

Review **Recent Development of Fluoroquinolone Derivatives as Anticancer Agents**

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Abstract: Cancer is the second leading cause of death in the world following cardiovascular disease. Its treatment, including radiation therapy and surgical removal of the tumour, is based on pharmacotherapy, which prompts a constant search for new and more effective drugs. There are high costs associated with designing, synthesising, and marketing new substances. Drug repositioning is an attractive solution. Fluoroquinolones make up a group of synthetic antibiotics with a broad spectrum of activity in bacterial diseases. Moreover, those compounds are of particular interest to researchers as a result of reports of their antiproliferative effects on the cells of the most lethal cancers. This article presents the current progress in the development of new fluoroquinolone derivatives with potential anticancer and cytotoxic activity, as well as structure–activity relationships, along with possible directions for further development.

Keywords: fluoroquinolones; anticancer; cytotoxic activity; metal ion complexes

1. Introduction

The National Cancer Institute (NCI) defines cancer as a disease in which some of the body's cells grow uncontrollably and spread throughout the body. Long periods of cell division, accumulation of oncogenic mutations, and favourable localisation of lesions allowing for the accumulation of abnormal cells appear to be necessary to initiate the carcinogenesis process [\[1\]](#page-17-0). According to statistical data, cancer is the second leading cause of death in the world following cardiovascular disease [\[2–](#page-17-1)[4\]](#page-17-2). Among the most lethal cancers, the World Health Organisation indicates lung, colon, liver, stomach, and breast cancers [\[5\]](#page-17-3). Cancer treatment is based on a reductionist approach. The methods used primarily encompass pharmacotherapy (chemotherapy, immunotherapy), radiotherapy, and surgical removal of the lesion [\[6\]](#page-17-4). Despite medical progress, the prognosis for patients may remain unfavourable. The lack of a perfect panacea and the high costs of designing, synthesising, and marketing new medicinal substances encourage the search for an anticancer drug among derivatives of available drugs through the so-called repositioning of the drug [\[7\]](#page-17-5).

Fluoroquinolones make up a group of synthetic antibacterial drugs with a broad spectrum of activity in invasive medical and veterinary infections. They are used in infections of the urogenital tract, respiratory tract, and gastrointestinal tract caused by Gram-positive and Gram-negative microorganisms [\[8\]](#page-17-6). They are also successfully used in the treatment of bacterial infections of the eyes, bones, joints, and soft tissues [\[7](#page-17-5)[–9\]](#page-17-7). They also have antiviral activity $[9,10]$ $[9,10]$. Chemically, fluoroquinolones are derivatives created by modifying the skeleton of 4-oxo-1,4-dihydroquinolones. In addition to the oxygen groups necessary for pharmacological activity, namely, carboxyl and ketone groups at positions 3 and 4 of the ring skeleton, respectively, they contain one or two fluorine atoms in their structure [\[11,](#page-17-9)[12\]](#page-17-10).

Nalidixic acid obtained as a byproduct of the synthesis of chloroquine has been the first compound classified as a quinolone antibiotic. The ring structure of that compound

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is 1,8-naphthyridine [\[13\]](#page-17-11). Nalidixic acid was a drug with a narrow spectrum of pharmacological action widely used in the 1960s in urinary tract infections. Modifications of the structure of nalidixic acid have led to the preparation of approximately 10,000 analogues classified into four generations of quinolones [\[13](#page-17-11)[,14\]](#page-17-12). In addition to nalidixic acid, repre-Figures and the first generation of quinolones $[19,14]$. In determined to handline acid, representatives of the first generation of quinolones also include piromidic acid and pipemidic acid. Replacing the carbon atom with a nitrogen atom at position 6 of the nalidixic acid ring skeleton and introducing heterocyclic substituents at position 7 (in piromidic acid, the
substituent is pyrrolidine with one pitrogen beteroatom, and in pinemidic acid, pinerazine substituent is pyrrolidine with one nitrogen heteroatom, and in pipemidic acid, piperazine containing two nitrogen heteroatoms) has resulted in the preparation of drugs with an extended spectrum of activity against Gram-negative bacteria and that are very well absorbed
after and administration 115,161. The second and third are entires include 4 guinelance after oral administration [\[15](#page-17-13)[,16\]](#page-17-14). The second and third generations include 4-quinolone derivatives containing one or two halogen atoms in the heterocyclic system (most often fluorine–fluoroquinolones), while the fourth-generation antibiotics differ mainly in terms
expansion of proper DNA replica-cells. In the proper DNA replica-cells. In the proper DNA replica-cells. In the proper DNA replicaof modifications within the heterocyclic substituent at position 7. Those structural changes or modifications which the neterocyclic substituent at position 7. Those structural enarges
have a positive impact on the antibacterial activity properties of quinolones, extending the spectrum of their action onto Gram-positive bacteria, as well as anaerobic and atypical bacteria [\[11\]](#page-17-9). The most important representatives of the second-generation quinolones are bacteria [11]. The most important representatives of the second-generation quinolones are
ciprofloxacin, norfloxacin, ofloxacin, pefloxacin, lomefloxacin, and fenoxacin. The third eneration includes levofloxacin, sparfloxacin, and grepafloxacin. The fourth-generation drugs include moxifloxacin and gemifloxacin.

drugs include moxifloxacin and gemifloxacin.
The mechanism of the antibacterial action of fluoroquinolones is based on the inhibition of bacterial enzymes involved in DNA replication. By inhibiting DNA gyrase, they prevent the binding of initiation proteins, thus inhibiting the initiation of the replication process. Inhibition of topoisomerase IV prevents decatenation, thereby preventing the process. Inhibition of topoisonierase TV prevents decatements, thereby preventing the separation of genetic material into two daughter cells. Inhibition of proper DNA replica-tion leads to apoptosis of bacterial cells [\[17\]](#page-17-15). Moreover, fluoroquinolones have anticancer
The chemical structure of the their deliberate is bibliotheaell structure of induces and side of concern potential, manifested by their ability to inhibit the cell cycle and induce apoptosis of cancer potential, mathemetically affect the may be made to determing the line matter apppend of cancer cells. In this review, we have decided to elaborate upon the current progress in the development of new fluoroquinolone derivatives showing anticancer properties and correlations between the structure of those compounds and their cytotoxic activity towards cancer cells.

2. Structure of Fluoroquinolones and Their Antiproliferative Activity

The chemical structure of fluoroquinolones is very diverse, and small structural modirhe chemical structure or huoroquinolones is very diverse, and small structural modifications may significantly affect their biological activity. Several structural elements of fluoroquinolones appear to be of particular importance in their mechanism of anticancer action.

As mentioned below, the core of fluoroquinolones is a modified quinoline ring σ (Figure [1\)](#page-1-0). Its structure is crucial in inhibiting topoisomerase II, which is an enzyme the potential $\frac{1}{2}$. The statement is created in material $\frac{1}{2}$ replication of genetic material $\frac{1}{8}$, $\frac{1}{9}$. The nitrogen at position 1 is necessary for activity, and the type of substituent on this atom determines the potency of the activity. The related literature data indicate the advantage In the cyclic substituent (cyclopropyl) over chain substituents in the inhibitory effect on
In the cyclic substituent (cyclopropyl) over chain substituents in the inhibitory effect on topoisomerase II [20]. In order to achieve anticancer activity, a fluorine atom at position 6 of the quinolone ring is necessary. The introduction of an additional fluorine atom or a position 8 increases the effectiveness of that action [18]. methoxy group at position 8 increases the effectiveness of that action [\[18\]](#page-17-16).

Figure 1. Structure of fluoroquinolones.

Company

Similarly to the carboxyl group at position 3, the heterocyclic substituent at position 7 has a significant impact on the antibacterial activity of fluoroquinolones and the interaction with the prokaryotic topoisomerase-DNA complex, but it does not have an affinity for eukaryotic topoisomerases. It thus gives fluoroquinolones selectivity towards bacterial enzymes. In order to change the profile of action toward cancer cells, it is important to **induce modifications within both functional groups [\[18,](#page-17-16)[20,](#page-17-18)[21\]](#page-17-19).**
 Properties this information of a property of decomposition der to change the profile of action toward cancer ce

Eurodinations which both functional groups $[10,20,21]$.
Based on this information, five representatives of fluoroquinolones were selected, and their tumorigenic activity against cancer cell lines was characterised. The data are summarised in Table 1, along with the structural formulas of the selected compounds.

Table 1. Anticancer activity of fluoroquinolones. noma cells/100 ivity of fluoroquinolones.

 \mathcal{B} information, finally representatives of fluoroquinolones were selected, for \mathcal{B}

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Table 1. *Cont.*

f Fluoroquinolones with Heavy Met 3. Complexes of Fluoroquinolones with Heavy Metals

 $\frac{1}{2}$, $\frac{1}{2}$, $\frac{1}{2}$, $\frac{1}{2}$, $\frac{1}{2}$, $\frac{1}{2}$

Among cytostatic drugs currently used in cancer pharmacotherapy, platinum-based drugs such as cisplatin, carboplatin, and oxaliplatin, which are coordination compounds and the Compounds of Pt²⁺ ions, are widely used. Platinum complexes interact with the DNA of cancer cells, division, which results in the induction of apoptosis. They are used within the framework of monotherapy or in combination with other anticancer drugs for the treatment of breast, ovarian, or colorectal cancer [37-[41\]](#page-18-11). \mathbf{S} and \mathbf{S} disrupting the structure of the DNA and interfering with its synthesis. They inhibit cell
Distinguish nitrogen in the piperazine of monoton Theorem and still in the feature or the piper

ovarian, or colorectal cancer [37–41].
Fluoroquinolones are able to form complex compounds through the coordination of *O,O'*-bidentate or through coordination through nitrogen atoms in the piperazine N , N' ring, so they constitute a chelate ligan[d](#page-3-0) of bidentate (Figure 2). The formation of a complex
involving the carbonyl and carboxyl oxygen atoms at positions 3 and 4 of the quipelene involving the carbonyl and carboxyl oxygen atoms at positions 3 and 4 of the quinolone skeleton in fluoroquinolones is associated with obtaining derivatives that exhibit a wide spectrum of biological activity, including higher antibacterial and cytotoxic potential comspectrum or biological activity, including nigher antibacterial and cytotoxic potential com-
pared to the original ligand. Chelation of metal ions by piperazine nitrogen atoms is a less common phenomenon, but it may result in derivatives with antiproliferative activity [42]. The biological activity and stability of metal–fluoroquinolone complexes vary and depend depend on both the type of bidentate and the ligand (Table 2). on both the type of bidentate and the ligand (Table [2\)](#page-4-0).

Figure 2. Scheme of metal ion chelation exemplified by the structure of ciprofloxacin, showing the formation of $O(G)$ and $I(M)$ and $I(M)$ and $I(M)$ formation of *O,O'* coordination (**a**) and *N,N'* coordination (**b**). formation of *O,O*′ coordination (**a**) and *N,N*′ coordination (**b**).

quinolones. The synthesis of platinum–fluoroquinolone complexes using ciprofloxacin, levofloxacin, sparfloxacin, gatifloxacin, and ofloxacin occurs through coordination in the
piperazine ring [43]. The results of the eytotoxicity tosts of the complex with sparfloxacin fluoro \mathcal{L} Knowing that platinum compounds are used successfully in the treatment of cancer, it seems beneficial to use this metal and other heavy metals in combination with fluoropiperazine ring [\[43\]](#page-18-13). The results of the cytotoxicity tests of the complex with sparfloxacin

<u>pancreatic control de la pro</u>

as the ligand indicate stronger inhibitory activity against breast cancer cells and greater selectivity as compared to cisplatin [\[44\]](#page-18-14). The same coordination is demonstrated by gold(III) complexes with norfloxacin, levofloxacin, or sparfloxacin. They exhibit similar cytotoxic activity against lymphoma, myeloid leukaemia, and melanoma cell lines: they inhibit the cell cycle in the G0/G1 phase and induce apoptosis in a concentration-dependent manner [\[45\]](#page-18-15). Replacing platinum(II) or gold(II) with another metal ion causes a change in coordination with the ligand from *N,N*′ to *O,O*′ . Among the coordination compounds of fluoroquinolones with semiprecious and noble metals, rhenium complexes are also known. Rhenium(I) in coordination with enrofloxacin shows stronger inhibition of DNA topoisomerase as compared to the ligand alone [\[46\]](#page-18-16). The results of cytotoxicity tests of complexes with enrofloxacin and levofloxacin indicate their activity against erythroleukaemia cells [\[47\]](#page-18-17).

