



Research article

Synthesis and characterization of some Metal ions complexes with mixed ligand of azo dye and Metformin and evaluation of its effectiveness on the growth of some pathogenic bacteria clinically isolated and study of its Toxicity on normal and cancerous Hepatocytes

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ABSTRACT

Four metal compounds mixed ligand of azo dye ligand (L) and metformin.(Met) were produced at aquatic ethanol for (1:1:1) (M:L:Met). The prepared compounds were identified by utilizing atomic absorption flame, FT.IR and UV-Vis spectrum manners as well as conductivity mensuration. These compounds was assayed of the gained datum the octahedral geometry was proposed into whole prepared complexes. Also in this research was studied represented examining the antibacterial and antifungal impact of the azo dye ligand (L), metformin.(Met) and (Co,Ni, Cu and Cd complexes) on four types of pathogenic, clinically isolated bacteria that are resistant to antibiotic, like *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Escherichia coli*, *Klebsiella pneumoniae*, and *fungi Candida albicans* from human in Iraq. The results of the sensitivity test showed the effectiveness of these compounds at a very low condensation of (10^{-3}) in inhibiting the isolated bacteria. On the other hand, cytotoxic effects of the ligand, Met and mix ligand complexes showed anticancer activity on HepG2 cells in a serial condensation 15.6, 31, 62, 125, 250, 500 $\mu\text{g/ml}$. As the effectiveness of the compounds increases with increasing their condensation, the most effective toxicant on hepatic cancer cells is Met and cd complex and with a rate of 68.5 and 68.3 % respectively.

1. Introduction

Azo dyes exemplify the largest volume for dye chemistry generated today, as well their proportional significance may raise at future. They play a critical role at managing the dye as well print market. Various methods also modulations are made to gain the desired color characteristics, yield as well particle size for dye to the improve dispersion [1]. Azo dyes are among the most widely utilized dyes, constituting more than 60 % of all dyes [2]. About 70 % of all dyes used at industry are azo dyes [3,4]. Azo dyes have

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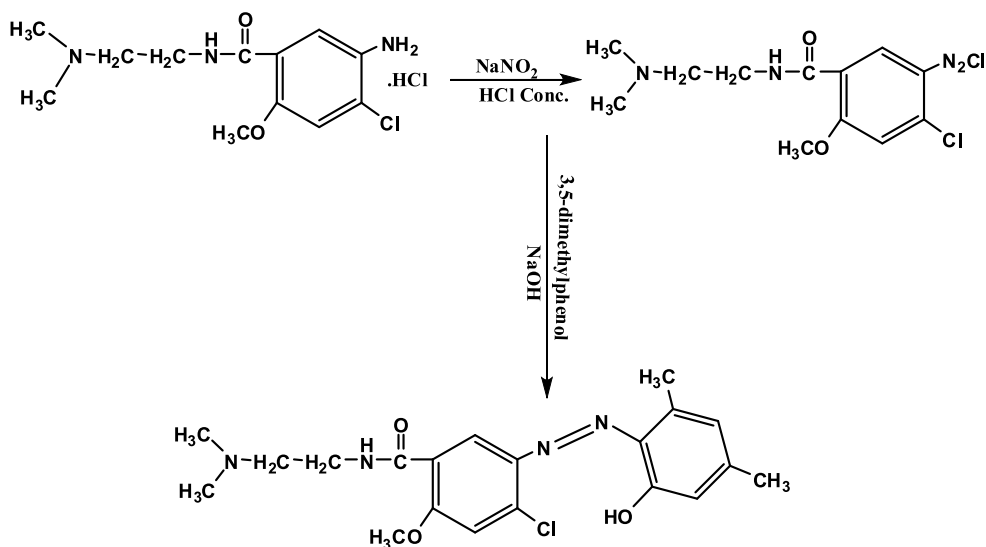
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Scheme 1. Synthesis for azo ligand (L).

been used as a very important component in the textile and printing industries [5]. Other than, used in several fields including acid-base indicators, food coloring, optical switches, plastics, cosmetics, optical data storage, nonlinear optics, liquid crystal displays, and electro-optical devices [6,7]. Further, azo dyes have been found to have biological applications, such as antioxidants, antibacterial, anticancer, anti-diabetic, and antiviral [8–10]. Transition metal ions have been contributed a diverse and rich field of research, its received much attention because of their applications in biology, medicine and industry [11]. Coordination compounds of azo dyes received much importance due to their ability to mimic complex molecules involved in vital biological process [12] such as oxygen transport, electron transfer, catalysis etc.,. At recent years, researchers have witnessed an interesting increase in the preparation of mixed-ligand complexes, particularly at biological domains [13]. The increase in the emergence of bacteria that are multidrug resistant and cancerous diseases has become a major global problem, and new compounds are needed to treat these diseases, so we studied the effectiveness of new substances to evaluate their inhibitory effects on bacteria and cancer cells. Metformin hydrochloride is an antihyperglycemic agent used to treat type 2 diabetes with a healthy diet, Metformin reduces blood glucose levels by decreasing glucose synthesis by the liver (gluconeogenesis).

It also increases insulin sensitivity by increasing glucose uptake and utilization by cells, and it also inhibits the activity of mitochondrial complex I and this mechanism is what gave metformin a strong effect in the treatment of diabetes and used to treat polycystic ovary syndrome. Metformin as antimicrobial and antifungal, antiprotozoal, and antituberculosis effect either alone or in combination [14]. Metformin showed a potential synergistic effect when combined with a tetracycline and fluoroquinolone group against a highly resistant strain of *E. coli*, *S. aureus* and *P. aeruginosa* [15,16] found metformin has anticancer activity and its mechanism of action is unknown. Azo ligand contain metoclopramide hydrochloride is antiemetic used to treat of gastroesophageal reflux disease, anticancer to expressed in a diversity of cancer tissue like lung, cervical, breast, and ovarian cancer by blocking the D2 receptor [17]. Metals at low concentration within the permissible limits of global health [18], and mixed with agent lead to use to treat cancer by specifically attacking cancer cells and interacting directly with DNA. The positive charge of most metals can interact with the negative charge of the phosphate portion of DNA [19]. Also, these elements and their complexes have antibacterial and antifungal effectiveness by eliminated or their growth prevented by preventing the formation of the cell wall, or through a defect in the physical or chemical structure of protein and nucleic acids, or a defect in the permeability of the cytoplasmic membranes. They can also be destroyed by a defect in the cellular enzymatic activity, as well as through Manufacturing is prohibited Proteins and nucleic acids [20].

