

Modified Delphi panel consensus recommendations for management of severe aplastic anemia

Daria V. Babushok,¹ Amy E. DeZern,² Carlos M. de Castro,³ Zora R. Rogers,⁴ David Beenhouwer,⁵ Michael S. Broder,⁵ Suzanne R. Fanning,⁶ Sarah N. Gibbs,⁵ Rabi Hanna,⁷ Jaroslaw P. Maciejewski,⁸ Bart L. Scott,⁹ Srinivas K. Tantravahi,¹⁰ Marcin W. Wlodarski,¹¹ Irina Yermilov,⁵ and Bhumika J. Patel⁶

¹Division of Hematology-Oncology, Department of Medicine, University of Pennsylvania, Philadelphia, PA; ²Sidney Kimmel Comprehensive Cancer Center at Johns Hopkins, Baltimore, MD; ³Department of Medicine, Division of Hematologic Malignancies and Cellular Therapy, Duke Cancer Institute, Durham, NC; ⁴Division of Pediatric Hematology-Oncology, University of Texas, Southwestern Medical Center, Dallas, TX; ⁵Partnership for Health Analytic Research, Beverly Hills, CA; ⁶Prisma Health Cancer Institute, University of South Carolina, Greenville, SC; ⁷Department of Pediatric Hematology Oncology and Bone Marrow Transplantation, Cleveland Clinic, Cleveland, OH; ⁸Translational Hematology and Oncology Research Department, Cleveland Clinic, Cleveland, OH; ⁹Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA; ¹⁰Division of Hematology and Hematologic Malignancies, Huntsman Cancer Institute, University of Utah, Salt Lake City, UT; and ¹¹Department of Hematology, St. Jude Children's Research Hospital, Memphis, TN

Key Points

- There is a need for nuanced consensus guidance on how to initiate and modify treatment for SAA based on individual patient characteristics.
- Expert panel provided recommendations for >600 varying scenarios of patients with SAA for initial and subsequent management of SAA.

Severe aplastic anemia (SAA) is a rare hematologic condition for which there is no clear management algorithm. A panel of 11 experts on adult and pediatric aplastic anemia was assembled and, using the RAND/University of California, Los Angeles modified Delphi panel method, evaluated >600 varying patient care scenarios to develop clinical recommendations for the initial and subsequent management of patients of all ages with SAA. Here, we present the panel's recommendations to rule out inherited bone marrow failure syndromes, on supportive care before and during first-line therapy, and on first-line (initial management) and second-line (subsequent management) therapy of acquired SAA, focusing on when transplant vs medical therapy is most appropriate. These recommendations represent the consensus of 11 experts informed by published literature and experience. They are intended only as general guidance for experienced clinicians who treat patients with SAA and are in no way intended to supersede individual physician and patient decision making. Current and future research should validate this consensus using clinical data. Once validated, we hope these expert panel recommendations will improve outcomes for patients with SAA.

Introduction

Acquired aplastic anemia (AA) is an immune-mediated bone marrow (BM) failure disorder, characterized by cytopenias and a hypocellular BM.¹⁻³ Although the precise etiology of autoimmunity in AA remains poorly understood, the last few decades have brought much progress to understanding dysregulated immune responses and autoimmune attack in AA.⁴⁻¹⁷ The estimated annual incidence of AA is ~2 cases per million globally.^{1,18-22} Although toxic exposures (eg, benzene), medications (eg, chloramphenicol and antiepileptics), and infections have been associated with some cases of acquired AA, in most cases an inciting event cannot be identified.¹ AA is most often diagnosed in middle to late childhood or late in life.¹

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A copy of the rating form may be found in the data supplement available with the online version of this article.

The full-text version of this article contains a data supplement.

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Acquired AA remains a diagnosis of exclusion, as there are other acquired or inherited bone marrow failure (IBMF) syndromes that may present similarly; a comprehensive diagnostic evaluation, including clinical and family history, physical examination, and laboratory testing with genetic diagnostics can help to establish the diagnosis.²³ Historically, AA has been classified using the modified Camitta criteria into nonsevere/moderate AA, severe AA (SAA), and very severe AA. SAA is defined by marrow cellularity <25% (or 25%-50% with <30% residual hematopoietic cells), plus at least 2 of the following peripheral blood findings: neutrophils of $<0.5 \times 10^9/L$, platelets of $<20 \times 10^9/L$, and reticulocytes of $<60 \times 10^9/L$.^{24,25} Very SAA fulfills the same criteria as SAA, except neutrophils are $<0.2 \times 10^9/L$.^{26,27} Without effective treatment, patients with SAA are at high risk of death from infection or hemorrhage.

Patients with SAA are typically treated with either allogeneic hematopoietic stem cell transplantation (allo-HSCT) or immunosuppressive therapy (IST) alongside a thrombopoietin receptor agonist (TPO-RA) eltrombopag. Allo-HSCT is generally recommended as first-line treatment for younger patients with a matched related donor (MRD), whereas IST is the recommended first line for older adults and those without a fully matched donor.²⁸ However, improving treatment outcomes with the use of unrelated and alternative donors suggest that earlier incorporation of allo-HSCT into the SAA treatment algorithm may be feasible.²⁹⁻⁴⁰

The recommended front-line IST consists of horse antithymocyte globulin (ATG) and cyclosporine (CsA), which, when used together with eltrombopag, results in an overall initial response rate of ~80%.⁴¹⁻⁴⁴ Among those who do not respond to front-line IST, allo-HSCT or a second course of IST with or without TPO-RAs can be considered. Eltrombopag was first approved in 2014 for the treatment of relapsed/refractory SAA and subsequently approved for use along with IST for treatment-naïve SAA in 2018.⁴²⁻⁴⁶ Other TPO-RAs are also in development, and emerging data demonstrate that these also have activity in SAA.⁴⁷⁻⁴⁹

Although previous SAA treatment recommendations exist,^{50,51} nuanced guidance on how to initiate and modify treatment for SAA based on individual patient characteristics (eg, medical fitness, age, and type of available donor) or disease characteristics (eg, response to prior treatment) is limited. Based on this need, we conducted a RAND/University of California, Los Angeles (UCLA) modified Delphi panel to develop clinical consensus on how to treat patients with SAA.⁵²⁻⁵⁷ Although the Grades of Recommendation, Assessment, Development, and Evaluation (GRADE) criteria are the gold standard criteria used to develop evidence-based guidelines,⁵⁸ the RAND/UCLA modified Delphi panel method⁵⁹ is a validated process for obtaining consensus in clinical management in areas in which optimal management is unclear because of limited, emerging, or changing clinical data, through the use of iterative structured surveys and group discussion of clinical experts.^{54,56,57} Using this process, we have assembled an 11-expert panel and analyzed 697 individual patient care scenarios across a spectrum of patient ages, physical fitness, initial vs subsequent therapy, and available donor type. Here, we present recommendations as a reference for use by the hematology community along with guidance summarizing consensus and various areas in which expert consensus could not be reached.

Materials and methods

The RAND/UCLA modified Delphi panel method⁵⁹ is a formal group consensus process that systematically and quantitatively combines expert opinion and evidence by asking physicians to rate, discuss, and then re-rate items. In brief, the steps include selecting a panel of physicians, summarizing the current evidence on the panel topic, and developing a survey collaboratively with panelists. Panelists then independently completed a first-round (premeeting) survey before attending a moderated meeting, during which areas of disagreement on the survey are discussed. Only aggregate results are presented. Panelists then repeat the survey, and these second-round ratings are used to develop clinical consensus recommendations. This process is consistent with elicitation methods recently recommended as best practice for healthcare decision making and for conducting health technology assessments.⁵²

For this report, a diverse 11-member physician panel, comprising 8 adult and 3 pediatric hematologists, was convened. The panel was double blinded; while work was ongoing, the sponsor did not know the identity of the physicians (except for the panel chair), and the physicians were not informed of the sponsor until after consensus recommendations were developed. The sponsor did not provide input on the methodology or results of the panel. The study did not involve human patients as defined by 45 Code of Federal Regulations part 46 and was therefore not subject to institutional review board approval.

Expert panelists were provided with a summary of current evidence on the therapeutic management of patients with SAA before the panel meeting: the etiological differences between inherited and acquired AA, diagnostic evaluation of patients with suspected SAA, and treatment trajectory of patients upon diagnosis. In the evidence review, we included 5 reviews,^{2,50,51,60,61} 1 prospective cohort study,⁶² 7 prospective randomized control trials,^{24,27,44,63-67} and 4 clinical trials served as significant resources for the literature review.^{42,45,46,68} Consensus recommendations in this manuscript were also informed by additional studies.^{41-44,69-74}

In collaboration with the expert panelists, a survey (rating form) comprising 697 patient scenarios that varied by factors identified as affecting therapy choice (eg, age, medical fitness defined by Eastern Cooperative Oncology Group [ECOG] criteria, and donor availability) was developed. A copy of the rating form is in the supplemental Appendix. For each scenario, experts rated the appropriateness of interventions for SAA on a 1-to-9 scale (9 being highly appropriate, and 1 being highly inappropriate), including treatments in first- and later-line settings. Panelists also rated the appropriateness of diagnostic testing to rule in SAA or rule out IBMF syndromes as well as rated various supportive care measures (including transfusion support and prophylactic antibiotics) through a patient's therapeutic lifetime. Physicians completed the ratings and a brief survey independently before a virtual group meeting (Round 1).

At the professionally moderated meeting, physicians reviewed Round 1 results and discussed areas of disagreement. After the meeting, physicians repeated their ratings (Round 2). A median rating was calculated for each item and categorized into 3 groups to represent inappropriate, uncertain, and appropriate ratings (1-3, 4-6, and 7-9). Agreement was defined as no more than 2 ratings outside the category with the group median. Using these classifications, and following several rounds of written review, we

developed the following consensus recommendations for the management of SAA, which were approved by all panels.

