Perspectives

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Mitochondrial Damage in Sepsis

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Mitochondria not only generate adenosine triphosphate (ATP) and act as the powerhouse of the cell but also contribute to host defense by producing reactive oxygen species. Therefore, mitochondrial damage in sepsis directly results in a shortage of energy currency and dysregulation of the immune system. Other than those, mitochondrial damage results in the release of highly dangerous mitochondrial DNA, facilitating acidosis by modulating the metabolism and inducing programmed cell death, thereby facilitating disease progression in sepsis. Various forms of cell death are induced by mitochondrial damage. Aponecrosis is a secondary conversion from apoptosis to necrosis. Although apoptosis is initially intended, it cannot be completed due to ATP depletion from mitochondrial damage, ultimately leading to inflammatory necrosis. Besides such accidental cell death, programmed inflammation-inducing cell deaths such as necroptosis, ferroptosis, and pyroptosis are induced by mitochondrial damage in sepsis. Based on these findings, the regulation of mitochondrial damage holds promise for the development of new therapeutic approaches for sepsis.

Key words: sepsis, mitochondria, programmed cell death, organ dysfunction, oxidative stress

Introduction

Mitochondria are essential components of living cells that generate energy currency, control metabolic pathways, regulate intracellular calcium levels, produce reactive oxygen species, and modulate programmed cell death¹⁾. Mitochondrial function is readily compromised in sepsis and significantly contributes to the worsening of the disease condition. In this perspective, we briefly overview recent advances in mitochondrial research related to sepsis.

Impaired cellular respiration

Mitochondria is responsible for producing adenosine triphosphate (ATP) via oxidative phosphorylation and functioning as the powerhouse of the cell (Figure 1). Damage to mitochondria impairs ATP production, leading to energy deficits in cells, particularly in high-energy-demanding organs such as the heart, kidneys, and liver. The count of mitochondria is considerably different between the cell types, and the mean volume for mitochondria from the rat was reportedly 0 60 μ^3 in the heart, 0.42 μ^3 in the liver, and 0.23 μ^3 in the kidney cortex²). Disrupted mitochondrial function is considered the fundamental mechanism of organ dysfunction, such as myocardial dysfunction and acute kidney injury in sepsis, and avoiding the decrease in the number and impaired function of mitochondria is crucial for maintaining cellular and organ functions^{3,4}).

Increase in oxidative stress

Reactive oxygen species (ROS) is a physiological byproduct of metabolism and is essential to the functions of immune systems. However, damaged mitochondria are known to produce excessive ROS,

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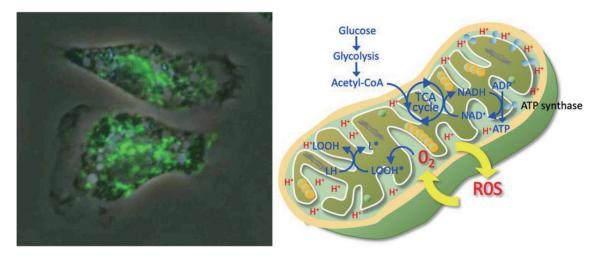


Figure 1 Immunofluorescent staining of mitochondria of the leukocytes and their functions Immunofluorescence of the leukocytes from rats displaying a mitochondrial fluorescein isothiocyanate (FITC) staining pattern of cellular distribution (left). Mitochondria are most densely distributed around the nucleus. The function of mitochondria is summarized in the right panel.

and overproduced ROS under pathophysiological conditions like sepsis are involved in the progression of the diseases. ROS can harm cellular components, including organelles, lipids, proteins, and DNA. Subsequent ROS-induced impaired cellular functions and inflammatory responses may result in organ damage. A complex system of interacting physiological antioxidant defenses normally diminishes oxidative stress and prevents damage to the mitochondria⁵⁾. However, since oxidative stressmediated injury increases in sepsis, overwhelming the antioxidant defenses and facilitating the development of cellular damage and organ failure, antioxidant supplementation has been considered a potentially beneficial treatment for sepsis. The effects of multiple agents, including vitamins E and C. melatonin, N-acetylcysteine, selenium, carnosine, fish oil, and glutathione, have been examined⁶. Nevertheless, the efficacy of these substances has not yet been established.

Release of mitochondrial DNA

Mitochondrial damage can trigger the release of mitochondrial DNA (mtDNA) and other mitochondrial components into the cytoplasm and extracellular space, which act as a damage-associated molecular pattern (DAMP), inducing systemic toxicity and damage to multiple organs⁷). Since mitochondria are suggested to originate from gram-negative bacteria and coexist in the cell (endosymbiotic theory), this endogenous 'enemy' acts as a patho-

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gen-associated molecular pattern (PAMP) to induce hyper-proinflammatory reactions⁸⁾. It was reported that plasma levels of mtDNA are associated with the clinical outcome of septic shock and were suggested to be a better biomarker than lactate in predicting mortality in severe sepsis⁹⁾. Recently, mitochondrial dysfunction and the release of mtDNA have attracted attention due to their relevance to skeletal muscle weakness, known as ICU-acquired weakness¹⁰⁾.

