

Analysis of Single Nucleotide Polymorphisms (SNPs) rs2234693 and rs9340799 of the *ESR1* Gene and the Risk of Breast Cancer

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Abstract. *Background/Aim:* The aim of this study was to analyze rs2234693 and rs9340799 polymorphisms of the *ESR1* gene in the context of breast cancer risk in Polish patients. *Materials and Methods:* The study involved a group of 117 patients with breast cancer and 106 controls. The analyses were carried out using the polymerase chain reaction – restriction fragments length polymorphism technique. *Results:* The presence of the CC genotype in rs2234693 more than doubled the risk of breast cancer ($p=0.04$), whereas the presence of the TT genotype in rs2234693 significantly reduced the risk of developing this type of cancer ($p=0.0002$). The presence of the GG genotype in rs9340799 more than doubled the risk of breast cancer ($p=0.04$), which was confirmed by the analysis of the recessive model ($p=0.04$). *Conclusion:* The polymorphisms rs2234693 and rs9340799 of the *ESR1* gene may be associated with the risk of breast cancer among Polish women.

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Hormone-dependent cancer of the breast is the most common cancer in women (1-3). Hormones are one of the main factors that regulate the function of individual organs of our body and sex hormones, including estrogens, can act as carcinogens (4, 5). They can cause chromosome segregation errors and structural chromosome changes. Estrogens can stimulate the uncontrolled proliferation of mutated breast cells, damage DNA and induce cancer (6, 7). As a result of binding of the steroid-receptor complex to DNA in the cell nucleus, estrogens have a direct effect on target cells (8). In this way, estrogens regulate gene expression and can cause proto-oncogenes to be converted into oncogenes (9). Cell proliferation is stimulated by estrogens by enhancing DNA synthesis in stromal and glandular tissues (10). Estrogen metabolism leads to the formation of several metabolites that, when converted into semiquinones and quinones, generate reactive oxygen species and directly damage DNA (11).

Literature data indicate an association of endogenous and exogenous circulating estrogen with the etiology of breast cancer. Premenopausal women have an increased risk of this cancer (12). Estrogen receptors (ERs) mediate the effects of estrogen on breast cells. Estrogen binding to its receptor promotes the growth and differentiation of normal breast cells and can lead to breast cancer (13-15). A recent research indicated that ER mediates the suppression of the expression of Capillary morphogenesis gene 2 (*CMG2*), which is involved in breast cancer progression, in relevant cancer cells. Studies have shown that *CMG2* expression is inversely correlated with ER status in breast cancer (16).

A distinction is made between two types of ER: ER α and ER β , encoded by two different genes, *ESR1* and *ESR2*,



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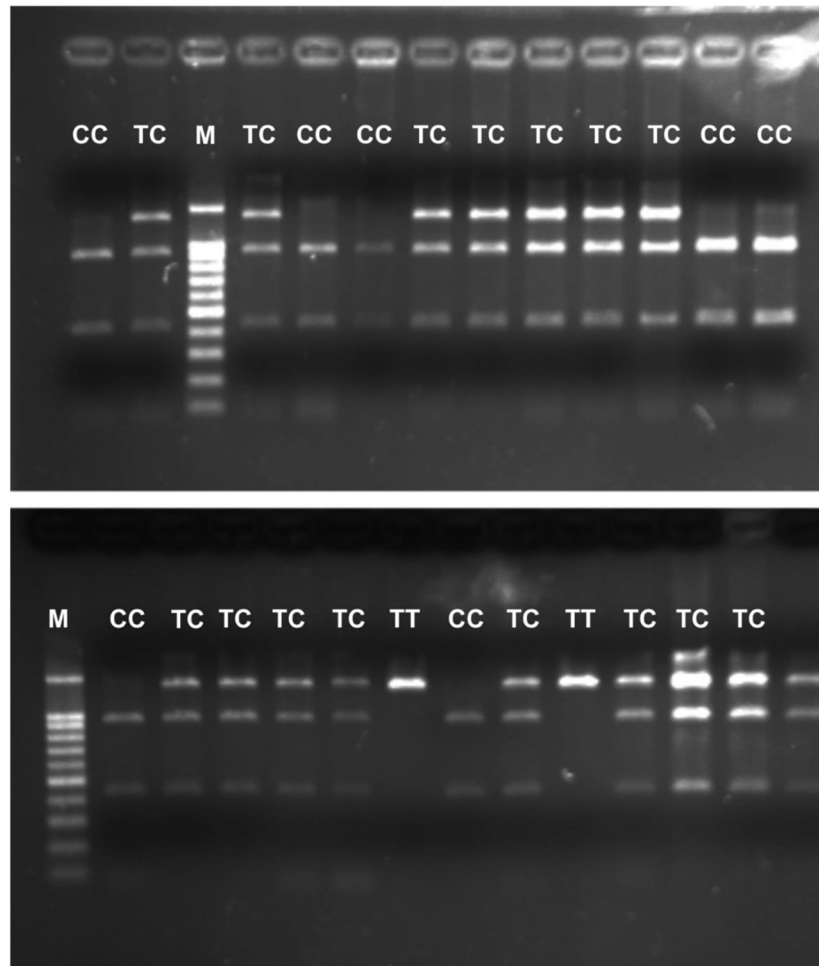


