

Cerebral ventriculitis associated with *Achromobacter xylosoxidans*

SHIRO SHIGETA, YOSHIO YASUNAGA, KEN HONZUMI, HIROKO OKAMURA, ROKURO KUMATA, AND SHINICHIRO ENDO

From the Central Clinical Laboratory and 2nd Department of Surgery, Fukushima Medical College, Fukushima, Japan

SUMMARY Six patients in the neurosurgical ward of Fukushima Medical College Hospital suffering from ventriculitis due to *Achromobacter xylosoxidans* infection had undergone craniotomy or cranial trepanation before the infection. The strains of *A. xylosoxidans* isolated from the patients were resistant to streptomycin, ampicillin, cephaloridine, gentamicin, and colistin. They were also resistant to chlorhexidine digluconate (Hibitane) in a concentration of 2%. When a study of the chlorhexidine used in the hospital was carried out four strains of *A. xylosoxidans* were isolated from 20 containers of chlorhexidine solution in the surgical ward but not from those in the operating theatre.

Achromobacter xylosoxidans, a nonfermentative, Gram-negative, peritrichous rod, was named and described by Yabuuchi and Ohyama (1971) in isolates from patients with chronic otitis media. The homogeneity of *A. xylosoxidans* was established as a result of Yabuuchi's subsequent study of 55 strains in which the uniformities of flagellar morphology, biochemical reaction, and percentage of guanine and cytosine mole in deoxyribonucleic acid were confirmed (Yabuuchi *et al.*, 1974). Although *A. xylosoxidans* has been recovered from a variety of human clinical sources, including blood, bile, and cerebrospinal fluid (CSF), its role as the primary aetiological agent of meningitis was proved and reported by us as a first case report (Shigeta *et al.*, 1974). This paper describes the recovery of *A. xylosoxidans* from the CSF of six patients with ventricular infection after neurosurgical operations.

Material and methods

BACTERIOLOGICAL SURVEY

All patients admitted to the neurosurgical ward of Fukushima Medical College Hospital, from whom *A. xylosoxidans* and other species of organisms were isolated from the CSF during the period May 1973 to February 1975, were included in this survey. CSF was the sole material for culture, and isolates

were identified using triple-sugar iron (TSI) agar, oxidation, and fermentation of sugars and decarboxylase media devised by Møller (1955).

In addition, production of water- or chloroform-soluble pigments, nitrate reduction, motility, and flagellar morphology were determined. The programme of identification was essentially that of Yabuuchi *et al.* (1974). Organisms were considered to cause infection only when a pure culture of *A. xylosoxidans* was obtained from samples of CSF taken by ventricular puncture or drained from the ventricles, and when there was clinical evidence of meningitis. Newly isolated strains of *A. xylosoxidans*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *Klebsiella pneumoniae* were subcultured on Dorset's egg medium (Nissui Ltd, Tokyo) and stored for one to three months. Before testing for sensitivity and resistance of the organisms, all strains were inoculated in brain-heart infusion (BHI) broth and cultured at 37°C for 24 hours.

SENSITIVITY TO ANTIBIOTICS

Sensitivity to antibiotics was tested using the single disc method of Bauer *et al.* (1966). Commercially purchased discs (Showa Pharmaceuticals Ltd, Tokyo) and Müller-Hinton agar (BBL) were used in the test.

RESISTANCE TO CHEMICAL DISINFECTION

The resistance of *A. xylosoxidans* to disinfection was

Received for publication 17 November 1976

tested by the addition of about 10^8 organisms to 1 ml of several dilutions of disinfectant. As a control, the same number of organisms was added to distilled water without disinfectant. The mixtures of the organisms and disinfectant or distilled water were kept at room temperature for 10 minutes and were then diluted to 10^{-2} with sterile saline, and 0.01 ml of the dilution was inoculated in BHI broth.

