax}-7); 1.72 (ddd, 1H, 2 J = 12.1, 3 J_{15β-16β} = 10.8, 3 J_{15β-16α} = 8.8, H_β-15); 1.81 (m, 1H, H_{eq}-12); 1.92 (m, 1H, H_β-16); 1.94 (ddd, 1H, ²J = 13.2, ³J_{1eq-2ax} = 7.6, ³J_{1eq-2eq} = 4.7, H_{eq}-1); 1.98 (ddd, 1H, ³J_{13-12ax} = 12.6, $^3J_{13\text{-}17}$ = 11.5, $^3J_{13\text{-}12\text{eq}}$ = 4.1, H-13); 2.00 (m, 1H, H_α-16); 2.43 (ddd, 1H, ²J = 15.6, $^3J_{2\text{eq-}1\text{ax}}$ = 7.8, $^3J_{\rm 2eq\text{-}1eq}$ = 4.7, H_{eq}-2); 2.47 (ddd, 1H, $^3J_{\rm 17\text{-}13}$ = 11.5, $^3J_{\rm 17\text{-}16\alpha}$ = 10.5, $^3J_{\rm 17\text{-}16\beta}$ = 6.5, H-17); 2.51 $(\text{ddd}, 1H, {}^{2}J = 15.6, {}^{3}J_{2ax\text{-}1ax} = 9.6, {}^{3}J_{2ax\text{-}1eq} = 7.6, H_{ax} - 2).$

3β-Acetoxy-22,23,24,25,26,27-hexanor-dammar-17-carboxylic acid (**11**)

White solid, yield: 91%, mp: 147–148 °C, $[\alpha]^{20}$ _D = +5 (c 0.05, CHCl₃). ¹³C NMR (CDCl₃, δ ppm): 15.56 (C18); 15.61 (C30); 16.30 (C19); 16.49 (C29); 18.14 (C6); 21.19 (C11); 21.29 (C2'); 23.69 (C2); 25.28 (C12); 26.25 (C16); 27.97 (C28); 31.54 (C15); 31.92 (C2); 35.46 (C7); 37.13 (C10); 37.90 (C4); 38.80 (C1); 40.50 (C8); 45.61 (C13); 46.51 (C14); 50.07 (C17); 50.63 (C9); 55.99 (C5); 80.85 (C3); 170.99 (C1′). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.81 (dd, 1H, 3 *J*5-6ax = 11.5, ³ *J*5-6eq = 1.8, H-5); 0.84 (s, 3H, H-29); 0.87 (s, 3H, H-28); 0.90 (s, 3H, H-30); 0.92 (s, 3H, H-19); 0.99 (s, 3H, H-18); 1.02 (ddd, 1H, ²J = 11.8, ³J_{1ax-2ax} = 11.6, ³J_{1ax-2eq} = 4.7, H_{ax} -1); 1.09 (qd, 1H, ²J = 12.4, ³J_{12ax-13} = 12.4, ³J_{12ax-11ax} = 12.4, ³J_{12ax-11eq} = 4.6, H_{ax}-12); 1.12 (m, 1H, H_α-15); 1.23 (qd, 1H, ²J = 12.7, ³J_{11ax-9} = 12.6, ³J_{11ax-12ax} = 12.6, ³J_{12ax-11eq} = 4.6, Hax-11); 1.29 (m, 1H, Heq-7); 1.32 (dd, 1H, ³ *J*9-11ax = 12.6, ³ *J*9-11eq = 2.6, H-9); 1.46 (m, 1H, H_{ax}-6); 1.53 (m, 1H, H_{eq}-6); 1.54 (m, 1H, H_{eq}-11); 1.58 (m, 1H, H_{ax}-7); 1.60 (m, 1H, H_{eq}-12); 1.61 (m, 1H, H_{ax}-2); 1.64 (m, 1H, H_{eq}-2); 1.65 (m, 1H, H_β-15); 1.71 (m, 1H, H_{eq}-1); 1.74 (m, 1H, H_β-16); 1.84 (ddd, 1H, ³J_{13-12ax} = 12.4, ³J₁₃₋₁₇ = 10.6, ³J_{13-12eq} = 3.7, H-13); 1.88 (m, 1H, H_{α} -16); 2.04 (s, 3H, H-2'); 2.48 (td, 1H, $^{3}J_{17\text{-}13} = 10.6$, $^{3}J_{17\text{-}16\alpha} = 10.6$, $^{3}J_{17\text{-}16\beta} = 6.2$, H-17); 4.50 (dd, $1H$, $3J_{3-2ax} = 10.7$, $3J_{3-2eq} = 5.8$, H-3).

Procedure of compound **12**

Through a solution of the acid 11 (0.406 g; 1 mmol) in 50 mL of anhydrous CH_2Cl_2 , 2 eq. of ozone at $-40\degree$ C was passed until the starting substance disappeared (TLC-control). The resulting acid chloride was dissolved in anhydrous CH_2Cl_2 (30 mL) and treated with pyrrolidine (0.150 g; 1.5 mmol) and three drops of $Et₃N$. The mixture was stirred at room temperature for 3 h, washed with 5% HCl solution (2×100 mL) and H₂O (100 mL), dried over CaCl₂, and the solvent was removed under reduced pressure. The product was purified by column chromatography with $SiO₂$ using CHCl₃ and a mixture of CHCl₃–EtOH (100:1) as eluents.

