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Dose Estimation Utility in a Population Pharmacokinetic Analysis of Inhaled Δ 9-Tetrahydrocannabinol Cannabis Market Products in Occasional and Daily Users

Thomas K. Henthorn, MD,*† George S. Wang, MD,‡ Greg Dooley, PhD,§ Ashley Brooks-Russell, MD, MPH,¶ Julia Wrobel, PhD,|| Sarah Limbacher, MPH,¶ and Michael Kosnett, MD**

Background: Unusually high variability in blood Δ 9tetrahydrocannabinol (THC) concentrations have been observed in subjects inhaling similar cannabis products over similar time periods when consumption is ad libitum. This makes simple gravimetric dose estimation a poor predictor of THC exposure. Population pharmacokinetic analyses of blood THC concentration versus time data are routinely used to estimate pharmacokinetic parameters. The aim of this study was to estimate the inhaled dose of THC in occasional and daily users of high potency market cannabis.

Methods: Blood THC concentrations were measured for 135 minutes from 29 participants who either smoked high concentration flower or inhaled concentrates ad libitum during a 15-minute session. Frequent blood samples were obtained over the following 135 minutes.

Results: The estimated central and rapidly equilibrating volumes of distribution of a 3-compartment model were 19.9 \pm 1.2 and 51.6 \pm 4.7 L whereas the intercompartmental clearances were 1.65 \pm 0.14 and 1.75 \pm 0.10 L/min, respectively. Covariate-

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- Correspondence: Thomas K. Henthorn, MD, Department of Anesthesiology and Department of Pharmaceutical Sciences, University of Colorado Anschutz Medical Campus, Aurora, CO 80045 (e-mail: thomas. henthorn@cuanschutz.edu).
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adjusted analysis revealed that the estimated inhaled THC dose was considerably less among occasional users compared with daily users.

Conclusions: Three-compartment pharmacokinetics of THC did not differ among the 3 user groups, and the early phase (first 135 minutes postinception of inhalation) kinetics were similar to those previously described after smoking low potency cannabis products. Therefore, inhaled THC dose can be estimated from pharmacokinetic data and covariate-driven adjustments can be used to estimate THC doses, based on the participant cannabis usage pattern (occasional versus daily), improving the accuracy of THC exposure estimates compared with those derived from weighed THC content alone.

Key Words: CB1 agonist, THC, cannabis, population pharmacokinetics, Δ9-tetrahydrocannabinol

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INTRODUCTION

The use of cannabis among adults in the United States continues to rise,^{1,2} with more states legalizing its use for recreational and medical purposes.³ Despite this widespread use, the pharmacokinetics (PK) of the main psychoactive compound of cannabis, Δ 9-tetrahydrocannabinol (THC), remains incomplete. Owing to federal restrictions on the source of cannabis products available to research institutions, most previous clinical trials used low-concentration cannabis made available from the National Institute on Drug Abuse (NIDA) Drug Supply Program and often adhered to paced smoking regimens intended to minimize variability in participants' delivered THC dose.⁴ Such uniformity was necessary to apply descriptive pharmacokinetic analyses (eg, Cmax, Tmax, Area-Under-the-Curve, and terminal half-life). However, the cannabis supplied by NIDA for these studies differed considerably, in cultivar and THC content, from that being sold in the current retail and medical marketplace.^{1,5,6}

The investigation of THC pharmacokinetics in studies using NIDA-supplied cannabis has been constrained by the Drug Enforcement Agency (DEA) requirements that the participants consume identical combusted or vaporized cannabis products over a relatively uniform time interval.^{4,7–9} Most cannabis consumers self-administer, ad libitum, in a

From the *Departments of Anesthesiology and †Pharmaceutical Sciences, University of Colorado Anschutz Medical Campus; ‡Department of Pediatrics, University of Colorado Anschutz Medical Campus, Children's Hospital Colorado, Aurora; §Department of Environmental and Radiological Health Sciences, Colorado State University, Fort Collins; ¶Injury and Violence Prevention Center, University of Colorado Anschutz Medical Campus, Aurora, Colorado; ||Department of Biostatistics and Bioinformatics, Rollins School of Public Health, Emory University, Atlanta, Georgia; and **Department of Environmental and Occupational Health, University of Colorado Anschutz Medical Campus, Aurora, Colorado.

