#### **REVIEW ARTICLE**



Heterocycles in Breast Cancer Treatment: The Use of Pyrazole Derivatives



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**Abstract:** Among the aromatic heterocycle rings, pyrazole –a five-membered ring with two adjacent nitrogen atoms in its structure has been postulated as a potent candidate in the pharmacological context. This moiety is an interesting therapeutic target covering a broad spectrum of biological activities due to its presence in many natural substances.

Hence, the potential of the pyrazole derivatives as antitumor agents has been explored in many investigations, showing promising results in some cases. In this sense, breast cancer, which is already the leading cause of cancer mortality in women in some countries, has been the topic selected for this review, which covers a range of different research from the earliest studies published in 2003 to the most recent ones in 2021.

Keywords: Breast, cancer, heterocycle, pyrazole, tumor, HDI.

### **1. INTRODUCTION**

It is well known that cancer is the second cause of mortality in the world after cardiovascular diseases [1]. Every year, millions of people get this disease, and half lose their lives. Although chemotherapy is one of the most effective and extensive treatments to combat this malady, the lack of selectivity of the drugs joined to the secondary effects caused by them or the development of drug-resistance experimented by the patients reduces the efficacy of this approach [2]. Consequently, this supposes a big problem of health in many developed countries, and a great number of research groups have focused their efforts on the study of this mortal illness [3-12]. Therefore, there is a current need for discovering new techniques and drug candidates overall due to the high mortality rates [13-15].

## 1.1. Breast Cancer

Among the different types of cancer, which overall represent approximately 30% of premature deaths

among the adult population aged 30-69 [16], breast cancer is postulated to be the predominant form of cancer in the female population, involving more than 50% of the reported cases of this disease [17].

Although hereditary and genetic factors such as personal or family history of breast or ovarian cancer and inherited mutations (in BRCA1, BRCA2 and other breast cancer susceptibility genes) account for 5-10% of breast cancer cases, studies in relation to migration processes have shown that non-hereditary factors are the main drivers of international and inter-ethnic differences. This has been demonstrated in comparative studies of low-risk populations that have migrated to high-risk populations, the movement of which has led to increased breast cancer incidence rates in subsequent generations [18].

These increased incidence rates in countries with higher HDI (Human Development Index, a summary measure of average achievement in dimensions of human development: a long and healthy life, being knowledgeable and having a decent standard of living) are associated with a higher prevalence of known risk factors. On the other hand, high incidence rates in transition countries are related to a higher prevalence of

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Fig. (1). Examples of pyrazole-based biologically active compounds.

known risk factors concerning menstruation, reproduction, nutrition or anthropometry, among others [19].

However, knowledge of this disease in terms of geographical or temporal variations in rates and specific etiological factors is still limited. Looking at different geographical areas, it can be observed that countries, where incidence rates have historically been low have experienced a marked increase during the last decades (e.g., in South American, African or Asian countries) [20]. These trends are probably due to a combination of demographic factors related to social and economic development. Similarly, among other aspects, the decline in incidence in the early 2000s in countries such as the United States, Canada, the United Kingdom and France was attributed to a decrease in the use of postmenopausal hormone treatment following the report of the Women's Health Initiative study linking the use of postmenopausal hormones with an increased risk of breast cancer [21].

The major risk factors for breast cancer are not easily modifiable because they derive from endogenous and prolonged hormonal exposures, although prevention by reducing alcohol consumption, engaging in physical activity, enhancing breastfeeding during the postpartum period or limiting hormone therapy may be beneficial.

#### 1.2. Heterocycle-Based Drugs

Within the plethora of anticancer drugs, those based on heterocycles have received special attention [22-27]. Among them, the pyrazole core is present in many of these potential active compounds (Fig. 1), and new synthetic methods have been developed in the last two decades to achieve a great scope of this interesting family of targets [28].

In view of its large pharmacological scope, the pyrazole skeleton is considered a privileged scaffold in medicinal chemistry because of its properties as antiinfluenza [29], antiviral [30-32], anti-inflammatory [33], antitubercular, antimicrobial, antioxidant, antidepressant, and insecticidal agent [34-36]. Related to cancer treatment, Crizotinib and Ruxolitinib [37] or Celecoxib [38-40] are important pyrazole-based anticancer drugs [41, 42]. Interestingly, there are also pyrazole-based drugs commercially available such as those described in Fig. (2) [43], among others.

Like many other drugs, pyrazoles exhibit activity due to their inhibitory capacity of diverse pivotal targets directly involved in the mechanism of cancer. In this work, we will also consider different aspects related to the role of pyrazole in the mechanism of cellular death. It is also remarkable that the rational design of new pyrazole scaffolds could be of interest in new drug discovery.

We would like to give the readers a broad overview of the plethora of pyrazole structures synthesized in the last two decades (2003-2021), emphasizing their utility as privileged scaffolds in the context of breast cancer, as previously mentioned, one of the most widespread cancers in women [44]. In this way, various representative examples of highly substituted pyrazole scaffolds



Fig. (2). Examples of pyrazole-based drugs are approved for commercialization [43].

and their pharmacological activity as potential anticancer drugs are illustrated and discussed in this work.

# 2. PYRAZOLES AS PROMISING ANTITUMOR AGENTS AGAINST BREAST CANCER

Hereafter, a great number of studies related to the different pyrazole derivatives with interesting results as antitumor agents are reported. Some of the sections in which this review has been structured are their capacity as kinase inhibitors, angiogenic agents, and those that act on estrogen-dependent factors or studies based on the synergistic effect with other structures of interest. Although the tumor lines studied in most research are diverse (and will be mentioned), only the results focused on breast cancer cells will be highlighted in this revision.

#### 2.1. Role as Kinase Inhibitors

Biological functions in eukaryotes are regulated by phosphorylation and dephosphorylation of proteins by kinases and phosphatases, respectively [45]. Therefore, the blocking of kinases is considered for possible therapies against cancer as well as inflammatory diseases. Hence, the human genome encodes many of these proteins, some of which are recognized as diseaserelevant, and small molecules targeting the ATPbinding site in these enzymes may inhibit the phosphorylation selectively, despite its ubiquitous nature [46, 47].

Moreover, selective inhibitors can be modelled by optimizing the peripheral groups, with the pyrazole moiety being a favored structure targeting kinases [48-51]. Over the last decade, much research has highlighted the use of pyrazole derivatives as kinase inhibitors with a broad spectrum of biological properties with a high pharmacological interest in different areas, like those referred to antimicrobial activity and neurodegenerative disorders or even cancer, among others [52].

In this context, Nielsen and co-workers carried out the synthesis of highly functionalized pyrazole



Fig. (3). Pyrazoles 1 with protein kinase inhibition (A) and antitumoral activity (B) [53].

carboxamides and carboxylic acids by saponification and transamidation, respectively, of esterfunctionalized pyrazoles [53]. Following this synthetic protocol, pyrazole scaffolds were tuned to optimize protein kinase inhibition, being tested on AKT1, CK2, PKA, PKC $\alpha$  and SAPK2a (p38) (each compound was studied using a final concentration of 10 µM). The most promising inhibitor compounds are depicted in Fig. (3). Thus, ester 1a showed significant inhibition of AKT1 and SAPK2a (p38) kinases, reducing activity by 40% and 35%, respectively (Fig. 3A), while carboxylic acid 1b inhibited AKT1 by 46% and PKA by 82% (Fig. 3A). Additionally, the most promising demonstration of inhibitory activity was observed for amide 1c with a lipophilic attachment, which inhibits several kinases (AKT1 by 61%, CK2 by 42% and PKA by 84%), indicating its multi-targeting approach (Fig. 3, A).

It is known that a drug targeting, for instance, AKT, is expected to affect cell growth and viability. Therefore, to further evaluate the physiological effect of these compounds, the authors first analyzed human MCF-7 breast cancer cells by phase-contrast microscopy either untreated or incubated for 48 h with the tested product at concentrations of 50 and 100  $\mu$ M. The morphological effects revealed that treatment with **1a** and **1c** had a clear inhibitory effect on cell growth (especially at the higher concentration), while treatment of the cells with **1b** caused no change in cell growth (Fig. **3B**, (a)). However, since no large numbers of floating dead cells were observed under the microscope for none of the compounds, it seems they did not cause severe cell death.

Later, the crystal violet binding method was used to quantify the effect of the selected inhibitors on cell growth (Fig. **3B**, (b)). Then, incubation of cells for 24 h with ester (**1a**) at 50 and 100  $\mu$ M concentrations resulted in a ~50% decrease in viability, reaching up to 24% of cells remaining with 100  $\mu$ M after 72 h; although the best result corresponded to amide **1c**, which induced the loss of 90% of cells under the same conditions (100  $\mu$ M after 72 h). However, carboxylic acid **1b** did not lead to a significant alteration. Since crystal violet staining does not distinguish between actively dividing and quiescent cells, the reduction in cell numbers could result from cytotoxic and cytostatic effects.

Moreover, the distribution of cells was analyzed in different phases of the MCF-7 cell cycle before and after treatment with 100  $\mu$ M of these pyrazole derivatives for 24 h and 48 h (Fig. **3B**, (c)). In this case, only incubation with **1c** for 48 h produced cell cycle arrest (S phase: 7.7% (control)  $\rightarrow$  20.3% (treated cells) and G2/M phase: 11.1% (control cells)  $\rightarrow$  22.4% (treated cells)), while the other two compounds **1a** and **1b** did not cause any variation in it. According to these data, the effect of amide **1c** over cell viability can be



Fig. (4). Selected compounds 2a-f to carry out the 60-cell-line screening at the National Cancer Institute (NCI), Bethesda, Maryland, USA and  $IC_{50}$  values in  $\mu$ M for six different cell lines of breast cancer [54].

attributed to arrested cells, while ester **1a** influence can be associated to deceleration of cell metabolism.

In a study reported in 2010 by Lee's group, 1*H*- and 2*H*-pyrazole derivatives were designed and synthesized for an extensive structure-activity relationship study [54] on the bases of the potent B-Raf inhibitor GDC-0879 (Fig. 1) [55]. In Fig. (4), six compounds out of thirteen (**2a-f**) showed very good activity in preliminary assays at 10  $\mu$ M and were derived in subsequent assays to determine the IC<sub>50</sub> in 60 cell lines. In particular, excellent values were obtained for six breast cancer cell lines (Fig. 4), which in some cases were in the nanomolar range (*i.e.*, 26 nM for **2c** in MDA-MB-468).

The results obtained from the *in vitro* assays, together with the structure-activity relationship (SAR) correlation studies, revealed that to obtain the best activity, the compounds should be substituted at the 4position of the pyrazole with a 4-(dimethylamino) phenyl group (**2a**, **2c** and **2d**) at the 2-position of the terminal pyridyl group, followed by 4-phenoxyphenyl (**2e**) and 4-acetyl (**2b**) substitution. The acetonitrile group at the 2-position of the pyrazole ring (2*H* isomers) was also enhanced over the 1-position. In addition, the effect of demethylation of the methoxy group (**2a**) to produce the hydroxyl analogues (**2d-e**) was not constant in the derivatives, suggesting the possibility of variable binding modes of these compounds at their target site or the presence of different targets, whose binding mode at each molecule would be different.

Moreover, following the interest in kinase inhibition and the understanding of the mechanism of action, compound **2c**, which had the highest potency in almost all cancer cell lines, was tested at 10  $\mu$ M concentration on a panel of 54 kinases at Reaction Biology Corporation, showing very good selectivity for FLT3 kinase with an inhibition percentage of 82.3%. This would mean that this compound could be used as a good new lead for developing selective FLT3 kinase inhibitors [56-58]. However, the moderate potency of the compound on FLT3 kinase (IC<sub>50</sub> = 1.74  $\mu$ M) could not justify its strong and broad-spectrum anticancer activity, and this suggested the presence of other underlying mechanisms that (in addition to moderate FLT3 kinase) might control the anticancer activity.

As well-known, the epidermal growth factor receptor (EGFR) kinase plays a crucial role in signal transduction pathways that regulate cell division, differentiation and migration. As usual, its abnormal function implies the development of illness. Thus, the overexpression of EGFR has been observed in many human cancers, such as bladder, breast, colon and lung cancers [59, 60]. In this context, small molecules may inhibit the kinase activity of EGFR by binding the ATP site of the intracellular tyrosine kinase domain to block



Fig. (5). Cinnamic acid metronidazole ester derivatives 3 were evaluated as anticancer agents, and  $IC_{50}$  values for the two most active molecules (3a-b) against the MCF-7 cell line [73].

	Inhibition of proliferation / MFC-7 (breast)							
	Compd.	R <sup>1</sup>	R <sup>2</sup>	IC <sub>50</sub> (μΜ)	Compd.	R <sup>1</sup>	R <sup>2</sup>	IC <sub>50</sub> (μM)
	4a	F	F	10.37	4s	Me	F	2.81
R <sup>2</sup>	4b		CI	8.57	4t		CI	2.35
``	4c		Br	8.22	4v		Br	1.50
	4d		Н	6.34	4w		Н	1.27
$\mathbb{N}^{\mathbb{P}}/\!/$	4e		Me	4.64 🖕	4x		Me	0.67 🚽
	4f		OMe	3.87	4у		OMe	0.32
_1	4g	CI	F	9.05	4z	OMe	F	2.02
	4h		CI	7.96	4aa		CI	1.97
) o	4i		Br	7.03	4ab		Br	1.15
$\langle A \rangle \rangle$	4j		Н	6.17	4ac		Н	0.92
	4k		Me	4.35 🖌	4ad		Me	0.56 🔺
// )	4		OMe	2.75	4ae		OMe	0.30
Ň	4m	н	F	7.45				
Ph	4n		CI	5.14				
4	4o		Br	4.74				
	4p		Н	4.05				
	4q		Ме	3.51 🖌				
	4r		OMe	2.11				

Fig. (6). Structure-activity relationship study of pyrazole-cinnamyl derivatives 4 [74].

tyrosine kinase (TK) autophosphorylation. This is a traditional area of research for developing anticancer drugs [61, 62] and many EGFR inhibitors have been used to treat human cancers [63-65].

Among other interesting structures, compounds containing pyrazole and cinnamyl moieties have shown high inhibitory activity against EGFR [66-70]. In particular, cinnamic acid ester derivatives have been postulated as structures with potential antitumor activity [71, 72]. For instance, Qian and co-workers found some metronidazole derivatives **3** to be potent inhibitors of EGFR and HER-2 (Fig. **5**), as well as they presented good antiproliferative activity against MCF-7 cell line, such as compounds **3a-b** which reached values of IC<sub>50</sub> of 0.36 and 0.98  $\mu$ M, respectively [73].

A few years later, the same group performed the synthesis and biological evaluation of other series of pyrazole-cinnamoyl derivatives **4** (Fig. **6**) [74]. Antiproliferative activity against MCF-7 (breast cancer) of the synthesized compounds was also tested in this case, showing remarkable effects. In addition, it was found that most of these derivatives presented good inhibitory activity against EGFR and HER-2. Among them, compound **4ae** exhibited the most potent activity (IC<sub>50</sub> = 0.30  $\mu$ M (MCF-7), IC<sub>50</sub> = 0.21  $\mu$ M (EGFR) and IC<sub>50</sub> = 1.08  $\mu$ M (HER-2)) (Fig. **6**).

Structure-activity relationship studies were performed by modification of the parent compound to determine how these substituents might affect the activity. Inspection of the chemical structures of compounds suggested that they could be divided into two subunits: The A-ring and the B-ring (Fig. 6). The introduction of an electron-donating group in both could enhance its antiproliferative activity, and the order of potency was OMe > Me. At the same time, compounds with electron-withdrawing groups in ring B caused a decrease in the activity.

		ОМе					
			Me				
R		N N	Me				
	5a-d 6a-g	R <sup>2</sup>		MDA-MB-468	IC <sub>50</sub> (μM)	AG01522	_
Compd.	1 <i>H</i> -pyrazole	R <sup>1</sup>	R <sup>2</sup>	24 h	48 h	24 h	
5a	-	Н	Н	19.35	11.03	31.19	
5b	-	OMe	Me	58.20	11.95	34.56	
5c	-	OMe	Cl	28.76	12.42	25.19	
5d	-	OMe	F	58.58	24.46	34.41	
6a	4,5-dihydro	Н	н	25.00	6.41	28.80	
6b	4,5-dihydro	Н	Me	19.20	9.29	29.82	
6c	4,5-dihydro	Н	CI	58.61	19.87	24.10	
6d	4,5-dihydro	Н	F	32.58	15.25	42.15	
6e	4,5-dihydro	OMe	н	21.31	10.81	42.79	
6f	4,5-dihydro	OMe	Me	14.97	6.45	28.74	
6g	4,5-dihydro	OMe	CI	25.83	8.50	43.30	

Fig. (7). Inhibitory growth on breast cancer cells (MDA-MB-468) and toxicity on normal fibroblast cell line (AGO1522) of synthesized compounds **5a-d** and **6a-g** [75].