Metabolic dysfunction

Mitochondrial dysfunction disrupts cellular metabolism, leading to metabolic imbalances. First, glycolysis is intensified in carbohydrate metabolism; the failure to enter pyruvate into the tricarboxylic acid (TCA) cycle increases lactate generation. Second, mitochondrial dysfunction also affects lipid metabolism, and lipolysis is upregulated, increasing fatty acids and triglycerides. Impaired fatty acid oxidation and shifting towards anaerobic glycolysis result in further accumulation of lactate and metabolic acidosis. Third, changes in lipid metabolism also affect the levels of ketone bodies and amino acids¹¹. Other than the above, mitochondrial damage also affects calcium metabolism. Since mitochondria store a large amount of calcium, mitochondrial damage affects cellular signaling pathways, leading to increased cytosolic calcium levels. Elevated cytosolic calcium can activate various calcium-dependent enzymes, inducing cell death pathways and

leading to reduced ATP production and further cellular stress¹²⁾. These metabolic changes are an important area of research for developing novel therapies for sepsis.

Regulation of programmed cell death

Mitochondria can pursue programmed apoptosis, which can convert secondary to necrosis (aponecrosis), a form of uncontrolled cell death that further promotes inflammation and tissue damage. The mechanisms of apoptosis are explained as follows: first, in response to pro-apoptotic signals, mitochondria release cytochrome c from the intermembrane space into the cytosol. Cytochrome c binds to apoptotic protease activating factor-1 in the cytosol, leading to the formation of the apoptosome¹³⁾. This complex then recruits and activates procaspase-9, which initiates a cascade of downstream caspase activations, including caspase-3, -6, and -7. These executioner caspases dismantle the cell by cleaving various structural and regulatory proteins. Apoptosis is a silent form of cell death that does not impair organ function. However, since the apoptotic process requires sufficient ATP, a lack of ATP production by the mitochondrial damage causes apoptosis to transition into necrosis¹⁴⁾. In other words, apoptosis and necrosis partially share the same pathway, and apoptosis relies on adequate intracellular ATP levels. Meanwhile, when ATP is depleted, the cell death mechanism shifts to necrosis. Ultimately, mitochondrial damage in sepsis leads to an increase in necrotic cell death.

Mitochondrial damage can contribute to the initiation of necroptosis, a form of regulated inflammatory cell death that shares features with both apoptosis and necrosis. Increased production of ROS, the release of DAMPs, such as mtDNA and other mitochondrial components, and receptor-interacting protein kinases (RIPK) 1 and RIPK3 are involved in the induction of necroptosis¹⁵. Necroptotic cells release DAMPs, triggering a robust inflammatory response similar to necrosis.

Mitochondrial damage also can contribute to the induction of ferroptosis, a form of regulated cell death distinct from apoptosis, necrosis, and necroptosis. Ferroptosis is characterized by iron-dependent lipid peroxidation. ROS produced by the damaged mitochondria promotes lipid peroxidation, a key feature of ferroptosis. Mitochondria play a crucial role in iron metabolism, including the storage and utilization of iron. Mitochondrial damage can disrupt iron homeostasis, leading to increased free iron within the cell. This free iron can catalyze the formation of highly reactive hydroxyl radicals via the Fenton reaction and induce ferroptosis¹⁶⁾.

Mitochondrial damage contributes to the induction of pyroptosis, which is characterized by the activation of inflammatory caspases. This leads to the cleavage of gasdermin D (GSDMD) and the subsequent formation of pores in the cell membrane. Increased ROS and DAMPs activate the NLRP3 (NACHT, LRR, and PYD domains-containing protein 3) inflammasome, which plays a critical role in the initiation of pyroptosis. mtDNA is recognized by NLRP3, leading to the activation of the inflammasome and subsequent pyroptosis¹⁷⁾.

Organ dysfunction

The combined effects of energy deficits, increased oxidative stress, upregulated inflammation, and cell death induction contribute to the dysfunction of vital organs such as the heart, kidneys, lungs, and liver, a hallmark of severe sepsis and septic shock. Therefore, protecting and maintaining mitochondrial function is a promising strategy to improve organ damage in sepsis.

Conclusion

In these two decades, the effects of impaired cellular function due to mitochondrial dysfunction on the development of organ dysfunction in sepsis have been unveiled. Impaired cellular respiration, increase in oxidative stress, release of mitochondrial DNA, metabolic dysfunction, and altered cell death mechanisms are the fundamental mechanisms of mitochondrial damage-induced cellular injury. Mitochondrial damage should be paid more attention to as a critical factor in the progression of sepsis. New research targeting the maintenance of mitochondrial function is warranted to pave the way for improved sepsis management.

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Author contributions

RF and TI wrote and reviewed the manuscript. Both authors read and approved the final manuscript.

Conflicts of interest statement

The authors declare that they have no conflict of interest.

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