

Review

# P-tau217 as a Reliable Blood-Based Marker of Alzheimer's Disease

Roy Lai <sup>1</sup>, Brenden Li <sup>1</sup>  and Ram Bishnoi <sup>2,3,4,\*</sup>

<sup>1</sup> Morsani College of Medicine, University of South Florida, Tampa, FL 33612, USA; bli20@usf.edu (B.L.)

<sup>2</sup> Department of Psychiatry and Behavioral Neurosciences, University of South Florida, Tampa, FL 33613, USA

<sup>3</sup> USF Health Byrd Alzheimer's Center and Research Institute, Tampa, FL 33613, USA

<sup>4</sup> USF Memory Disorder Clinic, Tampa, FL 33613, USA

\* Correspondence: bishnoi@usf.edu

**Abstract:** Amyloid plaques and tau tangles are the hallmark pathologic features of Alzheimer's disease (AD). Traditionally, these changes are identified in vivo via cerebrospinal fluid (CSF) analysis or positron emission tomography (PET) scans. However, these methods are invasive, expensive, and resource-intensive. To address these limitations, there has been ongoing research over the past decade to identify blood-based markers for AD. Despite the challenges posed by their extremely low concentrations, recent advances in mass spectrometry and immunoassay techniques have made it feasible to detect these blood markers of amyloid and tau deposition. Phosphorylated tau (p-tau) has shown greater promise in reflecting amyloid pathology as evidenced by CSF and PET positivity. Various isoforms of p-tau, distinguished by their differential phosphorylation sites, have been recognized for their ability to identify amyloid-positive individuals. Notable examples include p-tau181, p-tau217, and p-tau235. Among these, p-tau217 has emerged as a superior and reliable marker of amyloid positivity and, thus, AD in terms of accuracy of diagnosis and ability for early prognosis. In this narrative review, we aim to elucidate the utility of p-tau217 as an AD marker, exploring its underlying basis, clinical diagnostic potential, and relevance in clinical care and trials.

**Keywords:** Alzheimer's disease; biomarkers; tau; p-tau217; clinical utility



**Citation:** Lai, R.; Li, B.; Bishnoi, R. P-tau217 as a Reliable Blood-Based Marker of Alzheimer's Disease.

*Biomedicines* **2024**, *12*, 1836.

<https://doi.org/10.3390/biomedicines12081836>

Academic Editors: Simone Battaglia and Masaru Tanaka

Received: 17 July 2024

Revised: 8 August 2024

Accepted: 9 August 2024

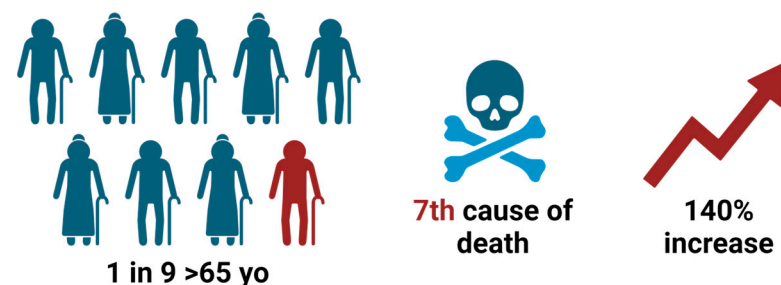
Published: 13 August 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder characterized by amyloid plaques and neurofibrillary tangles [1]. As the most common cause of dementia, AD affects 1 in 9 Americans aged 65 and older and was the seventh leading cause of death in the United States in 2021 [2] (Figure 1). While deaths from stroke and heart disease decreased from 2000 to 2021, deaths from AD increased more than 140% during this period, and AD is projected to affect 13.8 million Americans by 2060 [2] (Figure 1). Given that AD begins 20 years or more before symptom onset, there is a substantial window in which medical interventions may alter the disease progression [2]. Reliable biomarkers are needed for early detection of preclinical AD to delay or prevent the development of clinical AD.



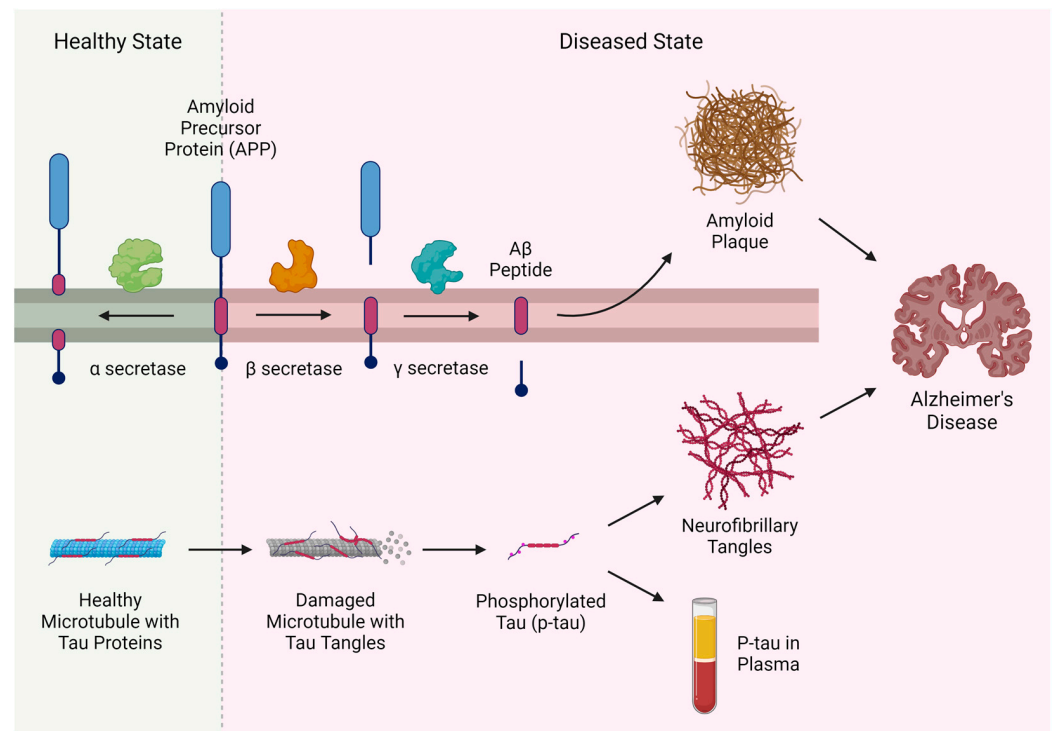
**Figure 1.** The significance of Alzheimer's disease. Created with BioRender.com.

Although the ultimate confirmation of AD diagnosis is based on postmortem examination of brain tissue for amyloid plaques consisting of amyloid-beta ( $A\beta$ ) peptides and neurofibrillary tangles containing phosphorylated tau (p-tau), *in vivo* diagnosis can be achieved using cerebrospinal fluid (CSF) or neuroimaging biomarkers [1,3]. However, these methods are limited by the invasiveness of lumbar punctures and the high cost and specialized facilities required for positron emission tomography (PET) imaging [4]. In contrast, blood-based biomarkers are easily accessible and available at a fraction of the cost of neuroimaging, which allows the expansion of access to AD biomarker research and clinical care [4,5]. Among various plasma amyloid and tau markers, phosphorylated tau at threonine 217 (p-tau217) is one of the emerging tau markers that has repeatedly shown more promise than other p-tau isoforms as a marker of AD pathology. This review evaluates the evidence for p-tau217 as a peripheral biomarker of AD, compares it to other phosphorylated tau biomarkers, explores its relationship with imaging methods, and examines its utility in AD diagnosis and prognosis.

Current literature overwhelmingly focuses on another p-tau isoform, p-tau181. A literature search of articles published between 2020 and 2023 revealed 458 publications mentioning p-tau 181 and only 69 publications mentioning p-tau217. However, there have been studies suggesting that p-tau217 is superior to p-tau181 in comparisons such as correlation with tau-PET measures and differentiating between AD and non-AD states [6,7]. Thus, it appears that p-tau217 may be an understudied and underappreciated potential clinical tool for AD care. This study aims to address this discrepancy in focus by synthesizing current knowledge on p-tau217 by asking how p-tau217 compares to other AD biomarkers in performance. The hypothesis is that p-tau217 performs at parity, if not better, than p-tau181 and other p-tau isoforms.

## 2. AD Pathology and Biomarkers

AD is characterized by the presence of extracellular  $A\beta$  plaques and intracellular neurofibrillary tangles, composed of proteins formed by the aberrant cleavage of transmembrane amyloid precursor protein (APP) and microtubule-associated tau protein, respectively [1,8] (Figure 2). The major peptide in amyloid plaques is the amyloid  $\beta$  peptide ( $A\beta$ ) which, as a hydrophobic molecule, is prone to aggregation [9].  $A\beta$  is a proteolytic fragment of APP, which is a transmembrane protein commonly found within neurons and involved in synapse generation [10]. APP can be cleaved in three distinct locations by various secretases which either produce  $A\beta$  or prevent its generation.  $\beta$ -secretase cleaves APP at the N-terminal of the  $A\beta$  domain, while  $\gamma$ -secretase cleaves it at the C-terminal of the  $A\beta$  domain [10]. Therefore, cleavage of APP by  $\beta$ - and  $\gamma$ -secretase produces the insoluble  $A\beta$  peptide that can accumulate to form plaques [10,11]. In contrast,  $\alpha$ -secretase cleaves APP within the  $A\beta$  domain and thus prevents the formation of complete  $A\beta$  and protects against AD [10,11]. Amyloid plaques are thought to disrupt synaptic communication, as well as trigger a local inflammatory response [12,13]. On the other hand, neurofibrillary tangles are thought to result from the breakdown of neuron microtubules [14]. Tau is a protein associated with microtubules that becomes hyperphosphorylated in the event of AD, forming p-tau [14]. In this form, p-tau dissociates from microtubules, aggregates intracellularly, and weakens microtubules, which in turn leads to a breakdown of axonal transport and even neuronal apoptosis [15]. The soluble building blocks of  $A\beta$  plaques and neurofibrillary tangles interact synergistically to drive neuronal death and synaptic dysfunction [1]. A growing body of evidence has suggested that  $A\beta$  functions upstream of tau in AD pathogenesis while tau exacerbates  $A\beta$  toxicity [16–18]. Given that soluble toxic  $A\beta$  and tau spread throughout the brain by prion-like mechanisms, with misfolded proteins converting their harmless counterparts into pathological forms, detection of these soluble peptides before they become observable plaques and tangles is essential for the early diagnosis and interventions of AD pathogenesis [19].