The authors further performed a molecular modeling study to explore the interaction with the active site of EGFR/HER-2, taking as a reference compound 4ae, which had provided the best inhibition values. It was found to bind very efficiently to the ATP binding site of EGFR through hydrophobic interactions and stabilized by a hydrogen bond and a  $\pi$ - $\sigma$  interaction. Thus, the oxygen atom (OMe group) at the A-ring was able to form a hydrogen bond with the amino hydrogen of Lys A: 721 (N-H···O = 2.477  $\Box$ ; 130.6°). A  $\pi$ - $\sigma$  interaction between Leu694 and the benzene ring (bond length: 2.48  $\square$ ) was also confirmed. Similarly, it was seen that compound 4ae bound to the active site of HER-2 via hydrogen bonding through the formation of a hydrogen bond between that oxygen atom and the amino hydrogen of Met A: 801 (N-H···O =  $2.278 \Box; 145.8^{\circ}$ ).

Among the most recent research, Shahsavari's group has focused on triple-negative breast cancer [75], which is approximately 15-20% of all breast carcinomas and is associated with an earlier age of onset, an aggressive clinical course and a poor prognosis [76].

During this study, the activity of a series of 1,3diaryl-5-(3,4,5-trimethoxyphenyl)-1*H*-pyrazoles (5) and 1,3-diaryl-5-(3,4,5-trimethoxyphenyl)-4,5-dihydro-1*H*-pyrazoles (6) was evaluated against MDA-MB-468, the triple-negative human breast cancer cell line, and the *in vitro* cytotoxicity assays revealed its dose- and time-dependent anti-proliferative properties (Fig. 7). 3-(4-Methoxyphenyl)-1-(*p*-tolyl)-5-(3,4,5-trimethoxyphenyl)-4,5-dihydro-1*H*-pyrazole (6f) resulted in being the most active compound against MDA-MB-468, with IC<sub>50</sub> values of 14.97  $\mu$ M and 6.45  $\mu$ M, after 24 and 48 hours, respectively, compared to the positive control (Paclitaxel, IC<sub>50</sub> values of 49.90  $\mu$ M and 25.19  $\mu$ M). This compound also induces the minimum toxicity on normal fibroblast cell lines (AGO1522), with IC<sub>50</sub> values of 28.74  $\mu$ M and 20.33  $\mu$ M, after 24 and 48 hours, respectively.

Additionally, a cell cycle study was performed after treatment with 14.97 µM of the most active compound (6f) after 24 h, concluding that the arrest occurred during the S phase. The cell death assays determined a significant increase in the early apoptotic phase in treated cells, concluding that apoptosis, and not necrosis, was the mechanism triggered by these compounds. Once this was known, the role of reactive oxygen species (ROS) in the induction of apoptosis was studied. The results of these assays led to the conclusion that ROS plays an important role in the apoptosis process. In addition, the fluorometric assay kit was used to measure the activity of caspase 3 as the main protease in the signaling of the apoptosis process, and a significant increase in its activity was observed in the cells treated with the candidate compound 6f compared to the control.

#### 2.2. Estrogen Dependant Anti-Cancer Derivatives

Estrogens control the normal function of female sex organs, secondary sex characteristics, and mammary glands. Moreover, some breast cancers are estrogendependent, where the binding of estrogens to their receptors activates the transcription of their target genes,



Fig. (8). General structures of 4-cyano-1,5-diphenylpyrazole derivatives 7 and 8 evaluated [87].

the last ones responsible for the proliferation of cancer cells in the tumor [77]. In this context, therapy based on anti-estrogens is used in several diseases, such as breast malignancies [78], including estrogen receptor antagonists, such as tamoxifen [79, 80] and toremifene [81, 82]. The discovery of the effect of cyclooxygen-ase-2 (COX-2) inhibitors, such as celecoxib (Fig. 2) or meloxicam [83] in many types of malignancies, especially in breast cancer [84-86], made 1,5-diphenylpyrazoles became important structures to work with.

Motivated by this background, Farag and coworkers were interested in synthesizing several 4cyano-1,5-diphenylpyrazoles linked to different heterocyclic ring systems (**7a-k**) and acyl hydrazide derivatives (**8a-i**) at the 3-position (Fig. **8**) to test their antiestrogenic effects *in vivo* and to evaluate their cytotoxic properties against estrogen-dependent tumors *in vitro* [87].

The *in vitro* assays were performed on seven breast tumor cell lines, namely MDA/MB-231 ATTC, MCF-7, T-47D, NCI/ADR-RES, HS-578T, MDA/MB-435, MiDA-N and BT549, as well as on additional ovarian cancer cells (6 lines), which were also addressed in this study.

Tables 1 and 2 collect the results as growth inhibitory concentration (GI<sub>50</sub>) values for all the breast cancer lines tested, showing good activity (below 10  $\mu$ M) in red. Regarding the effect of the structure, some correlations were found and are summarized as follows. Analyzing first the results obtained for a big family of triazoles (**7a-i**, Table 1), it is worth mentioning how variations on mercaptotriazole derivative **7a** affect, which revealed good activity (2.48-6.68  $\mu$ M) against five out of eight cell lines. Concerning the substitution on the N, increasing lipophilicity with a phenyl group (**7b**) resulted in total inactivity, while a more hydrophilic substituent, such as an amino group (**7c**), improved the activity against only one of the cell lines (HS-578T, 8.94  $\mu$ M) for which compound **7a** was inactive.

Additional changes on the inactive molecule 7b, such as methylation of the thiol group (7d), caused little improvement in the activity, getting  $GI_{50}$  up to 36.3 µM, whereas produced no effect in the case of replacement of the thiol group by a hydrazine moiety (7e), which was also totally inactive. The idea of generating fused ring systems (7f-i) out of the amino and thiol groups of molecule 7c had different effects depending on the cell line. Hence, in the case of the two derivatives with a second 5-membered ring (7h-i), aromatic substitution (Ph) rendered a totally inactive compound (7h), while aliphatic substitution (Bn) involved improvement only for two of the cell lines (MDA MB-231 ATTC and NCI ADR-RES), keeping similar activity for the other ones. The other two derivatives with a second 6-membered ring (7f-g) both contain aromatic substitution; a curious complementarity is observed in the case of a phenyl group (7f), compared to parent molecule 7c, meaning that the most sensitive cells (HS-578T, MDA MB-435 and MiDA-N) for the later (7c), are the most resistant ones for the former (7f). Exactly the same occurs when the comparison is between 7f and 7i. In general, introducing a methoxy group in the *para*-position of the phenyl substituent (7g) translates into the best activity, except against MCF-7, T-47D and BT549, compared to both its parent molecule 7c and its homologous 7f. In fact, the best activity within all newly synthesized compounds is achieved in derivative 7g with GI<sub>50</sub> values of 1.19 and 1.23 µM against MiDA-N and MDA MB-435, respectively.

Finally, the change of the N of the triazole by another heteroatom was examined. In this context, the activity of oxadiazole **7k** is worst compared to its homologous triazole **7a**, except against MCF-7, HS-578T and BT549. Thiadiazole **7j**, which is not exactly comparable with **7a** because of its different substitution at the C-position (**7j**: NHPh and **7a**: SH), gave good inhibition in the case of MDA MB-231 ATTC and MCF-7 (5.84 and 8.53  $\mu$ M, respectively). Table 1. Molecular structures 7a-k and *in vitro* results for different breast cancer cell lines (Growth inhibitory concentration (GI<sub>50</sub>, μM)).

	Compounds 7a-k	MDA MB-231 ATTC	MCF-7	T-47D	NCI ADR-RES	HS-578T	MDA MB-435	MiDA- N	BT 549
7a	N <sup>N</sup> SH	6.27	IA*	6.68	2.48	IA*	2.64	2.58	IA*
7b	N SH N N Vn Ph	IA*	IA*	IA*	IA*	IA*	IA*	IA*	IA*
7c	N SH	IA*	23.4	39.9	IA*	8.94	2.93	5.00	IA*
7d	N SMe N N Y	IA*	68.0	36.3	$\mathrm{IA}^*$	69.8	41.8	73.8	44.8
7e	N <sup>N</sup> NHNH <sub>2</sub> N <sup>2</sup> <sup>2</sup> <sup>2</sup> Ph	IA*	IA*	IA*	IA*	IA*	IA*	IA*	IA*
7f	N S N N N Ph	13.2	19.8	9.08	28.0	IA*	48.0	73.0	51.6
7g	N <sup>N</sup> S NN OMe	5.49	IA*	IA*	8.4	9.23	1.23	1.19	IA*
7h	N/N/S	IA*	IA*	IA*	IA*	IA*	IA*	IA*	IA*
7i	N/N/S	52.1	41.6	42.6	74.7	8.94	9.23	5.49	IA*
7j	N <sup>N</sup> NHPh S	5.84	8.53	18.0	99.8	IA*	55.8	93.8	IA*
7k	N SH	38.4	38.0	72.0	99.0	98.0	IA*	55.4	26.2

Note: \*IA: Inactive;  $GI_{50} > 100 \mu M$ .

Table 2. Molecular structures (8a-i) and *in vitro* results for different breast cancer cell lines (Growth inhibitory concentration (GI<sub>50</sub>, μM)).

	Compounds 8a-i	MDA MB-231 ATTC	MCF-7	T-47D	NCI ADR- RES	HS- 578T	MDA MB- 435	MiDA-N	ВТ 549
8a	O NH <sub>2</sub>	61.2	85.8	NT**	NT**	NT**	NT**	1.34	9.13
8b		IA*	99.8	45.6	IA*	61.8	42.8	34.8	37.7
8c		4.90	IA*	63.4	IA*	8.04	9.30	2.86	IA*
8d	O N N N	6.21	IA*	2.45	2.76	IA*	2.75	IA*	IA*
8e	O N N O	23.8	19.8	54.8	98.0	IA*	39.9	23.8	11.8
8f	O N S NH NH Ph	7.36	4.72	62.5	7.4	6.72	8.62	7.00	4.12
8g		28.0	40.8	22.2	IA*	61.8	61.0	56.8	11.8
8h		IA*	IA*	IA*	IA*	IA*	IA*	IA*	IA*
8i		IA*	79.8	26.5	28.0	IA*	IA*	26.5	11.8

\*IA: Inactive; GI<sub>50</sub> >100 µM, \*\*NT: not tested.

The second line of structural optimization was carried out based on the remarkable cytotoxicity of the carboxylic acid hydrazide derivative **8a** (GI<sub>50</sub> 1.34  $\mu$ M against MiDA-N, Table **2**). Thus, for instance, the thiosemicarbazide derivative **8b** was found to have moder-

ate to weak activity, whereas the potassium salt 8c provided good inhibition between 2.86 and 9.30  $\mu$ M against MDA MB-231 ATTC, HS-578T, MDA MB-435 and MiDA-N cell lines.

Additionally, the pyrazole derivative **8d** seemed cytotoxic against MDA MB-231 ATTC, T-47D, NCI ADR-RES and MDA MB-435 lines (6.21, 2.45, 2.76 and 2.75  $\mu$ M, respectively), while in general, its homologous pyrazolone **8e** had a moderate to weak cytotoxicity.

Finally, the authors also explored incorporating a 5membered ring containing S and N in the 2-N position of the acyl hydrazide (**8f-i**). Thus, the integration of phenylthiazolidene led to compound **8f** with the broadest scope, showing GI<sub>50</sub> values ranging between 4.12 and 8.62  $\mu$ M for all the cell lines except T-47D (62.5  $\mu$ M) and either replacing the phenyl group with a methyl group (**8i**) or incorporating a chlorine atom at the *para*-position of it (**8g**) allowed the improvement of the activity against this cell line (26.5 and 22.2  $\mu$ M, respectively), being the inhibition worse for the rest of tested cell lines. Curiously, substitution in the *para*position of the phenyl ring with a methoxy group turned the compound (**8h**) inactive.

Acute *in vivo* toxicity studies were performed on a representative sample of the novel compounds synthesized. Rats were orally treated with different doses of an aqueous suspension of a very fine powder of the tested compounds. Under these conditions, as shown in Table **3**, the tested compounds were found to be highly safe (LD<sub>50</sub> values) for albino rats compared to the reference drug letrozole [88].

Table 3.	Acute toxicity (LD <sub>50</sub> ) of the synthesized co	om-
	pounds and letrozole.	

Compounds	LD <sub>50</sub> (mg/kg)
Letrozole	$252.61\pm0.11$
7a	$434.71\pm0.13$
7b	$122.48\pm0.13$
7k	$323.06\pm0.11$
8a	$332.11\pm0.13$
8b	$311.52\pm0.12$
8c	$431.85\pm0.13$
8d	$873.06\pm0.19$
8e	$234.71\pm0.13$

Additionally, many pathological complications (breast cancer, endometrial cancer, osteoporosis, *etc.*) originate in aberrant expression of estrogen receptor alpha (ER- $\alpha$ ). It plays a key role in breast cancer initiation and progression and confers chemoresistance to

cancer cells by regulating the expression of antiapoptotic proteins (Bcl2) [89]. The overexpression of estrogen receptors is responsible for 70% of breast cancer cases, named estrogen receptor-positive tumors [90, 91].

On the base that pyrazoles are good ligands for estrogen receptors, Rangappa and coworkers have been working in the last years on the synthesis and study of the antitumoral activity of novel derivatives. From a series of novel 1-(4-methoxybenzyl)-3-cyclopropyl-1*H*-pyrazol-5-amine derivatives **9a-h**, some compounds exhibited interesting growth inhibitory effects against MCF-7 cell line (Fig. **9**) [92]. Structure-activity relationship studies revealed that electron donating groups on the phenyl ring, such as 3-methoxy (**9b**), 3methyl (**9g**), and 4-*tert*-butyl (**9d**), provided significant activity, whereas electrons withdrawing groups, such as 4-nitro (**9a**), 4-chloro (**9c**), 2-fluoro (**9e**) and 3-bromo (**9f**) delivered poor activity.

Later in 2017, these authors prepared 1-aryl-3,5bis(het)aryl pyrazole derivatives with complementary regioselectivity: **10a-e** *versus* **11a-e**, and also evaluated *in vitro* their activity against several breast cancer cell lines, such as MCF-7 and T47D (Fig. **10**) [93].

Compound **11d** showed the highest antiproliferative effects from this series, blocking cells in the sub-G1 phase of the cell cycle and promoting apoptosis. In addition, it was concluded that the compound decreased mitochondrial membrane potential, induced DNA fragmentation and chromatin condensation, and necessitated caspases cleavage in breast cancer cell lines [85].

Authors continued their work searching for new anti-estrogens or selective estrogen receptor modulators (SERMs) and published in 2018 further studies using compound **11d** (Fig. **10**) [94]. Molecular docking studies showed that fixes efficiently to the ligand binding domain of ER- $\alpha$  via hydrophobic interactions with a receptor pocket [95]. Additionally, *in vivo* assays were performed in Sprague-Dawley (SD) rats, carriers of a DMBA-induced mammary gland tumor. Twenty-five days after inoculation, animals were sacrificed, and the mammary glands were removed for evaluation. Treatment with the drug candidate reduced tumor size, indicating its ER- $\alpha$  antagonistic effect.

#### 2.3. Pyrazoles with Anti-Angiogenic Activity

The growth of blood vessels (a process known as angiogenesis) is essential for many physiological procedures (embryogenesis, organ development, wound healing, *etc.*) [96, 97]. However, unregulated



Fig. (9). 1-(4-Methoxybenzyl)-3-cyclopropyl-1*H*-pyrazol-5-amine derivatives 9a-h and IC<sub>50</sub> (µM) values against MCF-7 [92].



