

RESEARCH

Open Access

\*Correspondence:



# Relation between serum sclerostin and CTRP3 levels and bone mineral density in diabetic postmenopausal women

Inass Hassan Ahmad<sup>1</sup>, Sally Said Abd Elhamed Gbr<sup>2</sup>, Basma Mohamed Mohamed Ali El Naggar<sup>3</sup>, Marwa Khairy Abdelwahab<sup>4</sup>, Entesar Omar Ahmad El-saghier<sup>1</sup>, Doaa Sayed Mohammed<sup>1</sup>, Marwa Abdelmonim Mohamed<sup>2</sup>, Maha S. Mohamed<sup>3</sup>, Marwa Mohamed M. Ali Abd El-Rahim<sup>3</sup> and Shahinaz El Attar<sup>5\*</sup>

## Abstract

**Background** Osteoporosis (OP) is a common finding in diabetic patients especially high-risk populations such as postmenopausal women. Sclerostin is a glycoprotein chiefly secreted by mature osteocytes and is considered a main regulator of bone formation. The C1q/TNF-Related Protein 3 (CTRP3) was found to be significantly associated with OP in postmenopausal women. The effect of type 2 diabetes mellitus (T2DM) on sclerostin and CTRP3 levels in postmenopausal women is rarely investigated. The present study aimed to assess the impact of T2DM on sclerostin and CTRP3 levels and their relation to OP in postmenopausal women.

**Methods** The study included 60 postmenopausal women with T2DM and 60 age-matched postmenopausal non-diabetic women. Bone mineral density (BMD) was assessed using dual energy X-ray absorptiometry (DEXA). Serum levels of sclerostin and CTRP3 were assessed using enzyme-linked immunosorbent assay (ELISA) technique.

**Results** Diabetic group expressed significantly higher serum levels of sclerostin when compared with non-diabetic group ( $110.0 \pm 29.0$  versus  $51.5 \pm 23.2$  ng;  $p < 0.001$ ). Oppositely, CTRP3 were significantly lower in the diabetic group ( $3.5 \pm 3.5$  versus  $9.9 \pm 3.7$  ng/ml,  $p < 0.001$ ). Multivariate logistic regression analysis identified HbA1c levels [OR (95% CI): 0.49 (0.26–0.93),  $p = 0.028$ ], sclerostin levels [OR (95% CI): 1.06 (1.0–1.012),  $p = 0.041$ ] and CTRP3 levels [OR (95% CI): 1.64 (1.0–2.68),  $p = 0.047$ ] as significant predictors of OP in diabetic patients.

**Conclusions** Sclerostin and CTRP3 levels are involved in OP in postmenopausal diabetic patients.

**Keywords** Type 2 diabetes mellitus, Osteoporosis, CTRP3, Sclerostin

Shahinaz El Attar

shahinazattar@gmail.com

<sup>1</sup>Endocrinology and Metabolism Department, Faculty of Medicine for Girls, Al-Azhar University, Cairo, Egypt

<sup>2</sup>Internal Medicine Department, Faculty of Medicine for Girls, Al-Azhar University, Cairo, Egypt

<sup>3</sup>Rheumatology and Rehabilitation Department, Faculty of Medicine for Girls, Al-Azhar University, Cairo, Egypt

<sup>4</sup>Clinical Pathology Department, Faculty of Medicine for Girls, Al-Azhar University, Cairo, Egypt

<sup>5</sup>Medical Biochemistry and Molecular Biology Department, Faculty of Medicine for Girls, Al-Azhar University, Cairo, Egypt



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## Background

Type 2 diabetes mellitus (T2DM) is one of the most prevalent non-communicable diseases worldwide that substantially increase the risk of cardiovascular diseases, angiopathies, and various metabolic disorders [1–4]. Osteoporosis (OP) is a common finding in diabetic patients [5], especially high-risk populations such as postmenopausal women [6].

Sclerostin is a glycoprotein chiefly secreted by mature osteocytes and is considered a main regulator of bone formation through its effect on Wnt signaling [7] making its circulating level a promising biomarker for the diagnosis and prognosis of OP [8]. However, previous studies presented controversial results regarding the relation between sclerostin levels and OP [9–11].

