

HHS Public Access

Author manuscript Nat Med. Author manuscript; available in PMC 2024 August 26.

Published in final edited form as:

Nat Med. 2022 June ; 28(6): 1110–1115. doi:10.1038/s41591-022-01836-w.

An early warning system for emerging SARS-CoV-2 variants

A full list of authors and affiliations appears at the end of the article.

Short standfirst summary

Global sequencing and surveillance capacity for SARS-CoV-2 must be strengthened and combined with multidisciplinary studies of infectivity, virulence, and immune escape, in order to track the unpredictable evolution of the ongoing COVID-19 pandemic.

> In June 2020, the World Health Organization (WHO) SARS-CoV-2 evolution working group was established to track SARS-CoV-2 variants and their specific genetic changes and to monitor virus characteristics and their impact on medical and non-medical countermeasures, including COVID-19 vaccines. In November 2021, this working group transitioned to a formal WHO Technical Advisory Group on Virus Evolution (TAG-VE), with the aim to develop and implement a global risk-monitoring framework for SARS-CoV-2 variants, based on a multidisciplinary approach that includes in silico, virological, clinical, and epidemiological data.

Tracking variants

The main role of the TAG-VE is to function as an integrative forum for exchange of information from global surveillance and research studies to monitor early warning signals, and perform timely assessment of the need for public health action in response to emerging variants¹. It uses a Delphi consensus method to establish which emerging variants are considered variants of interest (VOI) or variants of concern (VOC; see Ref 2 for definitions)². To avoid stigmatizing countries that first identify and report variants, a naming scheme that follows WHO guidelines³ and assigns Greek letters to VOIs and VOCs was adopted for global discourse in June 2021⁴. As of 26 March 2022, eight VOIs and fiveVOCs have been designated, and are further described as previously or currently circulating variants to reflect changes in their epidemiology over time 2.5 .

The first available data on emerging variants that are assessed by TAG-VE are viral sequences and their associated metadata shared on publicly accessible genetic sequence databases (e.g. GISAID, Genbank, European Nucleotide Archive, DNA Database of Japan).

Author contributions

Competing interests

[#] corresponding author: vankerkhovem@who.int.

L.S., M.P. K. and A.A. drafted the initial manuscript. A.v.G, B.B.O.M., L.J.A., X.A., E.A. B.N.A, H.A.C., A.B., W.S.B., J.N.B.,L.C., Me.C., Ma.C., A.C., T.d.O., C.D., J.D., P.E., I.E.M., A.F., S.G., E.G., R.G., Ba.H.,Be.H., S.I., Z.K., M.K., R.J.K., J.L., Y.S.L., G.M.L., M.M, S.M., J.M., N.M., V.M., K.N., D.O., B.I.P., T.P.P., M.P., Z.P., L.P., A.R., J.S., Y.S., M.S., S.K.T.,E.V.,V.T., S.v.d.W., S.B., M.D.P.,M.D.v.K. critically read and contributed to the manuscript. N.W., L.T. and S.R. produced the figures. senior authors

No financial and non-financial competing interests declared

Because all current VOIs and VOCs originate from ancestral variants, available sequences are still compared to the 2019 index virus (GISAID Accession ID: EPI_ISL_402124) with the primary focus on regions of the genome that are known to encode important viral, immunity- or infection-associated proteins. Mutations in the gene encoding the spike protein aregiven highest priority,, as this contains the receptor binding domain (RBD), essential for docking of the virus on host cells, as well as major determinants of both virus transmissibility (e.g. polybasic cleavage site) and antigenic make-up (e.g. N-terminal domain and RBD), and have therefore the highest prior probability of being clinically important. However, non-spike mutations have also been found to be important and should not be neglected. The constellation of mutations detected in any emerging variant is compared to a list of annotated mutations, which are suspected or known to play a role in one or more of the viral characteristics included in the VOI/VOC definitions: transmissibility, immune escape, disease severity, detectability and susceptibility to available treatments. Figure 1 depicts the evolution of SARS-CoV-2 and the variant-defining sets of spike protein amino acid substitutions seen in VOCs. To date, researchers have studied more than two dozen spike proteins, including those with phenotypes conferring immune escape (Figure 2). The list mutations and associated phenotypic impact, directly demonstrated in laboratory studies or inferred using *in silico* methods, is updated regularly, based on timely research presentations from the WHO partner network, as well as searches of preprints and peer-reviewed published literature.² This annotated list of mutations has proven useful, as the same mutations have independently arisen in different lineages of SARS-CoV-2, strongly suggesting viral adaptation to the human host and selective pressure from population immunity.

