RESEARCH ARTICLE

Evaluation of polycyclic aromatic hydrocarbon contents in marine products in South Korea and risk assessment using the total diet study

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Abstract

This study investigated levels of eight polycyclic aromatic hydrocarbons (PAH8) compounds in both raw and processed marine products in South Korea. Katsuobushi exhibited the highest concentration of benzo[a]pyrene, at 14.22 µg/kg, exceeding the European Commission's regulation level of 5.0 µg/kg. The total PAH8 concentration in katsuobushi was 220.5 µg/ kg. Among the product categories, shellfsh had the highest detection rate (70%), followed by fsh (19%) and crustacea (8%), with chrysene being the most prominent PAH8 congener in all marine products. Grilled fish predominantly contained pyrogenic PAHs from combustion byproducts, while shellfsh primarily contained petrogenic ones from the aquatic environment. Grilling, smoking, and drying processes signifcantly contributed to the formation of PAH8 in these food products. Based on the results of a risk assessment using a margin of exposure approach through a total diet study, exposure to PAH8 from marine products is considered to pose low concern to the South Korean population.

Keywords Polycyclic aromatic hydrocarbon · Marine product · Total diet study · Risk assessment · Gas chromatographymass spectrometry

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Introduction

Polycyclic aromatic hydrocarbons (PAHs) can come from three sources: natural (biogenic), combustion (pyrogenic), and petroleum (petrogenic) (Abbas et al., [2018\)](#page-12-0). Biogenic PAHs stem from living organisms, while pyrogenic PAHs result from the high-temperature processing of organic matter. Petrogenic PAHs enter the environment through natural processes like gas leaks and fossil fuel seepage (Ofosu et al, [2022](#page-13-0)).

Many PAHs are mutagenic and genotoxic, capable of causing DNA adduct formation in both laboratory and living organisms (Zaidi, et al., [2021\)](#page-13-1). In addition to cancer, PAHs can lead to other adverse effects, including neurobehavioral changes in developing animals, decreased ovarian follicles and ovary weight, and altered thymus weight and serum immunoglobulin in rats (Kitts et al., [2012](#page-13-2); Kroese et al., [2002\)](#page-13-3).

The European Food Safety Authority (EFSA) indicated that utilizing the total PAH level of benzo[a]anthracene (B[a]A), chrysene (Chry), benzo[a]pyrene (B[a]P), and benzo[b]fuoranthene (B[b]F) represented for PAH4, or including B[a]A, chrysene, B[a]P, B[b]F, benzo[k]fuoranthene (B[k]F), indeno[1,2,3-cd]pyrene (I[c,d]P), dibenz[a,h]

anthracene ($D[a,h]A$), and benzo $[g,h,i]$ perylene ($B[g,h,i]P$) represented for PAH8 is more appropriate than using solely B[a]P level (EC, [2015\)](#page-12-1). The primary human exposure factor is known to be dietary intake, and the risk of exposure to PAHs in aquatic products has increased due to higher seafood consumption and pollution of the aquatic environment by human activities (Alomirah et al., [2011;](#page-12-2) Llobet et al., [2006\)](#page-13-4). Other studies also found that seafood is becoming the leading source of dietary PAH exposure (EFSA, [2008](#page-12-3); Habibullah-Al-Mamun et al., [2019](#page-13-5); Veyrand et al., [2013](#page-13-6)).

As a result, we employed a total diet study (TDS) approach to assess the levels of PAHs in marine products and investigate variations in PAH contents based on the preparation methods of these marine food items. TDS is one of the most effective and efficient methods for estimating dietary exposure and assessing health risk at the consumer level, considering the purchasing and eating habits of various populations (Ingenbleek et al., [2017\)](#page-13-7). There are limited studies that monitor the levels of benzopyrene, the total of PAH4 (∑PAH4), and the total of PAH8 (∑PAH8) generated during the processing of marine products. In the case of barbecued salmon fllet and grilled anchovies, it was observed that they exhibited elevated levels of benzo[a]pyrene, the total of PAH4 (Σ PAH4), and the total of PAH8 (Σ PAH8), with concentrations of 0.52 µg/kg, 2.41 µg/kg, and 2.88 µg/ kg for the former (Oz, [2020](#page-13-8)), and 0.73 µg/kg, 3.3 µg/kg, and 5.13 µg/kg for the latter (Sahin et al., [2020](#page-13-9)).

Because the toxicity of each PAH difers, the toxicity is assessed using a toxic equivalency factor (TEF) using benzo[a]pyrene as a reference molecule. EFSA adopted a margin of exposure (MOE) approach to estimate health risk to PAHs, based on benchmark dose lower-bound confdence limit 10% (BMDL₁₀) (EFSA, 2008). However, EFSA has concluded the toxic equivalent quantity (TEQ) approach is only suitable for compounds with the same toxicological efect, such as dioxins. Given that several PAHs have carcinogenic properties and produce tumors through diferent mechanisms, the TEQ approach may not be appropriate for PAHs (EFSA, [2008](#page-12-3)). Using toxicological values, rather than the TEQ, may be necessary for assessing the risks associated with PAHs.