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naturalistic manner. Cannabis plant material and concentrates may differ not only in THC content but also in moisture content, terpene profile, legal and illicit excipients, and residues (such as solvents, pesticides, or herbicides) that may influence the perceived quality (eg, harshness or irritancy) of the inhaled smoke or vapor. The temperature delivered by different pipes, vaping devices, and smoking techniques may also affect volatilization of cannabinoids and other constituents,7 which may contribute to relegating the starting point of most pharmacokinetic analyses, that is, the drug dose administered to unknowable variability. Yet, in a recent Request for Applications aimed at defining the pharmacokinetics and pharmacodynamics (PK/PD) of cannabinoids, NIDA stipulated that "results should be reported using the Standard THC Unit (STU), defined as 5 milligrams of THC (5 mg THC)"¹⁰ without first defining the imprecision and utility of undertaking the study of THC PK/PD with an assumed dose.

Assessing systemic exposure to THC is recognized as a key component to research examining the public health implications of cannabis consumption.¹¹ Cannabis legalization has increased the diversity of delivery methods and potency of products available which affects public health outcomes,¹² yet characterizing exposure with external techniques (ie, gravimetric and frequency) is difficult.¹³

Because of the diversity of cannabis delivery methods, observed user variability of inhalation efficiency, and titrated effects,^{9,14} a more accurate methodology for estimating THC dose is needed. It has been observed that distribution pharmacokinetics for a given drug show little interindividual variability in healthy adults because these processes are determined by physiological factors, such as tissue mass and blood flow,¹⁵ and physicochemical factors such as tissue:blood partitioning and protein binding.¹⁶ These principles allow for the construction of physiologically based pharmacokinetic models¹⁷ and form the basis of target-controlled infusions of anesthetic drugs¹⁸ in which plasma drug concentrations are held stationary by continuously adjusting the drug infusion rate to account for ongoing drug distribution. Therefore, it should be feasible to combine blood/plasma THC concentration data with a pharmacokinetic analysis to estimate the inhaled dose of THC.

To better understand real-world THC pharmacokinetics, a study should ideally allow research participants to bring their preferred cannabis product to a clinical laboratory and consume it in their usual manner. This study evaluated the pharmacokinetics of THC in occasional and daily cannabis users who smoked or vaped their preferred, commercially available cannabis product according to their usual pattern of consumption. It was hypothesized that these more denser, precisely timed blood THC concentrations versus time data would demonstrate, through a population pharmacokinetic analysis, small interindividual variability of parameters characterizing drug early distribution, and that the inhaled dose could be estimated and would show larger interindividual variability.

MATERIALS AND METHODS

Participants

We recruited 30 healthy adults (aged 21–55 years). Key inclusion criteria included cannabis use on a daily or weekly basis and willingness to consume a minimum of at least 2 inhalations of cannabis flower or concentrate products (eg, by dabbing or vaporizing wax or oil) containing less than 2% CBD. Participants were recruited into 3 groups: (1) occasional use characterized by an average of at least 2 days per month and no more than 3 days per week in the 90 days before enrollment, with typical use of flower products, (2) daily use with typical use of flower products, and (3) daily use with typical use of concentrate products (eg, vape pens or "dabs").

Participants were asked to refrain smoking or vaping inhaled cannabis products for at least 8 hours before the data collection study visit and ingesting edible cannabis products for at least 12 hours before the data collection study visit. This study was approved by the Colorado Multiple Institutional Review Board (Protocol 20-0949).

Cannabis Use Procedures

Participants were asked to supply a product purchased legally in Colorado, which require state laboratoryverified labeling of THC concentration, with total THC (THC and tetrahydrocannabinolic acid) concentrations ranging from 15% to 30% for flower products and 60%–90% for extract/concentrate products. The product package details were recorded. Participants were instructed to weigh the initial and residual cannabis containing product placed or contained within the smoking or vaping device before and after smoking or vaping using a digital scale provided by the study staff, and the weight before and after use was recorded.

During a 15-minute interval, participants in the user groups were instructed to smoke or vape ad libitum "the amount you most commonly use for the effect you most commonly desire." Start time was recorded as the inception of the first inhalation. The number of inhalations was counted, and the end time was noted as the last inhalation, which was no more than 15 minutes after the start time. Participants informed the research team which inhalation was intended to be their final inhalation.

