



Published in final edited form as:

Genes Chromosomes Cancer. 2024 January ; 63(1): e23206. doi:10.1002/gcc.23206.

When Molecular Outsmarts Morphology: Malignant Ossifying Fibromyxoid Tumors Masquerading as Osteosarcomas, Including a Novel *CREBZF::PHF1* Fusion

Aarti E Sharma^{1,2}, Josephine K Dermawan^{1,3}, Andy E Sherrod⁴, Shefali Chopra⁴, Robert G Maki⁵, Cristina R Antonescu¹

¹Department of Pathology and Laboratory Medicine, Memorial Sloan Kettering Cancer Center, New York, NY

²Department of Pathology and Laboratory Medicine, Hospital for Special Surgery, New York, NY

³Robert J. Tomsich Pathology and Laboratory Medicine Institute, Cleveland Clinic, Cleveland, OH

⁴Department of Pathology, University of Southern California, Keck School of Medicine, Los Angeles, CA

⁵Department of Medicine, Sarcoma Oncology, Memorial Sloan Kettering Cancer Center, New York, NY

Abstract

We present two cases of malignant ossifying fibromyxoid tumor (OFMT) which eluded diagnosis due to compelling clinicopathologic mimicry, compounded by similarly elusive underlying molecular drivers. The first is of a clavicle mass in a 69 year-old female, which histologically showed an infiltrative nested and trabeculated proliferation of monomorphic cells giving rise to scattered spicules of immature woven bone. Excepting SATB2 positivity, the lesion showed an inconclusive immunoprofile which along with negative *PHF1* FISH led to an initial diagnosis of high-grade osteosarcoma. Next generation sequencing revealed a particularly rare *CREBBP::BCORL1* fusion. The second illustrates the peculiar presentation of a dural-based mass in a 52 year-old female who presented with neurologic dyscrasias. Sections showed a sheeted monotonous proliferation of ovoid to spindle cells, but in contrast to Case #1, the tumor contained an exuberance of reticular osteoid and woven bone deposition mimicking malignant osteogenic differentiation. Next generation sequencing showed a novel *CREBZF::PHF1* fusion. Both tumors recurred locally less than one year post-operatively. As such we reiterate that careful morphologic examination is axiomatic to any diagnosis in this discipline, but this paradigm must shift to recognize that molecular diagnostics can provide closure where traditional tools have notable limitations.

Correspondence: Cristina R. Antonescu, MD, Department of Pathology and Laboratory Medicine, Memorial Sloan Kettering Cancer Center, 1275 York Ave, New York, NY, 10065, antonesc@mskcc.org.

Conflicts of Interest: The authors have disclosed that they have no significant relationships with, or financial interest in, any commercial companies pertaining to this article

Keywords

Malignant ossifying fibromyxoid tumor; fusion; *PHF1*; *CREBZF1*; *BCORL1*; *BCOR*; *CREBBP1*; Osteosarcoma

Introduction

Musculoskeletal oncologic literature describing ossifying fibromyxoid tumor (OFMT) has evolved over the years to recognize its recurrent molecular signature as well as its clinicopathologic and biologic heterogeneity. An entity of still ambiguous derivation, OFMT's inherently nonspecific immunoprofile along with its documentation in a variety of sites render it capable of masquerading as other benign as well as frankly malignant processes. Indeed, the diagnostic crux often rests upon molecular testing when all other tools – morphology included - have been exhausted.

The majority of OFMT pursue an innocuous clinical course with complete surgical excision. Local recurrence and distant metastases are observed rarely, a phenomenon usually confined to histologically 'atypical' or malignant subsets. In the latter case, morphology rather than molecular identity often predicts adverse behavior, although criteria across these variants are not well-defined - predicated on limited series and follow-up information.

Herein, we present the narrative of two clinicopathologically peculiar cases of malignant OFMT originally misclassified as high-grade osteosarcoma, including one of four reported cases primary to the brain which also harbored a heretofore undescribed *CREBZF::PHF1* fusion.¹⁻³ Pitfalls inherent to the differential diagnosis are discussed.

Case #1

A 69 year-old female presented with a right distal clavicle mass, appearing on pre-operative plain films as ill-defined mineralization over a portion of the distal clavicular circumference. Periosteal reaction of the superior cortex was observed, but there was otherwise no evidence of fracture. It was unclear at the time whether the lesion was intra- or extraosseous in etiology. The radiologic differential included osteosarcoma, tumoral calcinosis, or tophaceous pyrophosphate deposition (Figure 1).

Incisional biopsy showed an infiltrative soft tissue mass with alternating loose fibromyxoid to glassy hyaline stroma which imparted a distinctly trabeculated and nested cellular architecture (Figure 1). The lesional population was comprised by mildly atypical but overall monotonous epithelioid cells with variably abundant pale to clear cytoplasm and round nuclei with homogenous chromatin. A subset of cells showed a more fusiform morphology with amphophilic cytoplasmic processes. Embedded within the tumor were scattered minute foci of both unmineralized osteoid matrix elaborated by neoplastic cells, along with likely pre-existing remodeled lamellar bone with osteoblastic rimming and peripheral deposition of new matrix. Mitotic activity reached up to 10 per 10 high-power fields (all conventional forms), and necrosis was not identified.

