

Review



Thyroid Malignancy and Cutaneous Lichen Amyloidosis: Key Points Amid *RET* Pathogenic Variants in Medullary Thyroid Cancer/Multiple Endocrine Neoplasia Type 2 (MEN2)

Laura-Semonia Stanescu ^{1,2,†}, Adina Ghemigian ^{2,3,†}, Mihai-Lucian Ciobica ^{4,5,*}, Claudiu Nistor ^{6,7,*}, Adrian Ciuche ^{6,7}, Andreea-Maria Radu ^{8,9}, Florica Sandru ^{8,9} and Mara Carsote ^{2,3}

- ¹ PhD Doctoral School, "Carol Davila" University of Medicine and Pharmacy, 0505474 Bucharest, Romania; laura-semonia.stanescu@drd.umfcd.ro
- ² Department of Clinical Endocrinology V, C.I. Parhon National Institute of Endocrinology, 011863 Bucharest, Romania; adina.ghemigian@umfcd.ro (A.G.); carsote_m@hotmail.com (M.C.)
- ³ Department of Endocrinology, "Carol Davila" University of Medicine and Pharmacy, 020021 Bucharest, Romania
- ⁴ Department of Internal Medicine and Gastroenterology, "Carol Davila" University of Medicine and Pharmacy, 020021 Bucharest, Romania
- ⁵ Department of Internal Medicine I and Rheumatology, "Dr. Carol Davila" Central Military University Emergency Hospital, 010825 Bucharest, Romania
- ⁶ Department 4—Cardio-Thoracic Pathology, Thoracic Surgery II Discipline, "Carol Davila" University of Medicine and Pharmacy, 0505474 Bucharest, Romania; adrian.ciuche@umfcd.ro
- ⁷ Thoracic Surgery Department, "Dr. Carol Davila" Central Emergency University Military Hospital, 010825 Bucharest, Romania
- ⁸ Department of Dermatovenerology, Elias University Emergency Hospital, 011461 Bucharest, Romania; andreea.radu@rez.umfcd.ro (A.-M.R.); florica.sandru@umfcd.ro (F.S.)
- ⁹ Department of Dermatovenerology, "Carol Davila" University of Medicine and Pharmacy, 020021 Bucharest, Romania
- Correspondence: lucian.ciobica@umfcd.ro (M.-L.C.); claudiu.nistor@umfcd.ro (C.N.)
- These authors contributed equally to this work.

Abstract: We aimed to provide an updated narrative review with respect to the *RET* pathogenic variants and their implications at the clinical and molecular level in the diagnosis of medullary thyroid cancer (MTC)/multiple endocrine neoplasia (MEN) type 2, particularly with respect to the presence of cutaneous lichen amyloidosis (CLA). We searched English-language, in extenso original articles with no timeline nor study design restriction that were published on PubMed. A traditional interplay stands for CLA and MTC in MEN2 (not MEN3) confirmation. While the connection has been reported for more than three decades, there is still a large gap in understanding and addressing it. The majority of patients with MEN2A-CLA have RET pathogenic variants at codon 634; hence, it suggests an involvement of this specific cysteine residue in both disorders (most data agree that one-third of C634-positive subjects have CLA, but the ranges are between 9% and 50%). Females seem more prone to MEN2-CLA than males. Non-C634 germline RET pathogenic variants included (at a low level of statistical evidence) the following: RET V804M mutation in exon 14 for MTC-CLA (CLA at upper back); RET S891A mutation in exon 15 binding OSMR variant G513D (familial MTC and CLA comprising the lower legs to thighs, upper back, shoulders, arms, and forearms); and C611Y (CLA at interscapular region), respectively. Typically, CLA is detected at an early age (from childhood until young adulthood) before the actual MTC identification unless RET screening protocols are already applied. The time frame between CLA diagnosis and the identification of RET pathogenic variants was between 5 and 60 years according to one study. The same RET mutation in one family is not necessarily associated with the same CLA presentation. In MTC/MEN2 subjects, the most affected CLA area was the scapular region of the upper back. Alternatively, another hypothesis highlighted the fact that CLA is secondary to long-term prurit/notalgia paresthetica (NP) in MTC/MEN2. OSMR p. G513D may play a role in modifying the evolutionary processes of CLA in subjects co-harboring RET mutations (further studies are necessary to sustain this aspect). Awareness in CLA-positive patients is essential, including the decision of RET testing in selected cases.



Citation: Stanescu, L.-S.; Ghemigian, A.; Ciobica, M.-L.; Nistor, C.; Ciuche, A.; Radu, A.-M.; Sandru, F.; Carsote, M. Thyroid Malignancy and Cutaneous Lichen Amyloidosis: Key Points Amid *RET* Pathogenic Variants in Medullary Thyroid Cancer/Multiple Endocrine Neoplasia Type 2 (MEN2). *Int. J. Mol. Sci.* 2024, *25*, 9765. https://doi.org/ 10.3390/ijms25189765

Academic Editor: Malgorzata Gabriela Wasniewska

Received: 5 August 2024 Revised: 28 August 2024 Accepted: 5 September 2024 Published: 10 September 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Keywords:** thyroid; RET; amyloidosis; neuroendocrine; multiple endocrine neoplasia; skin; thyroid malignancy; lichen; codon; gene

1. Introduction

Medullary thyroid carcinoma (MTC), a rare neuroendocrine tumor, involves 1–2% of thyroid cancers (SEER—Surveillance, Epidemiology, and End Results) [1–4]. This rate is lower than 3–5% as previously shown, due to a relative increase in papillary-type (PTC) incidence over the last three decades [4], noting that differentiated thyroid malignancies remain the most frequent histological types all over the world nowadays [5–7].

We aimed to provide an updated narrative review with respect to the *RET* pathogenic variants and their implications at the clinical (dermatology and endocrinology assessments) and molecular level in the diagnosis of MTC/multiple endocrine neoplasia (MEN) type 2, particularly with respect to the presence of cutaneous lichen amyloidosis (CLA). We searched English-language (at least at the abstract level), in extenso original articles with no timeline nor study design restriction that were published on PubMed regarding the key points of the mentioned thyroid malignancy and its skin signature of the amyloid type.

2. Medullary Thyroid Carcinoma (MTC) and Multiple Endocrine Neoplasia (MEN) Harboring *RET* Pathogenic Variants

MTC may occur sporadically (sMTC) (75% of all cases) or hereditary as a manifestation of MEN type 2 (formerly named MEN2A) and MEN 3 (formerly called MEN2B) [8,9]. MEN2 has a prevalence of 1 in 25,000 people and accounts for over 95% of the hereditary MTC [10,11]. Four clinical variants of MEN2 have been described with various multidisciplinary elements on first presentation and long-term surveillance [12–14]. Firstly, classic MEN2A, featuring MTC in 100% of the patients and less frequent occurrence of phaeochromocytoma (PHEO) in 50% of the subjects, or primary hyperparathyroidism (HPTH) in 15% of the MEN2 individuals, or both; secondarily, MEN2A with CLA; thirdly, MEN2A with Hirschsprung disease (HD); and fourthly, isolated familial MTC (FMTC) that accounts for about ~15% of the patients with hereditary MTC and it is diagnosed only if MTC is present in the family members as well [1,15]. On the other hand, MEN3 involves a rare syndrome, also including MTC (5% of hereditary MTCs), etc. [11,16,17].

MTC is related to the Rearranged during Transfection (RET) gene [18–27] (Figure 1).

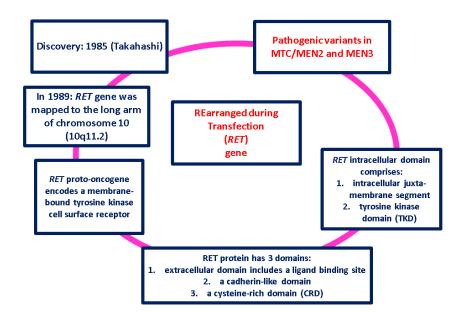
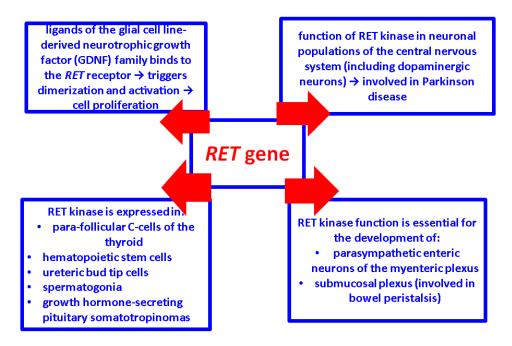
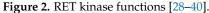


Figure 1. Main highlights with regard to RET gene [18-27].



RET functions are numerous [28–40] (Figure 2).



In the absence of the ligand, the RET protein is a single unphosphorylated tyrosine kinase receptor, while in cancer cells, *RET* proto-oncogene led to the auto-phosphorylation of the tyrosine residues [41–43].

Some *RET* pathogenic variants in thyroid malignancies are inherited and are present throughout every cell in the body (germline type) and others are acquired (somatic type) that are present only in certain cells in the body [44,45]. Even so, mutational screening is mandatory in all patients with MTC, allowing the detection of germline mutations in initially "so-called" sporadic MTC in up to 6.5% of the patients [46,47]. The etiology of sMTC has not been completely elucidated at this point [48–56] (Figure 3).

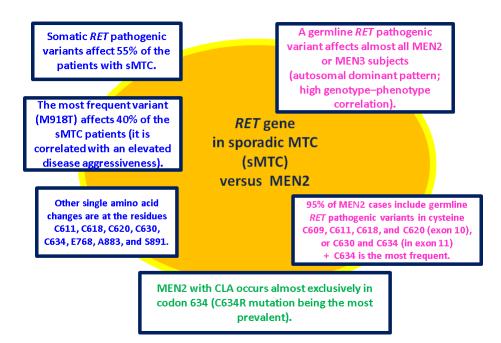


Figure 3. RET in sMTC versus MEN [48-56].

HD occurs in approximately 7% of MEN2 subjects; conversely, 2% to 5% of the patients diagnosed with HD have MEN2 [57–60]. *RET* pathogenic variants in individuals identified with MEN2 and HD are point mutations involving codons in exon 10: 609 (15%), 611 (5%), 618 (30%), and 620 (50%) [41,53,55]. It seems paradoxical that MEN2 and HD may occur together, since *RET* mutations associated with HD are loss-of-function, while those associated with MEN2A are gain-of-function. This dual occurrence could be explained by the fact that constitutive activation of *RET* is sufficient to trigger the neoplastic transformation of the C cells (at the level of thyroid cancer), yet insufficient to generate a trophic response in the precursor neurons due to a lack of expression of the RET protein at the cell surface [61–63]. Of note, the congenital absence of the ganglion cells at the level of the myenteric and submucosal intestinal plexus (namely, HD) has been found amidst other syndromes such as trisomy 21, congenital central hypoventilation syndrome, and Mowat–Wilson syndrome, while (other than *RET* proto-oncogene) endothelin receptor type B (*EDNRH*) gene was the most frequent gene incriminated in HD [63].

In isolated FMTC, the most common variants affect codons 768, 790, or 804 (exons 13 or 14), with 804 being the most frequent [11,15,64,65]. MEN3 is exclusively related to exon 16 (M918T). These mutations lead to increases in ATP-binding and autophosphorylation activity, thereby mediating a dimerization-independent activation of RET kinase [11,15,66–68]. In less than 10% of the patients, MEN3 is associated with an A883F mutation or with double mutations, which include V804M and either Y806C, S904C, E805K, or Q781R [69–72].

Current guidelines recommend germline *RET* testing in all patients with a new diagnosis of MTC, including family members. As a result of the link between the type of *RET* pathogenic variant and the aggressiveness of CMT in children with mutations at codon 918, exon 16 (harboring the highest risk) should undergo thyroidectomy in their first year of life. Those with mutations at codon 634, exon 11 or codon 883, or exon 15 (that are regarded as high risk) should have a thyroidectomy performed at the age of 5 years or earlier if the serum levels of calcitonin become elevated. In children with any of the other *RET* mutations (moderate risk), the decision regarding the age of prophylactic thyroidectomy is no longer based upon genotype alone, but is currently driven by the hormonal panel, specifically, by the increasing trend of the serum calcitonin levels [11,15].

