

1 Large scale serum proteomics identifies proteins associated with 2 performance decline and clinical milestones in Duchenne muscular dystrophy

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37 **Abstract**

38 Serum biomarkers are promising minimally invasive outcome measures in clinical studies in
39 Duchenne muscular dystrophy (DMD). However, biomarkers strongly associated with clinical
40 progression and predicting performance decline are lacking. In this study we aimed to identify serum
41 biomarkers associated with clinical performance and able to predict clinical milestones in DMD.
42 Towards this aim we present a retrospective multi-center cohort study including serum samples and
43 clinical data collected in research participants with DMD as part of a natural history study at the
44 University of Florida (UF) and real-world observations at Leiden University Medical Center (LUMC)
45 between 2009-2022. The 7K SomaScan[®] assay was used to analyse protein levels in individual
46 serum samples. Serum biomarkers predicted age at loss of ambulation (LoA), age at loss of overhead
47 reach (OHR) and age at loss of hand to mouth function (HTM). Secondary outcomes were the
48 association of biomarkers with age, corticosteroid (CS) usage, and clinical performance based on the
49 North Star Ambulatory Assessment (NSAA), 10 meter run velocity (10mrv), 6 minute walk (6MWT)
50 and Performance of the Upper Limb (PUL2.0). A total of 716 serum samples were collected in 79
51 participants at UF and 74 at LUMC (mean[SD] age; 10.9[3.2] vs 8.4[3.4]). 244 serum proteins showed
52 an association with CS usage in both cohorts independent of CS type and regimen, including MMP3
53 and IGLL1. 318 probes (corresponding to 294 proteins) showed significant associations with NSAA,
54 10mrv, 6MWT and/or PUL2.0 across both cohorts. The expression of 38 probes corresponding to 36
55 proteins such as RGMA, EHMT2, ART3, ANTXR2 and DLK1 was associated with risk of both lower and
56 upper limb clinical milestones in both the LUMC and UF cohort. In conclusion, multiple biomarkers
57 were associated with CS use, motor function and upper lower and upper limb disease milestones in
58 DMD. These biomarkers were validated across two independent cohorts, increasing their likelihood
59 of translation for use within the broader DMD population.

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71 Introduction

72 Duchenne muscular dystrophy (DMD) is the most common form of muscular dystrophy¹. It is caused
73 by variants in the *DMD* gene, resulting in an absence of functional dystrophin². Symptoms begin in
74 early childhood with delayed motor milestones and continue with progressive muscle weakness,
75 scoliosis, reduced pulmonary function, and dilated cardiomyopathy. Untreated, most patients will
76 lose ambulation by the age of 10 years^{3,4}.

77 Current treatment consists of long-term corticosteroid (CS), initiated at age 4-5 years and
78 recommended for life-long use⁵. Chronic use of CS has been shown to delay loss of ambulation,
79 onset and progression of scoliosis, and, in combination with non-invasive ventilation, it increases life
80 expectancy to 30-40 years of age^{3,4}. Yet CS are not without negative metabolic effects⁶. Despite the
81 huge progress in understanding the biology of the disease and the large investment in pre-clinical
82 research, there is currently no cure for DMD. Multiple investigational drugs have failed to show
83 significant improvement in participants' performance in clinical trials, with only a few drugs receiving
84 full and/or accelerated or conditional FDA approval such as microdystrophin gene therapy⁷, exon
85 skipping⁸, vamorolone⁹ and givinostat¹⁰. While failures may be related to limited drug potency, it has
86 also become clear that disease trajectories present high inter-individual variation, further reducing
87 the power to detect a significant and clinically meaningful treatment effect on a single outcome
88 measure in a typically short term, multi-center clinical trial. Several factors contribute to disease
89 heterogeneity, including mutation type¹¹⁻¹⁴, genetic modifiers¹⁵, and standards of care. Outcome
90 measures have been shown to be affected by patient motivation¹⁶. These factors complicate
91 accurate prediction of individual disease trajectories and thereby contribute to difficulty in clinical
92 trial design. Therefore, being able to objectively predict long-term clinical outcomes based on short
93 term evidence would greatly facilitate the evaluation of medicinal products.

94 Numerous serum biomarkers have been proposed for DMD¹⁷⁻¹⁹, however the focus has mostly been
95 in the identification of cross-sectional differences between unaffected controls and individuals with
96 DMD. Monitoring biomarkers, evaluated in series, are particularly lacking. Research efforts focused
97 on the analysis of muscle damage biomarkers²⁰⁻²², and in monitoring response to (micro-)dystrophin
98 restoration therapies^{23,24}. However, serum biomarkers with prognostic value are currently lacking.
99 The challenge and paradox relies on a disconnect between the early window of opportunity to treat
100 patients (roughly 3-8 years of age, in which most of the muscle mass is available for therapeutic
101 applications aimed at prevention vs reversal), the observable decline in patients function (typically
102 starting around 8-9 years of age), and the observation of disease milestones such as loss of
103 ambulation (between 10-16 years of age). Initial attempts to determine longitudinal trajectories in
104 serum biomarkers related to disease progression^{22,25,26} have been limited by the small number of
105 individuals, lack of follow-up samples, and incomplete collection of clinical data.

106 Therefore, discovery studies in large and well-characterized cohorts are needed to identify
107 associations between biomarker signatures and clinical performance and to define the context of
108 use for the biomarker signature. Importantly, prognostic biomarkers that can predict disease
109 milestones over longer time frames could enrich clinical trial design. Patients recruitment can be
110 tailored to individuals who are more likely to experience the milestone during the study. This is
111 particularly relevant as participants are usually enrolled in clinical trials before the age of 8, in early
112 stages of disease progression, but when little decline can be normally observed within the duration

113 of an interventional study of 12 months. Connecting short term changes in biomarker signatures to
114 the likelihood of long-term decline would help bridge this gap. Therefore, in this study we aimed to
115 identify serum biomarkers associated with clinical outcomes and their prognostic value for clinically
116 relevant milestones for both lower and upper limb function.

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119 **Methods**

120 Study Cohort, design and outcomes

121 This was a retrospective, multi-center, cohort study including serum samples and clinical data
122 collected from individuals with DMD participating in research protocols at the University of Florida
123 (UF) and at the Leiden University Medical Center (LUMC) between 2009-2022. We included 407
124 serum samples from 74 individuals with DMD aged 4 – 24 years at LUMC and 295 serum samples
125 from 79 individuals with DMD aged 5 – 22 years at UF. At LUMC, samples were collected during
126 yearly visits to the outpatient clinics as part of the standard of care. At UF, samples were collected as
127 part of an optional biosample addition to research visits conducted for the ImagingDMD natural
128 history study (NCT01484678). Written consent was obtained from all participants or their caregivers
129 as described in protocol B22.013 at LUMC and protocols IRB201500981 and IRB201700056 at UF,
130 which were approved by the respective regulatory boards at both sites.

131 Clinical data were obtained at the same clinic or research visit as serum sample collection. Data
132 included age at sample collection, CS use at the time of sample collection, and performance on tests
133 of function. CS information was categorized by use (treated or untreated), type (deflazacort,
134 prednisone, or other), regimen (daily or intermittent – defined as 10 days on/10 days off or weekend
135 dosing), and dose. Motor function tests included the North Star Ambulatory Assessment (NSAA), 10
136 meter run/walk velocity (10MRV), 6 minute walk test (6MWT), and Performance of the Upper Limb
137 (PUL2.0)^{27–30}. Three disease milestones were recorded: age at loss of ambulation (LoA), age at loss of
138 overhead reach (OHR), and age at loss of hand to mouth (HTM). LoA was defined similarly in both
139 cohorts with LoA based on patient reported inability to walk 5 meters unaided at home at LUMC and
140 inability to traverse 10 meters unaided within 45 seconds at UF. OHR and HTM were primarily
141 derived from PUL2.0 scores and occasionally from patient reported data.