In each test 4×10^3 to 3×10^4 organisms were inoculated in broth, and all test cultures were divided into two sets and then incubated at 25°C and 37°C. After 48 hours' incubation the cultures were examined for growth of organisms, and the resistance of the organisms was judged by their survival in the dilution of disinfectant after treatment. In another experiment the numbers of survivors were assessed by inoculation on a BHI agar plate after treatment.

INVESTIGATION FOR SOURCE OF INFECTION

Investigation of the chlorhexidine solution prepared in the operating theatre and in the surgical ward was carried out by inoculating a 1-ml aliquot of sample from the containers into 10 ml of BHI broth. Duplicate broth tubes were used and divided into two sets and were then incubated at 25°C and 37°C for 10 days. On the last day of culture, one drop of broth was subcultured on BHI agar containing 5% sheep erythrocytes and was then incubated at 37°C for 48 hours to isolate the contaminating organisms.

Results

CASE REPORTS

Case 1

A girl aged 9 was admitted to the neurosurgical ward on 7 March, 1973 for operation on an arachnoid cyst in the occipital region. On the tenth postoperative day she underwent a ventriculoperitoneal shunt to reduce the pressure of the CSF in the ventricles. On 19 May she developed fever (39°C) with continuous headache, frequent vomiting, and stiffness of the neck. On 29 May, ventricular puncture was performed at the site of the ventricular tap which connected the ventricles to the peritoneal shunt. The puncture yielded purulent CSF, and *A. xylosoxidans* was isolated from the fluid in pure culture. The patient received injections of chloramphenicol (1 g daily) and the infection improved temporarily but later recurred three times in September, November, and December of the same year. Three of the four relapses were associated with *A. xylosoxidans* infection. Sera tested for the

agglutinating antibody to the isolates on 15 June and 6 September had titres of 1 in 320 and 1 in 640.

Case 2

This 6-month-old boy had a haematoma in the occipital region after birth in October 1973. A swelling gradually developed associated with redness and fever. In February 1974 a cerebral abscess was diagnosed. Incision of the abscess yielded a purulent fluid from which *Proteus mirabilis* was recovered. After the incision, two drains were inserted in the abscess and in the corresponding ventricles of the cerebrum. In spite of extensive chemotherapy with daily injections of cephaloridine (1.0 g), the infection persisted for two months and on 16 April, *A. xylosoxidans* was isolated in pure culture not only from pus in the abscess but also from CSF drained from the ventricle. The organism was sensitive to chloramphenicol, ampicillin, carbenicillin, and nalidixic acid. Daily injections of chloramphenicol (500 mg) were continued for two weeks and he recovered from the infection.

Case 3

A 9-year-old boy with hydrocephalus had a cranial trepanation, and a drain was inserted in the ventricle on 10 July 1973. After the operation he was pyrexial (38.4°C) and three days after the operation *A. xylosoxidans* was isolated from CSF drained from the ventricle.

Case 4

This 8-year-old boy with a thalamic tumour had a ventriculoperitoneal shunt operation. After the operation he became pyrexial. One month later, a ventricular puncture yielded a turbid CSF from which *A. xylosoxidans* was isolated. After a 6-day course of chloramphenicol (500 mg daily), there was no growth of *A. xylosoxidans* but *Serratia marcescens* was isolated on culture.

Case 5

A 63-year-old man had a radical operation for multiple cerebral aneurysms performed on 10 July 1974. After the operation a drain was inserted in the ventricle. On the sixth day after operation, he had a fever of 39°C and *A. xylosoxidans* was isolated from the CSF in pure culture; on the following day the same organism was again isolated. These organisms were shown to be sensitive to tetracycline, chloramphenicol, and nalidixic acid but completely resistant to gentamicin, colistin, kanamycin, carbenicillin, and cephaloridine. He made a complete recovery from the infection after a two-week course of chloramphenicol injections (1.0 g daily).

Case 6

A man aged 49 was admitted to the surgical ward with a cerebral tumour on 9 September 1974. On 5 February 1975 he underwent craniotomy, and part of the tumour, shown histologically to be a meningioma, was resected. After the operation he had a fever of 39°C which continued for several days. On 11 February ventricular puncture yielded a turbid CSF from which *A. xylosoxidans* was isolated.

