

Original Article

RNF26-mediated ubiquitination of TRIM21 promotes bladder cancer progression

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Abstract: RNF26 is an important E3 ubiquitin ligase that has been associated with poor prognosis in bladder cancer. However, the underlying mechanisms of RNF26 in bladder cancer tumorigenesis are not fully understood. In the present study, we found that RNF26 expression level was significantly upregulated in the bladder cancer tissues, and higher RNF26 expression is closely associated with poorer prognosis, lower immune cell infiltration, and more sensitive to immune checkpoint blockade drugs and chemotherapy drugs, including cisplatin, VEGFR-targeting drugs and MET-targeting drugs. RNF26 knockdown in UMUC3 and T24 cell lines inhibited cell growth, colony formation and migratory capacity. Meanwhile, RNF26 overexpression had the opposite effects. Mechanistically, RNF26 exerts its oncogenic function by binding to TRIM21 and promoting its ubiquitination and subsequent degradation. Moreover, we revealed ZHX3 as a downstream target of RNF26/TRIM21 pathway in bladder cancer. Taken together, we identified a novel RNF26/TRIM21/ZHX3 axis that promotes bladder cancer progression. Thus, the RNF26/TRIM21/ZHX3 axis constitutes a potential efficacy predictive marker and may serve as a therapeutic target for the treatment of bladder cancer.

Keywords: RNF26, TRIM21, ZHX3, bladder cancer, ubiquitination

Introduction

Bladder cancer has become the second most common urological cancer, and the incidence and mortality rates are increasing every year [1, 2]. Although the clinical outcomes of bladder cancer have improved significantly with the development of surgical approaches and targeted therapy, the 5-year survival rate for patients with bladder cancer is still unsatisfactory [3]. In addition, bladder cancer tends to have a high rate of postoperative recurrence, possibly due to its multifocal and implantation characteristics [4]. However, the behind mechanisms are still unclear. Therefore, in-depth studies are urgently needed to further our understanding of bladder cancer tumorigenesis and progression and to identify novel diagnostic and therapeutic targets for bladder cancer.

Ubiquitination is an important post-transcriptional modification that can alter protein local-

ization and lead to protein degradation [5]. E3 ubiquitin ligase is the key enzyme for ubiquitination, transferring ubiquitin from the E2 enzyme to the underlying protein [6]. In addition, E3 ubiquitin ligases have been closely associated with tumorigenesis, progression, microenvironment, immunotherapy, and drug resistance in bladder cancer [7], and thus utilization of E3 ubiquitin ligase-targeted drugs has emerged as a novel approach for bladder cancer treatment [8]. ER-embedded RING finger protein 26 (RNF26) is an important E3 ubiquitin ligase that mediates ubiquitination of sequestosome 1 (SQSTM1) by interacting with the ubiquitin-binding enzyme UBE2J1, so as to anchor homologous vesicles in the perinuclear region of the cell and to facilitate the termination of epidermal growth factor (EGF)-induced AKT signaling [9, 10]. RNF26 can also collaborate with vimentin to maintain endoplasmic reticulum architecture and dynamics [11]. Moreover, RNF26 is prevalent in normal human tissues but is upregulated in several human

tumor cell lines (HL-60, HeLa S3 and SW480) [12]. Studies have shown that RNF26 could promote progression of pancreatic cancer and renal cell carcinoma through degrading RNA binding motif protein-38 (RBM38) and chromobox 7 (CBX7), respectively [13, 14]. Significantly higher expression of RNF26 was detected in bladder cancer cells and tissues, which was significantly associated with poor prognosis. Forkhead box M1 (FOXM1) promotes RNF26 expression at the transcriptional level, and RNF26 degrades p57 (CDKN1C) through a ubiquitination-dependent manner to regulate cell cycle processes [15]. Nevertheless, the relationship between RNF26 and the immune microenvironment of bladder cancer, the effect of chemotherapy, the benefit of immunotherapy, and how ubiquitination of candidate proteins may play a role in the progression of bladder cancer remains unclear.

In this study, we systematically investigated the relationship between RNF26 expression and bladder cancer prognosis and progression, immune microenvironment, chemotherapeutic drug efficacy, as well as the immune checkpoint blockade therapy benefits. The oncogenic effects of RNF26 and the candidate RNF26 binding proteins and their ubiquitination were also explored, suggesting existence of a novel tripartite motif containing 21 (TRIM21)/zinc fingers and homeoboxes 3 (ZHX3) axis in bladder cancer tumorigenesis and progression.

Materials and methods

Bioinformatics analysis

RNF26, TRIM21 and ZHX3 expression in normal and bladder cancer tissues from TCGA (<https://portal.gdc.cancer.gov/>). Cases were stratified according to median expression level into high and low expression groups for RNF26, TRIM21, and ZHX3. The relationship between RNF26, TRIM21, and ZHX3 expression and bladder cancer survival outcomes, including overall survival (OS), disease free survival (DFS) and progress free interval (PFI), was analyzed using Kaplan-Meier (KM) survival curve analyses. We also analyzed the differences in infiltration of 22 immune cell types between the two expression groups using the CIBERSORT algorithm based on the LM22 gene signature file [2, 16, 17]. In light of the importance of checkpoint inhibitor-based therapy, the expres-

sion patterns of immune checkpoint molecules in the high and low expression groups were further analyzed [18]. The half-maximal inhibitory concentration (IC₅₀) values of each TCGA-bladder cancer sample were assessed using the pRRophetic R package [19], and the IC₅₀ values of different therapeutic agents in the high-risk and low-risk groups were compared using the Wilcoxon signed-rank test to determine the sensitivity of the different expression groups to chemotherapeutic agents [20].

Cell culture and transfection

The human bladder cancer cell lines UMUC3 and T24 were obtained from the Chinese Academy of Cell Collection (Shanghai, China) and cultured in RPMI 1640 medium containing 10% foetal bovine serum, 100 U/ml penicillin-streptomycin, at 37°C under 5% CO₂ in a humidified incubator. For cell passages, cells were detached using 0.25% trypsin-EDTA (Invitrogen, USA) for 3-5 minutes at 37°C. The cell suspension was then centrifuged at 300 g for 5 minutes. Cells were counted using a hemocytometer, and the appropriate number of cells were seeded into new culture flasks at a 1:3 ratio. For transient knockdown, small interfering RNAs (siRNAs) (GenePharma, Suzhou, China) and overexpression plasmids (GenePharma) were transfected into UMUC3 and T24 cells with Lipofectamine 2000 (Invitrogen) according to the manufacturer's instructions. The nucleotide sequence of siRNA was as follows: si-NC, 5'-UUCUCCGAACGUGUC-AGGU-3'; si-RNF26 1#, 5'-UAGUGAAAUGUGC-ACAGUCUG-3'; si-RNF26 2#, 5'-UUGAGGUCC-AACACCAAGGUC-3'; si-ZHX3, 5'-AUGAUUGUCU-GGCUUGGCC-3'. To investigate potential forms of lysine ubiquitination, specifically focusing on the prevalent K48 ubiquitination-mediated protein degradation, T24 cells were transfected with Ub-WT, Ub-K48, Ub-K48R, and Ub-KO mutants. Cells were harvested for analysis at 48 h post-transfection.

RNA extraction and quantitative real-time (qRT)-PCR

Total RNA was extracted using the RNeasy Plus Micro Kit (Qiagen, Düsseldorf, Germany). RNA was reversely transcribed into cDNA using PrimeScript Reverse Transcription Kit (Vazyme, Nanjing, China). The mRNA expression was quantified on an ABI 7500 Real-Time PCR

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System (Applied Biosystems, Foster City, CA, USA) using SYBR Premix Ex Taq polymerase (Takara, Dalian, China). 18S ribosomal (r)RNA was chosen as an internal control and relevant expression was calculated by $2^{-\Delta\Delta Ct}$ method. The following primers were used in the qRT-PCR reaction: RNF26 forward 5'-ACTGAAAT-CCTGATGCGCCA-3' and reverse 5'-ACAGGGGT-AGGAGGGGATTC-3'; and 18S rRNA forward 5'-AAACGGCTACCACATCCAAG-3' and reverse 5'-CCTCCAATGGATCCTCGTTA-3'.

Cell proliferation, colony formation and migration assays

Cell viability was measured using the CCK-8 kit (Beyotime, Beijing, China), as previously described [21-24]. The optical density (OD) was measured at 450 nm wavelength using a microplate reader (Bio-Rad Model 680, Richmond, CA, USA). For colony formation, 1000 transfected cells were seeded in 6-well plates and maintained in RPMI-1640 medium containing 10% FBS. After two weeks, cell colonies were fixed with methanol for 15 min and stained with 0.1% crystal violet (Sigma-Aldrich) for 30 min. Subsequently, those visible colonies were photographed and counted using the ImageJ software.

Cell migration assay was performed by a transwell chamber (Corning, USA), as previously described [25-27]. In brief, transfected cells were suspended in 150 μ L of serum-free medium and cultured in the upper transwell chamber and the lower chamber was filled with 600 μ L of the medium supplemented with 10% FBS. After 48 h of incubation, cells inside the upper chamber were removed using cotton wool, and the cells moving to the lower side were fixed with 4% paraformaldehyde and dyed with 0.1% crystal violet (Beyotime). Finally, five randomly selected fields of view were photographed under an inverted microscope and counted in each well.

