

*Review*



# **The Role and Mechanism of Probiotics Supplementation in Blood Glucose Regulation: A Review**

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**Abstract:** With economic growth and improved living standards, the incidence of metabolic diseases such as diabetes mellitus caused by over-nutrition has risen sharply worldwide. Elevated blood glucose and complications in patients seriously affect the quality of life and increase the economic burden. There are limitations and side effects of current hypoglycemic drugs, while probiotics, which are safe, economical, and effective, have good application prospects in disease prevention and remodeling of intestinal microecological health and are gradually becoming a research hotspot for diabetes prevention and treatment, capable of lowering blood glucose and alleviating complications, among other things. Probiotic supplementation is a microbiologically based approach to the treatment of type 2 diabetes mellitus (T2DM), which can achieve anti-diabetic efficacy through the regulation of different tissues and metabolic pathways. In this study, we summarize recent findings that probiotic intake can achieve blood glucose regulation by modulating intestinal flora, decreasing chronic low-grade inflammation, modulating glucagon-like peptide-1 (GLP-1), decreasing oxidative stress, ameliorating insulin resistance, and increasing short-chain fatty acids (SCFAs) content. Moreover, the mechanism, application, development prospect, and challenges of probiotics regulating blood glucose were discussed to provide theoretical references and a guiding basis for the development of probiotic preparations and related functional foods regulating blood glucose.

**Keywords:** probiotics; blood glucose regulation; type 2 diabetes mellitus; gut microbiota; insulin resistance; intestinal flora

#### **1. Introduction**

With the improvement of people's living standards, a series of changes in dietary structure and lifestyle have occurred, leading to a rapid increase in the prevalence of metabolic diseases. Globally, the number of people with diabetes is expected to be 643 million by 2030 and 783 million by 2045 [\[1\]](#page-14-0). Diabetes mellitus is divided into three categories: type 1 diabetes mellitus (T1DM), type 2 diabetes mellitus (T2DM), and gestational diabetes mellitus (GDM), with the most significant number of patients suffering from T2DM, accounting for about 90% of the total number of diabetic patients, which has now become a major global crisis [\[2,](#page-14-1)[3\]](#page-14-2). Diabetes-related complications such as macrovascular lesions, microangiopathy, and tumors have become the leading cause of death and disability among diabetic patients in the country. Therapeutic measures at this stage cannot cure diabetes but can delay the onset and mitigate complications [\[4\]](#page-14-3). Currently, insulin therapy is effective but costly, and IDF data show that there are about 450 million people with diabetes worldwide, and the annual treatment cost is as high as USD 670 billion. There are many hypoglycemic drugs such as metformin, miglitol, acarbose, and glyburide, but each has different toxicity and side effects such as hypoglycemia, body mass increase, edema, nausea, etc. [\[5,](#page-14-4)[6\]](#page-14-5). Therefore, most of the current research on T2DM has focused on exploring novel therapeutic approaches without toxic side effects.



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Probiotics are a class of active microorganisms beneficial to the host. They can effectively colonize the human intestinal tract, improving the intestinal microecosystem when ingested in certain quantities, thus regulating the body's intestinal flora. Probiotic therapy has good application prospects in disease prevention and remodeling of intestinal health due to its low cost, high safety, and reliability [\[7](#page-14-6)[,8\]](#page-14-7). According to statistics, the global market of probiotic foods, dietary supplements, and probiotic raw materials is growing at 15% to 20% per year. Probiotics and their products have been shown to improve barrier function, reduce inflammation levels, and enhance immunomodulation. They are considered economical and safe alternatives for treating chronic diseases [\[9](#page-14-8)[–12\]](#page-14-9). Some probiotics and their main functions are shown in Table [1.](#page-1-0) In recent years, some studies have found that intestinal flora and homeostasis play an essential role in the treatment of T2DM, and progress has been made in both clinical studies and scientific research. For example, the results of clinical studies have shown that probiotics can regulate blood glucose in patients with T2DM and that probiotics reduce blood glucose and inflammatory responses by improving intestinal flora, leaky gut, and endotoxemia in patients with T2DM [\[13](#page-14-10)[,14\]](#page-14-11). Probiotics in scientific studies improved T2DM by mediating the gut microbial-SCFA-hormone/inflammatory pathway in mice [\[15\]](#page-14-12). Therefore, probiotics and their products will play an essential role in improving the health of T2DM patients in the future (Figure [1\)](#page-2-0).

<span id="page-1-0"></span>**Table 1.** Probiotics and their main functions.





**Figure 1.** Probiotics may relieve the symptoms of T2DM. **Figure 1.** Probiotics may relieve the symptoms of T2DM.

and challenges of probiotics in regulating T2DM, with a view to providing a theoretical basis for [th](#page-2-1)e development of microecological agents for regulating blood glucose (Figure 2). This paper reviews the mechanism of action, application, development trend, prospects,

<span id="page-2-0"></span>play an essential role in improving the health of T2DM patients in the future (Figure 1).

<span id="page-2-1"></span>

**Figure 2.** Schematic representation of the topic. **Figure 2.** Schematic representation of the topic.

#### **2. Methods**

This review was conducted by electronically searching the literature using the Web of Science. A total of 1599 papers that focused on probiotics and blood glucose from 2014 to 2024 were collected. The publication trend of the review topic keywords is shown in Figure [3.](#page-3-0) The keywords co-occurrence network illustrated the progress in research on the role and mechanism of probiotics supplementation in blood glucose regulation (Figure [4\)](#page-3-1).

<span id="page-3-0"></span>

<span id="page-3-1"></span>

**Figure 4.** Keyword co-occurrence network diagram of published articles that focused on probiotics **Figure 4.** Keyword co-occurrence network diagram of published articles that focused on probiotics and blood glucose (2014–2024). and blood glucose (2014–2024).

# **3. Pathogenesis and Research Status of T2DM 3. Pathogenesis and Research Status of T2DM**

As we all know, diabetes is a chronic metabolic disease that occurs when the body As we all know, diabetes is a chronic metabolic disease that occurs when the body cannot effectively use or produce insulin. The typical feature of diabetes mellitus is that cannot effectively use or produce insulin. The typical feature of diabetes mellitus is that the body cannot maintain normal blood sugar levels, mainly manifested as irritable thirst, the body cannot maintain normal blood sugar levels, mainly manifested as irritable thirst,

excessive eating, polyuria, weight loss, and so on [\[43\]](#page-15-16). This chronic metabolic disorder has become a global health problem due to the severe harm it causes to human health [\[44\]](#page-16-0). Accelerated urbanization, aging, and lifestyle changes have led to an increase in the rate of obesity, which in turn increases the prevalence of diabetes mellitus and cardiovascular diseases [\[45\]](#page-16-1). T2DM is considered as an intestinal disorder that predominates in diabetic patients and is mainly caused by disturbances in glucose metabolism regulated by pancreatic β-cells [46]. Insulin is a peptide hormone secreted by the pancreas by the β-cells which regulates the uptake of glucose in the corpuscular circulation by activating hepatocytes and myocytes as a source of energy or by storing it in the form of glycogen in hepatocytes or skeletal muscle cells. Due to insulin resistance, reduced insulin receptor function and instability of pancreatic β-cells result in the inability of the cells to take up blood glucose,<br> $\frac{1}{2}$  in addition to increase the side of glucose, which in turn leads to T2DM [\[47\]](#page-16-3) (Figure [5\)](#page-4-0). In addition to this, during the formation<br>exercises attention to distance with a complitude declination for the there had a condition to as well as development of diabetes mellitus, dysbiosis of intestinal flora and endotoxins produced by harmful flora further trigger inflammatory response, oxidative stress, and use of produced by harmful flora further trigger inflammatory response, oxidative stress, and  $\alpha$  between  $\alpha$  *j* minimum from the field ingger minimum term of period, destruction of pancreatic β-cells leading to T2DM and complications.

<span id="page-4-0"></span>

**Figure 5.** The pathogenesis of T2DM. The function of surviving cells is impaired due to the apoptosis **Figure 5.** The pathogenesis of T2DM. The function of surviving cells is impaired due to the apoptosis of islet β-cell. This significantly reduces the level of insulin circulating in the blood. In addition, of islet β-cell. This significantly reduces the level of insulin circulating in the blood. In addition, peripheral tissue insulin resistance impairs insulin action, and reduced insulin levels and action can peripheral tissue insulin resistance impairs insulin action, and reduced insulin levels and action can lead to hyperglycemia and hyperlipidemia. lead to hyperglycemia and hyperlipidemia.

**4. T2DM also leads to many complications in addition to the typical symptoms, such also leads to many complications in addition to the typical symptoms, such** increased risk of premature death, with cardiovascular disease being the leading cause of morbidity and mortality in diabetic patients  $[48-50]$  $[48-50]$ . In recent years, patients with T2DM have had an increased risk of death. These complications seriously affect the quality of life of patients and increase economic and family stress, making the treatment and prevention of diabetes urgent. The most effective therapeutic measure at this stage is as heart disease, stroke, kidney failure, amputation, loss of vision, nerve damage, and

insulin, but the cost of insulin therapy is high, and only about 50% of patients with T2DM receive the required insulin therapy [\[51\]](#page-16-6). Moreover, different hypoglycemic agents are tolerated differently (in terms of glycemic control) and have some side effects. In addition to increasing exercise, paying attention to diet, and keeping a comfortable mood, there has been some progress in the research on the use of probiotics to regulate T2DM. However, it is not without side effects.

#### **4. The Role and Mechanism of Probiotics in Blood Glucose Regulation**

Some studies have shown that probiotics regulate blood glucose by regulating intestinal flora balance, intestinal immunity, microbial-gut-brain axis, microbial-gut-hepatic axis, and other pathways (Figure [6\)](#page-5-0) [\[52](#page-16-7)[,53\]](#page-16-8). Research on the mechanism of probiotic action on host blood glucose regulation is gradually deepening, and the existing mechanisms of action may include the following pathways: (1) Probiotics can form a biological immune barrier with intestinal mucosa to maintain the balance of intestinal flora [\[54\]](#page-16-9). (2) Probiotics can reduce insulin resistance and repair oxidative damage to pancreatic β-cells to regulate blood glucose [\[55\]](#page-16-10). (3) Probiotics attach to the intestinal mucosa through adhesins on their surface, constituting a biological barrier and releasing immune factors, thus enhancing the body's immune function [\[56\]](#page-16-11). (4) Probiotics may reduce oxidative stress by decreasing chronic low-grade inflammation [\[57\]](#page-16-12). (5) Probiotics may increase autonomic activity and regulate the activity of enzymes related to glucolipid metabolism [\[58\]](#page-16-13). (6) Probiotics can produce substances such as bacteriocins, which protect the pancreatic islets, promote the function of pancreatic islets, and inhibit  $\alpha$ -glucosidase, playing a key role in regulating blood glucose [59]. (7) The peptides produced during the proliferation [of](#page-16-14) probiotics can inhibit the activity of postsynaptic neurons through the brain-gut axis and improve metabolism [\[60\]](#page-16-15).

<span id="page-5-0"></span>

**Figure 6.** Probiotics regulate blood glucose in a variety of ways. These include regulating the balance **Figure 6.** Probiotics regulate blood glucose in a variety of ways. These include regulating the balance of intestinal flora, intestinal immunity, microbe-gut-brain axis, microbe-gut-liver axis, and so on. of intestinal flora, intestinal immunity, microbe-gut-brain axis, microbe-gut-liver axis, and so on.

### *4.1. Probiotics Regulate Blood Glucose by Improving Intestinal Flora*

The gut microbiota is widely recognized as one of the most important components in maintaining homeostasis. It is closely associated with health and disease in humans and other mammals, and many diseases are accompanied by disorders of the gut microbiota [\[61\]](#page-16-16). More and more studies have shown that the development of T2DM is related to host genetics and environmental factors, and intestinal flora, as a significant environmental factor, is closely associated with the occurrence and development of T2DM [\[62\]](#page-16-17). Probiotics can colonize the human intestinal tract, improve human intestinal flora, regulate metabolism, and maintain the balance of the intestinal system. Probiotics alleviate and treat diabetes by improving the intestinal barrier function, changing the intestinal flora, increasing the content of SCFAs, decreasing oxidative stress and antioxidant effects, inhibiting the activity of enzymes related to glucose absorption, increasing the activity of bile salt hydrolase, and absorption or adsorption of cholesterol. The parts are interrelated and work together to achieve anti-diabetic efficacy [\[63–](#page-16-18)[65\]](#page-16-19). Probiotic complexes reduce *E. coli* and lipopolysaccharide levels and improve intestinal barrier function by increasing levels of SCFA-producing bacteria and SCFAs [\[66\]](#page-16-20). Park et al. [\[67\]](#page-16-21) found that treatment with *Lactobacillus flexneri* HY7601 and *Lactobacillus plantarum* KY1032 resulted in reduced body weight gain and changes in the intestinal flora of obese mice fed a high-fat diet. The probiotic blend, VSL#3, has been shown to inhibit weight gain and insulin resistance by altering the composition of the gut flora [\[68\]](#page-17-0). The combination of probiotics has a more pronounced effect on blood glucose regulation by improving the intestinal SCFA-producing flora, which in turn plays a role. Therefore, probiotics can have a preventive effect on the occurrence and development of metabolic diseases such as diabetes by improving the composition of intestinal flora, increasing beneficial bacteria, and inhibiting harmful bacteria.