Importantly, the detection of a new constellation of mutations in a variant does not necessarily translate into an increased public health threat, suggesting that some mutations, or combinations of mutations, may impart a fitness cost, rather than benefit. For example, the VOI Theta had a constellation of mutations that alerted scientists because it included top-ranked mutations E484K, N501Y, D614G and P681H in the spike protein (Figure 2), but there were other uncommon mutations in the spike and outside the spike that may have limited its potential to spread. To establish whether a new variant poses a serious threat, TAG-VE looks for early epidemiological signals of spread and clinical signals derived from surveillance data or specific studies. This includes assessing measures of how quickly cases are increasing, in what geographical areas and population subgroups a variant emerges and spreads, as well as changes to disease severity indicators, such as hospitalization. Careful consideration is also given to assessing evidence of relative transmissibility, compared to other circulating variants, including secondary attack rates observed in household transmission studies. Finally, the assessment of the threat posed by a variant also needs to consider vaccine- or infection-derived population immunity, which has progressively grown to high levels in many countries, as well as become more complex in terms of different permutations of hybrid immunity from vaccines, infection, or both)⁶. Therefore, studies that look at prevalence of re-infections or vaccine breakthroughs are also reviewed, where available.

Access to samples

The experience gained since the launch of the virus evolution working group, later becoming the TAG-VE, has shown that the speed of spread of new SARS-CoV-2 variants can outpace our current ability to assess their threat. Variant sequence determination has now become part of the global surveillance, and systematic phenotypic characterization of emerging variants is a critical component that needs to be added to the surveillance core toolbox. It is therefore important to define gaps that need to be addressed to enable better, and more timely, response to new variant threats.

While genomic indicators for transmissibility and immune escape of VOCs are conceptually relatively straightforward to assess, the wide diversity of assays, the lack of centralized biobanks of viruses and clinical specimens, the difficulties in international shipping of materials, and the globally fragmented funding landscape, makes the comparability of data amassed in real-time challenging. To improve the global capacity to rapidly perform phenotypic characterization, virus isolates of emerging variants should be promptly generated and shared to enable researchers in different laboratories to work with viruses that carry the same constellation of mutations. To address this challenge, the WHO is developing a bio hub system and recently established a Biohub Facility⁷. Its aim is to offer a reliable, safe, and transparent mechanism for WHO Member States to voluntarily share novel biological materials, without replacing or competing with existing systems such as EVAg and the NIH BEI repository.

Sequence data

Another important challenge concerns the availability, representativeness, and quality of genetic sequence data, which vary depending on sample quality, processing, experience, and the platforms, protocols, analytical tools used, and access to all of these. This challenge is exemplified in the large bias in the volume of sequences contributed by a small number of countries to global databases, as well as the release of sequences with lowconfidence genomic regions and gaps.. For example, amplicon-based methods are sensitive to mutations, leading to amplification errors that can lead to sequence errors or poor-quality sequence in certain regions of the genome. The consequences of viral diversification call for sustained, if not enhanced, investment from governments in the capacity of reference laboratories to match the global public health demand for high quality sequences and virus characterisation for SARS-CoV-2, which can then be used for future public health threats posed by any infectious disease.

Verification of sequence quality through analysis of raw reads is a critical quality check, which becomes even more critical for the detection of recombination between SARS-CoV-2 genomes. Verified detection of SARS-CoV-2 circulating recombinant forms since Omicron emerged has increased, most likely because of increased availability of genomic surveillance, as well as natural factors. The higher number of lineage-defining mutations in Omicron makes the detection of recombinant forms easier, and re-infection with immune escape variants increases the chances of co-infection and therefore recombination, especially if two variants co-circulate in the community at high levels. While it is impossible to predict

whether circulating recombinant forms with specific genomic breakpoints $(i.e.$ where the two genomes recombine) may become more transmissible, for a new variant to spread widely, it must inherit traits from the parental viruses that provide a selective advantage.

Infectivity and virulence

Immune escape of VOCs can be shown through live virus neutralisation studies, with results from different laboratories showing similar results. In contrast, there are many methodological gaps in how to assess infectivity and virulence in vitro, ahead of clinical and epidemiological data. Some critical sites and amino acid substitutions have been identified, but this list is not comprehensive; these sites influence determinants of infectivity such as receptor binding, S1/S2 cleavage, and cellular entry (Table 1). In vivo animal studies can help understand specific features that may be difficult to estimate from epidemiologic data, such as assessing cross-neutralisation between variants, which is challenging in human subjects, as, unlike animal models, these may have unknown prior exposures. Animal models can also help assess virulence without the confounding effects of background immunity, but it remains to be seen how animal data correlate with disease severity in humans $^{8-11}$.