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143 Samples collection and proteomic analysis

144 Serum samples were collected and prepared according to standard phlebotomy procedures.
145 Samples at LUMC were left to clot for ~30 minutes, followed by 10 min centrifugation at 2350g,
146 while clotting time was 30 minutes at UF, followed by centrifugation at 3000 rpm for 15 min. At the
147 LUMC, sample aliquots were frozen at -20°C for 1-2- months and then transferred to -80°C for long
148 term storage, while at UF, samples were immediately stored at -80°C. Serum samples were analysed
149 by the SomaScan® proteomic platform at SomaLogic (Boulder, Colorado, USA)³¹. SomaScan uses
150 aptamers as affinity reagents to detect and quantify proteins in complex mixtures such as serum.
151 Aptamers are nucleic acids that are able to bind specific protein targets with high specificity and
152 affinity. The pipeline is built to keep targets engaging aptamers, while washing away the unbound
153 ones. Detection of the aptamers is then performed using an array with probes complementary to the
154 aptamer sequence. At the time of this research, this platform included 7596 aptamers capable of
155 detecting 6628 proteins. As part of the assay, extensive quality control metrics are put in place
156 including internal and external controls. A total of 9 samples (2 LUMC / 7 UF) did not pass
157 SomaLogic's internal quality control and were excluded from further analysis, results, and data
158 tables.

193 **Results**

194 After retaining samples passing quality control, a total of 153 males with DMD and 693 serum
 195 samples were included in the study (Table 1). LUMC participants were significantly younger at the
 196 first sample visit compared to UF participants (mean[SD]; 8.4[3.4] vs 10.9[3.2], $p < 0.001$). An average
 197 of 4.3 samples per participant were analysed, with longer follow-up duration in the LUMC cohort
 198 (5.7[3.6] vs 3.4[2.5] years, $p < 0.001$). An intermittent (10 days on/10 days off of pred (or Deflazacort)
 199 CS regimen was most common at LUMC while daily dosing was most common at UF (prednisone and
 200 deflazacort); however, age at start of CS treatment was comparable across sites. Only a few patients
 201 remained CS naïve for the entire study period (4.1% vs 3.2%). The percentage of patients meeting
 202 one of the 3 clinical milestones at first sample was higher at LUMC compared to UF (LoA: 18.9% vs
 203 9.6%, loss of OHR: 16.0% vs 12.2%, loss of HTM: 5.3% vs 0.0%). For the LUMC cohort, 6660 probes
 204 passed quality control compared to 6690 for the UF cohort.

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	LUMC n = 74	UF n = 79
<i>Samples, n</i>	405	288
<i>Age at first sample, years, mean (SD)</i>	8.4 (3.4)	10.9 (3.2)
<i>Follow-up duration, years, mean (SD)</i>	5.7 (3.6)	3.4 (2.5)
<i>Samples per patient, n, mean (SD)</i>	5.5 (2.9)	3.6 (2.0)
<i>Age at start of corticosteroids, years, mean (SD)</i>	5.6 (1.6)	5.4 (1.7)
<i>Corticosteroid treatment at first sample, n (%)</i>		
• Not recorded	1 (1.4%)	3 (3.8%)
• Daily	2 (2.7%)	66 (83.5%)
• Intermittent	49 (66.2%)	4 (5.1%)
• None	22 (29.7%)	6 (7.6%)
<i>Dystrophin variants, n (%)</i>		
• Exonic deletion	49 (66.2%)	49 (62.0%)
• Exonic duplication	10 (13.5%)	8 (10.1%)
• Nonsense variant	13 (17.6%)	12 (15.2%)
• Splice site variant	2 (2.7%)	3 (3.8%)
• Other small point variants	—	5 (6.3%)
• Variant unknown	—	2 (2.5%)
<i>Weight at first sample, kg, mean (SD)</i>	32.6 (18.6)	35.1 (12.2)
<i>Height at first sample, cm, mean (SD)</i>	127.2 (19.2)	126.9 (10.7)
<i>BMI at first sample, mean (SD)</i>	18.3 (4.4)	21.2 (5.4)
<i>Ambulant patients at first sample, n (%)</i>	59 (79.7%)	68/78 (87.2%) (1 unknown)
<i>Able to perform overhead reach at first sample, n (%)</i>	64 (86.5%)	63/70 (90.0%) (9 unknown)
<i>Able to perform hand to mouth at first sample, n (%)</i>	73 (98.6%)	74/75 (98.7%) (4 unknown)

206 *Table 1. Cohort characteristics.*

207 Serum protein associations with age and corticosteroid usage

208 Given the progressive nature of DMD and the availability of longitudinal visits, we first identified
 209 proteins associated with age. 4,796 probes (4436 proteins) were significantly associated with age in
 210 the LUMC cohort and 2,668 probes (2498 proteins) were significantly associated with age in the UF
 211 cohort (FDR<0.05)(Fig. 1A-B); with overlap of 2,317 probes (2186 proteins) between both cohorts
 212 (Fig. 1C). 2,251 probes showed concordant directional change between cohorts, with the
 213 coefficients generally being higher in the LUMC cohort (Fig. 1D). Multiple muscle proteins were

214 negatively associated with age including, but not limited to, CK-MM, MYOM3, MYOM2, TRIM72,
215 MYL3, BIN1, TTN, LAMA2, TPM3, ACTN2, DES, CA3, MYL3 and TNNI2 (Fig1E-H). Markers of
216 fibroadipogenic progenitors (PDGFRA), fibrosis and extracellular matrix (collagens such as COL1A1,
217 COL2A1, COL3A1, COL6A1, COL6A2, COL6A3, TIMP1), several BMPs (BMP-1,4,5,6,7) and protein
218 synthesis pathways (AKT1, AKT2, IGF2), also decreased with increasing age.

219 Among the proteins with expression that significantly increased with age, we identified proteins
220 present in the central nervous system such as CNTN3 and CNDP1, proteins expressed in adipose
221 tissue (LEP, GHR, FABP4, ADIPOQ, CNTFR, CFD and TIMP4, PNLIPRP1, PNLIPRP2, PNLIP), and several
222 cytokines (CCL14, IFNA6, BMPR2, CXCL8, CCL3L1, IL26, IL5RA, CCL1, CCL16, CCL15, IL10RA, IL31RA,
223 IL17RE) (Fig1I-L). Two different probes for IGF1 had had opposing associations with age, and several
224 IGF binding proteins showed discordant trajectories, with IGFBP1 and IGFBP2 decreasing with age
225 and IGFBP5 and IGFBP6 increasing with age. Coefficients of all significant, concordant proteins
226 associated with age are provided in Supplementary Table 1.

227 Given that the majority of participants had chronic exposure to CS , we sought to identify a protein
228 expression signature related to exposure to CS while correcting for age. 846 probes (790 proteins) in
229 the LUMC cohort and 396 probes (364 proteins) in the UF cohort were significantly associated with
230 CS use (FDR < 0.05)(Fig. 2A-B). Furthermore, 244 probes (227 proteins) were shared across sites (Fig.
231 2C). The number of proteins associated with steroid use in the UF cohort was smaller due to the
232 small number of patients untreated (n=5) in this cohort compared to the untreated group in the
233 LUMC cohort (n=22), suggesting that the number of proteins identified as significantly associated
234 with the treatment is not a measure of the treatment effect. This is confirmed by the comparison of
235 the coefficients shown in Fig.2D, showing larger effects for the UF cohort in line with the higher
236 dosage (mostly daily) compared to the LUMC cohort (mostly intermittent). Among the proteins we
237 identified (CD23) and (MMP3)⁶ previously observed in DMD, as well as newly identified proteins
238 such as IGLL1 immunoglobulin lambda-like polypeptide 1 (IGLL1) and repulsive guidance molecule A
239 (RGMA)(Fig. 2E-G). To identify whether the identified associations were potentially related to
240 efficacy or safety effects, we assessed the directionality of the signature for age and steroids.
241 Proteins showing discordant association (such as declining with age as disease progresses and
242 increased by CS treatment) were considered to relate to the steroid efficacy signature, while
243 concordant effects were considered to belong to the safety signature (Fig.2I). A total of 44 proteins
244 showed discordant age and CS effects (potential efficacy signature) across the 2 cohorts
245 (Supplementary table 2); this list included previously identified proteins such as ANGPT2 (2
246 independent aptamers), proteins related to lipoproteins transportation (APOA4, APOC3, APOE) as
247 well as newly identified recognized by 2 independent aptamers such as DLD, ART3, NDUFA2, OLFM2
248 as well as 2 members of the RGM family (RGMA and RGMC)(Fig.2H). A total of 172 proteins showed
249 concordant sign for age and CS effects in both LUMC and UF cohorts (potential safety biomarkers;
250 Supplementary Table 3). This list included previously identified MMP3, Afamin, IGFBP-5, proteins
251 involved in SMAD signalling as well as new proteins belonging with IGF binding properties and
252 metalloproteinases.