#### *4.2. Probiotics Regulate Blood Glucose by Regulating Glucagon-like Peptide-1*

Glucagon-like peptide-1 (GLP-1) is a peptide composed of 31 amino acids, which is an enteric hypoglycemic hormone. When the human body eats, intestinal probiotics can utilize nutrients related to their metabolism to produce a variety of metabolites and stimulate the secretion of GLP-1 from intestinal L-cells, which helps to maintain blood glucose balance in the body, reduce food intake, inhibit obesity, and alleviate T2DM among other things [\[69,](#page-17-1)[70\]](#page-17-2). Long-term obesity not only destroys the function of pancreatic βcells and leads to abnormalities in glucose and lipid metabolism but also increases the synthesis of Dipeptidyl peptidase-IV (DPP-IV) in the body and promotes the degradation of GLP-1 [\[71](#page-17-3)[,72\]](#page-17-4). Therefore, diet-induced obesity may be a pre-sign of T2DM. Probiotics can influence the metabolites of the intestinal flora to activate metabolic pathways in the host and stimulate the secretion of GLP-1 from intestinal L-cells, which in turn alleviates T2DM and enhances the antioxidant capacity of host cells by increasing the levels of peroxide dismutase in vivo. This contributes to the scavenging of free radicals in vivo and reduces the incidence of complications from T2DM [\[73,](#page-17-5)[74\]](#page-17-6). Figure [7](#page-7-0) demonstrates that probiotic bacteria stimulate GLP-1 secretion in host cells by producing metabolites to stimulate the secretion of GLP-1 after the mitigating effect on T2DM. When the nutrients ingested by the body from the outside stimulate the signaling molecules on the intestinal cells, the intestinal L-cells will secrete and release GLP-1, which can repair the function of pancreatic β-cells. While doing this, it can also stimulate pancreatic  $\beta$ -cells to divide, gradually restoring their number and promoting insulin secretion back to the average level [\[75\]](#page-17-7). However, the gut is not the only source of GLP-1. It is also secreted in the brain. Recent studies have found that strains such as *Lactobacillus paracasei* 1F-20, *Lactobacillus fermentum* F40-4, and *Bifidobacterium animalis* subsp. *paracasei* promote the secretion of GLP-1 and peptide YY (PYY) by up-regulating the glucagon gene (GCG) and PYY genes in stanniocalcin-1 (STC-1) cells. Their metabolites can be regulated by peroxisome proliferator-activated receptor-α (PPARα), sterol regulatory element binding protein-1C (SREBP-1C), patatinlike phospholipase domain containing 3 (PNPLA3), and other regulatory genes reaching the liver to improve lipid accumulation and increase glucose uptake by up-regulating PI3K/AKT activity to restore the insulin signaling system [\[76\]](#page-17-8). Acetic acid, propionic acid, and butyric acid are SCFAs that play important probiotic roles in the human body [\[77\]](#page-17-9). Studies have shown that fecal levels of acetic acid and butyric acid were significantly

elevated after fecal transplantation interventions, which led to activation of the GLP-1 pathway by SCFAs and elevated GLP-1 protein expression in colonic tissues, thereby ameliorating glycolipid disorders [\[78\]](#page-17-10). In the future, by exploring the mechanism of action of probiotics and their metabolites on host intestinal cells and regulating their continuous production of GLP-1, we can further clarify the remission effect of probiotics on T2DM and finally provide universal, safe, and effective remission for T2DM patients and promote human health.

<span id="page-7-0"></span>

lites of intestinal flora by regulating the structure of intestinal flora, activating the metabolic pathway in the host body, and stimulating the secretion of GLP-1 by intestinal L-cells. This alleviates T2DM and enhances the antioxidant capacity of host cells by increasing the content of peroxide dismutase in the body, which helps to clear free radicals and reduce the occurrence of T2DM complications. Figure 7. Probiotics relieve diabetes by stimulating GLP-1 secretion. Probiotics can affect the metabo-

dismutase in the body, which helps to clear free radicals and reduce the occurrence of T2DM com-

## 4.3. Probiotics Regulate Blood Glucose by Lowering Inflammation Levels

and inflammatory proteins. Therefore, low-grade systemic inflammation is believed to play a vital role in the development of T2DM and its associated complications [\[79](#page-17-11)[,80\]](#page-17-12). The play a vital role in the development of T2DM and its associated complications [79,80]. The Therefore, the factor contribution in a static is below that the revers of interleukin-0 (IL-6), tumor necrosis factor-α (TNF-α), and interleukin-1β (IL-1β) are significantly higher in T2DM patients than in normal subjects  $[81]$ . Studies have shown that diabetic patients in T2DM patients than in normal subjects  $[81]$ . Studies have shown that diabetic patients have increased intestinal permeability, which correlates with the persistently low level of chronic inflammation present in their intestines, and that the intestinal mucus layer serves as an important barrier that prevents intestinal bacteria from invading the mucosa and causing inflammation [82,83]. Probiotics may ameliorate inflammation by exerting a positive effect on the dysfunction of the epithelial cells and mucosal immune system that form the basis of inflammation. Probiotics alleviate the body's symptoms in patients with prediabetes and T2DM by reducing inflammation and modulating immunity [\[84](#page-17-16)[,85\]](#page-17-17). Amar et al. [\[86\]](#page-17-18) in their study of the effects of probiotics on bacterial translocation and gluthe basis of inflammation. Probiotics alleviate the body's symptoms in patients with pre-*Lactobacillus* 420 application was able to alter the bacterial translocation in the early stages  $\Delta B$  of diabetes and reduce cytokines in tissues TNF-α, IL-1β, plasminogen activator inhibitor-1 et al.  $\mathcal{B}(\mathbf{S})$  in the effects of probiotics on bacterial translocation and glucoses on  $\mathcal{B}$ T2DM is associated with elevated levels of pro-inflammatory cytokines, chemokines, results of the latest clinical trial studies have also shown that the levels of interleukin-6 cose metabolism in high-fat diet-induced diabetic mice, found that *Bifidobacterium animalis*

(PAI-1), and IL-6 expression. *Lactobacillus plantarum* JY039 extracellular polysaccharide and *Lactobacillus paracasei* JY062 alleviated T2DM by balancing pro-inflammatory factor IL-6, TNF-α, and anti-inflammatory factor Interleukin-10 (IL-10) to reduce inflammation [\[87\]](#page-17-19). In addition, probiotics can maintain the stability of the intra-immune environment by balancing pro-inflammatory and anti-inflammatory immune responses, in which SCFAs play an important role in regulating T-cell function and exerting anti-inflammatory effects. The effects of probiotics on inflammatory pathways, weight gain, and glucose metabolism in animals are primarily attributed to the production of SCFAs, which promote the generation and differentiation of regulatory T-cells by directly activating G-protein-coupled receptors and inhibiting histone deacetylase which are potent anti-inflammatory factors. The inflammatory response is alleviated by inhibiting the release of pro-inflammatory factors in lamina propria macrophages [\[88](#page-17-20)[–90\]](#page-17-21).

#### *4.4. Probiotics Regulate Blood Glucose by Improving Oxidative Stress*

Oxidative damage and antioxidant capacity play essential roles in the pathogenesis of diabetes. Hyperglycemia can directly cause reactive oxygen species (ROS) to increase, and these oxygen free radicals induce oxidative stress, which in turn damages the endogenous antioxidant defense system [\[91](#page-18-0)[–93\]](#page-18-1). The intake of probiotics can reduce markers of inflammation and oxidative stress, and improve blood glucose and insulin metabolism [\[94\]](#page-18-2). Zhang et al. [\[95\]](#page-18-3) reported that the effect of probiotics on glucose metabolism could be achieved by reducing oxidative stress. Yadav et al. [\[96](#page-18-4)[,97\]](#page-18-5) showed that probiotics can improve the antioxidant content of glutathione, superoxide dismutase, catalase, and glutathione peroxidase in diabetic rats by inhibiting lipid peroxidation, thereby reducing oxidative damage, increasing insulin secretion, reducing glycosylated hemoglobin level, and reducing intestinal absorption of glucose. This can restore blood glucose to normal levels and ease the development of T2DM.

#### *4.5. Probiotics Regulate Blood Glucose by Improving Insulin Resistance*

Probiotics can improve insulin sensitivity in patients. The binding of SCFAs produced by them to their receptors can significantly reduce fasting blood glucose, fasting plasma insulin, and insulin resistance index levels. In addition, mixed probiotic supplementation can reduce hepatic transaminases and insulin resistance levels, among others [\[92,](#page-18-6)[98\]](#page-18-7). Siebler et al. [\[99\]](#page-18-8) demonstrated that oral administration of *Bifidobacterium bifidum* reduced intestinal endotoxin concentration, improved glucose tolerance, and alleviated insulin resistance in an animal model, thereby regulating blood glucose.

#### *4.6. Probiotics Regulate Blood Glucose by Raising Adiponectin Levels*

Adiponectin is an endogenous bioactive polypeptide or protein secreted by adipocytes, and its level is positively correlated with insulin, which can reflect the efficacy and prognosis of T2DM to a certain extent [\[100\]](#page-18-9). Compared with healthy people, adiponectin levels in T2DM patients are reduced, and the reduction is more obvious in T2DM patients with complications such as atherosclerosis [\[101\]](#page-18-10). Adiponectin improves insulin resistance and protects β-cells by increasing insulin sensitivity. Clinically, it shows hypoglycemic and antiinflammatory potential, alleviating insulin resistance and enhancing glucose metabolism, which are therapeutic targets for diabetes [\[102](#page-18-11)[,103\]](#page-18-12).

#### *4.7. Probiotics Regulate Blood Glucose by Increasing Levels of SCFAs*

By regulating the intestinal microenvironment, probiotics can increase the abundance of SCFA-producing bacteria and inhibit the growth of other pathogenic bacteria such as *Escherichia coli* in the intestine, thereby increasing the content of SCFAs in the intestine [\[104](#page-18-13)[,105\]](#page-18-14). This can help diabetic patients maintain blood glucose balance and effectively relieve T2DM (Figure [8\)](#page-9-0). SCFAs, including butyric acid, acetic acid, and propionic acid, are produced when gut bacteria ferment dietary fiber. SCFAs have a profound effect on insulin sensitivity and energy metabolism, altering the levels of several intestinal peptides involved

in glucose metabolism, intestinal barrier function, and energy homeostasis. For example, butyric acid and propionic acid can inhibit weight gain in obese mice induced by high-fat *Foods* **2024**, *13*, 2719 11 of 22 diets, and acetic acid can reduce food intake in healthy mice [\[106,](#page-18-15)[107\]](#page-18-16). The influence of probiotics on metabolic diseases may be partly due to the metabolic regulation of their metabolites—SCFAs and bile acids (BAs). In addition to providing energy substances for<br>the hadre SCFAs also related as a secondial rate in metabolism insuling considiribution of property the body, SCFAs also play an essential role in regulating insulin sensitivity and energy metabolism [\[108\]](#page-18-17). The effects of probiotics on inflammatory pathways, weight gain, and metabolism [108]. glucose metabolism in animals are primarily attributable to SCFA production  $[109]$ . It has glucose been found that SCFAs directly bind to free fatty acid receptor 2 (FFAR2) in mouse's white adipose tissue as a signaling molecule, inhibiting insulin signal transduction in adipose cells, thereby inhibiting fat accumulation and increasing energy consumption in the liver

> <span id="page-9-0"></span>and muscle [110]. On the other hand, SCFAs as an essential nutrient in the intestinal m[ucos](#page-18-19)a promote the growth and differentiation of intestinal epithelial cells and up-regulate the expression of the intestinal tight junction protein gene and proglucagon glucagon-like peptide-2 (GLP-2) gene in intestinal L-cells, thus strengthening the tight junction between intestinal epithelium, reducing intestinal permeability, and improving intestinal barrier



**Figure 8.** Probiotics relieve blood glucose levels by regulating SCFAs. SCFAs can induce L-cells to **Figure 8.** Probiotics relieve blood glucose levels by regulating SCFAs. SCFAs can induce L-cells to secrete GLP-1 and PYY, and promote insulin secretion to relieve T2DM. In addition, SFCAs reduce the level of inflammatory factors through intestinal epithelial cells, thereby reducing blood sugar content.

# **5. Application of Probiotics to Control Blood Glucose**

**5. Currently, in experimental and clinical studies, probiotics may regulate blood glucose** such as bacteriocins, decreasing inflammation, regulating the intestinal flora, increasing the content of SCFAs, enhancing immunity, and improving insulin resistance. In animal models (Table 2), prob[io](#page-10-0)tics can lower blood glucose and prevent damage to pancreatic β-cells by improving inflammation. *Lactobacillus* and *Bifidobacterium* have been shown to improve glucose tolerance and insulin resistance, and *Bifidobacterium* spp. can improve glucose homeostasis in mice induced by a high-fat diet [\[112](#page-18-21)[,113\]](#page-18-22). In clinical studies (Table 3), probiotic intervention studies have revealed positive effects on glucose metabolism. Among them, the hypoglycemic effects of *Lactobacillus* and *Bifidobacterium* have been demonstrated in several studies  $[114, 115]$  $[114, 115]$ . in T2DM through a combination of the following pathways: production of substances

In addition, in recent years, consumers have increasingly favored probiotic preparations and functional foods containing probiotics that improve gastrointestinal health and

other functions in the market. Due to the advantages of mild hypoglycemic effect, stable nature, and remarkable effect, probiotics for blood glucose regulation have broad market application prospects. Fruits and vegetables are rich in dietary fiber, vitamins, and various phytochemicals. Dietary modification based on fruits and vegetables is undoubtedly an important direction for the prevention and treatment of T2DM and other chronic diseases. Fermented foods can provide us with more probiotics and prebiotics than other types of supplements. Carrots contain hypoglycemic substances and are a good dietary supplement for diabetics. Xiong et al. [\[116\]](#page-18-25) prepared fermented carrot juice using *Lactobacillus plantarum* NCU116, which has a hypoglycemic function. Carrot juice fermented with probiotics can not only preserve the fermented flavor, but also meet the basic requirements of modern human healthcare. Johansson et al. [\[117\]](#page-19-0) applied *Lactobacillus reuteri* 180, which has a hypoglycemic effect on fermented fruit juice. In addition, probiotics can also cooperate with drugs or traditional Chinese medicine to play a role in lowering blood glucose. Studies have found that the combination of probiotics and metformin can enhance the hypoglycemic effect of metformin by regulating intestinal flora and up-regulating intestinal SCFAs [\[118\]](#page-19-1). Jang et al. [\[119\]](#page-19-2) showed that fermented red ginseng with oral probiotics could reduce fasting blood glucose, improve glucose tolerance, and alleviate symptoms of diabetic mice. Flavonoids, alkaloids, polysaccharides, terpenes, and polyphenols in probiotics-fermented fruit juice have an auxiliary hypoglycemic effect.