Immunohistochemical stains demonstrated the cells to be diffusely and strongly positive for SATB2, with equivocal reactivity for synaptophysin and myosin heavy chain. Epithelial (EMA, AE1/AE3, CAM5.2), myoepithelial (GFAP, SOX10, S100, calponin, p63), melanocytic (HMB45, MelanA), and myogenic/myofibroblastic (SMA, desmin) markers were all negative. FISH studies were negative for rearrangements in *EWSR1*, *NR4A3*, and *PHF1* genes. Due to the non-specific immunoprofile with only potential evidence of osteoblastic differentiation, the rendered diagnosis was most consistent with high-grade osteosarcoma. To elucidate any possible therapeutic targets, the patient's oncologist subsequently requested that the tumor undergo comprehensive DNA Next generation sequencing, which revealed an underlying in-frame *CREBBP::BCORL1* fusion (Figure 3A). FISH analysis using custom BAC break-apart probes for *BCORL1* and *CREBBP* confirmed rearrangements in both genes.⁴⁻⁶ In the resulting chimeric transcript, exons 1-30 (of 31) of *CREBBP* (NM_004380.2) were fused to exons 4-12 of *BCORL1* (NM_021946.4) [fusion breakpoint chr16:3,781,193::chrX:129,154,960]. The predicted fusion protein contained most of the coding sequence of CREBBP, including the CREB-binding protein domain.

Resection of the mass was performed three months later, showing an 11.2 cm mass focally present at the surgical margins. Mitotic rate was 21 per 10 high-power fields, with approximately 10% overall necrosis. External beam radiation was recommended but declined by the patient. The tumor recurred 3 months later in the acromion, detected initially at a size of 4 cm which expanded rapidly over the course of a month to 9 cm. She refused chemotherapy and immunotherapy, and was subsequently lost to follow-up.

Case #2

A 52 year-old female presented with a sudden-onset history of sensorineural abnormalities over the left upper extremity, which progressed to posturing of the arm, shaking, and ultimately loss of consciousness. In the emergency room, brain MRI demonstrated a 2.6 cm dural-based ring-enhancing lesion – hypointense on T1 and T2 sequences – in the right fronto-parietal region with peripheral vasogenic edema (Figure 2). The lesion appeared entirely contained within the brain parenchyma without dural breach or osseous involvement of the inner skull plate.

The patient underwent craniotomy and resection of the mass. The specimen consisted of a portion of tan-grey dura with an adherent calcified mass. Cut surface was focally fleshy but otherwise firm with a gritty consistency. Low-power appearance of the tumor, in comparison to that of Case #1, was remarkable for an internal meshwork of anastomotic and patchily mineralized, coarsely trabecular woven bone (Figure 2), the periphery of which was focally infiltrative into glial tissues. The intervening ovoid to polygonal cells showed moderate atypia without pleomorphism, along with amphophilic cytoplasm, and pale speckled chromatin. They were arranged in syncytial, streaming sheets between and appositional to the bone, but also focally molded into discrete small nests and cords by a dense hyaline matrix. Acellular reticular osteoid was also extensive. Other disparate morphologies were not identified to suggest heterologous osteosarcomatous differentiation from another histologic primary. Mitotic activity was brisk (up to 22 per 10 high-power fields, including atypical forms), and necrosis was focally identified. Immunohistochemically, the tumor cells

were strongly positive for vimentin, with weak, patchy reactivity for NKX2.2. They were negative for OSCAR, AE1/AE3, EMA, GFAP, MART1, SOX10, SMA, desmin, ERG, and synaptophysin. Surgical margins were close but free of tumor. The final diagnosis was that of a high-grade extraskeletal osteosarcoma involving the brain.

FoundationOne® sequencing revealed an in-frame *CREBZF::PHF1* fusion (Figure 3B), along with a targetable *PIK3CA* mutation (E545G) and *CCND3* amplification. In the chimeric transcript, exon 1 (of 1) of *CREBZF* (NM_001039618.2) was fused to exons 2-14 of *PHF1* (NM_002636.4) [fusion breakpoint chr11:85,368,608::chr6:33,380,025]. In the predicted fusion protein, the entire coding region of CREBZF was fused to the entire coding region of PHF1. The tumor was microsatellite stable with a tumor mutational burden of 3 Muts/Mb. Post-operatively, the patient reported persistent numbness over the lateral aspects of the left side of her body (upper extremity more severe, most pronounced in the fourth and fifth digits), but otherwise was regaining strength and ambulation. She was followed serially with MRI scans. Eleven months after initial resection, imaging revealed a 1.1 cm recurrence along the dura at the anterior aspect of the prior craniotomy. This was resected and interpreted once again as high-grade osteosarcoma (not available for review). Fourteen months after initial presentation, the patient is alive with disease and being considered for radiation therapy.

Discussion

Ossifying fibromyxoid tumor (OFMT) is a soft tissue neoplasm characterized by recurrent *PHF1* and rarely, *BCOR* and *BCORL1* genetic rearrangements.^{4,7-9} Classically encountered as primary in the superficial soft tissues of the extremities and trunk with a recognized male predilection, OFMT can also infrequently present intraosseously.¹⁰ Radiologic features are generally nonspecific and can include an inconsistently high signal on fluid-sensitive sequences due to either a predominance of collagenous or myxoid to fibromyxoid matrix.¹¹ Plain films might demonstrate a sclerotic rind of bone peripheral to an otherwise radiolucent mass. Aggressive features including infiltration and adjacent cortical destruction (when deep and abutting bone) can be observed, and may simulate an osteogenic or Ewing sarcoma.