Blood Collection

An upper arm vein was cannulated with an 18-gauge catheter for withdrawal of blood samples. Whole blood samples of approximately 5 mL were collected into tubes containing 100 mg sodium fluoride and 20 mg potassium oxalate additive at the following time points: baseline (minutes before the study's first inhalation); approximately 1, 5, and 10 minutes after the final inhalation; and then 30, 60, 90, and 145 minutes after the inception of the first inhalation. The actual midpoint of the blood withdrawal time relative to either the first or final inhalation of cannabis was recorded for pharmacokinetic analyses.

Sample Analysis

Reagents and Supplies

All cannabinoid standards and internal standards were purchased from Cerilliant (Round Rock, TX). Captiva EMR-Lipid columns (1 mL, 40 mg) were purchased from Agilent Technologies (Santa Clara, CA). Chromatography was performed with a 3.0×50 mm, 2.7 µm raptor biphenyl column (Restek, Inc., Bellefonte, PA).

Calibrators, Quality Controls, and Internal Standard Preparation

Matrix-matched calibrators and controls were prepared by the addition of appropriate volumes of methanolic stock standards mixes (0.01, 0.1, or 1.0 mcg/mL, or 10 mcg/mL of THC) to 100 μ L of cannabinoid free blood. These were used to produce calibrators for THC at 0.5, 1, 5, 10, 50, 100, and 500 ng/mL. Quality controls samples were prepared at 5 and 10 ng/mL for each analyte. Quality control samples were run after every 20 subject samples with an expected accuracy of $\pm 20\%$.

Cannabinoids Analysis by LC-MS/MS

Blood samples and matrix-matched standards and quality controls were prepared for LC-MS/MS analysis by protein precipitation and lipid removal. 10 μ L of the internal standard solution was added to 100 μ L of blood sample and mixed in a microcentrifuge tube, and then 600 μ L of acetonitrile/methanol (85%/15%) was added and vortexed for 30 seconds to precipitate proteins. Samples were centrifuged and supernatants transferred to Captiva EMR-Lipid columns for lipid removal. Eluents were collected into a clean glass test tube and dried under nitrogen at 45°C. Eluents were reconstituted in 75 μ L of water/methanol (50%/50%) with 0.1% acetic acid and transferred to autosampler vials for LC-MS/MS analysis.

Samples were analyzed with an Agilent 1290 UHPLC coupled to an Agilent 6460 triple quadruple mass spectrometer equipped with an Agilent Jet Stream electrospray ionization source (Agilent, Santa Clara, CA). Quantitation was performed using linear regression with 7-point calibration curves from 0.5 ng/mL to 500 ng/mL. Multiple cannabinoids were quantitated; however, only the values for THC are presented herein.

Each participant's baseline blood THC concentration was subtracted from the measured THC concentrations before pharmacokinetic analysis.

Pharmacokinetic Analyses

The pharmacokinetic analysis was performed using Phoenix NLME 8.3.4 with the FOCE ELS algorithm (Certara, Princeton, NJ). A 3-compartment pharmacokinetic model was used in keeping with models for rapid intravenous infusion and inhalation of lipid soluble, central nervous system–acting drugs such as ketamine^{19–21} and THC^{9,22} and in which the elimination clearance and slow equilibrating volume of distribution were fixed to previous estimates²² because no terminal phase pharmacokinetic data were collected in this study of 2.5 hours duration. In addition, we performed a 2-compartment analysis without linking to terminal phase pharmacokinetic estimates. The inhalation dose was arbitrarily assumed, for modeling purposes, to consist of a 60 mg THC joint smoked with 25% efficiency or bioavailability (ie, 15 mg fully bioavailable by inhalation). An individual's ad libitum inhaled dose was, therefore, estimated as a multiple of this assumed dose, $F_{inhaled}$. For instance, $F_{inhaled}$ estimates of 0.5, 1.0, and 1.5 would correspond to estimated inhaled doses of 7.5 mg, 15 mg, and 22.5 mg, respectively. Variability of $F_{inhaled}$ may reflect deviations from the measured weight of the cannabis product consumed, the actual inhalation efficiency, or both.