Disease severity in the clinical setting can be especially difficult to determine rapidly and accurately because of challenges posed by ascertainment, inclusion, quality of care, confounding and colliding biases from empirical observational studies. A key role will be played by the interoperability of electronic health records, which should allow easy linkage of sequencing data with patient clinical records. Unbiased, systematic clinical and epidemiological data collection with comprehensive biological sampling and rigorous characterization of virulence and correlates of protection remain the gold standard for variant threat assessment. The generation of such data from well-designed studies across diverse healthcare settings and different geographies is expected to remain challenging, especially once the COVID-19 response is de-escalated in the post-acute phase of the pandemic. Integration of such studies into newly established routine surveillance system workplans may help ensure that clinical and epidemiological studies continue in the future.

Until November 2021, the emergence of VOCs such as Alpha and Delta was mainly associated with an increase in transmissibility, most likely driven by viral adaptation to the human host, with modest degrees of immune escape¹². The emerging evidence on Omicron variants suggests that immune escape was a significant driver for its observed displacement over Delta, and its selective advantage was driven by increasing population immunity, in addition to the increased transmissibility¹³. While the lower virulence of Omicron was most likely a chance event, the preservation of cellular immunity that protects against severe infection is likely to be a recurring theme^{14,15}. In addition, so-called hybrid immunity in individuals who have experienced a breakthrough infection appears to broaden immune responses, which suggests that population immunity may be able to tolerate significant continued evolution in SARS-CoV-2. 16,17 However, because transmission and virulence are uncoupled for SARS-CoV-2, it cannot be assumed that the next variant will be less virulent. Future variants may be of similar, higher or lower virulence than Omicron.

An early warning system

A recent retrospective analysis suggested that some of the key variant-defining mutations could potentially have been detected much earlier, showing the importance of early warning bioinformatics tools from globally shared data, an approach in its infancy.¹⁸ The identification of key mutations could then trigger a subsequent virus-, variant- or mutationcharacterization pipeline. Machine learning algorithms to determine the potential impact of key mutations are being developed and validated, but a full assessment of variants still require epidemiological data and in vivo and /or in vitro experiments, as a VOC cannot be ascertained solely using genomic data.

The future of the pandemic is difficult to predict, for several reasons. Firstly, SARS-CoV-2 variants do not always emerge from the most recently dominant circulating virus, unlike other respiratory viruses such as human influenza viruses.19. Secondly, chronically infected patients can allow the acceleration of intra-host evolution $2⁰$. Thirdly, a wide number of susceptible mammals may act as secondary reservoirs, including cervids, mustelids and rodents, with potential for reverse zoonoses 21 . Fourthly, too much of the world's population remains unvaccinated to date, and current vaccines are suboptimal in preventing transmission (future vaccines may remain so). Given these challenges in predicting the evolution of the virus, targeted surveillance of suspected high-risk populations, such as chronically infected patients, or improving detection systems such as wastewater and a One Health approach to animal surveillance are critical for the early detection of future variants.

The evolving virus and the uncertainty of predicting the trajectory of the pandemic calls for strengthened surveillance and continued monitoring of SARS-CoV-2. The TAG-VE will continue to critically appraise state-of-the-art methodologies towards predicting further evolution of SARS-CoV-2 and to rapidly determine the threat levels posed by new variants. The pandemic is not over and SARS-CoV-2 is spreading at a high level globally. Now is the time to enhance global sequencing capacities, focusing on widening coverage to include previous geographical and populations' blind spots, and build a global consensus towards continued concerted multidisciplinary efforts, under the leadership of the WHO Research & Development Blueprint for action to prevent epidemics, to track and assess the threat posed by future SARS-CoV-2 variants.