The primary objectives of this research are to: (1) validate methods for extracting PAHs to evaluate the PAH8 concentration, (2) identify the main contributors of PAH8 exposure in humans in terms of each PAH compound and the origin of PAHs associated with processing methods, (3) determine the types of marine products generating most PAH8, (4) assess the health risks associated with PAH8 from the consumption of marine products in South Korea, and (5) to compared the risks estimated using toxicological values and traditional TEF values. This study focused on examining the PAH8 levels in marine products in a table-ready form within the framework of a TDS.

Materials and methods

Sample selection and preparation

Marine product samples were selected based on the results of the 6th Korean National Health and Nutrition Examination Survey (KNHANES-VI) conducted by Korea Disease Control and Prevention Agency (KDCA) from 2013 to 2015 (MOHW, [2013\)](#page-13-10). The raw data were obtained from the KNHANES website ([https://knhanes.kdca.go.kr/knhan](https://knhanes.kdca.go.kr/knhanes/main.do) [es/main.do\)](https://knhanes.kdca.go.kr/knhanes/main.do).

Supplementary Table 1 contains a list of 109 products with high consumption rates (covering more than 95% of total accumulated consumption), high frequency levels (covering more than 1% of the consumption rate), and fat contribution (containing the cumulative rate of fat intake up to 95%). The food samples were gathered from 21 supermarkets in ten large cities with a population of over one million, including Seoul, Incheon, Suwon, Gwangju, Daejeon, Cheongju, Daegu, Busan, Ulsan, and Changwon in South Korea.

An identical quantity of food samples were gathered to create each composite sample, and they were prepared in a table-ready form according to the sample preparation methods guidebook for TDS provided by the Ministry of Food and Drug Safety (MFDS, [2019](#page-13-11)). The cooked samples $(n = 287)$ are divided into eight groups: fish (139), shellfsh (46), cephalopoda (25), crustacea (37), sea algae (30), echinodermata (5), tunicata (3), and cnidaria (2). A total of 287 subsamples were homogenized and kept in a polyethylene bottle at −20 °C for PAH analysis.

Chemicals and materials

HPLC grade ethyl alcohol, methyl alcohol, n-hexane, and dichloromethane (DCM) were purchased from Burdick & Jackson (Muskegon, MI, USA). Younglin Instrument's AQUAMAX-Basic 363 water purifcation system (Dongan, Anyang, Republic of Korea) was used to produce distilled water. EPA 525 PAH Mix A, which contains 500 µg/mL of naphthalene (99% purity), acenaphthene (99% purity), fuoranthene (98% purity), acenaphthylene (99% purity), fuorene, phenanthrene, anthracene, pyrene, benz[a]anthracene, chrysene, benzo[b]fuoranthene (98% purity), benzo[k]fluoranthene, benzo[a]pyrene (96% purity), indeno[1,2,3-cd]pyrene, dibenz[a,h]anthracene, and benzo[g,h,i]perylene in dichloromethane was purchased from Sigma Aldrich (St. Louis, MO, USA). Benzo[a]pyrene-d₁₂ (98% purity), benzo[a]pyrene-d12 (98% purity), benzo[b]fuoranthene-d12 (98% purity), and chrysene-d₁₂ (98% purity) were purchased from Sigma

Aldrich for internal standards. Potassium hydroxide was purchased from Showa Denko (Tokyo, Japan), and anhydrous sodium sulfate was from Yakuri Pure Chemicals (Kyoto, Japan). Filter paper was purchased from Whatman (Kent, UK) and Bond Elut SI for solid phase extraction (SPE) was obtained from Agilent Technologies (Santa Clara, CA, USA).

Extraction of PAHs

Extraction steps are essential for determining PAHs. Depending on the properties of the sample matrices, two distinct extraction procedures were employed. These methods were adapted in accordance with the Korea Food Code and previous studies (Kim et al., [2021](#page-13-12); MFDS, [2019\)](#page-13-11).