Figure 1. An exemplary picture of the electrophoretic separation of the PCR-RFLP reaction products for the Pull polymorphism (rs2234693, T/C) in the *ESR1*. M: DNA fragment size standard (DNA Marker 100-1000 ladder, Sigma-Aldrich, Burlington, MA, USA); T: normal allele; C: mutant allele.

respectively. ER α shows a higher level of expression in breast tissue compared to ER β , which is why it is often associated with the development of breast cancer (17).

The aim of the presented work was to examine whether polymorphisms in *ESR1*, which is highly polymorphic, affect breast cancer incidence. Of the many polymorphisms in *ESR1*, the two best studied are rs2234693 (also known as PvuII or 397T>C) and rs9340799 (also known as XbaI or 351A>G). Both polymorphisms are located in intron 1, 1,397 bp and 351 bp, respectively, upstream of exon 2 of the gene. These polymorphisms may be associated with breast cancer and endometrial cancer (18-20). Studies indicate that *ESR1* amplification may be an early event in the development of endometrial cancer (21). Amplification of *ESR1* is independent of known clinical-pathological factors associated with poor prognosis and expression of certain proteins (PTEN, p53, HER2, MLH1, and ARID1A) (21). However,

many studies have found conflicting results as to the association of these two polymorphisms with breast cancer susceptibility (22-25). In order to explain these discrepancies, a meta-analysis of the polymorphism rs2234693 was performed in 2018. In that study, rs2234693 was shown to be associated with reduced susceptibility to breast cancer (26). As for rs9340799, Zhang *et al.* found no significant association between the polymorphism and breast cancer susceptibility (27). The polymorphisms rs2234693 and rs9340799 in the *ESR1* gene were not associated with breast cancer susceptibility in the Mexican population (27). However, the CTAG and CCGG haplogenotypes of rs2234693 and rs9340799 may significantly contribute to breast cancer risk susceptibility of patients who have had miscarriages or had significant tobacco consumption (27, 28).

The aim of our research was to determine: 1) the effect of two single nucleotide polymorphisms (SNPs - rs2234693,