3. Insights into Lichen Amyloidosis (LA)

Amyloidosis, characterized by abnormal misfolded protein-based deposits in addition to amyloid fibrils, affects various organ functions. It is divided into cutaneous and systemic forms (that may be accompanied by different co-morbidities) based on localization, but it can be sub-grouped via its biochemical structure [73–75]. The cutaneous form of amyloidosis was first recognized in the 1930s by Freudenthal [76,77], who noted the presence of Congo red-positive hyaline bodies within the epidermis [76,77]. The deposition of amyloid in previously apparently normal skin without deposits in the internal organs is known as primary localized cutaneous amyloidosis (PLCA), and secondary amyloidosis, which forms on a pre-existing dermatologic lesion (benign or malignant) [73,75]. Secondary PLCA is caused by rubbing/scrubbing ("friction amyloidosis") [76-78]. Its subtypes are macular (MA), lichen (LA)-the most common, and nodular (NA); biphasic amyloidosis (BA) is MA + LA [79–84]. The most common site involves the extensor surfaces of the extremities [85]. Clinically, it is difficult to distinguish different subtypes of PLCA, with the histological and immunohistochemistry characteristics being a very important tool for diagnosis. MA, LA, and BA are keratinocyte-derived, in which cytokeratins serve as amyloid precursors, while NA involves an immunoglobulin that is light-chain-derived and it is associated with dermal plasma cell infiltration [86–88]. With regard to LA, the amyloid deposits display immunoreactivity to antikeratin antibodies, but not to antibodies against A protein, pre-albumin, or fibronectin [78,88].

Generally, amyloid stands on >20 proteins [89–91]. In nature, proteins usually have both an alpha helix and β sheet structure. In amyloid, electron microscopy studies have shown the abnormal folding of proteins that are arranged predominantly as β -pleated sheets over the alpha helix [92–94]. Amyloidosis manifests when these sheets are misfolded, resulting in extracellular deposits. As for why these deposits form, the exact mechanism is not fully understood nowadays. Potential causes are as follows: protein over-production makes them fold into β -pleated sheets, abnormal protein structure, other contributors like low pH, metal ions, etc. [95–97]. Normally, certain mechanisms degrade the abnormal proteins but if they fail, amyloid can deposit in tissue. This process is highly variable and a lag phase is described in which the prerequisites are present, but no amyloid fibrils are formed yet. This lag phase can last from weeks to years. After a nucleus of amyloid is formed, aggregation occurs under fast kinetics and the development of the insoluble architecture soon follows [98–100].

Overall, the pathogenesis is considered multifactorial, involving both environmental and genetic factors. Regarding environmental factors, it is worth mentioning that Epstein–Barr virus (EBV) has been demonstrated in a high percentage of cases of MA, LA, or BA. The presence of EBV in the epidermis has been postulated as a contributing event in the degeneration of keratinocytes [101–103]. Recently, COVID-19 infection was suspected to exacerbate amyloidosis, potentially acting as a supplementary contributor or as a trigger for pre-existing histological lesions (as seen in other associated diseases following a coronavirus infection) [104–107]. When assessing if certain factors predispose individuals to develop PLCA, familial forms have been described; PLCA is common in Southeast Asia, South America, and Middle Eastern regions [108]. In addition, up to 10% of all PLCA cases are familial and an autosomal dominant inheritance may be observed [109,110]. Racial susceptibility and familial aggregation suggest that underlying genetic factors must play a role in the pathogenesis of PLCA. In cases of familial PLCA, pathogenic heterozygous missense mutations have been mapped to the oncostatin M receptor beta subunit (*OSMR*) gene [111–121] (Figure 4).

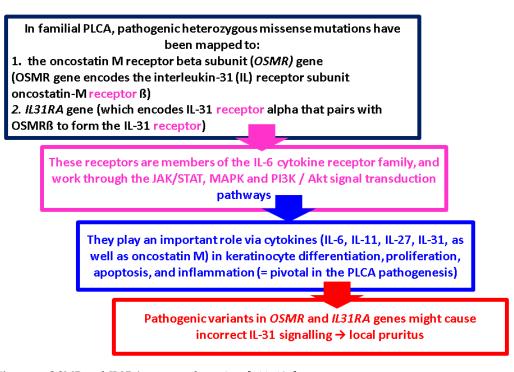


Figure 4. OSMR and IL3RA genes and pruritus [111-121].

4. CLA in MTC/MEN2

4.1. Traditional Pathogenic Aspects in CLA in Patients Diagnosed with MEN2

Traditionally, scratching appears to play an important role in the pathogenesis of CLA by causing epidermal cell damage that leads to filamentous degeneration of the keratinocytes, subsequent apoptosis, and conversion of the filamentous masses into typical amyloid material. This mechanism is supported by the fact that LA commonly presents as a pruriginous lesion. [121,122]. With regard to the majority of LA cases that are associated with MEN2, pruritus precedes the skin lesion, causing subsequent scratching and therefore cutaneous lesions. In an early study conducted by Chabre et al. [122] in 1992, a French MEN2A family (as the term was used at that moment) had three members with the same type of pruritic scapular skin lesion associated with paresthesia and hyperalgesia in the same area. Despite extensive analysis of the skin biopsies (special staining and immunohistochemistry analysis), no amyloid deposits were found at that time [122]. Moreover, these previous findings suggested a neurological origin of the cutaneous manifestation associated with MEN2A. Based on this hypothesis, CLA in MEN2 may be caused by a neurological condition called notalgia paresthetica (NP) [122–126]. NP refers to a neuropathy involving the dorsal primary divisions of spinal nerves, which causes localized pruritus. It was first described in 1934 and afterwards reviewed by Weber and Poulos [127,128], who extended the term to several entities previously described as "localized pruritus", "puzzling posterior pigmented patches", and "peculiar spotty pigmentation" [123–126]. The main symptom of NP is pruritus, and it is typically unilateral, localized on two-thirds of the scapula. In patients with long-standing pruritus, secondary lesions such as hyperpigmentation and lichen development may be visualized. The relationship between NP and MEN2 might be explained by the fact that neural crest cells, implicated in the embryological development of the adrenal medulla and para-follicular C cells of the thyroid, are also involved in the embryogenesis of the thoracic sensory fibers that explains the pruritus sites [127,128].

Another hypothesis relates to the *RET* ligand, namely, GDNF family receptor alpha (GFR α) proteins with a potential role in enteric neurons, parasympathetic and sympathetic neurons, and sensory neurons, but further evidence is necessary [30,129–135].

4.2. Sample-Based Data in MTC/MEN2-Related CLA

After Gagel et al. [136] described for the first time CLA in MEN2 (in 1989), it was found that multiple papules disposed as a well-demarcated plaque in the scapular area might accompany the endocrine malignancy as a skin signature (other than the presence of a carcinoid syndrome in metastatic MTC) [136]. Although the association of CLA with MEN2 has long been established, the reason why only a small percentage of MEN2 patients actually develop this manifestation remains an open matter nowadays. MEN2-related CLA occurs in patients with C634 RET pathogenic variants, and it has been estimated that more than 30% of the patients with this type of mutation will be CLA-positive across their life span [56,137]. Verga et al. [120] reported that CLA was found only in MEN2A/FMTC families harboring the RET pathogenic variant in codon 634, with an incidence of 36% (9/25 of the affected subjects), mentioning that two patients actually lacked these skin lesions, but the symptom of the neurological pruritus (namely, NP) in the upper back was present [120]. Moreover, in a cohort of 38 MEN2A cases harboring the C634 mutation, Scapineli et al. [121] reported CLA in 50% of these individuals, with the C634R mutation being the most prevalent (7 kindred, 35%) [121]. A smaller percentage was reported by Eng et al. [52], specifically, CLA was detected in approximately 9% (18/199) of MEN2 individuals, and all 18 families carried the RET codon 634 pathogenic variant [52]. Qi et al. [138] showed that the leading mutation at codon 634 in exon 11 was C634Y (affecting 43.6%, 24/55 of the subjects), followed by C634R (27.3%, 15/55 individuals), C634W (10.9%, 6/55), C634G (9.1%, 5/55), C634F (1.8%, 1/55), respectively, and C634 (5.5%, 3/55) [138]. The fact that the majority of the subjects diagnosed with MEN2A-related CLA harbor codon 634 suggests an involvement of this specific cysteine residue in both endocrine and

dermatologic disorders and this might be a key turning point in understanding common pathogenic features [52,138].

Yet, other *RET* germline mutations of the intracellular TKD have been described in MEN2-related CLA; for instance, Rothberg et al. reported a case of an American female with the germline *RET* V804M mutation within exon 14 with MTC and CLA on the upper back [56]. In a Chinese family with FMTC and CLA comprising the lower legs to thighs, upper back, shoulders, arms, and forearms, Qi et al. described (in 2015) the *RET* S891A mutation within exon 15 binding *OSMR* variant G513D [139]. In 2018, Qi et al. described a novel genotype–phenotype relationship between MEN2A and CLA. They reported a Chinese pedigree with 17 individuals carrying the C611Y *RET* mutation with one member (1/17, representing 5.9%) affected by CLA in the interscapular region [140].

To date, no family with MEN3 has been reported to be diagnosed with CLA. Moreover, families with CLA (but without MEN disease features) do not seem to harbor *RET* pathogenic variants [11,121,141]. To summarize, the genetic basis for MEN2A-related CLA remains obscure, with it being suggested that the primary cause is actually neurogenic, and the skin lesions are only a secondary phenomenon due to chronic pruritus and repeated scratching (as we already mentioned) [11,121,122,141].

Other potential contributors to MEN2-CLA have been studied. Gender-related predominance in the prevalence of CLA was observed by some authors. For instance, Qi et al. [139,140] described a higher prevalence in the female population with a male-tofemale ratio of approximately 2:9 (6:27), respectively, of 1.0:3.6 (12:43) [139,140]. Similarly, Scapianeli et al. [121] described a CLA prevalence of 1.0/2.3 in men/women in the mentioned cohort [121].

Interestingly, many literature data describe the presence of CLA earlier than other endocrine (clinical) elements of MEN2, thus delaying the diagnosis of MEN2-related CLA. For example, Qi et al. revealed that the mean age at diagnosis of CLA with the RET mutation was 29.5 years (range between 5 and 60 years) [140]. The age at onset for CLA for most MEN2 individuals was often in infancy or adolescence and most patients presented pruritic symptoms before the actual endocrine diagnosis of MTC [11,120,140,142–146]. Individuals with the same or different RET pathogenic variant typically presented with a variable clinical manifestation of CLA. The most described phenotypes are in the scapular region of the upper back, exhibiting on the side, midline, or bilateral extending across the midline, followed by the hyperpigmentation, respectively, by the papules developed in the same area after many years of itching and scratching [11,120,140,143–146]. Thus, much attention should be paid to the recognition of CLA located in the scapular region. The most recent guidelines for MTC recommend that subjects with CLA located in the scapular region should be investigated clinically and then undertake germline RET screening for MEN2 [11,15]. A more expanded affected region than previously described, encompassing the upper back, shoulders, arms, and legs manifesting as BA, harbored the RET S891A pathogenic variant and OSMR variant G513D in a Chinese FMTC family, as mentioned [139]. Conversely, MEN2-related CLA was strictly located in the scapular region and had the RET C634F/G mutation without the OSMR mutation in four Chinese patients [140]. Therefore, OSMR p.G513D may play a role in modifying the evolutionary process of CLA with RET pathogenic variants [139,140] (Table 1).

Table 1. Sample-focused analysis on published data with respect to MTC/MEN2 and CLA according to our methods (PubMed search based on key terms "lichen amyloidosis" and "thyroid cancer" without any timeline restriction); the display starts with the most recent publication date [56,121,128,136,139,142–156]. (Abbreviations: CLA = cutaneous lichen amyloidosis; F = female; GC = glucocorticoids; HPTH = primary hyperparathyroidism; PHEO = pheochromocytoma; MEN = multiple endocrine neoplasia; MTC = medullary thyroid carcinoma; M = male; NA = not available; n = number of family members/affected patients with MEN2A; a = two patients were not evaluated for the presence of CLA; b = all three patients with CLA had the*RET*p.S891A mutation and a novel*OSMR*variant p.G513D, which provides a possible new insight into the mechanism underlying FMTC/CA; c = data expressed as mean and standard deviation for age; of note, the data are displayed across a heterogeneous spectrum depending on the original reports).