Model parameters (θ_j , theta, for model parameter j) were assumed to be log-normally distributed across the population with a central, typical value (θ_{TV}) allowing for assessment of between subject variability ($\eta_{j,I}$, eta, for individual i and model parameter j) such that:

$$\theta_{\rm i} = \theta_{\rm TV} \times e^{\eta \rm j} \tag{1}$$

The residual within-subject error was calculated using the relative error method. Models were selected based on inspection of the data fits, standard errors of the parameter decrease estimates (SEE) а in the objective function, $-2 \cdot \text{Loglikelihood}$ (-2LL), by 6.63 for addition of a model parameter to be statistically significant (χ^2 , $P \leq 0.01$). THC concentration data were fit to a 3compartment model. To accomplish this and to compare the current results to previous pharmacokinetic analyses, which were performed with plasma THC concentration measurements, the THC blood:plasma concentration ratio of 0.63 was applied to the measured blood THC concentrations.²³ Values below the lower limit of quantitation (ie, <0.5ng/mL) were pharmacokinetically analyzed according to the M3 method of Beal.²⁴

Potential model covariates were sought through a stepwise search in which potential covariates were added when the $-2 \cdot \text{Loglikelihood}$ decrease was at the P < 0.05 significance level (χ^2 -test; 3.84) and removal did not result in an increase of the $-2 \cdot \text{Loglikelihood}$ at the P < 0.01 significance level (χ^2 -test; 6.63). Potential covariate effects of participant demographics that were continuous variables (ie, age and body mass index) were added to the base model in the following manner:

$$\theta_{\rm i} = \theta_{\rm TV} \times \left({\rm Var}_{\rm demographic} \right)^{\theta} {\rm cov} \times e^{\eta} {\rm i} \tag{2}$$

where θ_{TV} is the typical value for the parameter, $Var_{demographic}$ is the continuous demographic variable, θ_{cov} is the parameter which quantifies the covariate effect, and η_i is a random variable describing the variance between the individual (θ_i) and population mean (θ_{TV}). Categorical variables (ie, sex and category of usual use pattern, such as occasional flower, daily concentrate, and daily flower) were added in the following manner:

$$\theta_{\rm i} = \theta_{\rm TV} \times e^{\theta} {\rm cov} \times e^{\eta} {\rm i} \tag{3}$$

In addition, a modeled analysis was conducted, in which the pattern of use was dichotomized (daily use versus occasional use) without regard to whether flower or concentrate was consumed in the dosing protocol.

The "weighed cannabis inhalation dose" (estimated by multiplying the product's labeled THC concentration times the measured weight difference preinhalation to postinhalation) was treated as a continuous covariate:

$$\theta_{\rm TV} \times ({\rm THC}_{\rm wt}/15 \text{ mg})^{\theta} {\rm cov}_{\rm weighed_dose}$$
 (4)

RESULTS

A total of 30 participants completed this study. Twenty were daily users, among which 10 consumed inhaled cannabis concentrate products and 10 smoked cannabis flower. One participant in the occasional use group had no detectable THC in their blood at the first or subsequent blood draws, and was dropped from the analysis, resulting in 29 participants (Table 1) with highly variable cannabis usage patterns (see Table, Supplemental Digital Content, http:// links.lww.com/TDM/A754).

The average percent THC (wt/wt) of the products used in this study was 39.4%, which was 22.0% and 23.8% in the 2

groups using flower, and 72.1% in the group using concentrate products. The average number of inhalations taken during the consumption period was 9.5 over an average of 5.4 minutes Table 2 presents additional data on participants' observed cannabis consumption and perceived drug effect by group.

There were 203 blood samples collected from 29 research participants. Two occasional users had a total of 6 blood samples, in both cases at later collection points, for which the THC concentrations were below the limit of quantitation. Figure 1 shows the measured blood THC concentration versus time data for all 29 participants included in the analysis, color-coded according to their reported use patterns.

Figure 2 represents the final 3-compartment THC model. Table 3 includes the population pharmacokinetic parameter estimates. The value of the objective function $(-2 \cdot \text{Loglikelihood})$ of the 3-compartment pharmacokinetic model before inclusion of covariates was 1213.34. The 2compartment pharmacokinetic analysis -2.Loglikelihood was 1219.1. There was a significant correlation ($r^2 = 0.97$) of the individual F_{inhaled} parameter estimates of the 2- versus 3-compartment analyses with a slope of 1.41, indicating a systematic 40% underestimation of the inhaled dose when the terminal phase is not accounted for by inclusion of a deep peripheral compartment. Therefore, subsequent results will refer to only the 3-comparment population pharmacokinetic