Xenografts assay

The animal experiments were approved by the Soochow University Animal Ethics Committee. In brief, four-week-old male BALB/c nude mice were maintained in specific pathogen-free (SPF) animal room and manipulated according to protocols. ShRNF26 and negative control shRNA (GenePharma) were stably transfected

into UMUC3 cells. The transfected cells were subcutaneously injected into a single side of the posterior flank of each mouse. Tumor growth was examined every 3 days, and tumor volumes were calculated by the equation $V = 0.5 \times D \times d^2$ (V, volume; D, longitudinal diameter; d, latitudinal diameter). After 15 days, mice were euthanized under carbon dioxide exposure followed by cervical dislocation, the subcutaneous tumors were obtained and imaged, and tumor volumes and weight were examined. Tumor cell proliferation was assessed by Ki67 staining. The specimens were fixed with methanol/acetone and incubated with Ki67 antibody (1:200 dilution; Abcam, USA). The percentage of Ki67-positive cells was determined by cell counting at least five microscopic fields of view. This study was carried out in strict accordance with the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health.

Immunoprecipitation (IP) assays

Cell lysates were incubated with anti-Flag antibody (Sigma) or anti-GFP antibody (Abcam) for Co-IP experiments as previously described [28]. Briefly, cells were lysed using RIPA lysis buffer (Beyotime, Nantong, China) on ice for 30 min, followed by preclearing with protein A beads (Invitrogen). Lysates were incubated with the indicated primary antibodies or IgG antibody overnight at 4°C, followed by incubation with protein A beads for 3 h at 4°C. The immunoprecipitates were prepared for subsequent western blotting analysis by applying heat (95°C) to the beads with 2 \times SDS buffer for a duration of 5 minutes.

Liquid chromatography-tandem mass spectrometry analysis (LC-MS/MS)

The gel particles were washed sequentially with deionized water, 50% acetonitrile, and 100% ACN. The proteins contained in the gel particles were reduced and alkylated for 45 minutes at room temperature. The gel particles were then washed thoroughly and dried naturally. The gels were hydrated at 4°C for 30 minutes. The gels were then digested at 37°C for 12 hours. All extracts and supernatants were combined and dried using a SpeedVac. The digested peptides were desalted using a StageTip and analyzed using an LTQ Orbitrap Velos mass spectrometer (Thermo Scientific,

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Waltham, USA). Tandem mass spectra were searched using the UniProt protein database to identify proteins and peptides using a false discovery rate of 1%. All experiments were performed in triplicate. Protein expression levels were estimated using the iBAQ algorithm embedded in MaxQuant for label-free quantification.

Protein half-life assay

After transfection with Empty vector (EV) or pcDNA3.1-RNF26, T24 cells were treated with cyclohexanone (CHX, 100 µg/mL, Sigma) at different time points to block protein synthesis, and proteins were harvested for western blotting analysis to detect TRIM21 at 0, 3, and 6 hours.

Western blotting

For western blotting analysis, total protein was extracted from cell lines and tissue samples using RIPA buffer. Proteins were separated by 10% SDS-PAGE electrophoresis according to molecular weight and then transferred to a polyvinylidene difluoride membrane (Millipore, Bedford, MA, USA). Membranes were blocked in Tris-Tween (TBST) containing 5% nonfat dry milk for 1 hour at room temperature and incubated with the indicated primary antibody dilutions (anti-RNF26, 1:1000; anti-TRIM21, 1:1,000; anti-ZHX3, 1:1,000, and anti-tubulin, 1:3,000; all from Proteintech) overnight at 4°C. On the next day, the membranes were washed 3 times for 10 min each and incubated with horseradish peroxidase (HRP)-conjugated secondary antibody for 1 h at room temperature. Protein bands were detected using an enhanced chemiluminescent substrate (Thermo Scientific). In our study, bladder cancer samples and normal tissues for Western blotting were obtained from patients treated for bladder cancer at the Second People's Hospital of Lianyungang between February 2021 and February 2023 and were approved by the Second People's Hospital of Lianyungang's Ethics Committee (2021-007-01).

Immunofluorescence

For immunofluorescence, fresh 5 µm slides were obtained from paraffin-embedded bladder cancer and control tissue from xenografts assay. 5% bovine serum albumin (BSA, w/v;

Sunshine, Nanjing, China) is used for blocking to reduce non-specific binding of antibodies. The sections were incubated with 100 µl of primary antibody dilutions (anti-ZHX3, Proteintech; anti-TRIM21, Proteintech; anti-Ki67, Abcam) overnight at 4°C. Subsequent, Alexa-Fluor secondary antibodies staining, 4',6-diamidino-2-phenylindole (DAPI) staining, and photographed under a confocal laser microscope (Zeiss LSM710, Carl Zeiss, Oberkochen, Germany).

Statistical analysis

Data were analyzed using GraphPad Prism 9.0 (GraphPad, USA) and SPSS 22.0 (IBM, Chicago, IL, USA). Data from at least three independent replicate experiments are presented as mean ± standard deviation (SD). Differences between the two groups, or between more than two groups, were analyzed using Student's t-test or one-way ANOVA [29]. For multiple time points, a Tukey post hoc test following the Repeat measurement ANOVA was adopted to compare the two groups. $P < 0.05$ was set as the threshold for statistical significance.

Results

RNF26 expression significantly correlates with prognosis and response to targeted treatment in bladder cancer patients

To investigate whether RNF26 plays an oncogenic role in bladder cancer progression, RNF26 expression in para-cancerous tissue and tumor tissue was analyzed using TCGA data. RNF26 expression level was significantly upregulated in the bladder cancer tissues compared to para-cancerous tissue (**Figure 1A**). Then, we performed western blotting to detect RNF26 expression in patients with bladder cancer and found that RNF26 significantly overexpressed in bladder cancer tissues compared with normal counterparts (**Figure 1B, 1C**). KM survival analysis showed that low expression of RNF26 were related to the good overall survival (OS) and disease free survival (DFS) in bladder cancer from TCGA database (**Figure 1D, 1E**). The 22 immune cell proportions of bladder cancer patients in the RNF26 high- and low-expression groups were quantified by the CIBERSORT algorithm. Compared to high-expression group, B cell memory, B cell plasma, T cell follicular helper, T cell regulatory (Tregs), Monocyte, Myeloid dendritic cell acti-

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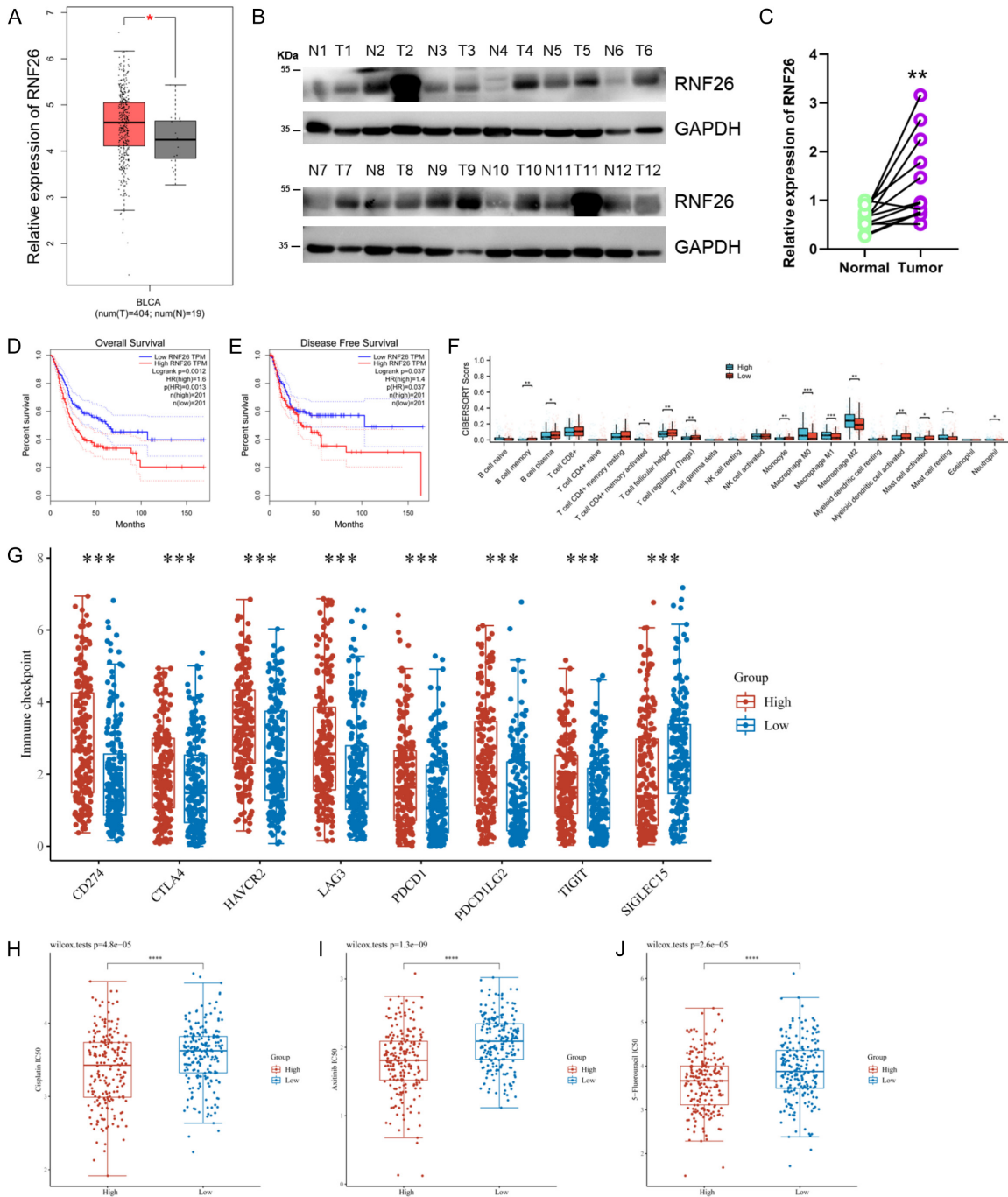


