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Robust differential gene expression patterns in the prefrontal cortex of male mice exposed to an occupationally relevant dose of laboratory-generated wildfire smoke

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Abstract

Wildfires have become common global phenomena concurrent with warmer and drier climates and are now major contributors to ambient air pollution worldwide. Exposure to wildfire smoke has been classically associated with adverse cardiopulmonary health outcomes, especially in vulnerable populations. Recent work has expanded our understanding of wildfire smoke toxicology to include effects on the central nervous system and reproductive function; however, the neurotoxic profile of this toxicant remains illexplored in an occupational context. Here, we sought to address this by using RNA sequencing to examine transcriptomic signatures in the prefrontal cortex of male mice modeling career wildland firefighter smoke exposure. We report robust changes in gene expression profiles between smoke-exposed samples and filtered air controls, evidenced by 2,862 differentially expressed genes (51.2% increased). We further characterized the functional relevance of these genes highlighting enriched pathways related to synaptic transmission, neuroplasticity, blood–brain barrier integrity, and neurotransmitter metabolism. Additionally, we identified possible contributors to these alterations through protein–protein interaction network mapping, which revealed a central node at ß-catenin and secondary hubs centered around mitochondrial oxidases, the Wnt signaling pathway, and gene expression machinery. The data reported here will serve as the foundation for future experiments aiming to characterize the phenotypic effects and mechanistic underpinnings of occupational wildfire smoke neurotoxicology.

Keywords: wildfire smoke; wildland firefighter; occupational health; transcriptomics

The wildfire landscape is rapidly changing across the globe with fires burning more intensely and for longer periods of time throughout the year, a trend which is expected to continue with warmer and drier climates [\(Higuera et al. 2023](#page-9-0); [Turco et al. 2023](#page-10-0)). In the United States, this has significantly increased the contribution of wildfire smoke to ambient air pollution in recent years ([Childs et al. 2022\)](#page-8-0). Although wildfire smoke is a complex mixture of chemical constituents that differ based on conditions surrounding each burn, it consistently comprised the largest percent of mass by volume particulate matter which is 2.5 microns or smaller in aerodynamic diameter (P $M_{2.5}$) [\(Kim et al. 2018\)](#page-9-0). This size fraction of air pollution particulate matter is especially important in a toxicologic context due to the fact that its exposure has been correlated with many adverse human health outcomes ([Reid et al. 2016](#page-10-0); [Reid and Maestas 2019](#page-10-0); [Thangavel et al.](#page-10-0) [2022\)](#page-10-0). Importantly, recent work has reported greater respiratory hospitalization incidence in individuals exposed to wildfire smoke $PM_{2.5}$ when compared with air pollution from other sources [\(Aguilera et al. 2021\)](#page-8-0). In addition to aberrancies in the cardiopulmonary system, wildfire smoke exposure has been

increasingly associated with cognitive decline and reproductive dysfunction [\(Rubin et al. 2021](#page-10-0); [Cleland et al. 2022;](#page-8-0) [Wen and](#page-10-0) [Burke 2022\)](#page-10-0). Still, these outcomes remain under-investigated in populations of individuals exposed to higher concentrations of wildfire smoke in an occupational setting (i.e. wildland firefighters).

Wildland firefighters are subjected to extreme and adverse work conditions throughout their shifts while performing diverse tasks ([Adetona et al. 2017;](#page-8-0) [Navarro et al. 2021a,](#page-9-0) [2021b;](#page-9-0) [Ruby et al.](#page-10-0) [2023](#page-10-0)). These exposures have been associated with negative acute cardiac and respiratory health outcomes, impaired kidney performance, and hearing loss [\(Adetona et al. 2016;](#page-8-0) [Navarro et al.](#page-9-0) [2019](#page-9-0); [Wu et al. 2021\)](#page-10-0); however, emphasis on long-term disease risk has recently been identified as a major concern in this population, especially in regard to the central nervous system (CNS) [\(Pelletier et al. 2022](#page-9-0)). Our group has previously reviewed the literature linking wildfire smoke exposure to Alzheimer's disease (AD) dementia ([Schuller and Montrose 2020\)](#page-10-0). Further, concentrated ambient wildfire smoke exposure in mice, mimicking a public health-relevant dose, has recently been shown to result in