**Figure 2.** The pathophysiological process of Alzheimer's disease. Created with BioRender.com.

AD has historically been diagnosed based on cognitive and behavioral symptoms, but it suffers from poor correlation between clinical and postmortem neuropathological diagnosis [20,21]. More recently, AD has been classified using the AT(N) system based on core AD pathophysiological features, namely A $\beta$  (A), tau (T), and neurodegeneration (N) [22,23]. A $\beta$  markers include A $\beta$ -PET and CSF A $\beta$ 42 or the ratio of A $\beta$ 42 to A $\beta$ 40 (A $\beta$ 42/A $\beta$ 40), while tau markers include tau-PET and CSF tau phosphorylated at various sites [23]. Markers of neurodegeneration include CSF total tau (t-tau), CSF neurofilament light chain (NfL), and hippocampal atrophy and/or cortical thinning determined by magnetic resonance imaging [23]. Newer studies have shown that blood-based markers may serve as less invasive and more affordable alternatives to neuroimaging and CSF biomarkers.

### 3. Tau as a Biomarker

Hyperphosphorylation of tau proteins is a characteristic pathological change in AD. There are many sites on the tau protein that may be phosphorylated at various disease stages from preclinical AD to mild cognitive impairment due to AD (MCI-AD) to clinical AD [24–27]. P-tau217, p-tau181, and p-tau231 have all been observed in neurofibrillary tangles and have been detected around A $\beta$  plaques in mice with A $\beta$  plaques without neurofibrillary tangles but not in mice with elevated A $\beta$  without A $\beta$  plaques, suggesting that these p-tau molecules are markers of postsynaptic pathology around A $\beta$  plaques [28]. P-tau181 has been extensively studied and has seen clinical applications such as the use of CSF p-tau181 in the prediction of cognitive decline in patients with MCI [29]. However, p-tau217 has been suggested to perform better as a biomarker than p-tau181, with p-tau217 being more closely associated with AD severity [6]. P-tau217 accumulation may differentiate AD from related diseases, as higher p-tau217 has been observed in the brains of those with AD than those with primary age-related tauopathy or other non-AD tauopathies [30]. In addition, the spread of p-tau217 in postmortem brain tissue has shown a correlation with antemortem plasma p-tau217 in those with amyloid plaques but not in those without, suggesting that plasma p-tau217 may reflect the presence of A $\beta$  plaques [30].

### *Tau in Distinct Brain Regions*

Hyperphosphorylation of tau has also been observed to show a differential association with neuronal loss from distinct brain regions. For example, nucleus basalis of Meynert (nbM) neuronal density was not correlated with p-tau concentrations, while lower locus coeruleus neuronal density correlated with higher p-tau217 [31]. In addition, norepinephrine depletion has been associated with increased p-tau217 and dose-dependent working memory deficits [32]. Together, these findings suggest that norepinephrine function may play a role in hyperphosphorylation of tau [32]. In the nbM, increased APP has been found to correlate with neurofibrillary tangle formation [33]. Furthermore, it has been found that 59% of the variability in plasma p-tau217 in the nbM can be predicted by a combination of global neuropathologic scales, especially tau and A $\beta$ . This contrasts with just 31% of plasma p-tau181 variability being predicted by the same scales [31]. These data suggest that in the nbM, tau and amyloid  $\beta$  pathological processes are closely related to p-tau217 formation.

P-tau217 is also unique among tau biomarkers in that it has been found to be persistently associated with the CA1 region of the hippocampus, particularly within neurofibrillary tangles and granulovacuolar bodies (GVBs) [30]. GVBs are prominent among pyramidal neurons in the hippocampus and associate with the severity of tauopathies and A $\beta$  pathology, spreading with tau pathology from the entorhinal cortex to the neocortex, hypothalamus, amygdala, and frontal and parietal cortices [34]. GVBs also correlate directly with neuronal cell loss and neurofibrillary tangle formation [34,35]. Taken together, p-tau217 may serve as a strong indicator of early-stage AD changes in the brain and may be more specific and better suited for clinical use than other tau biomarkers.

## **4. P-tau217 in CSF and Plasma**

### *4.1. CSF P-tau217*

The wide array of tau markers with different phosphorylation sites requires the identification of a specific p-tau biomarker that exhibits the highest accuracy in AD. CSF studies have suggested that p-tau217 may be the best tau biomarker for AD [36–39]. Levels of p-tau217, p-tau181, and p-tau231 in CSF have all been observed to increase in both prodromal AD and AD dementia [36–38]. Among these, p-tau217 showed the greatest change between non-AD controls and AD patients, with a 13-fold increase compared to a 1.9- to 5.4-fold increase for the other p-tau variants [36]. CSF p-tau217 has also shown a correlation with markers of AD, including medial temporal lobe atrophy, cognitive performance, and A $\beta$  burden [40]. Of all p-tau analytes, p-tau217 showed the strongest correlation with A $\beta$ -PET and tau-PET status, two highly reliable neuroimaging methods used to evaluate AD [36,41]. Overall, p-tau217 had the highest accuracy in distinguishing AD from non-AD as well as predicting A $\beta$ -PET and tau-PET positivity [36,38,39]. The higher increase in CSF levels and its strong correlation with other measures of AD suggest that CSF p-tau217 may be a better AD biomarker than other CSF p-tau biomarkers.

### *4.2. Plasma P-tau217*

Plasma p-tau217 has also been demonstrated as a robust and accurate biomarker for AD with the advent of improvements in detection technology. Several p-tau isoforms like p-tau217, p-tau181, p-tau205, p-tau231, and p-tau235 have been observed in plasma in clinically, radiologically, and pathologically diagnosed AD [42–47]. In addition to their increased concentrations, plasma p-tau217, p-tau181, and p-tau231 all showed correlation with amyloid-PET status and predicted amyloid positivity [44,47–49]. Certain p-tau species such as 217 and 181 showed strong correlations in their CSF and plasma levels [47,50]. This correlation has been the strongest in p-tau217, suggesting that plasma p-tau217 may serve as a less invasive alternative to its counterpart in CSF [47,50]. Similar to its levels in CSF, plasma p-tau217 showed a greater increase than that of p-tau181, 4.2 and 1.7 fold, respectively, in amyloid-positive individuals [43]. Additionally, plasma p-tau217 detected amyloid and tau pathology with comparable accuracy to CSF p-tau217, while

plasma p-tau181 and p-tau231 performed worse when predicting amyloid-PET status with respect to their CSF counterparts [51,52].

Studies comparing different plasma tau isoforms have suggested that p-tau217 may have the strongest predictive value due to its better dynamic range, discrimination of tau-PET positivity, and diagnostic performance compared to other p-tau biomarkers [53]. Stronger correlations have been reported between amyloid- and tau-PET status and plasma p-tau217 than any other isoform [41,46,54]. P-tau217 better identified amyloid-positivity and distinguished tau-positivity with higher accuracy [26,41,43,46,55]. P-tau217 has been shown to discriminate high and intermediate amyloid-PET scans with higher accuracy than p-tau181 [56]. Furthermore, p-tau217 was found to contribute to the identification of both amyloid and tau status, whereas p-tau231 was only found to contribute to the detection of tau positivity [57]. Notably, p-tau217 alone was noninferior to combining p-tau217 with A $\beta$ 42/A $\beta$ 40 when predicting A $\beta$  positivity [58]. In summary, plasma p-tau217 has been shown to be noninferior to CSF p-tau217 while being superior to other plasma p-tau isoforms in the identification of A $\beta$ - and tau-PET status.

## 5. P-tau217 as an AD Biomarker

### 5.1. Clinical AD

In addition to correlations with amyloid- and tau-PET status, plasma p-tau217 has been shown to differentiate pathologically diagnosed AD from non-AD with significantly higher accuracy than p-tau181, with an area under the curve (AUC) of 0.88–0.90 compared to p-tau181's AUC of 0.72–0.81 [26,59,60]. Only plasma p-tau217 was able to differentiate between all levels of AD neuropathological change (none, low, intermediate, and high), while p-tau181 and p-tau231 only differentiated between intermediate and high AD neuropathology [60]. Higher concentrations of p-tau217 in antemortem plasma and CSF as well as postmortem brain samples have been reported in clinical AD and MCI compared to cognitively unimpaired (CU) individuals [40,61]. Among six phosphorylation sites, antemortem plasma p-tau217 showed the greatest increase in autopsy-confirmed AD compared to non-AD participants and p-tau217 showed the strongest association with both Braak and Consortium to Establish a Registry for Alzheimer's Disease (CERAD) stages [59].

Limited data exist on the p-tau217 levels in atypical or non-amnesic presentations of AD such as logopenic variant primary progressive aphasia (lvPPA) or posterior cortical atrophy (PCA). One study found an average plasma p-tau217 concentration of  $0.72 \pm 0.4$  pg/mL in the amnesic presentation of AD,  $0.82 \pm 0.2$  pg/mL in lvPPA, and  $0.80 \pm 0.3$  pg/mL in PCA compared to  $0.17 \pm 0.1$  pg/mL in normal controls [46]. While no statistical tests were performed to compare the p-tau217 levels in different AD presentations, it appears that lvPPA and PCA show elevated p-tau217, similar to the typical presentation of AD.

### 5.2. Preclinical AD

Compared to other p-tau isoforms, p-tau217 showed larger increases at earlier disease stages, with notable elevations starting at PET-based Braak stage II and plateauing at stage IV [24,47]. In contrast, p-tau181 demonstrates greater increases at later stages [24]. P-tau217 levels increase before detectable tau aggregation, and p-tau217 can distinguish amyloid-positive tau-negative individuals from non-AD controls [47]. This early rise in p-tau217 is hypothesized to be due to its unique close association with the CA1 region of the hippocampus and localization with GVBs [30]. The formation of GVBs is driven by neuronal damage and neurofibrillary tangle formation. Interestingly, GVBs have been found to participate in exosomal secretion, disposing of cellular waste, such as p-tau217, extracellularly [62]. It is hypothesized that the secretion of GVBs in response to early-stage AD neurofibrillary tangle formation contributes to p-tau217's unique early staging [30].

The early rise in p-tau217 levels during the course of AD renders it an important biomarker of preclinical AD. For example, in an inherited form of AD, p-tau217 levels were found to rise 21 years before symptom onset, while p-tau181 and p-tau205 levels



increased 19 and 13 years, respectively, before symptom onset [25]. Elevated p-tau217 has also been reported in preclinical AD up to 20 years before MCI-AD onset [26,27]. In CU individuals, plasma p-tau217 correlates with amyloid- and tau-PET status as well as with CSF A $\beta$ 42/A $\beta$ 40 [58,63]. Additionally, p-tau217 independently identified A $\beta$  status and was not influenced by demographic factors in the detection of A $\beta$  positivity in CU individuals [27,57]. These findings underscore the potential of p-tau217 as a highly sensitive and specific biomarker for the early detection, diagnosis, and monitoring of preclinical AD.