In diabetic patients, the adipose tissue plays a vital role by secreting bioactive molecules called adipokines [12]. The chronic low-grade inflammation mediated by adipokines was shown as a potential therapeutic target for management of diabetic complications [13]. For instance, circulating levels of adiponectin are downregulated in T2DM patients. It is considered as the most beneficial adipokine in circulation, improving insulin sensitivity, endothelial functions, and inflammation [14].

The C1q/TNF-Related Protein (CTRP) family members are paralogues of adiponectin. They share favorable effects on inflammation, insulin sensitivity, and lipid metabolism [15]. Several studies observed the association between expression of CTRP family members and diabetes, coronary artery diseases, metabolic syndrome, non-alcoholic fatty liver disease, and polycystic ovary syndrome [16–18]. CTRP3 activates adenosine monophosphate-activated protein kinase (AMPK), thus enhancing insulin signaling and sensitivity [19]. Decline of circulating CTRP3 levels was linked to insulin resistance in diabetic patients and rise of its levels was noted after treatment with glucagon-like peptide-1 (GLP-1) receptor agonists [20]. Recently, serum CTRP3 was found to be significantly associated with OP in postmenopausal women [21]. The effect of T2DM on sclerostin and CTRP3 levels in postmenopausal women is rarely investigated. The present study aimed to assess the impact of T2DM on sclerostin and CTRP3 levels and their relation to OP in postmenopausal women.

## Methods

The present cross-sectional study was conducted at Al-Azhar University Hospitals, Cairo, Egypt. The study protocol was approved by the local ethical committee and all participants provided informed consent before enrollment. The study included 60 postmenopausal women with T2DM and 60 age-matched postmenopausal non-diabetic women. Menopause was identified through a history of menstruation cessation for at least one year

prior to enrollment. The diagnosis of T2DM was based on the criteria of the American Diabetes Association (ADA) [22]. We excluded women with diabetic nephropathy, morbid obesity, familial dyslipidemia, organ failure, malignancies, thyroid disorder, history of hormonal therapy, and/or associated immunological disorders. Patients were also excluded if they received medications known to impair bone metabolism e.g. thiazolidinediones or if they were under OP treatment.

Patients included in the study were submitted to careful history taking and through clinical examination. For biochemical assessment, 10 ml of venous blood were obtained from each participant after 8-hour fasting. Each collected sample was divided into two parts; 7 ml part for routine investigations and the other part was centrifuged for 10 min at 4000 rpm and was stored at  $-80^{\circ}\text{C}$ . Colorimetric enzymatic approaches were used for estimation of the lipid profile and blood glucose profile using Hitachi autoanalyzer 704 (Roche Diagnostics, Switzerland).

Automated Glycohemoglobin Analyzer (Tosoh Bioscience's HLC-723GX@, Tosoh, India) was used to estimate the HbA1c in blood. Chemiluminescent immunoassay (Immulate2000, Siemens, Germany) was utilized to assess serum insulin from blood samples. The following formula was used to estimate HOMA-IR:  $\text{HOMA-IR} = \text{fasting insulin (IU/mL)} \times \text{plasma glucose (mg/dL)} / 405$  [23].

Automated ELIZA (Thermo Scientific, Finland) and computer program (Scanlt for Multiscan FC 2.5.1) were used to measure serum levels of CTRP3. The device was set for CTRP3 sensitivity of 0.38 ng/ml. Serum sclerostin levels were assessed utilizing quantitative sandwich ELISA (Biomedica, Vienna, Austria).

Bone mineral density (BMD) was assessed through dual energy X-ray absorptiometry (Lunar Prodigy; General Electric Medical Systems; USA) at the level of the femur neck and L2-L4 spines. The diagnosis of OP was based on the findings of BMD, in which a T-score of less than  $-2.5$  was used as a cutoff value for the presence of OP; while a T-score ranging from  $-1$  to  $-2.5$  was used a definition of osteopenia.

Data analysis was conducted by SPSS software, version 22.0 (SPSS Inc., Chicago, Illinois, USA). Numerical variables were expressed as number and percent and compared using t test while categorical data were presented as number and percent and compared using Fisher's exact test or Chi-square test as appropriate. Correlation analysis was achieved using Pearson's correlation coefficient. Binary logistic regression was used to identify predictors of OP. *p* value less than 0.05 was considered statistically significant.