For solid food matrices such as salmon and oyster, alkali digestion was applied to extract PAHs from the products. In each flask, 10 g (wet weight) of samples or $1-2$ g (dry weight) of sample were placed. Then, 100 mL of 1 M KOH solution in ethanol was added, along with 1 mL of $13C$ -labeled internal standard solution (100 µg/kg of each benzo[a]pyrene- d_{12} and chrysene- d_{12}). The flask was connected to a refux condenser and placed into a water bath, WB-22, (Daihan Scientifc, Gangwon, Republic of Korea) at 80℃ for 3 h. After saponifcation, the fask was rapidly cooled down using cold water, and the refux condensers were rinsed with n-hexane. The extract in the fask was transferred into a separatory funnel through a flter paper. The fask was washed with 50 mL of n-hexane: ethanol (1:1, v/v) solution, and the washed solution was added into the funnel. The funnel was thoroughly shaken using a funnel shaker (Changshin Science, Seoul, Republic of Korea) with 300 rpm for 10 min after adding 50 mL of distilled water. After shaking, the separated organic phase fraction from the organic solvent was collected in each Erlenmeyer fask. Then, 50 mL of n-hexane was added to distilled water in the separatory funnel and shaken to separate two immiscible liquid phases. This procedure was repeated twice. All organic solvent phases were collected in another separatory funnel, and distilled water was added to remove watersoluble compounds. The water layer was eliminated after vigorously shaking the funnel. The obtained extract was filtered through 10 g of anhydrous $Na₂SO₄$ (Yakuri Pure Chemicals, Kyoto, Japan) to remove any remaining water. Next, the extracts were concentrated using a rotary evaporator (EYELA, Tokyo, Japan) until the fnal volume was below 2 mL. The concentrate was applied to activate SPE (Bond Elut SI) cartridges (Agilent technologies) and eluted with n-hexane and DCM. The eluant was concentrated under $N₂$ gas at 40 °C, and the residues were re-dissolved in 1 mL of DCM. The solution was filtered through 0.45 µm of PTFE membrane syringe flter for gas chromatography-mass spectrometry (GC–MS) analysis.

For liquid food matrices, such as fsh sauce or seafood stock, an ultrasound-assisted extraction method was used to quantify PAHs. In each fask, 10 g of samples were placed, and 50 mL of n-hexane spiked with 1 mL of internal standard solution was added. The mixture was then subjected to 20 min of ultrasonication. Subsequently, 35 mL of n-hexane was added, and ultrasound-assisted extraction was carried out for another 20 min. The following extraction steps for liquid food samples were conducted in the same manner as previously described methods for solid food samples.

GC–MS analysis

PAH levels were determined using gas chromatography-mass spectrometry (GC–MS, 7890B/5977B, Agilent Technologies) with a HP-5MS Ultra Inert column (30 m \times 0.25 mm \times 0.25 µm) (Agilent Technologies). Helium was used as a carrier gas with a flow rate 1.0 mL/min. The GC oven temperature was programmed as follows: initially set at 80 °C and held for 1 min, then heated at a rate of 20 °C/min up to 220 °C and held for 10 min, followed by an increase to 280 °C at a rate of 2 °C/min and maintained for 10 min. Mass spectra were generated using an electron ionization ion source at 70 eV in scan mode to determine a quantitative ion and two qualitative ions (Table [1\)](#page-3-0). PAH peaks were identifed using the ion in the selected ion monitoring mode.

Method validation

Salmon (solid-type) and fish sauce (liquid-type) were utilized as representative samples for each alkali digestion method and ultrasound-assisted extraction method to validate the efficacy of these two distinct PAH extraction procedures. Linearity, limit of detection (LOD), limit of quantifcation (LOQ), accuracy, precision, and measurement uncertainty were assessed to validate both PAH extraction methods. Additionally, proficiency testing using Food Analysis Performance Assessment Scheme (FAPAS) was also conducted.

Calibration curves for eight targets of PAHs (PAH8) were obtained through fve replicate experiments with 6 data points spanning the range of 0 to 20 µg/kg range, each spiked with the 100 μ g/kg of ¹³C-labeled internal standard solution. Linearity, LOD, and LOQ were determined by plotting the ratios of analyte compound peak area to their corresponding internal standards against nominal concentrations (0, 0.5, 1, 2, 5, 10, and 20 µg/kg). The linearities of each calibration curve were assessed as a coefficient of determination (R^2) . The LOD and LOQ were calculated using the following formula: LOD=3.3σ/S and LOQ=10σ/S, where σ represents the standard deviation of the response, and S is the slope of the calibration curve. Accuracy and precision were assessed

at three diferent nominal concentration levels (5, 10, and 20 µg/kg), with intraday accuracy and precision determined from five replicates and interday accuracy and precision validated in triplicate across three days.

To ensure the reliability of our analysis data, the measurement uncertainty was calculated, following the EURA-CHEM/CITAC Guide (EURACHEM/CITAC, [2012](#page-12-5)). This involved evaluating the standard uncertainty of each factor afecting the measurement value, including balance, pipette, volume of mass fask, external standard solution, internal standard solution, calibration curve, matrix effects, and GC–MS. For determining the measurement uncertainty, 10 µg/kg of external standard solution and 100 µg/ kg of internal standard solution were used. After assessing each standard uncertainty, they were integrated to obtain a combined standard uncertainty (*u'*). An expanded measurement uncertainty (U') was calculated by multiplying *u'* by a coverage factor $(k=2)$, representing a confidence level of approximately 95%.

Our laboratory's performance for PAH quantitation was evaluated trough FAPAS proficiency testing, with results falling within the range of $|z| < 2$.

Exposure assessment and risk characterization

Various statistical treatments were employed to handle fndings below the LOD depending on the detection rate. In accordance with JOINT GUIDANCE (FAO, [2011](#page-12-6)), it is typically recommended for risk assessment to use both a lower bound (LB) and an upper bound (UB). Results falling below LOD are substituted for zero at the LB and replaced by LOD at the UB (EFSA, [2008;](#page-12-3) WHO/IPCS, [2009\)](#page-13-13).