### 5.3. Role of P-tau217 in AD Prognosis

The early rise of p-tau217, prior to detectable tau aggregation or symptom onset, also makes it a promising candidate for the monitoring of AD progression. Baseline p-tau217 has been found to be associated with longitudinal A $\beta$ -PET increases in those without overt A $\beta$  pathology at baseline [44]. In addition, p-tau217, but not p-tau181 or p-tau231, showed an annual increase only in A $\beta$ -positive individuals, with the greatest change in A $\beta$ -positive tau-positive individuals [52,64]. With disease progression, p-tau217 area fraction has been shown to correlate strongly with amyloid-beta and neurofibrillary tangle brain load [30].

Not only is p-tau217 associated with amyloid plaque and neurofibrillary tangle accumulation, but it has also been shown to correlate with cognitive measures in both clinical and preclinical AD [38,39,63]. Among A $\beta$ -positive participants, p-tau217 predicted global cognition, memory, language, executive function, visuospatial skills, and cortical thickness with higher accuracy than p-tau181 [65]. Longitudinal increases in p-tau217 have shown an association with worsening cognition and brain atrophy [66].

P-tau217 may be useful in the prediction of progression from preclinical AD or MCI to clinical AD. Both higher baseline p-tau217 and more rapid longitudinal increase in p-tau217 levels have shown an association with future development of clinical AD in individuals with MCI [66–68]. In contrast, no longitudinal change in p-tau217 levels was observed in those with MCI who did not later develop clinical AD [66]. The combination of p-tau217 with memory, executive function, and APOE genotype has been shown to further increase the predictive accuracy of progression from MCI to clinical AD [68]. Among those with MCI, A $\beta$ -positive participants have been found to have an accelerated increase in p-tau217 compared to A $\beta$ -negative individuals, who did not show a longitudinal change in p-tau217 levels [66].

Among CU individuals, associations have been found between p-tau217 and worse clinical progression, as well as decreased cortical thickness [64,65]. A more rapid longitudinal increase in p-tau217 has been observed in A $\beta$ -positive CU participants compared to A $\beta$ -negative CU participants, who showed no significant longitudinal change in p-tau217 levels [66]. In CU individuals, p-tau217 was associated with an increased risk of MCI development in A $\beta$ -positive participants, while no such association was observed in their A $\beta$ -negative counterparts [69]. Across Braak stages, lower postmortem p-tau217 was associated with better baseline memory [70]. In contrast, higher p-tau217 at preclinical stages was associated with worse cognitive trajectories [63]. Taken together, these results suggest that higher baseline levels and faster longitudinal increases in p-tau217 are associated with more severe A $\beta$  pathology and cognitive impairments while also predicting worse clinical progression.

### 5.4. Role of P-tau217 in Differentiating AD from Non-AD Dementia

Tau biomarkers have been shown to differentiate tauopathies [71]. CSF p-tau has been observed to increase in AD but not in non-AD dementias including progressive supranuclear palsy and corticobasal syndrome [4,72,73]. In addition, individuals with Lewy body dementia showed lower CSF t-tau and p-tau levels than those with AD [74]. Similarly, lower t-tau and higher p-tau to t-tau ratios (p/t-tau) have been observed in frontotemporal dementia (FTD) than in AD [75]. In fact, FTD is associated with lower p/t-tau compared to non-AD controls [76].

P-tau217 has also shown utility in distinguishing AD from other neurodegenerative diseases [26,67]. Reduced plasma p-tau217 extracellular vesicles (EVs) have been observed in AD compared to non-AD controls while smaller p-tau217 EVs have been found in AD compared to non-AD dementia [77]. A model combining patient age with number and size of EVs carrying p-tau217 discriminated AD from non-AD controls and non-AD dementia with high accuracy [77]. While elevated p-tau217 has been observed in AD and other neurodegenerative diseases, p-tau217 distinguished AD from disorders such as primary progressive aphasia (specifically the nonfluent and semantic variants), frontotemporal dementia, progressive supranuclear palsy, and corticobasal degeneration with 90% accuracy, while p-tau181 showed a 78% accuracy [78]. In another study, p-tau217 performed better than p-tau181 when discriminating clinically diagnosed and pathology-confirmed AD from frontotemporal lobar degeneration [46].

## 6. Screening Assays for P-tau Analytes

Blood-based biomarkers for AD have generally been preferred to ones in CSF because of their accessibility. Plasma A $\beta$ 42/A $\beta$ 40 is correlated with pathologically identified A $\beta$  in the brain, while plasma NfL is associated with neurodegeneration in AD [60,79]. Plasma p-tau is especially promising due to its large increase in AD, strong correlation with established diagnostic methods, and ability to differentiate AD from other neurodegenerative disorders [4,53,67,80]. However, biomarkers of AD are inherently more abundant in the CSF due to its constant contact with the brain compared to biomarkers in plasma, leading to the historic difficulty of blood-based AD biomarker development [60]. Nevertheless, technological developments in recent decades, such as immunoprecipitation and mass spectrometry, and improvements in enzyme-linked immunosorbent assays (ELISA) have greatly increased the accuracy and viability of blood-based biomarkers.

P-tau levels detected via ELISA had been the mainstay of p-tau measurements, with CSF p-tau231 having a reported sensitivity of 90.2% and specificity and 80.0% in discriminating between AD and non-AD disorders [81]. With advances in immunoassay technology, the sensitivity and specificity were increased in CSF p-tau217 to 91% and 91% for discriminating between AD and non-AD disorders [6]. Plasma biomarkers of AD have been inferior to the CSF and neuroimaging methods due to technological limitations of assays to quantify significantly lower concentrations of markers in plasma [80].

Developments within the last decade have increasingly turned to mass spectrometry to quantify p-tau levels. Barthelemy et al. utilized mass spectrometry combined with nano-flow capillary liquid chromatography to analyze both plasma and CSF p-tau peptides [47,82]. Indeed, a comparison between multiple immunoassay methods and the mass spectrometry method outlined by Barthelemy found that mass spectrometry performed significantly better [83]. Mass spectrometry performed better than immunoassay methods in detecting abnormal A $\beta$  status, progression to AD, and showed the strongest correlation between plasma and CSF p-tau [83]. With regard to p-tau217, mass spectrometry was found to have the best performance out of any other p-tau biomarker in detecting abnormal A $\beta$  status and had strong predictive value for identifying patients with MCI and abnormal A $\beta$  levels, and those who progressed to AD [83]. Taken together, these results imply that mass spectrometry methods for p-tau detection, specifically p-tau217, may have strong future roles in AD detection and prognosis, edging out older immunoassay methods in terms of predictive value and accuracy.

Therriault et al. found that recent new developments in ultrasensitive immunoassays for p-tau perform comparably or better than previously described liquid chromatography-mass spectrometry methods [84]. Diagnostic performance between the two methods was similar for p-tau217, but mass spectrometry for p-tau181 and p-tau231 was inferior to immunoassay methods with regard to association with amyloid-PET and tau-PET [84].

More recently, a novel chemiluminescent enzyme immunoassay (CLEIA) to detect p-tau analytes has been developed. While ELISA measures optical density, CLEIA measures relative light units. In comparison, CLEIA methods have higher sensitivity and are less

time-consuming compared to ELISA but are more costly [85]. This, too, is true for p-tau: CLEIA detection of p-tau181 in CSF was found to be more accurate than methods using ELISA [86]. CLEIA kits for p-tau217 have been developed and have shown high accuracy in discerning AD from other neurodegenerative diseases [87]. Literature-reported sensitivities, specificities, and AUC for discerning between AD and non-AD controls for p-tau analytes in plasma and CSF can be found in Table 1. With recent advances, ELISA, mass spectrometry, and CLEIA methods have demonstrated consistent, similar testing characteristics across all analytes. However, p-tau217 tests perform notably better than those measuring p-tau181 or p-tau231, once more suggesting its superiority in utility for clinical AD testing.

In summary, developments in mass spectrometry and immunoassay methods have created near parity in their diagnostic performance, notably in the detection of p-tau217, with each having strengths and limitations. Mass spectrometry is able to provide absolute quantification of p-tau levels, whereas immunoassays cannot [84]. Furthermore, mass spectrometry is able to quantify multiple analytes, at the cost of decreased sensitivity, whereas immunoassays measure only a single prespecified analyte [80]. The ability to measure and quantify various p-tau analytes may prove vital to the prognosis and measurement of disease progression. As previously mentioned, distinct p-tau markers have been found to exist at different concentrations depending on the timescale of disease, and the ability to examine all levels of p-tau in one single assay may provide the best clinical picture [88]. Should a clinician wish to view p-tau217 levels specifically, immunoassay methods may be chosen instead to mitigate the slower result times and higher machine costs of mass spectrometry. Although CLEIA methods for p-tau are more recent compared to established ELISA and mass spectrometry assays, they represent a possibly quicker and more accurate alternative, although their current cost and novelty may hold them back from general use. For the widespread adoption of p-tau as a tool for AD diagnosis and monitoring in the clinic, however, universal cutoffs for p-tau levels are still required [46]. Additional research is needed to establish these values for clinical use.

**Table 1.** Reported sensitivities, specificities, and AUC of ELISA, mass spectrometry, and CLEIA detection methods for the p-tau analytes p-tau181, p-tau217, and p-tau231.

	ELISA	Mass Spectrometry	CLEIA
p-tau181 (plasma)	Sen: 86.6% [89] Spe: 80.0% [89] AUC: 0.889 [89]	Sen: No data Spe: No data AUC: 0.98 [47]	Sen: No data Spe: No data AUC: 0.910 [90]
p-tau217 (plasma)	Sen: 95% [91] Spe: 94% [91] AUC: 0.98 [91]	Sen: No data Spe: No data AUC: 0.98 [47]	Sen: No data Spe: No data AUC: 0.952 [87]
p-tau231 (plasma)	Sen: 81.2% [5] Spe: 93.3% [5] AUC: 0.94 [5]	Sen: No data Spe: No data AUC: 0.834 [59]	Sen: No data Spe: No data AUC: No data
p-tau181 (CSF)	Sen: 91.8% [89] Spe: 90.5% [89] AUC: 0.954 [89]	Sen: No data Spe: No data AUC: 0.95 [47]	Sen: 90% [86] Spe: 90% [86] AUC: 0.98 [86]
p-tau217 (CSF)	Sen: No data Spe: No data AUC: 0.95 [41]	Sen: No data Spe: No data AUC: 1.00 [47]	Sen: No data Spe: No data AUC: No data
p-tau231 (CSF)	Sen: No data Spe: No data AUC: 0.88 [41]	Sen: No data Spe: No data AUC: 0.9873 [92]	Sen: No data Spe: No data AUC: No data

## 7. Role of P-tau217 in Clinical Trials

Plasma p-tau217 may help improve future clinical trials by enhancing participant selection, distinguishing progressors from non-progressors, and monitoring the effects of



treatments. Plasma p-tau217 may be useful for the identification of preclinical AD among CU individuals. Given its strong correlation with tau-PET status, A $\beta$ -PET status, and CSF p-tau concentrations, plasma p-tau217 may be used to pre-screen clinical trial participants so that only individuals with high plasma p-tau217 receive PET scans and/or CSF collections. Though this may help to reduce cost and the number of invasive procedures during screening, clinical trials on anti-tau and anti-A $\beta$  treatments generally need to monitor target engagement throughout the study, so PET scans may still be necessary during enrollment. The TRAILBLAZER-ALZ 3 (NCT05026866) trial in a large sample of CU older adults is unique in being the first to use plasma p-tau217 as the only enrollment criterion, while A $\beta$ -PET is not performed in any part of enrollment or treatment monitoring [93].