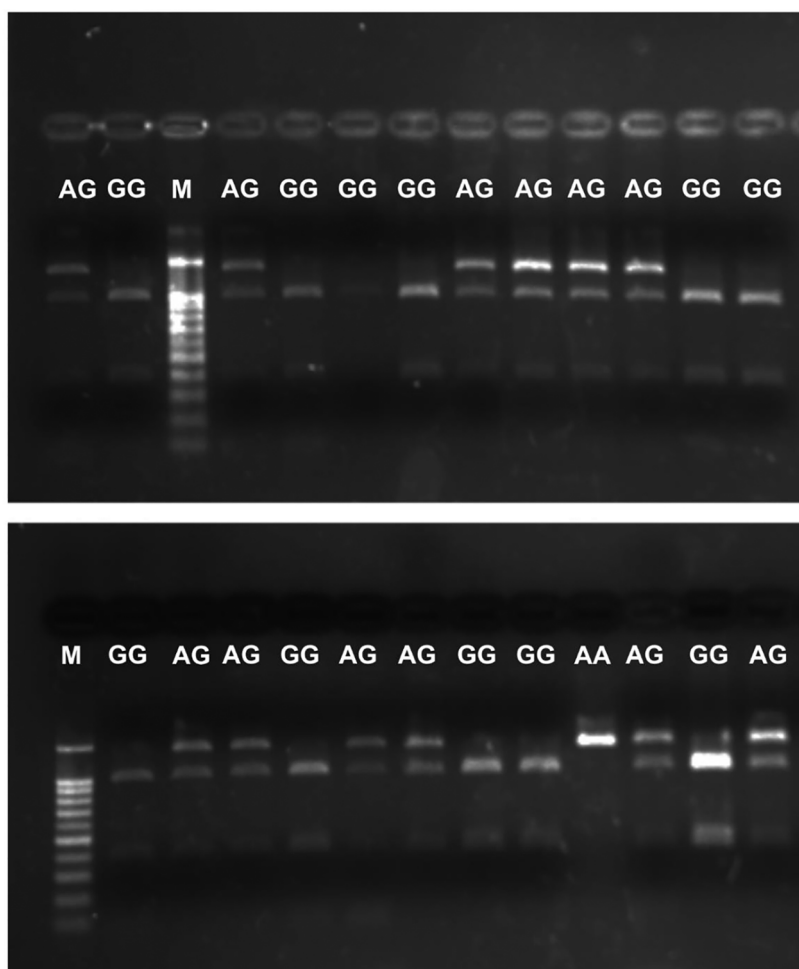


Figure 2. An exemplary picture of the electrophoretic separation of the PCR-RFLP reaction products for the Pull polymorphism (rs2234693, T/C) in the *ESR1*. M: DNA fragment size standard (DNA Marker 100-1000 ladder, Sigma-Aldrich, Burlington, MA, USA); T: normal allele; C: mutant allele.

rs9340799) of the *ESR1* gene on the risk of breast cancer in Polish women; 2) the effect of *ESR1* gene haplotypes in relation to the risk of breast cancer in women; 3) the association of the rs2234693 and rs9340799 polymorphisms of the *ESR1* gene with clinical-pathological features.

Materials and Methods

Patients. We used 117 venous blood samples from women diagnosed with breast cancer and 106 venous blood samples from women who were diagnosed with breast cancer. The samples were obtained from the Department of Surgical Oncology and Breast Diseases, Polish Mother's Memorial Hospital Research Institute, Lodz, Poland. The patients were operated on between 2015 and 2018. None of the patients had received prior neoadjuvant therapy, chemotherapy, or radiotherapy. The characteristics of the patients are presented in Table I. Approval of the Bioethics Committee at the Polish Mothers Memorial Hospital Research Institute in Lodz was obtained (Nr. 20/2015).

Application of histopathology scanner (3d Histech). Histological examination was performed by experienced pathologists using a digital slide scanner and slide review software (Case Viewer 2.3, 3D Histech, Budapest, Hungary). Histological sections were scanned with a panoramic scanner (3D Histech) and digital images were obtained. It is considered that the scanned digital form of the histological specimen, which can be evaluated using automated programmatic analysis modules, is an independent and objective method of histopathological examination. The study received funding from the Digital Poland Operational Programme MDB-BANK (grant no. POPC.02.03.01-00-0091/19).

DNA isolation from whole blood. DNA was isolated from whole blood using the GenElute Blood Genomic DNA Kit (Sigma-Aldrich, Steinheim am Albuch, Germany) according to the manufacturer's recommendations. DNA was stored at -20°C. The purity of the obtained DNA preparations was determined spectrophotometrically measuring the absorbance of each sample twice at wavelengths of 260 nm and 280 nm. The accepted criterion for DNA purity was a A260/A280 value within the range of 1.8-2.0. The concentration of

Table I. Characteristics of patients.

	Study group n (%)	Control group n (%)	
Group size	n=117	n=106	
Age, number of years	61.18±11.76	65.12±10.13	
<50 years	19 (16.2)	31 (29.2)	
≥50 years	98 (83.8)	75 (70.8)	0.03
Histological type	106 (90.8)		
Ductal cancer	7 (6.0)		
Lobular cancer	1 (0.8)		
Metaplastic cancer	1 (0.8)		
Mucinous cancer	2 (1.6)		
Ductal cancer <i>in situ</i>	0		
TNM classification			
pT1	98 (83.9)		
pT2	14 (12.1)		
pT3	1 (0.8)		
pT4	2 (1.6)		
pTis	2 (1.6)		
N0	91 (77.8)		
N1	23 (19.6)		
N2	3 (2.6)		
Histological grading			
G1	47 (40.9)		
G2	61 (53.0)		
G3	7 (6.1)		
Tumor size (mm)			
≤10	47 (40.9)		
10-30	61 (53.0)		
>30	7 (6.1)		
Expression of receptors			
ER+	21 (17.9)		
PR+	26 (22.2)		
HER2+	47 (40.9)		
ER+PR+HER2+	12 (10.4)		
ER-PR-HER2	9 (7.8)		

TNM: Tumor, nodules and metastases; ER: estrogen receptor; PR: progesterone receptor; HER: human epidermal growth factor receptor 2.