¹⁰ K-562/13.12 µM [44,4

Table 2. *Cont.*

$Table 2.$ *Cont.*

	Table 2. Cont.		
	The Structure of the Complex	Cell Line or DNA/Activity	Ref.
${\bf 20}$	F O	HL-60/induces apoptosis in 57.59% at concentration $100~\mu g \times mL^{-1}$	$[54]$
${\bf 21}$	H_{N} F $NH2$ $Q2$ $NH2$ 0 F_{\sim} F $H^{\tilde{\mathcal{N}}}$	$\rm BT20/12~\mu M$	$[55]$
${\bf 22}$	O Er -O- O O= Е	Ct DNA/Kb = 4.24×10^6 $\mathrm{M^{-1}}$	$[56]$

Table 2. *Cont.*

Among the transition metals, fluoroquinolones form comple biological activity as compared to the free ligand with ions such as Cu(I) [\[48,](#page-18-18)[57,](#page-19-8)[58\]](#page-19-9); Cu(II) [49-56,58-62]; Zn(II) [63-67]; Co(II) [68-72]; or Mn(II) [73]. Copper(II) coordination compounds are particularly interesting because of their broad spectrum of antiproliferative activity and much lower toxicity toward healthy cells than the platinum compounds currently used in chemotherapy. The chelation of copper ions by ciprofloxacin results in compounds that increase the production of reactive oxygen species and induce apoptosis μ using normal continuous discussion as the ligand μ as the ligand μ normal continuous copper (μ) normal copy ($\$ complexes proved and my electronic effects on our minute effects are active α as the ligand [57], Furthermore, $\text{copper}(\mu)$ introduced trouples enects on osteosarcoma and myelold leukaemia cells $[74,75]$. into sparifoxacin improves the inhibition of hormone-independe The use of pefloxacin as a ligand leads to the production of active copper complexes against breast cancer cells, and that activity exceeds the activity of analogous nickel(II) complexes [77]. Copper(II) complexes based on pefloxacin and statin derivatives effectively Among the transition metals, fluoroquinolones form complexes with similar or in human lung adenocarcinoma cells. Similar effects are achieved by using norfloxacin as the ligand [\[57\]](#page-19-8). Furthermore, $copper(II)$ norfloxacin complexes prove antiproliferative $\frac{1}{2}$ and changing the ligand into spartly the ligand into spartly into spartly $\frac{1}{2}$ $\frac{1}{2}$ independent and injective reasonable $\frac{1}{2}$. $\frac{1}{2}$. A mong the transition metals, fluoroquinolones form complexes with similar or \mathbf{r} Among the transition metals, fluoroquinolones form complexes with similar or higher effects on osteosarcoma and myeloid leukaemia cells $[74,75]$ $[74,75]$, and changing the ligand $\frac{1}{2}$ independent breast cancer cells $\frac{1}{2}$. The use of personal leads to the use of personal lea into sparfloxacin improves the inhibition of hormone-independent breast cancer cells [\[76\]](#page-20-3). inhibit the proliferation of human colorectal cancer cells, reduce the viability and clonogenic capacity of cells, and induce apoptosis [\[78\]](#page-20-5).

Lanthanide complexes are becoming increasingly important in cancer diagnosis and Landande compreses are becoming increasingly important in cancer diagnosis and therapy. They are used, among others, as contrast agents in magnetic resonance imaging therapy. They are used, among others, as contrast agents in magnetic resonance imaging
and in cancer radiotherapy [79]. The terbium(III)–ciprofloxacin complex, characterised by green fluorescence, may be successfully used as a fluorescent probe [80]. Lanthanide compounds with fluoroquinolone ligands, namely, ciprofloxacin [\[81](#page-20-8)[,82\]](#page-20-9), enrofloxacin [\[83\]](#page-20-10), gemifloxacin [\[66,](#page-19-15)[84\]](#page-20-11) levofloxacin [\[85\]](#page-20-12), norfloxacin [\[86\]](#page-20-13), and sparfloxacin [\[87\]](#page-20-14), show high gentification [60,04] revolutionally corrected [60], and sparticulate [67], show right
antimicrobial activity. The interest in erbium(III) complexes with fluoroquinolones has led to the preparation of compounds with cytotoxic and antimicrobial activity that exceeds the activity of other metal(II)-quinolone complexes found in the related literature [88]. Those reports may constitute a strong foundation for further attempts to obtain new lanthanide
complexes and study their attatexisity texted sensor sell lines. complexes and study their cytotoxicity toward cancer cell lines. green fluorescence, may be successfully used as a fluorescent probe [80]. Lanthanide high antimicrobial activity. The interest in erbium(III) complexes with fluoroquinolones Lanthanide complexes are becoming increasingly important in cancer diagnosis and anti-

and in cancer radiotherapy \overline{X} . The terminal complex, characterised by \overline{X}

 $\overline{}$ levoflowarian [85,84] levoflowarian [85,84], norfloxacin [85,84], and spartloxacin [87], show $\overline{}$

4. Fluoroquinolone Derivatives and Their Anticancer Properties

It is possible to modify the structure of a fluoroquinolone in many directions through substitutions at the 1 and 6 positions, modification of substituents at C3 and C7, and the addition of another fluorine atom at the 8 position. Modifications in the quinolone led to changes in the antimicrobial activity [89]. The attachment of substituents [70] core led to changes in the antimicrobial activity [\[89\]](#page-20-16). The attachment of substituents [\[70\]](#page-19-16) (Figure [3\)](#page-8-0) is associated with the spectrum expansion of drug activity beyond antibacterial, (Figure 3) is associated with the spectrum expansion of drug activity beyond antibacterial, to antiviral [90,91] and anticancer. to antivir[al](#page-20-17) [\[90](#page-20-18),91] and anticancer.

Figure 3. The main modification sites of the fluoroquinolone molecule exemplified by ciprofloxacin (modifications in position 7 are marked in red and modifications in position 3 are marked in blue). la regularity, bioavailability, and physicological properties in the properties of the properties α . As a second properties α

related literature about new fluoroquinolone derivatives that show anticancer activity in relation to human cell lines of the deadliest cancers. Substitution of the 7 position and substitution of the *N*-4-piperazinyl moiety have a large impact on the potency, bioavailability, and physicochemical properties [\[18](#page-17-16)[,92\]](#page-20-19). As a result of the synthetic work of researchers, there is more and more information in the

The latest literature reports expand the horizons in terms of the directions for obtaining
the latest literature reports expand the horizons in terms of the directions for obtaining a result of the condensation of a fluoroquinolone with fatty acids (Figure [4\)](#page-8-1). The selection of acids with varied chain lengths, degrees of unsaturation, and geometric isomerism ensured the structural variability of the compounds obtained. The synthetic route used crotonic (23), sorbic (24), geranium (25), oleic (26), elanic (27), linolenic (28), erucic (29), DHA (30), and paintific (51) acids. The entirely of the symmeses ranged from 44% to 59%. Bloogreat tests
allowed for the assessment of the cytotoxicity and apoptosis-inducing effect in primary and metastatic SW480 and SW620 colon cancer cells and in PC3 prostate cancer cells. isomerism ensured the structural variability of the compounds obtained. The synthetic Include the selection of all the selections of the selections for obtaining
derivatives. Chrzanowska et al. [\[93\]](#page-20-20) obtained various amide derivatives of ciprofloxacin as palmitic (**31**) acids. The efficiency of the syntheses ranged from 44% to 59%. Biological tests

Figure 4. Structure of ciprofloxacin amide derivatives obtained by conjugation with fatty acids.

The strongest antiproliferative properties were found in PC3 cells for derivatives **24, 26**, and 27, as compared to unsubstituted ciprofloxacin. The IC₅₀ values were 11.7, 7.7 and 15.3 µM, respectively. Conjugates **28** and **30** showed moderate activity (34.4 and respectively). The most promising inhibitory effect on the SW480 cell line was 27.8 μM, respectively). The most promising inhibitory effect on the SW480 cell line was demonstrated by conjugates 24, 25, 27, and 30 conjugated with polyunsaturated fatty acids, with IC_{50} values ranging between 20.1 and 35.7 μ M. The obtained ciprofloxacin amides showed high selectivity toward cancer cells, proving no cytotoxic effect towards normal human HaCaT keratinocytes. Furthermore, derivatives 24, 26, 27, and 30 showed noticeable apoptosis-inducing properties in the selected cell lines. These compounds significantly influenced the increase in the number of cells in the late phase of apoptosis at concentrations ranging from 600 to $1500 \mu M$, and the ciprofloxacin–oleic acid conjugate had the highest pro-apoptotic ability among those mentioned. had the highest pro-apoptotic ability among those mentioned. demonstrated by conjugate and product at the strongest and productative properties were found in PC3 cells for demonstratives $\frac{24}{50}$ values were 11.7, as compared to unsubstituted ciprofloxacin. The IC₅₀ values were 11.7, carignig from 600 to 1500 µm, and the cipronoxacin–oleic acid conjugate had the nighest

Akhtar et al. [\[94\]](#page-20-21) modified the structure of ciprofloxacin in two ways: esterifying the carboxyl group at position C3 and attaching a substituent to the nitrogen in the piperazine carboxyl group at position C3 and attaching a substituent to the nitrogen in the piperazine ring at position 7. The synthetic work gave rise to N-acylated derivatives 32–38 [\(F](#page-9-0)igure 5).

Figure 5. N-acylated and N-sulfonated derivatives of ciprofloxacin.

The obtained derivatives were tested for cytotoxic activity against the MCF-7 breast higher anticancer activity than unsubstituted ciprofloxacin. The highest cytotoxic potential tial was demonstrated by compounds **32** (IC₅₀ = 4.3 µM), **33** (IC₅₀ = 12.9 µM), and **35** $(IC_{50} = 60.9 \mu M)$, among which derivative 32 was singled out, and its usefulness in future syntheses of new N -alkylated fluoroquinolone derivatives with improved anticancer properties was indicated. The mechanism of the anticancer activity of compound 32 was investigated using in silico modelling methods. The results indicated topoisomerase II as a possible cytotoxic target; moreover, this derivative showed higher affinity for this enzyme cancer cell line using ciprofloxacin as the standard drug. All the compounds showed than the reference unsubstituted ciprofloxacin.

cancer cell line using ciprofloxacin as the standard drug. All the compounds showed

The syntheses conducted by Ahadi et al. [\[95\]](#page-20-22) led to the preparation of a series of $\frac{1}{2}$ corrected by Theorie of the preparation of a series of
ciprofloxacin derivatives 39–43 (Figure [6\)](#page-9-1). Modifications of the fluoroquinolone structure T_{ref} in N-(5-(benzylthio)-1.3.4-tiadazol-2-yl)carboxamide mojety at position 3 ciprofloxacin derivatives **39**–**43** (Figure 6). Modifications of the fluoroquinolone structure resulted in *N*-(5-(benzylthio)-1,3,4-tiadazol-2-yl)carboxamide moiety at position 3.

Figure 6. Ciprofloxacin derivatives containing N-(5-(benzylthio)-1,3,4-thiadiazol-2-yl)carboxamide moiety.

cancer cell lines: MCF-7 breast cancer, A549 lung cancer, and SKOV-3 ovarian cancer. Derivatives 39-48 showed anticancer activity against each of the selected cell lines and The synthesised compounds were assessed for their activity against selected human

were characterised by the stronger antiproliferative activity against MCF-7 cells than A549 and SKOV-3. The cytotoxic activity against the tested cell lines expressed in IC_{50} values for the derivatives is summarized in Table 3.