<span id="page-10-0"></span>**Table 2.** Experimental study on the effect and mechanism of probiotics on diabetes mellitus.



CFU, colony-forming unit; FBS, fasting blood glucose; HbA1c, glycated hemoglobin; OGTT, oral glucose tolerance test; HOMA-IR, homeostasis model assessment of insulin resistance; LPS, lipopolysaccharides; HDL-C, highdensity lipoprotein cholesterol; IGF1R, insulin-like growth factor 1 receptor; ZO-1, zona occludens protein-1; —, Not reported; ↑, rise; ↓, decline.

<b>Probiotics</b>	Sample	Dosage	Duration	<b>Results</b>	Reference
Bifidobacterium bifidum and Lactobacillus acidophilus	20 patients with T2DM	$1 \times 10^8$ CFU/mL	2 weeks	HDL-C $\uparrow$ Fasting glycemia $\downarrow$	Moroti et al. [129]
Lactobacillus paracasei HII01	50 patients with T2DM	$50 \times 10^9$ CFU/d	12 weeks	FBS, LPS, TNF- $\alpha$ , IL-6 and $hscRP \downarrow$	Toejing et al. [14]
Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum	60 patients with TDM	$2 \times 10^9$ CFU	12 weeks	Blood glucose and insulin sensitivity $\downarrow$	Soleimain et al. [130]
Lactobacillus acidophilus, Lactobacillus casei and Lactobacillus rhamnosus	54 patients with T2DM	$1 \times 10^9$ CFU/mL	8 weeks	TGL and HOMA-IR plasma levels $\uparrow$ Serum CRP L	Asemi et al. [114]
Lactobacillus acidophilus	136 patients with T2DM	$1 \times 10^8$ CFU	12 weeks	Blood glucose $\downarrow$ Activity of antioxidant enzymes $\uparrow$	Mirmiranpour et al. [131]
Lactobacillus reuteri <b>DSM 17938</b>	46 patients with T2DM	$1 \times 10^{10}$ CFU/d	12 weeks	FBS, HbA1c, insulin, TC, TG, LDL-C, CRP $\downarrow$ HDL-C $\uparrow$	Mobini et al. [132]
Lactobacillus sporogenes	81 patients with T2DM	$1 \times 10^8$ CFU	8 weeks	Serum insulin levels $\downarrow$	Tajadadi et al. [133]

<span id="page-11-0"></span>**Table 3.** Clinical studies on the effect and mechanism of probiotics on diabetes.

hsCRP, high-sensitivity c-reactive protein; TGL, total glutathione level; CRP, C-reactive protein; TC, total cholesterol; TG, triglycerides. ↑, rise; ↓, decline.

#### **6. Future Development Prospect of Probiotics to Regulate Blood Glucose**

There is an increasing interest in the study of probiotic regulation of blood glucose. Regulation of gut microbiology by probiotics is a potential mechanism. According to the different properties of probiotics, it is theoretically feasible to select different and appropriate combinations to regulate gut microecology in diabetic patients. With the research on the relationship between gut microbes, obesity, and T2DM, we need to focus on the future screening of potential hypoglycemic probiotics, the development of synthetic biology, the utilization of the next generation of probiotics, and the application of postbiotics and paraprobiotics.

#### *6.1. Screening of Potential Hypoglycemic Probiotics*

The screening probiotics need to meet the following three core characteristics: safe and harmless and have a healthy effect on the body, maintain a viable state of bacteria, and have sufficient quantities. At present, there are two main models for the isolation and identification of probiotics: (1) In vitro screening model of Caco-2 cells: simulates intestinal transport in vivo to screen probiotics with excellent performance rapidly and analyzes the bacterial strain's adhesion to the intestine and its impact on intestinal barrier function; (2) Mouse digestive model by gavage: dietary intervention (i.e., adding probiotics to food) was carried out with the help of sterile mouse model and the function of probiotics was explained through the apparent changes in mice [\[134](#page-19-17)[,135\]](#page-19-18). The relevant research contents mainly focused on anti-tumor, anti-cancer, diabetes prevention, intestinal inflammation treatment, and obesity prevention. Wang et al. [\[136\]](#page-19-19) took the world's characteristic foodborne substances as the source of lactic acid bacteria and determined the  $\alpha$ -glucosidase inhibition ability of the strain. Screened *Lactobacillus rhamnosus* LB1lac10 had the effect of lowering blood glucose. At the same time, it was determined that the exopolysaccharide extracellular polysaccharide (EPS1-1) produced by this strain may also act as a natural α-glycosidase inhibitor to regulate blood glucose concentration, and this strain and its exopolysaccharide have particular potential in the development of hypoglycemic foods in the future.

The rapid development of high-throughput sequencing technology in recent years has made it possible to conduct an in-depth analysis of complex samples of traditional fermented food, soil, feces, and so on, filling the data missing caused by technical defects, and promoting the comprehensive analysis of microbial community structure characteristics, functional genes, metabolic pathways, and other information. This lays a foundation for the efficient screening and development of probiotics and then provides new ideas and methods for the exploration and development of beneficial microorganisms, especially those regulating blood glucose [\[137](#page-19-20)[–139\]](#page-19-21).

#### *6.2. Development of Synthetic Biology*

In recent years, with the development of synthetic biology, the next generation of microbial therapies focuses on transforming probiotics into "drug synthesis factories" that can autonomously replicate and detect abnormal conditions to synthesize and release therapeutic factors in the human body. The engineering transformation of natural probiotics and the application of the obtained engineering probiotics to disease monitoring and targeted therapy is a very effective and feasible strategy with a broad application prospect. The therapeutic advantages of engineered probiotics are apparent, such as low cost, few side effects, and simple treatment mode [\[140\]](#page-20-0). Studies have shown that engineered symbiotic bacteria can reprogram intestinal cells into glucose-responsive insulin-secreting cells to treat diabetes [\[141\]](#page-20-1). Zhang et al. [\[142\]](#page-20-2) constructed optogenetically regulated engineered *Lactobacillus* by synthetic biology to achieve controlled secretion of GLP-1 in the organism's intestine under in vitro blue-light stimulation, thus exerting a role in regulating blood glucose. At present, medicine is moving towards the stage of individualized treatment, and probiotics as carriers can play a pivotal role in this field [\[143\]](#page-20-3). In the future, each probiotic could be used as a tailored biological therapy based on a patient's specific clinical situation [\[144\]](#page-20-4). With the continuous improvement and development of synthetic biology and other technologies, the exploration of probiotics will have more significant breakthroughs in the field of food and biomedicine.

#### *6.3. Next-Generation Probiotics*

Next-generation probiotics refer to microbial genera and species that have never been used in the food industry. Candidates have been searched for in health-related gut bacteria, including strains of the genera *Enterococcus faecalis*, *Clostridium*, *Bacteroides*, and *Ackermannia*, as well as genetically modified strains (usually *Lactococcus lactis* with novel health-beneficial properties) [\[145](#page-20-5)[,146\]](#page-20-6). In recent years, with the gradual deepening of research, the next generation of probiotics began to appear as a new prevention and treatment tool, and it is expected to provide a potential targeted pathway and a new direction for the prevention and treatment of diseases. *Akkermansia muciniphila* has been extensively studied in the treatment of metabolic disorders, which can improve insulin resistance and intestinal permeability, increase the energy consumption of obese mice after pasteurization, and thus alleviate diabetes [\[147](#page-20-7)[,148\]](#page-20-8). *Bifidobacterium harzianum* improved insulin sensitivity, increased energy expenditure, increased butyrate production, and regulated gut microbiota composition in diabetic mice [\[149\]](#page-20-9). *Pseudomonas hominis*, *Caulobacter* spp., and *Pseudomonas* spp. can prevent metabolic disorders and obesity by reducing serum leptin levels and fasting blood glucose concentration and improving glucose tolerance. They are considered potential therapeutic targets for T2DM patients [\[150](#page-20-10)[,151\]](#page-20-11). *Prevotella copr* can produce succinic acid in the TCA circulation to improve prediabetic syndrome [\[152](#page-20-12)[,153\]](#page-20-13).

Compared with traditional probiotics, the prominent function of next-generation probiotics is therapeutic, which is worthy of further exploration by researchers. However, the relevant experiments still remain at the level of animal testing and need to be verified in human trials. In the future, the next-generation probiotics are expected to be utilized to develop specific strains of bacteria that can treat diabetes or transplant the relevant strains into the intestinal tract. We hope to achieve better therapeutic effects and provide a new, safer, healthier, and more effective way to treat diabetes.

#### *6.4. Postbiotics and Paraprobiotics*

Probiotics are "living microbes" that can provide health benefits to their hosts. But their inactive ingredients are called paraprobiotics, which are more effective alternatives for susceptible individuals to use [\[53\]](#page-16-8). Postbiotics are defined as "inactivated bacteria and bacterial components that have a beneficial effect on the host". They include cellular components, secreted materials, metabolites, and non-viable microorganisms, which play vital roles in restoring gut microbiota [\[154\]](#page-20-14). Due to their potential to replace antibiotics, post-biologics have been widely used in general food, health food, and gastrointestinal

therapy [\[155\]](#page-20-15). Studies have shown that postbiotics can improve insulin sensitivity, reduce blood sugar levels, and shorten the course of diabetes [\[156\]](#page-20-16). Therefore, postbiotics and paraprobiotics have recently been used as better alternatives for the treatment and prevention of metabolic diseases and their complications. These are also the directions we need to study in the future.

#### **7. Current Challenges in the Regulation of Blood Glucose by Probiotics**

At present, there are some limitations in the clinical application of these novel live microbial therapies, and there are some concerns about their use in patients with immune dysfunction, intestinal barrier dysfunction, and newborns [\[157\]](#page-20-17). Given that probiotics are living microorganisms, many biological and biopharmaceutical barriers limit their clinical application [\[158\]](#page-20-18). The complexity of intestinal flora, the clinical application evaluation of novel therapeutic methods, and the lack of big clinical data all require further exploration and research in the future [\[159\]](#page-20-19). Although some probiotics have achieved good results in vitro and in vivo, research on genetic information, genetic stability, and safety of probiotics is not in-depth. Clinical promotion and wide application are still limited, and more research is needed to understand the impact of gut microbiota on the development of diabetes.

The results of this study can prove that probiotics improve diabetes, but this may fluctuate due to different strains and individual differences [\[160](#page-20-20)[,161\]](#page-20-21). At the same time, probiotics may also be resistant to some experimental subjects or have other unfavorable effects, which need to be comprehensively considered before conducting a comprehensive study on specific probiotic strains.

In addition, although some progress has been made in the research on the regulation of blood glucose by probiotics in recent years, the research on how probiotics play a role and its detailed mechanism is still not perfect. Furthermore, the research on the regulation of blood glucose by targeted probiotics is still not perfect. Therefore, further studies at the genetic and molecular levels are needed to develop probiotic products with solid targeting and precise regulation of blood glucose.

#### **8. Conclusions**

As a potential new intervention target for the treatment of diabetes, probiotics may participate in the regulation of energy metabolism through various mechanisms, namely, reducing chronic low-grade inflammation, regulating intestinal flora, increasing intestinal metabolites SCFAs, reducing oxidative stress, increasing bacterial bioactive peptides and improving insulin resistance, to achieve the purpose of regulating blood glucose. Probiotics are considered economical and safe alternatives to treat chronic diseases and improve human health. However, the prevention and alleviation of chronic diseases such as hyperglycemia through probiotics and their preparations is often a comprehensive synergistic effect of multi-factors, multi-links, multi-sites, and multi-mechanisms. The research on the mechanism of probiotics in lowering blood glucose levels and the interaction between probiotics and a variety of active substances is still insufficient. In the future, it is still necessary to strengthen the basic theory related to probiotics and the mechanism of action at the cellular and molecular levels. Although there are some deficiencies in probiotics at present, these deficiencies will be improved with the deepening of research and the development of science and technology. Therefore, the development of probiotics with blood glucose regulation function and related functional foods is of great significance for the development of the probiotic industry.