Results

In total, we evaluated 697 patient scenarios varied by age, donor availability, performance status, previous therapy, and patient response (supplemental Table 1; Figures 1-4). All Round 2 ratings

are provided in the supplemental Appendix. After a structured discussion, 11 panelists agreed on 87% of their Round 2 ratings (Table 1), a significant increase from the proportion of agreement in Round 1 (43%). Below, we summarize the expert recommendations based on areas of consensus. All recommendations below are intended only as general guidance for experienced clinicians who treat patients with SAA. They are in no way intended to supersede individual physician and patient decision making.

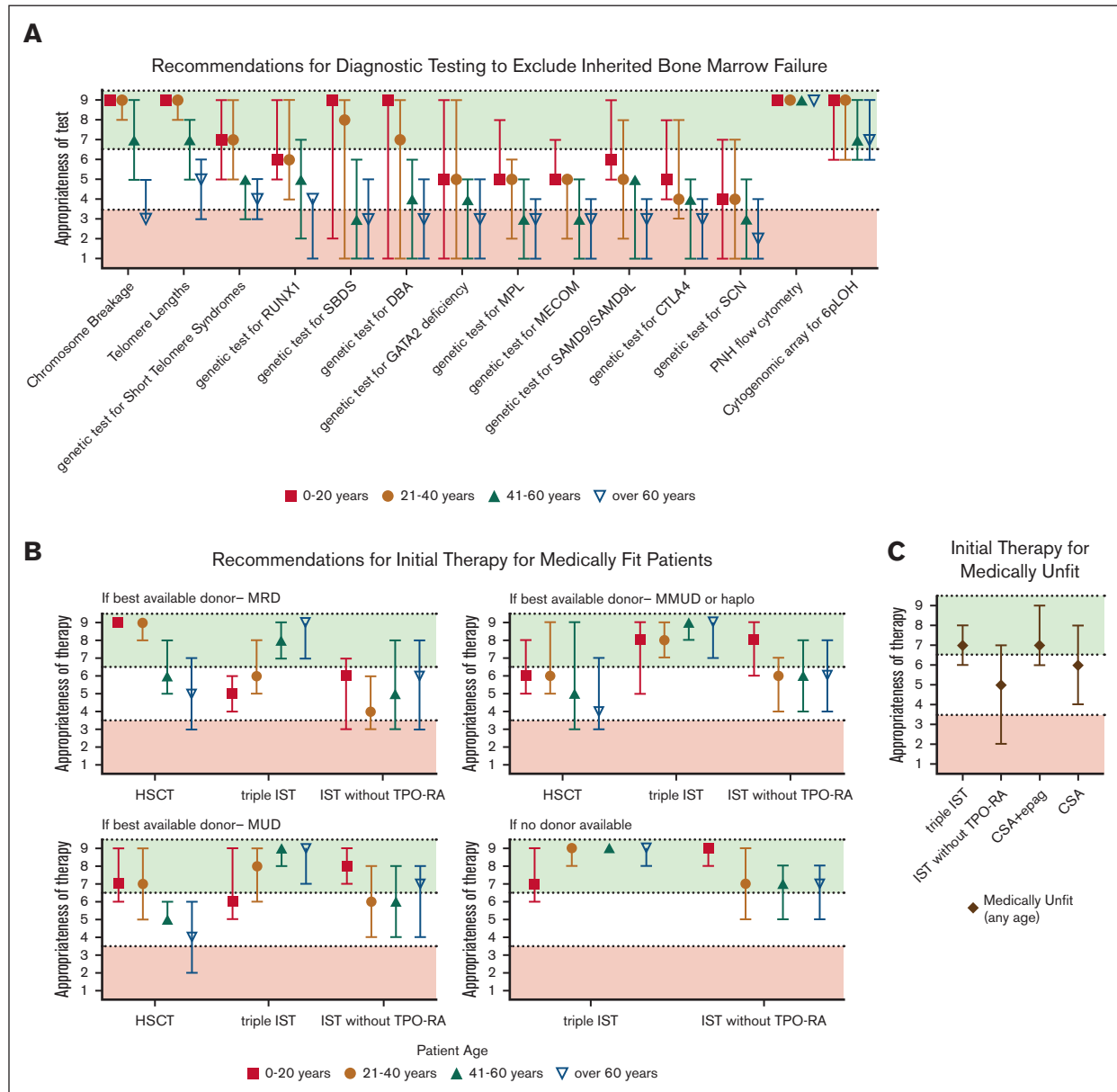


Figure 1. Summary of Delphi panel Round 2 expert consensus ratings for diagnostic testing and initial therapy for SAA. (A) Summary plot of recommended testing to exclude inherited BM failure and “rule in” acquired AA in the evaluation of patients presenting with SAA of different ages. (B) Summary plot of recommended initial therapy in medically fit patients whose best available potential allogeneic donor options are MRD, MUD, mismatched unrelated or haploidentical donor (MMUD or haplo), or no donor available (or unknown). (C) Summary plot of recommended initial treatment for medically unfit patients. In all cases, data represent median score \pm 95% confidence interval (CI). Scores in the range of 7 to 9 (green color) indicate treatments that would be highly appropriate. Scores in the range of 1 to 3 (red color) indicate treatments that would be less appropriate. Intermediate scores in the range of 4 to 6 (white color) indicate treatments that may be appropriate under selected circumstances.

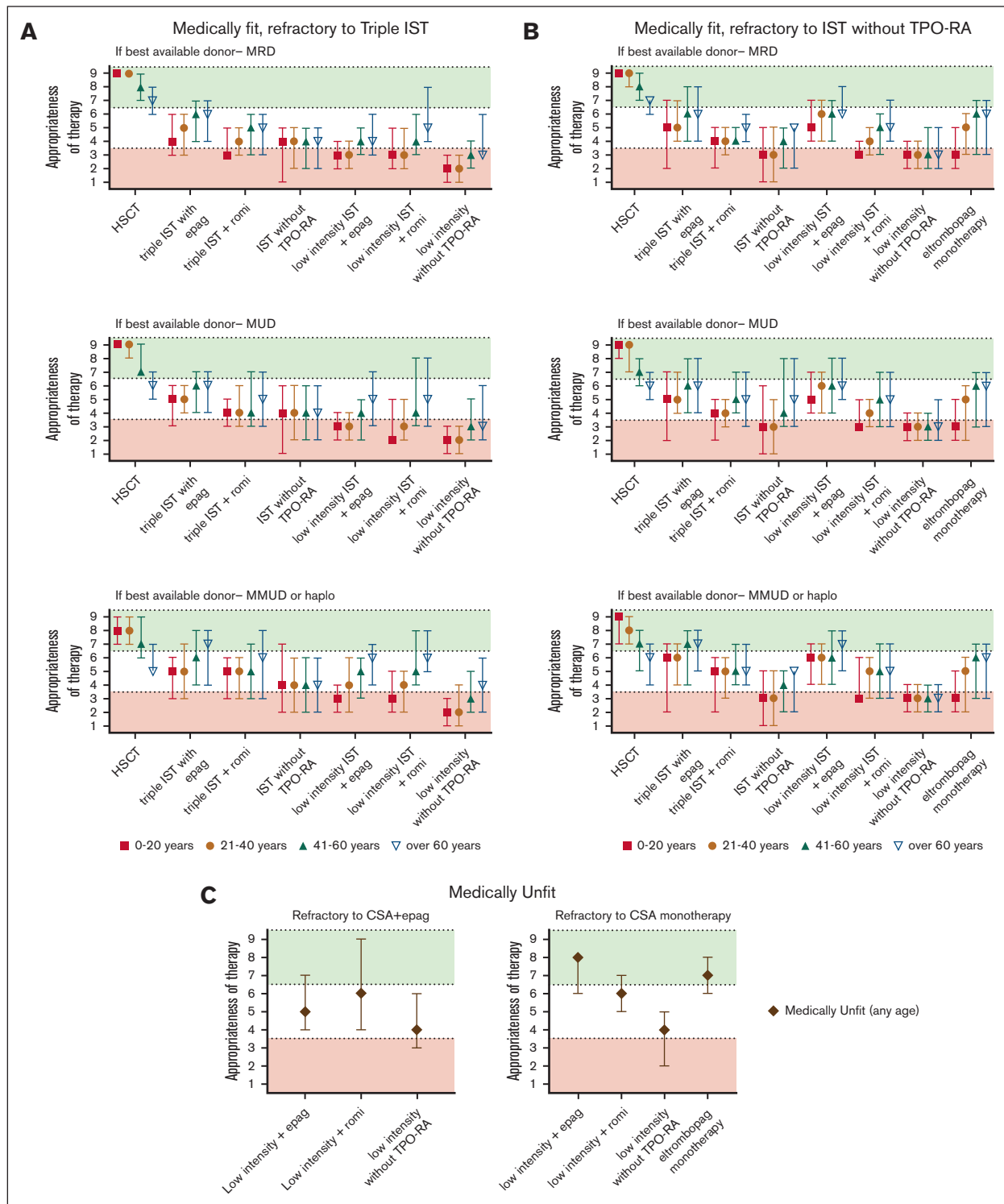


Figure 2. Summary of Delphi panel Round 2 expert consensus ratings for subsequent therapy in patients refractory to initial treatment course. (A) Summary plot of recommended subsequent therapy in medically fit patients who are refractory to “triple IST” (horse ATG, CsA, or eltrombopag) based on the best available allogeneic donor option. (B) Summary plot of recommended subsequent therapy in medically fit patients who are refractory to standard IST administered without TPO-RA stratified based on the best available allogeneic donor option. (C) Summary plot of recommended subsequent therapy in medically unfit patients refractory to the initial treatment of CsA eltrombopag (CsA + eltrombopag), or to CsA monotherapy. In all cases, data represent median score \pm 95% CI. Scores in the range of 7 to 9 (green color) indicate treatments that would be highly appropriate. Scores in the range of 1 to 3 (red color) indicate testing that would be less appropriate. Intermediate scores in the range of 4 to 6 (white color) may be appropriate under selected circumstances.

