



# *Review* **Revolutionizing Bone Regeneration with Grinder-Based Dentin Biomaterial: A Systematic Review**

 $\bf A$ nna Olchowy  $^{\bf 1}$   $\bf \bullet$  [,](https://orcid.org/0000-0002-4276-8275) Cyprian Olchowy  $^{\bf 2}$   $\bf \bullet$  , Ireneusz Zawiślak  $^{\bf 3}$   $\bf \bullet$  , Jacek Matys  $^{\bf 4,*}$  $^{\bf 4,*}$  $^{\bf 4,*}$   $\bf \bullet$  and Maciej Dobrzyński  $^{\bf 5}$  $^{\bf 5}$  $^{\bf 5}$ 

- <sup>1</sup> Department of Experimental Dentistry, Wroclaw Medical University, Krakowska 26, 50-425 Wroclaw, Poland; ania.olchowy@gmail.com
- <sup>2</sup> Collegium Medicum, Jan Dlugosz University in Czestochowa, Armii Krajowej 13/15, 42-217 Czestochowa, Poland; cyprian.olchowy@gmail.com
- <sup>3</sup> Faculty of Biotechnology and Food Sciences, Wrocław University of Environmental and Life Sciences, Chełmońskiego 37, 51-630 Wroclaw, Poland; ireneusz.zawislak@upwr.edu.pl
- <sup>4</sup> Oral Surgery Department, Wroclaw Medical University, Krakowska 26, 50-425 Wroclaw, Poland<br><sup>5</sup> Department of Podiatric Deptictry and Proclinical Deptictry, Wroclaw, Medical University, Krako
- <sup>5</sup> Department of Pediatric Dentistry and Preclinical Dentistry, Wroclaw Medical University, Krakowska 26, 50-425 Wroclaw, Poland; maciej.dobrzynski@umw.edu.pl
- **\*** Correspondence: jacek.matys@umw.edu.pl

**Abstract:** Bone tissue regeneration is a critical aspect of dental surgery, given the common occurrence of bone resorption leading to alveolar bone defects. The aim of this paper was to conduct a systematic review to provide a comprehensive summary of the evidence regarding the regenerative properties of dentin biomaterial. This systematic review was conducted through comprehensive searches in the PubMed, Scopus, and Web of Science databases, as well as an extensive exploration of the gray literature sources, including WorldCat, The New York Academy of Medicine Library, and Trip Database, following the established PRISMA protocol. Keywords such as tooth, dentin, grinder, and autograft guided the search, with a focus on a standardized procedure involving dentin grinders within laboratory, experimental, and clinical settings. Initially, a pool of 1942 articles was identified with 452 duplicates removed. An additional 1474 articles were excluded for not aligning with the predefined topics, and three more were excluded due to the unavailability of the full text. Ultimately, 13 articles met the strict inclusion criteria and were included in the review. The chemical composition of the dentin particles was similar to natural bone in terms of oxygen, carbon, calcium, phosphorus, sodium, and magnesium content, as well as in terms of the Ca/P ratio. In addition, the dentin also contained amide I and amide II structures, as well as aliphatic and hydroxyl functional groups. The chemically treated dentin was free of microorganisms. The dentin had characteristic tubules that opened after chemical treatment. At the cellular level, dentin released bone morphogenetic protein 2, induced significant cell growth, and stimulated the reorganization of the fibroblast cytoskeleton. Most clinical studies have focused on alveolar bone regeneration. After the transplantation of demineralized dentin particles, studies have observed new bone formation, a reduction in residual bone, and an increase in connective tissue. Clinical reports consistently indicate uncomplicated healing and recovery post-transplantation. However, there is a notable gap in the evidence concerning complication rates, patient-reported outcomes, and the presence of pro-inflammatory factors. In conclusion, dentin biomaterial emerges as a versatile bone substitute, demonstrating high biocompatibility and ease of acquisition. The preservation of its internal structure containing organic matter and growth factors enhances its potential for effective bone regeneration. Particularly, in dental surgery, dentin-derived materials present a promising alternative to traditional autologous bone autografts, offering the potential to reduce patient morbidity and treatment costs.

**Keywords:** bone regeneration; bone grafting; bone substitutes; demineralized dentin matrix; autogenous tooth graft; tooth grinder; dentin grinder



**Citation:** Olchowy, A.; Olchowy, C.; Zawiślak, I.; Matys, J.; Dobrzyński, M. Revolutionizing Bone Regeneration with Grinder-Based Dentin Biomaterial: A Systematic Review. *Int. J. Mol. Sci.* **2024**, *25*, 9583. [https://](https://doi.org/10.3390/ijms25179583) [doi.org/10.3390/ijms25179583](https://doi.org/10.3390/ijms25179583)

Academic Editor: Luigi Canullo

Received: 1 July 2024 Revised: 21 August 2024 Accepted: 3 September 2024 Published: 4 September 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

## **1. Introduction**

Bone tissue regeneration is a critical aspect of orthopedic and dental surgery. Conditions requiring bone tissue regeneration can be caused by trauma, infections, malignant lesions, or congenital defects or can develop as a consequence of applied treatment, such as following radiotherapy of malignant lesions [\[1,](#page-17-0)[2\]](#page-17-1). In dentistry, the occurrence of bone resorption leading to alveolar bone defects surpasses 90% and depends on patient demographics, oral hygiene practices, and general health [\[3\]](#page-17-2). Bone grafting and bone graft substitutes are frequently employed to provide structural support, protect from bone loss, secure capacity for future prosthetic implantation, and facilitate bone healing. A variety of bone grafts are available and are classified according to their origin into natural, synthetic, and composite categories. Natural bone graft materials encompass autogenic allogeneic and xenogeneic sources, while synthetic options include hydroxyapatite, tricalcium phosphate, bioactive glass, and polymers. Composite bone graft materials represent a hybrid of natural and synthetic components. Each type of bone graft material presents a distinct set of advantages and disadvantages [\[4\]](#page-17-3) (see Figure [1\)](#page-1-0). In the USA, the use of bone graft procedures has been on a downward trend over time. During the period from 2000 to 2003, autogenous bone grafts accounted for 83% of the procedures, while artificial bone grafts constituted 17%. Nonetheless, with the aid of contemporary advancements and novel surgical techniques, it is now feasible to extend the eligibility criteria for older patients, reduce the treatment burden, and shorten the duration of procedure-related hospitalization [\[4\]](#page-17-3).