Figure 1. RNF26 expression was significantly associated with prognosis and drug sensitivity in patients with bladder cancer. A. RNF26 expression was upregulated in the bladder cancer tissues in TCGA-BLCA dataset. B, C. RNF26 expression in bladder cancer patients using western blotting. D, E. Overall survival (OS) and disease-free survival (DFS) were significantly worse in bladder cancer patients with high RNF26 expression from TCGA database. F. Comparison of immune cell infiltration between the high and low RNF26 expression groups. G. The difference in immune checkpoint molecules between the high and low RNF26 expression groups. H-J. RNF26 high-expression group were more sensitive to cisplatin, VEGFR targeted drugs (Axitinib) and MET targeted drugs (5-Fluorouracil). *P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001.

vated and Mast cell activated significantly higher in low-expression group, whereas T cell

CD4+ memory activated, Macrophages M0, Macrophages M1, macrophages M2, Mast cell

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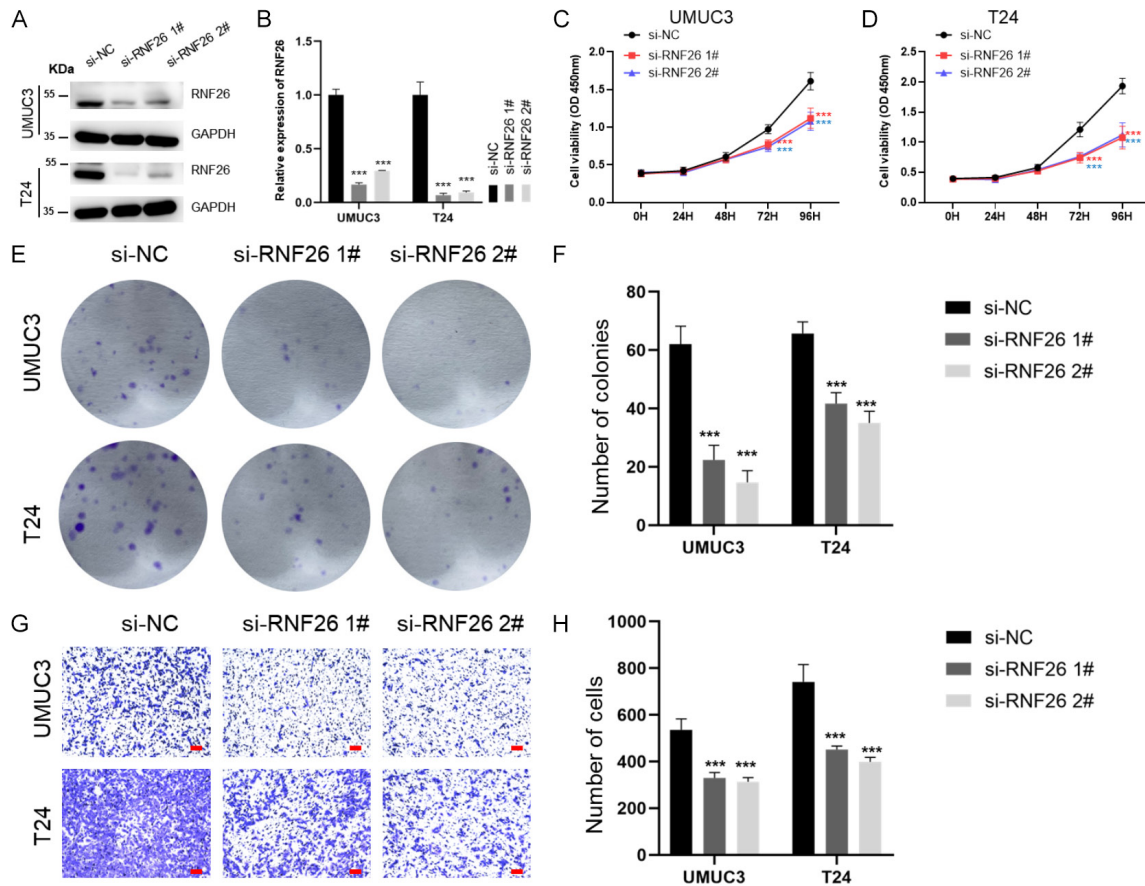


Figure 2. Knockdown of RNF26 inhibits bladder cancer cell growth. A, B. Cells were transfected with RNF26-siRNA to knockdown RNF26. C, D. CCK-8 assay indicated that the proliferation of UMUC3 and T24 cells was inhibited following RNF26 knockdown. $n = 6$ for each group. E-H. Colony formation and migrated cells were significantly reduced in RNF26-siRNA cells compared to control cells. $n = 3$ for each group. Scale bar: 100 μm . *** $P < 0.001$.

resting and Neutrophil were significant reduction in low-expression group (Figure 1F). Given the importance of checkpoint inhibitor-based therapy, we also compared the expression of immune checkpoint that differed between the two groups. Interestingly, the expression of all checkpoint molecules was remarkably higher in the RNF26 high-expression group than in the low-expression group, except for SIGLEC15 (Figure 1G), suggesting that patients in the RNF26 high-expression group may have greater immune escape and better benefit from immunotherapy. To investigate whether the response of chemotherapy correlates with RNF26 expression, the pRRophetic algorithm was performed. The patients of RNF26 high-expression group were more sensitive to cisplatin, VEGFR targeted drugs (Axitinib) and MET targeted drugs (5-Fluorouracil) (Figure 1H-J). These results suggest a correlation between

RNF26 and bladder cancer pathogenesis and have the potential to serve as a molecular marker to guide chemotherapy and immunotherapy in bladder cancer patients.

Knockdown of RNF26 inhibits bladder cancer cell growth in vitro and in vivo

Since the above results suggest that RNF26 is a potential bladder cancer oncogenic protein, we investigated the effect of RNF26 on bladder cancer tumorigenesis. Bladder cancer cell lines T24 and UMUC3 were transfected with si-RNF26, which leads to decreased expression of RNF26 (Figure 2A, 2B). CCK-8 assay results showed that knockdown of RNF26 significantly decreased cell proliferation in a time-dependent manner (Figure 2C, 2D). Colony formation assay and transwell assay explored the effect of RNF26 silencing on clonogenicity and migra-

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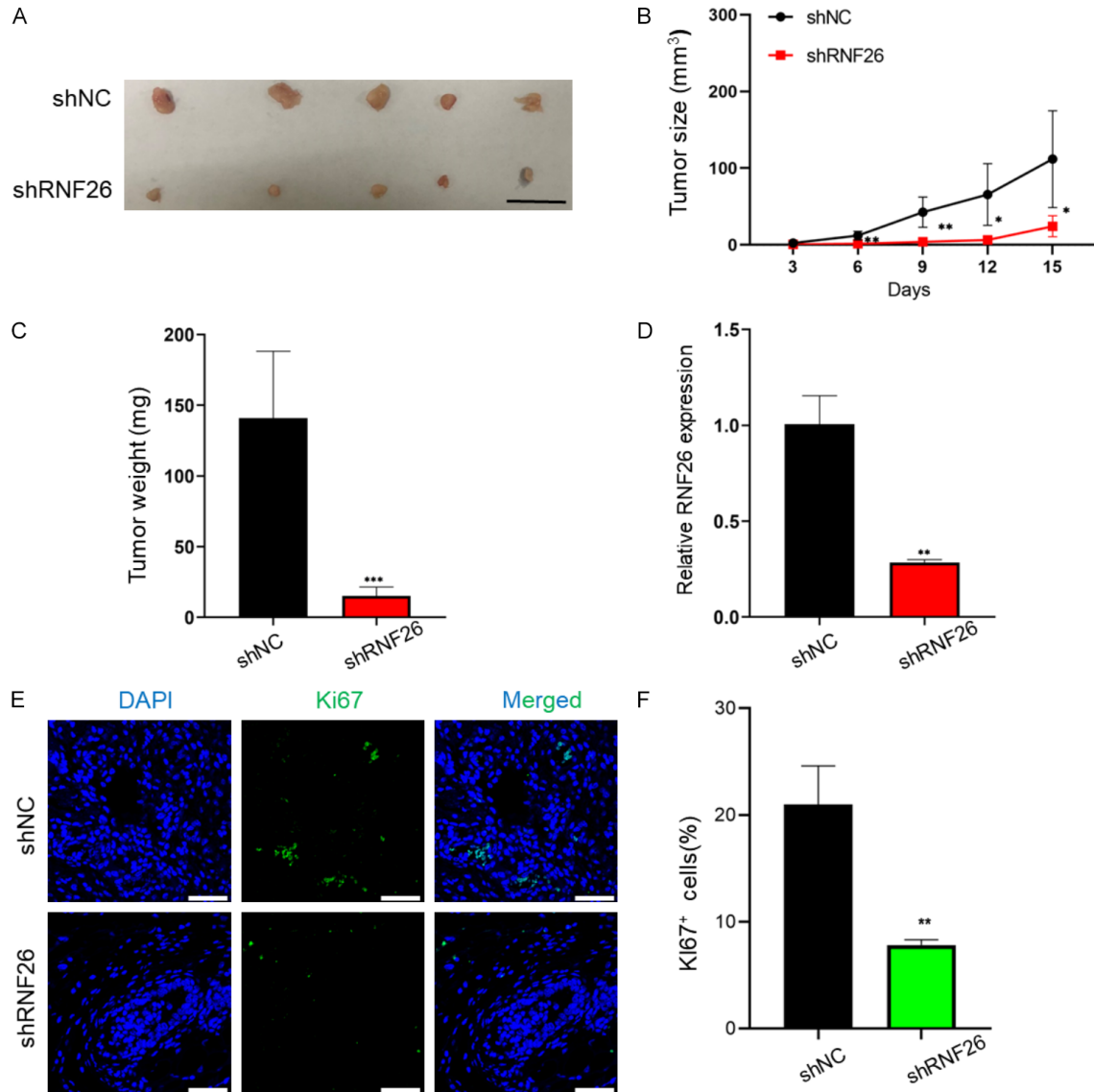


Figure 3. Knockdown of RNF26 inhibits bladder cancer cell tumorigenicity in xenograft tumor models. A-C. The tumor sizes and weight of mice were decreased in RNF26 knockdown group. $n = 5$ for each group. Scale bar: 1 cm. D. Relative expression of RNF26 in tumors transfected with shNC and shRNF26. E, F. Immunostaining of Ki67 indicates that RNF26 knockdown inhibits cell proliferation in xenograft tumor models. $n = 5$ for each group. Scale bar: 50 μm . * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

tion of bladder cancer cells and found that colony formation and migrated cells were significantly reduced in RNF26-siRNA cells compared to control cells (**Figure 2E-H**).