Publication Data Reference	Studied Population (MEN Subtype)	RET Mutation	CLA + [n (%)]	Sex CLA + M:F (F%)	Age CLA at Diagnosis (Years)	Clinical Features of CLA	Treatment	Associated Endocrine Lesions MTC [n (%)] PHEO [n (%)] HPTH [n (%)]	Other Observations
Fang X et al., 2022 [147]	28 MEN2A	C634G/F/R/S/W and C611Y	8/28	2:6	18.4 ± 4.6	Interscapular region	NA	NA	Incidence of CLA in C611Y lower than those in C634G/F/R/S/W
Tang HX et al., 2021 [148]	8 MEN2A	C634R	1/8	0:1 (100)	NA	Interscapular area of the left back corresponding to dermatomes T2-T6 level	Topical GC	РНЕО, МТС, НРТН	Pruritus before CLA CLA 22 years earlier than other endocrine symptoms of MEN2A Pruritus relieved temporarily by topical GS
Malhotra et al., 2020 [149]	A 33-year-old	634	1 (100)	0:1 (100)	NA	Right scapular region	NA	Metastatic MTC, PHEO	Pruritus before CLA
Pal R et al., 2018 [150]	A 45-year-old	C634R	1 (100)	0:1 (100)	41	Interscapular region	NA	Metastatic PHEO, MTC	
Zhang XW et al., 2016 [151]	73	634	14/73 ()		NA	NA	NA	NA	
Scapineli JO et al., 2016 [121]	38 ^a MEN2A	C634Y/R/W	18 (50)	5:13 (72)	19 ± 10	Interescapular region	NA	MTC 37 (97) Pheo 11 (31) HPTH 6 (19)	-15/18, 83% CLA before other endocrine symptoms of MEN2A Pruritus before CLA

		Table 1. Cont.							
Publication Data Reference	Studied Population (MEN Subtype)	RET Mutation	CLA + [n (%)]	Sex CLA + M:F (F%)	Age CLA at Diagnosis (Years)	Clinical Features of CLA	Treatment	Associated Endocrine Lesions MTC [n (%)] PHEO [n (%)] HPTH [n (%)]	Other Observations
Qi XP et al., 2015 [139]	6 MEN2A	S891A and G513D ^b	3 (50)	2:1 (33)	29 ± 2	Upper back and legs, arms, shoulders	All patients were treated with glucocorticoid cream, which resulted in a decreased period of itching, but the application was discontinued due to side effects.	MTC 3 (50) PHEO 0, HPTH 0	Pruritus before CLA
Rothberg AE et al., 2009 [56]	1 FMTC	V804M	1 (100)	0:1 (100)	40	Interscapular	Glucocorticoid cream provided minimal relief, but no other medication was palliative.	MTC 1 (100) PHEO 0, HPTH 0	Pruritus worsened by stress
Gullu S et al., 2005 [144]	1 MEN2A	C634Y	1 (100)	0:1 (100)	34	Interscapular region	NA	1 (100) MTC, 1 (100) bilateral PHEO, 0 (0) HPTH	Pruritus present
Abdullah F et al., 2004 [146]	4 MEN2A	634	3 (75)	0:3 (100)	31 ± 14	Interscapular region	NA	4 (100) MTC, PHEO NA, HPTH NA	
Lemos MC et al., 2003 [152]	5 MEN2A	C634W	4 (80)	1:3 (75)	33 ± 21	NA	NA	5 (100)CMT, 2(40) PHEO, 2(40)HPTH	
Vieira AE et al., 2002 [153]	4 MEN2A	C634Y/R	1 (25)	0:1 (100)	21	NA	NA	1 (100) MTC, 1 (100) PHEO, 1(100) HPTH	
Lemos MC et al., 2002 [154]	1 MEN2A	C634W	1 (100)	NA	5	NA	NA	1 (100) MTC, 0 (0) PHEO, 0 (0) HPTH	
Karga HJ et al., 1998 [145]	29 MEN2A	12 C634R/2 C620Y/15 C634Y	1 (3.4)	1:0 (0)	24	NA	NA	1 (100) MTC, 1 (50) PHEO, 0 (0) HPTH	CLA before the diagnosis of MEN 2A

Publication Data Reference	Studied Population (MEN Subtype)	RET Mutation	CLA + [n (%)]	Sex CLA + M:F (F%)	Age CLA at Diagnosis (Years)	Clinical Features of CLA	Treatment	Associated Endocrine Lesions MTC [n (%)] PHEO [n (%)] HPTH [n (%)]	Other Observations
Seri M et al., 1997 [142]	2 MEN2A	C634G	2 (100)	0:2 (100)	49 ± 7	Interscapular	NA	2 (100) MTC, 1 (50) PHEO, 0 (0) HPTH	Pruritus present
Pacini F et al., 1993 [155]	11 MEN2A	NA	4 (36)	0:4 (100)	NA	Interscapular region	NA	NA	Pruritus present
Kousseff BG et al., 1991 [128]	6 MEN2A	NA	2 (33)	0:2 (100)	NA	Interscapular region	NA	6 (100) 5 (83) 0 (0)	Pruritus present
Ferrer JP et al., 1991 [143]	7 MEN2A	C634W	5 (71)	2:3 (60)	16 ± 7	Interscapular region	NA	7 (100) 0 (0) 0 (0)	
Gagel RF et al., 1989 [136]	5 MEN2A	NA	3 (60)	0:4 (100)	19 ± 1	Upper back	Topical GS	5 (100) 1 (20) NA	Pruritus relieved temporarily by topical GS
Nunziata V et al., 1989 [156]	10 MEN2A	NA	5 (50)	1:4 (80)	7 ± 2	NA	NA	10 (100) 3 (30) NA	Prurit present

Table 1. Cont.	
----------------	--

5. Discussion

According to the searched data [56,121,128,136,139,142–156], a traditional interplay stands for CLA and MTC in MEN2 (not MEN3) confirmation. While the connection has been reported for more than three decades, there is still a large gap in understanding and addressing it from a practical multidisciplinary point of view and awareness is essential, especially since the timeline perspective includes an early CLA identification before the recognition of the thyroid cancer in subjects who are not already confirmed with *RET* pathogenic variants or they are not under endocrine surveillance protocols. Females seem more prone to MEN2-CLA than MEN2 males. The majority (but not all) of patients with MEN2A-related CLA have *RET* pathogenic variants at codon 634; hence, this represents a key point that suggests an involvement of the specific cysteine residue in both disorders, which may imply the possibility of additional mechanisms in the pathogenic signal transduction pathways for MEN2A-related CLA. Additionally, *OSMR* p.G513D may play a role in modifying the evolutionary process of CA in subjects co-harboring *RET* mutations (but the level of statistical evidence remains low in this specific matter) [56,121,128,136,139,142–156].

5.1. Unraveling Clinical CLA Insights

Regardless of the synchronous/asynchronous identification of MEN2 or confirmation of RET pathogenic variants at screening genetic testing on one patient, the diagnosis of PLCA should be confirmed by the histological exam in daily practice, whereas the deposition of eosinophil, amorphous globular material in the papillary dermis, melanophages, and increased pigmentation of the basal layer are typically observed [114]. The special stains routinely employed in the detection of amyloid deposits are Congo red and Crystal violet. Also, other stains such as Thiofavin T, periodic acid-Schiff method, and Sirius red may be used for a more defined visualization [80,85,109,114,157]. Of note, Congo red plays a particular role in the pathological confirmation of MTC as well [158–160]. Both macular and lichen amyloidosis display amyloid deposits restricted to the upper dermis, particularly the papillary dermis [80,85,109]. As previously reported by Nunziata et al., staining for amyloid may be negative in a skin biopsy and multiple biopsies are needed to demonstrate amyloid content [156]. Such negative results may be explained by the fact that in the initial stages of PLCA, there is a small amount of amyloid deposit. Perhaps, dermal amyloid might be demonstrated only after long-term scratching when the skin appears clearly hyperkeratotic and pigmented [80]. Currently, most of these techniques have lost some use in daily practice due to the availability of specific types of antibodies via immunohistochemistry analysis [161]. Electronic microscopy plays a limited role in the everyday diagnosis of CLA. Nevertheless, it might prove a useful tool when the mentioned special stains fail to demonstrate the amyloid deposit, as explained by [161,162].

To date, several treatment options for PLCA have been described: both topical and systemic agents as well as phototherapy and laser therapy. However, with few clinical trials available so far, there is no gold standard treatment. Despite the multitude of treatment options, none are curative and most of them aim to break the itch-scratch cycle. One treatment option could be systemic retinoid treatment, which revealed benefits in decreasing pruritus and the size of the cutaneous lesion in LA and BA [117,163]. Other systemic options were cyclophosphamide and colchicine, which showed benefits on pruritus, pigmentation, and decrease in the size of the lesions with few side effects according to some authors [164]. In cases of refractory PLCA, methotrexate was proposed as an alternative therapy in resolving skin manifestations and also in reducing pruritus [164,165]. Amitriptyline, a tricyclic antidepressant, was indicated in neuropathic itching and it has proved beneficial in the resolution of pruritus in PLCA with no change in the skin lesions [164]. Mild cases of PLCA responded to a topical treatment such as the application of capsaicin to some extent [165]. Topical corticosteroids have demonstrated long-term resolution in LA lesions and improvement in pruritus when combined with topical salicylic acid 25% ointment [166,167]. Dimethyl sulfoxide has been used in LA with varying degrees of success and notable side effects [117]. Alternatively, topical vitamin D3 analogue calcipotriol, the

calcineurin inhibitor cyclosporine, menthol, and tacrolimus have been used [117]. Recently, laser therapy was found to be efficient in PLCA (carbon dioxide laser and erbium:yttrium aluminum garnet laser) [168,169]. Different phototherapeutic modalities have also been described in the treatment of PLCA such as UVB irradiation and topical psoralen with UVA (PUVA) therapy with heterogeneous results until the present time [170,171]. Furthermore, additional interventions such as transcutaneous nerve stimulation and surgical procedures (such as electrodessication, dermabrasion) in LA have also been reported [117]. Currently, we have no statistically significant data to pinpoint whether MEN2-CLA should be approached differently than non-MTC cases and whether LCA in association with a thyroid malignancy/neuroendocrine neoplasm is more severe apart from an increased disease burden that comes with the clinical expression due to the *RET* pathogenic variants.

5.2. Integrating MTC/MEN2-Related CLA into the Larger Frame of Skin Lesions in Neuroendocrine Neoplasia and Endocrine Tumors/Malignancies

Rarely in the general population are there patients presenting with skin lesions that reflect an underlying endocrine disorder, typically a functionally active tumor. As seen in MEN2-CLA, awareness remains the key operating perspective. For instance, necrolytic migratory erythema (NME) was firstly described by Becker et al. (Becker's nevus) in 1942 in a woman with an islet cell type of pancreatic carcinoma that was revealed only post-mortem [172]. Further on, a connection with glucagonoma was established, regardless of if the neoplasia was a part of MEN1 in some patients (in addition to other endocrine tumors in the pituitary, parathyroid, thyroid, and adrenal glands) [173,174]. McCune-Albright syndrome (somatic activating mutations in GNAS gene) has cafe-au-lait spots, hyper-function of multiple endocrine glands, etc.; autonomous hyper-function most commonly involves the ovary, but also the thyroid (nodular hyperplasia with thyrotoxicosis), adrenal (multiple hyperplastic nodules with Cushing's syndrome), and pituitary glands (somatotropinomas and prolactinomas), and parathyroid tumors causing HPTH (another type of hereditary HPTH as found in MEN2) [175,176]. As mentioned, MEN1, with an autosomal dominant predisposition to tumors of the parathyroid glands, anterior pituitary, and pancreatic islet cells, associates with multiple cutaneous lesions such as angiofibromas, lipomas, and collagenomas, and, potentially, an increased risk of other non-endocrine malignancies [177,178]. The complex of spotty skin pigmentation, myxomas, endocrine gland over-activity due to various tumors, and schwannomas, namely, Carney complex, stands for an autosomal dominant syndrome involving the skin in terms of developing myxomas (that may have different non-cutaneous sites), and spotty pigmentation [179,180].

Finally, one of the most important aspects in the complex panel of skin signature with respect to the endocrine malignancies/neuroendocrine neoplasia is represented by the carcinoid syndrome in metastatic MTC. This is caused by the liver metastases that interfere with the serotonin (5-hydroxytriptamine), respectively, 5-hydroxyindolacetic acid metabolism as similarly seen in cancers of other primary origins [181,182]. Cutaneous involvement such as episodic flushing is the clinical hallmark in 85% of the patients. It primarily involves the face, neck, and upper chest, which become red to violaceous or purple, and are associated with a mild burning sensation [181,183,184]. Thus, CLA in MTC/MEN2 subjects should be regarded via the larger and complex frame of endocrine and skin interplay that involves a common genetic, epigenetic, molecular, immune, and hormonal background depending on the specific circumstance.

5.3. Current Limits and Further Expansion

We are aware of the limits of a non-systematic review, but, noting the current data available, we chose a more flexible approach under various perspectives, from genetic interplay to the clinical expression. There is still a matter of debate: if CLA is more severe in MEN2 versus non-MEN2 subjects; if MTC displays a more severe outcome in CLA-positive versus CLA-negative patients; if there is a distinct risk of non-MTC MEN2 components other than the indications provided by the mutation map in CLA-positive individuals. Overall,

early diagnosis of CLA should raise the question of a thyroid malignancy across the life span. Moreover, prurit before the actual skin lesion might pinpoint a pathogenic contributor to lichen, other than the actual *RET* gene influence. We still need large, longitudinal studies to address this crossroad between CLA and MEN2. Although the association of CLA with MEN2 has long been established, the reason why a small percentage of MEN2 patients actually develop this manifestation remains an open matter.