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254 Serum protein associations with motor performance

255 Longitudinal trajectories for the NSAA, 10MRW, 6MWT, and PUL 2.0 differed between the two sites,
256 with a shift towards earlier decline in motor performance in the LUMC cohort (Fig 3A-D). To assess
257 which proteins were associated with clinical severity, we evaluated associations with each functional
258 test in each cohort. 2,005 probes (1882 proteins) showed a significant association with at least one
259 motor function test in the LUMC cohort compared to 483 probes (454 proteins) in the UF cohort
260 (FDR < 0.05). 318 probes (294 proteins) were shared across the two sites, with the majority of
261 significant associations found for lower limb scales, consistent with the higher number of
262 observations and the larger degree of functional decline experienced in the cohorts. Cross-
263 correlation of the coefficients showed concordant directionality between the 2 cohorts, especially
264 for the lower limb scales (Fig. 3E).

265 We then assessed how generalizable models trained in each cohort were by using the other cohort
266 for validation, selecting the 5% most performant proteins for each scale. Validation reconstruction
267 accuracies were higher for lower limb function with Q^2 values comparable across both cohorts for
268 the 10MRV, while accuracies were higher for models trained on the LUMC cohort for the 6MWT and
269 higher for UF for the NSAA. 122 probes (116 proteins) in the LUMC cohort and 109 probes (105
270 proteins) in the UF cohort significantly and accurately predicted all 3 scales used to monitor lower
271 limb function (Fig. 3F). Supplementary Table 4 provides a list of proteins with significant associations
272 across scales and cohorts. ART3, RGMA and DLD showed highly significant associations and accurate
273 predictions across multiple motor function tests (Fig. 3G-H). To refine the efficacy and safety
274 signature identified in the steroid analysis we assessed whether cross-correlation between steroid
275 and clinical association coefficients; we identified 919 pairs where a protein was significantly
276 associated with a clinical score and with steroid treatment across both cohorts. Concordant
277 coefficients are in this case expected to be considered efficacy biomarkers, while discordant
278 coefficients are expected to be adverse effects (Fig. 3I). A chi-squared test showed that concordant
279 signs were significantly enriched in the previously defined efficacy signature, while discordant signs
280 were enriched in the safety signature ($P < 10^{-15}$), with only 835 of the 919 pairs being properly
281 assigned.

282 Prediction of clinical milestones

283 Given the association with continuous clinical scores, we sought to determine whether the identified
284 proteins had prognostic value for clinically meaningful disease milestones such as age at LoA, loss of
285 OHR, and loss of HTM. All clinical milestones were reached at significantly earlier ages and in a
286 greater percentage of patients in the LUMC cohort than in the UF cohort (Table 1, Fig. 4A-B). We
287 identified 41 probes / milestone associations. Among these, 3 of the probes targeting RGMA, ENPP5
288 and RGS21 were associated with 2 different milestones (FDR < 0.05). RGMA was associated with
289 lower and upper limb milestones across the 2 cohorts, , while RGS21 (UF) and ENPP5 (LUMC) were
290 cohort-specific. Associations for DLK1 and ART3 were confirmed by 2 independent probes: DLK1 was
291 found to be associated with OHR in the UF cohort and ART3 with LOA in the LUMC cohort (Fig.4C).
292 The direction of log Hazard Ratios (logHR) were conserved for these proteins across probes and
293 outcomes. Notably, RGMA, ANTXR2, EIF4G1, CAMK2A had large negative logHRs and PTPRD, NCAM
294 had large positive logHRs. Kaplan-Meier plots for RGMA are presented across all 3 milestones for
295 both LUMC and UF cohorts (Fig 4D). Supplementary Table 5 lists all proteins significantly predicting
296 lower and upper limb milestones. Since treatment with steroids is known to delay these milestones,
297 we assessed whether logHR and steroid effects show discordant effects. Figure 4E illustrates how

298 estimates are indeed anti-correlated for all but 2 proteins, supporting the use of these biomarkers to
299 evaluate the risk of meeting these milestones.

300 Among the multiple proteins associated with functional tests and disease milestones, we shortlisted
301 a few based on the consistency and strength of the associations across cohorts (Table 2). Proteins
302 ANTXR2, ART3, EHMT2 and RGMA showed a % risk increase for disease milestones between 136%
303 and 981% for a decrease of one unit (in log scale). Treatment with steroids corresponded to an
304 increase in the biomarker level corresponding to a reduced the risk % up to 90% with highest
305 normalization of risk for the upper limb HTM milestone; normalization for the risk % of LoA was in all
306 cases estimated between 22% and 39%. We further estimated how monitoring proteins in blood can
307 help evaluate the yearly increase in risk % for the milestones and observed yearly risk increase
308 between 43% and 215%.

Gene Symbol	Probe	Site	Milestone	Log HR	β CS	β age	Risk % increase with 1 log decrease	Risk % reduction with CS treat	Yearly risk % increase
ANTXR2	15559-5	LUMC	LoA	-1.14	0.43	-0.76	213.54	38.50	138.38
ART3	7970-315	LUMC	LoA	-0.89	0.47	-0.41	144.55	34.49	43.83
ART3	10970-3	LUMC	LoA	-0.86	0.30	-0.48	135.79	22.67	51.19
EHMT2	5843-60	LUMC	LoA	-1.02	0.47	-0.60	176.06	38.11	83.08
RGMA	5483-1	UF	HTM	-2.38	0.98	-0.48	981.79	90.38	215.03
RGMA	5483-1	LUMC	LoA	-1.00	0.51	-0.64	170.79	39.78	89.96

309

310 *Table 2. Shortlisted proteins with consistent association across performance tests, milestones and*
311 *cohorts. The coefficient estimates for the indicated milestone are obtained from the time to event*
312 *analysis, while the coefficient for CS and age were obtained by the linear mixed effect models*
313 *including both CS and age. We further show the risk % for the indicate milestone for a unit decrease*
314 *in the protein (in log scale) and how the risk is reduced by treatment with CS and increased per year.*

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316

317 Discussion

318 Large-scale serum biomarker discovery is now possible in DMD with the availability of high-
319 throughput proteomics platforms. In this retrospective study, we had the unique opportunity to pair
320 robust and comprehensive clinical data from two large, independent international cohorts of
321 individuals with DMD to serum levels of 6,628 proteins. The availability of extensive longitudinal
322 serum samples and clinical data not only allowed for analysis of protein signatures associated with
323 age and corticosteroid usage, but also allowed for identification of novel proteins predictive of
324 clinical function and clinical milestones. Proteins like RGMA were predictive of both lower and upper
325 limb clinical milestones such as LoA and loss of HTM. Furthermore, several proteins were associated
326 with increased risk of single clinical milestones such as ART3 for LoA. The ability to compare these
327 results across the two independent cohorts enabled for cross-validation of our findings.

328 Prior serum biomarker discovery in dystrophinopathies has largely been limited to smaller cohorts,
329 targeted protein selection for analysis, and limited longitudinal data. These studies have been crucial
330 for building our understanding of the disease's pathogenic processes. Key differences between
331 individuals with and without DMD have been reported, including several proteins associated with
332 age, shedding light on the ongoing pathogenic processes. It is well-known with DMD that serum CK
333 (activity and protein levels) decrease with age, reflecting muscle damage and loss of muscle mass.
334 Additional proteins decreasing with age have subsequently been identified including MDH2, MYL3,
335 CA3, MYOM3, COL1A1, TTN, TNNT2, ETFA, TNNT3, and MAP4^{26,35}. Proteins that have increasing
336 expression with age have included C4A, C4BPA, and GSN, among others³⁵. A small number of studies
337 have also utilized the SomaScan[®] platform to perform large-scale protein biomarker analysis in
338 DMD, identifying additional proteins associated with age/time such as LEP, CFTNR, FABP3, and
339 TNNI2^{25,36}. Complementing and largely validating the findings of these studies, we identified
340 previously described proteins such as CK and MYOM-3 showing steep decline in protein levels, but
341 also substitution of muscle with fibro-adipogenic tissue with markers such as Collagen 1 and 2
342 declining along with markers of fibro-adipogenic progenitors such as PDGFRA³⁷. We also observed
343 adipogenic markers such as LEP, GHR and ADIPOQ with increased expression along with a
344 complement and inflammation signature.