Histologically, conventional OFMT varies from a well-circumscribed solid mass to a more infiltrative, multinodular lesion. Appreciable on low-power, a histologic hallmark is a shell of peripheral bone which may be of woven or lamellar quality. This osseous tissue (conjectured as metaplastic rather than neoplastic in nature) can be completely absent or so exuberant as to obscure the underlying characteristic stromal elements and cellular architecture. Similar to other translocation-driven tumors, the constituent neoplastic population of OFMT is monomorphic with minimal frank atypia - predominantly ovoid or epithelioid but also spindled in some cases. Cells are arranged in patternless sheets to distinct cords and nests molded by intervening fibrocollagenous to myxoid matrix. OFMT may express S100 or desmin most frequently (more reliably in those with underlying *PHF1* rearrangement), along with unpredictable positivity for a host of other lineage- and or tumor-specific mesenchymal markers such as SMA, GFAP, cytokeratins, and even MUC4 and panTRK - rendering an overall nonspecific immunoprofile and definitive diagnosis consequently challenging on purely morphologic grounds.^{12,13} The continuum

from conventional to ‘atypical’ to ‘malignant’ OFMT is rather vaguely defined by a combination of severe atypia, increased cellularity with concomitant stromal rarefaction, and a proliferation index of >2 mitoses/50 high-power fields. Other attributes such as necrosis, destructive tissue invasion, and presence of metastasis, are calculated into this subjective gestalt.^{14–17}

Case #1 exemplifies the diagnostic challenges of OFMT at virtually any musculoskeletal site. The clavicle and its peripheral soft tissues are an uncommon location for any mesenchymal tumor, enabling perhaps a wider degree of plausible considerations – not the least of which was osteosarcoma. Complicating interpretation of the different qualities of bone matrix within the specimen was the uncertain location of the tumor – deriving from within the bone with soft tissue extension or vice versa. As such, fragments of both lamellar bone without features of remodeling and overlying matrix deposition, along with scattered islands of new bone formation produced by the cellular population could feasibly be interpreted as either a reparative/reactive, metaplastic, or genuinely neoplastic process. Together with this putatively neoplastic matrix, strong SATB2 positivity (a marker, albeit nonspecific, of osteoblastic differentiation which has not to our knowledge been systematically explored in the context of OFMT), and lack of any other convincing lineage, extraskeletal osteosarcoma became quite a plausible diagnosis of exclusion.

Areas of myxoid stroma with architecturally distinct clusters to trabeculae of ovoid cells, however, are typically not observed in osteosarcomas of any phenotype, and were rather more evocative of a myoepithelial neoplasm, extraskeletal myxoid chondrosarcoma, or *BCOR*-rearranged sarcoma. While the first two can be positive for S100 (like OFMT), salient ossification is not a characteristic feature of any of these three. Absence of immunohistochemical evidence of myoepithelial differentiation (cytokeratins, EMA, SOX10, GFAP, SMA) and lack of *EWSR1* rearrangement effectively excluded a malignant myoepithelial tumor.¹⁸ Extraskeletal myxoid chondrosarcoma can also show similar cytologic plasticity and structured architecture within glassy fibromyxoid matrix; they are otherwise characterized by a canonical *NR4A3* rearrangement (usually *EWSR1::NR4A3* fusion),¹⁹ which can be queried with FISH or RNA sequencing. Finally, we include *BCOR*-altered sarcoma as a potent diagnostic pitfall in most intraosseous malignancies of children and adolescents, displaying a distinct myxoid matrix populated by a variable combination of monomorphic ‘small round cells’ along with short fascicles of spindle cells. Like some OFMT, these should be unequivocally positive for the *BCOR* immunostain, and likewise for *BCOR* FISH – which can justifiably misdirect the diagnostic conclusion.²⁰

Indeed, OFMT might have been a strong contender synthesizing the body of evidence, but the negative FISH study for *PHF1* rearrangement (the most common gene involved in to 85% of these tumors) unfortunately deterred the diagnosis. Notably, comprehensive sequencing which led to the correct diagnosis and amended report, was requested for therapeutic rather than diagnostic purposes. Although in many settings this modality is financially prohibitive for solely the latter, the case highlights clinicopathologic inconsistencies which are occasionally reconciled only via comprehensive genomic interrogation. In a situation such as this where the diagnosis of OFMT is suspected but thwarted by negative *PHF1* FISH, FISH or even immunohistochemistry for *BCOR*

are alternative and less expensive tests that may confirm this less common fusion partner. To complicate matters further, *BCOR*-rearranged OFMT are distinct in that not only do they pursue a worse clinical trajectory, but also consistently show variant malignant histology without expression of S100 or desmin characteristic of their *PHF1*-rearranged counterparts.^{5,6,13,21} In contrast, one of the two previously described cases of *CREBBP::BCORL1*-fusion OFMT showed conventional ‘benign’ histology with rare S100-positive cells and negative desmin, which involved the same genetic breakpoint as that detected in the current case.⁶ The other *CREBBP::BCORL1*-fusion OFMT was considered malignant by virtue of brisk mitotic activity, necrosis, peripheral infiltration, and lymphovascular invasion; it also was focally positive for S100 and negative for desmin.⁶

