

Is raf-1 the 74 kDa signalling molecule in neutrophils?

The paper by Dusi et al. [1] reports the tyrosine phosphorylation of a number of proteins associated with neutrophil oxidase activation. Although the identity of MAP (mitogen-activated protein) kinase as a tyrosine-phosphorylation substrate in neutrophils may be secure, and in agreement with that reported by Thompson et al. [2], the identity of the 75 kDa protein as raf-1 (molecular mass 72-74 kDa) must remain questionable.

Activation of neutrophils by a number of agents, including many 'priming' agents such as $TNF-\alpha$ (tumour-necrosis factor α) and GMCSF (granulocyte-monocyte colony-stimulating factor) (which do so without any change in cytosolic free Ca^{2+} concentration), induces tyrosine phosphorylation of a number of proteins (reviewed in [3]). A common tyrosine-phosphorylated protein in a number of studies has a reported molecular mass of 72-76 kDa [3-5]. We have therefore postulated that this protein plays a key role in activation of the oxidase, by acting at a crosstalk site between tyrosine phosphorylation and Ca^{2+} signalling via serine/threonine phosphorylation [3-5]. Proteins, such as raf-1, whose activity is regulated by both serine/threonine and tyrosine phosphorylation are therefore attractive candidates. However, Grinstein et al. [6] report an inability to immunoprecipitate raf-l from neutrophils. We also have been unable to immunoprecipitate raf- ¹ from neutrophils, using either of two commercially available antibodies [4], one a monoclonal raised against the whole molecule (Oncogene Science, Uniondale, NY, U.S.A.) and the other a polyclonal raised against a peptide from raf-I (TCS Biologicals, Buckingham, U.K.). Johnson's group [7,8], who have reported detection of MEK-1 (MAP kinase kinase) phosphorylation activity in anti-raf-l immunoprecipitates in neutrophil lysates (using a polyclonal antibody raised against a peptide, from Santa Cruz Biotech), do not demonstrate that the immunoprecipitates contain 74 kDa protein. Dusi et al. [1] also fail to immunoprecipitate raf-1 from neutrophils (using the Santa Cruz Biotech antibody). Instead, they base their preference for identifying the 75 kDa protein as raf-1 on the 'exact coincidence' of superimposed blots for tyrosine phosphorylation and raf-I [1]. Because of the similarity in molecular masses, superimposition would be expected to occur, regardless of whether raf-I was the tyrosine-phosphorylation target. This can therefore provide no evidence that raf-1 becomes tyrosinephosphorylated during neutrophil activation.

Furthermore, the question arises whether 74 kDa raf-l is expressed in mature neutrophils. Unlike Dusi et al. [1], we cannot detect 74 kDa raf-1 in mature neutrophils by Western blotting, under conditions in which it can be readily identified in fibroblasts and the promyeloid cell line HL60 [4,9]. A cross-reacting protein of 56 kDa, however, was present. As HL60 cells are induced to differentiate towards granulocytes, there is a striking loss of detectable 74 kDa raf-1, but an increase in 56 kDa raf-like protein [9]. Despite the loss of 74 kDa raf-l during this time, signal coupling to the oxidase and to tyrosine phosphorylation of the 74 kDa target increases [9]. There was, however, a correlation between the increase in the 56 kDa raf-like protein and signal coupling.

The evidence is thus accumulating that a 74 kDa protein is an important signalling molecule in neutrophils, potentially a crosstalking point between tyrosine phosphorylation and Ca2+ induced serine-phosphorylation pathways which may underlie the mechanism of 'priming' of these cells [3]. We therefore agree with Dusi et al. [1] that tyrosine phosphorylation of a 74-75 kDa protein plays a key role in neutrophil activation, but suggest the identification of this protein as raf-l may be, at this stage, premature.

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- ¹ Dusi, S., Donini, M. and Rossi, F. (1994) Biochem. J. 304, 243-250
- 2 Thompson, H. L., Shiroo, M. and Saklatvala, J. (1993) Biochem. J. 290, 483-488
- 3 Halleft, M. B. and Lloyds, D. (1995) Immunol. Today 16, in the press
- Lloyds, D., Brindle, N. J. P. and Hallett, M. B. (1995) Immunology 84, 220-226
- 5 Lloyds, D. and Hallett, M. B. (1994) Biochem. Pharmacol. 48,15-23
- 6 Grinstein, S., Furaya, W., Butler, J. R. and Tseng, J. (1993) J. Biol. Chem. 268, 20223-20231
- 7 Worthen, G. S., Avdi, N., Buhl, A. M., Suzuki, N. and Johnson, G. L. (1993) J. Clin. Invest. 94, 815-823
- 8 Buhl, A. M., Avdi, N., Worthen, G. S. and Johnson, G. L. (1994) Proc. Natl. Acad. Sci. U.S.A. 91, 9190-9194
- 9 Lloyds, D. and Halleft, M. B. (1995) Biochim. Biophys. Acta, in the press

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