6. Conclusions

CLA in thyroid cancer, particularly, MTC (and mostly across the confirmation of MEN2 underlying the *RET* pathogenic variants) stands for the following key points:

- MEN2-related CLA occurs in patients with C634 RET pathogenic variants; most data agree that one-third of C634-positive subjects have CLA, but the ranges are between 9% and 50%.
- One single study showed the *RET* codons map (codon 634 in exon 11) in CLA: C634Y (affecting 43.6% of the subjects), followed by C634R (27.3%), C634W (10.9%), C634G (9.1%), C634F (1.8%), and C634 (5.5%).
- Non-C634 germline *RET* pathogenic variants included (at a low level of statistical evidence) the following: *RET* V804M mutation in exon 14 for MTC and CLA (CLA at upper back); *RET* S891A mutation in exon 15 binding *OSMR* variant G513D (FMTC and CLA comprising the lower legs to thighs, upper back, shoulders, arms, and forearms); and C611Y (CLA at interscapular region).
- Awareness in CLA-positive patients is essential, including the decision of *RET* testing in selected cases (Table 2).

Table 2. Key findings across our search according to the mentioned references [128,137,139,141,143, 146,149–155,157–162].

Number	Key Findings according to Our Search						
1.	MEN2-related CLA occurs in patients with C634 <i>RET</i> pathogenic variants; most data agree that one-third of the C634-positive subjects have CLA, but the ranges are between 9% and 50%.						
2.	One single study showed the <i>RET</i> codons map (codon 634 in exon 11) in CLA: C634Y (affecting 43.6% of the subjects) followed by C634R (27.3%), C634W (10.9%), C634G (9.1%), C634F (1.8%), and C634 (5.5%).						
3.	Non-C634 germline <i>RET</i> pathogenic variants included (at a low level of statistical evidence) the following: <i>RET</i> V804M mutation in exon 14 for MTC and CLA (CLA at upper back); <i>RET</i> S891A mutation in exon 15 binding <i>OSMR</i> variant G513D (FMTC and CLA comprising the lower legs to thighs, upper back, shoulders, arms, and forearms); and C611Y (CLA at interscapular region).						
4.	Typically, CLA is detected at an early age (from childhood until young adulthood) before the actual MTC identification unless <i>RET</i> screening protocols are already applied.						
5.	The time frame between CLA diagnosis and the identification of <i>RET</i> pathogenic variants varied between 5 and 60 years according to one study.						
6.	Females seem more prone to MEN2-CLA than males.						
7.	The same <i>RET</i> mutation is not necessarily associated with the same CLA presentation.						
8.	In MTC/MEN2 subjects, the most affected CLA area was the scapular region of the upper back.						
9.	Alternatively, another hypothesis highlights the fact that CLA is secondary to a long-term prurit/notalgia paresthetica in MTC/MEN2, and not to a distinct <i>RET</i> influence.						
10.	The relationship between NP and MEN2 might be explained by the fact that neural crest cells, implicated in the embryological development of the adrenal medulla and para-follicular C cells of the thyroid, are also involved in the embryogenesis of the thoracic sensory fibers that explains the pruritus sites.						
11.	While the NP hypothesis might explain some cases, pruritus may be largely absent in a high percentage of patients with PLCA, including some patients with LCA-MEN2; thus, other pathogenic loops might actually be involved.						
12.	Alternatively, it has been suggested that pathogenic variants in <i>OSMR</i> and <i>IL31RA</i> genes lead to incorrect IL-31 signaling, which is directly related to local pruritus (and the co-presence of other non-RET mutations should be taken into consideration to explain CLA in MTC/MEN2).						

Table 2. Cont.

Number	Key Findings according to Our Search							
13.	OSMR p.G513D may play a role in modifying the evolutionary processes of LA in the subjects that co-harbor <i>RET</i> mutations (further studies are necessary to sustain this aspect).							
14	Awareness in CLA-positive patients is essential, including the decision of <i>RET</i> testing in selected cases.							
	methodo A.C., F.S LS.S., <i>A</i> F.S. and LS.S., <i>A</i> (endocrin C.N., A.0 M.C.; su	Contributions: Conceptualization, LS.S., A.G., ML.C., C.N., A.C., AM.R., F.S. and M.C. logy, LS.S., A.G., ML.C., C.N., AM.R., F.S. and M.C.; software, LS.S., A.G., ML.C., C.N. and M.C.; validation, LS.S., A.G., ML.C., C.N., A.C., AM.R. and M.C.; formal analysis G.G., ML.C., C.N., A.C. and M.C.; investigation, LS.S., A.G., ML.C., C.N., A.C., AM.R. M.C.; resources, LS.S., A.G., ML.C., C.N., A.C., AM.R. and M.C.; data curatior G., ML.C., C.N., A.C. and M.C.; writing—original draft preparation, LS.S., A.G. nology), AM.R. and F.S. (dermatology); writing—review and editing, LS.S., A.G., ML.C. C., AM.R., F.S. and M.C.; visualization, LS.S., A.G., ML.C., C.N., A.C., AM.R., F.S. and pervision, LS.S., A.G., ML.C., C.N., A.C., AM.R., F.S. and M.C.; project administratior nding acquisition, ML.C. All authors have read and agreed to the published version o uscript.						
		•						
	-	: This research received no external funding.						
	Instituti	onal Review Board Statement: Not applicable.						
	Informe	d Consent Statement: Not applicable.						
	Data Av	ailability Statement: Not applicable.						
	Conflicts	s of Interest: The authors declare no conflicts of interest.						
	Abbrev BA	biphasic amyloidosis						
	CLA	cutaneous lichen amyloidosis						
	CRD EDNRH	cysteine-rich domain						
	EBV	endothelin receptor type B Epstein–Barr virus						
	FMTC	familial medullary thyroid carcinoma						
	GDNF	glial cell line-derived neurotrophic growth factor						
	GFRα	GDNF family receptor alpha						
	IL	interleukin						
	HD	Hirschsprung disease						
	HPTH	primary hyperparathyroidism						
	LA MTC	lichen amyloidosis						
	MEN	medullary thyroid carcinoma multiple endocrine neoplasia						
	MA	macular amyloidosis						
	NA	nodular amyloidosis						
	NP	notalgia paresthetica						
	NME	necrolytic migratory erythema						
	OSMR	oncostatin M receptor beta subunit						
	PTC	papillary thyroid carcinoma						
	PLCA	primary localized cutaneous amyloidosis						
	PHEO	phaeochromocytoma						
	PUVA	psoralen with UVA						
	RET	Rearranged during Transfection						
	sMTC SEER	sporadic medullary thyroid carcinoma Surveillance, Epidemiology, and End Results						
	TKD	tyrosine kinase domain						
		CHORE KHUSE COMUNI						

References

- 1. Frisco, N.A.; Gunn, A.H.; Wang, F.; Stang, M.T.; Kazaure, H.S.; Scheri, R.P. Guideline Adherence and Practice Patterns in the Management of Medullary Thyroid Cancer. J. Surg. Res. 2023, 281, 214–222. [CrossRef]
- Fritz, C.; De Ravin, E.; Suresh, N.; Romeo, D.; Shah, M.; Rajasekaran, K. Clinical practice guidelines for management of medullary thyroid carcinoma: An AGREE II appraisal. *Am. J. Otolaryngol.* 2022, 43, 103606. [CrossRef]
- 3. Williams, E.D. Histogenesis of medullary carcinoma of the thyroid. J. Clin. Pathol. 1966, 19, 114–118. [CrossRef]
- 4. Cancer of the Thyroid Invasive: Trends in SEERIncidence, U.S. Mortality Using the Joinpoint Regression Program, 1975–2011(SEER) Stat version 8.1.2 Rate Session. Available online: www.seer.cancer.gov (accessed on 27 July 2024).
- Asya, O.; Yumuşakhuylu, A.C.; Enver, N.; Gündoğdu, Y.; Abuzaid, G.; İncaz, S.; Gündoğmuş, C.A.; Ergelen, R.; Bağcı, P.; Oysu, Ç. A single-center multidisciplinary study analyzing thyroid nodule risk stratification by comparing the thyroid imaging reporting and data system (TI-RADS) and American thyroid association (ATA) risk of malignancy for thyroid nodules. *Auris Nasus Larynx*. 2023, *50*, 410–414. [CrossRef]
- Gordon, A.J.; Dublin, J.C.; Patel, E.; Papazian, M.; Chow, M.S.; Persky, M.J.; Jacobson, A.S.; Patel, K.N.; Suh, I.; Morris, L.G.T.; et al. American Thyroid Association Guidelines and National Trends in Management of Papillary Thyroid Carcinoma. *JAMA Otolaryngol. Head Neck Surg.* 2022, 148, 1156–1163. [CrossRef]
- 7. Gulec, S.A.; Ahuja, S.; Avram, A.M.; Bernet, V.J.; Bourguet, P.; Draganescu, C.; Elisei, R.; Giovanella, L.; Grant, F.; Greenspan, B.; et al. A Joint Statement from the American Thyroid Association, the European Association of Nuclear Medicine, the European Thyroid Association, the Society of Nuclear Medicine and Molecular Imaging on Current Diagnostic and Theranostic Approaches in the Management of Thyroid Cancer. *Thyroid* 2021, *31*, 1009–1019. [CrossRef]
- McDonnell, J.E.; Gild, M.L.; Clifton-Bligh, R.J.; Robinson, B.G. Multiple endocrine neoplasia: An update. Intern. Med. J. 2019, 49, 954–961. [CrossRef]
- 9. Rindi, G.; Mete, O.; Uccella, S.; Basturk, O.; La Rosa, S.; Brosens, L.A.A.; Ezzat, S.; de Herder, W.W.; Klimstra, D.S.; Papotti, M.; et al. Overview of the 2022 WHO Classification of Neuroendocrine Neoplasms. *Endocr. Pathol.* 2022, 33, 115–154. [CrossRef]
- 10. Romei, C.; Pardi, E.; Cetani, F.; Elisei, R. Genetic and clinical features of multiple endocrine neoplasia types 1 and 2. *J. Oncol.* 2012, 2012, 705036. [CrossRef]
- Wells, S.A., Jr.; Asa, S.L.; Dralle, H.; Elisei, R.; Evans, D.B.; Gagel, R.F.; Lee, N.; Machens, A.; Moley, J.F.; Pacini, F.; et al. Revised American Thyroid Association guidelines for the management of medullary thyroid carcinoma. *Thyroid* 2015, 25, 567–610. [CrossRef]
- 12. Carsote, M.; Valea, A.; Dumitru, N.; Terzea, D.; Petrova, E.; Albu, S.; Buruiana, A.; Ghemigian, A. Metastases in daily endocrine practice. *Arch. Balk. Med. Union* **2016**, *51*, 476–480.
- Nistor, C.; Ciuche, A.; Constantinescu, I. Emergency surgical tracheal decompression in a huge retrosternal goiter. *Acta Endocrinol.* 2017, 13, 370–374. [CrossRef]
- 14. Jinga, M.; Jurcut, C.; Vasilescu, F.; Becheanu, G.; Stancu, S.H.; Ciobaca, L.; Mircescu, G.; Jinga, V. A rare case of digestive hemorrhage in an elderly patient: Diagnosis and treatment difficulties. *Rom J. Morphol. Embryol.* **2012**, *53* (Suppl. 3), 831–834.
- 15. Wells, S.A., Jr. Advances in the management of MEN2: From improved surgical and medical treatment to novel kinase inhibitors. *Endocr. Relat. Cancer* **2018**, 25, T1–T13. [CrossRef]
- 16. Castinetti, F.; Moley, J.; Mulligan, L.; Waguespack, S.G. A comprehensive review on MEN2B. *Endocr. Relat. Cancer* **2018**, *25*, T29–T39. [CrossRef]
- 17. Castinetti, F.; Waguespack, S.G.; Machens, A.; Uchino, S.; Hasse-Lazar, K.; Sanso, G.; Else, T.; Dvorakova, S.; Qi, X.P.; Elisei, R.; et al. Natural history, treatment, and long-term follow up of patients with multiple endocrine neoplasia type 2B: An international, multicentre, retrospective study. *Lancet Diabetes Endocrinol.* **2019**, *7*, 213–220. [CrossRef]
- 18. Takahashi, M.; Ritz, J.; Cooper, G.M. Activation of a novel human transforming gene, ret, by DNA rearrangement. *Cell* **1985**, 42, 581–588. [CrossRef] [PubMed]
- 19. Greenberg, L.A. Multiple Endocrine Neoplasia Type 1, Type 2A, and Type 2B. Prim. Care 2024, 51, 483–494. [CrossRef]
- Loganathan, T.; George Priya Doss, C. Biomarker identification of medullary thyroid carcinoma from gene expression profiles considering without-treatment and with-treatment studies-A bioinformatics approach. *Adv. Protein Chem. Struct. Biol.* 2024, 142, 367–396. [CrossRef]
- Ishizaka, Y.; Itoh, F.; Tahira, T.; Ikeda, I.; Sugimura, T.; Tucker, J.; Fertitta, A.; Carrano, A.V.; Nagao, M. Human ret proto-oncogene mapped to chromosome 10q11.2. Oncogene 1989, 4, 1519–1521.
- 22. Simpson, N.E.; Kidd, K.K.; Goodfellow, P.J.; McDermid, H.; Myers, S.; Kidd, J.R.; Jackson, C.E.; Duncan, A.M.; Farrer, L.A.; Brasch, K. Assignment of multiple endocrine neoplasia type 2A to chromosome 10 by linkage. *Nature* **1987**, *328*, 528–530. [CrossRef]
- 23. Prodana, M.; Nistor, C.E.; Stoian, A.B.; Ionita, D.; Burnei, C. Dual nanofibrous bioactive coarings on TiZr implants. *Coatings* **2020**, 10, 526. [CrossRef]
- 24. Boucai, L.; Zafereo, M.; Cabanillas, M.E. Thyroid Cancer: A Review. JAMA 2024, 331, 425–435. [CrossRef]
- 25. Zhang, D.; Liang, N.; Sun, H.; Frattini, F.; Sui, C.; Yang, M.; Wang, H.; Dionigi, G. Critically evaluated key points on hereditary medullary thyroid carcinoma. *Front. Endocrinol.* **2024**, *15*, 1412942. [CrossRef]
- 26. Knowles, P.P.; Murray-Rust, J.; Kjaer, S.; Scott, R.P.; Hanrahan, S.; Santoro, M.; Ibáñez, C.F.; McDonald, N.Q. Structure and chemical inhibition of the RET tyrosine kinase domain. *J. Biol. Chem.* **2006**, *281*, 33577–33587. [CrossRef]