To calculate total PAH8 concentration $(TC_{PAH8}$ or ∑PAH8), the concentrations of individual congener were combined by using Eq. [\(1\)](#page-4-0). Meanwhile, to determine the total B[a]P toxic equivalent quantity (TEQ $_{B[a]p}$) of PAH8, the concentrations of each PAH compound were multiplied by their toxic equivalency factors (TEFs) and then summed using Eq. [\(2](#page-4-1)). The TEFs used in this study are based on those reported by Nisbet and LaGoy ([1992\)](#page-13-14), which are presented in Table [1](#page-3-0).

$$
TCPAHSi = \sum_{i=1}^{n} Ci\left(\frac{ng}{g}\right)
$$
 (1)

$$
TEQB[a]Pi = \sum_{i=1}^{n} Ci \times TEFi\left(\frac{ng}{g}\right)
$$
 (2)

where, TC_{PAH8} is the total concentration of the ith individual congener of PAH8, TEQ_{B[a]Pi} is the total B[a]P toxic equivalent concentration of the ith individual congener of PAH8, Ci is the measured concentration for the ith individual

congener of PAH8, and TEF_i is the toxic equivalency factor of the ith individual congener.

The daily intakes of TC_{PAH8} and TEQ_{BIaIP} from marine food exposure were calculated using Eq. ([3\)](#page-4-2) and [\(4](#page-4-3)), respectively. These equations multiply the food intake rate (IR_i) by the TC_{PAH8} or $TEQ_{B[a]P}$ value and divide by the body weight (b.w.). The respective IR_i and BW of total population and consumption group were obtained from the KNHANES published by the KDCA.

$$
\text{Dietary exposure} = \sum_{i=1}^{n} \frac{\text{TCPAHSi} \times \text{IRi}}{b.w} \left(\frac{ng}{kg b.w. day}\right)
$$
\n
$$
\frac{n}{\text{TPOPf}_0 \cdot p} \times p; \quad (98)
$$

$$
\text{Dietary exposure} = \sum_{i=1}^{n} \frac{\text{TEQB[a]Pi} \times \text{IRi}}{b.w.} \left(\frac{ng}{kg b.w. \text{ day}}\right)
$$
\n
$$
\tag{4}
$$

To determine the daily exposure to PAH8 through marine food, the MOE was recommended by a Scientifc Committee (EFSA, [2005](#page-12-7)). The MOE was calculated based on the TEQ approach, where a Benchmark Dose Lower Limit $(BMDL_{10})$ of PAH8 was divided by the daily dietary exposure expressed in total concentration of PAH8 (Σ PAH8) or total B[a]P equivalent quantity (TEQ $_{B[a]P}$)) using the following Eq. [\(5](#page-4-4)). The BMD10 and BMDL10 values for PAH8 in the experimental animal diet ranged from 0.87 to 1.93 mg/kg b.w. per day and 0.49 to 1.35 mg/kg b.w./day, respectively. The CONTAM Panel used the lowest BMDL10 value of 0.49 mg/kg b.w. per day to derive an MOE, which measures how safe a chemical is at a given exposure level (EFSA, [2008](#page-12-3)). Thus, The $BMDL_{10}$ of 0.49 mg/kg BW/d was chosen as the reference point of PAH8 in this study.

$$
MOE = \frac{BMDL10\left(\frac{ng}{kg b.w. day}\right)}{Daily exposure\left(\frac{ng}{kg b.w. day}\right)}
$$
(5)

An MOE value lower than 10,000 is considered as a possible concern for risk management while an MOE of 10,000 or higher is regarded to indicate a low concern (EFSA, [2005,](#page-12-7) [2008](#page-12-3)).

Results and discussion

Method validation and quality control

The linearities (R^2) of calibration curves after alkali digestion and ultrasound-assisted extraction, respectively were above 0.9983 (Table [1](#page-3-0)) satisfying the Codex guideline requirement which specifies that \mathbb{R}^2 should be 0.99 or above (Codex, [1993\)](#page-12-8). While the LOD of the former pretreatment for PAH8 congeners ranged from 0.070 to

0.175 µg/kg, that of the latter method ranged from 0.074 to 0.115 µg/kg. The measurement uncertainty ranged from 4.022 to 8.608% and 4.463 to 8.529% in alkali digestion and ultrasound-assisted extraction methods, respectively. This result met the Codex criteria, which state that the expanded uncertainty should be less than 44% when the nominal concentration was 100 µg/kg or less (Codex, [2011](#page-12-4)). Additionally, standard uncertainty of calibration curve was found to be the most influential factor in determining the measurement uncertainty of PAH8. As shown in Table [2](#page-5-0), the accuracy ranged from 91.83 to 111.8% and the precision were 0.07 to 8.75%. Based on these validation results, both extraction procedures were shown to be highly effective in determining the content of PAH8 in marine products.