3β-Acetoxy-22,23,24,25,26,27-hexanor-dammar-17-*N*-pyrrolidine amide (**12**)

White solid, yield: 89%, mp: 114–115 °C, $[\alpha]^{20}$ _D = +23 (c 0.05, CHCl₃). ¹³C NMR (CDCl3, δ ppm): 15.69 (C18); 16.01 (C30); 16.34 (C19); 16.50 (C29); 18.15 (C6); 21.26 (C11); 21.34 (C2′); 23.70 (C2); 24.36 (C4′′); 25.54 (C12); 26.11 (C3′′); 26.60 (C16); 27.97 (C28); 31.66 (C15); 35.53 (C7); 37.12 (C10); 37.89 (C4); 38.80 (C1); 40.53 (C8); 45.06 (C17); 45.81 (C5′′); 45.89 (C13); 46.48 (C2′′); 49.63 (C14); 50.77 (C9); 55.99 (C5); 80.94 (C3); 171.08 (C1′); 175.10 (C20). ¹⁵N NMR (CDCl3, δ ppm): 127.29 (N1′′). ¹H NMR (CDCl3, δ ppm, *J* Hz): 0.83 (dd, 1H, ³ *J*5-6ax = 11.7, ³ *J*5-6eq = 2.6, H-5); 0.84 (s, 3H, H-29); 0.85 (s, 3H, H-28); 0.86 (s, 3H, H-19); 0.87 (s, 3H, H-30); 1.01 (s, 3H, H-18); 1.04 (td, 1H, ²J = 12.2, ³J_{1ax-2ax} = 12.2, ³J_{1ax-2ax} = 5.6, H_{ax} -1); 1.15 (m, 1H, H_α-15); 1.17 (m, 1H, H_{ax}-12); 1.27 (m, 1H, H_{ax}-11); 1.30 (m, 1H, H_{eq}-7); 1.57 (m, 1H, H_{ax}-7); 1.31 (dd, 1H, ³J_{9-11ax} = 11.8, ³J_{9-11eq} = 3.3, H-9); 1.47 (m, 1H, H_{ax}-6); 1.49 (m, 1H, H_{eq}-11); 1.53 (m, 1H, H_{eq}-6); 1.57 (m, 1H, H_{ax}-7); 1.62 (m, 1H, H_{ax}-2); 1.64 (m, 1H, H_{eq}-12); 1.64 (m, 1H, H_{eq}-2); 1.71 (m, 1H, H_{eq}-1); 1.74 (m, 1H, H_β-15); 1.77 (m, 1H, H_β-16); 1.84 (m, 2H, H-4''); 1.92 (m, 1H, H_α-16); 1.92 (m, 2H, H-3''); 3.44 (s, 3H, H-2''); 2.16 (m, 1H, H-13); 2.52 (td, 1H, ${}^{3}J_{17\text{-}13} = 10.9, {}^{3}J_{17\text{-}16\alpha} = 10.9, {}^{3}J_{17\text{-}16\beta} = 6.2$, H-17); 3.44 (s, 3H, H-2''); 3.47 $(m, 1H, H-5'')$; 4.47 (dd, 1H, $3J_{3-2ax} = 10.7, 3J_{3-2eq} = 5.6, H-3$).

3. Discussion 3. Discussion 3. Discussion

We used hollongdiones 1 and 2 (for X-ray crystal structure, see Figure [3,](#page-7-0) for NMR data see Supplementary Material Figures S1-S8) in the reactions with POCl₃ and PCl₅ (Scheme [1\)](#page-7-1). As a result, a series of derivatives 3-8 was obtained and identified.

Figure 3. The molecular structure of compound **2**. **Figure 3.** The molecular structure of compound **2**. **Figure 3.** The molecular structure of compound **2**.

Scheme 1. Reagents and conditions: a. POCl₃/Py, Δ , 8 h; b. PCl₅/CHCl₃, Δ , 8 h; c. PCl₅/Py, DMAP (cat.), ∆, 2 h. (cat.), ∆, 2 h. (cat.), ∆, 2 h.

Alkyne 3 was synthesized by the interaction of hollongdione 1 with POCl_3 in pyridine, with the yield of 73% after purification by column chromatography, while the vinyl chloride 4 was also isolated as the minor (7%) product. The alkyne synthesis can be explained by dehydrohalogenation of a *gem*-dihalogen intermediate formation during thermal activation under reaction conditions; vinyl chloride's mechanism includes the cleavage of one HCl molecule step. It is worth noting that the composition of the products was different when lupane triterpenoids were used as the initial scaffolds: the presence of a molecule with a double C=C bond and tetrahydrofuran ring and a compound with a trichloroacetyl group besides the main alkyne derivative were confirmed, which differs from previous results [\[15\]](#page-14-13). Moreover, the *abeo*-lupene fragment was obtained under the action of POCl₃ in pyridine [\[29](#page-15-5)[,30\]](#page-15-6).

At the same time, the reaction of 1 with PCl_5 in CHCl₃ led to the vinyl chloride 4 as the main product (71%); alkyne 3 was obtained among the by-products (14%), and ring A was \sim transformed into a 3-chloro-2(3)-ene derivative $5(8\%)$, which could be explained as a result of the action of the Vilsmeyer-Haack reagent on the C3-oxo-group. The application of phosphorus oxychloride along with DMF to friedelin as the substrate with chlorofriedel-2 and 3-chlorofriedel-3-enes as the products was demonstrated [\[31\]](#page-15-7).

The protection of the C3 position by the acetoxy group (compound 2) made it possible to avoid the formation of A-ring by-products. In the case of reaction with POCl₃, compounds 6 and 7 were isolated only as the mixture of products, with the overall yield of 87%, and 8 became a new by-product (5%). The overall yield of compounds 6 and 7 reached 77% when the halogenating reagent was PCl₅ (Conditions *b* at Scheme [1\)](#page-7-1). Previously, the optimization of the synthetic strategy for the lup-20(30)-yne preparation by adding a catalytic amount of DMAP was proposed [\[30\]](#page-15-6). An attempt to streamline the synthesis in this way was also successful, yielding only compound 7 with 89% after purification by \mathbf{column} chromatography.