- Santoro, M.; Melillo, R.M.; Carlomagno, F.; Vecchio, G.; Fusco, A. Minireview: RET: Normal and abnormal functions. *Endocrinology* 2004, 145, 5448–5451. [CrossRef]
- 28. Ahmadi, S.; Landa, I. The prognostic power of gene mutations in thyroid cancer. Endocr. Connect. 2024, 13, e230297. [CrossRef]
- 29. Plaza-Menacho, I. Structure and function of RET in multiple endocrine neoplasia type 2. *Endocr. Relat. Cancer* **2018**, 25, T79–T90. [CrossRef]
- Goodman, K.M.; Kjær, S.; Beuron, F.; Knowles, P.P.; Nawrotek, A.; Burns, E.M.; Purkiss, A.G.; George, R.; Santoro, M.; Morris, E.P.; et al. RET recognition of GDNF-GFRα1 ligand by a composite binding site promotes membrane-proximal self-association. *Cell Rep.* 2014, *8*, 1894–1904. [CrossRef]
- 31. Conway, J.A.; Kramer, E.R. Is activation of GDNF/RET signaling the answer for successful treatment of Parkinson's disease? A discussion of data from the culture dish to the clinic. *Neural Regen. Res.* 2022, 17, 1462–1467. [CrossRef]
- 32. Conway, J.A.; Ince, S.; Black, S.; Kramer, E.R. GDNF/RET signaling in dopamine neurons in vivo. *Cell Tissue Res.* **2020**, *382*, 135–146. [CrossRef] [PubMed]
- 33. Richardson, D.S.; Rodrigues, D.M.; Hyndman, B.D.; Crupi, M.J.; Nicolescu, A.C.; Mulligan, L.M. Alternative splicing results in RET isoforms with distinct trafficking properties. *Mol. Biol. Cell* **2012**, *23*, 3838–3850. [CrossRef] [PubMed]
- 34. Ibáñez, C.F. Structure and physiology of the RET receptor tyrosine kinase. *Cold Spring Harb. Perspect. Biol.* **2013**, *5*, a009134. [CrossRef] [PubMed]
- Meng, X.; Lindahl, M.; Hyvönen, M.E.; Parvinen, M.; de Rooij, D.G.; Hess, M.W.; Raatikainen-Ahokas, A.; Sainio, K.; Rauvala, H.; Lakso, M.; et al. Regulation of cell fate decision of undifferentiated spermatogonia by GDNF. *Science* 2000, 287, 1489–1493. [CrossRef] [PubMed]
- 36. Lindfors, P.H.; Lindahl, M.; Rossi, J.; Saarma, M.; Airaksinen, M.S. Ablation of persephin receptor glial cell line-derived neurotrophic factor family receptor alpha4 impairs thyroid calcitonin production in young mice. *Endocrinology* **2006**, 147, 2237–2244. [CrossRef]
- 37. Cañibano, C.; Rodriguez, N.L.; Saez, C.; Tovar, S.; Garcia-Lavandeira, M.; Borrello, M.G.; Vidal, A.; Costantini, F.; Japon, M.; Dieguez, C.; et al. The dependence receptor Ret induces apoptosis in somatotrophs through a Pit-1/p53 pathway, preventing tumor growth. *EMBO J.* **2007**, *26*, 2015–2028. [CrossRef]
- Veiga-Fernandes, H.; Coles, M.C.; Foster, K.E.; Patel, A.; Williams, A.; Natarajan, D.; Barlow, A.; Pachnis, V.; Kioussis, D. Tyrosine kinase receptor RET is a key regulator of Peyer's patch organogenesis. *Nature* 2007, 446, 547–551. [CrossRef]
- Tizu, M.; Mărunțelu, I.; Cristea, B.M.; Nistor, C.; Ishkitiev, N.; Mihaylova, Z.; Tsikandelova, R.; Miteva, M.; Caruntu, A.; Sabliov, C.; et al. PLGA Nanoparticles Uptake in Stem Cells from Human Exfoliated Deciduous Teeth and Oral Keratinocyte Stem Cells. J. Funct. Biomater. 2022, 13, 109. [CrossRef]
- Ohgami, N.; Ida-Eto, M.; Shimotake, T.; Sakashita, N.; Sone, M.; Nakashima, T.; Tabuchi, K.; Hoshino, T.; Shimada, A.; Tsuzuki, T.; et al. c-Ret-mediated hearing loss in mice with Hirschsprung disease. *Proc. Natl. Acad. Sci. USA* 2010, 107, 13051–13056. [CrossRef]
- 41. Mulligan, L.M. RET revisited: Expanding the oncogenic portfolio. Nat. Rev. Cancer 2014, 14, 173–186. [CrossRef]
- 42. Eng, C. RET proto-oncogene in the development of human cancer. J. Clin. Oncol. 1999, 17, 380–393. [CrossRef] [PubMed]
- Prabhash, K.; Saldanha, E.; Patil, V.; Bal, M.; Reddy, P.S.; Sanjeev, A.; Kumar, R.; Poojary, D.; Noronha, V.; Menon, N.; et al. RET Alterations Differentiate Molecular Profile of Medullary Thyroid Cancer. *JCO Precis. Oncol.* 2024, *8*, e2300622. [CrossRef] [PubMed]
- 44. Greenman, C.; Stephens, P.; Smith, R.; Dalgliesh, G.L.; Hunter, C.; Bignell, G.; Davies, H.; Teague, J.; Butler, A.; Stevens, C.; et al. Patterns of somatic mutation in human cancer genomes. *Nature* **2007**, *446*, 153–158. [CrossRef]
- 45. Loewe, L.; Hill, W.G. The population genetics of mutations: Good, bad and indifferent. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 2010, 365, 1153–1167. [CrossRef] [PubMed]
- Rocha, A.P.; Magalhães, P.K.; Maia, A.L.; Maciel, L.M. Genetic polymorphisms: Implications in the pathogenesis of medullary thyroid carcinoma. *Arq. Bras. Endocrinol. Metabol.* 2007, 51, 723–730. [CrossRef]
- 47. Romei, C.; Cosci, B.; Renzini, G.; Bottici, V.; Molinaro, E.; Agate, L.; Passannanti, P.; Viola, D.; Biagini, A.; Basolo, F.; et al. RET genetic screening of sporadic medullary thyroid cancer (MTC) allows the preclinical diagnosis of unsuspected gene carriers and the identification of a relevant percentage of hidden familial MTC (FMTC). *Clin. Endocrinol.* **2011**, *74*, 241–247. [CrossRef]
- Ciampi, R.; Romei, C.; Ramone, T.; Prete, A.; Tacito, A.; Cappagli, V.; Bottici, V.; Viola, D.; Torregrossa, L.; Ugolini, C.; et al. Genetic Landscape of Somatic Mutations in a Large Cohort of Sporadic Medullary Thyroid Carcinomas Studied by Next-Generation Targeted Sequencing. *Iscience* 2019, 20, 324–336. [CrossRef]
- Elisei, R.; Cosci, B.; Romei, C.; Bottici, V.; Renzini, G.; Molinaro, E.; Agate, L.; Vivaldi, A.; Faviana, P.; Basolo, F.; et al. Prognostic significance of somatic RET oncogene mutations in sporadic medullary thyroid cancer: A 10-year follow-up study. J. Clin. Endocrinol. Metab. 2008, 93, 682–687. [CrossRef]
- 50. Machens, A.; Dralle, H. Long-term outcome after DNA-based prophylactic neck surgery in children at risk of hereditary medullary thyroid cancer. *Best Pract. Res. Clin. Endocrinol. Metab.* **2019**, *33*, 101274. [CrossRef]
- 51. Galofré, J.C.; Santamaría Sandi, J.; Capdevila, J.; Navarro González, E.; Zafón Llopis, C.; Ramón YCajal Asensio, T.; Gómez Sáez, J.M.; Jiménez-Fonseca, P.; Riesco Eizaguirre, G.; Grande, E. Consensus on the management of advanced medullary thyroid carcinoma on behalf of the Working Group of Thyroid Cancer of the Spanish Society of Endocrinology (SEEN) and the Spanish Task Force Group for Orphan and Infrequent Tumors (GETHI). *Endocrinol. Nutr.* 2015, 62, e37–e46. [CrossRef]