**Table 1** Comparison between groups according to demographic and laboratory data

	Diabetic group n=60	Non-diabetic group n=60	p-value
Age years	53.4±7.7	56.1±1.5	0.102
SBP mmHg	125.3±15.0	112.3±4.3	<0.001
DBP mmHg	81.3±10.1	73.7±4.9	<0.001
BMI kg/m <sup>2</sup>	33.5±4.6	29.1±3.0	<0.001
FBS mg/dL	207.0±68.0	89.2±8.0	<0.001
PPBS mg/dL	261.7±88.0	122.2±11.7	<0.001
HbA1c %	9.0±2.0	5.4±0.3	<0.001
Cholesterol mg/dL	207.3±49.2	182.6±23.6	<0.001
Triglycerides mg/dL	168.5±62.7	119.2±20.5	<0.001
HDL mg/dL	38.7±5.8	36.9±6.4	0.004
LDL mg/dL	139.4±44.8	122.8±21.3	<0.001
Insulin IU/mL	15.4±5.8	6.9±1.5	<0.001
HOMA-IR	6.6±2.1	1.5±0.4	<0.001
Sclerostin ng/ml	110.0±29.0	51.5±23.2	<0.001
CTRP3 ng/ml	3.5±3.5	9.9±3.7	<0.001
DEXA scan n (%)			
Normal	14 (23.3)	30 (50.0)	0.007
Osteopenia	18 (30.0)	15 (25.0)	
Osteoporosis	28 (46.7)	15 (25.0)	

BMI: Body mass index, CTRP3: C1q/TNF-Related Protein 3, DBP: Diastolic blood pressure, DEXA: dual energy X-ray absorptiometry, FBS: Fasting blood sugar, HDL: High-density lipoprotein, HOMA-IR: Homeostatic model assessment for insulin resistance, LDL: Low-density lipoprotein, PPBS: Postprandial blood sugar, SBP: Systolic blood pressure

**Results**

The present study included 60 diabetic postmenopausal women and 60 age matched postmenopausal non-diabetic women. Clinical findings in the studied groups are shown in tabl-1. Diabetic group expressed significantly higher serum levels of sclerostin when compared with non-diabetic group (110.0±29.0 versus 51.5±23.2 ng, *p*<0.001). Oppositely, CTRP3 were significantly lower

in the diabetic group (3.5±3.5 versus 9.9±3.7 ng/ml, *p*<0.001). BMD analysis identified osteopenia and OP in 30.0% and 46.7% respectively in the diabetic group while in the non-diabetic group, osteopenia and OP were identified in 25.0% and 25.0% of women respectively (*p*=0.007) (Table 1).

In the diabetic group, correlation analysis identified significant correlation between serum CTRP3 levels and age (*r*=0.369, *p*=0.045), SBP (*r*=0.369, *p*=0.045), DBP (*r*=0.328, *p*=0.037) and insulin (*r*=0.612, *p*=0.009). Also, there were significant correlations between serum sclerostin levels and HbA1c (*r*=-0.307, *p*=0.049) and HOMA-IR (*p*=-0.732, *p*<0.001). In the non-diabetic group, there were significant correlations between serum CTRP3 levels and age (*r*=0.683, *p*<0.001), SBP (*r*=0.557, *p*<0.001), FBS (*r*=0.632, *p*<0.001), HbA1c (*r*=-0.460, *p*=0.011), HDL (*r*=0.721, *p*<0.001), LDL (*r*=-0.477, *p*=0.008), insulin (*r*=0.466, *p*=0.010) and HOMA-IR (*r*=0.611, *p*<0.001). Also, there were significant correlations between sclerostin levels and age (*r*=-0.392, *p*=0.032), DBP (*r*=-0.411, *p*=0.024), FBS (*r*=-0.723, *p*<0.001), PPBS (*r*=-0.510, *p*=0.004), cholesterol (*r*=0.40, *p*=0.028), HDL (*r*=-0.680, *p*<0.001), LDL (*r*=0.548, *p*=0.002) and HOMA-RI (*r*=-0.399, *p*=0.029) (Table 2).

Multivariate logistic regression analysis identified HbA1c levels [OR (95% CI): 0.49 (0.26–0.93), *p*=0.028], sclerostin levels [OR (95% CI): 1.06 (1.0-1.012), *p*=0.041] and CTRP3 levels [OR (95%) CI: 1.64 (1.0-2.68), *p*=0.047] as significant predictors of OP in diabetic patients (Table 3).