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Methods Animals

Ethics statement

All experiments performed using animal models were conducted in accordance with NIH guidelines under protocols approved by the Institutional Animal Care and Use Committee (IACUC) at Northeastern University. Eight-wk-old male Apoe−/− mice, bred on a C57BL/6 background, were obtained from Jackson Laboratory (Bar Harbor, ME, United States). Apoe−/− mice represent an environmentally susceptible strain due to their impaired lipid metabolism and propensity to develop atherosclerosis ([Yamashita et al. 2014\)](#page-10-0). This strain has also been utilized to characterize the infiltration of ultrafine particulate matter into the CNS ([Peters et al. 2006](#page-9-0)). Importantly, even at aged time points these mice do not exhibit neuro-pathology consistent with AD dementia ([Ganor et al. 2018](#page-8-0)). All mice were housed in clear plastic cages, in groups of up to 5, with ad libitum food and water. Cages were housed in a temperature-controlled vivarium on a 12 h light/dark cycle.

Experimental design

Eight-wk-old male Apoe^{$-/-$} mice were randomly assigned to one of two groups: Smoke-exposed or filtered air control $(n=8$ mice/ group). Mice were exposed to filtered air or simulated wildfire smoke in a 3D printed exposure chamber, which has been experimentally validated to evenly distribute particulate matter [\(Eden](#page-8-0) [et al. 2023](#page-8-0)), for 2 h/d, 5 d/wk, for 16 wk. Simulated wildfire smoke was generated using a quartz-tube and ring furnace setup mounted on a linear actuator, as previously described ([Garg et al.](#page-8-0) [2021](#page-8-0)). Target smoke concentration was set to $40 \,\mathrm{mg/m^3}$ and maintained by diluting the simulated smoke with filtered indoor air. This dose and duration of exposure were determined to be occupationally relevant based on calculations of deposited particulate mass in the lungs of representative mice and men, normalized by lung surface area in the two species, as described previously ([Eden et al. 2023\)](#page-8-0). The average total PM concentration across the duration of the simulated smoke exposure was $39 \pm$ 13 mg/m³ and the average CO concentration across the 16 wk was 218 ± 46 ppm. Extensive characterization of the toxicologic profile of this smoke has been previously reported ([Garg et al.](#page-8-0) [2021](#page-8-0); [Eden et al. 2023\)](#page-8-0). All mice were monitored for signs of overt toxicity and all mice survived for the duration of the study with a final body mass percent change of $24 \pm 1\%$ for the filtered air controls and $19 \pm 1\%$ for the smoke-exposed animals across the 16wk study. Carboxyhemoglobin levels were measured across the study and are reported to be $1.7 \pm 0.9\%$ for the filtered air control group and $21.2 \pm 4.8\%$ for the smoke-exposed group. Mice were sacrificed within 48h of the final day of exposure at Northeastern University and tissue was dissected, flash frozen, and shipped to Boise State University for storage at –80°C (*n* = 8 mice/group).

RNA sequencing

Library preparation and sequencing

Approximately 30 to 35 mg of microdissected PFC per sample $(n=8$ mice/group) were RNA extracted and sequenced by technicians at Novogene (Beijing, China). Samples were subjected to quality control via nanodrop spectroscopy and agarose gel electrophoresis. Subsequently, mRNA was enriched using oligo(dT) beads, fragmented, and converted to cDNA using random hexamer primers via RT–PCR following a standard protocol. Library quality control was assessed using a Qubit fluorometer and libraries were diluted to 1 ng/µl concentration before insert size distribution was checked using Agilent 2100. Clustered libraries were sequenced using the Illumina HiSeq platform generating 150 bp paired-end reads.