Paclitaxel: 0.0096 µM (MCF7); not tested (T47D)

Fig. (10). 1-Aryl-3,5-bis(het)aryl pyrazole derivatives 10a-e and 11a-e and IC<sub>50</sub> ( $\mu$ M) values against MCF-7 and T47D [93, 94].

angiogenesis has been associated with different diseases, such as cancer, retinopathy, rheumatoid arthritis, psoriasis, atherosclerosis and hemangioma [98-101]. Antiangiogenic therapy targets activated endothelial cells and have gained increasing interest for its various advantages over targeted therapy. In addition, targets for anti-angiogenic therapy are easily accessible by systemic administration, as 10 out of 100 new tumor cells require at least one new endothelial cell (one gram of tumor contains approximately 20 million endothelial cells and 100 million to 1 billion tumor cells). Therefore, inhibiting angiogenesis and stopping the growth of an endothelial cell could have an amplified effect on tumor cells [102].

In this sense, and taking into account the good results achieved in terms of apoptosis (the programmed cell death that maintains the balance between cell survival and cell death) for some tumor cell lines previously reported using structures such as celecoxib and SC-236 (Figs. 1 and 2) [103-105], Abadi and coworkers carried out in 2003 the synthesis of a battery of 1,3,4-trisubstituted pyrazole derivatives. With a similar substitution pattern, those new structures proved to be potential antitumors and antiangiogenic agents [106].

The prepared compounds were evaluated in preliminary assays in 3 tumor cell lines (MCF-7 (breast), NCI-H460 (lung) and SF268 (central nervous system)) with a  $10^{-4}$  M drug concentration, and the results were provided as a percentage of cell growth after 48 h of incubation. Of thirty-four compounds tested, **12a-b** (Fig. **11**) were the most active in MCF-7, being compound **12a** the best one which provided a cell growth inhibition of 99%.



Fig. (11). Selected compounds 12a-b as the most active against MCF-7 cancer cell line [106].



Inhibition of proliferation / MFC-7 (breast)

R <sup>1</sup> – R <sup>4</sup>	Compd.	IC <sub>50</sub> (μΜ)	Compd.	IC <sub>50</sub> (μΜ)
$R^1 = R^2 = R^3 = R^4 = H$	13a	47	14a	5.8
$R^1 = R^2 = R^4 = H, R^3 = OH$	13b	29	14b	26
$R^1 = R^3 = R^4 = H, R^2 = OH$	13c	38	14c	25
R <sup>1</sup> = R <sup>4</sup> = H, R <sup>2</sup> = R <sup>3</sup> = OH	13d	24		
$R^2 = R^3 = H, R^1 = R^4 = OH$	13e	≥100		

Fig. (12). Cytostatic activity of compounds 13 and 14 [110].

A second test was performed for both compounds in in vitro screening, which provides the effect on cell growth in a panel of approximately 60 human tumor lines. These lines were divided into subpanels according to different representative tumor cell lines such as leukemia, ovary, prostate, breast or central nervous system. Although no comparatives were performed with normal cells, the ratio obtained by dividing the mean graph mid-point (MG-MID) (µM) of the full panel of the compound by the MG-MID  $(\mu M)$  of its subpanel is considered an indicator of selectivity. Ratios between 3 and 6 indicate moderate selectivity, while those above 6 indicate high selectivity towards the corresponding subpanel [107-109]. Compound 12a exhibited significant anticancer activity with a broad spectrum of action, although without being selective towards a specific tumor line.

Furthermore, subsequent studies determined that these 1,3,4-trisubstituted pyrazole derivatives showed interesting antiangiogenetic activity, even in compounds that did not present cytotoxicity in the preliminary assays.

In 2010 the group of Liekens and Haroutounian reported the synthesis of novel fused pyrazolo[4,3c]quinolone derivatives as potential anti-angiogenic agents, combining in a single backbone the known pharmacophore quinoline ring with a substituted pyrazole. In the course of this investigation, the authors also evaluated the cytotoxicity against various tumor cell lines, including MFC-7 (breast) (Fig. 12) [110]. Therefore, *in vitro* tumor cell proliferation inhibition studies were performed for hydroxyl pyrazolocarboxaldehydes 13a-e and hydroxyl pyrazoloquinolin-4-ones 14a-c. Notably, most of the tested compounds showed moderate IC<sub>50</sub> values for the MCF-7 line between 20 and 50  $\mu$ M, being compound 14a the most promising, with an IC<sub>50</sub> value of 5.8  $\mu$ M.

The advantages associated with steroid-based chemotherapeutics [111-113], including non-toxicity, reduced vulnerability to multidrug resistance (MDR)



Fig. (13). Pyrazole derivatives 15a-f and 16a-d synthesized from pregnenolone and their in vitro cytotoxic results [120, 121].

and high bioavailability, led to a growing interest in the modification of steroid molecules for their biological properties. This was demonstrated by different ring modification studies of steroid molecules involving the A- and D-rings, in which the incorporation of heteroatoms (N or O) was reported to enhance the biological activities of these molecules, including antimicrobial, anti-inflammatory, hypotensive, hypocholesterolemic and diuretic activities [114-117]. Among them, the natural steroid pregnenolone is a precursor of other hormones such as cortisone, estrogen, testosterone and progesterone [118, 119]. Under these conditions, in 2012, Mohareb's group used it as a template to develop new anticancer compounds [120, 121]. Modification of the D-ring by reaction with cyanoacetylhydrazine gave rise to the hydrazone derivative (non-shown), which finally underwent heterocyclization reactions to give the pyrazoles 15a-f (Fig. 13), pyridine (non-shown), thiazole (non-shown) and thiophene (non-shown) derivatives of pregnenolone [120]. In a parallel article, the same authors modified this precursor by reaction with 1-phenyl-3-thiosemicarbazide to give the thiosemicarbazone derivatives (non-shown), which, under the same hypothesis as above, were subjected to heterocyclization reactions to give the derivatives thiazolyl hydrazonoandrostane (non-shown) and pyrazolyl semicarbazidoandrostane derivatives 16a-d (Fig. 13) [121].

The cytotoxicity of the newly synthesized heterocyclic steroids was studied against three human tumor cell lines, namely non-small cell lung cancer (NCI-H460), central nervous system cancer (SF-268) and breast adenocarcinoma (MCF-7). The in vitro cytotoxicity assays showed little selectivity towards the different tumor lines studied, although very good GI<sub>50</sub> values were obtained. The data for the pyrazole derivatives of both studies against the MCF-7 line range between 0.01 and 50.1  $\mu$ M, as shown in Fig. (13). In fact, some of the tested compounds reached the nanomolar range and, in some cases, showed much higher inhibitory effects towards the three tumor cell lines than the reference drug, doxorubicin (MCF-7: 0.04 µM). Regarding the structure-activity relationship, it is possible to make some comments. For instance, in the case of pyrazoles 15a-f, it seems to be important the substitution of the phenyl group at the 5-position of pyrazole moiety, being the highly electronegative chlorine atom, the group that provided better GI<sub>50</sub> values (15c-d: 0.02 and 0.01  $\mu$ M), in comparison with the methoxy group (15e-f: 4.8 and 2.2  $\mu$ M) and the unsubstituted phenyl ring (15a-b: 30.0 and 22.3  $\mu$ M). On the other hand, in the case of pyrazoles 16c-d, their higher inhibitory effect may be due to the presence of an electron-withdrawing OH group -again at the 5-position of pyrazole moiety(0.01 and 0.02  $\mu$ M), instead of a methyl group (**16a-b**: 50.1 and 38.0  $\mu$ M).

#### 2.4. Candidates with Anti-Invasive Effects

Up to now, cancer treatment has mainly focused on growth reduction. Surgery, radiotherapy, and chemotherapy have all proven to be very effective in tackling tumor volume. However, cancer patients usually do not die from growth effects but the sequelae of invasion and metastasis.

The molecular mechanisms of invasion are not completely understood, but important contributions have been ascribed to proteases secreted by cancer cells, which can degrade the surrounding extracellular matrix (ECM) to cancer cell motility factors and factors that disrupt cell-cell adhesion [122, 123]. In view of the molecular cross-talks between cancer cells and host cells, receptor signaling through cytoplasmic tyrosine and serine/threonine kinases has been recognized as an important study target in cancer research. It was this growing interest in tissue invasion, a hallmark of malignant tumors responsible for poor prognosis in untreated cancer patients, that encouraged the group of Parmar to search for a new anti-invasive treatment, screening a total of ninety-five compounds from different classes of heterocyclic scaffolds, including lactones, coumarins, chromans and pyrazoles, among others [124]. These derivatives were tested against organotypic confrontation cultures of invasive human mammary carcinoma MCF-7/6 cells with embryonic chicken heart fragments in vitro. The assay consisted of three-dimensional confrontations between tumor cells and normal tissue.

In control cultures treated with solvent, MCF-7/6 cells invaded the PHF (precultured heart fragments) after 8 days of incubation. Many compounds showed no inhibition of invasion, and histologically their cultures were similar to controls. Other compounds were anti-invasive at the 10  $\mu$ M concentration. However, only three of the 95 compounds, including pyrazol (17) (Fig. 14), inhibited invasion of MCF-7/6 cells at a concentration as low as 1  $\mu$ M. Unfortunately, none of the compounds was active at 0.1  $\mu$ M. Furthermore, sulforhodamine B (SRB) assay showed no inhibition of MCF-7/6 cell growth by these three compounds, concluding that the compounds exhibited anti-invasive activity but were not cytotoxic to cancer cells.

Also, before performing the invasion assay on the synthesized compounds, several anti-invasive agents that inhibited both invasion and growth of tumor cells were screened. One of these compounds interfered with a common molecular target in mitosis and directional migration, namely vinca alkaloids, which bind to tubulin and prevent the assembly of the mitotic spindle and the cytoplasmic microtubule complex [125]. Another of the selected compounds was cytotoxic, with the antiinvasive effect resulting from a reduction in the number of intrinsically invasive cells in the challenged cultures.



Fig. (14). Pyrazole derivative 17 with anti-invasive activity [124].

However, the anti-invasive compounds in the study only limited invasion without affecting growth, which meant that their activity was not a result of cytotoxicity. These selective invasion inhibitors are structures of interest to understand which targets are involved in invasion mechanisms and which are involved in growth. The E-cadherin/catenin complex, for example, which is located in the cell membrane of normal epithelial cells, prevents epithelia from growing beyond their normal tissue boundaries [126]. In invasive tumors derived from epithelial tissues, the expression of this complex is down-regulated [127]. The complex in MCF-7/6 cells is inactive but is sensitive to functional regulation by several compounds shown to possess anti-invasive activity. Tangeretin, a citrus methoxyflavone, can increase E-cadherin-dependent cell-cell adhesion between MCF-7/6 cells and inhibit their invasion in the chicken heart invasion assay. This mechanism of action could be excluded for the anti-invasive compounds in the study, as no stimulation of cell aggregation was observed in vitro.

In summary, it was proved that the tested compounds did not inhibit tumor cell growth and did not have an effect through activation of the MCF-7/6 Ecadherin/catenin complex.

In addition, the broad battery of pyrazole-derived compounds synthesized allowed a further structureactivity study. Thirteen of the sixteen pyrazolyl acrylonitriles tested showed anti-invasive activity at 10  $\mu$ M on MCF-7/6 cells in the chicken heart invasion assay. Moreover, among the five pyrazolyl acrylonitriles synthesized with the 3,4-difluorophenyl substituent at the 3-position, three of them showed anti-invasive activity at a concentration of 10  $\mu$ M, but



Fig. (15). Compounds 18, 19 and 20 proved to be the most active ones in Rostom's study [128].

only compound 17 showed the activity at a concentration of 1  $\mu$ M.

These results revealed that the difluorophenyl moiety at the 3-position and the 4-chlorophenyl moiety at the 5-position of the pyrazole in compound **17** enhanced the anti-invasive activity of acrylonitriles.

### 2.5. Synergistic Effects Related to Structure-Activity Relationship (SAR) Studies

The growing interest in the biological activity of pyrazole derivatives and the study of pharmacophore groups in the cancer field led Rostom and co-workers to prepare compounds with a 4-hydroxypyrazole core and different heterocyclic systems such as triazoles and oxadiazoles, linked to it [128]. In this way, they aimed to achieve a chemotherapeutic synergy, increasing the selectivity and decreasing the toxicity of the substances. It seems that the substituent at the 4-position of the 1,2,4-triazolin-3-thiones influences the antitumor activity, as well as in the case of the 1,3,4-oxadiazoles this is closely related to the nature of the substituent at 2position. Thus, from the battery of synthesized compounds, 1,2,4-triazolin-3-thione 18 (4-cyclohexyl substituted) and 1,3,4-oxadiazoles 19 (2-one substituted) and 20 (2-phenyl substituted) (Fig. 15), presented very good GI<sub>50</sub> values for all the breast cancer lines tested: MCF-7, NCI/ADR-RES, HS 578T and MDA-MB-435, showing results below 0.01 µM in some cases. Among them, compound 20 showed an amazing profile of activity towards the rest of the cancer cell lines studied (26 lines) with GI<sub>50</sub> values lying in the nanomolar concentration range ( $< 0.01 \mu$ M).

Interestingly, for further *in vitro* studies, compounds **18** and **19** were selected by the National Cancer Institute (NCI) Development Therapeutic Program and compound **20** was taken to the NCI Biological Evaluation Committee.

In 2005, Lee's group became interested in heterocyclic cores restricted to a limited number of conformations and hindered in rotation by the substituents [129]. This led them to synthesize pyrazole oxime ether derivatives and to evaluate their inhibitory potential against different tumor cell lines through sulforhodamine B (SBR) assay [130]. Cytotoxicity results were shown as a percentage of growth inhibition after adding the different compounds at 1  $\mu$ M. Among the synthesized compounds differently substituted with alkyl or aromatic groups in the R<sup>1</sup> and R<sup>2</sup> positions, phenoxy in R<sup>3</sup> and benzyl or 2-aminoethyl group in R<sup>4</sup>, only three of them **21a-c** (with R<sup>1</sup>, R<sup>2</sup> = Me) had a moderate activity close to 40% inhibition at 1  $\mu$ M against MCF-7 (Fig. **16**).

Analyzing the structure-activity relationship, a decrease in activity was observed when a larger group replaced the methyl group. Compounds with a benzyl group at the  $R^4$  position showed potent cytotoxic activity against a wide range of tumor cell lines, which led to further investigation of the modification at the  $R^3$  position of the pyrazole framework, although it was not tested in breast cancer lines.

It is known that compounds based on a tricyclic planar structure consisting entirely or partially of anthraquinone, anthrapyrazole or acridine show interesting cytostatic and antitumor properties [131]. Furthermore, the presence of a five- or six-membered heterocyclic ring fused to the anthraquinone or acridine moiety can increase the activity and overcome the multidrug resistance of tumor cells [132, 133], there has been a growing interest in the development of these structures.



Fig. (16). Pyrazole oxime ether derivatives 21 as potential antitumor agents [129].



Fig. (17). Structures of acronycine and prepared xanthenone derivatives 22-24 [140-142].

Pouli's group has worked on the synthesis and biological studies of pyrano(thio)xanthenone derivatives [134-136], possessing structural similarity to the pyranoacridone alkaloid acronycine (Fig. 17), which showed promising antitumor properties in several murine solid tumor models [137] and was used as a lead for the synthesis of pharmacologically significantly improved analogues [138, 139]. A study of the structure-activity relationship in the pyranoxanthenone derivatives showed that substitution of the 6-methoxy group with a flexible (N,N-dialkylamino)ethylamino side chain substitution caused a strong enhancement of anti-proliferative activity towards the murine leukemia cell line L1210 (22, Fig. 17) [140]. Furthermore, pyrazole-fused homologues of these molecules (pyran moiety: 23 [141] and imidazole ring: 24 [142], (Fig. 17) were also synthesized and studied.

The results showed that incorporating a pyrazole ring in a xanthenone ring significantly increased antiproliferative activity. Fig. (17) presents the lowest IC<sub>50</sub> ( $\mu$ M) values obtained against the MDA-MB-231 breast cancer cell line for these pyrazole-fused xanthenone amino derivatives.

At the same time, Bandgar and coworkers synthesized a library of 3,5-diaryl pyrazole derivatives **25** to test them against five tumor cell lines, including breast cancer (Fig. **18**) [143]. The screening method selected to carry out these assays was the fluorescence test with propidium iodide [144]. Dyes such as propidium iodide (PI), which bind to DNA, provide a quick and accurate tool to quantify total nuclear DNA. The intensity of the PI fluorescence signal is directly proportional to the amount of DNA in each cell. Propidium iodide cannot penetrate an intact membrane, so the cells must first be permeabilized; therefore, there must be a cellular damaged.

