

## Echoviruses Bind Heparan Sulfate at the Cell Surface

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**Some echoviruses (EV) that bind decay-accelerating factor (DAF) also bind cells of human and murine origins in a DAF-independent manner. Pretreatment of cells with heparinase 1 or heparin blocks the binding of radiolabeled virus to the cell surface, and heparin prevents infection of rhabdomyosarcoma cells by certain EV, including several low-passage clinical isolates of EV 6 and some EV that do not bind DAF. These studies suggest that heparan sulfate may be of in vivo relevance as an attachment molecule for EV.**

Hemagglutination by picornaviruses of the genus *Enterovirus* is mediated by binding to decay-accelerating factor (DAF; CD55), a 70-kDa glycosylphosphatidylinositol-linked glycoprotein that is expressed on serum-exposed cells and that is involved in complement regulation and cell signaling (1, 14, 20, 22, 26, 30). The affinity of echovirus (EV) 11 (EV11) for DAF has previously been determined (15), although hemagglutination inhibition assays and the use of soluble DAF (sDAF) to block infection of rhabdomyosarcoma (RD) cells have suggested that DAF-binding enteroviruses exhibit a range of affinities for this receptor (20). Further studies also have suggested that certain enteroviruses require additional cell surface proteins for infection (2, 21, 27, 28).

It has been demonstrated that EV6, derived from the D'Amori type strain by serial passaging in RD cells, is a weakly hemagglutinating, DAF-binding virus (20). Cell infection required cell surface DAF and was blocked by 15  $\mu$ M sDAF, and a specific interaction with DAF was demonstrated by surface plasmon resonance (22). In contrast to a HeLa cell-passaged D'Amori-derived DAF-binding isolate reported by Bergelson et al. (1), which was found not to bind Chinese hamster ovary (CHO) cells, our EV6 strain bound murine NIH 3T3, CHO, and RD cells in a DAF-independent manner (20; R. M. Powell and D. J. Evans, unpublished results). This result implied that it might bind a widely expressed molecule conserved between humans and rodents. We considered the glycosaminoglycan (GAG) heparan sulfate (HS) a likely candidate, since it is present on the surface of most cells and is known to be bound by a number of viruses (5, 17, 29, 31), including the picornavirus foot-and-mouth disease virus (FMDV) (13).

We investigated whether EV6 interacted with HS by quantifying the binding of <sup>35</sup>S metabolically labeled EV6 particles (prepared and used as previously described [20]) to CHO *pgsA-745* cell lines, which are deficient in GAG synthesis (25) (Fig. 1). EV6 was bound for 2 h at 4°C, and unbound virus was removed by washing with cold serum-free Dulbecco modified Eagle medium (DMEM). Next, cells were subjected to quantification by scintillation counting and comparison with simi-

larly treated cells exposed to radiolabeled EV7, the binding of which is known to be DAF dependent (21, 30). As expected, EV7 bound RD cells but bound neither NIH 3T3 cells in the absence of DAF nor CHO cells (which do not express DAF). EV6 displayed a distinctly different cell interaction profile, binding NIH 3T3 cells in a DAF-independent manner (as previously shown [21]) and GAG-positive CHO cells but displaying only background levels of binding to CHO *pgsA-745* cells (Fig. 1). This analysis indicated that EV6 binding to CHO cells and, by inference, to NIH 3T3 cells might involve GAGs.

To further investigate a potential role for HS in EV attachment, we determined the effect of heparin—a highly sulfated analogue of cell surface HS—on virus binding to or infection of RD cells (Fig. 2). Heparin (porcine mucosal intestinal; Sigma) or de-*N*-sulfated heparin (Sigma) was prediluted in serum-free DMEM and mixed with 1,000 50% tissue culture infective doses (TCID<sub>50</sub>) of virus at room temperature for 15 min. Infection was monitored by adding the virus-heparin mixture to an 80% confluent layer of RD cells in a 96-well plate and incubating the plate for 24 h at 37°C in 5% CO<sub>2</sub> prior to staining with crystal violet. The binding of radiolabeled viruses in the presence of heparin was determined as described above. The range of EV6 strains tested was extended to include several clinical isolates (kind gifts from Brian Megson, Public Health Laboratory Service, Colindale, London, England) that had been passaged no more than twice in RD cells since isolation. The Hill strain of EV9, a receptor for which has yet to be identified, and poliovirus type 3 (PV3) were also included in these assays.