<span id="page-1-0"></span>

**Figure 1.** Classification of bone replacement materials [\[1](#page-17-0)[,2\]](#page-17-1).

Autologous bone refers to bone tissue harvested from the same patient, minimizing the risk of immune rejection and preserving robust osteogenic, osteoinductive, and osteoconductive properties [\[5\]](#page-17-4). In contrast to allogeneic and xenogeneic grafts, autologous bone exhibits lower resorption rates [\[6\]](#page-17-5). Despite these advantages, the harvesting of autogenous bone carries a potential risk of donor site morbidity [\[7\]](#page-17-6), prompting a recent trend toward substitutive grafts [\[4\]](#page-17-3). Allogeneic and xenogeneic bone grafts involve the use of bone material from one individual to another within the same species or between different species. Although they are associated with higher resorption rates and an increased risk of immunologic reactions and disease transmission, these grafts eliminate donor site morbidity, thereby reducing the treatment burden on the patient [\[8\]](#page-17-7). Given the limited efficacy of natural bone grafts, various synthetic bone substitutes have been investigated in clinical practice as alternative options. The objective was to offer an economical graft that closely mirrors the structure and strength of human bone while facilitating new bone

formation [\[9\]](#page-17-8). Despite their ability to replicate the structure and composition of natural  $\epsilon$  bone, be customized, and eliminate disease transmission, synthetic bone substitutes may be  $\epsilon$ . provoke a foreign body response. Additionally, their constrained biological activity and absence of cellular components found in natural bone can result in reduced efficiency in the regeneration process [\[10\]](#page-17-9). For these reasons, the focus has shifted toward osteoinductive materials containing growth factors, such as the demineralized bone matrix and bone materials containing grow in factors, such as the demineralized bone matrix and bone<br>morphogenetic proteins. Clinical outcomes may vary due to differences in osteoinductive properties, and considerations arise regarding the concentration of growth factors, tissue source, and potential variations in production process parameters. The high cost of the procedure further serves as a limiting factor [\[11\]](#page-17-10).

cellular components found in natural bone can result in reduced efficiency in the regener-

Human dentin materials from extracted teeth were traditionally deemed unsuitable for riuman denun materials from extracted teeth were traditionally deemed urisuitable for<br>use as a graft in bone regeneration. The first clinical case report documenting the utilization the interaction of patient control of patients and the control of patient-owned dentin for bone augmentation was published in 2002 [\[12\]](#page-17-11). Subsequently, researchers have started to develop diverse processing techniques for non-functional teeth, transforming them into a viable native resource for bone regeneration. Pang et al. [\[13\]](#page-17-12) conducted a comparative study assessing the efficacy of autogenous tooth graft material<br>300 and 800 percential articulate text Skytch underwenter in diameter. Subsequently, the particular was in a and anorganic bovine bone. The teeth were mechanically crushed into particles ranging and anorganic bovine bone. The teen were incentifieding erashed into particles ranging<br>between 300 and 800 µm in diameter. Subsequently, the particulate teeth underwent a series of processes, including washing, defatting, decalcification, and lyophilization. The resulting autogenous demineralized dentin matrix, as well as the anorganic bovine bone, were grafted into extraction sockets post-tooth extraction to address bone defects. The utilization of an autogenous demineralized dentin matrix demonstrated effectiveness The unization of an autogenous demineralized dentificantly demonstrated enectiveness<br>comparable to that achieved with anorganic bovine bone. Both groups displayed favorable wound healing, comparable implant stability, and histologically confirmed new bone formation. Similar results were reported in other studies [\[14](#page-17-13)[,15\]](#page-18-0) (Figure [2\)](#page-2-0).

<span id="page-2-0"></span>

Figure 2. Diagram of the specimen preparation and clinical application process: (A,B) debridement **Figure 2.** Diagram of the specimen preparation and clinical application process: (**A**,**B**) debridement of the third molar root following extraction; (C) removal of the tooth crown before grinding; (D) use of the Smart Dentin Grinder™ apparatus (KometaBio Inc., Cresskill, NJ, USA); (E) graft material of the Smart Dentin Grinder™ apparatus (KometaBio Inc., Cresskill, NJ, USA); (**E**) graft material post-grinding; (F) image of the alveolar socket before grafting; (G) image of the alveolar socket after grafting.

The aim of this project was to conduct a systematic review to provide a comprehensive if the anti of this project was to conduct a systematic feview to provide a comprenensive<br>summary of the evidence concerning the regenerative properties of dentin-biomaterial. Understanding the impact of dentin as a biomaterial, including at the molecular level, is crucial for clinicians. This knowledge can help clinicians in selecting the appropriate type of dentin-derived material based on the specific clinical scenario. level, is crucial for clinicians. This knowledge can help clinicians in selecting the appro-

# 2. Materials and Methods

# 2.1. Focus Question

With a particular emphasis on the outcomes associated with the utilization of a standardized grinding protocol utilizing a dentin grinder for the preparation of dentin biomaterial in various settings, including laboratory, experimental, and clinical contexts, what is the extensive molecular evidence concerning the regenerative attributes of dentin biomaterial?

#### *2.2. Protocol*

The selection process for articles in the systematic review was carefully outlined following the PRISMA flow diagram (Figure [3\)](#page-3-0). The systematic review was registered on the Open Science Framework at the following link: <https://osf.io/zj7p8> (accessed on 20  $\mu$ August 2024).  $T_{\text{max}}$  for articles in the systematic review was carefully outlined for a sys low selection process for articles in the systematic review was carefully on the selection process for all the systematic review was carefully on the systematic review was registered on the systematic review was carefully

# <span id="page-3-0"></span>PRISMA 2020 flow diagram for new systematic reviews which included searches of databases and registers only



Figure 3. PRISMA flowchart. tervention, OUTCOMES, PRISMA flowchart.<br>
Study Design (PICOS) framework framework framework framework and Study 2. Principal study of the Study of the<br>
Figure 3. PRISMA flowchart.

### *2.3. Eligibility Criteria*

The inclusion and exclusion criteria were developed according to the Population, Intervention, Comparison, Outcomes, and Study Design (PICOS) framework [\[16\]](#page-18-1). All inclusion and exclusion criteria are listed in Table [1.](#page-4-0) Our search was restricted to articles published in English; nevertheless, no limitations were applied regarding the publication timeframe or geographical scope. The search specifically focused on a specific dentin grinding method utilizing a dentin grinder in the human model. This limitation was justified by the availability of a standardized grinding procedure outlined in the manufacturer's instructions. The grinder is programmable to generate particles within a specified size range of  $300 \mu m$ to 1200 µm, ensuring standardization and reproducibility across all studies [\[17\]](#page-18-2).



<span id="page-4-0"></span>**Table 1.** The Population, Intervention, Comparison, Outcomes, and Study Design (PICOS) framework.

#### *2.4. Information Sources, Search Strategy, and Study Selection*

In May 2024, a thorough and systematic electronic exploration unfolded across prominent scholarly databases, including PubMed, Cochrane, Web of Science, and Scopus. In addition, the gray literature sources such as WorldCat, The New York Academy of Medicine Library, and Trip Database were searched. To ensure a comprehensive search of the relevant literature, the search strategy used MeSH terms, such as "tooth", "dentin", and "autograft", and also included non-MeSH keywords, such as "grinder". The search criteria were carefully designed, focusing on a strategic combination of the mentioned keywords. For PubMed, we used ((tooth[Title/Abstract]) OR (dentin[Title/Abstract])) AND ((grinder[Title/Abstract]) OR (autograft[Title/Abstract])). For WoS, we used  $AB =$  ((tooth OR dentin) AND (grinder OR autograft)). For Scopus, we used (TITLE-ABS-KEY (tooth) OR TITLE-ABS-KEY (dentin)) AND (TITLE-ABS-KEY (grinder) OR TITLE-ABS-KEY (autograft)). For WorldCat, we used ((tooth) OR (dentin)) AND ((grinder) OR (autograft)). For The New York Academy of Medicine Library, we used ((tooth) OR (dentin)) AND ((grinder) OR (autograft)). For Trip Database, we used ((tooth) OR (dentin)) AND ((grinder) OR (autograft)). The search methodology strictly adhered to the guidelines set forth in the Preferred Reporting Items for Systematic Review and Meta-analysis (PRISMA) statement [\[18\]](#page-18-3). Supplementary searches were conducted in journals pertinent to oral surgery and periodontology. Subsequent to the database search, a meticulous and exhaustive literature review was systematically carried out to identify any papers initially deemed potentially irrelevant to this study. Inclusion considerations were strictly reserved for articles with full-text versions.