Concentration of PAH8 in marine products

According to the results of the determination PAH8 in marine products prepared as ready-to-eat, the number of samples and the number of detected samples in the eight categories of marine samples were as follows: Fish $(n=26)$ out of 139; the number of detected samples.

out of total samples), Shellfish ($n=32$ out of 46), Cephalopods ($n=11$ out of 25), Crustacea ($n=3$ out of 37), Sea algae (n=13 out of 30), Echinodermata (n=5 out of 5), Tunicata ($n=2$ out of 3), and Cnidaria ($n=1$ out 2) (Supplementary Table 2). PAH8 levels were found to be above the limits of over the LODs in 93 of 287 subsamples. Among the tested samples, katsuobushi (dried and smoked bonito) exhibited the highest B[a]P levels, followed by dried sea cucumber (Supplementary Table 2). Katsuobushi showed the concentrations of B[a]P (14.22 μg/kg), B[a]A (70.95 μg/

Table 2 Accuracy and precision of PAH8 using two diferent extraction methods

Compounds	Nominal con- centration (µg/ kg)	Alkali digestion method				Ultrasonication method			
		Intra-day ^a $(n=5)$		Inter-day ^b $(n=3)$		Intra-day $(n=5)$		Inter-day $(n=3)$	
		Accuracy	Precision	Accuracy	Precision	Accuracy	Precision	Accuracy	Precision
		$(\%)$	$(\%$ RSD ^c)	$(\%)$	$(\%$ RSD)	$(\%)$	$(% \mathbb{R}$ (% RSD)	$(\%)$	$(% \mathbb{R}$ (% RSD)
B[a]A	5	97.35-100.00	$1.92 - 3.42$	99.74	1.09	98.43-101.39	$0.53 - 2.27$	100.70	0.84
	10	98.80-101.02	$1.07 - 3.41$	99.79	0.62	99.54-103.56	$1.87 - 2.17$	100.45	0.80
	20	99.98-106.03	1.87-4.08	102.13	2.28	102.18-106.83	$1.57 - 2.43$	102.11	1.50
Chry	5	98.30-100.27	$0.76 - 0.99$	99.85	0.68	98.38-100.72	$1.05 - 2.26$	99.77	0.53
	10	96.75-99.73	$1.72 - 3.13$	99.52	0.81	98.29-100.83	$0.82 - 2.21$	99.93	0.26
	20	99.50-106.62	$0.93 - 7.42$	100.01	0.24	102.41-103.05	$0.80 - 3.71$	100.77	3.32
B[b]F	5	97.92-98.47	$1.20 - 2.59$	99.93	0.47	96.82-98.15	$0.55 - 1.20$	98.60	0.66
	10	97.23-97.76	$2.66 - 4.86$	99.58	0.42	97.78-98.06	$0.60 - 0.76$	98.65	0.10
	20	98.24-105.74	$1.12 - 8.35$	100.15	0.08	98.35-98.92	$0.39 - 0.57$	99.20	0.31
B[k]F	5	99.07-100.30	$0.87 - 2.49$	99.99	0.07	96.29-100.39	$1.32 - 4.22$	98.82	0.60
	10	98.49-99.64	$2.32 - 3.22$	100.24	0.13	89.93-96.94	$2.83 - 6.72$	97.81	2.43
	20	99.95-101.91	$0.24 - 1.25$	100.23	0.38	96.26-97.18	$1.12 - 2.60$	98.74	0.89
B[a]P	5	98.80-100.52	$1.03 - 1.87$	99.96	0.25	98.79-101.00	$1.20 - 3.76$	100.60	0.68
	10	98.31-100.29	$1.31 - 3.92$	100.48	0.91	96.86-98.77	$0.75 - 2.95$	99.60	0.64
	20	99.98-111.30	$0.96 - 8.75$	100.41	0.36	97.84-99.72	$0.65 - 1.27$	99.86	0.48
I[c,d]P	5	98.92-102.63	1.13-4.87	99.76	0.75	91.83-93.76	$0.64 - 1.77$	94.44	1.71
	10	98.25-101.87	1.94-4.27	100.27	0.38	94.07-96.76	$2.29 - 2.85$	98.19	1.47
	20	101.47-105.78	1.38-3.31	101.17	1.55	94.20-95.55	$1.97 - 2.44$	97.54	1.69
D[a,h]A	5	99.85-105.53	$0.78 - 7.00$	101.37	0.71	94.62-96.16	$1.53 - 7.36$	98.88	1.65
	10	100.32-107.50	$0.51 - 5.29$	100.92	0.63	98.53-104.07	1.98-3.86	99.82	0.89
	20	102.97-111.82	$2.88 - 6.44$	103.99	3.43	99.54-106.69	$1.04 - 7.22$	100.47	1.12
B[g,h,i]P	5	99.23-105.51	$1.22 - 7.10$	101.03	1.41	92.71-93.11	$0.61 - 1.36$	94.01	0.69
	10	104.06-109.97	$2.77 - 4.49$	102.13	1.93	95.60-96.21	$1.15 - 2.50$	97.70	1.10
	20	100.83-108.58	$1.67 - 5.75$	101.38	0.87	98.29-100.09	$1.18 - 1.85$	99.68	1.15

Ranged from mean of 5 determinations performed daily for 3 days

Mean of 3 determinations

Relative standard deviation: 100×standard deviation/mean

kg), Chry (90.63 μg/kg), B[b]F (24.62 μg/kg), B[k]F (8.482 μg/kg), I[c,d]P (4.924 μg/kg), D[a,h]A (1.277 μg/ kg), and $B[g,h,i]P(5.367 \mu g/kg)$ (data not shown).