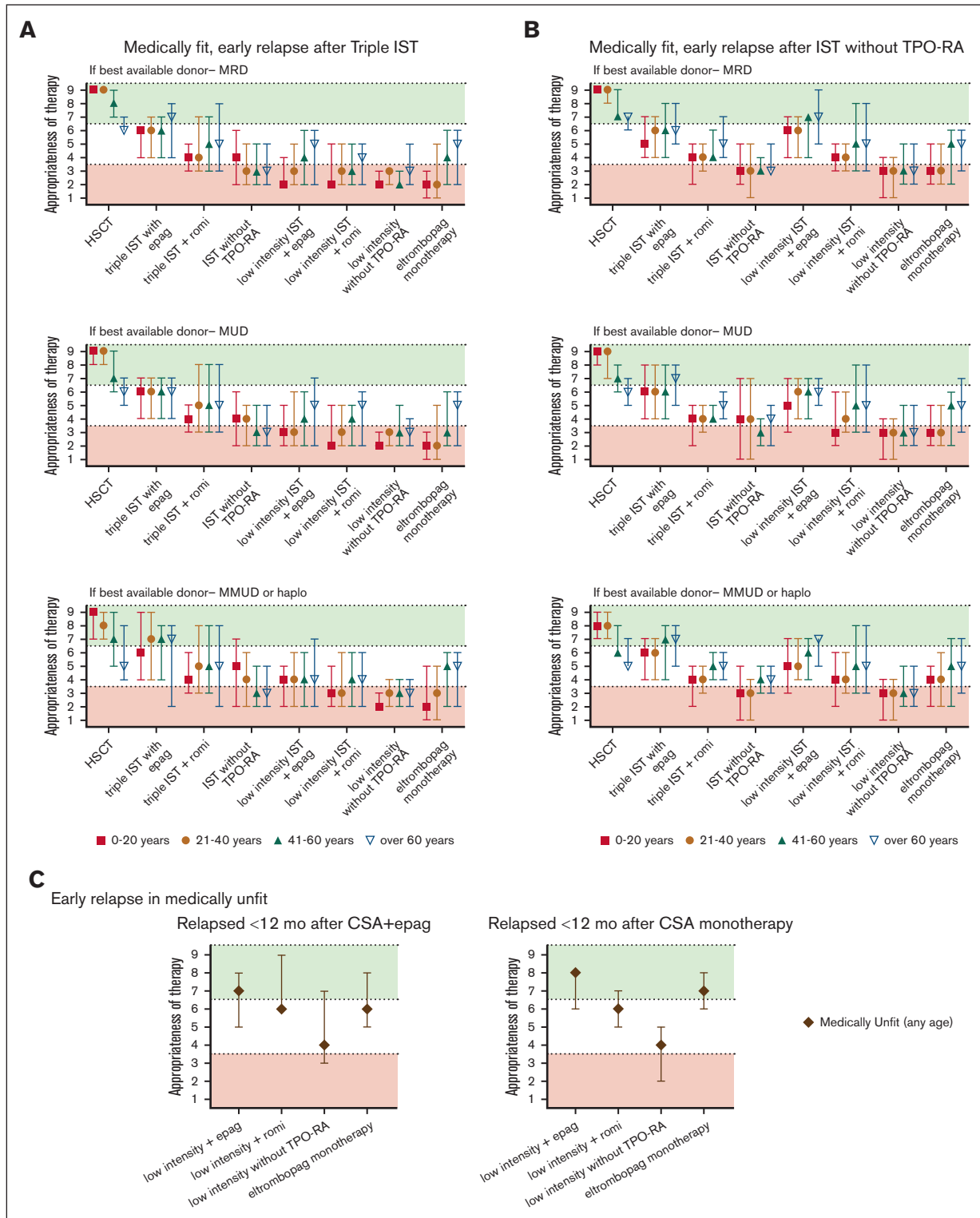


Figure 3. Summary of Delphi panel Round 2 expert consensus ratings for subsequent therapy in patients who relapse within 12 months of initial therapy and again develop severe cytopenias meeting the criteria for SAA. (A) Summary plot of recommended subsequent therapy in medically fit patients who relapse within 12 months of “triple IST” (horse ATG, CsA, or eltrombopag) based on the best available allogeneic donor option. (B) Summary plot of recommended subsequent therapy in medically fit patients who relapse within 12 months of standard IST administered without TPO-RA stratified based on the best available allogeneic donor option. (C) Summary plot of recommended subsequent therapy in medically unfit patients who relapse within 12 months of initial treatment of CsA eltrombopag (CsA + eltrombopag), or CsA monotherapy. In all cases, data represent median score \pm 95% CI. Scores in the range of 7 to 9 (green color) indicate treatments that would be highly appropriate. Scores in the range of 1 to 3 (red color) indicate testing that would be less appropriate. Intermediate scores in the range of 4 to 6 (white color) may be appropriate under selected circumstances.

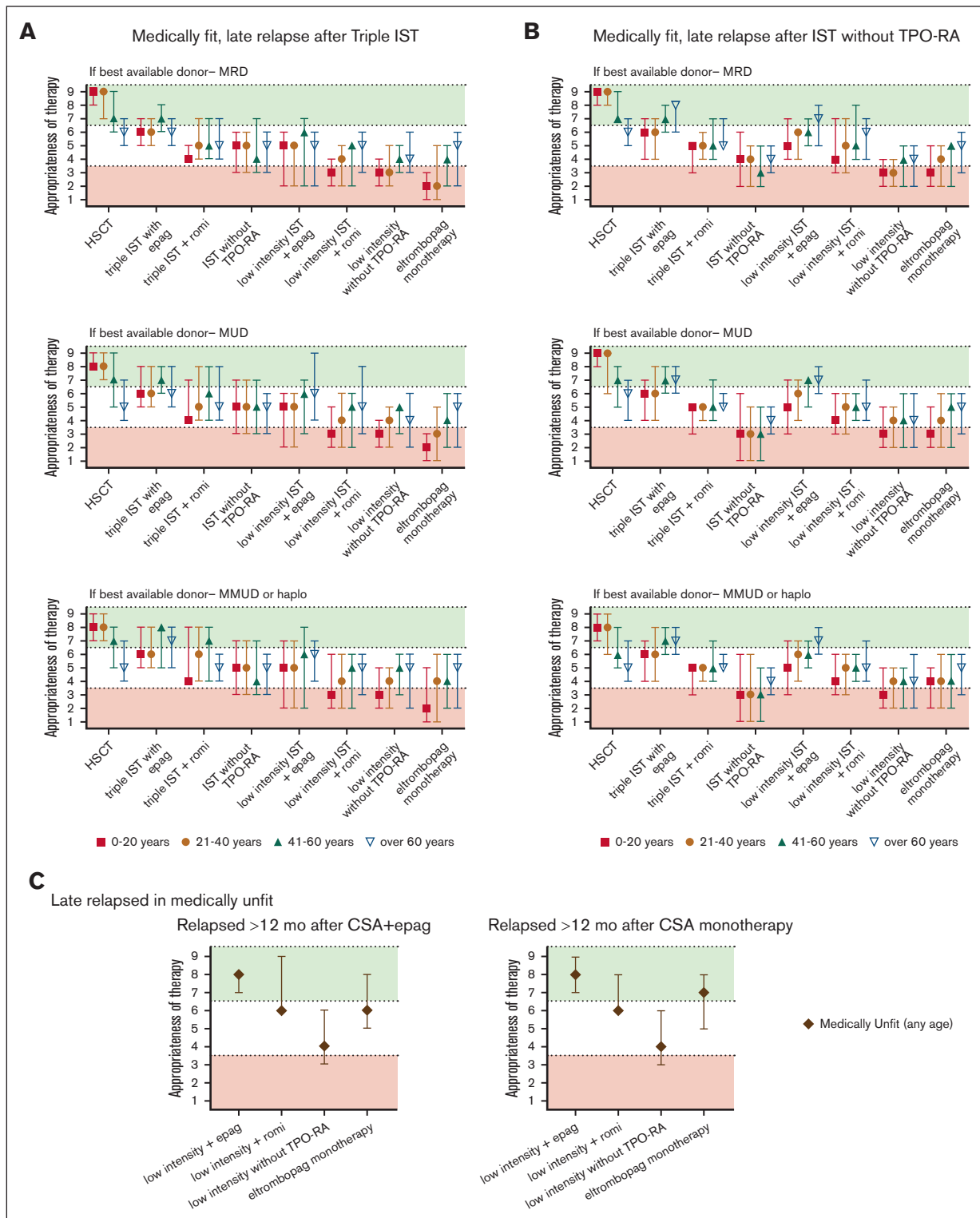


Figure 4. Summary of Delphi panel Round 2 expert consensus ratings for subsequent therapy in patients with late relapses (after 12 months of initial therapy) who again develop severe cytopenias meeting 3 criteria for SAA. (A) Summary plot of recommended subsequent therapy in medically fit patients who relapsed after 12 months of “triple IST” (horse ATG, CsA, or eltrombopag) based on the best available allogeneic donor option. (B) Summary plot of recommended subsequent therapy in medically fit patients who relapsed after 12 months of standard IST administered without TPO-RA stratified based on the best available allogeneic donor option. (C) Summary plot of recommended subsequent therapy in medically unfit patients who relapse after 12 months of initial treatment of CsA eltrombopag (CsA + eltrombopag), or CsA monotherapy. In all cases, data represent median score \pm 95% CI. Scores in the range of 7 to 9 (green color) indicate treatments that would be highly appropriate. Scores in the range of 1 to 3 (red color) indicate testing that would be less appropriate. Intermediate scores in the range of 4 to 6 (white color) may be appropriate under selected circumstances.