Radiolabeled EV6 binding to RD cells was reduced by 95% in the presence of 1 mg of heparin/ml (Fig. 2a), a result consistent with the observed block in infection observed with heparin concentrations above 0.5 mg/ml (Fig. 2b). In contrast, binding (Fig. 2a) or infection (Fig. 2b) by EV7, PV3, or EV6 clinical isolate #591 was unaffected by the presence of heparin at concentrations of up to 2 mg/ml. EV9 and the other EV6 clinical isolates displayed intermediate phenotypes. Binding by EV6 #21/97 and #488 and by EV9 was reduced by 50 to 75% in the presence of 1 mg of heparin/ml (Fig. 2a). For these clinical isolates, RD cell infection by #21/97 and #488 was more sensitive to inhibition by heparin, whereas the remaining clinical isolates were inhibited by heparin levels comparable to

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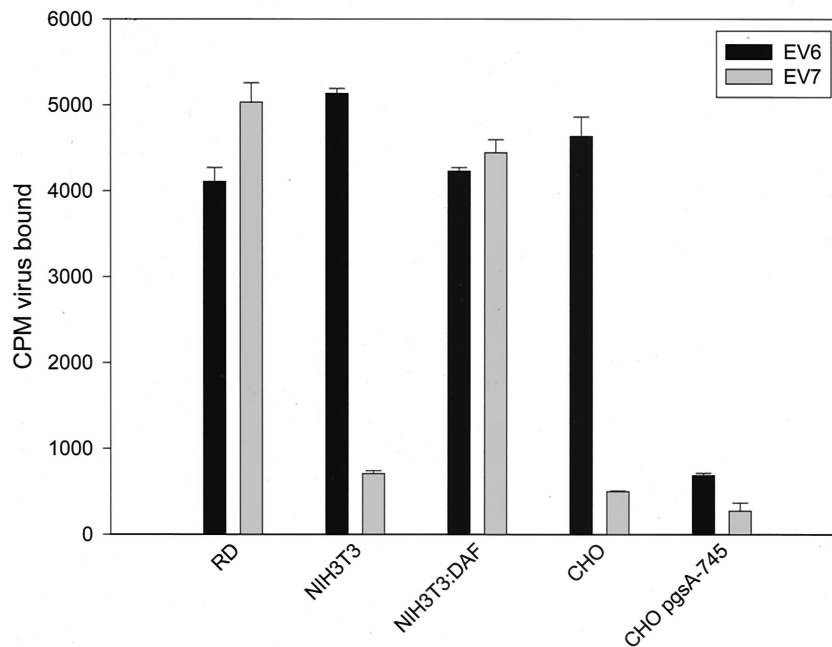


FIG. 1. Involvement of GAGs in the DAF-independent binding of EV6 to rodent cells.  $^{35}\text{S}$ -Cys-Met metabolically labeled EV6 or EV7 (10,000 counts) was bound to the indicated cell lines for 2 h at 4°C, unbound virus was removed by washing with cold serum-free DMEM, and retained virus was quantified by scintillation counting. The mean and standard deviation for three independent experiments are shown.

those that blocked wild-type EV6 infection (Fig. 2b). In all cases, de-*N*-sulfated heparin failed to block infection when added at 2 mg/ml, implying that the sulfation state of the heparin was critical for the observed block, as seen for other HS-binding viruses (3).

The DAF-binding phenotype of the viruses was confirmed by determining whether sDAF blocked infection of RD cells, as previously described (21). RD cell infection by 10,000 TCID<sub>50</sub> of wild-type EV6 and the EV6 clinical isolates was blocked by 15 μM sDAF, whereas 3.25 μM sDAF was required to block infection by EV7 (data not shown). EV9 and PV3 do not bind DAF and were unaffected by the presence of sDAF in this assay.

The enzymatic removal of HS by heparinase 1 and glycosylphosphatidylinositol-anchored proteins (including DAF) by phospholipase C (PIPLC) was used to further confirm a role for HS in EV binding and to quantify the relative contributions of HS and DAF to virus binding. RD cells were pretreated with heparinase 1 (Sigma) at 20 U/3.5 × 10<sup>7</sup> cells for 2 h at 31°C or PIPLC (used as described previously [6]). Cells were then tested for their ability to bind  $^{35}\text{S}$ -labeled virus particles as described previously (Fig. 3). Binding of the control PV3, which binds the transmembrane-anchored poliovirus receptor (16), was not reduced by PIPLC treatment, as expected; in addition, consistent with the data presented in Fig. 2, the binding of PV3 was not reduced by heparinase pretreatment. EV7 was similarly unaffected by heparinase treatment, but binding was significantly reduced following pretreatment with PIPLC, as expected for a DAF-binding virus. EV6 binding was reduced by over 70% with heparinase pretreatment but only slightly by PIPLC. These results reflect the weak and transient DAF-binding phenotype of this virus. The EV6 clinical isolate