Given the slow progression from preclinical to clinical AD, identification of individuals who will later develop clinical AD versus those who will remain stable may be important to shorten clinical trials and reduce the sample size needed to detect the effects of potential medications. Plasma p-tau217 has shown promise in its ability to predict cognitive decline and progression to clinical AD in both MCI and CU individuals and therefore may serve another important role in the selection of clinical trial participants.

Considering the ease of plasma biomarker collection, p-tau217 may also be used to monitor the effects of anti-tau and anti-A $\beta$  treatments more frequently. In the concluded TRAILBLAZER-ALZ (NCT03367403) trial, a significant decrease in plasma p-tau217 was observed compared with placebo after just 12 weeks of treatment and continued throughout the 76-week duration of the study [49]. Furthermore, lower plasma p-tau217 levels correlated with the reduction in tau- and A $\beta$ -PET in the treatment group, suggesting that plasma p-tau217 may be useful for monitoring brain neurofibrillary tangles and amyloid plaque accumulation during treatment [94,95].

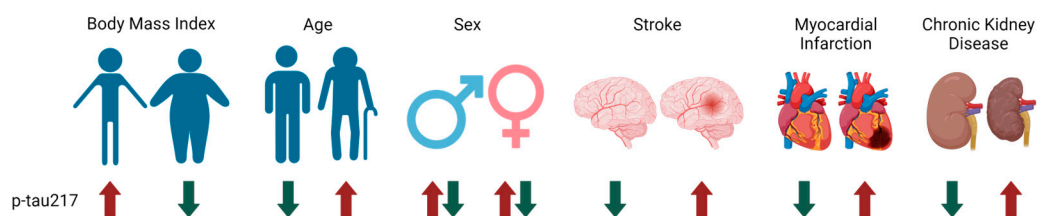
In addition to its role in enrollment and monitoring of treatments, p-tau217 itself may serve as a potential target for treatment in the future. For example, monoclonal antibodies against p-tau217 have been found to reduce tau aggregation, block apoptosis-associated neuronal loss, decrease brain atrophy, reverse cognitive deficits, and improve motor function in tauopathic mice [40].

## 8. Challenges and Limitations

Plasma p-tau levels may show variations within and between individuals based on factors unrelated to AD, such as body mass index (BMI), age, or sex, which poses a potential limitation in their utility in AD diagnosis and prognosis (Figure 3). A study found a 10.3% within-subject variation in p-tau217, while p-tau231 showed a 6.3% variation and p-tau181 showed a 16.7% variation [96]. Similarly, p-tau217, p-tau231, and p-tau181 varied by 21.1%, 17.2%, and 25.7%, respectively, among healthy individuals [96]. Higher p-tau217 has been observed in those with lower BMI [50]. When comparing biomarker levels among participants aged 30 to 98 years, an increase in plasma p-tau217 with age can be observed starting at ages 65 to 69 [97]. However, when participants were grouped by amyloid-PET status, the increase in p-tau217 with age was 12 times faster in amyloid-positive than amyloid-negative participants [97]. Mixed results have been reported regarding the relationship between sex and p-tau217 levels. Among CU individuals, males showed significantly higher p-tau217 than females, whereas males and females in the MCI or dementia groups showed no differences in p-tau217 levels [97]. No sex differences were found in the increase in p-tau217 with age in participants with or without *PSEN1* mutations, the most common cause of familial AD [98,99]. However, female *PSEN1* mutation carriers showed higher p-tau217 than male carriers [98]. In CU *PSEN1* mutation carriers, males had worse cognitive performance than females as p-tau217 concentrations increased, while no interaction between age and sex was observed in cognitively impaired mutation carriers [98].

Certain comorbidities such as stroke, myocardial infarction (MI), or chronic kidney disease (CKD) may also increase the concentrations of plasma p-tau and therefore decrease their utility in the diagnosis of AD (Figure 3). Increased p-tau217 levels have been reported in individuals with a history of stroke compared to those without [97]. Similarly,

those with a history of MI have shown higher p-tau217 than those without a history of MI [97]. Reduced kidney function and CKD were associated with increased p-tau217 in both cognitively impaired and unimpaired individuals [50,100]. More alarmingly, p-tau level differences between CKD and non-CKD individuals have been observed to be comparable to differences between amyloid-positive and amyloid-negative participants [97]. Adjusting for creatinine and BMI, however, has been found to improve the discrimination by plasma p-tau217 of stable CU and MCI individuals from those who later develop AD, especially in individuals with higher creatinine and BMI [50]. In addition, the ratio of phosphorylated to unphosphorylated tau (pT217/T217) could more accurately reflect pathological changes in AD. pT217/T217 was not correlated with kidney dysfunction in those with MCI and only weakly correlated in CU individuals [100]. Furthermore, in both MCI and CU cohorts, pT217/T217 showed a much larger difference between amyloid-positive and amyloid-negative participants than between CKD and non-CKD individuals [100]. Therefore, adjusting for creatinine and BMI or the use of pT217/T217 may serve to mitigate the variations in plasma p-tau217 due to factors unrelated to AD.



**Figure 3.** The effects of various demographic factors and comorbidities on plasma p-tau217 concentrations. The up arrows (↑) indicate an increase in levels, the down arrows (↓) point to a decrease in levels, and a combination of up and down arrows (↑↓) suggests an indeterminate effect. Created with BioRender.com.

Another factor of the viability of p-tau217’s use as a clinical tool will be its cost. As per the Brown University Center for Alzheimer’s Disease Research, a current Simoa® plasma p-tau217 assay is listed to cost USD 194 commercially. Comparisons of prices with other assays as well as imaging modalities can be found in Table 2. It must be noted that the prices of these assays do not include various extraneous hospital fees that may be incurred by the patient. Prices, especially for imaging studies, undoubtedly fluctuate greatly between institutions as well. Despite these limitations, the cost of plasma and CSF biomarker determination is still much lower than that of an amyloid-PET scan. As Alzheimer’s testing technology advances, it is hoped that the cost to the healthcare system and to the patient will decrease.

**Table 2.** Selected reference costs of AD laboratory tests and imaging. Simoa® and SMC™ assays are usable on both plasma and CSF samples. HD-X: HD-X Automated Immunoassay Analyzer; GFAP: glial fibrillated acidic protein; NF-light: neurofilament light.

Procedure	Cost
Simoa® pTau217 for HD-X	USD 194 [101]
Simoa® pTau181 for HD-X	USD 172 [101]
Simoa® pTau231 for HD-X	USD 141 [101]
Simoa® GFAP for HD-X	USD 145 [101]
Simoa® Neurofilament Light (NfL) for HD-X	USD 207 [101]
Simoa® Neurology 4-Plex E for HD-X (AB40, AB42, GFAP, NF-light)	USD 401 [101]
Simoa® Neurology 3-Plex A for HD-X (AB40, AB42, Tau)	USD 338 [101]
SMC™ Amyloid Beta 40	USD 137 [101]
SMC™ Amyloid Beta 42	USD 137 [101]
Amyloid-PET scan	USD 6487 [102]

## 9. Conclusions

Hyperphosphorylation of tau is a hallmark pathological change in AD present around A $\beta$  plaques and in neurofibrillary tangles. Among the many phosphorylated tau biomarkers, p-tau217 has attracted special attention in recent years due to its utility in AD diagnosis and prognosis. Its increase in the early stages of AD makes p-tau217 a useful biomarker for the diagnosis and prognosis of both clinical and preclinical AD. Elevated p-tau217 can be observed more than 20 years prior to symptom onset and is strongly associated with later diagnosis of AD. The field of p-tau detection and quantification has developed rapidly in the past several years, making less invasive and more cost-effective blood biomarkers for AD increasingly viable. Research on plasma p-tau217 has shown it to be an accurate and unique biomarker for AD and AD progression. However, further research is needed for its formal adoption as a clinical tool.

**Author Contributions:** Conceptualization, R.B.; methodology, literature search, R.L. and B.L.; writing—original draft preparation, R.L. and B.L.; writing—review and editing, R.L., B.L. and R.B.; visualization, R.L. and B.L.; supervision, R.B. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## Abbreviations

A $\beta$	amyloid-beta
A $\beta$ 42/A $\beta$ 40	ratio of A $\beta$ 42 to A $\beta$ 40
AD	Alzheimer's disease
APP	amyloid precursor protein
AUC	area under the curve
BMI	body mass index
CERAD	Consortium to Establish a Registry for Alzheimer's Disease
CKD	chronic kidney disease
CLEIA	chemiluminescent enzyme immunoassay
CSF	cerebrospinal fluid
CU	cognitively unimpaired
ELISA	enzyme-linked immunosorbent assay
EV	extracellular vesicle
FTD	frontotemporal dementia
GVB	granulovacuolar body
lvPPA	logopenic variant primary progressive aphasia
MCI-AD	mild cognitive impairment due to AD
MI	myocardial infarction
nbM	nucleus basalis of Meynert
NfL	neurofilament light chain
PCA	posterior cortical atrophy
PET	positron emission tomography
p-tau	phosphorylated tau
p-tau217	phosphorylated tau at threonine 217
p/t-tau	p-tau to t-tau ratio
pT217/T217	ratio of phosphorylated to unphosphorylated tau
t-tau	total tau