Table 1. Rating form results

| | Median ≥ 7 to 9 without disagreement % (n) | Median ≥ 4 to <7 without disagreement % (n) | Median 1 to <4 without disagreement % (n) | Disagreement (≥ 2 ratings of 1-3, and ≥ 2 ratings of 7-9) % (n) |
|------------------------|--|---|--|---|
| First round (N = 697) | 15 (105) | 12 (87) | 16 (110) | 57 (395) |
| Second round (N = 697) | 20 (138) | 47 (326) | 21 (145) | 13 (88) |

Percentages do not add up to 100 because of rounding.

Tests to diagnose acquired SAA and rule out IBMF syndromes

Therapy for acquired SAA should not be delayed while awaiting test results to rule out IBMF syndromes unless there are clinical features suggestive of an IBMF. The following should be obtained in all patients: a complete blood count with differential, review of peripheral blood smear, reticulocyte count, paroxysmal nocturnal hemoglobinuria (PNH) assessment by flow cytometry of peripheral blood, and BM biopsy and aspirate with karyotype. Given the poor growth of cytogenetics culture in cases of AA and to exclude hypocellular myelodysplastic syndrome (MDS), fluorescence in situ hybridization (FISH) for MDS-associated cytogenetic abnormalities is recommended. Somatic next-generation sequencing (NGS) gene panels for genes frequently mutated in hematologic malignancies have become standard in the evaluation of suspected BM failure.^{28,75} Although somatic mutations are commonly seen in both AA and in hypocellular MDS, the inclusion of somatic NGS sequencing panels, supported by commonly used practice guidelines, can help with baseline assessment, including identification of MDS-defining variants, and can be informative prognostically and in longitudinal follow-up of clonal evolution.^{62,76-79} HLA tissue typing should be ordered upfront for patients of all ages who are candidates for HSCT. Comprehensive ancillary studies to exclude transient etiologies of pancytopenia (eg, nutritional deficiencies or viral and other infections) should also be done in all patients.

We recommend obtaining PNH flow cytometry and single-nucleotide polymorphism array genetic testing for chromosome arm 6p loss of heterozygosity clone, because both of these findings are highly predictive of the diagnosis of immune-mediated AA and can be helpful for exclusion of an IBMF.^{80,81} PNH granulocyte clones of $>1\%$ have $\sim 100\%$ positive predictive value for AA, whereas PNH granulocyte clones of $>0.1\%$ were estimated to have 95% specificity and a 91% positive predictive value for the diagnosis of AA.^{80,82} In contrast, rare PNH-like cells and PNH populations of $<0.1\%$ are less specific and have been observed in patients with confirmed IBMF.⁸²

For testing for IBMF (Figure 1A), in patients aged ≤ 40 years, we recommend chromosome breakage analysis for Fanconi anemia (FA) and telomere length analysis for telomere biology disorders, 2 of the most common groups of IBMFs.⁸³ In patients aged 41- to 60 years, these tests can be considered depending on individual patient circumstances, family history, or clinical presentation. Germ line genetic testing is warranted in patients with a high index of suspicion for an IBMF. These include patients who have a clinical diagnosis of FA based on a positive chromosome breakage tests and patients that have a high clinical suspicion of FA but have a negative chromosome breakage test. An estimated 10% to 25% of patients with FA have somatic reversion mosaicism and have a less

severe hematologic presentation; these patients may be missed because of a false-negative chromosome breakage test on peripheral blood lymphocytes and require testing of chromosome breakage in skin fibroblasts.^{84,85}

We recommend IBMF genetic testing in patients with short telomeres suggestive of underlying telomere biology disorders and those with suspected germ line predisposition syndromes. Many known genes associated with IBMF are covered by germ line genetic testing panels offered by academic and commercial genetic laboratories. Depending on the gene content included in the specific NGS panel, testing of additional relevant IBMF genes may have to be ordered separately. Genetic testing should be performed using nonhematopoietic tissue, with gold standard being cultured skin fibroblasts; if hematopoietic tissue was used for initial mutation detection, germ line mutation status should be confirmed using constitutional DNA.

In patients aged >60 years with prior documented normal blood counts and no findings suggestive of IBMF, we do not recommend chromosome breakage analysis for FA or genetic testing for other IBMF syndromes.

Supportive care before and during first-line therapy

Regardless of planned treatment (IST or allo-HSCT), we recommend that a patient with hemoglobin (Hgb) of <7 g/L (or higher Hgb with symptomatic anemia) receive transfusion with leukoreduced, irradiated red blood cells. In patients with $<10 \times 10^9/L$ platelets (or $<50 \times 10^9/L$ if bleeding or if an invasive procedure is planned), we recommend leukoreduced, irradiated platelet transfusion (ideally with single donor platelets to reduce the risk of alloimmunization).

In patients treated with lymphocyte-depleting therapy (eg, ATG, allo-HSCT), we recommend the following prophylactic antimicrobials to be administered until adequate lymphocyte recovery (absolute CD4 T cells of $>200 \times 10^3/L$) and absolute neutrophil count of $>0.50 \times 10^9/L$: antibiotic for pneumocystis pneumonia prophylaxis (eg, atovaquone, trimethoprim/sulfamethoxazole, or pentamidine), antifungal prophylaxis (eg, mold active azole), and antiviral prophylaxis for herpes simplex virus and varicella-zoster virus (eg, acyclovir or valacyclovir).^{2,28,86-89} In line with the Infectious Disease Society of America recommendations for allo-HSCT recipients, we recommend broad-spectrum antibiotic with gram-negative and pseudomonas coverage (eg, fluoroquinolone) until neutrophil recovery in allo-HSCT recipients and patients treated with IST who have prolonged neutropenia (≥ 7 days).^{86,90}

Initial management

In this section, we present our recommendations for medically fit patients, stratified by age (pediatric [aged ≤ 20 years] and adult [aged 21-40, 41-60, and >60 years]; Figure 1B).

In medically fit (ECOG score of ≤ 2) patients⁹¹ aged ≤ 20 years, we recommend the following based on donor availability. In patients with a MRD, we recommend allo-HSCT. For patients without a MRD, we recommend horse ATG + CsA. Given the limited data on the benefits of adding eltrombopag in the pediatric setting, we recommend horse ATG + CsA without eltrombopag in this age group. If available, allo-HSCT from a well-matched unrelated donor (MUD) could also be considered. A randomized clinical trial of IST vs MUD for pediatric patients is currently ongoing.⁹² Upfront transplant from a haploidentical donor has published success and is currently under investigation in a multicenter Blood and Marrow Transplant Clinical Trials Network trial.^{31,93} If a haploidentical donor is available, the panel felt that a haploidentical transplant can also be an option with careful clinical consideration.^{31,93}

In patients aged 21 to 40 years, we recommend the following based on donor availability. In patients with a MRD, we recommend allo-HSCT. For other patients, based on the results of the RACE and the National Institute of Health studies in upfront AA, demonstrating the superior hematologic responses of IST with the inclusion of eltrombopag,^{42,44} we recommend horse ATG + CsA + eltrombopag. If a well-matched MUD is available, allo-HSCT can be considered. Decision making could be driven by various factors, including patients' health and performance status, weighing relative value for a given patient of the pros/cons of the short-term vs long-term risks of IST vs allo-HSCT, disease features and projected likelihood of response to IST (eg, in patients with favorable vs less favorable prognostic features),^{62,76,94-98} presence of deep neutropenia at diagnosis, fertility and family planning considerations, and psychosocial issues (eg, motivation/adherence to treatment regimen and availability of caregiver support).⁹⁹⁻¹⁰⁵ In patients aged >40 years, we recommend horse ATG + CsA + eltrombopag. If these patients have an available MRD or MUD, allo-HSCT can also be considered depending on individual patient circumstances. Upfront transplant from a haploidentical donor is available and, as above, currently under investigation.^{31,93}

In all medically unfit (ECOG score of >2) patients⁹¹ regardless of age, we recommend high- or low-intensity IST (Table 2) in combination with eltrombopag (Figure 1C).

Subsequent management

After medical therapy is initiated, eltrombopag should be continued for ~6 months, provided the patient has achieved a hematologic response, and then tapered or stopped.^{42,106} Patients should be maintained on full-dose CsA for 6 to 12 months. When adequate hematologic response is achieved, CsA should be tapered slowly

over the second year (ie, after the initial 6-12 months full-dose treatment, lower dose CsA maintenance should be maintained for at least 12-18 months, with taper guided by sustained hematologic response). A complete hematologic response is defined as neutrophils of $>1 \times 10^9/L$ (in some trials it is $1.5 \times 10^9/L$), platelets of $>100 \times 10^9/L$, and Hgb of >100 g/L.⁶³ Patients should be monitored for clonal evolution by BM biopsy while on TPO-RA and if there are clinical concerns (eg, a decline in blood counts or lack of response to therapy), with tests including karyotyping, FISH for MDS-associated cytogenetic rearrangements (in the setting of limited or no growth on cytogenetics), and NGS for malignancy-associated mutations.^{42-44,46,62,76,107,108}

Below we present recommendations for treating patients who are refractory (ie, no response at 3-6 months after initial therapy) or who have relapsed (ie, after obtaining a response). Unless the etiology of the patient's hematologic deterioration is clear with recent BM studies, we recommend repeating BM biopsy for karyotyping, NGS, and FISH to evaluate for MDS-associated clonal evolution. We recognize that relapsing patients will, in most cases, be treated initially by increasing the dose or resuming low-intensity therapy (eg, CsA and/or eltrombopag) to prevent recurrence of severe cytopenias, and that many patients will respond to these initial therapies. The recommendations below are for patients who did not respond to these initial approaches and progressed to meet SAA criteria despite therapeutic interventions.