Analysis workflow

Raw data (�54,750,000 reads/sample) were cleaned using fastp software by removing reads with adapter contamination, greater than 10% uncertain nucleotides, or base quality *<*5 for more than 50% of the read. Clean reads $(\sim$ 54,500,000/sample) were aligned to the mouse reference genome using Hisat2 v2.0.5 (96.7% total mapping efficiency) and assembled by StringTie v1.3.3b. Fragments per kilobase per million base pairs sequenced (FPKM) were calculated using featureCounts v1.5.0-p3. Gene expression analysis of smoke-exposed versus filtered air control groups was performed using the edgeR R package (v3.22.5) through one scaling normalized factor. Thresholds of false discovery rate (FDR) *<*0.05 and log2foldchange were used to determine differentially expressed genes (DEGs). DEGs were then analyzed for Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG), and Reactome pathways using the clusterProfiler R package, correcting for gene length bias. Significance was determined by FDR *<*0.05 when comparing between exposure groups. GO pathways enriched for biological process (BP) or cellular component (CC) classifications between groups were subsequently visualized using REVIGO ([Supek et al. 2011](#page-10-0)). Systems-level visualization of DEGs was accomplished via the construction of protein–protein interaction (PPI) networks via the STRING database plug-in for Cytoscape, including known and predicted interactions. To visualize these networks, we applied the Fruchterman– Reingold force-directed layout in Gephi (v0.10.1). Functional modules were further defined $(n = 3$ clusters) using the K means algorithm, with the top GO BP term being used to classify each cluster (FDR*<*0.05) via iDEP.96 ([Ge et al. 2018\)](#page-8-0).

Results

Overall transcriptional profile of wildfire smokeexposed and filtered air control samples

To begin to identify the effects of wildfire smoke exposure in the PFC of male mice, we utilized RNA-seq to compare gene expression profiles of smoke-exposed animals relative to filtered air controls ($n = 8$ /group). Across all samples, 93.75% of reads were found to be in exon regions which mapped to a total of 13,781 genes. Two hundred eighty-seven (2.1%) of those genes were found to be unique to wildfire smoke samples, whereas 227 (1.6%) genes were unique to filtered air controls [\(Fig. 1A](#page-2-0)). In order to assess variance across the sample population, we generated a PCA plot using the gene expression value (FPKM) of all samples ([Fig. 1B](#page-2-0)) which demonstrated two distinct clusters, one encompassing the wildfire smoke treated samples and the other containing the filtered air controls. DEG analysis revealed that 2,862 genes were differentially expressed between groups, with 1,396

Fig. 1. A) Venn diagram demonstrating overlap in the genes mapped to reads for each treatment group. Two hundred twenty-seven and 287 genes were found to be uniquely enriched in filtered air control or wildfire smoke-exposed cohorts, respectively. B) PCA plot comparing the variance of principal components 1 and 2 between overall gene expression profiles of all samples across both experimental groups. PCA analysis revealed distinct clustering by group. C) Bar graph highlighting the differentially expressed genes (DEGs) between smoke-exposed and filtered air control groups identified using edgeR (FDR*<*0.05 and log2foldchange). One thousand three hundred ninety-six genes were found to be decreased and 1,466 were found to be increased between groups. D) Heatmap generated by unsupervised clustering of genes with the highest variation across the dataset. Transcriptional profile of smoke-exposed samples is distinct when compared with filtered air control samples.

genes/transcripts increased and 1,466 decreased in wildfire smoke-exposed samples compared with filtered air controls (Fig. 1C). We additionally performed a hierarchical clustering analysis of the FPKM values of DEGs across the dataset to examine the effects of smoke exposure on transcriptional patterns (Fig. 1D) which demonstrated distinct clustering of samples by exposure group and broad distribution of DEGs across the genome.

To further characterize the effects of wildfire smoke exposure on specific genes across the transcriptome, we identified top DEGs across the dataset. [Figure 2A](#page-3-0) shows a volcano plot with all mapped genes across the dataset. DEGs were identified as genes with log2foldchange and FDR*<*0.05. Selected top hits were binned by increased or decreased expression and strength of log2foldchange or FDR ([Fig. 2B\)](#page-3-0). We also represent the FPKM of each of the top 5 strongest (by FDR) increased and decreased genes/transcripts ([Fig. 2C\)](#page-3-0). The most strongly increased genes included:

Vinculin (*Vcl*), natural killer tumor recognition sequence (*Nktr*), suppressor of Ty 16 (*Supt16*), a disintegrin-like and metallopeptidase (resprolysin type) with thrombospondin type 1 motif 3 (*Adamts3*), and protocadherin gamma subfamily C 3 (*Pcdhgc3*). Conversely, the most strongly decreased genes included: Pregnancy specific glycoprotein 16 (*Psg16*), ß-1,3-galactosaminyltransferase polypeptide 1 (*B3galnt1*), apolipoprotein O (*Apoo*), predicted gene 35161 (*Gm35161*), and sperm associated antigen 5 (*Spag5*). The relevance of these genes to the PFC and CNS disease states will be further explored in the discussion.