The structure of all compounds was determined by ¹H and ¹³C NMR spectroscopy data. For compound 3, the presence of the oxo-group at the C3 position was confirmed by HMBC cross-peaks of *gem*-dimethyl groups C28 (δ _H 1.08 ppm) and C29 (δ _H 1.04 ppm) along with δ_C 217.87 ppm. Also, HMBC cross-peaks of methylene protons at *C*1 and *C*2 were observed for the C3-oxo-group. For the ethynyl substituent at C17 with δ_C 89.04 ppm of the quaternary carbon atom and δ _C 67.33 ppm and δ _H 2.05 ppm of the methine group, a number of interactions were observed, confirming the position of substitution. Thus, HMBC proton cross-peaks of H-13, H-17, and H_{β} -16, with quaternary position and cross-peak H-17/C21 were confirmed. In addition, the acetylene proton signal has a doublet with the value 4 J = 2.4 Hz, with a methine proton *H*-17, which is confirmed in the spectrum COSY-DQF and its response splitting (Figure [4\)](#page-8-0), (Supplementary Material Figures S9–S16). S16).

Figure 4. The key NMR assignments and significant {¹H, ¹³C} HMBC, {¹H, ¹H} COSY, and NOESY correlations of compounds **3**–**7**. correlations of compounds **3**–**7**.

For the vinyl chloride group in compounds $4-7$ with characteristic values $\delta_C \sim 147$ ppm of the quaternary carbon atom *C*20 and δ_C ~112 ppm for the terminal methylene group, HMBC interactions with cycle *D* protons were observed, which confirms the position of substitution C17. In particular, HMBC cross-peaks of proton H_{α} -16, H_{β} -16, and H-13 with 13 with quaternary carbon atom *С*20 and НМВС cross-peaks of terminal protons with quaternary carbon atom *C*20 and HMBC cross-peaks of terminal protons with carbon signal C17 at δ_C ~50 ppm were observed. Additionally, in the NOESY spectrum, cross-peaks were observed between one of the terminal protons with H_{eq} -12, H -13 and H -17. The presence of acetate in the *C*3 position of cycle *A* for compound **4** is confirmed by the HMBC cross peak of the doublet-doublet signal *H*-3 (δ_H 4.48 ppm) with a carboxyl group signal at δ_C 170.95 [pp](#page-8-0)m (Figure 4).

For compound **5**, the presence of *C*2 = C3 double bond in cycle *A* was confirmed on For compound **5**, the presence of *C*2 = C3 double bond in cycle *A* was confirmed on the basis of HMBC cross-peaks *gem*-dimethyl *C*28 and *C*29 groups (δ_H 1.14 and 1.04 ppm, respectively) with a quaternary *Cl*-bearing position at δ_C 141.55 ppm, and *H*-2 protons ($δ$ _H 5.63 ppm) and *α*, $β$ -protons of the methylene group C1 ($δ$ _H 1.71, 2.09 ppm). A double

bond methine proton *H*-2 (δ_H 5.63 ppm) was represented by doublet-doublet splitting with $3J = 6.8$ and 2.2 Hz on neighboring methylene protons at *C*1. The position of the carbon signal C1 at δ_C 42.87 ppm was localized on the basis of HMBC cross-peaks with methine protons H -5 ($\delta_{\rm H}$ 1.26 ppm) and H -9 ($\delta_{\rm H}$ 1.36 ppm) and methyl group protons at C19 ($\delta_{\rm H}$ 0.92 ppm). The *α*-orientation of protons at *C*1 (δ_H 1.71 ppm) was confirmed by NOESY cross-peaks with *H*-5 and *H*-9; for the protons H_{β} -1, NOE interactions with H_{3} -19 and H_{eq} -11 ($\delta_{\rm H}$ 1.50 ppm) protons were also observed (Figure 5), (Supplementary Material Figures S17–S40). Protons were also observed (Figure 5), $\frac{1}{2}$ and $\frac{1}{2}$ a

Figure 5. The key NMR assignments and significant {¹H, ¹³C} HMBC, {¹H, ¹H} COSY, and NOESY **Figure 5.** The key NMR assignments and significant {1H, ¹³C} HMBC, {1H, ¹H} COSY, and NOESY correlations of compounds **5**, **8**. The oxygen atom is highlighted in red color in the structure of correlations of compounds **5**, **8**. The oxygen atom is highlighted in red color in the structure of compound **8**. compound **8**.

The formation of a 1,2-dichloro-1-hydroxyethyl group at the *C*17 of compound **8** is The formation of a 1,2-dichloro-1-hydroxyethyl group at the *C*17 of compound **8** is confirmed by the characteristic ¹³C NMR values: δ _C 97.32 ppm for the quaternary position *C*20 and δ_C 54.62 ppm for the methylene group *C*21. For *OH, Cl*-bearing quaternary carbon, three-bond HMBC correlations with protons H_{α} -16, H_{β} -16 ($\delta_{\rm H}$ 2.00 and 1.76 ppm, respectively), and *H*-13 (δ_H 1.95 ppm) are observed. The HMBC spectrum also contains cross peaks of diastereotopic methylene protons at *C*21 (δ_H 4.01 and 4.05 ppm) with carbons *C*20 and *C*17 (δ _C 51.18 ppm), and the protons themselves in the ¹H NMR spectrum are sented as two doublets with a geminal constant of 12.2 Hz. presented as two doublets with a geminal constant of 12.2 Hz.