Case #2 stands alone as an addition to the OFMT literature considering not only one but two singular features: 1) intracranial epicenter (only a few of which have ever been documented in the repertoire of this already uncommon tumor), and 2) underlying *CREBZF::PHF1* fusion, the latter of which has (to our knowledge) never been described in the context of OFMT. While bone tumors are not a first-line consideration when faced with an intracranial lesion, extensive ossification predominantly in the form of lace-like mineralized osteoid resembling that generated by a high-grade conventional osteoblastic osteosarcoma would certainly redirect diagnostic considerations (Figure 2). Perhaps reflective of the novel fusion, this particular matrix quality is not an expected feature of even atypical or malignant OFMT as outlined in the earliest series by Enzinger and Weiss (nor those subsequent). As mentioned, rearrangement of *PHF1* is characteristic of OFMT aligned with an assortment of partners including *EP400*, *EPC1*, and *TFE3*.^{4,8,21,22} Including these, other uncommon gene partners underlying OFMT share the common thread of involvement in histone modification. In contrast, the *CREBZF* locus encodes for a transcription factor which, among other regulatory roles, is involved in stabilization and transcriptional enhancement of the tumor suppressor TP53. It has only ever been implicated once in fusion-associated tumorigenesis prior to this case – a bladder urothelial carcinoma with a *CD44::CREBZF* fusion.²³

Admittedly, these two cases showed quite contrasting architectural features, but both demonstrated a critical cytomorphologic commonality: nuclear uniformity, which should intimate the presence of an underlying fusion-driven pathophysiology. Regardless of the subtype, most intra-osseous and extraskeletal osteosarcomas instead harbor complex genomic alterations as nonrecurrent copy number alterations and mutations, which translates to histology as frank sarcoma in the form of pleomorphism, high-grade nuclear atypia, and abnormal mitotic activity. With rare exceptions, osteosarcoma is not characterized by cytologic monotony disproportionate to other features of malignancy including destructive permeation, geographic coagulative necrosis, and neoplastic matrix deposition. It should be noted that all the aforementioned features of malignancy characteristic of osteosarcoma can be observed in ‘atypical’ and malignant OFMT, although none in isolation are necessary or sufficient for this qualification. These subsets have been described only in a few series, and as such our threshold for and understanding of this entity is somewhat tenuous.^{22, 23}

Conclusion

We describe two unique cases and expand the molecular profile of the already enigmatic ossifying fibromyxoid tumor, which were confounded not only by their clinicopathologic peculiarities – both elderly female patients, one with a clavicular and the other with a brain-based tumor and radiology favoring a malignant bone-forming lesion – but also by their documented but uncommon genomic identities. As molecular testing continues to more optimally partition historic mimics, knowledge of histologic overlap exemplified by the above narratives becomes crucial towards initiating these more granular examinations of tumor biology.

References:

- 1). Pisapia DJ, Ohara K, Bareja R, et al. Fusions involving BCOR and CREBBP are rare events in infiltrating glioma. *Acta Neuropathol Com-mun.* 2020;8(1):80. doi:10.1186/s40478-020-00951-4
- 2). Yamazaki A, Arai Y, Fukuoka K, et al. Diffusely infiltrating glioma with CREBBP-BCORL1 fusion showing overexpression of not only BCORL1 but BCOR: a case report. *Brain Tumor Pathol.* 2022;39(3):171–178. doi:10.1007/s10014-022-00435-4 [PubMed: 35596897]
- 3). Beyer S, Sebastian NT, Prasad RN, et al. Malignant ossifying fibromyxoid tumor of the brain treated with post-operative fractionated stereotactic radiation therapy: a case report and literature review. *SurgNeurol Int.* 2021;12:588. doi:10.25259/SNI_827_2021
- 4). Suurmeijer AJH, Song W, Sung YS, et al. Novel recurrent PHF1-TFE3 fusions in ossifying fibromyxoid tumors. *Genes Chromosomes Cancer.* 2019;58(9):643–649. doi:10.1002/gcc.22755 [PubMed: 30920708]
- 5). Antonescu CR, Sung YS, Chen CL, et al. Novel ZC3H7B-BCOR, MEAF6-PHF1, and EPC1-PHF1 fusions in ossifying fibromyxoid tumors—molecular characterization shows genetic overlap with endometrial stromal sarcoma. *Genes Chromosomes Cancer.* 2014;53(2):183–193. doi:10.1002/gcc.22132 [PubMed: 24285434]
- 6). Kao YC, Sung YS, Zhang L, Chen CL, Huang SC, Antonescu CR. Expanding the molecular signature of ossifying fibromyxoid tumors with two novel gene fusions: CREBBP-BCORL1 and KDM2A-WWTR1. *Genes Chromosomes Cancer.* 2017;56(1):42–50. doi:10.1002/gcc.22400 [PubMed: 27537276]
- 7). Graham RP, Weiss SW, Sukov WR, et al. PHF1 rearrangements in ossifying fibromyxoid tumors of soft parts: a fluorescence in situ hybridization study of 41 cases with emphasis on the malignant variant. *Am J Surg Pathol.* 2013;37(11):1751–1755. doi:10.1097/PAS.0b013e31829644b4 [PubMed: 23887158]
- 8). Gebre-Medhin S, Nord KH, Möller E, et al. Recurrent rearrangement of the PHF1 gene in ossifying fibromyxoid tumors. *Am J Pathol.* 2012;181(3):1069–1077. doi:10.1016/j.ajpath.2012.05.030 [PubMed: 22796436]
- 9). Srivastava P, Zilla ML, Naous R, et al. Expanding the molecular signatures of malignant ossifying fibromyxoid tumours with two novel gene fusions: PHF1::FOXR1 and PHF1::FOXR2. *Histopathology.* 2023;82(6):946–952. doi:10.1111/his.14868 [PubMed: 36648026]
- 10). Sbaraglia M, Bellan E, Gambarotti M, et al. Primary malignant ossifying fibromyxoid tumour of the bone. A clinicopathologic and molecular report of two cases. *Pathologica.* 2020;112(4):184–190. doi:10.32074/1591-951X-207 [PubMed: 33179613]
- 11). Finkelstein D, Foremny G, Singer A, et al. Differential diagnosis of T2 hypointense masses in musculoskeletal MRI. *Skeletal Radiol.* 2021;50(10):1981–1994. doi:10.1007/s00256-021-03711-0 [PubMed: 33651128]
- 12). Graham RP, Dry S, Li X, et al. Ossifying fibromyxoid tumor of soft parts: a clinicopathologic, proteomic, and genomic study. *Am J Surg Pathol.* 2011;35(11):1615–1625. doi:10.1097/PAS.0b013e3182284a3f [PubMed: 21997683]