Compound	IC_{50} [μ M]		
	MCF-7	A549	SKOV-3
39	3.84	10.24	9.66
40	3.58	9.97	7.17
41	3.90	6.49	8.50
42	3.31	8.52	7.60
43	3.26	10.53	5.08
44	5.71	14.80	4.14
45	3.34	9.69	5.43
46	9.48	6.95	3.58
47	7.71	5.50	10.57
48	15.79	23.51	16.58

Table 3. IC₅₀ values of derivatives 39–48 for MCF-7, A549, and SKOV-3 cell lines.

The introduction of fluorine into the benzyl group in compounds 45 and 46 resulted in favourable changes in the activity against SKOV-3, and the bromine-containing derivative
 47 showed higher activity against A549 cells. The nitro group in the para position of the induction o benzyl group reduced the activity of that compound against each of the selected cancer cell being group reduced the activity of that compound against each of the selected earlier centrical DNA fragmental DNA fragmentation-International cycle in sub-G1 phase and the induction of oligonucleosomal DNA fragmentation. compounds **43** and **45** showed a concentration-dependent pro-apoptotic effect.

Kassab et al. [\[96\]](#page-20-23) prepared a series of derivatives 49–70 with structural features of a topoisomerase II inhibitor, using ciprofloxacin substituted at position 4 of the piperazine ring with *N*-acetylarylhydrazone, oxadiazole, or its bioisosterotriazole scaffolds. Aryl and heteroaryl groups were introduced into the *N*-acetylarylhydrazone residue. The structures of the synthesised derivatives are shown in Figure [7.](#page-10-1)

Figure 7. Ciprofloxacin derivatives containing topoisomerase II inhibitor structural elements. **Figure 7.** Ciprofloxacin derivatives containing topoisomerase II inhibitor structural elements.

An assessment of the anticancer activity performed on 59 panels of human can-
all lines also we determined of derivatives \mathbb{F}^2 activity UO 21 (IC = 4.02 aM) and MDA-MB-468 ($IC_{50} = 2.16 \mu M$) cell lines, **54** and **56** relative to the U0-31 cell line $(N_{\rm B}T_{\rm B}T_{\$ $\overrightarrow{1}$ IGROV1 (IC₅₀ = 0.75 µM) and UO-31 (IC₅₀ = 0.72 µM), and **63** relative to the HL60 cell line (IC₅₀ = 1.55 μ M). The antiproliferative activity of the obtained compounds seemed to correlate effectively with their ability to inhibit topoisomerase II. The highest ability to inhibit that enzyme was demonstrated by derivatives 54 and 63, which was several cer cell lines showed strong activity of derivatives 53 against UO-31 (IC₅₀ = 4.92 μ M) times higher than the activity of the reference compounds: doxorubicin, amsacrine, and etoposide. The antiproliferative effect of the derivatives led to the inhibition of the cell cycle in the G2/M phase and the induction of the intrinsic mitochondrial apoptosis pathway.

Ezelarab et al. [97] obtained a series of ciprofloxacin and quinoline derivatives 65–70 as a result of a multi-stage synthetic process (Figure [8\)](#page-11-0). Those hybrids were assessed for their anticancer properties against a range of cell lines. Studies have shown significant cytotoxic activity of derivatives 65 and 66 against SR-leukaemia and UO-31 renal cell carcinoma cell lines. The antiproliferative activity was expressed as a percentage of growth inhibition. Derivatives **65** and **66** inhibited the growth of 33.25 and 52.62% of leukaemia cells, respectively, and 64.19 and 55.49% of renal cancer cells, respectively. Moreover, they inhibited the growth of LOX IMVI melanoma cells by 39.14 and 36.64%, respectively. $\,$

growth inhibition. Derivatives **65** and **66** inhibited the growth of 33.25 and 52.62% of

Figure 8. Structure of amidopiperazine derivatives of ciprofloxacin. **Figure 8.** Structure of amidopiperazine derivatives of ciprofloxacin.

The studies showed the significant cytotoxic activity of derivatives 65, 66, and 70 against the selected cell lines. These compounds inhibited the cell cycle and induced apoptosis of the cancer cells, thereby reducing their growth and viability. In addition to their antiproliferative properties, ciprofloxacin/quinoline derivatives demonstrated potent antifungal activity against Candida species, highlighting their potential to be multi-targeted therapeutic agents.

Fallica et al. [\[98\]](#page-21-0) presented the synthetic path and results of the research on the antiproliferative effect of photodonor hybrids of nitric oxide (NO) with fluoroquinolones ciprofloxacin and norfloxacin (Figure [9\)](#page-12-0). The synthesis involved modification of the fluoroquinolone structure with NO donor moieties, creating compounds that released NO upon exposure to light. Their antiproliferative effect was tested on the DU145 and PC3 prostate cancer, HCT116 colon cancer, and MDA-MB231 and MCF-7 breast cancer cell lines. In vitro experiments indicated that derivatives **71–74** were effective against the selected cell lines. The effect of the norfloxacin derivatives was stronger than that of the ciprofloxacin derivatives. The best results were achieved with norfloxacin derivative 73: the IC₅₀ values for the \overline{P} PC3, MCF7, and MDA-MB231 cell lines were 2.33 μ M, 2.27 μ M, and 1.52 μ M, respectively. In relation to the DU145 cell line, derivative **74** was the most effective $(IC_{50} = 1.56 \mu M)$. Cytotoxicity studies showed that regardless of the length of the carbon bridge, the carboxyl moiety was indispensable for the anticancer effect. At the same time, it was observed that these compounds exhibit similar cytotoxicity towards non-cancer cell lines HBL100 and WH1. The IC₅₀ values ranged from 3.51 to 13.2 μ M depending on the derivative and the cell line tested. Similarly, the norfloxacin derivatives showed higher cytotoxicity.

Hihh et al. [\[99\]](#page-21-1) modified the ciprofloxacin molecule by combining various derivatives through a urea linker and a piperazine ring at position 7 of the fluoroquinolone (Figure [10\)](#page-12-1). The cytotoxic activity of new hybrids **75**–**84** was assessed, among others, against the HCT-116 colon cancer and leukaemia-SR cell lines.

Figure 9. Photodonor hybrids of ciprofloxacin and norfloxacin. **Figure 9.** Photodonor hybrids of ciprofloxacin and norfloxacin.

Figure 10. Chalcone derivatives of ciprofloxacin. **Figure 10.** Chalcone derivatives of ciprofloxacin.

activity of the reference compounds camptothecin and topotecan. Cytotoxicity studies of cells. Compounds 76 and 81 showed very good growth inhibitory effects on leukaemia cells, with IC_{50} of 2.38 μ M and 3.22 μ M, respectively. Changing the position of the chlorine atom from 4 to 3 in compound 76 or replacing it with other halogens at position 4 in the case of derivatives 78 and 79 reduced the cytotoxic effect. Studies on the impact of compounds on the cell cycle progression indicated hybrid 84 to be the substance that inhibited the cell cycle in the $G2/M$ phase and apoptosis in leukaemia cells. Testing the cytotoxicity of compounds 77 and 84 towards the non-cancerous WI-38 cell line indicated the selectivity of the substance towards cancer cells. The IC_{50} values for the WI-38 cells were 15.96 μ M and 18.42 μ M, respectively, being 6 to even 20 times higher than the IC_{50} values determined for halogens at position 4 in the case of derivatives **78** and **79** and **79** and **79** *79* and **79** *79* and **79** *79* Among the chalcone derivatives of ciprofloxacin, compounds **77** and **84** showed Among the chalcone derivatives of ciprofloxacin, compounds **77** and **84** showed strong growth inhibitory effects on HCT-116 cells (IC₅₀ values of 2.53 μ M and 2.01 μ M, respectively) and leukaemia-SR cells (IC₅₀ values of 0.73 μ M and 0.63 μ M, respectively). Additionally, they showed a significant inhibitory activity against topoisomerase, comparable to the the derivatives indicated a stronger toxic effect on leukaemia cells than on colon cancer $\frac{1}{2}$ value that IC50 values determined for cancer cells. cancer cells.

Szostek et al. [100] designed and synthesised ciprofloxacin derivatives using menthol and thymol moieties attached to the fluoroquinolone using various carboxyl linkers Figure 11). Both N and O-derivatives were obtained, as well as di compounds substituted at position carboxyl group in the 3 position and at the nitrogen atom of the piperazine $\mathop{\mathrm{arg}}$ in position \mathcal{V} , but the assessment of the biological activity of the latter did not yield \mathcal{V} s atisfactory results. The best antiproliferative a I 16 colon cancer cells, and SW480 and SW620 colon cancer cells was demonstrated by derivative 87 , with IC_{50} values against the mentioned lines of 36.8, 24.2, 30.3 and 38.6 μ M, respectively; derivative **92**, with IC₅₀ values of 51.3, 39.1,33.7 and 43.5 μ M, respectively; and derivative **95**, with IC₅₀ values of 41.8, 30.5, 29.5 and 49.6 μ M, respectively, while being non-toxic towards non-cancer HaCaT cells ($IC_{50} = 45.5$, >100 and 64.9 μ M for derivatives **87, 92, and 95, respectively).** Szostek et al. [\[100\]](#page-21-2) designed and synthesised ciprofloxacin derivatives using men-
se al. thomas weight is a the dead to the flores weight and signed constructed well lighters. and thymol moieties attached to the fluoroquinolone using various carboxyl linkers (Figure 11). Both *N* and *O*-derivatives were obtained, as well as di compounds substituted (Figure 11). Both *N* and *O*-derivatives were obtained, as well as di compounds substituted at position carboxyl group in the 3 position and at the nitrogen atom of the piperazine at position carboxyl group in the 3 position and at the nitrogen atom of the piperazine ring in position 7, but the assessment of the biological activity of the latter did not yield ring in position 7, but the assessment of the biological activity of the latter did not yield satisfactory results. The best antiproliferative activity against HepG2 liver cancer cells, satisfactory results. The best antiproliferative activity against HepG2 liver cancer cells, HCT-116 colon cancer cells, and SW480 and SW620 colon cancer cells was demonstrated by derivative 87 , with IC₅₀ values against the mentioned lines of 36.8, 24.2, 30.3 and 38.6 μ M,

Figure 11. Menthol and thymol derivatives of ciprofloxacin. **Figure 11.** Menthol and thymol derivatives of ciprofloxacin. **Figure 11.** Menthol and thymol derivatives of ciprofloxacin.

64.9 µM for derivatives **87**, **92**, and **95**, respectively).

64.9 µM for derivatives **87**, **92**, and **95**, respectively).

Results proving the beneficial effects of substances of natural origin, such as curcumin, silybin, or berberine, on the inhibition of the development of cancer cells and their potential use in therapy have rece[ntly](#page-21-3) appeared more and more frequently $[101-108]$ $[101-108]$. Milata et al. [\[109\]](#page-21-5) synthesised a number of 9-O-substituted berberine derivatives, including the condensation product of berberine with ciprofloxac[in](#page-13-1) (Figure 12). The anticancer effects of the compounds were tested on HeLa and HL-60 cell lines. Derivative 96 (mean IC₅₀ = 19.3 μM) showed greater antiproliferative activity towards HL-60 cells than the nonsubstituted berberine and ciprofloxacin but had a weaker effect than the other compounds. That hybrid was found to have inhibited the cell cycle in the HL-60 leukaemia cell line in the G2/M phase.

Figure 12. Conjugate of berberine and ciprofloxacin. **Figure 12.** Conjugate of berberine and ciprofloxacin. **Figure 12.** Conjugate of berberine and ciprofloxacin.