The molecular structures of 3-oxo-22,23,24,25,26,27-hexanor-dammar-20(21)-in **3**, 3- The molecular structures of 3-oxo-22,23,24,25,26,27-hexanor-dammar-20(21)-in **3**, 3 oxo-22,23,24,25,26,27-hexanor-dammar-20-chlorо-20(21)-en **4,** and 22,23,24,25,26,27-hexa-oxo-22,23,24,25,26,27-hexanor-dammar-20-chloro-20(21)-en **4,** and 22,23,24,25,26,27-hexanordammar-3,20-dichloro-20(21), 2(3)-dien **5** were determined by X-ray analysis (Figure [6\)](#page-10-0). All the compounds have typical four-cycle moiety as in hollongdione [\[20\]](#page-14-18), including three sixand one five-membered trans-fused cycles. All the six-membered cycles in **3**, **4**, and **5** adopt chair conformation, except the chlorine substituted one in **5** with double bond C2 = C3 equaling 1.307(4) Å, which has a distorted half-chair conformation. The five-membered $\frac{1}{2}$ cycles show twist conformation in all the compounds. The length of the C≡C bond in 3 slightly differs, being the same within the limit of experimental accuracy (Table [1\)](#page-10-1) and very
all the same within the limit of experimental accuracy (Table 1) and very close to the average literature value of 1.174(11) [\[32\]](#page-15-8). The alkynyl substituent on C17 in **3** is in **3** is not ideally linear, so the angle C17C20C21 is not the 180° that is usual for such The bond lengths and orientation of the vinyl chloride group on C17 in **4** and **5**, describing moieties. The bond lengths and orientation of the vinyl chloride group on C17 in **4** and **5**, by torsion angle C16C17C20Cl1(Cl2), are the same in both structures (Table [1\)](#page-10-1). There are σ y torsion angle C16C17C20C11(Cl2), are the same in both structures (Table 1). There are not any specific intermolecular interactions in the crystals under consideration. The crystal There are not any specific intermolecular interactions in the crystals under consideration. structure of compounds is stabilized by van der Waals interactions. The main XRD data T_{total} structure of compounds μ states is stabilized by van der Waals interactions. The main value data XRD data and experimental details for compounds **2**–**5** are presented in Table 2. and experimental details for compounds **2**–**5** are presented in Table [2.](#page-11-0)not ideally linear, so the angle C17C20C21 is not the 180 $^{\circ}$ that is usual for such moieties.

* geometrical parameters for two independent molecules in **3**.

Figure 6. Molecular structures of compounds **3**, **4**, and **5**. **Figure 6.** Molecular structures of compounds **3**, **4**, and **5**.

	Table 1. Selected geometric parameters of compounds 3-5.	

* geometrical parameters for two independent molecules in 3.

	$\overline{2}$	3	$\overline{\mathbf{4}}$	5
Empirical formula	$C_{26}H_{42}O_3$	$C_{24}H_{35}O$	$C_{24}H_{37}ClO$	$C_{24}H_{36}Cl_2$
Formula weight	402.59	339.52	376.98	395.43
Crystal system	Monoclinic	Monoclinic	Triclinic	Monoclinic
Space group	$P2_1$	$P2_1$	P1	C ₂
Unit cell dimensions a, $b, c(\AA)$	25.332(2), 7.6508(5), 25.543(2)	$7.5834(6)$, $20.548(2)$, 12.9331(9)	$6.3333(3)$, $7.4257(5)$, 11.8452(8)	$13.880(1)$, 6.3288(4), 24.409(2)
α , β , γ (\circ)	90, 107.232(3), 90	90, 90.020(4), 90	89.026(2), 84.321(2), 69.960(2)	90, 97.004(4), 90
Volume (\AA^3)	4728.1(6)	2015.3(3)	520.69(6)	2128.1(3)
Ζ	$\,8\,$	4	$\mathbf{1}$	$\overline{4}$
Density (calcd.) $(Mg.mm^{-3})$	1.131	1.119	1.202	1.234
F(000)	1776	748	206	856
Absorption coefficient (mm^{-1})	0.07	0.07	0.19	0.311
Crystal size $(mm3)$	$1.00 \times 0.08 \times 0.05$	$1.00 \times 0.45 \times 0.01$	$1.00 \times 0.72 \times 0.20$	$1.00 \times 0.30 \times 0.02$
$T_{\rm min}$, $T_{\rm max}$	0.832, 0.928	0.770, 0.928	0.764, 0.813	0.762, 0.801
Θ range for data collection	$0.8 - 26.5$	$1.0 - 26.1$	$2.9 - 33.5$	$0.8 - 27.1$
Reflections collected	101,280	19,801	14,660	35,210
Independent reflections $[R_{\text{int}}]$	19,379 [0.088]	7303 [0.095]	6349 [0.024]	4471 [0.049]
Observed $[I > 2 \sigma (I)]$ reflections	11,585	4090	5142	4097
Completeness to $\theta = 25^{\circ}$ (%)	99.9	99.7	99.9	98.8
Data/restraints/parameters	193,79/1/1073	7303/2/462	6349/3/240	4471/1/240
Goodness-of-fit on F^2	1.01	0.99	1.02	1.107
Final R1, $[I > 2\sigma(I)]$	0.061	0.064	0.046	0.041
Final wR2 (all data)	0.155	0.148	0.138	0.109
Largest diff. peak/hole $e \text{A}^{-3}$	$0.20, -0.21$	$0.18, -0.17$	$0.36, -0.27$	$0.27, -0.201$
Absolute structure parameter	$-0.1(5)$	0.5(10)	0.039(17)	0.029(17)
CCDC	2,315,012	2,315,013	2,315,014	2,350,691

Table 2. XRD data and experimental details for compounds **2**–**5**.

It is known that alkynes are simple and valuable precursors that act as electrophiles due to the presence of $π$ -bonds and can be successfully utilized for the construction of various classes of carbocycles, heterocycles, complex molecular architectures, and natural products [\[33\]](#page-15-9). For example, the terminal carbon–carbon triple bond is an activating functional group that is required to render the substrate active in the Mannich reaction. In the last decade, new propargylamines were synthesized by the interaction of 19- and 28-alkynyltriterpenoids with *N*-methylpiperazine [\[34\]](#page-15-10). Propargylated oleanolic and glycyrrhetic acids were involved in aminomethylation by the Mannich reaction [\[35\]](#page-15-11). Moreover, this type of modification has been studied on the basis of steroid alkynes, and the interest keeps growing [\[36\]](#page-15-12).

Taking into account the above, we involved alkyne **3** in the Mannich reaction with diethylamine, pyrrolidine, or morpholine, giving the conjugates 9a-c in the yields of 78-87% (Scheme [2\)](#page-12-0).

Scheme 2. Reagents and conditions: a. diethylamine, pyrrolidine, or morpholine, paraformaldehyde, hyde, NaOAc, CuI, 1,4-dioxane, 10 h, 60 °C. NaOAc, CuI, 1,4-dioxane, 10 h, 60 ◦C.