345 The CS signature encompassed steroid engagement biomarkers such as MMP3 and IGLL1, which
346 showed the strongest association with CS usage, both in the daily and the intermittent regimen.
347 Previous research has also identified elevated MMP3 in patients with DMD treated with CS, while
348 IGLL1 has not yet been described in relation to DMD³⁸. However, it has very recently been shown
349 that weekend CS use in adults with limb girdle muscular dystrophy reduces IGLL1 in the presence of
350 MMP3 elevation³⁹. Given the role of IGLL1 in B cells, the reduction observed in combination with the
351 CS treatment could signal the immunosuppression expected in these patients due to the drug.

352 Treatment with corticosteroids contributed to normalize as well as exacerbate the disease
353 progression signature. An exacerbation of the signature related to age was observed for previously
354 reported safety biomarkers such as MMP3, Afamin, and IGFBP5⁵, and for certain apolipoproteins
355 such as APOA2, APOL1, APOA5. A normalization was observed for other apolipoproteins such as
356 APOE4, APOC3 and APOE, suggesting that steroid treatment could directly normalize the
357 dyslipidemia with effects on APOE4, APOC3, and APOE, while exacerbation of the APOA2, APOL1,
358 and APOA5 signature could highlight how steroids affect lipid metabolism and potentially

359 cardiovascular health. The compensatory effect of APOE is also underscored by the more severe
360 phenotype observed in *mdx* ApoE double knock-out mice^{40,41}. We also found that other previously
361 reported proteins such as ANGPT2⁶, and proteins (RGMA, DLK1, ANTXR2, and ART3) that were
362 decreased with disease progression in both UF and Leiden cohorts were higher in the CS groups
363 compared to untreated.

364 A major strength of this study was the ability to identify serum biomarkers predictive of clinical
365 function. RGMA and ART3 were all directly related to muscle performance as measured by both
366 upper (PUL2.0) and lower limb (NSAA, 6MWT, 10MRV) functional outcomes. RGMA, ANTXR2,
367 EHMT2 and ART3 were found to have large negative logHR when considering the loss of certain
368 clinical milestones. To further illustrate this, we stratified the population according to RGMA levels
369 and found that the decreasing levels of RGMA clearly shifted the Kaplan-Meier curve towards earlier
370 LoA and OHR.

371 RGMA is part of the repulsive guidance molecule family of glycoprotein-1 (GP1) anchor proteins that
372 is mostly expressed in the central nervous system and muscle tissue according to the human protein
373 atlas gene expression data. Originally described as playing an important role in neurogenesis by
374 guiding axonal growth and as an important target of neuronal survival, it has now also been
375 identified to play a role in myogenesis^{42,43}. RGMA has been proposed to play a central role in
376 regulating cellular hypertrophy and hyperplasia⁴⁴. RGMA has previously been found to have an
377 association with Spinal and bulbar muscular atrophy (SBMA)⁴⁵, Parkinson disease, Alzheimer disease,
378 multiple sclerosis, and cerebrovascular accidents, as well as an association with upper limb function
379 measured by elbow flexion in DMD^{25,42}.

380 ANTXR2, also known as capillary morphogenesis protein 2 (CMG2) plays an important role in cellular
381 interaction by binding collagen IV and laminin, suggesting involvement in extracellular matrix
382 adhesion. It is expressed in multiple tissues, including muscles. Loss of function variants in ANTXR2
383 cause hyaline fibromatosis syndrome and ANTXR2 knockout mice show collagen VI accumulation in
384 the uterus⁴⁶. This could implicate ANTXR2 as playing a role as a collagen VI regulator, and possible
385 involvement in muscle homeostasis.

386 ART3, known as ADP-Ribosyltransferase 3, is mainly expressed in skeletal muscle tissue according to
387 GTEx and human protein atlas databases. Gene expression data obtained from human individuals as
388 well as in Chinese Meishan pigs showed that ART3 is mostly expressed in muscles enriched in fast
389 twitch fibers^{47,48}, which are typically more prone to damage in DMD. ART3 has been reported to be
390 decreased in the serum across multiple dystrophies and myopathies³⁹ and in SBMA⁴⁵. It has been
391 recently reported in SBMA only 3 proteins, including RGMA, MSTN, and ART3 were found to have
392 decreased abundances in plasma and were significantly correlated with higher thigh MRI muscle fat
393 fraction akin to what is observed in DMD⁴⁵. Additional support for the relationship to fat deposition
394 has been obtained in Wannanhua pigs show that ART3 is involved in fat deposition in muscle⁴⁹,
395 further strengthening the biological rationale behind the association identified in this study. This is
396 the first study to show ART3 relationship to CS status and clinical function in DMD.

397 Finally, EHMT2, known as euchromatic histone lysine methyltransferase 2, and DLK1 known as Delta-
398 like 1 homolog as well as Pref-1 (preadipocyte factor 1) showed interesting association. They both
399 declined with age and were associated with disease milestone; EHMT2 was normalized by steroid
400 treatment. EHMT2 was previously associated with renal fibrosis⁵⁰, atrial fibrosis⁵¹, cardiomyocytes

401 hypertrophy⁵² and high fat diet induced obesity and hepatic insulin resistance⁵³, which align with the
402 pathogenic processes ongoing in DMD. The reduction of EHMT2 with age could potentially describe
403 the reduced magnitude of pathological processes as muscle mass is progressively lost. DLK1 plays a
404 multifaceted role in muscle development and regeneration. DLK1 is a transmembrane protein that
405 functions as a regulator of cell growth during development. In adults, its expression is low and
406 restricted to endocrine tissues. It is involved in the differentiation of multiple cell types, including
407 adipocytes and plays an important role in skeletal muscle biology during fetal development and
408 postnatal growth⁵⁴. While the role of DLK1 in adult skeletal muscle regeneration is less clear,
409 upregulated expression of DLK1 has previously been observed in DMD and Becker muscular
410 dystrophy⁵⁵ and reduction of DLK1 in fibroadipogenic progenitors corresponded to increased
411 adipogenic commitment⁵⁶. The reduction of the DLK1 levels may be associated with an increased
412 adipogenic signature in DMD.

413 One limitation of this study was the differences in cohort characteristics due to the participant pool
414 and standards of care. The LUMC samples consisted of participants seen in the clinic, were generally
415 younger in age, and were primarily undergoing a 10 days on, 10 days off CS regimen. No data were
416 available on whether a patient was in the “on” or “off” phase of treatment at the time of sample
417 collection. In contrast, the UF cohort samples were from participants in a natural history research
418 study, were generally older in age, and were primarily on a daily CS regimen. The differences in
419 steroid regimens may have led to smaller effect sizes associated with CS use in the LUMC cohort as
420 well as the earlier occurrence of clinical milestones. The smaller proportion of individuals reaching
421 milestones in the UF cohort may explain the smaller number of probes associated with clinical
422 outcomes and milestones in that cohort. As different regimes coincided with cohort effects, we were
423 unable to study the impact of intermittent CS compared to daily CS. These differences could
424 potentially be studied in prospective research such as in the FOR-DMD clinical trial. Lastly, due to the
425 retrospective nature of this study, we had a number of missing data points. Despite these
426 limitations, due to our relatively large patient population and number of samples, we were able to
427 identify multiple significant serum biomarkers which were validated across both cohorts.