#### *2.5. Data Collection Process and Data Items*

During the initial stage of study selection, the authors independently reviewed the titles and abstracts of each study. Any discrepancies in the inclusion or exclusion of articles were thoroughly discussed among the reviewers to ensure consensus. Three reviewers (C.O., A.O., I.Z.) autonomously performed data extraction from articles that satisfied the predefined inclusion criteria. The gathered information was then meticulously recorded in a standardized spreadsheet for systematic organization and analysis.

#### *2.6. Quality Assessment*

Two independent assessors (C.O., I.Z.) systematically evaluated the methodological rigor of each study to decide on the inclusion of the studies. In the event of a disagreement between reviewers regarding the inclusion of an article, a third reviewer was consulted to resolve the dispute and make the final decision. For the quality assessment, a set of critical appraisal tools was developed by the Joanna Briggs Institute (JBI) [\[19\]](#page-18-4).

To assess inter-rater reliability, Cohen's kappa test was conducted. The results of Cohen's kappa test indicated perfect agreement among the reviewers, with a kappa value of 1.0, demonstrating 100% consistency in their assessments.

#### **3. Results**

# *3.1. Study Selection*

Initially, 1942 articles were identified, with 452 duplicates removed. A further 1474 articles were excluded for not meeting the predefined topics, and three were excluded due to lack of access to the full text. In the end, 13 articles met the strict inclusion criteria and were included in the review. These comprised three in vitro studies [\[20–](#page-18-5)[22\]](#page-18-6) and ten human studies [\[14,](#page-17-13)[15,](#page-18-0)[17,](#page-18-2)[23–](#page-18-7)[29\]](#page-18-8). The human studies involved a total of 127 patients who were subjected to the procedure with a smart dentin grinder, with three case reports and studies ranging from nine to fifty-two patients.

### *3.2. Molecular Aspects of Ground Dentin Grafts*

Molecular aspects were exclusively evaluated in three studies [\[20](#page-18-5)[–22\]](#page-18-6). None of the studies focused solely on clinical outcomes. Ten studies presented results on the chemical composition or morphology evaluation of the material before grafting, as well as histomorphometric evaluation of bioptates obtained during implantation, alongside clinical data [\[14,](#page-17-13)[15](#page-18-0)[,17](#page-18-2)[,23–](#page-18-7)[29\]](#page-18-8). The understanding of the molecular aspects in studies involving bone tissue regeneration with ground dentin grafts is pivotal for the integration of a biomaterial into clinical practice. This aspect was explored through in vitro studies, morphometric analyses, and histological studies. This aspect was investigated through in vitro studies using the FTIR, ELISA, and XTT tests. The characteristics of the included studies are presented in Table [2.](#page-6-0)



# <span id="page-6-0"></span>**Table 2.** Characteristics of the included studies.



# **Table 2.** *Cont.*



**Table 2.** *Cont.*



**Table 2.** *Cont.*

ADM, autologous dental material; BCB, buccal cortical bone; DG, dentin grinder; hFOB; human fetal osteoblastic cell; hPLF, human periodontal ligament fibroblast; MDM, mineralized dentin matrix; TDBS, tooth-derived bone substitute; TT, demineralized dentin. \* Tooth-derived material processed with a dentin grinder marked in bold.

#### 3.2.1. Chemical Composition

Four studies provided insights into the composition of dentin after grinding. The analysis conducted by Khanijou et al. [\[21\]](#page-18-9) showed that tooth-derived material comprised oxygen (50.6%), carbon (22%), calcium (15.7%), phosphate (9.8%), sodium (1.4%), magnesium (0.6%), and a calcium-to-phosphate ratio (1.83), calculated based on the atomic percentage. Sarna-Bos et al. [\[22\]](#page-18-6) conducted a similar analysis of dentin composition, although with a different presentation of the results. Sarna-Bos et al. [\[22\]](#page-18-6), on the basis of icosahedral analysis of the description of functional groups using FTIR spectrometry, showed the presence of phosphate ions, carbonate anions, and amide I and amide II structures, as well as aliphatic and hydroxyl groups. Material obtained from premolars was characterized by a higher content of phosphate ions, carbonate anions, amides, and hydroxyl groups compared to material obtained from incisors and canines. However, a higher content of aliphatic groups was observed in incisors compared to other tooth types. Moreover, an analysis of the elemental composition of dentin by ICP OES and SEM-EDX was performed. The obtained conclusions were significant, as they demonstrated that all four groups of human teeth (incisors, canines, premolars, molars) showed a similar chemical composition. The overall composition included calcium (276.4  $\pm$  9.7 g/kg), magnesium (5.2  $\pm$  0.9 g/kg), sodium (2  $\pm$  0.1 g/kg), phosphate (147.6  $\pm$  4.9 g/kg), and a calcium-to-phosphate ratio of  $1.9 \pm 0.02$ . Dłucik et al. [\[24\]](#page-18-11) checked the elemental composition of crushed particles with three different devices (BonMaker; Korea Denta Solution Co., Ltd., Busan, Republic of Korea, Tooth Transformer, SRL, Milan, Italy, Smart Dentin Grinder, KometaBio Inc., Cresskil, NJ, USA) and compared them to the control dentin. The results obtained from energy-dispersive X-ray analysis showed significant differences. In the dentin specimen, the primary components were carbon, oxygen, phosphorus, and calcium. The notably

high levels of calcium and phosphorus indicated the mineral content. In Smart Dentin Grinder specimens, calcium dominated, constituting over 50% of the elements sampled. Carbon, oxygen, and phosphorus were also present, with percentages similar to those found in dentin. The Raman spectra also differed. The most pronounced band for all devices appeared in the region centered at 957 cm $^{-1}$ , attributed to the inorganic component. However, there were variations between devices in terms of the Raman signal in the wave number region of 2843–3005  $cm^{-1}$ , associated with the organic component. Finally, Del Canto-Díaz et al. [\[27\]](#page-18-10) using electron microscopy confirmed an atomic composition and a calcium-to-phosphate ratio similar to that of bone.

## 3.2.2. Evaluation of Cellular Responses

The evaluation of cellular responses was carried out in two in vitro studies. Bianchi et al. [\[20\]](#page-18-5) evaluated the impact of different dentin derivatives at the cellular level in a study with human periodontal ligament fibroblasts. They reported that mineralized dentin induced significant cellular growth in comparison to deproteinized bovine bone at 7 days. In addition, fibroblasts treated with mineralized dentin exhibited a thickening of the cellular membrane and showed strong vinculin and integrin signals at 72 h, while the actin signal remained constantly expressed during the entire follow-up. In their study, actin, vinculin, and integrin were considered markers, indicating the reorganization of the cytoskeleton. The observed differences in adhesion and proliferation between TT and DDP suggest an important role in the composition of these materials in cellular responses. TT, being a richer source of collagen and bone morphogenetic protein (BMP), exhibited stronger adhesion and proliferation effects compared to DDP, which was devoid of them. The presence of collagen in TT may influence better adhesion of gingival fibroblast cells (hPLFs), as collagen is known to promote cell–substrate interactions. Additionally, the presence of BMP in TT may affect proliferation processes, as BMP is a cell-inducing factor. In another study, Khanijou et al. [\[21\]](#page-18-9) investigated bone morphogenetic protein-2 (BMP2) from graft material. BMP2 belongs to the transforming growth factor beta (TGF-β) superfamily and plays a key role in osteogenesis. This study observed that both tooth-derived material and allograft began releasing BMP2 after graft preparation with a subsequent gradual decline observed until day 10. BMP2 was significantly more abundant in the release from dentin material compared to allograft and was measured at day 5  $(162.81 \pm 9.03 \text{ pg/mL} \text{ vs. } 132.15 \pm 3.85 \text{ pg/mL}; p < 0.05)$  and day 10 (212.53  $\pm$  9.11 pg/mL vs.  $175.75 \pm 2.25$  pg/mL;  $p < 0.05$ ). Nevertheless, there was a lower increase in the migration of human fetal osteoblastic cells when cultured with dentin material compared to allograft (44.4% vs. 59.2%) on day 3.