A well-known modification of fluoroquinolones to obtain anticancer compounds is the formation of Mannich bases. The ortho-phenol chalcone derivative of ciprofloxacin 97 obtained by Alaaeldin et al. [\[110\]](#page-21-6) showed beneficial properties. It inhibited topoisomerase
Lateral Ward distribution in the contract of the later distribution of the contract of the contract of the con I and II and had antiproliferative effects on A549 lung cancer (IC₅₀ = 27.71 μ M) and HepG2 hepatoma cells (IC₅₀ = 22.09 μ M). Furthermore, it inhibited the cell cycle in the G2/M μ and activities of indicated the apoptotic pathway. Additionally, exposure of the schemes tensor dependence to derivative 97 indicated high selectivity of the cytotoxic effect of the substance towards
sensor sells. The ICEO value for the WI32 sell line was 112.6 wM phase and activated the apoptotic pathway. Additionally, exposure of non-cancer cells to the apoptotic pathway. cancer cells. The IC50 value for the WI38 cell line was 118.65 μ M.

cancer cens. The IC50 value for the WI30 cell line was 118.65 µM. Fact $\frac{1}{2}$ sympaths and tested and tested and tested naphthological matrices of $\frac{1}{2}$ and $\frac{1}{2}$ a ovarian cancer and A-549 lung cancer cell lines by inhibiting the cell cycle in the S phase Fawzy et al. [111] synthesised and tested naphthol Mannich base **98** for anticancer Fawzy et al. [\[111\]](#page-21-7) synthesised and tested naphthol Mannich base **98** for anticancer activity (Figure 13). The compound showed antiproliferative activity against the OVCAR-activity (Figure [13\)](#page-14-0). The compound showed antiproliferative activity against the OVCAR-3 and inducing apoptosis through the mitochondrial pathway apoptotic.

Struga et al. [\[112\]](#page-21-8) obtained a series of *N*-acylated ciprofloxacin derivatives (Figure [14\)](#page-14-1) and conjugates of ciprofloxacin molecules connected with carbon linkers of various lengths. The antiproliferative activity of compounds **99**–**112** was proven against PC3 prostate cancer cells. Derivative **99** showed high antiproliferative activity against the selected cell line,

Figure 13. Ortho-phenol Mannich bases derived from ciprofloxacin.

Figure 14. *N*-acylated ciprofloxacin derivatives. **Figure 14.** *N*-acylated ciprofloxacin derivatives. **Figure 14.** *N*-acylated ciprofloxacin derivatives.

Recent literature reports also include data on norfloxacin and levofloxacin derivatives,
in addition to ciprofloxacin derivati[ves.](#page-21-9) Xi et al. [113] focused on obtaining inhibitors of microRNA-21, which is overexpressed in cancer cells. They designed and synthesised a $\frac{1}{100}$ in the micro $\frac{1}{100}$ of microscopic in cancer cells. They designed and cylindes set and contact $\frac{1}{100}$ or $\frac{1}{100}$ ($\frac{1}{100}$ $\frac{1}{100}$) $\frac{1}{100}$ series of benzamide derivatives of norfloxacin **113–120** (Figure [15\)](#page-14-2). Recent literature reports also include data on norfloxacin and levofloxacin derivatives,

Figure 15. Benzamide derivatives of norfloxacin. **Figure 15.** Benzamide derivatives of norfloxacin. **Figure 15.** Benzamide derivatives of norfloxacin.

floxacin derivatives among the compounds of other fluoroquinolones, namely, ciprofloxacin, comparable to the selected small molecule inhibitor. That compound suppressed the expression at the level of transcription of its original form. Evaluation of the antiproliferative activity against HCT-116 and HeLa cell lines confirmed the ability to inhibit colony formation and migration and induction of apoptosis in the case of compound 119. That work was a continuation of the research conducted and described by Hei et al. [\[114\]](#page-21-10), who indicated nor-Derivative **119** showed the best efficacy as a microRNA-21 inhibitor, and that effect Derivative **119** showed the best efficacy as a microRNA-21 inhibitor, and that effect was levofloxacin, gatifloxacin, and enoxacin, to be potential microRNA-21 inhibitors.

Wang et al. [\[115\]](#page-21-11) linked levofloxacin with hydroxamic acid using carbon linkers of varied lengths, resulting in dual-acting derivatives targeting histone deacetylase and tubulin polymerisation (Figure [16\)](#page-15-0). Modification of the length of the carbon chain had a significant impact on the inhibitory activity of the compound. The antiproliferative properties of the new hybrids were tested on A549 cell lines HepG2, MCF-7, PC-3, and HeLa. Compound 125 had the best anticancer properties and was more than 20 times

stronger than free levofloxacin. The IC $_{50}$ values of the derivative against these cell lines were 2.1 μ M, 2.3 μ M, 0.3 μ M, 4.9 μ M, and 1.1 μ M, respectively, while the IC₅₀ values of levofloxacin ranged from 64.2 to a value exceeding 100μ M in relation to individual cells of levofloxacin ranged from 64.2 to a value exceeding 100μ M in relation to individual cell lines. lines. $\mathbf{r} = \mathbf{r} - \mathbf{r} - \mathbf{r} - \mathbf{r}$ values of the derivative against the derivativ were $2.1 M_{\odot}$ $2.3 M_{\odot}$ M_{\odot} and $1.1 M_{\odot}$ and $1.1 M_{\odot}$ respectively, while the IC50 values of M_{\odot}

Figure 16. Levofloxacin and hydroxamic acid derivatives. **Figure 16.** Levofloxacin and hydroxamic acid derivatives.

To sum up, the examples described above show interesting directions of work ing to the preparation of anticancer compounds based on the fluoroquinolone skeleton. N-substituted compounds have higher cytotoxic potency compared to C-substituted derivatives, and the most frequently chosen substrate is ciprofloxacin. Nevertheless, these reports may constitute the basis for the syntheses of analogous derivatives using other fluoroquinolones in order to expand the range of potential therapeutics. To sum up, the examples described above show interesting directions of work lead-

5. Molecular Dockin<mark>g</mark>

5. Molecular Docking is associated with increasing research costs and stricter experimental procedures. In this context, modern computational methods are helpful, as they enable a large part of prelimi-
context, modern computational methods are helpful, as they enable a large part of prelimihas become a powerful tool for drug development (including the repositioning of known drugs) and is constantly evolving. Docking is a method that allows for determining the drugs) and is constantly evolving. Docking is a method that allows for determining the arage) and is consuming evening. Deciding is a method and the response concentrating the distribution and conformation of a ligand at the receptor binding site. It also enables the assessment of the binding strength of the complex. It is, of course, used in in silico research on the anticancer activity of fluoroquinolones and their derivatives. The search for new medicinal compounds is becoming increasingly difficult. This nary research to be carried out in silico. One of these methods is molecular docking, which

Mcl-1 is a potent anti-apoptotic protein that is amplified in many human cancers, while the microphthalmia-related transcription factor (MITF) promotes cell proliferation and plays a pro-survival role. Beberok et al. conducted in silico studies on the possible interaction of ciprofloxacin with MITF/Mcl-1 proteins. The analysis showed that ciprofloxacin has the ability to form complexes with MITF and Mcl-1 proteins and may thus have an apoptotic effect $[116]$.

Suresh et. al. performed docking of a series of ciprofloxacin derivatives to topoiso-merase II (Figure [17\)](#page-15-1) using the Glide program. These derivatives obtained docking score values ranging from -7.78 to -8.04 kcal/mol and were lower than the value obtained for the reference effective active [116]. The derivative complexes 127–134 than the reference [\[117\]](#page-21-13). the reference ciprofloxacin (−7.57 kcal/mol), indicating higher stability of the obtained

Figure 17. Ciprofloxacin amide derivatives. **Figure 17.** Ciprofloxacin amide derivatives.

In silico studies by Allaka et al. also used topoisomerase II as a protein target. A series In silico studies by Allaka et al. also used topoisomerase II as a protein target. A series of hydrazone derivatives of pefloxacin were used as ligands (Figure [18\)](#page-16-0). The obtained dock-

ing score values for hydrazones **135–142** ranged from −7.29 kcal/mol to −6.27 kcal/mol, and for the reference compound, this value was -5.80 kcal/mol [\[118\]](#page-21-14). This result may indicate the potential anticancer activity of the tested derivatives.

of hydrazone derivatives of performance of performance $\mathcal{L}_{\mathcal{A}}$. The obtained as ligands (Figure 18). T

Figure 18. Pefloxacin hydrazones. **Figure 18.** Pefloxacin hydrazones.

6. Methods 6. Methods

A literature review of reviews and studies was performed from 2000 to 2024. Using terms such as 'anticancer drugs', 'fluoroquinolone', and 'fluoroquinolones derivatives' and using the structural formula of ciprofloxacin modified with R substituent, PubMed, Scopus, and Reaxys databases were searched. The articles selected were in English, with free access to the full content, discussing the mechanisms of the action of anticancer drugs, and presenting synthetic protocols for fluoroquinolone derivatives and the results of tests of those compounds for antiproliferative effects on human cancer cells. Furthermore, a snowball approach was used, a result of which the reports relevant to the topic of this article were extracted from the documents cited in this article selected in the main search. A literature review of reviews and studies was performed from 2000 to 2024. Using

article were extracted in this article selected in this article selected in this article selected in the main search. **7. Conclusions and Future Directions**

Cancer pharmacotherapy is constantly being optimised to improve effectiveness and reduce the risk of inclusions and compiled tors. Modern selement reports indicate that the search for new anticancer drugs may be based on the fluoroquinolone pharmacophore are search for new annealises analysis of and complications in the masses procedure that the critical one, and one analysis of available data shows that the critical one, from the point of view of the activity of those derivatives, is the *N*-alkylated pyridone system with two carbonyl oxygen atoms: one at position 4 of the pyridone ring and the other in the carboxyl group present at the C3 atom of the pyridone system. Possible modifications in the structure of fluoroquinolone derivatives are primarily applicable to substituents at position 7 of the quinolone system and modifications within the carboxyl group (esterification, amidation). Complexes with metals also seem to be an important direction in the modification of fluoroquinolones to obtain derivatives with anticancer activity. This is confirmed by the complexes of fluoroquinolones obtained so far with $Cu(I)$, $Cu(II)$, $Zn(II)$, $Co(II)$, or Mn(II). It seems that modifications of the fluoroquinolone system at the above-mentioned positions should be taken into account in further research on new anticancer drugs. Attempts to use in silico methods in the design or repositioning

on the fluoroquinology of the fluoroqui reduce the risk of metastases and complications. Modern scientific reports indicate that of fluoroquinolones as compounds with anticancer activity are also promising.

Author Contributions: Conceptualization, J.N. and K.M.; methodology, J.N. and K.M.; validation, J.N and K.M; formal analysis, J.N., K.M., R.C. and D.R.; investigation, J.N.; resources, J.N. and K.M.; data curation, J.N. and K.M.; writing—original draft preparation, J.N., K.M., R.C. and D.R.; writing review and editing, J.N. and K.M.; visualization, J.N.; supervision, K.M.; project administration, K.M. All authors have read and agreed to the published version of the manuscript.

