DNA sequences of a bovine gene and of two related pseudogenes for the proteolipid subunit of mitochondrial ATP synthase

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The dicyclohexylcarbodi-imide-reactive proteolipid is a membrane subunit of mitochondrial ATP synthase. In cows it is encoded by two different nuclear genes known as P1 and P2. These genes are expressed in a tissue-specific fashion which reflects the embryonic origin of the tissues. The proteins that they encode are synthesized in the cytosol, and are precursors of the proteolipid that have different mitochondrial import sequences of 61 and 68 amino acids respectively. By use of gene-specific probes derived from the bovine P2 cDNA, regions containing corresponding parts of the bovine P2 gene have been isolated from a bovine genomic library, and their DNA sequences and those of flanking and intervening regions have been determined. The sequence contains four exons, which represent the cDNA sequence, spread over 3.8 kb of the bovine genome. Two of the introns are in the DNA sequence coding for the mitochondrial import sequence, and a third intron is in a sequence encoding an extramembranous structure between the two putative transmembrane α -helical domains of the mature proteolipid. An *Alu*-type repetitive element was detected at the extreme 5' end of the sequence. The bovine P1 and P2 genes for the dicyclohexylcarbodiimide-reactive proteolipid of ATP synthase are members of a multiple gene family that also contains many pseudogenes. The bovine P1 gene has not been isolated, but two distinct P1 pseudogenes have been cloned and their DNA sequences have been determined. Both of them contain 'in-phase' stop codons and frameshift mutations, and one of them bears the hallmarks of retroposition; it has no introns, it contains a poly(A) tract at its 3' end and it is flanked by direct DNA sequence repeats. The second P1 pseudogene is very unusual. It appears to be derived from a partially processed transcript and contains an intervening DNA sequence of 861 bp that corresponds in position with an intron in the human P1 gene. This pseudogene also could have been introduced by retroposition since its sequence is flanked by short direct repeats. However, it does not contain a poly(A) tract at its 3' end. An alternative, but less likely, explanation is that rather than being a retroposon, this sequence arose by duplication of an expressed gene at a time when it had only one intron.

INTRODUCTION

The dicyclohexylcarbodi-imide (DCCD)-reactive proteolipid is an essential membrane protein component of the proton channel of bovine mitochondrial ATP synthase. It is 75 amino acids long (Sebald & Hoppe, 1981), and is a nuclear gene product, as are all but thirteen bovine mitochondrial proteins (Anderson et al., 1982). Import into the organelle of these nuclear coded proteins is usually directed by an N-terminal extension known as the mitochondrial import sequence. This is removed during entry into the mitochondrion (Schatz & Butow, 1983). The DCCD-proteolipid is unique among mitochondrial proteins so far investigated in having two different, but weakly homologous, import presequences, which, when removed from the precursors, produce an identical mature protein. The precursors are the products of two different genes, P1 and P2, that are expressed in different ratios in various bovine tissues (Gay & Walker, 1985). In the present paper the characterization of the bovine P2 gene is presented. It is split into at least four exons, as are the human homologues (M. R. Dyer & J. E. Walker, unpublished work), and their sequence is spread over about 4 kb of bovine DNA. This gene is a member of a multigene family which includes at least two expressed genes and several pseudogenes. The sequences of two related pseudogenes are also presented. One of them has no intervening sequences and appears to be a retroposon; the other is a partly spliced pseudogene.

MATERIALS AND METHODS

Preparation of bovine DNA and genomic libraries

The preparation of bovine liver DNA has been described previously (Walker *et al.*, 1987). A phage library of partial *Sau*3AI fragments of bovine genomic DNA was made in λ 2001 (Karn *et al.*, 1984).

DNA hybridization

Digests of DNA were fractionated by electrophoresis in 0.6% agarose gels, and fragments were transferred and fixed to nitrocellulose as described by Southern (1975). After transfer, the nitrocellulose filters were incubated at 65 °C for 1 h in a solution containing $6 \times SSC$ (1 × SSC is 0.15 M-NaCl/0.015 M-trisodium

Abbreviation used: DCCD, dicyclohexylcarbodi-imide.

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citrate), 0.2% bovine serum albumin (fraction V), 0.2% Ficoll, 0.2% polyvinylpyrrolidone, 0.5% *N*-laurylsarcosine and sonicated salmon testis DNA ($100 \mu g/ml$). Then filters were hybridized for 15–20 h at the same temperature in the presence of radioactive 'prime-cut' probes (Farrell *et al.*, 1983) dissolved in the same solution as used for pre-hybridization, except that it contained also 10% dextran sulphate. Subsequently, the filters were washed four times for 30 min at 65 °C in either 0.2 or $2 \times SSC$, each containing 0.5% laurylsarcosine. Autoradiographs of filters were exposed for 1–3 h at -70 °C in the presence of an intensifying screen.

Screening the genomic library

Plaques (about 10^6) were produced on *Escherichia coli* Q358 grown on 20 cm diameter agar plates, and were screened by the plaque hybridization method (Benton & Davis, 1977). Phage from each plate were transferred to two nitrocellulose filters placed sequentially on the agar. The preparation of 'prime-cut' probes and the hybridization conditions employed were the same as those described above. Recombinant phages were grown on *E. coli* Q358 in 500 ml cultures, and DNA was prepared from them according to Maniatis *et al.* (1982).

Identification and manipulation of genomic clones

Hybridization studies indicated that the recombinant λ P4.21 contained the bovine P2 gene. From its DNA were excised a 5.3 kb NcoI fragment and an overlapping 2.8 kb XbaI fragment. They were purified by electrophoresis in low melting point agarose, and then broken up by sonication. The resultant fragments were fractionated by electrophoresis, and those that were greater than about 500 bp were cloned into the SmaI site of M13mp8 (Deininger, 1983). Two other recombinants, λ P3.9 and λ P3.17, contained sequences related to the bovine P1 cDNA. By sequence analysis it seemed that they both contained pseudogenes. From $\lambda P3.9$ and $\lambda P3.17$ respectively, a SacI fragment of 5.2 kb and a 4.0 kb EcoRI fragment were purified, and libraries of random fragments produced by sonication were prepared from them as above.