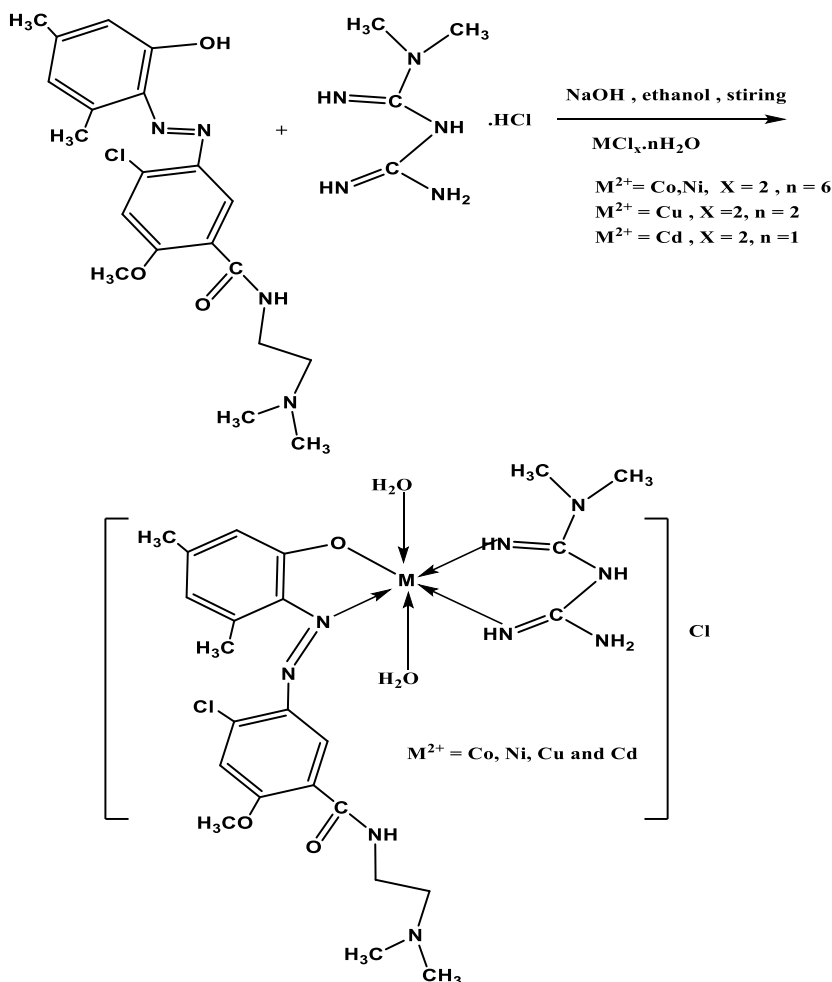
2. Experimental

2.1. Materials

In this study, all the reagents used in this research were of the highest quality analar, which consist of Metformin.HCl, NaOH ethanol and DMSO (Merck and Aldrich chemicals). The metal chlorides, $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ and $\text{CdCl}_2 \cdot \text{H}_2\text{O}$ (Merck, BDH and Riedel) are utilized unpurified.

2.2. Physical Measurements

UV-Vis spectrum have been recorded at a Shimadzu-U.V-160. Ultraviolet spectrometer utilizing DMSO solution 10^{-3} at range (200–1000) nm for the (Met) as well as mixed ligand metal complexes. The molar conductance for mixed ligand metal complexes have been



Scheme 2. Preparation of mixed ligand complexes.

measured employing a CON 510 Conductivity at dry DMSO (10^{-3} M) solution on room temperature. While metal includes into complexes were determined through Atomic Absorption (A.A) Technique/Flame Emission Spectrophotometer using AA – 680. NMR spectrum (^1H , ^{13}C NMR) were acquired at DMSO- d_6 solution employing NMR Bruker 500MHz Germany at DMSO- d_6 for the cadmium complex. Melting point have been obtained at Stuart Melting Point Apparatus for the (Met) and mixed ligand complexes. Infrared (FT-IR) spectral have been registered at $4000\text{--}400\text{ cm}^{-1}$ range to the (Met) as well metal complexes at a Shimadzu IR-470 Spectrophotometer employing KBr. The mass spectra for the all complexes were recorded at the Agilent 6211 mass spectrometer. Sherwood Applying the Auto Magnetic Susceptibility at 25°C allowed scientists to refine the magnetic characteristics. Elemental (C, H, N) micro-analysis, to search using the EA 3000 single V.3 model vector device.

2.3. Preparation of the azo dye ligand

A solution was produced [21], of metoclopramide hydrochloride (0.84 gm, 1 mM) in mixture (10 ml ethanol, 2 ml conc. HCl), and diazotized at 5°C with 10% solution of NaNO_2 . Diazotized solution has been added colligium wise for stirring into a cooled ethanolic solution at (0.305 gm, 1 mM) for 3,5-dimethylphenol. Then 25 ml at 1M NaOH solution has been followed into dusky colored mix and precipitation for azo ligand has been noticed. This deposit have been filtrated, washed number ounces for (1:1) $\text{C}_2\text{H}_5\text{OH}:\text{H}_2\text{O}$, mixture subsequently left into dry, the reaction is appear at Scheme 1.

2.4. Preparation for mixed ligand Metal complexes

A public method has been utilized with the research for all coordinations complexes. The complexes have been prepared at a comparable method around synthesis at the molar ratio from M:L:Met (1:1:1). In the aqueous solution (10ml) for the appropriate cobalt(II) chloride hexahydrate (0.238g, 1 mM), nickel(II) chloride hexahydrate (0.238g, 1 mM), copper(II) chloride dihydrate

Table 1
Elemental analysis and physical data for (L), (Met) and mixed ligand complexes.

Compounds	Empirical	Molecular weight (g/mol)	Color (Yield %)	M.P (°C)	Analysis Calc (Found)			
	Formula				M%	C%	H%	N%
Azo dye ligand (L)	C ₂₀ H ₂₅ ClN ₄ O ₃	403.50	Orange (81)	<300	–	59.25 (58.93)	6.17 (6.03)	13.82 (12.96)
Metformin.HCl (Met)	C ₄ H ₁₂ ClN ₅	165.50	white	221	–	–	–	–
[Co(L)(Met)(H ₂ O) ₂] Cl	[CoC ₂₄ H ₃₉ N ₉ O ₅ Cl]Cl	663	Deep green (73)	>350	8.89 (8.37)	43.43 (42.85)	5.88 (4.91)	19.00 (18.77)
[Ni(L)(Met)(H ₂ O) ₂] Cl	[NiC ₂₄ H ₃₉ N ₉ O ₅ Cl]Cl	662	Orange (82)	>350	8.76 (7.32)	43.50 (42.97)	5.89 (4.87)	19.33 (18.72)
[Cu(L)(Met) (H ₂ O) ₂] Cl	[CuC ₂₄ H ₃₉ N ₉ O ₅ Cl]Cl	668	Light brown (77)	>350	9.58 (7.56)	43.11 (42.97)	5.83 (4.75)	18.86 (17.92)
[Cd(L)(Met)(H ₂ O) ₂] Cl	[CdC ₂₄ H ₃₉ N ₉ O ₅ Cl]Cl	716	Orange (81)	>350	15.64 (13.52)	40.22 (39.87)	5.44 (4.97)	17.59 (16.88)

(0.171 g, 1 mM) as well cadmium(II) chloride monohydrate (0.201 g, 1 mM). Has been added into a ethanolic solution (20 ml) for azo dye ligand (L), C₂₀H₂₅ClN₄O₃ (0.404 gm, 1 mM) that was previously prepared [1] as well added (0.04 gm, 1 mM) for the NaOH was added to the previous solution as well as (0.1656 gm, 1 mM) for the metformine.HCl (C₄H₁₂ClN₅) in (20 ml) in an ethyl alcohol solution. The solution was refluxed about 2hr., then the precipitated solid complex has been filtrated, washed completely with (C₂H₅)₂O as well dried at vacuo, see Scheme 2.

2.5. Antimicrobial susceptibility test

Preparation of bacterial and fungus suspension by transferring 2–3 colonies of *E. coli*, *K.pneumoniae* grew on MacConkey agar and *S. epidermidis*, *S.aureus* grew on blood agar base as for the *C. albicans* fungus it was grown on amedium bromocresol green agar, into 5 ml of normal saline, the turbidity of suspension was compared to 0.5 Mcfarland using Densicheck plus technique. The agar well diffusion method was used to evaluate the effect of azo dye ligand (L), Metformine.HCl (Met) and (Co, Ni, Cu and Cd) complexes on the growth of the *E. coli*, *K.pneumoniae*, *S.aureus*, *S. epidermidis* and *C. albicans* at concentration 10⁻³. The muller hinton agar 20 ml was cultured by spreading the bacterial and fungus suspension. Well, were made by cork borer and volume 0.05 ml was added to each well in addition to DMSO as control treatment. This plates were left at room temperature for 15–20 min then incubated at 37 °C for 24 h. Three replicates from each compounds were done. measured the inhibition zone diameter by ruler [22].

2.6. Chloride content analysis (Cl %)

Chloride content of the prepared complexes was determined and analysed using the standard method [23], using Moore's method by titrating the complex against the silver nitrate present in the burette.

2.7. Cell culture

HepG2 human hepatic cell carcinoma was obtained from aliver cancer of 42 years old male and Hdfn (Human dermal fibroblast, neonatal) normal cell line the same age, were obtained from the university of malya, Malaysia. The cell sheet washed with phosphate buffer saline. 2–3 ml/trypsin/EDTA solution was added to the cell in the vessels and incubated 37 C for 2 min to detach from the vessel then [15–20] ml RPMI medium added Penicillin G 10³ IU, Streptomycin 0.001 g, Sodium Bicarbonate 1 %, Fetal Bovine Serum 10 % and incubated at 37 °C in CO₂ incubator to grow the cells. Cell count was calculated by haematocytometer [24].