### *3.3. Morphology*

Dentin material underwent evaluation using various microscopy techniques immediately after grinding. Based on SEM analysis, the researchers consistently observed a comparable size range of dentin particles post-grinding, ranging from  $300 \mu m$  to  $1200 \mu m$  [ $21,23,26-28$  $21,23,26-28$  $21,23,26-28$  $21,23,26-28$ ], 250 to 1200  $\mu$ m [\[14\]](#page-17-13), or 300  $\mu$ m to 1300  $\mu$ m [\[24\]](#page-18-11). Sarna-Bos et al. [\[22\]](#page-18-6) evaluated chemically untreated dentin particles using scanning electron microscopy (SEM) and reported that dentin particles appeared crumbled but not crushed. These particles contained cylindrical prisms of enamel, dentin with characteristic tubules, and innervated radicular pulp. The peak maximum for pores was observed at a diameter of about 55 µm. The total porosity did not exceed 29%, and the pore area was  $1.89 \text{ m}^2/\text{g}$ . Based on SEM images, Khanijou et al. [\[21\]](#page-18-9) reported that the crushing force of the dentin grinder opened the dentinal tubules, but their visibility and homogeneity became apparent only after chemical treatment. Before the chemical treatment, dentin particles showed a regular pattern of dentinal tubules with occluded organic contents between them, resulting in an unclear tubular pattern. After chemical treatment, the area surrounding the dentinal tubules exhibited an irregular and coarse surface structure. Although the dentinal tubules seemed more blocked before chemical treatment, the surrounding area appeared smoother. The conclusion drawn was that chemical treatment played a crucial

role in revealing the homogeneity of dentinal tubules and influencing surface characteristics. Binderman et al. [\[15\]](#page-18-0) present the progressive opening of tubuli openings during cleansing and their wide opening at 10 min. Dłucik et al., 2023 [\[24\]](#page-18-11) compared demineralized dentin produced with three different devices (BonMaker, Tooth Transformer, Smart Dentin Grinder). SEM showed important differences in the size and shape of granules. Particles prepared with a Smart Dentin Grinder were larger and had sharp margins. The particles' surfaces were mostly flat, occasionally displaying linear reliefs. At higher magnification, these reliefs exhibited a lamellar shape, featuring dentinal tubules and orifices. Some particles displayed deeper and orderly rows, especially on their lateral surfaces.

In a study by Del Canto-Díaz et al. [\[27\]](#page-18-10), the prismatic and irregular appearance of dentin fragments was observed using an optical microscope.

#### *3.4. Histomorphometric Outcomes*

The study carried out by Cervera-Maillo et al. [\[23\]](#page-18-7) selected 10 healthy patients for the treatment of their atrophic edentulous mandible. After their teeth were ground, particulate dentin was placed in empty sockets and bone defects after teeth extractions. Histomorphometric analysis showed the continuous formation of new bone (16.3 at 3 months vs. 59.4 at 12 months), a decrease in residual bone (37.1 at 3 months vs. 15.6 at 12 months), and a decrease in the amount of connective tissue (46.6 at 3 months vs. 25 at 12 months). Illustratively, initially, the teeth particles were immersed in a developing connective tissue with minimal new bone formation. Subsequently, small particles of dentin became integrated into new, immature bone. Then, a substantial amount of bone formation surrounding the tooth particles became apparent. Finally, a new bone structure with incorporated dentin particles was observed. A randomized controlled trial conducted by Santos et al. [\[14\]](#page-17-13) enrolled 52 patients (66 implants). Primary implant stability at baseline and secondary stability at 2 months after implant placement were similar between mineralized dentin and xenograft. The percentage of newly formed bone was higher for mineralized dentin than for xenograft (47.3% vs. 34.9%;  $p < 0.001$ ), while the percentage of the residual graft was lower for mineralized dentin than for xenograft (12.2% vs. 22.1%; *p* < 0.001).

Four studies examined the histology of bioptats collected during implantation. In the study by Pohl et al. [\[26\]](#page-18-14), the presence of new bone tissue that was in direct contact with dentin particles was noted. Additionally, the presence of osteoblasts was confirmed using specific antibodies. In the study conducted by Binderman et al. [\[15\]](#page-18-0), dentin with its tubules was observed, surrounded by a newly formed bone matrix. In Dłucik et al.'s study [\[28\]](#page-18-12), histological biopsies were taken 4 to 5 months after grafting ground in the alveolar bone in the maxilla. Examination of the images revealed that the majority of dentin fragments were enveloped by newly formed bone, while some fragments remained enclosed by soft connective tissue. Stained images displayed osteocytes within the bone tissue and osteoblasts on its surface, clearly indicating the distinction between newly formed and fully mineralized bone. No signs of inflammation or other pathological processes were noted.

Histometric analysis showed a significant difference in osteoid thickness between the groups. The osteoid thickness was  $8.35 \mu m$  in the Bio-Oss group compared to  $13.12 \mu m$ in the autogenous tooth bone graft (AutoBT) group. This result suggests that there are noticeable changes in bone mineralization between the groups. The reduced osteoid thickness in the Bio-Oss group may indicate a more advanced mineralization process compared to the AutoBT group. In addition, computed tomography (CT) analysis showed no significant differences between the groups in total bone volume, new bone volume, and new bone mineral density. There was, however, a significant difference in trabecular thickness, which was  $0.07 \mu m$  in the Bio-Oss group compared to  $0.08 \mu m$  in the AutoBT group [\[29\]](#page-18-8).

#### *3.5. Bacteriological Purity*

Three studies provided information on bacterial purity. Binderman et al. [\[15\]](#page-18-0) reported that their bacteriological test showed no bacterial growth after a 10 min treatment with the cleanser, while Khanijou et al. [\[21\]](#page-18-9) revealed that the initial colony forming unit (CFU) per 0.1 g was  $2.3 \times 10^6$  before chemical treatment. However, after the chemical treatment, no microorganisms were detected. Dłucik et al. [\[24\]](#page-18-11) confirmed obtaining negative results in microbiological testing.

#### *3.6. Clinical Outcomes*

Clinical outcomes were limited, with a focus on radiographic imaging for evaluating tissue density and in vivo structural changes in the examined bone tissue. Few studies addressed complication rates and patient-reported outcomes.

### 3.6.1. Bone Tissue Regeneration

The study by Pohl et al. [\[26\]](#page-18-14), involving 13 patients, utilized radiographic comparative analysis in order to evaluate dimensional ridge changes 4 months after tooth extraction and immediate grafting with mineralized dentin particulate autograft and chopped platelet-rich fibrin. Extraction sockets with up to 2 mm of missing buccal bone in the coronal aspect were compared to the lingual bone. At the end of the study, cone beam computed tomography revealed an average mid-buccal bone height gain of +1.1% and a mid-lingual height gain of 5.6%.