The best results regarding anticancer activity for the MCF-7 cell line were found with those compounds with the R substitution at positions 2 or 4 in the aromatic ring, preferably with Cl or Br substituents. Subsequent structure-activity studies revealed that only the halosubstituents (Cl and Br) were directly implicated in enhancing the anticancer activity. In contrast, fluorine,



Fig. (18). 3,5-Diaryl pyrazole derivatives 25 synthesized for the biological assays [143].

methyl and methoxy groups as substituents produced a marked decrease in activity (63% inhibition in the case of 2-Cl (**25a**) *versus* 15% for 2-F (**25e**), and 11% for 2-CH<sub>3</sub> (**25f**) substitution, Fig. **18**).

However, it is not just the binding of the pyrazole structure to organic compounds that have been of interest in studying its biological properties. It is well known that ferrocene core has provided a versatile scaffold for the preparation of interesting structures useful in many areas such as catalysis, materials science, or bioinorganic chemistry [145-148]. In addition, numerous studies support that combining a ferrocenyl moiety with a heterocycle could increase their biological activity or give rise to new properties [149, 150]. Moreover, it has been demonstrated that ferrocenes could act in the activation of reactive oxygen species (ROS) formed in tumor cells, partially contributing to the antiproliferative effects of cancer cells [151-153]. On the other hand, ferrocenium salts were the first type of organometallic compounds reported to have antiproliferative properties, and nowadays, many ferrocenebased compounds are used as therapeutics [154, 155].

In this context, Ruan and co-workers, encouraged by their previous works [156, 157] and the properties of ferrocene derivatives, developed a variety of ferrocenes containing the pyrazole moiety **26** and **27** to study their activity as antitumor agents (Fig. **19**) [158].

In vitro assays were performed to study three tumor cell lines, including breast cancer MDA-MB-45, using 5-fluorouracil and cisplatin as positive controls (Fig. **19**). Although none of the synthesized compounds displayed IC<sub>50</sub> values lower than these references, the results were below 9  $\mu$ g/mL in most cases. In particular, compounds **26e,h,l** showed the strongest inhibitory activities against the MDA-MB-45 cell line, with IC<sub>50</sub> values lower than 6  $\mu$ g/mL. Therefore, these promising results, together with the low toxicity and the possibility of oral administration, positioned these compounds as good candidates for developing new anti-tumor drugs.

In search for new bioactive compounds, the pharmacophore hybridization strategy has been proven effective in modern medicinal chemistry since combining two molecules with different mechanisms of action often results in improved effects [159, 160]. In this context, and inspired by hybrid molecules containing benzimidazole (an interesting heterocycle for its biological properties and applications in medicine [161-163]) with enhanced anticancer activities [164, 165], Kamal, Shukla and coworkers undertook the synthesis of pyrazole derivatives **28** which incorporate that structural motif to produce promising new anticancer agents (Fig. **20**) [166].

The authors assessed the cytotoxicity of the prepared hybrids against HaCaT (normal keratinocyte cells) and three different cancer cell lines: A549 (lung), HeLa (cervix) and MCF-7 (breast, Fig. **20**). In general, growth inhibition was highly satisfactory, finding in many cases better results than those of controls used such as 5-fluorouracil (IC<sub>50</sub> = 2.36  $\mu$ M) and nocodazole (IC<sub>50</sub> = 1.6  $\mu$ M). In the case of the breast cancer line, values of IC<sub>50</sub> below 1  $\mu$ M were reached for several compounds (**28d,l,p**). In addition, the line corresponding to human epidermal keratinocyte provided numbers above 50  $\mu$ M in most of the assays, which indicated selectivity of the pyrazoles **28** towards cancer cells in contrast to normal HaCaT cells.

Structure-activity relationships generally showed that mono-substitution with an F, Cl, Br and OMe group on the benzimidazole B ring exhibited more potent activities than CF<sub>3</sub> groups and Me. In addition, di-substitution with Cl and Me groups demonstrated a lower inhibitory effect. On the other hand, it seems that substitution on the A ring did not influence significantly.



Positive controls: 5-fluorouracil (2.8  $\mu M)$  and cisplatin (1.14  $\mu M).$ 

Fig. (19). Ferrocene derivatives 26 and 27 used for the *in vitro* assays [158].



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Compd.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	IC <sub>50</sub> (μM)	Compd.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	IC <sub>50</sub> (μΜ)
28a	Н	Н	н	3.19	28u	CI	н	Н	3.03
28b		F		2.43	28v		F		2.83
28c		CI		1.96	28w		CI		1.17
28d		Br		0.83	28x		Br		1.92
28e		Me		5.94	28y		Me		8.23
28f		OMe		1.59	28z		OMe		1.76
28g		$CF_3$		2.66	28aa		CF <sub>3</sub>		4.21
28h		Me	Me	>50	28ab		Me	Me	13.0
28i		CI	CI	7.56	28ac		CI	CI	48.3
28j		cbz		12.6	28ad		cbz		13.9
28k	F	Н	Н	3.20	28ae	OMe	Н	Н	2.54
281		F		0.95	28af		F		1.51
28m		CI		2.23	28ag		CI		1.76
28n		Br		3.21	28ah		Br		1.83
280		Me		1.12	28ai		Me		6.61
28p		OMe		0.97	28aj		OMe		3.13
28q		$CF_3$		2.13	28ak		$CF_3$		5.73
28r		Me	Me	27.2	28al		Me	Me	12.8
28s		CI	CI	6.93	28am		CI	CI	13.2
28t		cbz		9.24	28an		cbz		7.46

Inhibition of proliferation / MFC-7 (breast)

Positive controls: 5-fluorouracil (2.36 µM) and nocodazole (1.6 µM).

Fig. (20). In vitro antiproliferative activity (IC<sub>50</sub>) of pyrazolo-benzimidazole derivatives 28 [166].

Moreover, the authors conducted further mechanistic studies with MCF-7 cells using the most potent compounds. For instance, it was found that the number of viable MCF-7 cells was significantly reduced after treatment with these compounds with concentrations corresponding to their  $IC_{50}$  for 48 h, compared to control cells.

Additional evaluation of the long-term cytotoxicity by measuring the ability of a single cell to grow into a colony revealed that these compounds inhibited MCF-7 cell colony formation, and all of them reduced clonogenic survival by about 50%.

As a potential metastatic measurement, the migration capacity of cancer cells is also a property of interest to be estimated. In this case, after treatment with pyrazolo-benzimidazoles, the amount of invasive breast cancer cells penetrating the respective wound areas was inhibited.

In addition, flow cytometry assays indicated that cell cycle arrest in the G0/G1 phase contributed to cytotoxicity. Studies conducted to elucidate the responsible mechanisms concluded that these compounds could decrease the levels of cyclins and cyclin-dependent kinases (CDKs), cell cycle regulators, leading to cell cycle arrest and, consequently, inhibiting the cell growth.

The apoptotic effect was studied through morphological changes found in MCF-7 cells treated with the IC<sub>50</sub> concentration of the corresponding compounds for 24 h and after staining with Hoechst 33242. While control cells treated with DMSO showed uniformly dispersed chromatin, 25-34% of the cells treated with the different compounds showed apoptotic features, including chromatin condensation and nuclear degradation. Moreover, MCF-7 cells were treated with the IC<sub>50</sub> concentration of the different compounds for 24 hours and chromosomal DNA was extracted to perform DNA fragmentation assays which may give extra information about the apoptosis-inducing effect of these products. The DNA-smeared ladder pattern observed was compatible with DNA breaks at multiple positions across the chromosomal DNA. Further contributions to the apoptotic effect of these compounds were identified, such as the 44-62% loss of mitochondrial membrane potential (DYm) and the 2-4 times increased levels of reactive oxygen species (ROS) in the mitochondria, exhibited by compound-treated cells compared to control cells.

A failure to regulate apoptosis can lead to disease, including cancer. During this process, cysteine protease enzymes, called caspases, are activated, making caspase 3 a key regulator. Using those proteins as a target to induce apoptosis in cancer cells has emerged as an attractive strategy in cancer therapy [167]. Moreover, the introduction of a trifluoromethyl group, which increases the lipophilicity of the final products, has been of collective interest during the last decades for its effect on pharmacological properties [168]. Increased absorption across cell membranes improves the selectivity, efficacy and bioavailability of the compound in the body [169]. In addition, incorporating an azo group has also been of interest following the discovery of its ability to raise the biological activity of heterocyclic compounds [170].

Given these comments, Fayed group has recently designed a new class of pyrazoles 29 and pyrazolopyrimidines 30 bearing trifluoromethyl and azo groups into their molecular structures to examine the effect of this combination on the anticancer activity of the system [171]. In particular, the authors studied three different cancer cell lines: HepG-2 (liver), HCT-116 (colon) and MCF-7 (breast). Fig. (21) shows the predicted and experimental activity expressed as IC<sub>50</sub> values obtained for the latter cancer cell line. The efficacy of the most active compounds (29, 30e and 30g) in activating caspase 3 in the MCF-7 cell line was tested to elucidate the pathway leading to cell death through these new derivatives. Thus, treatment of the cells with 29 and **30e** resulted in an almost 5-fold increase in the level of caspase 3 compared to the control, whereas in the case of 30g, this increase was only 3.6-fold, suggesting a caspase-dependent pathway to explain the activity of these compounds (Fig. 21).

The further analyses of a structure-activity relationship based on modeling studies led to conclude that a CN group at the 6-position, a (substituted)phenyl group at the 5-position and a free NH<sub>2</sub> group at the 2-position of pyrazolopyrimidine-containing compounds **30**, as well as the latter group at 3-position of the non-fused pyrazole scaffold **29**, could be a requirement for anticancer activity. Moreover, these studies confirmed the importance of fluorine building blocks in substituting the pyrazolopyrimidine scaffold.

An important aspect to consider in medicine is the concept of "drug repurposing" as an alternative to developing drugs from the ground up [172]. This approach provides the opportunity to find new drugs from pre-existing ones or molecules with proven good activities, which is a substantial time and cost saver. With this premise, Zheng, Zhu and coworkers decided to synthesize novel derivatives (**31** and **32**) of Sorafenib,

FC

	H <sub>2</sub> N <sub>3</sub> N	<b>29</b> <sub>5</sub> , NH <sub>2</sub> <sup>5</sup> , <sup>5,4</sup> H	<b>30a<sup>-</sup>h</b>	Predicted activity	Experimental activity	Caspase 3
Compd.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	IC <sub>50</sub> (	(μg/mL)	conc. (pg/mL)
29	-	-	-	16.77	7.73	564
30a	Me	Н	Me	17.03	14.2	-
30b	Me	Н	Ph	17.51	11.4	-
30c	Ph	Н	4-CIPh	20.94	23.6	-
30d	Ph	CN	NH <sub>2</sub>	16.54	23.5	-
30e	4-MePh	CN	$NH_2$	3.91	2.79	542.5
30f	4-OMePh	CN	$NH_2^-$	16.89	23.8	-
30g	Ph	CN	OH	5.02	3.09	392.7
30h	4-MePh	CN	OH	9.45	11.2	-
Taxol	-	-	-	-	4.7	-
control	-	-	-	-	-	110.1

Fig. (21). Molecular structures of 29 and 30a-h, and results of the activity against MCF-7 cells: predicted activity, experimental activity and caspase 3 enzyme assay [171].



Fig. (22). Sorafenib molecule-based substitution pattern developed [175].

bis-aryl urea with remarkable biological properties [173, 174], replacing this group with a pyrazole moiety to evaluate the influence of the change on the cytotoxicity against several cancer cell lines (A549, HepG2, MCF-7, and PC-3) and kinases (VEGFR-2/KDR, BRAF, CRAF, c-Met, EGFR and Flt-3) (Fig. 22) [175].

The results of the biological tests showed that the substitution of the urea fraction by the pyrazole moiety did benefit the antitumor activity. Moreover, the different substitutions in the ring also involved a change in the cytotoxicity of the final compound. Regarding the results obtained for the breast cancer line MCF-7, among other observations, it was concluded that the presence of Br in the 3-position of the aromatic ring

(31a) or the lack of substituent (31d) was the most effective pattern through which the best  $IC_{50}$  values were obtained (1.96 and 1.24  $\mu$ M, respectively), even lower than Sorafenib (Fig. 22). However, strongly electro-attracting groups, such as NO<sub>2</sub> in the 3-position (31b), decreased activity (Fig. 22).

To further explore the mode of binding of these compounds to vascular endothelial growth factor (VEGFR-2), molecular simulation studies were performed with compound **31a**. These analyses showed that the mode of binding of the synthesized compound **31a** produced a new H-bond with the ASN923 residues of the carbonyl group of the *N*-methylpicolinamide that was not present in the reference compound, playing an

important role in the enhancement of the inhibitory potency.

#### CONCLUSION

It has been proven that the pyrazole core and the infinite number of derivatives to which it can be linked represent a privileged structure of great pharmacological interest.

Their action as kinase inhibitors or potent antitumor agents against breast cancer lines (MCF-7 or MDA-MB-231, among others) is highly effective, reaching in some cases a level of efficacy compared to current treatments. The activity of these derivatives has not only been reflected in breast tumors but also in other cancer cell lines such as HeLa (cervix) or A549 (lung) and even as antimicrobial agents, reflecting the wide spectrum of biological activities that can be achieved.

Among the many publications collected, we highlight the preclinical *in vivo* assays with some of them, and the promising results obtained, encouraging us to continue working on these structures, providing a new pathway towards effective and less invasive therapies.

# LIST OF ABBREVIATIONS

- HDI = Human Development Index
- SD = Sprague-Dawley
- MDR = Multidrug Resistance
- ECM = Extracellular Matrix
- SRB = Sulforhodamine B
- ROS = Reactive Oxygen Species
- CDKs = Cyclin-Dependent Kinases

# **CONSENT FOR PUBLICATION**

Not applicable.

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# **CONFLICT OF INTEREST**

The authors declare no conflict of interest, financial or otherwise.