2.8. MTT cytotoxicity Protocol

The cytotoxicity effect of chemical compounds (azo dye ligand (L), Metformine.HCl (Met), (Co, Ni, Cu and Cd) complexes were performed by using MTT kit. 200 µL of medium added to the each well of microtiter plate, incubated at 37 °C, 5 % CO₂ for 24 h, 200 µL of serial concentration of (Met), (L) and mix ligand complexes (15.6, 31, 62, 125, 250, 500) µg/ml. Triplicate were used for each concentration were added to the wells. The control was the cells treated with serum free medium. 10 µL of the MTT was added to well and the microplate incubated at 37 °C, 5%CO₂ for 4 h. The medium was removed and 100 µL of solubilization solution was added and incubated 5 min. The absorbance was measured by Elisa at 575 nm [25,26].

3. Results and discussion

The solid compounds were generated through reaction from alcoholic solution of the azo dye ligand (L) for the aqueous solution from the metal ions and Metformin (Met) into a (M:L:Met) from (1:1:1). The metal contents of these complexes were into good correspondence for the calculated values (Table 1) contains the physical characteristics. The molar conductance from the compounds as (10⁻³ M) on DMSO indicating there electrolyte nature at ratio (1:1) [27], and the data recorded in (Table 2).

Table 2
UV spectra data and molar conductivity for(met) and mixed ligand complexes.

Compounds	λ (nm)	ABS	Wave number (cm ⁻¹)	ϵ_{\max} (L.mol ⁻¹ .cm ⁻¹)	Assignments	Molar conductivity Λ_m .cm ² .Mol ⁻¹ 10 ⁻³ M in DMSO	μ_{eff} (B. M)
Metformin. HCl (Met)	260	0.539	38462	539	$\pi-\pi^*$	–	
	369	0.040	27100	40	$n-\pi^*$		
Azo dye ligand (L)	263	1.207	38022	1207	$\pi-\pi^*$	–	
	312	0.858	32051	858	$\pi-\pi^*$		
	386	1.919	25907	1919	$n-\pi^*$		
[Co(L)(Met)(H₂O)₂] Cl	265	0.883	37736	883	intra ligand	60.2	4.86
	386	0.807	25907	807	C.T		
	543	0.023	18416	23	${}^4T_{1g(F)} \rightarrow {}^4T_{1g(P)}$		
[Ni(L)(Met)(H₂O)₂] Cl	266	1.363	37594	1363	intra ligand	53.1	2.88
	390	1.687	25641	1687	C.T		
	520	0.024	19230	24	${}^3A_{2g} \rightarrow {}^3T_{1g(P)}$		
[Cu(L)(Met)(H₂O)₂] Cl	263	0.659	38023	659	intra ligand	74.3	1.71
	388	0.548	25773	548	C.T		
	580	0.011	10163	11	${}^2E_g \rightarrow {}^2T_{2g}$		
[Cd(L)(Met)(H₂O)₂] Cl	264	0.645	37879	645	intra ligand	52.8	Dia
	391	1.378	25575	1378	C.T.		
	527	0.520	18975	520	MLCT		

3.1. Mass spectra

Mass spectra of the complexes displays peaks centered at $m/z = 663, 662, 668$ and 716 due to the formulas $[C_{24}H_{39}N_9O_5ClCo]Cl$, $[C_{24}H_{39}N_9O_5ClNi]Cl$, $[C_{24}H_{39}N_9O_5ClCu]Cl$ and $[C_{24}H_{39}N_9O_5ClCd]Cl$ respectively. The general pattern of fragmentation are summarized in Schemes- 3,4,5 and 6, see Figs. 1–4.

3.1.1. NMR spectra

¹HNMR spectrum of Cd²⁺ complex at dimethylsulfoxide (Fig. 5) display various signals at ($\delta = 7.67-7.96$) ppm refers to aromatic protons [28]. Signal obtained at ($\delta = 8.39$) ppm lead to proton of amide [29]. Finding, the signals at ($\delta = 1.78$) and ($\delta = 1.27$) ppm were assigned to $\delta(CH_3)$ of phenol and (N-(CH₃)₂), signals at ($\delta = 2.65, 3.35$) and ($\delta = 3.94$) ppm which were assigned to (CH₂) and (OCH₃) groups [30]. The signals at ($\delta = 2.93$) and ($\delta = 6.59$) due to protons of (N-(CH₃)₂) and (NH,NH₂)in metformin [31]. signals at ($\delta = 7.30$) and ($\delta = 2.5$) ppm which were assigned to(=NH) groups and DMSO- d₆ [32].

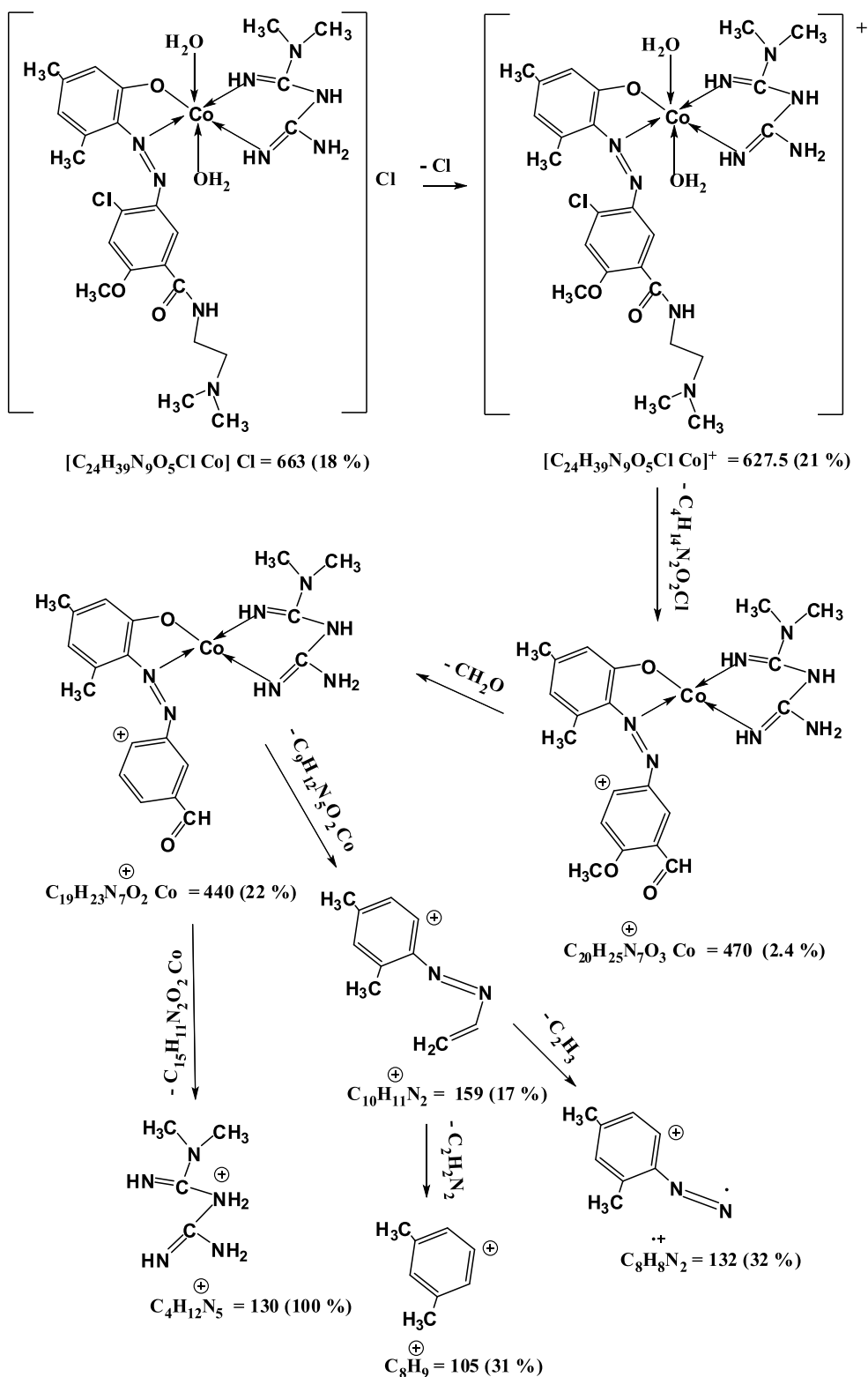
¹³CNMR Cd²⁺ complex spectrum display resonance at ($\delta = 12.48$) and ($\delta = 22.47$) ppm due to carbon of (CH₃) in phenol ring. Signals at ($\delta = 37.80, 56.46$) and ($\delta = 46.82$) ppm lead to carbon of (CH₂) and (N-(CH₃)₂) respectively. The resonance at ($\delta = 37.92$) and ($\delta = 51.54$) ppm indicative to carbon of (N-(CH₃)₂) for metformin and methoxy group (OCH₃) for azo dye. The various signals at ($\delta = 158.67, 156.89, 152.65, 139.56, 132.20, 120.33, 99.82$ and 73.29) ppm attributed to carbon atoms of aromatic rings. Signals at ($\delta = 163.38$) ppm and ($\delta = 159.69$) ppm due to carbon of (C=O) and (C=NH) groups and the indicative in ($\delta = 39.83$) ppm due into DMSO-d₆ [33,34], see Fig. 6.