Functional enrichment analysis

In order to better understand the functional trends of DEGs across the dataset, we employed multiple pathways analyses. Firstly, GO terms were assessed across the entire set of 2,862 DEGs. Pathways relevant to neuronal function ([Fig. 3A,](#page-4-0) e.g. metabolic processes, neurotransmitter synthesis/release, synaptic

Fig. 2. A) Volcano plot demonstrating expression of all of the genes mapped from the dataset. Annotations for the top genes (up and down) by strongest FDR and/or log2foldchange. A) Expression data for the top genes annotated in volcano plot in panel (A). C) Bar graphs depicting the FPKM value (mean± SD) of each of the top 5 increased (top) and decreased (bottom) genes by strongest FDR. Abbreviations: Vinculin (*Vcl*), natural killer tumor recognition sequence (*Nktr*), suppressor of Ty 16 (*Supt16*), a disintegrin-like and metallopeptidase (resprolysin type) with thrombospondin type 1 motif 3 (*Adamts3*), protocadherin gamma subfamily C 3 (*Pcdhgc3*), pregnancy specific glycoprotein 16 (*Psg16*), ß-1,3-galactosaminyltransferase polypeptide 1 (*B3galnt1*), apolipoprotein O (*Apoo*), predicted gene 35161 (*Gm35161*), sperm associated antigen 5 (*Spag5*). Value over each comparison represents FDR.

transmission) as well as neuronal compartmentalization [\(Fig. 3B](#page-4-0), e.g. presynapse, cytoplasmic vesicle membrane, synaptic membrane) were found to be most strongly enriched between groups. This was additionally confirmed by k-means clustering which distinctly identified 3 clusters with Biological Adhesion, Neuron Differentiation, and RNA Processing as the most strongly associated GO BP terms ([Fig. 4\)](#page-5-0). To further assess functional enrichment across the dataset, we analyzed the KEGG and Reactome pathways associated with either the 1,396 decreased DEGs [\(Fig. S1A](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data) and [C\)](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data) or the 1,466 increased DEGs [\(Fig. S1B](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data) and [D](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data)) uniquely. The top 5 strongest KEGG pathways associations across either dataset were: Synaptic vesicle cycle (down), neuroactive ligand–receptor interaction (up), insulin secretion (down), retrograde endocannabinoid signaling (down), and oxidative

Gene Ontology Biological Processes A

Gene Ontology Cellular Components B

Fig. 3. A) Multidimensional scaling plot representing the top gene ontology (GO) biological process (BP) terms by semantic similarity which are enriched between smoke-exposed and filtered air control samples (*q*-value*<*0.05) across all 2,862 DEGs identified in our dataset. B) Pathway interaction map highlighting GO pathways in the cellular component (CC) category between smoke-exposed and filtered air control samples (*q*-value*<*0.05) across the entire DEG dataset. For both panels, color scale corresponds to log(*q*-value), whereas symbol size corresponds to log(no. of terms).

phosphorylation (down). The five strongest Reactome pathways associations across either dataset were: Amine ligand-binding receptors (up), neuronal system (down), transmission across chemical synapses (down), serotonin neurotransmitter release (down), and glutamate neurotransmitter release (down). Lastly, we visualized the PPI network map generated using the STRING plug-in for cytoscape via Gephi. This resulted in a network [\(Fig. 5](#page-6-0)) with a central node at ß-catenin and secondary hubs around gene expression machinery, Wnt signaling, and mitochondrial oxidases.