To further determine the role of RNF26 in bladder cancer development, we conducted xenograft assays in nude mice. The results showed that significant downregulation of RNF26 in xenograft tumour models and that RNF26 knockdown inhibited tumor growth and weight (**Figure 3A-D**). Moreover, immunofluorescence

staining indicated that Ki67-positive cells were significantly reduced in the RNF26 knockdown group (**Figure 3E, 3F**), suggesting that knockdown of RNF26 can inhibit bladder cancer cell proliferation in vivo.

Overexpression of RNF26 promotes bladder cancer cells proliferation and migration

To further validate the oncogenic effect of RNF26 in bladder cancer, T24 and UMUC3 were transfected with pcDNA3.1-RNF26. CCK-8

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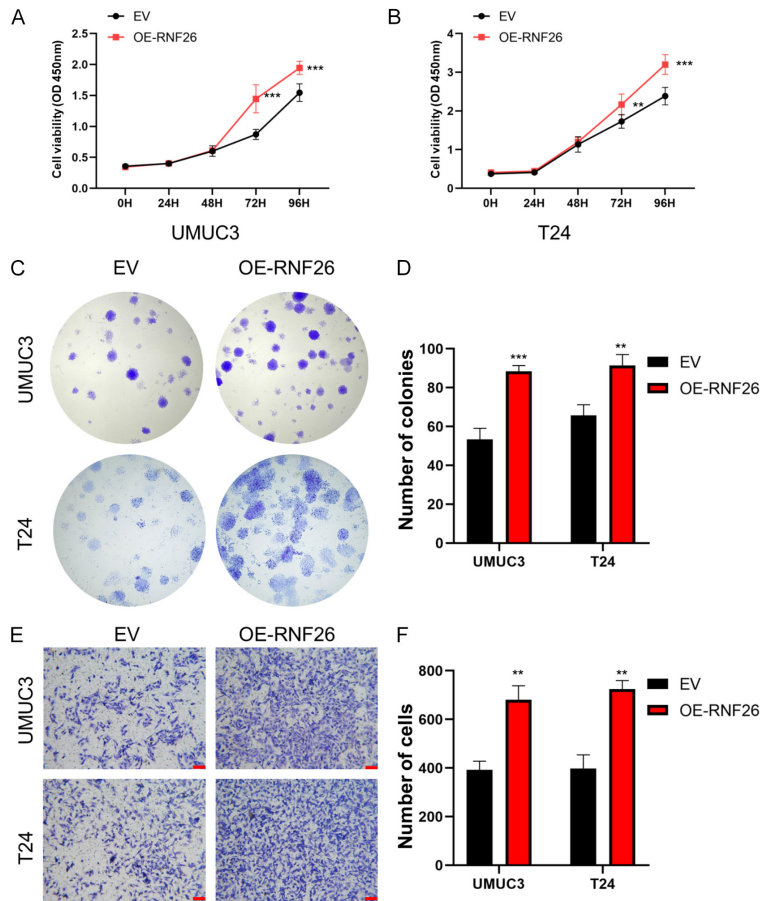


Figure 4. Overexpression of RNF26 promoted the malignant level of bladder cancer cells in vitro. A, B. CCK-8 assay indicated that the proliferation of UMUC3 and T24 cells was promoted following RNF26 overexpression. $n = 6$ for each group. C-F. Clonogenicity and migrated cells were increased in RNF26 overexpression cells compared to control cells. $n = 3$ for each group. Scale bar: 50 μm . ** $P < 0.01$; *** $P < 0.001$.

assay was performed and the results showed that overexpression of RNF26 significantly enhanced the growth ability of bladder cancer cells (Figure 4A, 4B). Moreover, colony formation and migrated cells were significantly increased in RNF26-overexpression cells compared to control cells (Figure 4C-F).

RNF26 interacts with TRIM21 and promotes its K48-linked ubiquitination

To clarify the downstream substrates of RNF26, we performed three independent immunoprecipitation-liquid chromatography-tandem mass spectrometry (IP-LC-MS/MS) analyses using RNF26 overexpressing T24 bladder cancer cells. As a result, 259 potential target proteins were identified from three inde-

pendent experiments (Figure 5A, 5B; Table S1). Among them, TRIM21 plays a key regulatory function in bladder cancer progression and is in the TOP10 candidates according to the iBAQ ranking (Figure 5C) [30]. Therefore, TRIM21 was selected as a candidate target for RNF26. First, Co-IP experiments were performed to confirm the interaction between RNF26 and TRIM21 (Figure 5D). In bladder cancer cells, TRIM21 works as an E3 ligase to mediate ZHX3 ubiquitination and subsequent degradation [30]. We therefore supposed that ZHX3 may be involved in the RNF26/TRIM21 pathway. Indeed, western blotting results showed that RNF26 overexpressing in T24 and UMUC3 cells reduced TRIM21 protein expression and promotes ZHX3 expression (Figure 5E). We also examined TRIM21 and ZHX3 expression following knockdown of RNF26 in the xenograft tumour model described above and found that TRIM21 expression increased and ZHX3 expression decreased following RNF26 knockdown (Figure 5F-H). Cycloheximide

(CHX) treatment significantly shortened the half-life of TRIM21 protein and increased the half-life of ZHX3 protein after overexpression of RNF26 in T24 cells (Figure 5I-K). Moreover, overexpression of RNF26 increased the level of polyubiquitination of TRIM21 in T24 cells (Figure 5L). In particular, the level of polyubiquitination was promoted by the addition of MG132, a well-known proteasome inhibitor. To explore potential lysine ubiquitination types, the most common form of K48 ubiquitination-mediated protein degradation, we subsequently transfected T24 cells with Ub-K48-only, Ub-K48R and Ub-KO mutants [31]. The results showed that wild-type (WT) Ub and Ub-K48-only could be linked to TRIM21 by RNF26, suggesting that RNF26 promoted K48-linked ubiquitination on TRIM21 (Figure 5M).

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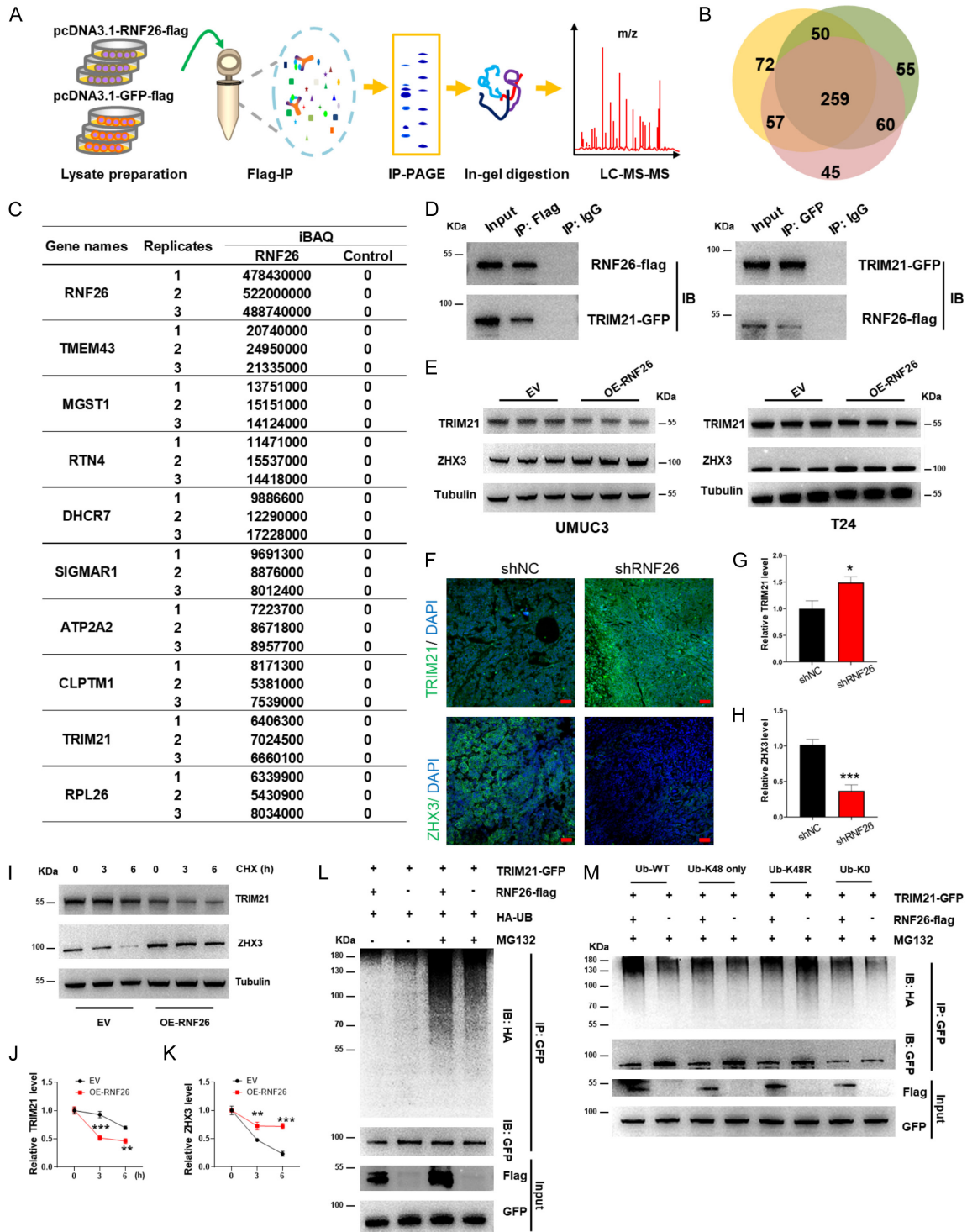