DNA was determined spectrophotometrically based on absorbance values measured at a wavelength of 260 nm considering that 1OD=50 µg DNA/ml.

Analysis of selected polymorphisms of the *ESR1* gene. On the basis of data available in the SNP database NCBI (National Center for Biotechnology Information, <http://www.ncbi.nlm.nih.gov/snp>), two single nucleotide polymorphisms located in the *ESR1* gene were selected. The SNPs selected are NC_000006.11:g.152163335T>C (rs2234693) and NC_000006.11:g.152163381A>G (rs9340799). The analysis of the incidence of the above-mentioned polymorphisms in the group of patients with breast cancer as well as in the control group was carried out using PCR reaction combined with restriction fragment analysis, PCR-RFLP (Polymerase Chain Reaction – Restriction Fragments Length Polymorphism). Selected fragments of the *ESR1* gene were amplified and then digested using restriction enzymes - rs2234693 (PvuII); rs9340799 (XbaI). The nucleotide sequence of the

Table II. Nucleotide sequence of the amplified area and sequences of the primers used to carry out the amplification reaction of the *ESR1* gene fragment (27).

Primer F
F:5'-CTG CCA CCC TAT CTG TAT CTT TTC CTA TTC TCC-3'
Primer R
R:5'-TCT TTC TCT GCC ACC CTG GCG TCG ATT ATC TGA-3'
Product size
1,387 bp
Total nucleotide sequence with marking of cleavage sites by restriction enzymes used
ctgccaccctatctgtatcttttctctctccaagtctcaaaaggaatgcctccctcatfctctctt acattccatctgtgaatttggctgtgctgtacgtgtctaaagaaactcctgtaagatctgcttgc aggctgaattcaacttaaccagcttctgtttggtgactctctgcccaattgccattctgacatcctc cataaaccttctactaaagcatttactctctatttcttggtttctcagaatcctctactgttcatttccagc ttccttctgttctctctctctctctacatttttttagctttctactttctaaagcatttactctctatttctgtg gttttctagaatttctactgttcatttccagttctctgtgttcctctgattgtctctttctacatttttttt ctgtgttcctctgattttcacgcagctctggagttgcatgatcaatcatagcctactgcagcctcgacatcct aggctcaagtgtattctccacctcagccttacaagtagctaggactacagtcacacatcaccattctcag ctaatttttaagaagcattttatagagatggagctgtctatattgtgcaggctgggctcaactacagg gcttaaacattctctctgttggcctccaaaagtctgggattccaggcatgaaccaccatgctcagctct ctacatgtcctaagaaggagtttgaattgaagaacagattttcaattacatttcaagttataaaaa ctgatatccagggttatgtggcaatgacgtaaaaaattgaaattgtattttttgacacatgttctgtgtcc atcagttcatctgagttccaaatgtccag(PvuII)ctgttttatgctttgtctgtttccagagaccctg agtggtg(XbaI)ctagatgtgggatgagcattgtctcctaattgttctgaaataattgtatattctgca aaaacattaaagctattagaaccagctaattcattttgtcatttttataggtaacattctgtgtagcaggtag tatgttttttaaacagtttgcaataaacattcccctcaaggtaataataggcaacaccttttctgca acagacggcaagaggaatgaaagattagcttaccattgattcattttcaaaatgtcagataaagtg gatctgtcgtcctccagagagtgcatgttttcttttctaattgtaattgatttactgtttttccccccag gccaattcagataatcgacgccagggtggcagagaaaga

amplified fragments and the characteristics of the primers used are presented in Table II.