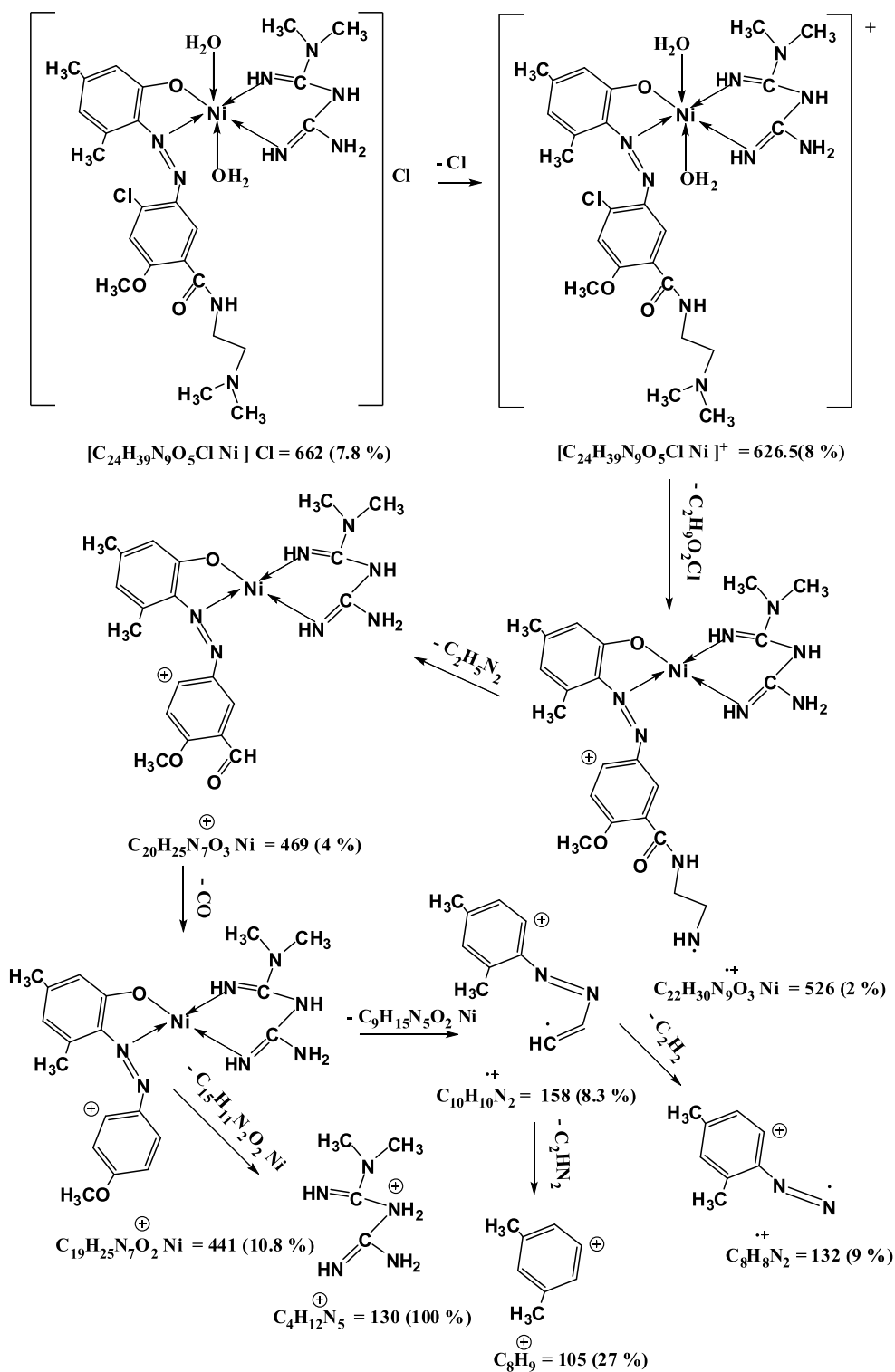
3.1.2. Electronic spectra

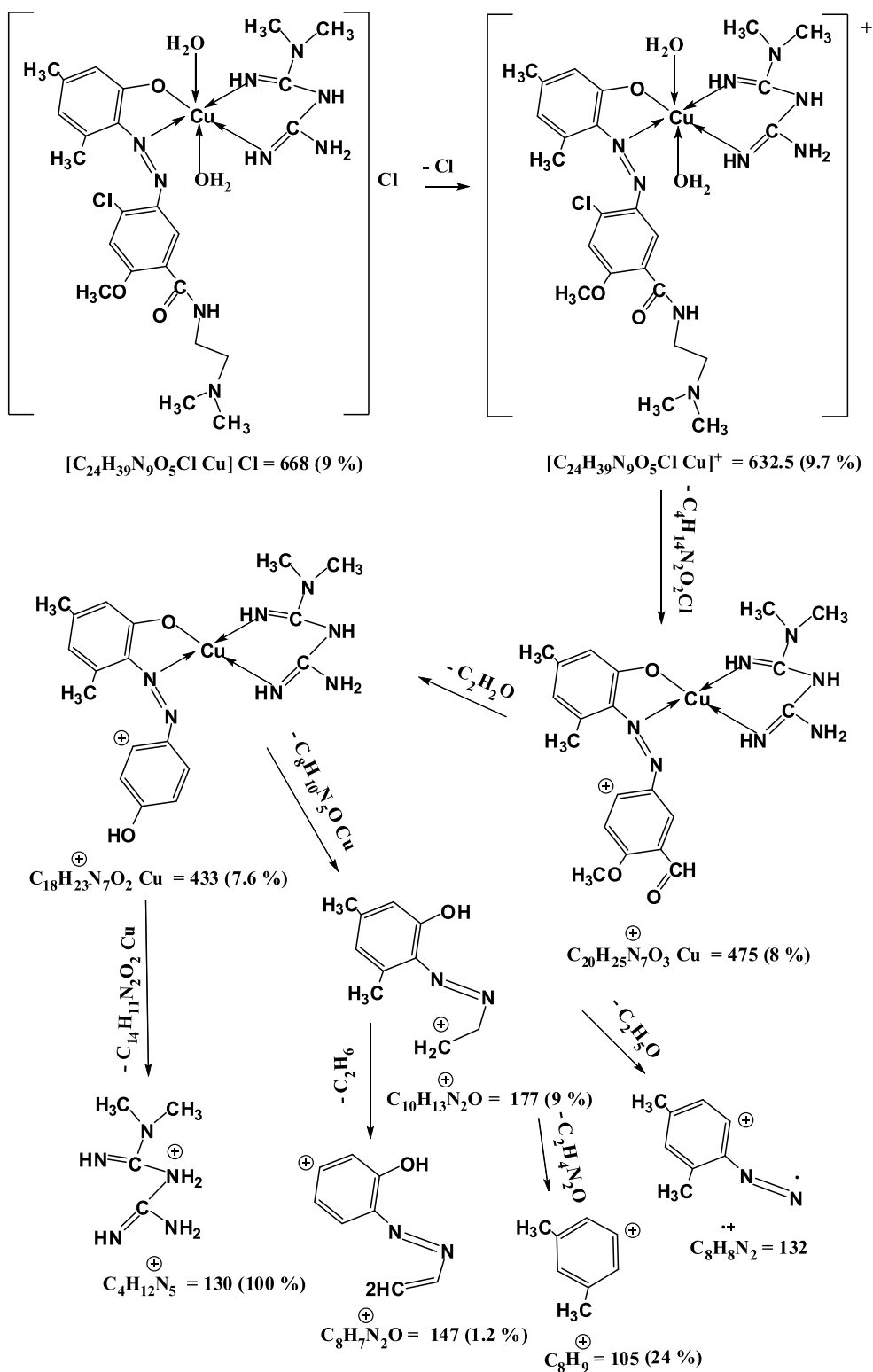
UV-Vis spectra to the produced compounds melted at DMSO (10⁻³ M/L) were gauged as well the datum formed is listed at Table 2. UV-Vis spectrum to the azo ligand shows peaks at 263, 312 and 386 nm were appointed into mild energy ($\pi-\pi^*$) and ($n-\pi^*$) transition, metformin spectrum shows to peaks at 260 and 369 nm due to ($\pi-\pi^*$) and ($n-\pi^*$) transition [35]. Co(II) spectrum appeared two peaks at 265 and 386 nm due to intra ligand and charge transfer. Peak at 543 nm assigned to electronic transition type ${}^4T_{1g(F)} \rightarrow {}^4T_{1g(P)}$, also the value from the magnetic moment at 4.86 B M may be possessed as extra confirmation to octahedral [36]. The spectrum for Ni(II) complex display peaks in 266 and 390 nm whom were qualified into intra ligand and charge transfer. Else peak at 520 nm whom was refereed into electronic transition type ${}^3A_{2g(F)} \rightarrow {}^3T_{1g(P)}$, the magnetic moment for this complex was discover in 2.88 B M whom was much close to the octahedral [37]. Cu(II) spectrum display peaks at 263 and 388 nm due to intra ligand and charge transfer. Other peak at 580 nm lead to electronic transition type ${}^2E_g \rightarrow {}^2T_{2g}$, magnetic moment for this complex was found at 1.71 B M whom was much close to the octahedral environment [38]. The electronic spectrum of Cd(II) complex do offer the charge transferz and the magnetic susceptibility seemed the complex has diamagnetic moments, result to (d-d) transition are not likely hence electronic spectrum did not confer any productive datum, on fact this outcome are a good agreement for former work from octahedral [39].