Binderman et al. [\[15\]](#page-18-0) shared their broad experience in grafting the demineralized dentin matrix; however, they presented two challenging case reports in detail. In case 1, the surgically removed tooth exposed the distal root surface of the neighboring tooth. A follow-up after 4 months revealed a normal pattern of marginal gingiva around the neighboring tooth, with normal probing depths of 1–2 mm. On the X-ray, new bone and dentin were integrated, fully restoring the extraction site and supporting adjacent teeth. In case 2, two teeth were removed due to poor periodontal attachment, increased mobility, and bone loss. Following extraction, granulation tissue was cleared, exposing bone walls, and dentin particles were inserted into sockets and covered with a platelet-rich fibrin membrane. Implants were placed after a 2-month interval. A 2-year follow-up X-ray showed highly radiopaque bone integrated into the implants providing robust support.

Del Canto-Díaz et al. [\[27\]](#page-18-10) evaluated the outcomes of their split-mouth study conducted on nine patients requiring the extraction of two single-rooted teeth and subsequent rehabilitation with osseointegrated implants. The condition of post-extraction sockets after filling with autologous dentin material was compared to this without filling with any material. The dentin graft exhibited a lower mean vertical bone loss (4.2% vs. 16.87%) and a reduced mean horizontal loss of buccal cortical bone at 16 weeks (0.16 mm vs. 2.22 mm; *p* = 0.067) compared to no graft. Additionally, densitometric analysis of the coronal alveolar area indicated higher bone density for the dentin graft compared to no graft (922.68  $\pm$  250.82 HU vs.  $564.35 \pm 288.73$ ;  $p = 0.045$ ).

Matsuzawa et al. [\[25\]](#page-18-13) reported a case involving the repair of a unilateral maxillary alveolar cleft during mixed dentition. At the 6-month follow-up X-ray, the grafted demineralized dentin was identifiable as radiopaque granules, distinguishable from the original bone by both its structure and radiopacity. After 6 years, the graft composed of demineralized dentin from autogenous primary teeth was entirely replaced by bone, without affecting tooth formation or impeding spontaneous tooth eruption.

In the split-mouth case report by De Biase et al. [\[17\]](#page-18-2), the X-ray results indicated less bone loss at the site treated with a demineralized dentin matrix compared to the control site. Specifically, when measuring the distance between the cementoenamel junction (CEJ) and the bone peak, the control site exhibited a reduction of 0.94 mm, whereas in the grafted site, the reduction was 2.32 mm. The clinical probing depth decreased by 1 mm in the grafted site, while no changes were observed in the control site.

Dłucik et al. [\[28\]](#page-18-12) reported the absence of significant bone resorption during the followup period, irrespective of the method employed.

#### 3.6.2. Periprocedural and Long-Term Complications

In a randomized controlled trial conducted by Santos et al. [\[14\]](#page-17-13) with 52 patients, a comparison between mineralized dentin and xenograft resulted in the lack of significant differences in terms of bleeding, marginal bone loss, the amount of keratinized gingival width, and the occurrence of peri-implant mucositis. In a study by Pohl et al. [\[26\]](#page-18-14) involving 13 patients who received mineralized dentin with platelet-rich fibrin to fill post-extraction sockets, there were no observed signs of inflammation or fibrosis formation around the autologous augmentation material. Several other studies reported an uncomplicated healing process and recovery following the surgical procedure and grafting [\[15](#page-18-0)[,25\]](#page-18-13), with one study reporting no clinical and radiographic signs of complications at six months post-procedure [\[17\]](#page-18-2). In all alveolar bone regeneration and sinus lift procedures, Dłucik et al. [\[24\]](#page-18-11) reported the absence of significant complications during the recovery period, ensuring uneventful healing.

### 3.6.3. Patients' Reported Outcomes

Only one study focused on patient-reported outcomes. Among the 52 participants in the study by Santos et al. [\[14\]](#page-17-13), patients reported significantly lower pain experience the next day after the procedure with demineralized dentin in comparison to xenograft (*p* = 0.014). Furthermore, drawing from the aggregated outcomes across groups employing various graft preparation devices, Dłucik et al. [\[28\]](#page-18-12) concluded that none of the patients reported additional pain or discomfort.

### *3.7. Quality Assessment*

Three case reports were evaluated with a checklist. One case report received a maximum score of 8 points [\[25\]](#page-18-13), while two others received 6 points [\[17,](#page-18-2)[25\]](#page-18-13). Only one case series was included [\[27\]](#page-18-10) that received a maximum score of 10 points. Only one cohort study was included [\[23\]](#page-18-7) with 8 points out of 11 possible. Of the two randomized controlled trials, one had a low risk of bias with 11 points out of 13 possible [\[14\]](#page-17-13), while the other had a moderate risk of bias with 7 points out of 13 possible [\[29\]](#page-18-8). The risk of bias in three in vitro studies was not assessed due to the lack of a relevant checklist [\[20–](#page-18-5)[22\]](#page-18-6). Table [3](#page-13-0) presents the scoring for the evaluated studies.



<span id="page-13-0"></span>**Table 3.** Risk of bias evaluation of the included studies.



# **Table 3.** *Cont.*



## **Table 3.** *Cont.*

#### **4. Discussion**

Our systematic review aimed to provide a thorough insight into the potential application of ground dentin grafts for bone tissue regeneration. Commencing with a focus on molecular aspects explored in both in vitro and histomorphometric studies, progressing through experimental models, and culminating in the assessment of clinical outcomes, this review bridges the gap between understanding the underlying biological mechanisms and evaluating clinical effectiveness. Various studies in the literature have demonstrated the significant role of autologous dentin in bone reconstruction [\[14,](#page-17-13)[15\]](#page-18-0). In a randomized controlled trial, Santos et al. [\[14\]](#page-17-13) compared autogenous mineralized dentin to xenograft granules for ridge preservation in delayed implantation. The percentage of newly formed bone within the mineralized dentin matrix was significantly higher than that observed with xenograft  $(47.3\% \text{ vs. } 34.9\%; p < 0.001)$ , with a lower percentage of residual graft  $(12.2\% \text{ in } 10^{-19})$ mineralized dentin matrix vs. 22.1% in xenograft; *p* < 0.001). No significant differences were identified in clinical, radiographic, and patient-related outcomes. The mineralized dentin matrix was prepared using a standardized protocol with a grinder generating particles with diameters between 250 and 1200  $\mu$ m. Although various protocols were employed by other researchers, potentially influencing the results, the outcomes remained promising despite the divergence in dentin biomaterial processing methods [\[15\]](#page-18-0).