- 13). Linos K, Kerr DA, Sumegi J, Bridge JA. Pan-Trk immunoexpression in a superficial malignant ossifying fibromyxoid tumor with ZC3H7B-BCOR fusion: a potential obfuscating factor in the era of targeted therapy. *J Cutan Pathol.* 2021;48(2):340–342. doi:10.1111/cup.13915 [PubMed: 33169434]
- 14). Enzinger FM, Weiss SW, Liang CY. Ossifying fibromyxoid tumor of soft parts. A clinicopathological analysis of 59 cases. *Am J Surg Pathol.* 1989;13(10):817–827. doi:10.1097/0000478-198910000-00001 [PubMed: 2476942]
- 15). Folpe AL, Weiss SW. Ossifying fibromyxoid tumor of soft parts: a clinicopathologic study of 70 cases with emphasis on atypical and malignant variants. *Am J Surg Pathol.* 2003;27(4):421–431. doi:10.1097/0000478-200304000-00001 [PubMed: 12657926]
- 16). Dantey K, Schoedel K, Yergiyev O, McGough R, Palekar A, Rao UNM. Ossifying fibromyxoid tumor: a study of 6 cases of atypical and malignant variants. *Hum Pathol.* 2017;60:174–179. doi:10.1016/j.humpath.2016.10.012 [PubMed: 27816723]
- 17). Kilpatrick SE, Ward WG, Mozes M, Miettinen M, Fukunaga M, Fletcher CD. Atypical and malignant variants of ossifying fibromyxoid tumor. Clinicopathologic analysis of six cases. *Am J Surg Pathol.* 1995;19(9):1039–1046. doi:10.1097/0000478-199509000-00007 [PubMed: 7661277]
- 18). Hornick JL, Fletcher CD. Myoepithelial tumors of soft tissue: a clinicopathologic and immunohistochemical study of 101 cases with evaluation of prognostic parameters. *Am J Surg Pathol.* 2003;27(9):1183–1196. doi:10.1097/0000478-200309000-00001 [PubMed: 12960802]
- 19). Suurmeijer AJH, Dickson BC, Swanson D, et al. A morphologic and molecular reappraisal of myoepithelial tumors of soft tissue, bone, and viscera with EWSR1 and FUS gene rearrangements. *Genes Chromosomes Cancer.* 2020;59(6):348–356. doi:10.1002/gcc.22835 [PubMed: 31994243]
- 20). Kao YC, Owosho AA, Sung YS, et al. BCOR-CCNB3 fusion positive sarcomas: a clinicopathologic and molecular analysis of 36 cases with comparison to morphologic spectrum and clinical behavior of other round cell sarcomas. *Am J Surg Pathol.* 2018;42(5):604–615. doi:10.1097/PAS.0000000000000965 [PubMed: 29300189]
- 21). Linos K, Kerr DA, Baker M, et al. Superficial malignant ossifying fibromyxoid tumors harboring the rare and recently described ZC3H7B-BCOR and PHF1-TFE3 fusions. *J Cutan Pathol.* 2020;47(10):934–945. doi:10.1111/cup.13728 [PubMed: 32352579]
- 22). Schneider N, Fisher C, Thway K. Ossifying fibromyxoid tumor: morphology, genetics, and differential diagnosis. *Ann Diagn Pathol.* 2016;20:52–58. doi:10.1016/j.anndiagpath.2015.11.002 [PubMed: 26732302]
- 23). Gao Q, Liang WW, Foltz SM, et al. Driver fusions and their implications in the development and treatment of human cancers. *Cell Rep.* 2018;23(1):227–238.e3. doi:10.1016/j.celrep.2018.03.050 [PubMed: 29617662]