#591 was almost indistinguishable from EV7 in this assay; binding was DAF dependent and was reduced only marginally after the heparinase cleavage of HS, in agreement with the failure of heparin to block infection or binding to RD cells (Fig. 2). The remaining clinical isolates tested were intermediate in phenotype, being affected by both enzymatic treatments. EV9 binding was PIPLC resistant, as would be expected for a virus that does not bind DAF, and was reduced only slightly in the presence of heparinase. Under the conditions used, both heparinase and PIPLC were less efficient at blocking binding than heparin (Fig. 2a) or sDAF (data not shown). We ascribe this result to the incomplete removal of HS or DAF from the cell surface or the recycling of HS or DAF proteins in cells at the nearly physiological temperatures used in these assays.

These studies demonstrated that the genus *Enterovirus* of the family *Picornaviridae* contains representatives that bind the cell surface GAG HS. We extended these studies to determine the concentrations of heparin required to block RD cell infection by 1,000 TCID<sub>50</sub> of a range of enteroviruses (Table 1). This experiment demonstrated that the phenotype is widespread within the human enterovirus B species and that there is no correlation with the ability of the virus to bind DAF. HS binding by FMDV is likely to be an adaptation to cell cultures (12, 13, 19). This is probably not the case for HS-binding EV; the majority of the isolates tested in the present study have been serially passaged in RD cells, and low-passage clinical isolates of EV6 can also exhibit this phenotype (Fig. 2a).

These observations suggest that HS binding by EV may be of in vivo relevance. It is possible that the ability to bind HS provides an additional means of cell association (as outlined in reference 10) that facilitates an interaction with the cell surface