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References

- 1. Brown, J.S.; Amend, S.R.; Austin, R.H.; Gatenby, R.A.; Hammarlund, E.U.; Pienta, K.J. Updating the Definition of Cancer. *Mol. Cancer Res.* **2023**, *21*, 1142–1147. [\[CrossRef\]](https://doi.org/10.1158/1541-7786.MCR-23-0411) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37409952)
- 2. Health at a Glance: Europe 2022. Available online: <https://www.oecd.org/health/health-at-a-glance-europe> (accessed on 11 June 2024).
- 3. Global Burden of Disease 2019 Cancer Collaboration; Kocarnik, J.M.; Compton, K.; Dean, F.E.; Fu, W.; Gaw, B.L.; Harvey, J.D.; Henrikson, H.J.; Lu, D.; Pennini, A.; et al. Cancer Incidence, Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life Years for 29 Cancer Groups from 2010 to 2019: A Systematic Analysis for the Global Burden of Disease Study 2019. *JAMA Oncol.* **2022**, *8*, 420–444. [\[CrossRef\]](https://doi.org/10.1001/jamaoncol.2021.6987) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34967848)
- 4. Siegel, R.L.; Miller, K.D.; Fuchs, H.E.; Jemal, A. Cancer Statistics, 2022. *CA A Cancer J. Clin.* **2022**, *72*, 7–33. [\[CrossRef\]](https://doi.org/10.3322/caac.21708)
- 5. Breast Cancer. Available online: <https://www.who.int/news-room/fact-sheets/detail/breast-cancer> (accessed on 11 June 2024).
- 6. Zugazagoitia, J.; Guedes, C.; Ponce, S.; Ferrer, I.; Molina-Pinelo, S.; Paz-Ares, L. Current Challenges in Cancer Treatment. *Clin. Ther.* **2016**, *38*, 1551–1566. [\[CrossRef\]](https://doi.org/10.1016/j.clinthera.2016.03.026)
- 7. Yadav, V.; Talwar, P. Repositioning of Fluoroquinolones from Antibiotic to Anti-Cancer Agents: An Underestimated Truth. *Biomed. Pharmacother.* **2019**, *111*, 934–946. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2018.12.119)
- 8. Bhatt, S.; Chatterjee, S. Fluoroquinolone Antibiotics: Occurrence, Mode of Action, Resistance, Environmental Detection, and Remediation—A Comprehensive Review. *Environ. Pollut.* **2022**, *315*, 120440. [\[CrossRef\]](https://doi.org/10.1016/j.envpol.2022.120440)
- 9. Scroggs, S.L.P.; Offerdahl, D.K.; Flather, D.P.; Morris, C.N.; Kendall, B.L.; Broeckel, R.M.; Beare, P.A.; Bloom, M.E. Fluoroquinolone Antibiotics Exhibit Low Antiviral Activity against SARS-CoV-2 and MERS-CoV. *Viruses* **2020**, *13*, 8. [\[CrossRef\]](https://doi.org/10.3390/v13010008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33374514)
- 10. Yacouba, A.; Olowo-okere, A.; Yunusa, I. Repurposing of Antibiotics for Clinical Management of COVID-19: A Narrative Review. *Ann. Clin. Microbiol. Antimicrob.* **2021**, *20*, 37. [\[CrossRef\]](https://doi.org/10.1186/s12941-021-00444-9)
- 11. Pham, T.D.M.; Ziora, Z.M.; Blaskovich, M.A.T. Quinolone Antibiotics. *Med. Chem. Commun.* **2019**, *10*, 1719–1739. [\[CrossRef\]](https://doi.org/10.1039/C9MD00120D)
- 12. Tang, K.; Zhao, H. Quinolone Antibiotics: Resistance and Therapy. *Infect. Drug Resist.* **2023**, *16*, 811–820. [\[CrossRef\]](https://doi.org/10.2147/IDR.S401663) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36798480)
- 13. Bisacchi, G.S. Origins of the Quinolone Class of Antibacterials: An Expanded "Discovery Story". *J. Med. Chem.* **2015**, *58*, 4874–4882. [\[CrossRef\]](https://doi.org/10.1021/jm501881c)
- 14. Asif, M. Antimicrobial and Anti-Tubercular Activity of Quinolone Analogues. *Sci. Int.* **2013**, *1*, 336–349. [\[CrossRef\]](https://doi.org/10.17311/sciintl.2013.336.349)
- 15. Shimizu, M.; Takase, Y.; Nakamura, S.; Katae, H.; Minami, A. Pipemidic Acid, a New Antibacterial Agent Active against Pseudomonas Aeruginosa: In Vitro Properties. *Antimicrob. Agents Chemother.* **1975**, *8*, 132–138. [\[CrossRef\]](https://doi.org/10.1128/AAC.8.2.132)
- 16. Shimizu, M.; Nakamura, S.; Takase, Y.; Kurobe, N. Pipemidic Acid: Absorption, Distribution, and Excretion. *Antimicrob. Agents Chemother.* **1975**, *7*, 441–446. [\[CrossRef\]](https://doi.org/10.1128/AAC.7.4.441) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1147580)
- 17. Blondeau, J.M. Fluoroquinolones: Mechanism of Action, Classification, and Development of Resistance. *Surv. Ophthalmol.* **2004**, *49* (Suppl. S2), S73–S78. [\[CrossRef\]](https://doi.org/10.1016/j.survophthal.2004.01.005)
- 18. Abdel-Aal, M.A.A.; Abdel-Aziz, S.A.; Shaykoon, M.S.A.; Abuo-Rahma, G.E.-D.A. Towards Anticancer Fluoroquinolones: A Review Article. *Arch. Pharm.* **2019**, *352*, e1800376. [\[CrossRef\]](https://doi.org/10.1002/ardp.201800376)
- 19. McClendon, A.K.; Osheroff, N. DNA Topoisomerase II, Genotoxicity, and Cancer. *Mutat. Res.* **2007**, *623*, 83–97. [\[CrossRef\]](https://doi.org/10.1016/j.mrfmmm.2007.06.009)
- 20. Kohlbrenner, W.E.; Wideburg, N.; Weigl, D.; Saldivar, A.; Chu, D.T. Induction of Calf Thymus Topoisomerase II-Mediated DNA Breakage by the Antibacterial Isothiazoloquinolones A-65281 and A-65282. *Antimicrob. Agents Chemother.* **1992**, *36*, 81–86. [\[CrossRef\]](https://doi.org/10.1128/AAC.36.1.81) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1317151)
- 21. Idowu, T.; Schweizer, F. Ubiquitous Nature of Fluoroquinolones: The Oscillation between Antibacterial and Anticancer Activities. *Antibiotics* **2017**, *6*, 26. [\[CrossRef\]](https://doi.org/10.3390/antibiotics6040026)
- 22. Beberok, A.; Wrześniok, D.; Minecka, A.; Rok, J.; Delijewski, M.; Rzepka, Z.; Respondek, M.; Buszman, E. Ciprofloxacin-Mediated Induction of S-Phase Cell Cycle Arrest and Apoptosis in COLO829 Melanoma Cells. *Pharmacol. Rep.* **2018**, *70*, 6–13. [\[CrossRef\]](https://doi.org/10.1016/j.pharep.2017.07.007)
- 23. Beberok, A.; Wrześniok, D.; Rok, J.; Rzepka, Z.; Respondek, M.; Buszman, E. Ciprofloxacin Triggers the Apoptosis of Human Triple-Negative Breast Cancer MDA-MB-231 Cells via the P53/Bax/Bcl-2 Signaling Pathway. *Int. J. Oncol.* **2018**, *52*, 1727–1737. [\[CrossRef\]](https://doi.org/10.3892/ijo.2018.4310) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29532860)
- 24. Beberok, A.; Rzepka, Z.; Respondek, M.; Rok, J.; Sierotowicz, D.; Wrześniok, D. GSH Depletion, Mitochondrial Membrane Breakdown, Caspase-3/7 Activation and DNA Fragmentation in U87MG Glioblastoma Cells: New Insight into the Mechanism of Cytotoxicity Induced by Fluoroquinolones. *Eur. J. Pharmacol.* **2018**, *835*, 94–107. [\[CrossRef\]](https://doi.org/10.1016/j.ejphar.2018.08.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30086267)
- 25. Jemel-Oualha, I.; Elloumi-Mseddi, J.; Beji, A.; Hakim, B.; Aifa, S. Controversial Effect on Erk Activation of Some Cytotoxic Drugs in Human LOVO Colon Cancer Cells. *J. Recept. Signal Transduct.* **2016**, *36*, 21–25. [\[CrossRef\]](https://doi.org/10.3109/10799893.2014.975246) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25343691)
- 26. Kloskowski, T.; Szeliski, K.; Fekner, Z.; Rasmus, M.; Dąbrowski, P.; Wolska, A.; Siedlecka, N.; Adamowicz, J.; Drewa, T.; Pokrywczyńska, M. Ciprofloxacin and Levofloxacin as Potential Drugs in Genitourinary Cancer Treatment-The Effect of Dose-Response on 2D and 3D Cell Cultures. *Int. J. Mol. Sci.* **2021**, *22*, 11970. [\[CrossRef\]](https://doi.org/10.3390/ijms222111970) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34769400)
- 27. Fan, M.; Chen, S.; Weng, Y.; Li, X.; Jiang, Y.; Wang, X.; Bie, M.; An, L.; Zhang, M.; Chen, B.; et al. Ciprofloxacin Promotes Polarization of CD86+CD206- Macrophages to Suppress Liver Cancer. *Oncol. Rep.* **2020**, *44*, 91–102. [\[CrossRef\]](https://doi.org/10.3892/or.2020.7602) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32377744)
- 28. Kloskowski, T.; Gurtowska, N.; Olkowska, J.; Nowak, J.M.; Adamowicz, J.; Tworkiewicz, J.; D˛ebski, R.; Grzanka, A.; Drewa, T. Ciprofloxacin Is a Potential Topoisomerase II Inhibitor for the Treatment of NSCLC. *Int. J. Oncol.* **2012**, *41*, 1943–1949. [\[CrossRef\]](https://doi.org/10.3892/ijo.2012.1653) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23042104)
- 29. Mondal, E.R.; Das, S.K.; Mukherjee, P. Comparative Evaluation of Antiproliferative Activity and Induction of Apoptosis by Some Fluoroquinolones with a Human Non-Small Cell Lung Cancer Cell Line in Culture. *Asian Pac. J. Cancer Prev.* **2004**, *5*, 196–204. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15244525)
- 30. Beberok, A.; Wrześniok, D.; Szlachta, M.; Rok, J.; Rzepka, Z.; Respondek, M.; Buszman, E. Lomefloxacin Induces Oxidative Stress and Apoptosis in COLO829 Melanoma Cells. *Int. J. Mol. Sci.* **2017**, *18*, 2194. [\[CrossRef\]](https://doi.org/10.3390/ijms18102194)
- 31. Beberok, A.; Rzepka, Z.; Rok, J.; Banach, K.; Wrześniok, D. UVA Radiation Enhances Lomefloxacin-Mediated Cytotoxic, Growth-Inhibitory and Pro-Apoptotic Effect in Human Melanoma Cells through Excessive Reactive Oxygen Species Generation. *Int. J. Mol. Sci.* **2020**, *21*, 8937. [\[CrossRef\]](https://doi.org/10.3390/ijms21238937)
- 32. Perucca, P.; Savio, M.; Cazzalini, O.; Mocchi, R.; Maccario, C.; Sommatis, S.; Ferraro, D.; Pizzala, R.; Pretali, L.; Fasani, E.; et al. Structure-Activity Relationship and Role of Oxygen in the Potential Antitumour Activity of Fluoroquinolones in Human Epithelial Cancer Cells. *J. Photochem. Photobiol. B Biol.* **2014**, *140*, 57–68. [\[CrossRef\]](https://doi.org/10.1016/j.jphotobiol.2014.07.006)
- 33. Nakai, S.; Imaizumi, T.; Watanabe, T.; Iwase, Y.; Nishi, K.; Okudaira, K.; Yumita, N. Photodynamically-Induced Apoptosis Due to Ultraviolet A in the Presence of Lomefloxacin in Human Promyelocytic Leukemia Cells. *Anticancer Res.* **2017**, *37*, 6407–6413. [\[CrossRef\]](https://doi.org/10.21873/anticanres.12094) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29061826)