| | Total N = 29 | $\frac{\text{Daily, Flower}}{n=10}$ | $\frac{\text{Daily, Concentrate}}{n = 10}$ | Occasional, Flower n = 9 | |
|--------------------------|-----------------|-------------------------------------|--------------------------------------------|-----------------------------|--|
| | | | | | |
| Gender | | | | | |
| Male | 10 (34.5%) | 5 (50%) | 3 (30%) | 2 (22.2%) | |
| Female | 19 (65.5%) | 5 (50%) | 7 (70%) | 7 (77.8%) | |
| Age | | | | | |
| Mean (SD) | 32.8 (7.6) | 34.7 (8.3) | 32.2 (6.5) | 31.4 (7.8) | |
| 21–29 | 12 (41.4%) | 4 (40%) | 3 (30%) | 5 (55.6%) | |
| 30–39 | 12 (40%) | 3 (30%) | 6 (60%) | 3 (33.3%) | |
| 40-49 | 3 (10%) | 2 (20%) | 1 (1%) | 0 (0%) | |
| 50-53 | 2 (6.7%) | 1 (10%) | 0 (0%) | 1 (11.1%) | |
| Race | | | | | |
| White | 25 (86.2%) | 9 (90%) | 8 (80%) | 8 (88.9%) | |
| Black/African American | 3 (10.3%) | 1 (10%) | 2 (20%) | 0 (0%) | |
| Other/no response | 1 (3.4%) | 0 (0%) | 0 (0%) | 1 (11.1%) | |
| Ethnicity | | | | | |
| Hispanic/Latino | 5 (17.2%) | 1 (10%) | 3 (30%) | 1 (11.1%) | |
| Non-Hispanic/Latino | 22 (75.9%) | 8 (80%) | 7 (70%) | 7 (77.8%) | |
| Declined to respond | 2 (6.9%) | 1 (10%) | 0 (0%) | 1 (11.1%) | |
| Body mass index (BMI) | | | | | |
| Mean (SD) | 26.9 (6.2) | 26.6 (6.4) | 27.8 (7.5) | 26.3 (4.5) | |
| Age at first use | | | | | |
| Mean (SD) | 15.6 (2.5) | 15.2 (2.5) | 14.4 (2.2) | 17.3 (2.1) | |
| Days used (past 30 days) | | | | | |
| Mean (SD) | 22.8 (9.8) | 28.8 (1.8) | 29.7 (0.5) | 9.1 (3.8) | |
| Times used per day | | | | | |
| Mean (SD) | 3.7 (2.8) | 4.6 (3.5) | 5.2 (1.4) | 1.2 (0.2) | |
| | | | | | |

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| | Total N = 29 Mean (SD) | Daily, Flower n = 10 Mean (SD) | Daily, Concentrate n = 10 Mean (SD) | Occasional, Flower n = 9 Mean (SD) | |
|------------------------------------|------------------------------|--------------------------------------|-------------------------------------------|------------------------------------------|--|
| | | | | | |
| % Total THC (w/w) | 39.8 (26.1) | 22.0 (4.9) | 72.1 (18.7) | 23.8 (4.1) | |
| Weight change (mg) | 178.5 (203.6) | 288.9 (266.6) | 50.0 (34.8) | 198.8 (169.8) | |
| No. of inhalations | 9.5 (6.3) | 12.1 (5.7) | 6.4 (5.1) | 10.1 (6.9) | |
| Consumption time (min) | 5.4 (3.6) | 7.6 (2.5) | 4.1 (3.3) | 4.3 (3.9) | |
| Preconsumption drug effect (mm)* | 1.4 (2.4) | 0.6 (1.4) | 1.5 (2.9) | 2.1 (2.7) | |
| Postconsumption, drug effect (mm)* | 74.9 (18.5) | 85.0 (9.4) | 77.9 (16.5) | 60.4 (20.5) | |

| TABLE 2. Descriptiv | e Summary | / of Participant | Cannabis | Consum | otion |
|---------------------|-----------|------------------|----------|--------|-------|
|---------------------|-----------|------------------|----------|--------|-------|

*The drug effect questionnaire asked, "Do you feel a drug effect right now?" Participants marked on a 100-mm line visual analog scale from "not at all (0 mm)" to "extremely (100 mm)."