DNA sequence analysis

DNA sequences were determined by the dideoxy chain termination method (Sanger *et al.*, 1977) as modified by Biggin *et al.* (1983) and a random strategy was employed. All sequences were determined minimally at least once in both senses of the DNA. This required that both clone turn-arounds and some long-runs be performed on selected clones. 'Compressed' sequences were resolved by substituting deoxyinosine triphosphate for deoxy-GTP in the appropriate sequencing reactions (Mills & Kramer, 1979). DNA sequences were compiled with the help of the computer programs DBUTIL and DBAUTO (Staden, 1982), and were analysed with ANALYSEQ (Staden, 1985).

RESULTS AND DISCUSSION

Gene cloning

About 10⁶ recombinants in the bovine genomic library were screened with probes derived from the 3' noncoding regions of the bovine P1 and P2 cDNAs. In contrast to the coding regions, these sequences are poorly conserved and would be expected to distinguish between the P1 and P2 genes. The probes employed were nucleotides 404-558 and 406-615 respectively of P1 and P2 cDNAs (Gay & Walker, 1985). After re-screening of initial isolates, 20 recombinants, $\lambda P3.1 - \lambda P3.20$, were selected which hybridized with the P1 probe, and also 21 recombinants, $\lambda P4.1 - \lambda P4.21$, which hybridized with the P2 probe. DNA was prepared from each isolate, and further restriction analysis and hybridization experiments showed that the P1 isolates appeared to fall into three related groups, and the P2 isolates could be put in four groups. Each group contained overlapping, but nonidentical, DNA inserts. From the P1 isolates, λ P3.9 and λ P3.17 were subsequently found to contain different pseudogenes. Another isolate, $\lambda P3.3$, which is not described in this paper, appears to contain part of exon IV of the P1 gene and a flanking sequence. However, part of the clone seems to result from a re-ligation event with unrelated DNA (N. J. Gay, unpublished work). No further attempts have been made to find other clones containing the bovine P1 gene. Restriction fragments in recombinant λ P4.21 hybridized to the P2 probe derived from the non-coding region of the bovine cDNA, and other restriction fragments hybridized also to a second probe taken from the coding region. From these data it was deduced that this recombinant contained the regions of the expressed gene that correspond to the P2 cDNA, as was proved subsequently by sequence analysis.

DNA sequence of the bovine P2 gene

A digest with *NcoI* of DNA from λ P4.21 was fractionated in a 0.6% agarose gel, and the fragments were hybridized to a 'prime-cut' probe containing nucleotides 120-409 of the cDNA sequence of bovine P2 (Gay & Walker, 1985). As shown in Fig. 1, this revealed a 5.3 kb *NcoI* restriction fragment, suggesting that most of the DNA sequence in the probe is within this fragment. This conclusion is based on the assumption that the *NcoI* site within the cDNA sequence was also present in genomic DNA, and that it had not been created by splicing of the primary transcript. The DNA sequences encoding the 15 C-terminal amino acids of the protein and 3' untranslated region of the mRNA were not expected to be found in this restriction fragment. The DNA sequence of this fragment (see Fig. 2) contained the 5' non-coding region present in the cDNA, and sequences representing the coding sequence of the cDNA up to and including the first base of the codon for amino acid 61 of the mature protein; as anticipated, the remainder of the 3' end of the gene was absent. Therefore, in order to extend this sequence, an XbaI restriction fragment derived from λ P4.21 DNA that overlaps the 3' region of the NcoI fragment was identified by hybridization to a 'prime-cut' probe derived from the 3' region of the P2 cDNA. This XbaI fragment was sequenced completely and corresponds to nucleotides 4831-7647 in Fig. 2. The extended sequence contained the missing 3' region encoding 15 amino acids at the C-terminal end of the proteolipid, and the 3' untranslated region present in the mRNA.

Each nucleotide in the sequence of the bovine P2 gene was determined six times on average, and at least once on each strand. The G+C content of this 7647 bp segment is 44%, in reasonable agreement with the estimated G+C content of 42% for the bovine genome (Chargaff & Lipshitz, 1953).



Fig. 1. Hybridizations of bovine DNA in a recombinant phage with probes for the bovine P2 gene

In panel (a), DNA from recombinant λ P4.21 (0.5 µg of DNA/digest) was restricted with NcoI, and in panel (b) DNA (0.5 µg of DNA/digest) from the same phage was digested with SacI and EcoRI (lane a), EcoRI and XbaI (lane b), StuI and EcoRI (lane c), NciI and SacI (lane d), AhaI and SacI (lane e), XbaI (lane f), TaqI (lane g), StuI (lane h), NciI (lane i), and AhaII (lane j). The DNA digests were fractionated on a 0.6% agarose gel and the restriction fragments were transferred to nitrocellulose. In panels (a) and (b), the DNA fragments were hybridized with 'prime-cut' probes containing nucleotides 120–409 and 406–615 respectively from the bovine P2 cDNA (Gay & Walker, 1985). Both of these blots were washed at 65 °C in 0.2 × SSC, and then autoradiographed for 1 h at -70 °C using Fuji X-ray film. In panels (a) and (b), bands that correspond to a 5.3 kb NcoI and 2.8 kb XbaI restriction fragments, respectively, are labelled.