- 34. He, X.; Yao, Q.; Hall, D.D.; Song, Z.; Fan, D.; You, Y.; Lian, W.; Zhou, Z.; Duan, L.; Chen, B. Levofloxacin Exerts Broad-Spectrum Anticancer Activity via Regulation of THBS1, LAPTM5, SRD5A3, MFAP5 and P4HA1. *Anticancer Drugs* **2022**, *33*, e235–e246. [\[CrossRef\]](https://doi.org/10.1097/CAD.0000000000001194) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34419964)
- 35. Yu, M.; Li, R.; Zhang, J. Repositioning of Antibiotic Levofloxacin as a Mitochondrial Biogenesis Inhibitor to Target Breast Cancer. *Biochem. Biophys. Res. Commun.* **2016**, *471*, 639–645. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2016.02.072) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26902121)
- 36. Yadav, V.; Sultana, S.; Yadav, J.; Saini, N. Gatifloxacin Induces S and G2-Phase Cell Cycle Arrest in Pancreatic Cancer Cells via P21/P27/P53. *PLoS ONE* **2012**, *7*, e47796. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0047796)
- 37. Zhang, C.; Xu, C.; Gao, X.; Yao, Q. Platinum-Based Drugs for Cancer Therapy and Anti-Tumor Strategies. *Theranostics* **2022**, *12*, 2115–2132. [\[CrossRef\]](https://doi.org/10.7150/thno.69424)
- 38. Romani, A.M.P. Cisplatin in Cancer Treatment. *Biochem. Pharmacol.* **2022**, *206*, 115323. [\[CrossRef\]](https://doi.org/10.1016/j.bcp.2022.115323)
- 39. Rottenberg, S.; Disler, C.; Perego, P. The Rediscovery of Platinum-Based Cancer Therapy. *Nat. Rev. Cancer* **2021**, *21*, 37–50. [\[CrossRef\]](https://doi.org/10.1038/s41568-020-00308-y)
- 40. López-Gresa, M.P.; Ortiz, R.; Perelló, L.; Latorre, J.; Liu-González, M.; García-Granda, S.; Pérez-Priede, M.; Cantón, E. Interactions of Metal Ions with Two Quinolone Antimicrobial Agents (Cinoxacin and Ciprofloxacin): Spectroscopic and X-ray Structural Characterization. Antibacterial Studies. *J. Inorg. Biochem.* **2002**, *92*, 65–74. [\[CrossRef\]](https://doi.org/10.1016/S0162-0134(02)00487-7)
- 41. Chohan, Z.H.; Supuran, C.T.; Scozzafava, A. Metal Binding and Antibacterial Activity of Ciprofloxacin Complexes. *J. Enzym. Inhib. Med. Chem.* **2005**, *20*, 303–307. [\[CrossRef\]](https://doi.org/10.1080/14756360310001624948)
- 42. Azéma, J.; Guidetti, B.; Dewelle, J.; Le Calve, B.; Mijatovic, T.; Korolyov, A.; Vaysse, J.; Malet-Martino, M.; Martino, R.; Kiss, R. 7-((4-Substituted)Piperazin-1-Yl) Derivatives of Ciprofloxacin: Synthesis and in Vitro Biological Evaluation as Potential Antitumor Agents. *Bioorg. Med. Chem.* **2009**, *17*, 5396–5407. [\[CrossRef\]](https://doi.org/10.1016/j.bmc.2009.06.053)
- 43. Vieira, L.M.M.; de Almeida, M.V.; de Abreu, H.A.; Duarte, H.A.; Grazul, R.M.; Fontes, A.P.S. Platinum(II) Complexes with Fluoroquinolones: Synthesis and Characterization of Unusual Metal–Piperazine Chelates. *Inorganica. Chimica. Acta* **2009**, *6*, 2060–2064. [\[CrossRef\]](https://doi.org/10.1016/j.ica.2008.08.018)
- 44. de Oliveira, L.P.; Carneiro, Z.A.; Ribeiro, C.M.; Lima, M.F.; Paixão, D.A.; Pivatto, M.; de Souza, M.V.N.; Teixeira, L.R.; Lopes, C.D.; de Albuquerque, S.; et al. Three New Platinum Complexes Containing Fluoroquinolones and DMSO: Cytotoxicity and Evaluation against Drug-Resistant Tuberculosis. Available online: [https://www.americanelements.com/three-new-platinum](https://www.americanelements.com/three-new-platinum-complexes-containing-fluoroquinolones-and-dmso-cytotoxicity-and-evaluation)[complexes-containing-fluoroquinolones-and-dmso-cytotoxicity-and-evaluation](https://www.americanelements.com/three-new-platinum-complexes-containing-fluoroquinolones-and-dmso-cytotoxicity-and-evaluation) (accessed on 11 June 2024).
- 45. Gouvea, L.R.; Garcia, L.S.; Lachter, D.R.; Nunes, P.R.; de Castro Pereira, F.; Silveira-Lacerda, E.P.; Louro, S.R.W.; Barbeira, P.J.S.; Teixeira, L.R. Atypical Fluoroquinolone Gold(III) Chelates as Potential Anticancer Agents: Relevance of DNA and Protein Interactions for Their Mechanism of Action. *Eur. J. Med. Chem.* **2012**, *55*, 67–73. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2012.07.004)
- 46. Kydonaki, T.E.; Tsoukas, E.; Mendes, F.; Hatzidimitriou, A.G.; Paulo, A.; Papadopoulou, L.C.; Papagiannopoulou, D.; Psomas, G. Synthesis, Characterization and Biological Evaluation of (99m)Tc/Re-Tricarbonyl Quinolone Complexes. *J. Inorg. Biochem.* **2016**, *160*, 94–105. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2015.12.010)
- 47. Pagoni, C.-C.; Xylouri, V.-S.; Kaiafas, G.C.; Lazou, M.; Bompola, G.; Tsoukas, E.; Papadopoulou, L.C.; Psomas, G.; Papagiannopoulou, D. Organometallic Rhenium Tricarbonyl-Enrofloxacin and -Levofloxacin Complexes: Synthesis, Albumin-Binding, DNA-Interaction and Cell Viability Studies. *J. Biol. Inorg. Chem.* **2019**, *24*, 609–619. [\[CrossRef\]](https://doi.org/10.1007/s00775-019-01666-1)
- 48. Komarnicka, U.K.; Starosta, R.; Kyzioł, A.; Płotek, M.; Puchalska, M.; Jezowska-Bojczuk, M. New Copper(I) Complexes Bearing ˙ Lomefloxacin Motif: Spectroscopic Properties, in Vitro Cytotoxicity and Interactions with DNA and Human Serum Albumin. *J. Inorg. Biochem.* **2016**, *165*, 25–35. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2016.09.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27764707)
- 49. Galani, A.; Efthimiadou, E.K.; Mitrikas, G.; Sanakis, Y.; Psycharis, V.; Raptopoulou, C.; Kordas, G.; Karaliota, A. Synthesis, Crystal Structure and Characterization of Three Novel Copper Complexes of Levofloxacin. Study of Their DNA Binding Properties and Biological Activities. *Inorganica Chim. Acta* **2014**, *423*, 207–218. [\[CrossRef\]](https://doi.org/10.1016/j.ica.2014.08.005)
- 50. Xiao, Y.; Wang, Q.; Huang, Y.; Ma, X.; Xiong, X.; Li, H. Synthesis, Structure, and Biological Evaluation of a Copper(II) Complex with Fleroxacin and 1,10-Phenanthroline. *Dalton Trans.* **2016**, *45*, 10928–10935. [\[CrossRef\]](https://doi.org/10.1039/C6DT00915H) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27301999)
- 51. Tewes, F.; Bahamondez-Canas, T.F.; Smyth, H.D.C. Efficacy of Ciprofloxacin and Its Copper Complex against Pseudomonas Aeruginosa Biofilms. *AAPS PharmSciTech* **2019**, *20*, 205. [\[CrossRef\]](https://doi.org/10.1208/s12249-019-1417-9)
- 52. Tewes, F.; Bahamondez-Canas, T.F.; Moraga-Espinoza, D.; Smyth, H.D.C.; Watts, A.B. In Vivo Efficacy of a Dry Powder Formulation of Ciprofloxacin-Copper Complex in a Chronic Lung Infection Model of Bioluminescent Pseudomonas Aeruginosa. *Eur. J. Pharm. Biopharm.* **2020**, *152*, 210–217. [\[CrossRef\]](https://doi.org/10.1016/j.ejpb.2020.05.014)
- 53. Gałczy ´nska, K.; Drulis-Kawa, Z.; Arabski, M. Antitumor Activity of Pt(II), Ru(III) and Cu(II) Complexes. *Molecules* **2020**, *25*, 3492. [\[CrossRef\]](https://doi.org/10.3390/molecules25153492)
- 54. Guo, R.-F.; Yan, H.-T.; Liu, R.-X.; Li, H.-C.; Liu, Y.-C.; Chen, Z.-F.; Liang, H. Structural Characterization and Pharmacological Assessment in Vitro/in Vivo of a New Copper(II)-Based Derivative of Enrofloxacin. *Metallomics* **2020**, *12*, 2145–2160. [\[CrossRef\]](https://doi.org/10.1039/d0mt00155d) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33300517)
- 55. Mubarak, A.; Abu Ali, H.; Metani, M. Two Novel Cu (II) Levofloxacin Complexes with Different Bioactive Nitrogen-Based Ligands; Single-Crystal X-Ray and Various Biological Activities Determinations. *Appl. Organomet. Chem.* **2021**, *35*, e6428. [\[CrossRef\]](https://doi.org/10.1002/aoc.6428)
- 56. Bashir, M.; Yousuf, I. Synthesis, Structural Characterization and in Vitro Cytotoxic Evaluation of Mixed Cu(II)/Co(II) Levofloxacin– Bipyridyl Complexes. *Inorganica Chim. Acta* **2022**, *532*, 120757. [\[CrossRef\]](https://doi.org/10.1016/j.ica.2021.120757)
- 57. Bykowska, A.; Komarnicka, U.K.; Jeżowska-Bojczuk, M.; Kyzioł, A. CuI and CuII Complexes with Phosphine Derivatives of Fluoroquinolone Antibiotics—A Comparative Study on the Cytotoxic Mode of Action. *J. Inorg. Biochem.* **2018**, *181*, 1–10. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2018.01.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29348049)
- 58. Guz-Regner, K.; Komarnicka, U.K.; Futoma-Kołoch, B.; Wernecki, M.; Cal, M.; Kozieł, S.; Ziółkowska, A.; Bugla-Płoskońska, G. Antibacterial Activity and Action Mode of Cu(I) and Cu(II) Complexes with Phosphines Derived from Fluoroquinolone against Clinical and Multidrug-Resistant Bacterial Strains. *J. Inorg. Biochem.* **2020**, *210*, 111124. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2020.111124) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32534287)
- 59. Mjos, K.D.; Polishchuk, E.; Abrams, M.J.; Orvig, C. Synthesis, Characterization, and Evaluation of the Antimicrobial Potential of Copper(II) Coordination Complexes with Quinolone and p-Xylenyl-Linked Quinolone Ligands. *J. Inorg. Biochem.* **2016**, *162*, 280–285. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2016.02.026) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26979255)
- 60. Bhatt, B.S.; Gandhi, D.H.; Vaidya, F.U.; Pathak, C.; Patel, T.N. Cell Apoptosis Induced by Ciprofloxacin Based Cu(II) Complexes: Cytotoxicity, SOD Mimic and Antibacterial Studies. *J. Biomol. Struct. Dyn.* **2021**, *39*, 4555–4562. [\[CrossRef\]](https://doi.org/10.1080/07391102.2020.1776641) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32476567)
- 61. Ude, Z.; Flothkötter, N.; Sheehan, G.; Brennan, M.; Kavanagh, K.; Marmion, C.J. Multi-Targeted Metallo-Ciprofloxacin Derivatives Rationally Designed and Developed to Overcome Antimicrobial Resistance. *Int. J. Antimicrob. Agents* **2021**, *58*, 106449. [\[CrossRef\]](https://doi.org/10.1016/j.ijantimicag.2021.106449)
- 62. Liu, Q.-Y.; Qi, Y.-Y.; Cai, D.-H.; Liu, Y.-J.; He, L.; Le, X.-Y. Sparfloxacin—Cu(II)—Aromatic Heterocyclic Complexes: Synthesis, Characterization and in Vitro Anticancer Evaluation. *Dalton Trans.* **2022**, *51*, 9878–9887. [\[CrossRef\]](https://doi.org/10.1039/D2DT00077F)