analyses. No demographic covariates (ie, age, body weight, or sex), applied to any of the parameter estimates, reduced the objective function by more than 3.84 and, therefore, did not reach statistical significance in the forward segment of the stepwise covariate selection process. Inclusion of pattern of use (daily versus occasional) resulted in a $-2 \cdot \text{Loglikelihood}$ of 1202.11, showing a significant (P < 0.001) reduction of 11.23 (see Figure 1, Supplemental Digital Content, http:// links.lww.com/TDM/A755). Further delineation of usage pattern to include daily concentrate and daily flower use was significant at a lower level and did not further decrease the $-2 \cdot \text{Loglikelihood}$, so it was not included in the final model. Neither weighed dose nor inhalation device used showed a significant reduction in $-2 \cdot \text{Loglikelihood}$ in the forward step. The variance (ω^2) of the estimated F_{inhaled} parameter (ie, the dose estimate) was 1.92 ± 0.022 without the usage pattern covariate and was 0.82 ± 0.23 with the covariate, indicating that approximately 50% of the interindividual variance of inhaled, bioavailable THC was explained by the usage pattern. Diagnostic plots appear as Supplemental Digital Content (see Figure 2, http://links. lww.com/TDM/A755).

The mean weighed, used THC dose was 53.0 ± 58.5 mg, whereas the estimated inhaled amount of THC in all participants was 11.1 ± 11.1 mg. The daily users of concentrates had smaller weighed doses than the daily or occasional flower users (Fig. 3A). However, the estimated inhaled doses of THC were similar for both the daily users of concentrates and flower whereas the estimated inhaled THC doses were much smaller for the occasional users (Fig. 3B), mirroring the relationships among the observed concentration versus time profiles presented in Figure 1.

The weight of the consumed cannabis product multiplied by the product labeled THC concentration for a given inhalation event was a very poor predictor of the actual, inhaled THC amount or exposure, as presented in Figure 4.

DISCUSSION

We described the population pharmacokinetics of THC in 29 subjects during the first 135 minutes after ad libitum inhalation of participant-selected Colorado commercial market cannabis flower and concentrates. Despite the vast differences in product composition and concentration between this study and 2 previous THC population pharmacokinetic analyses (15%–30% THC for cannabis flower and 60%–90% THC for the concentrates in this study—versus 1.75%, 3.55%, and 9% THC cannabis flower as modeled by Sempio et al,²² from the cannabis clinical trial published by Huestis et al⁴ as well as the study by Alvarez et al,²⁵ the corrected blood-to-plasma and blood THC concentrations in this study were similar to those observed in the earlier studies in which the inhalation consumption of low concentration, NIDA-sourced cannabis,²² and THC and CBD powder added to tobacco²⁵ were both rigidly controlled by prompted paced inhalation.

Because there were no blood samples obtained beyond 2.25 hours after inhalation of cannabis in this study, we fixed the values of the deep pharmacokinetic compartment (V3) and elimination clearance (Cle) according to Sempio et al. The pharmacokinetic parameters estimated from the 2 hours of plasma THC concentration versus time data correspond well with previous parameter estimates^{9,22} for the VC, V2, Q2, and Q3 of THC, although these previous studies included time points beyond 12 hours and this study did not include a formal washout period; instead subtracting baseline blood THC concentrations from all measurements to allow direct comparison with these previous studies9,22 and the THC plasma concentration profiles among our subjects without regard for their baseline starting point. Furthermore, using a parsimonious, reduced model (ie, 2-compartment), while fitting plasma THC concentration versus time data, resulted in a marked systematic underestimation of the inhaled dose $(r^2 = 0.97, slope = 1.41)$. This difference is in the same direction as the ratio of the sum of clearances for the respective models (1.83), suggesting a smaller dose is estimated for the 2-compartment model in which concentration versus time data is confined to the early distribution phase before full model identifiability of the ongoing rapid distribution to deep compartments and elimination clearance is apparent. Thus, it is reasonable to use the current THC population pharmacokinetic model parameters to simulate plasma and blood THC concentration profiles, and their variability in similar populations, of various market-available cannabis products for the



FIGURE 1. Observed blood THC concentration versus time, beginning with the first postinhalation blood sample, for all 29 subjects (each line connects the measured concentrations from one subject) after subtraction of the baseline (before inhalation) blood THC concentration. Blue lines are occasional flower users, brown lines are daily concentrate users, and green lines are daily flower users.

first 2 hours after consumption when neuropsychological effects are most apparent.^{14,26} Further study and simulation may be required to best determine precise requirements for estimating the inhaled dose from limited concentration versus time data.