**Discussion**

While the current published literature demonstrates significant association between the serum sclerostin and CTRP3 with OP, little is known about the additional

**Table 2** Correlation between CTRP3 and sclerostin levels and clinical data

	Diabetic group				Non-diabetic group			
	CTRP3		Sclerostin		CTRP3		Sclerostin	
	r	p-value	r	p-value	r	p-value	r	p-value
Age (years)	0.369	0.045	0.095	0.619	0.683	<0.001	-0.392	0.032
BMI	0.260	0.166	0.004	0.983	-0.300	0.107	0.055	0.773
Disease duration	-0.302	0.105	-0.009	0.964	-	-	-	-
SBP	0.369	0.045	0.171	0.367	0.557	<0.001	-0.220	0.243
DBP	0.328	0.037	0.132	0.487	0.207	0.273	-0.411	0.024
FBS	0.137	0.470	-0.205	0.277	0.632	<0.001	-0.723	<0.001
PPBS	0.013	0.947	-0.131	0.490	0.207	0.271	-0.510	0.004
HbA1c	0.010	0.957	-0.307	0.049	-0.460	0.011	0.113	0.552
Cholesterol	-0.112	0.554	-0.075	0.693	-0.248	0.186	0.400	0.028
Triglycerides	0.121	0.525	0.059	0.757	0.122	0.519	0.042	0.827
HDL	0.060	0.753	-0.106	0.577	0.721	<0.001	-0.680	<0.001
LDL	-0.232	0.217	0.012	0.951	-0.477	0.008	0.548	0.002
Insulin	0.612	0.009	-0.286	0.265	0.466	0.010	-0.174	0.356
HOMA-IR	0.046	0.859	-0.732	<0.001	0.611	<0.001	-0.399	0.029

**Table 3** Predictors of osteoporosis in diabetic patients

	Univariate analysis			Multivariate analysis		
	OR	95% CI	p value	OR	95% CI	p value
Age	1.16	1.04–1.31	0.011	1.12	0.87–1.45	0.36
Disease duration	1.21	1.08–1.35	0.001	1.09	0.87–1.36	0.46
HbA1c	0.53	0.38–0.75	<0.001	0.49	0.26–0.93	0.028
Sclerostin	1.06	1.03–1.1	<0.001	1.06	1.0–1.012	0.041
CTRP3	2.07	1.41–3.03	<0.001	1.64	1.0–2.68	0.047

impact of T2DM on these biomarkers. In the present study, multivariate logistic regression analysis identified sclerostin levels and CTRP3 levels as significant predictors of OP in diabetic patients.

Recently, emerging evidence highlighted a significant role of serum CTRP3 in regulation of bone hemostasis; the role of CTRP3 in regulation of bone structure appears to stem from its ability to maintain normal turnover of chondrocytes and cartilaginous structure through regulation of ERK1/2 and PI3K pathways [24, 25]. Thus, authors linked downregulation of serum CTRP3 to defective bone metabolism and features of OP [26]. On the other hand, the association between CTRP3 and T2DM is well-established with reported decline in serum CTRP3 levels among cases with insulin resistance and poor glycemic control [20]. Our analysis demonstrated that serum CTRP3 was independent predictor of OP in T2DM women and correlated significantly with metabolic parameters. To our knowledge, this is the first report that addressed the impact of T2DM on serum CTRP3 among women with OP.

The impact of T2DM on CTRP3 levels can be mediated through multiple mechanisms. Previous studies found an association between dyslipidemia and reduced CTRP3 levels in diabetic patients [27, 28]. Also, experimental evidence suggests a link between increased oxidative stress and inflammation related to the diabetic state and low levels of CTRP3 [29–31]. Importantly, the study of Wagner et al. [32] found that only female diabetic patients had reduced CTRP3 levels suggesting a possible relation between circulating CTRP3 levels and the hormonal status.