The PCR reaction was carried out in a final volume of 10 µl. The reaction mixture contained approximately 50 ng of genomic DNA, 1 µl of 5µM solution of each primer, 1 µl of GeneAmp 10× PCR Buffer with MgCl₂ (Applied Biosystems, Foster City, CA, USA), 0.2 µl of AmpliTaq Gold® polymerase (5U/µl) (Applied Biosystems), 1 µl of 10 µM dNTPs, and water. Initial denaturation was carried out for 5 min at 95°C, then the selected fragment of the tested gene was amplified during 40 cycles with the following parameters: 30 s at 95°C, 1 min at 62°C and 1 min at 72°C.

The products of DNA amplification were digested for 16 h using commercially available restriction enzymes - PvuII and XbaI (New England Biolabs, Ipswich, MA, USA) (29). The reaction products were then electrophoresed in 2% agarose gel, stained with ethidium bromide, and visualized under ultraviolet light.

The size of the products for the analysis of the PvuII polymorphism (rs2234693, T/C) and XbaI polymorphism (rs9340799, A/G) is summarized in Table III. The selected PCR products were additionally sequenced using the Sanger method at

the Department of Cytobiochemistry (University of Lodz, Poland) to corroborate the genotypes previously estimated using the PCR-RFLP method. An example of the electrophoretic separation of PCR-RFLP reaction products is shown in Figure 1 and Figure 2.

Statistical analysis. Statistical analysis was performed with the use of using STATISTICA 11 (StatSoft, Poznań, Poland), Chaplin 1.2 (genetics.emory.edu) and THESIAS (www.genecanvas.org). The effect of *ESR1* gene polymorphisms on breast cancer risk was assessed using odds ratio (OR) and a 95% confidence interval (CI) using a logistic regression model. In this case, OR allows assessment of how many times the risk of disease increases/decreases with an increase/decrease by one unit of the variable treated as a risk factor.

In order to examine whether the Hardy-Weinberg law of equilibrium is observed in the studied populations, the calculation program on the website www.ihg.gfs.de (Institute of Human Genetics, Technical University Munich and Helmholtz Center Munich) was used. To investigate the effect of *ESR1* gene haplotypes on cancer risk, Chaplin 1.2 and THESIAS were used.

All statistical tests were performed at a significance level of $\alpha=0.05$. The assessment of the consistency of the distribution of the values of the studied variables with the normal distribution was carried out using the Shapiro-Wilk test. In order to verify the hypothesis of age significance in the study and control groups, the χ^2 analysis was used.

Results

The obtained results indicate that the presence of the CC genotype in rs2234693 more than doubles the risk of breast cancer (OR=2.19; 95%CI=1.00-4.78); $p=0.04$ (Table IV), while the presence of the TT genotype in rs2234693 significantly reduces the risk of developing this type of cancer (OR=0.31; 95%CI=0.16-0.60); $p=0.0002$ (Table V). The age of the women had no effect on the prevalence of the individual genotypes or the associated risk of disease. The results of the analysis are also presented in the form of adjusted odds ratios.

In the case of the second polymorphism, rs9340799, it was found, analogously to the previous studies, that the presence of the GG genotype more than doubled the risk of breast cancer (OR=2.48; 95%CI=1.01-6.03) $p=0.04$ (Table VI), which was confirmed by the analysis of the recessive model (OR=2.36; 95%CI=1.02-5.41; $p=0.049$) (Table VII).

Haplotypes analysis. The analysis of *ESR1* gene haplotypes in relation to the rs2234693 and rs9340799 polymorphisms showed that among the examined patients, only the occurrence of the T-C-A haplogenotypes was associated with a significant reduction in the risk of cancer in women (OR=0.31; 95%CI=0.14-0.65; $p=0.001$). Detailed results of the analyses are summarized in Table VIII.

Analysis of the association of the rs2234693 and rs9340799 polymorphisms of the *ESR1* gene with clinical-pathological features. It was found that the presence of the TC or CC

Table III. *The size of the products obtained in the analysis of the PvuII polymorphism (rs2234693, T/C) and in the analysis of the XbaI polymorphism (rs9340799, A/G).*

PvuII (rs2234693); bp	
TT – 1374	
TC – 1374, 934, 440	
CC – 934, 440	
XbaI (rs9340799); bp	
AA – 1374	
AG – 1374, 981, 393	
GG – 981, 393	

genotype (presence of the C allele) in rs2234693 was significantly associated with a more than seven-fold increase in the tendency of breast cancers to metastasize (OR=7.79; 95%CI=3.21-18.90; $p=0.001$) and with a higher degree of anaplasia (OR=2.42; 95%CI=1.35-4.33; $p=0.003$) (Table IX).