The formation of aminomethylated products $9a-c$ is confirmed by NMR spectra due to the presence of methylene group signals in the range δ_C 41.07 \div 47.74 ppm (δ_H) $13.36 \div 3.62$ ppm), for which HMBC cross-peaks with carbon signals of the acetylene group (δ C 73.42÷74.89 and δ C 89.28 ÷ 89.88 ppm) are observed. In addition, cross-peaks of protons of the *α*-position of amines (δ _H 2.60 ÷ 2.69 ppm) with the carbon signal of the newly formed methylene group *C*1' are observed in the HMBC spectrum (Supplementary Material Figures S41–S63).

In the next stage of our research, we decided to involve the hollongdione vinyl chloride for transformation to carboxylic acid. It is known that androstane methylketones are transformed to carboxylic acids by a haloform reaction [37,38][. O](#page-15-13)[n th](#page-15-14)e other hand, dichlorovinyl insecticides by ozone oxidation in water-containing solvents afford carboxy-derivatives [\[39\]](#page-15-15).

Vinyl chloride is a well-known monomer for the synthesis of copolymers [\[40\]](#page-15-16), but no instances of its oxidation to acid have been found in the case of triterpenoid or steroid scaf-instances of its oxidation to acid have been found in the case of triterpenoid or steroid folds. We propose here a new approach for the synthesis of acids from C17-methylketones. For this, vinyl chlorides **4** and **6** were oxidized with ozone in acetone-water medium to tones. For this, vinyl chlorides **4** and **6** were oxidized with ozone in acetone-water medium give acids **10** and **11**. So, these acids could be called "hollongdionoic acid"-like, androstane-17-carboxylic acids were given the trivial name "etianic acids" [\[41\]](#page-15-17). One-pot synthesis of amide **12** in the yield of 89% was realized by the oxidation of vinyl chloride **6** in CH₂Cl₂ followed by the addition of pyrrolidine [\(Sc](#page-12-1)heme 3).

Scheme 3. Reagents and conditions: a. O_{3} , (CH₃)₂C(O):H₂O (20:1), rt; b. *i* O₃, CH₂Cl₂, -40 °C, *ii* pyrrolidine, CH_2Cl_2 , rt, 3 h.

4. Materials and Methods

The formation of a carboxyl group at the C17 position of compound 10 was established based on the presence of a signal at δ_C 181.26 ppm in the ¹³C NMR spectrum. The HMBC spectrum showed cross-peaks of the *H*-13, H_β -16, and *H*-17 (δ _H 1.98, 1.92, and 2.47 ppm, respectively) protons of cycle *D* with a quaternary carbon signal at δ_C 181.26 ppm, (Supplementary Material Figures S64–S79).

4. Materials and Methods

NMR spectra were recorded on a Bruker "*Avance-III*" 500 MHz spectrometer (Bruker, Billerica, MA, USA), (500, 125, and 50 MHz for ¹H, ¹³C, and ¹⁵N, respectively, δ, ppm, *J*, Hz) in CDCl₃ with tetramethylsilane as the internal standard. Mass spectra were obtained on a liquid chromatograph–mass spectrometer LCMS-2010 EV (Shimadzu, Kyoto, Japan). Melting points were detected on the micro table "Rapido PHMK05" (Nagema, Dresden, Germany). Optical rotations were measured on the polarimeter "Perkin-E lmer 241 MC" (Perkin Elmer, Waltham, MA, USA) in a tube length of 1 dm. Thin-layer chromatography analyses were performed on Sorbfil plates (Sorbpolimer, Krasnodar, Russian Federation) using the solvent system chloroform-ethyl acetate, 40:1. Substances were detected by 10% H_2SO_4 with subsequent heating to 100–120 °C for 2–3 min. The X-ray diffraction experiments for compounds **2**–**5** were carried out at 296(2) K on a Bruker KAPPA APEX II diffractometer (graphite-monochromated Mo Kα radiation). Reflection intensities were corrected for absorption by the SADABS-2016 program [\[42\]](#page-15-18). The structure of compounds was solved by direct methods using the SHELXT-2014 program [\[43\]](#page-15-19) and refined by the anisotropic (isotropic for all H atoms) full-matrix least-squares method against *F* ² of all reflections by SHELXL-2018 [\[44\]](#page-15-20). The positions of the hydrogen atoms were calculated geometrically and refined in the riding model. The asymmetric unit of **2** contained four molecules, the unit of **3**—two molecules, and structure of this compound was refined as a twin. Crystallographic data for **2**–**5** have been deposited at the Cambridge Crystallographic Data Centre as supplementary publication Nos. CCDC 2315012, 2315013, 2315014, and 2350691. A copy of the data can be obtained, free of charge, on application to the CCDC, 12 Union Road, Cambridge CB21EZ, UK (fax: +44 122 3336033 or e-mail: deposit@ccdc.cam.ac.uk; internet: [www.ccdc.cam.ac.uk,](www.ccdc.cam.ac.uk) accessed on 14 October 2023). Compound **1** was obtained according to the method described previously [\[20\]](#page-14-18).

5. Conclusions

Hollongdione is the naturally occurring "triterpenoid-steroid" hybrid. Its design and synthesis is deemed to be very interesting from the perspective of potent activities. Its reactivity with an emphasis on dehydrohalogenation reactions such as phosphorus-chlorine derivatives (POCl3, PCl5) was studied for the first time. As a result, compounds with *C*17 alkynyl and vinyl chloride substituents were synthesized, the structures of which were confirmed by NMR spectra and X-ray analysis. The choice of a dehydrohalogenating agent made it possible to regulate the structure, composition, and the yield of the products. Hollongdione C17-alkyne was successfully transformed into aminomethylated products by the Mannich reaction. An opportunity for a one-stage pathway from hollongdione vinyl chloride to the carboxylic acid with the following amide preparation is also demonstrated.