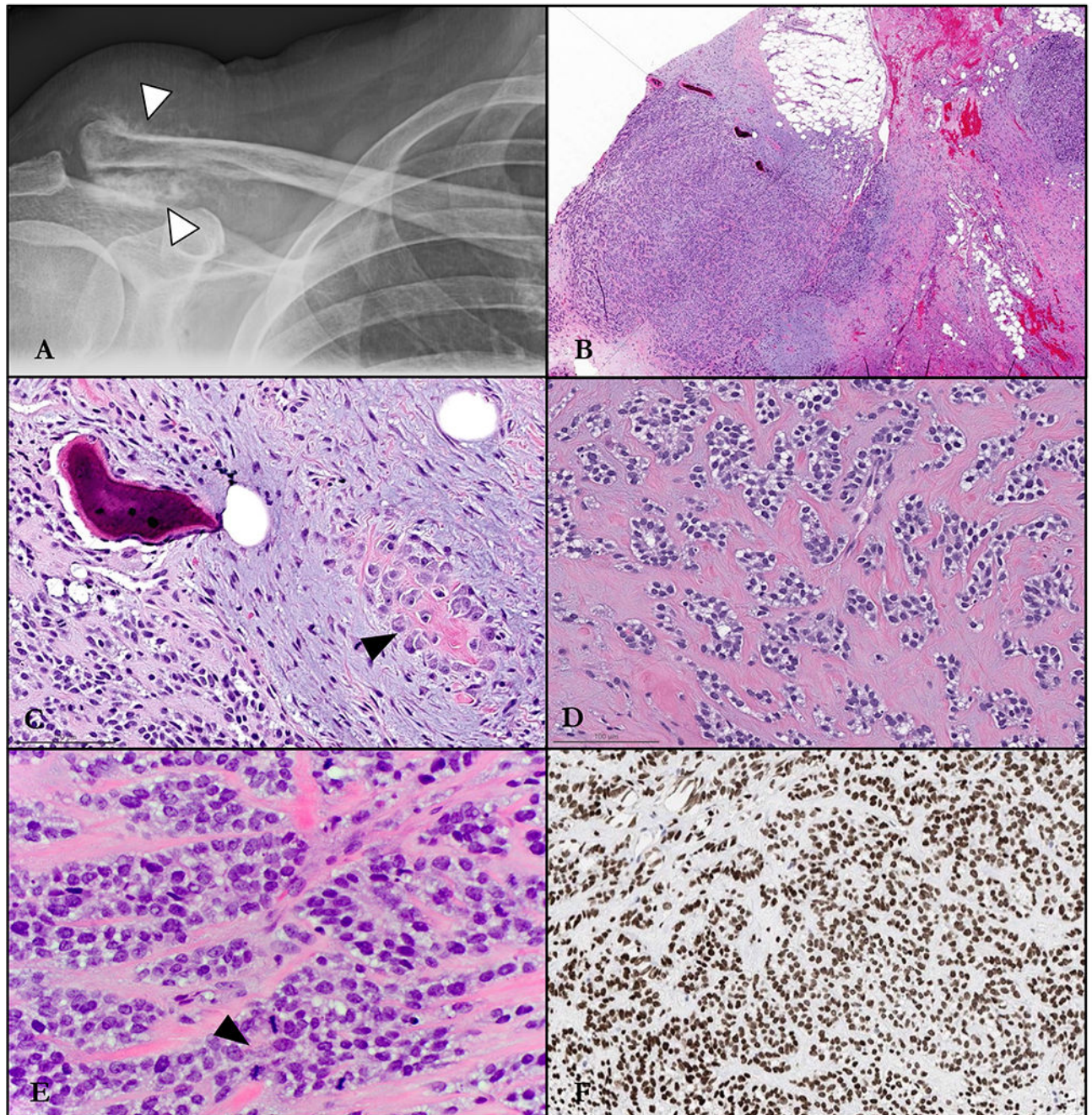


Figure 1.

Radiology and histomorphology of Case #1. **A.** Plain film showing slight lucency within and flocculent calcifications surrounding the distal clavicle (arrowhead). **B.** Destructive infiltration into soft tissue and bone. **C.** Spicule of immature matrix (arrowhead) elaborated by neoplastic ovoid cells which are embedded in a dense myxoid stroma containing shards of lamellar bone. **D.** Irregular nests of epithelioid cells moulded by a fibrocollagenous matrix. **E.** Nested and corded epithelioid cells with increased mitotic activity (arrowhead).