Discussion

With warmer and dryer climates increasing wildfire prevalence worldwide, smoke exposure is of increasing concern regarding public and occupational health, including recently associated adverse CNS outcomes ([Oudin et al. 2018](#page-9-0); [Sosedova et al. 2021;](#page-10-0) [Scieszka et al. 2022](#page-10-0), [2023](#page-10-0); [Zhang et al. 2023](#page-10-0)). To address this, we explored the effects of chronic exposure to laboratory-generated wildfire smoke, estimated to be equivalent to a career wildland firefighter exposure, on the gene expression profile of the PFC in male mice using RNA-seq. We report robust changes in gene

process (BP) term used to classify each cluster (FDR*<*0.05) as associated with cluster bar color on the left side of the image: Biological adhesion (yellow), neuron differentiation (pink), and RNA processing (blue). Color scale on the right side of the image represents the log-transformed FPKM value of each gene/transcript.

expression across the genome and functionally characterized DEGs between groups using a battery of bioinformatic approaches. Here, we will discuss the contextual relevance of those disruptions given our experimental paradigm and previously published work.

The PFC was selected as the brain region of interest due to its dynamic involvement in development and cognitive function ([Koechlin 2016;](#page-9-0) [Xu et al. 2019;](#page-10-0) [Kolk and Rakic 2022](#page-9-0)), its high level of conservation between rodents and humans ([Chini and](#page-8-0) [Hanganu-Opatz 2021](#page-8-0)), and its documented susceptibility to environmental toxicant exposure [\(Tooley et al. 2021\)](#page-10-0). Human PFC has been shown to be reduced in volume following ambient $PM_{2.5}$ exposure [\(Gale et al. 2020](#page-8-0); [Petkus et al. 2022\)](#page-9-0). Further, both male and female mice have been demonstrated to possess neuroinflammatory and neurometabolic phenotypes following public health-relevant doses of wildfire smoke or biomass combustion smoke exposure, respectively ([Scieszka et al. 2022,](#page-10-0) [2023\)](#page-10-0). Here, our results indicate that there are robust trends of dysregulation in synaptic transmission ([Fig. 3\)](#page-4-0), and suggest specific alterations in serotonergic, cholinergic, and dopaminergic neurotransmitter metabolism pathways following chronic occupational smoke

exposure ([Fig. 3](#page-4-0) and [Fig. S1\)](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data). This is consistent with previous work demonstrating lower serotonergic metabolite levels in human urine following occupational air pollution exposure ([Tomei et al. 2004\)](#page-10-0) as well as dysregulation of serotonin across the limbic system in both mice and rats exposed to ambient air pollution [\(Mokoena et al. 2015;](#page-9-0) [Yokota et al. 2016\)](#page-10-0). Atypical serotonin signaling in the PFC has been broadly reviewed previously ([Puig and Gulledge 2011](#page-9-0)), highlighting the involvement of this disrupted transmission in aberrant neuropsychiatric outcomes. It is worth noting that increased prevalence of PTSD, suicide risk, and other neuropsychiatric symptoms have been reported in career wildland firefighters and in volunteer firefighters taking part in wildfire containment efforts [\(Stanley et al. 2018](#page-10-0); [Schnell](#page-10-0) [et al. 2020\)](#page-10-0). Although behavioral assessment was not included in our study, previous groups have demonstrated the ability of biomass combustion smoke to drive aberrant exploratory behavior, locomotor patterns, spatial navigation, and anxiety even into subsequent generations of paternally exposed rats ([Sosedova](#page-10-0) [et al. 2021\)](#page-10-0). This locomotor disruption is especially interesting in the context of the differential cholinergic and dopaminergic metabolic pathways enrichment in our current study ([Fig. 3A](#page-4-0)),

Fig. 5. Protein–protein interaction (PPI) network map assembled using STRING database plug-in for cytoscape and visualized using Gephi (Freuchterman–Reingold) for the 2,862 DEGs identified in our dataset between smoke-exposed and filtered air controls. The size of each node corresponds to the degree and the color of each edge corresponds to the string strength score (integrating known and predicted interactions).

because the recruitment of both of these neurotransmitters in the PFC has been shown to modulate movement governing signaling in the striatum [\(Adrover et al. 2020\)](#page-8-0).