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#### REFERENCES

- Miller, K.D.; Nogueira, L.; Mariotto, A.B.; Rowland, J.H.; Yabroff, K.R.; Alfano, C.M.; Jemal, A.; Kramer, J.L.; Siegel, R.L. Cancer treatment and survivorship statistics, 2019. *CA Cancer J. Clin.*, **2019**, *69*(5), 363-385. http://dx.doi.org/10.3322/caac.21565 PMID: 31184787
- Wu, Q.; Yang, Z.; Nie, Y.; Shi, Y.; Fan, D. Multi-drug resistance in cancer chemotherapeutics: Mechanisms and lab approaches. *Cancer Lett.*, **2014**, *347*(2), 159-166. http://dx.doi.org/10.1016/j.canlet.2014.03.013 PMID: 24657660
- Brown, S.B.; Brown, E.A.; Walker, I. The present and future role of photodynamic therapy in cancer treatment. *Lancet Oncol.*, 2004, 5(8), 497-508. http://dx.doi.org/10.1016/S1470-2045(04)01529-3 PMID: 15288239
- [4] Peer, D.; Karp, J.M.; Hong, S.; Farokhzad, O.C.; Margalit, R.; Langer, R. Nanocarriers as an emerging platform for cancer therapy. *Nat. Nanotechnol.*, 2007, 2(12), 751-760. http://dx.doi.org/10.1038/nnano.2007.387 PMID: 18654426
- [5] Dasari, S.; Tchounwou, P.B. Cisplatin in cancer therapy: Molecular mechanisms of action. *Eur. J. Pharmacol.*, 2014, 740, 364-378. http://dx.doi.org/10.1016/j.ejphar.2014.07.025 PMID: 25058905
- [6] West, A.C.; Johnstone, R.W. New and emerging HDAC inhibitors for cancer treatment. J. Clin. Invest., 2014, 124(1), 30-39.
  - http://dx.doi.org/10.1172/JCI69738 PMID: 24382387 Zhao, J. Cancer stem cells and chemoresistance: The smart-
- Zhao, J. Cancer stem cells and chemoresistance: The smartest survives the raid. *Pharmacol. Ther.*, **2016**, *160*, 145-158. http://dx.doi.org/10.1016/j.pharmthera.2016.02.008 PMID: 26899500
- [8] Keibler, M.A.; Wasylenko, T.M.; Kelleher, J.K.; Iliopoulos, O.; Vander Heiden, M.G.; Stephanopoulos, G. Metabolic requirements for cancer cell proliferation. *Cancer Metab.*, 2016, 4, 16. http://dx.doi.org/10.1186/s40170-016-0156-6 PMID:

http://dx.doi.org/10.1186/s401/0-016-0156-6 PMID: 27540483

- [9] Nilsson, A.; Nielsen, J. Genome scale metabolic modeling of cancer. *Metab. Eng.*, 2017, 43(Pt B), 103-112. http://dx.doi.org/10.1016/j.ymben.2016.10.022 PMID: 27825806
- [10] Hason, M.; Bartůněk, P. Zebrafish models of cancer-new insights on modeling human cancer in a non-mammalian vertebrate. *Genes (Basel)*, **2019**, *10*(11), 935. http://dx.doi.org/10.3390/genes10110935 PMID: 31731811
- [11] Andrei, L.; Kasas, S.; Ochoa Garrido, I.; Stanković, T.; Suárez Korsnes, M.; Vaclavikova, R.; Assaraf, Y.G.; Pešić, M. Advanced technological tools to study multidrug resistance in cancer. *Drug Resist. Updat.*, **2020**, *48*, 100658. http://dx.doi.org/10.1016/j.drup.2019.100658 PMID: 31678863
- [12] Afshar, N.; English, D.R.; Thursfield, V.; Mitchell, P.L.; Te Marvelde, L.; Farrugia, H.; Giles, G.G.; Milne, R.L. Differences in cancer survival by sex: A population-based study using cancer registry data. *Cancer Causes Control*, **2018**, 29(11), 1059-1069. http://dx.doi.org/10.1007/s10552-018-1079-z PMID:

30194549

 [13] Vanneman, M.; Dranoff, G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat. Rev. Cancer*, 2012, 12(4), 237-251. http://dx.doi.org/10.1038/nrc3237 PMID: 22437869

- [14] Jeon, J.S.; Bersini, S.; Gilardi, M.; Dubini, G.; Charest, J.L.; Moretti, M.; Kamm, R.D. Human 3D vascularized organotypic microfluidic assays to study breast cancer cell extravasation. *Proc. Natl. Acad. Sci. USA*, **2015**, *112*(1), 214-219. http://dx.doi.org/10.1073/pnas.1417115112 PMID: 25524628
- [15] Kaushik, A.K.; De Berardinis, R.J. Applications of metabolomics to study cancer metabolism. *Rev. Cancer*, 2018, 1870(1), 2-14. http://dx.doi.org/10.1016/j.bbcan.2018.04.009
- [16] WHO. Who report on cancer: Setting priorities, investing wisely and providing care for all. 2020. Available from: https://www.who.int/publications/i/item/9789240001299
- [17] Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.*, **2018**, *68*(6), 394-424.

http://dx.doi.org/10.3322/caac.21492 PMID: 30207593

- [18] Ziegler, R.G.; Hoover, R.N.; Pike, M.C.; Hildesheim, A.; Nomura, A.M.; West, D.W.; Wu-Williams, A.H.; Kolonel, L.N.; Horn-Ross, P.L.; Rosenthal, J.F.; Hyer, M.B. Migration patterns and breast cancer risk in Asian-American women. J. Natl. Cancer Inst., 1993, 85(22), 1819-1827. http://dx.doi.org/10.1093/jnci/85.22.1819 PMID: 8230262
- Brinton, L.A.; Gaudet, M.M.; Gierach, G.L. Breast cancer. In: *Cancer Epidemiology and Prevention*, 4<sup>th</sup> ed; Thun, M.J.; Linet, M.S.; Cerhan, J.R.; Haiman, C.A.; Schottenfeld, D., Eds.; Oxford University Press: New York, **2018**; pp. 861-888.

http://dx.doi.org/10.1093/oso/9780190238667.003.0045

- [20] Bray, F.; McCarron, P.; Parkin, D.M. The changing global patterns of female breast cancer incidence and mortality. *Breast Cancer Res.*, 2004, 6(6), 229-239. http://dx.doi.org/10.1186/bcr932 PMID: 15535852
- [21] Rossouw, J.E.; Anderson, G.L.; Prentice, R.L.; LaCroix,
- A.Z.; Kooperberg, C.; Stefanick, M.L.; Jackson, R.D.; Beresford, S.A.A.; Howard, B.V.; Johnson, K.C.; Kotchen, J.M.; Ockene, J. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: Principal results From the Women's Health Initiative randomized controlled trial. *JAMA*, **2002**, *288*(3), 321-333.
- http://dx.doi.org/10.1001/jama.288.3.321 PMID: 12117397
  [22] Ali, I.; Lone, M.N.; Al-Othman, Z.A.; Al-Warthan, A.; Sanagi, M.M. Heterocyclic scaffolds: Centrality in anticancer drug development. *Curr. Drug Targets*, 2015, 16(7), 711-734. http://dx.doi.org/10.2174/1389450116666150309115922

PMID: 25751009 Martins, P.; Jesus, J.; Santos, S.; Raposo, L.R.; Roma-

- Martins, P.; Jesus, J.; Santos, S.; Raposo, L.R.; Roma-Rodrigues, C.; Baptista, P.V.; Fernandes, A.R. Heterocyclic anticancer compounds: Recent advances and the paradigm shift towards the use of nanomedicine's tool box. *Molecules*, 2015, 20(9), 16852-16891. http://dx.doi.org/10.3390/molecules200916852 PMID: 26389876
- [24] Lang, D.K.; Kaur, R.; Arora, R.; Saini, B.; Arora, S. Nitrogen-containing heterocycles as anticancer agents: An overview. *Anticancer. Agents Med. Chem.*, 2020, 20(18), 2150-2168.
   http://dx.doi.org/10.2174/1871520620666200705214017

http://dx.doi.org/10.2174/1871520620666200705214917 PMID: 32628593

[25] Auria-Luna, F.; Marqués-López, E.; Romanos, E.; Fernández-Moreira, V.; Gimeno, M.C.; Marzo, I.; Herrera, R.P. Novel ureido-dihydropyridine scaffolds as theranostic agents. *Bioorg. Chem.*, **2020**, *105*, 104364. http://dx.doi.org/10.1016/j.bioorg.2020.104364 PMID: 33113409

[26] Rodrigues, J.M.; Calhelha, R.C.; Nogueira, A.; Ferreira, I.C.F.R.; Barros, L.; Queiroz, M.R.P. Synthesis of novel methyl 7-[(hetero)arylamino]thieno[2,3-b]pyrazine-6carboxylates and antitumor activity evaluation: Effects in human tumor cells growth, cell cycle analysis, apoptosis and toxicity in non-tumor cells. *Molecules*, **2021**, *26*(16), 4823.

http://dx.doi.org/10.3390/molecules26164823 PMID: 34443411

- [27] Lenis-Rojas, O.A.; Cordeiro, S.; Horta-Meireles, M.; Fernández, J.A.A.; Fernández Vila, S.; Rubiolo, J.A.; Cabezas-Sainz, P.; Sanchez, L.; Fernandes, A.R.; Royo, B. N- Heterocyclic carbene iron complexes as anticancer agents: In vitro and in vivo biological studies. Molecules, 2021, 26(18), 5535. http://dx.doi.org/10.3390/molecules26185535 PMID: 34577006
- [28] Fustero, S.; Sánchez-Roselló, M.; Barrio, P.; Simón-Fuentes, A. From 2000 to mid-2010: A fruitful decade for the synthesis of pyrazoles. *Chem. Rev.*, 2011, 111(11), 6984-7034.

http://dx.doi.org/10.1021/cr2000459 PMID: 21806021

- [29] Shih, S-R.; Chu, T-Y.; Reddy, G.R.; Tseng, S-N.; Chen, H-L.; Tang, W-F.; Wu, M-S.; Yeh, J-Y.; Chao, Y-S.; Hsu, J.T.; Hsieh, H-P.; Horng, J-T. Pyrazole compound BPR1P0034 with potent and selective anti-influenza virus activity. J. Biomed. Sci., 2010, 17(13), 13. http://dx.doi.org/10.1186/1423-0127-17-13 PMID: 20178582
- [30] Hashem, A.I.; Youssef, A.S.A.; Kandeel, K.A.; Abou-Elmagd, W.S.I. Conversion of some 2(3*H*)-furanones bearing a pyrazolyl group into other heterocyclic systems with a study of their antiviral activity. *Eur. J. Med. Chem.*, 2007, 42(7), 934-939. http://dx.doi.org/10.1016/j.ejmech.2006.12.032 PMID: 17321008
- [31] Rashad, A.E.; Hegab, M.I.; Abdel-Megeid, R.E.; Micky, J.A.; Abdel-Megeid, F.M.E. Synthesis and antiviral evaluation of some new pyrazole and fused pyrazolopyrimidine derivatives. *Bioorg. Med. Chem.*, 2008, 16(15), 7102-7106. http://dx.doi.org/10.1016/j.bmc.2008.06.054 PMID: 18635363
- [32] Morsy, A.R.I.; Ramadan, S.K.; Elsafty, M.M. Synthesis and antiviral activity of some pyrrolonyl substituted heterocycles as additives to enhance inactivated Newcastle disease vaccine. *Med. Chem. Res.*, 2020, 29, 979-988. http://dx.doi.org/10.1007/s00044-020-02538-z
- [33] Chandna, N.; Kumar, S.; Kaushik, P.; Kaushik, D.; Roy, S.K.; Gupta, G.K.; Jachak, S.M.; Kapoor, J.K.; Sharma, P.K. Synthesis of novel celecoxib analogues by bioisosteric replacement of sulfonamide as potent anti-inflammatory agents and cyclooxygenase inhibitors. *Bioorg. Med. Chem.*, 2013, 21(15), 4581-4590. http://dx.doi.org/10.1016/j.bmc.2013.05.029 PMID: 23769654
- [34] Steinbach, G.; Lynch, P.M.; Phillips, R.K.; Wallace, M.H.; Hawk, E.; Gordon, G.B.; Wakabayashi, N.; Saunders, B.; Shen, Y.; Fujimura, T.; Su, L.K.; Levin, B.; Godio, L.; Patterson, S.; Rodriguez-Bigas, M.A.; Jester, S.L.; King, K.L.; Schumacher, M.; Abbruzzese, J.; DuBois, R.N.; Hittelman, W.N.; Zimmerman, S.; Sherman, J.W.; Kelloff, G. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N. Engl. J. Med.*, **2000**, *342*(26), 1946-1952.

http://dx.doi.org/10.1056/NEJM200006293422603 PMID: 10874062

- [35] Padmaja, A.; Payani, T.; Reddy, G.D.; Padmavathi, V. Synthesis, antimicrobial and antioxidant activities of substituted pyrazoles, isoxazoles, pyrimidine and thioxopyrimidine derivatives. *Eur. J. Med. Chem.*, **2009**, *44*(11), 4557-4566. http://dx.doi.org/10.1016/j.ejmech.2009.06.024 PMID: 19631423
- [36] Ramadan, S.K.; El-Helw, E.A.E. Synthesis and antimicrobial activity evaluation of some novel heterocycles derived from chromonyl-2(3*H*)-furanone. J. Chem. Res., 2018, 42(6), 332-336. http://dx.doi.org/10.3184/174751918X15295796734379
- [37] Bronson, J.; Dhar, M.; Ewing, W.; Lonberg, N. Chapter thirty-one -to market, to market-2011. *Annu. Rep. Med. Chem.*, 2012, 47, 499-569. http://dx.doi.org/10.1016/B978-0-12-396492-2.00031-X
- [38] Hsu, A-L.; Ching, T-T.; Wang, D-S.; Song, X.; Rangnekar, V.M.; Chen, C-S. The cyclooxygenase-2 inhibitor celecoxib
  - induces apoptosis by blocking Akt activation in human prostate cancer cells independently of Bcl-2. *J. Biol. Chem.*, **2000**, *275*(15), 11397-11403. http://dx.doi.org/10.1074/jbc.275.15.11397 PMID:

10753955

- [39] Williams, C.S.; Watson, A.J.; Sheng, H.; Helou, R.; Shao, J.; DuBois, R.N. Celecoxib prevents tumor growth *in vivo* without toxicity to normal gut: Lack of correlation between *in vitro* and *in vivo* models. *Cancer Res.*, **2000**, *60*(21), 6045-6051.
  - PMID: 11085526
- [40] Kulp, S.K.; Yang, Y-T.; Hung, C-C.; Chen, K-F.; Lai, J-P.; Tseng, P-H.; Fowble, J.W.; Ward, P.J.; Chen, C-S. 3phosphoinositide-dependent protein kinase-1/Akt signaling represents a major cyclooxygenase-2-independent target for celecoxib in prostate cancer cells. *Cancer Res.*, 2004, 64(4), 1444-1451. http://dx.doi.org/10.1158/0008\_5472\_CAN\_02\_2206\_PMID:

http://dx.doi.org/10.1158/0008-5472.CAN-03-2396 PMID: 14973075

- [41] Nitulescu, G.M.; Draghici, C.; Missir, A.V. Synthesis of new pyrazole derivatives and their anticancer evaluation. *Eur. J. Med. Chem.*, 2010, 45(11), 4914-4919. http://dx.doi.org/10.1016/j.ejmech.2010.07.064 PMID: 20728965
- [42] Ramadan, S.K.; El-Helw, E.A.E.; Sallam, H.A. Cytotoxic and antimicrobial activities of some novel heterocycles employing 6-(1,3-diphenyl-1*H*-pyrazol-4-yl)-4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carbonitrile. *Heterocycl. Commun.*, 2019, 25(1), 107-115. http://dx.doi.org/10.1515/hc-2019-0008
- [43] FDA. Drug approval date and data were obtained from the following sources. Available from: https://www.ema. europa.eu/.
- [44] Torre, L.A.; Islami, F.; Siegel, R.L.; Ward, E.M.; Jemal, A. Global cancer in women: Burden and trends. *Cancer Epidemiol. Biomarkers Prev.*, 2017, 26(4), 444-457. http://dx.doi.org/10.1158/1055-9965.EPI-16-0858 PMID: 28223433
- [45] Manning, G.; Whyte, D.B.; Martinez, R.; Hunter, T.; Sudarsanam, S. The protein kinase complement of the human genome. *Science*, 2002, 298(5600), 1912-1934. http://dx.doi.org/10.1126/science.1075762 PMID: 12471243
- [46] Noble, M.E.M.; Endicott, J.A.; Johnson, L.N. Protein kinase inhibitors: Insights into drug design from structure. *Science*, 2004, 303(5665), 1800-1805.

http://dx.doi.org/10.1126/science.1095920 PMID: 15031492

- [47] Cherry, M.; Williams, D.H. Recent kinase and kinase inhibitor X-ray structures: Mechanisms of inhibition and selectivity insights. *Curr. Med. Chem.*, 2004, 11(6), 663-673. http://dx.doi.org/10.2174/0929867043455792 PMID: 15032722
- [48] Furet, P.; Meyer, T.; Strauss, A.; Raccuglia, S.; Rondeau, J-M. Structure-based design and protein X-ray analysis of a protein kinase inhibitor. *Bioorg. Med. Chem. Lett.*, 2002, 12(2), 221-224. http://dx.doi.org/10.1016/S0960-894X(01)00715-6 PMID: 11755359
- [49] Ikuta, M.; Kamata, K.; Fukasawa, K.; Honma, T.; Machida, T.; Hirai, H.; Suzuki-Takahashi, I.; Hayama, T.; Nishimura, S. Crystallographic approach to identification of cyclin-dependent kinase 4 (CDK4)-specific inhibitors by using CDK4 mimic CDK2 protein. *J. Biol. Chem.*, 2001, 276(29), 27548-27554. http://dx.doi.org/10.1074/jbc.M102060200 PMID:

11335721

- [50] Sawyer, J.S.; Beight, D.W.; Britt, K.S.; Anderson, B.D.; Campbell, R.M.; Goodson, T., Jr; Herron, D.K.; Li, H-Y.; McMillen, W.T.; Mort, N.; Parsons, S.; Smith, E.C.; Wagner, J.R.; Yan, L.; Zhang, F.; Yingling, J.M. Synthesis and activity of new aryl- and heteroaryl-substituted 5,6-dihydro-4*H*-pyrrolo[1,2-*b*]pyrazole inhibitors of the transforming growth factor-β type I receptor kinase domain. *Bioorg. Med. Chem. Lett.*, **2004**, *14*(13), 3581-3584. http://dx.doi.org/10.1016/j.bmcl.2004.04.007 PMID: 15177479
- [51] Sawyer, J.S.; Anderson, B.D.; Beight, D.W.; Campbell, R.M.; Jones, M.L.; Herron, D.K.; Lampe, J.W.; McCowan, J.R.; McMillen, W.T.; Mort, N.; Parsons, S.; Smith, E.C.R.; Vieth, M.; Weir, L.C.; Yan, L.; Zhang, F.; Yingling, J.M. Synthesis and activity of new aryl- and heteroarylsubstituted pyrazole inhibitors of the transforming growth factor-β type I receptor kinase domain. J. Med. Chem., 2003, 46(19), 3953-3956.