In medically fit patients⁹¹ who received horse ATG + CsA + eltrombopag in a first-line setting, we recommend the following (Figure 2A): in patients aged ≤ 60 years refractory to initial therapy, we recommend allo-HSCT with the best available donor (eg, MRD, MUD, or haploidentical). In patients aged >60 years refractory to initial therapy, we also recommend allo-HSCT with a MRD, and depending on individual patient circumstances, allo-HSCT with the best-available alternative donor (eg, MUD or haploidentical) may also be considered.

The panel considered clinical scenarios of early relapses (within 12 months of IST; Figure 3) separately from late relapses after 12 months of therapy (Figure 4). This was because early relapses at the time when most patients remain on CsA but discontinue eltrombopag may have a distinct pathogenesis from later immunologic relapses that occur at the time of CsA taper and discontinuation.^{106,109} However, our recommendations for managing early and late relapses are similar. In patients aged ≤ 60 years who relapsed after obtaining a response (Figure 3), we recommend either allo-HSCT with the best available donor (MRD, MUD, or haploidentical) or repeating high-intensity medical therapy with

Table 2. High- and low-intensity therapeutic options

| High-intensity therapy | Low-intensity therapy |
|---|---|
| <ul style="list-style-type: none"> • Horse or rabbit ATG + CsA w/wo eltrombopag • Horse or rabbit ATG + other non-CsA immunosuppressant (eg, tacrolimus) w/wo eltrombopag • Alemtuzumab • Horse or rabbit ATG + CsA + androgens or growth factors | <ul style="list-style-type: none"> • CsA w/wo eltrombopag • Other non-CsA immunosuppressant (eg, tacrolimus) • CsA + androgens • Androgens (single agent) • Growth factors (single agent; eg, erythropoietin or granulocyte colony-stimulating factor) • Eltrombopag (single agent) |

w/wo, with or without.

These are examples of common therapy combinations. Cyclophosphamide has also shown efficacy for AA but is associated with high levels of toxicity and is no longer routinely recommended.⁷²

Romiplostim can also be combined with horse or rabbit ATG + CsA or non-CsA immunosuppressant or used as a single agent.

eltrombopag or romiplostim. When allo-HSCT is not an option in the refractory or relapsed setting, we recommend high-intensity medical therapy (Table 1; eg, rabbit ATG + CsA, horse ATG + CsA, alemtuzumab, in combination with eltrombopag). Recent data showed that romiplostim also has activity in relapsed/refractory AA.¹¹⁰⁻¹¹³ We also recommend evaluating all patients for available clinical trials.

In medically fit patients who received horse ATG + CsA as first-line therapy, we recommend the following: in patients aged ≤ 60 years refractory to initial therapy (Figure 2B) or relapsed after obtaining a response, we recommend allo-HSCT with the best available donor (MRD, MUD, or haploidentical). In patients aged > 60 years refractory to initial therapy or relapsed after obtaining a response, we also recommend allo-HSCT with an MRD; allo-HSCT with a best available alternative donor (eg, MUD or haploidentical) may be considered in patients depending on individual patient circumstances. For patients who relapsed after obtaining a response (Figure 3B), a second course of high-intensity therapy or resuming CsA with eltrombopag can be attempted to reinduce response; we recommend this option particularly in patients with late immune-type AA relapses. In patients aged > 20 years, when allo-HSCT is not a medical option, we recommend high-intensity or low-intensity medical therapy in combination with eltrombopag (Table 2). In patients aged ≤ 20 years, when allo-HSCT is not a medical option, we did not agree on whether eltrombopag should be provided with second-line medical therapy because of limited data for this option in young patients. We also recommend evaluating all patients for available clinical trials.

In all medically unfit patients⁹¹ regardless of age (Figures 2C and 3C), we recommend the following: low-intensity therapy (Table 2) with eltrombopag; single-agent eltrombopag in patients who did not receive eltrombopag in the first-line setting; romiplostim can also be considered depending on patient-specific clinical context. Switching to romiplostim may be particularly advantageous in patients intolerant to eltrombopag, such as those with Gilbert syndrome, difficulty with dietary restrictions, and those with insufficient response on eltrombopag for whom dose escalation with romiplostim can be helpful.¹¹¹⁻¹¹³

Discussion

In this study, we used a RAND/UCLA modified Delphi panel process to develop expert consensus recommendations for the treatment of SAA. SAA is a relapsing/remitting autoimmune disease that can be managed with differing therapies ranging from allo-HSCT to single-agent low-intensity therapies. Recent advances in transplant and nontransplant therapies and supportive care have improved patient outcomes, but evidence comparing therapeutic modalities and their optimal use over the disease course is lacking. Given the rarity of AA and changing clinical landscape of available therapies, we convened a panel of 11 AA experts to consider various aspects of initial and subsequent management of patients with SAA to develop consensus recommendations. Through the iterative consensus process, we evaluated 697 individual patient care scenarios spanning the spectrum of patient age, medical fitness, best donor availability, initial or subsequent line of therapy, and response to prior therapies, and present our expert assessments of the management of diverse AA clinical scenarios. Our expert panel recommendations presented

here summarize areas consensus and areas of active discussion for which expert consensus on optimal management was not reached.

During the Delphi process, treatment options were considered individually within each clinical scenario. Therapies with proven efficacy were rated as appropriate with varying strengths of the recommendation based on whether this was a preferred or a less-preferred option based on a patient's clinical characteristics and response to prior therapies. For example, in considering initial therapies in younger medically fit patients with an available MRD donor, allo-HSCT was the preferred option, but IST-based regimens were also rated as potentially appropriate under selected circumstances. In doing so, we recognized that, in practice, patients may do well with IST-based regimens, which they may select over allo-HSCT because of personal considerations (eg, fertility, educational, financial, or psychological factors).

Improved allo-HSCT outcomes with unrelated and haploidentical donors are reflected in the panel's recommendation of allo-HSCT using MRDs or MUDs as the preferred initial therapy in younger, medically fit patients. The panel recommended salvage allo-HSCT using either MRDs, MUDs, or haploidentical donors in younger medically fit patients who were refractory or relapsed to meet SAA criteria after initial therapy. Our panel took place before the availability of the more recent studies reporting encouraging outcomes using haploidentical transplant donors in the upfront setting^{31,35,36,114-116} and we are not yet able to provide consensus recommendations on the role haploidentical donor allo-HSCT in upfront management of AA. Similarly, the panel could not clearly recommend allo-HSCT over IST outright in patients whose best donor was a MUD because there is not yet clear clinical support for this recommendation, but the panel felt that in practice upfront MUD allo-HSCT could be appropriate in younger fit patients with careful clinical consideration. A multi-center randomized study comparing MUD vs IST for pediatric and young adults with SAA is ongoing (ClinicalTrials.gov identifier: NCT05600426).⁹² Of course, the use of each of these therapies varies by patient characteristics, including age, donor availability, medical fitness, response to prior therapies, and physician input. Detailed recommendations on when different therapeutic options should be initiated or changed based on these characteristics will facilitate better patient care.

The addition of eltrombopag to upfront IST has improved overall response rates to nearly 80%, with faster and more complete hematologic recovery.^{42,44,106} Our panel recommended the inclusion of eltrombopag in high- and low-intensity IST regimens as the preferred nontransplant treatment option. At the time of our panel, romiplostim had been shown to also have efficacy in SAA, particularly during second- or later-line treatment¹¹¹⁻¹¹³; and the panel recommended romiplostim to be considered as an alternative, particularly during subsequent therapy and in cases of insufficient response or intolerance of eltrombopag. Since the time of the panel, early data demonstrate encouraging efficacy of other TPO-RAs (eg, avatrombopag and hetrombopag) in AA therapy.⁴⁷⁻⁴⁹ Further research is needed to provide clear guidance on comparative effectiveness and safety of the newer TPO-RA and optimal sequencing in first-line vs subsequent therapy. The evidence on the efficacy of eltrombopag in subsequent management has been less clear in the pediatric population,¹¹⁷ and our panel acknowledged

that limitation by not uniformly recommending it in patients aged ≤ 20 years.

There are gaps in our recommendations. We did not specify the amount of time that patient blood counts may be trending down after loss of hematologic response before initiating high-intensity therapy. Depending on clinical scenario at the time of relapse (eg, whether loss of hematologic response was temporally related to the taper or discontinuation of CsA or discontinuation of eltrombopag) we recommend early therapeutic intervention (eg, resuming eltrombopag or CsA, or intensifying CsA dose), before patients once again meet the criteria for SAA. We did not attempt to develop a comprehensive list of all genetic tests needed to rule out IBMF or germ line predisposition syndromes. Our panel did agree that for patients who do not have overt findings of IBMF, the care of AA should not be delayed while awaiting the results of genetic testing, and recommended functional screening with chromosome breakage and telomere length measurements for all but the oldest patients. Recent publications have presented recommendations for additional genetic testing,^{23,51,118-120} the specifics of which may depend on institutional availability, patient age, or other presenting characteristics. Additional work is needed to determine optimal genetic testing in the evaluation of suspected idiopathic SAA in adults.

To generate our recommendations, we used the RAND/UCLA modified Delphi panel method, which has been used extensively to develop quality measures and clinical guidance in a variety of areas. This method is distinct from evidence-based recommendations produced using the GRADE system, which rely on the availability of evidence and the classification of such evidence into 4 levels (high, moderate, low, and very low). The quality of evidence helps determine the strength of resulting guidelines.¹²¹ Consensus recommendations developed using the Delphi panel method have content, construct, and predictive validity,⁵³ and have been shown to improve health outcomes.^{54,55} Nevertheless, the method still relies on physician opinion. Although all physicians had significant experience treating patients with SAA and represented diverse backgrounds, practice settings, and geographic regions, 11 people cannot represent the full experience of clinicians who work in this field. Furthermore, our panel included only experts from the United States, so the consensus recommendations may not be generalizable to care in other countries. Although the modified Delphi method does have reasonable reproducibility,⁵⁶ different groups of experts may have reached different conclusions. We hope that our Delphi consensus ratings for individual scenarios along with the spread of scores within the panel (Figures 1-4) will be useful to the community as an expert discussion of the relative appropriateness of treatment options and areas of disagreement in the field.