Sclerostin is usually secreted by osteocytes and late osteoblasts to mediate physiological bone metabolism [33]. The changes in the levels of circulating sclerostin may reflect the changes in bone activity, making it a biomarker for the diagnosis and prognosis of OP [34]. On the other hand, previous animal models demonstrated high expression of sclerostin gene, SOST, in the setting of T2DM [35]. Thus, it is logical to assume higher degree of dysregulated levels of sclerostin in patients with combined T2DM and OP. We found that serum sclerostin is a significant predictor of OP in diabetic women. In support of our conclusions, Wang and colleagues [36] showed that the combination of T2DM and OP led to

higher increase in serum sclerostin than OP alone; moreover, serum sclerostin correlated with BMD parameters, HbA1c, and serum glucose level. The increased levels of sclerostin levels in diabetic patients are related to higher oxidative stress [37], poor glycemic control [38] and some genetic polymorphisms [39].

In the present study, we acknowledge the presence of some methodological limitations. The cross-sectional nature of the present study limits the validity of the observed associations and further long-term studies are still needed to confirm the sequential role of T2DM on osteoporosis biomarkers. In addition, the lack of pre-planned samples size calculation and being a single-center experience are additional limitations of the present study.

## Conclusions

In conclusion, the present study provides novel evidence about the impact of T2DM on OP biomarkers, serum CTRP3 and sclerostin. The results indicated that women with combined T2DM and OP/osteopenia exhibited more dysregulation in both biomarkers than non-diabetic women. Thus, serum CTRP3 and sclerostin can be used as biomarkers for early detection of OP in diabetic patients. Further experiments are warranted to confirm our findings and to understand the mechanistic processes behind the additional impact of T2DM on the OP biomarkers. In addition, further investigations about the link between adipose tissue and bone hemostasis are recommended.

## Acknowledgements

We heartfully thank all patients who participated in this study.

## Author contributions

Conceptualization: IHA, SSAG, BMMAE, MKA, EOAE, DSM, MAM, MSM, MAMMAA, SE; patients' recruitment: IHA, SSAG, BMMAE, MKA, EOAE, DSM, MAM, MSM, MAMMAA, SE data acquisition and analysis: IHA, SSAG, BMMAE, MKA, EOAE, DSM, MAM, MSM, MAMMAA, SE; manuscript drafting and writing: IHA, SSAG, BMMAE, MKA, EOAE, DSM, MAM, MSM, MAMMAA, SE. All authors revised the final manuscript.

## Funding

None.

Open access funding provided by The Science, Technology & Innovation Funding Authority (STDF) in cooperation with The Egyptian Knowledge Bank (EKB).

## Data availability

No datasets were generated or analysed during the current study.

## Declarations

### Ethics approval and consent to participate

Protocol of the present study was approved by the ethical committee of Al-Azhar Faculty of Medicine and all participants provided informed consent in accordance with the Helsinki Declaration on clinical research involving human subjects.

### Consent for publication

NA.

### Competing interests

The authors declare no competing interests.