http://dx.doi.org/10.1021/jm0205705 PMID: 12954047

- [52] El-Gamal, M.I.; Zaraei, S-O.; Madkour, M.M.; Anbar, H.S. Evaluation of substituted pyrazole-based kinase inhibitors in one decade (2011-2020): Current status and future prospects. *Molecules*, 2022, 27(1), 330. http://dx.doi.org/10.3390/molecules27010330 PMID: 35011562
- [53] Persson, T.; Yde, C.W.; Rasmussen, J.E.; Rasmussen, T.L.; Guerra, B.; Issinger, O-G.; Nielsen, J. Pyrazole carboxamides and carboxylic acids as protein kinase inhibitors in aberrant eukaryotic signal transduction: Induction of growth arrest in MCF-7 cancer cells. *Org. Biomol. Chem.*, 2007, 5(24), 3963-3970.
- http://dx.doi.org/10.1039/b711279c PMID: 18043801 [54] El-Deeb, I.M.; Lee, S.H. Design and synthesis of new po-
- tent anticancer pyrazoles with high FLT3 kinase inhibitory selectivity. *Bioorg. Med. Chem.*, **2010**, *18*(11), 3961-3973. http://dx.doi.org/10.1016/j.bmc.2010.04.029 PMID: 20472440
- [55] Shankar, D.B.; Li, J.; Tapang, P.; Owen McCall, J.; Pease, L.J.; Dai, Y.; Wei, R-Q.; Albert, D.H.; Bouska, J.J.; Osterling, D.J.; Guo, J.; Marcotte, P.A.; Johnson, E.F.; Soni, N.; Hartandi, K.; Michaelides, M.R.; Davidsen, S.K.; Priceman, S.J.; Chang, J.C.; Rhodes, K.; Shah, N.; Moore, T.B.; Sakamoto, K.M.; Glaser, K.B. ABT-869, a multitargeted receptor tyrosine kinase inhibitor: Inhibition of FLT3 phos-

phorylation and signaling in acute myeloid leukemia. *Blood*, **2007**, *109*(8), 3400-3408. http://dx.doi.org/10.1182/blood-2006-06-029579 PMID: 17209055

- [56] Schmidt-Arras, D.; Schwäble, J.; Böhmer, F-D.; Serve, H. Flt3 receptor tyrosine kinase as a drug target in leukemia. *Curr. Pharm. Des.*, 2004, 10(16), 1867-1883. http://dx.doi.org/10.2174/1381612043384394 PMID: 15180525
- [57] Gazit, A.; Yee, K.; Uecker, A.; Böhmer, F-D.; Sjöblom, T.; Östman, A.; Waltenberger, J.; Golomb, G.; Banai, S.; Heinrich, M.C.; Levitzki, A. Tricyclic quinoxalines as potent kinase inhibitors of PDGFR kinase, Flt3 and Kit. *Bioorg. Med. Chem.*, 2003, 11(9), 2007-2018. http://dx.doi.org/10.1016/S0968-0896(03)00048-8 PMID: 12670652
- [58] Mahboobi, S.; Uecker, A.; Cénac, C.; Sellmer, A.; Eichhorn, E.; Elz, S.; Böhmer, F-D.; Dove, S. Inhibition of FLT3 and PDGFR tyrosine kinase activity by bis(benzo[b]furan-2-yl)methanones. *Bioorg. Med. Chem.*, 2007, 15(5), 2187-2197. http://dx.doi.org/10.1016/j.bmc.2006.12.011 PMID: 17210255
- [59] Woodburn, J.R. The epidermal growth factor receptor and its inhibition in cancer therapy. *Pharmacol. Ther.*, **1999**, 82(2-3), 241-250. http://dx.doi.org/10.1016/S0163-7258(98)00045-X PMID: 10454201
- [60] Wells, A. Tumor invasion: Role of growth factor-induced cell motility. *Adv. Cancer Res.*, 2000, 78, 31-101. http://dx.doi.org/10.1016/S0065-230X(08)61023-4 PMID: 10547668
- [61] Bridges, A.J. The rationale and strategy used to develop a series of highly potent, irreversible, inhibitors of the epidermal growth factor receptor family of tyrosine kinases. *Curr. Med. Chem.*, **1999**, *6*(9), 825-843. http://dx.doi.org/10.2174/092986730609220401151141
   PMID: 10495354
- [62] Boschelli, D.H. Small molecule inhibitors of receptor tyrosine kinases. *Drugs Future*, **1999**, 24(5), 515-537. http://dx.doi.org/10.1358/dof.1999.024.05.858622
- [63] Liu, Y.; Gray, N.S. Rational design of inhibitors that bind to inactive kinase conformations. *Nat. Chem. Biol.*, 2006, 2(7), 358-364.

http://dx.doi.org/10.1038/nchembio799 PMID: 16783341

- [64] Prossnitz, E.R.; Arterburn, J.B.; Smith, H.O.; Oprea, T.I.; Sklar, L.A.; Hathaway, H.J. Estrogen signaling through the transmembrane G protein-coupled receptor GPR30. *Annu. Rev. Physiol.*, 2008, 70, 165-190. http://dx.doi.org/10.1146/annurev.physiol.70.113006.10051 8 PMID: 18271749
- [65] Sawai, A.; Chandarlapaty, S.; Greulich, H.; Gonen, M.; Ye, Q.; Arteaga, C.L.; Sellers, W.; Rosen, N.; Solit, D.B. Inhibition of Hsp90 down-regulates mutant epidermal growth factor receptor (EGFR) expression and sensitizes EGFR mutant tumors to paclitaxel. *Cancer Res.*, **2008**, 68(2), 589-596. http://dx.doi.org/10.1158/0008.5472.CAN.07.1570.DMID:

http://dx.doi.org/10.1158/0008-5472.CAN-07-1570 PMID: 18199556

[66] Wei, F.; Zhao, B-X.; Huang, B.; Zhang, L.; Sun, C-H.; Dong, W-L.; Shin, D-S.; Miao, J-Y. Design, synthesis, and preliminary biological evaluation of novel ethyl 1-(2'hydroxy-3'-aroxypropyl)-3-aryl-1*H*-pyrazole-5-carboxylate. *Bioorg. Med. Chem. Lett.*, **2006**, *16*(24), 6342-6347. http://dx.doi.org/10.1016/j.bmcl.2006.09.008 PMID: 17000107

- [67] Das, J.; Pany, S.; Panchal, S.; Majhi, A.; Rahman, G.M. Binding of isoxazole and pyrazole derivatives of curcumin with the activator binding domain of novel protein kinase C. *Bioorg. Med. Chem.*, 2011, 19(21), 6196-6202. http://dx.doi.org/10.1016/j.bmc.2011.09.011 PMID: 21975067
- [68] Xie, Y-S.; Pan, X-H.; Zhao, B-X.; Liu, J-T.; Shin, D-S.; Zhang, J-H.; Zheng, L-W.; Zhao, J.; Miao, J-Y. Synthesis, structure characterization and preliminary biological evaluation of novel 5-alkyl-2-ferrocenyl-6,7-dihydropyrazolo [1,5-a]pyrazin-4(5H)-one derivatives. J. Organomet. Chem., 2008, 693(7), 1367-1374. http://dx.doi.org/10.1016/j.jorganchem.2008.01.043

[69] Insuasty, B.; Tigreros, A.; Orozco, F.; Quiroga, J.; Abonía, R.; Nogueras, M.; Sanchez, A.; Cobo, J. Synthesis of novel pyrazolic analogues of chalcones and their 3-aryl-4-(3-aryl-4,5-dihydro-1*H*-pyrazol-5-yl)-1-phenyl-1*H*-pyrazole derivatives as potential antitumor agents. *Bioorg. Med. Chem.*, 2010, 18(14), 4965-4974. http://dx.doi.org/10.1016/j.bmc.2010.06.013 PMID:

20594863

- [70] Li, D-D.; Lv, P-C.; Zhang, H.; Zhang, H-J.; Hou, Y-P.; Liu, K.; Ye, Y-H.; Zhu, H-L. The combination of 4-anilinoquinazoline and cinnamic acid: A novel mode of binding to the epidermal growth factor receptor tyrosine kinase. *Bioorg. Med. Chem.*, 2011, 19(16), 5012-5022. http://dx.doi.org/10.1016/j.bmc.2011.06.044 PMID: 21763148
- [71] Cárdenas, M.; Marder, M.; Blank, V.C.; Roguin, L.P. Antitumor activity of some natural flavonoids and synthetic derivatives on various human and murine cancer cell lines. *Bioorg. Med. Chem.*, 2006, 14(9), 2966-2971. http://dx.doi.org/10.1016/j.bmc.2005.12.021 PMID: 16412650
- [72] Dallavalle, S.; Cincinelli, R.; Nannei, R.; Merlini, L.; Morini, G.; Penco, S.; Pisano, C.; Vesci, L.; Barbarino, M.; Zuco, V.; De Cesare, M.; Zunino, F. Design, synthesis, and evaluation of biphenyl-4-yl-acrylohydroxamic acid derivatives as histone deacetylase (HDAC) inhibitors. *Eur. J. Med. Chem.*, 2009, 44(5), 1900-1912. http://dx.doi.org/10.1016/j.ejmech.2008.11.005 PMID: 19084294
- [73] Qian, Y.; Zhang, H-J.; Zhang, H.; Xu, C.; Zhao, J.; Zhu, H-L. Synthesis, molecular modeling, and biological evaluation of cinnamic acid metronidazole ester derivatives as novel anticancer agents. *Bioorg. Med. Chem.*, **2010**, *18*(14), 4991-4996.

http://dx.doi.org/10.1016/j.bmc.2010.06.003 PMID: 20594859

- [74] Zhang, W-M.; Xing, M.; Zhao, T-T.; Ren, Y-J.; Yang, X-H.; Yang, Y-S.; Lv, P-C.; Zhu, H-L. Synthesis, molecular modeling and biological evaluation of cinnamic acid derivatives with pyrazole moieties as novel anticancer agents. *RSC Advances*, **2014**, *4*(70), 37197-37207. http://dx.doi.org/10.1039/C4RA05257A
- [75] Ashourpour, M.; Mostafavi Hosseini, F.; Amini, M.; Saeedian Moghadam, E.; Kazerouni, F.; Arman, S.Y.; Shahsavari, Z. Pyrazole derivatives induce apoptosis *via* ROS generation in the triple negative breast cancer cells, MDA-MB-468. *Asian Pac. J. Cancer Prev.*, **2021**, *22*(7), 2079-2087.

http://dx.doi.org/10.31557/APJCP.2021.22.7.2079 PMID: 34319030

[76] Tao, Z.; Shi, A.; Lu, C.; Song, T.; Zhang, Z.; Zhao, J. Breast cancer: Epidemiology and etiology. *Cell Biochem. Biophys.*, 2015, 72(2), 333-338. http://dx.doi.org/10.1007/s12013-014-0459-6 PMID: 25543329

- [77] Hong, Y.; Cho, M.; Yuan, Y-C.; Chen, S. Molecular basis for the interaction of four different classes of substrates and inhibitors with human aromatase. *Biochem. Pharmacol.*, 2008, 75(5), 1161-1169. http://dx.doi.org/10.1016/j.bcp.2007.11.010 PMID: 18184606
- [78] Leonard, G.D.; Swain, S.M. Ductal carcinoma *in situ*, complexities and challenges. J. Natl. Cancer Inst., 2004, 96(12), 906-920. http://dx.doi.org/10.1093/inci/djh164 PMID: 15199110
- [79] Gusberg, S.B. Tamoxifen for breast cancer: Associated endometrial cancer. *Cancer*, **1990**, *65*(7), 1463-1464. http://dx.doi.org/10.1002/1097-0142(19900401)65:7<1463::AID-CNCR2820650702>3.0.CO;2-S PMID: 2311060
- [80] Neven, P.; Vergote, I. Should tamoxifen users be screened for endometrial lesions? *Lancet*, **1998**, *351*(9097), 155-157. http://dx.doi.org/10.1016/S0140-6736(05)78216-7 PMID: 9449866
- [81] Wiseman, L.R.; Goa, K.L. Toremifene. A review of its pharmacological properties and clinical efficacy in the management of advanced breast cancer. *Drugs*, 1997, 54(1), 141-160. http://dx.doi.org/10.2165/00003495-199754010-00014
   PMID: 9211086
- [82] Diasio, R.B. The role of dihydropyrimidine dehydrogenase (DPD) modulation in 5-FU pharmacology. Oncology (Williston Park), 1998, 12(10)(Suppl. 7), 23-27.
   PMID: 9830621
- [83] Naruse, T.; Nishida, Y.; Ishiguro, N. Synergistic effects of meloxicam and conventional cytotoxic drugs in human MG-63 osteosarcoma cells. *Biomed. Pharmacother.*, 2007, *61*(6), 338-346. http://dx.doi.org/10.1016/j.biopha.2007.02.011 PMID: 17395421
- [84] Dang, C.T.; Dannenberg, A.J.; Subbaramaiah, K.; Dickler, M.N.; Moasser, M.M.; Seidman, A.D.; D'Andrea, G.M.; Theodoulou, M.; Panageas, K.S.; Norton, L.; Hudis, C.A.; Phase, I.I. Phase II study of celecoxib and trastuzumab in metastatic breast cancer patients who have progressed after prior trastuzumab-based treatments. *Clin. Cancer Res.*, **2004**, *10*(12 Pt 1), 4062-4067. http://dx.doi.org/10.1158/1078-0432.CCR-03-0463 PMID: 15217939
- [85] Reardon, D.A.; Quinn, J.A.; Vredenburgh, J.; Rich, J.N.; Gururangan, S.; Badruddoja, M.; Herndon, J.E., II; Dowell, J.M.; Friedman, A.H.; Friedman, H.S. Phase II trial of irinotecan plus celecoxib in adults with recurrent malignant glioma. *Cancer*, 2005, 103(2), 329-338. http://dx.doi.org/10.1002/cncr.20776 PMID: 15558802
- [86] Csiki, I.; Morrow, J.D.; Sandler, A.; Shyr, Y.; Oates, J.; Williams, M.K.; Dang, T.; Carbone, D.P.; Johnson, D.H. Targeting cyclooxygenase-2 in recurrent non-small cell lung cancer: A phase II trial of celecoxib and docetaxel. *Clin. Cancer Res.*, 2005, 11(18), 6634-6640. http://dx.doi.org/10.1158/1078-0432.CCR-05-0436 PMID: 16166442
- [87] Farag, A.M.; Mayhoub, A.S.; Eldebss, T.M.A.; Amr, A-G.E.; Ali, K.A.K.; Abdel-Hafez, N.A.; Abdulla, M.M. Synthesis and structure-activity relationship studies of pyrazole-based heterocycles as antitumor agents. *Arch. Pharm. (Weinheim)*, **2010**, *343*(7), 384-396. http://dx.doi.org/10.1002/ardp.200900176 PMID: 20397210