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#### **References**

- <span id="page-14-0"></span>1. Saeedi, P.; Petersohn, I.; Salpea, P.; Malanda, B.; Karuranga, S.; Unwin, N.; Colagiuri, S.; Guariguata, L.; Motala, A.A.; Ogurtsova, K.; et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9th edition. *Diabetes Res. Clin. Pract.* **2019**, *157*, 107843. [\[CrossRef\]](https://doi.org/10.1016/j.diabres.2019.107843)
- <span id="page-14-1"></span>2. Salkar, M.; Rosenthal, M.; Thakur, T.; Arnold, A. Patient Centered Studies Focusing on Diabetes Self-Management: A Scoping Review. *Curr. Diabetes Rev.* **2020**, *16*, 557–569. [\[CrossRef\]](https://doi.org/10.2174/1573399816666191230112657) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31886751)
- <span id="page-14-2"></span>3. Dai, M.Y.; Guo, W.J.; Zhu, S.; Gong, G.D.; Chen, M.; Zhong, Z.L.; Guo, J.L.; Zhang, Y.Y. Type 2 diabetes mellitus and the risk of abnormal spermatozoa: A Mendelian randomization study. *Front. Endocrinol.* **2022**, *13*, 1035338. [\[CrossRef\]](https://doi.org/10.3389/fendo.2022.1035338)
- <span id="page-14-3"></span>4. Zhang, X.H.; Liu, L.S.; Campbell, N.R.C.; Niebylski, M.L.; Nilsson, P.; Lackland, D.T.; World Hypertension, L. Implementation of World Health Organization Package of Essential Noncommunicable Disease Interventions (WHO PEN) for Primary Health Care in Low-Resource Settings: A Policy Statement From the World Hypertension League. *J. Clin. Hypertens.* **2016**, *18*, 5–6. [\[CrossRef\]](https://doi.org/10.1111/jch.12749)
- <span id="page-14-4"></span>5. Iqbal, J.; Wu, H.X.; Hu, N.; Zhou, Y.H.; Li, L.; Xiao, F.; Wang, T.; Jiang, H.L.; Xu, S.N.; Huang, B.L.; et al. Effect of glucagon-like peptide-1 receptor agonists on body weight in adults with obesity without diabetes mellitus-a systematic review and meta-analysis of randomized control trials. *Obes. Rev.* **2022**, *23*, e13435. [\[CrossRef\]](https://doi.org/10.1111/obr.13435) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35194917)
- <span id="page-14-5"></span>6. Amjad, S.; Jafri, A.; Sharma, A.K.; Serajuddin, M. A novel strategy of nanotized herbal drugs and their delivery in the treatment of diabetes: Present status and future prospects. *J. Herb. Med.* **2019**, *17–18*, 100279. [\[CrossRef\]](https://doi.org/10.1016/j.hermed.2019.100279)
- <span id="page-14-6"></span>7. Shen, X.Y.; Xie, A.J.; Li, Z.J.; Jiang, C.X.; Wu, J.Q.; Li, M.H.; Yue, X.Q. Research Progress for Probiotics Regulating Intestinal Flora to Improve Functional Dyspepsia: A Review. *Foods* **2024**, *13*, 151. [\[CrossRef\]](https://doi.org/10.3390/foods13010151) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38201179)
- <span id="page-14-7"></span>8. Xie, A.J.; Zhao, S.S.; Liu, Z.F.; Yue, X.Q.; Shao, J.H.; Li, M.H.; Li, Z.W. Polysaccharides, proteins, and their complex as microencapsulation carriers for delivery of probiotics: A review on carrier types and encapsulation techniques. *Int. J. Biol. Macromol.* **2023**, *242*, 124784. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2023.124784) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37172705)
- <span id="page-14-8"></span>9. Hernández-González, J.C.; Martínez-Tapia, A.; Lazcano-Hernández, G.; García-Pérez, B.E.; Castrejón-Jiménez, N.S. Bacteriocins from Lactic Acid Bacteria. A Powerful Alternative as Antimicrobials, Probiotics, and Immunomodulators in Veterinary Medicine. *Animals* **2021**, *11*, 979. [\[CrossRef\]](https://doi.org/10.3390/ani11040979)
- 10. Peng, X.Q.; Ed-Dra, A.; Song, Y.; Elbediwi, M.; Nambiar, R.B.; Zhou, X.; Yue, M. *Lacticaseibacillus rhamnosus* alleviates intestinal inflammation and promotes microbiota-mediated protection against *Salmonella* fatal infections. *Front. Immunol.* **2022**, *13*, 973224. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.973224)
- 11. Smith, D.; Jheeta, S.; Fuentes, H.V.; Palacios-Pérez, M. Feeding Our Microbiota: Stimulation of the Immune/Semiochemical System and the Potential Amelioration of Non-Communicable Diseases. *Life* **2022**, *12*, 1197. [\[CrossRef\]](https://doi.org/10.3390/life12081197)
- <span id="page-14-9"></span>12. Yang, Y.; Song, X.; Wang, G.Q.; Xia, Y.J.; Xiong, Z.Q.; Ai, L.Z. Understanding *Ligilactobacillus salivarius* from Probiotic Properties to Omics Technology: A Review. *Foods* **2024**, *13*, 895. [\[CrossRef\]](https://doi.org/10.3390/foods13060895) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38540885)
- <span id="page-14-10"></span>13. Martin, A.M.; Yabut, J.M.; Choo, J.M.; Page, A.J.; Sun, E.W.; Jessup, C.F.; Wesselingh, S.L.; Khan, W.I.; Rogers, G.B.; Steinberg, G.R.; et al. The gut microbiome regulates host glucose homeostasis via peripheral serotonin. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 19802–19804. [\[CrossRef\]](https://doi.org/10.1073/pnas.1909311116) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31527237)
- <span id="page-14-11"></span>14. Toejing, P.; Khampithum, N.; Sirilun, S.; Chaiyasut, C.; Lailerd, N. Influence of *Lactobacillus paracasei* HII01 Supplementation on Glycemia and Inflammatory Biomarkers in Type 2 Diabetes: A Randomized Clinical Trial. *Foods* **2021**, *10*, 1455. [\[CrossRef\]](https://doi.org/10.3390/foods10071455) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34201653)
- <span id="page-14-12"></span>15. Gu, Y.X.; Chen, H.R.; Li, X.; Li, D.; Sun, Y.; Yang, L.; Ma, Y.; Chan, E.C.Y. *Lactobacillus paracasei* IMC 502 ameliorates type 2 diabetes by mediating gut microbiota-SCFA-hormone/inflammation pathway in mice. *J. Sci. Food Agric.* **2023**, *103*, 2949–2959. [\[CrossRef\]](https://doi.org/10.1002/jsfa.12267)
- <span id="page-14-13"></span>16. Colston, J.M.; Taniuchi, M.; Ahmed, T.; Ferdousi, T.; Kabir, F.; Mduma, E.; Nshama, R.; Iqbal, N.T.; Haque, R.; Ahmed, T.; et al. Intestinal Colonization with *Bifidobacterium longum* Subspecies Is Associated with Length at Birth, Exclusive Breastfeeding, and Decreased Risk of Enteric Virus Infections, but Not with Histo-Blood Group Antigens, Oral Vaccine Response or Later Growth in Three Birth Cohorts. *Front. Pediatr.* **2022**, *10*, 804798.
- 17. Chichlowski, M.; Shah, N.; Wampler, J.L.; Wu, S.S.; Vanderhoof, J.A. *Bifidobacterium longum* Subspecies *infantis* (*B. infantis*) in Pediatric Nutrition: Current State of Knowledge. *Nutrients* **2020**, *12*, 1581. [\[CrossRef\]](https://doi.org/10.3390/nu12061581)
- <span id="page-14-14"></span>18. Feng, X.B.; Su, Y.; Jiang, J.; Li, N.; Ding, W.W.; Wang, Z.M.; Hu, X.H.; Zhu, W.Y.; Li, J.S. Changes in Fecal and Colonic Mucosal Microbiota of Patients with Refractory Constipation after a Subtotal Colectomy. *Am. Surg.* **2015**, *81*, 198–206. [\[CrossRef\]](https://doi.org/10.1177/000313481508100235) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25642885)
- <span id="page-15-0"></span>19. Hsieh, F.C.; Lee, C.L.; Chai, C.Y.; Chen, W.T.; Lu, Y.C.; Wu, C.S. Oral administration of *Lactobacillus* reuteri GMNL-263 improves insulin resistance and ameliorates hepatic steatosis in high fructose-fed rats. *Nutr. Metab.* **2013**, *10*, 35. [\[CrossRef\]](https://doi.org/10.1186/1743-7075-10-35)
- 20. Huang, H.Y.; Korivi, M.; Tsai, C.H.; Yang, J.H.; Tsai, Y.C. Supplementation of *Lactobacillus plantarum* K68 and Fruit-Vegetable Ferment along with High Fat-Fructose Diet Attenuates Metabolic Syndrome in Rats with Insulin Resistance. *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 943020. [\[CrossRef\]](https://doi.org/10.1155/2013/943020)
- 21. Andreasen, A.S.; Larsen, N.; Pedersen-Skovsgaard, T.; Berg, R.M.G.; Moller, K.; Svendsen, K.D.; Jakobsen, M.; Pedersen, B.K. Effects of *Lactobacillus acidophilus* NCFM on insulin sensitivity and the systemic inflammatory response in human subjects. *Br. J. Nutr.* **2010**, *104*, 1831–1838. [\[CrossRef\]](https://doi.org/10.1017/S0007114510002874) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20815975)
- 22. Simon, M.C.; Strassburger, K.; Nowotny, B.; Kolb, H.; Nowotny, P.; Burkart, V.; Zivehe, F.; Hwang, J.H.; Stehle, P.; Pacini, G.; et al. Intake of *Lactobacillus reuteri* Improves Incretin and Insulin Secretion in Glucose-Tolerant Humans: A Proof of Concept. *Diabetes Care* **2015**, *38*, 1827–1834. [\[CrossRef\]](https://doi.org/10.2337/dc14-2690) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26084343)
- 23. Tabuchi, M.; Ozaki, M.; Tamura, A.; Yamada, N.; Ishida, T.; Hosoda, M.; Hosono, A. Antidiabetic effect of *Lactobacillus* GG in streptozotocin-induced diabetic rats. *Biosci. Biotechnol. Biochem.* **2003**, *67*, 1421–1424. [\[CrossRef\]](https://doi.org/10.1271/bbb.67.1421)
- <span id="page-15-1"></span>24. Ejtahed, H.S.; Mohtadi-Nia, J.; Homayouni-Rad, A.; Niafar, M.; Asghari-Jafarabadi, M.; Mofid, V. Probiotic yogurt improves antioxidant status in type 2 diabetic patients. *Nutrition* **2012**, *28*, 539–543. [\[CrossRef\]](https://doi.org/10.1016/j.nut.2011.08.013)
- <span id="page-15-2"></span>25. Meucci, A.; Zago, M.; Rossetti, L.; Fornasari, M.E.; Bonvini, B.; Tidona, F.; Povolo, M.; Contarini, G.; Carminati, D.; Giraffa, G. *Lactococcus hircilactis* sp nov and *Lactococcus laudensis* sp nov., isolated from milk. *Int. J. Syst. Evol. Microbiol.* **2015**, *65*, 2091–2096. [\[CrossRef\]](https://doi.org/10.1099/ijs.0.000225)
- <span id="page-15-3"></span>26. Firmani, S.E.; Maples, H.D.; Balamohan, A. *Lactococcus* Species Central Line-Associated Bloodstream Infection in Pediatrics: A Case Series. *Front. Med.* **2022**, *9*, 802493. [\[CrossRef\]](https://doi.org/10.3389/fmed.2022.802493) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35186991)
- <span id="page-15-4"></span>27. Xu, Z.Y.; Guo, Q.B.; Zhang, H.; Wu, Y.; Hang, X.M.; Ai, L.Z. Exopolysaccharide produced by *Streptococcus thermophiles S-3*: Molecular, partial structural and rheological properties. *Carbohydr. Polym.* **2018**, *194*, 132–138. [\[CrossRef\]](https://doi.org/10.1016/j.carbpol.2018.04.014)
- <span id="page-15-5"></span>28. Lee, S.H.; Park, M.S.; Jung, J.Y.; Jeon, C.O. *Leuconostoc miyukkimchii* sp nov., isolated from brown algae (*Undaria pinnatifida*) kimchi. *Int. J. Syst. Evol. Microbiol.* **2012**, *62*, 1098–1103. [\[CrossRef\]](https://doi.org/10.1099/ijs.0.032367-0)
- 29. Olsen, K.N.; Brockmann, E.; Molin, S. Quantification of Leuconostoc populations in mixed dairy starter cultures using fluorescence in situ hybridization. *J. Appl. Microbiol.* **2007**, *103*, 855–863. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2672.2007.03298.x)
- <span id="page-15-6"></span>30. Yue, X.Q.; Li, M.H.; Liu, Y.M.; Zhang, X.M.; Zheng, Y. Microbial diversity and function of soybean paste in East Asia: What we know and what we don't. *Curr. Opin. Food Sci.* **2021**, *37*, 145–152. [\[CrossRef\]](https://doi.org/10.1016/j.cofs.2020.10.012)
- <span id="page-15-7"></span>31. Cao, J.; Yu, Z.M.; Liu, W.Y.; Zhao, J.X.; Zhang, H.; Zhai, Q.X.; Chen, W. Probiotic characteristics of *Bacillus coagulans* and associated implications for human health and diseases. *J. Funct. Foods* **2020**, *64*, 103643. [\[CrossRef\]](https://doi.org/10.1016/j.jff.2019.103643)