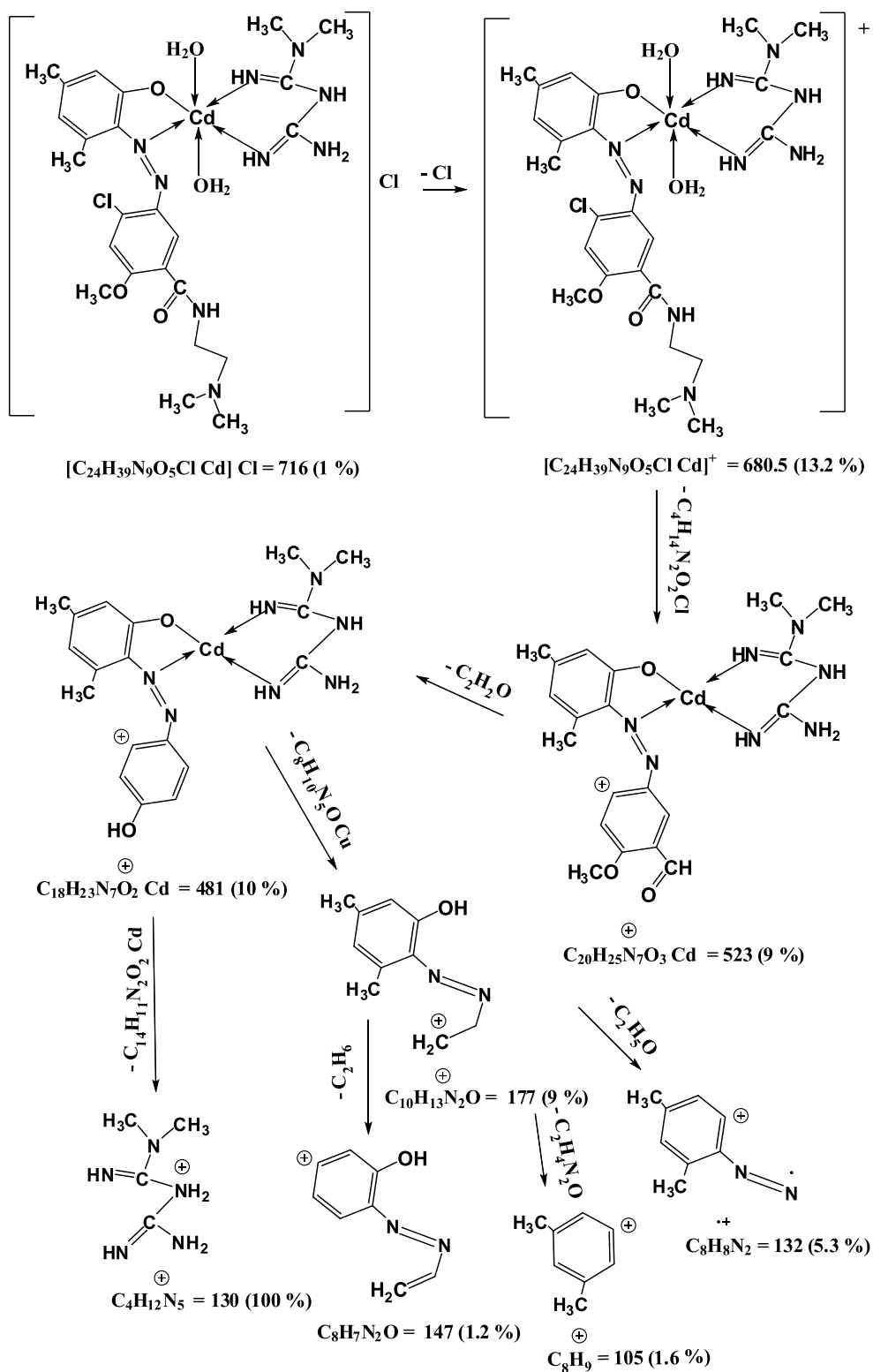
3.1.3. FTIR spectra

FTIR spectra to the azo ligand and their metal chelates have been collated, and the data was scheduled in Table 3. The broad band in the spectrum of the azo ligand at 3371 cm⁻¹, that was described into the stretching vibration from $\nu(OH)$ phenol, the disappearance of this band at the spectra with all produced compounds pointed out the deprotonation for phenol group to coordination with metal ion [40]. Band for azo group at 1570 cm⁻¹ displaced into lower wave number for change during shape at spectra for all produced compounds [41]. The spectrum of metformin exhibited bands at 3371,3170 and 3294 cm⁻¹due to $\nu(NH_2)$ and $\nu(NH)$ [42]. Band at

Scheme 3. Fragmentation Pattern for $[Co(L)(Met)(H_2O)_2]Cl$ Complex.

Scheme 4. Fragmentation Pattern for $[Ni(L)(Met)(H_2O)_2]Cl$ Complex.

Scheme 5. Fragmentation Pattern for $[Cu(L)(Met)(H_2O)_2]Cl$ Complex.

Scheme 6. Fragmentation Pattern for $[Cd(L)(Met)(H_2O)_2]Cl$ Complex.