Gene structure

Comparison of the sequence with that of the bovine P2 cDNA showed that the gene is split into at least four exons (see Fig. 3), and the sequences of the exons together agree exactly with that of the bovine P2 cDNA (Gay & Walker, 1985). The exons are in precisely the same positions as those in the human P1 and P2 genes (M. R. Dyer & J. E. Walker, unpublished work). The exact location of the 5' end of exon I is not known, as the transcriptional start site(s) of bovine P2 has (have) not been determined experimentally. However, eukaryotic promoters contain DNA sequences which regulate the rate of transcription by RNA polymerase II, and these elements are usually located within a 100 bp to the 5' side of the transcriptional initiation site (Cochran & Weissmann, 1984; McKnight & Kingsbury, 1982). Many eukaryotic promoters contain a TATA box, a conserved AT-rich sequence which is centred about 25 bp to the 5' side of the cap site. The TATA box has been shown to control the precise position of transcriptional initiation in some eukaryotic genes (Grosschedl & Birnstiel, 1980; Benoist & Chambon, 1981; McKnight & Kingsbury, 1982). A second conserved sequence, the CCAAT box, is located 70-90 bp to the 5' side of the cap site in many genes. Its precise function is unknown but in certain genes it has been shown to bind different cellular factors (McKnight & Tijian, 1986; Dorn *et al.*, 1987). Both of these sequences are present in the region to the 5' side of the furthest established extent of exon I (see Fig. 2). Nonetheless, the possibility remains that the 5' noncoding region present in the mRNA is more extensive than that characterized in the bovine P2 cDNA, and that additional intervening sequences are present towards or beyond the 5' end of the present sequence.

Two of the introns that have been detected are found in sequences encoding the mitochondrial import sequence, and none is found at the boundary between the import sequence and the mature proteolipid. A third intron, however, is at a position which is within the protein sequence ARNP (amino acids 37-40 of the mature protein). This is thought to form a β -turn at the membrane periphery, and to link the two transmembrane α -helices into which the proteolipid is probably folded. The location of introns in genes encoding membrane proteins in sequences that are believed to be extramembranous links between transmembrane segments has been noted also in rhodopsins (Nathans & Hogness, 1984), in the band III protein from mouse red cell membranes (Kopito *et al.*, 1987) and in the mitochondrial

	ATTTCAGAAC	ACCAGGCTT	CCCTGTCCAT 30	CACCAACTCC 10	TGGAGCTTAT 50	GGAAACTCAT 60	GTTCATCGA 70	GTCAGTGATG 80	CCATCCAACC 90	ATCTCATCCT	CTGTCGTCCC	<u>111CTC</u> 120
				hovine rene	titive elem	ent						
CIGCOL	TCAGICITIC	00000000000	GGTCTTTTC	BAATGAGTCA	TOTOTTTOCA	TCAGGTGGCC	AAGGTATTG	GAGTTTTAGC	TTCAGCATCA	GICCCICCAR	TGAAGATICA	GGACTG
	130	140	150	160	170	180	190	200	210	220	230	240
ATCTCC	TTTAGGATGO	ATTGGTTGG	ATCICCTICC	AGTCCAAGGG	ACTOTOAAGA	GTCTTCTCCA	GCACCACAG	TTCARAAGCA	TCRATITIC	AGCACTCAGC	TITCTTTATA	TTCCRA
meree	250	260	270	280	290	300	310	320	330	310	350	360
CTCTCA		TGACTACTO			ACATCCACCT	1161666088	AGTARTOTO	1016011111	AATACACTGI	CTAGGTTGGT	CATAACTIT	611CC8
creren	370	380	390	100	410	120	130	110	150	160	470	180
											COCATCAACT	CATCCC
AGGAGC	AGGCATCTTT 490	S00	GGCTGCAGTC 510	S20	530	NGCCCHAMAA 540	550	560	570	580	590	600
ACCHGH	610	620	630	610	650	660	670	680	690	700	710	720
ATCTGC	ATATCTGAGG 730	740	750	760	770	780	790	800	810	820	830	840
CAGCCT	TGATGAACTO 850	860	TTTGGAACCA 870	ATCTGTTGTT 880	CCATGTCCAG 890	TTCTAACTTG 900	OTTETTGAC 910	CTGCATACAG 920	ATTTCTCAGO 930	SAGGAAGGTGA 940	166T66TCT66 950	960
CATCTC	TTTAAGAATI 970	980	TTGTTGTGAT 990	CCACATGGTC 1000	AAAGGCTTTG 1010	GCGTAGTCAG 1020	TAAAGCAGC 1030	AGTAGATGTT 1040	TTTCTGGAAC 1050	1060	1070	CCAGCA
ATTGAT	CCTCTGCCTI	ITTCTARATC	CAGCTTGAAC 1110	ATCTTTAACA 1120	AATCTTTTGA 1130	CGTGTTTTGA 1140	ICAAATGCTT 1150	CCTGGTAATT 1160	CAATAAGAGO 1170	GATTGTTAGAC 1180	ACCGTTTCAC 1190	AGCCTT 1200
CACCTT	CCGCACGTA1	ICTTCCAAGA	GTCTTTTTA 1230	TTTTTTTGA 1240	CTTGTGCTGT 1250	ATGGCTTATG 1260	GGATCTTAG	TTCCCCAGTO 1280	AGGGATGGAA 1290	ACCCATTCCCC 1 300	CTACAGTTGA 1310	IAGCTCA 1320
RAGTCC	TAACTACTGO 1330	GACTGCCAGG 1340	GAATTACCCT 1350	GAGGGTCTTA 1360	AAGGGCAACA 1370	ITTTTGAGATC	CAGGAAATG 1390	AAGGCTTGAG 1400	AAGAGTTCC1	TTTTCAGTAAC 1420	GAGACTGGGGG 1430	GGAAAC 1440
TTGGGC	AAAAGATGCI 1450	IGGGAAAATT 1460	CGTCTATAAA 1470	CCCACCTCAT	GAGGCTATGG 1 490	AAAGAGATAA 1500	ICATGTCTTA 1510	AAGCTGGGAC 1520	CAAGTGGCTI 1530	TCTTGTTCAA 1540	ICCAAGAGAAG 1550	TCACTT 1560
GATTAT	AAGGAAAGCI	ITAAAAGCTG 1580	TTTGGAGGAA 1590	ATCCTGACTT	CAGGGTTCTC 1610	ATTCCATCTO	TAAGACTTG	AGGTCAGTCO 1640	CTGCTTTCT(1650	GGGCCATTTA 1660	AGTTTTCCAGA 1670	AGCACTT 1680
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GCTAAT	ACAGCGGTC1 =	ITTTGATGAG 1700	CAAGAAAAAA 1710	CATTTACACT 1720	GCTGCTTCTG 1730	TAATTGCAGO 1740	TTTCCTGCC 1750	GTAGCCCTTC 1760	ATCCCCTGAI 1770	ARATGTACACI 1780	TGCGCCAAG 1790	TCGTCT 1800
T CCACCC	P S L CCTCCTTO	IGAGTACCCA	CCTTTCCCAA	GAAAGTTTTT	AAGGAGAGGT	GTTTTGTCTT	TTCCTTCTC	AGACCTTATO	CGTCACAGT	TGGGCCCTTT	SCTTIGCTAG	CTGACG
	1010	-	1830	1810	1050	1860	1870	1880	1890	1900	1910	1920
AGAGTG	AATACTTCCI	AGGCGTTAC	TTGCCTTGTG	TCCCATGAAG	GCCAGTTGAT	TTTTCTCATO	CAGTCATAA	TAGAAGAGCE	TAAATGAGCI	ATCTAGGCTA	IGACAGGTTA	TCCTAT
	1930	1940	1950	1960	1970	1980	1990	2000	2010	2020	2030	2040
CCHGHR	ACACTGTGTG 2050	CTTGTARAC 2060	TGGAGCCCCA 2070	2080	TAAACATGAA 2090	ATAAATAGCC 2100	2110	CTGAGTATGO 2120	ACTGTGTGTACI 2130	AGGATCTTGTO 2140	2150	AGAGAGC 2160
TGTAGC	TAGGAGTTCC 2170	TCTGGAAGC 2180	TTAAATTTGG 2190	GTGGGAAGCT 2200	AAAATACACA 2210	TAGGAAAAAT 2220	2230	AAAAGCAACF 2210	AGATGGAAGI 2250	AAATGGAAGA 2260	STATTAGATAG 2270	5TGGAGG 2280
CAATCG	GTTCATTCAC 2290	TCGTAAGTG 2300	AAAAGGAGTG 2310	ATGCTGTTAG 2320	TTGGTATTAT 2330	TTATTGAGCA 2340	CTTGCTATG 2350	CCTGGCATTA 2360	AGTCTACCA 2370	GTTGATGTACO 2380	2390	TACTCC 2400
TCCAGT	CAACCCTGAG 2410	ATATGTTAT 2420	TCCCATTTGA 2430	TAGATGAGAA 2110	AGCAGGCTTA 2450	AAAAGATTTA 2460	CATGACTTG 2470	TCCTAAGATO 2180	ACACAACTCI 2190	RGATAGAGCTI 2500	AGGATTTGAA1 2510	ICTAGTT 2520
ATATAT	GACTAAAGCO 2530	CTGGCTGTT 2540	AACCACCATG 2550	TTGTCTATTT 2560	TGGAGACGCT 2570	GGCTTGGAGT 2580	TAGAAAAGG 2590	GCTTCCAGGF 2600	IGGGGGTACA 2610	TTTTCAGCACI 2620	AGGTCTTCATO 2630	GTCTGTT 2640
TTCCAG	AACAGTGTG1 2650	TCAGGGAAA 2660	GATGTTCCAG 2670	TCATTCACTT 2680	CCTTCTAATG 2690	AGCTGGCCA1 2700	TACCAGTAT 2710	ATGTGTAGA1 2720	TTCTTATTG 2730	AGTGAAGTAA 2740	ATACTITAAA 2750	TGAATGA 2760
TTCCAG	ATTCTTTTT 2770	IGAGGTTCCA 2780	ATTTAAGTTC 2790	TAGTGAATAA 2800	ATGGGGTATC 2810	TCAGTAATAA 2820	ATTTGCAGTA 2830	IGTTCTAAAA 2840	TCTCCACAG 2850	CGTTTCTTGA 2860	CCTTGTATTT 2870	TTCCGGC 2880