However, both katsuobushi (smoked and dried bonito) and dried sea cucumber were found to contain levels of B[a] P, a known carcinogen (EFSA, [2008](#page-12-3)), that exceeded both EC [\(2015](#page-12-9)) and MFDS ([2019\)](#page-13-11) regulations. The EC regulation for B[a]P in smoked fish is 5 µg/kg, and the MFDS regulation is 5 µg/kg for smoked fsh and 10 µg/kg for smoked and dried fsh. Katsuobushi contained 14.22 µg/kg of B[a]P, and dried sea cucumber contained 11.35 µg/kg of B[a]P. Tsut-sumi et al. [\(2019](#page-13-15)) demonstrated that 21 µg/kg B[a]P in dried bonito fakes demonstrated 1.5 times higher than the concentration in the present study. PAHs can form in the smoke that is produced during cooking, and they can also be formed on the surface of the food itself (Alomirah et al., [2011](#page-12-2)). Another study by Kafouris et al. ([2020\)](#page-13-16) found that smoked fish contained significantly higher levels of PAHs compared to fresh fsh. The fndings of these studies suggest that open fame cooking methods can elevate the B[a]P content of the smoked and dried katsuobushi. The sea cucumber samples in this study are dried. Dried sea cucumber is the most popular form of sea cucumber, accounting for 80% of the market. To reduce processing time, several drying methods have been developed, including hot-air drying at temperatures of 60–100 °C and vacuum cooking at 95 °C (Fan et al., [2022](#page-12-10)). The thermal processing of dried sea cucumber samples in this study may have increased the levels of B[a]P in the samples. For non-processed products other than smoked fsh, the B[a]P level in the grilled salted mackerel was substantially higher (8.5 µg/kg). Open flame cooking methods, like grilling, are known to generate elevated levels of levels of PAHs in food, as PAHs are a class of organic compounds formed during the incomplete combustion of carbon-based materials such as wood, charcoal, and fat (Sampaio et al., [2017\)](#page-13-17). How PAHs form in food during open fame cooking is a complex process that depends on a number of factors, such as the type of fuel used, the cooking temperature, and the cooking time. For instance, a study by Alomirah et al. (2011) (2011) (2011) found that grilled meat contained signifcantly higher levels of PAHs than baked meat. Consumers should be aware of the potential health risks associated with the consumption of PAHs and should consider alternative cooking methods, such as smoking, baking or roasting, whenever possible.

However, the shellfsh category had the highest detection rate (70%) above LOD, while the fsh and crustacea categories had low detection rates (19% and 8%, respectively). The diferent detection rates between shellfsh and crustaceans were explained by their difering feeding habits. Shelfsh feeds on suspended nutritional components in water, whereas crustaceans are scavengers (Veyrand et al., [2013](#page-13-6)). PAHs were found in all mussel subsamples, and the toxicants are thought to have come from contaminated sea

water (EFSA, [2008\)](#page-12-3). Although shellfish were frequently tested for contamination, none of the subsamples exceeded the regulatory limits set by EC and MFDS. According to the regulations from MFDS ([2019\)](#page-13-11), the maximum permissible levels of B[a]P in bivalvia and cephalopoda are 10.0 µg/kg and 5.0 µg/kg, respectively. Overall, there was a variation in PAH levels depending on cooking methods, with the smoke produced during the heating process appearing to be the primary cause of excessive PAH levels.

PAH8 profling in marine products

The relative proportions of PAH8 in the medium-bound state within eight categories are described in Fig. [1.](#page-7-0) Chrysene was a primary component accounting for 33% of an amount of PAH8 in total marine products, followed by B[a]A at 25%. In PAH8, D[a,h]A has the lowest percentage at 3%, followed by I[c,d]P at 5%. These distributions showed the similar tendency in which EFSA ([2008\)](#page-12-3) reported that chrysene was a dominant element showing 33% followed by B[a]A at 20%, while D[a,h]A made up the lowest rate at 2% followed by B[k]F at 6% (EFSA, [2008](#page-12-3)). Chrysene showed the highest contribution in French TDS as well, followed by B[b]F (Veyrand et al., [2013\)](#page-13-6).