Supplementary Materials: The following supporting information can be downloaded at: [https:](https://www.mdpi.com/article/10.3390/ijms25158356/s1) [//www.mdpi.com/article/10.3390/ijms25158356/s1.](https://www.mdpi.com/article/10.3390/ijms25158356/s1)

Author Contributions: Conceptualization, O.K.; methodology, O.K. and I.S.; validation, I.S. and Z.G.; formal analysis, A.L., T.R. and D.P.; investigation, A.L. and T.R.; writing—original draft preparation, O.K., I.S. and Z.G.; writing—review and editing, O.K. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the Federal Program No. 1021062311392-9-1.4.1 and No. 123011300044-5.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data available within the article or its Supplementary Materials.

Acknowledgments: The authors would like to acknowledge the Center for the Collective Use "Chemistry" of the Ufa Institute of Chemistry of the UFRC RAS, the RCCU "Agidel" of the UFRC RAS, and the "Chemical service center for collective use" of the SB RAS for carrying out spectral and analytical measurements.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Chávez-Hernández, A.L.; Sánchez-Cruz, N.; Medina-Franco, J.L. Fragment library of natural products and compound databases for drug discovery. *Biomolecules* **2020**, *10*, 1518. [\[CrossRef\]](https://doi.org/10.3390/biom10111518) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33172012)
- 2. Siddiq, A.; Dembitsky, V. Acetylenic anticancer agents. *Anti-Cancer Agents Med. Chem.* **2008**, *8*, 132–170. [\[CrossRef\]](https://doi.org/10.2174/187152008783497073) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18288919)
- 3. Pecak, P.; Świtalska, M.; Chrobak, E.; Boryczka, G.; Bebenek, E. Betulin Acid Ester Derivatives Inhibit Cancer Cell Growth by Inducing Apoptosis through Caspase Cascade Activation: A Comprehensive In Vitro and In Silico Study. *Int. J. Mol. Sci.* **2022**, *24*, 196. [\[CrossRef\]](https://doi.org/10.3390/ijms24010196) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36613643)
- 4. Heller, L.; Schwarz, S.; Obernauer, A.; Csuk, R. Allobetulin derived seco-oleananedicarboxylates act as inhibitors of acetylcholinesterase. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 2654. [\[CrossRef\]](https://doi.org/10.1016/j.bmcl.2015.04.086)
- 5. Pereira, V.V.; Pereira, N.R.; Pereira, R.C.G.; Duarte, L.P.; Takahashi, J.A.; Silva, R.R. Synthesis and antimicrobial activity of ursolic acid ester derivatives. *Chem. Biodivers.* **2022**, *19*, e202100566. [\[CrossRef\]](https://doi.org/10.1002/cbdv.202100566) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34793623)
- 6. Tsepaeva, O.V.; Salikhova, T.I.; Ishkaeva, R.A.; Kundina, A.V.; Abdullin, T.I.; Laikov, A.V.; Tikhomirova, M.V.; Idrisova, L.R.; Nemtarev, A.V.; Mironov, V.F. Bifunctionalized Betulinic Acid Conjugates with C-3-Monodesmoside and C-28- Triphenylphosphonium Moieties with Increased Cancer Cell Targetability. *J. Nat. Prod.* **2023**, *86*, 1939–1949. [\[CrossRef\]](https://doi.org/10.1021/acs.jnatprod.3c00304) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37497692)
- 7. Nazaryan, S.; Bruguière, A.; Hovhannisyan, N.; Miyamoto, T.; Dias, A.M.M.; Bellaye, P.S.; Collin, B.; Briand, L.; Mitaine-Offer, A.C. Oleanolic Acid Glycosides from Scabiosa caucasica and Scabiosa ochroleuca: Structural Analysis and Cytotoxicity. *Molecules* **2023**, *28*, 4329. [\[CrossRef\]](https://doi.org/10.3390/molecules28114329) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37298806)
- 8. Gauthier, C.; Legault, J.; Piochon-Gauthier, M.; Pichette, A. Advances in the synthesis and pharmacological activity of lupane-type triterpenoid saponins. *Phytochem. Rev.* **2011**, *10*, 521–544. [\[CrossRef\]](https://doi.org/10.1007/s11101-010-9176-y)
- 9. Augustin, J.M.; Kuzina, V.; Andersen, S.B.; Bak, S. Molecular activities, biosynthesis and evolution of triterpenoid saponins. *Phytochemistry* **2011**, *72*, 435–457. [\[CrossRef\]](https://doi.org/10.1016/j.phytochem.2011.01.015)
- 10. Akhmetova, V.R.; Shakurova, E.R.; Khalilova, A.Z.; Khalilov, L.M.; Dzhemilev, U.M. Synthesis and transformations of 20-oxo-30 nortaraxasteryl acetate derivatives. *Russ. J. Org. Chem.* **2007**, *43*, 363–369. [\[CrossRef\]](https://doi.org/10.1134/S1070428007030050)
- 11. Pokorný, J.; Olejníková, D.; Frydrych, I.; Lišková, B.; Gurská, S.; Benická, S.; Šarek, J.; Kotulová, J.; Hajdúch, M.; Džubák, P.; et al. Substituted dienes prepared from betulinic acid–synthesis, cytotoxicity, mechanism of action, and pharmacological parameters. *Eur. J. Med. Chem.* **2021**, *224*, 113706. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2021.113706) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34311159)