## References

1. Bloom, G.S. Amyloid- $\beta$  and Tau: The Trigger and Bullet in Alzheimer Disease Pathogenesis. *JAMA Neurol.* **2014**, *71*, 505. [[CrossRef](#)] [[PubMed](#)]
2. 2024 Alzheimer's Disease Facts and Figures. *Alzheimer's Dement.* **2024**, *20*, 3708–3821. [[CrossRef](#)]
3. Nelson, P.T.; Alafuzoff, I.; Bigio, E.H.; Bouras, C.; Braak, H.; Cairns, N.J.; Castellani, R.J.; Crain, B.J.; Davies, P.; Tredici, K.D.; et al. Correlation of Alzheimer Disease Neuropathologic Changes With Cognitive Status: A Review of the Literature. *J. Neuropathol. Exp. Neurol.* **2012**, *71*, 362–381. [[CrossRef](#)] [[PubMed](#)]
4. Gonzalez-Ortiz, F.; Kac, P.R.; Brum, W.S.; Zetterberg, H.; Blennow, K.; Karikari, T.K. Plasma Phospho-Tau in Alzheimer's Disease: Towards Diagnostic and Therapeutic Trial Applications. *Mol. Neurodegener.* **2023**, *18*, 18. [[CrossRef](#)] [[PubMed](#)]
5. Pais, M.V.; Forlenza, O.V.; Diniz, B.S. Plasma Biomarkers of Alzheimer's Disease: A Review of Available Assays, Recent Developments, and Implications for Clinical Practice. *J. Alzheimer's Dis. Rep.* **2023**, *7*, 355–380. [[CrossRef](#)] [[PubMed](#)]
6. Janelidze, S.; Stomrud, E.; Smith, R.; Palmqvist, S.; Mattsson, N.; Airey, D.C.; Proctor, N.K.; Chai, X.; Shcherbinin, S.; Sims, J.R.; et al. Cerebrospinal Fluid P-Tau217 Performs Better than p-Tau181 as a Biomarker of Alzheimer's Disease. *Nat. Commun.* **2020**, *11*, 1683. [[CrossRef](#)] [[PubMed](#)]
7. Wildsmith, K.R.; Barthélemy, N.R.; Bohorquez, S.S.; Manser, P.T.; Teng, E.; Ward, M.; Keeley, M.; Bateman, R.J.; Weimer, R. O3-14-02: Tau Burden Measured Using [18F]GTP1 Correlates with CSF TAU Phosphorylation at Sites T217 and T205 More Closely than T181. *Alzheimer's Dement.* **2018**, *14*, P1059–P1060. [[CrossRef](#)]
8. Hampel, H.; Hardy, J.; Blennow, K.; Chen, C.; Perry, G.; Kim, S.H.; Villemagne, V.L.; Aisen, P.; Vendruscolo, M.; Iwatsubo, T.; et al. The Amyloid- $\beta$  Pathway in Alzheimer's Disease. *Mol. Psychiatry* **2021**, *26*, 5481–5503. [[CrossRef](#)]
9. Lichtenthaler, S.F. Alpha-Secretase in Alzheimer's Disease: Molecular Identity, Regulation and Therapeutic Potential: Alpha-Secretase in Alzheimer's Disease. *J. Neurochem.* **2011**, *116*, 10–21. [[CrossRef](#)]
10. MacLeod, R.; Hillert, E.-K.; Cameron, R.T.; Baillie, G.S. The Role and Therapeutic Targeting of  $\alpha$ -,  $\beta$ - and  $\gamma$ -Secretase in Alzheimer's Disease. *Future Sci. OA* **2015**, *1*, FSO11. [[CrossRef](#)]
11. Chow, V.W.; Mattson, M.P.; Wong, P.C.; Gleichmann, M. An Overview of APP Processing Enzymes and Products. *Neuromol. Med.* **2010**, *12*, 1–12. [[CrossRef](#)]
12. Karisetty, B.C.; Bhatnagar, A.; Armour, E.M.; Beaver, M.; Zhang, H.; Elefant, F. Amyloid- $\beta$  Peptide Impact on Synaptic Function and Neuroepigenetic Gene Control Reveal New Therapeutic Strategies for Alzheimer's Disease. *Front. Mol. Neurosci.* **2020**, *13*, 577622. [[CrossRef](#)]
13. Kinney, J.W.; Bemiller, S.M.; Murtishaw, A.S.; Leisgang, A.M.; Salazar, A.M.; Lamb, B.T. Inflammation as a Central Mechanism in Alzheimer's Disease. *AD Transl. Res. Clin. Interv.* **2018**, *4*, 575–590. [[CrossRef](#)] [[PubMed](#)]
14. Metaxas, A.; Kempf, S. Neurofibrillary Tangles in Alzheimer's Disease: Elucidation of the Molecular Mechanism by Immunohistochemistry and Tau Protein Phospho-Proteomics. *Neural Regen. Res.* **2016**, *11*, 1579. [[CrossRef](#)] [[PubMed](#)]
15. Beharry, C.; Cohen, L.S.; Di, J.; Ibrahim, K.; Briffa-Mirabella, S.; Alonso, A.D.C. Tau-Induced Neurodegeneration: Mechanisms and Targets. *Neurosci. Bull.* **2014**, *30*, 346–358. [[CrossRef](#)] [[PubMed](#)]
16. Götz, J.; Chen, F.; Van Dorpe, J.; Nitsch, R.M. Formation of Neurofibrillary Tangles in P301L Tau Transgenic Mice Induced by A $\beta$ 42 Fibrils. *Science* **2001**, *293*, 1491–1495. [[CrossRef](#)] [[PubMed](#)]
17. Lewis, J.; Dickson, D.W.; Lin, W.-L.; Chisholm, L.; Corral, A.; Jones, G.; Yen, S.-H.; Sahara, N.; Skipper, L.; Yager, D.; et al. Enhanced Neurofibrillary Degeneration in Transgenic Mice Expressing Mutant Tau and APP. *Science* **2001**, *293*, 1487–1491. [[CrossRef](#)] [[PubMed](#)]
18. Leroy, K.; Ando, K.; Laporte, V.; Dedecker, R.; Suain, V.; Authelet, M.; Héraud, C.; Pierrot, N.; Yilmaz, Z.; Octave, J.-N.; et al. Lack of Tau Proteins Rescues Neuronal Cell Death and Decreases Amyloidogenic Processing of APP in APP/PS1 Mice. *Am. J. Pathol.* **2012**, *181*, 1928–1940. [[CrossRef](#)]
19. Nussbaum, J.M.; Seward, M.E.; Bloom, G.S. Alzheimer Disease: A Tale of Two Prions. *Prion* **2013**, *7*, 14–19. [[CrossRef](#)]
20. McKhann, G.; Drachman, D.; Folstein, M.; Katzman, R.; Price, D.; Stadlan, E.M. Clinical Diagnosis of Alzheimer's Disease: Report of the NINCDS-ADRDA Work Group under the Auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* **1984**, *34*, 939–944. [[CrossRef](#)]
21. Kovacs, G.G.; Milenkovic, I.; Wöhrer, A.; Höftberger, R.; Gelpi, E.; Haberler, C.; Hönigschnabl, S.; Reiner-Concin, A.; Heinzl, H.; Jungwirth, S.; et al. Non-Alzheimer Neurodegenerative Pathologies and Their Combinations Are More Frequent than Commonly Believed in the Elderly Brain: A Community-Based Autopsy Series. *Acta Neuropathol.* **2013**, *126*, 365–384. [[CrossRef](#)] [[PubMed](#)]
22. Jack, C.R.; Bennett, D.A.; Blennow, K.; Carrillo, M.C.; Feldman, H.H.; Frisoni, G.B.; Hampel, H.; Jagust, W.J.; Johnson, K.A.; Knopman, D.S.; et al. A/T/N: An Unbiased Descriptive Classification Scheme for Alzheimer Disease Biomarkers. *Neurology* **2016**, *87*, 539–547. [[CrossRef](#)] [[PubMed](#)]
23. Hampel, H.; Cummings, J.; Blennow, K.; Gao, P.; Jack, C.R.; Vergallo, A. Developing the ATX(N) Classification for Use across the Alzheimer Disease Continuum. *Nat. Rev. Neurol.* **2021**, *17*, 580–589. [[CrossRef](#)] [[PubMed](#)]
24. Therriault, J.; Pascoal, T.A.; Lussier, F.Z.; Tissot, C.; Chamoun, M.; Bezgin, G.; Servaes, S.; Benedet, A.L.; Ashton, N.J.; Karikari, T.K.; et al. Biomarker Modeling of Alzheimer's Disease Using PET-Based Braak Staging. *Nat. Aging* **2022**, *2*, 526–535. [[CrossRef](#)] [[PubMed](#)]