In Fig. [2,](#page-7-1) the ratios of $B[a]A/(B[a]A+Chry)$ and $I[c,d]$ $P/(I[c,d]P+B[g,h,i]P$ were displayed. These ratios were employed as a marker to determine the origin of PAHs (Yan et al., [2005](#page-13-18); Yunker et al., [2002\)](#page-13-19). Medium bound was applied to obtain the ratios in the present study. When PAH8 were found in fsh subsamples above LODs, they typically underwent heating processes like grilling or drying (Supplementary Table 2). Most of the dots in the plot's zone (A) are grilled fsh, whereas those on the plot's zone (B) are often dried fsh (Fig. [2](#page-7-1)). These patterns, specifcally, demonstrate that whereas dried fsh contains petrogenic PAHs from the environment, grilled fsh contains PAHs produced by burning. In general, it has been noted that grilled food is one of the major contributors to PAH consumption (EFSA, [2008](#page-12-3)).

In the shellfsh category, however, PAH8 concentrations were frequently above LODs in raw materials, and the variance of PAH8 levels depending on the heating methods was smaller than in the fsh category (Supplementary Table 2). According to zone (C) in the scatter plot, the PAH8 in the shellfish samples might be derived from petrogenic origin rather than from incomplete combustion (Fig. [2\)](#page-7-1). Fernando et al ([2019](#page-12-11)) reported that shellfsh products were contaminated from spilt crude oil in sea water (Fernando et al., [2019\)](#page-12-11). In this research, cephalopods had a slightly larger $B[a]A/(B[a]A+Chry)$ ratio (0.3) than the 0.2 from the Second French TDS (Veyrand et al., [2013\)](#page-13-6) based on mean distributions of B[a]A and Chry.

Relative proportion (%)

Fig. 1 Relative proportions of mean concentrations of 8 polycyclic aromatic hydrocarbons (PAH8) in 287 marine product subsamples under medium-bound state

Fig. 2 $B[a]A/(B[a]A+Chry)$ ratio plotted against the I[c,d] $P/(I[c,d]P+B[g,h,i]P$ ratio on a scatter plot for marine product subsamples. Reference ranges for diagnostic ratios were applied (Yan et al., [2005](#page-13-18)). B[a] A stands for Benz[a]pyrene, whereas Chry, Indeno[1,2,3-cd] pyrene, and B[g,h,i]P stand for Benzo[g,h,i]perylene

Dietary exposure and risk assessment

To calculate daily dietary exposure, the daily food intake rate was multiplied by the total PAH8 concentration $(\Sigma PAH8)$ value or the $TEQ_{B[a]P}$ value of PAH8 in the marine products using diferent cooking methods, and then divided the result by body weight. The outcomes are represented in Table [3](#page-8-0) with both LB and UB values. On average, South Koreans are exposed to 1.278 ng per kg of body weight per day (ng/ kg bw/d) of ∑PAH8 and 0.399 ng/kg bw/d of TEQB[a]P from total marine products under the LB scenario. Under the UB scenario, the average exposure is 2.164 ng/kg bw/d for ∑PAH8 and 1.347 ng/kg bw/d for TEQB[a]P. The 1–2 age group's mean dietary exposure to PAH8 from total marine products utilizing the UB scenario had the greatest mean daily exposure (3.212 ng/kg bw/d for ∑PAH8 and 2.071 ng/ kg bw/d for $TEQ_{B[a]P}$) due to their lowest body weights across all age groups.

In this study, dietary exposure to PAHs from fish is higher than from shellfsh, cephalopods and crustaceans. The average dietary intake for the fsh category was found to be 0.805 ng/kg bw/d for ∑PAH8 and 0.334 ng/kg bw/d

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Total population $(n=20,671)$

over 64 years $(n=4268)$

 $20-64$ years (n = 11,592)

 \mathbf{B}

 \mathbf{B}

 \mathbf{B}

 \mathbf{B}

 \mathbf{B}

 \mathbf{B}

407.306.792

6.469.460.509

809,287,807

9.865.394.107

336,404,931

न्त्र

Category

 $1-2$ years (n=519)

(Number of composite samples)

 \Box

Total marine products (109)

18,580,015,331

Category 1–2 years (n=519) 3–6 years (n=1062) 7–12 years (n=1601) 13–19 years (n=1629) 20–64 years (n=11,592) over 64 years (n=4268) Total population (n=20,671)

 $7-12$ years (n = 1601)

 $3-6$ years (n = 1062)

 $13-19$ years (n = 1629)

LB UB LB UB LB UB LB UB LB UB LB UB LB UB

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18,580,015,331 350,409,060 2,244,569,374 123,460,013 38,849,793,033 4,002,250,430 2,368,503,149,506 3,364,40,493,00, 386,300,000,000,000,000,000,000,000,009,400,509 407,306,792,009,400,400,400,400,400,509 407,306,792,000,0