In the case of rs9340799 polymorphism, the presence of AG or GG genotype (presence of the G allele) was associated with a larger tumor size (OR=16.66; 95%CI=2.13-130.05; $p=0.007$) and a higher degree of anaplasia (OR=38.73; 95%CI=13.34-112.37; $p=0.001$) (Table X).

Discussion

Genetic polymorphisms in the estrogen receptor- α (*ESR1*) gene are involved in alterations in receptor expression and function, evolving as important determinants of breast cancer susceptibility. The associations of the single nucleotide polymorphisms (SNPs) rs2234693 and rs9340799 in the *ESR1* gene with breast cancer have been studied. However, the data and conclusions are inconsistent and controversial. The rs2234693 polymorphism is found in the first base of intron 397 bp upstream exon 2, in which cytosine is replaced by thymine, and this change is identified by the PvuII endonuclease (30, 31).

It is known that localization of a polymorphism in intron regions may lead to modification of splicing in mRNA transcripts. The consequence of these changes is a change in the function of genes (31). There are differences in the incidence of this polymorphisms in different populations. It has been shown that the T or C allele may be a protective or risk factor for breast cancer (32-34, 27).

The C allele is considered to be an ancestral allele that is present at a lower frequency in the general population. Studies in the Mexican population have shown a different frequency of the C allele compared to other populations. The first studies were conducted in postmenopausal women (age range=46-80 years) and the C allele was found to be present

Table IV. Distribution of genotypes of the rs2234693 polymorphism of the ESR1 gene in the study and control groups and analysis of the risk of breast cancer.

SNP/genotype	Patients (%) / control (%)	OR (95%CI) ^a	p-Value	OR (95%CI) ^b	p-Value*
rs2234693 (PvuII)					
TT	28 (23.9)/38 (35.8)	1.00		1.00	
TC	60 (51.3)/50 (47.2)	1.63 (0.87-3.03)	0.12	1.63 (0.84-3.03)	0.12
CC	29 (24.8)/18 (17.0)	2.19 (1.00-4.78)	0.04	2.18 (1.00-4.75)	0.04
p-trend ^c	0.04				

^aRaw; ^bage-adjusted; ^cadditive genetic model (Cochran-Armitage test). SNP: Single nucleotide polymorphism; OR: odds ratio; p: probability; rs: polymorphism reference number.

Table V. Distribution of genotypes of the rs2234693 polymorphism of the ESR1 gene in the study and control groups – dominant and recessive model.

SNP/genotype	Patients (%) / control (%)	OR (95%CI) ^a	p-Value	OR (95%CI) ^b	p-Value*
rs2234693 (PvuII)					
TC lub CC vs. TT ^c	89 (76.1)/68 (64.1)	0.31 (0.16-0.60)	0.0002	0.30 (0.15-0.60)	0.0002
TC lub TT vs. CC ^d	88 (75.2)/88 (83.0)	1.61 (0.83-3.12)	0.15	1.61 (0.83-3.14)	0.16

^aRaw; ^bage-adjusted; ^cdominant genetic model; ^drecessive genetic model. SNP: Single nucleotide polymorphism; OR: odds ratio; p: probability; rs: polymorphism reference number.

Table VI. Distribution of genotypes of the rs9340799 polymorphism of the ESR1 gene in the study and control groups and analysis of the risk of breast cancer.

SNP/genotype	Patients (%) / control (%)	OR (95%CI) ^a	p-Value	OR (95%CI) ^b	p-Value*
Prs9340799 (XbaI)					
AA	49 (41.9)/52 (49.0)	1.00		1.00	
AG	47 (40.2)/45 (42.5)	1.12 (0.63-1.95)	0.72	1.12 (0.63-1.98)	0.71
GG	21 (17.9)/9 (8.5)	2.48 (1.01-6.03)	0.04	2.49 (1.02-6.05)	0.04
p-trend ^c	0.07				

^aRaw; ^bage-adjusted; ^cadditive genetic model (Cochran-Armitage test). SNP: Single nucleotide polymorphism; OR: odds ratio; p: probability; rs: polymorphism reference number.

Table VII. Distribution of genotypes of the rs9340799 polymorphism of the ESR1 gene in the study and control groups – dominant and recessive model.