**INCIDENCE OF BACTERIAL VENTRICULITIS
IN THE NEUROSURGICAL WARD**

From May 1973 to February 1975, 11 patients were admitted to the neurosurgical ward of Fukushima Medical College Hospital suffering from bacterial ventriculitis. Craniotomy or cranial trepanation had been performed on all the patients for resection of a tumour or incision of an abscess cavity. Each patient also had a ventriculoperitoneal shunt or drain from the ventricle.

Six of the 11 patients were infected with *A. xylosoxidans*, and the details of those cases have been cited above. Other species of organisms recovered from the CSF of these patients were *Enterobacter*, *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Ps. aeruginosa*, *P. mirabilis*, *S. marcescens*, *Acinetobacter calcoaceticus*, and *K. pneumoniae*.

Some patients relapsed with the same infection or with an infection caused by different species of organisms. If such relapses are considered as individual cases, there were 18 cases of ventriculitis among 11 patients during the period of the survey. The causative organisms in these cases are shown in Table 1. Eight of 18 were *A. xylosoxidans* and three others were nonfermentative rods (2 *A. calcoaceticus* and 1 *Ps. aeruginosa*). Eleven *A. xylosoxidans* strains were isolated from the CSF of these patients and some characteristics of the isolated strains and type strain ATCC-27061 of

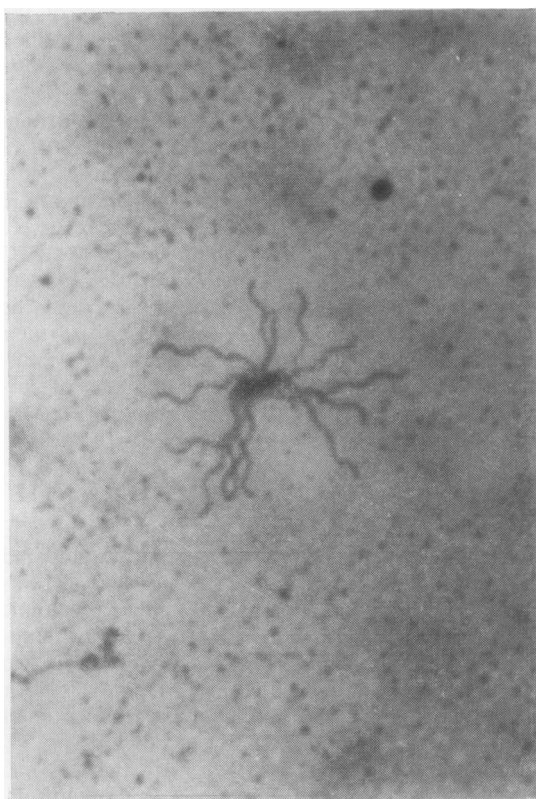


Figure *Achromobacter xylosoxidans* F-1482 isolated from cerebrospinal fluid of case 1. Leifson's flagellar staining after formalin fixation. $\times 1000$.

A. xylosoxidans are shown in Table 2 and the Figure.

**SENSITIVITY OF *A. XYLOSOXIDANS* TO
ANTIBIOTICS**

Eleven strains of *A. xylosoxidans* isolated from CSF were tested for sensitivity to antibiotics. As shown in Table 3, all strains were resistant to streptomycin, cephaloridine, and ampicillin. Ten strains were resistant to gentamicin and eight strains were resistant to colistin. However, more than half of all the strains were sensitive to tetracycline, chloramphenicol, nalidixic acid, and carbenicillin.