25. The Dominantly Inherited Alzheimer Network; Barthélemy, N.R.; Li, Y.; Joseph-Mathurin, N.; Gordon, B.A.; Hassenstab, J.; Benzinger, T.L.S.; Buckles, V.; Fagan, A.M.; Perrin, R.J.; et al. A Soluble Phosphorylated Tau Signature Links Tau, Amyloid and the Evolution of Stages of Dominantly Inherited Alzheimer's Disease. *Nat. Med.* **2020**, *26*, 398–407. [[CrossRef](#)] [[PubMed](#)]
26. Palmqvist, S.; Janelidze, S.; Quiroz, Y.T.; Zetterberg, H.; Lopera, F.; Stomrud, E.; Su, Y.; Chen, Y.; Serrano, G.E.; Leuzy, A.; et al. Discriminative Accuracy of Plasma Phospho-Tau217 for Alzheimer Disease vs Other Neurodegenerative Disorders. *JAMA* **2020**, *324*, 772–781. [[CrossRef](#)] [[PubMed](#)]
27. Suárez-Calvet, M.; Karikari, T.K.; Ashton, N.J.; Lantero Rodríguez, J.; Milà-Alomà, M.; Gispert, J.D.; Salvadó, G.; Minguillon, C.; Fauria, K.; Shekari, M.; et al. Novel Tau Biomarkers Phosphorylated at T181, T217 or T231 Rise in the Initial Stages of the Preclinical Alzheimer's Continuum When Only Subtle Changes in A $\beta$  Pathology Are Detected. *EMBO Mol. Med.* **2020**, *12*, e12921. [[CrossRef](#)] [[PubMed](#)]
28. Hirota, Y.; Sakakibara, Y.; Ibaraki, K.; Takei, K.; Iijima, K.M.; Sekiya, M. Distinct Brain Pathologies Associated with Alzheimer's Disease Biomarker-Related Phospho-Tau 181 and Phospho-Tau 217 in *App* Knock-in Mouse Models of Amyloid- $\beta$  Amyloidosis. *Brain Commun.* **2022**, *4*, fca286. [[CrossRef](#)] [[PubMed](#)]
29. Hansson, O. Biomarkers for Neurodegenerative Diseases. *Nat. Med.* **2021**, *27*, 954–963. [[CrossRef](#)]
30. Wennström, M.; Janelidze, S.; Nilsson, K.P.R.; The Netherlands Brain Bank; Serrano, G.E.; Beach, T.G.; Dage, J.L.; Hansson, O. Cellular Localization of P-Tau217 in Brain and Its Association with p-Tau217 Plasma Levels. *Acta Neuropathol. Commun.* **2022**, *10*, 3. [[CrossRef](#)]
31. Murray, M.E.; Moloney, C.M.; Kouri, N.; Syrjanen, J.A.; Matchett, B.J.; Rothberg, D.M.; Tranovich, J.F.; Sirmans, T.N.H.; Wiste, H.J.; Boon, B.D.C.; et al. Global Neuropathologic Severity of Alzheimer's Disease and Locus Coeruleus Vulnerability Influences Plasma Phosphorylated Tau Levels. *Mol. Neurodegener.* **2022**, *17*, 85. [[CrossRef](#)] [[PubMed](#)]
32. Prinzi, C.; Kostenko, A.; De Leo, G.; Gulino, R.; Leanza, G.; Caccamo, A. Selective Noradrenaline Depletion in the Neocortex and Hippocampus Induces Working Memory Deficits and Regional Occurrence of Pathological Proteins. *Biology* **2023**, *12*, 1264. [[CrossRef](#)]
33. Murphy, G.M.; Greenberg, B.D.; Ellis, W.G.; Forno, L.S.; Salamat, S.M.; Gonzalez-DeWhitt, P.A.; Lowery, D.E.; Tinklenberg, J.R.; Eng, L.F. Alzheimer's Disease. Beta-Amyloid Precursor Protein Expression in the Nucleus Basalis of Meynert. *Am. J. Pathol.* **1992**, *141*, 357–361.
34. Thal, D.R.; Del Tredici, K.; Ludolph, A.C.; Hoozemans, J.J.M.; Rozemuller, A.J.; Braak, H.; Knippschild, U. Stages of Granulovacuolar Degeneration: Their Relation to Alzheimer's Disease and Chronic Stress Response. *Acta Neuropathol.* **2011**, *122*, 577–589. [[CrossRef](#)]
35. Koper, M.J.; Van Schoor, E.; Ospitalieri, S.; Vandenbergh, R.; Vandenbulcke, M.; Von Arnim, C.A.F.; Tousseyn, T.; Balusu, S.; De Strooper, B.; Thal, D.R. Necrosome Complex Detected in Granulovacuolar Degeneration Is Associated with Neuronal Loss in Alzheimer's Disease. *Acta Neuropathol.* **2020**, *139*, 463–484. [[CrossRef](#)]
36. Leuzy, A.; Janelidze, S.; Mattsson-Carlsson, N.; Palmqvist, S.; Jacobs, D.; Cicognola, C.; Stomrud, E.; Vanmechelen, E.; Dage, J.L.; Hansson, O. Comparing the Clinical Utility and Diagnostic Performance of CSF P-Tau181, P-Tau217, and P-Tau231 Assays. *Neurology* **2021**, *97*, e1681–e1694. [[CrossRef](#)] [[PubMed](#)]
37. Karikari, T.K.; Emeršič, A.; Vrillon, A.; Lantero-Rodríguez, J.; Ashton, N.J.; Kramberger, M.G.; Dumurgier, J.; Hourregue, C.; Čučnik, S.; Brinkmalm, G.; et al. Head-to-head Comparison of Clinical Performance of CSF Phospho-tau T181 and T217 Biomarkers for Alzheimer's Disease Diagnosis. *Alzheimer's Dement.* **2021**, *17*, 755–767. [[CrossRef](#)] [[PubMed](#)]
38. Barthélemy, N.R.; Toth, B.; Manser, P.T.; Sanabria-Bohórquez, S.; Teng, E.; Keeley, M.; Bateman, R.J.; Weimer, R.M.; Wildsmith, K.R. Site-Specific Cerebrospinal Fluid Tau Hyperphosphorylation in Response to Alzheimer's Disease Brain Pathology: Not All Tau Phospho-Sites Are Hyperphosphorylated. *J. Alzheimer's Dis.* **2022**, *85*, 415–429. [[CrossRef](#)]
39. Meeker, K.L.; Luckett, P.H.; Barthélemy, N.R.; Hobbs, D.A.; Chen, C.; Bollinger, J.; Ovod, V.; Flores, S.; Keefe, S.; Henson, R.L.; et al. Comparison of Cerebrospinal Fluid, Plasma and Neuroimaging Biomarker Utility in Alzheimer's Disease. *Brain Commun.* **2024**, *6*, fcae081. [[CrossRef](#)]
40. Zhang, D.; Zhang, W.; Ming, C.; Gao, X.; Yuan, H.; Lin, X.; Mao, X.; Wang, C.; Guo, X.; Du, Y.; et al. P-Tau217 Correlates with Neurodegeneration in Alzheimer's Disease, and Targeting p-Tau217 with Immunotherapy Ameliorates Murine Tauopathy. *Neuron* **2024**, *112*, 1676–1693.e12. [[CrossRef](#)]
41. Mendes, A.J.; Ribaldi, F.; Lathuiliere, A.; Ashton, N.J.; Janelidze, S.; Zetterberg, H.; Scheffler, M.; Assal, F.; Garibotto, V.; Blennow, K.; et al. Head-to-Head Study of Diagnostic Accuracy of Plasma and Cerebrospinal Fluid p-Tau217 versus p-Tau181 and p-Tau231 in a Memory Clinic Cohort. *J. Neurol.* **2024**, *271*, 2053–2066. [[CrossRef](#)] [[PubMed](#)]
42. Brickman, A.M.; Manly, J.J.; Honig, L.S.; Sanchez, D.; Reyes-Dumeyer, D.; Lantigua, R.A.; Lao, P.J.; Stern, Y.; Vonsattel, J.P.; Teich, A.F.; et al. Plasma P-tau181, P-tau217, and Other Blood-based Alzheimer's Disease Biomarkers in a Multi-ethnic, Community Study. *Alzheimer's Dement.* **2021**, *17*, 1353–1364. [[CrossRef](#)] [[PubMed](#)]
43. Arranz, J.; Zhu, N.; Rubio-Guerra, S.; Rodríguez-Baz, Í.; Ferrer, R.; Carmona-Iragui, M.; Barroeta, I.; Illán-Gala, I.; Santos-Santos, M.; Fortea, J.; et al. Diagnostic Performance of Plasma pTau 217, pTau 181, A $\beta$  1-42 and A $\beta$  1-40 in the LUMIPULSE Automated Platform for the Detection of Alzheimer Disease. *Res. Sq.* **2023**, preprint. [[CrossRef](#)]
44. Milà-Alomà, M.; Ashton, N.J.; Shekari, M.; Salvadó, G.; Ortiz-Romero, P.; Montoliu-Gaya, L.; Benedet, A.L.; Karikari, T.K.; Lantero-Rodríguez, J.; Vanmechelen, E.; et al. Plasma P-Tau231 and p-Tau217 as State Markers of Amyloid- $\beta$  Pathology in Preclinical Alzheimer's Disease. *Nat. Med.* **2022**, *28*, 1797–1801. [[CrossRef](#)] [[PubMed](#)]