- <span id="page-15-8"></span>32. Angelopoulou, A.; Alexandraki, V.; Georgalaki, M.; Anastasiou, R.; Manolopoulou, E.; Tsakalidou, E.; Papadimitriou, K. Production of probiotic Feta cheese using *Propionibacterium freudenreichii* subsp shermanii as adjunct. *Int. Dairy J.* **2017**, *66*, 135–139. [\[CrossRef\]](https://doi.org/10.1016/j.idairyj.2016.11.011)
- <span id="page-15-9"></span>33. Reiche, A.; Sivell, J.L.; Kirkwood, K.M. Electricity generation by *Propionibacterium freudenreichii* in a mediatorless microbial fuel cell. *Biotechnol. Lett.* **2016**, *38*, 51–55. [\[CrossRef\]](https://doi.org/10.1007/s10529-015-1944-8)
- <span id="page-15-10"></span>34. Lara, Z.B.; Amoranto, M.B.C.; Elegado, F.B.; Dalmacio, L.M.M.; Balolong, M.P. Draft genome sequence of Pediococcus acidilactici 3G3 isolated from Philippine fermented pork. *Microbiol. Resour. Announc.* **2024**, *13*, e0129923. [\[CrossRef\]](https://doi.org/10.1128/mra.01299-23) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38526097)
- 35. Zommiti, M.; Boukerb, A.M.; Feuilloley, M.G.J.; Ferchichi, M.; Connil, N. Draft Genome Sequence of Pediococcus pentosaceus MZF16, a Bacteriocinogenic Probiotic Strain Isolated from Dried Ossban in Tunisia. *Microbiol. Resour. Announc.* **2019**, *8*, 19. [\[CrossRef\]](https://doi.org/10.1128/MRA.00285-19)
- <span id="page-15-11"></span>36. Xie, A.J.; Dong, Y.S.; Liu, Z.F.; Li, Z.W.; Shao, J.H.; Li, M.H.; Yue, X.Q. A Review of Plant-Based Drinks Addressing Nutrients, Flavor, and Processing Technologies. *Foods* **2023**, *12*, 3952. [\[CrossRef\]](https://doi.org/10.3390/foods12213952)
- <span id="page-15-12"></span>37. Suzuki, K.; Ohsumi, Y. Molecular machinery of autophagosome formation in yeast, Saccharomyces cerevisiae. *FEBS Lett.* **2007**, *581*, 2156–2161. [\[CrossRef\]](https://doi.org/10.1016/j.febslet.2007.01.096)
- 38. Borren, E.; Tian, B. The Important Contribution of Non-*Saccharomyces* Yeasts to the Aroma Complexity of Wine: A Review. *Foods* **2021**, *10*, 13. [\[CrossRef\]](https://doi.org/10.3390/foods10010013)
- <span id="page-15-13"></span>39. Yarimizu, T.; Nonklang, S.; Nakamura, J.; Tokuda, S.; Nakagawa, T.; Lorreungsil, S.; Sutthikhumpha, S.; Pukahuta, C.; Kitagawa, T.; Nakamura, M.; et al. Identification of auxotrophic mutants of the yeast *Kluyveromyces marxianus* by non-homologous end joining-mediated integrative transformation with genes from *Saccharomyces cerevisiae*. *Yeast* **2013**, *30*, 485–500. [\[CrossRef\]](https://doi.org/10.1002/yea.2985)
- <span id="page-15-14"></span>40. Stepanovic, S.; Dakic, I.; Hauschild, T.; Vukovic, D.; Morrison, D.; Jezek, P.; Cirkovic, I.; Petras, P. Supplementary biochemical tests useful for the differentiation of oxidase positive staphylococci. *Syst. Appl. Microbiol.* **2007**, *30*, 316–318. [\[CrossRef\]](https://doi.org/10.1016/j.syapm.2006.11.002)
- 41. Nandhini, P.; Kumar, P.; Mickymaray, S.; Alothaim, A.S.; Somasundaram, J.; Rajan, M. Recent Developments in Methicillin-Resistant *Staphylococcus aureus* (MRSA) Treatment: A Review. *Antibiotics* **2022**, *11*, 606. [\[CrossRef\]](https://doi.org/10.3390/antibiotics11050606)
- <span id="page-15-15"></span>42. Vernozy-Rozand, C.; Mazuy, C.; Meugnier, H.; Bes, M.; Lasne, Y.; Fiedler, F.; Etienne, J.; Freney, J. *Staphylococcus fleurettii* sp. nov., isolated from goat's milk cheeses. *Int. J. Syst. Evol. Microbiol.* **2000**, *50 Pt 4*, 1521–1527. [\[CrossRef\]](https://doi.org/10.1099/00207713-50-4-1521) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10939659)
- <span id="page-15-16"></span>43. Herrera-Balandrano, D.D.; Chai, Z.; Hutabarat, R.P.; Beta, T.; Feng, J.; Ma, K.Y.; Li, D.J.; Huang, W.Y. Hypoglycemic and hypolipidemic effects of blueberry anthocyanins by AMPK activation: In vitro and in vivo studies. *Redox Biol.* **2021**, *46*, 102100. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2021.102100)
- <span id="page-16-0"></span>44. Cho, N.H.; Shaw, J.E.; Karuranga, S.; Huang, Y.; Fernandes, J.D.D.; Ohlrogge, A.W.; Malanda, B. IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045. *Diabetes Res. Clin. Pract.* **2018**, *138*, 271–281. [\[CrossRef\]](https://doi.org/10.1016/j.diabres.2018.02.023)
- <span id="page-16-1"></span>45. Jeon, D.; Kim, S.; Kim, M.A.; Chong, Y.P.; Shim, T.S.; Jung, C.H.; Kim, Y.J.; Jo, K.W. Type 2 Diabetes Mellitus- and Complication-Related Risk of Nontuberculous Mycobacterial Disease in a South Korean Cohort. *Microbiol. Spectr.* **2023**, *11*, 4511–4522. [\[CrossRef\]](https://doi.org/10.1128/spectrum.04511-22) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36975830)
- <span id="page-16-2"></span>46. Patil, P.; Mandal, S.; Tomar, S.K.; Anand, S. Food protein-derived bioactive peptides in management of type 2 diabetes. *Eur. J. Nutr.* **2015**, *54*, 863–880. [\[CrossRef\]](https://doi.org/10.1007/s00394-015-0974-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26154777)
- <span id="page-16-3"></span>47. Huang, H.; Yang, K.Y.; Wang, R.N.; Han, W.H.; Kuny, S.; Zelmanovitz, P.H.; Sauvé, Y.; Chan, C.B. β-Cell compensation concomitant with adaptive endoplasmic reticulum stress and β-cell neogenesis in a diet-induced type 2 diabetes model. *Appl. Physiol. Nutr. Metab.* **2019**, *44*, 1355–1366. [\[CrossRef\]](https://doi.org/10.1139/apnm-2019-0144)
- <span id="page-16-4"></span>48. Tomic, D.; Shaw, J.E.; Magliano, D.J. The burden and risks of emerging complications of diabetes mellitus. *Nat. Rev. Endocrinol.* **2022**, *18*, 525–539. [\[CrossRef\]](https://doi.org/10.1038/s41574-022-00690-7)
- 49. Liu, G.; Li, Y.P.; Pan, A.; Hu, Y.; Chen, S.Y.; Qian, F.; Rimm, E.B.; Manson, J.E.; Stampfer, M.J.; Giatsidis, G.; et al. Adherence to a Healthy Lifestyle in Association with Microvascular Complications Among Adults with Type 2 Diabetes. *JAMA Netw. Open* **2023**, *6*, 52239. [\[CrossRef\]](https://doi.org/10.1001/jamanetworkopen.2022.52239)
- <span id="page-16-5"></span>50. Ikeda, S.; Shinohara, K.; Enzan, N.; Matsushima, S.; Tohyama, T.; Funakoshi, K.; Kishimoto, J.; Itoh, H.; Komuro, I.; Tsutsui, H. A higher resting heart rate is associated with cardiovascular event risk in patients with type 2 diabetes mellitus without known cardiovascular disease. *Hypertens. Res.* **2023**, *46*, 1090–1099. [\[CrossRef\]](https://doi.org/10.1038/s41440-023-01178-1)
- <span id="page-16-6"></span>51. Pieber, T.R.; Bajaj, H.S.; Heller, S.R.; Jia, T.; Khunti, K.; Klonoff, D.C.; Ladelund, S.; Leiter, L.A.; Wagner, L.; Philis-Tsimikas, A. Impact of kidney function on the safety and efficacy of insulin degludec versus insulin glargine U300 in people with type 2 diabetes: A post hoc analysis of the CONCLUDE trial. *Diabetes Obes. Metab.* **2022**, *24*, 332–336. [\[CrossRef\]](https://doi.org/10.1111/dom.14564) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34605127)
- <span id="page-16-7"></span>52. Zhou, Y.D.; Liang, F.X.; Tian, H.R.; Luo, D.; Wang, Y.Y.; Yang, S.R. Mechanisms of gut microbiota-immune-host interaction on glucose regulation in type 2 diabetes. *Front. Microbiol.* **2023**, *14*, 1121695. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2023.1121695)
- <span id="page-16-8"></span>53. Pintaric, M.; Langerholc, T. Probiotic Mechanisms Affecting Glucose Homeostasis: A Scoping Review. *Life* **2022**, *12*, 1187. [\[CrossRef\]](https://doi.org/10.3390/life12081187)
- <span id="page-16-9"></span>54. Liu, D.L.; Zhang, S.B.; Li, S.J.; Zhang, Q.; Cai, Y.; Li, P.; Li, H.; Shen, B.C.; Liao, Q.F.; Hong, Y.J.; et al. Indoleacrylic acid produced by *Parabacteroides distasonis* alleviates type 2 diabetes via activation of AhR to repair intestinal barrier. *BMC Biol.* **2023**, *21*, 90. [\[CrossRef\]](https://doi.org/10.1186/s12915-023-01578-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37072819)
- <span id="page-16-10"></span>55. Shahali, A.; Soltani, R.; Akbari, V. Probiotic Lactobacillus and the potential risk of spreading antibiotic resistance: A systematic review. *Res. Pharm. Sci.* **2023**, *18*, 468–477. [\[CrossRef\]](https://doi.org/10.4103/1735-5362.383703)
- <span id="page-16-11"></span>56. Ewaschuk, J.B.; Backer, J.L.; Churchill, T.A.; Obermeier, F.; Krause, D.O.; Madsen, K.L. Surface expression of Toll-like receptor 9 is upregulated on intestinal epithelial cells in response to pathogenic bacterial DNA. *Infect. Immun.* **2007**, *75*, 2572–2579. [\[CrossRef\]](https://doi.org/10.1128/IAI.01662-06)
- <span id="page-16-12"></span>57. Ahmadi, S.; Wang, S.H.; Nagpal, R.; Wang, B.; Jain, S.; Razazan, A.; Mishra, S.P.; Zhu, X.W.; Wang, Z.; Kavanagh, K.; et al. A humanorigin probiotic cocktail ameliorates aging-related leaky gut and inflammation via modulating the microbiota/taurine/tight junction axis. *JCI Insight* **2020**, *5*, 132055. [\[CrossRef\]](https://doi.org/10.1172/jci.insight.132055)
- <span id="page-16-13"></span>58. Rebolledo, M.; Rojas, E.; Salgado, F. Effect of Two Probiotics Containing *Lactobacillus casei rhamnosus* and *Lactobacillus johnsonii* variety on the in vitro Growth of Streptococcus mutans. *Int. J. Odontostomatol.* **2013**, *7*, 415–419. [\[CrossRef\]](https://doi.org/10.4067/S0718-381X2013000300013)
- <span id="page-16-14"></span>59. Miraghajani, M.; Dehsoukhteh, S.S.; Rafie, N.; Hamedani, S.G.; Sabihi, S.; Ghiasvand, R. Potential mechanisms linking probiotics to diabetes: A narrative review of the literature. *Sao Paulo Med. J.* **2017**, *135*, 169–178. [\[CrossRef\]](https://doi.org/10.1590/1516-3180.2016.0311271216) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28538869)
- <span id="page-16-15"></span>60. Gabanyi, I.; Lepousez, G.; Wheeler, R.; Vieites-Prado, A.; Nissant, A.; Wagner, S.; Moigneu, C.; Dulauroy, S.; Hicham, S.; Polomack, B.; et al. Bacterial sensing via neuronal Nod2 regulates appetite and body temperature. *Science* **2022**, *376*, eabj3986. [\[CrossRef\]](https://doi.org/10.1126/science.abj3986)
- <span id="page-16-16"></span>61. Staniszewski, A.; Kordowska-Wiater, M. Probiotic and Potentially Probiotic Yeasts-Characteristics and Food Application. *Foods* **2021**, *10*, 1306. [\[CrossRef\]](https://doi.org/10.3390/foods10061306)
- <span id="page-16-17"></span>62. Delzenne, N.M.; Cani, P.D.; Everard, A.; Neyrinck, A.M.; Bindels, L.B. Gut microorganisms as promising targets for the management of type 2 diabetes. *Diabetologia* **2015**, *58*, 2206–2217. [\[CrossRef\]](https://doi.org/10.1007/s00125-015-3712-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26224102)
- <span id="page-16-18"></span>63. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [\[CrossRef\]](https://doi.org/10.1038/nrgastro.2014.66)
- 64. Cani, P.D.; Osto, M.; Geurts, L.; Everard, A. Involvement of gut microbiota in the development of low-grade inflammation and type 2 diabetes associated with obesity. *Gut Microbes* **2012**, *3*, 279–288. [\[CrossRef\]](https://doi.org/10.4161/gmic.19625) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22572877)
- <span id="page-16-19"></span>65. Okubo, H.; Nakatsu, Y.; Kushiyama, A.; Yamamotoya, T.; Matsunaga, Y.; Inoue, M.K.; Fujishiro, M.; Sakoda, H.; Ohno, H.; Yoneda, M.; et al. Gut Microbiota as a Therapeutic Target for Metabolic Disorders. *Curr. Med. Chem.* **2018**, *25*, 984–1001. [\[CrossRef\]](https://doi.org/10.2174/0929867324666171009121702) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28990516)