**RESISTANCE OF *A. XYLOSOXIDANS* TO
CHEMICAL DISINFECTION**

It was conceivable that the *A. xylosoxidans* infection in these patients might have been due to the resistance of organisms to, and failure of, chemical disinfection. In this hospital, chlorhexidine (Hibitane) was the most popular disinfectant for washing the

Table 1 *Causative organisms in 18 cases of bacterial meningitis*

Causative organism	Cases of infection*	
	No.	%
<i>Achromobacter xylosoxidans</i>	8	44.4
<i>Enterobacter</i> spp	2	11.1
<i>Acinetobacter calcoaceticus</i>	2	11.1
<i>Serratia marcescens</i>	1	5.6
<i>Staphylococcus aureus</i>	1	5.6
<i>Staphylococcus epidermidis</i>	1	5.6
<i>Pseudomonas aeruginosa</i>	1	5.6
<i>Proteus mirabilis</i>	1	5.6
<i>Klebsiella pneumoniae</i>	1	5.6
Total	18	

*Relapses of infection with the same or different species of organisms were considered as individual cases.

Table 2 Characteristics of the type strain of *A. xylosoxidans* and of 11 isolated strains

Substrates and tests	Type strain ATCC-27061	No. of positive strains
Gram negative, rod shaped	+	11
Motility	+	11
Peritrichous flagella	+	11
Growth on NAC agar	+	11
Indophenol oxidase	+	11
Citrate, Simmons	+	11
OF-glucose medium open, acid	+	11 ₃₋₄
OF-glucose medium sealed, acid	-	0
Oxidative acid production from*		
Fructose	-	2 ₄₋₇
Mannose	-	2 ₇
Galactose	-	0
D-arabinose	-	0
Xylose	+	11 ₁₋₃
Maltose	-	0
Ethanol (3%)	+	11 ₃₋₄
L-lysine decarboxylase	-	0
L-arginine dihydrolase	-	0
Amidase for propionate	+	9

Subscripts indicate days required for positive reaction.

*Used OF basal medium (Difco) containing 1% carbohydrate.

Table 3 Results of antibiotic sensitivity test on 11 strains of *A. xylosoxidans*

Antibiotics		Sensitive strains	
		No.	%
Tetracycline	200 µg	7	63.6
Chloramphenicol	100 µg	8	72.7
Streptomycin	50 µg	0	0.0
Kanamycin	50 µg	5	45.5
Cephaloridine	30 µg	0	0.0
Ampicillin	30 µg	0	0.0
Carbenicillin	30 µg	8	72.7
Gentamicin	30 µg	1	9.1
Colistin	150 units	3	27.3
Nalidixic acid	50 µg	9	81.8

Showa sensitivity disk was used.

surgeon's hands and for the preparation of the patient's skin before operation. Therefore the resistance of *A. xylosoxidans* to chlorhexidine and two other disinfectants, ie, benzalkonium chloride (Osban, Takeda Chemical Industries Ltd, Osaka) and Cresol solution (Dainippon Pharmaceutical Ltd, Tokyo) was examined. Seven of 11 *A. xylosoxidans* strains were resistant to 2.0% chlorhexidine and three other strains were resistant to 1.0% chlorhexidine after 10 minutes' treatment (Table 4). This

Table 4 Resistance of *A. xylosoxidans* and other organisms to concentrations of chlorhexidine

Organism	No. of strains tested	No. of strains resistant to chlorhexidine with a concentration of:				
		2.0%	1.0%	0.1%	0.01%	<0.01%
<i>A. xylosoxidans</i>	11	7	3	1	0	0
<i>Ps. aeruginosa</i>	5	0	0	0	2	3
<i>E. coli</i>	2	0	0	0	0	2
<i>K. pneumoniae</i>	2	0	0	0	0	2

Table 5 Percentage of surviving organisms in 0.1% chlorhexidine

Organism	Strain No.	Time for treatment (minutes)			
		1	5	10	20
<i>A. xylosoxidans</i>	1948	32.3	32.0	27.5	18.7
<i>A. xylosoxidans</i>	4473	35.0	24.7	25.0	27.5
<i>Staph. aureus</i> *		1.3	0.7	0.07	<0.05
<i>E. coli</i> *		<0.05	<0.05	<0.05	<0.05
<i>Ps. aeruginosa</i> *		<0.05	<0.05	<0.05	<0.05

*Isolates from clinical specimens.