- 52. Eng, C.; Clayton, D.; Schuffenecker, I.; Lenoir, G.; Cote, G.; Gagel, R.F.; van Amstel, H.K.; Lips, C.J.; Nishisho, I.; Takai, S.I.; et al. The relationship between specific RET proto-oncogene mutations and disease phenotype in multiple endocrine neoplasia type 2. International RET mutation consortium analysis. *JAMA* **1996**, *276*, 1575–1579. [CrossRef] [PubMed]
- Mulligan, L.M.; Eng, C.; Healey, C.S.; Clayton, D.; Kwok, J.B.; Gardner, E.; Ponder, M.A.; Frilling, A.; Jackson, C.E.; Lehnert, H. Specific mutations of the RET proto-oncogene are related to disease phenotype in MEN 2A and FMTC. *Nat. Genet.* 1994, 6, 70–74. [CrossRef] [PubMed]
- Santoro, M.; Carlomagno, F.; Romano, A.; Bottaro, D.P.; Dathan, N.A.; Grieco, M.; Fusco, A.; Vecchio, G.; Matoskova, B.; Kraus, M.H. Activation of RET as a dominant transforming gene by germline mutations of MEN2A and MEN2B. *Science* 1995, 267, 381–383. [CrossRef] [PubMed]
- 55. Frank-Raue, K.; Rondot, S.; Raue, F. Molecular genetics and phenomics of RET mutations: Impact on prognosis of MTC. *Mol. Cell. Endocrinol.* **2010**, 322, 2–7. [CrossRef] [PubMed]
- 56. Rothberg, A.E.; Raymond, V.M.; Gruber, S.B.; Sisson, J. Familial medullary thyroid carcinoma associated with cutaneous lichen amyloidosis. *Thyroid* 2009, *19*, 651–655. [CrossRef] [PubMed]
- 57. Verdy, M.; Weber, A.M.; Roy, C.C.; Morin, C.L.; Cadotte, M.; Brochu, P. Hirschsprung's disease in a family with multiple endocrine neoplasia type 2. *J. Pediatr. Gastroenterol. Nutr.* **1982**, *1*, 603–607. [CrossRef]
- 58. Decker, R.A.; Peacock, M.L. Occurrence of MEN 2a in familial Hirschsprung's disease: A new indication for genetic testing of the RET proto-oncogene. *J. Pediatr. Surg.* **1998**, 33, 207–214. [CrossRef]
- Sijmons, R.H.; Hofstra, R.M.; Wijburg, F.A.; Links, T.P.; Zwierstra, R.P.; Vermey, A.; Aronson, D.C.; Tan-Sindhunata, G.; Brouwers-Smalbraak, G.J.; Maas, S.M.; et al. Oncological implications of RET gene mutations in Hirschsprung's disease. *Gut* 1998, 43, 542–547. [CrossRef]
- Bussières, V.; Roy, S.; Deladoey, J.; Rousseau, É.; St-Vil, D.; Piché, N. Prophylactic thyroidectomies in MEN2 syndrome: Management and outcomes. J. Pediatr. Surg. 2018, 53, 283–285. [CrossRef]
- 61. Gietz, R.; Armando, R.; Lobos, P.; Liberto, D. Non-syndromic Hirschsprung's disease as a result of a RET gene variant. *Cir. Pediatr.* **2024**, *37*, 89–92. [CrossRef]
- Asai, N.; Jijiwa, M.; Enomoto, A.; Kawai, K.; Maeda, K.; Ichiahara, M.; Murakumo, Y.; Takahashi, M. RET receptor signaling: Dysfunction in thyroid cancer and Hirschsprung's disease. *Pathol. Int.* 2006, 56, 164–172. [CrossRef] [PubMed]
- 63. Montalva, L.; Cheng, L.S.; Kapur, R.; Langer, J.C.; Berrebi, D.; Kyrklund, K.; Pakarinen, M.; de Blaauw, I.; Bonnard, A.; Gosain, A. Hirschsprung disease. *Nat. Rev. Dis. Primers* **2023**, *9*, 54. [CrossRef]
- 64. Donis-Keller, H.; Dou, S.; Chi, D.; Carlson, K.M.; Toshima, K.; Lairmore, T.C.; Howe, J.R.; Moley, J.F.; Goodfellow, P.; Wells, S.A., Jr. Mutations in the RET proto-oncogene are associated with MEN, 2A and FMTC. *Hum. Mol. Genet.* **1993**, *2*, 851–856. [CrossRef]
- 65. Puñales, M.K.; Graf, H.; Gross, J.L.; Maia, A.L. RET codon 634 mutations in multiple endocrine neoplasia type 2: Variable clinical features and clinical outcome. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 2644–2649. [CrossRef]
- 66. Pavlidis, E.; Sapalidis, K.; Chatzinikolaou, F.; Kesisoglou, I. Medullary thyroid cancer: Molecular factors, management and treatment. *Rom. J. Morphol. Embryol.* 2020, *61*, 681–686. [CrossRef] [PubMed]
- 67. Gujral, T.S.; Singh, V.K.; Jia, Z.; Mulligan, L.M. Molecular mechanisms of RET receptor-mediated oncogenesis in multiple endocrine neoplasia 2B. *Cancer Res.* 2006, *66*, 10741–10749. [CrossRef] [PubMed]
- Plaza-Menacho, I.; Barnouin, K.; Goodman, K.; Martínez-Torres, R.J.; Borg, A.; Murray-Rust, J.; Mouilleron, S.; Knowles, P.; McDonald, N.Q. Oncogenic RET kinase domain mutations perturb the autophosphorylation trajectory by enhancing substrate presentation in trans. *Mol. Cell* 2014, 53, 738–751. [CrossRef] [PubMed]
- 69. Miyauchi, A.; Futami, H.; Hai, N.; Yokozawa, T.; Kuma, K.; Aoki, N.; Kosugi, S.; Sugano, K.; Yamaguchi, K. Two germline missense mutations at codons 804 and 806 of the RET proto-oncogene in the same allele in a patient with multiple endocrine neoplasia type 2B without codon 918 mutation. *Jpn. J. Cancer Res.* **1999**, *90*, 1–5. [CrossRef]
- Menko, F.H.; van der Luijt, R.B.; de Valk, I.A.; Toorians, A.W.; Sepers, J.M.; van Diest, P.J.; Lips, C.J. Atypical MEN type 2B associated with two germline RET mutations on the same allele not involving codon 918. *J. Clin. Endocrinol. Metab.* 2002, *87*, 393–397. [CrossRef]
- 71. Cranston, A.N.; Carniti, C.; Oakhill, K.; Radzio-Andzelm, E.; Stone, E.A.; McCallion, A.S.; Hodgson, S.; Clarke, S.; Mondellini, P.; Leyland, J.; et al. RET is constitutively activated by novel tandem mutations that alter the active site resulting in multiple endocrine neoplasia type, 2B. *Cancer Res.* 2006, *66*, 10179–10187. [CrossRef]
- 72. Nakao, K.T.; Usui, T.; Ikeda, M.; Mori, Y.; Yamamoto, T.; Kawashima, S.T.; Nanba, K.; Yuno, A.; Tamanaha, T.; Tagami, T.; et al. Novel tandem germline RET proto-oncogene mutations in a patient with multiple endocrine neoplasia type 2B: Report of a case and a literature review of tandem RET mutations with in silico analysis. *Head Neck* **2013**, *35*, E363–E368. [CrossRef] [PubMed]
- Kaltoft, B.; Schmidt, G.; Lauritzen, A.F.; Gimsing, P. Primary localised cutaneous amyloidosis-a systematic review. *Dan. Med. J.* 2013, 60, A4727. [PubMed]
- 74. Zenoaga-Barbăroșie, C.; Berca, L.; Vassu-Dimov, T.; Toma, M.; Nica, M.I.; Alexiu-Toma, O.A.; Ciornei, C.; Albu, A.; Nica, S.; Nistor, C.; et al. The Predisposition for Type 2 Diabetes Mellitus and Metabolic Syndrome. *Balk. J. Med. Genet.* 2023, 26, 21–26. [CrossRef] [PubMed]
- 75. Sanchorawala, V. Systemic Light Chain Amyloidosis. N. Engl. J. Med. 2024, 390, 2295–2307. [CrossRef]
- 76. Konigstein, H. Amyloid der Haut, Jadassohn's Handbuch der Haut und Geschlechtskrankheiten; Verlag von Julius Springer: Berlin, Germany, 1932.

- 77. Black, M.M.; Jones, E.W. Macular amyloidosis. A study of 21 cases with special reference to the role of the epidermis in its histogenesis. *Br. J. Dermatol.* **1971**, *84*, 199–209. [CrossRef]
- 78. Wong, C.K.; Lin, C.S. Friction amyloidosis. Int. J. Dermatol. 1988, 27, 302–307. [CrossRef] [PubMed]
- Bandhlish, A.; Aggarwal, A.; Koranne, R.V. A clinico-epidemiological study of macular amyloidosis from north India. *Indian J. Dermatol.* 2012, 57, 269–274. [CrossRef]
- 80. Fernandez-Flores, A. Cutaneous amyloidosis: A concept review. Am. J. Dermatopathol. 2012, 34, 1–14, quiz 15–17. [CrossRef]
- Haász, C.; Kuroli, E.; Anker, P.; Márton, D.F.; Szigeti, Á.; Nagy, Z.; Demeter, J.; Sárdy, M.; Medvecz, M. Periorbitalis bőrtünetekkel járó könnyűlánc-amyloidosis. *Case report of amyloid light-chain amyloidosis with periocular cutaneous involvement*. Orv. Hetil. 2021, 162, 1303–1308. [CrossRef] [PubMed]
- 82. Aung, T.; Noakes, R.; Murrell, D.F.; Daniel, B.S.; Kelati, A. Primary cutaneous amyloidosis: A review of the available studies and gaps in data. *Australas. J. Dermatol.* 2023, 64, e121–e124. [CrossRef]
- 83. Wang, W.J. Clinical features of cutaneous amyloidoses. Clin. Dermatol. 1990, 8, 13–19. [CrossRef] [PubMed]
- Zychowska, M.; Pięta, K.; Rudy, I.; Skubisz, A.; Reich, A. Dermoscopic Features of Lichen Amyloidosis in Caucasians-A Case Series and Literature Review. *Medicina* 2021, 57, 1027. [CrossRef] [PubMed]
- Hamie, L.; Haddad, I.; Nasser, N.; Kurban, M.; Abbas, O. Primary Localized Cutaneous Amyloidosis of Keratinocyte Origin: An Update with Emphasis on Atypical Clinical Variants. *Am. J. Clin. Dermatol.* 2021, 22, 667–680. [CrossRef]
- 86. Kobayashi, H.; Hashimoto, K. Amyloidogenesis in organ-limited cutaneous amyloidosis: An antigenic identity between epidermal keratin and skin amyloid. *J. Investig. Dermatol.* **1983**, *80*, 66–72. [CrossRef] [PubMed]
- 87. Kitano, Y.; Okada, N.; Kobayashi, Y.; Tanigaki, T.; Okano, M.; Yoshikawa, K. A monoclonal anti-keratin antibody reactive with amyloid deposit of primary cutaneous amyloidosis. *J. Dermatol.* **1987**, *14*, 427–429. [CrossRef] [PubMed]
- Aoki, K.; Ohyama, M.; Mizukawa, Y. A case of lichen amyloidosis associated with atopic dermatitis successfully treated with dupilumab: A case report and literature review. *Dermatol. Ther.* 2021, 34, e15005. [CrossRef] [PubMed]
- Puchtler, H.; Sweat, F. A review of early concepts of amyloid in context with contemporary chemical literature from 1839 to 1859. J. Histochem. Cytochem. 1966, 14, 123–134. [CrossRef]
- Virchow, R.L.K. Weitere Mitteilungen uber das Vorkommen der pflanzlichen cellulose beim Menschen. Arch. Pathol. Anat. Physiol. Klin. Med. 1853, 6, 268–278. [CrossRef]
- 91. Sipe, J.D.; Cohen, A.S. Review: History of the amyloid fibril. J. Struct. Biol. 2000, 130, 88–98. [CrossRef]
- 92. Bonar, L.; Cohen, A.S.; Skinner, M.M. Characterization of the amyloid fibril as a cross-beta protein. *Proc. Soc. Exp. Biol. Med.* **1969**, 131, 1373–1375. [CrossRef]
- 93. Glenner, G.G.; Eanes, E.D.; Bladen, H.A.; Linke, R.P.; Termine, J.D. Beta-pleated sheet fibrils. A comparison of native amyloid with synthetic protein fibrils. *J. Histochem. Cytochem.* **1974**, *22*, 1141–1158. [CrossRef] [PubMed]
- Sunde, M.; Blake, C. The structure of amyloid fibrils by electron microscopy and X-ray diffraction. *Adv. Protein Chem.* 1997, 50, 123–159. [CrossRef] [PubMed]
- Malone, M.A.V.; Castillo, D.A.A.; Santos, H.T.; Kaur, A.; Elrafei, T.; Steinberg, L.; Kumar, A. A systematic review of the literature on localized gastrointestinal tract amyloidosis: Presentation, management and outcomes. *Eur. J. Haematol.* 2024. [CrossRef] [PubMed]
- 96. Abbas, A.K. Diseases of immunity. In *Robbins and Cotran Pathologic Basis of Disease;* Kumar, V., Abbas, A.K., Fausto, N., Eds.; Elsevier Saunders: Philadelphia, PA, USA, 1999; pp. 193–267.
- 97. Ghiso, J.; Frangione, B. Amyloidosis and Alzheimer's disease. Adv. Drug Deliv. Rev. 2002, 54, 1539–1551. [CrossRef]
- van der Hilst, J.; van der Meer, J.; Drenth, J.; Simon, A. AL amyloidosis enhances development of amyloid A amyloidosis. Br. J. Dermatol. 2007, 156, 748–749. [CrossRef]
- 99. Merlini, G.; Bellotti, V. Molecular mechanisms of amyloidosis. N. Engl. J. Med. 2003, 349, 583–596. [CrossRef]
- Roy, S.; Srinivasan, V.R.; Arunagiri, S.; Mishra, N.; Bhatia, A.; Shejale, K.P.; Prajapati, K.P.; Kar, K.; Anand, B.G. Molecular insights into the phase transition of lysozyme into amyloid nanostructures: Implications of therapeutic strategies in diverse pathological conditions. *Adv. Colloid. Interface Sci.* 2024, 331, 103205. [CrossRef]
- 101. Drago, F.; Ranieri, E.; Pastorino, A.; Casazza, S.; Crovato, F.; Rebora, A. Epstein-Barr virus-related primary cutaneous amyloidosis. *Successful treatment with acyclovir and interferon-alpha. Br. J. Dermatol.* **1996**, *134*, 170–174. [PubMed]
- Chang, Y.T.; Liu, H.N.; Wong, C.K.; Chow, K.C.; Chen, K.Y. Detection of Epstein-Barr virus in primary cutaneous amyloidosis. *Br. J. Dermatol.* 1997, 136, 823–826. [CrossRef]
- 103. Patra, P.; Rani, A.; Sharma, N.; Mukherjee, C.; Jha, H.C. Unraveling the Connection of Epstein-Barr Virus and Its Glycoprotein M(146-157) Peptide with Neurological Ailments. ACS Chem. Neurosci. 2023, 14, 2450–2460. [CrossRef]
- Nistor, C.E.; Pantile, D.; Gavan, C.S.; Ciuche, A. Pneumothorax on COVID-19 patients-retrospective clinical observations. *Rom. J. Leg. Med.* 2022, 30, 112–116. [CrossRef]
- 105. Beretti, F.; Gatti, M.; Ricchi, F.; Lipani, F.; Cortelli, P.; Cermelli, C.; Maraldi, T. Neurotoxic effects of coronavirus: Potential implications in Alzheimer's onset and progression. *Exp. Neurol.* **2024**, 114908. [CrossRef] [PubMed]
- Milton, N.G.N. SARS-CoV-2 amyloid, is COVID-19-exacerbated dementia an amyloid disorder in the making? *Front. Dement.* 2023, 2, 1233340. [CrossRef]
- 107. Nistor, C.E.; Gavan, C.S.; Pantile, D.; Tanase, N.V.; Ciuche, A. Cervico-Thoracic Air Collections in COVID-19 Pneumonia Patients-Our Experience and Brief Review. *Chirurgia* **2022**, *117*, 317–327. [CrossRef]