428 In conclusion, we found proteins associated with upper and/or lower limb outcomes in individuals
429 with DMD across two independent cohorts. Significant consideration should be given to the use of
430 RGMA, DLK1, ANTXR2, EHMT2 and ART3 as potential biomarkers based on the strength of their
431 associations, significance of the findings across scales and milestones and biological plausibility in
432 connection to the disease. A panel that accurately and precisely detects these proteins in serum
433 could enable the connection of short-term changes to disease stabilization and a decreased risk of
434 decline in the mid- to long-term. Furthermore, measuring these proteins in a clinical setting would
435 help predict individual disease trajectories, assess treatment options before milestones, and inform
436 patient inclusion criteria in clinical trials. These findings are of importance for future clinical trial
437 design, paving the way for innovative outcome measures such as serum biomarkers.

438

439

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449

450 **Potential Conflicts of Interest**

451 SWME, SVH, and KCHH are employees of BioSymetrics, which has a commercial interest in the
452 results. The remaining authors have no competing interests.

453

454 **References**

- 455 1. Mercuri, E. & Muntoni, F. Muscular dystrophies. *Lancet (London, England)* **381**, 845–60
456 (2013).
- 457 2. Duan, D., Goemans, N., Takeda, S., Mercuri, E. & Aartsma-Rus, A. Duchenne muscular
458 dystrophy. *Nature Reviews Disease Primers* vol. 7 1–19 (2021).
- 459 3. Bello, L. *et al.* Prednisone/prednisolone and deflazacort regimens in the CINRG Duchenne
460 Natural History Study. *Neurology* **85**, 1048–1055 (2015).
- 461 4. McDonald, C. M. *et al.* Long-term effects of glucocorticoids on function, quality of life, and
462 survival in patients with Duchenne muscular dystrophy: a prospective cohort study. *Lancet*
463 *(London, England)* **391**, 451–461 (2018).
- 464 5. Birnkrant, D. J. *et al.* Review Diagnosis and management of Duchenne muscular dystrophy,
465 part 1: diagnosis, and neuromuscular, rehabilitation, endocrine, and gastrointestinal and
466 nutritional management. *Lancet Neurol.* **17**, 251–267 (2018).
- 467 6. Conklin, L. S. *et al.* Serum biomarkers of glucocorticoid response and safety in anti-neutrophil
468 cytoplasmic antibody-associated vasculitis and juvenile dermatomyositis. *Steroids* **140**, 159–
469 166 (2018).
- 470 7. Hoy, S. M. Delandistrogene Moxeparvec: First Approval. *Drugs* **83**, 1323–1329 (2023).
- 471 8. Aartsma-Rus, A. The Future of Exon Skipping for Duchenne Muscular Dystrophy. *Hum. Gene*
472 *Ther.* **34**, 372–378 (2023).
- 473 9. Keam, S. J. Vamorolone: First Approval. *Drugs* **84**, 111–117 (2024).
- 474 10. FDA Approves Nonsteroidal Treatment for Duchenne Muscular Dystrophy | FDA.
475 [https://www.fda.gov/news-events/press-announcements/fda-approves-nonsteroidal-](https://www.fda.gov/news-events/press-announcements/fda-approves-nonsteroidal-treatment-duchenne-muscular-dystrophy)
476 [treatment-duchenne-muscular-dystrophy.](https://www.fda.gov/news-events/press-announcements/fda-approves-nonsteroidal-treatment-duchenne-muscular-dystrophy)
- 477 11. Aartsma-Rus, A., Ginjaar, I. B. & Bushby, K. The importance of genetic diagnosis for Duchenne
478 muscular dystrophy. *J. Med. Genet.* **53**, 145–151 (2016).
- 479 12. Wang, R. T. *et al.* DMD genotype correlations from the Duchenne Registry: Endogenous exon
480 skipping is a factor in prolonged ambulation for individuals with a defined mutation subtype.
481 *Hum. Mutat.* **39**, 1193–1202 (2018).
- 482 13. Bello, L. *et al.* DMD genotypes and loss of ambulation in the CINRG Duchenne Natural History
483 Study. *Neurology* **87**, 401–409 (2016).
- 484 14. Waldrop, M. A. *et al.* Intron mutations and early transcription termination in Duchenne and
485 Becker muscular dystrophy. *Hum. Mutat.* **43**, 511–528 (2022).
- 486 15. Bello, L. & Pegoraro, E. The “Usual Suspects”: Genes for Inflammation, Fibrosis, Regeneration,
487 and Muscle Strength Modify Duchenne Muscular Dystrophy. *J. Clin. Med.* **8**, 649 (2019).
- 488 16. Alfano, L. *et al.* Role of motivation on performance of the 6-minute walk test in boys with
489 Duchenne muscular dystrophy. *Dev. Med. Child Neurol.* **57**, 57–58 (2015).
- 490 17. Al-Khalili Szigyarto, C. & Spitali, P. Biomarkers of Duchenne muscular dystrophy: current
491 findings. *Degener. Neurol. Neuromuscul. Dis.* **Volume 8**, 1–13 (2018).

- 492 18. Molinaro, M., Torrente, Y., Villa, C. & Farini, A. Advancing Biomarker Discovery and
493 Therapeutic Targets in Duchenne Muscular Dystrophy: A Comprehensive Review. *Int. J. Mol.*
494 *Sci.* **25**, (2024).
- 495 19. Hathout, Y. *et al.* Disease-specific and glucocorticoid-responsive serum biomarkers for
496 Duchenne Muscular Dystrophy. *Sci. Rep.* **9**, 12167 (2019).
- 497 20. Wagner, K. R. *et al.* Safety and disease monitoring biomarkers in Duchenne muscular
498 dystrophy: Results from a Phase II trial. *Biomark. Med.* **15**, 1389–1396 (2021).
- 499 21. Zygmunt, A. M. *et al.* A longitudinal study of creatine kinase and creatinine levels in
500 Duchenne muscular dystrophy. *Muscle Nerve* **67**, 138–145 (2023).
- 501 22. Awano, H. *et al.* Longitudinal data of serum creatine kinase levels and motor, pulmonary, and
502 cardiac functions in 337 patients with Duchenne muscular dystrophy. *Muscle Nerve* **69**, 604–
503 612 (2024).
- 504 23. Boehler, J. F. *et al.* Clinical potential of microdystrophin as a surrogate endpoint.
505 *Neuromuscul. Disord.* **33**, 40–49 (2023).
- 506 24. Chamberlain, J. S. *et al.* Microdystrophin Expression as a Surrogate Endpoint for Duchenne
507 Muscular Dystrophy Clinical Trials. *Hum. Gene Ther.* **34**, 404 (2023).
- 508 25. Spitali, P. *et al.* Tracking disease progression non-invasively in Duchenne and Becker muscular
509 dystrophies. *J. Cachexia. Sarcopenia Muscle* **9**, 715–726 (2018).
- 510 26. Strandberg, K. *et al.* Blood-derived biomarkers correlate with clinical progression in Duchenne
511 muscular dystrophy. *J. Neuromuscul. Dis.* **7**, 231–246 (2020).
- 512 27. Scott, E. *et al.* Development of a functional assessment scale for ambulatory boys with
513 Duchenne muscular dystrophy. *Physiother. Res. Int.* **17**, 101–109 (2012).
- 514 28. Arora, H. *et al.* Longitudinal timed function tests in Duchenne muscular dystrophy:
515 ImagingDMD cohort natural history. *Muscle Nerve* **58**, 631–638 (2018).
- 516 29. McDonald, C. M. *et al.* The 6-minute walk test and other clinical endpoints in duchenne
517 muscular dystrophy: Reliability, concurrent validity, and minimal clinically important
518 differences from a multicenter study. *Muscle Nerve* **48**, 357–368 (2013).
- 519 30. Pane, M. *et al.* Upper limb function in Duchenne muscular dystrophy: 24 month longitudinal
520 data. *PLoS One* **13**, (2018).
- 521 31. Candia, J., Daya, G. N., Tanaka, T., Ferrucci, L. & Walker, K. A. Assessment of variability in the
522 plasma 7k SomaScan proteomics assay. *Sci. Rep.* **12**, (2022).
- 523 32. Jolly, E. Pymer4: Connecting R and Python for Linear Mixed Modeling. *J. Open Source Softw.*
524 **3**, 862 (2018).
- 525 33. Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting Linear Mixed-Effects Models Using
526 lme4. *J. Stat. Softw.* **67**, (2015).
- 527 34. Davidson-Pilon, C. lifelines: survival analysis in Python. *J. Open Source Softw.* **4**, 1317 (2019).
- 528 35. Signorelli, M. *et al.* Longitudinal serum biomarker screening identifies malate dehydrogenase
529 2 as candidate prognostic biomarker for Duchenne muscular dystrophy. *J. Cachexia.*
530 *Sarcopenia Muscle* **11**, 505–517 (2020).