Bone regeneration is a crucial aspect of oral and maxillofacial surgery, particularly in procedures like placing dental implants, bone grafting, and the surgical treatment of periodontal diseases [\[30\]](#page-18-15). This process involves the restoration of bone tissue in areas where bone has been lost due to disease, trauma, or tooth extraction [\[30\]](#page-18-15). Advances in regenerative techniques, such as guided bone regeneration (GBR), the use of biomaterials, like bone grafts (autografts, allografts, xenografts), and growth factors, such as platelet-rich plasma (PRP) or bone morphogenetic proteins (BMPs), have significantly enhanced the ability to promote bone healing and regeneration [\[30](#page-18-15)[–33\]](#page-18-16). Among these, autologous dentin grafts, derived from a patient's own extracted teeth, offer unique advantages [\[25](#page-18-13)[,26\]](#page-18-14). These grafts are biocompatible, eliminating the risk of immune rejection, and they closely mimic the natural composition of bone, facilitating more efficient integration and regeneration. Additionally, using a patient's own dentin reduces the need for external donor materials, making the process more cost effective and reducing the risk of disease transmission [\[14](#page-17-13)[,29\]](#page-18-8). These methods not only reconstruct bone in the maxillofacial region but also increase the

success rates of dental implants and other restorative procedures, ultimately leading to better patient chewing, improved aesthetics, and enhanced quality of life.

Nevertheless, it is important to note that despite the use of aseptic techniques during the harvesting and transplantation of bone grafts, the risk of contamination persists. Microbiotas can develop on the material between the time of harvesting and transplantation [\[34\]](#page-18-17). Research by S. Ghanaati et al. [\[35\]](#page-18-18), which analyzed allogenic bone blocks available on the market, found that three out of five materials were contaminated with organic and cellular residues. J. Lorenz et al. [\[36\]](#page-18-19) conducted histological analysis of the Maxgraft material, uncovering cellular debris, intertrabecular fat, connective tissue, and collagen. In a study by T. Fretwurst et al. [\[37\]](#page-18-20), evaluations of allografts from various donors showed that the cleaning techniques used for bone blocks did not fully eliminate contaminants. Further histological examinations by Lorenz J. et al. [\[38\]](#page-19-0) of allogeneic bone blocks implanted in patients revealed the presence of donor cellular remnants; however, these remnants did not adversely affect alveolar process regeneration. These findings suggest that more effective cleaning and decontamination methods for graft materials should be adopted by both manufacturers and clinicians.

Although the number of studies identified by our search is relatively small, the studies present a variety of outcomes. During this decade, laboratory studies, experimental models, and clinical trials have been conducted, providing data on the morphology of ground dentin and the composition of the dentin material and showcasing how the graft integrates with adjacent tissues providing support for implants. Clinical studies used demineralized dentin grafts in the following indications: the regeneration of alveolar bone and the prevention of its loss after extraction [\[14](#page-17-13)[,15](#page-18-0)[,17](#page-18-2)[,23](#page-18-7)[,26,](#page-18-14)[27\]](#page-18-10), the sinus lift procedure [\[24\]](#page-18-11), and unilateral alveolar cleft [\[25\]](#page-18-13). The regeneration of alveolar bone has not been extensively studied in patients with specific pathologies and has primarily been explored in small patient populations. Furthermore, these indications are currently limited to dentistry and maxillofacial surgery, reflecting a natural progression toward using extracted teeth as a material in maxillofacial procedures. Although typically demineralized dentin is not considered a suitable material for skeletal bone regeneration due to the scarcity of autologous grafting material, it holds potential as a source of growth factors for bone regeneration [\[39\]](#page-19-1). It is worth noting that, while there is scientific interest and ongoing research in this area, the use of dentin for regenerating bones other than the alveolar bone is far from being a standard clinical practice. Clinical trials and further research are necessary to determine the safety and efficacy of such approaches before they can be widely adopted.

A literature gap has been identified, revealing a lack of emphasis on periprocedural and long-term complications in the existing studies. Patient-reported outcomes were notably absent, and the evaluation of satisfaction has not been conducted, underscoring the need for further research in these areas. The strength of this review lies in its focus on a specific device. Dłucik et al. [\[24\]](#page-18-11) demonstrated that grinding with various devices yields dentin particles of varying quality in terms of size and shape. Additionally, dentin preparation protocols significantly impact the osteoinductivity and osteoconductivity of dentin [\[40\]](#page-19-2). The limitation of our systematic review was its intentionally focused and narrow scope; however, this approach aimed to cover a sufficient range of study types, providing data on both molecular and clinical aspects, thereby connecting laboratory studies with clinical research. In line with this intention, we also limited the search to papers published in English to avoid potential errors that could arise from misinterpretation due to limited proficiency in other languages. In future research on the use of dentin biomaterial in bone reconstruction, it would be worthwhile to expand the molecular aspect with studies indicating the presence and quantity of pro-inflammatory factors, such as cytokines and chemokines, as well as the hydrophobic enzyme ALP and adhesion proteins, which play a key role in the process of bone regeneration. In addition, the determination of additional growth factors, such as IGF, may also prove important in understanding, at the molecular level, the process of bone regeneration using dentin biomaterial. Additionally,

further research should be conducted to enable the synthesis of the obtained results through meta-analysis.

#### **5. Conclusions**

In conclusion, dentin biomaterial emerges as a novel and versatile bone substitute, demonstrating high biocompatibility and ease of acquisition. The preservation of its internal structure, enriched with substantial organic matter, enhances its potential for effective bone regeneration. Particularly, in dental surgery, materials derived from teeth, with inherent growth factors, present a promising alternative to traditional bone autografts, offering the potential to reduce patient morbidity and treatment costs. Examining the molecular aspects of bone tissue regeneration with ground dentin provides a comprehensive understanding, paving the way for targeted and optimized approaches in the realm of regenerative medicine and maxillofacial surgery.

**Author Contributions:** Conceptualization, C.O., A.O., J.M. and M.D.; methodology, C.O., A.O., J.M. and M.D.; validation, C.O., A.O. and I.Z.; formal analysis, C.O., A.O., I.Z., J.M. and M.D.; data curation, C.O., A.O., I.Z. and J.M.; writing—original draft preparation, C.O., A.O. and I.Z.; writing review and editing, C.O., A.O., J.M. and M.D. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was financed by a subsidy from Wroclaw Medical University, number SUBZ.B180.24.058.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Data supporting the findings of this study are available within the article.