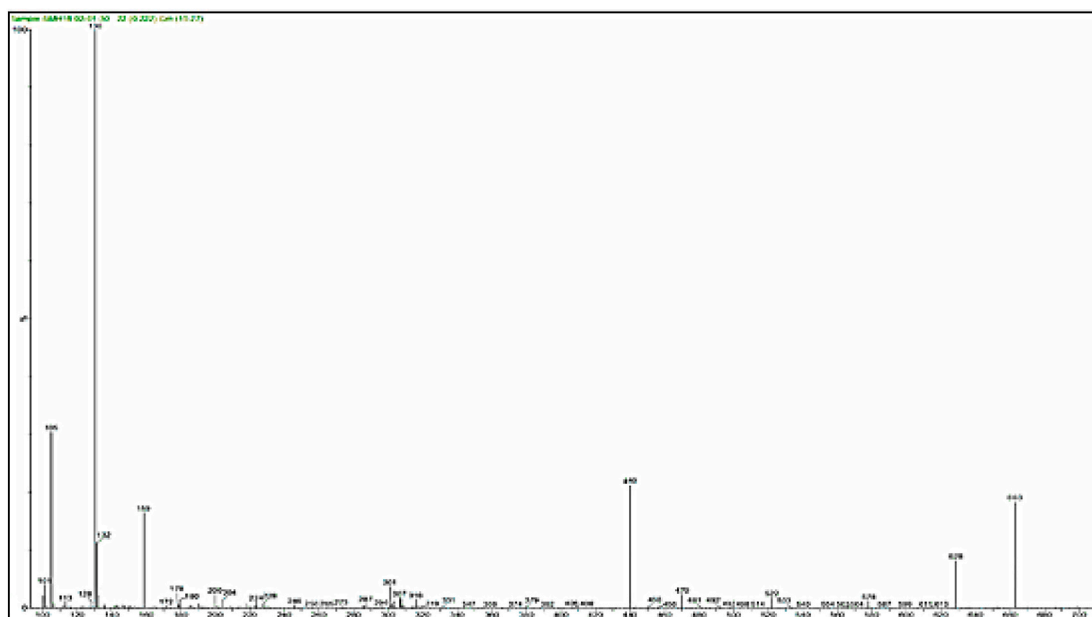
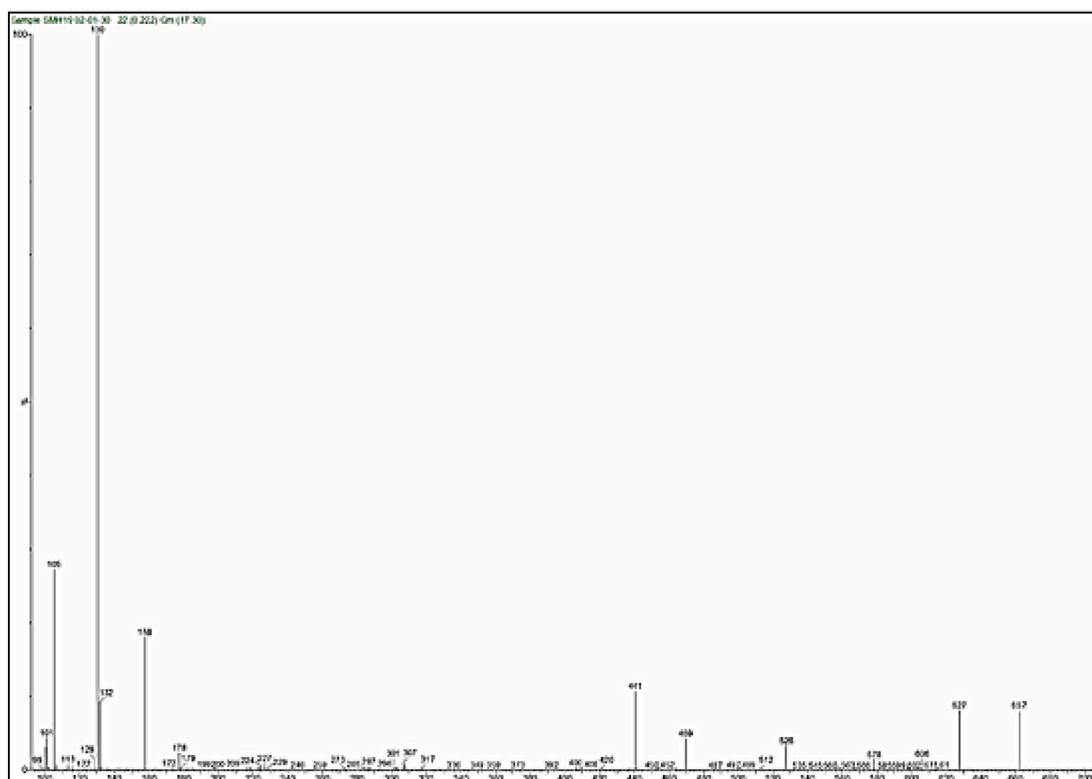


Fig. 1. Mass spectrum for $[\text{Co}(\text{L})(\text{Met})(\text{H}_2\text{O})_2]\text{Cl}$ complex.



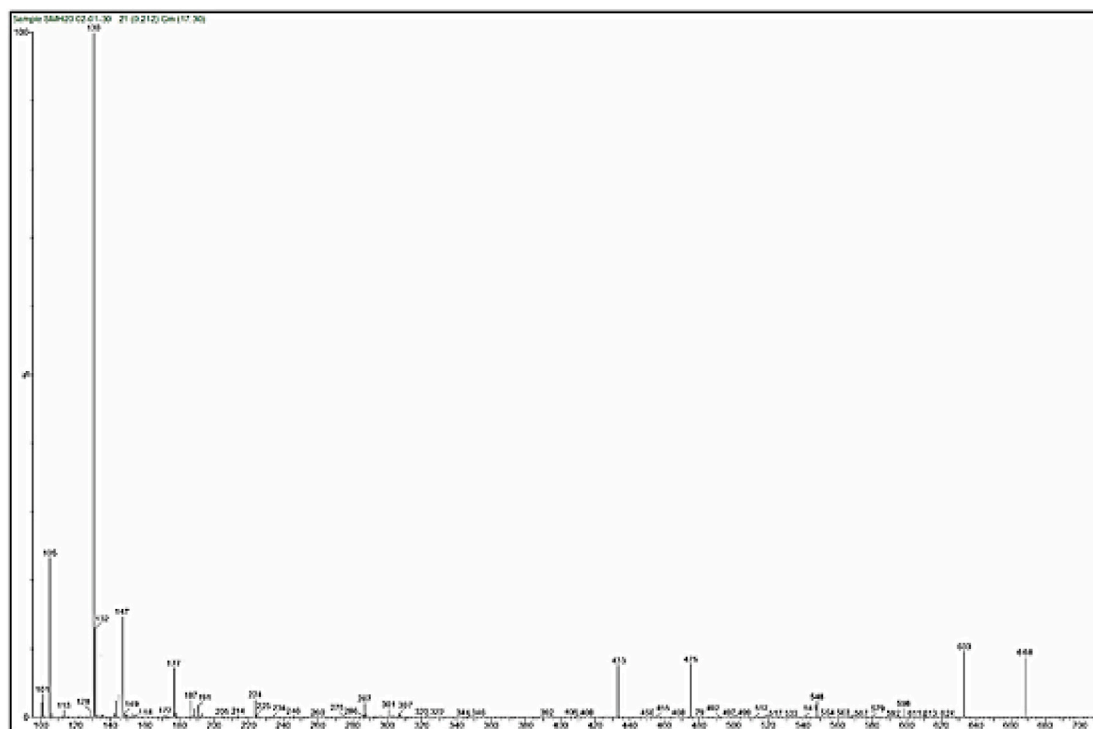


Fig. 3. Mass spectrum for [Cu(L)(Met)(H₂O)₂]Cl complex.