There was a significant difference in the estimated inhaled THC dose based on product labeling and precannabis to postcannabis product weight change between users of

| TABLE 3. Parameters of the Final Pharmacokinetic Model | | | | |
|--------------------------------------------------------|-------------------------|--------------------|-----------|--|
| | Typical value \pm SEE | $\omega^2 \pm SEE$ | Shrinkage | |
| VC (L) | 17.9 ± 1.20 | 0.044 ± 0.044 | 0.29 | |
| V2 (L) | 51.6 ± 4.66 | 0.16 ± 0.16 | 0.10 | |
| V3* (L) | 3327 | | | |
| Q2 (L/min) | 1.65 ± 0.14 | 0.13 ± 0.046 | 0.25 | |
| Q3 (L/min) | 1.75 ± 0.10 | | | |
| CLe* (L/min) | 0.72 | | | |
| Fi | 0.12 ± 0.03 | 0.82 ± 0.02 | 0.004 | |
| Covariate (daily user) | 1.79 ± 0.29 | | | |
| δ | 0.15 ± 0.02 | | | |

*Parameters fixed to values from Sempio et al.22

δ, proportional (relative) intrasubject variability; $ω^2$, intersubject variability; CLe, elimination clearance of THC; Covariate_{daily user}, value of the covariate for the estimated inhaled THC dose for both concentrate and flower users where the default is Covariate_{occasional user} = 0; Fi, fraction of nominal inhaled (bioavailable) THC 'dose' (15 mg); Q2 and Q3, intercompartmental clearances to the fast and slow compartments, respectively; SEE, standard error of the estimate; VC, V2, and V3, volumes of the central, fast, and slow equilibrating THC compartments, respectively.



FIGURE 2. Final compartmental model used to fit plasma THC concentrations versus time. Drug is inhaled into VC of a 3-compartment THC model with rapidly and slowly equilibrating peripheral compartments V2 and V3, intercompartmental clearances Q2 and Q3, and elimination clearance CLe. Fi is the inhaled dose estimated as the fraction of a "standard" 15 mg absolute bioavailable THC dose. The *s signify model parameters which were fixed to the typical value estimates of those parameters from a previous study.²²

cannabis flower and concentrate, with the concentrate group using much less weighed dose. Despite this difference in dose, similar blood THC concentrations were observed for the 2 hours after their inhalation sessions in the 2 daily user groups, regardless of whether flower or concentrate was inhaled. The occasional cannabis flower users had similar weighed THC doses as the daily cannabis flower users. However, the model-estimated inhaled THC dose between the occasional users and both the 2 daily user groups was lower. This suggests that the occasional users are either much less efficient in their ability to extract the active drug from the cannabis product or may self-titrate to smaller inhaled doses and effects. Similar results between daily and occasional users were found by Alvarez et al.²⁵ despite that study's rigidly prompted and paced smoking regimen. This difference associated with cannabis use history was verified by the population pharmacokinetic analysis because occasional versus daily cannabis use (without regard to daily use of flower or concentrate) was a statistically significant covariate for the estimates of THC inhaled dose. The parameter estimates for the covariate of usage pattern (occasional 0. daily = 1.79 ± 0.67) translates to an approximate 5-fold difference in inhaled THC consumption on the day of the study. No other tested covariate met statistical inclusion criterion in the stepwise analysis for modeling of THC compartment volumes, clearances, and dose.

The interindividual variance terms, ω^2 , clearly indicate that the biggest contributor to variance among research participants is the estimated inhaled dose. This variability far exceeds any of the pharmacokinetic parameters found in this study. Nonetheless, the derived parametric indicators, along with observation of the raw data presented in Figures 1 and 4, are consistent with the notion that plasma or blood THC concentration data can reliably be used to estimate the inhaled THC dose. A previous study, using only 2 plasma THC concentration measurements after an unwitnessed cannabis use session,²⁷ was able to estimate inhaled dose, but required many pharmacokinetic modeling assumptions. The current results with detailed pharmacokinetic data for approximately 2.25 hours, beginning approximately 60 seconds after a



FIGURE 3. A, Box plots of weighed THC doses with colors coded to match Figure 1. The dashed lines are the median values; the solid lines are the mean values. The ends of the "box" are the 25th and 75th percentiles. The whiskers show the lowest data value still within 1.5 IQR of the lower quartile and the highest value within 1.5 IQR of the upper quartile, whereas IQR is the interquartile range. B, Box plots of model-estimated inhaled THC doses with colors coded to match Figure 1. The dashed lines are median values; the solid lines are mean values. The ends of the "box" are the 25th and 75th percentiles. The whiskers show the lowest data value still within 1.5 IQR of the lower quartile and the highest value within 1.5 IQR of the upper quartile, whereas IQR is the interquartile range. The value still within 1.5 IQR of the lower quartile and the highest value within 1.5 IQR of the upper quartile, whereas IQR is the interquartile range.