F. Immunohistochemical stain for SATB2, showing strong and unequivocal positivity in the trabeculated tumor cells.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

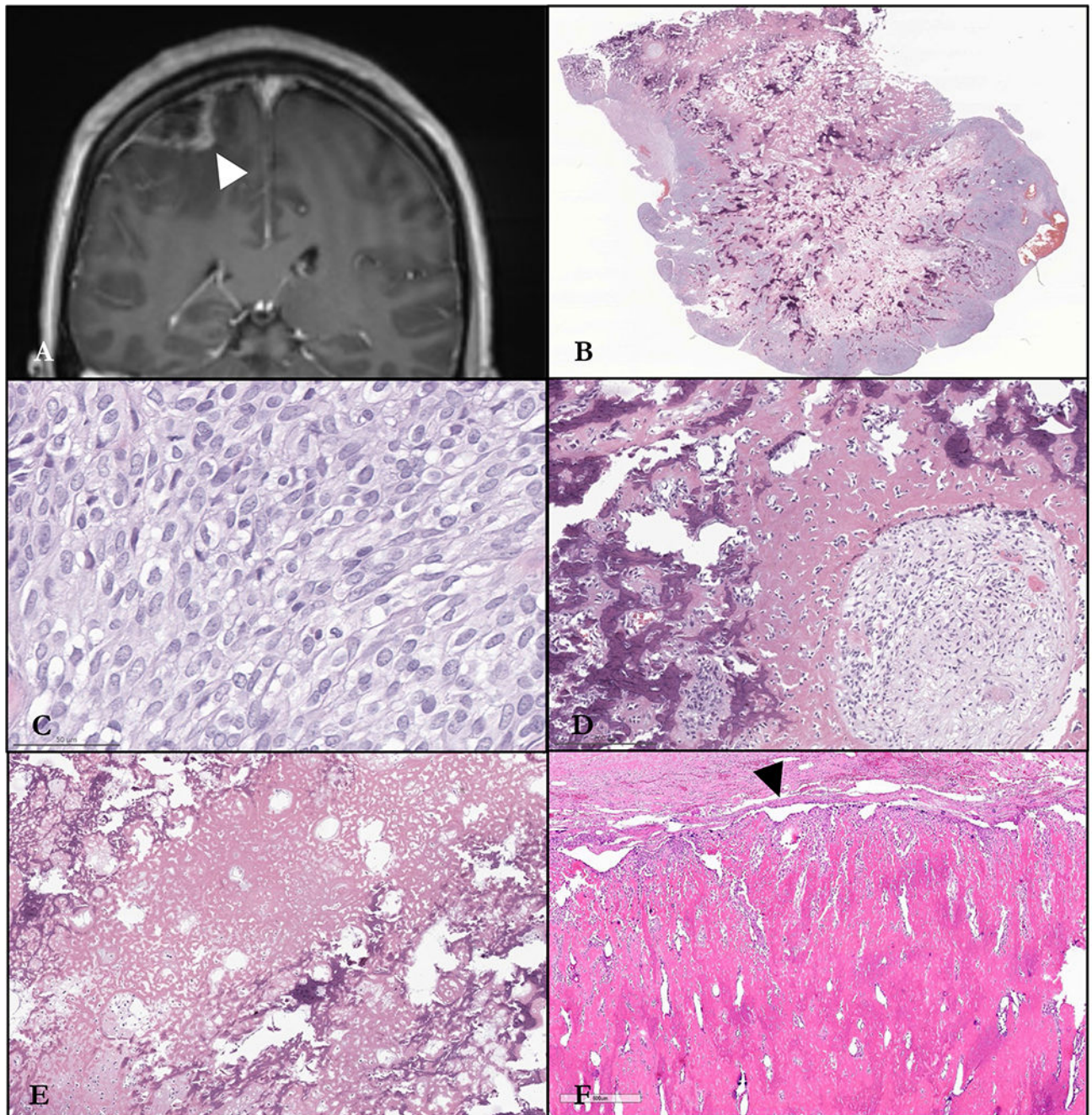


Figure 2. Radiology and histomorphology of Case #2. **A.** T1 coronal MRI showing a hypointense frontoparietal dural-based ring-enhancing lesion. **B.** Low-power image illustrating central and peripheral mineralized bony trabeculae. **C.** Syncytial sheets of monomorphic ovoid to round cells with pale amphophilic cytoplasm. **D.** Thick anastomotic woven bone with neoplastic cells in myxoid matrix. **E.** Extensive deposition of reticular osteoid matrix reminiscent of a high-grade osteoblastic osteosarcoma. **F.** Dural-based recurrence eroding

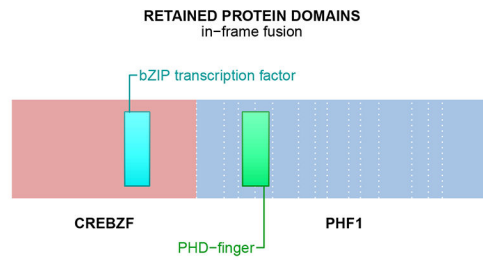
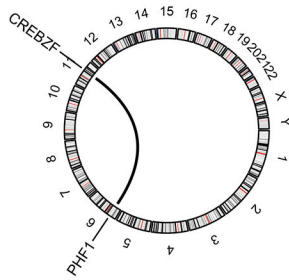
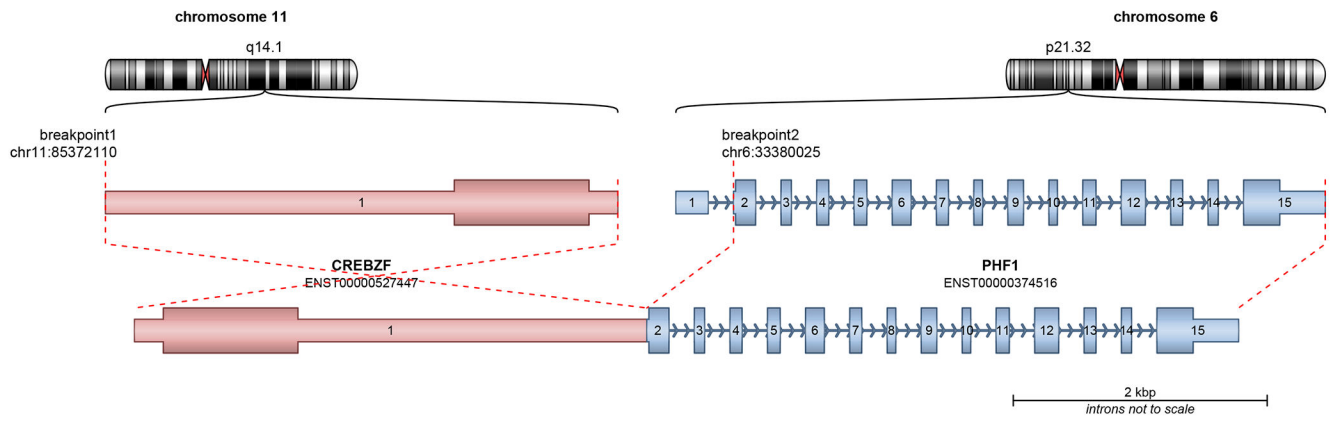
into the skull plate, with extensive deposition of sclerotic bone matrix with a peripheral rind of neoplastic cells (arrowhead).