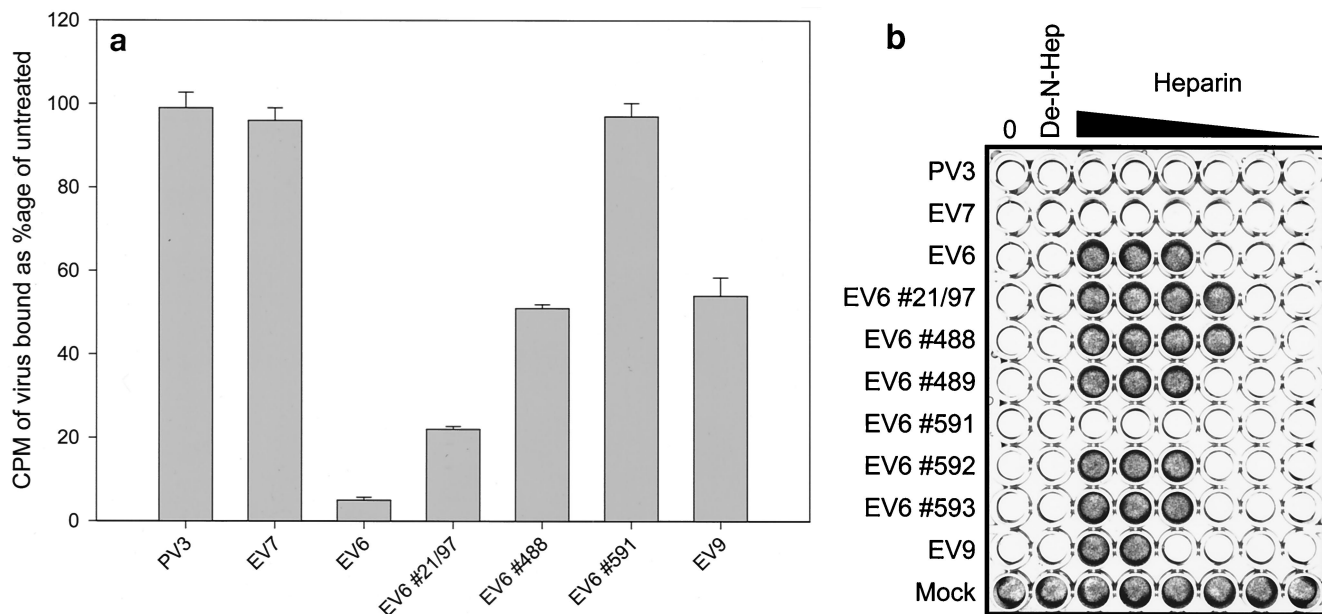


FIG. 2. Heparin inhibits the binding (a) and infection (b) of RD cells by EV6. (a) Binding of radiolabeled EV6, clinical isolates of EV6 (indicated by the number symbol), EV7, EV9, and PV3 to RD cells was monitored in the presence of 1 mg of heparin/ml. Results are presented as the percentage of virus bound in comparison to binding of the untreated virus control (mean and standard deviation). (b) The inhibitory effect of heparin on infection of RD cells was determined by preincubating 1,000 TCID<sub>50</sub> of the indicated viruses with dilutions of heparin (doubling dilutions from 2 mg/ml to 62.5 ng/ml) for 15 min before addition to 80% confluent RD cells. Infection was determined by staining with crystal violet after 24 h. The row labeled “Mock” indicates the effect of heparin in the absence of virus; lanes labeled 0 and De-N-Hep contained no heparin or 1 mg of de-N-sulfated heparin/ml, respectively.

molecules that mediate virus entry. This situation resembles that seen for herpes simplex virus type 1, human immunodeficiency virus type 1, dengue virus, and adeno-associated virus (11, 17, 18, 23, 24). Characterized HS-binding motifs contain a

small number of predominantly basic residues, and structural analysis of the FMDV-HS interface indicates that a limited number of electrostatic interactions are critical for binding (4, 7–9). This information suggests that the genotypic plasticity of

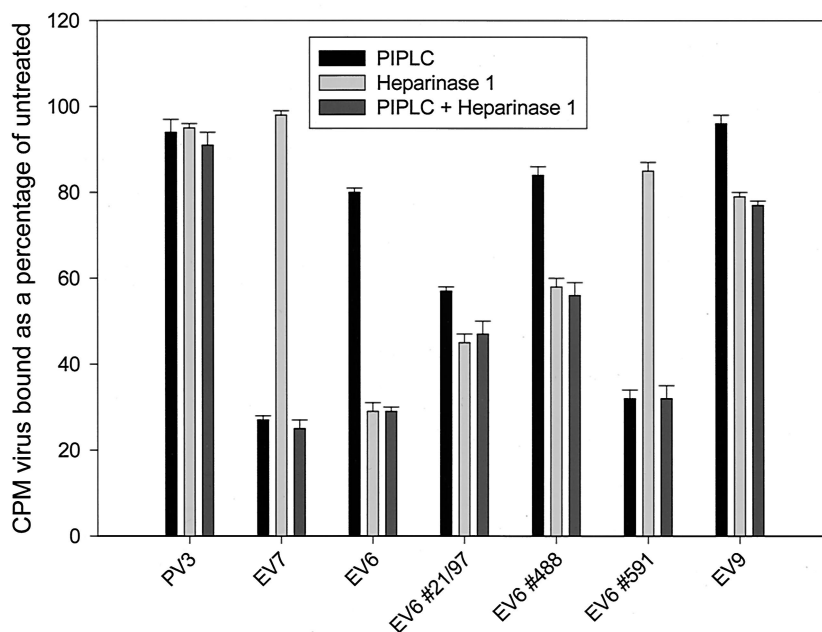


FIG. 3. Relative effects of heparinase 1 and PIPLC on enterovirus binding to RD cells. Metabolically labeled, gradient-purified virus particles (10,000 counts) were bound to RD cells following pretreatment of the cell monolayer with heparinase 1 or PIPLC. The results presented are the mean and standard deviation for three independent assays and are expressed as the percentage bound in comparison to binding to untreated RD cells.

TABLE 1. Heparin concentrations required to block infection

| Viruses <sup>a</sup>   | Heparin ( $\mu\text{g/ml}$ ) required to block 1,000 TCID <sub>50</sub> |
|--|---|
| 9, 25  | 125   |
| 2, 3, 5, 6, 15   | 500   |
| 30   | 1,000   |
| 19, 26   | 2,000   |
| 1, 4, 7, 12, 13, 14, 16, 20, 24, 27, 29, 31, 32, 33, PV3, CBV2, CBV3 | — <sup>b</sup>  |

<sup>a</sup> EV, unless otherwise indicated. CBV2 and CBV3, coxsackie B2 and B3 viruses, respectively. Boldfacing indicates DAF-binding viruses.

<sup>b</sup> —, Insensitive to heparin at 2 mg/ml.

picornaviruses could lead to the acquisition or loss of an HS-binding phenotype relatively rapidly. The *in vivo* relevance of HS binding remains to be determined and will require the further analysis of clinical virus isolates. Studies are under way to investigate the adaptation to HS binding of EV6 isolates such as #591 to provide a better molecular understanding of this phenotype.

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