These consensus statements were developed with the goal of improving the quality of care delivered to patients with SAA, informed by currently available evidence and expert clinical opinion. In this setting, the guidance is unable to capture the nuance that will vary in individual patients (eg, insurance coverage, cost of therapy, practice setting, and other social determinants of health) and are intended as general consensus recommendations only. In practice, other clinical and nonclinical factors beyond those addressed here may affect optimal SAA management. Still, the hope is to ultimately offer the best available therapy to each patient with SAA.

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ORCID profiles: C.M.d.C., [0000-0002-4485-6025](https://orcid.org/0000-0002-4485-6025); Z.R.R., [0000-0003-1478-2368](https://orcid.org/0000-0003-1478-2368); B.L.S., [0000-0001-9620-7839](https://orcid.org/0000-0001-9620-7839); M.W.W., [0000-0001-6638-9643](https://orcid.org/0000-0001-6638-9643); I.Y., [0000-0001-6035-2815](https://orcid.org/0000-0001-6035-2815); B.J.P., [0009-0008-4984-7751](https://orcid.org/0009-0008-4984-7751).

Correspondence: Bhumika J. Patel, Prisma Health, 701 Grove Rd, Greenville, SC 29605; email: bhumika.patel@prismahealth.org.

References

1. Young NS, Kaufman DW. The epidemiology of acquired aplastic anemia - PubMed. *Haematologica*. 2008;93(4):489-492.
2. Peslak SA, Olson T, Babushok D. Diagnosis and treatment of Aplastic Anemia. *Curr Treat Options Oncol*. 2017;18(12):1-22.
3. Young NS, Bacigalupo A, Marsh JCW. Aplastic anemia: pathophysiology and treatment. *Biol Blood Marrow Transplant*. 2010;16(suppl 1):S119-S125.
4. Jang J, Kim H, Park SS, et al. Single-cell RNA sequencing reveals novel cellular factors for response to immunosuppressive therapy in aplastic anemia. *Hemasphere*. 2023;7(11):e977.
5. Zhang J, Liu T, Duan Y, et al. Single-cell analysis highlights a population of Th17-polarized CD4+ naïve T cells showing IL6/JAK3/STAT3 activation in pediatric severe aplastic anemia. *J Autoimmun*. 2023;136:103026.
6. Solimando AG, Desantis V, Palumbo C, et al. STAT1 overexpression triggers aplastic anemia: a pilot study unravelling novel pathogenetic insights in bone marrow failure. *Clin Exp Med*. 2023;23(6):2687-2694.
7. Vaht K, Brenner J, Ednersson SB, Ljungman P, Brune M, Andersson P. Bone marrow expression of CD68/CD163 macrophages, IL-17 and FOXP3 cells in aplastic anemia and their relation to prognosis. *Eur J Haematol*. 2023;110(3):313-321.
8. Pagliuca S, Gurnari C, Hercus C, et al. Molecular landscape of immune pressure and escape in aplastic anemia. *Leukemia*. 2023;37(1):202-211.

9. Olson TS, Frost BF, Duke JL, et al. Pathogenicity and impact of HLA class I alleles in aplastic anemia patients of different ethnicities. *JCI Insight*. 2022; 7(22):e163040.
10. Dong P, Chen L, Wu H, et al. Impaired immunosuppressive role of myeloid-derived suppressor cells in acquired aplastic anemia. *Haematologica*. 2022; 107(12):2834-2845.
11. You X, Yang Q, Yan K, et al. Multi-omics profiling identifies pathways associated with CD8+ T-cell activation in severe aplastic anemia. *Front Genet*. 2021;12:790990.
12. Zaimoku Y, Patel BA, Adams SD, et al. HLA associations, somatic loss of HLA expression, and clinical outcomes in immune aplastic anemia. *Blood*. 2021;138(26):2799-2809.
13. Lim SP, Costantini B, Mian SA, et al. Treg sensitivity to FasL and relative IL-2 deprivation drive idiopathic aplastic anemia immune dysfunction. *Blood*. 2020;136(7):885-897.
14. Savage SA, Viard M, O'hUigin C, et al. Genome-wide association study identifies HLA-DPB1 as a significant risk factor for severe aplastic anemia. *Am J Hum Genet*. 2020;106(2):264-271.
15. Kelkka T, Tyster M, Lundgren S, et al. Anti-COX-2 autoantibody is a novel biomarker of immune aplastic anemia. *Leukemia*. 2022;36(9):2317-2327.
16. Sklarz T, Hurwitz SN, Stanley NL, Juusola J, Bagg A, Babushok D. Aplastic anemia in a patient with COVID due to NFKB1 haploinsufficiency. *Cold Spring Harb Mol Case Stud*. 2020;6(6):a005769.
17. Braudeau C, Delbos L, Couec ML, et al. System-level immune monitoring reveals new pathophysiological features in hepatitis-associated aplastic anemia. *Blood Adv*. 2023;7(15):4039-4045.
18. Kaufman D, Kelly J, Levy M, Shapiro S. *The Drug Etiology of Agranulocytosis and Aplastic Anemia*. Oxford University Press; 1991.
19. Mary JY, Baumelou E, Guiguet M. Epidemiology of aplastic anemia in France: a prospective multicentric study. the French Cooperative Group for epidemiological study of aplastic Anemia. *Blood*. 1990;75(8):1646-1653.
20. Cartwright RA, McKinney PA, Williams L, et al. Aplastic anaemia incidence in parts of the United Kingdom in 1985. *Leuk Res*. 1988;12(6):459-463.
21. Clausen N, Kreuger A, Salmi T, Storm-Mathisen I, Johannesson G. Severe aplastic anaemia in the Nordic countries: a population based study of incidence, presentation, course, and outcome. *Arch Dis Child*. 1996;74(4):319-322.
22. Maluf EMCP, Pasquini R, Eluf JN, Kelly J, Kaufman DW. Aplastic anemia in Brazil: incidence and risk factors. *Am J Hematol*. 2002;71(4):268-274.
23. DeZern AE, Churpek JE. Approach to the diagnosis of aplastic anemia. *Blood Adv*. 2021;5(12):2660-2671.
24. Camitta BM, Thomas D, Nathan DG. A prospective study of androgens and bone marrow transplantation for treatment of severe aplastic anemia. *Blood*. 1979;53(3):504-14.
25. Yoon HH, Huh SJ, Lee JH, et al. Should we still use Camitta's criteria for severe aplastic anemia? *Korean J Hematol*. 2012;47(2):126-130.
26. Camitta B, Storb R, Thomas E. Aplastic anemia (first of two parts): pathogenesis, diagnosis, treatment, and prognosis. *N Engl J Med*. 1982;306(11):645-652.
27. Camitta BM, Storb R, Thomas D. Aplastic anemia: second of two parts: pathogenesis, diagnosis, treatment and prognosis. *N Engl J Med*. 1982; 306(12):712-718.
28. Killick SB, Bown N, Cavenagh J, et al. Guidelines for the diagnosis and management of adult aplastic anaemia. *Br J Haematol*. 2016;172(2):187-207.
29. Prata PH, Eikema DJ, Afansyev B, et al. Haploidentical transplantation and posttransplant cyclophosphamide for treating aplastic anemia patients: a report from the EBMT Severe Aplastic Anemia Working Party. *Bone Marrow Transplant*. 2020;55(6):1050-1058.
30. Zhang Y, Mo W, Zuo Y, et al. Comparable survival outcome between transplantation from haploidentical donor and matched related donor or unrelated donor for severe aplastic anemia patients aged 40 years and older: a retrospective multicenter cohort study. *Clin Transplant*. 2020;34(3):e13810.
31. DeZern AE, Zahurak ML, Symons HJ, et al. Haploidentical BMT for severe aplastic anemia with intensive GVHD prophylaxis including posttransplant cyclophosphamide. *Blood Adv*. 2020;4(8):1770-1779.
32. ElGohary G, El Fakih R, de Latour R, et al. Haploidentical hematopoietic stem cell transplantation in aplastic anemia: a systematic review and meta-analysis of clinical outcome on behalf of the severe aplastic anemia working party of the European group for blood and marrow transplantation (SAAWP of EBMT). *Bone Marrow Transplant*. 2020;55(10):1906-1917.
33. Li Y, Wang N, Li L, et al. Haploidentical transplantation with modified post-transplantation cyclophosphamide for patients with primary aplastic anemia: a multicenter experience. *Transplant Cell Ther*. 2021;27(4):331.e1-331.e7.
34. Yang K, Gong S, Jiang T, et al. Haploidentical peripheral stem cell transplantation for young patients with severe aplastic anemia using post-transplantation cyclophosphamide and methotrexate. *Transplant Cell Ther*. 2021;27(5):429.e1-429.e7.
35. Alotaibi H, Aljurf M, De Latour R, et al. Upfront alternative donor transplant versus immunosuppressive therapy in patients with severe aplastic anemia who lack a fully HLA-matched related donor: systematic review and meta-analysis of retrospective studies, on behalf of the Severe Aplastic Anemia Working Party of the European Group for Blood and Marrow Transplantation. *Transplant Cell Ther*. 2022;28(2):105.e1-105.e7.
36. DeZern AE, Eapen M, Wu J, et al. Haploidentical bone marrow transplantation in patients with relapsed or refractory severe aplastic anaemia in the USA (BMT CTN 1502): a multicentre, single-arm, phase 2 trial. *Lancet Haematol*. 2022;9(9):e660-e669.
37. Xu LP, Xu ZL, Wang SQ, et al. Long-term follow-up of haploidentical transplantation in relapsed/refractory severe aplastic anemia: a multicenter prospective study. *Sci Bull (Beijing)*. 2022;67(9):963-970.