SNP/genotype	Patients (%) / control (%)	OR (95%CI) ^a	p-Value	OR (95%CI) ^b	p-Value*
rs9340799 (XbaI)					
AG lub GG vs. AA ^c	68 (58.1)/54 (50.9)	0.75 (0.44-1.27)	0.28	0.75 (0.44-1.26)	0.28
AG lub AA vs. GG ^d	96 (99.2)/97 (100.0)	2.36 (1.02-5.41)	0.04	2.36 (1.02-5.41)	0.04

^aRaw; ^bage-adjusted; ^cdominant genetic model; ^drecessive genetic model. SNP: Single nucleotide polymorphism; OR: odds ratio; p: probability; rs: polymorphism reference number.

at a frequency of 30.9% (33), while in another study conducted in women under 45 years of age with C allele was present in 25.8% (34).

Recent studies in the Mexican population regarding these polymorphisms have shown that they are not associated with susceptibility to breast cancer. However,

Table VIII. *ESR1* gene haplotypes and the risk of breast cancer.

Haplotypes (rs2234693; rs9340799)	Patients (%) / control (%)	OR (95%CI) ^a	p-Value
T-T-A	48 (41.0)/27 (25.5)	1.00 (reference)	
A-A-T	29 (24.8)/24 (22.6)	0.68 (0.33-1.40)	0.29
T-C-A	20 (17.0)/37 (34.9)	0.31 (0.14-0.65)	0.001
A-G-T	13 (11.1)/12 (11.3)	0.53 (0.17-0.83)	0.15

^aRaw. OR: Odds ratio; p: probability; rs: polymorphism reference number.

Table IX. Association of the *ESR1* gene rs2234693 polymorphism with clinical-pathological features.

Variable	rs2234693		OR (95%CI) ^a	p-Value
	TT n (%)	TC or CC n (%)		
pT1 vs. pT2-T4	22 (18.8)/6 (5.1)	78 (66.7)/11 (9.4)	0.53 (0.18-1.59)	0.26
N0 vs. N1-N2	23 (19.6)/5 (4.3)	60 (51.3)/29 (24.8)	7.79 (3.21-18.90)	0.001
G1 vs. G2-G3	21 (62.3)/7 (13.1)	26 (16.9)/63 (7.7)	2.42 (1.35-4.33)	0.003

^aRaw. OR: Odds ratio; p: probability; rs: polymorphism reference number.

Table X. Association of the rs9340799 polymorphism of the *ESR1* gene with clinical-pathological features.

Variable	rs9340799		OR (95%CI) ^a	p-Value
	AA n (%)	AG or GG n (%)		
pT1 vs. pT2-T4	49 (41.9)/1 (0.8)	50 (42.7)/17 (14.6)	16.66 (2.13-130.05)	0.007
N0 vs. N1-N2	42 (35.9)/7 (6.0)	49 (41.9)/19 (16.2)	2.32 (0.89-6.073)	0.08
G1 vs. G2-G3	40 (34.2)/9 (7.7)	7 (6.0)/61 (52.1)	38.73 (13.34- 112.37)	0.001

^aRaw. OR: Odds ratio; p: probability; rs: polymorphism reference number.

the CTAG and CCGG haplogenotypes of the rs2234693 and rs9340799 polymorphisms may significantly contribute to breast cancer risk susceptibility in women who have had a miscarriage and those who have consumed tobacco (28).

Numerous meta-analyses indicate that women who carry the T allele have a low risk of developing breast cancer. In Caucasian populations, the risk of breast cancer is higher in carriers of the TT genotype, whereas in Asians the CC genotype (32, 35).

Kornats *et al.* found no association between *ESR1* gene variants and the risk of developing mastopathy in infertile women, although heterozygous *ESR1* gene variants increased the “protective” effect of FSHR (estrogen-stimulating hormone receptor) gene variants and reduced the risk of mastopathy (36).

The *ESR1* and *ESR2* gene variants have been studied in triple-negative breast cancer (TNBC), where unique genetic variants *ESR1* and *ESR2* have been shown to be associated with the risk of TNBC. This suggests a possible diagnostic and prognostic role of these polymorphisms in TNBC (37).