Xba I

Exon II R R T S T U L S R S L S A U U U R R P E T L T D E

GCACATGCCCCTGTGTGCTGCTTATAACTCCCATTTTACTTGATCTAGTGCGCTGACTTGAGTTGGGRGCATGTAGCCCAGGAAGATGGTCTCCTCAAGGCATCAGGAACAGGCTAGGGC TGTAGTCTTCCATTTCTAGACTGTACCTGAGTCATGAAGATATTTGACATCCACCTCTTGCATTTAAGCGTTCTGCTTGGCAGTAGAGTTTTCTGGGGAGTTAAAAGATTTTTTGTGTGG SHSSL AGGAGAAACGATGGGATTTGATAGCATAAAGGGATTAAGTGGTACAGAAGATTTCCAAAGCAAAAAAAGGGTTTCCTAAAAACTTGCTATCTGCTGCTTCTCACAGGAGCCACAGCAGCAGCA Exon III A U U P R P L T T S L T P S R S F Q T S A I S R D I D T A A K F I G A G A A T U ANTAT IT I GGGG TT CTGGAGCTGCAGTATCC TATACAGTAGCCACCTCACCCTAGTGCTATTCANATGTGAGTTTTAACTAAAAGTACACTGACTTAAGTATACTTTGTATACTTTCAG TGTAGCTCAGTAGCCACATGTGACTCGTGGCTACCATTTTGGAAAGTTCTATTGGACAGCTTTGGTATGAGTAGGAGTTAGCAAAGCCAGGAGGTCTGTATTGGGCCAGCCTGGT GTTTGCCATGTCCACCCCCAGTCTAAAGCCTCAGTCAGCATAGCTTCGCTGCTGTTTTTCCCCGTCCCGGGAATTCCCTCATCACTCCTGGGAAAACTGCTTCTGCATCCAGCCCCAG TGCTCCCTGRCRCTTGRGRCTCTTTGGTCCCCRRCRCTCGGGCRRCTCRGTGTTAGRGCTARATTRATCCTTTRGRGRCCTGCCRGTTCTCTCRGTGCCTTTGGGGRARTARGTTTAT