Figure 5. RNF26 interacts with TRIM21 in bladder cancer cells. **A.** The flowchart of immunoprecipitation-liquid chromatography tandem mass spectrometry (IP-LC-MS/MS) analyses in T24 cells. **B.** Venn diagram of three independent IP-LC-MS/MS assays of proteins. **C.** List of Top 10 proteins based on iBAQ quantification. **D.** The interaction between RNF26 and TRIM21 was confirmed by co-immunoprecipitation in T24 cells. **E.** RNF26 overexpressing in UMUC3 and T24 cells reduced TRIM21 protein expression and promotes ZHX3 expression. **F-H.** Immunostaining of xenograft tumor shows that TRIM21 expression was increased and ZHX3 expression was decreased with RNF26 knockdown. $n = 3$ for each group. Scale bar: 50 μm . **I-K.** Cycloheximide (CHX) treatment significantly shortened the half-life of TRIM21 protein and increased the half-life of ZHX3 protein after overexpression of RNF26 in T24 cells. **L.** Overexpression of RNF26 increased the level of polyubiquitination of TRIM21 in T24 cells. **M.** RNF26 promoted K48-linked ubiquitination on TRIM21 in T24 cells. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

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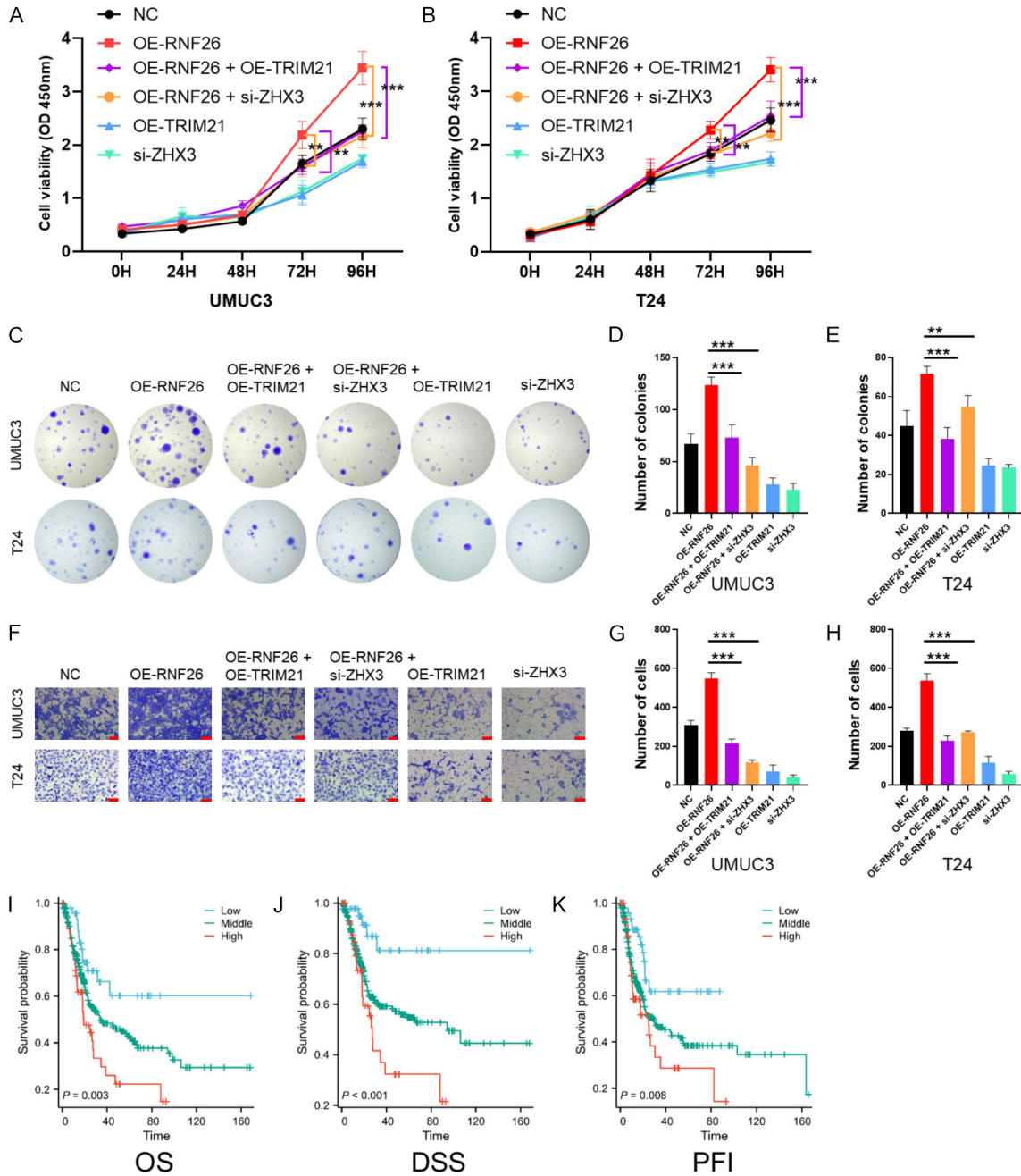


Figure 6. RNF26/TRIM21 induced bladder cancer cell proliferation and migration through ZHX3. TRIM21 overexpressing or ZHX3 silencing rescued the effects of RNF26 overexpressing on cell proliferation (A, B), migration (C-E), and colony formation (F-H) capacities of T24 and UMUC3 cells. (I-K) low risk group (low RNF26+high TRIM21+low ZHX3) were related to the good overall survival (OS) (I), disease specific survival(DSS) (J) and progress free interval (PFI) (K) in bladder cancer. Low risk group: low RNF26+high TRIM21+low ZHX3, high risk group: high RNF26+low TRIM21+high ZHX3, middle risk group: except for low RNF26+high TRIM21+low ZHX3 and high RNF26+low TRIM21+high ZHX3. Scale bar: 100 μ m. * $P < 0.05$, ** $P < 0.01$; *** $P < 0.001$.

RNF26/TRIM21 induced bladder cancer cell proliferation and migration through ZHX3

To further explore the functional association between RNF26 and TRIM21, rescue experi-

ments were performed and analyzed. The results revealed a decrease in T24 and UMUC3 cell proliferation (Figure 6A, 6B), migration (Figure 6C-E), and colony formation (Figure 6F-H) following the overexpression of TRIM21.

Simultaneously, TRIM21 overexpressing rescued the effects of RNF26 overexpressing on cell proliferation (**Figure 6A, 6B**), migration (**Figure 6C-E**), and colony formation (**Figure 6F-H**) capacities of T24 and UMUC3 cells. Moreover, TRIM21 silencing rescued the effects of RNF26 silencing on cell proliferation (**Figure S1A, S1B**), migration (**Figure S1C-E**), and colony formation (**Figure S1F-H**) capacities of T24 and UMUC3 cells. To explore whether ZHX3 is a target molecule of RNF26/TRIM21 for their oncogenic function, cell rescue assays were also performed. As expected, ZHX3 silencing decreased T24 and UMUC3 cell proliferation (**Figure 6A, 6B**), migration (**Figure 6C-E**), and colony formation (**Figure 6F-H**). Simultaneously, ZHX3 silencing reversed the effect of RNF26 overexpressing on cell proliferation (**Figure 6A, 6B**), migration (**Figure 6C-E**), and colony formation (**Figure 6F-H**) capacities of T24 and UMUC3 cells. Moreover, ZHX3 overexpressing reversed the effect of RNF26 silencing on cell proliferation (**Figure S1A, S1B**), migration (**Figure S1C-E**), and colony formation (**Figure S1F-H**) capacities of T24 and UMUC3 cells.

Finally, we also validated the clinical significance of RNF26/TRIM21/ZHX3 axis in bladder cancer patients. KM survival analysis showed that patients with low risk group (low RNF26+high TRIM21+low ZHX3) were related to the good overall survival (OS) (**Figure 6I**), disease free survival (DFS) (**Figure 6J**) and progress free interval (PFI) (**Figure 6K**) in bladder cancer. In contrast, patients with high risk (high RNF26+low TRIM21+high ZHX3) had the worst prognosis.

Discussion

Bladder cancer is one of the prevalent urological malignancies, and existing therapeutic strategies including immunotherapy and immune checkpoint inhibitors are effective but with limitations in the treatment of bladder cancer patients [32]. New molecular mechanisms remain to be explored to improve the therapeutic outcome of bladder cancer. It has been demonstrated that E3 ubiquitin ligases are closely related to bladder carcinogenesis, metastasis, drug resistance and immune escape [33, 34]. RNF26, a key E3 ubiquitin ligase, has been reported to promote bladder cancer progression by regulating the cell cycle [15]. However, its role and related mechanisms

in immune infiltration, immunotherapy, chemotherapeutic drug efficacy and tumor progression in bladder cancer remain to be elucidated. In this study, RNF26 expression was found to be closely associated with the efficacy of immunochemical drugs by biomimetic analysis, and Co-immunoprecipitation (Co-IP) assays and mass spectrometry (MS) analysis identified 259 potential RNF26 binding proteins. Among these, TRIM21 was an important binding protein of RNF26 and its ubiquitination was found mediated by RNF26 to affect its stability and to upregulate ZHX3, so as to influence bladder cancer progression.