45. Therriault, J.; Vermeiren, M.; Servaes, S.; Tissot, C.; Ashton, N.J.; Benedet, A.L.; Karikari, T.K.; Lantero-Rodriguez, J.; Brum, W.S.; Lussier, F.Z.; et al. Association of Phosphorylated Tau Biomarkers With Amyloid Positron Emission Tomography vs Tau Positron Emission Tomography. *JAMA Neurol.* **2023**, *80*, 188–199. [[CrossRef](#)] [[PubMed](#)]
46. Thijssen, E.H.; La Joie, R.; Strom, A.; Fonseca, C.; Iaccarino, L.; Wolf, A.; Spina, S.; Allen, I.E.; Cobigo, Y.; Heuer, H.; et al. Plasma Phosphorylated Tau 217 and Phosphorylated Tau 181 as Biomarkers in Alzheimer’s Disease and Frontotemporal Lobar Degeneration: A Retrospective Diagnostic Performance Study. *Lancet Neurol.* **2021**, *20*, 739–752. [[CrossRef](#)] [[PubMed](#)]
47. Barthélemy, N.R.; Horie, K.; Sato, C.; Bateman, R.J. Blood Plasma Phosphorylated-Tau Isoforms Track CNS Change in Alzheimer’s Disease. *J. Exp. Med.* **2020**, *217*, e20200861. [[CrossRef](#)]
48. Mohs, R.C.; Beaugregard, D.; Dwyer, J.; Gaudioso, J.; Bork, J.; MaGee-Rodgers, T.; Key, M.N.; Kerwin, D.R.; Hughes, L.; Cordell, C.B.; et al. The Bio-Hermes Study: Biomarker Database Developed to Investigate Blood-based and Digital Biomarkers in Community-based, Diverse Populations Clinically Screened for Alzheimer’s Disease. *Alzheimer’s Dement.* **2024**, *20*, 2752–2765. [[CrossRef](#)]
49. Pontecorvo, M.J.; Lu, M.; Burnham, S.C.; Schade, A.E.; Dage, J.L.; Shcherbinin, S.; Collins, E.C.; Sims, J.R.; Mintun, M.A. Association of Donanemab Treatment With Exploratory Plasma Biomarkers in Early Symptomatic Alzheimer Disease: A Secondary Analysis of the TRAILBLAZER-ALZ Randomized Clinical Trial. *JAMA Neurol.* **2022**, *79*, 1250–1259. [[CrossRef](#)]
50. Pichet Binette, A.; Janelidze, S.; Cullen, N.; Dage, J.L.; Bateman, R.J.; Zetterberg, H.; Blennow, K.; Stomrud, E.; Mattsson-Carlgrén, N.; Hansson, O. Confounding Factors of Alzheimer’s Disease Plasma Biomarkers and Their Impact on Clinical Performance. *Alzheimer’s Dement.* **2023**, *19*, 1403–1414. [[CrossRef](#)]
51. Therriault, J.; Servaes, S.; Tissot, C.; Rahmouni, N.; Ashton, N.J.; Benedet, A.L.; Karikari, T.K.; Macedo, A.C.; Lussier, F.Z.; Stevenson, J.; et al. Equivalence of Plasma P-tau217 with Cerebrospinal Fluid in the Diagnosis of Alzheimer’s Disease. *Alzheimer’s Dement.* **2023**, *19*, 4967–4977. [[CrossRef](#)] [[PubMed](#)]
52. Ashton, N.J.; Brum, W.S.; Di Molfetta, G.; Benedet, A.L.; Arslan, B.; Jonaitis, E.; Langhough, R.E.; Cody, K.; Wilson, R.; Carlsson, C.M.; et al. Diagnostic Accuracy of a Plasma Phosphorylated Tau 217 Immunoassay for Alzheimer Disease Pathology. *JAMA Neurol.* **2024**, *81*, 255. [[CrossRef](#)] [[PubMed](#)]
53. Ossenkoppele, R.; Van Der Kant, R.; Hansson, O. Tau Biomarkers in Alzheimer’s Disease: Towards Implementation in Clinical Practice and Trials. *Lancet Neurol.* **2022**, *21*, 726–734. [[CrossRef](#)]
54. Barthélemy, N.R.; Salvadó, G.; Schindler, S.E.; He, Y.; Janelidze, S.; Collij, L.E.; Saef, B.; Henson, R.L.; Chen, C.D.; Gordon, B.A.; et al. Highly Accurate Blood Test for Alzheimer’s Disease Is Similar or Superior to Clinical Cerebrospinal Fluid Tests. *Nat. Med.* **2024**, *30*, 1085–1095. [[CrossRef](#)]
55. Woo, M.S.; Tissot, C.; Lantero-Rodriguez, J.; Snellman, A.; Therriault, J.; Rahmouni, N.; Macedo, A.C.; Servaes, S.; Wang, Y.; Arias, J.F.; et al. Plasma pTau-217 and N-terminal Tau (NTA) Enhance Sensitivity to Identify Tau PET Positivity in Amyloid- $\beta$  Positive Individuals. *Alzheimer’s Dement.* **2023**, *20*, 1166–1174. [[CrossRef](#)] [[PubMed](#)]
56. Jack, C.R.; Wiste, H.J.; Algeciras-Schimnich, A.; Figdore, D.J.; Schwarz, C.G.; Lowe, V.J.; Ramanan, V.K.; Vemuri, P.; Mielke, M.M.; Knopman, D.S.; et al. Predicting Amyloid PET and Tau PET Stages with Plasma Biomarkers. *Brain* **2023**, *146*, 2029–2044. [[CrossRef](#)] [[PubMed](#)]
57. Ferreira, P.C.L.; Therriault, J.; Tissot, C.; Ferrari-Souza, J.P.; Benedet, A.L.; Povala, G.; Bellaver, B.; Leffa, D.T.; Brum, W.S.; Lussier, F.Z.; et al. Plasma P-tau231 and P-tau217 Inform on Tau Tangles Aggregation in Cognitively Impaired Individuals. *Alzheimer’s Dement.* **2023**, *19*, 4463–4474. [[CrossRef](#)] [[PubMed](#)]
58. Janelidze, S.; Palmqvist, S.; Leuzy, A.; Stomrud, E.; Verberk, I.M.W.; Zetterberg, H.; Ashton, N.J.; Pesini, P.; Sarasa, L.; Allué, J.A.; et al. Detecting Amyloid Positivity in Early Alzheimer’s Disease Using Combinations of Plasma A $\beta$ 42/A $\beta$ 40 and P-tau. *Alzheimer’s Dement.* **2022**, *18*, 283–293. [[CrossRef](#)]
59. Montoliu-Gaya, L.; Alosco, M.L.; Yhang, E.; Tripodis, Y.; Sconzo, D.; Ally, M.; Grötschel, L.; Ashton, N.J.; Lantero-Rodriguez, J.; Sauer, M.; et al. Optimal Blood Tau Species for the Detection of Alzheimer’s Disease Neuropathology: An Immunoprecipitation Mass Spectrometry and Autopsy Study. *Acta Neuropathol.* **2024**, *147*, 5. [[CrossRef](#)]
60. Salvadó, G.; Ossenkoppele, R.; Ashton, N.J.; Beach, T.G.; Serrano, G.E.; Reiman, E.M.; Zetterberg, H.; Mattsson-Carlgrén, N.; Janelidze, S.; Blennow, K.; et al. Specific Associations between Plasma Biomarkers and Postmortem Amyloid Plaque and Tau Tangle Loads. *EMBO Mol. Med.* **2023**, *15*, e17123. [[CrossRef](#)]
61. Zabala-Findlay, A.; Penny, L.K.; Lofthouse, R.A.; Porter, A.J.; Palliyil, S.; Harrington, C.R.; Wischik, C.M.; Arastoo, M. Utility of Blood-Based Tau Biomarkers for Mild Cognitive Impairment and Alzheimer’s Disease: Systematic Review and Meta-Analysis. *Cells* **2023**, *12*, 1184. [[CrossRef](#)] [[PubMed](#)]
62. Yamoah, A.; Tripathi, P.; Sechi, A.; Köhler, C.; Guo, H.; Chandrasekar, A.; Nolte, K.W.; Wruck, C.J.; Katona, I.; Anink, J.; et al. Aggregates of RNA Binding Proteins and ER Chaperones Linked to Exosomes in Granulovacuolar Degeneration of the Alzheimer’s Disease Brain. *J. Alzheimer’s Dis.* **2020**, *75*, 139–156. [[CrossRef](#)] [[PubMed](#)]
63. Jonaitis, E.M.; Janelidze, S.; Cody, K.A.; Langhough, R.; Du, L.; Chin, N.A.; Mattsson-Carlgrén, N.; Hogan, K.J.; Christian, B.T.; Betthausen, T.J.; et al. Plasma Phosphorylated Tau 217 in Preclinical Alzheimer’s Disease. *Brain Commun.* **2023**, *5*, fcad057. [[CrossRef](#)] [[PubMed](#)]
64. Ashton, N.J.; Janelidze, S.; Mattsson-Carlgrén, N.; Binette, A.P.; Strandberg, O.; Brum, W.S.; Karikari, T.K.; González-Ortiz, F.; Di Molfetta, G.; Meda, F.J.; et al. Differential Roles of A $\beta$ 42/40, p-Tau231 and p-Tau217 for Alzheimer’s Trial Selection and Disease Monitoring. *Nat. Med.* **2022**, *28*, 2555–2562. [[CrossRef](#)] [[PubMed](#)]

65. Mielke, M.M.; Aakre, J.A.; Algeciras-Schimnich, A.; Proctor, N.K.; Machulda, M.M.; Eichenlaub, U.; Knopman, D.S.; Vemuri, P.; Graff-Radford, J.; Jack, C.R.; et al. Comparison of CSF Phosphorylated Tau 181 and 217 for Cognitive Decline. *Alzheimer's Dement.* **2022**, *18*, 602–611. [[CrossRef](#)] [[PubMed](#)]
66. Mattsson-Carlgrén, N.; Janelidze, S.; Palmqvist, S.; Cullen, N.; Svenningsson, A.L.; Strandberg, O.; Mengel, D.; Walsh, D.M.; Stomrud, E.; Dage, J.L.; et al. Longitudinal Plasma P-Tau217 Is Increased in Early Stages of Alzheimer's Disease. *Brain* **2020**, *143*, 3234–3241. [[CrossRef](#)] [[PubMed](#)]
67. Hansson, O.; Blennow, K.; Zetterberg, H.; Dage, J. Blood Biomarkers for Alzheimer's Disease in Clinical Practice and Trials. *Nat. Aging* **2023**, *3*, 506–519. [[CrossRef](#)] [[PubMed](#)]
68. Palmqvist, S.; Tideman, P.; Cullen, N.; Zetterberg, H.; Blennow, K.; the Alzheimer's Disease Neuroimaging Initiative; Dage, J.L.; Stomrud, E.; Janelidze, S.; Mattsson-Carlgrén, N.; et al. Prediction of Future Alzheimer's Disease Dementia Using Plasma Phospho-Tau Combined with Other Accessible Measures. *Nat. Med.* **2021**, *27*, 1034–1042. [[CrossRef](#)] [[PubMed](#)]
69. Mattsson-Carlgrén, N.; Salvadó, G.; Ashton, N.J.; Tideman, P.; Stomrud, E.; Zetterberg, H.; Ossenkoppele, R.; Betthausen, T.J.; Cody, K.A.; Jonaitis, E.M.; et al. Prediction of Longitudinal Cognitive Decline in Preclinical Alzheimer Disease Using Plasma Biomarkers. *JAMA Neurol.* **2023**, *80*, 360–369. [[CrossRef](#)]
70. Gicas, K.M.; Honer, W.G.; Petyuk, V.A.; Wilson, R.S.; Boyle, P.A.; Leurgans, S.E.; Schneider, J.A.; De Jager, P.L.; Bennett, D.A. Primacy and Recency Effects in Verbal Memory Are Differentially Associated with Post-Mortem Frontal Cortex p-Tau 217 and 202 Levels in a Mixed Sample of Community-Dwelling Older Adults. *J. Clin. Exp. Neuropsychol.* **2023**, *45*, 770–785. [[CrossRef](#)]
71. Campese, N.; Palermo, G.; Del Gamba, C.; Beatino, M.F.; Galgani, A.; Belli, E.; Del Prete, E.; Della Vecchia, A.; Vergallo, A.; Siciliano, G.; et al. Progress Regarding the Context-of-Use of Tau as Biomarker of Alzheimer's Disease and Other Neurodegenerative Diseases. *Expert Rev. Proteom.* **2021**, *18*, 27–48. [[CrossRef](#)] [[PubMed](#)]
72. Arai, H.; Morikawa, Y.; Higuchi, M.; Matsui, T.; Clark, C.M.; Miura, M.; Machida, N.; Lee, V.M.-Y.; Trojanowski, J.Q.; Sasaki, H. Cerebrospinal Fluid Tau Levels in Neurodegenerative Diseases with Distinct Tau-Related Pathology. *Biochem. Biophys. Res. Commun.* **1997**, *236*, 262–264. [[CrossRef](#)]
73. Schoonenboom, N.S.M.; Reesink, F.E.; Verwey, N.A.; Kester, M.I.; Teunissen, C.E.; Van De Ven, P.M.; Pijnenburg, Y.A.L.; Blankenstein, M.A.; Rozemuller, A.J.; Scheltens, P.; et al. Cerebrospinal Fluid Markers for Differential Dementia Diagnosis in a Large Memory Clinic Cohort. *Neurology* **2012**, *78*, 47–54. [[CrossRef](#)] [[PubMed](#)]
74. Mukaetova-Ladinska, E.B.; Monteith, R.; Perry, E.K. Cerebrospinal Fluid Biomarkers for Dementia with Lewy Bodies. *Int. J. Alzheimer's Dis.* **2010**, *2010*, 536538. [[CrossRef](#)] [[PubMed](#)]
75. Abu-Rumeileh, S.; Mometto, N.; Bartoletti-Stella, A.; Polisch, B.; Oppi, F.; Poda, R.; Stanzani-Maserati, M.; Cortelli, P.; Liguori, R.; Capellari, S.; et al. Cerebrospinal Fluid Biomarkers in Patients with Frontotemporal Dementia Spectrum: A Single-Center Study. *J. Alzheimer's Dis.* **2018**, *66*, 551–563. [[CrossRef](#)] [[PubMed](#)]
76. Meeter, L.H.H.; Vijverberg, E.G.; Del Campo, M.; Rozemuller, A.J.M.; Donker Kaat, L.; De Jong, F.J.; Van Der Flier, W.M.; Teunissen, C.E.; Van Swieten, J.C.; Pijnenburg, Y.A.L. Clinical Value of Neurofilament and Phospho-Tau/Tau Ratio in the Frontotemporal Dementia Spectrum. *Neurology* **2018**, *90*, e1231–e1239. [[CrossRef](#)] [[PubMed](#)]
77. Guo, Z.; Tian, C.; Shi, Y.; Song, X.-R.; Yin, W.; Tao, Q.-Q.; Liu, J.; Peng, G.-P.; Wu, Z.-Y.; Wang, Y.-J.; et al. Blood-Based CNS Regionally and Neuronally Enriched Extracellular Vesicles Carrying pTau217 for Alzheimer's Disease Diagnosis and Differential Diagnosis. *Acta Neuropathol. Commun.* **2024**, *12*, 38. [[CrossRef](#)] [[PubMed](#)]
78. Hanes, J.; Kovac, A.; Kvartsberg, H.; Kontsekova, E.; Fialova, L.; Katina, S.; Kovacech, B.; Stevens, E.; Hort, J.; Vyhnaek, M.; et al. Evaluation of a Novel Immunoassay to Detect P-Tau Thr217 in the CSF to Distinguish Alzheimer Disease from Other Dementias. *Neurology* **2020**, *95*, e3026–e3035. [[CrossRef](#)] [[PubMed](#)]
79. Ashton, N.J.; Janelidze, S.; Al Khleifat, A.; Leuzy, A.; Van Der Ende, E.L.; Karikari, T.K.; Benedet, A.L.; Pascoal, T.A.; Lleó, A.; Parnetti, L.; et al. A Multicentre Validation Study of the Diagnostic Value of Plasma Neurofilament Light. *Nat. Commun.* **2021**, *12*, 3400. [[CrossRef](#)]
80. Korecka, M.; Shaw, L.M. Mass Spectrometry-based Methods for Robust Measurement of Alzheimer's Disease Biomarkers in Biological Fluids. *J. Neurochem.* **2021**, *159*, 211–233. [[CrossRef](#)]
81. Buerger, K.; Zinkowski, R.; Teipel, S.J.; Tapiola, T.; Arai, H.; Blennow, K.; Andreasen, N.; Hofmann-Kiefer, K.; DeBernardis, J.; Kerkman, D.; et al. Differential Diagnosis of Alzheimer Disease With Cerebrospinal Fluid Levels of Tau Protein Phosphorylated at Threonine 231. *Arch. Neurol.* **2002**, *59*, 1267–1272. [[CrossRef](#)] [[PubMed](#)]
82. Barthélemy, N.R.; Mallipeddi, N.; Moiseyev, P.; Sato, C.; Bateman, R.J. Tau Phosphorylation Rates Measured by Mass Spectrometry Differ in the Intracellular Brain vs. Extracellular Cerebrospinal Fluid Compartments and Are Differentially Affected by Alzheimer's Disease. *Front. Aging Neurosci.* **2019**, *11*, 121. [[CrossRef](#)] [[PubMed](#)]
83. Janelidze, S.; Bali, D.; Ashton, N.J.; Barthélemy, N.R.; Vanbrabant, J.; Stoops, E.; Vanmechelen, E.; He, Y.; Dolado, A.O.; Triana-Baltzer, G.; et al. Head-to-Head Comparison of 10 Plasma Phospho-Tau Assays in Prodromal Alzheimer's Disease. *Brain* **2023**, *146*, 1592–1601. [[CrossRef](#)] [[PubMed](#)]
84. Therriault, J.; Woo, M.S.; Salvadó, G.; Gobom, J.; Karikari, T.K.; Janelidze, S.; Servaes, S.; Rahmouni, N.; Tissot, C.; Ashton, N.J.; et al. Comparison of Immunoassay- with Mass Spectrometry-Derived p-Tau Quantification for the Detection of Alzheimer's Disease Pathology. *Mol. Neurodegener.* **2024**, *19*, 2. [[CrossRef](#)] [[PubMed](#)]
85. Khan, M.; Shah, S.H.; Salman, M.; Abdullah, M.; Hayat, F.; Akbar, S. Enzyme-Linked Immunosorbent Assay versus Chemiluminescent Immunoassay: A General Overview. *Glob. J. Med. Pharm. Biomed. Update* **2023**, *18*, 1. [[CrossRef](#)]