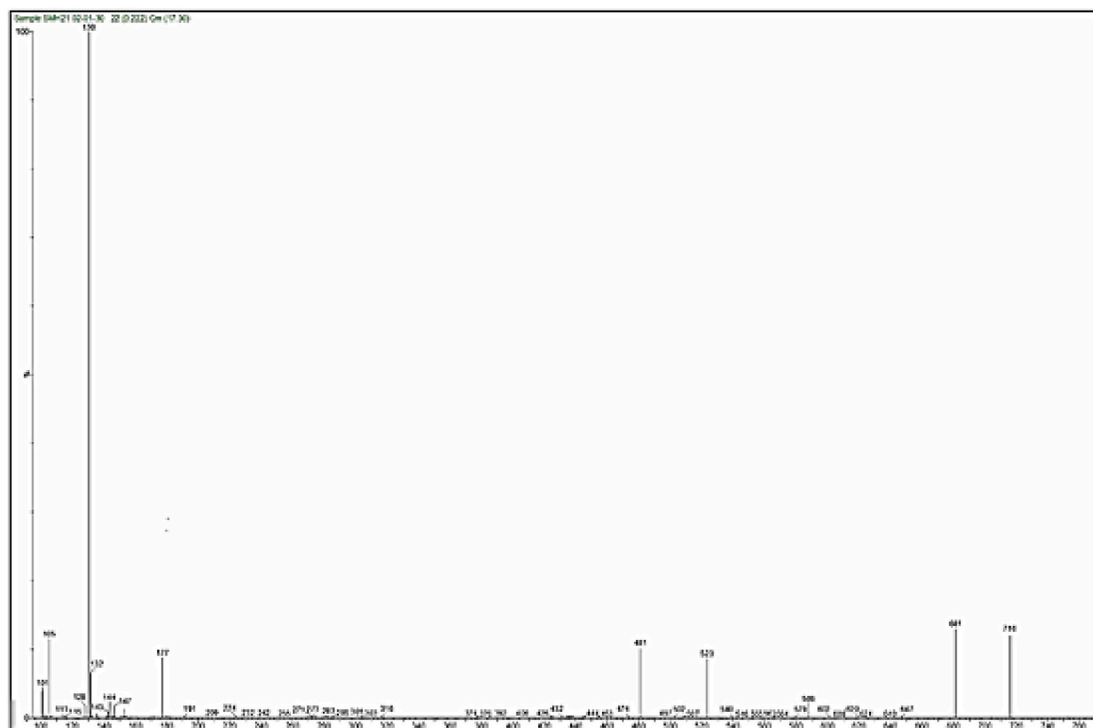
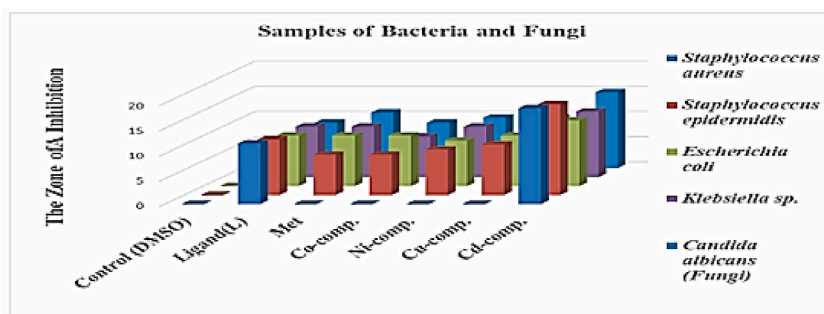


Fig. 4. Mass spectrum for [Cd(L)(Met)(H₂O)₂]Cl complex.

Table 3The main frequencies for azo ligand and metal chelate (cm^{-1}).

Compounds	$\nu(\text{NH}_2)$	$\nu(\text{C}=\text{N})$	$\nu(\text{H}_2\text{O})$	$\nu(\text{M}-\text{N})$
	+	+	+	+
	(NH) ν	$\nu(\text{N}=\text{N})$	$\nu(\text{OH})$	(M-O) ν
Metformin. HCl (Met)	3371 s. 3170 s. 3294 s.	1627 s. -	-	-
Azo dye ligand (L)	-	-	-	-
[Co(L)(Met)(H ₂ O) ₂] Cl	3470 br. 3394 br. 3167 sh.	1585 sh. 1631 s. 1500 s.	3371 br. 937 sh. 775 s.	547 w. 513 w.
[Ni(L)(Met)(H ₂ O) ₂] Cl	3414 br. 3367 br. 3181 sh.	1639 sh. 1535 s.	3462 br. 891 s. 833 s.	493 w. 462 w.
[Cu(L)(Met)(H ₂ O) ₂] Cl	3482 br. 3340 br. 3190 sh.	1643 s. 1516 s.	3491 br. 879 sh. 802 sh.	563 w. 478 w.
[Cd(L)(Met)(H ₂ O) ₂] Cl	3452 br. 3382 br. 3259 sh.	1635 sh. 1531 s.	3549 br. 860 sh. 748 s.	555 w. 459 w.

**Fig. 7.** Bioactivity for (L), (Met) and Mixed Ligand Complexes Against the Bacteria Species as well Fungi.**Table 4**

Effect of (L), (Met) and mixed ligand complexes for growth of bacteria and fungi clinically isolated.

Compounds	<i>Staphylococcus aureus</i>	<i>Staphylococcus epidermidis</i>	<i>Escherichia coli</i>	<i>Klebsiella pneumoniae</i>	<i>Candida albicans</i>
	(G + ve)	(G + ve)	(G-ve)	(G-ve)	(Fungi)
Control (DMSO)	-	-	-	-	-
Ligand(L)	12	11	10	10	9
Met	-	8	10	10	11
Co-comp.	-	8	10	8	9
Ni-comp.	-	9	9	10	10
Cu-comp.	-	10	10	10	10
Cd-comp.	19	18	13	13	15

Schiff base ligand [46]. The principle of action of these compounds may be to inhibit the growth of isolated bacteria by their effect on the production of the bacterial cell wall that protects cells from the external environment, or to interfere in protein synthesis by linking the mechanism that builds amino acids and protein, or to affect metabolic processes such as folic acid synthesis and vitamin B is necessary for the growth of bacteria or preventing the synthesis of DNA [47]. Or it may be attributed to hydroxyl groups present in these complexes that could bind with active groups of microorganism enzymes by hydrogen bonds also precipitate proteins due to the formation of hydrogen bonds with those proteins, thus inhibiting necessary enzymes in organisms [48]. The resistance of *S.aureus* to most of the compounds was due to its possession of virulence factors such as gelatinase and beta-lactamase enzyme, and it was surrounded by biofilm and capsule that protects it from external influences [49,50].

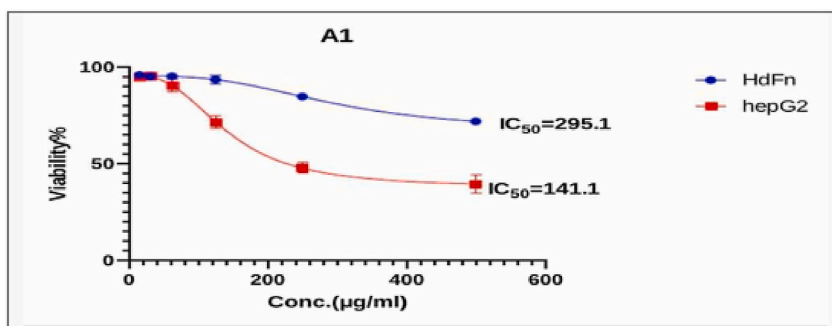


Fig. 8. Cytotoxicity effect of ligand azo dye on cancer HepG2 and normal HdFn cell Lne.

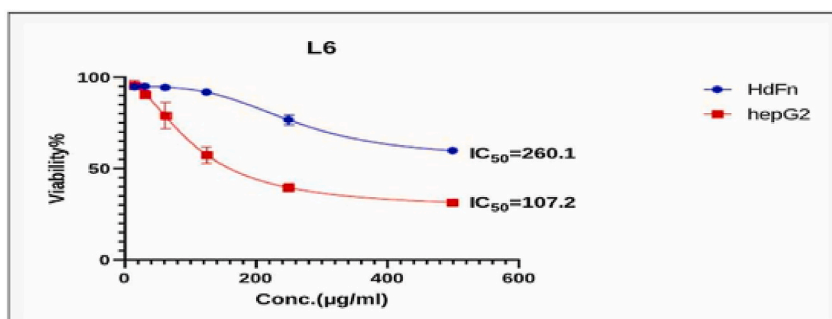


Fig. 9. Cytotoxicity effect of Met on cancer HepG2 and normal HdFn cell line.