dose of chlorhexidine is 10 times that of the concentration (0.1%) used in this hospital. On the other hand, *Ps. aeruginosa*, *E. coli*, and *K. pneumoniae* were completely destroyed by benzalkonium chloride and Cresol at the concentrations used in this hospital (1.0% and 3.0%, respectively). In order to examine the survival of *A. xylosoxidans* in the dilution of chlorhexidine used in the hospital representative strains of *A. xylosoxidans* and other organisms were treated with 0.1% chlorhexidine and titrated, following the time course shown in Table 5. The results showed that almost one-quarter of the *A. xylosoxidans* organisms survived in the chlorhexidine after 10 minutes' treatment.

ISOLATION OF *A. XYLOSOXIDANS* FROM CHLORHEXIDINE SOLUTION

In order to investigate the source of the infection, chlorhexidine used in the vicinity of patients was tested for contaminants. In April 1975 (2 months after the last outbreak of infection) and in February 1976, a study was carried out in the operating theatre and in the surgical ward for the isolation of organisms from each washbasin. One-tenth percent chlorhexidine had been prepared with sterilised distilled water in the operating theatre and with unsterilised tap water in the surgical ward. In both areas samples were taken from 10 to 20 separate basins. The results are shown in Table 6. Four *A. xylosoxidans* strains were isolated from basins in the surgical ward. One *P. cepacia* strain was isolated from a bowl used for the disinfection of surgical instruments. No organisms were isolated from basins in the operating theatre.

Table 6 Isolation of *A. xylosoxidans* from 0.1% chlorhexidine in washbasins and other containers

Unit for investigation	Date	Container	No. of specimens		Organism
			Tested	Positive	
Operating theatre	Apr. 1975	Washbasin*	20	0	
	Feb. 1976	"	20	0	
Surgical ward (sick room) (treatment room)	Apr. 1975	"	6	2	<i>A. xylosoxidans</i>
	Feb. 1976	"	6	0	
	Apr. 1975	"	3	1	<i>A. xylosoxidans</i>
	"	Bowl†	1	1	<i>P. cepacia</i>
	Feb. 1976	Washbasin	2	1	<i>A. xylosoxidans</i>
	"	Bowl	2	0	

*In the operating theatre chlorhexidine was changed in the basin after each hand washing but in the surgical ward it was changed only twice a day.

†The bowl contained chlorhexidine for the disinfection of used surgical instruments.

Discussion

The case reports presented concern bacterial ventriculitis due to *A. xylosoxidans* infection. All patients suffered from the infection after craniotomy or cranial trepanation had been performed. *A. xylosoxidans* is the pathogen in this context because it was the only species isolated from the CSF in these patients during the relevant ventriculitis. In each case *A. xylosoxidans* was the sole aerobic isolate of cultured organisms and the clinical process of infection corresponded well with the isolation of this agent. In case 1, the patient relapsed three times with ventriculitis associated with the recovery of *A. xylosoxidans*. In addition, the patient's serum agglutinated the isolate up to a dilution of 1 in 640. Except in case 1, the duration of infection with *A. xylosoxidans* was relatively short and patients recovered quickly with appropriate chemotherapy.

Ten of 11 strains of *A. xylosoxidans* isolated were resistant to 1.0% chlorhexidine, which was 10 times the concentration of that used to disinfect the surgeon's and nurse's hands in the hospital. Although the study of chlorhexidine used in this hospital was carried out two months after the last infection with *A. xylosoxidans* the organisms still persisted in the surgical ward and were isolated from the washbasins. The observations cited above suggest that the infection described was of nosocomial origin.