86. Arcaro, M.; Fenoglio, C.; Serpente, M.; Arighi, A.; Fumagalli, G.G.; Sacchi, L.; Floro, S.; D’Anca, M.; Sorrentino, F.; Visconte, C.; et al. A Novel Automated Chemiluminescence Method for Detecting Cerebrospinal Fluid Amyloid-Beta 1-42 and 1-40, Total Tau and Phosphorylated-Tau: Implications for Improving Diagnostic Performance in Alzheimer’s Disease. *Biomedicines* **2022**, *10*, 2667. [[CrossRef](#)]
87. Pilotto, A.; Quaresima, V.; Trasciatti, C.; Tolassi, C.; Bertoli, D.; Mordenti, C.; Galli, A.; Rizzardi, A.; Caratozzolo, S.; Zancanaro, A.; et al. Plasma P-Tau217 in Alzheimer’s Disease: Lumipulse and ALZpath SIMOA Head-to-Head Comparison. *medRxiv* **2024**. [[CrossRef](#)]
88. Therriault, J.; Zimmer, E.R.; Benedet, A.L.; Pascoal, T.A.; Gauthier, S.; Rosa-Neto, P. Staging of Alzheimer’s Disease: Past, Present, and Future Perspectives. *Trends Mol. Med.* **2022**, *28*, 726–741. [[CrossRef](#)] [[PubMed](#)]
89. Baiardi, S.; Quadalti, C.; Mammana, A.; Dellavalle, S.; Zenesini, C.; Sambati, L.; Pantieri, R.; Polischi, B.; Romano, L.; Suffritti, M.; et al. Diagnostic Value of Plasma P-Tau181, NfL, and GFAP in a Clinical Setting Cohort of Prevalent Neurodegenerative Dementias. *Alzheimer’s Res. Ther.* **2022**, *14*, 153. [[CrossRef](#)]
90. Giacomucci, G.; Mazzeo, S.; Crucitti, C.; Ingannato, A.; Bagnoli, S.; Padiglioni, S.; Galdo, G.; Emiliani, F.; Frigerio, D.; Moschini, V.; et al. Plasma P-Tau181 as a Promising Non-Invasive Biomarker of Alzheimer’s Disease Pathology in Subjective Cognitive Decline and Mild Cognitive Impairment. *J. Neurol. Sci.* **2023**, *453*, 120805. [[CrossRef](#)]
91. Kivisäkk, P.; Fatima, H.A.; Cahoon, D.S.; Otieno, B.; Chacko, L.; Minooei, F.; Demos, C.; Stengelin, M.; Sigal, G.; Wohlstadter, J.; et al. Clinical Evaluation of a Novel Plasma pTau217 Electrochemiluminescence Immunoassay in Alzheimer’s Disease. *Sci. Rep.* **2024**, *14*, 629. [[CrossRef](#)]
92. Gobom, J.; Benedet, A.L.; Mattsson-Carlgrén, N.; Montoliu-Gaya, L.; Schultz, N.; Ashton, N.J.; Janelidze, S.; Servaes, S.; Sauer, M.; Pascoal, T.A.; et al. Antibody-Free Measurement of Cerebrospinal Fluid Tau Phosphorylation across the Alzheimer’s Disease Continuum. *Mol. Neurodegener.* **2022**, *17*, 81. [[CrossRef](#)] [[PubMed](#)]
93. Tariot, P.N.; Reiman, E.M.; Alexander, R.C.; Langbaum, J.B.; Holdridge, K.; Ferguson, M.B.; Yaari, R.; Sims, J.R. TRAILBLAZER-ALZ 3 Trial Design and Rationale. *J. Prev. Alzheimer’s Dis.* **2021**, *8* (Suppl. 1), S3.
94. Shcherbinin, S.; Andersen, S.W.; Evans, C.D.; Lo, A.C.; Lu, M.; Navitsky, M.; Collins, E.C.; Sims, J.R.; Brooks, D.A.; Mintun, M.A. TRAILBLAZER-ALZ Study: Dynamics of Amyloid Reduction after Donanemab Treatment. *Alzheimer’s Dement.* **2021**, *17*, e057492. [[CrossRef](#)]
95. Sims, J.R.; Lu, M.; Schade, A.E.; Brooks, D.A.; Mintun, M.A. TRAILBLAZER-ALZ Studies: Plasma P-Tau Assays and the Initial Performance in Clinical Trials. *J. Prev. Alzheimer’s Dis.* **2021**, *8* (Suppl. 1), S2.
96. Brum, W.S.; Ashton, N.J.; Simrén, J.; Di Molfetta, G.; Karikari, T.K.; Benedet, A.L.; Zimmer, E.R.; Lantero-Rodriguez, J.; Montoliu-Gaya, L.; Jeromin, A.; et al. Biological Variation Estimates of Alzheimer’s Disease Plasma Biomarkers in Healthy Individuals. *Alzheimer’s Dement.* **2024**, *20*, 1284–1297. [[CrossRef](#)]
97. Mielke, M.M.; Dage, J.L.; Frank, R.D.; Algeciras-Schimmich, A.; Knopman, D.S.; Lowe, V.J.; Bu, G.; Vemuri, P.; Graff-Radford, J.; Jack, C.R.; et al. Performance of Plasma Phosphorylated Tau 181 and 217 in the Community. *Nat. Med.* **2022**, *28*, 1398–1405. [[CrossRef](#)]
98. Vila-Castelar, C.; Chen, Y.; Langella, S.; Lopera, F.; Zetterberg, H.; Hansson, O.; Dage, J.L.; Janelidze, S.; Su, Y.; Chen, K.; et al. Sex Differences in Blood Biomarkers and Cognitive Performance in Individuals with Autosomal Dominant Alzheimer’s Disease. *Alzheimer’s Dement.* **2023**, *19*, 4127–4138. [[CrossRef](#)] [[PubMed](#)]
99. Kelleher, R.J.; Shen, J. Presenilin-1 Mutations and Alzheimer’s Disease. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 629–631. [[CrossRef](#)]
100. Janelidze, S.; Barthélemy, N.R.; He, Y.; Bateman, R.J.; Hansson, O. Mitigating the Associations of Kidney Dysfunction With Blood Biomarkers of Alzheimer Disease by Using Phosphorylated Tau to Total Tau Ratios. *JAMA Neurol.* **2023**, *80*, 516–522. [[CrossRef](#)]
101. Rates. Center for Alzheimer’s Disease Research. Available online: <https://alz.carney.brown.edu/facilities-resources/biomarkers-facility/rates> (accessed on 1 July 2024).
102. Schindler, S.E.; Li, Y.; Li, M.; Despotis, A.; Park, E.; Vittert, L.; Hamilton, B.H.; Womack, K.B.; Saef, B.; Holtzman, D.M.; et al. Using Alzheimer’s Disease Blood Tests to Accelerate Clinical Trial Enrollment. *Alzheimer’s Dement.* **2023**, *19*, 1175–1183. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.