TTCTTAGGATTTGTATGARATAGARACTTCTATGTCCACAATTCTATTAAAAGCTAGTGTTGCTTTACCTTATCCATCAAGTCTTCCGGAGGCGACAGTAAGGGARATAGAAATTATAT

GTGATGGACAGGGAGGCCTGGCGTGCCGCATTCATGGGGTCGCAAAGAGTCGGACGCGACTGATCTGATCTGATCTGATCTATCCCATTTTGATCTATGTCTAAATGTGTGCCCCC

ACTCCCCRAGGCGGTTTGTAGCCCARAATCTTTTGACTATTTGARATAAGAGAAGTTATTTGTCCCTCTGATGTCCCTAGTAGAATAGTATTTCTCTAATTGTGTTACACAAGCC

4710 4720

CTRGGGTTCAGAGGGTACACCCCAGAGACGTCTAGAGAACGAAGGGGAGACCAAACAAGACTTCCAACTCCGCCACTCCATTACACTTTAGCAGTTACAAGTTTTGTATTATGGGGTTCT

ACTGCTTARAGTTTGGAAACGCTGCTCGTATAGACCCTGCCACCCAGAGACGTCTTCCATCTTGGTTGAGCCCTGGATAGTTAAGTGTCCAGACCCAGGGAAGATGTGGTGAGATACAGTAG

AGGAGGTAATGTTTGTACCTGGGCTGTGTTGGAGAATTTTGGATTGTCCCCTCCCCCCTTCATCTCTGTAGAGACCTTAATGTGTCGGGCAAGAAATGGGGCATAATGGTATCACCCTG

Exon IV

S L K O O L F S Y A I L G F A L S E A M G L F C L M U A F L I L F A M 4

5320 5330 NCO 1

TACTGTATTAATAAGATGTTTCTTGAGTCTCCTGTGTATATTTCTTTTCCACAATTGGCTGAATGCCTTGGTGAAAGTATAAGGCCAUAUGTTAGTGATGGTTTTAAAACTCAATGTGGAT

 CTGGGCTCACTTATTTTCATTCTCTATGTTGGGAAAGCTTTATTCAAATCAGTGCTCTTTTTTAATAATTTTCACTTGGGATGGCCTATTACCATGGAGAATTGGGACAGAAGCAACAAT TTAGATTGTTAGTTTCCTGAGGCAGGGGCTCTGTCATACTCGTATCCTTAGAAGTATCTTAGCATGACACATCTCAGCAATATCTTAGTCAAATTTTCAATTTAATTGTCTAAACCAGCC ACCTCACTTCAGAAGTGAGGGAACAGACTTTGAGATATCAAAGGACTCTCCCAGGAACATTTATTGCAGAGCTGAGTGGGAGCAGGGGTCACGTGTTGTAGTCCAGGGTGTACGACTA CCGCRCRGTTGGTCTCGTTCTCTCCRGTGTTGCTGGTGRCTGCTATTTTCTCRRGCRATCCARRAAGRGCRGAATAGGCRATGGTARACCTTCTTCCTTARARATGCCTCARGAGGGAC TTCTGGRCRGGGGTTRGTATARRGGTARRGGATTATTATTTTTCTARRGGCTARATTTTCGGGTCTTTCCTGGTGGTCGTGTGTTGTTARGARTCCACCTACCGGTGRGGTGACATGGGTTCC ATCCCT6GTCCA6GAATCTGACAT6CCTT6GAGCAACTAA6CCT6T6T6CCACAACTATTGA6CCTACGA6CCCACACCCT6A6CCCACTT6CT6CAACTATTGAA6CCCAT6CTCT6C ARCARGAGAAGCCACTGCAATTATGARGCCCCCGCTCGCCACACTAGAGARAGCCCCACATGCAGCAATAAAGACCCAARATAAAATTAAAAATTAAAAATTAAACAAAAA

RATAAAATTTTTTAAAGGGCTAAAACTTTAGAAGTGGATGGCGCAGTGGCGTAAGAATTTATCTGCCACCGCAGGGGACATGGGTTCCATCCCTGGTCTGGGGGGGAATTCCACATGTTGCAG 7090 7100 7110 7120 7130 7140 7150 7160 7170 7180 7190 7200

AGCRATGGAGCCCACGTGCCACAGCTAGTGAGTCCACATGCCTGGAGCCTGTGCTCCTCAACAAGAGAAGCCCATGCCATGAGAAGCCTGAGCACCGCAACTAGTTGGCCCTGCTCACTG 7210 7220 7230 7240 7250 7260 7270 7280 7290 7300 7310 7320 CAACTAGAGAARAGCCCAAGCGAAGCAACGAAGCTCAGCCAAAACTGAATTTTTTCTTAAATCTTTTTTGAAGAAAAAAGTTAAACTTCAAGAGCTTAACTTCAATATGCTCG

7330 7340 7350 7360 7370 7380 7390 7400 7410 7420 7430 7440

RRGTTGGGTATARARGTCCTTTCATGATAATGTTGATGATGTTCATGAGTTTTTCTTTTGGTTTAATCAGTAATAARAGCCTAGAGCAGATACTTGACCCCCACAATAARCTGATGGTTAGG 7450 7460 7470 7480 7490 7500 7510 7520 7530 7540 7550 7560

ARGCAGGTCACCTCGAACTATTCACCCTGTCCTTTCGCTATTGTCATCAAGGATTGAATGGGATCGTATCCTCAATTTGCTTCAAGA

Fig. 2. DNA sequence of a segment of bovine DNA containing the bovine P2 gene for the proteolipid subunit of ATP synthase

The numbers refer to the nucleotide sequence. The *NcoI* and *XbaI* restriction sites that were used to clone the two overlapping DNA fragments are shown. *Cis*-acting DNA sequences that are of importance in control of gene expression have not as yet been identified. However, the double- and triple-underlined sequences are conserved at corresponding positions in the human P2 gene (M. R. Dyer & J. E. Walker, unpublished work) and might contain TATA boxes (Breathnach & Chambon, 1981). The overlined sequence is found 63 bp upstream from one of these conserved AT-rich elements and might be a functional CCAAT box (Efstratiadis *et al.*, 1980). The single underlined sequence denotes a potential signal for polyadenylation (Proudfoot & Brownlee, 1977). Intron–exon boundaries are denoted by arrows. A bovine repetitive element is shown also. It is related to the repetitive sequence in the bovine foetal β -globin gene (Duncan, 1987; see Fig. 4).