**Conflicts of Interest:** The authors declare no conflicts of interest.

# **References**

- <span id="page-17-0"></span>1. De Santis, R.; Guarino, V.; Ambrosio, L. Composite biomaterials for bone repair. In *Bone Repair Biomaterials*; Woodhead Publishing: Cambridge, UK, 2019; pp. 273–299. [\[CrossRef\]](https://doi.org/10.1016/b978-0-08-102451-5.00010-x)
- <span id="page-17-1"></span>2. Sheikh, Z.; Hamdan, N.; Abdallah, M.-N.; Glogauer, M.; Grynpas, M. Natural and synthetic bone replacement graft materials for dental and maxillofacial applications. In *Advanced Dental Biomaterials*; Woodhead Publishing: Cambridge, UK, 2019; pp. 347–376. [\[CrossRef\]](https://doi.org/10.1016/b978-0-08-102476-8.00015-3)
- <span id="page-17-2"></span>3. Dominiak, M.; Hnitecka, S.; Olchowy, C.; Olchowy, A.; Gedrange, T. Analysis of alveolar ridge width in an area of central lower incisor using cone-beam computed tomography in vivo. *Ann. Anat.* **2021**, *236*, 151699. [\[CrossRef\]](https://doi.org/10.1016/j.aanat.2021.151699)
- <span id="page-17-3"></span>4. Kinaci, A.; Neuhaus, V.; Ring, D.C. Trends in bone graft use in the United States. *Orthopedics* **2014**, *37*, e783–e788. [\[CrossRef\]](https://doi.org/10.3928/01477447-20140825-54)
- <span id="page-17-4"></span>5. Bernardi, S.; Macchiarelli, G.; Bianchi, S. Autologous Materials in Regenerative Dentistry: Harvested Bone, Platelet Concentrates and Dentin Derivates. *Molecules* **2020**, *25*, 5330. [\[CrossRef\]](https://doi.org/10.3390/molecules25225330)
- <span id="page-17-5"></span>6. Barone, A.; Toti, P.; Menchini-Fabris, G.B.; Felice, P.; Marchionni, S.; Covani, U. Early volumetric changes after vertical augmentation of the atrophic posterior mandible with interpositional block graft versus onlay bone graft: A retrospective radiological study. *J. Cranio-Maxillofac. Surg.* **2017**, *45*, 1438–1447. [\[CrossRef\]](https://doi.org/10.1016/j.jcms.2017.01.018)
- <span id="page-17-6"></span>7. Schmidt, A.H. Autologous bone graft: Is it still the gold standard? *Injury* **2021**, *52*, S18–S22. [\[CrossRef\]](https://doi.org/10.1016/j.injury.2021.01.043)
- <span id="page-17-7"></span>8. Ya¸sar Mahsut, D. Bone Graft Types. In *Bone Grafting*; Raja, K., Ed.; IntechOpen: Rijeka, Croatia, 2018; Chapter 3. [\[CrossRef\]](https://doi.org/10.5772/intechopen.79449)
- <span id="page-17-8"></span>9. Archunan, M.W.; Petronis, S. Bone Grafts in Trauma and Orthopaedics. *Cureus* **2021**, *13*, e17705. [\[CrossRef\]](https://doi.org/10.7759/cureus.17705)
- <span id="page-17-9"></span>10. Sohn, H.S.; Oh, J.K. Review of bone graft and bone substitutes with an emphasis on fracture surgeries. *Biomater. Res.* **2019**, *23*, 9. [\[CrossRef\]](https://doi.org/10.1186/s40824-019-0157-y)
- <span id="page-17-10"></span>11. Hinsenkamp, M.; Collard, J.F. Growth factors in orthopaedic surgery: Demineralized bone matrix versus recombinant bone morphogenetic proteins. *Int. Orthop.* **2015**, *39*, 137–147. [\[CrossRef\]](https://doi.org/10.1007/s00264-014-2562-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25338109)
- <span id="page-17-11"></span>12. Murata, M.; Nezu, T.; Takebe, H.; Hirose, Y.; Okubo, N.; Saito, T.; Akazawa, T. Human dentin materials for minimally invasive bone regeneration: Animal studies and clinical cases. *J. Oral Biosci.* **2023**, *65*, 13–18. [\[CrossRef\]](https://doi.org/10.1016/j.job.2022.10.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36336319)
- <span id="page-17-12"></span>13. Pang, K.M.; Um, I.W.; Kim, Y.K.; Woo, J.M.; Kim, S.M.; Lee, J.H. Autogenous demineralized dentin matrix from extracted tooth for the augmentation of alveolar bone defect: A prospective randomized clinical trial in comparison with anorganic bovine bone. *Clin. Oral Implant. Res.* **2017**, *28*, 809–815. [\[CrossRef\]](https://doi.org/10.1111/clr.12885) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27279547)
- <span id="page-17-13"></span>14. Santos, A.; Botelho, J.; Machado, V.; Borrecho, G.; Proença, L.; Mendes, J.J.; Mascarenhas, P.; Alcoforado, G. Autogenous Mineralized Dentin versus Xenograft granules in Ridge Preservation for Delayed Implantation in Post-extraction Sites: A Randomized controlled clinical trial with an 18 months follow-up. *Clin. Oral Implant. Res.* **2021**, *32*, 905–915. [\[CrossRef\]](https://doi.org/10.1111/clr.13765)
- <span id="page-18-0"></span>15. Binderman, I.; Hallel, G.; Nardy, C.; Yaffe, A.; Sapoznikov, L. A Novel Procedure to Process Extracted Teeth for Immediate Grafting of Autogenous Dentin. *J. Interdiscipl Med. Dent. Sci.* **2014**, *7*, 1000154. [\[CrossRef\]](https://doi.org/10.4172/2376-032X.1000154)
- <span id="page-18-1"></span>16. Amir-Behghadami, M.; Janati, A. Population, Intervention, Comparison, Outcomes and Study (PICOS) design as a framework to formulate eligibility criteria in systematic reviews. *Emerg. Med. J.* **2020**, *37*, 387. [\[CrossRef\]](https://doi.org/10.1136/emermed-2020-209567) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32253195)
- <span id="page-18-2"></span>17. De Biase, A.; Mazzucchi, G.; Di Nardo, D.; Lollobrigida, M.; Serafini, G.; Testarelli, L. Prevention of Periodontal Pocket Formation after Mandibular Third Molar Extraction Using Dentin Autologous Graft: A Split Mouth Case Report. *Case Rep. Dent.* **2020**, *2020*, 1762862. [\[CrossRef\]](https://doi.org/10.1155/2020/1762862)
- <span id="page-18-3"></span>18. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *BMJ* **2021**, *372*, n71. [\[CrossRef\]](https://doi.org/10.1136/bmj.n71)
- <span id="page-18-4"></span>19. Critical Appraisal Tools for Use in JBI Systematic Reviews. Joanna Briggs Institute (JBI). Available online: [https://jbi.global/](https://jbi.global/critical-appraisal-tools) [critical-appraisal-tools](https://jbi.global/critical-appraisal-tools) (accessed on 28 February 2024).
- <span id="page-18-5"></span>20. Bianchi, S.; Mancini, L.; Torge, D.; Cristiano, L.; Mattei, A.; Varvara, G.; Macchiarelli, G.; Marchetti, E.; Bernardi, S. Bio-Morphological Reaction of Human Periodontal Ligament Fibroblasts to Different Types of Dentinal Derivates: In Vitro Study. *Int. J. Mol. Sci.* **2021**, *22*, 8681. [\[CrossRef\]](https://doi.org/10.3390/ijms22168681) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34445386)
- <span id="page-18-9"></span>21. Khanijou, M.; Zhang, R.; Boonsiriseth, K.; Srisatjaluk, R.L.; Suphangul, S.; Pairuchvej, V.; Wongsirichat, N.; Seriwatanachai, D. Physicochemical and osteogenic properties of chairside processed tooth derived bone substitute and bone graft materials. *Dent. Mater. J.* **2021**, *40*, 173–183. [\[CrossRef\]](https://doi.org/10.4012/dmj.2019-341)