Previous studies have demonstrated that RNF26 is upregulated in bladder cancer tissues and is strongly associated with poor prognosis [15]. Moreover, immune cell infiltration, such as macrophages, is strongly associated with tumor progression [35]. Corresponding to this, we have found in our study that low RNF26 expression displayed more immune cell infiltration, which may antagonize tumor cells, leading to better prognosis of bladder cancer. Meanwhile, immune checkpoint molecules were highly expressed in patients with high level RNF26, suggesting that such patients might be sensitive to immune checkpoint drugs. In addition, chemotherapy drug sensitivity analysis revealed that the RNF26 high expression group was more sensitive to cisplatin, VEGFR-targeted drugs and MET-targeted drugs. The fact that high level RNF26 expression (even with a poor prognosis), was sensitive to immune checkpoint drugs and multiple chemotherapeutic agents suggests that RNF26 may serve as a better predictive marker for the efficacy, and thus, better treatment strategy for bladder cancer.

To explore the effect of RNF26 in bladder cancer, functional assays were performed. A significant reduction in cell proliferation migration and colony-forming capacity was observed in the RNF26 knocking-down group compared to the control group. Meanwhile, overexpression of RNF26 in T24 and UMUC3 cells confirmed its oncogenic function. Xenograft assays further confirmed that knockdown of RNF26 resulted in reduced tumor growth. Previous studies have suggested that RNF26 binds to p57 and affects its stability through ubiquitination which is involved in the aggressiveness of bladder cancer [15]. Moreover, this study iden-

tified many other candidate proteins that may bind to RNF26 to influence its function. Indeed, 259 potential RNF26 binding proteins were identified by mass spectrometry analysis in this study. Interestingly, among the 259 identified proteins, TRIM21, an RNF26-binding protein with an E3 ubiquitin ligase active RING domain at its N-terminal end, could degrade different proteins by ubiquitination [36]. TRIM21 has been shown to have oncogenic effects on a variety of tumors through the ubiquitination pathway. For example, TRIM21 increases the ubiquitination and degradation of octamer-binding transcription factor 1 (Oct-1), which is self-renewed by cancer stem cells, resulting in stem cell reduction [37]. The cell cycle protein-dependent kinase 2 (CDK2) complex is hyperactivated in most cancers [38, 39], and TRIM21 can promote CDK2 autophagic degradation, thereby inhibiting leukemia progression [40]. All these suggest that RNF26 may promote bladder cancer progression by inhibiting TRIM21 function. Indeed, Co-IP experiments, Cycloheximide (CHX) treatment and ubiquitination assays, confirmed that RNF26 exerts an oncogenic function by binding to TRIM21 and promoting its ubiquitination. In addition, it has been reported that ZHX3 is an important target molecule of TRIM21 in bladder cancer and is closely associated with bladder cancer invasion, migration and poor prognosis. To clarify whether ZHX3 is the downstream target of RNF26/TRIM21. Rescue experiments were performed and the results showed that ZHX3 silencing reversed the effects of RNF26 over-expression on the proliferation, migration and colony-forming ability of bladder cancer cells. In addition, the RNF26/TRIM21/ZHX3 axis was also confirmed to be closely associated with poor prognosis of bladder cancer patients.

In summary, our study provides evidence that RNF26 is closely associated with poor prognosis, chemotherapeutic drug efficacy and immune checkpoint blockade therapy benefits of bladder cancer. RNF26 may bind to TRIM21 to regulate its protein stability and play an oncogenic role in bladder cancer through ZHX3. Together, these data support that RNF26 is a potential efficacy predictive marker and a therapeutic target for patients with bladder cancer.

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Disclosure of conflict of interest

None.

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Table S1. Putative interactors identified from three independent experiments in this study

Gene names	Unique peptides control_rep1	Unique peptides control_rep2	Unique peptides control_rep3	Unique peptides RNF26_rep1	Unique peptides RNF26_rep2	Unique peptides RNF26_rep3	iBAQ control_rep1	iBAQ control_rep2	iBAQ control_rep3	iBAQ RNF26_rep1	iBAQ RNF26_rep2	iBAQ RNF26_rep3
RNF26	0	0	0	21	18	18	0	0	0	478430000	522000000	488740000
TMEM43	0	0	0	17	22	19	0	0	0	20740000	24950000	21335000
MGST1	0	0	0	4	4	4	0	0	0	13751000	15151000	14124000
RTN4	0	0	0	6	9	7	0	0	0	11471000	15537000	14418000
DHCR7	0	0	0	4	5	8	0	0	0	9886600	12290000	17228000
SIGMAR1	0	0	0	3	3	3	0	0	0	9691300	8876000	8012400
ATP2A2	0	0	0	23	32	27	0	0	0	7223700	8671800	8957700
CLPTM1	0	0	0	14	13	15	0	0	0	8171300	5381000	7539000
TRIM21	0	0	0	15	18	14	0	0	0	6406300	7024500	6660100
RPL26	0	0	0	4	4	6	0	0	0	6339900	5430900	8034000
RPL27A	0	0	0	4	5	4	0	0	0	6133000	6617300	6483400
RPL23	0	0	0	3	2	4	0	0	0	4335700	6096900	8473600
LGALS3BP	0	0	0	10	13	12	0	0	0	4383000	5316700	4814500
RPS13	0	0	0	7	5	5	0	0	0	4894600	4284400	4334100
GNB2	0	0	0	3	4	3	0	0	0	2087800	3353000	6767900
EMC4	0	0	0	4	4	3	0	0	0	4331300	3104300	2971500
BCAP31	0	0	0	6	8	8	0	0	0	2853700	3213200	4171400
RER1	0	0	0	2	2	4	0	0	0	3145400	3120900	3800400
SPCS3	0	0	0	3	3	2	0	0	0	3904800	3205600	1872700
HACD3	0	0	0	5	9	7	0	0	0	2513400	3266800	3118600
DHCR24	0	0	0	7	8	8	0	0	0	2966600	2769000	3036800
CAV2	0	0	0	3	3	2	0	0	0	2025300	3486600	3153400
SLC25A11	0	0	0	7	8	7	0	0	0	3266000	2766800	2608200
RPL34	0	0	0	3	3	4	0	0	0	1718100	1730400	4647500
CPT1A	0	0	0	13	11	12	0	0	0	2729900	2231400	2830800
TMED10	0	0	0	3	3	3	0	0	0	2559000	2436800	2785400
EBP	0	0	0	2	2	2	0	0	0	2534700	2183000	2878400
RPL5	0	0	0	5	6	4	0	0	0	2579100	2644000	2247200
RPL35A	0	0	0	4	2	3	0	0	0	2976100	1636500	2720700
COPA	0	0	0	23	29	26	0	0	0	2224500	2498200	2565900
SEC62	0	0	0	2	2	3	0	0	0	1294700	2506300	3475800
UQCRI10	0	0	0	2	2	2	0	0	0	2022600	2573000	2203100
ATP5F1	0	0	0	4	4	7	0	0	0	1662100	2161700	2914000
OPA1	0	0	0	23	24	18	0	0	0	2343100	2413600	1966300
ERLIN2	0	0	0	4	5	5	0	0	0	2049800	1940300	2618200
MYO1C	0	0	0	14	19	19	0	0	0	1528900	2369400	2493200

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RPL10A	0	0	0	4	5	4	0	0	0	1753400	2379900	2233100
CRMP1	0	0	0	5	4	3	0	0	0	418070	2807800	3119400
FITM2	0	0	0	3	3	3	0	0	0	2126400	1875500	2070300
SNX2	0	0	0	10	11	10	0	0	0	1598700	2103600	1896500
TAP1	0	0	0	9	8	9	0	0	0	1991800	1562000	1957000
PFN2	0	0	0	3	2	3	0	0	0	2497000	887140	2117800
CANX	0	0	0	7	10	10	0	0	0	1382200	1964400	2047700
RPS18	0	0	0	4	6	4	0	0	0	1407600	2685200	754050
SLC25A1	0	0	0	5	3	5	0	0	0	1877200	956210	1770900
NUP205	0	0	0	34	30	33	0	0	0	1522300	1537700	1493200
PRKDC	0	0	0	50	58	61	0	0	0	1220800	1515600	1703200
EMC3	0	0	0	2	5	4	0	0	0	851920	1941400	1619300
THEM6	0	0	0	4	6	5	0	0	0	1214900	1602500	1495100
CYFIP1	0	0	0	12	14	16	0	0	0	1405200	1449600	1404400
TAP2	0	0	0	7	5	6	0	0	0	1735700	953240	1534100
CLN6	0	0	0	2	2	2	0	0	0	1520700	1366900	1331200
LBR	0	0	0	5	4	4	0	0	0	1437200	1284900	1476600
FDPS	0	0	0	5	5	5	0	0	0	1047600	1413100	1732300
ATL3	0	0	0	4	6	7	0	0	0	1080000	1354000	1750600
WARS	0	0	0	8	10	8	0	0	0	1372600	1444400	1195600
VAR5	0	0	0	18	15	18	0	0	0	1397100	1194500	1359000
C14orf1	0	0	0	2	3	2	0	0	0	1305200	1127300	1371700
DHX9	0	0	0	16	15	14	0	0	0	1258000	1250500	1255500
PLIN3	0	0	0	5	7	6	0	0	0	1117100	1319700	1244800
CTPS1	0	0	0	10	9	9	0	0	0	1032800	1259400	1251900
TMEM159	0	0	0	2	2	2	0	0	0	1196600	1018600	1091300
XPO1	0	0	0	9	13	14	0	0	0	832640	1097400	1230400
ACOT9	0	0	0	5	6	5	0	0	0	1199800	1037000	885010
JAGN1	0	0	0	2	2	2	0	0	0	1047900	1016200	1052500
LPCAT1	0	0	0	6	5	4	0	0	0	1259000	1064500	762700
HSD17B12	0	0	0	2	5	4	0	0	0	1150900	905200	1021900
PRPF19	0	0	0	4	4	4	0	0	0	858810	1082200	1132200
EIF3E	0	0	0	7	6	8	0	0	0	1095500	1154700	801100
EMC2	0	0	0	3	3	3	0	0	0	1037700	1068200	939180
SGPL1	0	0	0	5	5	4	0	0	0	993770	1071500	941570
TRIP13	0	0	0	8	7	8	0	0	0	910340	858530	1025400
NCLN	0	0	0	8	6	8	0	0	0	1109400	611550	1071400
EMD	0	0	0	4	3	3	0	0	0	1010300	863970	883520
TOMM22	0	0	0	2	2	3	0	0	0	507430	780060	1469000
HSDL1	0	0	0	2	3	4	0	0	0	852110	858130	1037800