ADP/ATP translocase (Cozens et al., 1989). These observations support the view that some exons encode structural domains (Gilbert, 1978; Blake, 1979).

The nucleotide sequences adjacent to the 5' and 3' boundaries of the introns in the bovine P2 gene are conserved (see Table 1). They all begin with the dinucleotide GT and end with the dinucleotide AG, and so they

conform exactly with the consensus sequences adjacent to splice junctions (Breathnach & Chambon, 1981). Furthermore, the conservation extends for 8–10 bp beyond the splice junctions in the sequences of the introns, and these extended sequences agree rather well with the consensus for sequences around splice sites (Mount, 1982).



Fig. 3. Structure of the bovine P2 gene encoding the mitochondrial import precursor of the proteolipid subunit of ATP synthase

Exons I–IV and introns A–C are denoted by black boxes and solid lines, respectively. The sizes of exons and introns are given in base pairs. The site of initiation of transcription, and therefore the structure of the gene in the 5' non-coding region, are not known at present.

An Alu-type repeated sequence in the bovine P2 gene

In mammalian DNA the largest class of intermediate repeated sequences is known as the Alu family. In humans and other primates the Alu sequences are well conserved (Deininger et al., 1981; Daniels et al., 1983), although in the prosimian species Galago a second distinct but related repeated sequence is also present (Daniels & Deininger, 1983). The bovine genome, in common with those of other ruminants, contains Alu-type elements which can be composed of monomers, dimers and trimers of a 120 bp DNA sequence, and the bovine Alu-type repeat shares similarity with the human Alu sequence over a 40 bp segment of DNA (Watanabe et al., 1982). Recently, DNA sequencing studies of the bovine β -globin locus have demonstrated a second and distinct bovine Alu-type family (Duncan, 1987). Members of this family of repeated DNA sequences are about 500 bp long and there are about 10⁵ members in the bovine genome. They are related to the original bovine Alu-type repeated sequence described by Watanabe et al. (1982) over a 75 bp segment of DNA (Duncan, 1987), but the remainder of the sequences from these two families are quite different. This second family of bovine Alu repeats is not similar in its DNA sequence or structure to human Alu repeats, nor is it composed of two related monomers, and it lacks a poly(A) tract near its 3' end. The sequence that encompasses the bovine P2 gene contains most of

the nucleotide sequence of an *Alu*-type repeat belonging to the family described by Duncan (1987; see Fig. 4). It lacks 45 bp found in the full-length repeat, presumably because it lies at the extremity of the sequence that has been determined.

DNA sequences of two bovine pseudogenes

As mentioned above, a 4.0 kb EcoRI fragment in λ P3.17 and a 5.2 kb SacI fragment in isolate λ P3.9 hybridized with the bovine P1 specific gene probe. Sequencing experiments showed that each fragment appears to contain a different pseudogene, and their DNA sequences are presented in Figs. 5 and 6. These sequences have been determined in both senses of the DNA. The first fragment appears to contain the sequence of a spliced pseudogene related to the bovine P1 cDNA sequence. However, it differs in 56 nucleotide positions from the cDNA, and some of these changes result in 29 amino acid differences. The changes also give rise to two 'in-phase' stop codons, and the sequence contains a frame-shift and a 6 bp deletion. These features form part of the basis of the assignment of this sequence as a pseudogene. In addition, this pseudogene has a poly(A) tract near to its 3' end which is preceded by a potential polyadenylation signal (Proudfoot & Brownlee, 1977), and the sequence is flanked by a direct repeat of 10 base pairs. These latter characteristics indicate that this sequence was introduced into the bovine genome by retroposition (Rogers, 1985; Weiner et al., 1986).

The second sequence (Fig. 6) also is related to that of the bovine P1 cDNA, and in all probability it is a pseudogene, although other explanations of its origin are possible. In common with the spliced pseudogene described above, it contains an 'in-phase' stop-codon, and it has four small deletions relative to the bovine P1 cDNA sequence, from which it differs by 50 nucleotides; some of these differences give rise to 31 changes in amino acid sequence. However, it differs in a number of ways from the other pseudogenes for P1 and P2 that have been characterized from the human (M. R. Dyer & J. E. Walker, unpublished work) and bovine genomes. Firstly, no poly(A) tract is found near to the 3' end of the sequence, although a potential polyadenylation signal is present; secondly, it is flanked by a shorter than usual direct repeat. This is six bases long, with the sequence CTGGGA, and the position of the direct repeat at the 3'

			Sequence				
Gene	Intron	Size	Class	5' boundary	3' boundary		
Bovine P2	А	1069	0	tcc.ttg.GIGAGIACC	CTTCCGGCTAG.atc.agg I R		
Bovine P2	В	506	0	gat.gag.GIACCTTAC	ATICTICACAG.agc.cac S H		
Bovine P2	С	1607	2	gcc.ag.GTAAGATGG A R	GCCCCTCCCAG.g.aac N		