- 531 36. Hathout, Y. *et al.* Large-scale serum protein biomarker discovery in Duchenne muscular
532 dystrophy. *Proc. Natl. Acad. Sci.* **112**, 7153–7158 (2015).
- 533 37. Alonso-Jiménez, A. *et al.* Platelet Derived Growth Factor-AA Correlates With Muscle Function
534 Tests and Quantitative Muscle Magnetic Resonance in Dystrophinopathies. *Front. Neurol.* **12**,
535 659922 (2021).
- 536 38. Hathout, Y. *et al.* Serum pharmacodynamic biomarkers for chronic corticosteroid treatment
537 of children. *Sci. Rep.* **6**, 31727 (2016).
- 538 39. Willis, A. B. *et al.* Serum protein and imaging biomarkers after intermittent steroid treatment
539 in muscular dystrophy. 1–24 (2024) doi:10.1101/2024.06.14.24308858.
- 540 40. Milad, N. *et al.* Increased plasma lipid levels exacerbate muscle pathology in the mdx mouse
541 model of Duchenne muscular dystrophy. *Skelet. Muscle* **7**, 19 (2017).
- 542 41. White, Z. *et al.* Cholesterol absorption blocker ezetimibe prevents muscle wasting in severe
543 dysferlin-deficient and mdx mice. *J. Cachexia. Sarcopenia Muscle* **13**, 544–560 (2022).
- 544 42. do Carmo Costa, A. *et al.* RGMa can induce skeletal muscle cell hyperplasia via association
545 with neogenin signalling pathway. *Vitr. Cell. Dev. Biol. - Anim.* **57**, 415–427 (2021).
- 546 43. Copola, A. G. L. *et al.* Transcriptomic characterization of the molecular mechanisms induced
547 by RGMa during skeletal muscle nuclei accretion and hypertrophy. *BMC Genomics* **23**, (2022).
- 548 44. Neto, J. X. *et al.* Repulsive Guidance Molecules a, b and c Are Skeletal Muscle Proteins, and
549 Repulsive Guidance Molecule a Promotes Cellular Hypertrophy and Is Necessary for Myotube
550 Fusion Aline Fagundes Martins¹. *Cells Tissues Organs* **200**, 326–338 (2015).
- 551 45. Tebbenkamp, A. T. N. *et al.* Protein biomarker signature in patients with spinal and bulbar
552 muscular atrophy. *JCI Insight* **9**, (2024).
- 553 46. Bürgi, J. *et al.* CMG2/ANTXR2 regulates extracellular collagen VI which accumulates in hyaline
554 fibromatosis syndrome. *Nat. Commun.* 2017 81 **8**, 1–10 (2017).
- 555 47. Abbassi-Daloui, T. *et al.* A transcriptome atlas of leg muscles from healthy human volunteers
556 reveals molecular and cellular signatures associated with muscle location. *Elife* **12**, 1–29
557 (2023).
- 558 48. Li, Y., Xu, Z., Li, H., Xiong, Y. & Zuo, B. Differential transcriptional analysis between red and
559 white skeletal muscle of Chinese Meishan pigs. *Int. J. Biol. Sci.* **6**, 350–360 (2010).
- 560 49. Li, X. *et al.* Integrated 4D Analysis of Intramuscular Fat Deposition: Quantitative Proteomic
561 and Transcriptomic Studies in Wannanhua Pig Longissimus Dorsi Muscle. *Animals* **14**, 167
562 (2024).
- 563 50. Irifuku, T. *et al.* Inhibition of H3K9 histone methyltransferase G9a attenuates renal fibrosis
564 and retains klotho expression. *Kidney Int.* **89**, 147–157 (2016).
- 565 51. Xiao, Z. *et al.* MicroRNA-205-5p plays a suppressive role in the high-fat diet-induced atrial
566 fibrosis through regulation of the EHMT2/IGFBP3 axis. *Genes Nutr.* **17**, 11 (2022).
- 567 52. Papait, R. *et al.* Histone Methyltransferase G9a Is Required for Cardiomyocyte Homeostasis
568 and Hypertrophy. *Circulation* **136**, 1233–1246 (2017).
- 569 53. Xue, W. *et al.* Histone methyltransferase G9a modulates hepatic insulin signaling via

- 570 regulating HMGA1. *Biochim. Biophys. Acta - Mol. Basis Dis.* **1864**, 338–346 (2018).
- 571 54. Zhang, L. *et al.* Expression and Functional Analyses of Dlk1 in Muscle Stem Cells and
572 Mesenchymal Progenitors during Muscle Regeneration. *Int. J. Mol. Sci.* **20**, (2019).
- 573 55. Andersen, D. C. *et al.* Characterization of DLK1+ cells emerging during skeletal muscle
574 remodeling in response to myositis, myopathies, and acute injury. *Stem Cells* **27**, 898–908
575 (2009).
- 576 56. Garcia, S. M. *et al.* Distinct human stem cell subpopulations drive adipogenesis and fibrosis in
577 musculoskeletal injury. *bioRxiv Prepr. Serv. Biol.* (2024) doi:10.1101/2023.07.28.551038.
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581 **Figure legends**

582 **Figure 1.** Associations of Serum Proteins with Age in Individuals with DMD. Volcano plots
583 demonstrate the proteins from DMD serum that are associated with age in A) the LUMC cohort, and
584 B) the UF cohort. C) Venn diagram showing the number of proteins associated with age in common
585 between the LUMC and UF cohorts, with D) a plot of coefficients, indicating the direction of protein
586 association with age, in the LUMC versus UF cohorts. Plots E-H highlight the longitudinal trajectories
587 of protein levels over time in select proteins that decrease with increasing age such as CKM,
588 MYOM3, TTN and TNNI2, while plots I-L show trajectories of select proteins that increase with
589 increasing age such as CNTN3, CNDP1, LEP and TIMP4. Each trajectory plot includes the gene symbol
590 and aptamer identification number. DMD, Duchenne muscular dystrophy; LUMC, Leiden University
591 Medical Center; UF, University of Florida.

592 **Figure 2.** Associations of Serum Proteins with Corticosteroid use in Individuals with DMD. Volcano
593 plots demonstrate the proteins from DMD serum that are associated with corticosteroid use in A)
594 the LUMC cohort, and B) the UF cohort. C) Venn diagram showing the number of proteins associated
595 with corticosteroid use in common between the LUMC and UF cohorts, with D) a plot of coefficients,
596 indicating the direction of protein association with corticosteroid use, in the LUMC versus UF
597 cohorts. Plots E-H show differences in protein levels between serum from CS treated versus
598 untreated individuals with DMD in two previously identified proteins and two proteins identified in
599 this study. I) Scatterplot showing the relationship between age and steroid treatment coefficients for
600 proteins significantly associated with CS treatment. Proteins showing discordant coefficients for age
601 and CS treatments were considered as efficacy biomarkers, while with discordant coefficients were
602 considered as safety biomarkers (grey shaded). Orange dots represent estimates for the LUMC
603 cohort, while blue dots represent estimates for the UF cohort. CS, corticosteroid; DMD, Duchenne
604 muscular dystrophy; LUMC, Leiden University Medical Center; UF, University of Florida.