Chlorhexidine, one of the derivatives of bis-diguanide, was reported to be effective on several kinds of microorganisms, including Gram-positive and -negative bacteria (Davies *et al.*, 1954; Calman and Murray, 1956). *Pseudomonas cepacia*, recently documented as a cause of nosocomial infection (Ederer and Matsen, 1972), was reported to be resistant to Savlon, which contained chlorhexidine and cetrimide (Bassett *et al.*, 1970). *Alcaligenes fecalis* which is similar in morphology and biochemical characteristics to *A. xylosoxidans*, was also reported to be resistant to chlorhexidine (Kim

et al., 1973). Recently, analytical work on the fatty acid composition of *Pseudomonas* and *Alcaligenes* was performed by some workers using gas chromatography (Dees and Moss, 1975; Moss *et al.*, 1972; Samuels *et al.*, 1973; Yabuuchi *et al.*, 1974). The results of these analyses show the similarity of the cellular fatty acid composition in the three species of organisms cited above and their possession of a relatively high percentage of 3-hydroxy-tetradecanoic acid. Further investigation on the comparison of fatty acid composition of chlorhexidine-sensitive and -resistant strains of the same species of organisms and on the neutralising effect of specific fatty acid on disinfection by chlorhexidine may explain the relationship between the cellular fatty acid component and resistance to this disinfectant.

A. xylosoxidans was resistant to gentamicin and colistin, which were commonly said to be effective against *Ps. aeruginosa* infection. Prolonged treatment of the patient with these antibiotics may play a significant role in the colonisation and infection with *A. xylosoxidans*.

We thank Dr E. Yabuuchi for kindly supplying the type strain ATCC-27061 of *A. xylosoxidans*.

References

- Bassett, D. C. J., Stokes, K. J., and Thomas, W. R. G. (1970). Wound infection with *Pseudomonas multivorans*: a water-borne contaminant of disinfectant solution. *Lancet*, **1**, 1188-1191.
- Bauer, A. W., Kirby, W. M. M., Sherris, J. C., and Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology*, **45**, 493-496.
- Calman, R. M., and Murray, J. (1956). Antiseptics in midwifery. *British Medical Journal*, **2**, 200-204.
- Davies, G. E., Francis, J., Martin, A. R., Rose, F. L., and Swain, G. (1954). 1;6-Di-4'-chlorophenyldiguanidohexane (Hibitane): laboratory investigation of a new antibacterial agent of high potency. *British Journal*

- of *Pharmacology and Chemotherapy*, **9**, 192-196.
- Dees, S. B., and Moss, C. W. (1975). Cellular fatty acids of *Alcaligenes* and *Pseudomonas* species isolated from clinical specimens. *Journal of Clinical Microbiology*, **1**, 414-419.
- Ederer, G. M., and Matsen, J. M. (1972). Colonization and infection with *Pseudomonas cepacia*. *Journal of Infectious Disease*, **125**, 613-618.
- Kim, E., Akashi, T., Shimizu, K., and Ohguro, Y. (1973). A contamination of an ultrasonic washing machine with a hititane resistant organism (in Japanese). *Igaku No Ayumi*, **85**, 355-357.
- Møller, V. (1955). Simplified tests for some amino acid decarboxylases and for the arginine dihydrolase system. *Acta Pathologica Microbiologica Scandinavica*, **36**, 158-172.
- Moss, C. W., Samuels, S. B., and Weaver, R. E. (1972). Cellular fatty acid composition of selected *Pseudomonas* species. *Applied Microbiology*, **24**, 596-598.
- Samuels, S. B., Moss, C. W., and Weaver, R. E. (1973). The fatty acid of *Pseudomonas multivorans* (*Pseudomonas cepacia*) and *Pseudomonas kingii*. *Journal of General Microbiology*, **74**, 275-279.
- Shigeta, S., Higa, K., Ikeda, M., and Endo, S. (1974). A purulent meningitis caused by *Achromobacter xylosoxidans* (in Japanese). *Igaku No Ayumi*, **88**, 336-337.
- Yabuuchi, E., and Ohyama, A. (1971). *Achromobacter xylosoxidans* n sp. from human ear discharge. *Japanese Journal of Microbiology*, **15**, 477-481.
- Yabuuchi, E., Yano, I., Goto, S., Tanimura, E., Ito, T., and Ohyama, A. (1974). Description of *Achromobacter xylosoxidans* Yabuuchi and Ohyama 1971. *International Journal of Systematic Bacteriology*, **24**, 470-477.