Exposure to wildfire smoke has been recently associated with cognitive dysfunction, including a significantly heightened risk of AD dementia in a human epidemiologic cohort when compared with other air pollutants ([Zhang et al. 2023](#page-10-0)). Further, exposures commonly faced during wildland firefighting tasks have also been recently correlated with cognitive decline and dementia risk ([Genuis and Kelln 2015\)](#page-8-0). Although the mechanism underlying the aforementioned associations is not well understood, our study highlights functionally enriched pathways and specific gene targets to explore in this context. In our present dataset, the transcriptomic profile is highly similar to those previously identified in AD ([Nativio et al. 2020\)](#page-9-0), advanced chronologic age ([Dillman et al. 2017;](#page-8-0) [Mohan et al. 2018\)](#page-9-0), and accelerated aging

cohorts ([Cavalier et al. 2021\)](#page-8-0). This is reflected by the differential enrichment for KEGG pathways including Huntington's disease, Parkinson's disease, and AD in this dataset [\(Fig. S1](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data)). Mechanistically, one term that stands out in our dataset in the context of AD pathogenesis is the serine/threonine kinase signaling pathway ([Fig. 3A\)](#page-4-0). These kinases are directly responsible for the phosphorylation of microtubule-associated protein tau in AD, as well as other tauopathies, and contribute to pathogenic aggregation and subsequent neuronal loss ([Perluigi et al. 2016](#page-9-0)). Further, ß-catenin (*Ctnnb1*) presents as a central node in our PPI network map (Fig. 5). This protein is known to modulate neuroplasticity ([Maguschak and Ressler 2012](#page-9-0)) and, when activated, has been shown to curb blood–brain barrier (BBB) dysfunction [\(Wang](#page-10-0) [et al. 2022\)](#page-10-0). Similarly, dysregulation of ß-catenin has been implicated in accelerated aging and AD [\(Palomer et al. 2019\)](#page-9-0). Strongly associated with this central node, we also see enrichment of the Wnt signaling pathway [\(Fig. 5](#page-6-0)). Interestingly, the Wnt/ß-catenin signaling pathway has been previously demonstrated to be enriched in an analysis of the effects of wildfire smoke on the nonhuman primate nasal epigenome [\(Brown et al. 2022\)](#page-8-0). This signaling pathway has also been implicated in the pathogenesis of tauopathies and AD ([Dengler-Crish et al. 2018\)](#page-8-0). In the same PPI network in our present study, we report a hub centered around mitochondrial oxidases ([Fig. 5\)](#page-6-0). Mitochondrial dysfunction has been previously reported in the male rat brain following exposure to smoldering biomass combustion smoke ([Lee et al. 2010\)](#page-9-0) and aberrant metabolomic profiles have been reported in the hippocampus of female mice exposed to smoke from wood chip combustion [\(Scieszka et al. 2023](#page-10-0)). Mitochondrial dysfunction has further been implicated in the pathogenesis of AD, and is currently being explored as a potential druggable target in this disease ([Bhatia et al. 2022](#page-8-0)). Given the dynamic nature of these previous reports and our work, it is necessary to further explore the effects of duration of exposure to wildfire smoke, as well as to expand the assessment strategy to include additional brain regions. It is also worth examining these molecular cascades at a cellular resolution either by implementing single-cell transcriptomic approaches or by assessing protein expression with histopathologic techniques (i.e. immunofluorescent imaging modalities).

On a gene-specific basis, vinculin (*Vcl*) was found to be the most significantly increased in smoke-exposed compared with filtered air control samples ([Fig. 2\)](#page-3-0). Interestingly, vinculin levels have also been shown to increase in the PFC of rats exposed to a single prolonged stressor [\(Li et al. 2015](#page-9-0)). Further, MAPK-vinculin signaling has been shown to modulate extracellular matrix (ECM) stiffness in response to environmental signals [\(Garakani](#page-8-0) [et al. 2017](#page-8-0)). ECM remodeling was also implicated in our k-means clustering analysis which revealed a major cluster around biological adhesion ([Fig. 4](#page-5-0)). Wildfire smoke exposure has been shown to increase the expression of matrix metalloproteinases (MMPs) in guinea pigs, including those known to digest tight junctions at the BBB (MMP2/9) ([Ramos et al. 2021](#page-9-0)). More broadly, ECM degradation at the neurovascular unit is hypothesized as a mechanism through which air pollution might bypass the BBB to reach the CNS [\(Liu et al. 2021](#page-9-0)). On the other hand, research has previously demonstrated that iron soot is capable of being taken up directly through olfactory nerve bundles and tracts [\(Hopkins et al. 2018\)](#page-9-0). This is also interesting in the context of recent work demonstrating changes in DNA methylation patterns in the nasal epithelia following wildfire smoke exposure in nonhuman primates, even persisting long after exposure ceased ([Brown et al. 2022\)](#page-8-0). In our study, whereas the route of wildfire smoke exposure to the CNS was not assessed, we did see evidence of other epigenetic alterations such as the hub centered around transcriptional and translational machinery visible in the PPI network map ([Fig. 5](#page-6-0)) and the k-means cluster centered around RNA processing ([Fig. 4\)](#page-5-0). Our group has also previously shown that exposure to simulated wildfire smoke at a lesser dose and duration is sufficient to significantly disrupt DNA methylation patterns in the germ cells of male mice [\(Schuller et al. 2021\)](#page-10-0). Taken together, this warrants future assessment of epigenetic alterations both in the context of CNS exposure route and in a mechanistic capacity once the toxicant, or other factors produced following inhalation, reach the brain parenchyma.