605 **Figure 3.** Associations of Serum Proteins with Performance on Tests of Motor Function in Individuals
606 with DMD. A-D) Longitudinal trajectory plots of NSAA results, 10MRV, 6MWD, and PUL 2.0 score
607 from both cohorts. E) Plots of coefficients, indicating the direction of protein association with motor
608 function test performance in the LUMC versus UF cohorts. Highlighted data points represent
609 significant associations. The number of significant associations is mentioned for each panel. F) UpSet
610 plot demonstrating the intersection of significantly associated probes with each of the four tests of
611 motor function. G-H) Trajectory plots showing the relationship between ART3 and either NSAA (G) or
612 6MWT (H) over the age in the LUMC cohort. I) Scatterplot showing the relationship between the
613 coefficient estimates for proteins significantly associated with steroid treatment and performance
614 test. Proteins showing concordant coefficients for performance tests and CS treatments were
615 considered as efficacy biomarkers (e.g. a protein associated with both high functional scores and
616 further elevated by steroid treatment), while with concordant coefficients were considered as safety
617 biomarkers (grey shaded). Orange dots represent estimates for the LUMC cohort, while blue dots
618 represent estimates for the UF cohort. Shapes indicate the association for each performance test.
619 6MWD, 6-minute walk distance; 10MRV, 10-meter run velocity; CS, corticosteroid; DMD, Duchenne
620 muscular dystrophy; LUMC, Leiden University Medical Center; NSAA, North Star Ambulatory
621 Assessment; PUL, Performance of the Upper Limb; UF, University of Florida.

622

623 **Figure 4.** Associations of Serum Proteins with Clinical Milestones. A) Kaplan Meier curves showing
624 the probability of LoA, loss of OHR, and loss of HTM by age in each cohort. B) UpSet plot of the
625 intersection of significant probes with each clinical milestone. C) Log hazard ratios of the protein
626 probes significantly associated with at least 1 clinical milestone. D) Kaplan Meier curves for loss of
627 each clinical milestone for RGMA by years since first sample collection. E) Correlations between
628 protein probe log hazard ratios and use of corticosteroids. HTM, hand-to-mouth; LoA, loss of
629 ambulation; OHR, overhead reach.