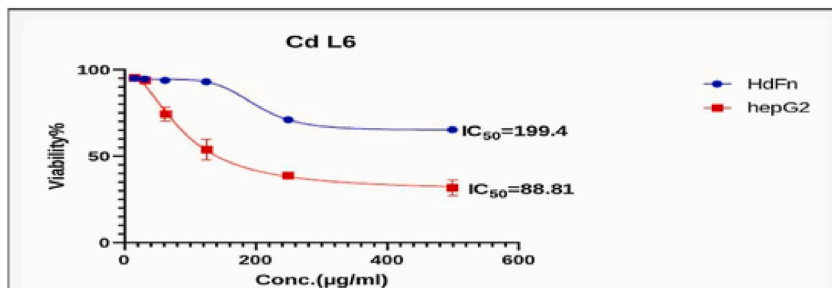


Fig. 10. Cytotoxicity effect of Cd complex on cancer HepG2 and normal HdFn cell line.

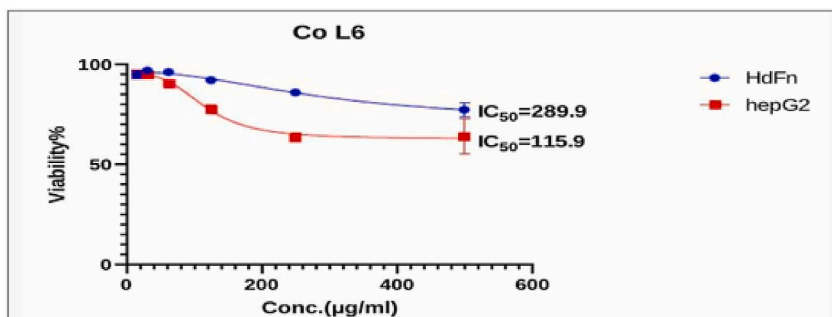


Fig. 11. Cytotoxicity effect Co complex on cancer HepG2 and normal HdFn cell line.

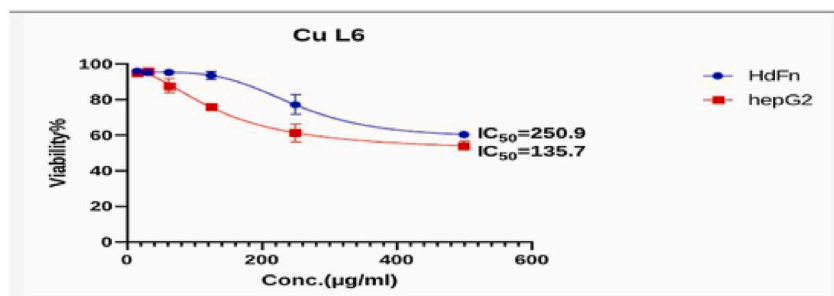


Fig. 12. Cytotoxicity effect of Cu complex on cancer HepG2 and normal Hdfn cell line.

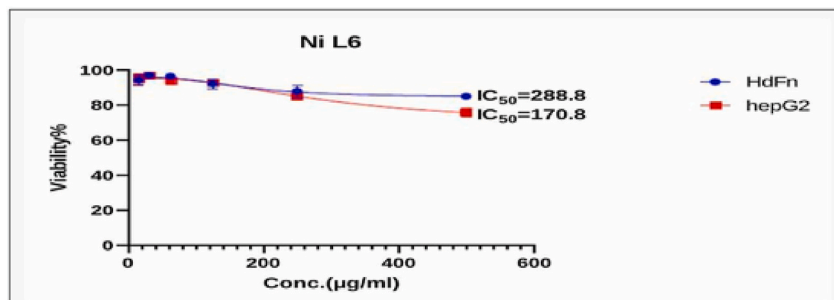


Fig. 13. Cytotoxicity effect of Ni complex on cancer HepG2 and normal Hdfn cell line.

Table 5

The cytotoxic effect of A1 (ligand azo dye) on Hdfn and HepG2 cell line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	Hdfn	HepG2
500	71.95 \pm 0.81	39.352 \pm 4.78
250	84.79 \pm 1.2	48.03 \pm 2.5
125	93.5 \pm 2.10	71.48 \pm 3.39
62	95.33 \pm 1.1	90.70 \pm 3.18
31	95.21 \pm 0.8	95.71 \pm 0.80
15.6	95.949 \pm 1.02	95.17 \pm 1.28

Table 6

The cytotoxic effect of L6 (Met) on Hdfn and HepG2 cell line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	Hdfn	HepG2
500	59.8 \pm 1.10	31.55 \pm 1.3
250	76.69 \pm 2.84	39.53 \pm 2.36
125	91.85 \pm 0.9	57.48 \pm 4.63
62	94.36 \pm 1.44	79.05 \pm 7.34
31	95.02 \pm 0.9	90.50 \pm 2.10
15.6	94.83 \pm 1.18	95.679 \pm 2.65

3.3. Cytotoxic effect of ligand, Met and Metal complexes on cell line

Figs. 8–13 and Tables 5–10 shows the result of the effect of serial concentration of ligand, Met and metal mix ligand complexes on the normal hepatic cell Hdfn and carcinoma cell line HepG2. All complexes, ligand and Met had an effective inhibition of cancer cells. The highest inhibitory activity was met and cd complex with a percentage of (68.5 and 68.3) % respectively, then ligand azo dye 60.65 %, Cu complex 45.92 %, Co complex 35.85 and finally Ni complex 24.23 %. And compared with their effect on normal cells, their effectiveness on inhibiting cancer cells was higher than normal cells. There is no research on their inhibitory effectiveness, but it may be the effect of these complexes on DNA and change the internal structure of each cancer cell or loss of cellular communication and

Table 7

The cytotoxic Effect of CdL6-(Cd Complex) on HdFn and HepG2 Cell Line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	HdFn	HepG2
500	65.31 \pm 1.8	31.7 \pm 4.4
250	71.02 \pm 1.74	38.92 \pm 1.01
125	92.978 \pm 1.3	53.89 \pm 5.87
62	93.78 \pm 0.24	74.34 \pm 4.13
31	94.6 \pm 1.62	94.05 \pm 1.07
15.6	95.06 \pm 1.6	95.25 \pm 1.7

Table 8

The cytotoxic effect of Co L6 (Co complex) on HdFn and HepG2 cell line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	HdFn	HepG2
500	77.27 \pm 3.65	64.15 \pm 8.9
250	86.034 \pm 0.85	63.69 \pm 2.1
125	92.129 \pm 1.55	77.62 \pm 2.41
62.5	96.18 \pm 1.2	90.58 \pm 1.8
31.2	96.95 \pm 1.14	94.79 \pm 1.33
15.6	94.90 \pm 2.19	95.40 \pm 1.4

Table 9

The Cytotoxic Effect of CuL6 (Cu complex) on HdFn and HepG2 Cell Line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	HdFn	HepG2
500	60.378 \pm 0.8	54.08 \pm 2.56
250	77.08 \pm 5.52	61.381 \pm 5.20
125	93.59 \pm 2.10	75.8 \pm 2.05
62	95.33 \pm 1.18	87.73 \pm 3.9
31	95.216 \pm 0.82	95.94 \pm 0.90
15.6	95.949 \pm 1.02	94.86 \pm 0.29

Table 10

The cytotoxic effect of NiL6 (Ni complex) on HdFn and HepG2 cell line.

Concentration $\mu\text{g mL}^{-1}$	Mean viability (%) \pm SD	
	HdFn	HepG2
500	85.03 \pm 1.03	75.77 \pm 2.65
250	87.92 \pm 3.57	85.108 \pm 1.2
125	92.20 \pm 2.77	92.93 \pm 0.8
62	96.48 \pm 1.29	94.05 \pm 1.38
31	96.95 \pm 1.14	96.79 \pm 0.9
15.6	94.2 \pm 2.9	95.02 \pm 3.0

starvation of amino acids and interference with intracellular [51] especially when DNA binding of the copper II compound as anti-oxidant [52], or inhibits the electron transport chain and ATP synthesis like metformin agent [53].

CRediT authorship contribution statement

Shatha M.H. Obaid: Resources, Project administration, Formal analysis, Conceptualization. **Afnan E. Abd-Almonuim:** Writing – original draft, Visualization, Conceptualization. **Hanan Adnan Shaker Al -Naymi:** Validation, Methodology, Data curation. **Amer J. Jarad:** Validation, Supervision, Conceptualization. **Marwan Mahmood Saleh:** Writing – review & editing, Supervision.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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