- [88] Weil, C.S. Tables for convenient calculation of medianeffective dose (LD<sub>50</sub> or ED<sub>50</sub>) and instructions in their use. *Biometrics*, **1952**, 8(3), 249-263. http://dx.doi.org/10.2307/3001557
- [89] Giuliano, M.; Hu, H.; Wang, Y-C.; Fu, X.; Nardone, A.; Herrera, S.; Mao, S.; Contreras, A.; Gutiérrez, C.; Wang, T.; Hilsenbeck, S.G.; De Angelis, C.; Wang, N.J.; Heiser, L.M.; Gray, J.W.; López-Tarruella, S.; Pavlick, A.C.; Trivedi, M.V.; Chamness, G.C.; Chang, J.C.; Osborne, C.K.; Rimawi, M.F.; Schiff, R. Upregulation of ER signaling as an adaptive mechanism of cell survival in HER2positive breast tumors treated with anti-HER2 therapy. *Clin. Cancer Res.*, 2015, 21(17), 3995-4003. http://dx.doi.org/10.1158/1078-0432.CCR-14-2728 PMID: 26015514
- [90] Dunnwald, L.K.; Rossing, M.A.; Li, C.I. Hormone receptor status, tumor characteristics, and prognosis: A prospective cohort of breast cancer patients. *Breast Cancer Res.*, 2007, 9(1), R6.
  - http://dx.doi.org/10.1186/bcr1639 PMID: 17239243
- [91] Ali, S.; Coombes, R.C. Estrogen receptor alpha in human breast cancer: Occurrence and significance. J. Mammary Gland Biol. Neoplasia, 2000, 5(3), 271-281. http://dx.doi.org/10.1023/A:1009594727358 PMID: 14973389
- [92] Raju, H.; Chandrappa, S.; Prasanna, D.S.; Ananda, H.; Nagamani, T.S.; Byregowda, S.M.; Rangappa, K.S. Synthesis, characterization and *in-vitro* antiproliferative effects of novel 5-amino pyrazole derivatives against breast cancer cell lines. *Anti-Cancer Drug Discov.*, **2011**, *6*(2), 186-195. http://dx.doi.org/10.2174/157489211795328459 PMID: 21247401
- [93] Ananda, H.; Sharath Kumar, K.S.; Nishana, M.; Hegde, M.; Srivastava, M.; Byregowda, R.; Choudhary, B.; Raghavan, S.C.; Rangappa, K.S. Regioselective synthesis and biological studies of novel 1-aryl-3, 5-bis (het) aryl pyrazole derivatives as potential antiproliferative agents. *Mol. Cell. Biochem.*, 2017, 426(1-2), 149-160. http://dx.doi.org/10.1007/s11010-016-2887-7 PMID: 27882441
- [94] Ananda, H.; Sharath Kumar, K.S.; Sudhanva, M.S.; Rangappa, S.; Rangappa, K.S. A trisubstituted pyrazole derivative reduces DMBA-induced mammary tumor growth in rats by inhibiting estrogen receptor-α expression. *Mol. Cell. Biochem.*, 2018, 449(1-2), 137-144. http://dx.doi.org/10.1007/s11010-018-3350-8 PMID: 29777335
- [95] Vajdos, F.F.; Hoth, L.R.; Geoghegan, K.F.; Simons, S.P.; LeMotte, P.K.; Danley, D.E.; Ammirati, M.J.; Pandit, J. The 2.0 A crystal structure of the ERalpha ligand-binding domain complexed with lasofoxifene. *Protein Sci.*, 2007, 16(5), 897-905.

http://dx.doi.org/10.1110/ps.062729207 PMID: 17456742

- [96] Folkman, J. Tumor angiogenesis: Therapeutic implications. *N. Eng. J. Med.*, **1971**, 285(21), 1182-1186. http://dx.doi.org/10.1056/NEJM197111182852108 PMID: 4938153
- [97] Liekens, S.; De Clercq, E.; Neyts, J. Angiogenesis: Regulators and clinical applications. *Biochem. Pharmacol.*, 2001, 61(3), 253-270.
   http://dx.doi.org/10.1016/S0006-2952(00)00529-3 PMID: 11172729
- [98] Carmeliet, P. Angiogenesis in life, disease and medicine. *Nature*, 2005, 438(7070), 932-936. http://dx.doi.org/10.1038/nature04478 PMID: 16355210

- [99] Folkman, J. Angiogenesis: An organizing principle for drug discovery? *Nat. Rev. Drug Discov.*, 2007, 6(4), 273-286. http://dx.doi.org/10.1038/nrd2115 PMID: 17396134
- [100] Folkman, J. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat. Med.*, **1995**, *1*(1), 27-31. http://dx.doi.org/10.1038/nm0195-27 PMID: 7584949
- [101] Wu, Y.; Sun, W.L.; Feng, J.F. Antiangiogenic therapy in the management of breast cancer. *Asia Pac. J. Clin. Oncol.*, **2013**, *9*(2), 110-116. http://dx.doi.org/10.1111/j.1743-7563.2012.01569.x PMID: 22898270
- [102] Kerbel, R.S. Clinical trials of antiangiogenic drugs: Opportunities, problems, and assessment of initial results. *J. Clin. Oncol.*, 2001, 19(18)(Suppl.), 45S-51S.
   PMID: 11560971
- [103] Leahy, K.M.; Koki, A.T.; Masferrer, J.L. Role of cyclooxygenases in angiogenesis. *Curr. Med. Chem.*, 2000, 7(11), 1163-1170. http://dx.doi.org/10.2174/0929867003374336 PMID: 11032965
- [104] Connolly, E.M.; Harmey, J.H.; O'Grady, T.; Foley, D.; Roche-Nagle, G.; Kay, E.; Bouchier-Hayes, D.J. Cyclooxygenase inhibition reduces tumour growth and metastasis in an orthotopic model of breast cancer. *Br. J. Cancer*, 2002, 87(2), 231-237. http://dx.doi.org/10.1038/sj.bjc.6600462 PMID: 12107848
- [105] Blanke, C.D. Celecoxib with chemotherapy in colorectal cancer. *Oncology (Williston Park)*, 2002, 16(4)(Suppl. 3), 17-21.
  - PMID: 12014863
- [106] Abadi, A.H.; Eissa, A.A.H.; Hassan, G.S. Synthesis of novel 1,3,4-trisubstituted pyrazole derivatives and their evaluation as antitumor and antiangiogenic agents. *Chem. Pharm. Bull. (Tokyo)*, **2003**, *51*(7), 838-844. http://dx.doi.org/10.1248/cpb.51.838 PMID: 12843591
- [107] Boyd, M.R.; Paull, K.D. Some practical considerations and applications of the national cancer institute *in vitro* anticancer drug discovery screen. *Drug Dev. Res.*, **1995**, *34*(2), 91-109.
  - http://dx.doi.org/10.1002/ddr.430340203
- [108] Monks, A.; Scudiero, D.; Skehan, P.; Shoemaker, R.; Paull, K.; Vistica, D.; Hose, C.; Langley, J.; Cronise, P.; Vaigro-Wolff, A.; Gray-Goodrich, M.; Campbell, H.; Mayo, J.; Boyd, M. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. *J. Natl. Cancer Inst.*, **1991**, *83*(11), 757-766. http://dx.doi.org/10.1093/jnci/83.11.757 PMID: 2041050
- [109] Grever, M.R.; Schepartz, S.A.; Chabner, B.A. The National Cancer Institute: Cancer drug discovery and development program. *Semin. Oncol.*, **1992**, *19*(6), 622-638. PMID: 1462164
- [110] Christodoulou, M.S.; Liekens, S.; Kasiotis, K.M.; Haroutounian, S.A. Novel pyrazole derivatives: Synthesis and evaluation of anti-angiogenic activity. *Bioorg. Med. Chem.*, **2010**, *18*(12), 4338-4350. http://dx.doi.org/10.1016/j.bmc.2010.04.076 PMID: 20493716
- [111] Elmegeed, G.A.; Khalil, W.K.B.; Mohareb, R.M.; Ahmed, H.H.; Abd-Elhalim, M.M.; Elsayed, G.H. Cytotoxicity and gene expression profiles of novel synthesized steroid derivatives as chemotherapeutic anti-breast cancer agents. *Bioorg. Med. Chem.*, **2011**, *19*(22), 6860-6872. http://dx.doi.org/10.1016/j.bmc.2011.09.033 PMID: 22000946
- [112] El-Far, M.; Elmegeed, G.A.; Eskander, E.F.; Rady, H.M.; Tantawy, M.A. Novel modified steroid derivatives of an-

drostanolone as chemotherapeutic anti-cancer agents. *Eur. J. Med. Chem.*, **2009**, *44*(10), 3936-3946. http://dx.doi.org/10.1016/j.ejmech.2009.04.020 PMID: 19447526

- [113] Chiang, K-C.; Yeh, C-N.; Chen, H-Y.; Lee, J.M.; Juang, H-H.; Chen, M-F.; Takano, M.; Kittaka, A.; Chen, T.C. 19-Nor-2α-(3-hydroxypropyl)-1α,25-dihydroxyvitamin D<sub>3</sub> (MART-10) is a potent cell growth regulator with enhanced chemotherapeutic potency in liver cancer cells. *Steroids*, 2011, 76(13), 1513-1519. http://dx.doi.org/10.1016/j.steroids.2011.08.006 PMID: 21888924
- Troisi, L.; Florio, S.; Granito, C. Chemoselective construction of novel steroid derivatives. *Steroids*, 2002, 67(8), 687-693. http://dx.doi.org/10.1016/S0039-128X(02)00032-6 PMID:

http://dx.doi.org/10.1016/S0039-128X(02)00032-6 PMID: 12117615

[115] Mohareb, R.M.; Elmegeed, G.A.; Abdel-Salam, O.M.E.; Doss, S.H.; William, M.G. Synthesis of modified steroids as a novel class of non-ulcerogenic, anti-inflammatory and anti-nociceptive agents. *Steroids*, **2011**, *76*(10-11), 1190-1203. http://dx.doi.org/10.1016/j.steroids.2011.05.011.PMID;

http://dx.doi.org/10.1016/j.steroids.2011.05.011 PMID: 21664368

- Banday, A.H.; Mir, B.P.; Lone, I.H.; Suri, K.A.; Kumar, H.M. Studies on novel D-ring substituted steroidal pyrazolines as potential anticancer agents. *Steroids*, 2010, 75(12), 805-809. http://dx.doi.org/10.1016/j.steroids.2010.02.014 PMID: 20206644
- [117] Shinkawa, T.; Nakajima, H.; Nishijima, K.; Yamasaki, F.; Kato, K.; Ohzawa, N.; Mizota, M. A novel quinolinone diuretic, M12285, and its activation mechanism through sulfate conjugation. *Eur. J. Pharmacol.*, **1992**, *219*(2), 217-224. http://dx.doi.org/10.1016/0014-2999(92)90299-J PMID: 1330606
- [118] Maurice, T.; Urani, A.; Phan, V-L.; Romieu, P. The interaction between neuroactive steroids and the σ<sub>1</sub> receptor function: Behavioral consequences and therapeutic opportunities. *Brain Res. Brain Res. Rev.*, **2001**, *37*(1-3), 116-132. http://dx.doi.org/10.1016/S0165-0173(01)00112-6 PMID: 11744080
- [119] Vajda, F.J.E. Neuroprotection and neurodegenerative disease. J. Clin. Neurosci., 2002, 9(1), 4-8. http://dx.doi.org/10.1054/jocn.2001.1027 PMID: 11749009
- [120] Mohareb, R.M.; Al-Omran, F. Reaction of pregnenolone with cyanoacetylhydrazine: Novel synthesis of hydrazide-hydrazone, pyrazole, pyridine, thiazole, thiophene derivatives and their cytotoxicity evaluations. *Steroids*, 2012, 77(14), 1551-1559. http://dx.doi.org/10.1016/j.steroids.2012.09.007 PMID: 23064008
- [121] Mohareb, R.M.; Wardakhan, W.W.; Elmegeed, G.A.; Ashour, R.M.S. Heterocyclizations of pregnenolone: Novel synthesis of thiosemicarbazone, thiophene, thiazole, thieno[2,3-b]pyridine derivatives and their cytotoxicity evaluations. *Steroids*, **2012**, 77(14), 1560-1569. http://dx.doi.org/10.1016/j.steroids.2012.09.004 PMID: 23064007
- [122] Stetler-Stevenson, W.G.; Aznavoorian, S.; Liotta, L.A. Tumor cell interactions with the extracellular matrix during invasion and metastasis. *Annu. Rev. Cell Biol.*, **1993**, *9*, 541-573. http://dx.doi.org/10.1146/annurev.cb.09.110193.002545
   PMID: 8280471

- [123] Van Aken, E.; De Wever, O.; Correia da Rocha, A.S.; Mareel, M. Defective E-cadherin/catenin complexes in human cancer. *Virchows Arch.*, 2001, 439(6), 725-751. http://dx.doi.org/10.1007/s004280100516 PMID: 11787845
- [124] Parmar, V.S.; Sharma, N.K.; Husain, M.; Watterson, A.C.; Kumar, J.; Samuelson, L.A.; Cholli, A.L.; Prasad, A.K.; Kumar, A.; Malhotra, S.; Kumar, N.; Jha, A.; Singh, A.; Singh, I.; Himanshu, ; Vats, A.; Shakil, N.A.; Trikha, S.; Mukherjee, S.; Sharma, S.K.; Singh, S.K.; Kumar, A.; Jha, H.N.; Olsen, C.E.; Stove, C.P.; Bracke, M.E.; Mareel, M.M. Synthesis, characterization and *in vitro* anti-invasive activity screening of polyphenolic and heterocyclic compounds. *Bioorg. Med. Chem.*, **2003**, *11*(6), 913-929. http://dx.doi.org/10.1016/S0968-0896(02)00539-4 PMID: 12614877
- [125] Mareel, M.M.; De Mets, M. Effect of microtubule inhibitors on invasion and on related activities of tumor cells. *Int. Rev. Cytol.*, **1984**, *90*, 125-168. http://dx.doi.org/10.1016/S0074-7696(08)61489-8 PMID: 6389412
- [126] Takeichi, M. Morphogenetic roles of classic cadherins. *Curr. Opin. Cell Biol.*, **1995**, 7(5), 619-627. http://dx.doi.org/10.1016/0955-0674(95)80102-2 PMID: 8573335
- [127] Birchmeier, W.; Behrens, J. Cadherin expression in carcinomas: Role in the formation of cell junctions and the prevention of invasiveness. *Biochim. Biophys. Acta*, 1994, 1198(1), 11-26. http://dx.doi.org/10.1016/0304-419X(94)90003-5 PMID: 8199193
- [128] Rostom, S.A.F.; Shalaby, M.A.; El-Demellawy, M.A. Polysubstituted pyrazoles, part 5. Synthesis of new 1-(4chlorophenyl)-4-hydroxy-1*H*-pyrazole-3-carboxylic acid hydrazide analogs and some derived ring systems. A novel class of potential antitumor and anti-HCV agents. *Eur. J. Med. Chem.*, 2003, 38(11-12), 959-974. http://dx.doi.org/10.1016/j.ejmech.2003.08.003 PMID: 14642328
- Park, H-J.; Lee, K.; Park, S-J.; Ahn, B.; Lee, J-C.; Cho, H.; Lee, K-I. Identification of antitumor activity of pyrazole oxime ethers. *Bioorg. Med. Chem. Lett.*, 2005, 15(13), 3307-3312. http://dx.doi.org/10.1016/j.bmcl.2005.03.082 PMID: 15922597
- [130] Rubinstein, L.V.; Shoemaker, R.H.; Paull, K.D.; Simon, R.M.; Tosini, S.; Skehan, P.; Scudiero, D.A.; Monks, A.; Boyd, M.R. Comparison of *in vitro* anticancer-drugscreening data generated with a tetrazolium assay *versus* a protein assay against a diverse panel of human tumor cell lines. *J. Natl. Cancer Inst.*, **1990**, *82*(13), 1113-1118. http://dx.doi.org/10.1093/jnci/82.13.1113 PMID: 2359137
- [131] Krapcho, A.P.; Menta, E.; Oliva, A.; Di Domenico, R.;
  Fiocchi, L.; Maresch, M.E.; Gallagher, C.E.; Hacker, M.P.;
  Beggiolin, G.; Giuliani, F.C.; Pezzoni, G.; Spinelli, S. Synthesis and antitumor evaluation of 2,5-disubstitutedindazolo[4, 3-gh]isoquinolin-6(2H)-ones (9-aza-anthrapyrazoles). J. Med. Chem., 1998, 41(27), 5429-5444. http://dx.doi.org/10.1021/jm9804432 PMID: 9876113
- [132] Bontemps-Gracz, M.M.; Kupiec, A.; Antonini, I.; Borowski, E. The ability to overcome multidrug resistance of tumor cell lines by novel acridine cytostatics with condensed heterocyclic rings. *Acta Biochim. Pol.*, 2002, 49(1), 87-92. http://dx.doi.org/10.18388/abp.2002\_3824 PMID: 12136960