Authors

Lorenzo Subissi¹, Anne von Gottberg^{2,3}, Lipi Thukral⁴, Nathalie Worp⁵, Bas B. Oude Munnink⁵, Surabhi Rathore⁴, Laith J. Abu-Raddad⁶, Ximena Aguilera⁷, Erik Alm⁸, Brett N. Archer¹, Homa Attar Cohen¹, Amal Barakat⁹, Wendy S. Barclay¹⁰, Jinal N. Bhiman^{2,3}, Leon Caly¹¹, Meera Chand¹², Mark Chen¹³, Ann Cullinane¹⁴, Tulio De Oliveira¹⁵, Christian Drosten¹⁶, Julian Druce⁷, Paul Effler¹⁷, Ihab El Masry¹⁸, Adama Faye¹⁹, Simani Gaseitsiwe²⁰, Elodie Ghedin²¹, Rebecca Grant¹, Bart Haagmans⁵, Belinda L. Herring²², Shilpa Iyer²³, Zyleen Kassamali¹, Manish Kakkar²⁴, Rebecca J. Kondor²⁵, Juliana Leite²⁶, Yee Sin Leo¹³, Gabriel M. Leung²⁷, Marco Marklewitz²⁸, Sikhulile Moyo²⁰, Jairo Mendez-Rico²⁶, Nada Melhem²⁹, Vincent Munster³⁰, Karen Nahapetyan²⁸, Djin-Ye Oh³¹, Boris I. Pavlin¹, Thomas

P. Peacock^{10,12}, Malik Peiris²⁷, Zhibin Peng³², Leo Poon²⁷, Andrew Rambaut³³, Jilian Sacks¹, Yinzhong Shen³⁴, Marilda M. Siqueira³⁵, Sofonias K. Tessema³⁶, Erik Volz¹⁰, Volker Thiel^{37,38}, Sylvie van der Werf³⁹, Sylvie Briand¹, Mark D. Perkins¹, Maria D. Van Kerkhove^{1,#,*}, Marion P.G. Koopmans^{5,*}, Anurag Agrawal^{4,*}

Affiliations

1.World Health Organization, Geneva, Switzerland

2.National Institute for Communicable Diseases, Johannesburg, South Africa,

3.School of Pathology, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa,

4.CSIR Institute of Genomics and Integrative Biology, Delhi, India

5.Erasmus Medical Centre, Rotterdam, The Netherlands, and Pandemic and Disaster Preparedness Research Centre, Rotterdam/Delft, The Netherlands

6.Weill Cornell Medicine – Qatar of Cornell University, Doha, Qatar

7.Epidemiology and Health Policy Centre, Universidad del Desarollo, Santiago, Chile

8.European Centre for Disease Prevention and Control, Stockholm, Sweden

9. World Health Organization Regional Office for the Eastern Mediterranean, Cairo, Egypt

10.Imperial College London, London, United Kingdom

11.Victorian Infectious Diseases Reference Laboratory (VIDRL), Melbourne, Australia

12.UK Health Security Agency, London, United Kingdom

13.National Centre for Infectious Diseases, Singapore, Singapore

14.World Organisation for Animal Health, Paris, France

15.Centre for Epidemic Response and Innovation (CERI), Stellenbosch University, Stellenbosch, South Africa

16. Charité Medical Center, Berlin, Germany

17.University of Western Australia, Perth, Australia

18. Food and Agriculture Organization, Rome, Italy

19.Health and Development Institute, Dakar, Senegal

20.Botswana Harvard AIDS Institute Partnership, Gaborone, Botswana

21.National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, United States of America

22.World Health Organization Regional Office for Africa, Brazzaville, Republic of Congo

Philippines

24.World Health Organization Regional Office for South-East Asia, New Delhi, India

25.United States Centers for Disease Control and Prevention, Atlanta, United States of America

26.World Health Organization Regional Office for the Americas, Washington, United States of America

^{27.}School of Public Health, LKS Faculty of Medicine, The University of Hong Kong, Hong Kong SAR, The People's Republic of China

28.World Health Organization Regional Office for Europe, Copenhagen, Denmark

29.American University of Beirut, Beirut, Lebanon

³⁰ National Institute of Allergy and Infectious Diseases, National Institutes of Health, Rocky Mountain Laboratories, Hamilton, Montana, United States of America

31.Robert Koch Institute, Berlin, Germany,

³² Chinese Center for Disease Control and Prevention, Beijing, The People's Republic of China

33.University of Edinburgh, Edinburgh, United Kingdom

³⁴ Fudan University, Shanghai, The People's Republic of China

35.Oswaldo Cruz Institute, Fiocruz, Rio de Janeiro, Brazil

36.Africa Centers for Disease Control and Prevention, Addis Ababa, Ethiopia

³⁷ Multidisciplinary Center for Infectious Diseases, University of Bern, Bern, **Switzerland**

38. Institute of Virology and Immunology, Mittelhäusern and Bern, Switzerland

39.Institut Pasteur, Université Paris Cité, Paris, France

Acknowledgments

We acknowledge scientists, public health professionals and Ministries of Health across the world for early generation and sharing of data on SARS-CoV-2 variants.