- <span id="page-16-20"></span>66. Wang, Y.M.; Dilidaxi, D.; Wu, Y.C.; Sailike, J.; Sun, X.; Nabi, X.H. Composite probiotics alleviate type 2 diabetes by regulating intestinal microbiota and inducing GLP-1 secretion in db/db mice. *Biomed. Pharmacother.* **2020**, *125*, 109914. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2020.109914) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32035395)
- <span id="page-16-21"></span>67. Park, D.Y.; Ahn, Y.T.; Park, S.H.; Huh, C.S.; Yoo, S.R.; Yu, R.; Sung, M.K.; McGregor, R.A.; Choi, M.S. Supplementation of *Lactobacillus curvatus* HY7601 and *Lactobacillus plantarum* KY1032 in Diet-Induced Obese Mice Is Associated with Gut Microbial Changes and Reduction in Obesity. *PLoS ONE* **2013**, *8*, e59470. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0059470)
- <span id="page-17-0"></span>68. Yadav, H.; Lee, J.H.; Lloyd, J.; Walter, P.; Rane, S.G. Beneficial Metabolic Effects of a Probiotic via Butyrate-induced GLP-1 Hormone Secretion. *J. Biol. Chem.* **2013**, *288*, 25088–25097. [\[CrossRef\]](https://doi.org/10.1074/jbc.M113.452516)
- <span id="page-17-1"></span>69. Wang, X.L.; Ye, F.; Li, J.; Zhu, L.Y.; Feng, G.; Chang, X.Y.; Sun, K. Impaired secretion of glucagon-like peptide 1 during oral glucose tolerance test in patients with newly diagnosed type 2 diabetes mellitus. *Saudi Med. J.* **2016**, *37*, 48–54. [\[CrossRef\]](https://doi.org/10.15537/smj.2016.1.12035)
- <span id="page-17-2"></span>70. Sun, X.L.; Zhang, Z.Y.; Liu, M.Y.; Zhang, P.; Nie, L.Q.; Liu, Y.Q.; Chen, Y.; Xu, F.J.; Liu, Z.H.; Zeng, Y.L. Small-molecule albumin ligand modification to enhance the anti-diabetic ability of GLP-1 derivatives. *Biomed. Pharmacother.* **2022**, *148*, 112722. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2022.112722)
- <span id="page-17-3"></span>71. Babu, S.N.; Govindarajan, S.; Vijayalakshmi, M.A.; Noor, A. Role of zonulin and GLP-1/DPP-IV in alleviation of diabetes mellitus by peptide/polypeptide fraction of *Aloe vera* in streptozotocin- induced diabetic wistar rats. *J. Ethnopharmacol.* **2021**, *272*, 113949. [\[CrossRef\]](https://doi.org/10.1016/j.jep.2021.113949) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33610707)
- <span id="page-17-4"></span>72. Ashraf, A.; Mudgil, P.; Palakkott, A.; Iratni, R.; Gan, C.Y.; Maqsood, S.; Ayoub, M.A. Molecular basis of the anti-diabetic properties of camel milk through profiling of its bioactive peptides on dipeptidyl peptidase IV (DPP-IV) and insulin receptor activity. *J. Dairy Sci.* **2021**, *104*, 61–77. [\[CrossRef\]](https://doi.org/10.3168/jds.2020-18627)
- <span id="page-17-5"></span>73. Xiao, J.B.; Högger, P. Dietary Polyphenols and Type 2 Diabetes: Current Insights and Future Perspectives. *Curr. Med. Chem.* **2015**, *22*, 23–38. [\[CrossRef\]](https://doi.org/10.2174/0929867321666140706130807)
- <span id="page-17-6"></span>74. Stefanou, M.I.; Theodorou, A.; Malhotra, K.; de Sousa, D.A.; Katan, M.; Palaiodimou, L.; Katsanos, A.H.; Koutroulou, I.; Lambadiari, V.; Lemmens, R.; et al. Risk of major adverse cardiovascular events and stroke associated with treatment with GLP-1 or the dual GIP/GLP-1 receptor agonist tirzepatide for type 2 diabetes: A systematic review and meta-analysis. *Eur. Stroke J.* **2024**, *9*, 530–539. [\[CrossRef\]](https://doi.org/10.1177/23969873241234238) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38400569)
- <span id="page-17-7"></span>75. Pegah, A.; Abbasi-Oshaghi, E.; Khodadadi, I.; Mirzaei, F.; Tayebinai, H. Probiotic and resveratrol normalize GLP-1 levels and oxidative stress in the intestine of diabetic rats. *Metab. Open* **2021**, *10*, 100093. [\[CrossRef\]](https://doi.org/10.1016/j.metop.2021.100093) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33997755)
- <span id="page-17-8"></span>76. Zhao, F.Q.; Liu, Q.B.; Cao, J.; Xu, Y.S.; Pei, Z.S.; Fan, H.F.; Yuan, Y.Q.; Shen, X.R.; Li, C. A sea cucumber (*Holothuria leucospilota*) polysaccharide improves the gut microbiome to alleviate the symptoms of type 2 diabetes mellitus in Goto-Kakizaki rats. *Food Chem. Toxicol.* **2020**, *135*, 110886. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2019.110886)
- <span id="page-17-9"></span>77. Stadtman, E.R.; Berlett, B.S. Reactive oxygen-mediated protein oxidation in aging and disease. *Drug Metab. Rev.* **1998**, *30*, 225–243. [\[CrossRef\]](https://doi.org/10.3109/03602539808996310)
- <span id="page-17-10"></span>78. Han, X.; Wang, Y.; Zhang, P.P.; Zhu, M.L.; Li, L.; Mao, X.M.; Sha, X.T.; Li, L.L. Kazak faecal microbiota transplantation induces short-chain fatty acids that promote glucagon-like peptide-1 secretion by regulating gut microbiota in *db/db* mice. *Pharm. Biol.* **2021**, *59*, 1077–1087. [\[CrossRef\]](https://doi.org/10.1080/13880209.2021.1954667)
- <span id="page-17-11"></span>79. Kobyliak, N.; Falalyeyeva, T.; Mykhalchyshyn, G.; Kyriienko, D.; Komissarenko, I. Effect of alive probiotic on insulin resistance in type 2 diabetes patients: Randomized clinical trial. *Diabetes Metab. Syndr.* **2018**, *12*, 617–624. [\[CrossRef\]](https://doi.org/10.1016/j.dsx.2018.04.015)
- <span id="page-17-12"></span>80. Cimini, F.A.; D'Eliseo, D.; Barchetta, I.; Bertoccini, L.; Velotti, F.; Cavallo, M.G. Increased circulating granzyme B in type 2 diabetes patients with low-grade systemic inflammation. *Cytokine* **2019**, *115*, 104–108. [\[CrossRef\]](https://doi.org/10.1016/j.cyto.2018.11.019)
- <span id="page-17-13"></span>81. Fadaei, R.; Bagheri, N.; Heidarian, E.; Nouri, A.; Hesari, Z.; Moradi, N.; Ahmadi, A.; Ahmadi, R. Serum levels of IL-32 in patients with type 2 diabetes mellitus and its relationship with TNF-α and IL-6. *Cytokine* **2020**, *125*, 154832. [\[CrossRef\]](https://doi.org/10.1016/j.cyto.2019.154832) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31479874)
- <span id="page-17-14"></span>82. Jayashree, B.; Bibin, Y.S.; Prabhu, D.; Shanthirani, C.S.; Gokulakrishnan, K.; Lakshmi, B.S.; Mohan, V.; Balasubramanyam, M. Increased circulatory levels of lipopolysaccharide (LPS) and zonulin signify novel biomarkers of proinflammation in patients with type 2 diabetes. *Mol. Cell. Biochem.* **2014**, *388*, 203–210. [\[CrossRef\]](https://doi.org/10.1007/s11010-013-1911-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24347174)
- <span id="page-17-15"></span>83. Zhang, Y.M.; Guo, Y.Z.; Jiang, C.X.; Xie, A.J.; Yue, X.Q.; Li, M.H. A review of casein phosphopeptides: From enrichment identification to biological properties. *Food Biosci.* **2024**, *59*, 104217. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2024.104217)
- <span id="page-17-16"></span>84. Naseri, K.; Saadati, S.; Ghaemi, F.; Ashtary-Larky, D.; Asbaghi, O.; Sadeghi, A.; Afrisham, R.; de Courten, B. The effects of probiotic and synbiotic supplementation on inflammation, oxidative stress, and circulating adiponectin and leptin concentration in subjects with prediabetes and type 2 diabetes mellitus: A GRADE-assessed systematic review, meta-analysis, and meta-regression of randomized clinical trials. *Eur. J. Nutr.* **2023**, *62*, 543–561.
- <span id="page-17-17"></span>85. You, S.Y.; Ma, Y.C.; Yan, B.W.; Pei, W.H.; Wu, Q.M.; Ding, C.; Huang, C.X. The promotion mechanism of prebiotics for probiotics: A review. *Front. Nutr.* **2022**, *9*, 1000517. [\[CrossRef\]](https://doi.org/10.3389/fnut.2022.1000517)
- <span id="page-17-18"></span>86. Amar, J.; Chabo, C.; Waget, A.; Klopp, P.; Vachoux, C.; Bermúdez-Humarán, L.G.; Smirnova, N.; Bergé, M.; Sulpice, T.; Lahtinen, S.; et al. Intestinal mucosal adherence and translocation of commensal bacteria at the early onset of type 2 diabetes: Molecular mechanisms and probiotic treatment. *Embo Mol. Med.* **2011**, *3*, 559–572. [\[CrossRef\]](https://doi.org/10.1002/emmm.201100159)
- <span id="page-17-19"></span>87. Zhao, J.Y.; Wang, L.H.; Cheng, S.S.; Zhang, Y.; Yang, M.; Fang, R.X.; Li, H.X.; Man, C.X.; Jiang, Y.J. A Potential Synbiotic Strategy for the Prevention of Type 2 Diabetes: *Lactobacillus paracasei* JY062 and Exopolysaccharide Isolated from *Lactobacillus plantarum* JY039. *Nutrients* **2022**, *14*, 377. [\[CrossRef\]](https://doi.org/10.3390/nu14020377) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35057558)
- <span id="page-17-20"></span>88. Koh, A.; De Vadder, F.; Kovatcheva-Datchary, P.; Bäckhed, F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. *Cell* **2016**, *165*, 1332–1345. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2016.05.041)
- 89. Singh, N.; Gurav, A.; Sivaprakasam, S.; Brady, E.; Padia, R.; Shi, H.D.; Thangaraju, M.; Prasad, P.D.; Manicassamy, S.; Munn, D.H.; et al. Activation of Gpr109a, Receptor for Niacin and the Commensal Metabolite Butyrate, Suppresses Colonic Inflammation and Carcinogenesis. *Immunity* **2014**, *40*, 128–139. [\[CrossRef\]](https://doi.org/10.1016/j.immuni.2013.12.007)
- <span id="page-17-21"></span>90. Chang, P.V.; Hao, L.M.; Offermanns, S.; Medzhitov, R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 2247–2252. [\[CrossRef\]](https://doi.org/10.1073/pnas.1322269111)
- <span id="page-18-0"></span>91. Lenzen, S. Chemistry and biology of reactive species with special reference to the antioxidative defence status in pancreatic β-cells. *Biochim. Biophys. Acta-Gen. Subj.* **2017**, *1861*, 1929–1942. [\[CrossRef\]](https://doi.org/10.1016/j.bbagen.2017.05.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28527893)
- <span id="page-18-6"></span>92. Xiao, R.; Wang, L.L.; Tian, P.J.; Jin, X.; Zhao, J.X.; Zhang, H.; Wang, G.; Zhu, M.M. The Effect of Probiotic Supplementation on Glucolipid Metabolism in Patients with Type 2 Diabetes: A Systematic Review and Meta-Analysis. *Nutrients* **2023**, *15*, 3240. [\[CrossRef\]](https://doi.org/10.3390/nu15143240)
- <span id="page-18-1"></span>93. Shen, X.Y.; Yue, J.Z.; Fu, J.; Guo, Y.Z.; Yang, H.Y.; Liu, Q.M.; Xu, N.; Yue, X.Q.; Li, M.H. Nutritional evaluation of almond protein-whey protein double system and its effect on lipid metabolism in HepG2 cells. *Food Biosci.* **2024**, *61*, 104670. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2024.104670)
- <span id="page-18-2"></span>94. Jadzinsky, M.; Pfuetzner, A.; Paz-Pacheco, F.; Xu, Z.; Allen, F.; Chen, R. Saxagliptin given in combination with metformin as initial therapy improves glycaemic control in patients with type 2 diabetes compared with either monotherapy: A randomized controlled trial. *Diabetes Obes. Metab.* **2009**, *11*, 611–622, Erratum in *Commun. Math. Phys.* **2010**, *12*, 462. [\[CrossRef\]](https://doi.org/10.1111/j.1463-1326.2009.01056.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19515181)
- <span id="page-18-3"></span>95. Zhang, Q.Q.; Wu, Y.C.; Fei, X.Q. Effect of probiotics on glucose metabolism in patients with type 2 diabetes mellitus: A meta-analysis of randomized controlled trials. *Medicina* **2016**, *52*, 28–34. [\[CrossRef\]](https://doi.org/10.1016/j.medici.2015.11.008)
- <span id="page-18-4"></span>96. Yadav, H.; Jain, S.; Sinha, P.R. Antidiabetic effect of probiotic dahi containing *Lactobacillus acidophilus* and *Lactobacillus casei* in high fructose fed rats. *Nutrition* **2007**, *23*, 62–68. [\[CrossRef\]](https://doi.org/10.1016/j.nut.2006.09.002)