- [133] Stefańska, B.; Bontemps-Gracz, M.M.; Antonini, I.; Martelli, S.; Arciemiuk, M.; Piwkowska, A.; Rogacka, D.; Borowski, E. 2,7-Dihydro-3*H*-pyridazino[5,4,3-*kl*]acridin-3-one derivatives, novel type of cytotoxic agents active on multidrug-resistant cell lines. Synthesis and biological evaluation. *Bioorg. Med. Chem.*, **2005**, *13*(6), 1969-1975. http://dx.doi.org/10.1016/j.bmc.2005.01.023 PMID: 15727851
- [134] Ghirtis, K.; Pouli, N.; Marakos, P.; Skaltsounis, A-L.; Leonce, S.; Gaignard, D.H.; Atassi, G. Synthesis and conformational analysis of some new Pyrano[2,3-c]xanthen-7-one and Pyrano[3,2-b]xanthen-6-one derivatives with cytotoxic activity. *Heterocycles*, 2000, 53(1), 93-106. http://dx.doi.org/10.3987/COM-99-8727
- [135] Ghirtis, K.; Pouli, N.; Marakos, P.; Skaltsounis, A-L.; Leonce, S.; Atassi, G.; Caignard, D.H. Design and synthesis of some new pyranoxanthenones with cytotoxic activity. *J. Heterocycl. Chem.*, 2001, 38(1), 147-152. http://dx.doi.org/10.1002/jhet.5570380121
- [136] Kostakis, I.K.; Pouli, N.; Marakos, P.; Mikros, E.; Skaltsounis, A-L.; Leonce, S.; Atassi, G.; Renard, P. Synthesis, cytotoxic activity, NMR study and stereochemical effects of some new pyrano[3,2-b]thioxanthen-6-ones and pyrano[2,3-c]thioxanthen-7-ones. *Bioorg. Med. Chem.*, 2001, 9(11), 2793-2802. http://dx.doi.org/10.1016/S0968-0896(01)00130-4 PMID: 11597459
- [137] Svoboda, G.H.; Poore, G.A.; Simpson, P.J.; Boder, G.B. Alkaloids of *Acronychia baueri* schott I. isolation of the alkaloids and a study of the antitumor and other biological properties of acronycine. *J. Pharm. Sci.*, **1966**, *55*(8), 758-768.

http://dx.doi.org/10.1002/jps.2600550803 PMID: 5975286

- [138] Michel, S.; Gaslonde, T.; Tillequin, F. Benzo[b]acronycine derivatives: A novel class of antitumor agents. *Eur. J. Med. Chem.*, 2004, 39(8), 649-655. http://dx.doi.org/10.1016/j.ejmech.2004.05.001 PMID: 15276298
- [139] Elomri, A.; Mitaku, S.; Michel, S.; Skaltsounis, A-L.; Tillequin, F.; Koch, M.; Pierré, A.; Guilbaud, N.; Léonce, S.; Kraus-Berthier, L.; Rolland, Y.; Atassi, G. Synthesis and cytotoxic and antitumor activity of esters in the 1,2dihydroxy-1,2-dihydroacronycine series. J. Med. Chem., 1996, 39(24), 4762-4766.

http://dx.doi.org/10.1021/jm9602975 PMID: 8941390

- [140] Kostakis, I.; Ghirtis, K.; Pouli, N.; Marakos, P.; Skaltsounis, A-L.; Leonce, S.; Caignard, D.H.; Atassi, G. Synthesis and cytotoxic activity of 2dialkylaminoethylamino substituted xanthenone and thioxanthenone derivatives. *Farmaco*, 2000, 55(6-7), 455-460. http://dx.doi.org/10.1016/S0014-827X(00)00068-9 PMID: 11204746
- [141] Kostakis, I.K.; Magiatis, P.; Pouli, N.; Marakos, P.; Skaltsounis, A.L.; Pratsinis, H.; Léonce, S.; Pierré, A. Design, synthesis, and antiproliferative activity of some new pyrazole-fused amino derivatives of the pyranoxanthenone, pyranothioxanthenone, and pyranoacridone ring systems: A new class of cytotoxic agents. J. Med. Chem., 2002, 45(12), 2599-2609.

http://dx.doi.org/10.1021/jm011117g PMID: 12036369

[142] Giannouli, V.; Kostakis, I.K.; Pouli, N.; Marakos, P.; Kousidou, O.Ch.; Tzanakakis, G.N.; Karamanos, N.K. Design, synthesis, and evaluation of the antiproliferative activity of a series of novel fused xanthenone aminoderivatives in human breast cancer cells. *J. Med. Chem.*, **2007**, *50*(7), 1716-1719. 20638287

http://dx.doi.org/10.1021/jm061410m PMID: 17335189

[143] Bandgar, B.P.; Totre, J.V.; Gawande, S.S.; Khobragade, C.N.; Warangkar, S.C.; Kadam, P.D. Synthesis of novel 3,5-diaryl pyrazole derivatives using combinatorial chemistry as inhibitors of tyrosinase as well as potent anticancer, anti-inflammatory agents. *Bioorg. Med. Chem.*, 2010, 18(16), 6149-6155. http://dx.doi.org/10.1016/j.bmc.2010.06.046 PMID:

 [144] Dengler, W.A.; Schulte, J.; Berger, D.P.; Mertelsmann, R.; Fiebig, H.H. Development of a Propidium Iodide fluorescence assay for proliferation and cytotoxicity assays. *Anticancer Drugs*, **1995**, 6(4), 522-532. http://dx.doi.org/10.1097/00001813-199508000-00005
 PMID: 7579556

- [145] Hu, P.; Zhao, K-Q.; Xu, H-B. (4-Hydroxybenzylidene)-4ferrocenylaniline. *Molecules*, 2001, 6(12), M251. http://dx.doi.org/10.3390/M251
- [146] Togni, A.; Halterman, R.L., Eds.; *Metallocenes*; Wiley-VCH Verlag GmbH: Weinheim, Germany, 1998. http://dx.doi.org/10.1002/9783527619542
- [147] Drent, E. Ferrocene: Homogenous catalysis, organic synthesis, material. VCH: Weinheim, 1995.
- [148] Stepnicka, P., Ed.; Ferrocenes: Ligands, Material and Biomolecules; John Wiley and Sons: NJ: USA, 2008.
- Bruijnincx, P.C.A.; Sadler, P.J. New trends for metal complexes with anticancer activity. *Curr. Opin. Chem. Biol.*, 2008, 12(2), 197-206. http://dx.doi.org/10.1016/j.cbpa.2007.11.013 PMID: 18155674
- [150] Yu, H.; Shao, L.; Fang, J. Synthesis and biological activity research of novel ferrocenyl-containing thiazole imine derivatives. J. Organomet. Chem., 2007, 692(5), 991-996. http://dx.doi.org/10.1016/j.jorganchem.2006.10.059
- [151] Tabbi, G.; Cassino, C.; Cavigiolio, G.; Colangelo, D.; Ghiglia, A.; Viano, I.; Osella, D. Water stability and cytotoxic activity relationship of a series of ferrocenium derivatives. ESR insights on the radical production during the degradation process. J. Med. Chem., 2002, 45(26), 5786-5796.
  - http://dx.doi.org/10.1021/jm021003k PMID: 12477361
- [152] Hillard, E.; Vessières, A.; Thouin, L.; Jaouen, G.; Amatore, C. Ferrocene-mediated proton-coupled electron transfer in a series of ferrocifen-type breast-cancer drug candidates. *Angew. Chem. Int. Ed.*, **2005**, *45*(2), 285-290. http://dx.doi.org/10.1002/anie.200502925 PMID: 16312004
- [153] Hamels, D.; Dansette, P.M.; Hillard, E.A.; Top, S.; Vessières, A.; Herson, P.; Jaouen, G.; Mansuy, D. Ferrocenyl quinone methides as strong antiproliferative agents: Formation by metabolic and chemical oxidation of ferrocenyl phenols. *Angew. Chem. Int. Ed. Engl.*, **2009**, 48(48), 9124-9126.
  - http://dx.doi.org/10.1002/anie.200903768 PMID: 19876986
- [154] van Staveren, D.R.; Metzler-Nolte, N. Bioorganometallic chemistry of ferrocene. *Chem. Rev.*, 2004, 104(12), 5931-5985.

http://dx.doi.org/10.1021/cr0101510 PMID: 15584693

- [155] Köpf-Maier, P.; Köpf, H.; Neuse, E.W. Ferrocenium salts the first antineoplastic iron compounds. *Angew. Chem. Int. Ed. Engl.*, **1984**, *23*(6), 456-457. http://dx.doi.org/10.1002/anie.198404561
- [156] Sun, M-L.; Ruan, B-F.; Zhang, Q.; Liu, Z-D.; Li, S-L.; Wu, J-Y.; Jin, B-K.; Yang, J-X.; Zhang, S-Y.; Tian, Y-P. Synthesis, crystal structures, electrochemical studies and antitumor activities of three polynuclear organotin(IV) carbox-

ylates containing ferrocenyl moiety. *J. Organomet. Chem.*, **2011**, *696*(20), 3180-3185.

http://dx.doi.org/10.1016/j.jorganchem.2011.06.045
[157] Huang, X-F.; Tang, J-F.; Ji, J-L.; Wang, X-L.; Ruan, B-F. Synthesis, characterization and antitumor activity of novel amide derivatives containing ferrocenyl pyrazol-moiety. J. Organomet. Chem., 2012, 706-707, 113-123.

http://dx.doi.org/10.1016/j.jorganchem.2012.02.001

[158] Huang, X-F.; Wang, L-Z.; Tang, L.; Lu, Y-X.; Wang, F.; Song, G-Q.; Ruan, B-F. Synthesis, characterization and antitumor activity of novel ferrocene derivatives containing pyrazolyl-moiety. *J. Organomet. Chem.*, **2014**, *749*, 157-162.

http://dx.doi.org/10.1016/j.jorganchem.2013.08.043

- [159] Viegas-Junior, C.; Danuello, A.; da Silva Bolzani, V.; Barreiro, E.J.; Fraga, C.A.M. Molecular hybridization: A useful tool in the design of new drug prototypes. *Curr. Med. Chem.*, 2007, 14(17), 1829-1852. http://dx.doi.org/10.2174/092986707781058805 PMID: 17627520
- [160] Gediya, L.K.; Njar, V.C. Promise and challenges in drug discovery and development of hybrid anticancer drugs. *Expert Opin. Drug Discov.*, 2009, 4(11), 1099-1111. http://dx.doi.org/10.1517/17460440903341705 PMID: 23480431
- Shah, K.; Chhabra, S.; Shirvastava, S.K.; Mishra, P. Benzimidazole: A promising pharmacophore. *Med. Chem. Res.*, 2013, 22, 5077-5104. http://dx.doi.org/10.1007/s00044-013-0476-9
- [162] Sarhan, A.A.O.; Al-Dhfyan, A.; Al-Mozaini, M.A.; Adra, C.N.; Aboul-Fadl, T. Cell cycle disruption and apoptotic activity of 3-aminothiazolo[3,2-*a*]benzimidazole-2-carbonitrile and its homologues. *Eur. J. Med. Chem.*, **2010**, *45*(6), 2689-2694. http://dx.doi.org/10.1016/j.ejmech.2010.02.025 PMID:

http://dx.doi.org/10.1016/j.ejmech.2010.02.025 PMID: 20226574

- [163] El Rashedy, A.A.; Aboul-Enein, H.Y. Benzimidazole derivatives as potential anticancer agents. *Mini Rev. Med. Chem.*, 2013, 13(3), 399-407. http://dx.doi.org/10.2174/138955713804999847 PMID: 23190032
- [164] Husain, A.; Rashid, M.; Shaharyar, M.; Siddiqui, A.A.; Mishra, R. Benzimidazole clubbed with triazolothiadiazoles and triazolo-thiadiazines: New anticancer agents. *Eur. J. Med. Chem.*, 2013, 62, 785-798. http://dx.doi.org/10.1016/j.ejmech.2012.07.011 PMID: 23333063
- [165] Paul, K.; Bindal, S.; Luxami, V. Synthesis of new conjugated coumarin-benzimidazole hybrids and their anticancer activity. *Bioorg. Med. Chem. Lett.*, **2013**, 23(12), 3667-3672. http://dx.doi.org/10.1016/j.bmcl.2012.12.071 PMID: 23642480
- [166] Reddy, T.S.; Kulhari, H.; Reddy, V.G.; Bansal, V.; Kamal, A.; Shukla, R. Design, synthesis and biological evaluation of 1,3-diphenyl-1*H*-pyrazole derivatives containing benzimidazole skeleton as potential anticancer and apoptosis inducing agents. *Eur. J. Med. Chem.*, **2015**, *101*, 790-805. http://dx.doi.org/10.1016/j.ejmech.2015.07.031 PMID: 26231080
- [167] Cotter, T.G. Apoptosis and cancer: The genesis of a research field. *Nat. Rev. Cancer*, 2009, 9(7), 501-507. http://dx.doi.org/10.1038/nrc2663 PMID: 19550425
- [168] Aggarwal, R.; Kumar, R.; Kumar, S.; Garg, G.; Mahajan, R.; Sharma, J. Synthesis and antibacterial activity of some 5-hydroxy-5-trifluoromethyl-4,5-dihydropyrazol-1-thiocarboxamides, 3-trifluoromethylpyrazol-1-thiocarboxamides

and 4-aryl-2-(5(3)-trifluoromethyl-1-pyrazolyl)thiazoles. *J. Fluor. Chem.*, **2011**, *132*(11), 965-972. http://dx.doi.org/10.1016/j.jfluchem.2011.07.029

- [169] Usachev, B.I.; Obydennov, D.L.; Röschenthaler, G-V.; Sosnovskikh, V.Y. 2-Cyano-6-(trifluoromethyl)-4H-pyran-4-one: A novel versatile CF<sub>3</sub>-containing building block. J. Fluor. Chem., 2012, 137, 22-26. http://dx.doi.org/10.1016/j.jfluchem.2012.01.006
- [170] Aggarwal, R.; Bansal, A.; Mittal, A. Synthesis and antimicrobial activity of 3-(2-thienyl)-4-arylazo-5-hydroxy-5trifluoromethyl-Δ<sup>2</sup>-isoxazolines and 3-(2-thienyl)-4-arylazo-5-trifluoromethylisoxazoles. J. Fluor. Chem., 2013, 145, 95-101.

http://dx.doi.org/10.1016/j.jfluchem.2012.10.005

- [171] Fayed, E.A.; Eissa, S.I.; Bayoumi, A.H.; Gohar, N.A.; Mehany, A.B.M.; Ammar, Y.A. Design, synthesis, cytotoxicity and molecular modeling studies of some novel fluorinated pyrazole-based heterocycles as anticancer and apoptosis-inducing agents. *Mol. Divers.*, 2019, 23(1), 165-181. http://dx.doi.org/10.1007/s11030-018-9865-9 PMID: 30099687
- [172] Ashburn, T.T.; Thor, K.B. Drug repositioning: Identifying and developing new uses for existing drugs. *Nat. Rev. Drug Discov.*, 2004, 3(8), 673-683.

http://dx.doi.org/10.1038/nrd1468 PMID: 15286734

- [173] Lu, W.; Li, P.; Shan, Y.; Su, P.; Wang, J.; Shi, Y.; Zhang, J. Discovery of biphenyl-based VEGFR-2 inhibitors. Part 3: Design, synthesis and 3D-QSAR studies. *Bioorg. Med. Chem.*, 2015, 23(5), 1044-1054. http://dx.doi.org/10.1016/j.bmc.2015.01.006 PMID: 25637123
- [174] Kamal, A.; Faazil, S.; Ramaiah, M.J.; Ashraf, M.; Balakrishna, M.; Pushpavalli, S.N.C.V.L.; Patel, N.; Pal-Bhadra, M. Synthesis and study of benzothiazole conjugates in the control of cell proliferation by modulating Ras/MEK/ERKdependent pathway in MCF-7 cells. *Bioorg. Med. Chem. Lett.*, **2013**, *23*(20), 5733-5739. http://dx.doi.org/10.1016/j.bmcl.2013.07.068 PMID: 23999041
- [175] Wang, M.; Xu, S.; Lei, H.; Wang, C.; Xiao, Z.; Jia, S.; Zhi, J.; Zheng, P.; Zhu, W. Design, synthesis and antitumor activity of Novel Sorafenib derivatives bearing pyrazole scaffold. *Bioorg. Med. Chem.*, 2017, 25(20), 5754-5763. http://dx.doi.org/10.1016/j.bmc.2017.09.003 PMID: 28927801