Received: 22 May 2024 / Accepted: 12 August 2024

Published online: 05 September 2024

## References

- Viigimaa M, Sachinidis A, Toumpourleka M, Koutsampasopoulos K, Alliksoo S, Titma T. Macrovascular Complications of Type 2 Diabetes Mellitus. *Curr Vasc Pharmacol*. 2020;18(2):110–16. <https://doi.org/10.2174/1570161117666190405165151>. PMID: 30961498.
- Horton WB, Barrett EJ. Microvascular Dysfunction in Diabetes Mellitus and Cardiometabolic Disease. *Endocr Rev*. 2021;42(1):29–55. <https://doi.org/10.1210/edrv/bnaa025>. PMID: 33125468; PMCID: PMC7846151.
- Tripodiadis F, Xanthopoulos A, Bargiota A, Kitai T, Katsiki N, Farmakis D, Skoularigis J, Starling RC, Iliodromitis E. Diabetes Mellitus and Heart failure. *J Clin Med*. 2021;10(16):3682. <https://doi.org/10.3390/jcm10163682>. PMID: 34441977; PMCID: PMC8396967.
- Kane JP, Pullinger CR, Goldfine ID, Malloy MJ. Dyslipidemia and Diabetes Mellitus: role of lipoprotein species and interrelated pathways of lipid metabolism in diabetes mellitus. *Curr Opin Pharmacol*. 2021;61:21–7. Epub 2021 Sep 22. PMID: 34562838.
- Brandt IAG, Starup-Linde J, Andersen SS, Viggers R. Diagnosing osteoporosis in Diabetes-A systematic review on BMD and fractures. *Curr Osteoporos Rep*. 2024;22(2):223–44. <https://doi.org/10.1007/s11914-024-00867-1>. Epub 2024 Mar 21. PMID: 38509440.
- Ali D, Tencerova M, Figeac F, Kassem M, Jafari A. The pathophysiology of osteoporosis in obesity and type 2 diabetes in aging women and men: the mechanisms and roles of increased bone marrow adiposity. *Front Endocrinol (Lausanne)*. 2022;13:981487. <https://doi.org/10.3389/fendo.2022.981487>. PMID: 36187112; PMCID: PMC9520254.
- Omran A, Atanasova D, Landgren F, Magnusson P, Sclerostin. From molecule to clinical biomarker. *Int J Mol Sci*. 2022;23(9):4751. <https://doi.org/10.3390/ijms23094751>. PMID: 35563144; PMCID: PMC9104784.
- Anastasilakis AD, Tsoardi E. The story of sclerostin inhibition: the past, the present, and the future. *Hormones (Athens)*. 2024 Jan 3. <https://doi.org/10.1007/s42000-023-00521-y>. Epub ahead of print. PMID: 38170438.
- Starup-Linde J, Lykkeboe S, Gregersen S, Hauge EM, Langdahl BL, Handberg A, Vestergaard P. Bone structure and predictors of fracture in type 1 and type 2 diabetes. *J Clin Endocrinol Metab*. 2016;101(3):928–36. <https://doi.org/10.1210/jc.2015-3882>. Epub 2016 Jan 12. PMID: 26756117.
- Yamamoto M, Yamauchi M, Sugimoto T. Elevated sclerostin levels are associated with vertebral fractures in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab*. 2013;98(10):4030–7. <https://doi.org/10.1210/jc.2013-2143>. Epub 2013 Jul 26. PMID: 23894157.
- Ardawi MS, Akhbar DH, Alshaikh A, Ahmed MM, Qari MH, Rouzi AA, Ali AY, Abdulrafee AA, Saeda MY. Increased serum sclerostin and decreased serum IGF-1 are associated with vertebral fractures among postmenopausal women with type-2 diabetes. *Bone*. 2013;56(2):355–62. Epub 2013 Jul 9. PMID: 23845326.
- Sakaguchi M. Adipose tissue plasticity and insulin signaling in the pathogenesis of type 2 diabetes. *Diabetol Int*. 2023;15(1):28–33. <https://doi.org/10.1007/s13340-023-00676-4>. PMID: 38264220; PMCID: PMC10800324.
- Bahadoran Z, Mirmiran P, Ghasemi A. Adipose organ dysfunction and type 2 diabetes: Role of nitric oxide. *Biochem Pharmacol*. 2024;221:116043. <https://doi.org/10.1016/j.bcp.2024.116043>. Epub 2024 Feb 5. PMID: 38325496.
- Begum M, Choubey M, Tirumalasetty MB, Arbee S, Mohib MM, Wahiduzzaman M, Mamun MA, Uddin MB, Mohiuddin MS. Adiponectin: a Promising Target for the treatment of diabetes and its complications. *Life (Basel)*. 2023;13(11):2213. <https://doi.org/10.3390/life13112213>. PMID: 38004353; PMCID: PMC10672343.
- Guo B, Zhuang T, Xu F, Lin X, Li F, Shan SK, Wu F, Zhong JY, Wang Y, Zheng MH, Xu QS, Ehsan UMH, Yuan LQ. New insights into implications of CTRP3 in obesity, metabolic dysfunction, and Cardiovascular diseases: potential of therapeutic interventions. *Front Physiol*. 2020;11:570270. <https://doi.org/10.3389/fphys.2020.570270>. PMID: 33343381; PMCID: PMC7744821.