- 12. Heise, N.V.; Kahnt, M.; Wagner, C.; Al-Harrasi, A.; Csuk, R. An unprecedented epimerization and annelation reaction of platanic acid amides. *J. Mol. Struct.* **2020**, *1220*, 128718. [\[CrossRef\]](https://doi.org/10.1016/j.molstruc.2020.128718)
- 13. Habrant, D.; Rauhala, V.; Koskinen, A.M.P. Conversion of carbonyl compounds to alkynes: General overview and recent developments. *Chem. Soc. Rev.* **2010**, *39*, 2007–2017. [\[CrossRef\]](https://doi.org/10.1039/b915418c) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20502799)
- 14. Kazakova, O.B.; Medvedeva, N.I.; Tolstikov, G.A.; Kukovinets, O.S.; Yamansarov, E.Y.; Spirikhin, L.V.; Gubaidullin, A.T. Synthesis of terminal acetylenes using POCl3 in pyridine as applied to natural triterpenoids. *Mendeleev Commun.* **2010**, *20*, 234–236. [\[CrossRef\]](https://doi.org/10.1016/j.mencom.2010.06.018)
- 15. Kazakova, O.B.; Yamansarov, E.Y.; Spirikhin, L.V.; Yunusov, M.S.; Baikova, I.P.; Kukovinets, O.S.; Musin, R.Z. Effective synthesis and transformations of alkyne betulin derivatives. *Russ. J. Org. Chem.* **2011**, *47*, 456460. [\[CrossRef\]](https://doi.org/10.1134/S1070428011030249)
- 16. Shakhmaev, R.N.; Sunagatullina, A.S.; Abdullina, E.A.; Zorin, V.V. Pd-catalyzed synthesis of 2-alkynyl derivatives of 19β, 28-epoxy-18α-olean-1-en-3-one. *Russ. J. Org. Chem.* **2017**, *53*, 1705–1709. [\[CrossRef\]](https://doi.org/10.1134/S1070428017110173)
- 17. Kotovshchikov, Y.N.; Latyshev, G.V.; Lukashev, N.V.; Beletskaya, I.P. Alkynylation of steroids via Pd-free Sonogashira coupling. *Org. Biomol. Chem.* **2015**, *13*, 5542–5555. [\[CrossRef\]](https://doi.org/10.1039/C5OB00559K) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25880697)
- 18. Ngoc, T.D.; Dehaen, W.; Meervelt, L.V.; Balzarini, J. Synthesis of Heterocyclic Triterpene Derivatives with Biological Activities via Click Reaction. *Curr. Org. Chem.* **2019**, *23*, 2969–2974. [\[CrossRef\]](https://doi.org/10.2174/1385272823666191212110411)
- 19. Khusnutdinova, E.F.; Petrova, A.V.; Kukovinets, O.S.; Kazakova, O.B. Synthesis and Cytotoxicity of 28-N-Propargylaminoalkylated 2,3-Indolotriterpenic acids. *Nat. Prod. Commun.* **2018**, *13*, 1934578X1801300. [\[CrossRef\]](https://doi.org/10.1177/1934578X1801300603)
- 20. Smirnova, I.E.; Kazakova, O.B.; Huong, D.T.T.; Minnibaeva, E.M.; Lobov, A.N.; Suponitsky, K.Y. One-pot synthesis of hollongdione from dipterocarpol. *Nat. Prod. Commun.* **2014**, *9*, 1934578X1400901005. [\[CrossRef\]](https://doi.org/10.1177/1934578X1400901005)
- 21. Grougnet, R.; Magiatis, P.; Mitaku, S.; Skaltsounis, A.L.; Cabalion, P.; Tillequin, F.; Michel, S. Dammarane Triterpenes from Gardenia aubryi Vieill. *Helv. Chim. Acta* **2011**, *94*, 656–661. [\[CrossRef\]](https://doi.org/10.1002/hlca.201000286)
- 22. Phongmaykin, J.; Kumamoto, T.; Ishikawa, T.; Suttisri, R.; Saifah, E. A new sesquiterpene and other terpenoid constituents of Chisocheton penduliflorus. *Arch. Pharm. Res.* **2008**, *31*, 21–27. [\[CrossRef\]](https://doi.org/10.1007/s12272-008-1115-8)
- 23. Chen, H.T.; Chuang, C.W.; Cheng, J.C.; Yeh, Y.J.; Chang, T.H.; Shi, Y.T.; Chao, C.H. Terpenoids with anti-influenza activity from the leaves of Euphorbia leucocephala. *Nat. Prod. Res.* **2023**, *37*, 936–943. [\[CrossRef\]](https://doi.org/10.1080/14786419.2022.2098739) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35838448)
- 24. Révész, L.; Hiestand, P.; La Vecchia, L.; Naef, R.; Naegeli, H.U.; Oberer, L.; Roth, H.J. Isolation and synthesis of a novel immunosuppressive 17α-substituted dammarane from the flour of the Palmyrah palm (*Borassus flabellifer*). *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1521–1526. [\[CrossRef\]](https://doi.org/10.1016/S0960-894X(99)00220-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10386928)
- 25. Giaginis, C.; Theocharis, S. Current evidence on the anticancer potential of Chios mastic gum. *Nutr. Cancer* **2011**, *63*, 1174–1184. [\[CrossRef\]](https://doi.org/10.1080/01635581.2011.607546) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22044444)
- 26. Smirnova, I.; Drăghici, G.; Kazakova, O.; Vlaia, L.; Avram, S.; Mioc, A.; Mioc, M.; Macaşoi, I.; Dehelean, C.; Voicu, A.; et al. Hollongdione arylidene derivatives induce antiproliferative activity against melanoma and breast cancer through pro-apoptotic and antiangiogenic mechanisms. *Bioorg. Chem.* **2022**, *119*, 105535. [\[CrossRef\]](https://doi.org/10.1016/j.bioorg.2021.105535) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34906859)
- 27. Shao, L.D.; Bao, Y.; Shen, Y.; Su, J.; Leng, Y.; Zhao, Q.S. Synthesis of selective 11β-HSD1 inhibitors based on dammarane scaffold. *Eur. J. Med. Chem.* **2017**, *135*, 324–338. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2017.04.059) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28458137)