630

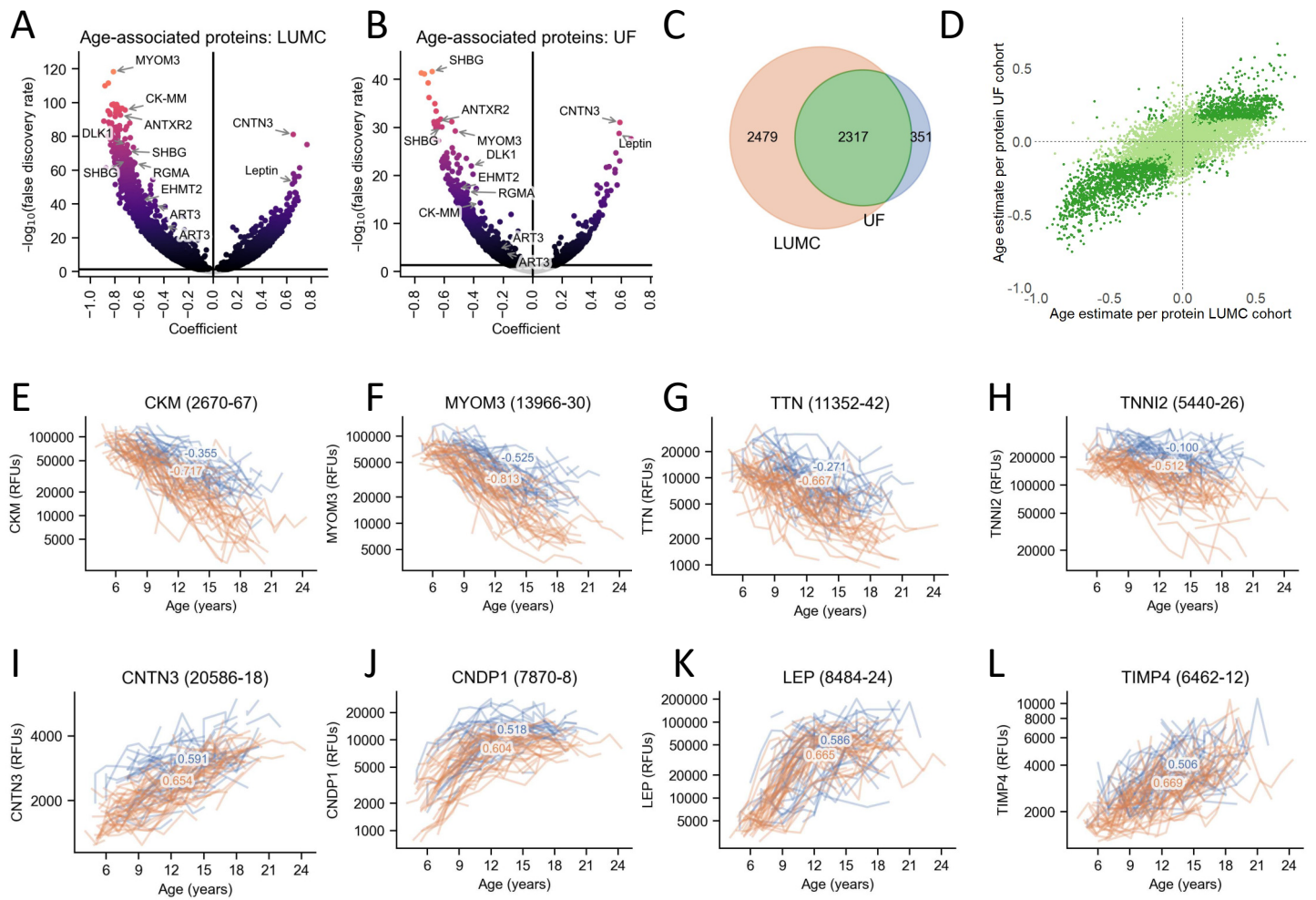


Figure 1

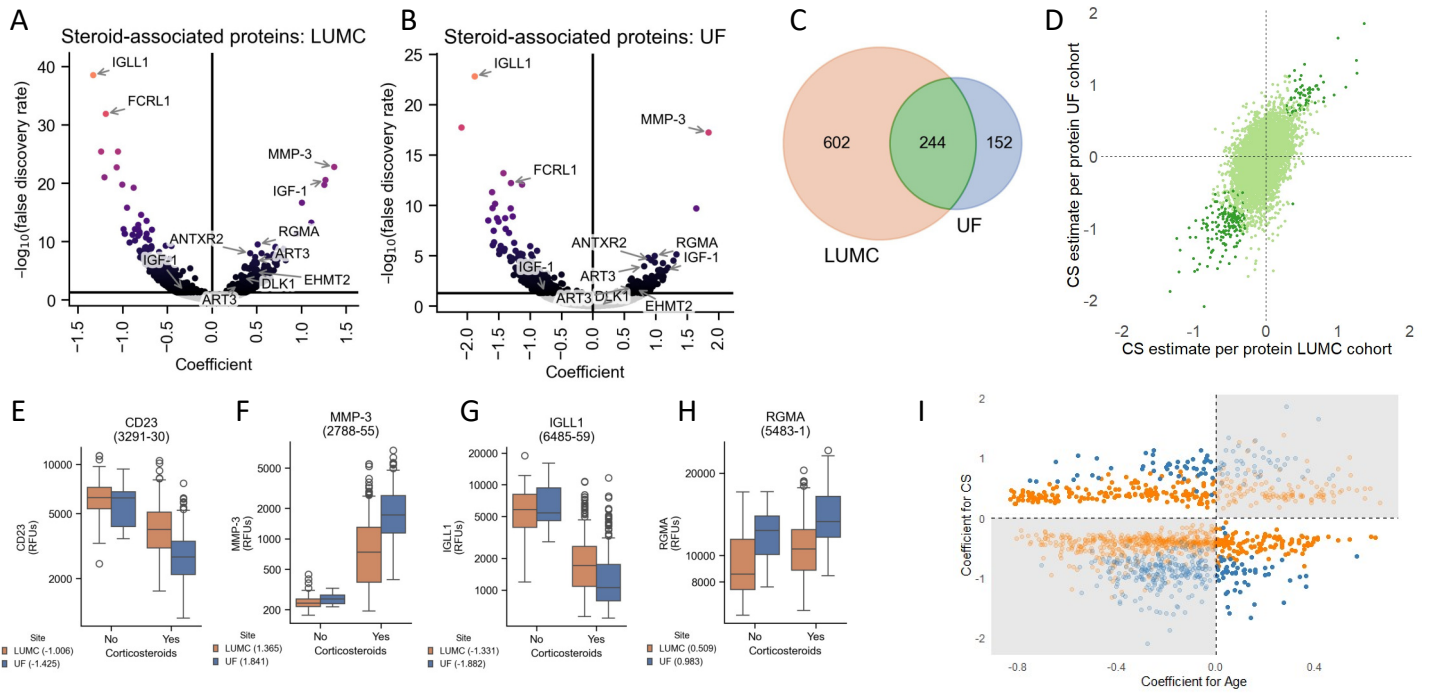


Figure 2

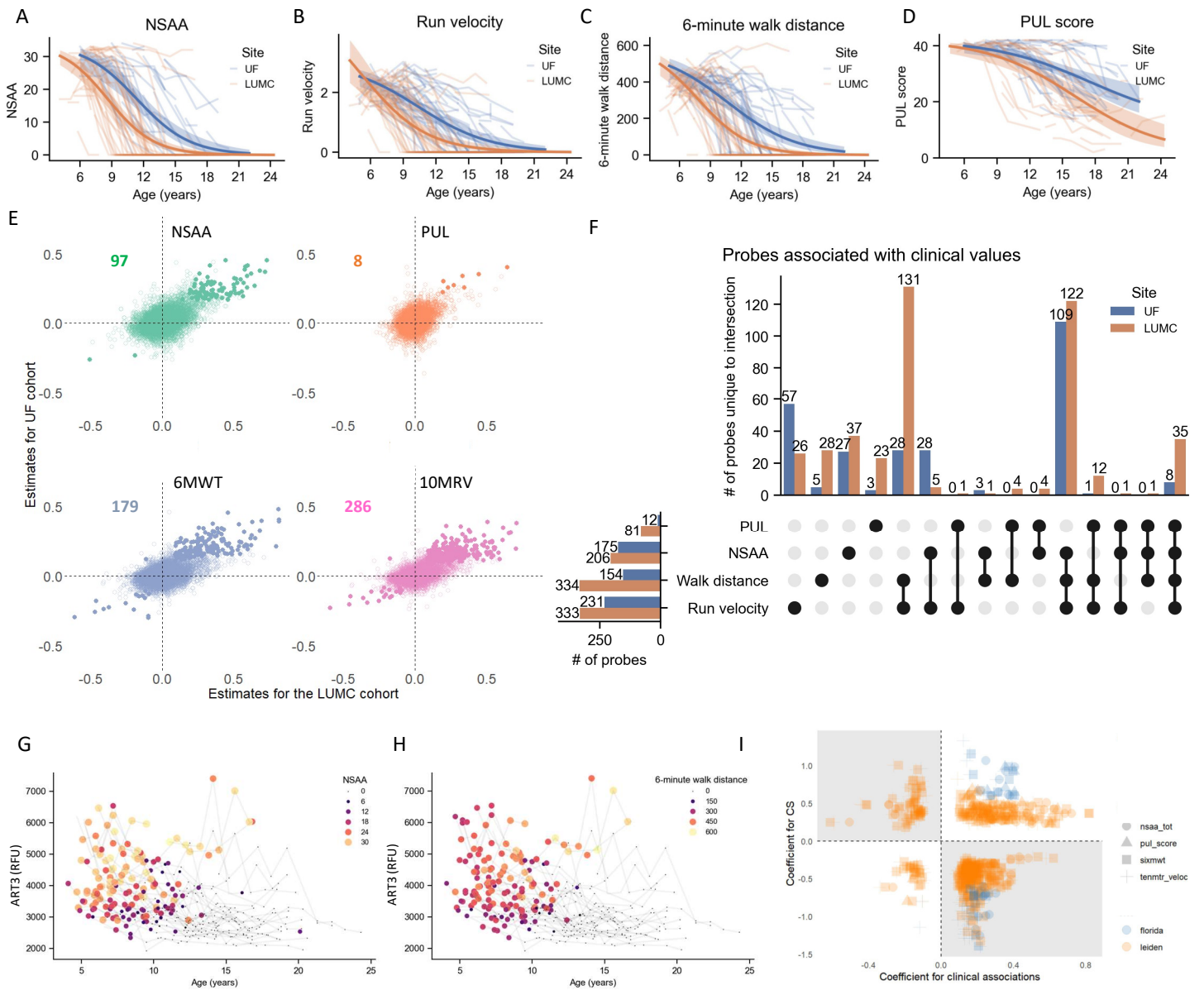


Figure 3

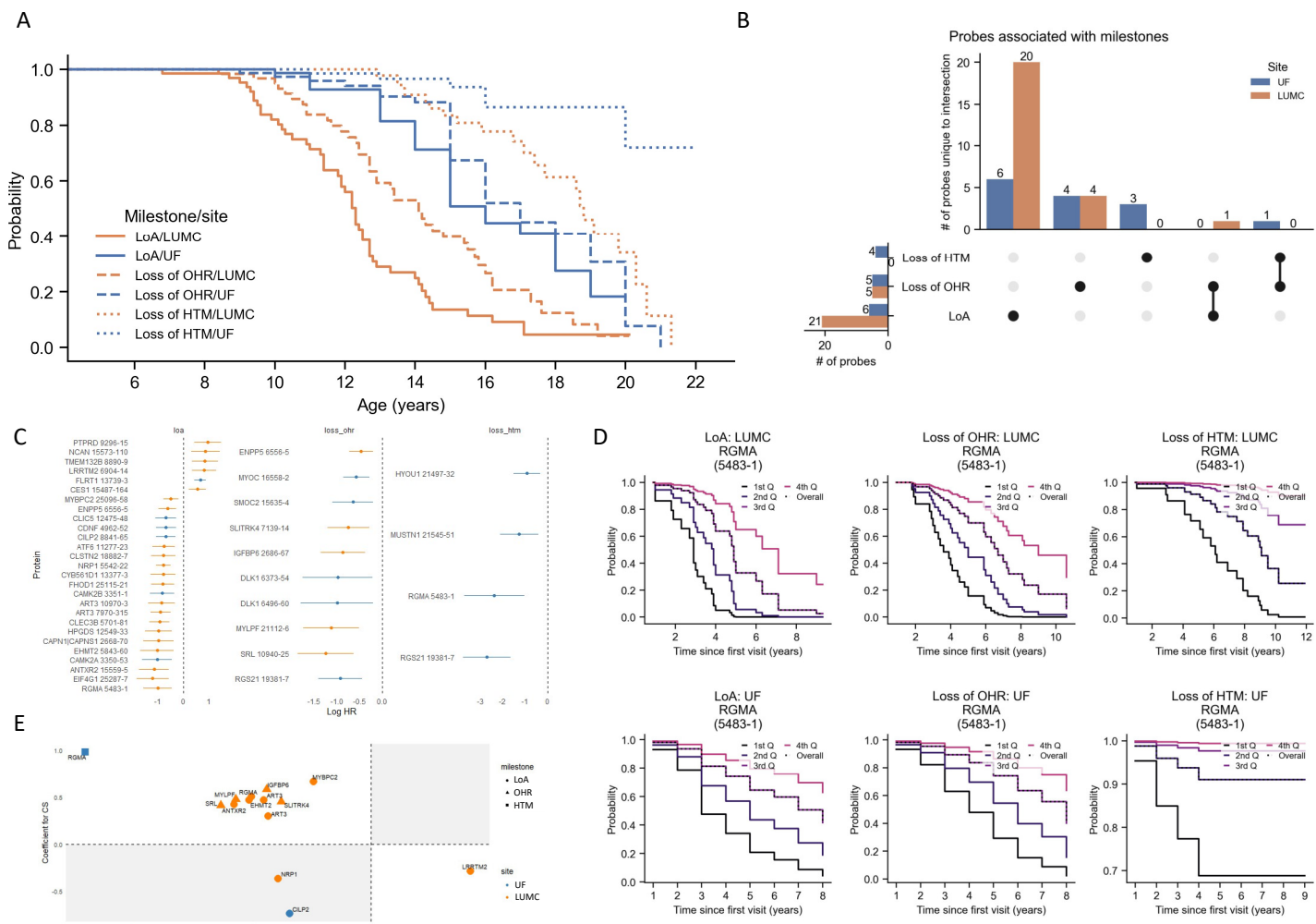


Figure 4