- <span id="page-18-5"></span>97. Yadav, H.; Jain, S.; Sinha, P.R. Oral administration of dahi containing probiotic *Lactobacillus acidophilus* and *Lactobacillus casei* delayed the progression of streptozotocin-induced diabetes in rats. *J. Dairy Res.* **2008**, *75*, 189–195. [\[CrossRef\]](https://doi.org/10.1017/S0022029908003129)
- <span id="page-18-7"></span>98. Wu, H.J.; Shang, H.; Wu, J. Effect of ezetimibe on glycemic control: A systematic review and meta-analysis of randomized controlled trials. *Endocrine* **2018**, *60*, 229–239. [\[CrossRef\]](https://doi.org/10.1007/s12020-018-1541-4)
- <span id="page-18-8"></span>99. Siebler, J.; Galle, P.R.; Weber, M.M. The gut-liver-axis: Endotoxemia, inflammation, insulin resistance and NASH. *J. Hepatol.* **2008**, *48*, 1032–1034. [\[CrossRef\]](https://doi.org/10.1016/j.jhep.2008.03.007)
- <span id="page-18-9"></span>100. Nie, Z.H.; Xu, L.M.; Li, C.Y.; Tian, T.; Xie, P.P.; Chen, X.; Li, B.J. Association of endothelial progenitor cells and peptic ulcer treatment in patients with type 2 diabetes mellitus. *Exp. Ther. Med.* **2016**, *11*, 1581–1586. [\[CrossRef\]](https://doi.org/10.3892/etm.2016.3114)
- <span id="page-18-10"></span>101. Malenica, M.; Prnjavorac, B.; Causevic, A.; Dujic, T.; Bego, T.; Semiz, S. Use of Databases for Early Recognition of Risk of Diabetic Complication by Analysis of Liver Enzymes in Type 2 Diabetes Mellitus. *Acta Inform. Medica* **2016**, *24*, 90–93. [\[CrossRef\]](https://doi.org/10.5455/aim.2016.24.90-93) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27147797)
- <span id="page-18-11"></span>102. Kadowaki, T.; Yamauchi, T.; Kubota, N.; Hara, K.; Ueki, K.; Tobe, K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J. Clin. Investig.* **2006**, *116*, 1784–1792. [\[CrossRef\]](https://doi.org/10.1172/JCI29126) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16823476)
- <span id="page-18-12"></span>103. Ruan, H.; Dong, L.Q. Adiponectin signaling and function in insulin target tissues. *J. Mol. Cell Biol.* **2016**, *8*, 101–109. [\[CrossRef\]](https://doi.org/10.1093/jmcb/mjw014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26993044)
- <span id="page-18-13"></span>104. Roelofsen, H.; Priebe, M.G.; Vonk, R.J. The interaction of short-chain fatty acids with adipose tissue: Relevance for prevention of type 2 diabetes. *Benef. Microbes* **2010**, *1*, 433–437. [\[CrossRef\]](https://doi.org/10.3920/BM2010.0028)
- <span id="page-18-14"></span>105. Hong, R.; Xie, A.; Jiang, C.; Guo, Y.; Zhang, Y.; Chen, J.; Shen, X.; Li, M.; Yue, X. A review of the biological activities of lactoferrin: Mechanisms and potential applications. *Food Funct.* **2024**, *15*, 8182–8199. [\[CrossRef\]](https://doi.org/10.1039/D4FO02083A)
- <span id="page-18-15"></span>106. Tolhurst, G.; Heffron, H.; Lam, Y.S.; Parker, H.E.; Habib, A.M.; Diakogiannaki, E.; Cameron, J.; Grosse, J.; Reimann, F.; Gribble, F.M. Short-Chain Fatty Acids Stimulate Glucagon-Like Peptide-1 Secretion via the G-Protein-Coupled Receptor FFAR2. *Diabetes* **2012**, *61*, 364–371. [\[CrossRef\]](https://doi.org/10.2337/db11-1019)
- <span id="page-18-16"></span>107. Lachmandas, E.; van den Heuvel, C.; Damen, M.; Cleophas, M.C.P.; Netea, M.G.; van Crevel, R. Diabetes Mellitus and Increased Tuberculosis Susceptibility: The Role of Short-Chain Fatty Acids. *J. Diabetes Res.* **2016**, *2016*, 6014631. [\[CrossRef\]](https://doi.org/10.1155/2016/6014631)
- <span id="page-18-17"></span>108. Hur, K.Y.; Lee, M.S. Gut Microbiota and Metabolic Disorders. *Diabetes Metab. J.* **2015**, *39*, 198–203. [\[CrossRef\]](https://doi.org/10.4093/dmj.2015.39.3.198)
- <span id="page-18-18"></span>109. Gibson, G.R.; Roberfroid, M.B. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.* **1995**, *125*, 1401–1412. [\[CrossRef\]](https://doi.org/10.1093/jn/125.6.1401)
- <span id="page-18-19"></span>110. Kimura, I.; Ozawa, K.; Inoue, D.; Imamura, T.; Kimura, K.; Maeda, T.; Terasawa, K.; Kashihara, D.; Hirano, K.; Tani, T.; et al. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. *Nat. Commun.* **2013**, *4*, 1829. [\[CrossRef\]](https://doi.org/10.1038/ncomms2852)
- <span id="page-18-20"></span>111. Cani, P.D.; Possemiers, S.; Van de Wiele, T.; Guiot, Y.; Everard, A.; Rottier, O.; Geurts, L.; Naslain, D.; Neyrinck, A.; Lambert, D.M.; et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* **2009**, *58*, 1091–1103. [\[CrossRef\]](https://doi.org/10.1136/gut.2008.165886) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19240062)
- <span id="page-18-21"></span>112. Al-Salami, H.; Butt, G.; Fawcett, J.P.; Tucker, I.G.; Golocorbin-Kon, S.; Mikov, M. Probiotic treatment reduces blood glucose levels and increases systemic absorption of gliclazide in diabetic rats. *Eur. J. Drug Metab. Pharmacokinet.* **2008**, *33*, 101–106. [\[CrossRef\]](https://doi.org/10.1007/BF03191026) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18777945)
- <span id="page-18-22"></span>113. Musso, G.; Gambino, R.; Cassader, M. Obesity, Diabetes, and Gut Microbiota The hygiene hypothesis expanded? *Diabetes Care* **2010**, *33*, 2277–2284. [\[CrossRef\]](https://doi.org/10.2337/dc10-0556) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20876708)
- <span id="page-18-23"></span>114. Asemi, Z.; Zare, Z.; Shakeri, H.; Sabihi, S.S.; Esmaillzadeh, A. Effect of Multispecies Probiotic Supplements on Metabolic Profiles, hs-CRP, and Oxidative Stress in Patients with Type 2 Diabetes. *Ann. Nutr. Metab.* **2013**, *63*, 1–9. [\[CrossRef\]](https://doi.org/10.1159/000349922)
- <span id="page-18-24"></span>115. Eslamparast, T.; Zamani, F.; Hekmatdoost, A.; Sharafkhah, M.; Eghtesad, S.; Malekzadeh, R.; Poustchi, H. Effects of synbiotic supplementation on insulin resistance in subjects with the metabolic syndrome: A randomised, double-blind, placebo-controlled pilot study. *Br. J. Nutr.* **2014**, *112*, 438–445. [\[CrossRef\]](https://doi.org/10.1017/S0007114514000919)
- <span id="page-18-25"></span>116. Xiong, T.; Song, S.H.; Huang, X.H.; Feng, C.; Liu, G.Q.; Huang, J.Q.; Xie, M.Y. Screening and Identification of Functional Lactobacillus Specific for Vegetable Fermentation. *J. Food Sci.* **2013**, *78*, M84–M89. [\[CrossRef\]](https://doi.org/10.1111/j.1750-3841.2012.03003.x)
- <span id="page-19-0"></span>117. Johansson, S.; Diehl, B.; Christakopoulos, P.; Austin, S.; Vafiadi, C. Oligosaccharide Synthesis in Fruit Juice Concentrates Using a Glucansucrase From *Lactobacillus reuteri* 180. *Food Bioprod. Process.* **2016**, *98*, 201–209. [\[CrossRef\]](https://doi.org/10.1016/j.fbp.2016.01.013)
- <span id="page-19-1"></span>118. Zhang, W.; Jia, X.Z.; Xu, Y.H.; Xie, Q.L.; Zhu, M.Z.; Zhang, H.S.; Zhao, Z.F.; Hao, J.Y.; Li, H.Q.; Du, J.R.; et al. Effects of Coix Seed Extract, *Bifidobacterium* BPL1, and Their Combination on the Glycolipid Metabolism in Obese Mice. *Front. Nutr.* **2022**, *9*, 939423. [\[CrossRef\]](https://doi.org/10.3389/fnut.2022.939423)
- <span id="page-19-2"></span>119. Jang, S.H.; Park, J.; Kim, S.H.; Choi, K.M.; Ko, E.S.; Cha, J.D.; Lee, Y.R.; Jang, H.; Jang, Y.S. Red ginseng powder fermented with probiotics exerts antidiabetic effects in the streptozotocin-induced mouse diabetes model. *Pharm. Biol.* **2017**, *55*, 317–323. [\[CrossRef\]](https://doi.org/10.1080/13880209.2016.1237978)
- <span id="page-19-3"></span>120. Lee, Y.S.; Lee, D.; Park, G.S.; Ko, S.H.; Park, J.; Lee, Y.K.; Kang, J. *Lactobacillus plantarum* HAC01 ameliorates type 2 diabetes in high-fat diet and streptozotocin-induced diabetic mice in association with modulating the gut microbiota. *Food Funct.* **2021**, *12*, 6363–6373. [\[CrossRef\]](https://doi.org/10.1039/D1FO00698C)
- <span id="page-19-4"></span>121. Liu, Y.; Zheng, S.J.; Cui, J.L.; Guo, T.T.; Zhang, J.T. *Lactiplantibacillus plantarum* Y15 alleviate type 2 diabetes in mice via modulating gut microbiota and regulating NF-κB and insulin signaling pathway. *Braz. J. Microbiol.* **2022**, *53*, 935–945. [\[CrossRef\]](https://doi.org/10.1007/s42770-022-00686-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35150432)
- <span id="page-19-5"></span>122. Rodrigues, R.R.; Gurung, M.; Li, Z.P.; García-Jaramillo, M.; Greer, R.; Gaulke, C.; Bauchinger, F.; You, H.; Pederson, J.W.; Vasquez-Perez, S.; et al. Transkingdom interactions between *Lactobacilli* and hepatic mitochondria attenuate western diet-induced diabetes. *Nat. Commun.* **2021**, *12*, 101. [\[CrossRef\]](https://doi.org/10.1038/s41467-020-20313-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33397942)
- <span id="page-19-6"></span>123. Jiang, H.R.; Cai, M.M.; Shen, B.Y.; Wang, Q.; Zhang, T.C.; Zhou, X. Synbiotics and Gut Microbiota: New Perspectives in the Treatment of Type 2 Diabetes Mellitus. *Foods* **2022**, *11*, 2438. [\[CrossRef\]](https://doi.org/10.3390/foods11162438)
- <span id="page-19-7"></span>124. Zhang, L.; Qin, Q.Q.; Liu, M.N.; Zhang, X.L.; He, F.; Wang, G.Q. Akkermansia muciniphila can reduce the damage of gluco/lipotoxicity, oxidative stress and inflammation, and normalize intestine microbiota in streptozotocin-induced diabetic rats. *Pathog. Dis.* **2018**, *76*, fty028. [\[CrossRef\]](https://doi.org/10.1093/femspd/fty028)
- <span id="page-19-8"></span>125. Koay, K.P.; Tsai, B.C.K.; Kuo, C.H.; Kuo, W.W.; Luk, H.N.; Day, C.H.; Chen, R.J.; Chen, M.Y.C.; Padma, V.V.; Huang, C.Y. Hyperglycemia-Induced Cardiac Damage Is Alleviated by Heat-Inactivated *Lactobacillus reuteri* GMNL-263 via Activation of the IGF1R Survival Pathway. *Probiotics Antimicrob. Proteins* **2021**, *13*, 1044–1053. [\[CrossRef\]](https://doi.org/10.1007/s12602-021-09745-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33527184)
- <span id="page-19-9"></span>126. Gulnaz, A.; Nadeem, J.; Han, J.H.; Lew, L.C.; Son, J.D.; Park, Y.H.; Rather, I.A.; Hor, Y.Y. *Lactobacillus* Sps in Reducing the Risk of Diabetes in High-Fat Diet-Induced Diabetic Mice by Modulating the Gut Microbiome and Inhibiting Key Digestive Enzymes Associated with Diabetes. *Biology* **2021**, *10*, 348. [\[CrossRef\]](https://doi.org/10.3390/biology10040348)
- <span id="page-19-10"></span>127. Zhou, X.R.; Shang, G.S.; Tan, Q.; He, Q.; Tan, X.Y.; Park, K.Y.; Zhao, X. Effect of *Lactobacillus fermentum* TKSN041 on improving streptozotocin-induced type 2 diabetes in rats. *Food Funct.* **2021**, *12*, 7938–7953. [\[CrossRef\]](https://doi.org/10.1039/D1FO01571K)
- <span id="page-19-11"></span>128. Archer, A.C.; Muthukumar, S.P.; Halami, P.M. *Lactobacillus fermentum* MCC2759 and MCC2760 Alleviate Inflammation and Intestinal Function in High-Fat Diet-Fed and Streptozotocin-Induced Diabetic Rats. *Probiotics Antimicrob. Proteins* **2021**, *13*, 1068, Erratum in *Probiotics Antimicrob. Proteins* **2023**, *15*, 1078. [\[CrossRef\]](https://doi.org/10.1007/s12602-023-10122-1)
- <span id="page-19-12"></span>129. Moroti, C.; Magri, L.F.S.; Costa, M.D.; Cavallini, D.C.U.; Sivieri, K. Effect of the consumption of a new symbiotic shake on glycemia and cholesterol levels in elderly people with type 2 diabetes mellitus. *Lipids Health Dis.* **2012**, *11*, 29. [\[CrossRef\]](https://doi.org/10.1186/1476-511X-11-29)