- 63. Ahmadi, F.; Ebrahimi-Dishabi, N.; Mansouri, K.; Salimi, F. Molecular Aspect on the Interaction of Zinc-Ofloxacin Complex with Deoxyribonucleic Acid, Proposed Model for Binding and Cytotoxicity Evaluation. *Res. Pharm. Sci.* **2014**, *9*, 367–383.
- 64. Ahmadi, F.; Saberkari, M.; Abiri, R.; Motlagh, H.M.; Saberkari, H. In Vitro Evaluation of Zn-Norfloxacin Complex as a Potent Cytotoxic and Antibacterial Agent, Proposed Model for DNA Binding. *Appl. Biochem. Biotechnol.* **2013**, *170*, 988–1009. [\[CrossRef\]](https://doi.org/10.1007/s12010-013-0255-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23636652)
- 65. Shahabadi, N.; Asadian, A.A.; Mahdavi, M. Intercalation of a Zn(II) Complex Containing Ciprofloxacin Drug between DNA Base Pairs. *Nucleosides Nucleotides Nucleic Acids* **2017**, *36*, 676–689. [\[CrossRef\]](https://doi.org/10.1080/15257770.2017.1388394) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29185900)
- 66. Sakr, S.H.; Elshafie, H.S.; Camele, I.; Sadeek, S.A. Synthesis, Spectroscopic, and Biological Studies of Mixed Ligand Complexes of Gemifloxacin and Glycine with Zn(II), Sn(II), and Ce(III). *Molecules* **2018**, *23*, 1182. [\[CrossRef\]](https://doi.org/10.3390/molecules23051182)
- 67. Elshafie, H.S.; Sakr, S.H.; Sadeek, S.A.; Camele, I. Biological Investigations and Spectroscopic Studies of New Moxifloxacin/Glycine-Metal Complexes. *Chem. Biodivers.* **2019**, *16*, e1800633. [\[CrossRef\]](https://doi.org/10.1002/cbdv.201800633) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30629800)
- 68. Psomas, G.; Kessissoglou, D.P. Quinolones and Non-Steroidal Anti-Inflammatory Drugs Interacting with Copper(II), Nickel(II), Cobalt(II) and Zinc(II): Structural Features, Biological Evaluation and Perspectives. *Dalton Trans.* **2013**, *42*, 6252–6276. [\[CrossRef\]](https://doi.org/10.1039/c3dt50268f) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23529676)
- 69. Protogeraki, C.; Andreadou, E.G.; Perdih, F.; Turel, I.; Pantazaki, A.A.; Psomas, G. Cobalt(II) Complexes with the Antimicrobial Drug Enrofloxacin: Structure, Antimicrobial Activity, DNA- and Albumin-Binding. *Eur. J. Med. Chem.* **2014**, *86*, 189–201. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2014.08.043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25151581)
- 70. Kouris, E.; Kalogiannis, S.; Perdih, F.; Turel, I.; Psomas, G. Cobalt(II) Complexes of Sparfloxacin: Characterization, Structure, Antimicrobial Activity and Interaction with DNA and Albumins. *J. Inorg. Biochem.* **2016**, *163*, 18–27. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2016.07.022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27501348)
- 71. Kozsup, M.; Farkas, E.; Bényei, A.C.; Kasparkova, J.; Crlikova, H.; Brabec, V.; Buglyó, P. Synthesis, Characterization and Biological Evaluation of Co(III) Complexes with Quinolone Drugs. *J. Inorg. Biochem.* **2019**, *193*, 94–105. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2019.01.005)
- 72. Singh, B.; Kisku, T.; Das, S.; Mukherjee, S.; Kundu, A.; Rath, J.; Das, R.S. Refashioning of the Drug-Properties of Fluoroquinolone through the Synthesis of a Levofloxacin-Imidazole Cobalt (II) Complex and Its Interaction Studies on with DNA and BSA Biopolymers, Antimicrobial and Cytotoxic Studies on Breast Cancer Cell Lines. *Int. J. Biol. Macromol.* **2023**, *253*, 127636. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2023.127636)
- 73. Zampakou, M.; Balala, S.; Perdih, F.; Kalogiannis, S.; Turel, I.; Psomas, G. Structure, Antimicrobial Activity, Albumin- and DNA-Binding of Manganese(II)–Sparfloxacinato Complexes. *RSC Adv.* **2015**, *5*, 11861–11872. [\[CrossRef\]](https://doi.org/10.1039/C4RA11682H)
- 74. Di Virgilio, A.L.; León, I.E.; Franca, C.A.; Henao, I.; Tobón, G.; Etcheverry, S.B. Cu(Nor)² ·5H2O, a Complex of Cu(II) with Norfloxacin: Theoretic Approach and Biological Studies. Cytotoxicity and Genotoxicity in Cell Cultures. *Mol. Cell Biochem.* **2013**, *376*, 53–61. [\[CrossRef\]](https://doi.org/10.1007/s11010-012-1548-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23238873)
- 75. Katsarou, M.E.; Efthimiadou, E.K.; Psomas, G.; Karaliota, A.; Vourloumis, D. Novel Copper(II) Complex of N-Propyl-Norfloxacin and 1,10-Phenanthroline with Enhanced Antileukemic and DNA Nuclease Activities. *J. Med. Chem.* **2008**, *51*, 470–478. [\[CrossRef\]](https://doi.org/10.1021/jm7013259) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18205294)
- 76. Shingnapurkar, D.; Butcher, R.; Afrasiabi, Z.; Sinn, E.; Ahmed, F.; Sarkar, F.; Padhye, S. Neutral Dimeric Copper–Sparfloxacin Conjugate Having Butterfly Motif with Antiproliferative Effects against Hormone Independent BT20 Breast Cancer Cell Line. *Inorg. Chem. Commun.* **2007**, *10*, 459–462. [\[CrossRef\]](https://doi.org/10.1016/j.inoche.2006.12.016)
- 77. He, X.; Yao, Q.; Hall, D.D.; Song, Z.; Fan, D.; You, Y.; Lian, W.; Zhou, Z.; Duan, L.; Chen, B. Theoretical, in Vitro Antiproliferative, and in Silico Molecular Docking and Pharmacokinetics Studies of Heteroleptic Nickel(II) and Copper(II) Complexes of Thiosemicarbazone-Based Ligands and Pefloxacin. *Chem. Biodivers.* **2023**, *20*, e202300702. Available online: [https:](https://onlinelibrary.wiley.com/doi/full/10.1002/cbdv.202300702) [//onlinelibrary.wiley.com/doi/full/10.1002/cbdv.202300702](https://onlinelibrary.wiley.com/doi/full/10.1002/cbdv.202300702) (accessed on 11 June 2024).
- 78. Gandhi, D.H.; Vaidya, F.U.; Pathak, C.; Patel, T.N.; Bhatt, B.S. Mechanistic Insight of Cell Anti-Proliferative Activity of Fluoroquinolone Drug-Based Cu(II) Complexes. *Mol. Divers.* **2022**, *26*, 869–878. [\[CrossRef\]](https://doi.org/10.1007/s11030-021-10199-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33646502)
- 79. Teo, R.D.; Termini, J.; Gray, H.B. Lanthanides: Applications in Cancer Diagnosis and Therapy. *J. Med. Chem.* **2016**, *59*, 6012–6024. [\[CrossRef\]](https://doi.org/10.1021/acs.jmedchem.5b01975)
- 80. Nghia, N.N.; Huy, B.T.; Phong, P.T.; Han, J.S.; Kwon, D.H.; Lee, Y.-I. Simple Fluorescence Optosensing Probe for Spermine Based on Ciprofloxacin-Tb3+ Complexation. *PLoS ONE* **2021**, *16*, e0251306. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0251306)
- 81. Li, J.-B.; Yang, P.; Gao, F.; Han, G.-Y.; Yu, K.-B. Novel Lanthanide Complexes of Ciprofloxacin: Synthesis, Characterization, Crystal Structure and in Vitro Antibacterial Activity Studies. *Chin. J. Chem.* **2001**, *19*, 598–605. [\[CrossRef\]](https://doi.org/10.1002/cjoc.20010190611)
- 82. Shaban, S.Y.; El-Kemary, M.A.; Samir, G.; Elbaradei, H. Synthesis, Characterization, Antibacterial Activities Testing and the Interaction of DNA with Ciprofloxacin and Its La(III)-Based Complex. *J. Chin. Adv. Mater. Soc.* **2018**, *6*, 123–133. [\[CrossRef\]](https://doi.org/10.1080/22243682.2018.1425907)
- 83. Wang, Y.-J.; Hu, R.-D.; Jiang, D.-H.; Zhang, P.-H.; Lin, Q.-Y.; Wang, Y.-Y. Synthesis, Crystal Structure, Interaction with BSA and Antibacterial Activity of La(III) and Sm(III) Complexes with Enrofloxacin. *J. Fluoresc.* **2011**, *21*, 813–823. [\[CrossRef\]](https://doi.org/10.1007/s10895-010-0775-1)
- 84. Sadeek, S.A.; Abd El-Hamid, S.M.; El-Aasser, M.M. Synthesis, Characterization, Antimicrobial and Cytotoxicity Studies of Some Transition Metal Complexes with Gemifloxacin. *Monatsh. Chem.* **2015**, *146*, 1967–1982. [\[CrossRef\]](https://doi.org/10.1007/s00706-015-1507-7)
- 85. Sadeek, S.; El-Shwiniy, W. Preparation, Structural Characterization and Biological Studies of Some New Levofloxacin Metal Complexes. *J. Iran. Chem. Soc.* **2017**, *14*, 1711–1723. [\[CrossRef\]](https://doi.org/10.1007/s13738-017-1112-2)
- 86. Refat, M.; El-Hawary, W.; Mohamed, M. Study of the chemical chelates and anti-microbial effect of some metal ions in nanostructural form on the efficiency of antibiotic therapy "norfloxacin drug". *J. Mol. Struct.* **2012**, *1013*, 45–54. [\[CrossRef\]](https://doi.org/10.1016/j.molstruc.2011.12.010)
- 87. el-Gamel, N.E.A.; Zayed, M.A. Synthesis, Structural Characterization and Antimicrobial Activity Evaluation of Metal Complexes of Sparfloxacin. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2011**, *82*, 414–423. [\[CrossRef\]](https://doi.org/10.1016/j.saa.2011.07.072) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21856215)
- 88. Arnaouti, E.; Georgiadou, C.; Hatizdimitriou, A.G.; Kalogiannis, S.; Psomas, G. Erbium(III) Complexes with Fluoroquinolones: Structure and Biological Properties. *J. Inorg. Biochem.* **2024**, *255*, 112525. [\[CrossRef\]](https://doi.org/10.1016/j.jinorgbio.2024.112525) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38522216)
- 89. Castro, W.; Navarro, M.; Biot, C. Medicinal Potential of Ciprofloxacin and Its Derivatives. *Future Med. Chem.* **2013**, *5*, 81–96. [\[CrossRef\]](https://doi.org/10.4155/fmc.12.181) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23256815)
- 90. Cardoso-Ortiz, J.; Leyva-Ramos, S.; Baines, K.M.; Gómez-Durán, C.F.A.; Hernández-López, H.; Palacios-Can, F.J.; Valcarcel-Gamiño, J.A.; Leyva-Peralta, M.A.; Razo-Hernández, R.S. Novel Ciprofloxacin and Norfloxacin-Tetrazole Hybrids as Potential Antibacterial and Antiviral Agents: Targeting, S. Aureus Topoisomerase and SARS-CoV-2-MPro. *J. Mol. Struct.* **2023**, *1274*, 134507. [\[CrossRef\]](https://doi.org/10.1016/j.molstruc.2022.134507)
- 91. Alaaeldin, R.; Mustafa, M.; Abuo-Rahma, G.E.-D.A.; Fathy, M. In Vitro Inhibition and Molecular Docking of a New Ciprofloxacin-Chalcone against SARS-CoV-2 Main Protease. *Fundam. Clin. Pharmacol.* **2022**, *36*, 160–170. [\[CrossRef\]](https://doi.org/10.1111/fcp.12708)