- 108. Lu, P.; Wu, F.F.; Rong, Z.L.; Fang, C.; Deng, C.C.; Bin, L.H.; Yang, B. Clinical and genetic features of Chinese patients with lichen and macular primary localized cutaneous amyloidosis. *Clin. Exp. Dermatol.* **2019**, *44*, e110–e117. [CrossRef]
- 109. Breathnach, S.M. Amyloid and amyloidosis. J. Am. Acad. Dermatol. 1988, 18 1 Pt 1, 1–16. [CrossRef]
- 110. Ueo, D.; Utani, A.; Okubo, Y.; Yozaki, M.; Mine, Y.; Anan, T.; Nishida, H.; Takahashi, D.; Sakai, T.; Hatano, Y.; et al. Familial primary localized cutaneous amyloidosis in a Japanese family. *J. Dermatol. Sci.* **2016**, *83*, 162–164. [CrossRef] [PubMed]
- 111. Arita, K.; South, A.P.; Hans-Filho, G.; Sakuma, T.H.; Lai-Cheong, J.; Clements, S.; Odashiro, M.; Odashiro, D.N.; Hans-Neto, G.; Hans, N.R.; et al. Oncostatin M receptor-beta mutations underlie familial primary localized cutaneous amyloidosis. *Am. J. Hum. Genet.* 2008, *82*, 73–80. [CrossRef] [PubMed]
- 112. Lin, M.W.; Lee, D.D.; Liu, T.T.; Lin, Y.F.; Chen, S.Y.; Huang, C.C.; Weng, H.Y.; Liu, Y.F.; Tanaka, A.; Arita, K.; et al. Novel IL31RA gene mutation and ancestral OSMR mutant allele in familial primary cutaneous amyloidosis. *Eur. J. Hum. Genet.* 2010, *18*, 26–32. [CrossRef]
- 113. Chia, B.; Tan, A.; Tey, H.L. Primary localized cutaneous amyloidosis: Association with atopic dermatitis. *J. Eur. Acad. Dermatol. Venereol.* **2014**, *28*, 810–813. [CrossRef]
- 114. Pálla, S.; Kuroli, E.; Tóth, E.A.; Hidvégi, B.; Holló, P.; Medvecz, M. Primary Localized Cutaneous Amyloidosis in Central Europe: A Retrospective Monocentric Study on Epidemiology and Therapy. J. Clin. Med. **2023**, 12, 7672. [CrossRef] [PubMed]
- 115. Adams, R.; Colmont, C.; Mukhtar, A.; Morgan, H.; Patel, G.K. A novel oncostatin M/interleukin-31 receptor mutation in familial primary localized cutaneous amyloidosis. *Clin. Exp. Dermatol.* **2020**, *45*, 254–256. [CrossRef]
- 116. Wali, A.; Liu, L.; Takeichi, T.; Jelani, M.; Rahman, O.U.; Heng, Y.K.; Thng, S.; Lee, J.; Akiyama, M.; McGrath, J.A.; et al. Familial primary localized cutaneous amyloidosis results from either dominant or recessive mutations in OSMR. *Acta Derm. Venereol.* 2015, 95, 1005–1007. [CrossRef]
- 117. Weidner, T.; Illing, T.; Elsner, P. Primary Localized Cutaneous Amyloidosis: A Systematic Treatment Review. *Am. J. Clin. Dermatol.* **2017**, *18*, 629–642. [CrossRef]
- 118. Ruzicka, T.; Hanifin, J.M.; Furue, M.; Pulka, G.; Mlynarczyk, I.; Wollenberg, A.; Galus, R.; Etoh, T.; Mihara, R.; Yoshida, H.; et al. Anti-Interleukin-31 Receptor A Antibody for Atopic Dermatitis. *N. Engl. J. Med.* **2017**, *376*, 826–835. [CrossRef]
- 119. Tidman, M.J.; Wells, R.S.; MacDonald, D.M. Pachyonychia congenita with cutaneous amyloidosis and hyperpigmentation--a distinct variant. *J. Am. Acad. Dermatol.* **1987**, *16 5 Pt 1*, 935–940. [CrossRef]
- 120. Verga, U.; Fugazzola, L.; Cambiaghi, S.; Pritelli, C.; Alessi, E.; Cortelazzi, D.; Gangi, E.; Beck-Peccoz, P. Frequent association between MEN 2A and cutaneous lichen amyloidosis. *Clin. Endocrinol.* **2003**, *59*, 156–161. [CrossRef] [PubMed]
- 121. Scapineli, J.O.; Ceolin, L.; Puñales, M.K.; Dora, J.M.; Maia, A.L. MEN 2A-related cutaneous lichen amyloidosis: Report of three kindred and systematic literature review of clinical, biochemical and molecular characteristics. *Fam. Cancer* 2016, 15, 625–633. [CrossRef] [PubMed]
- 122. Chabre, O.; Labat, F.; Pinel, N.; Berthod, F.; Tarel, V.; Bachelot, I. Cutaneous lesion associated with multiple endocrine neoplasia type 2A: Lichen amyloidosis or notalgia paresthetica? *Henry Ford Hosp. Med. J.* **1992**, *40*, 245–248.
- Astwasturov, M.I. Uber paresthetische neuralgien un eine besondere form derselben-notalgia paresthetica. Dtsch. Z. Nervenheilkd. 1934, 133, 198. [CrossRef]
- 124. Weber, P.J.; Poulos, E.G. Notalgia paresthetica: Case reports and histologic appraisal. J. Am. Acad. Dermatol. 1988, 18 1 Pt 1, 25–30. [CrossRef]
- 125. Comings, D.E.; Comings, S.N. Hereditary localized pruritus. Arch. Dermatol. 1965, 92, 236–237. [CrossRef] [PubMed]
- 126. Gibbs, R.C.; Frank, S.B. A peculiar, spotty pigmentation: Report of five cases. Dermatol. Int. 1969, 8, 14–16. [CrossRef] [PubMed]
- 127. Le Douarin, N.M. Cell line segregation during peripheral nervous system ontogeny. *Science* **1986**, *231*, 1515–1522. [CrossRef] [PubMed]
- 128. Kousseff, B.G.; Espinoza, C.; Zamore, G.A. Sipple syndrome with lichen amyloidosis as a paracrinopathy: Pleiotropy, heterogeneity, or a contiguous gene? *J. Am. Acad. Dermatol.* **1991**, 25, 651–657. [CrossRef] [PubMed]
- 129. Airaksinen, M.S.; Titievsky, A.; Saarma, M. GDNF family neurotrophic factor signaling: Four masters, one servant? *Mol. Cell. Neurosci.* **1999**, *13*, 313–325. [CrossRef]
- Bennett, D.L.; Michael, G.J.; Ramachandran, N.; Munson, J.B.; Averill, S.; Yan, Q.; McMahon, S.B.; Priestley, J.V. A distinct subgroup of small DRG cells express GDNF receptor components and GDNF is protective for these neurons after nerve injury. *J. Neurosci.* 1998, *18*, 3059–3072. [CrossRef]
- 131. Molliver, D.C.; Wright, D.E.; Leitner, M.L.; Parsadanian, A.S.; Doster, K.; Wen, D.; Yan, Q.; Snider, W.D. IB4-binding DRG neurons switch from NGF to GDNF dependence in early postnatal life. *Neuron* **1997**, *19*, 849–861. [CrossRef]
- Butler, D.C.; Berger, T.; Elmariah, S.; Kim, B.; Chisolm, S.; Kwatra, S.G.; Mollanazar, N.; Yosipovitch, G. Chronic Pruritus: A Review. JAMA 2024, 331, 2114–2124. [CrossRef]
- 133. Robinson, C.; Downs, E.; De la Caridad Gomez, Y.; Nduaguba, C.; Woolley, P.; Varrassi, G.; Gill, J.; Simopoulos, T.T.; Viswanath, O.; Yazdi, C.A. Notalgia Paresthetica Review: Update on Presentation, Pathophysiology, and Treatment. *Clin. Pract.* 2023, 13, 315–325. [CrossRef]
- 134. Ansari, A.; Weinstein, D.; Sami, N. Notalgia paresthetica: Treatment review and algorithmic approach. *J. Dermatol. Treat.* **2020**, *31*, 424–432. [CrossRef] [PubMed]
- 135. Šitum, M.; Kolić, M.; Franceschi, N.; Pećina, M. Notalgia paresthetica. Acta Clin. Croat. 2018, 57, 721–725. [CrossRef]