38. Wu L, Zhou M, Li Y, et al. Prospective study of a modified post-transplantation cyclophosphamide regimen for severe aplastic anemia patients with HLA-haploidentical transplantation. *Transplant Cell Ther.* 2023;29(7):463.e1-463.e7.
39. Quintero V, Bueno-Sánchez D, Mozo-Del-Castillo Y, et al. Haploidentical hematopoietic stem cell transplantation in pediatric patients with acquired hypocellular bone marrow failure. *Transplant Cell Ther.* 2023;29(10):621.e1-621.e6.
40. Storb R. Allogeneic bone marrow transplantation for aplastic anemia. *Int J Hematol.* 2024;119(3):220-230.
41. Scheinberg P, Scheinberg P, Young NS. Horse versus rabbit antithymocyte globulin in acquired aplastic anemia. *N Engl J Med.* 2011;365(5):430-8.
42. Townsley DM, Scheinberg P, Winkler T, et al. Eltrombopag added to standard immunosuppression for aplastic Anemia. *N Engl J Med.* 2017;376(16):1540-1550.
43. Assi R, Garcia-Manero G, Ravandi F, et al. Addition of eltrombopag to immunosuppressive therapy in patients with newly diagnosed aplastic anemia. *Cancer.* 2018;124(21):4192-4201.
44. Peffault de Latour R, Kulasekararaj A, Iacobelli S, et al. Eltrombopag added to immunosuppression in severe aplastic anemia. *N Engl J Med.* 2022;386(1):11-23.
45. Desmond R, Townsley DM, Dumitriu B, et al. Eltrombopag restores trilineage hematopoiesis in refractory severe aplastic anemia that can be sustained on discontinuation of drug. *Blood.* 2014;123(12):1818-1825.
46. Olnes MJ, Scheinberg P, Calvo KR, et al. Eltrombopag and improved hematopoiesis in refractory aplastic anemia. *N Engl J Med.* 2012;367(1):11-19.
47. Yang W, Zhao X, Liu X, et al. Hetrombopag plus porcine ATG and cyclosporine for the treatment of aplastic anaemia: early outcomes of a prospective pilot study. *Exp Hematol Oncol.* 2023;12(1):1-4.
48. Zhang Z, Hu Q, Yang C, Chen M, Han B. Comparison of eltrombopag and avatrombopag in the treatment of refractory/relapsed aplastic anemia: a single-center retrospective study in China. *Ther Adv Hematol.* 2023;14:20406207231191310.
49. Wan Z, Chen M, Han B. Avatrombopag, a promising novel thrombopoietin receptor agonist for refractory/relapsed/intolerant non-severe aplastic anemia: a phase 2 single-arm clinical trial. *Ann Med.* 2023;55(1):2224044.
50. Scheinberg P. Acquired severe aplastic anaemia: how medical therapy evolved in the 20th and 21st centuries. *Br J Haematol.* 2021;194(6):954-969.
51. Young NS. Aplastic Anemia. *N Engl J Med.* 2018;379(17):1643-1656.
52. Bojke L, Soares MO, Claxton K, et al. Reference case methods for expert elicitation in health care decision making. *Med Decis Making.* 2022;42(2):182-193.
53. Kravitz RL, Laouri M, Kahan JP, Sherman T, Hilborne L, Brook RH. Validity of criteria used for detecting underuse of coronary revascularization. *JAMA.* 1995;274(8):632-638.
54. Patel MR, Calhoon JH, Dehmer GJ, et al. ACC/AATS/AHA/ASE/ASNC/SCAI/SCCT/STS 2017 appropriate use criteria for coronary revascularization in patients with stable ischemic heart disease : a report of the American College of Cardiology Appropriate Use Criteria Task Force, American Association for Thoracic Surgery, American Heart Association, American Society of Echocardiography, American Society of Nuclear Cardiology, Society for Cardiovascular Angiography and Interventions, Society of Cardiovascular Computed Tomography, and Society of Thoracic Surgeons. *J Nucl Cardiol.* 2017;24(5):1759-1792.
55. Bradley SM, Chan PS, Hartigan PM, et al. Validation of the appropriate use criteria for percutaneous coronary intervention in patients with stable coronary artery disease (from the COURAGE trial). *Am J Cardiol.* 2015;116(2):167-173.
56. Shekelle PG, Kahan JP, Bernstein SJ, Leape LL, Kamberg CJ, Park RE. The reproducibility of a method to identify the overuse and underuse of medical procedures. *N Engl J Med.* 1998;338(26):1888-1895.
57. Hemingway H, Crook AM, Feder G, et al. Underuse of coronary revascularization procedures in patients considered appropriate candidates for revascularization. *N Engl J Med.* 2001;344(9):645-654.
58. Schünemann H, Brożek J, Guyatt G, Oxman. GRADE handbook: handbook for grading the quality of evidence and the strength of recommendations using the GRADE approach. Updated October 2013. Accessed 12 March 2024. <https://gdt.gradeapro.org/app/handbook/handbook.html>
59. Fink A, Kossecoff J, Chassin M, Brook RH. Consensus methods: characteristics and guidelines for use. *Am J Public Health.* 1984;74(9):979-983.
60. Drexler B, Passweg J. Current evidence and the emerging role of eltrombopag in severe aplastic anemia. *Ther Adv Hematol.* 2021;12:1-10.
61. Stanley N, Olson TS, Babushok DV. Recent advances in understanding clonal haematopoiesis in aplastic anaemia. *Br J Haematol.* 2017;177(4):509-525.
62. Yoshizato T, Dumitriu B, Hosokawa K, et al. Somatic mutations and clonal hematopoiesis in aplastic anemia. *N Engl J Med.* 2015;373(1):35-47.
63. Scheinberg P, Wu CO, Nunez O, et al. Treatment of severe aplastic anemia with a combination of horse antithymocyte globulin and cyclosporine, with or without sirolimus: a prospective randomized study. *Haematologica.* 2009;94(3):348-354.
64. Frickhofen N, Kaltwasser JP, Schrezenmeier H, et al. Treatment of aplastic anemia with antilymphocyte globulin and methylprednisolone with or without cyclosporine. The German Aplastic Anemia Study Group. *N Engl J Med.* 1991;324(19):1297-1304.
65. Frickhofen N, Heimpel H, Kaltwasser JP, Schrezenmeier H. Antithymocyte globulin with or without cyclosporin A: 11-year follow-up of a randomized trial comparing treatments of aplastic anemia. *Blood.* 2003;101(4):1236-1242.
66. Tichelli A, Schrezenmeier H, Socié G, et al. A randomized controlled study in patients with newly diagnosed severe aplastic anemia receiving antithymocyte globulin (ATG), cyclosporine, with or without G-CSF: a study of the SAA Working Party of the European Group for Blood and Marrow Transplantation. *Blood.* 2011;117(17):4434-4441.