- 92. Ahadi, H.; Emami, S. Modification of 7-Piperazinylquinolone Antibacterials to Promising Anticancer Lead Compounds: Synthesis and in Vitro Studies. *Eur. J. Med. Chem.* **2020**, *187*, 111970. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2019.111970)
- 93. Chrzanowska, A.; Roszkowski, P.; Bielenica, A.; Olejarz, W.; Stępień, K.; Struga, M. Anticancer and Antimicrobial Effects of Novel Ciprofloxacin Fatty Acids Conjugates. *Eur. J. Med. Chem.* **2020**, *185*, 111810. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2019.111810)
- 94. Akhtar, R.; Noreen, R.; Raza, Z.; Rasul, A.; Zahoor, A.F. Synthesis, Anticancer Evaluation, and In Silico Modeling Study of Some N-Acylated Ciprofloxacin Derivatives. *Russ. J. Org. Chem.* **2022**, *58*, 541–548. [\[CrossRef\]](https://doi.org/10.1134/S107042802204011X)
- 95. Ahadi, H.; Shokrzadeh, M.; Hosseini-Khah, Z.; Ghassemi barghi, N.; Ghasemian, M.; Emadi, E.; Zargari, M.; Razzaghi-Asl, N.; Emami, S. Synthesis and Biological Assessment of Ciprofloxacin-Derived 1,3,4-Thiadiazoles as Anticancer Agents. *Bioorganic Chem.* **2020**, *105*, 104383. [\[CrossRef\]](https://doi.org/10.1016/j.bioorg.2020.104383) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33130342)
- 96. Kassab, A.E.; Gedawy, E.M. Novel Ciprofloxacin Hybrids Using Biology Oriented Drug Synthesis (BIODS) Approach: Anticancer Activity, Effects on Cell Cycle Profile, Caspase-3 Mediated Apoptosis, Topoisomerase II Inhibition, and Antibacterial Activity. *Eur. J. Med. Chem.* **2018**, *150*, 403–418. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2018.03.026) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29547830)
- 97. Ezelarab, H.A.A.; Hassan, H.A.; Abuo-Rahma, G.E.-D.A.; Abbas, S.H. Design, Synthesis, and Biological Investigation of Quinoline/Ciprofloxacin Hybrids as Antimicrobial and Anti-Proliferative Agents. *J. Iran. Chem. Soc.* **2023**, *20*, 683–700. [\[CrossRef\]](https://doi.org/10.1007/s13738-022-02704-7)
- 98. Fallica, A.N.; Barbaraci, C.; Amata, E.; Pasquinucci, L.; Turnaturi, R.; Dichiara, M.; Intagliata, S.; Gariboldi, M.B.; Marras, E.; Orlandi, V.T.; et al. Nitric Oxide Photo-Donor Hybrids of Ciprofloxacin and Norfloxacin: A Shift in Activity from Antimicrobial to Anticancer Agents. *J. Med. Chem.* **2021**, *64*, 11597–11613. [\[CrossRef\]](https://doi.org/10.1021/acs.jmedchem.1c00917) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34319100)
- 99. Hhh, M.; Sh, A.; Am, H.; Gea, A.-R.; Ya, M. Novel Urea Linked Ciprofloxacin-Chalcone Hybrids Having Antiproliferative Topoisomerases I/II Inhibitory Activities and Caspases-Mediated Apoptosis. *Bioorganic Chem.* **2021**, *106*. [\[CrossRef\]](https://doi.org/10.1016/j.bioorg.2020.104422)
- 100. Szostek, T.; Szulczyk, D.; Szymańska-Majchrzak, J.; Koliński, M.; Kmiecik, S.; Otto-Ślusarczyk, D.; Zawodnik, A.; Rajkowska, E.; Chaniewicz, K.; Struga, M.; et al. Design and Synthesis of Menthol and Thymol Derived Ciprofloxacin: Influence of Structural Modifications on the Antibacterial Activity and Anticancer Properties. *Int. J. Mol. Sci.* **2022**, *23*, 6600. [\[CrossRef\]](https://doi.org/10.3390/ijms23126600)
- 101. Ming, T.; Lei, J.; Peng, Y.; Wang, M.; Liang, Y.; Tang, S.; Tao, Q.; Wang, M.; Tang, X.; He, Z.; et al. Curcumin Suppresses Colorectal Cancer by Induction of Ferroptosis via Regulation of P53 and Solute Carrier Family 7 Member 11/Glutathione/Glutathione Peroxidase 4 Signaling Axis. *Phytother. Res.* **2024**. Available online: <https://onlinelibrary.wiley.com/doi/abs/10.1002/ptr.8258> (accessed on 18 June 2024). [\[CrossRef\]](https://doi.org/10.1002/ptr.8258)
- 102. Li, J.; Feng, S.; Wang, X.; Zhang, B.; He, Q. Exploring the Targets and Molecular Mechanisms of Curcumin for the Treatment of Bladder Cancer Based on Network Pharmacology, Molecular Docking and Molecular Dynamics. *Mol. Biotechnol.* **2024**. [\[CrossRef\]](https://doi.org/10.1007/s12033-024-01190-x)
- 103. Mokbel, K.; Mokbel, K. Harnessing Micronutrient Power: Vitamins, Antioxidants and Probiotics in Breast Cancer Prevention. *Anticancer Res.* **2024**, *44*, 2287–2295. [\[CrossRef\]](https://doi.org/10.21873/anticanres.17036)
- 104. Schiavoni, V.; Emanuelli, M.; Sartini, D.; Salvolini, E.; Pozzi, V.; Campagna, R. Curcumin and Its Analogues in Oral Squamous Cell Carcinoma: State-of-the-Art and Therapeutic Potential. *Anti-Cancer Agents Med. Chem.* **2024**, *24*, 1–18. [\[CrossRef\]](https://doi.org/10.2174/0118715206297840240510063330) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38757321)
- 105. Karimzadeh, M.R.; Masoudi Chelegahi, A.; Shahbazi, S.; Reiisi, S. Co-Treatment of Silymarin and Cisplatin Inhibited Cell Proliferation, Induced Apoptosis in Ovarian Cancer. *Mol. Biol. Rep.* **2024**, *51*, 118. [\[CrossRef\]](https://doi.org/10.1007/s11033-023-09026-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38227082)
- 106. Fu, W.; Liu, L.; Tong, S. Berberine Inhibits the Progression of Breast Cancer by Regulating METTL3-Mediated m6A Modification of FGF7 mRNA. *Thorac. Cancer* **2024**, *15*, 1357–1368. [\[CrossRef\]](https://doi.org/10.1111/1759-7714.15321) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38709912)
- 107. Ruan, L.; Jiao, J.; Cheng, C.; Zhang, Y.; Cao, Z.; He, B.; Chen, Z. Berberine Chloride Suppresses Pancreatic Adenocarcinoma Proliferation and Growth by Targeting Inflammation-Related Genes: An in Silico Analysis with in Vitro and Vivo Validation. *Cancer Chemother. Pharmacol.* **2024**, 1–13. [\[CrossRef\]](https://doi.org/10.1007/s00280-024-04663-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38502348)
- 108. Khaled, A.M.; Othman, M.S.; Obeidat, S.T.; Aleid, G.M.; Aboelnaga, S.M.; Fehaid, A.; Hathout, H.M.R.; Bakkar, A.A.; Moneim, A.E.A.; El-Garawani, I.M.; et al. Green-Synthesized Silver and Selenium Nanoparticles Using Berberine: A Comparative Assessment of In Vitro Anticancer Potential on Human Hepatocellular Carcinoma Cell Line (HepG2). *Cells* **2024**, *13*, 287. [\[CrossRef\]](https://doi.org/10.3390/cells13030287) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38334679)
- 109. Milata, V.; Svedova, A.; Barbierikova, Z.; Holubkova, E.; Cipakova, I.; Cholujova, D.; Jakubikova, J.; Panik, M.; Jantova, S.; Brezova, V.; et al. Synthesis and Anticancer Activity of Novel 9-O-Substituted Berberine Derivatives. *Int. J. Mol. Sci.* **2019**, *20*, 2169. [\[CrossRef\]](https://doi.org/10.3390/ijms20092169) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31052469)
- 110. Alaaeldin, R.; Abdel-Rahman, I.M.; Ali, F.E.M.; Bekhit, A.A.; Elhamadany, E.Y.; Zhao, Q.-L.; Cui, Z.-G.; Fathy, M. Dual Topoisomerase I/II Inhibition-Induced Apoptosis and Necro-Apoptosis in Cancer Cells by a Novel Ciprofloxacin Derivative via RIPK1/RIPK3/MLKL Activation. *Molecules* **2022**, *27*, 7993. [\[CrossRef\]](https://doi.org/10.3390/molecules27227993) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36432094)
- 111. Fawzy, M.A.; Abu-baih, R.H.; Abuo-Rahma, G.E.-D.A.; Abdel-Rahman, I.M.; El-Sheikh, A.A.K.; Nazmy, M.H. In Vitro Anticancer Activity of Novel Ciprofloxacin Mannich Base in Lung Adenocarcinoma and High-Grade Serous Ovarian Cancer Cell Lines via Attenuating MAPK Signaling Pathway. *Molecules* **2023**, *28*, 1137. [\[CrossRef\]](https://doi.org/10.3390/molecules28031137) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36770806)
- 112. Struga, M.; Roszkowski, P.; Bielenica, A.; Otto-Ślusarczyk, D.; Stępień, K.; Stefańska, J.; Zabost, A.; Augustynowicz-Kopeć, E.; Koliński, M.; Kmiecik, S.; et al. N-Acylated Ciprofloxacin Derivatives: Synthesis and In Vitro Biological Evaluation as Antibacterial and Anticancer Agents. *ACS Omega* **2023**, *8*, 18663–18684. [\[CrossRef\]](https://doi.org/10.1021/acsomega.3c00554)
- 113. Xi, X.-X.; Hei, Y.-Y.; Guo, Y.; Zhao, H.-Y.; Xin, M.; Lu, S.; Jiang, C.; Zhang, S.-Q. Identification of Benzamides Derivatives of Norfloxacin as Promising microRNA-21 Inhibitors via Repressing Its Transcription. *Bioorg. Med. Chem.* **2022**, *66*, 116803. [\[CrossRef\]](https://doi.org/10.1016/j.bmc.2022.116803)
- 114. Hei, Y.-Y.; Wang, S.; Xi, X.-X.; Wang, H.-P.; Guo, Y.; Xin, M.; Jiang, C.; Lu, S.; Zhang, S.-Q. Design, Synthesis, and Evaluation of Fluoroquinolone Derivatives as microRNA-21 Small-Molecule Inhibitors. *J. Pharm. Anal.* **2022**, *12*, 653–663. [\[CrossRef\]](https://doi.org/10.1016/j.jpha.2021.12.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36105166)
- 115. Wang, X.; Jiang, X.; Sun, S.; Liu, Y. Synthesis and Biological Evaluation of Novel Quinolone Derivatives Dual Targeting Histone Deacetylase and Tubulin Polymerization as Antiproliferative Agents. *RSC Adv.* **2018**, *8*, 16494–16502. [\[CrossRef\]](https://doi.org/10.1039/C8RA02578A) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35540517)
- 116. Beberok, A.; Rok, J.; Rzepka, Z.; Marciniec, K.; Boryczka, S.; Wrześniok, D. The Role of MITF and Mcl-1 Proteins in the Antiproliferative and Proapoptotic Effect of Ciprofloxacin in Amelanotic Melanoma Cells: In Silico and in Vitro Study. *Toxicol. Vitr.* **2020**, *66*, 104884. [\[CrossRef\]](https://doi.org/10.1016/j.tiv.2020.104884) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32437906)
- 117. Suresh, N.; Suresh, A.; Yerramsetty, S.; Bhadra, M.P.; Alvala, M.; Sekhar, K.V.G.C. Anti-Proliferative Activity, Molecular Modeling Studies and Interaction with Calf Thymus DNA of Novel Ciprofloxacin Analogues. *J. Chem. Sci.* **2018**, *130*, 121. [\[CrossRef\]](https://doi.org/10.1007/s12039-018-1528-y)
- 118. Allaka, T.R.; Katari, N.K.; Veeramreddy, V.; Anireddy, J.S. Molecular Modeling Studies of Novel Fluoroquinolone Molecules. *Curr. Drug Discov. Technol.* **2018**, *15*, 109–122. [\[CrossRef\]](https://doi.org/10.2174/1570163814666170829142044)

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