- 136. Gagel, R.F.; Levy, M.L.; Donovan, D.T.; Alford, B.R.; Wheeler, T.; Tschen, J.A. Multiple endocrine neoplasia type 2a associated with cutaneous lichen amyloidosis. *Ann. Intern. Med.* **1989**, *111*, 802–806. [CrossRef] [PubMed]
- 137. Singh, D.N.; Gupta, S.; Agarwal, A. Cutaneous lichen amyloidosis associated with multiple endocrine neoplasia type 2A: An early clinical marker. *World J. Endocr. Surg.* **2012**, *4*, 41. [CrossRef]
- 138. Qi, X.-P.; Zhao, J.-Q.; Chen, Z.-G.; Cao, J.-L.; Du, J.; Liu, N.-F.; Li, F.; Sheng, M.; Fu, E.; Guo, J.; et al. The clinical spectrum of multiple endocrine neoplasia type 2A with cutaneous lichen amyloidosis in ethnic Han Chinese. *Cancer Investig.* 2018, 36, 141–151. [CrossRef] [PubMed]
- Qi, X.P.; Zhao, J.Q.; Chen, Z.G.; Cao, J.L.; Du, J.; Liu, N.F.; Li, F.; Sheng, M.; Fu, E.; Guo, J.; et al. RET mutation pS891A in a Chinese family with familial medullary thyroid carcinoma associated cutaneous amyloidosis binding OSMR variant, p.G513D. *Oncotarget* 2015, *6*, 33993–34003. [CrossRef]
- 140. Qi, X.P.; Peng, J.Z.; Yang, X.W.; Zao, Z.L.; Yu, X.H.; Fang, X.D.; Zhang, D.H.; Zhao, J.Q. The RET C611Y mutation causes MEN 2A and associated cutaneous. *Endocr. Connect.* 2018, 7, 998–1005. [CrossRef]
- 141. Elisei, R.; Alevizaki, M.; Conte-Devolx, B.; Frank-Raue, K.; Leite, V.; Williams, G.R. 2012 European thyroid association guidelines for genetic testing and its clinical consequences in medullary thyroid cancer. *Eur. Thyroid J.* **2013**, *1*, 216–231. [CrossRef]
- 142. Seri, M.; Celli, I.; Betsos, N.; Claudiani, F.; Camera, G.; Romeo, G. A Cys634Gly substitution of the RET proto-oncogene in a family with recurrence of multiple endocrine neoplasia type 2A and cutaneous lichen amyloidosis. *Clin. Genet.* **1997**, *51*, 86–90. [CrossRef]
- 143. Ferrer, J.P.; Halperin, I.; Conget, J.I.; Alsina, M.; Martinez-Osaba, M.J.; Palou, J.; Bombi, J.A.; Vilardell, E. Primary localized cutaneous amyloidosis and familial medullary thyroid carcinoma. *Clin. Endocrinol.* **1991**, *34*, 435–439. [CrossRef]
- 144. Gullu, S.; Gursoy, A.; Erdogan, M.F.; Dizbaysak, S.; Erdogan, G.; Kamel, N. Multiple endocrine neoplasia type 2A/localized cutaneous lichen amyloidosis associated with malignant pheochromocytoma and ganglioneuroma. *J. Endocrinol. Investig.* 2005, 28, 734–737. [CrossRef] [PubMed]
- 145. Karga, H.J.; Karayianni, M.K.; Linos, D.A.; Tseleni, S.C.; Karaiskos, K.D.; Papapetrou, P.D. Germ line mutation analysis in families with multiple endocrine neoplasia type 2A or familial medullary thyroid carcinoma. *Eur. J. Endocrinol.* **1998**, 139, 410–415. [CrossRef] [PubMed]
- 146. Abdullah, F.; Udelsman, R. Cutaneous lichen amyloidosis in a family with familial medullary thyroid cancer. *Surgery* **2004**, *135*, 563–564. [CrossRef] [PubMed]
- 147. Fang, X.; Wang, H.; Dong, F.; Lian, B.; Li, F.; Jin, H.; Yu, Y.; Zhang, N.; Qi, X. [Clinical and genetic analysis of seven Chinese pedigrees affected with multiple endocrine neoplasia type 2A with cutaneous lichen amyloidosis]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* 2022, 39, 938–943. [CrossRef]
- Tang, H.X.; Yang, H.; Li, F.; Cao, Z.L.; Huang, Y.T.; Qi, X.P. Elevated basal serum levels of calcitonin and simultaneous surgery of MEN2A-specific tumors. *Neoplasma* 2021, 68, 1098–1106. [CrossRef]
- 149. Malhotra, R.; Boro, H.; Shamim, S.A.; Khadgawat, R. Multiple endocrine neoplasia type 2A with cutaneous lichen amyloidosis. BMJ Case Rep. 2020, 13, e238423. [CrossRef]
- 150. Pal, R.; Rastogi, A.; Kumar, S.; Bhansali, A. Metastatic pheochromocytoma in MEN 2A: A rare association. *BMJ Case Rep.* 2018, 2018, bcr2017222758. [CrossRef]
- 151. Zhang, X.W.; Wang, J.Y.; Zhang, Y.B.; Wan, H.F.; Zhang, B.; Yan, D.G.; Liu, W.S.; Xu, Z.G.; Tang, P.Z. Genotype-phenotype correlations in multiple endocrine neoplasia type 2. *Zhonghua Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 2016, *51*, 538–541. [CrossRef]
- 152. Lemos, M.C.; Carrilho, F.; Rodrigues, F.J.; Cavalheiro, M.; Regateiro, F.J.; Ruas, M.M. Caracterização molecular de uma família com nem2a e suas implicações clínicas Molecular characterisation of a kindred with MEN2A and clinical implications. *Acta Med. Port.* **2003**, *16*, 245–250.
- 153. Vieira, A.E.; Mello, M.P.; Elias, L.L.; Lau, I.F.; Maciel, L.M.; Moreira, A.C.; Castro, M. Molecular and biochemical screening for the diagnosis and management of medullary thyroid carcinoma in multiple endocrine neoplasia type 2A. *Horm. Metab. Res.* 2002, 34, 202–206. [CrossRef]
- Lemos, M.C.; Carrilho, F.; Rodrigues, F.J.; Santos, P.; Carvalheiro, M.; Ruas, M.A.; Regateiro, F.J. Early onset of medullary thyroid carcinoma in a kindred with multiple endocrine neoplasia type iia associated with cutaneous lichen amyloidosis. *Endocr. Pract.* 2002, *8*, 19–22. [CrossRef] [PubMed]
- 155. Pacini, F.; Fugazzola, L.; Bevilacqua, G.; Viacava, P.; Nardini, V.; Martino, E. Multiple endocrine neoplasia type 2A and cutaneous lichen amyloidosis: Description of a new family. *J. Endocrinol. Investig.* **1993**, *16*, 295–296. [CrossRef] [PubMed]
- 156. Nunziata, V.; di Giovanni, G.; Lettera, A.M.; D'Armiento, M.; Mancini, M. Cutaneous lichen amyloidosis associated with multiple endocrine neoplasia type 2A. *Henry Ford Hosp. Med J.* **1989**, *37*, 144–146. [PubMed]
- 157. Csikós, M.; Orosz, Z.; Bottlik, G.; Szöcs, H.; Szalai, Z.; Rozgonyi, Z.; Hársing, J.; Török, E.; Bruckner-Tuderman, L.; Horváth, A.; et al. Dystrophic epidermolysis bullosa complicated by cutaneous squamous cell carcinoma and pulmonary and renal amyloidosis. *Clin. Exp. Dermatol.* **2003**, *28*, 163–166. [CrossRef]
- 158. Li, C.; Zhang, P.; Nie, R.; Gong, X.; Xie, J.; Yu, Z.; Wang, C.; Zhang, H.; Yan, R.; Lu, Z. Targeting Amyloids with [(18)F]AV-45 for Medullary Thyroid Carcinoma Positron Emission Tomography/Computed Tomography Imaging: A Pilot Clinical Study. *Mol. Pharm.* 2022, 19, 584–591. [CrossRef] [PubMed]
- 159. Pinto, A.; Nosé, V. Localized amyloid in thyroid: Are we missing it? Adv. Anat. Pathol. 2013, 20, 61–67. [CrossRef]

- 160. Lacout, A.; Isaac, S.; Marcy, P.Y. Micro-medullary thyroid carcinoma: A diagnosis not to be missed. *Postgrad. Med. J.* 2015, *91*, 236–237. [CrossRef]
- 161. Cheung, S.T.; Maheshwari, M.B.; Tan, C.Y. A comparative study of two Congo red stains for the detection of primary cutaneous amyloidosis. J. Am. Acad. Dermatol. 2006, 55, 363–364. [CrossRef] [PubMed]
- 162. Schepis, C.; Siragusa, M.; Gagliardi, M.E.; Torre, V.; Cicciarello, R.; Albiero, F.; Cavallari, V. Primary macular amyloidosis: An ultrastructural approach to diagnosis. *Ultrastruct. Pathol.* **1999**, *23*, 279–284. [CrossRef]
- 163. Terao, M.; Nishida, K.; Murota, H.; Katayama, I. Clinical effect of tocoretinate on lichen and macular amyloidosis. *J. Dermatol.* **2011**, *38*, 179–184. [CrossRef]
- 164. Stull, C.M.; Tey, H.L.; Yosipovitch, G. Methotrexate for the Treatment of Recalcitrant Primary Localized Cutaneous Amyloidosis: A Case Series. *Acta Derm. Venereol.* **2018**, *98*, 900–901. [CrossRef] [PubMed]
- 165. Zeidler, C.; Metze, D.; Ständer, S. Successful treatment of lichen amyloidosis using capsaicin 8% patch. *J. Eur. Acad. Dermatol. Venereol.* **2016**, *30*, 1236–1238. [CrossRef] [PubMed]
- 166. Lee, J.Y.; Park, M.Y.; Ahn, J.Y. A case of lichen amyloidosis improved by topical salicylic acid and topical corticosteroid. *Korean J. Dermatol.* **2010**, *48*, 33–536.
- 167. Shoen, E.; Hou, A.; Zahn, J.; Friedman, A. Successful Treatment of Lichen Amyloidosis Using a Fixed Combination of Halobetasol-Propionate and Tazarotene Lotion. *J. Drugs Dermatol.* **2021**, *20*, 336–337. [CrossRef]
- 168. Esmat, S.M.; Fawzi, M.M.; Gawdat, H.I.; Ali, H.S.; Sayed, S.S. Efficacy of different modes of fractional CO₂ laser in the treatment of primary cutaneous amyloidosis: A randomized clinical trial. *Lasers Surg. Med.* **2015**, *47*, 388–395. [CrossRef]
- Norisugi, O.; Yamakoshi, T.; Shimizu, T. Successful treatment of lichen amyloidosis using a CO₂ surgical laser. *Dermatol. Ther.* 2014, 27, 71–73. [CrossRef] [PubMed]
- 170. Kalkan, G.; Markoç, F.; Bas, Y. An alternative treatment model: The combination therapy of narrow band ultraviolet B phototherapy and tacrolimus ointment 0.1% in biphasic amyloidosis. *J. Pak. Med. Assoc.* **2014**, *64*, 579–582.
- 171. Alonso-González, J.; Rodríguez-Granados, M.T.; Toribio, J. Satisfactory response to narrowband UV-B therapy in generalized lichen amyloidosis. *Actas Dermosifiliogr.* **2013**, *104*, 527–529. [CrossRef]
- 172. Becker, S.; Kahn, D.; Rothman, S. Cutaneous manifestations of internal malignant tumors. *Arch. Dermatol.* **1942**, *45*, 1069–1080. [CrossRef]
- 173. Yu, S.; Ge, M.; Zhang, C.; Chen, L.; Zhao, L. Diagnosis and comprehensive treatment of a glucagonoma in a patient with residual intrahepatic metastases postoperatively: A case report and literature review. *Oncol. Lett.* **2024**, *27*, 202. [CrossRef]
- 174. Anghel, A.; Stanciu, S.; Ciobica, M.L.; Stoicescu, D.; Muresan, M.M. Contrast-enhanced ultrasound-clinical applications. *Rom. J. Mil. Med.* **2011**, *114*, 25–30.
- 175. Anderson, S. Café au Lait Macules and Associated Genetic Syndromes. J. Pediatr. Health Care 2020, 34, 71–81. [CrossRef] [PubMed]
- 176. Nistor, C.E.; Ciuche, A.; Cucu, A.P.; Serban, B.; Cursaru, A.; Cretu, B.; Cirstoiu, C. Clavicular Malignancies: A Borderline Surgical Management. *Medicina* 2022, 58, 910. [CrossRef] [PubMed]
- 177. Waguespack, S.G. Beyond the "3 Ps": A critical appraisal of the non-endocrine manifestations of multiple endocrine neoplasia type 1. *Front. Endocrinol.* **2022**, *13*, 1029041. [CrossRef] [PubMed]
- 178. Nistor, C.; Ranetti, A.E.; Ciuche, A.; Pantile, D.; Constatin, L.M.; Brincoveanu, R. Betadine in chemical pleurodesis. *Farmacia* **2014**, 62, 897–906.
- Pierotti, L.; Pardi, E.; Dinoi, E.; Piaggi, P.; Borsari, S.; Della Valentina, S.; Sardella, C.; Michelucci, A.; Caligo, M.A.; Bogazzi, F.; et al. Cutaneous lesions and other non-endocrine manifestations of Multiple Endocrine Neoplasia type 1 syndrome. *Front. Endocrinol.* 2023, 14, 1191040. [CrossRef]
- 180. Stanciu, S.; Enciu, C.; Raduta, I.; Stoicescu, D.; Anghel, A.; Anghel, D.; Olan, B.; Ciobica, L. The role of contrast-enhanced ultrasound in risk assessment of carotid atheroma. *Rom. J. Mil. Med.* **2016**, *119*, 9–11. [CrossRef]
- 181. Rastogi, V.; Singh, D.; Mazza, J.J.; Parajuli, D.; Yale, S.H. Flushing Disorders Associated with Gastrointestinal Symptoms: Part 1, Neuroendocrine Tumors, Mast Cell Disorders and Hyperbasophila. *Clin. Med. Res.* **2018**, *16*, 16–28. [CrossRef]
- Nistor, C.E.; Găvan, C.S.; Ciritel, A.A.; Nemes, A.F.; Ciuche, A. The Association of Minimally Invasive Surgical Approaches and Mortality in Patients with Malignant Pleuropericarditis-A 10 Year Retrospective Observational Study. *Medicina* 2022, 58, 718. [CrossRef]
- 183. Grozinsky-Glasberg, S.; Davar, J.; Hofland, J.; Dobson, R.; Prasad, V.; Pascher, A.; Denecke, T.; Tesselaar, M.E.T.; Panzuto, F.; Albåge, A.; et al. European Neuroendocrine Tumor Society (ENETS) 2022 Guidance Paper for Carcinoid Syndrome and Carcinoid Heart Disease. J. Neuroendocrinol. 2022, 34, e13146. [CrossRef]
- 184. Hannah-Shmouni, F.; Stratakis, C.A.; Koch, C.A. Flushing in (neuro) endocrinology. Rev. Endocr. Metab. Disord. 2016, 17, 373–380. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.