25,388,266,009

4,002,250,430

123,460,013 38,849,793,033

350.409.060 2.244.569.374

2,131,642,668 5,863,149,596

for $TEQ_{B[a]P}$ in the LB scenario, while in the UB scenario, it was 1.152 ng/kg bw/d for ∑PAH8 and 0.735 ng/kg bw/d for TEQ_{BfaIP} for the total population, whereas the overall dietary exposure for the shellfish, cephalopods, and crustaceans was 0.184 ng/kg bw/d for ∑PAH8 and 0.013 ng/kg bw/d for TEQ $_{\text{BfaIP}}$ under the LB scenario, and 0.391 ng/kg bw/d for Σ PAH8 and 0.287 ng/kg bw/d for TEQ_{B[a]P} under the UB scenario for total population (Table [3\)](#page-8-0). However, it is important to note that shellfsh, cephalopods, and crusta ceans constituted the primary contributors to PAH exposure during the Second French TDS (Veyrand et al., [2013](#page-13-6)), which is believed to be due to varying consumption habits among country. In their investigation, the average dietary exposure from fsh was 0.03 ng/kg bw/d for adults and 0.07 ng/kg bw/d for children, compared to 0.193 ng/kg bw/d for adults and 0.098 ng/kg bw/d for children from mollusks and crus taceans. As seen in Table [3](#page-8-0), fsh products were the highest contributor to dietary exposure in this study, ranging from 63.0% for Σ PAH8 (83.7% for TEQ_{B[a]P}) in the total population to 88.8% for Σ PAH8 (97.3% for TEQ_{B[alP}) in the 3–6 year old population using the LB value, and from 53.2% for Σ PAH8 (54.6% for TEQ_{B[a]P}) in the total population to 65.6% for \sum PAH8 (55.9% for TEQ_{B[a]P}) in the 3–6 year old population using the UB value. Sea algae was the next major contributor, ranging from 15.4% for ∑PAH8 (19.8% for TEQ_{B[a]P}) in the total population to for 18.5% ∑PAH8 (23.7% for TEQ $_{\text{BfaIP}}$) in 3–6 years population under the UB scenario, while its contribution was smaller under the LB scenario. The high consumption level of sea algae appears to result from its signifcant contribution, despite its low contamination level. Particularly in the 20–64 age group under the LB scenario, Echinodermata, including dried sea cucumber, also made a considerable contribution of 24.3% for Σ PAH8 (14.2% for TEQ_{B[a]P}). The marine subsample in Supplementary Table 1 and 2 with the highest dietary expo sure level was dried sea cucumber, which the adult group consumed more than the other groups.

The MOE approach was used for the risk assessment. According to the fndings of this study, independent of age groups or marine products, all MOEs were above 10,000 (Table [4\)](#page-10-0). The calculated MOEs were about 1,055,476,358 for \sum PAH8 (6,469,460,509 for TEQ_{B[a]P}) at the LB scenario and 346,669,319 for ∑PAH8 (407,307,792) for $TEQ_{B[a]P}$) at the UB scenario for the total population. It's important to note that the MOEs for the calculated equiva lent B[a]P, as determined by TEQ (TEQ $_{B[a]P}$), in the case of the other 7 PAHs, ranged from 1.2 times higher in the UB scenario to 6.1 times higher in the LB scenario compared to those of \sum PAH8. All MOEs exceeded 1.0×10^4 , indicating that the risk assessment results indicate a low level of concern from a public perspective regarding health risks associated with dietary exposure to PAH8 from marine products. In the 1–2 age group under the UB

scenario, the risk of PAH8 from fsh diet had the lowest MOE value (257,240 for ∑PAH8 and 446,434 for $TEQ_{B[a]P}$, followed by the risk of PAH8 through sea algae (MOE value: 576,548 for ∑PAH8 and 724,207 for $TEQ_{B[a]P}$).

Individual MOE values for composite samples are further included in Supplementary Tables 1 and 2. Considering the total population based on KNHANES (MOHW, [2013](#page-13-10)), the MOE for all individual samples was above 10,000. When considering only those people in the consumption group who consumed the specifc food samples listed in the KNHANES under the UB scenario, dried sea cucumber had the lowest MOE value, with 6,201 for Σ PAH8 and 38,651 for $TEQ_{B[a]P}$. Salted mackerel had the second lowest MOE values, with 36,481 for Σ PAH8 and 87,244 for TEQ_{B[alP}. This is attributed to their elevated levels of PAH8 and substantial consumption rates. On the other hand, katsuobushi showed relatively high MOE value (758,956 for ∑PAH8 and 4,934,786 for TEQ $_{B[a]P}$), even though it contained high PAH8 level. Katsuobushi is typically used as a stock ingredient or spice in Korean cuisine, resulting in modest daily intakes. Another study also reported that Katsuobushi had comparably low daily exposure to PAHs because to its low consumption rate (Lee et al., [2018\)](#page-13-20).

In conclusion, this study found that fish is the primary source of PAH exposure in marine products, and that grilling, smoking, and drying processes increase PAH levels in food. The estimated risk of exposure to eight PAHs using TEFs may be an overestimation. It is advisable for many studies using TEFs to consider using the total concentration and the toxicological values for PAH mixtures. Following a risk assessment using a MOE approach in a TDS, PAH8 exposure from marine products is considered low for the South Korean population. The fndings of this study can serve as a foundation for developing strategies to reduce PAH levels in food and to identify the most efective ways for consuming marine products.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10068-023-01491-y>.

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Data availability The data that support the fndings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

Declarations

Conflict of interest On behalf of all authors, the corresponding author states that there is no confict of interest.

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