- 28. Shao, L.D.; Xu, J.; Li, X.N.; Zhang, Z.J.; Shi, X.; Ren, J.; He, J.; Zhao, Y.; Leng, Y.; Xia, C.F.; et al. Synthesis of hupehenols A, B, and E from protopanaxadiol. *RSC Adv.* **2016**, *6*, 35792–35803. [\[CrossRef\]](https://doi.org/10.1039/C6RA04236H)
- 29. Flekhter, O.B.; Medvedeva, N.I.; Kukovinets, O.S.; Spirikhin, L.V.; Galkin, E.G.; Galin, F.Z.; Golovanov, D.G.; Pavlova, N.I.; Savinova, O.V.; Boreko, E.I.; et al. Synthesis and antiviral activity of lupane triterpenoids with modified cycle E. *Russ. J. Bioorg. Chem.* **2007**, *33*, 584–588. [\[CrossRef\]](https://doi.org/10.1134/S1068162007060088)
- 30. Khusnutdinova, E.F.; Bremond, P.; Petrova, A.V.; Kukovinets, O.S.; Kazakova, O.B. Synthesis of lupane mono-and bis-C19-(1, 2, 3-triazolyl)-triterpenoids by "Click" reaction. *Lett. Org. Chem.* **2017**, *14*, 743–747. [\[CrossRef\]](https://doi.org/10.2174/1570178614666170918120624)
- 31. Das, J.; Sarkar, A.; Ghosh, P. Friedelane triterpenoids: Transformations toward A-ring modifications including 2-homo derivatives. *New J. Chem.* **2018**, *42*, 6673–6688. [\[CrossRef\]](https://doi.org/10.1039/C8NJ00009C)
- 32. Allen, F.H.; Kennard, O.; Watson, D.G.; Brammer, L.; Orpen, A.G.; Taylor, R.J. Tables of bond lengths determined by X-ray and neutron diffraction. Part 1. Bond lengths in organic compounds. *J. Chem. Soc. Perkin Trans.* **1987**, *2*, S1–S19. [\[CrossRef\]](https://doi.org/10.1039/p298700000s1)
- 33. Shaw, R.; Elagamy, A.; Althagafi, I.; Pratap, R. Synthesis of alkynes from non-alkyne sources. *Org. Biomol. Chem.* **2020**, *18*, 3797–3817. [\[CrossRef\]](https://doi.org/10.1039/D0OB00325E) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32393951)
- 34. Khusnutdinova, E.F.; Apryshko, G.N.; Petrova, A.V.; Kukovinets, O.S.; Kazakova, O.B. The synthesis and selective cytotoxicity of new Mannich bases, derivatives of 19-and 28-alkynyltriterpenoids. *Russ. J. Bioorg. Chem.* **2018**, *44*, 123–127. [\[CrossRef\]](https://doi.org/10.1134/S1068162018010090)
- 35. Petrova, A.; Tretyakova, E.; Khusnutdinova, E.; Kazakova, O.; Slita, A.; Zarubaev, V.; Ma, X.; Jin, H.; Xu, H.; Xiao, S. Antiviral opportunities of Mannich bases derived from triterpenic N-propargylated indoles. *Chem. Biol. Drug Des.* **2024**, *103*, e14370. [\[CrossRef\]](https://doi.org/10.1111/cbdd.14370)
- 36. Hirai, S.; Harvey, R.G.; Jensen, E.V. The Mannich reaction: Improved conditions and application to 20-ketosteroids. *Tetrahedron Lett.* **1963**, *4*, 1123–1126. [\[CrossRef\]](https://doi.org/10.1016/S0040-4039(01)90787-7)
- 37. Zhu, N.; Ling, Y.; Lei, X.; Handratta, V.; Brodie, A.M.H. Novel P45017α inhibitors: 17-(2'-oxazolyl)-and 17-(2'-thiazolyl)-androstene derivatives. *Steroids* **2003**, *68*, 603–611. [\[CrossRef\]](https://doi.org/10.1016/S0039-128X(03)00082-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12957665)
- 38. Kovács, D.; Wölfling, J.; Szabó, N.; Szécsi, M.; Kovács, I.; Zupkó, I.; Frank, É. An efficient approach to novel 17-5'-(1',2',4')oxadiazolyl androstenes via the cyclodehydration of cytotoxic O-steroidacylamidoximes, and an evaluation of their inhibitory action on 17α-hydroxylase/C17,20-lyase. *Eur. J. Med. Chem.* **2013**, *70*, 649–660. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2013.10.038) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24211641)
- 39. Nagendrappa, G.; Griesbaum, K. Degradation of cyclodiene insecticide-like vinylic chloro compounds by ozone. *J. Agric. Food Chem.* **1978**, *26*, 581–583. [\[CrossRef\]](https://doi.org/10.1021/jf60217a022)
- 40. Kazakova, O.B.; Medvedeva, N.I.; Yamansarov, E.Y.; Spirikhin, L.V.; Khusnutdinova, E.F.; Kukovinets, O.S.; Tolstikov, G.A. Synthesis of Vinyl Chloride Derivatives on the Basis of Betulin. *Chem. Sustain. Dev.* **2011**, *19*, 335–338.
- 41. Monder, C.; Bradlow, H.L. Carboxylic acid metabolites of steroids. *J. Steroid Biochem.* **1977**, *8*, 897–908. [\[CrossRef\]](https://doi.org/10.1016/0022-4731(77)90101-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/338990)
- 42. Krause, L.; Herbst-Irmer, R.; Sheldrick, G.M.; Stalke, D.J. Comparison of silver and molybdenum microfocus X-ray sources for single-crystal structure determination. *Appl. Cryst.* **2015**, *48*, 3–10. [\[CrossRef\]](https://doi.org/10.1107/S1600576714022985) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26089746)
- 43. Sheldrick, G.M. SHELXT—Integrated space-group and crystal-structure determination. *Acta Crystallogr. Sect. A* **2015**, *71*, 3–8. [\[CrossRef\]](https://doi.org/10.1107/S2053273314026370) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25537383)
- 44. Sheldrick, G.M. Crystal Structure Refinement with SHELXL. *Acta Crystallogr. Sect. C* **2015**, *71*, 3–8. [\[CrossRef\]](https://doi.org/10.1107/S2053229614024218)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.