References

- 1. World Health Organization. Terms of Reference for the Technical Advisory Group on SARS-CoV-2 Virus Evolution (TAG-VE).
- 2. World Health Organization. Tracking SARS-CoV-2 variants. [https://www.who.int/en/activities/](https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/) [tracking-SARS-CoV-2-variants/](https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/).
- 3. World Health Organization Best Practices for the Naming of New Human Infectious Diseases. [https://apps.who.int/iris/bitstream/handle/10665/163636/WHO_HSE_FOS_15.1_eng.pdf.](https://apps.who.int/iris/bitstream/handle/10665/163636/WHO_HSE_FOS_15.1_eng.pdf)
- 4. Konings F et al. Nat Microbiol 6, 821–823 (2021). [PubMed: 34108654]
- 5. World Health Organization. Weekly Epidemiological Updates. [https://www.who.int/emergencies/](https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports) [diseases/novel-coronavirus-2019/situation-reports.](https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports)

Nat Med. Author manuscript; available in PMC 2024 August 26.

- doi:10.1101/2021.12.14.21267791.
- 7. World Health Organization. WHO Biohub System. [https://www.who.int/initiatives/who-biohub.](https://www.who.int/initiatives/who-biohub)
- 8. CMMID COVID-19 Working Group et al. Nature 593, 270–274 (2021). [PubMed: 33723411]
- 9. Twohig KA et al. The Lancet Infectious Diseases 22, 35–42 (2022). [PubMed: 34461056]
- 10. Munster VJ et al. Sci. Adv 7, eabj3627 (2021). [PubMed: 34678071]
- 11. Muñoz-Fontela C et al. PLoS Pathog 18, e1010161 (2022). [PubMed: 35025969]
- 12. Oude Munnink BB et al. Nat Med 27, 1518–1524 (2021). [PubMed: 34504335]
- 13. Viana R et al. Nature (2022) doi:10.1038/s41586-022-04411-y.
- 14. Gao Y et al. Nat Med (2022) doi:10.1038/s41591-022-01700-x.
- 15. Keeton R et al. Nature (2022) doi:10.1038/s41586-022-04460-3.
- 16. Kitchin D et al. Cell Reports Medicine 100535 (2022) doi:10.1016/j.xcrm.2022.100535.
- 17. He W et al. <http://biorxiv.org/lookup/doi/10.1101/2021.09.08.459480> (2021) doi:10.1101/2021.09.08.459480.
- 18. Maher MC et al. Sci. Transl. Med 14, eabk3445 (2022). [PubMed: 35014856]
- 19. Jones S et al. Sci Rep 9, 14690 (2019). [PubMed: 31604969]
- 20. Corey L et al. N Engl J Med 385, 562–566 (2021). [PubMed: 34347959]
- 21. Oude Munnink BB et al. Science 371, 172–177 (2021). [PubMed: 33172935]

Nat Med. Author manuscript; available in PMC 2024 August 26.

Author Manuscript

Figure 1. Overview of VOCs and spike amino acid substitutions.

The spike structure of the index virus is shown, together with key domains, including N-terminal domain (NTD), Receptor Binding Domain (RBD), the S1/S2 junction (including the polybasic cleavage site), and the heptapeptide repeat domains (HR1, HR2) highlighted in different colours. The five VOCs and the characterising amino acids are marked with blue lines, both on the spike protein schematic and protein structure. VOCs are plotted chronologically according to date of identification.

Figure 2. Schematic of spike amino acid substitutions with known impact.

The circular plot shows key amino acid substitutions and their associated lineage, and presence in N-terminal domain, RBD, and S1/S2 polybasic cleavage site. The red are marked to emphasize mutations involved in immune evasion. The complete list of mutations is shown in Table 1.

Nat Med. Author manuscript; available in PMC 2024 August 26.

 Author ManuscriptAuthor Manuscript

Author Manuscript

Author Manuscript

Τ

Т

Nat Med. Author manuscript; available in PMC 2024 August 26.

Abbreviations: mAbs - monoclonal antibodies; NAbs - neutralising antibodies. NTD - N-terminal domain; RBD - Receptor Binding Domain; ACE2 - Angiotensin converting enzyme 2 Abbreviations: mAbs – monoclonal antibodies; NAbs – neutralising antibodies. NTD – N-terminal domain; RBD – Receptor Binding Domain; ACE2 – Angiotensin converting enzyme 2

ı

Г

Т

Author Manuscript

Author Manuscript

Author Manuscript