FIGURE 4. Plot of weighed THC doses and model-estimated inhaled THC doses for all subjects. Circles represent the values for each individual and the black line is the linear regression through the origin.

witnessed last inhalation, demonstrates both the feasibility and improved accuracy of using modeled pharmacokinetic data to estimate the THC dose.

The typical inhaled THC dose for daily users estimated in this study (10.78 mg) is only slightly larger than the assumed (fixed) value of 8.40 mg used by Sempio et al²² arrived at by assuming a bioavailability of 0.25 for a 33.6 mg low-concentration (3.55%) NIDA cannabis cigarette.^{28,29} Further studies with larger sample sizes and stratification according to dosage form may allow for covariate refinements of inhaled dose estimates according to product form (ie, flower versus concentrate) and method of inhalation (eg, joint, bowl, vape pen, and dabbing).

In this pharmacokinetic study of 29 subjects, focusing on the time of onset and dissipation of THC neuropsychological effects (ie, within 2.25 hours after inhalation), the estimates of the population pharmacokinetic model parameters most affecting the early time course of plasma and blood THC concentrations were verified and refined. The estimated interindividual variances for the central and rapidly equilibrating volumes of distribution along with the variance estimate for rapid intercompartmental clearance are similar to those reported by Sempio et al²² and Alvarez et al²⁵ and demonstrate, in this larger sample, that the variability of early phase pharmacokinetics among research participants may be small. This observation is supported by Figure 1 in which the shapes of the individual observed blood THC concentrations are similar. Thus, it would be reasonable to apply Bayesian estimation techniques³⁰ to sparse data (1–3 blood or plasma THC concentrations of known intervals) to both estimate the relevant pharmacokinetic parameters for the individual and use the Bayesian parameter estimates to simulate a full blood or plasma THC concentration versus time profile from the time of first inhalation to 2.25 hours postinhalation. Further studies and application of these concepts could have both clinical and forensic interests.

This population pharmacokinetic study of THC had several unique features: enrolled participants represented a wide range of cannabis usage patterns, varying from 2 times per week to 12 times per day; they used different cannabis products (flower and concentrates) from the commercial Colorado market, and the cannabis inhalation pattern on the study visit was ad libitum. This design yielded a diverse dose range which, when combined with an adaptable, but uniform and relatively dense blood sampling regimen, allowed comparison of pharmacokinetics across various patterns of usage and estimated inhaled THC dose. However, the participants in this study were mostly young adult White females (aged 21–39 years), so extrapolations to other populations will benefit from further study.

Certain study limitations were related to the regulatory environment regarding clinical research with this DEA Schedule I drug. The investigators were unable to take possession of and temporarily store a sample of the cannabis products self-supplied by the subjects for independent analysis; however, the product package label was the sole source for THC potency information. Baseline whole blood THC concentrations were subtracted from all subsequent postsmoking measurements. This introduced a small error (<5% over the 2 hours measurement interval), based on the assumption that after a minimum of 8 hours of cannabis abstinence, all subjects would be in the terminal elimination pharmacokinetic phase of THC, which is associated with a 30-100-h half-life.^{9,22,25,31}

CONCLUSION

This population pharmacokinetic analysis of 2.25 hours of blood THC concentration versus time data following ad libitum inhalation of high potency cannabis flower and concentrate products compared closely with the findings of previous THC pharmacokinetic modeling^{9,22,25} and descriptive data.^{7,8} This study demonstrates how this populationbased pharmacokinetic approach facilitates estimation of inhaled THC dose from the pattern of blood THC concentrations over time. Furthermore, the population pharmacokinetic analysis suggests that covariate-driven adjustment to weighed THC dose based on a subject's cannabis usage pattern (occasional versus daily) greatly improves the accuracy of estimating the inhaled THC dose.

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