- Fadaei R, Moradi N, Baratchian M, Aghajani H, Malek M, Fazaeli AA, Fallah S. Association of C1q/TNF-Related Protein-3 (CTRP3) and CTRP13 serum levels with coronary artery disease in subjects with and without type 2 diabetes Mellitus. *PLoS ONE*. 2016;11(12):e0168773. <https://doi.org/10.1371/journal.pone.0168773>. PMID: 28033351; PMCID: PMC5199067.
- Fadaei R, Moradi N, Kazemi T, Chamani E, Azdaki N, Moezibady SA, Shah-mohamadnejad S, Fallah S. Decreased serum levels of CTRP12/adipolin in patients with coronary artery disease in relation to inflammatory cytokines and insulin resistance. *Cytokine*. 2019;113:326–31. <https://doi.org/10.1016/j.cyto.2018.09.019>. Epub 2018 Oct 15. PMID: 30337217.
- Shanaki M, Moradi N, Fadaei R, Zandieh Z, Shabani P, Vatannejad A. Lower circulating levels of CTRP12 and CTRP13 in polycystic ovarian syndrome: irrespective of obesity. *PLoS ONE*. 2018;13(12):e0208059. <https://doi.org/10.1371/journal.pone.0208059>. PMID: 30540803; PMCID: PMC6291267.
- Yaribeygi H, Rashidfarrokhi F, Atkin SL, Sahebkar A. C1q/TNF-related protein-3 and glucose homeostasis. *Diabetes Metab Syndr*. 2019 May-Jun;13(3):1923–7. <https://doi.org/10.1016/j.dsx.2019.04.047>. Epub 2019 Apr 25. PMID: 31235116.
- Li X, Jiang L, Yang M, Wu Y, Sun S, Sun J. GLP-1 receptor agonist increases the expression of CTRP3, a novel adipokine, in 3T3-L1 adipocytes through PKA signal pathway. *J Endocrinol Invest*. 2015;38(1):73–9. <https://doi.org/10.1007/s40618-014-0156-8>. Epub 2014 Aug 23. PMID: 25149084.
- Zhang XJ, Zhang D, Qin J, Zhou YR, Guo JL, Zhang YZ. Serum CTRP3 concentrations are positively correlated with disease severity in women with postmenopausal osteoporosis: a population-based cross-sectional study. *Eur Rev Med Pharmacol Sci*. 2023;27(22):10868–74. [https://doi.org/10.26355/eur-rev\\_202311\\_34454](https://doi.org/10.26355/eur-rev_202311_34454). PMID: 38039016.
- American Diabetes Association. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes-2019. *Diabetes Care*. 2019;42(Suppl 1):S13-S28. <https://doi.org/10.2337/dc19-S002>. PMID: 30559228.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985;28(7):412–9. <https://doi.org/10.1007/BF00280883>. PMID: 3899825.
- Maeda T, Abe M, Kurisu K, Jikko A, Furukawa S. Molecular cloning and characterization of a novel gene, CORS26, encoding a putative secretory protein and its possible involvement in skeletal development. *J Biol Chem*. 2001;276(5):3628–34. <https://doi.org/10.1074/jbc.M007898200>. Epub 2000 Nov 8. PMID: 11071891.
- Maeda T, Jikko A, Abe M, Yokohama-Tamaki T, Akiyama H, Furukawa S, Takigawa M, Wakisaka S. Cartducin, a paralog of Acrp30/adiponectin, is induced during chondrogenic differentiation and promotes proliferation of chondrogenic precursors and chondrocytes. *J Cell Physiol*. 2006;206(2):537–44. <https://doi.org/10.1002/jcp.20493>. PMID: 16155912.
- Musso G, Paschetta E, Gambino R, Cassader M, Molinaro F. Interactions among bone, liver, and adipose tissue predisposing to diabetes and fatty liver. *Trends Mol Med*. 2013;19(9):522–35. <https://doi.org/10.1016/j.mol-med.2013.05.006>. Epub 2013 Jun 28. PMID: 23816817.
- Elsaid HH, Elgohary MN, Elshabrawy AM. Complement c1q tumor necrosis factor-related protein 3 a novel adipokine, protect against diabetes mellitus in young adult Egyptians. *Diabetes Metab Syndr*. 2019 Jan-Feb;13(1):434–38. <https://doi.org/10.1016/j.dsx.2018.10.004>. Epub 2018 Oct 12. PMID: 30641739.
- Trogen G, Alamian A, Peterson JM. High molecular weight, but not total, CTRP3 levels are associated with serum triglyceride levels. *Physiol Rep*. 2019;7(23):e14306. <https://doi.org/10.14814/phy2.14306>. PMID: 31814309; PMCID: PMC6900490.
- Ma ZG, Yuan YP, Xu SC, Wei WY, Xu CR, Zhang X, Wu QQ, Liao HH, Ni J, Tang QZ. CTRP3 attenuates cardiac dysfunction, inflammation, oxidative stress and cell death in diabetic cardiomyopathy in rats. *Diabetologia*. 2017;60(6):1126–37. <https://doi.org/10.1007/s00125-017-4232-4>. Epub 2017 Mar 3. PMID: 28258411.
- Zhang J, He J. CTRP3 inhibits high glucose-induced oxidative stress and apoptosis in retinal pigment epithelial cells. *Artif Cells Nanomed Biotechnol*. 2019;47(11):3758–64. <https://doi.org/10.1080/21691401.2019.1666864>. PMID: 31556307.