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript



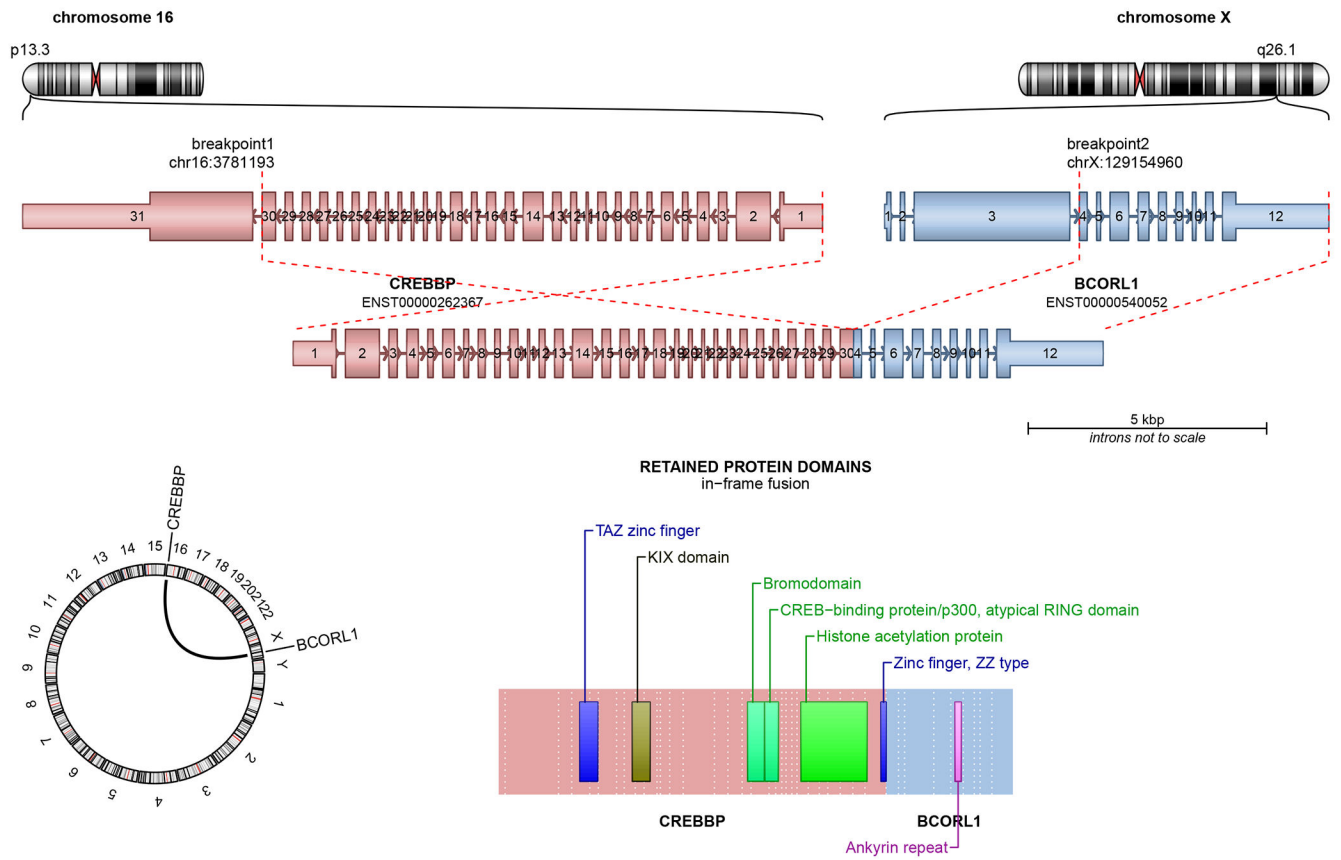


Figure 3. Structural schematics of CREBZF::PHF1 (top) and CREBBP::BCORL1 (bottom) fusions in the malignant ossifying fibromyxoid tumors.