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COPE	0	0	0	3	5	5	0	0	0	553640	944030	1171000
SF3B1	0	0	0	10	13	12	0	0	0	597790	1037300	1008400
PDCD6IP	0	0	0	12	13	17	0	0	0	708450	756450	1149200
ACTR2	0	0	0	4	2	3	0	0	0	840660	869490	897760
ACTR3	0	0	0	4	4	3	0	0	0	997810	884970	719700
HNRNPA0	0	0	0	3	4	5	0	0	0	747550	855420	995730
MBOAT7	0	0	0	3	4	3	0	0	0	817470	908240	837080
PSMA7	0	0	0	2	3	3	0	0	0	587600	938510	1020200
AP2M1	0	0	0	5	7	4	0	0	0	942230	1102400	495020
DYNC1H1	0	0	0	52	54	51	0	0	0	815640	859830	831370
NDUFA9	0	0	0	5	9	7	0	0	0	575810	1019300	876790
RAE1	0	0	0	3	4	2	0	0	0	693470	1031700	687790
TEX10	0	0	0	9	9	8	0	0	0	902340	805340	654810
OAS3	0	0	0	11	10	12	0	0	0	812430	697720	843100
SEC22B	0	0	0	5	3	3	0	0	0	1050200	546840	685860
WDR1	0	0	0	5	5	4	0	0	0	764090	800450	710550
IQGAP1	0	0	0	19	21	19	0	0	0	707010	757530	777310
EIF2S2	0	0	0	3	4	5	0	0	0	644130	722550	838920
SMPD4	0	0	0	5	6	5	0	0	0	883280	774160	546570
NAMPT	0	0	0	3	8	7	0	0	0	281500	1002100	875770
LMAN1	0	0	0	3	3	3	0	0	0	474110	869760	782230
PSMD6	0	0	0	7	7	6	0	0	0	860590	621990	620830
CYP51A1	0	0	0	5	8	6	0	0	0	595900	928740	550390
DNM1L	0	0	0	9	10	7	0	0	0	616000	825320	612340
VDAC1	0	0	0	4	5	4	0	0	0	618470	659710	760390
Septin-7	0	0	0	5	5	5	0	0	0	692230	640610	677020
BLVRA	0	0	0	3	4	4	0	0	0	569980	621380	774360
MRPS22	0	0	0	3	6	7	0	0	0	371550	683450	910650
FBL	0	0	0	2	2	3	0	0	0	373160	881420	709430
RBM14	0	0	0	4	6	6	0	0	0	611220	703000	640480
EEF1E1	0	0	0	2	2	2	0	0	0	611760	616420	724870
TUBG1	0	0	0	2	3	2	0	0	0	429760	1034000	470130
PDHB	0	0	0	3	5	2	0	0	0	935860	733440	251240
AIMP2	0	0	0	2	4	4	0	0	0	451080	710360	739450
PIGT	0	0	0	6	5	3	0	0	0	652620	708640	524850
TMEM194A	0	0	0	3	3	3	0	0	0	463980	720970	694390
NOP56	0	0	0	7	7	6	0	0	0	570160	701910	581110
DLD	0	0	0	4	5	6	0	0	0	638280	615690	595430
PEX11B	0	0	0	4	4	3	0	0	0	501940	655090	688970
EIF3B	0	0	0	7	6	10	0	0	0	571680	548370	695080

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ADSL	0	0	0	4	2	3	0	0	0	693210	476700	645180
GTPBP4	0	0	0	7	6	5	0	0	0	765830	621990	406700
Septin-2	0	0	0	3	5	4	0	0	0	412350	786150	590010
SF3B6	0	0	0	2	3	3	0	0	0	224820	832720	726030
EIF3A	0	0	0	9	7	9	0	0	0	616940	527760	613210
PIGK	0	0	0	4	3	4	0	0	0	741000	412740	592530
ARCN1	0	0	0	7	8	4	0	0	0	689760	589260	449010
P4HA1	0	0	0	5	5	5	0	0	0	615940	522520	573990
HSD17B11	0	0	0	3	3	2	0	0	0	572330	614420	524310
MYO1B	0	0	0	8	11	11	0	0	0	461220	640090	579850
VPS26A	0	0	0	3	2	2	0	0	0	705550	457670	513190
ACTR1A	0	0	0	2	4	4	0	0	0	219470	743520	704980
PSMC2	0	0	0	6	7	5	0	0	0	541160	595430	528210
PTRF	0	0	0	2	2	2	0	0	0	456120	572100	594380
ARPC2	0	0	0	4	4	4	0	0	0	549710	550970	519960
NUP93	0	0	0	7	8	9	0	0	0	438270	467010	709900
MAT2B	0	0	0	2	2	2	0	0	0	514970	544690	504020
UBAC2	0	0	0	3	2	4	0	0	0	508130	399460	636060
SYVN1	0	0	0	2	2	2	0	0	0	523610	571520	446200
ABCD3	0	0	0	6	8	5	0	0	0	538380	617610	381980
VKORC1L1	0	0	0	2	2	3	0	0	0	422830	456040	640010
MTA2	0	0	0	7	7	6	0	0	0	508650	486910	520700
PES1	0	0	0	6	5	5	0	0	0	411070	527350	514970
EIF3I	0	0	0	4	3	3	0	0	0	512160	458320	451010
ATIC	0	0	0	6	8	5	0	0	0	391840	617070	403100
ATP13A1	0	0	0	10	14	10	0	0	0	371120	552310	482400
QARS	0	0	0	9	10	9	0	0	0	543900	477290	382590
MRPS34	0	0	0	2	2	3	0	0	0	383010	431100	572490
SRPRB	0	0	0	5	3	4	0	0	0	540490	387680	439590
ALG1	0	0	0	3	3	2	0	0	0	465700	453720	444450
CLTC	0	0	0	13	18	14	0	0	0	413940	533670	412010
AAAS	0	0	0	4	3	3	0	0	0	494210	486020	375370
TMX1	0	0	0	2	2	2	0	0	0	458090	419880	468230
VAPB	0	0	0	2	4	3	0	0	0	452280	456380	421890
TARDBP	0	0	0	2	3	2	0	0	0	447900	557900	320170
TFRC	0	0	0	7	10	5	0	0	0	366190	603550	355420
GART	0	0	0	3	6	3	0	0	0	363210	586740	368870
DDX58	0	0	0	9	12	8	0	0	0	416120	479030	421120
SRP68	0	0	0	5	5	5	0	0	0	386510	419410	473500
EPRS	0	0	0	13	8	14	0	0	0	469100	310210	498910

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PGM1	0	0	0	6	7	3	0	0	0	498120	519760	252290
COPB1	0	0	0	5	9	8	0	0	0	305290	511490	437610
ACP1	0	0	0	2	2	2	0	0	0	427600	432680	392630
OASL	0	0	0	3	5	5	0	0	0	346240	391940	511650
PAFAH1B1	0	0	0	3	2	3	0	0	0	397690	376490	469260
NUP54	0	0	0	4	5	4	0	0	0	262490	501330	464080
NCKAP1	0	0	0	9	7	8	0	0	0	387420	332640	481930
MCM7	0	0	0	7	7	6	0	0	0	379750	429170	385620
MMTAG2	0	0	0	2	2	2	0	0	0	324980	423130	441290
HM13	0	0	0	3	2	2	0	0	0	383500	242850	537010
TUBGCP3	0	0	0	5	6	7	0	0	0	214920	543860	346260
NOP58	0	0	0	3	2	3	0	0	0	409430	276120	418370
FAM120A	0	0	0	7	9	6	0	0	0	357140	426510	289410
EIF3G	0	0	0	3	2	4	0	0	0	480560	243510	346340
NHP2L1	0	0	0	2	2	2	0	0	0	212840	431760	420780
ZMPSTE24	0	0	0	2	3	2	0	0	0	163500	435780	456140
EIF3C	0	0	0	4	6	5	0	0	0	342130	357380	330580
NOC2L	0	0	0	5	4	3	0	0	0	470430	343830	208490
DDX47	0	0	0	2	4	4	0	0	0	202600	366920	452680
OXA1L	0	0	0	2	2	2	0	0	0	253860	314760	409340
PON2	0	0	0	2	2	3	0	0	0	226890	208530	542430
PHGDH	0	0	0	3	3	2	0	0	0	256680	393580	316250
TUBB2A	0	0	0	2	2	2	0	0	0	361580	245290	357160
PSMD12	0	0	0	3	4	2	0	0	0	335960	388940	231720
HP1BP3	0	0	0	4	3	4	0	0	0	362610	229480	354070
NCAPD2	0	0	0	9	11	10	0	0	0	296120	291550	340880
PICALM	0	0	0	3	4	2	0	0	0	378700	387900	158960
XPO7	0	0	0	7	8	8	0	0	0	273340	314840	318770
FAR1	0	0	0	3	3	3	0	0	0	280560	330050	292070
ALDH18A1	0	0	0	6	6	5	0	0	0	298960	335370	266750
KRT18	0	0	0	2	3	2	0	0	0	334490	187190	372670
UBE3C	0	0	0	6	6	8	0	0	0	313180	269080	310760
ACSL3	0	0	0	4	3	4	0	0	0	323920	213480	355250
EMC1	0	0	0	3	5	5	0	0	0	229620	348460	313790
TNPO3	0	0	0	6	2	3	0	0	0	427880	198850	264720
PTPN1	0	0	0	3	4	3	0	0	0	176050	376890	337420
EIF2AK2	0	0	0	2	3	4	0	0	0	133570	258550	480940
STAT3	0	0	0	6	4	5	0	0	0	305600	262770	302200
TMCC3	0	0	0	3	4	3	0	0	0	250410	326220	289010
MX2	0	0	0	3	5	5	0	0	0	238910	260910	332880