- <span id="page-19-13"></span>130. Soleimani, A.; Motamedzadeh, A.; Mojarrad, M.Z.; Bahmani, F.; Amirani, E.; Ostadmohammadi, V.; Tajabadi-Ebrahimi, M.; Asemi, Z. The Effects of Synbiotic Supplementation on Metabolic Status in Diabetic Patients Undergoing Hemodialysis: A Randomized, Double-Blinded, Placebo-Controlled Trial. *Probiotics Antimicrob. Proteins* **2019**, *11*, 1248–1256. [\[CrossRef\]](https://doi.org/10.1007/s12602-018-9499-3)
- <span id="page-19-14"></span>131. Mirmiranpour, H.; Huseini, H.F.; Derakhshanian, H.; Khodaii, Z.; Tavakoli-Far, B. Effects of probiotic, cinnamon, and synbiotic supplementation on glycemic control and antioxidant status in people with type 2 diabetes; a randomized, double-blind, placebo-controlled study. *J. Diabetes Metab. Disord.* **2020**, *19*, 53–60. [\[CrossRef\]](https://doi.org/10.1007/s40200-019-00474-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32550156)
- <span id="page-19-15"></span>132. Mobini, R.; Tremaroli, V.; Stahlman, M.; Karlsson, F.; Levin, M.; Ljungberg, M.; Sohlin, M.; Perkins, R.; Perkins, R.; Bäackhed, F.; et al. Metabolic effects of *Lactobacillus reuteri* DSM 17938 in people with type 2 diabetes: A randomized controlled trial. *Diabetes Obes. Metab.* **2017**, *19*, 579–589. [\[CrossRef\]](https://doi.org/10.1111/dom.12861) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28009106)
- <span id="page-19-16"></span>133. Tajadadi-Ebrahimi, M.; Bahmani, F.; Shakeri, H.; Hadaegh, H.; Hijijafari, M.; Abedi, F.; Asemi, Z. Effects of Daily Consumption of Synbiotic Bread on Insulin Metabolism and Serum High-Sensitivity C-Reactive Protein among Diabetic Patients: A Double-Blind, Randomized, Controlled Clinical Trial. *Ann. Nutr. Metab.* **2014**, *65*, 34–41. [\[CrossRef\]](https://doi.org/10.1159/000365153)
- <span id="page-19-17"></span>134. Kozawa, T.; Aoyagi, H. Novel method for screening probiotic candidates tolerant to human gastrointestinal stress. *J. Microbiol. Methods* **2024**, *222*, 106945. [\[CrossRef\]](https://doi.org/10.1016/j.mimet.2024.106945) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38729266)
- <span id="page-19-18"></span>135. Ramlucken, U.; Roets, Y.; Ramchuran, S.O.; Moonsamy, G.; van Rensburg, C.J.; Thantsha, M.S.; Lalloo, R. Isolation, selection and evaluation of *Bacillus* spp. as potential multi-mode probiotics for poultry. *J. Gen. Appl. Microbiol.* **2020**, *66*, 228–238. [\[CrossRef\]](https://doi.org/10.2323/jgam.2019.11.002)
- <span id="page-19-19"></span>136. Wang, J.B.; Yu, L.Y.; Zeng, X.; Zheng, J.W.; Wang, B.; Pan, L. Screening of probiotics with efficient α-glucosidase inhibitory ability and study on the structure and function of its extracellular polysaccharide. *Food Biosci.* **2022**, *45*, 101452. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2021.101452)
- <span id="page-19-20"></span>137. Oberhardt, M.A.; Zarecki, R.; Gronow, S.; Lang, E.; Klenk, H.P.; Gophna, U.; Ruppin, E. Harnessing the landscape of microbial culture media to predict new organism-media pairings. *Nat. Commun.* **2015**, *6*, 8493. [\[CrossRef\]](https://doi.org/10.1038/ncomms9493)
- 138. Oliveira, D.; Vidal, L.; Ares, G.; Walter, E.H.M.; Rosenthal, A.; Deliza, R. Sensory, microbiological and physicochemical screening of probiotic cultures for the development of non-fermented probiotic milk. *Lwt-Food Sci. Technol.* **2017**, *79*, 234–241. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2017.01.020)
- <span id="page-19-21"></span>139. Xie, W.W.; Wu, Y.; Tian, Y.; Li, S.Y.; Zhang, H.Y.; Liu, J.T. Screening for protential new probiotic based on probiotic proper ties and cholesterol degradation activity from Douchi. *Asia-Pac. J. Clin. Oncol.* **2022**, *18*, 80.
- <span id="page-20-0"></span>140. Bijukumar, G.; Somvanshi, P.R. Reverse Engineering in Biotechnology: The Role of Genetic Engineering in Synthetic Biology. *Methods Mol. Biol.* **2024**, *2719*, 307–324.
- <span id="page-20-1"></span>141. Duan, F.F.; Liu, J.H.; March, J.C. Engineered Commensal Bacteria Reprogram Intestinal Cells Into Glucose-Responsive Insulin-Secreting Cells for the Treatment of Diabetes. *Diabetes* **2015**, *64*, 1794–1803. [\[CrossRef\]](https://doi.org/10.2337/db14-0635)
- <span id="page-20-2"></span>142. Zhang, X.Y.; Ma, N.; Ling, W.; Pang, G.J.; Sun, T.; Liu, J.; Pan, H.Z.; Cui, M.H.; Han, C.L.; Yang, C.; et al. A micro-nano optogenetic system based on probiotics for in situ host metabolism regulation. *Nano Res.* **2023**, *16*, 2829–2839. [\[CrossRef\]](https://doi.org/10.1007/s12274-022-4963-5)
- <span id="page-20-3"></span>143. Sampson, K.; Sorenson, C.; Adamala, K.P. Preparing for the future of precision medicine: Synthetic cell drug regulation. *Synth. Biol.* **2024**, *9*, ysae004. [\[CrossRef\]](https://doi.org/10.1093/synbio/ysae004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38327596)
- <span id="page-20-4"></span>144. Komera, I.; Chen, X.L.; Liu, L.M.; Gao, C. Microbial Synthetic Epigenetic Tools Design and Applications. *Acs Synth. Biol.* **2024**, *13*, 1621–1632. [\[CrossRef\]](https://doi.org/10.1021/acssynbio.4c00125) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38758631)
- <span id="page-20-5"></span>145. Khan, M.T.; Dwibedi, C.; Sundh, D.; Pradhan, M.; Kraft, J.D.; Caesar, R.; Tremaroli, V.; Lorentzon, M.; Backhed, F. Synergy and oxygen adaptation for development of next-generation probiotics. *Nature* **2023**, *620*, 7973. [\[CrossRef\]](https://doi.org/10.1038/s41586-023-06378-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37532933)
- <span id="page-20-6"></span>146. Torp, A.M.; Bahl, M.I.; Boisen, A.; Licht, T.R. Optimizing oral delivery of next generation probiotics. *Trends Food Sci. Technol.* **2022**, *119*, 101–109. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2021.11.034)
- <span id="page-20-7"></span>147. Kobyliak, N.; Falalyeyeva, T.; Kyriachenko, Y.; Tseyslyer, Y.; Kovalchuk, O.; Hadiliia, O.; Eslami, M.; Yousefi, B.; Abenavoli, L.; Fagoonee, S.; et al. *Akkermansia muciniphila* as a novel powerful bacterial player in the treatment of metabolic disorders. *Minerva Endocrinol.* **2022**, *47*, 242–252. [\[CrossRef\]](https://doi.org/10.23736/S2724-6507.22.03752-6)
- <span id="page-20-8"></span>148. Jian, H.F.; Liu, Y.T.; Wang, X.M.; Dong, X.Y.; Zou, X.T. *Akkermansia muciniphila* as a Next-Generation Probiotic in Modulating Human Metabolic Homeostasis and Disease Progression: A Role Mediated by Gut-Liver-Brain Axes? *Int. J. Mol. Sci.* **2023**, *24*, 3900. [\[CrossRef\]](https://doi.org/10.3390/ijms24043900)
- <span id="page-20-9"></span>149. Udayappan, S.; Manneras-Holm, L.; Chaplin-Scott, A.; Belzer, C.; Herrema, H.; Dallinga-Thie, G.M.; Duncan, S.H.; Stroes, E.S.G.; Groen, A.K.; Flint, H.J.; et al. Oral treatment with Eubacterium hallii improves insulin sensitivity in db/db mice. *NPJ Biofilms Microbiomes* **2016**, *2*, 16009. [\[CrossRef\]](https://doi.org/10.1038/npjbiofilms.2016.9)
- <span id="page-20-10"></span>150. del Pulgar, E.M.G.; Benítez-Páez, A.; Sanz, Y. Safety Assessment of *Bacteroides Uniformis* CECT 7771, a Symbiont of the Gut Microbiota in Infants. *Nutrients* **2020**, *12*, 551. [\[CrossRef\]](https://doi.org/10.3390/nu12020551)
- <span id="page-20-11"></span>151. Wang, T.Y.; Zhang, X.Q.; Chen, A.L.; Zhang, J.; Lv, B.H.; Ma, M.H.; Lian, J.; Wu, Y.X.; Zhou, Y.T.; Ma, C.C.; et al. A comparative study of microbial community and functions of type 2 diabetes mellitus patients with obesity and healthy people. *Appl. Microbiol. Biotechnol.* **2020**, *104*, 7143–7153. [\[CrossRef\]](https://doi.org/10.1007/s00253-020-10689-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32623494)
- <span id="page-20-12"></span>152. Fu, G.-F.; Li, X.; Hou, Y.-Y.; Fan, Y.-R.; Liu, W.-H.; Xu, G.-X. *Bifidobacterium longum* as an oral delivery system of endostatin for gene therapy on solid liver cancer. *Cancer Gene Ther.* **2005**, *12*, 133–140. [\[CrossRef\]](https://doi.org/10.1038/sj.cgt.7700758) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15565182)
- <span id="page-20-13"></span>153. Li, C.; Chen, X.; Kou, L.; Hu, B.; Zhu, L.P.; Fan, Y.R.; Wu, Z.W.; Wang, J.J.; Xu, G.X. Selenium-*Bifidobacterium longum* as a delivery system of endostatin for inhibition of pathogenic bacteria and selective regression of solid tumor. *Exp. Ther. Med.* **2010**, *1*, 129–135. [\[CrossRef\]](https://doi.org/10.3892/etm_00000022)
- <span id="page-20-14"></span>154. Chen, Q.; Li, X.J.; Xie, W.; Su, Z.A.; Qin, G.M.; Yu, C.H.N. Postbiotics: Emerging therapeutic approach in diabetic retinopathy. *Front. Microbiol.* **2024**, *15*, 1359949. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2024.1359949)
- <span id="page-20-15"></span>155. Tsai, Y.L.; Lin, T.L.; Chang, C.J.; Wu, T.R.; Lai, W.F.; Lu, C.C.; Lai, H.C. Probiotics, prebiotics and amelioration of diseases. *J. Biomed. Sci.* **2019**, *26*, 3. [\[CrossRef\]](https://doi.org/10.1186/s12929-018-0493-6)
- <span id="page-20-16"></span>156. Mishra, S.P.; Wang, B.; Jian, S.; Ding, J.Z.; Rejeski, J.; Furdui, C.M.; Kitzman, D.W.; Taraphder, S.; Brechot, C.; Kumar, A.; et al. A mechanism by which gut microbiota elevates permeability and inflammation in obese/diabetic mice and human gut. *Gut* **2023**, *72*, 1848–1865. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2022-327365) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36948576)
- <span id="page-20-17"></span>157. Piqué, N.; Berlanga, M.; Miñana-Galbis, D. Health Benefits of Heat-Killed (Tyndallized) Probiotics: An Overview. *Int. J. Mol. Sci.* **2019**, *20*, 2534. [\[CrossRef\]](https://doi.org/10.3390/ijms20102534) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31126033)
- <span id="page-20-18"></span>158. Baral, K.C.; Bajracharya, R.; Lee, S.H.; Han, H.K. Advancements in the Pharmaceutical Applications of Probiotics: Dosage Forms and Formulation Technology. *Int. J. Nanomed.* **2021**, *16*, 7535–7556. [\[CrossRef\]](https://doi.org/10.2147/IJN.S337427)
- <span id="page-20-19"></span>159. Koczarski, M. The Effect of Probiotic Supplementation on Glycemic Control in Women with Gestational Diabetes Mellitus A Systematic Review. *Top. Clin. Nutr.* **2020**, *35*, 270–276. [\[CrossRef\]](https://doi.org/10.1097/TIN.0000000000000219)
- <span id="page-20-20"></span>160. Falcinelli, S.; Rodiles, A.; Hatef, A.; Picchietti, S.; Cossignani, L.; Merrifield, D.L.; Unniappan, S.; Carnevali, O. Influence of Probiotics Administration on Gut Microbiota Core: A Review on the Effects on Appetite Control, Glucose, and Lipid Metabolism. *J. Clin. Gastroenterol.* **2018**, *52*, S50–S56. [\[CrossRef\]](https://doi.org/10.1097/MCG.0000000000001064)
- <span id="page-20-21"></span>161. Wu, L.; Gao, Y.; Su, Y.; Li, J.; Ren, W.C.; Wang, Q.H.; Kuang, H.X. Probiotics with anti-type 2 diabetes mellitus properties: Targets of polysaccharides from traditional Chinese medicine. *Chin. J. Nat. Med.* **2022**, *20*, 641–655. [\[CrossRef\]](https://doi.org/10.1016/S1875-5364(22)60210-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36162950)

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