Table 1. Introns in the bovine P2 pre-proteolipid gene

Consensus sequence

		20	40	60	80		
P2			CCATGG	ATTTCAGAACACCAGGCTTC	CCTGTCCATCACCAACTCCTG	GAGCTTATGGAAACTCATGTTCATCGA	G
	TTAAGTTCAGTTCAG	TCACTCAGTCATGTCC	GACTCTTTGCGACCCCATGA	ATCACAGCACGCCAGGCCTC	CCTGTCCATCTCCTCCCC	GAGTTCACCCAACGTCACGTCCATCGA	ŝ
B	TTCTCTTCACTCCAC	TCCCTCACTTCTCTAC	A APPCPTTCC A ACCCC APCC	ACTCC ACC ATCCC ACCCTTC	CCTCTCCATCACCAACTCCTC	CACCTTCCTCAAATTCATCCCCATTCA	õ
-	** ******						
-	120	140	160	180	200	220	
12	TCAGTGATGCCATCC	AACCATCTCATCCTCI	IGTCGTCCCTTTCTCCTG	CCTTCAGTCTTTCCCAGCGT	CAGGGTCTTTTCAAATGAGTC	ATCTCTTTGCATCAGGTGGCCAAGGT	T
A	TCGGTGATGCCATCC	AGCCATCTCATCCTCT	GTCGTCCCCTTTTCCTCCTG	CCCCCAATCCCTCCCAGCAT	CAGGGTCTTTTCCAGTGAGTC	AACTCTTCACATGAGGTGGCCAAAGTA	S,
в	TTGGTGATGCCATTC	AACCATCTCATCCTCI	GTTATCCCCTTCTCTTG	CCTTCAATCTTTCCCAGCAT	CAGGGTCTTTTCCAATGAGTC	AGTTCTTCACATCACGTGTCCAAAGTA	Т
	* ********	* ***********	*** **** ** *** **	** ** ** *******	************ * ******	* **** *** * *** **** ***	
	240	260	280	300	320	340	
82	TCCACTTANACCTA	200 ACCATCACTCCCTCC	200 ATCA ACATT_CACCACTCAT	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	320 TCC 3 TOTO COTTOC 3 CTCC 3 3C		
	TOGAGITITAGCITC	AGCAICAGICCCICCA		CICCIIIAGGAIGGAIIGGI	TGGATCICCIIGCAGICCAAG	GGACICICAAGAGICIICICCAGCACC	
8	IGGAGIIICAGIIII	AGCAICATICCTICCA	MACAACACCCCAGGGCTTAT	CICCITIAGAAIGGACIGGI	TGGATCTCCCTGCAGTCCAAG	GGGUTUGUAAGAATUTTUTUUAAUAUU	A
D	IGAAGCTTCACGTTC	GGTATCAGTCCTTCCA	ATGAATATT-CAGGACTAAT	ATCCTTCAGGTTTGATTGGI	TTGATCTCCTTGCAGTCCAAG	GGACTCTCAGGAGTCTTCTCCAACATC	A
	** ** ** * **	* **** *** ***	* ** * * **** ** **	***** ** * ** ****	******** ***********	** *** ** ** ********* ** *	*
	360	380	400	420	440	460	
P2	CAGTTCAAAAGCATC	AATTTTTCAGCACTC#	GCTTTCTTTATATTCCAACT	CTCACATCCATACATGACTA	CTGGAGAAACCATAGCTTTGA	CTAGATGGACGTTTGTGGGCAAAGTAA	Т
	CAGTTCAAAAGCATC	AATTCTTCGGCACTC	ACTTTCTTCACAGTCCAACT	CTCACATCCGTACATGACCA	CTGGAAAAACCATAGCCTTGA	CTAGACGGACCTTTGTTGGAAC-GTA	Т
в	CAGTTCAAAAGCATC	AATTCTTCAGCGGTC	CCTTTCTTCATTGTCCAACT	CTCACATCCACACATGACTA	CTGGAAAAACCATAGCCTTGA	CTAGATGGACATTTGTTGATAAAGTAG	т
	***********	**** *** ** ***	******* * ******	********	***** ********* ****	***** **** ***** * * ***	*
	480	500	520				
P2	GTCTCTGCTTTTTAA	TACACTGTCTAGGTTC	GTCATAACTTTTCTTCCAAG	SAGCAGGCATC			
A	GTCTCTGCTTTTCAA	TATECTATCTAGETTE	GTCATAGCTTTCCTTCCAAG	GAGTAAGCA			
Ř	CTCTCTCCCTTTTT	TATCOTOTACA	CTC ATTACCTTTTTTTTTTTTTTTTTTTTTTTTTTTTT				

Fig. 4. An Alu-type repeat in the bovine P2 gene

The 5' region of the fragment of bovine genome containing the P2 gene contains most of the DNA sequence of an Alu-type repeat first described by Duncan (1987). This sequence is aligned with two other members of this family of repeats from the cow β -globin locus. Sequences A and B are Alu-type elements from the 5' and 3' flanking regions of the cow foetal globin gene respectively (Duncan, 1987). The nucleotide sequences are numbered according to the Alu-type element A. –, Insertions needed to improve these alignments; *, conserved nucleotides. The underlined sequence is conserved in the original cow Alu-type family described by Watanabe et al. (1982).



Fig. 5. DNA sequence and translation of a processed P1 pseudogene

The numbers refer to the nucleotide sequence. :, Nucleotide differences with the bovine P1 cDNA sequence (Gay & Walker, 1985); +, amino acid changes with the P1 pre-proteolipid. The potential polyadenylation signal is underlined. This gene is unlikely to encode a functional polypeptide since it has two in-phase stops, a frame-shift and a 6 bp deletion within the potential protein coding region. This pseudogene has two features which are diagnostic of retroposition (Rogers, 1985; Weiner *et al.*, 1986); it is flanked by 10 bp direct repeats and has a poly(A) tract at a position corresponding with the poly(A) tail in the mRNA.