Interestingly, there did not appear to be significant alterations in canonical neuroinflammatory pathways in our primary analyses. This was unexpected given the reports of robust neuroinflammation in prior smoke exposure studies ([Scieszka et al. 2022](#page-10-0), [2023\)](#page-10-0). Still, there are a variety of differences between this previous work and our current experiment which might explain the lack of conserved pathways disruption. For example, the dose and duration of smoke exposure in our study are both much greater than in previous analyses. Additionally, the carbon monoxide levels mice were subjected to in this study are far greater than in prior reports. Although the COHb levels observed in the present work are below the threshold of CO toxicity ([Palmeri and](#page-9-0) [Gupta 2023\)](#page-9-0), it is possible that this co-exposure did exacerbate the effects associated with wildfire smoke PM inhalation. Still, the levels of CO ppm exposed to animals in this study are within plausible exposure levels observed in wildland firefighters performing various occupational tasks [\(Cone et al. 2005;](#page-8-0) [Semmens](#page-10-0) [et al. 2021\)](#page-10-0). The increased CO and PM levels exposed to mice in this study are a direct consequence of our goal to model longitudinal wildland firefighter service as opposed to acute public health exposure. Moreover, these previous studies have employed targeted approaches to measure neuroinflammatory gene expression (e.g. RT–qPCR) which do not survey pathway enrichment across the entirety of the transcriptome. It is well established in the literature that when neuroinflammation is induced by environmental toxicant exposure, there are temporal dynamics associated with various stages of activation and resolution of specific glial cell responses [\(Rocha et al. 2022](#page-10-0)). For these reasons, we did follow-up with a brief targeted assessment of specific proteins identified in previous work. Interestingly, this highlighted concordant shifts in directionality for each of the genes/transcripts differentially expressed in our study mirroring prior reports ([Fig. S2\)](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data). Additionally, one major node in the center of a hub on our PPI network map ([Fig. 5](#page-6-0)), *Rps27a*, has been recently implicated in microglial activation in neurodegeneration via control of neuroinflammation ([Khayer et al. 2020\)](#page-9-0). Although we cannot confirm that this is representative of expression changes in glia specifically in our study, this does warrant future exploration of neuroinflammatory cascades following smoke exposure, especially at a cell-specific resolution, by more diverse time points, and across other brain regions of interest implicated in neurodegenerative disease pathogenesis.

Conclusion

We have here demonstrated that exposure to an occupationally relevant dose of simulated wildfire smoke results in robust disruption of gene expression patterns in the male mouse PFC. Future work is necessary to explore the phenotypic relevance of these molecular changes, as well as to tease apart the effects of administering similar doses of smoke across different patterns of exposure duration and at varied combustion parameters. Further, the integration of this work with other occupationally relevant exposure types through the implementation of multi-hit models, and an exploration of the sex differences in this context, would better advance our understanding of the neurotoxic risks associated with conducting wildland firefighting duties. This initial report of functionally relevant gene expression differences will guide downstream mechanistic analyses which can now apply targeted, evidence-based approaches.

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Supplementary material

[Supplementary material](https://academic.oup.com/toxsci/article-lookup/doi/10.1093/toxsci/kfae097#supplementary-data) is available at *Toxicological Sciences* online.

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Conflicts of interest

None declared.

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