67. Scheinberg P, Nunez O, Weinstein B, Scheinberg P, Wu CO, Young NS. Activity of alemtuzumab monotherapy in treatment-naïve, relapsed, and refractory severe acquired aplastic anemia. *Blood*. 2012;119(2):345-354.
68. Afaible MG, Shaik M, Sugimoto Y, et al. Efficacy of rabbit anti-thymocyte globulin in severe aplastic anemia. *Haematologica*. 2011;96(9):1269-1275.
69. Champlin R, Ho W, Gale RP. Antithymocyte globulin treatment in patients with aplastic anemia: a prospective randomized Trial. *N Engl J Med*. 1983;308(3):113-118.
70. Champlin RE, Ho WG, Feig SA, Winston DJ, Lenarsky C, Gale RP. Do androgens enhance the response to antithymocyte globulin in patients with aplastic anemia? a prospective randomized trial. *Blood*. 1985;66(1):184-188.
71. Brodsky RA, Sensenbrenner LL, Jones RJ. Complete remission in severe aplastic anemia after high-dose cyclophosphamide without bone marrow transplantation. *Blood*. 1996;87(2):491-494.
72. Tisdale JF, Dunn DE, Geller N, et al. High-dose cyclophosphamide in severe aplastic anaemia: a randomised trial. *Lancet*. 2000;356(9241):1554-1559.
73. Gluckman E, Rokicka-Milewska R, Hann I, et al. Results and follow-up of a phase III randomized study of recombinant human-granulocyte stimulating factor as support for immunosuppressive therapy in patients with severe aplastic anaemia. *Br J Haematol*. 2002;119(4):1075-1082.
74. Tichelli A, de Latour RP, Passweg J, et al. Long-term outcome of a randomized controlled study in patients with newly diagnosed severe aplastic anemia treated with antithymocyte globulin and cyclosporine, with or without granulocyte colony-stimulating factor: a Severe Aplastic Anemia Working Party Trial from the European Group of Blood and Marrow Transplantation. *Haematologica*. 2020;105(5):1223-1231.
75. Bacigalupo A. How I treat acquired aplastic anemia. *Blood*. 2017;129(11):1428-1436.
76. Gurnari C, Pagliuca S, Prata PH, et al. Clinical and molecular determinants of clonal evolution in aplastic anemia and paroxysmal nocturnal hemoglobinuria. *J Clin Oncol*. 2023;41(1):132-142.
77. Khoury JD, Solary E, Abla O, et al. The 5th edition of the World Health Organization classification of haematolymphoid tumours: myeloid and histiocytic/dendritic neoplasms. *Leukemia*. 2022;36(7):1703-1719.
78. Groarke EM, Patel BA, Shalhoub R, et al. Predictors of clonal evolution and myeloid neoplasia following immunosuppressive therapy in severe aplastic anemia. *Leukemia*. 2022;36(9):2328-2337.
79. Kulasekararaj AG, Jiang J, Smith AE, et al. Somatic mutations identify a subgroup of aplastic anemia patients who progress to myelodysplastic syndrome. *Blood*. 2014;124(17):2698-2704.
80. Shah YB, Priore SF, Li Y, et al. The predictive value of PNH clones, 6p CN-LOH, and clonal TCR gene rearrangement for aplastic anemia diagnosis. *Blood Adv*. 2021;5(16):3216-3226.
81. Zaimoku Y, Katagiri T, Nakagawa N, et al. HLA class I allele loss and bone marrow transplantation outcomes in immune aplastic anemia. *Transplant Cell Ther*. 2024;30(3):281.e1-281.e13.
82. Narita A, Miwata S, Imaya M, et al. Minor PNH clones do not distinguish inherited bone marrow failure syndromes from immune-mediated aplastic anemia. *Blood Adv*. 2022;6(8):2517-2519.
83. Alder JK, Hanumanthu VS, Strong MA, et al. Diagnostic utility of telomere length testing in a hospital-based setting. *Proc Natl Acad Sci U S A*. 2018;115(10):E2358-E2365.
84. Fargo JH, Rochowski A, Giri N, Savage SA, Olson SB, Alter BP. Comparison of chromosome breakage in non-mosaic and mosaic patients with Fanconi anemia, relatives, and patients with other inherited bone marrow failure syndromes. *Cytogenet Genome Res*. 2014;144(1):15-27.
85. Gregory JJ, Wagner JE, Verlander PC, et al. Somatic mosaicism in Fanconi anemia: evidence of genotypic reversion in lymphohematopoietic stem cells. *Proc Natl Acad Sci U S A*. 2001;98(5):2532-2537.
86. Tomblyn M, Chiller T, Einsele H, et al. Guidelines for preventing infectious complications among hematopoietic cell transplantation recipients: a global perspective. *Biol Blood Marrow Transplant*. 2009;15(10):1143-1238.
87. Höchsmann B, Moicean A, Risitano A, Ljungman P, Schrezenmeier H. Supportive care in severe and very severe aplastic anemia. *Bone Marrow Transplant*. 2013;48(2):168-173.
88. DeZern AE, Brodsky RA. Clinical management of aplastic anemia. *Expert Rev Hematol*. 2011;4(2):221-230.
89. Valdez JM, Scheinberg P, Nunez O, Wu CO, Young NS, Walsh TJ. Decreased infection-related mortality and improved survival in severe aplastic anemia in the past two decades. *Clin Infect Dis*. 2011;52(6):726-735.
90. Taplitz RA, Kennedy EB, Bow EJ, et al. Antimicrobial prophylaxis for adult patients with cancer-related immunosuppression: ASCO and IDSA clinical practice guideline update. *J Clin Oncol*. 2018;36(30):3043-3054.
91. Oken MM, Creech RH, Tormey DC, et al. Toxicity and response criteria of the Eastern Cooperative Oncology Group. *Am J Clin Oncol*. 1982;5(6):649-655.
92. US National Library of Medicine. A trial comparing unrelated donor BMT with IST for pediatric and young adult patients with severe aplastic anemia (TransIT, BMT CTN 2202) (TransIT). Accessed 12 March 2024. <https://clinicaltrials.gov/ct2/show/NCT05600426>
93. DeZern AE, Zahurak M, Symons HJ, et al. Alternative donor BMT with posttransplant cyclophosphamide as initial therapy for acquired severe aplastic anemia. *Blood*. 2023;141(25):3031-3038.
94. Chang MH, Kim KH, Kim HS, et al. Predictors of response to immunosuppressive therapy with antithymocyte globulin and cyclosporine and prognostic factors for survival in patients with severe aplastic anemia. *Eur J Haematol*. 2010;84(2):154-159.

95. Kulagin A, Lisukov I, Ivanova M, et al. Prognostic value of paroxysmal nocturnal haemoglobinuria clone presence in aplastic anaemia patients treated with combined immunosuppression: results of two-centre prospective study. *Br J Haematol.* 2014;164(4):546-554.
96. Scheinberg P, Wu CO, Nunez O, Young NS. Predicting response to immunosuppressive therapy and survival in severe aplastic anaemia. *Br J Haematol.* 2009;144(2):206-216.
97. Narita A, Kojima S. Biomarkers for predicting clinical response to immunosuppressive therapy in aplastic anemia. *Int J Hematol.* 2016;104(2):153-158.
98. Fattizzo B, Ireland R, Dunlop A, et al. Clinical and prognostic significance of small paroxysmal nocturnal hemoglobinuria clones in myelodysplastic syndrome and aplastic anemia. *Leukemia.* 2021;35(11):3223-3231.
99. Blume KG, Krance RA. *The evaluation and counseling of candidates for hematopoietic cell transplantation. Thomas' Hematopoietic Cell Transplantation.* 1st ed. Wiley; 2009:443-460.
100. Scott BL, Sandmaier BM. The evaluation and counseling of candidates for hematopoietic cell transplantation. In: Forman SJ, Negrin RS, Antin JH, Appelbaum FR, eds. *Thomas' Hematopoietic Cell Transplantation: Stem Cell Transplantation.* 5th ed. John Wiley & Sons, Ltd; 2016:349-365. Accessed 4 January 2024. <https://onlinelibrary.wiley.com/doi/10.1002/9781118416426.ch29>
101. Sorror ML, Storb RF, Sandmaier BM, et al. Comorbidity-Age Index: A clinical measure of biologic age before allogeneic hematopoietic cell transplantation. *J. Clin. Oncol.* 2014 Oct 10;32(29):3249-56.
102. Fedele R, Salooja N, Martino M. Recommended screening and preventive evaluation practices of adult candidates for hematopoietic stem cell transplantation. *Expert Opin Biol Ther.* 2016;16(11):1361-1372.
103. Abel GA, Klepin HD. Frailty and the management of hematologic malignancies. *Blood.* 2018;131(5):515-524.
104. Sorror ML. How I assess comorbidities before hematopoietic cell transplantation. *Blood.* 2013;121(15):2854-2863.
105. Sorror ML, Maris MB, Storb R, et al. Hematopoietic cell transplantation (HCT)-specific comorbidity index: a new tool for risk assessment before allogeneic HCT. *Blood.* 2005;106(8):2912-2919.
106. Patel BA, Groarke EM, Lotter J, et al. Long-term outcomes in patients with severe aplastic anemia treated with immunosuppression and eltrombopag: a phase 2 study. *Blood.* 2022;139(1):34-43.
107. Dufour C, Svahn J, Bacigalupo A. on behalf of the Severe Aplastic Anemia–Working Party of the EBMT. Front-line immunosuppressive treatment of acquired aplastic anemia. *Bone Marrow Transplant.* 2013;48(2):174-177.
108. Passweg JR, Tichelli A. Immunosuppressive treatment for aplastic anemia: are we hitting the ceiling? *Haematologica.* 2009;94(3):310-312.
109. Babushok DV. Eltrombopag: a springboard to early responses in SAA. *Blood.* 2022;139(1):1-2.
110. Mitani K, Lee JW, Jang JH, et al. Long-term efficacy and safety of romiplostim in refractory aplastic anemia: follow-up of a phase 2/3 study. *Blood Adv.* 2024;8(6):1415-1419.
111. Jang JH, Tomiyama Y, Miyazaki K, et al. Efficacy and safety of romiplostim in refractory aplastic anaemia: a Phase II/III, multicentre, open-label study. *Br J Haematol.* 2021;192(1):190-199.
112. Lee JW, Lee SE, Jung CW, et al. Romiplostim in patients with refractory aplastic anaemia previously treated with immunosuppressive therapy: a dose-finding and long-term treatment phase 2 trial. *Lancet Haematol.* 2019;6(11):e562-e572.
113. Yamazaki H, Lee JW, Jang JH, et al. P827: efficacy and safety of romiplostim added to immunosuppressive therapy as a first-line treatment in patients with aplastic anemia: a phase 2/3 clinical trial. *Hemasphere.* 2022;6(S3):721-722.
114. Xu ZL, Xu LP, Wu DP, et al. Comparable long-term outcomes between upfront haploidentical and identical sibling donor transplant in aplastic anemia: a national registry-based study. *Haematologica.* 2022;107(12):2918-2927.
115. DeZern AE, Zahurak M, Jones RJ, Brodsky RA. Uniform conditioning regardless of donor in bone marrow transplantation for severe aplastic anemia. *Haematologica.* 2024;109(2):657-660.
116. DeZern AE, Zahurak M, Symons HJ, et al. Alternative donor BMT with posttransplant cyclophosphamide as initial therapy for acquired severe aplastic anemia. *Blood.* 2023;141(25):3031-3038.
117. Rogers ZR, Nakano TA, Olson TS, et al. Immunosuppressive therapy for pediatric aplastic anemia: a North American Pediatric Aplastic Anemia Consortium study. *Haematologica.* 2019;104(10):1974-1983.
118. National Comprehensive Cancer Network. NCCN guidelines for patients. Myelodysplastic Syndromes. Accessed 12 March 2024. <https://www.nccn.org/patients/guidelines/content/PDF/mds-patient.pdf>
119. Kallen ME, Dulau-Florea A, Wang W, Calvo KR. Acquired and germline predisposition to bone marrow failure: diagnostic features and clinical implications. *Semin Hematol.* 2019;56(1):69-82.
120. Keel SB, Scott A, Sanchez-Bonilla M, et al. Genetic features of myelodysplastic syndrome and aplastic anemia in pediatric and young adult patients. *Haematologica.* 2016;101(11):1343-1350.
121. Guyatt GH, Oxman AD, Kunz R, Vist GE, Falck-Ytter Y, Schünemann HJ. What is "quality of evidence" and why is it important to clinicians? *BMJ.* 2008; 336(7651):995-998.