A total of 25 case-control studies (meta-analysis) were analyzed to investigate the association between the *ESR1* gene polymorphisms rs2234693, rs9340799, and rs1801132 and breast cancer risk in different populations. Three different comparative models were used to assess the association – the dominant model, the recessive model, and the homozygote comparative model. The results showed that regarding the rs2234693 polymorphism, individuals with the TT+TC or TT genotype were at a higher risk of developing breast cancer than those with the CC genotype. However, in the case of the rs9340799 and

rs1801132 polymorphisms, none of the 3 models found significant associations.

Analysis of polymorphisms was also performed according to ethnicity (white or Asian) and source of control (hospital or population). An assessment of ethnic subgroups showed that the TT rs2234693 genotype, compared to the CC genotype, conferred a higher risk of breast cancer only in Asians, but not in white populations. In the source-stratified subgroup analysis, individuals with the TT+TC genotype were found to be at higher risk of breast cancer than those with the CC genotype in the hospital subgroup (27).

Polymorphisms in genes involved in estrogen synthesis (*UGT2B17* - *UGT2B17**2, *CYP19A1* rs10046/rs4646 and *ESR1* rs2077647/rs2234693/rs9340799) have been correlated with breast cancer risk, prognosis, and response to treatment (38). The studies involved women with ER-positive postmenopausal breast cancer. The *UGT2B17**2 polymorphism was associated with higher levels of 17-hydroxyexemestane and a better prognosis compared to homozygous *UGT2B17*. The rs10046 A and rs4646 C alleles in *CYP19A1* were associated with higher estrogen levels. The presence of the rs10046 G and rs4646 A alleles was associated with low estrogen levels and a better prognosis compared to homozygous wild type (median follow-up of 7 years). Thus, postmenopausal hormone-responsive breast cancer is influenced by *UGT2B17* and *CYP19A1*. Carriers of *UGT2B17**2 and *CYP19A1* variants with low estrogen levels may have a better prognosis. Polymorphisms may be important in optimizing hormone therapy.

Three polymorphisms in *ESR1*, rs2881766, rs9383951, rs9340799 and one in *ESR2*, rs3020449, were studied in 459 patients and 549 healthy controls (38). The rs2881766 polymorphism was associated with a reduced risk of breast cancer. Whereas the rs3020449 was associated with an increased risk of this cancer. The other two polymorphisms were not associated with susceptibility to breast cancer. In addition, rs2881766 was correlated with lymph node metastases and ER expression, and rs3020449 was associated with tumor size, histological grade, and ER expression. The authors of the study emphasize that these findings require further validation in a large population (39).

A recent meta-analysis of 23 studies involving 34,721 patients, found no significant association between the rs9340799 *ESR1* polymorphism and breast cancer susceptibility (40). Subgroup analyses by ethnicity, menopausal status, and study quality also found no statistically significant association (40).

In our study, women who were carriers of the CC rs2234693 and GG rs9340799 genotypes had a higher risk of developing breast cancer. The TT rs2234693 genotype had a protective role. The T-C-A haplogenotype of the rs2234693 and rs9340799 polymorphisms in the *ESR1* gene in relation to was associated with a lower risk of breast cancer. Our

results differ from the previously cited literature data and shed new light on this topic in the Polish population. We would like to emphasize that this is the first study conducted in Poland. Of course, we are aware of the limitations of our study. First of all, the study population was small (117 patients, 106 controls). The cited research articles are few and the results of the research are contradictory. Further research is needed to confirm or reject the importance of *ESR1* polymorphisms in breast cancer.

Conclusion

The CC and GG genotypes of rs2234693 and rs9340799, respectively, of the *ESR1* gene increase the risk of breast cancer in women: 1) The TT genotype of the rs2234693 polymorphism in the *ESR1* gene lowers the risk of breast cancer in women; 2) The occurrence of the T-C-A haplotype regarding the rs2234693 and rs9340799 polymorphisms of the *ESR1* gene is associated with a lower risk of breast cancer in women; 3) In the case of *ESR1* gene polymorphisms rs2234693 and rs9340799, the presence of the mutant alleles C and G, respectively, is associated with a higher degree of anaplasia in female breast cancers.

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Conflicts of Interest

The Authors declare no conflicts of interest in relation to this study.

Authors' Contributions

Research concept: B.S., AZN.; MB.; EF. and H.R.; Execution of the experiments: EF; patient collection: AZN.; analysis of data: B.S., H.L., D.S.; contribution to the writing of manuscript: B.S., S.L. All Authors have read and agreed to the published version of the manuscript.

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