- <span id="page-18-6"></span>22. Sarna-Boś, K.; Boguta, P.; Skic, K.; Wiącek, D.; Maksymiuk, P.; Sobieszczański, J.; Chałas, R. Physicochemical Properties and Surface Characteristics of Ground Human Teeth. *Molecules* **2022**, *27*, 5852. [\[CrossRef\]](https://doi.org/10.3390/molecules27185852)
- <span id="page-18-7"></span>23. Cervera-Maillo, J.M.; Morales-Schwarz, D.; Morales-Melendez, H.; Mahesh, L.; Calvo-Guirado, J.L. Autologous Tooth Dentin Graft: A Retrospective Study in Humans. *Medicina* **2021**, *58*, 56. [\[CrossRef\]](https://doi.org/10.3390/medicina58010056) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35056364)
- <span id="page-18-11"></span>24. Dłucik, R.; Orzechowska-Wylęgała, B.; Dłucik, D.; Puzzolo, D.; Santoro, G.; Micali, A.; Testagrossa, B.; Acri, G. Comparison of clinical efficacy of three different dentin matrix biomaterials obtained from different devices. *Expert Rev. Med. Devices* **2023**, *20*, 313–327. [\[CrossRef\]](https://doi.org/10.1080/17434440.2023.2190512)
- <span id="page-18-13"></span>25. Matsuzawa, Y.; Okubo, N.; Tanaka, S.; Kashiwazaki, H.; Kitagawa, Y.; Ohiro, Y.; Mikoya, T.; Akazawa, T.; Murata, M. Primary Teeth-Derived Demineralized Dentin Matrix Autograft for Unilateral Maxillary Alveolar Cleft during Mixed Dentition. *J. Funct. Biomater.* **2022**, *13*, 153. [\[CrossRef\]](https://doi.org/10.3390/jfb13030153)
- <span id="page-18-14"></span>26. Pohl, S.; Binderman, I.; Tomac, J. Maintenance of Alveolar Ridge Dimensions Utilizing an Extracted Tooth Dentin Particulate Autograft and PlateletRich Fibrin: A Retrospective Radiographic ConeBeam Computed Tomography Study. *Materials* **2020**, *13*, 1083. [\[CrossRef\]](https://doi.org/10.3390/ma13051083)
- <span id="page-18-10"></span>27. Del Canto-Díaz, A.; de Elío-Oliveros, J.; Del Canto-Díaz, M.; Alobera-Gracia, M.A.; Del Canto-Pingarrón, M.; Martínez-González, J.M. Use of autologous tooth-derived graft material in the post-extraction dental socket. Pilot study. *Med. Oral Patol. Oral Cir. Bucal* **2019**, *24*, e53–e60. [\[CrossRef\]](https://doi.org/10.4317/medoral.22536)
- <span id="page-18-12"></span>28. Dłucik, R.; Orzechowska-Wylęgała, B.; Dłucik, D.; Bogus, K. Histological examination of tooth-derived biomaterials obtained from different devices. *Expert. Rev. Med. Devices* **2023**, *20*, 979–988. [\[CrossRef\]](https://doi.org/10.1080/17434440.2023.2251891) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37639725)
- <span id="page-18-8"></span>29. Jun, S.H.; Ahn, J.S.; Lee, J.I.; Ahn, K.J.; Yun, P.Y.; Kim, Y.K. A prospective study on the effectiveness of newly developed autogenous tooth bone graft material for sinus bone graft procedure. *J. Adv. Prosthodont.* **2014**, *6*, 528–538. [\[CrossRef\]](https://doi.org/10.4047/jap.2014.6.6.528)
- <span id="page-18-15"></span>30. Zwittnig, K.; Mukaddam, K.; Vegh, D.; Herber, V.; Jakse, N.; Schlenke, P.; Zrnc, T.A.; Payer, M. Platelet-rich fibrin in oral surgery and implantology: A narrative review. *Transfus. Med. Hemotherapy* **2023**, *50*, 348–359. [\[CrossRef\]](https://doi.org/10.1159/000527526)
- 31. Funato, A.; Moroi, H.; Ogawa, T. Guided bone regeneration assisted by tooth roots with periodontal ligament: Case reports of immediate and staged approaches to implant therapy. *Int. J. Esthet. Dent.* **2022**, *17*, 276–291.
- 32. Sun, J.; Jiang, X.; Luo, J.; Zhao, L.; Xu, Z.; Xiao, W. Effect of platelet-derived growth factor (PDGF-BB) and bone morphogenic protein 2 (BMP-2) transfection of rBMSCs compounded with platelet-rich plasma on adipogenic differentiation. *Braz. J. Med. Biol. Res.* **2020**, *54*, e9944. [\[CrossRef\]](https://doi.org/10.1590/1414-431x20209944)
- <span id="page-18-16"></span>33. Kim, B.J.; Kim, S.K.; Lee, J.H. Bone regeneration of demineralized dentin matrix with platelet-rich fibrin and recombinant human bone morphogenetic protein-2 on the bone defects in rabbit calvaria. *Maxillofac. Plast. Reconstr. Surg.* **2021**, *43*, 34. [\[CrossRef\]](https://doi.org/10.1186/s40902-021-00320-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34499280)
- <span id="page-18-17"></span>34. Baseri, N.; Meysamie, A.; Campanile, F.; Hamidieh, A.A.; Jafarian, A. Bacterial Contamination of Bone Allografts in the Tissue Banks: A Systematic Review and Meta-Analysis. *J. Hosp. Infect.* **2022**, *123*, 156–173. [\[CrossRef\]](https://doi.org/10.1016/j.jhin.2021.10.020) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34752801)
- <span id="page-18-18"></span>35. Ghanaati, S.; Barbeck, M.; Booms, P.; Lorenz, J.; Kirkpatrick, C.J.; Sader, R.A. Potential Lack of "Standardized" Processing Techniques for Production of Allogeneic and Xenogeneic Bone Blocks for Application in Humans. *Acta Biomater.* **2014**, *10*, 3557–3562. [\[CrossRef\]](https://doi.org/10.1016/j.actbio.2014.04.017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24769111)
- <span id="page-18-19"></span>36. Lorenz, J.; Schlee, M.; Al-Maawi, S.; Chia, P.; Sader, R.A.; Ghanaati, S. Variant Purification of an Allogeneic Bone Block. *Acta Stomatol. Croat.* **2017**, *51*, 141–147. [\[CrossRef\]](https://doi.org/10.15644/asc51/2/7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28827851)
- <span id="page-18-20"></span>37. Fretwurst, T.; Spanou, A.; Nelson, K.; Wein, M.; Steinberg, T.; Stricker, A. Comparison of Four Different Allogeneic Bone Grafts for Alveolar Ridge Reconstruction: A Preliminary Histologic and Biochemical Analysis. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol.* **2014**, *118*, 424–431. [\[CrossRef\]](https://doi.org/10.1016/j.oooo.2014.05.020)
- <span id="page-19-0"></span>38. Lorenz, J.; Kubesch, A.; Al-Maawi, S.; Schwarz, F.; Sader, R.A.; Schlee, M.; Ghanaati, S. Allogeneic Bone Block for Challenging Augmentation—A Clinical, Histological, and Histomorphometrical Investigation of Tissue Reaction and New Bone Formation. *Clin. Oral Investig.* **2018**, *22*, 3159–3169. [\[CrossRef\]](https://doi.org/10.1007/s00784-018-2407-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29524026)
- <span id="page-19-1"></span>39. Minetti, E.; Palermo, A.; Malcangi, G.; Inchingolo, A.D.; Mancini, A.; Dipalma, G.; Inchingolo, F.; Patano, A.; Inchingolo, A.M. Dentin, Dentin Graft, and Bone Graft: Microscopic and Spectroscopic Analysis. *J. Funct. Biomater.* **2023**, *14*, 272. [\[CrossRef\]](https://doi.org/10.3390/jfb14050272)
- <span id="page-19-2"></span>40. Tabatabaei, F.S.; Tatari, S.; Samadi, R.; Moharamzadeh, K. Different methods of dentin processing for application in bone tissue engineering: A systematic review. *J. Biomed. Mater. Res. A* **2016**, *104*, 2616–2627. [\[CrossRef\]](https://doi.org/10.1002/jbm.a.35790)

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.