TGGGGAATTCCCATCA	TATGAGTCTTGGTGAGAAGA	ARAGAGTACCACTTAATGT	CCTGCTGAAAAAACCACCTT	TCCAAGTCCTAGAGCTAAA	N Q T SCTGGGAGATTGAAAAACATGCAGAC
	20	10	60	80	120
+ P G A L L CCCTGGGGCACTACTO	+ + I P P A F CATTCCTCCTGCTTTTATCTC	CTTTTTCTGTTGTTCAGTT	GCTAAGTCCTGTCCAACTCT	TTGTGACCCCACAAACTGC	AGCACACCAGGCTTCCCTGTCCTTCA
::	: :: - 1 1 0	160	180	200	220 210
CTATCTCCCAGAGTT	TGCTCARACTCATGTCCACT(260	AGTCAGTGATGCCATGTCA 280	TCCTCTGTTATCCCCTTCTC 300	CTGCCTTCACTCTTTCCCA 320	GCATCAGGGTCTTTTCCAATGAGTCA 310 360
GCTCTTCAAATCAGG	TGGCTAGAGTTTTGGAGCTTC 380	CAGCAGCAGTCCTTCTGATG	AATATTCAGGACTGATTTCC 120	TTTAGGATTGGCAGGTTTG 11 0	ATCTTCTTGCAGTCCAAGGGATTCTC 460 480
		Inter	rvening DNA sequence	of 861bp	******
AAGAGTTTCCTCCAG	TACCACAGTCTGAAAGCATCI 500	S20	540	560	580 600
ANATGATGTCTCTGC	TTTTTARTATGTTGTCTAGG 620	ITTGTCATAGCTTTATCTTC 640	CAAGGAGTTCAGTTCAGTCA 660	CTCAGTCATGTCCGAGTCT 680	TTGCGACCCCATGAACTGCAGCACGC 700 720
CAGGCCTCCCTGTCC	ATCACCAACTCCTGGAGTCCI 740	ACCCARACCCATGTCCATTG 760	AGTCAGTAATGCCATCTAAC 780	CATCTCATTCTCTGTTGTC 800	CCCTTCTCCTCCTGCCCTCAATCTTT 820 840
CCCAGCATCAGGGTC	TTTTCAAATGAGTCAGCTCT 860	ICACATCAGGTGGCGTCATT 880	TAATTTTGTGGCTATAGTCA 900	CCATCCACAGTAACTTTGG 920	AGCCCAAGAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
GTTTCCATCACTTCA	TGGCAAATAGATTCAGAAAAA	I ANGTGGANACAGTGACC G AT	+ + + Y S C P K G Ctactcttgtcccaagggtc	+ L I R L V S A Tgatcaggcttgtgtctgc	+ + + + + S F L R K P A U P Ctccttcctgaggargccagcggtcc
	980	1000	1020	: 10 1 0	::::: 1060 1080
+ S E Q P S Crtctgracagcctte	+ S Y S S U P L CCTACAGCAGTGTCCCACTC	+ + + * E A R R E S IAGGAGGCTAGGCGGGAGTC	+ Q T S R D I CCRGRCCAGTGCTGATATTG	D T A A K F I Acacagcagccaagttat	G A G A A T U G U Tggcgctggggctgccacagttggtg
	1100	• • • • • • •	11 1 0 ∳	: 1160	1180 1200
	in-phase	l s stop codon	9bp deletion	• •	• • •
A G S G I Tggctggttcaggag	+ A S I G T V F CCAGCATTGGAACAGTCTTTG	+ + G S L I A D Y GCAGCTTGATCGCTGACTA	A R N P S L Tgccaggaacccgtccctga	+ P C K Q R L F Agcagcgactcttccctgt	A U L G F A U A E
: :	1220	1240	:	1280	: : : 1300 1320
* *	• •	•		 frame-shift	
ARRLF	CLMAPFI	. I F T H +			
GGCTAGGAGGCTCTT	CTGTCTGATGGCCCCCTTCC	ICATCTTCACCATGTGAGAC	TCCGTGGAGGTCACCTACTT	ATCCTTGCTGCTCAACTCC	AGGCCATGCCCAGTACTTTACCATTA
::	1340	Ť	1380	1100	1120
	•	3bp deletion		1bp deleti	on 15bp deletion
AACACAGCATTTCTG	GGACTT				

Fig. 6. DNA sequence and translation of a partially processed P1 pseudogene

The numbers refer to the nucleotide sequence. :, Nucleotide differences with the sequence of the bovine P1 cDNA (Gay & Walker, 1985); +, amino acid changes in comparison with the P1 pre-proteolipid. A potential signal for polyadenylation is underlined. This pseudogene is unusual since it contains an 861 bp intervening DNA sequence corresponding in position to an intron in the human P1 gene (M. R. Dyer & J. E. Walker, unpublished work). Boundaries between this intervening sequence and the P1 coding sequence are denoted by arrows. This pseudogene is unlikely to encode a functional polypeptide as it contains an in-phase stop codon, frame-shifts and 9 bp and 3 bp deletions within its potential protein coding region. Two other deletions are found in the DNA sequence corresponding to the 3' untranslated region of the message. This pseudogene is flanked by short direct repeats of 6 bp.

end of the pseudogene suggests that the poly(A) sequence might have been removed during retroposition (Rogers, 1985; Weiner *et al.*, 1986). A third difference is that it contains an intervening sequence of 861 bp. This sequence is found in a position that corresponds to that of an intron in the human P1 gene (M. R. Dyer & J. E. Walker, unpublished work), but the sequence is not related to that of the equivalent intron in human P1. This bovine intervening sequence is flanked at its 5' and 3' ends respectively by the dinucleotides, AT and CG; these could be mutated canonical splice junction sequences, GT and AG (Breathnach & Chambon, 1981).

Another mammalian gene which could have been formed from a partially processed mRNA encodes insulin in mice and rats (Soares *et al.*, 1985). Both species have two expressed non-allelic insulin genes. The gene for preproinsulin II contains two introns and therefore is similar in its structure to the single copy genes for this protein in other mammalian species. However, the gene for preproinsulin I contains only a single intron and has distinctive features that are diagnostic of a retrogene; its sequence is flanked by 41 bp direct repeats and there is the remnant of a poly(A) tract downstream from the polyadenylation signal. There is no evidence that suggests expression of a protein from the partially spliced bovine P1 gene.

A second explanation for the origin of the partially spliced P1 sequence can be advanced. It is possible that this sequence is not a retroposon, but rather that it is a relic of a duplicated version of an early expressed P1 gene, which at the time of duplication had only one intron. However, since its sequence is closer to the bovine P1 cDNA than it is to the human P1 cDNA sequence (M. R. Dyer & J. E. Walker, unpublished work) this explanation is less plausible.

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