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AGK	0	0	0	2	2	2	0	0	0	309770	315010	202350
GNAS	0	0	0	2	2	2	0	0	0	270180	290250	266290
PRPF6	0	0	0	6	8	6	0	0	0	254540	298500	269930
RAB32	0	0	0	2	2	2	0	0	0	261540	294150	264180
ABHD16A	0	0	0	4	6	3	0	0	0	274240	340920	195400
SUN1	0	0	0	4	6	6	0	0	0	220590	262140	313130
POR	0	0	0	5	4	5	0	0	0	294360	237420	255110
POM121C	0	0	0	5	3	5	0	0	0	274120	146410	353650
PDLIM7	0	0	0	3	3	4	0	0	0	204870	211610	354360
TMX2	0	0	0	2	2	2	0	0	0	245750	257090	241100
TBL2	0	0	0	3	4	5	0	0	0	261440	177920	281600
GOT1	0	0	0	3	2	2	0	0	0	323500	156620	216090
NCEH1	0	0	0	2	3	3	0	0	0	181980	236430	268060
SEC23A	0	0	0	2	2	6	0	0	0	158580	150070	371560
GPS1	0	0	0	3	2	2	0	0	0	193770	252240	230870
AP2A1	0	0	0	5	6	7	0	0	0	202970	187930	267690
G6PD	0	0	0	3	2	2	0	0	0	200970	141450	296550
NAE1	0	0	0	2	3	2	0	0	0	158510	314320	163630
TUBGCP2	0	0	0	4	3	4	0	0	0	215700	174910	235770
IPO5	0	0	0	6	5	5	0	0	0	296340	121440	180280
KLHL12	0	0	0	3	2	3	0	0	0	155300	221270	221020
RNH1	0	0	0	4	3	3	0	0	0	256640	125780	214270
ERO1L	0	0	0	2	2	3	0	0	0	234220	142110	210360
ACAD9	0	0	0	4	2	3	0	0	0	215900	113810	252730
MARS	0	0	0	2	3	5	0	0	0	85068	163580	331810
SEC63	0	0	0	4	3	3	0	0	0	208380	197860	171230
OAS2	0	0	0	3	2	2	0	0	0	288190	238350	42586
RECQL	0	0	0	3	3	3	0	0	0	204550	172470	168740
TMEM214	0	0	0	2	4	3	0	0	0	137120	229290	170990
EIF3D	0	0	0	2	2	2	0	0	0	216290	105960	195540
LRRC40	0	0	0	2	4	2	0	0	0	114830	262960	133830
KIAA0196	0	0	0	6	7	6	0	0	0	218670	159230	133500
TOR4A	0	0	0	2	4	2	0	0	0	122500	259400	122390
DDX19A	0	0	0	2	3	3	0	0	0	147800	180840	163790
ANXA6	0	0	0	3	3	3	0	0	0	168910	134360	177230
IGF2BP2	0	0	0	2	3	3	0	0	0	111140	105940	260830
ZW10	0	0	0	3	4	3	0	0	0	139310	182190	156230
ANXA11	0	0	0	2	2	2	0	0	0	131180	110210	231290
AP1G1	0	0	0	2	2	4	0	0	0	139610	165220	131100
PAK2	0	0	0	2	3	3	0	0	0	67339	138540	212320

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DYNC1LI1	0	0	0	2	2	2	0	0	0	143770	141360	131090
SEC24C	0	0	0	2	3	3	0	0	0	125650	125190	157430
TELO2	0	0	0	4	3	4	0	0	0	160350	110840	120170
MYO1E	0	0	0	3	4	5	0	0	0	106610	122340	150110
PCID2	0	0	0	2	3	3	0	0	0	108770	100790	164250
USO1	0	0	0	2	5	3	0	0	0	72602	204980	80935
TBCD	0	0	0	2	6	5	0	0	0	33511	139460	170090
WFS1	0	0	0	2	2	2	0	0	0	124930	94586	118210
NPEPPS	0	0	0	2	3	2	0	0	0	84219	128780	94134
EXOC4	0	0	0	2	3	3	0	0	0	88863	96864	113410
DCTN1	0	0	0	2	2	2	0	0	0	102510	61673	97020
SAMD9	0	0	0	3	4	4	0	0	0	66456	88167	96438
PSMD5	0	0	0	2	2	2	0	0	0	55123	79879	107540
FLNA	0	0	0	4	6	5	0	0	0	59594	79548	96120
PLEC	0	0	0	9	7	11	0	0	0	70849	42336	114330
XPO6	0	0	0	2	3	2	0	0	0	77047	107670	40856
RARS2	0	0	0	2	2	2	0	0	0	111510	42462	55471
KIF5B	0	0	0	2	3	3	0	0	0	49604	68717	76528
VPS51	0	0	0	2	2	2	0	0	0	103040	46976	42454
KIAA1524	0	0	0	2	3	2	0	0	0	54855	74574	53827
KIAA1033	0	0	0	2	3	2	0	0	0	36330	86521	21366
GCN1L1	0	0	0	3	2	5	0	0	0	35162	21426	62894
RANBP2	0	0	0	2	4	3	0	0	0	17896	36344	26198

RNF26 promotes bladder cancer progression

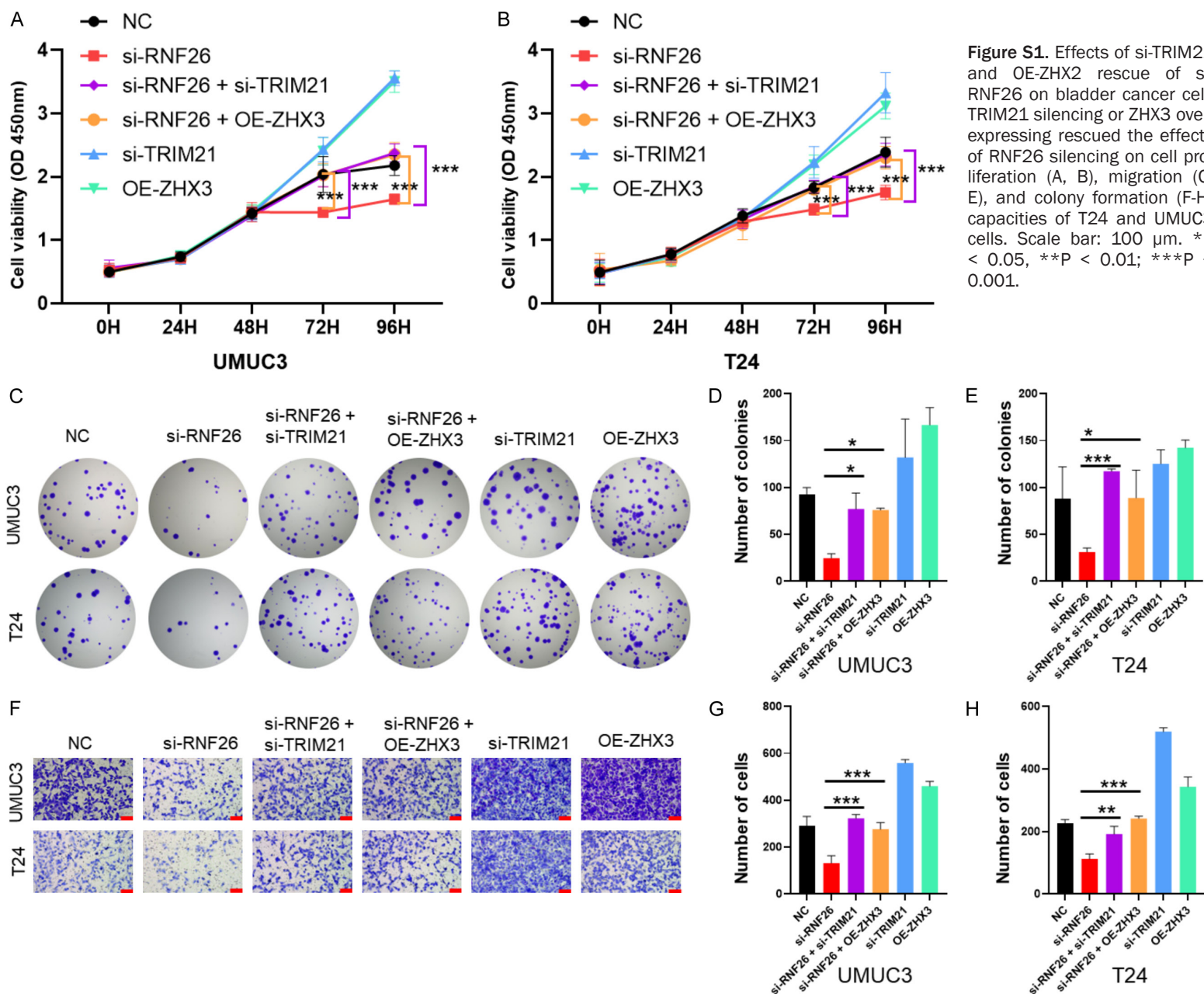


Figure S1. Effects of si-TRIM21 and OE-ZHX2 rescue of si-RNF26 on bladder cancer cell. TRIM21 silencing or ZHX3 over-expressing rescued the effects of RNF26 silencing on cell proliferation (A, B), migration (C-E), and colony formation (F-H) capacities of T24 and UMC3 cells. Scale bar: 100 μ m. *P < 0.05, **P < 0.01; ***P < 0.001.