31. Zeng X, Peng Y, Wang Y, Kang K. C1q/tumor necrosis factor-related protein-3 (CTRP3) activated by forkhead box O4 (FOXO4) down-regulation protects retinal pericytes against high glucose-induced oxidative damage through nuclear factor erythroid 2-related factor 2 (Nrf2)/Nuclear factor-kappaB (NF-κB) signaling. *Bioengineered*. 2022;13(3):6080–91. PMID: 35196182; PMCID: PMC8974204.
32. Wagner RM, Sivagnanam K, Clark WA, Peterson JM. Divergent relationship of circulating CTRP3 levels between obesity and gender: a cross-sectional study. *PeerJ*. 2016;4:e2573. <https://doi.org/10.7717/peerj.2573>. PMID: 27781167; PMCID: PMC5075694.
33. Voorzanger-Rousselot N, Journe F, Doriath V, Body JJ, Garnero P. Assessment of circulating Dickkopf-1 with a new two-site immunoassay in healthy subjects and women with breast cancer and bone metastases. *Calcif Tissue Int*. 2009;84(5):348–54. <https://doi.org/10.1007/s00223-009-9225-y>. Epub 2009 Feb 28. PMID: 19252761.
34. Drake MT, Srinivasan B, Mödder UI, Peterson JM, McCreedy LK, Riggs BL, Dwyer D, Stolina M, Kostenuik P, Khosla S. Effects of parathyroid hormone treatment on circulating sclerostin levels in postmenopausal women. *J Clin Endocrinol Metab*. 2010;95(11):5056–62. <https://doi.org/10.1210/jc.2010-0720>. Epub 2010 Jul 14. PMID: 20631014; PMCID: PMC2968729.
35. Nuche-Berenguer B, Moreno P, Portal-Nuñez S, Dapia S, Esbrit P, Villanueva-Peñacarrillo ML. Exendin-4 exerts osteogenic actions in insulin-resistant and type 2 diabetic states. *Regul Pept*. 2010;159(1–3):61–6. <https://doi.org/10.1016/j.regpep.2009.06.010>. PMID: 19586609.
36. Wang N, Xue P, Wu X, Ma J, Wang Y, Li Y. Role of sclerostin and dkk1 in bone remodeling in type 2 diabetic patients. *Endocr Res*. 2018;43(1):29–38. doi: 10.1080/07435800.2017.1373662. Epub 2017 Oct 3. PMID: 28972408.
37. Sabancilar I, Unsal V, Demir F, Toprak G, Pekkolay Z. Does oxidative status affect serum sclerostin levels in patients with type 2 diabetes mellitus? *Folia Med (Plovdiv)*. 2023;65(1):46–52. <https://doi.org/10.3897/foimed.65.e72953>. PMID: 36855973.
38. Singh PK, Naitthani M, Pathania M, Mirza AA, Saha S. An Insight into the Association of Sclerostin with Insulin Sensitivity and glycemic parameters in male Indian Prediabetic and Diabetic Population. *Cureus*. 2022;14(7):e27123. <https://doi.org/10.7759/cureus.27123>. PMID: 36004027; PMCID: PMC9392653.
39. Li J, Ren Y, Li S, Li J. Relationship between sclerostin (SOST) expression and genetic loci rs851056, rs1230399 polymorphisms and bone Mineral Density in Postmenopausal Women with type 2 diabetes in Xinjiang. *Diabetes Metab Syndr Obes*. 2021;14:4443–50. PMID: 34764662; PMCID: PMC8575445.

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.